

# The effects of growth rate and biomechanical loading on bone laminarity within the emu skeleton (#27991)

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# The effects of growth rate and biomechanical loading on bone laminarity within the emu skeleton

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The orientation of vascular canals in primary bone may reflect differences in growth rate and/or adaptation to biomechanical loads. Previous studies link specific canal orientations to bone growth rates, but results between different taxa are contradictory. Circularly-oriented vascular canals (forming laminar bone) have been hypothesized to reflect either (or both) rapid growth rate or locomotion-induced torsional loading. Previous work on the hindlimb biomechanics in the emu shows that the femur and tibiotarsus experience large shear strains, likely resulting from torsional loads, that increase through ontogeny. Here, we test how growth rate and biomechanical loading affect bone laminarity in wing and hindlimb elements from growing emu (2 - 416 wks). If bone laminarity is purely an expression of growth rate, it should be most elevated at the growth spurt and decrease with age at a time when shear strains increase in the emu femur and tibiotarsus. Alternatively, if laminar bone reflects biomechanical accommodation, it should become more abundant with age. Transverse mid-shaft histological sections from the limb bones (femur, tibiotarsus, humerus, ulna, and radius) were prepared and imaged. Growth rates were measured using fluorescent bone labels. Vascular canal orientation was quantified using laminarity index (proportion of circularly oriented canals). Growth rates were found to be highest in young individuals for all five skeletal elements. Laminarity significantly decreased with increasing growth rate in hindlimb elements, but no relationship was seen between laminarity and growth rate in forelimb elements. In the femur and tibiotarsus, laminarity significantly increased with increasing shear strain, which supports the hypothesis that laminar bone is a feature of torsionally loaded bone. A moderate amount of bone laminarity was found in the humerus and ulna, despite emu wings being vestigial and likely to experience minimal biomechanical loading, suggesting that laminar bone may in some cases be a retained ancestral feature. In conclusion, torsional loading appears to have a greater impact on vascular canal orientation than growth rate in the femur and

tibiotarsus of the emu.

# **The effects of growth rate and biomechanical loading on bone laminarity within the emu skeleton**

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# 24 ABSTRACT

25       The orientation of vascular canals in primary bone may reflect differences in growth rate  
 26 and/or adaptation to biomechanical loads. Previous studies link specific canal orientations to  
 27 bone growth rates, but results between different taxa are contradictory. Circularly-oriented  
 28 vascular canals (forming laminar bone) have been hypothesized to reflect either (or both) rapid  
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 30 biomechanics in the emu shows that the femur and tibiotarsus experience large shear strains,  
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 46 minimal biomechanical loading, suggesting that laminar bone may in some cases be a retained

47 ancestral feature. In conclusion, torsional loading appears to have a greater impact on vascular  
48 canal orientation than growth rate in the femur and tibiotarsus of the emu.

49

## 50 INTRODUCTION


51 Avian bone tissue is highly vascularized with a fibrolamellar structure, which allows for  
52 rapid growth by depositing randomly arranged woven bone initially, then **filling in** with stronger  
53 more organized lamellar bone (Curry, 2002). Primary osteons are deposited around central canals  
54 that house **blood vessels and nerves**. These vascular canals vary in orientation and bones can be  
55 classified based on the predominant canal orientation. Laminar bone **has an abundance of**  
56 **circular canals**, those that in a bone cross section are oriented parallel to the periosteal surface of  
57 the bone. Additional canal orientations include: radial, those orthogonal to the periosteal surface;  
58 longitudinal, those running parallel to the long axis of the bone; **and oblique, all other**  
59 **orientations** (de Ricqlès et al., 1991). Avian bone tissue largely retains its primary structure  
60 throughout growth and adulthood (Enlow and Brown, 1957). It has been hypothesized that  
61 differences in primary vascular canal orientation might be a reflection **of the functional demands**  
62 **placed on the bone, growth rate, or phylogenetic relationships**.

63 Amprino (1947) first suggested that the organization of bone microstructure may be  
64 influenced by bone growth rate, such that woven bone is deposited during rapid growth and  
65 lamellar bone during slow growth (de Ricqlès et al., 1991). Further studies have investigated  
66 whether specific primary vascular canal orientations in fibrolamellar bone are also associated  
67 with slow or fast growth by directly comparing microstructure with bone growth rates measured  
68 through the use of injectable fluorochromes (Castanet et al., 2000; de Margerie, 2002; de  
69 Margerie et al., 2004). Rapidly growing hindlimb bones of ratites have been found to exhibit




70 structure that is laminar and reticular (bone with an abundance of obliquely oriented canals)  
 71 (Castanet et al., 2000), whereas the much slower growing wing elements of the ratites exhibit  
 72 reticular and longitudinal canal structure. This suggests that laminar bone, in part, may reflect  
 73 faster growth rates. This result was supported in a recent study of pigeon wing elements, which  
 74 showed that peak laminarity (proportion of laminar bone) coincides with the growth spurt in each  
 75 element (Ourfalian, Ezell & Lee, 2016). However, work on mallard long bones showed no  
 76 relationship between growth rate and predominant vascular canal orientation (de Margerie,  
 77 2002). And in the king penguin, radially-oriented canals dominated in the fastest growing  
 78 sections, not circular canals (laminar bone) as was hypothesized. (de Margerie et al., 2004).  
 79 Likewise, chickens selected for fast growth showed tibiotarsi with more radial canals (Williams  
 80 et al., 2004).

81 Laminar bone has been hypothesized to better resist torsional loading. In laminar bone,  
 82 the bone tissue is arranged in 'sheets' or 'plates' between layers of circular canals. Shear strain is  
 83 thought to flow continuously within these 'sheets', and thus the concentrated stresses on the bone  
 84 tissue surrounding the canals is reduced (de Margerie et al., 2004). Indeed, elements that are  
 85 predominantly experiencing torsional loads have been found to exhibit laminar bone. Laminar  
 86 bone is found to be most abundant in the humerus, ulna, and femur in a large sample of flighted  
 87 bird species (de Margerie, 2002; de Margerie et al., 2005). *In vivo* strain gauge studies have  
 88 shown that at least two of these elements, the humerus in the pigeon during flapping flight and  
 89 the femur in the chicken and emu during terrestrial running, experience predominantly torsional  
 90 loads (Biewener and Dial, 1995; Carrano and Biewener, 1999; Main and Biewener, 2007).  
 91 Changes in posture affect the amount of torsion experienced, such that when the center of mass  
 92 of white leghorn chickens was modified, the femur experienced higher torsional loads due to its

 more horizontal orientation. In addition, it has been hypothesized that specific wing shape and the locomotor style associated with it may affect how much torsional load is placed on limb elements. For example, laminarity in the humerus of birds that utilize flapping and static soaring is higher than in the humerus of birds that use dynamic soaring (de Margerie et al, 2005; Simons and O'Connor, 2012).

