

Three-dimensional scoring of zebrafish response to psychoactive drugs questions the predictive validity of two-dimensional analyses

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Zebrafish (*Danio rerio*) have recently emerged as a valuable laboratory species in the field of behavioral pharmacology, where they afford rapid and precise high-throughput drug screening. Although the behavioral repertoire of this species manifests along three dimensions (3D), most of the efforts in behavioral pharmacology rely on 2D projections acquired from a single overhead or front camera. We recently showed that, compared to a 3D scoring approach, 2D analyses could lead to inaccurate claims regarding individual and social behavior of drug-free experimental subjects. Here, we examined whether this conclusion extended to the field of behavioral pharmacology by phenotyping adult zebrafish, acutely exposed to citalopram (30 mg/L, 50 mg/L, and 100 mg/L) or ethanol (0.25%, 0.50%, and 1.00%), in the novel tank diving test over a six-minute experimental session. We observed that both compounds modulated the time course of general locomotion and anxiety-related profiles, the latter being represented by specific behaviors (erratic movements and freezing) and avoidance of anxiety-eliciting areas of the test tank (top half and distance from the side walls). We observed that 2D projections of 3D trajectories (ground truth data) may introduce a source of unwanted variation in zebrafish behavioral phenotyping. Predictably, both 2D views underestimate absolute levels of general locomotion. Additionally, while data obtained from a camera positioned on top of the experimental tank are similar to those obtained from a 3D reconstruction, 2D front view data yield false negative findings.

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18 **Abstract**

19 Zebrafish (*Danio rerio*) have recently emerged as a valuable laboratory species in the field of
20 behavioral pharmacology, where they afford rapid and precise high-throughput drug screening.

21 Although the behavioral repertoire of this species manifests along three dimensions (3D), most
22 of the efforts in behavioral pharmacology rely on 2D projections acquired from a single overhead
23 or front camera. We recently showed that, compared to a 3D scoring approach, 2D analyses
24 could lead to inaccurate claims regarding individual and social behavior of drug-free
25 experimental subjects. Here, we examined whether this conclusion extended to the field of
26 behavioral pharmacology by phenotyping adult zebrafish, acutely exposed to citalopram (30
27 mg/L, 50 mg/L, and 100 mg/L) or ethanol (0.25%, 0.50%, and 1.00%), in the novel tank diving
28 test over a six-minute experimental session. We observed that both compounds modulated the
29 time course of general locomotion and anxiety-related profiles, the latter being represented by
30 specific behaviors (erratic movements and freezing) and avoidance of anxiety-eliciting areas of

31 the test tank (top half and distance from the side walls). We observed that 2D projections of 3D
32 trajectories (ground truth data) may introduce a source of unwanted variation in zebrafish
33 behavioral phenotyping. Predictably, both 2D views underestimate absolute levels of general
34 locomotion. Additionally, while data obtained from a camera positioned on top of the
35 experimental tank are similar to those obtained from a 3D reconstruction, 2D front view data
36 yield false negative findings.

37

38 **Keywords:** anxiety; automated tracking; citalopram; ethanol; novel tank diving test.

39

40 1. Introduction

41 Preclinical animal models constitute a central tool to detail the fundamental mechanisms
42 underlying the expression of human emotions in physiological and pathological conditions
43 (Haller & Alicki 2012). Within this framework, several experimental models have been proposed
44 to investigate the neurobiological processes underlying anxiety (Davis et al. 2010; Hart et al.
45 2010), an evolutionarily preserved adaptive emotion, normally occurring as an anticipatory
46 response to a potential threat (Bateson et al. 2011). The adaptive value of anxiety resides in the
47 fact that it limits the negative outcomes associated with a potential threat (Nesse 1999).
48 Notwithstanding its adaptive nature, inappropriate (context-independent) or excess anxiety may
49 often culminate in anxiety-related disorders that require medical attention (Bateson et al. 2011).

50 In parallel with the aforementioned evolutionary roots, the underlying biological
51 determinants of anxiety are very well conserved across different taxa. For example, the
52 neuroendocrine machinery activated in response to external stressors exhibits striking
53 homologies and analogies among fish (Bernier & Peter 2001), birds (Lynn & Kern 2018),
54 rodents (Macri & Wurbel 2006), monkeys (Parker et al. 2012), and humans (Rodrigues et al.
55 2009). Likewise, neurotransmitters such as serotonin have been associated with anxiety-related
56 behaviors in species as diverse as fish (Fossat et al. 2014), birds (Hogg et al. 1994), humans
57 (Caspi et al. 2003), and sheep (Lee et al. 2016).

58 Although rodents have traditionally constituted the species of choice in this field (Hart et
59 al. 2010; Kalueff et al. 2007), zebrafish have recently emerged as an extremely promising
60 experimental species (Fontana et al. 2018; Shams et al. 2018; Stewart et al. 2014). The success of

61 this freshwater species rests upon several advantages that range from genetic and neuroanatomic
62 isomorphism between zebrafish and humans (Howe et al. 2013), to their small size and high
63 reproductive rates favoring the execution of high-throughput studies (Kalueff et al. 2014). In
64 addition, the possibility to dissolve substances in water allows for the non-invasive
65 administration of drugs readily absorbed through the gills (Tran & Gerlai 2013). These
66 characteristics designate zebrafish as a fundamental tool in the field of psychopharmacology
67 whereby they allow the preliminary screening of numerous drugs within spaces and time frames
68 much smaller than those required by laboratory mammals (McCarroll et al. 2016).

69 High-throughput behavioral experiments on zebrafish generally share the following
70 methodological structure: administration of water-soluble drugs, videorecording of observable
71 phenotypes, offline scoring of video, coding of the observed behaviors, and data analysis
72 (Stewart et al. 2014). Traditional behavioral phenotyping leveraged the use of a single camera
73 positioned on top or in front of the experimental tank and the subsequent use of behavioral
74 scoring software, in which the phenotype of interest had to be input by a trained observer
75 (Cianca et al. 2013; Spinello et al. 2013). Albeit extremely productive, this approach was prone
76 to observer bias and has been recently complemented by tracking algorithms capable of
77 automatically coding and scoring zebrafish behavior with limited human supervision (Delcourt et
78 al. 2018; Franco-Restrepo et al. 2019; Nema et al. 2016; Perez-Escudero et al. 2014).

79 However, from the two-dimensional (2D) view offered by a single video-camera it is
80 impossible to phenotype the 3D swimming pattern exhibited by zebrafish. This consideration
81 prompted the design and development of experimental platforms capable of investigating
82 zebrafish behavior adopting a 3D approach (Cachat et al. 2011; Maaswinkel et al. 2013; Macrì et
83 al. 2017; Stewart et al. 2015). We recently demonstrated that the limitation of 2D scoring
84 methods extends beyond the geometrical underestimation of swimming paths (3D trajectories
85 being longer than their 2D projections by definition), and may result in numerous false positive
86 and false negative findings (Macrì et al. 2017). Specifically, we first tested zebrafish in
87 conventional binary choice behavioral assays, and then analyzed group differences based on 3D
88 or 2D (top and front views) trajectories. This analysis demonstrated that 2D views generated
89 approximately 20% of false findings, being represented by inappropriate reporting of significant
90 inter-group differences in spite of undistinguishable ground truth data (false positives) or failure

91 to detect significant results in instances in which experimental groups belonged to different
92 populations (false negatives) (Macri et al. 2017).

93 In the present study, we aimed at prospectively investigating whether 3D scoring of
94 zebrafish behavior may also benefit pharmacological research. To this aim, we exposed
95 experimentally naïve zebrafish to drugs capable of modulating anxiety-related behaviors in both
96 humans and zebrafish (Cianca et al. 2013; Sackerman et al. 2010), and then analyzed their
97 phenotype in response to an anxiety-provoking test paradigm in 3D or in 2D (top and front
98 views). Specifically, we investigated the behavior of zebrafish in a novel tank diving test in
99 response to the administration of the selective serotonin reuptake inhibitor citalopram (30 mg/L,
100 50 mg/L, and 100 mg/L) or ethanol (0.25%, 0.50%, and 1.00%). The goal of this study was
101 twofold: first, we sought to replicate existing findings indicating that ethanol (Pannia et al. 2014)
102 and citalopram (Sackerman et al. 2010) modulate anxiety in zebrafish (predictive validity of the
103 assay), and then we aimed at testing whether the experimental advantages afforded by 3D
104 scoring in drug-free states (Macri et al. 2017) also extend to zebrafish psychopharmacology.

