Molecular characterization of G-protein-coupled receptor (GPCR) and protein kinase A (PKA) cDNA in *Perinereis aibuhitensis* and expression during benzo(a)pyrene exposure

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Background. G-protein-coupled receptors (GPCRs) are one of the most important molecules that transfer signals across the plasma membrane, and play central roles in physiological systems. The molecular architecture of GPCRs allows them to bind to diverse chemicals, including environmental contaminants.

Methods. To investigate the effects of benzo(a)pyrene (B[a]p) on GPCR signaling, GPCR and the protein kinase A (PKA) catalytic subunit of *Perinereis aibuhitensis* were cloned. The expression patterns of these two genes during B(a)p exposure were determined with real-time fluorescence quantitative PCR.

Results. The full-length cDNAs of PaGPCR and the PaPKA catalytic subunit were 1,514 and 2,662 nucleotides, respectively, encoding 339 and 351 amino acids, respectively. Multiple sequence alignments indicated that the deduced amino acid sequence of PaGPCR shared a low level of similarity with the orphan GPCRs of polychaetes and echinoderms, whereas PaPKA shared a high level of identify with the PKA catalytic subunits of other invertebrates. B(a)p exposure time-dependently elevated the expression of *PaGPCR* and *PaPKA*. The expression of both *PaGPCR* and *PaPKA* was also dose-dependent, except at a dose of 10 μg/L B(a)p.

Discussion. These results suggested that GPCR signaling in *P. aibuhitensis* was involved in the polychaete's response to environmental contaminants.

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Abstract

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Introduction

Benzo(a)pyrene (B(a)P), a kind of polycyclic aromatic hydrocarbon (PAH), can cause genetic damage, immune and endocrine dysfunction, and malformation in humans and other organisms. Its high lipophilicity allows it to absorb to organic matter and other particulate matter and thus accumulate in sediments. Recent increases in offshore oil production and transportation and the sewerage discharge of domestic and industrial wastewater have led to environmental pollution in coastal regions, and B(a)P has been widely detected in sediments around the world, even in China. The levels of B(a)P in the sediments of Dalian Bay vary from 10.5 to 3421.2 ng/g (*Zhang,*

2008), and in the sediments around the drilling platform in the Bohai Sea, the concentration of B(a)p is up to 27.69 ng/g (*Yang et al, 2016*). PAHs such as B(a)p can be absorbed by benthic organisms via ingestion and through their body surfaces, and B(a)p is reported to have serious effects on deposit feeders. Therefore, the toxicity and bioavailability of B(a)p are important factors in the assessment of sediment pollution.

G-protein-coupled receptors (GPCRs) are the largest superfamily of cellmembrance proteins (*Fredriksson et al, 2003*). The molecular architecture of the GPCRs allows them to bind to diverse organic and inorganic molecules. GPCRs mediate cell proliferation and survival by transmitting signals from a range of extracellular ligands across the cell membrane to signaling pathways. In vertebrates, they are key regulators of the innate and adaptive immune responses and have been investigated as potential targets in drug discovery (*Garland, 2013*). However, examples of GPCRs in invertebrates are limited. *Miller et al.* (*2015*) reported that *Caenorhabditis elegans* with mutations in the GPCR follicle-stimulating hormone receptor 1 (FSHR-1) died significantly more quickly in the presence of cadmium than wild-type nematodes, which suggests that this GPCR pathway protects the nematode against cadmium-induced damage. They also found that FSHR-1 antagonizes the capacity of *C. elegans* to resist cold stress, and the mutants lacking *fshr-1* survived better than wild-type worms at low temperatures. *Dong and Zhang* (*2012*) identified a putative GPCR gene, *HP1R*, in the red swamp crayfish Procambarus clarkia, and the expression of HP1R was significantly increased in the presence of Gram-negative bacteria.

Because of the aromatic structures present a number of GPCR ligands, GPCRs are potential targets of aromatic pollutants such as B(a)p *(Ferrec and Ovrevik, 2018)*. *Mayati et al.* (*2012*) reported the interaction between B(a)p and the β_2-adrenergic receptor (β_2ADR) in endothelial HMEC-1 cells and the consequent increase in intracellular Ca^{2+}, which influenced the expression of cytochrome P450 B1. This suggests that β_2ADR, a kind of GPCR, is potentially involved in the deleterious effects of PAHs. *Factor et al.* (*2011*) also observed the reduced expression and function of β_2ADR in airway epithelial cells and smooth muscle cells after their exposure to a mixture of PAHs. This implies that the β_2ADR signal transduction pathway is affected by PAHs. These data indicate that PAHs, including B(a)p, modulate the concentrations of intracytosolic cyclic adenosine monophosphate (cAMP) or Ca^{2+} via G-protein-dependent mechanisms (*Bainy, 2007; Nadal et al., 2000*).

The marine polychaete *Perinereis aibuhitensis* is widely distributed in the mudflats and estuarine sediments that occur widely along the coasts of Southeast Asia. They spend most of their lives within the sediments, ensuring their continuous contact with any sediment-associated contaminants. *Chen et al.* (*2012*) identified a *CYP4* gene of *P. aibuhitensis* and showed that exposure to petroleum hydrocarbons significantly induce the expression of this gene. To clarify whether GPCRs were involved in modulating the toxicity of aromatic pollutants, the full-length GPCR and protein kinase A (PKA) cDNAs were cloned and the expression patterns of these two genes were determined in this study. Our results provide important information on the function of GPCRs in polychaetes.

Materials & Methods

B(a)p exposure

Perinereis aibuhitensis specimens (10–15 cm, 2.0 ± 0.5 g wet weight) were collected from an estuary of Jinzhou Bay in Dalian, China. The animals were transferred to the laboratory and acclimatized in filtered seawater (salinity 31–32, temperature 16 ± 0.5°C) for a week before the experiment. During acclimatization, the *P. aibuhitensis* were fed a powdered mix containing kelp powder, gulfweed powder, fishmeal, yeast, and spirulina. The worms were deprived of food during their exposure to B(a)P.

Based on the standard seawater quality of the People's Republic of China (GB 3097-1997), four B(a)P concentration groups were established: 0.5, 5, 10, and 50 μg/L. A blank control (seawater only) group and an acetone solvent group (100 μL/L) were also established. Three repetitions of each concentration group were set up. Ten worms were randomly placed in 2L beakers containing different concentrations of B(a)P. During the experiment, the temperature of the seawater was 16 ± 0.5°C, and the seawater was renewed every 24h. On days 4, 7, and 14 of the experiment, four individuals were randomly sampled from each concentration group, and the body wall was removed and preserved at -80°C.

Cloning the full-length GPCR and PKA cDNAs of *P. aibuhitensis*

Each sample was ground to powder and the total RNA was extracted with RNAiso™ Plus (TaKaRa, Dalian, China). The quality of the RNA was determined with 1% agarose gel electrophoresis. The RNA (500 ng) was reverse transcribed to cDNA for the rapid amplification of cDNA ends (RACE) using the SMARTer® RACE Kit (Clontech, Palo Alto, CA, USA). The 3′ and 5′ RACE primers were designed with the Primer 5.0 software (PREMIER Biosoft,Palo Alto, CA, USA) according to the confirmed partial sequences of GPCR and PKA obtained from *P. aibuhitensis* transcriptome sequences. The primers used in this study are shown in Table 1.