A limitation of previous studies of laminar bone is the indirect comparison of bone histology in one species with bone growth rates and/or *in vivo* strain gauge measures taken from different species. In this study, we present an analysis of laminar bone in a species in which bone growth rate and *in vivo* bone strain data were directly measured. The emu (*Dromaius novaehollandiae*, Order Struthioniformes, Family Dromaiidae) is a flightless bird endemic to Australia, but widely farmed in the US. The individuals included in this study comprise a growth series that were previously injected with fluorescent bone labels and surgically implanted with gauges to measure *in vivo* locomotor strains in the femur and tibiotarsus (Main and Biewener, 2007). These strains were predominantly shear (produced by torsional loads) and increased from juveniles to adults (Main and Biewener, 2007). Although the wings of these individuals were not measured for bone strain, they are extremely reduced and have no known function other than being raised to aid thermoregulation (del Hoyo et al., 1992; Maxwell and Larsson, 2007).

 Presumably, shear strains are negligible in the wing elements. Therefore, if laminarity is an adaptation to torsion-induced shear strains, we predict that hindlimb bone laminarity will increase from juveniles to adults. Moreover, we expect little to no laminar bone in the vestigial wing elements. Alternatively, if bone laminarity reflects rapid growth, it should be abundant in juveniles and decrease with age as growth slows in adults. In addition, hindlimb elements in emu grow at a faster rate than wing elements (Castanet et al., 2000), so we predict overall greater



116 laminarity in hindlimb than forelimb elements. Having access to growth rate measurements,  
 117 direct biomechanical data, and direct histological work to classify laminarity makes this study a  
 118 first of its kind that will be able to clarify which factors have the most impact on vascular canal  
 119 orientation in emu limbs.

120

## 121 MATERIALS & METHODS

122 This study samples forelimb and hindlimb elements from ten emus ranging in age from 2  
 123 to 416 weeks (Table 1). Birds used in this study were euthanized as part of a previous study  
 124 (Main and Biewener, 2007) and the selected elements stored frozen. Emus were originally  
 125 obtained as hatchlings by R.P. Main (at the time at Harvard University) from commercial farms  
 126 (Songline Emu Farm, Gill, MA, USA; Scattered Oaks Emu Farm, Iola, TX, USA; Deep Hollow  
 127 Farm, Oakdale, CT, USA) and raised at Harvard University's Concord Field Station (Bedford,  
 128 MA, USA)(Harvard FAS IACUC AEP 23-15). For the first eight weeks of life the emus were



129 held in indoor enclosures, and then moved into large outdoor enclosures. All birds had free  
 130 access to commercial ratite diet (Mazuri, PMI Nutrition International, LLC, Brentwood, MO,  
 131 USA) and water. Male and female birds were included based on availability. Emus exhibit a  
 132 minor degree of sexual dimorphism, with females being slightly larger on average (del Hoyo et  
 133 al., 1992). The difference in size is not large enough to be considered a confounding factor for  
 134 this study.

135 As a part of a previous study (Main and Biewener, 2007), each bird was given a single  
 136 intramuscular injection of xylenol orange (80mg/kg) followed by calcein (30mg/kg) according to  
 137 an injection schedule. Injections were given one week apart in birds less than 16 weeks of age,  
 138 two weeks apart in birds between 16 and 65 weeks of age, and four weeks apart in birds older

139 than 65 weeks. Xylenol and calcein are fluorescent labels that incorporated rapidly into newly  
 140 mineralizing surfaces of bone at the time of injection (An and Martin, 2003). Thus, the time  
 141 elapsed and the space between xylenol and calcein labels allows the calculation of periosteal  
 142 (radial) growth rate. One week after the last injection, surgery was performed to attach strain  
 143 gauges to the cranial, caudal, and lateral aspects of the left femur and the cranial, caudal, and  
 144 medial aspects of the left tibiotarsus. Single element strain gauges were used on the lateral femur  
 145 and cranial and medial tibiotarsus. Rectangular rosette gauges were used on the cranial and  
 146 caudal femur and caudal tibiotarsus. Rosette strain gauges allow both tensile and compressive  
 147 principal strains and their orientations to be measured, and were placed so the central element of  
 148 the gauge was parallel to the long axis of the bone. One day after surgery, the birds were run on a  
 149 treadmill over a wide range of speeds and gaits. The raw data produced from the strain gauges  
 150 was converted from voltages into microstrain using a custom MATLAB program. Shear strains  
 151 were calculated from the rosette strain gauges using standard equations (Biewener and Dial,  
 152 1995). High quality shear strain data were most consistently collected from the caudal cortices of  
 153 the femur and tibiotarsus and that is what is reported here. Trials in which the birds ran with a  
 154 duty factor near 0.50 are included in the shear strain analysis (mean  $\pm$  SD:  $0.50 \pm 0.02$ ). Each  
 155 trial was represented by five footfalls and, generally, two trials were collected for each bird.  
 156 Following bone strain data collection, animals were euthanized. After death, whole wings were  
 157 removed from the individuals and stored frozen. Histological sections of the femora and  
 158 tibiotarsi were prepared (see Main and Biewener, 2007 for details) and shipped with the frozen  
 159 wings to Midwestern University.

## 160 Histological Preparation

161 Emu wings were thawed and feathers, skin, muscles, and tendons were reflected to

expose the skeletal elements. Both right and left wings were used based on availability. Using digital calipers, total length of each bone was measured and recorded. A 37-mm segment was removed using a Dremel tool from the mid-shaft region of the humerus, ulna, and radius. For two and four week old individuals whole elements were harvested due to their small size. Segments were marked to maintain orientation. Dissected bone segments were placed in 10% neutral buffered formalin for fixation, then dehydrated in a graded ethanol series (70%, 85%, 100%) under vacuum. Segments were cleared with a xylene-substitute (Histo-clear; National Diagnostics, Atlanta, Georgia, USA). The bone segments were then vacuum- infiltrated and embedded in glass vials using Osteo-Bed Plus Resin, a two-part methyl methacrylate (Polysciences Inc.). Vials were placed in a 32° C bead bath to fully harden.

