# Peer

# Sargassum blooms in the Caribbean alter the trophic structure of the sea urchin Diadema antillarum

Nancy Cabanillas-Terán<sup>1</sup>, Héctor A. Hernández-Arana<sup>2</sup>, Miguel-Ángel Ruiz-Zárate<sup>2</sup>, Alejandro Vega-Zepeda<sup>2</sup> and Alberto Sanchez-Gonzalez<sup>3</sup>

<sup>1</sup> Consejo Nacional de Ciencia y Tecnología México- El Colegio de la Frontera Sur, Chetumal, Quintana Roo, México

<sup>2</sup> Departamento de Sistemática y Ecología Acuática, El Colegio de la Frontera Sur, Chetumal, Quintana Roo, México

<sup>3</sup> Centro Interdisciplinario de Ciencias Marinas del Instituto Politécnico Nacional, La Paz, Baja California, México

# ABSTRACT

The arrival of large masses of drifting Sargassum since 2011 has caused changes in the natural dynamics of Caribbean coastal ecosystems. In the summer of 2015, unprecedented and massive mats of S. fluitans and S. natans have been observed throughout the Mexican Caribbean including exceptional accumulations ashore. This study uses stable isotopes to assess the impact of Sargassum blooms on the trophic dynamics of the Diadema antillarum sea urchin, a keystone herbivore on many Caribbean reefs. Bayesian models were used to estimate the variations in the relative proportions of carbon and nitrogen of assimilated algal resources. At three lagoon reef sites, the niche breadth of D. antillarum was analysed and compared under massive influx of drifting Sargassum spp. vs. no influx of Sargassum blooms. The effects of the leachates generated by the decomposition of Sargassum led to hypoxic conditions on these reefs and reduced the taxonomic diversity of macroalgal food sources available to D. antillarum. Our trophic data support the hypothesis that processes of assimilation of carbon and nitrogen were modified under Sargassum effect. Isotopic signatures of macroalgae associated with the reef sites exhibited significantly lower values of  $\delta^{15}N$ altering the natural herbivory of D. antillarum. The Stable Isotopes Analysis in R (SIAR) indicated that, under the influence of Sargassum blooms, certain algal resources (Dictyota, Halimeda and Udotea) were more assimilated due to a reduction in available algal resources. Despite being an abundant available resource, pelagic Sargassum was a negligible contributor to sea urchin diet. The Stable Isotope Bayesian Ellipses in R (SIBER) analysis displayed differences between sites, and suggests a reduction in trophic niche breadth, particularly in a protected reef lagoon. Our findings reveal that Sargassum blooms caused changes in trophic characteristics of D. antillarum with a negative impact by hypoxic conditions. These dynamics, coupled with the increase in organic matter in an oligotrophic system could lead to reduce coral reef ecosystem function.

Subjects Ecology, Ecosystem Science, Marine Biology, Natural Resource Management, Environmental Impacts

**Keywords** Echinoids, Pelagic macroalgae, Stable isotopes, Trophic ecology, Coral reefs, Niche breadth, Mexican caribbean

Submitted 6 March 2019 Accepted 30 July 2019 Published 30 August 2019

Corresponding authors Nancy Cabanillas-Terán, ncabanillas@ecosur.mx Héctor A. Hernández-Arana, hhernand@ecosur.mx

Academic editor Salvador Sánchez-Carrillo

Additional Information and Declarations can be found on page 22

DOI 10.7717/peerj.7589

© Copyright 2019 Cabanillas-Terán et al.

Distributed under Creative Commons CC-BY 4.0



# INTRODUCTION

The arrival of massive amounts of pelagic *Sargassum* spp. has caused changes in the natural benthic dynamics of Caribbean coastal ecosystems for the last nine years (*Gower*, *Young & King*, 2013; *Schell*, *Goodwin & Siuda*, 2015). Pelagic *Sargassum* is a complex of two species, namely *S. fluitans* and *S. natans* (*Oyesiku & Egunyomi*, 2014). Since 2011, extensive masses of *Sargassum* appeared in unusual ways in oceanic waters off northern Brazil (*De Széchy et al.*, 2012; *Sissini et al.*, 2017), along the West Indies and Caribbean coasts (*Gower*, *Young & King*, 2013) from Trinidad to the Dominican Republic (*Rodríguez-Martínez*, *van Tussenbroek & Jordán-Dahlgren*, 2016; *van Tussenbroek et al.*, 2017), and along the west African coast from Sierra Leone to Ghana (*Smetacek & Zingone*, 2013). *Wang et al.* (2019) recorded that for June 2018, wet biomass reached more than 20 million tons in the Caribbean Sea and Central Atlantic Ocean.