105 The novel tank diving test has already been validated as a locomotion- and anxiety-
106 related behavioral test (Cachat et al. 2010). Therein, anxiety is measured through the evaluation
107 of fish position in the water column, swimming speed, erratic movements, and freezing, as
108 functions of the time spent in the experimental tank from the initial release. In order to detail the
109 specific information that can be potentially inferred from these measurements, we preliminarily
110 conducted a principal component analysis (PCA) on nine behavioral measures, objectively
111 scored from 3D trajectories (average speed, average peak speed, average angular speed, average
112 peak angular speed, average acceleration, average peak acceleration, time spent freezing, time
113 spent in the top half of the tank, and time spent in the vicinity of the walls). The PCA was aimed
114 at detecting potential correlations among the variables and identifying underlying orthogonal
115 factors associated with independent domains.

116 Grounded in our previous work, we anticipated 2D views to be characterized by reduced
117 absolute locomotion values compared to 3D trajectories. Most importantly, in the light of the
118 high rate of false findings observed in drug-free conditions (Macri et al. 2017), we expected the
119 predictive validity of 2D trajectories to be potentially jeopardized. This hypothesis rests on the
120 fact that, when exposed to psychoactive substances, fish may exhibit a series of responses that
121 vary in space and time. For example, increased anxiety may reflect in a progressive reduction in

122 general locomotion, increased freezing, erratic movements, and preference for the bottom of the
123 experimental tank. These patterns manifest differentially depending on the time spent in the
124 experimental apparatus (with preference for the bottom varying with the prolonged exposure),
125 and on the view (i.e., top or front view). For example, while horizontal erratic movements are
126 best detected through a top view, geotaxis can be appropriately scored only from a side view.
127 Therefore, we hypothesized that the specific view may reflect into a bias in detecting time-
128 dependent effects of psychoactive drugs, thereby potentially generating view \times drug \times
129 experimental-progression effects.

130

131 **2. Materials and methods**

132 *2.1 Animal care and maintenance*

133 The experiments and analysis were performed and reported according to the ARRIVE guidelines
134 (Kilkenny et al. 2010). A total of 112 wild-type adult zebrafish (*Danio rerio*), with a 1:1
135 male/female ratio were used in this study. The fish were purchased from Carolina Biological
136 Supply Co. (Burlington, NC, USA), and housed in 10 L (2.6 gallons) vivarium tanks (Pentair
137 Aquatic Eco-Systems Locations, Cary, NC, USA), with a density of no more than 10 fish per
138 tank. Fish were kept under a 12 h light/12 h dark photoperiod (Cahill 1996), and fed with
139 commercial flake food (Hagen Corp. Nutrafin max, Mansfield, MA, USA) once a day,
140 approximately at 7 PM. Water parameters of the holding tanks were regularly checked, and
141 temperature and pH were maintained at 26 °C and 7.2 pH, respectively. Regular tap water was
142 used with the addition of a stress coat to remove chlorine and chloramines. Prior to the beginning
143 of the experiments, fish were acclimatized in the holding facility for a period of 12-15 days.

144 The number of fish used in the study – compatible with obtaining sufficiently reliable and
145 biologically relevant data – was estimated through a power analysis. Briefly, we computed the
146 minimum required sample size considering the two-tailed Student t test for independent groups
147 using the following values, based on the results of previous studies (Abaid et al. 2012; Cianca et
148 al. 2013; Spinello et al. 2013): (i) standard deviation homogeneous among groups $s = 0.23$; (ii)
149 Type I error probability $\alpha = 0.05$ and power $1 - \beta = 0.80$ (conventional values); and (iii) minimum
150 difference between control and treatment group means $D = 0.17$. The sample size resulting from
151 this calculation was 15 subjects per group. To promote the generalizability of our findings, we

152 conducted experiments on both males and females. We thus increased the sample size to 16 per
153 group (eight males and eight females). We estimated that a sample size of 16 subjects (per
154 group) would have 80% power to detect a 0.60 effect size on the principal outcome measures
155 with a two-tailed significance level of 0.05.

156

157 *2.2 Experimental setup*

158 To obtain 3D trajectories, we used two Flea 3 high resolution cameras (one overhead and one in
159 front). The dimensions of the test tank were 29 cm (length) × 14 cm (height) × 8.5 cm (width)
160 and water 13 cm deep, similar to tanks used in comparable studies (Egan et al. 2009). To
161 maximize the visual contrast and ease automatic tracking, the bottom of the tank was lined with
162 white contact paper. The two short sides of the tank were covered with black contact paper to
163 prevent reflection. On the other hand, the two long sides were kept transparent to allow data
164 acquisition and avoid position bias (i.e., a potential side preference had one side been kept
165 transparent for data acquisition and the other kept opaque). The experimental arena was
166 surrounded by black curtains to prevent light reflection and visual disturbance from the outside.

167

168 *2.3 Experimental procedure*

169 Experiments, performed in June 2018, were conducted on seven groups, each consisting of 16
170 subjects (eight males and eight females). Specifically, the experimental design entailed one
171 control group exposed to vehicle (water), three groups treated with citalopram (30 mg/L, 50
172 mg/L, 100 mg/L), and three groups treated with ethanol (0.25%, 0.50%, 1.00% ethanol/water
173 solution in volume/volume %). The fish were randomly allocated to each of the seven conditions
174 in the following way. The conditions were randomly distributed over several weeks, testing eight
175 subjects per day (four in the morning and four in the afternoon). We balanced sex across
176 conditions, and conditions across mornings and afternoons. Male and female fish were kept in
177 separate tanks; in total, fish were housed in 12 tanks. At the beginning of each test session, we
178 sampled one subject from a tank. Such a tank was different from that out of which we chose the
179 previous subject tested in the same condition. This procedure guaranteed that potential tank
180 effects were distributed evenly across all experimental groups.

181 Due to technical issues, four trials had to be discarded: this resulted in a slight reduction
182 in the number of subjects in the 100 mg/L citalopram group (15 subjects instead of 16) and in

183 both the 0.25% and 1.00% ethanol groups (14 and 15 subjects instead of 16, respectively).
184 Following (Sackerman et al. 2010), we measured the effect of acute exposure to citalopram by
185 treating the fish to the substance for five minutes before testing it. Following previous work on
186 the effect of ethanol by our group (Cianca et al. 2013), we measured the effect of exposure to
187 ethanol over a one-hour period. In the interest of reducing the number of subjects used in animal
188 experimentation, the same control subjects were used to test the effects of citalopram and
189 ethanol. Fish were treated and tested in isolation.

190 Since these substances required a differential pre-exposure time (five minutes for
191 citalopram and one hour for ethanol), we devised a common procedure for vehicle, citalopram
192 and ethanol. Thus, one hour before testing, fish were placed in a 500 mL beaker filled with 450
193 mL of the following fluid: water for control and citalopram groups, or a solution of ethanol
194 (0.25%, 0.50%, and 1.00%) for the other groups. Five minutes before testing, an additional 50
195 mL of fluid were slowly added to the beaker over a period of 20-30 seconds. These 50 mL were
196 constituted by either water (the control group), an ethanol solution of the concentration already
197 present in the beaker (the ethanol groups), or a concentrated solution of citalopram that resulted
198 in a final concentration in the beaker of 30 mg/L, 50 mg/L, or 100 mg/L (the citalopram
199 condition). Fish were left in the beaker for five minutes, at the end of which they were
200 transferred to the test tank and recorded for six minutes.

201 Simultaneous recording from both cameras was initiated before transferring the fish into
202 the test tank. In addition, at the beginning of the recording, a laser beam, visible from both
203 cameras, was pointed into the test tank in order to ensure later synchronization of both video
204 streams. At the end of the experiment, the fish was hand-netted into a separate tank.

205 All the experiments were performed at the New York University Tandon School of
206 Engineering in Brooklyn NY (USA) in accordance with relevant guidelines and regulations, with
207 National Institutes of Health guide for the care and use of Laboratory animals (NIH Publications
208 No. 8023, revised 1978), and was approved by the University Animal Welfare Committee
209 (UAWC) of New York University under protocol number 13-1424.