Once the resin hardened, vials were broken and two roughly 800- $\mu$ m transverse sections were cut using a diamond blade saw (Isomet 1000; Buehler, Lake Bluff, Illinois, USA). These sections were glued onto frosted glass slides using two-ton epoxy (Devcon, Milpitas, California, USA), keeping consistent spatial orientation. Slides were then ground to a thickness of 100 $\pm$ 10 $\mu$ m using a graded scale of grit paper on a stand grinder (Metaserv 250; Buehler, Lake Bluff, Illinois, USA) and coverslipped with Permount (Fisher Scientific). The histological preparation was modified from An and Martin (2003) and closely followed Lee and Simons (2015).


## Image Collection

The undecalcified sections contain xylenol (orange) and calcein (green) fluorochromes that were incorporated into newly mineralizing bone at the time of injection (see above for injection schedule). These fluorochromes create stable long lasting tags (van Gaalen et al., 2010) and were examined under bright-field and fluorescent illumination with a motorized

185 epifluorescent microscope (IX73, Olympus). The xyleneol (orange) and calcein (green) tags were  
 186 revealed using the TRITC and FITC filter cubes, respectively, and a multichannel (red, green,  
 187 bright-field) image of each section was generated with imaging software (cellSens, Olympus).  
 188 Sufficient optical resolution (10X UPlanAPO  $\approx 0.84 \mu\text{m}$ ; 20X UPlan S-APO  $\approx 0.45 \mu\text{m}$ ) allowed  
 189 the dual color-monochrome camera (DP80, Olympus) to capture high quality images (10X =  
 190  $1.02 \mu\text{m}/\text{pixel}$ ; 20X =  $0.51 \mu\text{m}/\text{pixel}$ ).

# 191 Data Analysis

192 Bright-field and fluorescent images were obtained from the wing and hindlimb elements  
 193 (Figs. 1 and 2) and divided into equal octants from the estimated bone centroid. Four octants  
 194 representing the cardinal anatomical positions (wing elements: cranial, caudal, dorsal, ventral;  
 195 hindlimb elements: cranial, caudal, lateral, medial) were extracted. Using ImageJ, each extracted  
 196 octant was then uncurved using the “Straighten” function. The purpose of straightening was to  
 197 standardize the periosteal tangent line so that appropriate measurements could be made in  
 198 classifying the orientation of the vascular canals (Lee and Simons, 2015). To ensure there was  
 199 minimal deformation of the image during the straightening process, known test angles were  
 200 placed upon the image and measured in relation to the periosteal surface after the straightening  
 201 function had been applied. Only those images with an average deformation less than or equal to  
 202  $10^\circ$  were accepted.

203 Within each of the four octants, the calcein green and xyleneol orange tags were outlined  
 204 with two reference lines.  The distance between reference lines was measured at 10 random  
 205 points in each octant. Growth rate was measured by taking the mean distance between  
 206 consecutive fluorescent tags divided by number of days between injections (Fig. 2).

207 Degree of laminarity (Laminarity Index, LI) for each octant was assessed in two distinct




sample areas. To address the growth hypothesis, laminarity was measured in the area between the fluorochrome reference lines in all four octants. To address the biomechanical hypothesis, laminarity was measured in four sample boxes (1.5 x 1.5 mm each) placed in the caudal bone cortex to represent the entire bone cortex (hindlimbs only). To measure laminarity, an ellipse was drawn in each primary vascular canal in the designated sample area of the straightened octant. The angle at which the ellipse sat in relation to the straightened periosteal surface was measured. We used the criteria set forth by de Margerie (2002) to classify the orientation of the vascular canals: (1) circular canals are oriented parallel ( $0^\circ \pm 22.5^\circ$ ) to the periosteal surface of the bone; (2) radial canals are orthogonal ( $90^\circ \pm 22.5^\circ$ ) to the periosteal surface; (3) longitudinal canals run parallel to the long axis of the bone and have ellipses with an aspect ratio of less than 3; (4) oblique canals are all other orientations. Only primary vascular canals were measured. Any secondary osteons in the sample area were excluded. Laminarity index (proportion of circular canals to the total number of canals) was calculated using the Wilson estimate to account for bias from small sample sizes (Lee and Simons, 2015).


Statistical analyses were performed using R (R Development Team, 2017). Logistic regression was used because laminarity index is a proportion and may not be normally distributed; logistic regression relaxes the assumption that the data are normally distributed (Warton and Hui, 2011; Ourfalian, Ezell & Lee, 2016). To test the growth hypothesis, we performed logistic regression between laminarity indices of each bone and growth rate. To test for correlation between laminarity and shear strain, we focused on the data obtained from the caudal cortex in the femur and tibiotarsus. Average laminarity indices from the four sample boxes placed in the caudal octant of the hindlimb elements for each bird were logistically regressed against a mean for the caudally measured shear strains from two experimental trials,

231 previously collected by Main and Biewener (2007).

232

## 233 RESULTS

234 Measured growth rates ranged from 1.33  um/day (radius of 48 week old individual) to  
 235 162.62  um/day (femur of 4.6 week old individual) (Table 2). Negligible growth was recorded in  
 236 the eight-year-old individuals. Laminarity indices range from 0.02 to 0.58 as measured for the  
 237 growth hypothesis (Table 2) and 0.03 to 0.79 as measured for the biomechanical hypothesis  
 238 (Table 3). 

239 Logistic regression between LI and growth rate (GR) showed a significant negative  
 240 relationship in the femur ( $p < 0.001$ ) and tibiotarsus ( $p < 0.001$ ) (Fig. 3, Table 2). No significant  
 241 relationship was found between laminarity and growth rate in wing elements (Fig. 4, humerus:  
 242  $p = 0.08$ , ulna:  $p = 0.29$ , radius:  $p = 0.52$ ). 

243 Logistic regression between caudal octant LI and shear strain showed a significant  
 244 positive relationship in both the femur ( $p < 0.001$ ) and tibiotarsus ( $p < 0.001$ ) (Fig. 5, Table 3).