The Mexican Caribbean shores faced atypical massive mats of pelagic Sargassum in the summer of 2015 (van Tussenbroek et al., 2017; Cuevas, Uribe-Martínez & Liceaga-Correa, 2018; Arellano-Verdejo, Lazcano-Hernandez & Cabanillas-Terán, 2019). There was a subsequent decrease during 2016 and 2017, but for most of 2018 and thus far in 2019 influx has increased again. Several studies revealed that these massive mats of Sargassum have a new possible distribution source different from the historic North Atlantic Recirculation Region (NARR) known as "The Sargasso Sea" (Schell, Goodwin & Siuda, 2015). Instead, the most probable origin of the massive influx on the Caribbean shores is the North Equatorial Recirculation Region (NERR) (Johnson et al., 2013; Schell, Goodwin & Siuda, 2015). High oceanic temperatures and nutrient inputs (Franks, Johnson & Ko, 2016; Wang et al., 2018), among other oceanographic coupled patterns such as changes of surface currents, are the most probable causes of this new region of Sargassum flourishment (Johnson et al., 2013; Gower, Young & King, 2013; Sissini et al., 2017). A recent study by Wang et al. (2019) revealed that increases of pelagic Sargassum are driven by upwelling off West Africa during the boreal winter and by Amazon River discharge during the spring and summer. The authors state that recurrent blooms in the Caribbean Sea and tropical Atlantic are likely, highlighting the importance for understanding their effects on existing ecosystems for future planning.

Changes in habitat structure can directly influence trophic dynamics (*Hunter & Price,* 1992; *Sweatman, Layman & Fourqurean, 2017*) and have been shown to cause synergistic effects on coral reefs (*Smetacek & Zingone, 2013*). For example, harmful macroalgae blooms have been recognized as drivers of degradation in coral reef habitats (*Lapointe et al., 2005*). This has effects on the diversity of reef biota (*Bauman et al., 2010; Louime, Fortune & Gervais, 2017*) like variations in the sea urchin populations (*Lapointe et al., 2010*). The carbon and nitrogen flow by macroalgae blooms likely has adverse effects at different scales. Such disturbances from *Sargassum*, coupled with pre-existing threats on coral reefs, add to the drivers of Anthropocene reef degradation (*Alvarez-Filip et al., 2011; Cramer et al., 2012*).

The massive decomposition of *Sargassum* has negative impacts not only on tourism and local fisheries, but on nearshore ecosystems (*Solarin et al., 2014; Louime, Fortune &* 

*Gervais, 2017*). However, few studies assess the trophic impact of *Sargassum* blooms on benthic communities. Pelagic *Sargassum* and their attached epiphytic algae can contribute new organic matter to these communities (*Rooker, Turner & Holt, 2006; Wang et al., 2018*). Therefore, we consider whether or not these new sources of nitrogen and carbon act in a detrimental manner on the trophic chain of benthic communities. The beaching and decomposing of massive *Sargassum* mats produce hypoxia in near-shore coral reef communities (*Rodríguez-Martínez et al., 2019*). This effect coupled with high hydrogen sulfide and ammonium concentrations have been shown to cause faunal mortality in the Mexican Caribbean (*Rodríguez-Martínez et al., 2019*). As a consequence, the coastal environment becomes even more sensitive to degradation agents. To assess these issues, we included measurements of dissolved oxygen in our study.

Evaluating consumers and resources through a trophic approach by tracking the relationships between consumers and prey provides relevant information on the trophic structure and dynamics of a benthic community (*Minagawa & Wada, 1984; Vanderklift, Kendrick & Smit, 2006; Behmer & Joern, 2008*). Stable isotopes of carbon ( $\delta^{13}$ C) and nitrogen ( $\delta^{15}$ N) have been used in marine ecosystems to determine the feeding habits of species (*Peterson & Fry, 1987*), nutrient migrations within food webs, trophic position of organisms and their contribution at all trophic levels (*Vander Zanden & Rasmussen, 1996*). It is also possible to trace the origin and transformation of the ingested organic matter and to detect changes in the trophic positions of organisms that coexist in the same habitat (*Hobson, 1999; Vanderklift, Kendrick & Smit, 2006; Rodríguez-Barreras et al., 2016*).