210

211 *2.4 Tracking and 3D reconstruction*

212 Images recorded from the high-resolution cameras were processed through an in-house
213 developed tracking software, see (Butail et al. 2013) for a detailed description. The top and front

214 view cameras provided time series of the trajectory projected onto the x-y and x-z planes,
215 respectively. Each pair of tracks were automatically synchronized using the common x
216 coordinate along length of the tank. The time-series for each x coordinate of the pair were shifted
217 relative to each other and the relative shift producing the smallest difference was selected. Once
218 synchronized, the tracks from the top view and from the front views were combined to construct
219 the x, y, and z coordinates of the trajectory in the three-dimensional space (see Fig. 1 for a
220 representative trajectory exhibited by a control subject).

221 Reconstructed trajectories were used to quantify the following ethogram: time spent
222 freezing (percentage of time that the fish moved less than 2 cm anywhere in the tank over a
223 rolling period of 2 s), time spent wall following (percentage of time that the fish spent within 3
224 cm of any side wall or the bottom of the tank), average speed (time-average of the first-order
225 numerical differentiation of the position time series), average peak speed (time-average of the
226 speed values greater than the 90th percentile), average acceleration (time-average of the
227 magnitude of the first order numerical differentiation of the velocity time series), average peak
228 acceleration (time-average of the acceleration values greater than the 90th percentile), average
229 angular speed (time-computed on the basis of a finite difference approximation of the curvature
230 of fish trajectories), average peak angular speed (time-average of the angular velocity values
231 greater than the 90th percentile), and time spent in the top half of the water column. These nine
232 measures were selected from the technical literature on zebrafish behavior in novel tank tests
233 (Cachat et al. 2010) and their objective scoring from 3D trajectories follows our previous work
234 (Macrì et al. 2017; Mwaffo et al. 2015).

235

236 *2.5 Statistical analyses*

237 Experiments with ethanol and citalopram were analyzed separately, but both were compared to
238 the same control condition.

239

240 *2.5.1 Principal component analysis on 3D data*

241 Using raw data on all the nine measures of our ethogram identified from 3D trajectories, we
242 conducted a PCA to identify correlation structure of behavior and potentially reduce the number
243 of variables analyzed. Only principal components with eigenvalues larger than one were retained
244 in the analysis. For each compound (citalopram or ethanol), the loadings were varimax-rotated,

245 and the resulting scores for each principal component were used as dependent variables in a four
246 (citalopram: vehicle, 30 mg/L, 50 mg/L, and 100 mg/L, or ethanol: vehicle, 0.25%, 0.50%, and
247 1.00%) \times six (time bins, one minute each) \times two (sex: male, female) repeated measures analysis
248 of variance (ANOVA) for split-plot designs. Testing males and females served the aim to access
249 a heterogeneous experimental population and therefore improve the generalizability of our
250 findings.

251 Principal components, derived from 3D observations, were used to test the efficacy of
252 ethanol and citalopram in modifying zebrafish behavior. It was not possible to use PCA to
253 compare the different views since the number of variables that construct the principal
254 components in 3D was greater than that in 2D.

255

256 *2.5.2 Statistical model to compare 2D and 3D analyses*

257 To investigate whether 2D projections of 3D trajectories may introduce a bias in the predictive
258 validity of behavioral data on each of the nine measures, we conducted another repeated
259 measures ANOVA for split-plot designs. In this analysis, the two general models for citalopram
260 and ethanol were, respectively: three (view: 3D, 2D top, 2D front) \times four (treatment: vehicle, 30
261 mg/L, 50 mg/L, 100 mg/L) \times six (time bins, one minute each) \times two (sex: male, female), and
262 three (view: 3D, 2D top, 2D front) \times four (treatment: vehicle, 0.25%, 0.50%, 1.00%
263 ethanol/water solution) \times six (time bins, one minute each) \times two (sex: male, female) repeated
264 measures ANOVAs. Similar to the PCA, predictions of the effect of sex were not considered.

265 For all ANOVAs, the distribution of the model residuals was visually inspected to verify
266 that they were close to normality (Quinn & Keough 2002). Statistical analyses were performed
267 using R 3.5.0, with the aov function for ANOVAs, the prcomp function for the PCA, and the
268 emmeans 1.3.0 package for post-hoc comparisons using the Dunnett's multiple comparisons test,
269 comparing control to other conditions and first minute to other minutes.

270 This statistical model allowed testing the hypothesis that 2D views yielded spurious
271 results compared to 3D data. While main effects of the view factor allowed assessing whether
272 absolute values differed depending on the tracking method, significant interactions between view
273 and any other factor suggested that the effects of the latter were moderated by the tracking
274 method. For example, a significant view \times treatment interaction would suggest that the effects of
275 a given compound may vary as a function of how the behavior of the animal was scored (i.e.,

276 using 2D projections from top or front, or resorting to 3D trajectories). Upon detecting a
277 significant interaction, we performed post-hoc comparisons, correcting for type-I errors, to detail
278 whether and which pairwise comparisons were significant. Among these comparisons, those
279 contrasting 2D and 3D were germane to the key question of the study.

280

281 **3. Results**

282 *3.1 Ethanol and citalopram alter individual habituation to the test*

283 For citalopram and ethanol treatments, three principal components with eigenvalue larger than
284 one were extracted by the PCA (Table 1), accounting for 87% of the total variance. The first
285 principal component, accounting for 47% of the variance, reflected locomotion, with positive
286 loadings for average speed, average peak speed, average acceleration, and average peak
287 acceleration, and a modest negative loading for the time spent freezing. The second principal
288 component, accounting for 26% of the variance, reflected anxiety-related behavioral patterns
289 (behavioral anxiety) with positive loadings for average angular speed, average peak angular
290 speed, and the time spent freezing. The third principal component, accounting for 11% of
291 variance, reflected anxiety-related spatial preference (positional anxiety), with positive loadings
292 for the time spent wall following, and negative loadings for the time spent in top half.

293 When analyzing the three components identified by PCA, we observed that absolute
294 levels of locomotion were indistinguishable between control and citalopram-treated subjects
295 (condition: $F_{3,55} = 0.52$, $P = 0.668$) (Fig. 2a). Additionally, general locomotion steadily declined
296 throughout the experimental session in all subjects (time: $F_{5,275} = 3.03$, $P = 0.011$; $t_{275} > 3.12$, P
297 < 0.009), regardless of the specific experimental group (time bins \times condition: $F_{15,275} = 1.13$, $P =$
298 0.329). Absolute values of behavioral anxiety did not significantly vary across citalopram
299 conditions (condition: $F_{3,55} = 0.76$, $P = 0.524$) (Fig. 2b). Yet, it significantly decreased over the
300 trial (time: $F_{5,275} = 3.52$, $P = 0.004$; $t_{275} > 2.69$, $P < 0.033$), albeit at a different rate (time bins \times
301 condition: $F_{15,275} = 1.85$, $P = 0.029$). Specifically, while behavioral anxiety remained constant
302 throughout the experimental session in citalopram 50 mg/L and 100 mg/L conditions, it
303 significantly declined in control and citalopram 30 mg/L conditions ($t_{275} > 2.60$, $P < 0.043$).

304 While positional anxiety did not significantly vary across citalopram conditions (condition: $F_{3,55}$
305 $= 0.95$, $P = 0.421$) (Fig. 2c), it significantly increased over time (time: $F_{5,275} = 3.51$, $P = 0.004$;

306 $t_{275} > 2.82, P < 0.023$). Such time-dependent profile varied depending on the experimental
307 treatment (time bins \times condition: $F_{15,275} = 1.79, P = 0.036$). Thus, while it remained constant in
308 control and citalopram 100 mg/L, it was low at the beginning of the test session and steadily
309 increased in citalopram 30 mg/L and citalopram 50 mg/L subjects ($t_{275} > 2.74, P < 0.029$).

