

Long-term study of behaviors of two cohabiting sea urchin species, *Mesocentrotus nudus* and *Strongylocentrotus intermedius*, under conditions of high food quantity and predation risk *in situ*

Peter M. Zhadan ^{Corresp., Equal first author, 1}, Marina A. Vaschenko ^{Corresp., Equal first author, 2}

¹ Department of Geochemistry and Ecology of the Ocean, V. I. Il'ichev Pacific Oceanological Institute FEB RAS, Vladivostok, Russia

² Laboratory of Physiology, National Scientific Center of Marine Biology FEB RAS, Vladivostok, Russia

Corresponding Authors: Peter M. Zhadan, Marina A. Vaschenko
Email address: pzhadan@poi.dvo.ru, mvaschenko@mail.ru

Background. In the predator–sea urchin–macrophyte trophic cascade, the ecological effect of sea urchins as grazers depends both on their density and the changes in foraging activity, which are influenced by various disturbing factors. However, the complete duration of the alarm reactions of echinoids has not been studied until now. Here, we tested a hypothesis that two cohabiting sea urchins, *Mesocentrotus nudus* and *Strongylocentrotus intermedius*, which differ morphologically, might display different behavioral responses to high hydrodynamic activity and predation. **Methods.** We used continuous time-lapse video recording to clarify behavioral patterns of *M. nudus* and *S. intermedius* in presence of a large quantity of food (the kelp *Saccharina japonica*) but under different weather conditions and different types of predation threat: (1) calm weather conditions, (2) stormy weather conditions, (3) predation risk associated with the presence of several sea star species, and (4) predation risk associated with an alarm stimulus (crushed conspecifics or heterospecifics). Three separate video recording experiments (134 days in total) were conducted under field conditions. Video recording analysis was performed to determine the number of specimens of each sea urchin species in the cameras' field of view, size of sea urchins' groups, movement patterns and the duration of the alarm responses of both sea urchin species. **Results.** We showed that in the presence of kelp, *M. nudus* and *S. intermedius* exhibited both similar and different behavioral responses to hydrodynamics and predation threat. Under calm weather, movement patterns of both echinoids were similar but *M. nudus* exhibited the higher locomotion speed and distance travelled. Furthermore, *S. intermedius* but not *M. nudus* tended to group near the food substrate. The stormy weather caused a sharp decrease in movement activity followed by escape response in both echinoids. Six starfish species failed to predate on healthy sea urchins of either species, and only a few attacks on ailing

S. intermedius specimens were successful. The alarm response of *S. intermedius* lasted approximately 90 h and 20 h for starfish attacks on ailing conspecifics and for simulated attacks (crushed conspecifics or heterospecifics), respectively, and involved several phases: (1) flight response, (2) grouping close to the food, (3) leaving the food, and (4) return to the food. Phase 3 was the more pronounced in a case of starfish attack. *M. nudus* only responded to crushed conspecifics and exhibited no grouping behavior but displayed fast escape (during 4 h) and prolonged (up to 19 days) avoidance of the food source. This outcome is the longest alarm response reported for sea urchins. **Discussion.** The most interesting finding is that two cohabiting sea urchin species, *M. nudus* and *S. intermedius*, display different alarm responses to predation threat. Both alarm responses are interpreted as defensive adaptations against visual predators.

1 **Long-term study of behaviors of two cohabiting sea urchin species, *Mesocentrotus nudus***
2 **and *Strongylocentrotus intermedius*, under conditions of high food quantity and predation**
3 **risk *in situ***

4
5 Peter M. Zhadan¹ and Marina A. Vaschenko²

6 ¹Department of Geochemistry and Ecology of the Ocean, V. I. Il'ichev Pacific Oceanological
7 Institute FEB RAS, Vladivostok, Russia

8 ²Laboratory of Physiology, National Scientific Center of Marine Biology FEB RAS,
9 Vladivostok, Russia

10 Corresponding Authors: Peter M. Zhadan, Marina A. Vaschenko
11 Email address: pzhadan@poi.dvo.ru, mvaschenko@mail.ru

12

13 **ABSTRACT**

14 **Background.** In the predator–sea urchin–macrophyte trophic cascade, the ecological effect of
15 sea urchins as grazers depends both on their density and the changes in foraging activity, which
16 are influenced by various disturbing factors. However, the complete duration of the alarm
17 reactions of echinoids has not been studied until now. Here, we tested a hypothesis that two
18 cohabiting sea urchins, *Mesocentrotus nudus* and *Strongylocentrotus intermedius*, which differ
19 morphologically, might display different behavioral responses to high hydrodynamic activity and
20 predation. **Methods.** We used continuous time-lapse video recording to clarify behavioral
21 patterns of *M. nudus* and *S. intermedius* in presence of a large quantity of food (the kelp
22 *Saccharina japonica*) but under different weather conditions and different types of predation
23 threat: (1) calm weather conditions, (2) stormy weather conditions, (3) predation risk associated
24 with the presence of several sea star species, and (4) predation risk associated with an alarm
25 stimulus (crushed conspecifics or heterospecifics). Three separate video recording experiments
26 (134 days in total) were conducted under field conditions. Video recording analysis was
27 performed to determine the number of specimens of each sea urchin species in the cameras' field
28 of view, size of sea urchins' groups, movement patterns and the duration of the alarm responses
29 of both sea urchin species. **Results.** We showed that in the presence of kelp, *M. nudus* and *S.*

30 *intermedius* exhibited both similar and different behavioral responses to hydrodynamics and
31 predation threat. Under calm weather, movement patterns of both echinoids were similar but *M.*
32 *nudus* exhibited the higher locomotion speed and distance travelled. Furthermore, *S. intermedius*
33 but not *M. nudus* tended to group near the food substrate. The stormy weather caused a sharp
34 decrease in movement activity followed by escape response in both echinoids. Six starfish
35 species failed to predate on healthy sea urchins of either species, and only a few attacks on ailing
36 *S. intermedius* specimens were successful. The alarm response of *S. intermedius* lasted
37 approximately 90 h and 20 h for starfish attacks on ailing conspecifics and for simulated attacks
38 (crushed conspecifics or heterospecifics), respectively, and involved several phases: (1) flight
39 response, (2) grouping close to the food, (3) leaving the food, and (4) return to the food. Phase 3
40 was the more pronounced in a case of starfish attack. *M. nudus* only responded to crushed
41 conspecifics and exhibited no grouping behavior but displayed fast escape (during 4 h) and
42 prolonged (up to 19 days) avoidance of the food source. This outcome is the longest alarm
43 response reported for sea urchins. **Discussion.** The most interesting finding is that two cohabiting
44 sea urchin species, *M. nudus* and *S. intermedius*, display different alarm responses to predation
45 threat. Both alarm responses are interpreted as defensive adaptations against visual predators.

46

47 **Subjects** Animal behavior, Ecology, Marine Biology, Zoology

48 **Keywords** Predator–prey interactions, Sea urchin behavior, Chemical alarm cues, Alarm
49 response duration, Grouping behavior, Escape response, Defensive adaptation

50

51 INTRODUCTION

52 The need to forage and the need to avoid predation are considered as the most important
53 evolutionary forces in the selection of morphological and behavioral characteristics of animals
54 (*Lima, 1998; Lima & Dill, 1990*). Sea urchins (class Echinoidea), which have a long evolutionary
55 history and a worldwide distribution, have developed a number of defense mechanisms against
56 predators promoting survival and reproduction. Possessing a locomotion speed that is
57 significantly inferior to that of many potential predators, sea urchins have a hard, internal
58 calcium carbonate skeleton (test) covered by spines. Some species are additionally protected by
59 pedicellaria with poison glands (*Jensen, 1966; Cannone, 1970*). Sea urchins exhibit mostly
60 nocturnal activity for both the movement/migration (*Crook et al., 2000; Dance, 1987;*

61 *Hasegawa, 2014; Hereu et al., 2005*) and spawning (*Zhadan et al., 2018*), appearing to be an
62 adaptation for enemy avoidance (*Fricke, 1973*).

63 Sea urchins lack image-forming eyes (*Ullrich-Lüter et al., 2011*); therefore, chemical
64 senses are an essential source of information on predation risk. The emergence of chemical cues
65 (substances emitted by predators and/or physically damaged con- or heterospecific prey) can
66 cause various behavioral reactions in sea urchins: ‘cryptic behavior’, i.e. hiding in a crack or hole
67 (*Fricke, 1973; Kintzing & Butler, 2014a; Spyksma et al., 2017*); ‘associative behavior’, i.e.
68 aggregation into dense groups (*Bernstein et al., 1983; Hagen et al., 2002; Kintzing & Butler,*
69 *2014a*), and ‘dispersion behavior’, i.e. escape (*Hagen et al., 2002; Urriago et al., 2011; Vadas &*
70 *Elnor, 2003*). For some species, it has been shown that the emergence of a chemical signal from
71 predator presence was associated with a decrease in the intensity of feeding (the so-called ‘fear
72 effect’) (*Freeman, 2006; Kintzing & Butler, 2014b; Matassa, 2010; Spyksma et al., 2017*). The
73 changes in behavior, including a decrease in foraging activity, were more pronounced in young
74 individuals (*Clemente et al., 2013; Freeman, 2006*), and these behaviors may not occur in large
75 well-armed adults (*Parker & Shulman, 1986; Wirtz & Duarte, 2012*).

76 Being consumers of macrophytes, sea urchins severely affect the structure of coastal
77 benthic communities (see for review *Estes & Duggins, 1995; Mann, 1982; Steneck, 2013*). There
78 is growing evidence that in the predator–prey–plant trophic cascade, the ecological effect can be
79 not only prey density-mediated (i.e., associated with the direct influence of predators on prey
80 number) but also prey behavior-mediated (i.e., associated with inhibition of prey foraging
81 activity by predators) (*Abrams, 1995; Dill et al., 2003; Pearson, 2010; Schmitz et al., 2004;*
82 *Trussell et al., 2006; Werner & Peacor, 2003*), prompting the suggestion that the grazing effect
83 of sea urchins on macrophytes’ abundance may depend on the duration of their alarm response.
84 However, the complete duration of the alarm responses of sea urchin species to waterborne
85 chemical signals emitted either from other prey (conspecific or non-conspecific), the predator
86 itself, or both has remained unknown until now. Most studies of sea urchins’ responses to
87 chemical alarm signals have been conducted under laboratory conditions (*Chivers & Smith,*
88 *1998; Hagen et al., 2002; Kintzing & Butler, 2014a, 2014b; Manzur & Navarrete, 2011;*
89 *Matassa, 2010; Spyksma et al., 2017*), and field experiments conducted under conditions of calm
90 water and under unidirectional water flows (*Parker & Shulman, 1986; Snyder & Snyder, 1970;*
91 *Vadas & Elnor, 2003; Wirtz & Duarte, 2012*; but see *Manzur & Navarrete, 2011; Urriago et al.,*

92 2011). All these studies have focused on the initial stage of sea urchins' alarm response, whereas
93 the entire sequence of behavioral events in sea urchins in their natural habitat, starting from the
94 onset of the alarm reaction to its complete extinction, has not been investigated.

95 The present study was undertaken to identify the temporal and spatial parameters of the
96 complete alarm reactions in two species of sea urchins, *Strongylocentrotus intermedius* (A.
97 Agassiz, 1864) and *Mesocentrotus nudus* (A. Agassiz, 1864) (= *Strongylocentrotus nudus*) under
98 conditions of food abundance using continuous time-lapse video recording (during 1.5–2 mo) of
99 animal behavior in the field. These echinoids are abundant in coastal environments of the
100 northwestern Pacific; however, their ranges overlap only partially, in the southern part of the Sea
101 of Japan washing the coasts of Korea, Japan and the Primorye region of Russia, where the lower
102 and upper geographical boundaries of *S. intermedius* and *M. nudus* occur, respectively (see for
103 review [Agatsuma, 2013a, 2013b](#); [Kafanov & Pavlyuchkov, 2001](#)). In the Sea of Japan, these sea
104 urchins are common species in benthic communities inhabiting hard substrates at a depth of up to
105 25 m ([Kafanov & Pavlyuchkov, 2001](#)). Morphologically, *S. intermedius* and *M. nudus* differ from
106 each other in respect to the color (brownish-black in *M. nudus* and brownish-grey in *S.*
107 *intermedius*) and spine length and hardness (hard sharp spines of up to 30 mm in *M. nudus* and
108 less hard spines of up to 8 mm in *S. intermedius*). We hypothesized that these sea urchin species,
109 presumably sharing an evolutionary history but differing morphologically, might display
110 different behavioral responses to various disturbing factors such as high hydrodynamic activity
111 and predation.

112

113 MATERIALS AND METHODS

114 Study areas, sea urchins and video recording experimental setup

115 Field experiments were conducted in the northwestern Sea of Japan: Kievka Bay (42.830° N,
116 133.691° E) and Alekseev Bay (42.981° N, 131.730° E). Kievka Bay, with a width of 8.3 km and
117 a length of 3.3 km, is typical of the southeastern coast of the Primorye region of Russia. Being
118 open to prevailing summer winds from the southeast to the southwest, the bay is characterized by
119 high wave activity. The studies here were performed at a depth of 6 m on relatively flat bedrock
120 surrounded by large stones. Alekseev Bay has a width of 0.8 km and a length of 1.3 km, and
121 wave activity here is high only under winds from the north, a phenomenon that is quite rare in
122 summer. The studies here were performed at a depth of 2 m on a flat bottom covered with

123 medium sized gravel. Within both bays, water depth is weakly influenced by tidal activity: an
124 irregular semidiurnal tide has a maximum amplitude of approximately 50 cm (*Zhadan et al.,*
125 *2018*).

126 In both bays, the density of *S. intermedius* and *M. nudus* in the areas adjacent to the
127 experimental installations was 1 to 2 ind. m⁻² (*Zhadan et al., 2018*). In addition, to balance the
128 density of the species in the study areas, approximately 300 specimens of *S. intermedius* were
129 collected in the bays in a radius of 100–200 m from the experimental installations and placed at a
130 distance of 10–20 m from them. The video observation of sea urchins' behaviors was performed
131 with TLC200 Pro (Brinno Incorporated) time-lapse video cameras mounted on steel stanchions
132 approximately 1 m above the bottom. Time-lapse videos were taken in 1 min intervals at a
133 resolution of 1280 × 720 pixels. The cameras were installed in such a way that the size of the
134 field of view was approximately 1.5 × 1.0 m. During the night, the cameras' fields of view were
135 illuminated by LED lamps (1 W) which were synchronized with the cameras by flash LED
136 indicator. The illumination duration was 1 s.

137 To attract sea urchins in the field of view of video cameras, flat mesh containers filled
138 with the kelp *Saccharina japonica* (Laminariales, Phaeophyta; hereafter simply laminaria) were
139 used. It is known that laminaria stimulates foraging activity of *M. nudus* and *S. intermedius*
140 (*Machiguchi, 1987; Machiguchi et al., 1994*). The containers, each composed of 2 steel frames
141 and mesh stretched on them, 1.1 × 0.75 × 0.01 m in size, were filled with laminaria and placed in
142 the cameras' field of view. Each container (hereafter feeder) contained approximately 30 kg of
143 the kelp. Sea urchins of both species themselves found the feeders and populated them within 2–
144 3 days. All sea urchins that were in the cameras' field of view on all the feeders were taken into
145 account for further video analysis. The steel frames of the containers were pressed down by
146 stones with a diameter of 20–40 cm to protect against wave activity.

147 The indicator of the onset of sea urchins' alarm response was the escape or redistribution
148 of sea urchins in the field of view of the cameras, and the indicator of the end of the alarm
149 reaction was the return of sea urchins to their original spatial distribution pattern. Such a design
150 of the experiments allowed us to clarify (1) the features of sea urchins' behavior in the presence
151 of abundant palatable food and under different weather conditions, (2) the frequency of predator
152 attacks under natural conditions, and (3) the temporal patterns of the complete behavioral
153 response of sea urchins to a natural predator attack and simulated predator attack (crushed

154 conspecifics or heterospecifics) in sea urchin species with different morphological
155 characteristics.