Stable carbon and nitrogen isotope ratios provide time-integrated information regarding feeding relationships and energy flow through food webs (DeNiro & Epstein, 1981; Peterson & Fry, 1987; Vander Zanden & Rasmussen, 2001). Moreover, stable isotopes can be used to study the trophic niche breadth of a species (Bearhop et al., 2004; Parnell et al., 2010; *Phillips et al.*, 2014). This is directly influenced by consumers and resource input, providing a quantitative assessment of trophic conditions (*Newsome et al.*, 2007; *Boecklen et al.*, 2011). Stable isotope analyses are useful for assessing the health of ecosystems because it is possible to associate the consumers trophodynamics and niche breadth with habitat disturbances (Layman et al., 2007b; Hamaoka et al., 2010). It is also possible to detect changes in the trophic spectrum from anthropogenic impacts or unusual conditions that cause shifts in ecosystems (Wing et al., 2008; Prado, Alcoverro & Romero, 2010; Tomas, Box & Terrados, 2011). In light of the massive arrival of pelagic macroalgae, sea urchin herbivory is a good model to understand variability in the benthic trophic chain, as sea urchins are considered generalist consumers with a plastic feeding habit (Lawrence, 1975; Vanderklift, Kendrick & Smit, 2006). Echinoids have the capability to modify the community structure through foraging behaviour (Carpenter, 1986; Hay & Fenical, 1988; Sala et al., 1998; Eklöf et al., 2008). Thus, the relative position of  $\delta^{13}$ C vs.  $\delta^{15}$ N echinoids displayed in a bi-plot can give insights about organism responses to niche shifts, diet variability and habitat modification (Layman et al., 2007a; Layman et al., 2007b; Layman et al., 2012; Sweatman, Layman & Fourgurean, 2017).

The effect of *Sargassum* and their leachates on the diet of *D. antillarum* can improve our understanding on the impact on trophic ecology of one of the most important sea

urchins of the Mexican Caribbean. The main reason to focus this study on *D. antillarum* is that this species is and was the major shallow-hard-bottom grazer in our study sites (*Jorgensen, Espinoza-Ávalos & Bahena-Basave, 2008; Jordán-Garza et al., 2008*). One of the most dramatic events in the Caribbean resulted from the pathogen-driven reduction in the populations of *D. antillarum* (*Lessios et al., 1984*) with detrimental ecological consequences like coral-algal phase-shifts. The southern part of Quintana Roo is not an exception encompassing with the effects of the abrupt coastal development and watershed pollution as key drivers along the Costa Maya (*Arias-González et al., 2017*).

The overarching aim of this study was to determine variations in the relative proportions of carbon and nitrogen of assimilated algal resources and the niche breadth of *D. antillarum* under massive influx of drifting *Sargassum* spp. vs. no influx of *Sargassum* at back reefs. We also aimed to determine whether pelagic *Sargassum* was a substantial source of energy for *D. antillarum*. To do this, we compared  $\delta^{15}$ N and  $\delta^{13}$ C values of *D. antillarum* with and without influx of *Sargassum* to track changes in this species trophic ecology (diet, trophic position and niche breadth). Ultimately, we tested the hypothesis that an influx in *Sargassum* in coastal ecosystem creates a significant change in the available algal sources and a shift in the trophic structure.