The 3′ RACE amplification of *P. aibuhitensis GPCR (PaGPCR)* was performed using the 3′ RACE cDNA as the template. The PCR system (50 μL) for *PaGPCR* contained 15.5 μL of PCR-grade water, 25.0 μL of 2× SeqAmp Buffer, 1.0 μL of SeqAmp DNA Polymerase, 2.5 μL of 3′ RACE cDNA, 5.0 μL 10 × UPM (Universal Primer Mixture), and 1.0 μL of primer GPCR-F1 (10 μM). The thermal cycling conditions were: 35 cycles of denaturation at 94°C for 30 s, annealing at 65°C for 30s, and extension at 72°C for 3 min. The 3′ RACE amplification of *PaPKA* was performed with nested PCR. The outer PCR reaction system for *P. aibuhitensis PKA (PaPKA)* was the same as that for *PaGPCR*, except that a specific primer was used. The reaction conditions for the outer PCR were: 35 cycles of denaturation at 94°C for 30 s, annealing at 63.9°C for 30 s, and extension at 72°C for 3 min. The outer PCR product (5.0 μL) was diluted with 245 μL of TE buffer, and 5.0 μL of the diluted product was used as the template for the inner PCR. The reaction conditions and system for the inner PCR were the same as for the outer PCR of *PaPKA*.

The 5′ RACE product of *PaGPCR* was amplified with nested PCR. The outer PCR reaction system (50 μL) for *PaGPCR* contained 15.5 μL of PCR-grade water, 25.0 μL of 2× SeqAmp

Buffer, 1.0 μL of SeqAmp DNA Polymerase, 2.5 μL of 5′ RACE DNA, 5.0 μL of 10 × UPM, and 1.0 μL of primer GPCR-R1 (10 μM). The reaction conditions for the outer PCR were: 35 cycles of denaturation at 94°C for 30 s, annealing at 60°C for 30 s, and extension at 72°C for 3 min. The outer PCR product (5.0 μL) of *PaGPCR* was diluted with 245 μL of TE buffer and 5.0 μL of the diluted product was used as the template for the inner PCR. The reaction system (50 μL) for the inner PCR of *PaGPCR* contained 5.0 μL of the diluted outer PCR product, 17.0 μL of PCR-grade water, 25.0 μL of 2 × SeqAmp Buffer, 1.0 μL of SeqAmp DNA Polymerase, 1.0 μL of UPM Short, and 1.0 μL of primer GPCR-R2 (10 μM). The reaction conditions were: 20 cycles of denaturation at 94°C for 30 s, annealing at 60°C for 30 s, and extension at 72°C for 3 min. The 5′ RACE product of PaPKA was amplified with ordinary PCR, and the reaction system and conditions were the same as those for PaGPCR.

The PCR products were detected with 1% agarose gel electrophoresis and purified with the Agarose Gel DNA Purification Kit (Tiangen, Beijing, China), according to the manufacturer's instructions. The PCR products were sequenced by TaKaRa Biotechnology Co. Ltd.

Bioinformatic analysis of PaGPCR and PaPKA

The amino acid sequences of PaGPCR and PaPKA were deduced with the Expert Protein Analysis System (http://www.us.expasy.org/tools). The conserved domain in each amino acid sequence was analyzed with the Motif Scan (http://www.hits.isbsib.ch/cgi-bin/PESCAN) and Expasy (http://www.au.expasy.org/prosite/). The protein localization sites in the cell were predicted with the Psort software (http://psort.hgc.jp/form2.html). The transmembrane (TM) helices in the protein were predicted with the TMHMM software (http://www.cbs.dtu.dk/services/TMHMM/). The tertiary structures of GPCR and PKA were predicted with the Swiss-Model software (http://swissmodel.expasy.org/interactive). Multiple sequences were aligned with the Clustal W software (http://www.ebi.ac.uk/clustalW). Phylogenetic analyses of GPCR and PKA were performed in MEGA 5.0. The tree topologies were evaluated with bootstrapping, using 1,000 replicates.

Expression of *PaGPCR* and *PaPKA* genes during B(a)p exposure

Real-time fluorescence quantitative PCR was used to investigate the expression of the two genes in *P. aibuhitensis* during B(a)p exposure. The β-actin gene was used as the reference gene, according to our previous study (*Li et al., 2018*). The primer information is shown in Table 1. Amplification was performed in 20 μL reactions containing 10 μL of SYBR Premix Ex Taq II (Tli RNaseH Plus)(TaKaRa, Dalian, China), 0.8 μL of each primer (10 μM), 0.4 μL of 50× ROX Reference Dye II, 2.0 μL of cDNA, and 6.0 μL of H_2O. The reaction conditions were: 95°C for 30 s, then 40 cycles of 95°C for 5 s and 60°C for 34 s. The melting curves were analyzed after the real-time quantitative analyses. The standard curves were tested with serial 10-fold sample dilutions. The slopes of the standard curves and the PCR efficiency were calculated to confirm the accuracy of the real-time PCR data.

Statistical analysis

The relative quantitative ($2^{-\Delta\Delta Ct}$) method was used to analyze the expression of the *GPCR* and *PKA* genes. The data are expressed as means ± standard deviations (SD), and one-way analysis

of variance was used to analyze the significance of the differences among the different concentration groups at each sampling point, with the SPSS 19.0 software. P values ≤ 0.05 were considered statistically significant.

Results

Molecular characterization of *PaGPCR*

The full-length cDNA of *PaGPCR* was 1,514 bp and included a 5′ untranslated region (UTR) of 213 bp, a 3′ UTR of 284 bp, and an open reading frame (ORF) of 1,017 bp, encoding 338 amino acids with a predicted molecular weight of 38.799 kDa and a theoretical isoelectric point of 9.38 (Figure 1). This nucleotide sequence was deposited in the GenBank database under accession number KX792261. The seven-transmembrane (7TM)-helix bundle (304–1,146 bp) that defines the GPCR protein family was present in PaGPCR. The glutamic acid/aspartic acid-arginine-tyrosine (E/DRY) motif (amino acids 124–126) at the border between TM III and intracellular loop 2 and the NPXXY motif (amino acids 298–302) of TM VII near the inner cell membrance were detected in the deduced protein sequence, indicating that the protein sequence belonged to the rhodopsin family. In an amino acid comparison, PaGPCR shared 33% similarity with the orphan GPCR of *Platynereis dumerilii* and 30%–33% similarity with galanin receptor type 2 of echinoderms (Figure 2).

The predicted cellular localization of the PaGPCR protein showed it mostly located on the cell membrane (52.2%), and seven TM helices were predicted in the deduced protein sequence (Figure 3). The three-dimensional structural analysis of PaGPCR showed that it contained seven α-helices, similar to the GPCRs of other animals (Figure 4). Its three-dimensional structure and protein localization confirmed that this protein sequence was a GPCR.

Molecular characterization of PaPKA

The total length of *P. aibuhitensis* PKA cDNA was 2662 bp, containing a 3′ UTR of 1,483 bp, a 5′ UTR of 126 bp, and an ORF of 1,053 bp encoding 350 amino acids (Figure 5). The predicted molecular weight of PaPKA was 40.28 kDa and its theoretical isoelectric point was 8.35. The nucleotide sequence was deposited in GenBank under accession number KX839259. A glycine-rich loop GTGSFGRV (amino acids 50–57), Ser/Thr active site RDLKPEN (amino acids 165–171), PKA-regulatory-subunit-binding site LCGTPEY (amino acids 198–204), DFG triplet (Asp–Phe–Gly) for orienting the γ-phosphates of adenosine triphosphate (ATP) for transfer, APE motif (Ala-Pro-Glu) to stabilize the structure of the large lobe of PKA, and conserved phosphorylation site (Thr197) were detected in this deduced amino acid sequence. The presence of these conserved regions indicated that PaPKA was the catalytic subunit of PKA. An amino acid comparison indicated that PaPKA was highly similar to other PKA catalytic subunits (Figure 6).