156

157 **Video recording analysis**

158 The recorded videos were viewed frame by frame. We counted the numbers of specimens of
159 each sea urchin species in the each camera field of view both in the absence and in the presence
160 of disturbing factors. To access the grouping behavior of the sea urchins, we calculated the mean
161 group size as the ratio of the total number of individuals in the cameras' field of view to the
162 number of associations (*Hagen & Mann, 1994*). Following *Vadas et al. (1986)*, we distinguished
163 between sea urchin associations and aggregations. Each group of sea urchins in two-dimensional
164 groupings, including individuals suspected of being in tactile contact (there was no visible space
165 between them) and single individuals, was considered a separate association.

166 To determine the mean group size for both sea urchin species, the video frames were
167 randomly selected over the periods of calm weather. Only the video frames showing not more
168 than 30 specimens (43 frames for each species) were chosen for the mean group size calculation
169 in order to avoid crowding effects when sea urchins might be in tactile contact due to their high
170 density on the feeders. Under such a limitation, no groups of 3 or more sea urchins in cohesive
171 three-dimensional groupings (aggregations, according to *Vadas et al., 1986*) were observed in
172 our study.

173 Sea urchins' movement was analyzed using the free software, 'Tracker', for video
174 analysis (www.opensourcephysics.org/items/detail.cfm?ID=7365). The cell size (2×2 cm) of
175 the feeder mesh was used as a scale. We tracked and measured sea urchin displacement with an
176 interval of 1 min. Following *Lauzon-Guay et al. (2006)*, we defined a step as the distance
177 between two successive positions of the sea urchin (1 min apart), a stop as an interval when sea
178 urchin remains stationary during at least 1 min (2 successive frames) and a move as the distance
179 between two successive stops which can be composed of one or more steps. The mean
180 locomotion speed was calculated as total distance passed divided by total time.

181

182 **Long-term video recording of sea urchins' behavior**

183 We conducted 3 separate video recordings in Kievka Bay (during 51 d in August–September
184 2014 and 37 d in August–October 2015) and in Alekseev Bay (during 46 d in July–September

185 2016). In these experiments, 4 cameras were used, and 4 feeders were arranged in pairs in such a
186 way that the direction of the tidal currents coincided with the direction of the long sides of the
187 feeders. The distances between the long sides of the feeders were 0.4 m, and the distance
188 between the pairs of the feeders was 2 m. The feeders were replaced every 15–20 days (see Fig.
189 1) when approximately 80% of laminaria was consumed by sea urchins. Sea urchins were
190 carefully transferred from the old feeders to the feeders with fresh laminaria. It took
191 approximately 5 min to change one feeder.

192 We tested for behavioral differences between *M. nudus* and *S. intermedius* in presence of
193 a large quantity of palatable food but under different weather conditions and different types of
194 predation threat: (1) under calm weather conditions, (2) under stormy weather conditions, (3)
195 under predation risk associated with the presence of several sea star species, and (4) under
196 predation risk associated with simulated predator attack (crushed individuals of *M. nudus*). The
197 numbers of specimens of each sea urchin species were counted per each 6 h of observations
198 throughout all 3 long-term experiments, both in the absence and in the presence of disturbing
199 factors (see [Table S1](#) for original data).

200 The stormy periods were determined by several signs which are clearly visible on video
201 recordings: (1) the oscillation of the feeder surface and (2) the increase in suspended particles
202 movement and water turbidity. In addition, an increase in wave height during storm was recorded
203 by the depth sensor of a multi-parameter RBRXR-620 datalogger (Sea and Land Technologies
204 Pte, Singapore) which measured the sea level every 10 min (see for example [Fig. S1](#)).

205 We compared movement parameters (the number and length of the steps and moves, the
206 entire displacement, the number and duration of the stops, the entire stationary period and mean
207 speed) of both sea urchin species under calm and stormy weather conditions. For this purpose,
208 the distances traversed by randomly selected 10 sea urchins of each species during 240 min
209 before the storm and in the beginning of the storm ([Fig. S1](#)) were tracked and measured with an
210 interval of 1 min (see [Tables S2](#) and [S3](#) for raw data). Further tracking showed that a half of
211 observable sea urchins left the cameras' field of view during approximately 10 h after the
212 beginning of measurements. Locomotion speed during escape was calculated as total distance
213 passed during the period from the beginning of active movement of the sea urchin to its
214 disappearance from the field of view divided by the total time of escape (see [Tables S2](#) and [S3](#)
215 for raw data).

216 Predation risk was associated with several species of sea stars that were present
217 simultaneously with sea urchins on the feeders or with simulation of predation attack (crushed
218 individuals of *M. nudus*). To determine behavioral responses of *S. intermedius* to starfish attacks
219 on the conspecifics which took place on September 5 and 21, 2015(see [Table S1](#)), we measured
220 the distances of 13 sea urchins closest to the site of the attack (i.e., their entire displacement)
221 during approximately 100 h after the beginning of the attack with the intervals from 2 h to 4–16
222 h. For these periods, the total numbers of *S. intermedius* specimens as well as the individuals of 2
223 starfish species, *P. pectinifera* and *L. fusca*, which were in contact with the prey, were counted.

224 Our pilot experiments showed that *M. nudus* rapidly left the area of the bottom where
225 conspecifics were crushed and avoided it for a long time. There was no similar reaction to
226 crushed *S. intermedius* specimens. Furthermore, *S. intermedius* exhibited no visible responses
227 both to crushed conspecifics and heterospecifics. Therefore, we focused on the study of the
228 phenomenon of *M. nudus* response to crushed conspecifics. We compared how quickly *M. nudus*
229 populated the feeders in the presence and in the absence of crushed conspecifics. In the first case,
230 10 *M. nudus* specimens with test diameters of 50–60 mm were crushed in the middle between the
231 pairs of the feeders when *M. nudus* were present on them (5 experiments in total; we did not
232 include the data of 1 experiment into statistical analysis because it might be influenced by the
233 experiment on storm imitation conducted 3 days earlier). In the second case, all *M. nudus*
234 specimens were removed from the cameras' fields of view and transferred to a distance of
235 approximately 10 m from the feeders (7 experiments in total; we did not include the data of 3
236 experiments into statistical analysis because they might be influenced by the previous or next
237 storm). In both cases, we estimated the temporal dynamics of the numbers of sea urchins (with
238 time intervals of 1 h and 6 h in the first and second cases, respectively). To test our assumption
239 that the storm contributes to the return of sea urchins *M. nudus* to the feeders, two experiments
240 mimicking stormy conditions were conducted several days (6 and 12 in 2014 and 2016,
241 respectively, see [Table S1](#)). After *M. nudus* left the feeders in response to crushed conspecifics,
242 the feeders were removed, and several scuba divers actively swam in the area of the experimental
243 installation. After that, the feeders with fresh laminaria were placed in their original place. These
244 procedures took 30–40 min. The average sea urchin numbers for 1 day before crushing and 2
245 days after storm imitation were determined.

246

247 **Short-term video recording of sea urchins' behavior**

248 A set of experiments, each with a duration of approximately 6 d, was conducted in Alekseev Bay
249 from July–September 2016. The experiments were designed to elucidate behavioral patterns of
250 *M. nudus* and *S. intermedius* in response to an alarm stimulus (crushed conspecifics or
251 heterospecifics). In each of these experiments, 2 video cameras and 2 feeders were used. Two
252 days after the feeder placement, when 30–40 sea urchins of both species were gathered on the
253 feeder surface, 5 specimens of *S. intermedius* or *M. nudus* were crushed at a distance of 20 cm
254 from the feeder's short side. Three to four days after simulation of the predation threat, all the
255 sea urchins were removed from the cameras' field of view, and the experiment was repeated on
256 the other feeder with fresh laminaria and with sea urchins not previously used. In total, 4
257 experiments of this kind were carried out with each of sea urchin species used as simulated prey.
258 During 22 h before and 55 h after the treatment, we estimated the numbers of sea urchins as well
259 as the temporal dynamics of the mean group size (with time intervals from 1 to 4 h) with one
260 exception: we failed to assess the temporal dynamics of the mean group size of sea urchins *M.*
261 *nudus* in response to crushed conspecifics due to fast escape of *M. nudus*.

262 To determine movement patterns of sea urchins in response to crushed con- and
263 heterospecifics, we measured step length and locomotion speed before and after treatment.
264 Additionally, we measured with intervals of 6–9 min the distances of 10 sea urchins closest to
265 the site where simulated attack was performed. The exception was a case with response of *M.*
266 *nudus* to crushed conspecifics because sea urchins rapidly left the cameras' field of view.
267 Duration of sea urchin tracking was from 200 to 1200 min due to the species specific response of
268 sea urchins to the alarm signal.

269

270 **Statistical analysis**

271 To analyze the species-specific and treatment-specific differences in *M. nudus* and *S. intermedius*
272 behavior, the data sets on the numbers of sea urchins, mean group sizes as well as sea urchins'
273 movement parameters were formed and tested for normal distribution (D'Agostino and Pearson
274 omnibus normality test, $P < 0.05$). Normally distributed data were further analyzed by parametric
275 tests (unpaired *t*-test, 1-way ANOVA). In a case of abnormally distributed data, non-parametric
276 tests were used (Mann-Whitney test, Kruskal–Wallis test followed by Dunn's multiple
277 comparisons). To analyze the temporal dynamics of the numbers of both sea urchins in the

278 experiments on the response to crushed conspecifics or heterospecifics as well as in the
279 experiments on the repopulation the feeders by sea urchins *M. nudus* after their removal, linear
280 or nonlinear regression (curve fit) were used. All statistical analyses were run using GraphPad
281 Prism v. 6.0. The details regarding raw data and statistics are presented in the Supplementary
282 Materials. In all the Figures, the data relating to *M. nudus* is in blue color and the data relating to
283 *S. intermedius* is in red color.

284

285 **Procedural controls**

286 In our *in situ* experiments we did not use the procedural controls which are usually applied in the
287 experiments with animals contained in tanks or cages, such as ‘food presence – food absence’,
288 ‘animal treatment/manipulation – imitation of animal treatment/manipulation’. First, both sea
289 urchin species exhibited clear food search behavior, and the absence of food strongly stimulated
290 them to migrate to the place where the food is present. Second, we minimized manipulations in
291 our experiments that could affect the results and conclusions. A negligible displacement of water
292 masses near a feeder associated with sea urchins’ crushing as well as the swimming of the diver,
293 who served the installation, above the feeder did not cause escape reactions of both sea urchin
294 species similar to those during storm. The experiments mimicking stormy conditions were
295 conducted in the absence of the feeders. Both in long-term and short-term experiments, we used
296 short-term periods (from 1 to 3 days) just before the treatments as proper procedural controls,
297 and compared quantitative parameters (sea urchins’ number and mean group size) obtained for
298 these periods with those obtained for the periods after the treatments (storms, starfish attacks or
299 crushed sea urchins).

300

301 **RESULTS**

302 **General characteristics of sea urchins’ behavior**

303 During the recording periods in 2014, 2015 and 2016 (134 days in total), 24 ± 25 (mean \pm SD) of
304 *M. nudus* specimens and 78 ± 42 of *S. intermedius* specimens were in the field of view of the
305 video cameras (Fig. 1A–C, Table S1). During the periods without any treatment (storms, *M.*
306 *nudus* specimens removal or crushing, starfish attacks), the numbers of *M. nudus* and *S.*
307 *intermedius* were higher, 50 ± 15 and 85 ± 42 , respectively. The size composition of the two
308 echinoid species was slightly different. Only large adults of *M. nudus* with a test diameter of 62.2

309 ± 7.5 mm (mean \pm SD) were present in the cameras' field of view, whereas among the adult *S.*
310 *intermedius* with test diameters from 37 to 74 mm (64.4 ± 4.3 mm), there was a small number
311 (up to 12%) of juveniles with test diameters of 10–15 mm. On the surface of the feeders, both sea
312 urchin species were relatively evenly distributed in one plane and did not form aggregates (three-
313 dimensional groups); however, they formed associations (dense two-dimensional groups).
314 Grouping behavior in *S. intermedius* was expressed to a greater extent than in *M. nudus*: when
315 from 14 to 25 of individuals were present on the surface of the feeders, the mean group size of *S.*
316 *intermedius* was approximately 2 times higher than that of *M. nudus* (2.27 ± 0.4 versus $1.12 \pm$
317 0.1 , Mann–Whitney test, $U = 0$, $P < 0.0001$; see [Table S4](#) for raw data and statistics).

318 Both sea urchin species displayed so called 'covering behavior' but it was more
319 pronounced in *S. intermedius* than in *M. nudus*. The debris covering sea urchin aboral surfaces
320 consisted mainly of pieces of the algae such as *Ulva fenestrata* Ruprecht, 1840 and *Desmarestia*
321 *viridis* (O.F. Müller) J.V. Lamouroux, 1813 in June–July, and seagrass *Zostera marina* L. in the
322 end of September. During these periods, from 87 to 100% of *S. intermedius* individuals were
323 decorated compared to from 0 to 18% for *M. nudus*.

324

325 **Behavioral responses of sea urchins to increased wave activity**

326 During the storms, the number of sea urchins of both species in the cameras' field of view
327 sharply decreased ([Fig. 1A–C](#), see [Table S1](#) for raw data). On the eve of the storm periods, there
328 were 54 ± 9 (mean \pm SD) of *M. nudus* specimens and 76 ± 37 of *S. intermedius* specimens
329 whereas during the storms, the average numbers for both species (26 ± 15 and 34 ± 21 for *M.*
330 *nudus* and *S. intermedius*, respectively) were significantly lower ([Fig. 2](#), see [Tables S5, S6](#) for
331 raw data and statistics). Approximately one day after the storm, sea urchins of both species
332 restored their numbers on the feeders ([Fig. 1A–C](#), [Fig. 2](#), [Tables S5, S6](#)).

333 Both sea urchin species exhibited similar patterns of movement under conditions of calm
334 weather. The average numbers of moves and stops were not significantly different between
335 species, however, the moves of *S. intermedius* were shorter and consisted of higher number of
336 shorter steps whereas the locomotion speed and entire distance travelled were significantly
337 higher in *M. nudus* ([Tables 1 and 2](#), see also [Table S7](#) for interspecies comparison).

338 Both sea urchin species responded to increased wave activity by a sharp decrease in the
339 number of steps, length of one move and entire distance travelled ([Tables 1 and 2](#), see also [Table](#)

340 S7 for interspecies comparison). At the same time, the duration of one stop increased
341 approximately 3 and 4 times in *S. intermedius* and *M. nudus*, respectively, and the average
342 proportions of time sea urchins spent stationary were 76 and 91%, respectively, against 33 and
343 54% under calm weather. The average locomotion speeds during storm conditions were as low
344 as 0.07 and 0.03 cm min⁻¹ in *S. intermedius* and *M. nudus*, respectively. During escape, sea urchin
345 speeds sharply increased and averaged 0.82 ± 0.19 (range of 0–7.02) cm min⁻¹ in *S. intermedius*
346 and 1.76 ± 0.30 (range of 0–11.81) cm min⁻¹ in *M. nudus*, these were, respectively, 4.3 and 5.5
347 times higher than those under calm weather (see [Tables S2](#) and [S3](#) for raw data).

348

349 **Behavioral response of *S. intermedius* to starfish attack**

350 Over 3 periods of our studies, only three cases of the attacks of the sea stars (*Patiria pectinifera*
351 (Muller & Troschel, 1842) and *Lethasterias fusca* Djakonov, 1931) on single individuals of *S.*
352 *intermedius* were recorded (5, 10 and 21 September, 2015, see [Table S1](#)) whereas no attacks of
353 predators on sea urchins *M. nudus* were observed. Judging by the presence of injuries and
354 abnormal behavior (low motor activity and body position with the oral surface upward), only
355 sick or damaged individuals of *S. intermedius* have been attacked by sea stars ([Fig. S2A](#)). It is
356 possible that the appearance of damaged *S. intermedius* specimens that have lost a significant
357 part of the spines was due to typhoon ‘Goni’, which occurred on August 27–29, 2015 in the
358 northwestern Sea of Japan.