# **MATERIAL & METHODS**

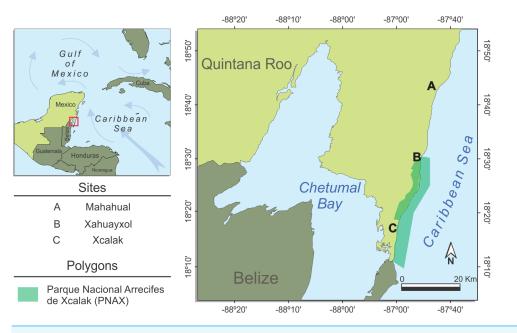
#### Study sites

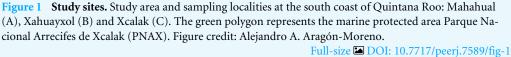
We determined the stable isotopes of carbon and nitrogen for *D. antillarum* at three reef lagoons (Mahahual, Xahuayxol, and Xcalak) with different distances from the beach to the reef crest (Fig. 1). The main strategy implemented by local authorities at some beaches with the massive arrival of macroalgae included the removal and disposal of *Sargassum* in the highest part of the beach or in places determined *ex profeso*. This contributed to a continuous accumulation of *Sargassum* masses on the beach. However, the *Sargassum* removal was not quantified and the information regarding removal included here is only preliminary.

Mahahual (18°42'16.96''N 87°42.619'W) is located in the northern part of the Mesoamerican Barrier Reef System (MBRS) in the state of Quintana Roo. Mahahual is a former fishing village but during the last two decades has undergone reef degradation due to anthropogenic impact (*Martínez-Rendis et al., 2016*). It has a narrow reef lagoon (230–450 m). *Sargassum* management in this locality was active through removing it from the beach and *ex situ* disposition.

Xahuayxol (18°30'21.78"N; 87°45'24.84"W) located south of Mahahual, has a larger reef lagoon measuring 300 to 500 m from the beach to the reef crest. *Sargassum* was not removed from the beach in any systematic way and remained accumulated on the shore. This reef is the northern limit of the marine protected area Parque Nacional Arrecifes de Xcalak (PNAX) and human activities are less salient than in Mahahual (*Schmitter-Soto et al., 2018*).

Xcalak (18°14′7.68″N; 87°50′1.46″W), at the southern limit of the Mexican Caribbean, is part of PNAX since 2000. It is also part of the MBRS (*Hoffman*, 2009). It has a wide reef



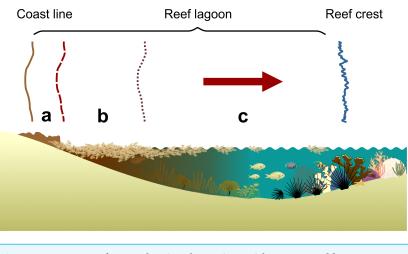


lagoon (950–1,200 m), and *Sargassum* was accumulated along the shore in large amounts. There was active but less intense *Sargassum* management in place at Xcalak, where final disposal was *in situ* on the highest part of beach.

At all sampled sites, the dominant forcing mechanism was reef lagoon circulation from wave action (*Mariño-Tapia et al., 2010*). In our study area, during the period from June to August has the wave orbital velocity over the threshold of motion (*Maldonado-Sánchez et al., 2019*), indicating active circulation in the reef lagoons.

#### Collecting and processing data

This study covers two periods: Under *Sargassum* effect (USE) during the months of July– August 2015 and without *Sargassum* effect (WSE) in July–August 2016. USE sampling for stable isotope analysis included drifting *Sargassum* (mixture of S. *fluitans* and S. *natans*), turf associated pelagic *Sargassum*, benthic macroalgae, local turf and 19 individuals of *D. antillarum*. WSE sampling included benthic macroalgae, local turf and 15 individuals of *D. antillarum* (see sampling details ST1). Samples sizes were based on previous studies to obtain sufficient data for statistical analysis (*Rodríguez, 2003; Tomas et al., 2006; Wing et al., 2008; Rodríguez-Barreras et al., 2016*). The sampling sites were at coastal lagoons in the back reef zone (section c, Fig. 2), zone with no visible presence of *Sargassum* leachates (*van Tussenbroek et al., 2017*) and where *D. antillarum* is distributed (*Steneck & Lang, 2003; Jorgensen, Espinoza-Ávalos & Bahena-Basave, 2008; Jordán-Garza et al., 2008; Maldonado-Sánchez, 2018*).