310 In response to ethanol administration, absolute levels of locomotion failed to reach a
311 statistically significant variation across experimental groups (condition: $F_{3,52} = 2.43, P = 0.072$)
312 (Fig. 3a). When analyzing the time course of general locomotion, we observed that it
313 significantly decreased over time (time: $F_{5,260} = 3.02, P = 0.011; t_{260} > 2.67, P < 0.035$), and that
314 such a decrease was indistinguishable across all experimental groups (time bins \times condition:
315 $F_{15,260} = 1.50, P = 0.105$). Behavioral anxiety did not significantly vary across ethanol conditions
316 (condition: $F_{3,52} = 0.80, P = 0.500$) (Fig. 3b), neither did it apparently change over time (time:
317 $F_{5,260} = 1.66, P = 0.144$). However, we observed that the habituation profile varied depending on
318 the specific experimental group (time bins \times condition: $F_{15,260} = 1.83, P = 0.031$). Specifically,
319 while behavioral anxiety remained constant in most experimental groups, it significantly
320 declined over time in the ethanol 0.5% condition ($P < 0.050; t_{260} = 2.89; P = 0.019$). Finally,
321 positional anxiety failed to reach a statistically significant variation across ethanol conditions
322 (condition: $F_{3,52} = 2.49, P = 0.071$) (Fig. 3c), although it significantly decreased over time (time:
323 $F_{5,260} = 3.25, P = 0.007; t_{260} > 2.75; P < 0.029$). Specifically, it significantly decreased for the
324 ethanol 1.0% condition (time bins \times condition: $F_{15,260} = 2.33, P = 0.004; t_{260} > 3.76; P < 0.001$).

325

326 *3.2 The scoring view influences the validity of experimental outcomes*

327 Herein, we report data concerning the effects of the views on all the experimental variables
328 measured in the study. For the sake of clarity, in this section, we only report statistical findings
329 associated with the scoring view (3D, 2D top, and 2D front) and its interactions with time or
330 condition. Results concerning the main effects of condition, time, and their interaction
331 irrespective of view are available in the supplementary material.

332 Before delving into detailed comparisons between the three different views for all the
333 considered behavioral measures, we present an aggregated assessment of potentially inaccurate
334 conclusions that would be drawn from 2D projections against 3D trajectories. Briefly, we

335 identified that the specific view selected to quantify the behavioral repertoire reverberated in
336 both false negative (erroneous reporting of absence of differences *in lieu* of significant findings
337 in 3D) and false positive (erroneous reporting of significant differences *in lieu* of non-
338 significantly different findings in 3D) results. The rate of false negative and false positive
339 findings is synoptically reported in Tables 2 and 3.

340

341 3.3.1. Citalopram

342 Average speed: Predictably, average speed varied significantly depending on which view was
343 used to compute it (view: $F_{2,110} = 118.46$, $P < 0.001$) (Fig. 4a). Specifically, both 2D front and
344 top views underestimated absolute levels of locomotion compared to 3D data ($t_{291.7} = 9.87$, $P <$
345 0.001 ; and $t_{291.7} = 6.88$, $P < 0.001$, respectively); additionally, 2D front view resulted in reduced
346 average speed compared to top view ($t_{291.7} = 2.98$, $P = 0.009$). Experimental subjects did not
347 show a habituation profile to the test, yet 2D top projections indicated that the average speed
348 decreased from the first to the last minute (time bins \times view: $F_{10,550} = 14.08$, $P < 0.001$; $t_{317.5} =$
349 3.03 , $P = 0.012$).

350 Average peak speed: Average peak speed was significantly underestimated in both 2D front and
351 top views in comparison with 3D data (view: $F_{2,110} = 81.93$, $P < 0.001$; $t_{346.6} > 6.64$, $P < 0.001$)
352 (Fig. 4b). While the average peak speed decreased over time in all subjects (supplementary
353 material), experimental groups showed a differential habituation profile (time bins \times view: $F_{10,550}$
354 $= 15.64$, $P < 0.001$; $t_{286.4} > 2.73$, $P < 0.029$).

355 Average angular speed: Average angular speed was underestimated in the 2D front view
356 compared to both 3D and 2D top views (view: $F_{2,110} = 88.45$, $P = 0.001$; $t_{295.3} > 5.33$, $P < 0.001$)
357 (Fig. 4c). Furthermore, a decrease in average angular speed over time was observed in all views
358 (time bins \times view: $F_{10,550} = 7.40$, $P < 0.001$; $t_{275} > 2.69$, $P < 0.034$).

359 Average peak angular speed: Average peak angular speed was underestimated when scored from
360 2D top view (view: $F_{2,110} = 10.09$, $P < 0.001$; $t_{358.6} = 3.62$, $P = 0.001$) (Fig. 4d). A decrease in
361 average peak angular speed over time was recorded from all views, but not at the same times
362 (time bins \times view: $F_{2,550} = 2.72$, $P = 0.003$; $t_{275} > 2.64$, $P < 0.038$).

363 Average acceleration: Average acceleration was underestimated in both front and top 2D views
364 compared to 3D (view: $F_{2,110} = 126.62$, $P < 0.001$; $t_{284.4} = 9.24$, $P < 0.001$; and $t_{284.4} = 5.57$, $P <$
365 0.001 , respectively); additionally, 2D front view underestimated average acceleration compared
366 to 2D top view ($t_{284.4} = 3.67$, $P < 0.001$). Average acceleration varied over time depending on the
367 view adopted to score fish behavior (time bins \times view: $F_{10,550} = 13.40$, $P < 0.001$) (Fig. 4e).
368 Specifically, although average acceleration steadily declined from the third minute in ground
369 truth 3D data ($t_{371.0} = 2.57$, $P < 0.046$), such a decline was observable also from 2D top view
370 ($t_{371.0} > 2.80$, $P < 0.025$), but only during the last minute in 2D front view ($t_{275.0} = 2.70$, $P <$
371 0.033).

372 Average peak acceleration: Average peak acceleration significantly decreased over time,
373 regardless of the specific view adopted to compute this measure (time bins \times view: $F_{10,550} = 4.42$,
374 $P < 0.001$; $t_{344.7} > 3.15$, $P < 0.008$) (Fig. 4f). Yet, average peak acceleration was underestimated
375 in 2D front and top views compared to 3D (view: $F_{2,110} = 79.67$, $P < 0.001$; $t_{402.3} = 6.05$, $P <$
376 0.001 ; and $t_{402.3} = 3.46$, $P = 0.002$, respectively). Additionally, 2D front view yielded a lower
377 average peak acceleration compared to top view ($t_{402.3} = 2.59$, $P = 0.027$).

378 Wall following: Time spent wall following was significantly underestimated in both 2D front
379 view compared to 3D data (view: $F_{2,110} = 237.90$, $P < 0.001$; $t_{234.4} = 17.55$; $P < 0.001$).
380 Additionally, this metric was lower in 2D front view compared to 2D top view ($t_{234.4} = 15.95$, P
381 < 0.001) (Fig. 4g). While 3D and 2D top view data indicated that wall following increased
382 between the first and fifth minute of the experimental session (time bins \times view: $F_{10,550} = 2.16$, P
383 $= 0.019$, $t_{824.7} > 2.56$, $P < 0.05$), 2D front view data failed to identify this time dependent pattern
384 of thigmotaxis.

385 Position in the water column (proportion of time spent in the top half): Since this metric takes
386 into account only the vertical position of the fish, it cannot be scored from 2D top view and there
387 is no difference between values from 2D front view and 3D reconstructed trajectories (Fig. 4h
388 and 5h).

389 Freezing: Although the time spent freezing seemed to vary depending on which view was used to
390 compute it (view: $F_{2,110} = 4.19$, $P = 0.018$) (Fig. 4i), post-hoc tests revealed no pairwise

391 difference. Similarly, although an interaction between view and time was registered (time bins \times
392 view: $F_{10,550} = 2.35$, $P = 0.010$), post-hoc comparisons did not indicate any specific difference.

393

394 3.3.2. Ethanol

395 Average speed: The different scoring views resulted in variable average speed values (view:
396 $F_{2,104} = 90.45$, $P < 0.001$) (Fig. 5a). Both 2D top and front views underestimated average speed
397 compared to 3D ($t_{174.3} = 3.79$; $P < 0.001$; and $t_{174.3} = 9.47$; $P < 0.001$, respectively).
398 Additionally, average speed was lower in 2D front view compared to top view ($t_{174.3} = 5.69$; $P <$
399 0.001). While data inspection suggested that habituation profiles were skewed by the view
400 adopted to score individual behavior (time bins \times view: $F_{10,520} = 10.14$, $P < 0.001$), post-hoc
401 analyses failed to show significant view-dependent variations in this parameter.