359 On September 10, sea urchin reaction to a predator attack could not be traced due to low
360 visibility caused by the storm. The behaviors of sea urchins *S. intermedius* during starfish attacks
361 on September 5 and 21, 2015 were slightly different. On September 5, one *S. intermedius*
362 specimen was consumed by 1–5 individuals of *P. pectinifera* and 1 individual of *L. fusca* in the
363 centre of one of the 4 feeders for 45 h ([Fig. 3A](#), see also [Fig. S2](#)). Within 2 h after the beginning
364 of the attack, most of the sea urchins left the surface of the feeder and formed several groups on
365 the tops and at the base of nearby stones ([Fig. S2B](#)). The number of sea urchins in the cameras’
366 field of view was relatively stable during the first 12 h, and sea urchin distances from the site of
367 attack did not change much ([Fig. 3A](#)). Then, the distances began to increase sharply, and their
368 maximum coincided with maximum number of sea stars consuming an ailing specimen ([Fig.](#)
369 [3A](#)). 26 h after the beginning of starfish attack, the number of sea urchins in the cameras’ field of
370 view began to decrease, and after 50 h, there remained approximately 30% of sea urchins ([Figs.](#)

371 3A and S2C). The number of sea urchins on the feeder began to increase between 16 and 28
372 hafter the starfish left the sea urchin remains, and this coincided with gradual decrease of sea
373 urchin distances from the site of attack (Fig. 3A). In general, the alarm reaction of *S. intermedius*
374 from the onset of the starfish attack to restoration of the initial sea urchin population on the
375 surface of the feeder (Figs. 3A and S2D) lasted for approximately 90 h.

376 On September 21, a starfish attack occurred at the short edge of the feeder. Sea urchin
377 distances from the site of attack were almost unchanged during the first 9 h and then sharply
378 increased, and this coincided with maximum number of sea stars (*P. pectinifera* and *L. fusca*)
379 consuming an ailing specimen (Fig. 3B). After 24 h, no *S. intermedius* specimens remained
380 closer than 40 cm from the site of attack. They formed associations on the feeder and the nearest
381 stones. Eight hours after the beginning the attack, the number of sea urchins in the cameras' field
382 of view began to decrease, and after 55 h, there remained approximately 50% of sea urchins.
383 Consumption of the prey by the sea stars lasted 70 h. Restoration of sea urchin abundance and
384 distribution on the feeder began 10 h after the sea stars left the remains of the prey. The total
385 duration of the sea urchin alarm reaction was 88 h (Fig. 3B).

386 It should be noted that the sea stars *P. pectinifera* and *L. fusca* were constantly present on
387 the feeders. The starfish *Asterias amurensis* Lutken, 1871, *Distolasterias nipon* (Döderlein,
388 1902), *Lysastrosoma anthosticta* Fisher, 1992 and *Aphelasterias japonica* Bell, 1881 also often
389 appeared on the feeders. With a few exceptions, these starfish did not cause visible reactions in
390 healthy sea urchins. The behavior of *P. pectinifera* was the most aggressive. In one case, during
391 28 min, *P. pectinifera* attacked an *S. intermedius* specimen, which lost approximately 20% of its
392 spines, but finally, it was left alone. In two cases, sea stars *P. pectinifera* completely crawled on
393 *S. intermedius* individuals in such a way that starfish mouth was located directly above the sea
394 urchin's anal orifice. After 11 and 15 min in the first and second cases, respectively, the sea stars
395 left the potential prey, which indicates that a healthy sea urchin can effectively resist the
396 penetration of a starfish stomach through the anus.

397

398 **Behavioral responses of *M. nudus* and *S. intermedius* to crushed *M. nudus*** 399 **specimens**

400 Four experiments conducted during long-term recordings of 2014–2016 (Fig. 1A–C, Table S1)
401 showed that after the conspecifics were crushed near the feeders, approximately 90% of *M.*

402 *nudus* individuals left the cameras' field of view during 4 h (Fig. 4, see Table S8 for raw data
403 and statistics). During this period, a sharp increase in the average step length just after the
404 treatment was recorded (Fig. 5). There were 2 time intervals with the highest locomotion speed,
405 the first 55 min after the treatment when a half of *M. nudus* specimens left the cameras' field of
406 view, and the last 128 min when the rest of sea urchins escaped (Fig. S3A). These intervals were
407 interrupted by the relatively stable 1 hour period when sea urchins almost stopped moving (Fig.
408 5).

409 Nine hours after the beginning of the experiment, there were no *M. nudus* specimens on
410 the feeders (Fig. 4, Table S8), and then, during a much longer period (7–19 days), only single *M.*
411 *nudus* individuals appeared (Fig. 1A–C). The restoration of the initial sea urchin numbers on the
412 feeders occurred only after the next storm event.

413 The imitation of stormy weather also contributed to the return of *M. nudus* after sea
414 urchins left the feeders in response to presentation of crushed conspecifics. Sea urchins were
415 absent on the feeders during approximately 5 and 13 days in 2014 and 2016, respectively, and
416 came back within 2 days after intensive swimming and replacement of the feeders; however,
417 their number was lower than that before the experiments (Figs. 1A, C and S4, Table S1).

418 Four experiments conducted during long-term recordings of 2014–2016 (Fig. 1A–C,
419 Table S1) demonstrated that, in the absence of crushed conspecifics, sea urchins *M. nudus* came
420 back 2–3 days after their removal from the surfaces of the feeders (Fig. 6, see Table S9 for raw
421 data and statistics).

422 Sea urchins *S. intermedius* did not leave the cameras' field of view during the
423 experiments with presentation of crushed *M. nudus* specimens (Fig. 1A–C, Table S1); moreover,
424 statistical analysis revealed small but significant increase in *S. intermedius* numbers in 3 cases
425 when *M. nudus* was absent on the feeders (see Table S10 for statistics). Spatial pattern of *S.*
426 *intermedius* remained almost unchanged with the exception of one case: after the *M. nudus*
427 specimens were crushed, sea urchins *S. intermedius* avoided the surface of the feeder for 24 h
428 and were among the stones opposite of the crushed *M. nudus*.

429

430 **Behavioral responses of *M. nudus* and *S. intermedius* to crushed conspecifics**
431 **and heterospecifics**

432 In the 6-d short-term experiments, *M. nudus* exhibited a strong avoidance reaction in response to
433 crushed conspecifics similar to that in long-term observations (Fig. S5A, see Table S11 for raw
434 data and statistics). When *S. intermedius* individuals were crushed near the feeders, behavior of
435 sea urchins *M. nudus* remained unchanged, as evidenced by the absence of significant changes in
436 such indicators as the number of sea urchins in the cameras' field of view (Fig. S5B, see Table
437 S12 for raw data and statistics) and mean group size (Fig. S5C, see Table S13 for raw data and
438 statistics). Analysis of *M. nudus* movement activity also revealed no differences in the average
439 step length (Fig. 7A), distance from the site of simulated attack (Fig. 7B) and locomotion speed
440 (Fig. S3C).

441 In response to crushed conspecifics, *S. intermedius* moved towards the opposite side of
442 the feeder and formed associations there. Before the experiment, the mean group size was $2.25 \pm$
443 0.75 (mean \pm SD for 21 h of observation), and this parameter was 2.5-fold higher (5.85 ± 2.18)
444 during the 14 h period after the stimulus was presented (Fig. 8A, see Table S14 for raw data and
445 statistics). The changes in the number of sea urchins in the cameras' field of view were not very
446 obvious (Fig. 9A); however, linear regression showed significant decrease in this parameter ($P <$
447 0.0001 , see Table S15 for raw data and statistics). The associations were positioned in such a
448 way that sea urchins were partly on the feeder and partly outside it. On average, approximately
449 40% of the sea urchins, both grouped and alone, were located in the immediate vicinity outside
450 the feeders. Judging by the restoration of the original spatial distribution of sea urchins on the
451 feeder (the initial mean group size), the duration of the alarm reaction of *S. intermedius* was
452 approximately 18 h (Fig. 8A).

453 Analysis of *S. intermedius* movement revealed 2 time intervals of the highest activity,
454 each approximately 1.5 h in duration: the first when sea urchins formed associations and the
455 second when these associations dispersed (Figs. 10 and S3B, see Table S16 for raw data).
456 Between these 2 peaks of activity, there was prolonged period of approximately 16 h, when the
457 average step length and locomotion speed were relatively low, and the distance from the site of
458 simulated attack was almost unchanged.

459 Judging by the dynamics of the mean group size, response of *S. intermedius* to crushed
460 individuals of *M. nudus* was generally similar to the response to crushed conspecifics (Fig. 8B,
461 see Table S17 for raw data and statistics). Movement analysis showed that just after the
462 treatment, there was an increase in the average step length (Fig. 7A), distance from the site of

463 simulated attack (Fig. 7B) and locomotion speed (Fig. S3D). A vast majority of sea urchins did
464 not leave the cameras' field of view (Fig. 9B), and linear regression showed no significant
465 decrease in this parameter ($P = 0.0822$; see Table S18 for raw data and statistics). The original
466 spatial distribution of sea urchins on the feeder was restored within approximately 20 h after the
467 beginning of simulated attack (Fig. 8B).

468

469 DISCUSSION

470 The present work is the first study analyzing long-term, around-the-clock behavior of sea urchins
471 exposed under field conditions to both calm and stormy weather and to presentation of two
472 competing stimuli, food and predation threat, which are assumed to be key factors influencing
473 species survival. Due to the natural turbidity of sea water, video camera cannot register the initial
474 stages of the sea urchin alarm reaction, namely, extension of the tube feet and movement of
475 spines, which are manifested in the first seconds or minutes after sea urchin exposure to an alarm
476 signal (Morishita & Barreto, 2011; Urriago et al., 2011). At the same time, the method of
477 continuous time-lapse video recording allowed analysis of the long-term dynamics of the spatial
478 distributions of sea urchins under natural conditions when both stimuli were presented.
479 Previously, several authors applied video recording in the field to quantify sea urchins at the kelp
480 grazing front (Lauzon-Guay & Scheibling, 2007) and to determine sea urchin movement patterns
481 (Dumont et al., 2007; Lauzon-Guay et al., 2006); however, they used separate time-lapse video
482 sequences, each lasting several hours. We showed that two sea urchin species, different in
483 morphology and living under similar conditions, exhibited distinctly different strategies for
484 avoiding predation in terms of response duration and behavioral patterns.

485

486 Sea urchin response to hydrodynamics

487 In the absence of predation threat, *S. intermedius* and *M. nudus* also showed somewhat different
488 behavioral patterns. Under calm weather, *S. intermedius* much more often than *M. nudus*
489 exhibited the covering behavior and tended to group on the food substrate. Despite general
490 patterns of movement (intermittent locomotion, characterized by moves interspersed with
491 pauses) in both species, *M. nudus* spent more time stationary but moved at approximately 2 times
492 higher speed than *S. intermedius*, resulting in the higher distance traversed. However, both
493 species are highly mobile, and under conditions of calm sea and presence of food, *M. nudus* and

494 *S. intermedius* would be able to overcome on average 4.6 and 2.7 m per day, respectively
495 (calculated based on the average locomotion speeds). These average distances are similar to
496 those observed for the sea urchins *Strongylocentrotus droebachiensis* (0.4–1.72 m; [Dumont et](#)
497 [al., 2006](#)), *Toxopneustes roseus* (1.65 and 2.49 m; [James, 2000](#)), *Tripneustes ventricosus* (3.7
498 and 8.8 m; [Tertschnig, 1989](#)) and *Diadema antillarum* (3.7 m; [Tuya et al., 2004](#)).

499 The results of both laboratory and field experiments evidence that sea urchins are capable
500 to sense a change in hydrodynamic activity and react by changing the behavior. In our study,
501 both sea urchins responded to the stormy weather, firstly, by decreasing the movement activity
502 up to almost complete stop and secondly, by leaving the food. It is noteworthy that during escape
503 under the stormy conditions, sea urchins can move on average 4–5-fold faster than during
504 feeding under calm weather.

505 It is known that with increasing water velocity (higher than approximately 15 cm s⁻¹), sea
506 urchins decrease their displacement and cease feeding both under laboratory ([Kawamata, 1998](#);
507 [Frey & Gagnon, 2016](#); [Cohen-Rengifo et al., 2018](#); [Tamaki et al., 2018](#)) and field ([Lissner, 1980](#);
508 [Dance, 1987](#); [Siddon & Witman, 2003](#); [Dumont et al., 2006, 2007](#)) conditions. Escape behavior
509 was also observed in laboratory flume experiments: at flow velocity ≤ 30 cm s⁻¹, sea urchins
510 moved in a downstream direction whereas at 35–45 cm s⁻¹, individuals moved in an upstream
511 direction ([Morse & Hunt, 2013](#); [Cohen-Rengifo et al., 2018](#)). However, escape response of sea
512 urchins to wave-induced benthic water flow *in situ* has been poorly documented. [Dance \(1987\)](#)
513 observed that during a period of turbulence lasting several hours, movement of *P. lividus* was
514 significantly oriented to the deep water with lower hydrodynamic activity. [Lauzon-Guay &](#)
515 [Scheibling \(2007\)](#) found that *S. droebachiensis* density at the grazing front decreased when wave
516 action increased and suggested that the ‘whiplash effect’ of the swaying kelp prevented sea
517 urchins from climbing onto kelp plants. In our study, the kelp was packed into mesh containers
518 and formed a kind of soft substrate. We believe that sea urchins *M. nudus* and *S. intermedius*,
519 being able to sense an oscillation of the substratum and/or increasing water flow, reduce the
520 movement activity and then make a decision to stay close to the food or escape. We noted that in
521 the case of the weaker storm, only a part of sea urchins left the food source. Remaining *S.*
522 *intermedius* individuals gathered into groups at the base of the feeders whereas *M. nudus*
523 specimens were on the surface of the feeders. However, unfortunately, we did not measure a

524 velocity of water flow and, therefore, cannot correlate it with sea urchin movement activity.
525 Further studies are thus required to elucidate such a correlation.

526

527 **Sea urchin response to predation threat**

528 During this study, no successful starfish attacks on healthy sea urchins were observed for either
529 species, and only 3 cases of consuming of single ailing *S. intermedius* individuals by several
530 starfish specimens (*P. pectinifera* and *L. fusca*) were recorded. Taking into account the
531 laboratory experimental data that the starfish, *L. anthosticta* and *P. pectinifera* in particular, are
532 predators of *M. nudus* (see for review [Agatsuma, 2013b](#)), we further analyzed the video records
533 of 2017 captured by 6 video cameras in the course of another project. During 44 days, 132 ± 46
534 of *M. nudus* individuals were in the cameras' field of view and no cases of predator attacks were
535 recorded (*P.M. Zhadan pers. comm., 2018*). Considering also that both sea urchin species
536 showed only a weak response to sea stars even during direct contact, we can conclude that none
537 of 6 starfish species observed on the feeders (*P. pectinifera*, *L. fusca*, *A. amurensis*, *D. nipon*, *L.*
538 *anthosticta* and *A. japonica*) are specialized predators of *S. intermedius* and *M. nudus*. Most
539 likely, starfish perform the function of scavengers.

540 The alarm response of *S. intermedius* to predation depended on the type of alarm signal
541 and involved several phases. The sea urchins: (1) moved away from a source of threat (so called
542 'flight response'), (2) exhibited grouping behavior forming dense two-dimensional groups close
543 to the food source, (3) left the food source, and (4) restored the initial spatial distribution on the
544 food source. Phase 3 was most pronounced when the alarm signals were the attack of starfish on
545 ailing specimens or simulated attack with crushed conspecifics: approximately 50 h after the
546 beginning of starfish attack, there remained only 30–50% of sea urchins on the feeders. When
547 the alarm signal was a simulated attack with crushed heterospecifics, phase 3 was much weakly
548 pronounced or absent.

549 Sea urchins *M. nudus* exhibited a fast (during 4 h) escape (flight response) and prolonged
550 (up to 19 days) avoidance of the source of attractive food near which the conspecifics were
551 crushed. Such a long fear effect of the alarm signal associated with crushed conspecifics is
552 probably due to the marking of the area with substances released by injured sea urchins. At the
553 same time, *M. nudus* exhibited no responses to damaged heterospecifics.