**Figure 2** Lagoon reef-scape showing the sections with *Sargassum* blooms. Lagoon reef-scape showing the sections: a: decomposing *Sargassum* spp., Section b, leachates (dark brown water) and section c, back reef, areas without visible leachates. Based on *van Tussenbroek et al.* (2017). Full-size DOI: 10.7717/peerj.7589/fig-2

#### Under Sargassum effect (USE) measurements

USE included measurements of dissolved oxygen (mg  $l^{-1}$ ) recorded with a calibrated Multi-parameter water quality checker HORIBA 50 at Mahahual, Xahuayxol and Xcalak. Measurements of dissolved oxygen were made at points distributed in three sections from areas with decomposing *Sargassum* (section a), leachates (section b -dark brown water-) and reef lagoon areas without *Sargassum* leachates (section c) (Fig. 2).

Pelagic *Sargassum* spp., turf (benthic turf and the associated turf to pelagic *Sargassum*) and macroalgae samples were collected in coral reef patches of section c (back reef zone) for each sampling site.

#### Under and without Sargassum effect (USE and WSE) measurements

We collected algal samples to obtain biomass, and for stable isotope analysis using nine quadrats ( $50 \times 50$  cm) per site. Pelagic *Sargassum* biomass was calculated based on sunken thalli and overlaid on reef substrates inside the quadrats. The quadrats were located randomly within the sea urchin habitat (radius of 15 m from collected echinoids). The substrate inside each quadrat was scrapped, carefully removed, collected in bags, and frozen for later analysis.

Macroalgae were identified according to *Littler & Littler (2000)*. Analyses were performed to genus level. For biomass estimates samples were dried for 48 h in an oven at 60 °C. Samples were weighed with a digital balance (standard error = 0.0001 g). To determine *D. antillarum* differential algae assimilation considering USE and WSE, algae samples were pooled per site. The sampled echinoids and algal species for this study are not threatened. The collection permit was obtained from the Comisión Nacional de Acuacultura y Pesca (CONAPESCA, PPF/DGOPA-002/17).

The collected individuals of *D. antillarum* were at the same depth range (1.5–2.5 m) and only individuals greater than 5.0 cm in test diameter were collected to avoid any ontogenic effect. Samples were frozen shortly after collection and processed later at the

laboratory. The muscles of Aristotle's lanterns were carefully removed and washed from the stomach contents to estimate algal assimilation by *D. antillarum* because this tissue offers a time-integrated measure of carbon and nitrogen assimilated sources (*Polunin et al., 2001; Ben-David & Schell, 2001; Phillips & Koch, 2002*).

Macroalgae and local turf, pelagic *Sargassum* species (*S. fluitans* and *S. natans*), turf associated to pelagic *Sargassum*, and echinoids muscle samples were rinsed with filtered water, dried at 50 °C during 48 h, grounded to a fine powder and placed in glass vial for isotope analyses. To remove carbonates from some algal species (eg., *Halimeda* spp. *Penicillus* spp., etc.), the samples were washed with diluted HCl at 1 N prior to drying to avoid disturbance in the mass spectrometer reading.

A subsample of each algae and muscle (1mg) was taken to evaluate the <sup>13</sup>C/<sup>12</sup>C and <sup>15</sup>N/<sup>14</sup>N ratios using a Delta V Plus Mass Spectrometer. Catalyzers silvered cobaltous/cobaltic oxide and chromium oxide were used. Carbon and nitrogen samples were analysed in a dual isotope mode at the Centro Interdisciplinario de Ciencias Marinas from Instituto Politécnico Nacional. Isotope samples were loaded into tin-capsules and placed in a 50-position automated Zero Blank sample carousel on a COSTECH 4020 elemental analyzer. The carbon and nitrogen isotopic results were expressed in standard delta notation relative to Vienna Pee Dee Belemnite (VPDB) and to atmospheric air.

$$\delta^{13}C = \left[ \left( \frac{\left(\frac{^{13}C}{^{12}C}\right) \text{Sample}}{\left(\frac{^{13}C}{^{12}C}\right) \text{Standard}} \right) - 1 \right] \times 1000 (\%)$$

and

$$\delta^{15} \mathrm{N} = \left[ \left( \frac{\left(\frac{15}{14}\mathrm{N}\right)}{\left(\frac{15}{14}\mathrm{N}\right)} \mathrm{Sample}}{\left(\frac{15}{14}\mathrm{N}\right)} - 1 \right] \times 1000 (\%).$$

The standard deviations of  $\delta^{13}$ C and  $\delta^{15}$ N replicate analyses were estimated; the precision values were 0.2‰ for carbon and nitrogen isotope measurements. In addition, we calculated the trophic level (TL) according to *Hobson & Welch (1992)* for every individual of *D. antillarum* in each site, expressed as:

$$TL = \frac{1 + (Nm - Nb)}{TEF}$$

Where Nm is the mean  $\delta^{15}$ N ratio of each sea urchin, Nb is average basis  $\delta^{15}$ N value of the algal community, and TEF is the given value for the trophic enrichment factor (TEF). We assumed a TEF of 2.4 following *Moore & Semmens (2008)*.