The predicted location of PaPKA in the cell was predominantly in the cytoplasm (47.8%). The three-dimensional structural analysis of PaPKA showed that it folded into a two-lobed structure (Figure 7). The small lobe had a predominantly β-sheet structure, which was responsible for anchoring and orienting the nucleotide, and the large lobe had a predominantly α-helix structure,

and was primarily involved in binding the peptide substrate and initiating phosphotransfer (*Hanks and Hunter, 1995*). Ser53, Phe54, and Gly55 formed hydrogen bonds with ATP β-phosphate oxygens, and Leu49 and Val57 formed a hydrophobic pocket enclosing the adenine ring of ATP.

Phylogenetic analysis of PaGPCR and PaPKA

Phylogenetic trees were constructed from the amino acid sequences of GPCR and PKA, (Figure 8 and Figure 9, respectively). Figure 8 indicates that PaGPCR shared great identity with the orphan GPCRs of other polychaetes. Figure 9 shows that PaPKA shared identity with mollusk PKAs, which clustered together on a single branch.

Effects of B(a)P on *PaGPCR* and *PaPKA* expression in *P. aibuhitensis*

Figure 10A shows the expression of the *PaGPCR* gene of *P. aibuhitensis* during B(a)p exposure. There was no difference in its expression between the acetone control group and the blank control group, indicating that acetone as a solvent had no toxic effect on the nematodes. During exposure to B(a)p, the expression of the *PaGPCR* gene increased both time- and approximately dose-dependently. On day 4, *PaGPCR* expression was significantly upregulated ($P < 0.05$) in all but the 0.5 μg/L B(a)P group. The expression of *PaGPCR* in the 5, 10, and 50 μg/L B(a)P groups was 2.32-, 3.46-, and 3.15-fold higher than in the control group, respectively. On day 7, the expression of *PaGPCR* in the 0.5, 5, 10, and 50 μg/L B(a)P groups was 3.10-, 2.91-, 3.59-, and 3.28-fold higher than in the control group, respectively ($P < 0.05$). The expression of *PaGPCR* in each concentration group reached its highest level on day 14, at 4.30-, 6.60-, 4.79-, and 6.36-fold higher than the control group, respectively ($P < 0.01$).

The expression pattern of the *PaPKA* gene during B(a)P exposure was the same as that of *PaGPCR* (Figure 10B). The expression of *PaPKA* increased as the time of exposure increased. On day 4, the expression of *PaPKA* was slightly higher in all but the 0.5 μg/L B(a)p concentration group, at 1.79-, 1.21-, and 2.21-fold higher in the 5, 10, and 50 μg/L B(a)P groups, respectively, than in the control group. The expression of *PaPKA* in each concentration group was higher on day 7 than on day 4, at 1.25-, 1.90-, 2.52-, and 2.89-fold higher in the 0.5, 5, 10, and 50 μg/L B(a)P groups, respectively, than in the control group ($P < 0.05$). On day 14, the expression of *PaPKA* reached its highest level in each concentration group, at 3.19-, 5.03-, 3.10-, and 5.02-fold higher than the control group, respectively ($P < 0.05$).

Discussion

To investigate the reaction of GPCR to B(a)p exposure in *P. aibuhitensis*, the full-length cDNAs of the *PaGPC*R and *PaPKA* were isolated and characterized in *P. aibuhitensis* for the first time. The sequence of *PaGPCR* contained 1514 bp, encoding 338 amino acids. The deduced protein sequence of PaGPCR contained a 7TM helix bundle domain, flanked by the extracellular N-terminal region and the intracellular C-terminus. As part of the functional mechanism of GPCR, the E/DRY motif (amino acids 124–126), which plays an important role in regulating the conformational state of GPCR, occurred at the border between TM III and intracellular loop 2 in this sequence. The protein sequence of PaGPCR also contained the NPXXY motif (amino acids 298–302) in TM VII, which confirmed that it belonged to the rhodopsin family, the largest of the

five families involved in many signaling processes (*Fredriksson et al., 2003*). A multiple protein sequence alignment showed that PaGPCR shared almost 33% homology with the galanin receptor of echinoderms. A phylogenetic analysis showed that it clustered most closely with the orphan GPCRs of other polychaetes, and the galanin receptor of echinoderms. It is well-known that proteins with similar sequences often display comparable functions if the sequence identify exceeds 30% (*Kakarala and Jamil, 2014*). However, a prediction based on sequence identity alone may not be reliable. Therefore, further study is needed to prove the function of PaGPCR. In contrast to the low sequence identify of GPCR, PKA in *P. aibuhitensis* shared high sequence identify with the PKA catalytic subunits of other species. *PaPKA* contained 2,662 bp, which encoded 350 amino acids. The deduced amino acid sequence of PaPKA contained all the conserved domains that were necessary for kinase activity, such as the conserved Thr in the activation loop, the ATP-binding site (GTGSFGRV), the serine/threonine kinase active site (RDLKPEN), and the PKA-regulatory-subunit-binding site (LCGTPEY). The highly conserved amino acids at the ATP-binding site played important roles in ATP binding and phosphotransfer. The high homology among the PKA catalytic subunits suggested that they have a conserved role in intracellular signaling in both vertebrates and invertebrates.

GPCRs comprise the largest and most important family of cell-surface proteins, transmitting signals from extracellular ligands. In vertebrates, they are key regulators of the innate and adaptive immune responses and have been used as potential targets in drug discovery (*Garland, 2013*). However, they have been inadequately investigated in invertebrates. *Miller et al.* (*2015*) reported that FSHR-1 mutants of *C. elegans* died significantly more quickly during cadmium exposure than wild-type nematodes, which suggests that the GPCR pathway protects *C. elegans* against pollutant damage. *Dong and Zhang* (*2012*) reported that Gram-negative bacterial infection induced the expression of the *HP1R* gene in *Procambarus clarkia*. In the present study, we found that B(a)p exposure induced the expression of *PaGPCR*, which increased with time in each concentration group. This result was similar to reports in other invertebrates, and implied that GPCR played an important role in reducing the deleterious effects of B(a)p in *P. aibuhitensis.* GPCRs interact with diverse chemical structures, which increases cAMP production, which then stimulates phospholipase C activity and the subsequent mobilization of Ca^{2+}. *Mayati et al.* (*2012*) observed an interaction between B(a)p and β_2ADR in endothelial HMEC-1 cells, which altered the levels of intracellular Ca^{2+} and the expression of cytochrome P450 B1. *Factor et al.* (*2011*) observed the reduced expression and function of β_2ADR in airway epithelial cells and smooth muscle cells after their exposure to a mixture of PAHs. In the present study, we also observed that the expression of *PaPKA* in *P. aibuhitensis*, was higher during B(a)p exposure than the control level in all but the 0.5 μg/L B(a)p concentration group. The expression of *PaPKA* was significantly and exposure-time-dependently induced by 50 μg/L B(a)p. The increased expression of *PaPKA* and *PaGPCR* suggested that the GPCR signal transduction pathway in *P. aibuhitensis* was affected by PAHs.