554 Escape of sea urchins from an alarm source is the most well-documented first phase of
555 the alarm response of sea urchins (*Snyder & Snyder, 1970; Parker & Shulman, 1986; Sheibling*
556 *& Hamm, 1991; Campbell et al., 2001; Hagen et al., 2002; Vadas & Elner, 2003; Urriago et al.,*
557 *2011; Wirtz & Duarte, 2012*). It has been shown that the alarm reaction in sea urchins started
558 within a few minutes of exposure to waterborne chemosensory cues from some potential
559 predators (fish, lobsters, crabs, sea stars, gastropods), as well as from crushed prey, conspecifics
560 or heterospecifics. Generally, escape response was most pronounced in the experiments with
561 damaged conspecifics.

562 To date, responses to chemosensory cues from damaged conspecifics have been
563 demonstrated in several sea urchin species: *Diadema antillarum* (*Kintzing & Butler, 2014a;*
564 *Snyder & Snyder, 1970*), *S. droebachiensis* (*Hagen et al., 2002; Mann, 1982*), *Echinometra*
565 *viridis* and *Lytechinus williamsi* (*Parker & Shulman, 1986*), *Echinometra lucunter* (*Morishita &*
566 *Barreto, 2011; Parker & Shulman, 1986*), *Echinus esculentus* and *Psammechinus miliaris*
567 (*Campbell et al., 2001*), *Lytechinus variegatus* and *Tripneustes ventricosus* (*Vadas & Elner,*
568 *2003*), and *Arbacia lixula* and *Sphaerechinus granularis* (*Wirtz & Duarte, 2012*). It should be
569 noted, however, that *Parker & Shulman (1986)* did not find an alarm reaction to extracts of
570 conspecifics when analyzing sea urchin motion for 1 min in *Eucidaris tribuloides*, *T. ventricosus*
571 and *L. variegatus* living in long, dense seagrass that provided protection from detection by
572 predators, and *D. antillarum* occupying crevices.

573 In our studies, both *M. nudus* and *S. intermedius* exhibited a phased escape response to
574 crushed conspecifics: (1) a sharp increase in locomotion speed just after presentation of an alarm
575 signal lasting for 1–1.5 h and resulting in an increase of a distance from a threat source and
576 grouping close to a food source (*S. intermedius*) or escape of approximately half of specimens
577 (*M. nudus*), (2) deceleration lasting for approximately 1 h in *M. nudus* and 16 h in *S. intermedius*,
578 and (3) the second increase in locomotion speed lasting for approximately 1.5 h and resulting in
579 association dispersion and repopulation of feeders (*S. intermedius*) or escape of all remaining
580 specimens (*M. nudus*). Previously, *Vadas & Elner (2003)* found in the field experiments that two
581 sympatric tropical sea urchin species, *L. variegatus* and *T. ventricosus*, demonstrated an initial
582 burst of speed followed by a gradual deceleration up to relatively stable level in response to
583 conspecific alarm cues. However, these movement reactions were much shorter and lasted only 2
584 min each.

585 The most striking difference of these sea urchins in the escape responses to conspecific
586 alarm cues was that *M. nudus* exhibited fast and total escape whereas only a part of *S.*
587 *intermedius* individuals left a food source, and the remaining specimens formed associations on
588 the feeder and in close vicinity to it. Our results showed that grouping behavior is a common
589 phase of the alarm response in *S. intermedius* but not in *M. nudus*. As it was shown in laboratory
590 and field studies, a number of sea urchin species are capable of forming dense groups on a food
591 substrate (*Bernstein et al., 1981, 1983; Garnick, 1978; Pearse, 2006; Vadas & Elner, 2003*).
592 *Bernstein et al. (1981, 1983)* found that the presence of predators (lobsters *Homarus americanus*
593 or crabs *Cancer irroratus*) in laboratory aquariums or in field cages triggered the formation of *S.*
594 *droebachiensis* aggregations that were larger than groups of feeding and non-feeding sea urchins
595 in the absence of predators. The researchers interpreted such aggregation behavior as a defense
596 mechanism of *S. droebachiensis* against predation. *Vadas et al. (1986)*, however, did not find a
597 tendency to form aggregations in the same species in the presence of predators (decapods *H.*
598 *americanus*, *C. irroratus* and sea star *Asterias vulgaris*) and suggested that grouping of sea
599 urchins in tank corners or on tank walls/cage mesh may be an experimental artefact caused by
600 the accumulation of sea urchins near artificial obstacles that prevented them from escaping a
601 predator. This point of view was supported by other studies on interactions between sea urchins
602 *S. droebachiensis* and their predators in field and laboratory experiments (*Harding & Scheibling,*
603 *2015; Scheibling & Hamm, 1991*). In addition, *Vadas & Elner (2003)*, investigating the reactions
604 of sympatric sea urchins *L. variegatus* and *T. ventricosus* to simulated predator attacks in field
605 experiments, also found no formation of sea urchin groups in response to an alarm signal and
606 concluded that the flight response is the primary, and perhaps only defensive behavior employed
607 by these species. However, our results showed that two other sympatric sea urchin species (*M.*
608 *nudus* and *S. intermedius*) exhibit distinctly different behavioral response strategies to predation
609 risk: *M. nudus* employs fast escape and prolonged avoidance of dangerous area while *S.*
610 *intermedius* employs both grouping and escape behaviors.

611 The duration of the alarm response of *S. intermedius* (from the appearance of the alarm
612 signal to the return to feeding) was different under different conditions. It was the longest
613 (approximately 90 h) after the attack of sea stars on ailing individuals. The eating of prey lasted
614 from 45 to 70 h, and after that, from 20 to 45 h passed before sea urchins restored their original
615 arrangement on the feeder. Considering that semidiurnal tidal cycles and constant wave activity

616 took place in the study areas, there is little reason to believe that a waterborne chemical cue from
617 the primary source (injured prey) could have persisted. The secondary source of the alarm signal
618 could be the products of predator metabolism (*Scherer & Smee, 2016*). For example, black sea
619 urchin *E. lucunter* is able to distinguish sea stars feeding on conspecifics or closely related
620 species (*Morishita & Barreto, 2011*). For predatory fish, it has been shown that substances that
621 cause the alarm response in the prey can remain active after passing through the digestive tract
622 (*Manassa & McCormick, 2012*). In addition, it is likely that waterborne chemical cues from
623 predators and/or injured prey may be sorbed on the bottom sediments and gradually released,
624 thereby increasing the time of the alarm reaction in prey.

625 Based on the above data, it may be assumed that the duration of the alarm reaction of sea
626 urchins depends on two main factors: (1) the duration of the release of substances from predators
627 and/or injured prey to the environment, and (2) the time during which sea urchins can detect
628 these substances sorbed on the sediment. In addition, species-specific previous learning might
629 also be a factor determining different behavioral patterns in sea urchin species (*Ferrari et al.,*
630 *2010*).

631 In our study, sea urchins *M. nudus* exhibited unique prolonged avoidance behavior in
632 response to crushed conspecifics, and this behavior has not been described before in sea urchins
633 and other echinoderms. The ability of *M. nudus* to avoid a site of predation for up to 19 days and
634 return to the food source only after a storm indicates that crushed *M. nudus* specimens released
635 some stable substances that marked the bottom for a long time and served as an alarm signal and
636 that the disruption and removal of the upper sediment layer during the storm probably
637 contributed to the removal of the alarm signal. The experiments with storm imitation support this
638 suggestion.

639

640 **Possible mechanisms underlying the difference in sea urchin alarm responses**

641 The mechanisms underlying different patterns of the alarm responses in cohabiting sea urchin
642 species are not yet understood. We believe that both the ability of *S. intermedius* to form
643 associations close to a food source and the ability of *M. nudus* to leave the area of predation risk
644 for a long period are useful evolutionary adaptations that enhance the likelihood of species
645 survival under permanent pressure from visual predators.

646 We suggest that sea urchin *S. intermedius* uses camouflage to protect itself from visual
647 predators because a group of these sea urchins forms a grey spot of irregular shape decorated
648 with algae, which is more difficult to be identified from air or under water than a single object
649 with a regular round shape. Furthermore, a solitary sea urchin is easier to be captured by a diving
650 predator. At the same time, such camouflage cannot be effective for sea urchin *M. nudus* because
651 its black color is in high contrast to the color of the bottom, whereas leaving the area occupied by
652 a predator increases the chances of *M. nudus* survival.

653 It is well known that in temperate waters, the most active consumers of sea urchins that
654 are able to control their abundance are the sea otter *Enhydra lutris* (Duggins, 1980; Estes &
655 Duggins, 1995; Watson & Estes, 2011) and a number of bird species, mainly gulls (Guillemette
656 et al., 1992; Himmelman & Steele, 1971; Hori & Noda, 2007; Merkel et al., 2007; Wootton,
657 1995). Wootton (1995) compared the densities of sea urchin *Strongylocentrotus purpuratus* in
658 several places in a lower intertidal zone both exposed to bird predators (glaucous-winged gulls
659 *Larus glaucescens*, American black oyster catchers *Haematopus bachmani* and northwestern
660 crows *Corvus caurinus*) and protected from birds by cages and showed that sea urchin
661 abundance was 59% lower after 1 year and 45% lower after 2 years in the presence of bird
662 predators compared to the absence of bird predators. For *S. intermedius*, the most abundant avian
663 predators are carrion crow *Corvus corone* and a few gull species that are able to consume a large
664 number of sea urchins, more than 4,000 specimens per 1 ha (Hori & Noda, 2007).

665 Data on the geographical distributions of *S. intermedius* and *M. nudus* (Agatsuma, 2013a,
666 2013b; Bazhin, 1998; Kafanov & Pavlyuchkov, 2001) and the sea otter (Kenyon, 1969) give
667 evidence that the ranges of these species may have partially overlapped in the past, but at
668 present, the overlapping of this predator–prey habitat seems more likely for temperate-boreal
669 species, *S. intermedius*, which inhabits the Asian Pacific coastal waters from the Kamchatka
670 Peninsula southward to the Korean Peninsula and from the Russian coast eastward to the
671 Japanese Islands. The sea urchin *M. nudus* is a subtropical species, and coastal waters of the Sea
672 of Japan near Russia (Primorye Region) and Japan (northern Hokkaido) represent the northern
673 part of its range, whereas for the sea otter, the northern Hokkaido represented the southern
674 boundary of its range in the northwestern Pacific until the 18th century, before fur hunting began
675 (Wilson et al., 1991).

676 Sea otters and predatory birds prefer sea urchins of medium and large size, i.e., adult
677 specimens contributing to population reproduction (*Estes & Duggins, 1995; Guillemette et al.,*
678 *1992; Himmelman & Steele, 1971; Hori & Noda, 2007*). The defensive behaviors of sea urchins
679 *S. intermedius* and *M. nudus* could have formed mainly under the pressure of these predators.
680 Due to natural selection, the individuals that could avoid predation attacks survived, and useful
681 genetic traits have been passed from generation to generation in the form of different defensive
682 behaviors.

683

684 CONCLUSION

685 Our results show that cohabiting sea urchin species, *S. intermedius* and *M. nudus*, which were
686 monitored in their natural environment under conditions of food abundance display both similar
687 and different behavioral responses to hydrodynamics and predation threat. The most interesting
688 findings are the following: (1) under calm weather, *S. intermedius* but not *M. nudus* tended to
689 group on the food substrate; movement patterns of both sea urchins were similar but *M. nudus*
690 exhibited the higher locomotion speed and distance travelled; (2) both sea urchins responded to
691 increased wave activity by a sharp decrease in the movement activity up to almost complete stop
692 and then made a decision to stay close to the food or escape; (3) several sea star species failed to
693 predate on healthy sea urchins of both species, and only a few starfish attacks on ailing *S.*
694 *intermedius* specimens were successful; (4) the alarm response of *S. intermedius* depended on the
695 type of alarm signal (consumption of ailing conspecifics by starfish or simulated attack) and
696 included the formation of dense groups close to the food source; (5) the alarm response of *S.*
697 *intermedius* lasted approximately 90 h and 20 h for starfish attacks on ailing conspecifics and for
698 simulated attacks (crushed conspecifics or heterospecifics), respectively; (6) *M. nudus* responded
699 to crushed conspecifics only and exhibited no grouping behavior but displayed fast escape
700 (during 4 h) and prolonged (up to 19 days) avoidance of the food source; (7) both sea urchins
701 exhibited a phased escape response to crushed conspecifics consisting of a sharp increase in
702 locomotion speed just after presentation of the alarm signal followed by deceleration, and the
703 second increase in locomotion speed associated with repopulation of feeders (*S. intermedius*) or
704 complete escape (*M. nudus*); (8) damaged specimens of *M. nudus* released some stable alarm
705 substances. Furthermore, our results show the benefits of using continuous time-lapse video
706 recording to study the long-term behavioral responses of sea urchins to different disturbing

707 factors such as high hydrodynamic activity and predation threat. Considering the important
708 ecological role of sea urchins as grazers of marine plants, data on the duration of the fear
709 response in sea urchin species, i.e., the periods when their foraging activity is inhibited, may be
710 of greatest use in mathematical modelling of the marine ecosystem.

711

712 **ACKNOWLEDGEMENTS**

713 We thank M.Yu. Cheranov for the help in making the underwater installation for video recording
714 and L.Yu. Pavin for assistance in the field.

715

716 **REFERENCES**

- 717 **Abrams PA. 1995.** Implications of dynamically variable traits for identifying, classifying, and
718 measuring direct and indirect effects in ecological communities. *American Naturalist*
719 **146(1):** 112–134 [DOI 10.1086/285789](https://doi.org/10.1086/285789).
- 720 **Agatsuma Y. 2013a.** *Strongylocentrotus intermedius*. In: Lawrence JM, ed. *Sea Urchins:*
721 *Biology and Ecology*. Elsevier B.V., 438–447 [DOI 10.1016/B978-0-12-396491-5.00028-](https://doi.org/10.1016/B978-0-12-396491-5.00028-9)
722 [9](https://doi.org/10.1016/B978-0-12-396491-5.00028-9).
- 723 **Agatsuma Y. 2013b.** *Strongylocentrotus nudus*. In: Lawrence JM, ed. *Sea Urchins: Biology and*
724 *Ecology*. Elsevier B.V., 449–460 [DOI 10.1016/B978-0-12-396491-5.00029-0](https://doi.org/10.1016/B978-0-12-396491-5.00029-0).
- 725 **Bazhin AG. 1998.** The sea urchin genus *Strongylocentrotus* in the seas of Russia: taxonomy and
726 ranges. In: Mooi R, Telford M, eds. *Echinoderms: Proceedings of the 9th International*
727 *Echinoderm Conference, San Francisco, August 1996*. Rotterdam: Balkema, 563–566.
- 728 **Bernstein BB, Schroeter SC, Mann KH. 1983.** Sea urchin (*Strongylocentrotus droebachiensis*)
729 aggregating behavior investigated by a subtidal multifactorial experiment. *Canadian*
730 *Journal of Fisheries and Aquatic Sciences* **40:**1975–1986 [DOI 10.1139/f83-227](https://doi.org/10.1139/f83-227).

- 731 **Bernstein BB, Williams BE, Mann KH. 1981.** The role of behavioral responses to predators in
732 modifying urchins' (*Strongylocentrotus droebachiensis*) destructive grazing and seasonal
733 foraging patterns. *Marine Biology* **63**:39–49 DOI [10.1007/BF00394661](https://doi.org/10.1007/BF00394661).
- 734 **Campbell AC, Coppard S, Abreo CD, Tudor-Thomas R. 2001.** Escape and aggregation
735 responses of three echinoderms to conspecific stimuli. *Biological Bulletin* **201**:175–185
736 DOI <https://www.journals.uchicago.edu/doi/abs/10.2307/1543332?journalCode=bbl>.
- 737 **Cannone AJ. 1970.** The anatomy and venom-emitting mechanism of the globiferous
738 pedicellariae of the urchin *Parechinus angulosus* (Leske) with notes on their behaviour.
739 *Zoologica Africana* **5**:179–190 DOI
740 <https://www.ajol.info/index.php/az/article/view/152472/142062>
- 741 **Chivers DP, Smith JF. 1998.** Chemical alarm signalling in aquatic predator-prey systems: a
742 review and prospectus. *Ecoscience* **5**:338–352 DOI [10.1080/11956860.1998.11682471](https://doi.org/10.1080/11956860.1998.11682471).
- 743 **Clemente S, Hernandez JC, Montano-Moctezuma G, Russell M, Ebert TA. 2013.** Predators
744 of juvenile sea urchins and the effect of habitat refuges. *Marine Biology* **160(3)**:579–590
745 DOI [10.1007/s00227-012-2114-3](https://doi.org/10.1007/s00227-012-2114-3).
- 746 **Crook AC, Long M, Barnes DKA. 2000.** Quantifying daily migration in the sea urchin
747 *Paracentrotus lividus*. *Journal of the Marine Biological Association of the UK*
748 **80(1)**:177–178 DOI [10.1017/S0025315499001721](https://doi.org/10.1017/S0025315499001721).
- 749 **Cohen-Rengifo M, Agüera A, Detrain C, Bouma TJ, Dubois P, Flammang P. 2018.**
750 Biomechanics and behaviour in the sea urchin *Paracentrotus lividus* (Lamarck, 1816)
751 when facing gradually increasing water flows. *Journal of Experimental Marine Biology*
752 *and Ecology* **506**:61–71 DOI [10.1016/j.jembe.2018.05.010](https://doi.org/10.1016/j.jembe.2018.05.010).