245

## 246 DISCUSSION

### 247 Does bone laminarity reflect fast growth?

248 The fastest growth rate in all elements was found in the 4.6-week old individual (Table  
 249 2). As expected, hindlimb elements had higher growth rates than forelimb elements, reaching a  
 250 maximum of 163 um/day in the femur and 99 um/day in the tibiotarsus. The humerus grew the  
 251 fastest of the wing elements, reaching a maximum rate of 25 um/day measured in the 2.3 and 4.6  
 252 week old individual. Birds older than 8 weeks experienced a drastic decrease in bone growth rate  
 253 in both hindlimb and forelimb elements. Previous analysis of emu somatic growth rate showed

254 the maximum velocity of growth (inflection point) to be about 15-17 weeks of age  
 255 (Goonewardene et al., 2003). Our results suggest maximum bone growth occurs much earlier  
 256 (about 5 weeks) and is not an accurate proxy for the somatic growth inflection.

257 There was a significant negative relationship between bone laminarity and growth rate in  
 258 both hindlimb elements (Fig. 3). In the hindlimb, the lowest laminarity was found in the  
 259 youngest birds, where the growth rate was the fastest. This is consistent with findings in the king  
 260 penguin that also reported laminar bone to be associated with slower growth rates in four limb  
 261 bones: femur, tibiotarsus, humerus, and radius (de Margerie et al., 2004). Notably, our results  
 262 differ from those previously reported for young emu bones in which laminar and reticular bone  
 263 was found in the fastest growing hindlimb bones (Castanet et al., 2000). In particular, Castanet et  
 264 al. (2000) found laminar bone to be most abundant in the femur and tibiotarsus of emu less than  
 265 2 months of age, which corresponds to the youngest individuals in our study. The emus included  
 266 in our study grew in mass about 3 times faster than the emus in the Castanet et al. (2000) study.

267 If laminarity is associated with slower growth rates, the youngest emus we studied may have  
 268 been growing too fast for laminar bone to form. In contrast, Castanet et al. (2000) found reticular  
 269 bone to be more abundant in the humerus. Our study did not specifically address reticular bone,  
 270 but by taking the proportion of oblique vascular canals (a “reticular index”), we found the  
 271 amount of reticular bone in the fastest growing individual to be low in the hindlimb elements  
 272 (femur and tibiotarsus: 0.17), and moderate to high in the wing elements (humerus: 0.62, ulna:  
 273 0.58, radius: 0.45). This result is, at least, consistent with the previous study. The data analysis in  
 274 the earlier emu study (Castanet et al., 2000) was conducted before a more rigorous method for  
 275 measuring vascular canals was developed (de Margerie, 2002), which may also affect differences  
 276 seen between the two separate emu populations. Regardless, the results of the direct laminarity

277 indices and growth rates measured here do not support the hypothesis that high laminarity  
278 reflects rapid growth in the emu.

# 279 **Does bone laminarity reflect biomechanical load?**

280 Within the caudal octants of the hindlimb elements, we found a positive correlation  
281 between laminarity and shear strain (Fig. 5). This result supports the hypothesis that laminar  
282 bone may reflect a specific type of biomechanical load - torsion. The amount of shear strain  
283 occurring at the midshaft of the emu femur and tibiotarsus has been found to significantly  
284 increase with age by 2-3 times, whereas body mass increases by 46 times. Laminar bone, in  
285 combination with increased bone mineralization and decreased bone curvature during growth,  
286 may help mitigate shear strains despite the large increase in mass (Main and Biewener, 2007).

287 Our results support the hypothesis that laminar bone is a feature of torsionally loaded  
288 bone and strengthens the premise that bone histology can be used to estimate the degree of shear  
289 strain experienced by bones during different types of locomotion, such as flight modes (de  
290 Margerie et al., 2005; Simons and O'Connor, 2012). Although strain gauge technology is  
291 improving, shear strains are not easily measured for some types of locomotion, especially flight.  
292 In addition, this correlation between laminarity and shear strain could help researchers better  
293 understand locomotive patterns of extinct animals that are closely related to Aves, such as other  
294 extinct maniraptorans (the clade of dinosaur that includes birds). Not all members of the  
295 maniraptoran clade were flighted animals (Makovicky and Zanno, 2011), but as one moves up  
296 the phylogeny from non-flighted to flighted animals we may expect to see an increase in  
297 laminarity within at least the humerus an adaptation to the larger expected torsional loads  
298 experienced during active flight. Indeed, the humerus of Archaeopteryx appears to have  
299 predominately longitudinal canals despite having cross-sectional geometry similar to birds that

300 use active flapping (Voeten et al., 2018), but further up the paravian tree, *Confuciusornis* shows  
301 laminar-reticular bone in the humerus (de Ricqlès et al. 2003).

302 However, this histological proxy for torsional load should be used with caution. Previous  
303 studies have shown that when wings have the same general shape, laminarity in wing bones is  
304 similar despite differences in primary flight mode (Simons and O'Connor, 2012; Marelli and  
305 Simons, 2014). In addition, preferred flight mode may only have subtle effects on overall  
306 loading of the bones, with the dominant loads being the high strains present during take-off  
307 (Biewener and Dial, 1995). There is also at least one example of a bone loaded in predominantly  
308 torsion that does not exhibit laminar bone. The humeri of bats show an absence of laminar bone  
309 despite being loaded in torsion during flapping flight, presumably due to the slow somatic  
310 growth rates of bats (Swartz, Bennet & Carrier, 1992; Lee and Simons, 2015). Additionally,  
311 laminar bone may be present in the absence of dominant torsional loads. The radius of the goat  
312 qualitatively appears laminar during growth and early adulthood despite experiencing  
313 predominantly bending loads (Main, 2007).

314 The mean laminarity across all ages of emu was statistically larger for the femur (0.41)  
315 than the tibiotarsus (0.28,  $p=0.007$ ). Generally, the femur has also been shown to experience a  
316 larger magnitude of shear strain than the tibiotarsus (Main and Biewener, 2007). Both elements  
317 exhibited a large degree of variation, with the laminarity indices of the femur ranging from 0.02 -  
318 0.56 and the tibiotarsus from 0.02 - 0.58 through ontogeny (Table 2). Interestingly, in the  
319 tibiotarsus, a wide range of variation in laminarity can be seen for similar shear strain values  
320 (Fig. 5). Laminarity measured on a histological section is a 2-dimensional representation of a 3-  
321 dimensional meshwork of vascular canals in cortical bone. This research is limited by the  
322 assumption that one or two closely placed mid-shaft histological sections are an accurate

323 representation of vascular canal structure. Future studies should incorporate new methods, such  
 324 as microCT (e.g., Pratt and Cooper, 2017), to assess how well laminarity measured on  
 325 histological sections represents actual biological structure.