B(a)p can be metabolized by organisms through a series of enzymatic and nonenzymatic reactions. Typically, B(a)p is metabolized by the phase I enzyme cytochrome P450 (CYP) and

phase II enzymes such as glutathione S-transferase (GST). The induction of CYP gene expression has been detected in *P. aibuhitensis* (*Chen et al., 2012; Zhao et al., 2014*). In vertebrates, the induction of the CYP enzymes involved in the biotransformation of PAHs is mediated by the aryl hydrocarbon receptor (AhR) pathway. Both α- and β-type AhR proteins have been reported in bivalves (*Fabbri and Capuzzo, 2010*). However, no AhR homologues have been identified in other invertebrates, including marine polychaetes (*Jorgensen et al., 2008*). *Ferrec and Ovrevik* (*2018*) reported that a number of GPCR ligands contain aromatic structures, and that B(a)p modulates the concentration of intracytosolic cAMP through the GPCR pathway without the involvement of conventional nuclear receptors. In this study, we demonstrated the induction of *GPCR* and *PKA* expression during B(a)p exposure, so we hypothesized that the GPCR pathway is also involved in the biotransformation of PAHs in *P. aibuhitensis*. Further study of the relationship between GPCR and CYP expression is required to test our hypothesis.

Conclusions

GPCR represents a critical point of contact between cells and their surrounding environments. This is the first study in which *P. aibuhitensis* GPCR and PKA cDNAs have been cloned. We have also demonstrated that the expression of *GPCR* and *PKA* was induced in *P. aibuhitensis* by B(a)p exposure, and that their expression was affected, to some extent, by the B(a)p concentration and the exposure time. These results should be useful in investigating the biotransformation of PAHs by marine polychaetes.

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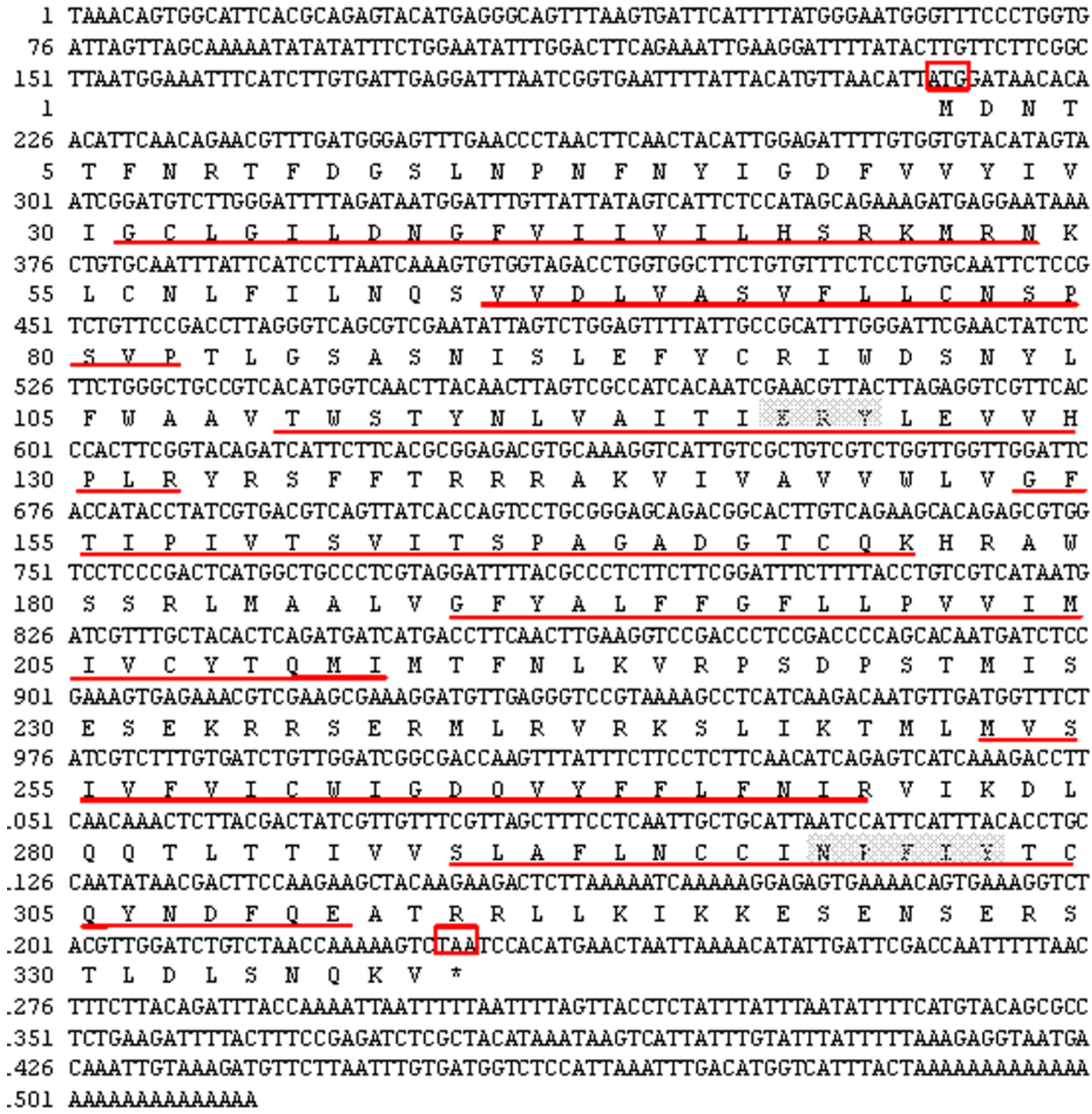
Figure 1

Nucleotide sequence and deduced amino acid sequence of GnRHR from *perinereis aibuhitensis.*

Initiation codon and termination codon are included in the boxes;Terminator is showed as *;