- 753 **Dance C. 1987.** Patterns of activity of the sea urchin *Paracentrotus lividus* in the Bay of Port-
754 Cros (Var, France, Mediterranean). *Marine Ecology* **8**:131–142 DOI [10.1111/j.1439-](https://doi.org/10.1111/j.1439-0485.1987.tb00179.x)
755 [0485.1987.tb00179.x](https://doi.org/10.1111/j.1439-0485.1987.tb00179.x).
- 756 **Dill LM, Heithaus MR, Walters CJ. 2003.** Behaviorally mediated indirect interactions in
757 marine communities and their conservation implications. *Ecology* **84(5)**:1151–1157 DOI
758 [10.1890/0012-9658\(2003\)084\[1151:BMIIIM\]2.0.CO;2](https://doi.org/10.1890/0012-9658(2003)084[1151:BMIIIM]2.0.CO;2).
- 759 **Duggins DO. 1980.** Kelp beds and sea otters: an experimental approach. *Ecology* **61**: 447–453
760 DOI [10.2307/1937405](https://doi.org/10.2307/1937405).
- 761 **Dumont CP, Himmelman JH, Russell MP. 2006.** Daily movement of the sea urchin
762 *Strongylocentrotus droebachiensis* in different subtidal habitats in eastern Canada. *Marine*
763 *Ecology Progress Series* **317**: 87–99 DOI [0.3354/meps317087](https://doi.org/0.3354/meps317087).
- 764 **Dumont CP, Himmelman JH, Robinson SMC. 2007.** Random movement pattern of the sea
765 urchin *Strongylocentrotus droebachiensis*. *Journal of Experimental Marine Biology and*
766 *Ecology* **340**:80–89 DOI [10.1016/j.jembe.2006.08.013](https://doi.org/10.1016/j.jembe.2006.08.013).
- 767 **Estes JA, Duggins DO. 1995.** Sea otters and kelp forests in Alaska: generality and variation in a
768 community ecological paradigm. *Ecological Monographs* **65**:75–100 DOI
769 [10.2307/2937159](https://doi.org/10.2307/2937159).
- 770 **Ferrari MCO, Wisenden BD, Chivers DP. 2010.** Chemical ecology of predator-prey
771 interactions in aquatic ecosystems: a review and prospectus. *Canadian Journal of*
772 *Zoology* **88**:698–724 DOI [10.1139/Z10-029](https://doi.org/10.1139/Z10-029).
- 773 **Frey DL, Gagnon P. 2016.** Spatial dynamics of the green sea urchin *Strongylocentrotus*
774 *droebachiensis* in food-depleted habitats. *Marine Ecology Progress Series* **552**:223–240
775 DOI <https://doi.org/10.3354/meps11787>.

- 776 **Freeman A. 2006.** Size-dependent trait-mediated indirect interactions among sea urchin
777 herbivores. *Behavioral Ecology* **17**:182–187 DOI<https://doi.org/10.1093/beheco/arj014>.
- 778 **Fricke HW. 1973.** Behaviour as part of ecological adaptation. *In situ* studies in the coral reef.
779 *Helgoländer Wissenschaftliche Meeresuntersuchungen* **24**:120–144
780 DOI<https://link.springer.com/article/10.1007/BF01609505>.
- 781 **Garnick E. 1978.** Behavioral ecology of *Strongylocentrotus droebachiensis* (Muller)
782 (Echinodermata: Echinoidea). Aggregating behavior and chemotaxis. *Oecologia (Berl.)*
783 **37**:77–84 DOI[10.1007/BF00349993](https://doi.org/10.1007/BF00349993).
- 784 **Guillemette M, Ydenberg RC, Himmelman JH. 1992.** The role of energy intake rate in prey
785 and habitat selection of common eiders *Somateria mollissima* in winter: a risk-sensitive
786 interpretation. *Journal of Animal Ecology* **61**:599–610 DOI[10.2307/5615](https://doi.org/10.2307/5615).
- 787 **Hagen NT, Andersen A, Stabell OB. 2002.** Alarm responses of the green sea urchin,
788 *Strongylocentrotus droebachiensis*, induced by chemically labeled durophagus predators
789 and simulated acts of predation. *Marine Biology* **140**:365–374
790 DOI[10.1007/s002270100694](https://doi.org/10.1007/s002270100694).
- 791 **Hagen NT, Mann KH. 1994.** Experimental analysis of factors influencing the aggregating
792 behavior of the green sea urchin *Strongylocentrotus droebachiensis* (Müller). *Journal of*
793 *Experimental Marine Biology and Ecology* **176**:107–126 DOI[10.1016/0022-](https://doi.org/10.1016/0022-0981(94)90200-3)
794 [0981\(94\)90200-3](https://doi.org/10.1016/0022-0981(94)90200-3).
- 795 **Harding APC, Scheibling RE. 2015.** Feed or flee: effect of a predation-risk cue on sea urchin
796 foraging activity. *Journal of Experimental Marine Biology and Ecology* **466**:59–69
797 DOI<https://doi.org/10.1016/j.jembe.2015.02.005>.

- 798 **Hasegawa K. 2014.** Analysis of circadian rhythm in *Strongylocentrotus nudus* using a
799 commercial digital camera with an interval function. *Reports of Marine Ecology*
800 *Research Institute (MERI, Tokyo)* **18**:63–65 (in Japanese).
801 http://www.kaiseiken.or.jp/english/meri_report2017.pdf.
- 802 **Hereu B, Zabala M, Linares C, Sala E. 2005.** The effects of predator abundance and habitat
803 structural complexity on survival of juvenile sea urchins. *Marine Biology* **146**:293–299
804 DOI <https://link.springer.com/article/10.1007/s00227-004-1439-y>.
- 805 **Himmelman JH, Steele DH. 1971.** Foods and predators of the green sea urchin
806 *Strongylocentrotus droebachiensis* in Newfoundland waters. *Marine Biology* **9**:315–322
807 DOI [10.1007/BF00372825](https://doi.org/10.1007/BF00372825).
- 808 **Hori M, Noda T. 2007.** Avian predation on wild and cultured sea urchin *Strongylocentrotus*
809 *intermedius* in a rocky shore habitat. *Fisheries Science* **73(2)**:303–313
810 DOI [10.1111/j.1444-2906.2007.01336.x](https://doi.org/10.1111/j.1444-2906.2007.01336.x).
- 811 **James DW. 2000.** Diet, movement, and covering behavior of the sea urchin *Toxopneustes roseus*
812 in rhodolith beds in the Gulf of California, México. *Marine Biology* **137**:913–923 DOI
813 [10.1007/s002270000423](https://doi.org/10.1007/s002270000423).
- 814 **Jensen M. 1966.** The response of two sea-urchins to the sea-star *Marthasterias glacialis* (L.) and
815 other stimuli. *Ophelia* **3**:209–219 DOI <https://doi.org/10.1080/00785326.1966.10409643>.
- 816 **Kafanov AI, Pavlyuchkov VA. 2001.** Ecology of the commercial sea urchins (genus
817 *Strongylocentrotus*) of continental Japan Sea. *Proceedings of the Pacific Research*
818 *Institute of Fisheries and Oceanography (TINRO-Center)* **128(2)**:349–373 (in Russian
819 with English summary).

- 820 **Kawamata S. 1998.** Effect of wave-induced oscillatory flow on grazing by a subtidal sea urchin
821 *Strongylocentrotus nudus* (A. Agassiz). *Journal of Experimental Marine Biology and*
822 *Ecology* **224**:31–48 DOI[https://doi.org/10.1016/S0022-0981\(97\)00165-2](https://doi.org/10.1016/S0022-0981(97)00165-2).
- 823 **Kenyon KW. 1969.** The sea otter in the eastern Pacific Ocean. *North American Fauna* **68**:1–352
824 DOI[10.3996/nafa.68.0001](https://doi.org/10.3996/nafa.68.0001).
- 825 **Kintzing MD, Butler MJ. 2014a.** The influence of shelter, conspecifics, and threat of predation
826 on the behavior of the long-spined sea urchin (*Diadema antillarum*). *Journal of Shellfish*
827 *Research* **33**:3781–3785 DOI[10.2983/035.033.0312](https://doi.org/10.2983/035.033.0312).
- 828 **Kintzing MD, Butler MJ. 2014b.** Effects of predation upon the long-spined sea urchin *Diadema*
829 *antillarum* by the spotted spiny lobster *Panulirus guttatus*. *Marine Ecology Progress*
830 *Series* **495**:185–191 DOI[10.3354/meps10568](https://doi.org/10.3354/meps10568).
- 831 **Lauzon-Guay J-S, Scheibling RE, Barbeau MA. 2006.** Movement patterns in the green sea
832 urchin, *Strongylocentrotus droebachiensis*. *Journal of the Marine Biological Association*
833 *of the UK* **86**:167–174 DOI<https://doi.org/10.1017/S0025315406012999>.
- 834 **Lauzon-Guay J-S, Scheibling RE. 2007.** Behaviour of sea urchin *Strongylocentrotus*
835 *droebachiensis* grazing fronts: food-mediated aggregation and density-dependent
836 facilitation. *Marine Ecology Progress Series* **329**:191–204
837 DOI<http://dx.doi.org/10.3354/meps329191>.
- 838 **Lima SL. 1998.** Non lethal effects in the ecology of predator-prey interactions. What are the
839 ecological effects of anti-predator decision-making? *BioScience* **48(1)**:25–34
840 DOI<https://doi.org/10.2307/1313225>.

- 841 **Lima SL, Dill LM. 1990.** Behavioral decisions made under the risk of predation: a review and
842 prospectus. *Canadian Journal of Zoology* **68**:619–640 DOI[https://doi.org/10.1139/z90-](https://doi.org/10.1139/z90-092)
843 [092](https://doi.org/10.1139/z90-092).
- 844 **Lissner AL. 1980.** Some effects of turbulence on the activity of the sea urchin *Centrostephanus*
845 *coronatus* Verrill. *Journal of Experimental Marine Biology and Ecology* **48**:185–193
846 DOI [https://doi.org/10.1016/0022-0981\(80\)90016-7](https://doi.org/10.1016/0022-0981(80)90016-7).
- 847 **Machiguchi Y. 1987.** Feeding behavior of sea urchin *Strongylocentrotus intermedius* (A.
848 Agassiz) observed in Y-shaped chamber. *Bulletin of the Hokkaido Regional Fisheries*
849 *Research Institute* **51**:33–37 (in Japanese with English abstract).
- 850 **Machiguchi Y, Mizutori S, Sanbonsuga Y. 1994.** Food preference of sea urchin
851 *Strongylocentrotus nudus* in laboratory. *Bulletin of the Hokkaido National Fisheries*
852 *Research Institute* **58**:35–43 (in Japanese with English abstract).
- 853 **Manassa RP, McCormick MI. 2012.** Risk assessment via predator diet cues in a coral reef
854 goby. *Journal of Experimental Marine Biology and Ecology* **426–427**:48–52
855 DOI<https://doi.org/10.1016/j.jembe.2012.05.023>.
- 856 **Mann KH. 1982.** Kelp, sea urchins and predators: a review of strong interactions in rocky
857 subtidal systems of eastern Canada, 1970–1980. *Netherlands Journal of Sea Research*
858 **16**:414–423 DOI[https://doi.org/10.1016/0077-7579\(82\)90047-3](https://doi.org/10.1016/0077-7579(82)90047-3).
- 859 **Manzur T, Navarrete SA. 2011.** Scales of detection and escape of the sea urchin *Tetrapygus*
860 *niger* in interactions with the predatory sun star *Heliaster helianthus*. *Journal of*
861 *Experimental Marine Biology and Ecology* **407**:302–308
862 DOI<https://doi.org/10.1016/j.jembe.2011.06.025>.

- 863 **Matassa CM. 2010.** Purple sea urchins *Strongylocentrotus purpuratus* reduce grazing rates in
864 response to risk cues from the spiny lobster *Panulirus interruptus*. *Marine Ecology*
865 *Progress Series* **400**:283–288 DOI <https://doi.org/10.3354/meps08425>.
- 866 **Merkel FR, Mosbech A, Jamieson SE, Falk K. 2007.** The diet of king eiders wintering in
867 Nuuk, Southwestern Greenland, with reference to sympatric wintering common eiders.
868 *Polar Biology* **30**:1593–1597 DOI <https://doi.org/10.1007/s00300-007-0321-z>.
- 869 **Morishita VR, Barreto RE. 2011.** Black sea urchins evaluate predation risk using chemical
870 signals from a predator and injured con- and heterospecific prey. *Marine Ecology*
871 *Progress Series* **435**:173–181 DOI <http://dx.doi.org/10.3354/meps09253>.
- 872 **Morse B, Hunt H. 2013.** Effect of unidirectional water currents on displacement behaviour of
873 the green sea urchin *Strongylocentrotus droebachiensis*. *Journal of the Marine Biological*
874 *Association of the UK* **93(7)**:1923–1928
875 DOI <https://doi.org/10.1017/S002531541300060X>.
- 876 **Parker DA, Shulman MJ. 1986.** Avoiding predation: alarm responses of Caribbean sea urchins
877 to simulated predation on conspecific and heterospecific sea urchins. *Marine Biology*
878 **93**:201–208 DOI <https://doi.org/10.1007/BF00508257>.
- 879 **Pearse JS. 2006.** Ecological role of purple sea urchins. *Science* **314**:940–941 DOI
880 [10.1126/science.1131888](https://doi.org/10.1126/science.1131888).
- 881 **Pearson DE. 2010.** Trait- and density-mediated indirect interactions initiated by an exotic
882 invasive plant autogenic ecosystem engineer. *American Naturalist* **176(4)**:394–403 DOI
883 <https://doi.org/10.1086/656274>.

- 884 **Scheibling RE, Hamm J. 1991.** Interactions between sea urchins (*Strongylocentrotus*
885 *droebachiensis*) and their predators in field and laboratory experiments. *Marine Biology*
886 **110**:105–116 DOI <https://doi.org/10.1007/BF01313097>.
- 887 **Scherer AE, Smee DL. 2016.** A review of predator diet effects on prey defensive responses.
888 *Chemoecology* **26(3)**:83–100 DOI [10.1007/s00049-016-0208-y](https://doi.org/10.1007/s00049-016-0208-y).
- 889 **Schmitz OJ, Krivan V, Ovadia O. 2004.** Trophic cascades: the primacy of trait-mediated
890 indirect interactions. *Ecology Letters* **7**:153–163 DOI [https://doi.org/10.1111/j.1461-](https://doi.org/10.1111/j.1461-0248.2003.00560.x)
891 [0248.2003.00560.x](https://doi.org/10.1111/j.1461-0248.2003.00560.x).
- 892 **Siddon CE, Witman JD. 2003.** Influence of chronic, low-level hydrodynamic forces on subtidal
893 community structure. *Marine Ecology Progress Series* **261**:99–110
894 DOI [10.3354/meps261099](https://doi.org/10.3354/meps261099).
- 895 **Snyder N, Snyder H. 1970.** Alarm response of *Diadema antillarum*. *Science* **168**:276–278
896 DOI [10.1126/science.168.3928.276](https://doi.org/10.1126/science.168.3928.276).
- 897 **Spyksma AJP, Taylor RB, Shears NT. 2017.** Predation cues rather than resource availability
898 promote cryptic behaviour in a habitat-forming sea urchin. *Oecologia* **183(3)**:821–829
899 DOI [10.1007/s00442-017-3809-4](https://doi.org/10.1007/s00442-017-3809-4).
- 900 **Steneck RS. 2013.** Sea urchins as drivers of shallow benthic marine community structure. In:
901 Lawrence JM, ed. *Sea Urchins: Biology and Ecology*. Elsevier B.V., 195–212
902 DOI <https://doi.org/10.1016/B978-0-12-396491-5.00014-9>.
- 903 **Tamaki H, Muraoka D, Inoue T. 2018.** Effect of water flow on grazing by the sea urchin
904 (*Strongylocentrotus nudus*) in the presence of refuge habitat. *Journal of Water and*
905 *Environment Technology* **16(1)**:30–39 DOI [10.2965/jwet.17-010](https://doi.org/10.2965/jwet.17-010).