402 Average peak speed: Average peak speed varied in all subjects, and this profile was apparently
403 influenced by the view adopted to score individual trajectories (time bins \times view: $F_{10,520} = 9.36$,
404 $P < 0.001$; $t_{260.0} > 2.64$, $P < 0.039$). This variation was manifested as a robust decline in subjects
405 treated with ethanol 0.50% concentration (time bins \times condition \times view: $F_{30,520} = 1.83$, $P =$
406 0.005 ; $t_{260.0} > 2.57$, $P < 0.047$, see Fig. S2, supplementary information). Furthermore, average
407 peak speed was significantly underestimated in both 2D front and top views compared to 3D data
408 (view: $F_{2,104} = 83.2$, $P < 0.001$; $t_{270.2} > 4.41$; $P < 0.001$) (Fig. 5b), as well as from the front view
409 compared to the top view ($t_{260.0} = 2.60$; $P = 0.027$). Although ANOVA reported a significant
410 interaction between view and condition (condition \times view: $F_{6,104} = 2.33$, $P = 0.037$), post-hoc
411 tests failed to reveal any significant pairwise difference.

412 Average angular speed: Predictably, both 2D front and top views underestimated average angular
413 speed compared to the 3D view (view: $F_{2,104} = 63.33$, $P < 0.001$; $t_{265.7} = 7.73$, $P < 0.001$; and
414 $t_{265.7} = 4.78$, $P < 0.001$, respectively). Additionally, average angular speed was smaller in 2D
415 front view compared to 2D top view ($t_{265.7} = 2.95$, $P < 0.010$) (Fig. 5c). Furthermore, 3D data
416 demonstrated that average angular speed declined between the first and the last minute of
417 observation. Such a decline, observable in 2D top view data, was not detected in 2D front view
418 (time bins \times view: $F_{10,520} = 5.19$, $P < 0.001$, $t_{730.9} = 3.902$, $P < 0.005$).

419 Average peak angular speed: Although average peak angular speed appeared significantly
420 different depending on which view was used (view: $F_{2,104} = 4.37$, $P = 0.015$), such a difference
421 failed to emerge in post-hoc comparisons. From all views, average peak angular speed declined
422 throughout the experimental session (time bins \times view: $F_{10,520} = 3.53$, $P < 0.001$; $t_{707.1} > 2.66$, P
423 < 0.035) (Fig. 5d).

424 Average acceleration: In line with most of the locomotion-related variables, average acceleration
425 was underestimated in both 2D front and top views compared to 3D view (view: $F_{2,104} = 64.41$, P
426 < 0.001 ; $t_{164.6} = 7.45$, $P < 0.001$; and $t_{164.6} = 3.20$, $P = 0.005$, respectively, see Fig. 5e).
427 Furthermore, 2D front view yielded lower values of the average acceleration compared to the top
428 view ($t_{164.6} = 4.25$, $P < 0.001$). Individual habituation profile was differentially expressed by
429 experimental subjects depending on the specific view (time bins \times view: $F_{10,520} = 11.87$, $P <$
430 0.001). Specifically, while 3D and 2D top view data indicated a general decrease in average
431 acceleration throughout the experimental session ($t_{312.7} = 2.94$, $P < 0.016$), such a profile was not
432 visible in 2D front view, showing only a reduction during the third minute of the test ($t_{260.0} =$
433 2.70 , $P = 0.033$). Data analysis suggested that the habituation profile varied depending on both
434 the view and the ethanol treatment (time bins \times condition \times view: $F_{30,520} = 1.50$, $P = 0.046$).
435 Specifically, we observed that the reduction in average acceleration was significant in ethanol
436 0.5% ($t_{260.0} > 2.80$, $P < 0.025$), and that this decrease occurred regardless of the specific view
437 from which data were scored.

438 Average peak acceleration: Average peak acceleration varied depending on the specific scoring
439 view (view: $F_{2,104} = 61.85$, $P < 0.001$); specifically it was underestimated in both 2D front and
440 top views compared to 3D ($t_{273.9} = 5.69$, $P < 0.001$; and $t_{273.9} = 2.41$, $P = 0.044$, respectively) and
441 was also less in 2D front view compared to 2D top view ($t_{273.9} = 3.28$, $P = 0.003$) (Fig. 5f).
442 Furthermore, although data inspection suggested that the time-dependent habituation profile
443 varied depending on the specific view (time bins \times view: $F_{10,520} = 5.70$, $P < 0.001$; $t_{312.6} = 2.57$,
444 $P < 0.047$), post-hoc tests did not support this suggestion. Thus, acceleration decreased with time
445 in experimental subjects regardless of the specific view adopted.

446 Wall following: The time spent in the proximity of the walls significantly varied depending on
447 the specific view used to compute it (view: $F_{2,104} = 56.09$, $P < 0.001$). Wall following was
448 significantly underestimated in both 2D front and top views compared to 3D ($t_{178.8} = 9.96$; $P <$

449 0.001; and $t_{178.8} = 3.49$; $P = 0.002$, respectively), and this parameter was less in 2D front view
450 compared to 2D top ($t_{178.8} = 6.47$, $P < 0.001$). The individual habituation profiles varied
451 depending on the view (time bins \times view: $F_{10,520} = 3.37$, $P = 0.001$) (Fig. 5g). Specifically, wall
452 following remained constant in 3D and 2D top view, and decreased in 2D front view ($t_{260.0} =$
453 2.79 ; $P < 0.025$). While wall following was apparently differed between conditions depending on
454 the scoring view (condition \times view: $F_{2,104} = 5.54$, $P < 0.001$), such difference was not statistically
455 significant in pairwise comparisons.

456 Freezing: While the time spent freezing seemed to vary depending on the specific scoring view,
457 (view: $F_{2,104} = 5.35$, $P = 0.006$) (Fig. 5i), such a difference was not confirmed by post-hoc tests
458 performed between the first and the sixth minute.

459

460 4. Discussion

461 The methodological nature of the present study first reverberated in the systematic evaluation of
462 the correlation among the variables that constitute the ethogram exhibited in the novel tank
463 diving test. The PCA revealed the presence of three orthogonal factors, reflecting general
464 locomotion (average speed, average peak speed, average acceleration, and average peak
465 acceleration), anxiety-related behavioral patterns (average angular speed, average angular peak
466 speed, and freezing), and anxiety-related spatial preference (time spent close to the side walls
467 and time spent in the upper half of the water column). The first principal component relates to
468 the translational motion within the water tank. The behavioral patterns loading on the second
469 principal component have been consistently associated with anxiety, in the form of erratic
470 movements (zig-zagging) and freezing (Kalueff et al. 2013). From the catalog of Kalueff and
471 colleagues (Kalueff et al. 2013), anxiety-related behavior is also related to thigmotaxis and
472 geotaxis, which are the two behavioral measures that load on the third principal component.

473 While this analysis aligns with previous evidence indicating that anxiety can be expressed
474 through different modalities, it also points at potential pitfalls of common practice in the
475 construction of the ethogram of the novel tank diving test from 2D views. Specifically, the fact
476 that variables contributing to the same principal component require different perspectives further

477 corroborates the need for a 3D approach. For example, while position in the water column
478 requires a front camera, wall distance and erratic movements need an overhead camera.

479 The analysis conducted on the aforementioned principal components revealed that both
480 citalopram and ethanol influenced anxiety-related behaviors, thus corroborating the predictive
481 validity of the novel tank diving test. Importantly, while citalopram concentration-dependently
482 reduced locomotion and predictably reduced anxiety, ethanol resulted in increased anxiety, but
483 only at a medium concentration (Tran et al. 2016c). Higher and lower ethanol concentrations
484 were apparently ineffective. Low and medium concentrations of citalopram did not influence
485 general locomotion but were associated with the exhibition of reduced anxiety, selectively during
486 the first three minutes of testing. High concentrations of citalopram were associated with reduced
487 locomotion and reduced anxiety throughout the entire test session. The anxiolytic effects of
488 citalopram have already been reported in several studies. For example, (Sackerman et al. 2010)
489 reported that zebrafish treated with 100 mg/L citalopram spent significantly more time than
490 control fish in the top two thirds of the tank, suggesting a decrease in anxiety compared to the
491 control.

492 It is worth noticing that, when analyzing discrete parameters rather than focusing on the
493 principal components, some anxiety-related behavioral parameters seemed unaffected by the
494 anxiolytic treatments applied. Specifically, we failed to observe a significant effect of citalopram
495 on the time spent in the upper portion of the tank, a classical measure of anxiety. We note that
496 such absence of a concentration-dependent behavioral response to anxiolytic compounds has also
497 been reported in other studies. For example, (Sackerman et al. 2010) reported that acute exposure
498 to 0.5% ethanol failed to alter the time spent in the upper portion of the test tank in zebrafish.
499 Similarly, (Maximino et al. 2011) failed to observe significant anxiety-related behavioral
500 alterations in response to fluoxetine. Finally, in a previous study, we also observed that 0.25%
501 and 0.5% ethanol did not modulate anxiety-related behaviors in the light-dark test (Cianca et al.
502 2013). These false negative findings further corroborate the potential heuristic value of
503 conducting PCA in zebrafish behavioral pharmacology.