326 Although there is no direct biomechanical data for the forelimb elements of these birds,  
 327 the wing elements presumably experience minimal loading. The emu wing is extremely reduced  
 328 in size, even when compared to other ratites, and no wing function has been observed except to  
 329 be raised for thermoregulation (del Hoyo et al., 1992). Wing muscles of emu are primarily slow  
 330 acting tonic muscle fibers that may not allow much wing movement (Maxwell and Larsson,  
 331 2007), which suggests the underlying wing elements would experience minimal biomechanical  
 332 loading. Interestingly, despite the assumption that the emu wing is under minimal load, a  
 333 moderate to high degree of laminarity was found in at least the humerus and ulna (Table 2).

334 Within the paleognaths, it has been hypothesized that at least three independent flight  
 335 losses have occurred, with only one order (the tinamous) still retaining the ability (Harshman et  
 336 al., 2008; Mitchell et al., 2014). The moderate/high wing bone laminarity may be a feature of the  
 337 flighted common ancestor of paleognaths that is retained in the flightless descendants. Indeed,  
 338 significant phylogenetic signal has been found in some osteohistological features in a sample of  
 339 paleognaths (Legendre et al., 2014). Future studies should investigate the histological and *in vivo*  
 340 loading of the flighted relatives of emus to better understand the potential influence of phylogeny  
 341 on bone laminarity.

342

## 343 CONCLUSIONS

344 For the emu skeleton, torsional loading appears to have a greater impact on vascular  
 345 canal orientation than growth rate. The increase of laminarity with increasing shear strain

suggests that laminar bone may be an adaptation to resist torsional loads in the femur and tibiotarsus. For the humerus and ulna, moderate to high laminarity may have been inherited from a flighted ancestor. Future studies should investigate laminarity in other palaeognathous birds to better understand the interplay among phylogeny, ontogeny, and torsional loading on bone laminarity. Other future work should focus on the experimental manipulation of biomechanical loads to observe the effects on vascular canal orientation in limb bones. This could be done using a similar study design, but with the addition of individuals that have been weighted (to increase normal biomechanical loads) and/or undergone limb immobilization (to reduce normal biomechanical loads) to better understand to what extent the amount of torsional load has on the development of limb bone laminarity. It is also important that variation found between different populations be addressed and studied further. Emu body mass growth rates vary among populations (e.g. Goonewardene et al., 2003), but it is unknown if bone histology also varies with geography.

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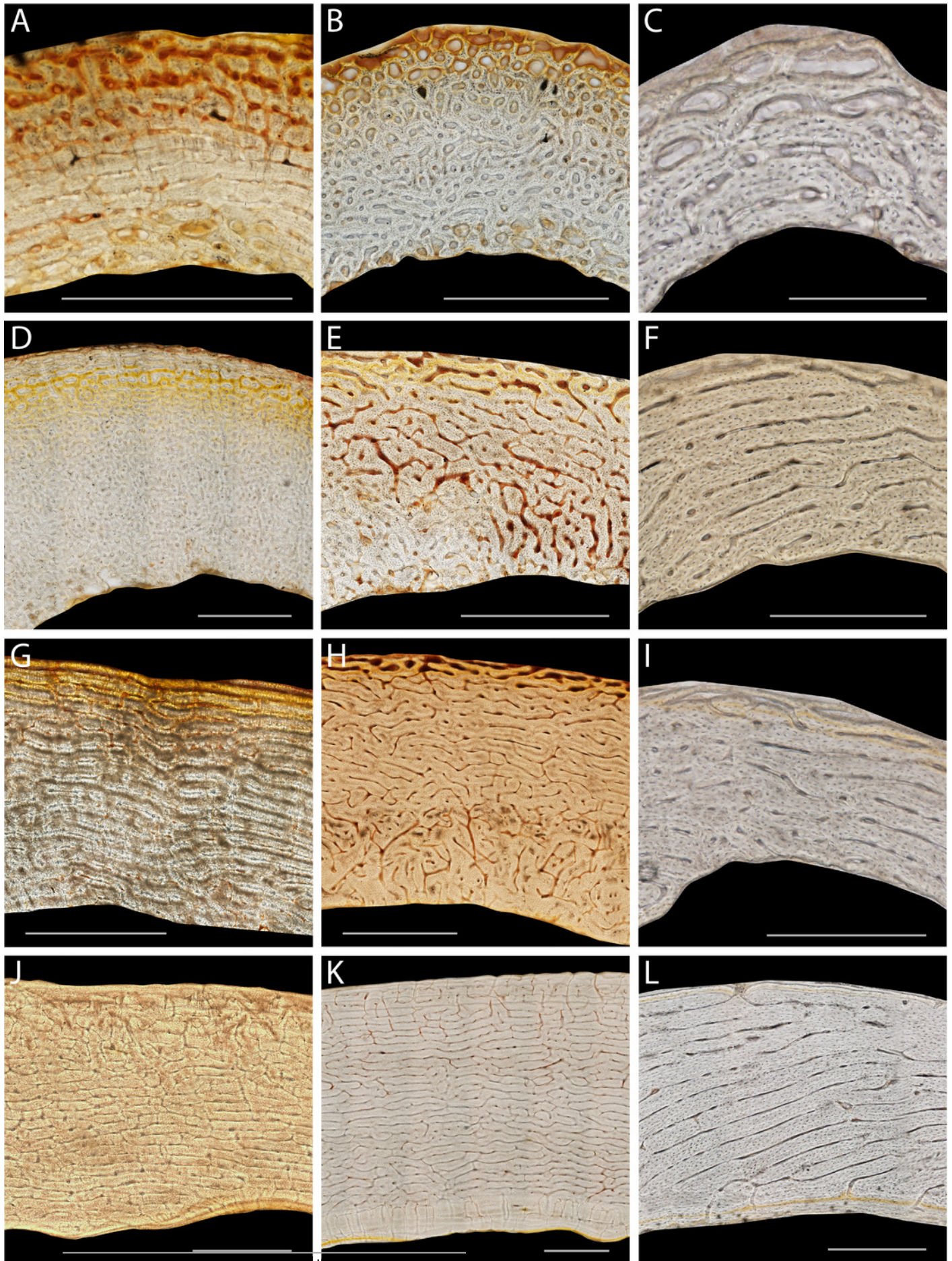
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# Figure 1

Representative histological sections of emu femora, tibiotarsi, and humeri from a range of ages.