- 906 **Tertschnig WP. 1989.** Diel activity patterns and foraging dynamics of the sea urchin
907 *Tripneustes ventricosus* in a tropical seagrass community and a reef environment (Virgin
908 Islands). *Marine Ecology* **10(1)**:3–21 DOI[https://doi.org/10.1111/j.1439-](https://doi.org/10.1111/j.1439-0485.1989.tb00063.x)
909 [0485.1989.tb00063.x](https://doi.org/10.1111/j.1439-0485.1989.tb00063.x).
- 910 **Trussell GC, Ewanchuk PJ, Matassa CM. 2006.** Habitat effects on the relative importance of
911 trait and density-mediated indirect interactions. *Ecology Letters* **9**:1245–1252.
912 DOI[10.1111/j.1461-0248.2006.00981.x](https://doi.org/10.1111/j.1461-0248.2006.00981.x).
- 913 **Tuya F, Martin JA, Luque A. 2004.** Patterns of nocturnal movement of the long-spined sea
914 urchin *Diadema antillarum* (Philippi) in Gran Canaria (the Canary Islands, central East
915 Atlantic Ocean). *Helgoland Marine Research* **58**:26–31 DOI
916 <https://doi.org/10.1007/s10152-003-0164-0>.
- 917 **Ullrich-Lüter EM, Dupon TS, Arboleda E, Hausen H, Arnone MI. 2011.** Unique system of
918 photoreceptors in sea urchin tube feet. *Proceedings of the National Academy of Sciences*
919 *of the United States of America* **108(20)**:8367–8372
920 DOI<https://doi.org/10.1073/pnas.1018495108>.
- 921 **Urriago JD, Himmelman JH, Gaymer CF. 2011.** Responses of the black sea urchin
922 *Tetrapygus niger* to its sea-star predators *Heliaster helianthus* and *Meyenaster*
923 *gelatinosus* under field conditions. *Journal of Experimental Marine Biology and Ecology*
924 **399**:17–24 DOI<https://doi.org/10.1016/j.jembe.2011.01.004>.
- 925 **Vadas RL, Elnor RW, Garwood PE, Babb IG. 1986.** Experimental evaluation of aggregation
926 behavior in the sea urchin *Strongylocentrotus droebachiensis*. A reinterpretation. *Marine*
927 *Biology* **90**:433–448 DOI <https://doi.org/10.1007/BF00428567>.

- 928 **Vadas RL, Elnor RW. 2003.** Responses to predation cues and food in two species of sympatric,
929 tropical sea urchins. *Marine Ecology* **24(2)**:101–121 DOI [https://doi.org/10.1046/j.1439-](https://doi.org/10.1046/j.1439-0485.2003.03817.x)
930 [0485.2003.03817.x](https://doi.org/10.1046/j.1439-0485.2003.03817.x).
- 931 **Watson JC, Estes JA. 2011.** Stability, resilience and phase shifts in rocky subtidal communities
932 along the west coast of Vancouver Island. *Ecological Monographs* **81(2)**:215–239
933 DOI <https://doi.org/10.1890/10-0262.1>.
- 934 **Werner EE, Peacor SD. 2003.** A review of trait-mediated indirect interactions in ecological
935 communities. *Ecology* **84(5)**:1083–1100 DOI [https://doi.org/10.1890/0012-](https://doi.org/10.1890/0012-9658(2003)084[1083:AROTII]2.0.CO;2)
936 [9658\(2003\)084\[1083:AROTII\]2.0.CO;2](https://doi.org/10.1890/0012-9658(2003)084[1083:AROTII]2.0.CO;2).
- 937 **Wilson DE, Bogan MA, Brownell RLJ, Burdin AM, Maminov MK. 1991.** Geographic
938 variation in sea otters, *Enhydra lutris*. *Journal of Mammalogy* **72(1)**:22–36
939 DOI <https://doi.org/10.2307/1381977>.
- 940 **Wirtz P, Duarte J. 2012.** Alarm reaction and absence of alarm reaction to simulated predation
941 on conspecifics in two temperate sea urchin species. *Arquipelago – Life and Marine*
942 *Sciences* **29**:59–62 URI <http://hdl.handle.net/10400.3/1440>.
- 943 **Wootton J. 1995.** Effects of birds on sea urchin and algae: a lower-intertidal trophic cascade.
944 *Ecoscience* **2(4)**:321–328.
- 945 **Zhadan PM, Vaschenko MA, Ryazanov SD. 2018.** Assessing the effect of environmental
946 factors on the spawning activity of the sea urchin *Strongylocentrotus intermedius* through
947 video recording observations. *Marine Ecology Progress Series* **588**:101–119
948 DOI <https://doi.org/10.3354/meps12436>.

Figure 1

Temporal dynamics of the numbers of sea urchins *Mesocentrotus nudus* and *Strongylocentrotus intermedius* in long-term experiments.

The experiments were conducted in: (A) Kievka Bay, 2014; (B) Kievka Bay, 2015; (C) Alekseev Bay, 2016. Blue and red circles connected by lines denote the numbers of *M. nudus* and *S. intermedius*, respectively, presented as the sum of all sea urchins of the given species on 4 feeders per each 6 h of observations (see Table S1 for original data). Triangles indicate the time points when sea urchins *M. nudus* were removed from the feeders. Upside down triangles denote the time points when sea urchins *M. nudus* were crushed near the feeders. Green squares denote the time points when the feeders were changed. Green rhombuses indicate the time points when the feeders were changed after the mimicking of stormy weather conditions. Shaded areas denote storm periods. Solid violet horizontal lines indicate periods of poor visibility because of high water turbidity. X-axis: month and date.

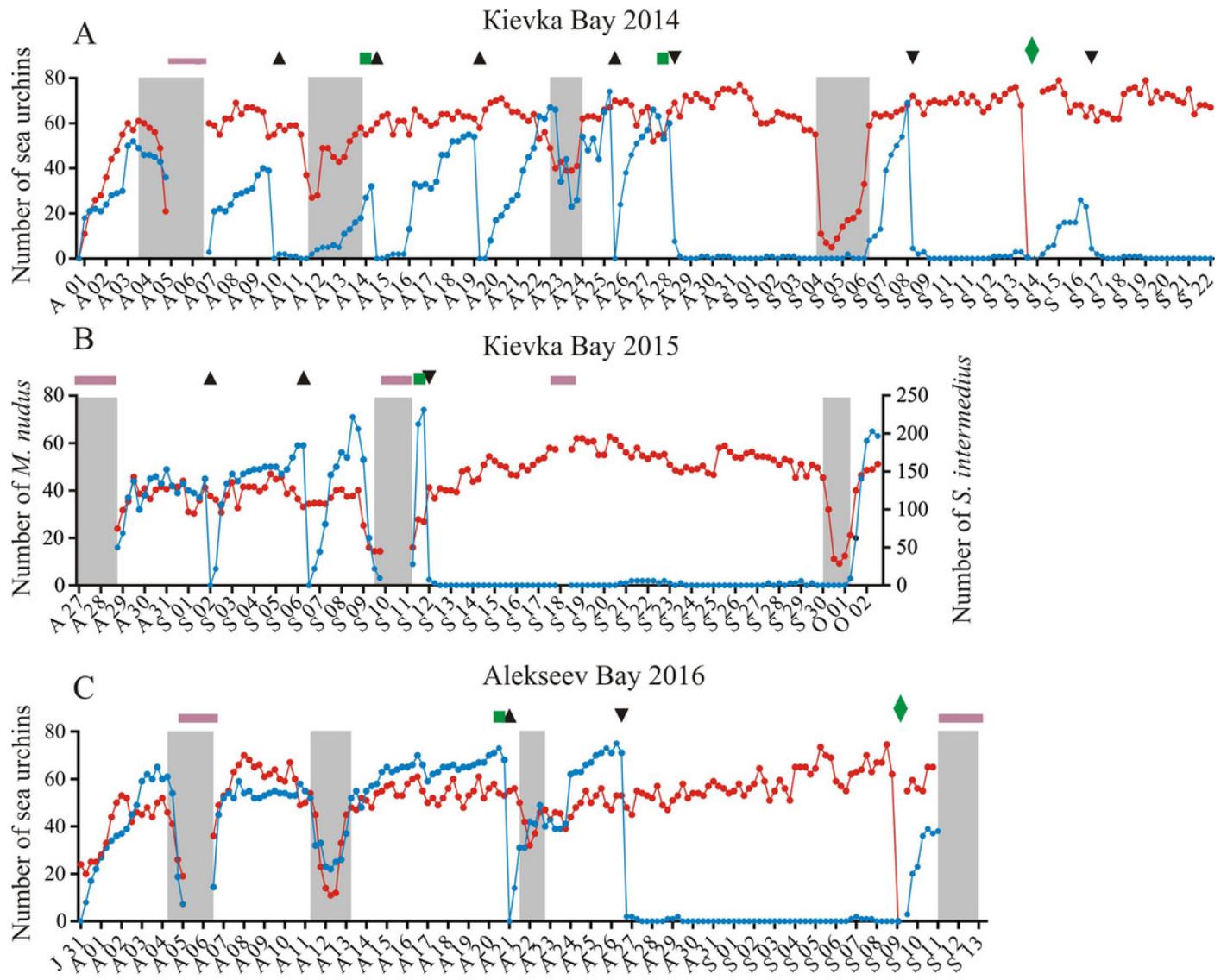


Figure 2

Changes in the numbers of sea urchins *Mesocentrotus nudus* (A) and *Strongylocentrotus intermedius* (B) in response to the stormy weather.

Range, upper and lower quartile (box), mean (+), and median (solid line) of the numbers of sea urchins before, during and after the storm periods are presented. Different lowercase letters above the boxes indicate significant differences in sea urchin numbers: (A) the differences between 'before the storm' and 'during the storm' numbers are significant at $P < 0.0001$, the differences between 'during the storm' and 'after the storm' numbers are significant at $P < 0.001$ (1-way ANOVA followed by Tukey's multiple comparisons test), (B) the differences between 'before the storm' and 'during the storm' numbers, 'during the storm' and 'after the storm' numbers are significant at $P < 0.0001$ (Kruskal-Wallis test followed by Dunn's multiple comparisons test) . See Table S5 and Table S6 for raw data and statistics.

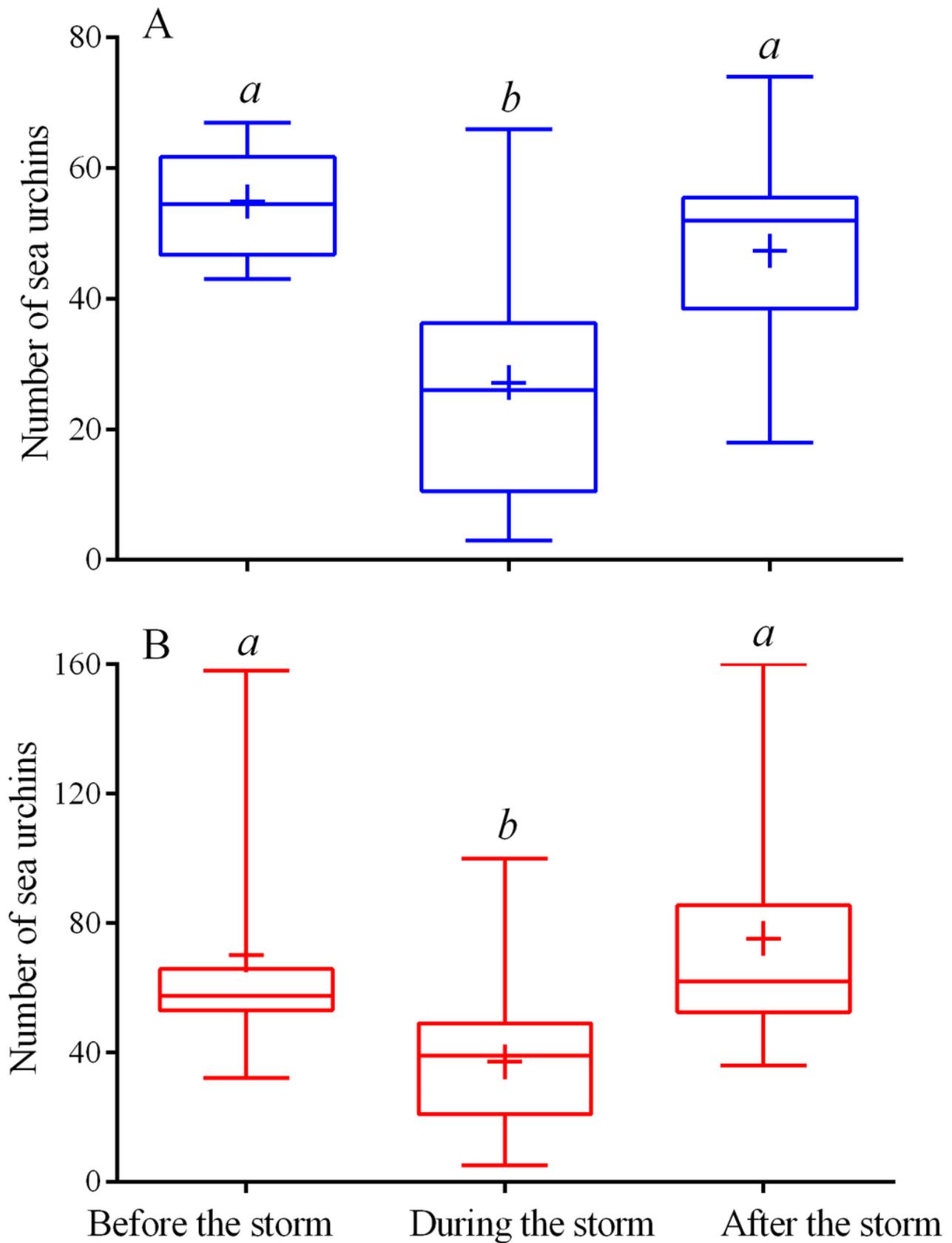


Figure 3

Movement activity of the sea urchins *Strongylocentrotus intermedius* in response to starfish attacks.

(A) Starfish attack on September 5, 2015. (B) Starfish attack on September 21, 2015. Red rhombuses denote sea urchin distances from the site of the attack, Mean \pm SD (n = 13). Black squares connected by dashed line denote the sum number of 2 species of sea stars (*Patiria pectinifera* and *Lethasterias fusca*) at the site of the attack. Red circles connected by solid line denote the number of sea urchins in the cameras' field of view. Time of the beginning of the attack is indicated by black asterisk.