504 The anxiolytic effects of citalopram are likely related to its direct influence on
505 serotonergic concentrations. For example, handling stress has been shown to increase anxiety-
506 like behavior and reduce brain concentrations of the serotonin metabolite 5-HIAA (Tran et al.
507 2016b). Furthermore, (Maximino et al. 2014) observed that acute administration of the 5-HT_{1a}

508 receptor agonist buspirone reduced behavioral anxiety in the light-dark test. Finally, in
509 accordance with the present study, the acute administration of the selective serotonin reuptake
510 inhibitor fluoxetine resulted in reduced anxiety in the geotaxis test (Maximino et al. 2013).

511 With respect to ethanol, available literature (Gerlai et al. 2000) indicates that its effects
512 vary depending on the concentration, administration schedule, and methodological issues. (Tran
513 et al. 2016a) reported that ethanol can have either anxiogenic or anxiolytic effects on zebrafish
514 depending on whether the water in the test tank comes from the individual's holding tank or from
515 a tank that did not hold any fish. Further, since ethanol influences general locomotion, some of
516 its effects on anxiety may be spurious and potentially related to locomotor effects. For example,
517 a lack of vertical exploration may reflect a decrease in swimming behavior due to the sedative
518 effect of high concentration of ethanol, rather than an anxiety response (Rosemberg et al. 2012).
519 In our previous study (Cianca et al. 2013), we observed that high ethanol concentration resulted
520 in reduced anxiety, associated with reduced motility and increased freezing. Likewise, (Gebauer
521 et al. 2011) observed that ethanol administration resulted in reduced anxiety in the light/dark test,
522 but not in the novel tank diving test. In contrast with these findings, Tran and collaborators (Tran
523 et al. 2016c) reported that acute exposure to high ethanol concentration resulted in increased
524 preference for the bottom of the test tank, and that such a variation related to alterations in brain
525 monoamines. Specifically, alcohol-treated subjects showed reduced concentrations of the
526 dopamine metabolite DOPAC, of serotonin and its metabolite 5-IAA (Tran et al. 2016c). Thus,
527 while the effects of ethanol on anxiety are more variable compared to those exerted by
528 citalopram, they apparently impinge on the same neurochemical pathways modulated by
529 citalopram. Ultimately, the complementary use of these substances served the aim to address the
530 validity of 2D approaches in zebrafish pharmacology of anxiety.

531 In order to compare 3D and 2D approaches, all experimental variables were also analyzed
532 independently from one-another. This comparison was aimed at confirming the intuition that
533 locomotion is underestimated when scoring the behavior in 2D and at assessing whether 2D
534 views yielded incorrect conclusions regarding the effects of anxiolytics on individual behavior.
535 Working with raw experimental variables rather than aggregated principal components allowed
536 for a direct comparison of our findings with available literature, where the selected metrics are
537 routinely assessed in pharmacological phenotyping of zebrafish (Kalueff et al. 2013). With
538 respect to absolute values of locomotion, predictably, they were higher in 3D than 2D, regardless

539 of whether the latter referred to the frontal or the horizontal plane. This can be easily explained
540 by recognizing that 2D trajectories correspond to the projection of the full 3D motion on
541 independent views, which would, by definition, abolish movement along a third dimension. This
542 evidence echoes our previous findings obtained in drug-free states (Macri et al. 2017).

543 The core objective of the present study was to evaluate whether 2D views may result in
544 inaccurate rejection of null hypotheses or acceptance of alternative ones. We observed that the
545 specific view consistently skewed the time course of the behavioral response to the novel tank.
546 This was reflected in the presence of ubiquitous significant view \times time bins interactions across
547 most of the variables, and only few instances of view \times condition interactions. Thus, these data
548 could preliminarily suggest a relative robustness of current scoring methods in zebrafish
549 pharmacology. Yet, in the light of the paucity of drug-dependent effects and of the nature of the
550 statistical model required to test the suitability of the 2D approaches compared to 3D, we argue
551 that this assessment only reflects a partial consideration of the observed results.

552 Specifically, in our previous study, we demonstrated that 2D experiments are
553 underpowered compared to 3D and therefore more prone to false negative findings than false
554 positive ones (Macri et al. 2017). While in situations characterized by few significant main
555 effects of a given variable the likelihood to observe false negatives is intrinsically limited, data
556 with numerous significant main effects shall be amenable to the identification of numerous false
557 reporting instances. Accordingly, in the present study, the sporadic main effects of the condition
558 have apparently masked view-dependent false negatives; complementarily, the ubiquitous
559 presence of main effects of time bins allowed the detection of numerous view \times time bins
560 interactions. Thus, the specific view from which data were scored influenced the observed
561 individual habituation patterns to the experimental paradigm. For example, while 3D data
562 indicated that locomotion-related parameters (e.g., speed, angular speed, and acceleration)
563 declined throughout the experimental session, 2D front view data failed to capture such a time-
564 dependent habituation pattern. While this aspect may simply indicate the limited heuristic
565 potential of the front view and advocate in favor of the use of a top view camera, we nonetheless
566 note that a front camera is indispensable to quantify the position in the water column, which
567 contributes to the anxiety-related phenotype.

568 These considerations extrapolate to zebrafish pharmacology, whereby our and others'
569 data (Cachat et al. 2010) indicate that anxiety-modulating compounds often alter habituation

570 profiles rather than absolute values averaged across different time points (Wong et al. 2010). For
571 example, we reported that anxiety-related behaviors in control subjects appear relatively constant
572 throughout the entire course of the experimental session. Conversely, experimental subjects
573 treated with low and medium concentrations of citalopram exhibit reduced anxiety-related
574 behaviors during the early stages of the task, which gradually rise to attain control values
575 towards the end of the session. Similar to (Watts et al. 2017), we found that although 3D
576 measures offer higher precision, the benefit of using 3D compared to a view from the top is
577 limited regarding general behavioral pattern. The use of a front view remains necessary to
578 capture specific behaviors linked to the position of the fish in the water column.

579

580 **5. Conclusions**

581 It is important to emphasize that in the present study we primarily focused on anxiolytic drugs
582 and we thus cannot extrapolate our findings to the entire spectrum of anxiety-related behaviors.
583 Future studies are needed to test whether the considerations outlined in this study also extend to
584 anxiogenic compounds (e.g., caffeine) and non-pharmacological anxiety-eliciting stimuli (e.g.,
585 predators).

586

587 **Author contributions**

588 SM and MP designed the research; RC and CS performed the experiments; RC and CS scored
589 animal behavior; SM, RC, and MP performed statistical analyses; RC and CS provided a
590 preliminary draft; SM and MP wrote the final draft; and all the authors reviewed the final draft
591 and offered comments.

592

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603

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Figure 1

Trajectory for a single fish from a control trial.

(A) top view, (B) 3D reconstructed trajectory obtained from synchronizing trajectories from top and front views, and (C) front view. The color of the trajectory denotes the evolution of the position of the fish along the six-minute trial. The axes dimensions are 29 cm \times 8.5 cm \times 13 cm.

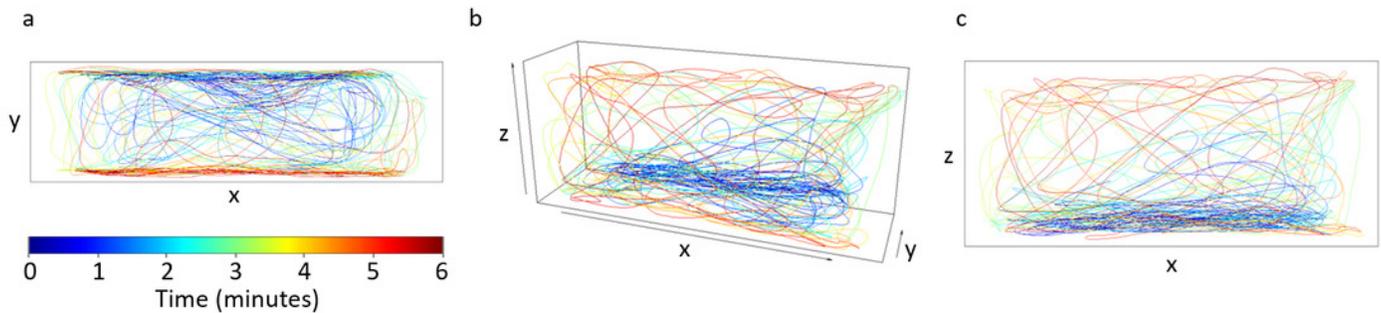


Figure 2

Principal components for the citalopram conditions.