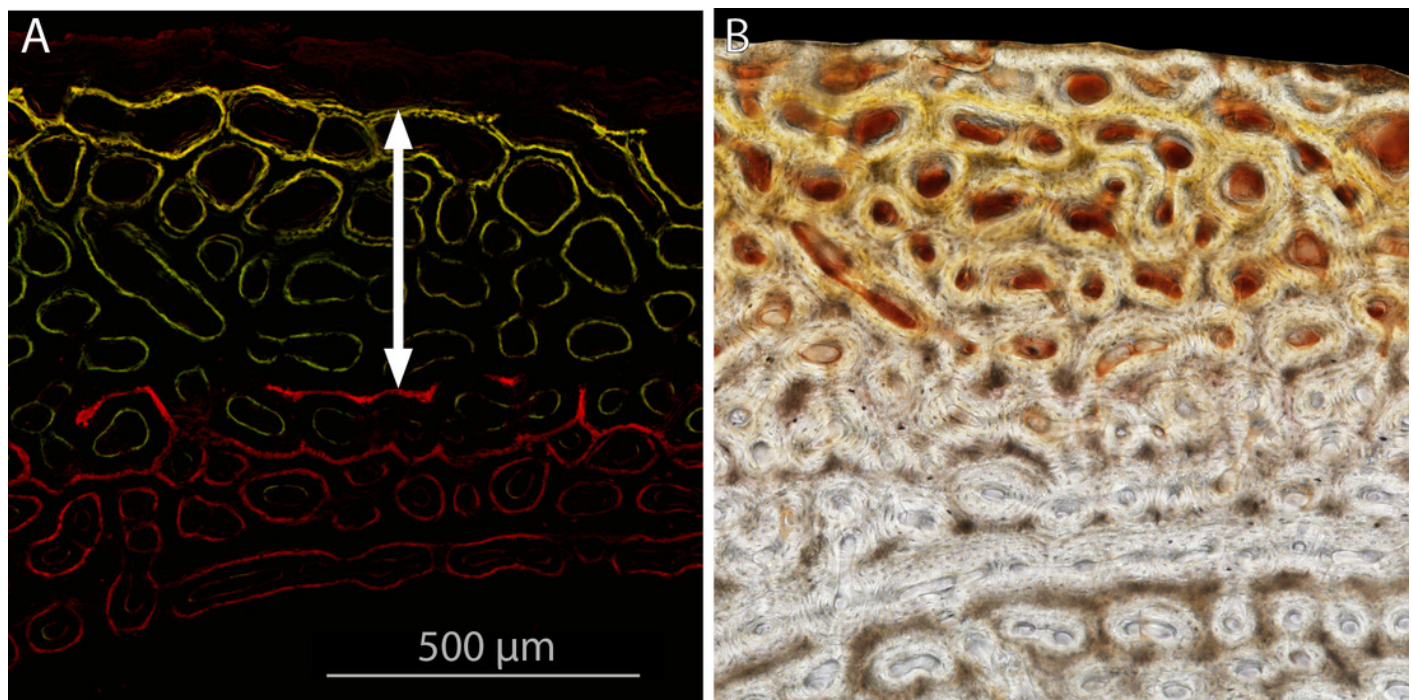
2.3 weeks (A, B, C), 8.1 weeks (D, E), 12 weeks (F), 16 weeks (G, H, I), 60 weeks (L), 416 weeks (J, K). Femora (A, D, G, J), tibiotarsus (B, E, H, K) and humeri (C, F, I, L). Scale bars equal 1000  $\mu\text{m}$  for femora and tibiotarsi, and 250  $\mu\text{m}$  for humeri. Bright field images of non-straightened caudal or medial octants (10x magnification).



# Figure 2

Histology of fluorescent-labelled bone.

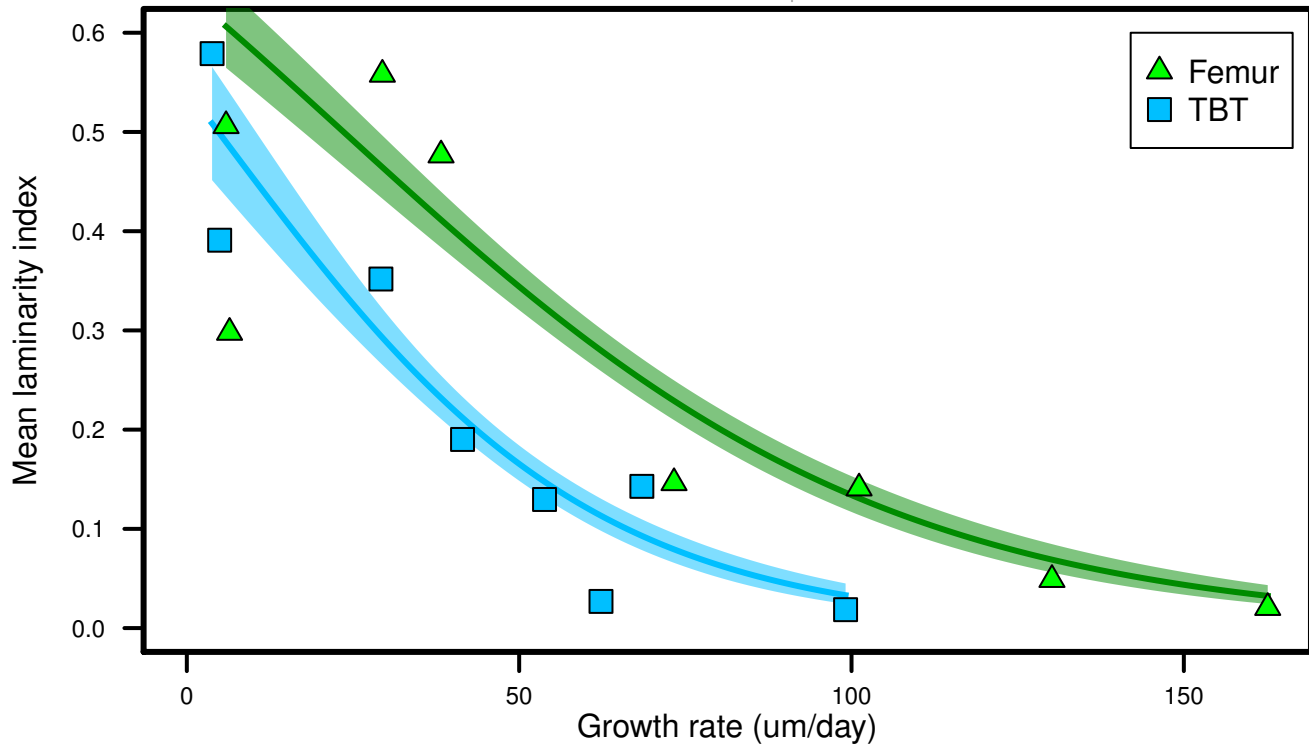
On fluorescent images (A), the sample area was defined by outlining the periosteal extent of the xylenol (appears red) and calcein (green/yellow) tags. Growth rate was measured by taking the mean distance between consecutive fluorescent tags (white arrow) divided by number of days between injections. On brightfield images taken under the same magnification (B), laminarity was measured in the corresponding sample area.