The approximate locations of predicted transmembrane domains 1 through 7 are noted.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151 TTAATGGAAATTTCATCTTGTGATTGAGGATTTAATCGGTGAATTTTATTACATGTTAACATTATGGATAACACA
1 M D N T
226 ACATTCAACAGAACGTTTGATGGGAGTTTGAACCCTAACTTCAACTACATTGGAGATTTTGTGGTGTACATAGTA
5 T F N R T F D G S L N P N F N Y I G D F V V Y I V
301 ATCGGATGTCTTGGGATTTTAGATAATGGATTTGTTATTATAGTCATTCTCCATAGCAGAAAGATGAGGAATAAA
30 I G C L G I L D N G F V I I V I L H S R K M R N K
376 CTGTGCAATTTATTCATCCTTAATCAAAGTGTGGTAGACCTGGTGGCTTCTGTGTTTCTCCTGTGCAATTCTCCG
55 L C N L F I L N Q S V V D L V A S V F L L C N S P
451 TCTGTTCCGACCTTAGGGTCAGCGTCGAATATTAGTCTGGAGTTTTATTGCCGCATTTGGGATTCGAACTATCTC
80 S V P T L G S A S N I S L E F Y C R I W D S N Y L
526 TTCTGGGCTGCCGTCACATGGTCAACTTACAACTTAGTCGCCATCACAATCGAACGTTACTTAGAGGTCGTTCAC
105 F W A A V T W S T Y N L V A I T I E R Y L E V V H
601 CCACTTCGGTACAGATCATTCTTCACGCGGAGACGTGCAAAGGTCATTGTCGCTGTCGTCTGGTTGGTTGGATTC
130 P L R Y R S F F T R R R A K V I V A V V W L V G F
676 ACCATACCTATCGTGACGTCAGTTATCACCAGTCCTGCGGGAGCAGACGGCACTTGTCAGAAGCACAGAGCGTGG
155 T I P I V T S V I T S P A G A D G T C Q K H R A W
751 TCCTCCCGACTCATGGCTGCCCTCGTAGGATTTTACGCCCTCTTCTTCGGATTTCTTTTACCTGTCGTCATAATG
180 S S R L M A A L V G F Y A L F F G F L L P V V I M
826 ATCGTTTGCTACACTCAGATGATCATGACCTTCAACTTGAAGGTCCGACCCTCCGACCCCAGCACAATGATCTCC
205 I V C Y T Q M I M T F N L K V R P S D P S T M I S
901 GAAAGTGAGAAACGTCGAAGCGAAAGGATGTTGAGGGTCCGTAAAAGCCTCATCAAGACAATGTTGATGGTTTCT
230 E S E K R R S E R M L R V R K S L I K T M L M V S
976 ATCGTCTTTGTGATCTGTTGGATCGGCGACCAAGTTTATTTCTTCCTCTTCAACATCAGAGTCATCAAAGACCTT
255 I V F V I C W I G D Q V Y F F L F N I R V I K D L
.051 CAACAAACTCTTACGACTATCGTTGTTTCGTTAGCTTTCCTCAATTGCTGCATTAATCCATTCATTTACACCTGC
280 Q Q T L T T I V V S L A F L N C C I N F F I Y T C
.126 CAATATAACGACTTCCAAGAAGCTACAAGAAGACTCTTAAAAATCAAAAAGGAGAGTGAAAACAGTGAAAGGTCT
305 Q Y N D F Q E A T R R L L K I K K E S E N S E R S
.201 ACGTTGGATCTGTCTAACCAAAAAGTCTAATCCACATGAACTAATTAAAACATATTGATTCGACCAATTTTTAAC
330 T L D L S N Q K V *
.276 TTTCTTACAGATTTACCAAAATTAATTTTTAATTTTAGTTACCTCTATTTATTTAATATTTTCATGTACAGCGCC
.351 TCTGAAGATTTTACTTTCCGAGATCTCGCTACATAAATAAGTCATTATTTGTATTTATTTTTAAAGAGGTAATGA
.426 CAAATTGTAAAGATGTTCTTAATTTGTGATGGTCTCCATTAAATTTGACATGGTCATTTACTAAAAAAAAAAAAA
.501 AAAAAAAAAAAAA

Figure 2

Multiple alignment analysis of PaGPCR with other GPCR protein.

The identical amino acid are shaded. The GenBank accession number for these proteins are as following: *(A) Platynereis dumerilii*,AKQ63061.1. *(B) Strongylocentrotus purpuratus*, XP_003727596.1. *(C) Acanthaster planci*,XP022098630.1. *(D) Apostichopus japonicus*,PIK48567.1.