Mean \pm standard error for (A) locomotion, (B) behavioral anxiety, and (C) positional anxiety, over six-minute trials, showing overall variation, as well as for each concentration of citalopram (control 0 mg/L, 30 mg/L, 50 mg/L, and 100 mg/L) based on the reconstructed trajectories in 3D. Filled symbols denote a significant difference from the first minute within each condition. Horizontal bar denotes a significant overall difference in time.

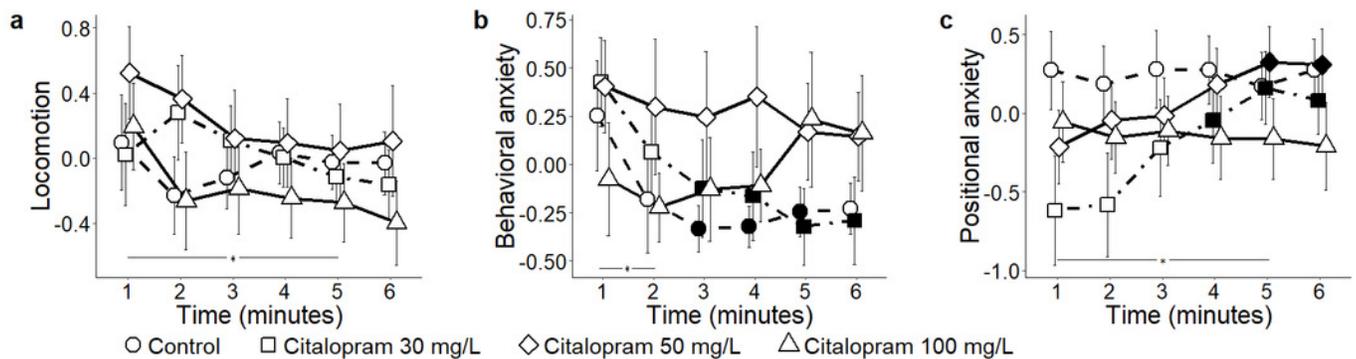


Figure 3

Principal components for the ethanol conditions.

Mean \pm standard error for (A) locomotion, (B) behavioral anxiety, and (C) positional anxiety, over six-minute trials, showing overall variation, as well as for each concentration of ethanol (control 0%, 0.25%, 0.50%, and 1.0%) based on the reconstructed trajectories in 3D. Filled symbols denote a significant difference from the first minute within each condition.

Horizontal bar denotes a significant overall difference in time.

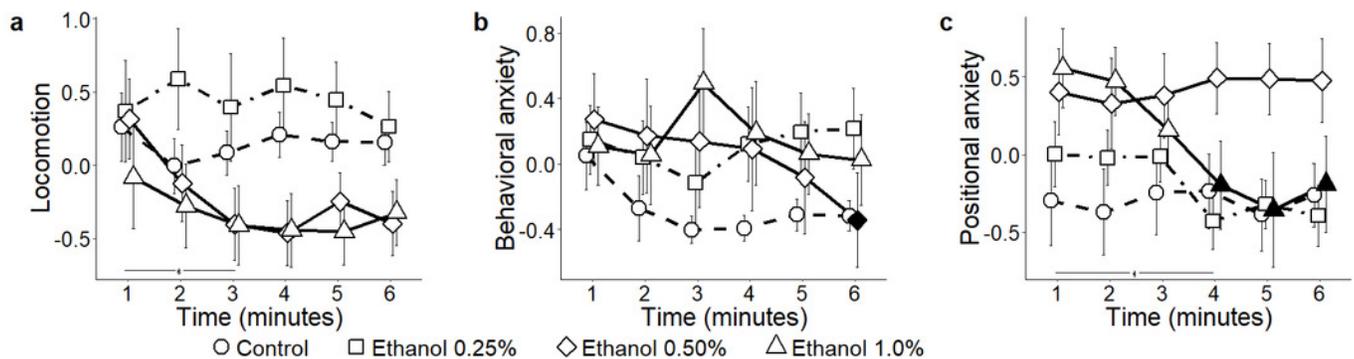


Figure 4

Behavioral parameters for the citalopram conditions.

Mean \pm standard error for (A) average speed, (B) average peak speed, (C) average angular speed, (D) average peak angular speed, (E) average acceleration, (F) average peak acceleration, (G) proportion of time spent within 3 cm of walls, (H) proportion of time spent in the top half of the tank, and (I) proportion of time spent freezing, over six-minute trials aggregated for all citalopram conditions, computed from 2D front and top views, and 3D reconstructed trajectories. Filled symbols denote a significant difference from the first minute within each condition. Horizontal bar denotes a significant overall difference over time. Filled symbols in the top right corner of each panel indicate a significant overall difference with respect to 3D data.

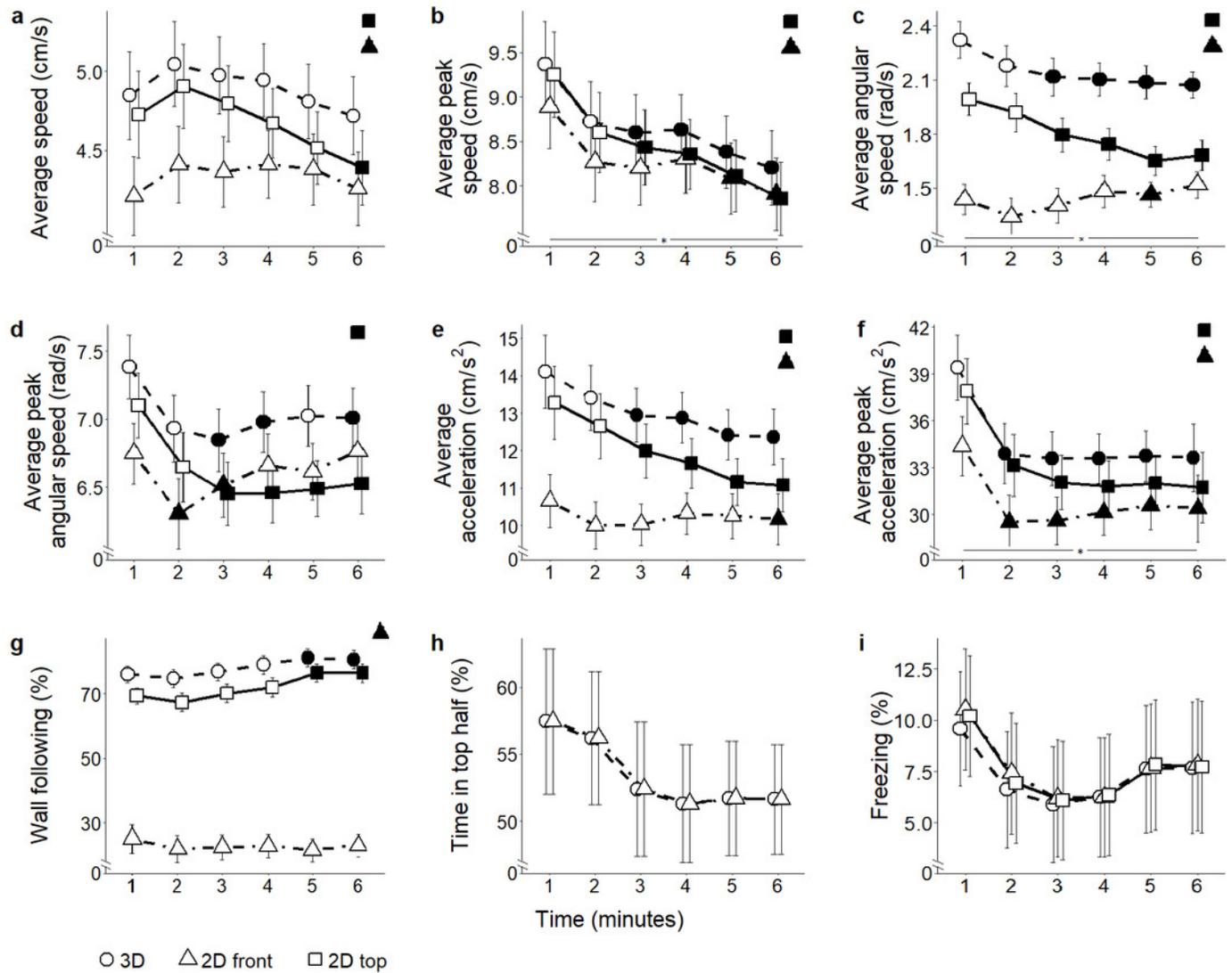


Figure 5

Behavioral parameters for the ethanol conditions.