#### Data analysis

Dissolved oxygen data were summarized to obtain average values ( $\pm$  standard error) by section (sections a, b, c in Fig. 2) and reef lagoons (Mahahual, Xahuayxol, and Xcalak). We evaluated differences among sections and at the reef lagoons (sections a, b, c, in Fig. 2). We plotted raw data of dissolved oxygen as a function of distance to coast to visualize the low to high values gradient related to that distances in every reef lagoon.

The relative contribution of algae to the diet of the sea urchins *D. antillarum* was estimated with a Bayesian isotopic mixing model (SIAR *Parnell & Jackson, 2013*), which included the isotopic signatures, fractionation and variability to estimate the probability distribution of the contribution of the food source to a mixture. This procedure supplied accurate information about the contribution of algal species to the sea urchin tissues, as it provided the proportion for every source and recognized the main sources as important components of the diet (*Peterson, 1999*; *Fry, 2006*; *Wing et al., 2008*) at three different sites, and under and without *Sargassum* effect. To run the model, the isotopic discrimination factor values used were  $2.4 \pm 1.6\%$  (mean  $\pm$  SD) for  $\delta^{15}$ N, and  $0.4 \pm 1.3\%$  (mean  $\pm$  SD) for  $\delta^{13}$ C (*Minagawa & Wada, 1984*; *Fry & Sherr, 1989*; *Moore & Semmens, 2008*; *Cabanillas-Terán et al., 2016*).

The following algal taxa/groups were considered for the mixing models analyses: *Caulerpa, Codium, Dictyota, Halimeda, Laurencia, Lobophora, Padina, Penicillus, Sargassum polyceratum, Stypopodium,* turf, and *Udotea.* The sources for the model were selected following the theoretical geometric assumptions of the mixing model according to *Phillips et al.* (2014) and *Rodríguez-Barreras et al.* (2015) to ensure reliable resources. Samples of *D. antillarum* did not require lipid extraction since C:N ratios of Aristotle lantern's muscle were lower than 3.5 (*Post et al., 2007*).

We performed a comparison USE and WSE between the niche width and overlap for *D. antillarum* by using Stable Isotope Bayesian Ellipses in R (SIBER) (*Jackson et al., 2011*) from the SIAR package (*Parnell & Jackson, 2013*). This procedure performs metrics based on ellipses and provides the standard ellipse corrected area (SEAc) used as the trophic niche breadth and the overlap between ellipses, presuming that values close to 1 exhibit a higher trophic overlap. Models were run with 200,000 iterations and a burn in of 50,000.

Homogeneity and normality of variance were tested by performing a Kolmogorov– Smirnov and a Cochran's test (*Zar*, 1999). Nitrogen data followed the premises of parametric analysis, but the carbon, dissolved oxygen and biomass data required a power transformation for reaching normality and homogeneity of variance (*Box & Cox*, 1964). We ran two-way ANOVA to evaluate dissolved oxygen data differences among sections in the reef lagoons and we performed a post hoc comparison using Tukey-HSD test. The functions aov and glm from the Gaussian family were used to test the differences in isotopic ratios of carbon and nitrogen values to compare the effect (WSE and USE) between sites and their interaction. Statistics were performed with  $\alpha < 0.05$  (R Core Team, 1.0.153, 2017).