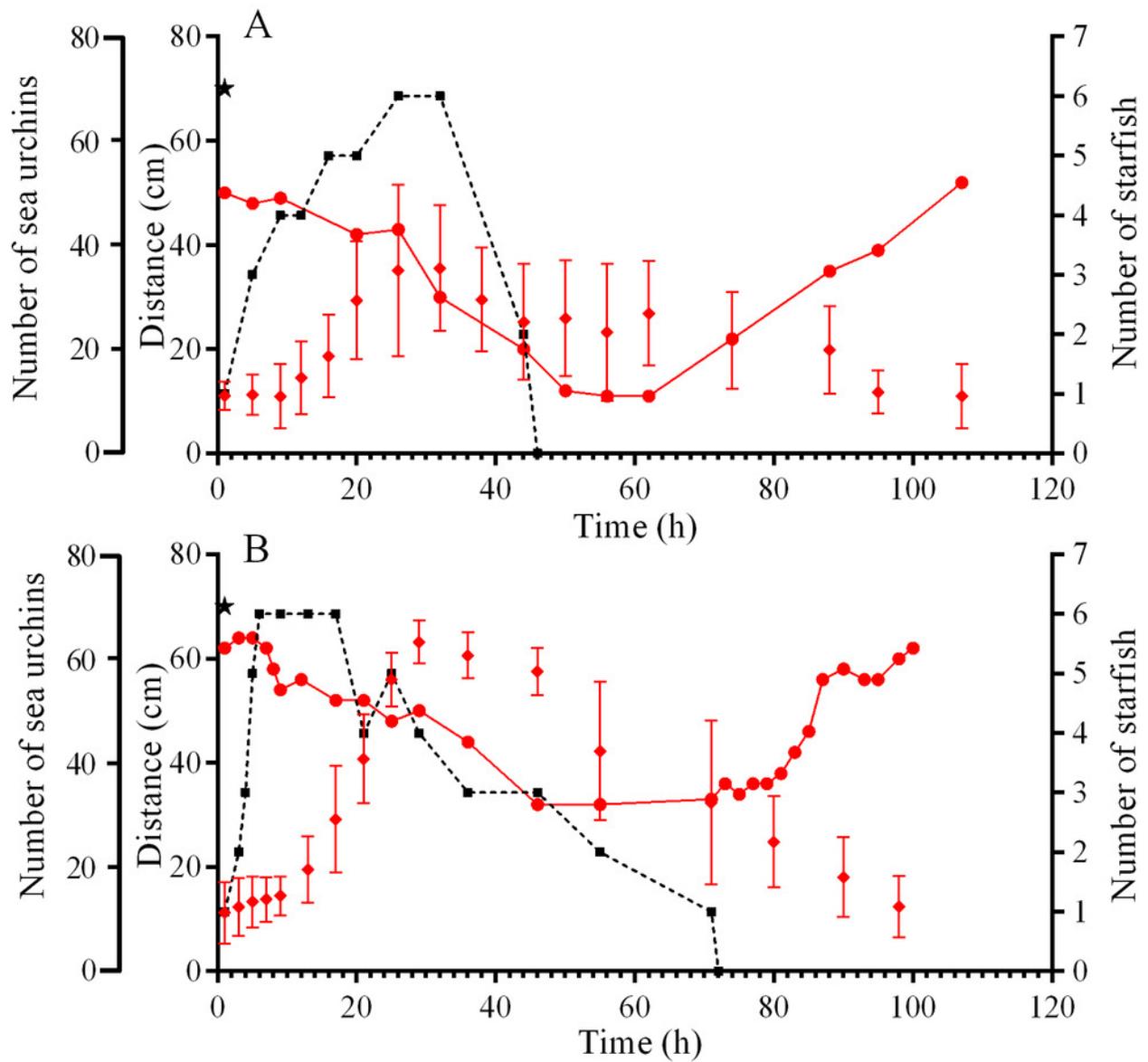


Figure 4

Changes in the numbers of sea urchins *Mesocentrotus nudus* during escape in response to crushed conspecifics in long-term experiments of 2014–2016.

The data of 4 experiments conducted during long-term recordings of 2014–2016 (see Fig. 1A–C, Tables S1, S8) are presented as median and range of the number of *M. nudus* per 1 h after crushing of conspecifics. Nonlinear regression is significant ($R^2 = 0.9454$, see Table S8 for raw data and statistics).

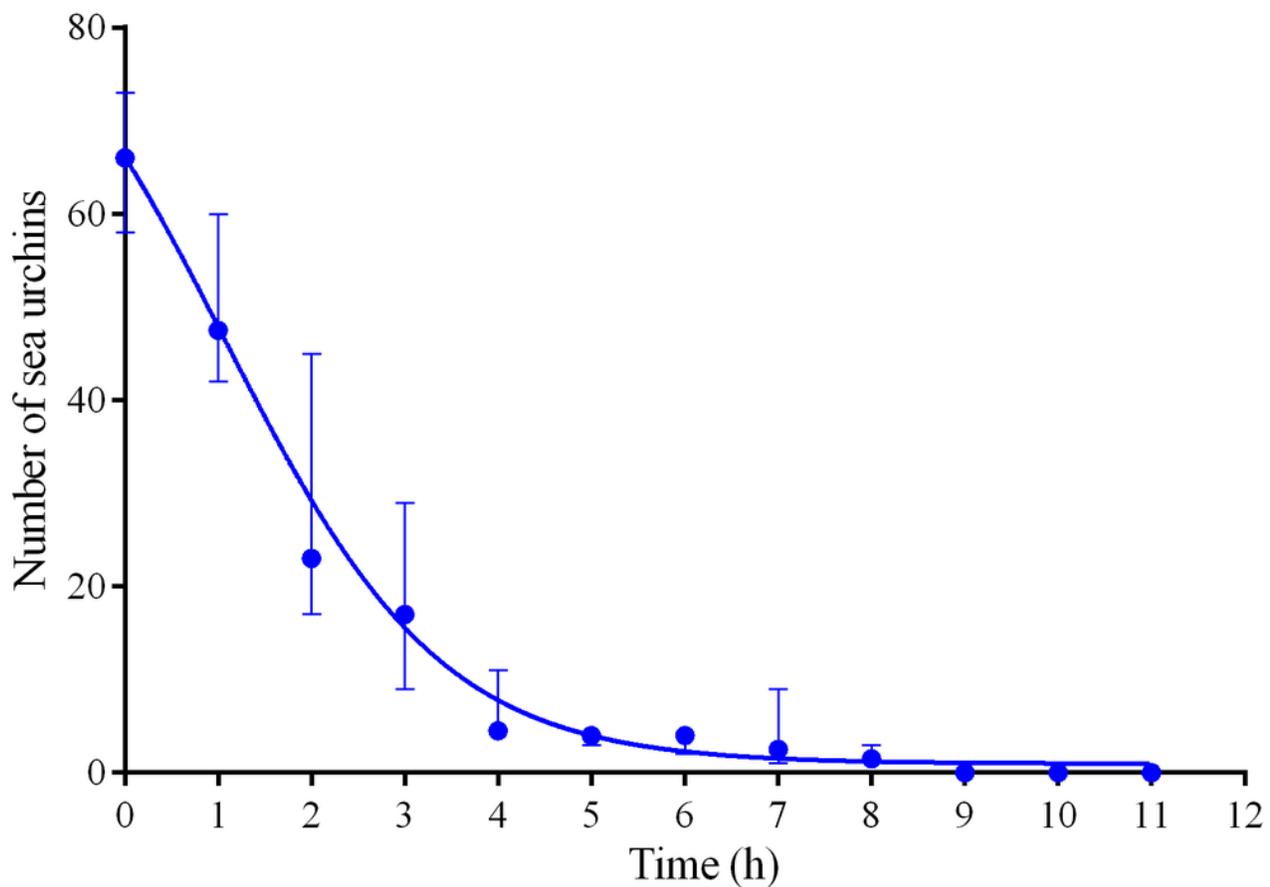


Figure 5

Movement activity of the sea urchins *Mesocentrotus nudus* in response to crushed conspecifics.

Blue solid line denotes the average step length of sea urchins ($n = 10$). Blue circles connected by dashed line denotes the number of sea urchins in the cameras' field of view. Time point of treatment is denoted by upside down triangle.

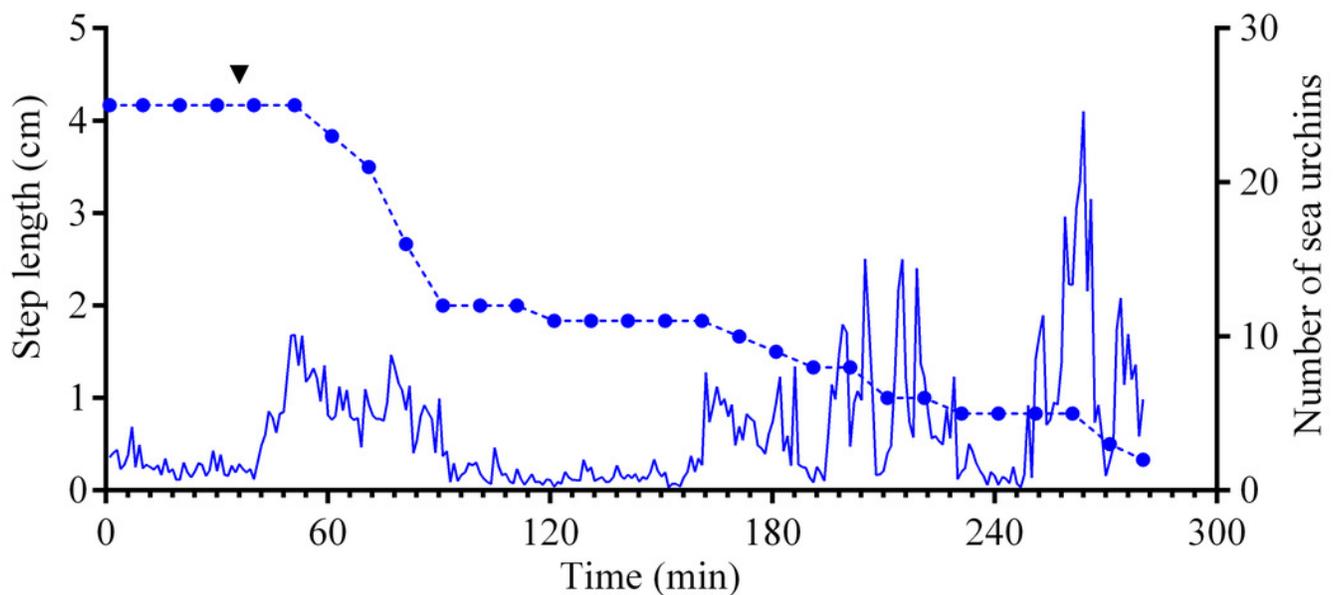


Figure 6

Temporal dynamics of repopulation of the feeders by sea urchins *Mesocentrotus nudus* after their removal in long-term experiments of 2014–2016.

The data of 4 experiments conducted during long-term recordings of 2014–2016 (Fig. 1A–C, Tables S1, S9) are presented as median and range of the number of *M. nudus* per 6 h.

Nonlinear regression is significant ($R^2 = 0.731$, see Table S9 for statistics).

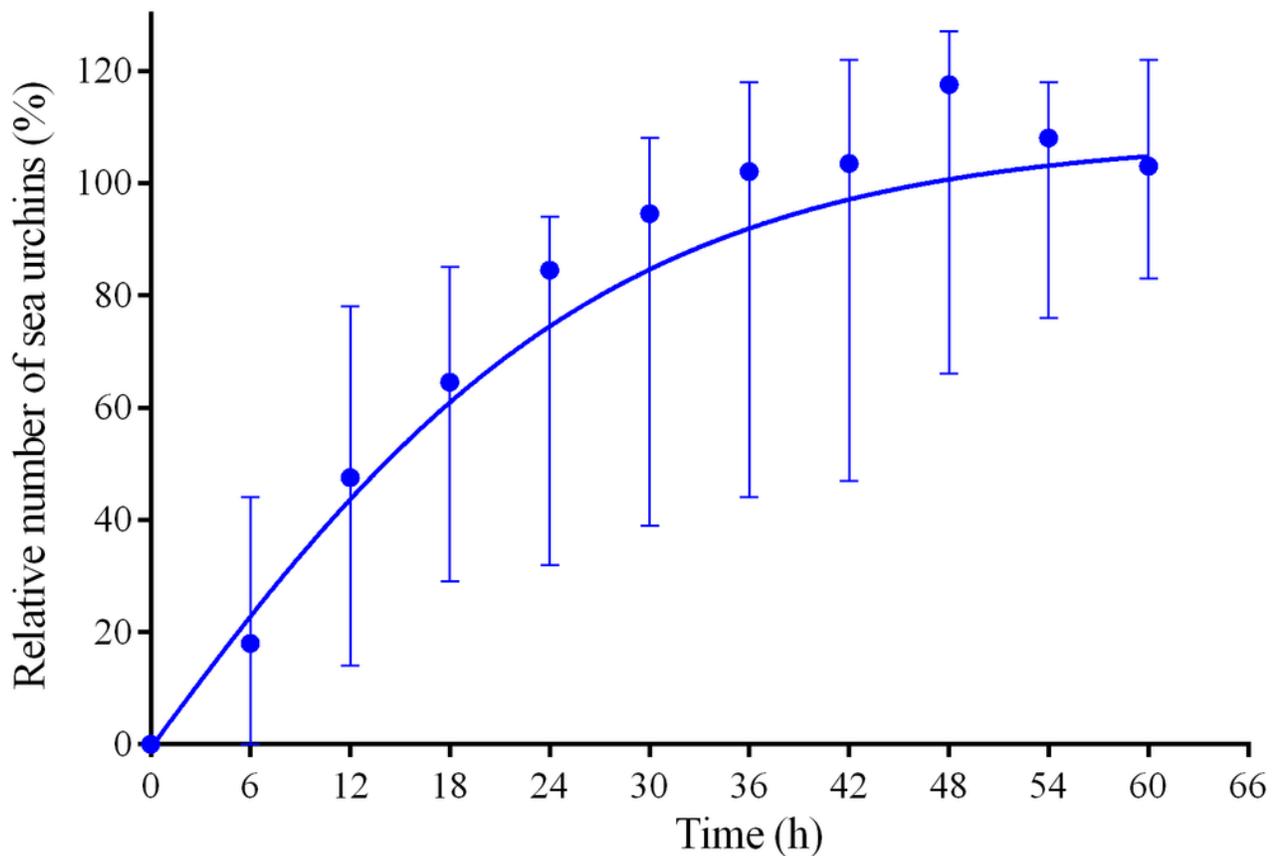


Figure 7

Movement activity of the sea urchins *Mesocentrotus nudus* (blue lines) and *Strongylocentrotus intermedius* (red lines) in response to crushed heterospecifics.

(A) The changes in the average step length of sea urchins (n = 10). (B) The changes in sea urchin distances from the site of simulated attack, mean \pm SD (n = 10). SD is shown for every sixth measurement. Time of the treatment is denoted by upside down triangle.

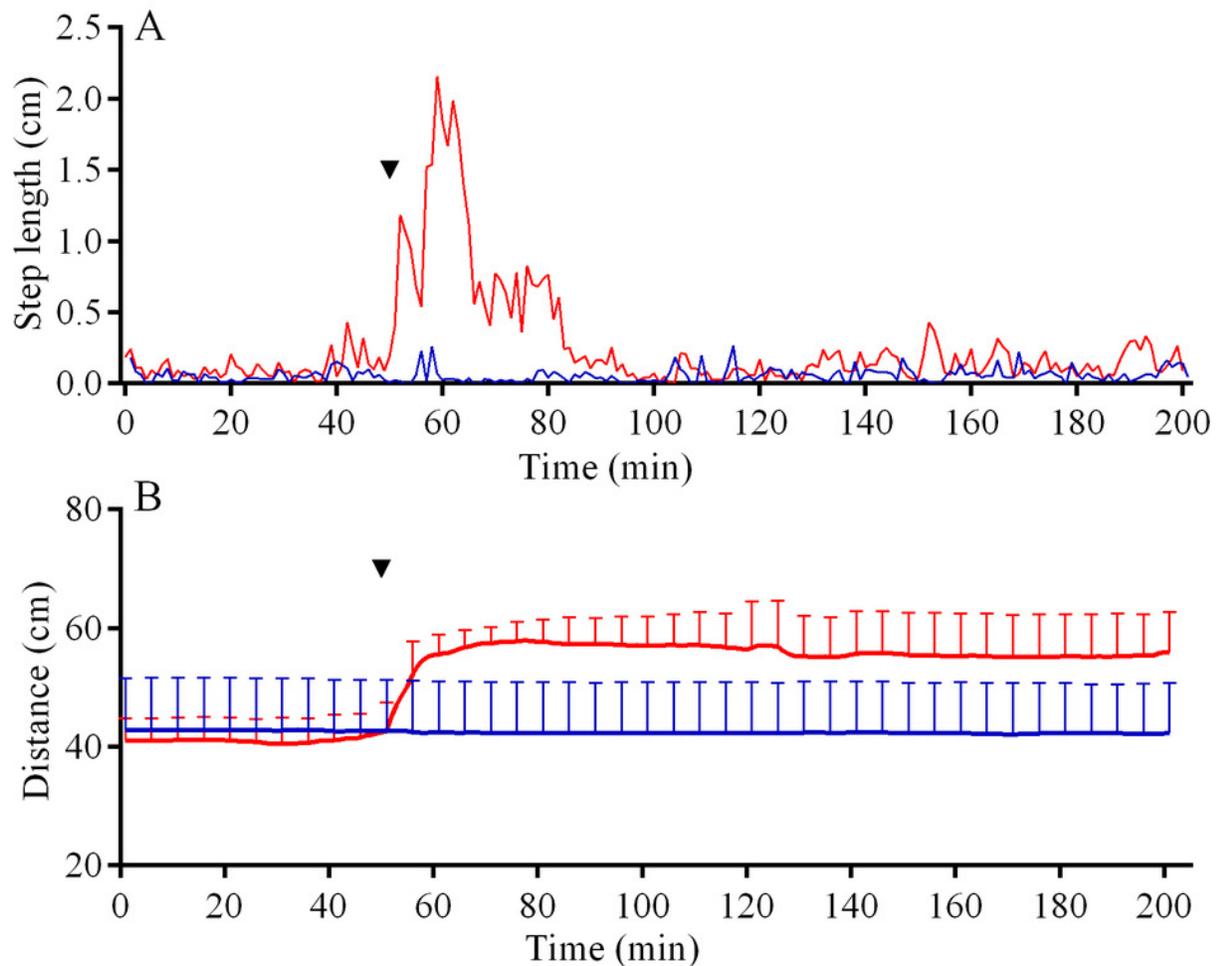


Figure 8

Temporal dynamics of the mean group size of sea urchins *Strongylocentrotus intermedius* in response to simulated predator attack.