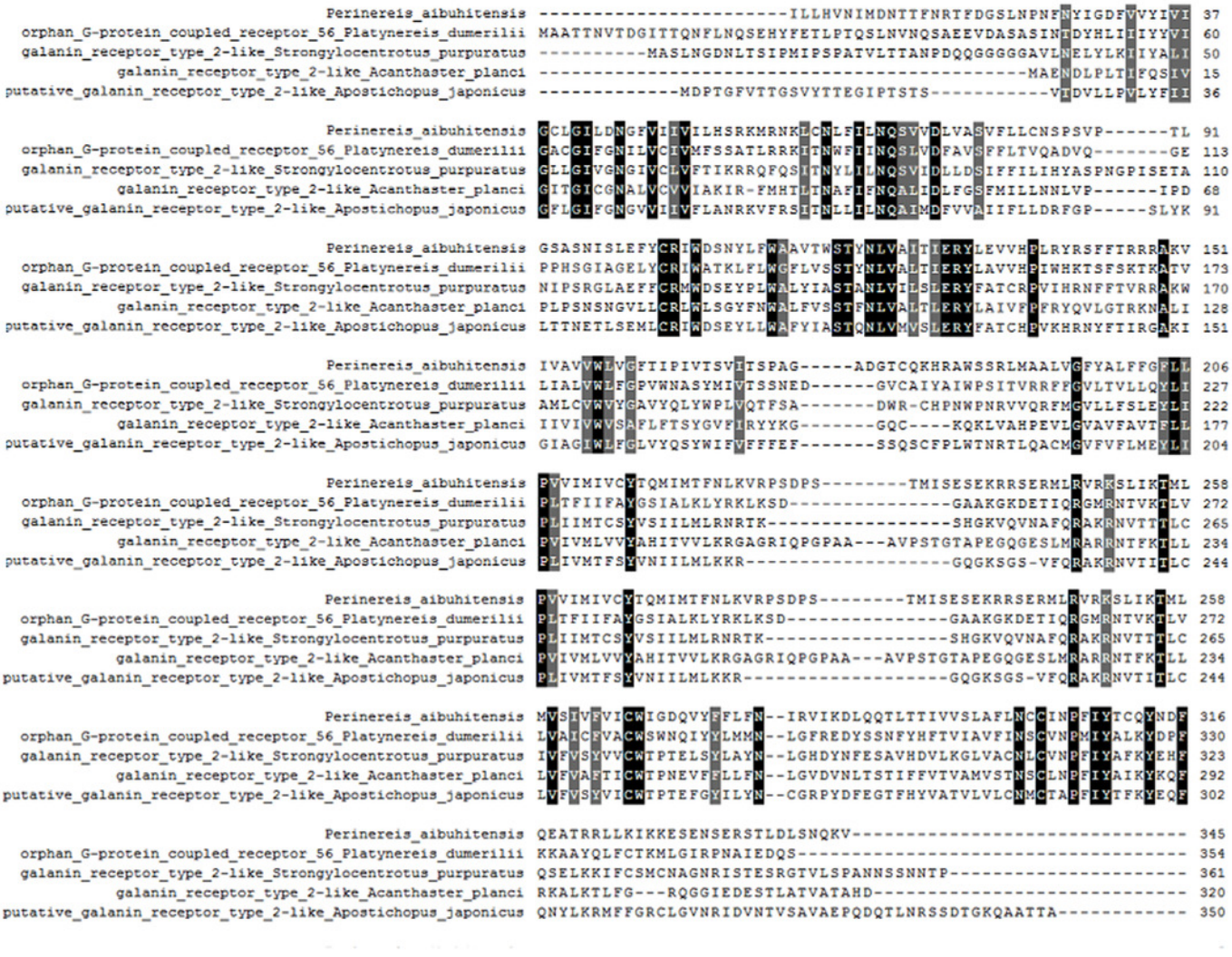

Figure 3

Analysis of transmembrane region of PaGPCR

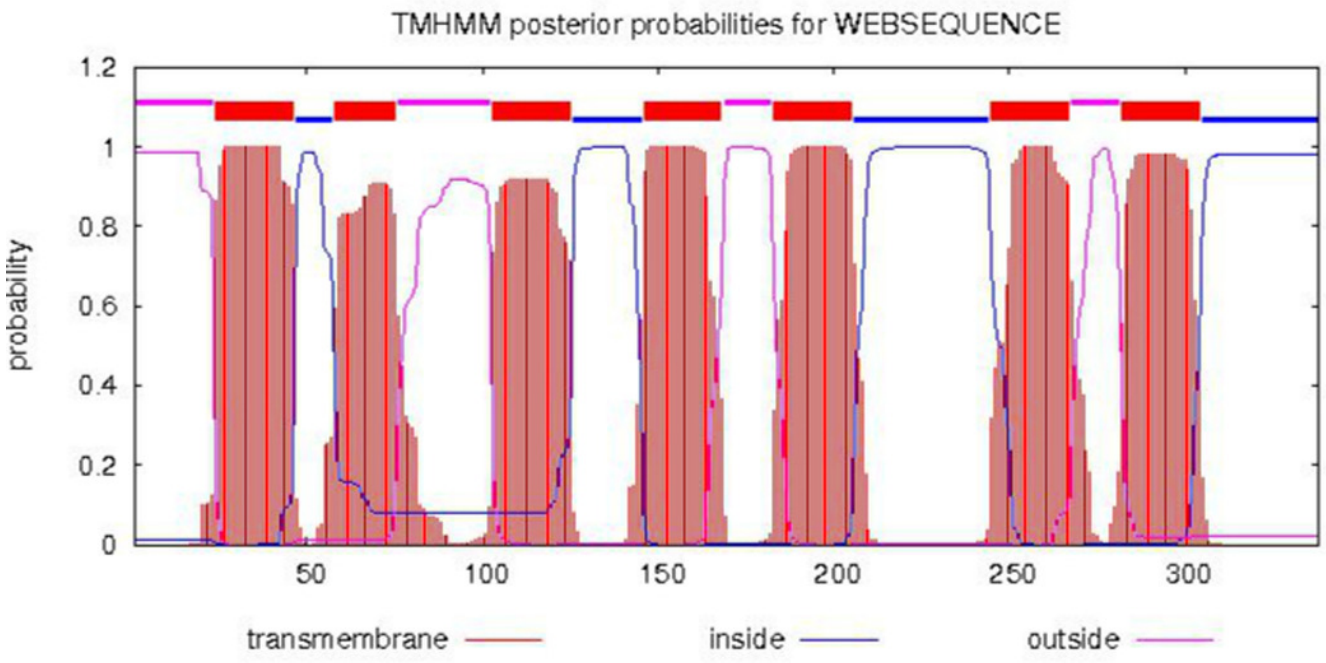

Figure 4

The three dimensional structure of PaGPCR

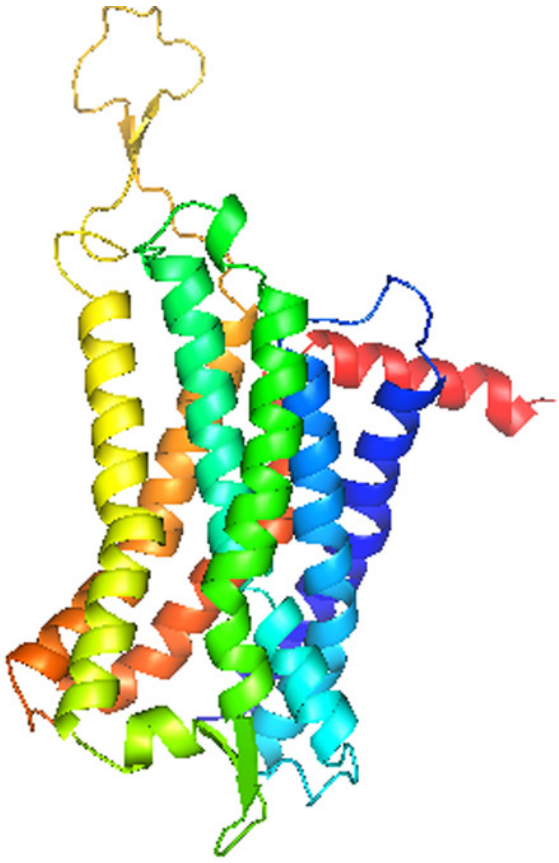

Figure 5

the full cDNA sequence and deduced amino acid sequence of PaPKA.

Red fonts show initiation codon and termination codon; Terminator is showed as*; The red bars under the sequences indicated the ATP active site, Ser /Thr active site and PKA regulatory subunit binding site was underlined with red line, conservative phosphorylation site is boxed in green.

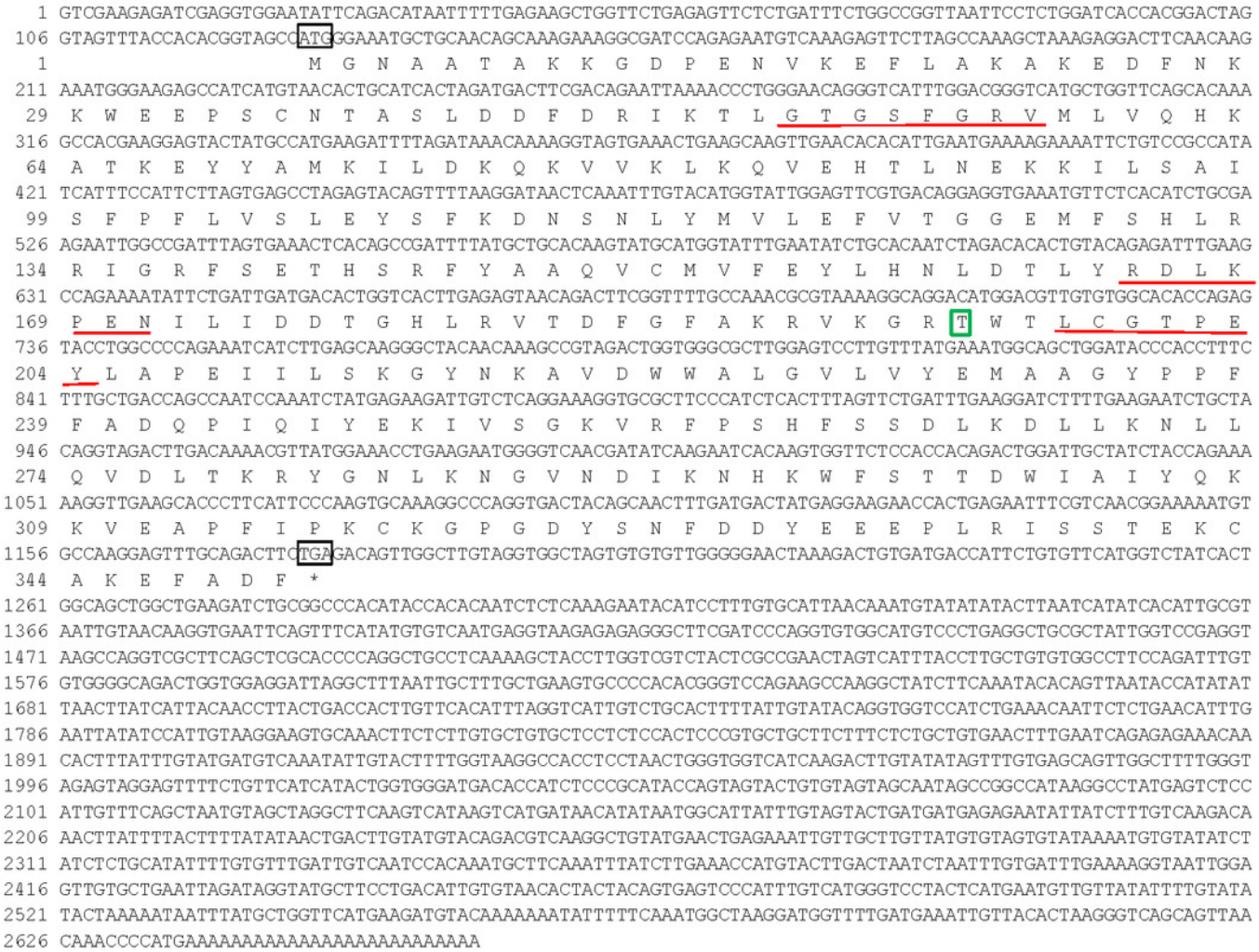

Figure 6

Multiple alignment of PaPKA with other PKA.