# Figure 3(on next page)

Laminarity of the femur and tibiotarsus (tbt) plotted against growth rate.

Dark green and blue lines represent mean plot lines, shaded areas reflect 95% confidence interval bands. Logistic regression shows significant negative relationship (femur,  $p < 0.001$ ; tbt,  $p < 0.001$ ; pseudo  $R^2 = 0.78$ ). As growth rate increases, bone laminarity decreases. These results do not support the hypothesis that high laminarity is a result of fast growth rate in the emu hindlimb.

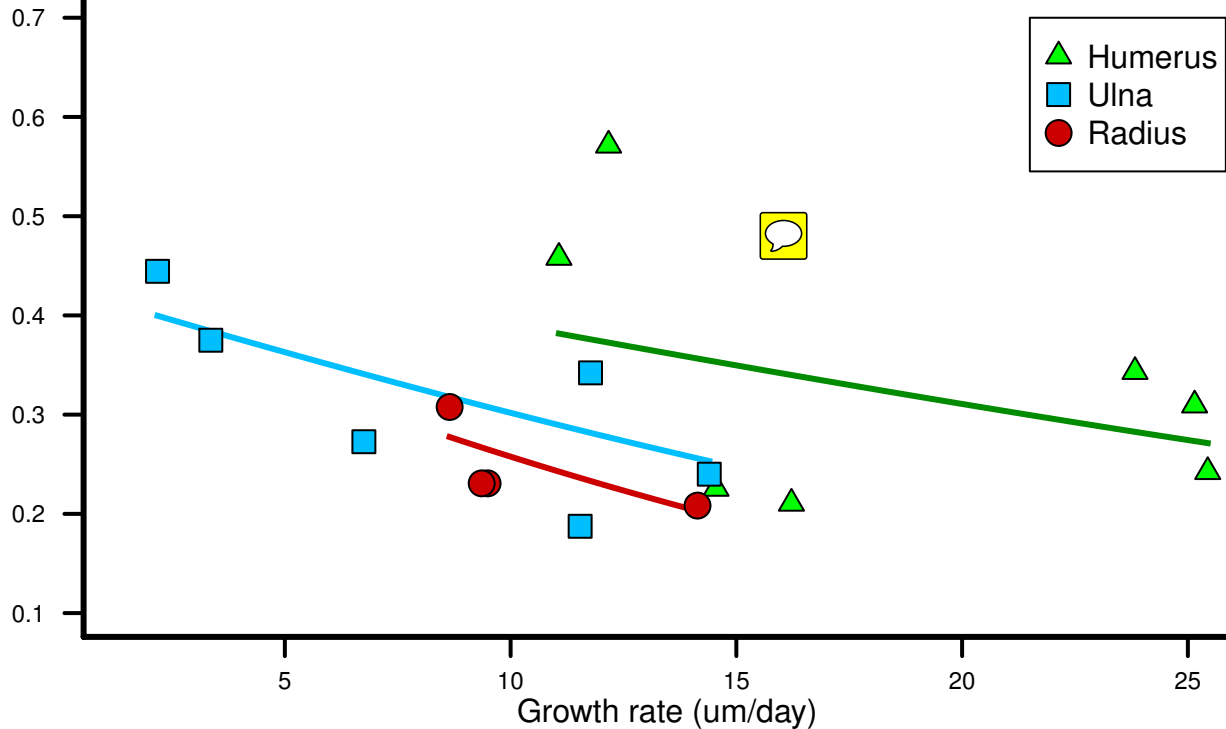


# Figure 4(on next page)

Laminarity of the humerus, ulna, and radius plotted against growth rate.

Dark green, blue, and red lines represent mean plot lines. Logistic regression shows no significant relationship between laminarity and growth rate in wing elements (humerus:  $p=0.08$ , ulna:  $p=0.29$ , radius:  $p=0.52$ ). These results do not support the hypothesis that high laminarity is a result of fast growth rate in the emu forelimb.