(A) Response of *S. intermedius* to crushed conspecifics. (B) Response of *S. intermedius* to crushed specimens of the sea urchin *Mesocentrotus nudus*. Upside down triangles denote the time points when sea urchins were crushed near the feeders. Black solid lines indicate the medians. See Table S14 and Table S17 for raw data and statistics.

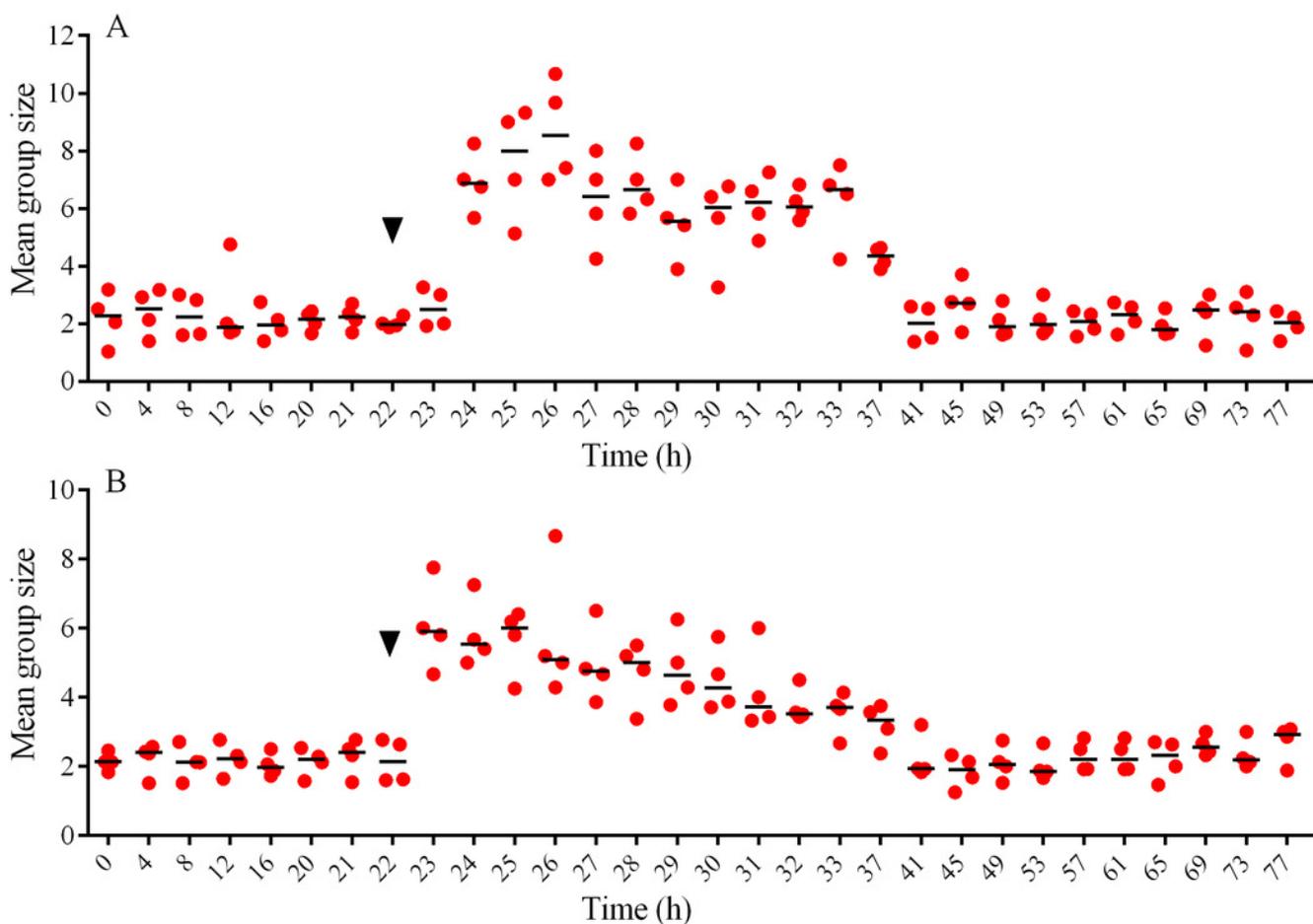


Figure 9

Temporal dynamics of the numbers of sea urchins *Strongylocentrotus intermedius* in response to simulated predator attack.

(A) Response of *S. intermedius* to crushed conspecifics. Linear regression is significant ($P < 0.0001$, see Table S15 for raw data and statistics). (B) response of *S. intermedius* to crushed specimens of the sea urchin *Mesocentrotus nudus*. Linear regression is not significant ($P = 0.0822$, see Table S18 for raw data and statistics). The data are presented as median and range. Time of the treatment is indicated by upside down triangle.

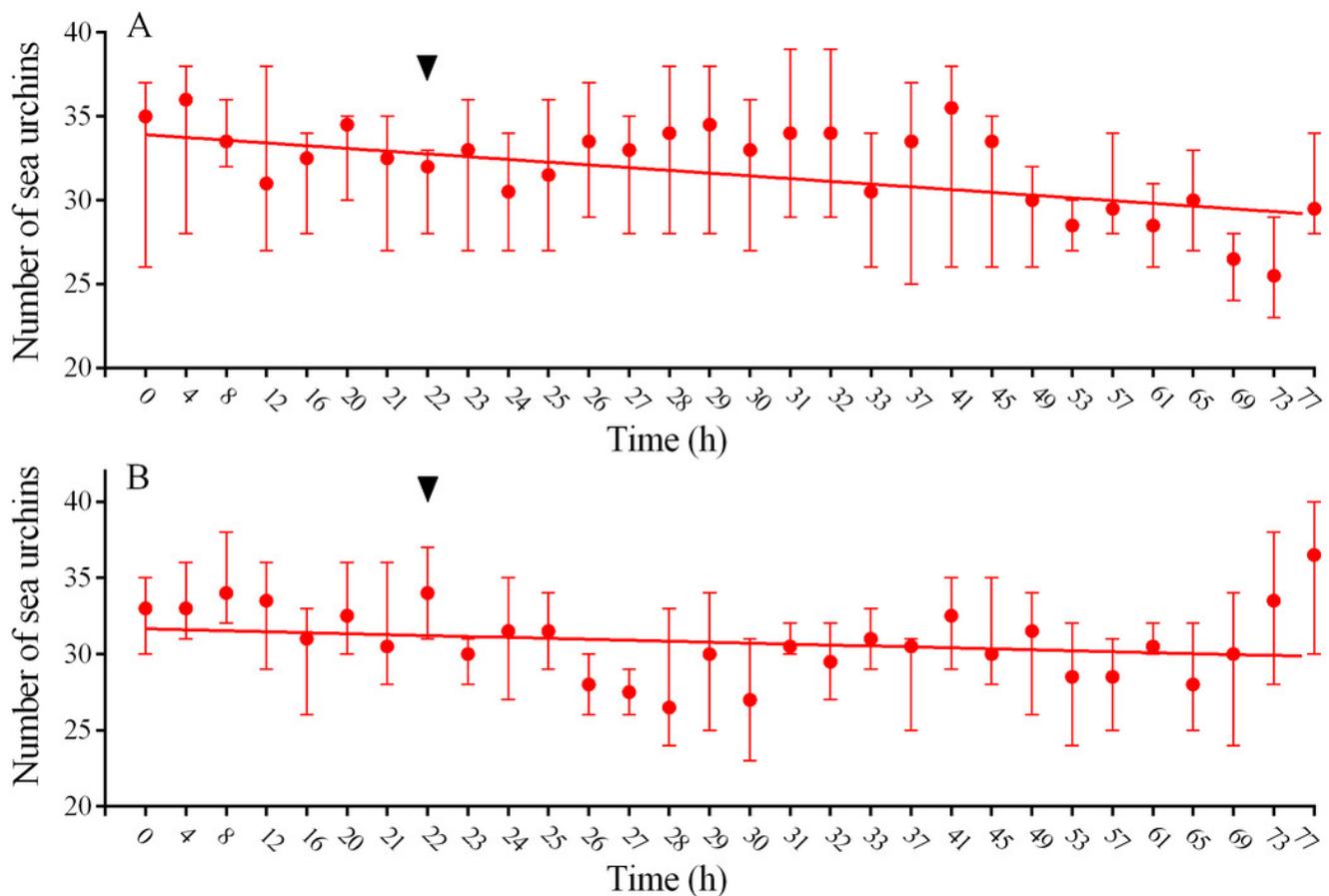


Figure 10

Movement activity of the sea urchins *Strongylocentrotus intermedius* in response to crushed conspecifics.

Red dotted line denotes sea urchin distance from the site of simulated attack, mean \pm SD (n = 10). SD is shown for every ninth measurement. Red solid line denotes the average step length of sea urchins (n = 10). Time of the treatment is indicated by upside down triangle.

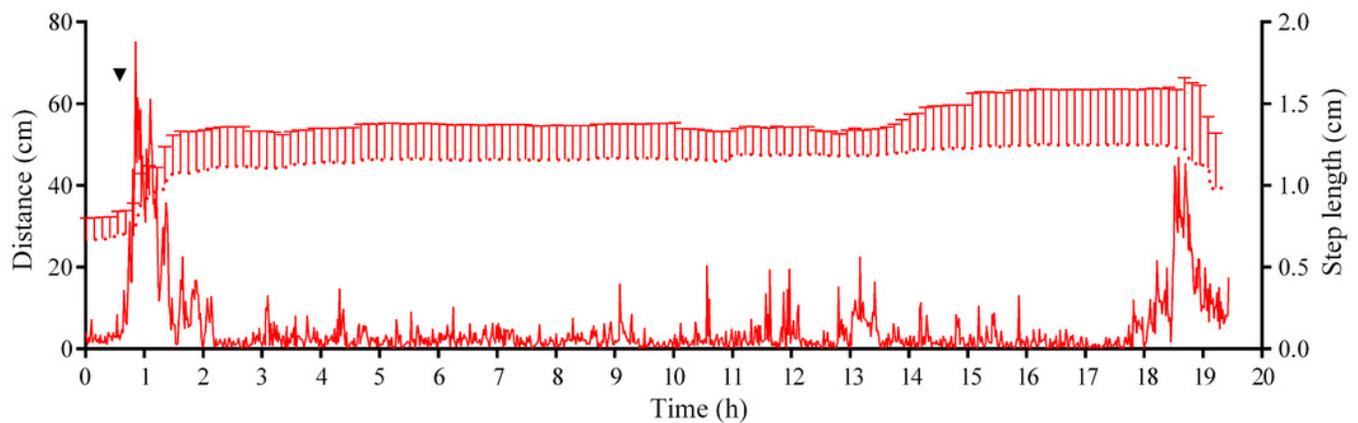


Table 1 (on next page)

Parameters of movement activity of the sea urchin *Mesocentrotus nudus* under calm and stormy weather.

Data are presented as Mean \pm SEM (n = 10) and the range (in the parentheses) for 240-min interval.

1

Parameter	Calm weather	Stormy weather	Statistics
Number of steps	110 ± 12 (50–160)	21 ± 3 (9–35)	$t_{10} = 7.314$; df = 9.883; $p < 0.0001$
Step length, cm	0.71 ± 0.08 (0.10–9.54)	0.29 ± 0.02 (0.10–1.34)	$t_{10} = 4.966$; df = 10.26; $p = 0.0005$
Number of moves	28 ± 2 (22–35)	13 ± 2 (5–23)	$t_{10} = 5.915$; df = 18; $p < 0.0001$
Move length, cm	2.99 ± 0.58 (0.10–56.5)	0.57 ± 0.16 (0.10–8.37)	Mann-Whitney U = 5.0; $p = 0.0002$
Entire distance traversed, cm	76.90 ± 11.86 (34.91–146.40)	5.87 ± 0.7 (3.91–9.97)	$t_{10} = 5.98$; df = 9.063; $p = 0.0002$
Number of stops	28 ± 2 (22–35)	14 ± 2 (5–24)	$t_{10} = 5.773$; df = 17.41; $p < 0.0001$
Stop duration, min	4.82 ± 0.54 (1–43)	19.69 ± 3.02 (1–87)	$t_{10} = 4.845$; df = 9.566; $p = 0.0008$
Entire stop duration, min	130.0 ± 11.87 (80–190)	217.9 ± 2.73 (205–231)	$t_{10} = 7.219$; df = 9.95; $p < 0.0001$
Speed, cm min ⁻¹	0.32 ± 0.05 (0.15–0.61)	0.03 ± 0.003 (0.02–0.04)	$t_{10} = 6.054$; df = 9.056; $p = 0.0002$

2

Table 2 (on next page)

Parameters of movement activity of the sea urchin *Strongylocentrotus intermedius* under calm and stormy weather.

Data are presented as Mean \pm SEM (n = 10) and the range (in the parentheses) for 240-min interval.

1

Parameter	Calm weather	Stormy weather	Statistics
Number of steps	160 ± 10 (110–211)	57 ± 12 (18–141)	$t_{10} = 6.606$; $df = 17.49$; $p < 0.0001$
Step length, cm	0.28 ± 0.03 (0.10–5.46)	0.33 ± 0.08 (0.10–4.14)	Mann-Whitney U = 33.5; $p = 0.2233$
Number of moves	31 ± 2 (16–39)	26 ± 3 (13–39)	$t_{10} = 1.147$; $df = 18$; $p = 0.2662$
Move length, cm	1.65 ± 0.35 (0.10–39.72)	0.65 ± 0.17 (0.10–37.7)	Mann-Whitney U = 13.0; $p = 0.0038$
Entire distance traversed, cm	44.28 ± 5.56 (29.73–88.85)	16.10 ± 4.3 (3.80–45.76)	Mann-Whitney U = 12.0; $p = 0.0029$
Number of stops	31 ± 2 (18–39)	26 ± 3 (12–38)	$t_{10} = 1.331$; $df = 15.56$; $p = 0.2023$
Stop duration, min	2.54 ± 0.26 (1–29)	8.63 ± 1.70 (1–80)	$t_{10} = 3.402$; $df = 9.431$; $p = 0.0058$
Entire stop duration, min	80.1 ± 10.1 (29–130)	183.2 ± 11.97 (99–222)	$t_{10} = 6.592$; $df = 17.49$; $p < 0.0001$
Speed, cm min ⁻¹	0.19 ± 0.02 (0.12–0.37)	0.07 ± 0.02 (0.02–0.19)	Mann-Whitney U = 11.0; $p = 0.0019$

2