Mean \pm standard error for (A) average speed, (B) average peak speed, (C) average angular speed, (D) average peak angular speed, (E) average acceleration, (F) average peak acceleration, (G) proportion of time spent within 3 cm of walls, (H) proportion of time spent in the top half of the tank, and (I) proportion of time spent freezing, over six-minute trials aggregated for all ethanol conditions, computed from 2D front and top views, and 3D reconstructed trajectories. Filled symbols denote a significant difference from the first minute within each condition. Horizontal bar denotes a significant overall difference over time. Filled symbols in the top right corner of each panel indicate a significant overall difference with respect to 3D data.

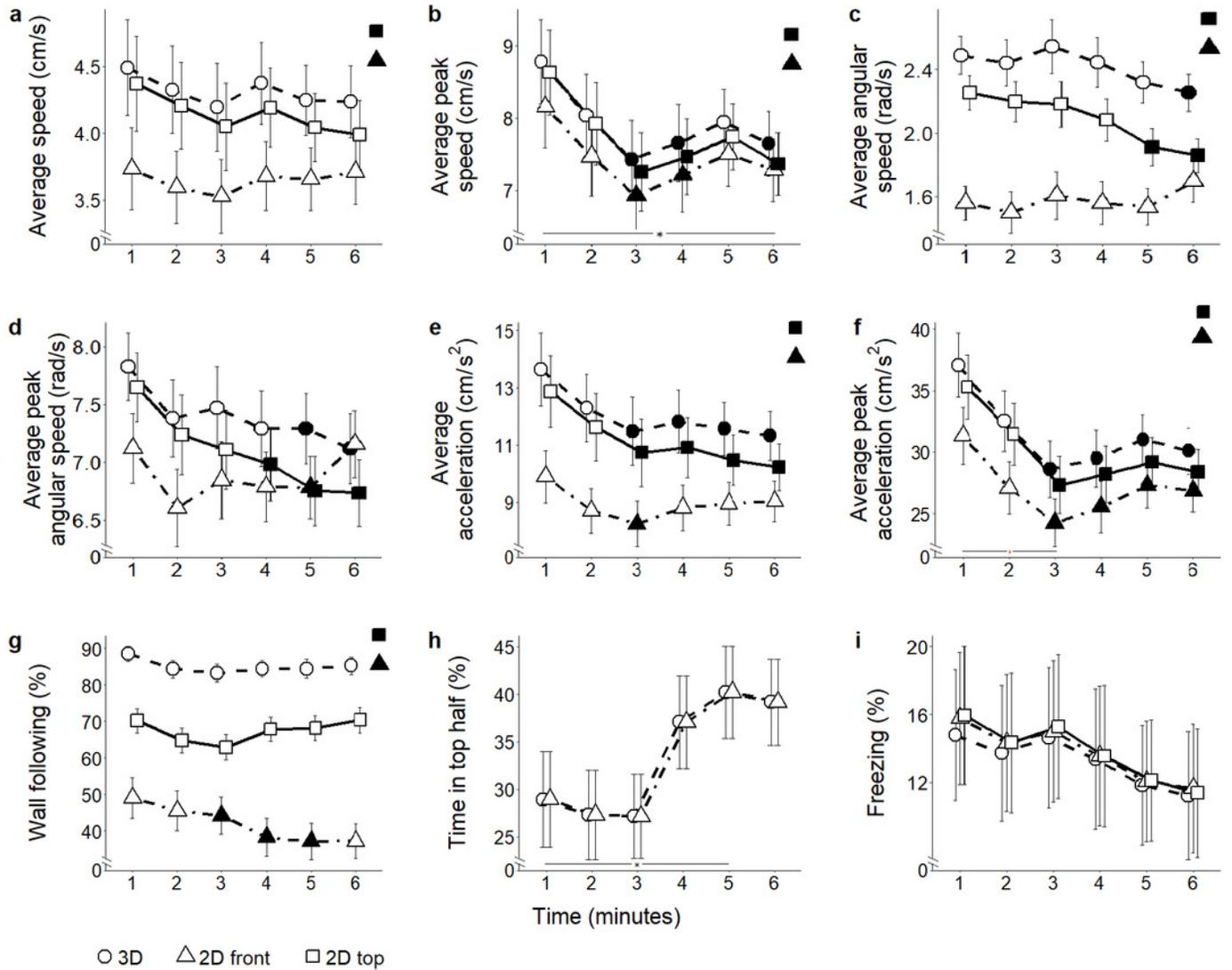


Table 1 (on next page)

Principal component analysis.

Summary results from the principal component analysis for citalopram and ethanol conditions. Principal components with eigenvalue larger than 1 are shown. Loadings greater than 0.7 or smaller than -0.7 are emboldened; loadings smaller than 0.1 in magnitude are not displayed.

	Citalopram			Ethanol		
	Locomotion	Behavioral anxiety	Positional anxiety	Locomotion	Behavioral anxiety	Positional anxiety
Eigenvalues	4.29	2.34	1.19	4.29	2.47	1.06
Explained variance (%)	47.7	26.0	13.3	47.7	27.4	11.8
Cumulative variance (%)	47.7	73.6	86.9	47.7	75.1	86.9
Varimax-rotated loadings						
Speed	0.938	-0.261		0.924	-0.289	
Average peak speed	0.948	-0.172		0.953	-0.122	
Average angular speed		0.939			0.946	0.106
Average peak angular speed		0.971			0.976	0.113
Average acceleration	0.977			0.976		
Average peak acceleration	0.942			0.969		
Freezing	-0.542	0.760		-0.524	0.746	0.170
Wall following		0.176	0.817		0.309	0.670
Time in top half	-0.213		-0.788	-0.118		-0.873

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Table 2 (on next page)

Number of false positive and false negative findings for citalopram.

Number of false positives and false negatives produced for each parameter when computed based on 2D top view and front view data, for the citalopram conditions. A false positive indicates that the 2D view (top or front) yields a significant result that is not supported by the 3D scoring approach. A false negative indicates that the 2D view (top or front) fails to detect a significant result that is instead evident from the 3D scoring approach.

Parameters	Differences between 3D and 2D top view			Differences between 3D and 2D front view		
	False positives	False negatives	Total	False positives	False negatives	Total
Average speed	1	0	1	0	0	0
Average peak speed	0	0	0	0	2	2
Average angular speed	0	0	0	0	3	3
Average peak angular speed	1	0	1	0	2	2
Average acceleration	0	0	0	0	3	3
Average peak acceleration	0	0	0	0	2	2
Wall following	0	0	0	0	2	2
Time in top half	-	-	-	0	0	0
Freezing	0	0	0	0	0	0
Total	2	0	2	0	14	14

1

Table 3(on next page)

Number of false positive and false negative findings for ethanol.

Number of false positives and false negatives produced for each parameter when computed based on 2D top view and front view data, for the citalopram conditions. A false positive indicates that the 2D view (top or front) yields a significant result that is not supported by the 3D scoring approach. A false negative indicates that the 2D view (top or front) fails to detect a significant result that is instead evident from the 3D scoring approach.

Ethanol	Differences between 3D and 2D top view			Differences between 3D and 2D front view		
	Parameters	False positives	False negatives	Total	False positives	False negatives
Average speed	0	0	0	0	0	0
Average peak speed	0	0	0	0	1	1
Average angular speed	1	0	1	0	1	1
Average peak angular speed	1	0	1	0	1	1
Average acceleration	0	0	0	3	0	3
Average peak acceleration	0	0	0	0	0	0
Wall following	0	0	0	3	0	3
Time in top half	-	-	-	0	0	0
Freezing	0	0	0	0	0	0
Total	2	0	2	6	3	9

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