# RESULTS

The dissolved oxygen values USE indicated that the effects of the leachates generated by the decomposition process, together with the organic material carried in their vegetal structures, reduced the values of dissolved oxygen in the reef lagoon water. The decomposing *Sargassum* area (section a, Fig. 2) showed an average range from 1.01 (S.E.  $\pm$  0.30) mg l<sup>-1</sup>at Xcalak to 1.88 (S.E.  $\pm$  0.37) mg l<sup>-1</sup> at Mahahual. The leachates area (section b, Fig. 2) showed an average range from 2.42 (S.E.  $\pm$  0.32) mg l<sup>-1</sup> at Xahuayxol to 3.66 (S.E.  $\pm$  0.42)

mg l<sup>-1</sup>at Mahahual. The back reef area (section c, Fig. 2) showed an average range from 4.1 (S.E.  $\pm$  0.34) mg l<sup>-1</sup>at Mahahual to 4.8 (S.E.  $\pm$  0.22) mg l<sup>-1</sup>at Xcalak. The two-way ANOVA indicated significant differences between reef lagoons (p < 0.05) and sections (p < 0.01); Mahahual was significantly different to Xcalak, but Mahahual and Xcalak were not significantly different to Xahuayxol (Post-hoc HSD of Tukey test, 95% confidence). The three sections at the three reefs were significantly different, except the sections b and c of Mahahual (Post-hoc HSD of Tukey test, 95% confidence). Therefore dissolved oxygen data showed a gradient significantly different between sections. The overall values of dissolved oxygen displayed the lowest concentrations for section a, near the shoreline and higher values beyond the back reef section c (Fig. 3).

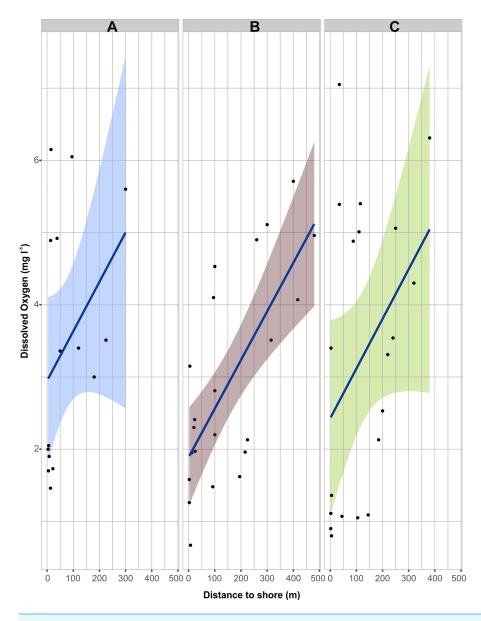
# Biomass, $\delta^{15}N$ and $\delta^{13}C$ of macroalgae

Biomass data for benthic taxa displayed no significant differences between USE and WSE, but significant differences were found among localities (ANOVA, df = 2, F = 8.24, p < 0.0001). Mahahual had the highest mean benthic biomass values (55.2 dry weight m<sup>-2</sup>) followed by Xahuayxol with (38.8 dry weight m<sup>-2</sup>) and Xcalak (16 dry weight m<sup>-2</sup> ±). WSE biomass average values for local benthic algae ranged from 3.01 dry weight m<sup>-2</sup> ±0.95 (*Codium* spp. at Xcalak) to 133.50 dry weight m<sup>-2</sup> ±30.29 (*Halimeda* spp. at Mahahual). USE values ranged from 7.75 dry weight m<sup>-2</sup> ±5.4 (*Caulerpa* at Xcalak) to 145.99 dry weight m<sup>-2</sup> ± 36.21 (*Halimeda* spp. at Mahahual, Table 1). Genus-level biomass of pelagic taxa showed no significant differences per site neither at genus level, however *Sargassum fluitans* displayed the highest biomass values.

Under and without *Sargassum* effect values revealed significant differences in overall benthic algae values of  $\delta^{15}$ N (ANOVA, df = 1, F = 20.27, p < 0.0001). Specifically under *Sargassum* blooms most of the algae exhibited isotopic signatures with significantly depleted  $\delta^{15}$ N like *Dictyota* and turf across the lagoon reef sites (Table 2). The overall macroalgal  $\delta^{15}$ N under *Sargassum* fluctuated from 0.023 to 2.08‰. At Xcalak *Caulerpa* displayed the highest mean values of nitrogen with 2.02 ± 0.08‰. Local Turf USE displayed negative values and overall turf values fluctuated from -0.97% to 0.42%. Xahuayxol displayed the most negative  $\delta^{15}$ N mean value of local turf ( $-0.51 \pm 0.02\%$ ). Without *Sargassum* effect the mean algal genus  $\delta^{15}$ N fluctuated from  $0.06 \pm 0.08$  with *Penicillus* at Xcalak, and Xahuayxol displayed the highest mean value of  $\delta^{15}$ N with *Caulerpa* (5.68 ± 0.01‰) (Table 2).