The identical amino acid are shaded. The GenBank accession number for these proteins are as following: *(A) Aplysia californica,*NP_001191420.1. *(B) Xenopus tropicalis,* NP_001164667.1. (C) *Branchiostoma floridae,* XP_002600447.1.*(D) Danio rerio,* NP_001030148.1. *(E) Octopus bimaculoides,* XP_014777153.1. *(F) Lingula anatina* ,XP_013409439.1.*(G) Crassostrea gigas,* XP_011439335.1. *(H) Biomphalaria glabrata,* XP_013072294.1. *(I) Salmo salar,* XP_014071121.1. *(J) Gallus gallus,* XP_015146370.1.

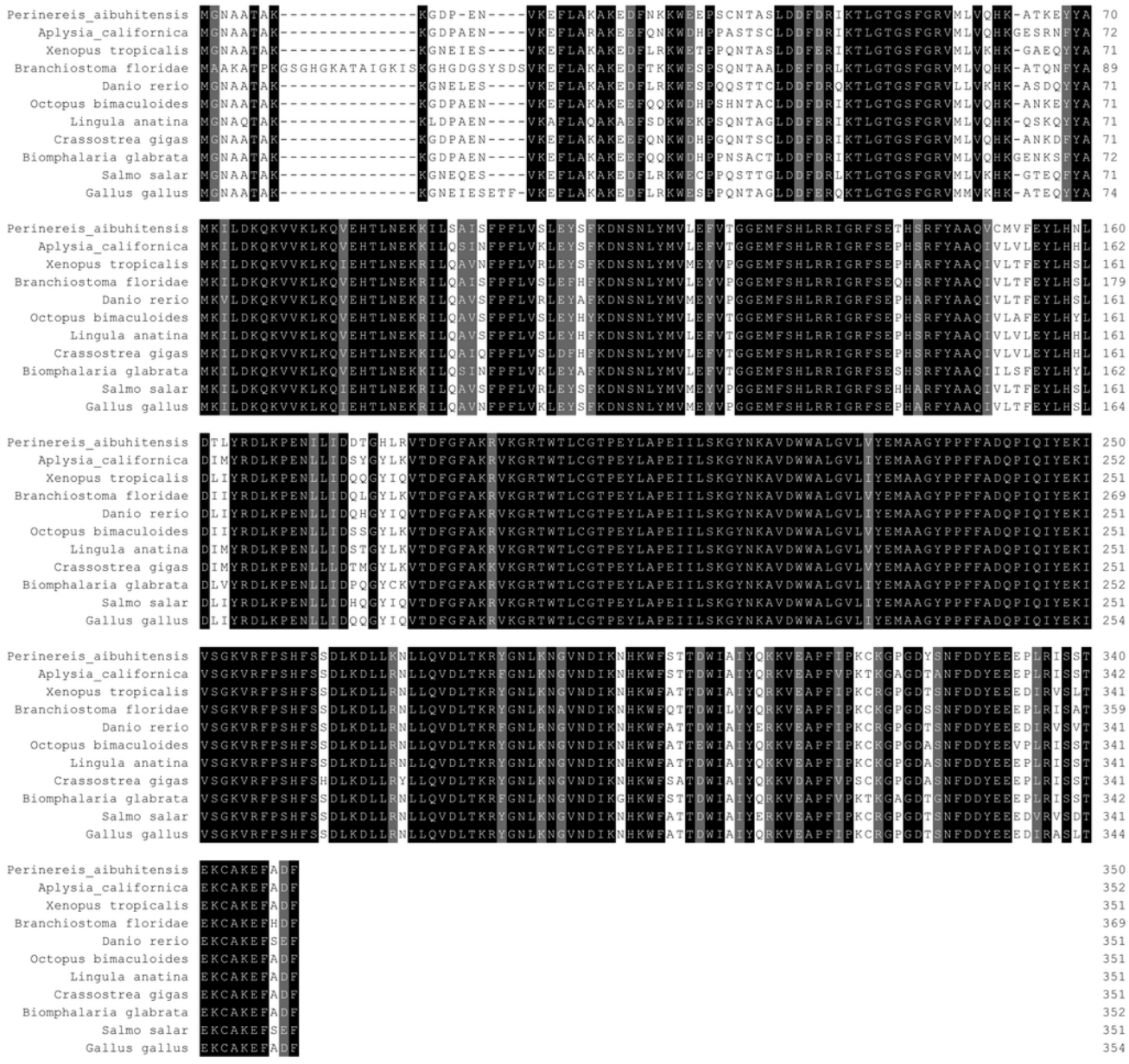
Perinereis_aibuhitensis
Aplysia_californica
Xenopus tropicalis
Branchiostoma floridae
Danio rerio
Octopus bimaculoides
Lingula anatina
Crassostrea gigas
Biomphalaria glabrata
Salmo salar
Gallus gallus

Figure 7

The three dimensional structure of PKA from *P. aibuhitensis*

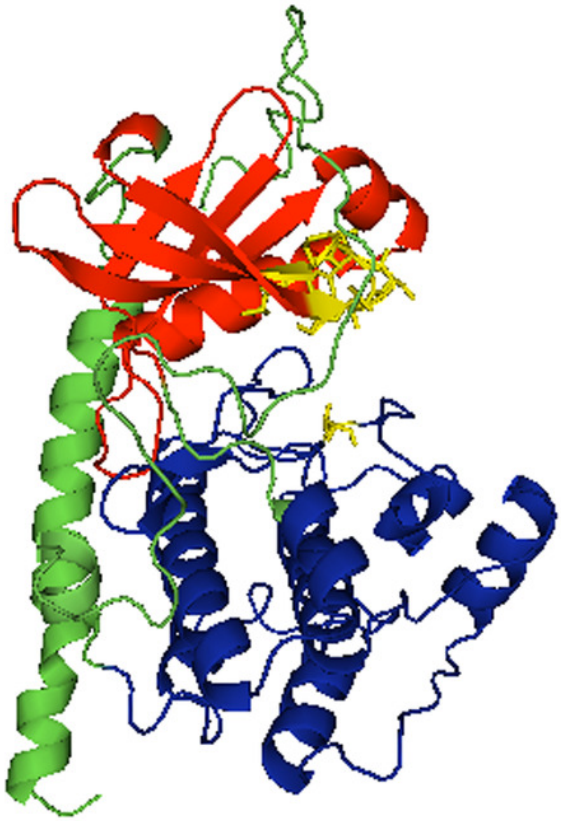

Figure 8

Molecular phylogenetic analysis of PaGPCR related to GPCR of other invertebrates and vertebrates