Mean lamnarity index

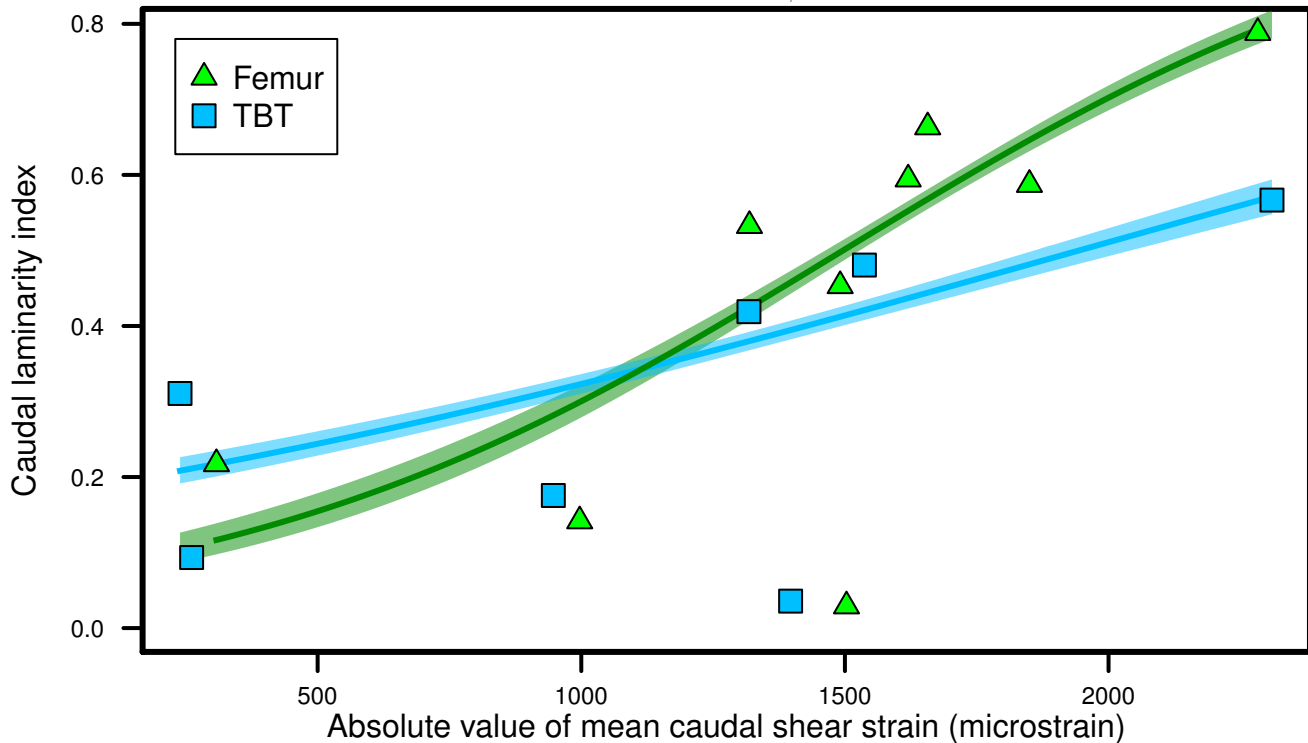


# Figure 5(on next page)

Bone laminarity from the caudal octant of femur and tibiotarsus (tbt) plotted against shear strains measured at the caudal periosteal surface.



Dark green and blue lines represent mean plot lines, shaded areas reflect 95% confidence interval bands. Logistic regression shows significant positive relationship (femur:  $p < 0.001$ ; tbt:  $p < 0.001$ ; pseudo  $R^2 = 0.60$ ). As shear strain increases, bone laminarity increases. This result supports the hypothesis that high laminarity is a feature of bones experiencing greater shear strains.



**Table 1**(on next page)

Emu identification number, age at sacrifice, and mass.

1

<b>Specimen</b>	<b>Age (weeks)</b>	<b>Mass (kg)</b>
15	2.3	0.74
1c	2.4	0.94
17	4.6	1.53
14b	8.1	4.73
16	12	6.85
2a	15.9	11.13
21	48	28.9
23	60.1	29.4
27	416	50.9
28	416	51.7



2

3

4

## Table 2 (on next page)

Growth rate and bone laminarity for specimens analyzed to address the growth hypothesis.

Laminarity Index (LI, calculated using the Wilson's estimate)  was measured in a sample area outlined by the periosteal extent of two fluorescent tags. No growth was present in adult specimens #27 and 28.  n/a indicates a specimen with no recorded circular canals.

1

Specimen	Age (weeks)	Element	Growth Rate (um/day)	LI
15	2.3	Femur	130.16	0.05
		Tibiotarsus	62.31	0.03
		Humerus	25.44	0.24
		Ulna	6.75	0.27
		Radius	15.22	0.20
1c	2.4	Femur	73.3	0.15
		Tibiotarsus	53.82	0.13
		Humerus	16.22	0.21
		Ulna	11.57	0.29
		Radius	9.49	0.25
17	4.6	Femur	162.62	0.02
		Tibiotarsus	99.11	0.02
		Humerus	25.15	0.31
		Ulna	11.54	0.19
		Radius	9.36	0.23
14b	8.1	Femur	101.14	0.14
		Tibiotarsus	68.46	0.14
		Humerus	23.83	0.34
		Ulna	14.4	0.24
		Radius	14.14	0.21
16	12	Femur	38.25	0.48
		Tibiotarsus	41.48	0.19
		Humerus	12.17	0.57
		Ulna	11.77	0.34
		Radius	8.65	0.31
2a	15.9	Femur	29.42	0.56
		Tibiotarsus	29.2	0.35
		Humerus	11.07	0.46
		Ulna	3.36	0.38
		Radius	2.37	0.29
21	48	Femur	6.43	0.30
		Tibiotarsus	4.94	0.39
		Humerus	14.55	0.23
		Ulna	2.18	0.44
		Radius	1.33	0.33

23	60.1	Femur	5.9	0.51
		Tibiotarsus	3.8	0.58
		Humerus	1.7	0.25
		Ulna	1.62	0.33
		Radius	1.67	0.40

2

### **Table 3**(on next page)

Bone laminarity and caudal shear strain for specimens analyzed to address the biomechanical hypothesis.

Shear strain data was previously collected by Main and Biewener (2007). Laminarity Index (LI, calculated using the Wilson's estimate) was measured in four representative sample boxes (1.5 x 1.5 mm each) placed throughout the cortex on each caudal octant.

1

Specimen	Age (weeks)	Element	LI	Caudal Shear Strain (microstrain)
15	2.3	Femur	0.13	N/A
		Tibiotarsus	0.10	N/A
1c	2.4	Femur	0.22	-308
		Tibiotarsus	0.12	N/A
17	4.6	Femur	0.03	-1503
		Tibiotarsus	0.04	-1397
14b	8.1	Femur	0.14	-997
		Tibiotarsus	0.09	-261
16	12	Femur	0.45	-1491
		Tibiotarsus	0.18	-947
2a	15.9	Femur	0.59	-1620
		Tibiotarsus	0.31	-293
21	48	Femur	0.66	-1657
		Tibiotarsus	0.42	-1318
23	60.1	Femur	0.79	-2283
		Tibiotarsus	0.55	N/A
27	416	Femur	0.53	-1319
		Tibiotarsus	0.48	-1537
28	416	Femur	0.59	-1850
		Tibiotarsus	0.57	-2310

2

3