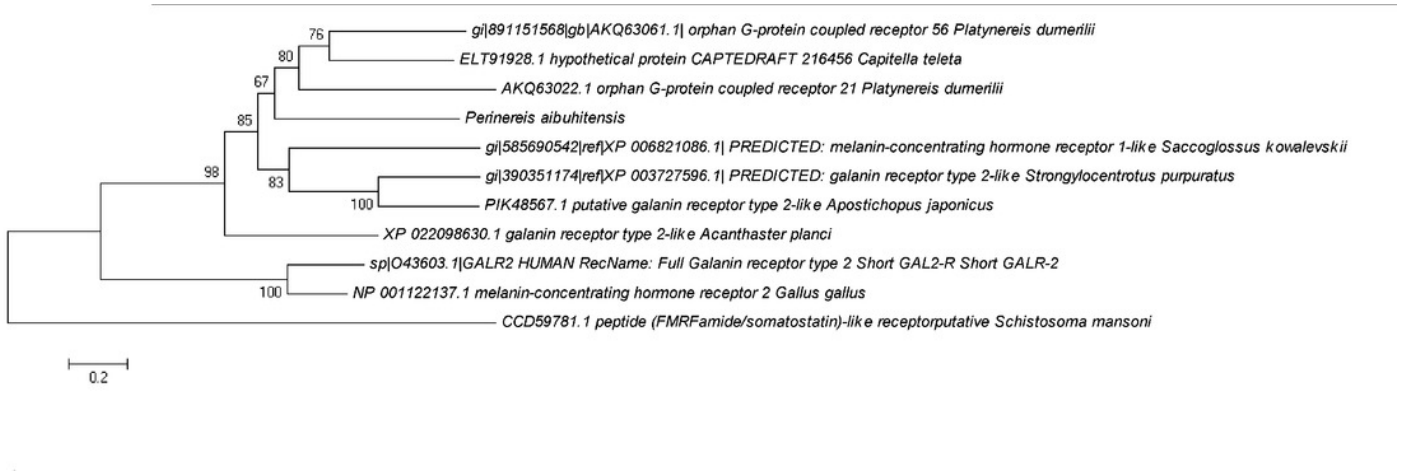

Figure 9

Molecular phylogenetic analysis of PaPKA related to PKA of other invertebrates and vertebrates

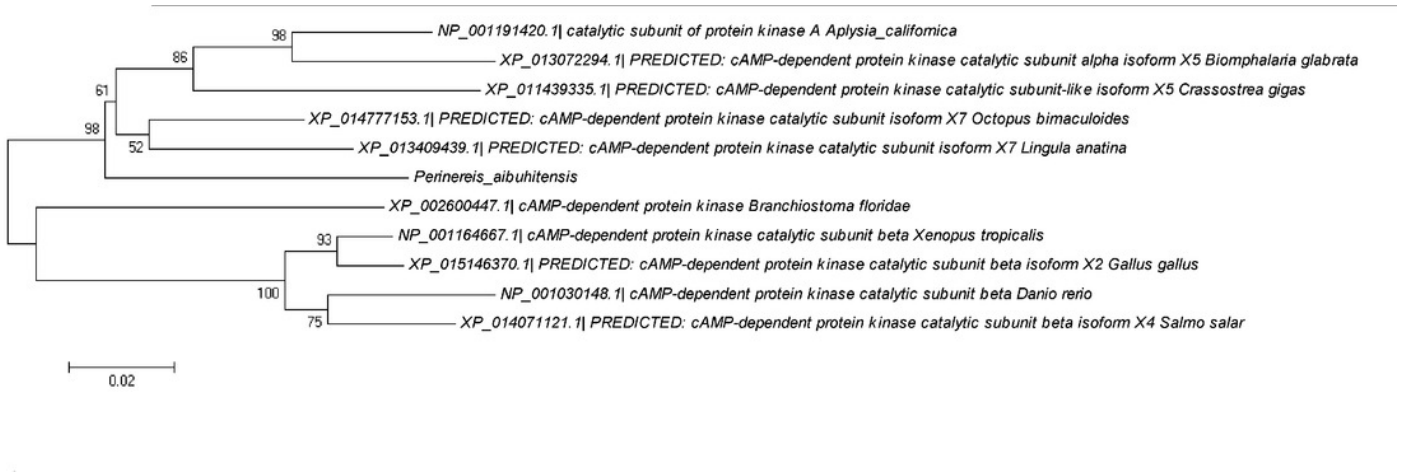

Figure 10

The relative expression level of PaGPCR and PaPKA cDNAs under various B(a)p concentration exposure. A PaGPCR, B PaPKA.Different lowercase letters indicate significant difference ($P< 0.05$). all data as mean+SD.N= four worms.

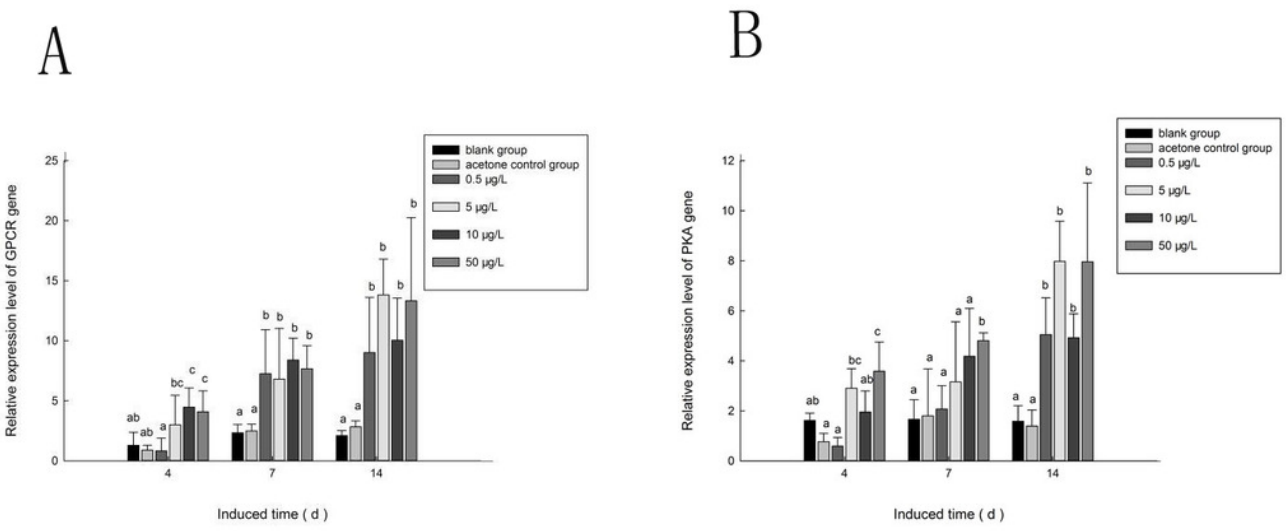

Table 1(on next page)

The primers used in this study

Primer name		sequence（5'-3'）
RACE	GPCR－F1	TGAGAAACGTCGAAGCGAAAGG
	GPCR－R1	ATATTCGACGCTGACCCTAAGG
	PKA-F1	GATACCCACCTTTCTTTGCTGACC
	PKA-F2	GGTGCGCTTCCCATCTCACTTT
	PKA-R1	CAATAGCGCAGCCTCAGGGACA
	UPM Long	CTAATACGACTCACTATAGGGCAAGCAG TGGTATCAACGCAGAGT
	UPM Short	CTAATACGACTCACTATAGGGC
Real time PCR	β -actin-R	CGAAGTCCAGAGCAACATAG
	β -actin-F	GGGCTACTCCTTCACCACCA
	GPCR-R3	CCGTAAAAGCCTCATCAAGACA
	GPCR-F3	TTGGCAGGTGTAAATGAATGG
	PKA-F3	GACCAGCCAATCCAAATCTATG
	PKA-R3	GACCCCATTCTTCAGGTTTCC

1

2

3

4