As for  $\delta^{13}$ C USE ratios fluctuated from -21.98 to -9.23% and WSE from -20.90 to -5.65%. Considering only the algae presented in both sampling periods (WSE and USE) there was no significant difference in  $\delta^{13}$ C among sites (ANOVA, df = 2, F = 0.55, p > 0.05) neither was significant difference analysing the effect (ANOVA, df = 1, F = 1.14, p > 0.05) and their interaction (ANOVA, df = 2, F = 0.86, p > 0.05).

Overall USE pelagic *Sargassum*  $\delta^{13}$ C values fluctuated from -17.95% to -15.24%. *S. natans* exhibited the most negative mean values of  $\delta^{13}$ C ( $-17.44 \pm 0.71\%$ ) at Mahahual (Table 2). There was no difference in  $\delta^{13}$ C among sites (ANOVA, df = 2, F = 0.05, p > 0.05) but there were significant differences  $\delta^{13}$ C between species (ANOVA, df = 2, F = 7.57, p = 0.01). *Sargassum*'s associated turf  $\delta^{13}$ C values fluctuated from -18.65%



**Figure 3** Dissolved Oxygen values under Sargassum effect (USE). Dissolved oxygen  $(mgl^{-1})$  values along the distance to shoreline at (A) Mahahual (blue), (B) Xahuayxol (purple) and (C) Xcalak (green) considering the sections depicted in Fig. 2: dissolved oxygen  $< 2 mgl^{-1}$ : decomposing *Sargassum* spp; dissolved oxygen between 2–4 mgl<sup>-1</sup>: leachates (dark brown water) and dissolved oxygen  $> 4 mgl^{-1}$ : back reef, areas without visible leachates.

Full-size DOI: 10.7717/peerj.7589/fig-3

to -15.37%. The most negative  $\delta^{13}$ C mean value was displayed at Mahahual ( $-18.3 \pm 0.5\%$ ) for *Sargassum*'s associated turf.

Overall pelagic *Sargassum*  $\delta^{15}$ N values ranged from -2.87% to -0.30%. The less negative mean value was exhibited at Mahahual ( $-0.53 \pm 0.26\%$ ) for *S. fluitans*. There was no significant difference for  $\delta^{15}$ N among sites (ANOVA, df = 2, F = 3.90, p = 0.05), but there was a remarkable trend to depleted  $\delta^{15}$ N at Xcalak where *S. fluitans* displayed

Mahahual Xahuayxol Xcalak WSE USE Genus WSE USE Genus WSE USE Genus Caulerpa  $39.49 \pm 20.79$  $19.82\pm 6.48$ Caulerpa  $5.38\pm0.93$ Caulerpa  $7.97\pm3.51$  $7.75\pm5.4$ Codium Dictyota  $19.92 \pm 11.69$ Dictyota  $6.61 \pm 2.49$  $3.01\pm0.95$  $20.40 \pm 5.41$  $20.36\pm5.96$ Halimeda  $133.50 \pm 30.29$  $145.99 \pm 36.21$ Halimeda  $118.07\pm29.43$  $89.18 \pm 9.998$ Dictyota  $21.99 \pm 5.99$  $17.76 \pm 2.34$ Lobophora Laurencia  $14.73\pm22.15$ Laurencia  $8.49 \pm 4.10$  $26.96 \pm 4.30$ Lobophora Padina Stypopodium  $95.41 \pm 66.10$ 19.933 ±11.50  $12.62 \pm 4.30$ Turf  $24.69 \pm 9.17$ Penicillus  $12.88\pm3.94$ Penicillus  $27.49 \pm 3.51$  $19.042 \pm 6.045$  $26.23\pm2.45$ Udotea  $59.79 \pm 45.74$ Sargassum  $14.26\pm4.42$ Sargassum  $15.01\pm4.30$ Stypopodium  $10.06\pm12.13$ Turf  $12.00\pm3.51$  $11.40 \pm 4.21$ Turf  $5.886 \pm 2.83$  $14.26\pm7.84$ Udotea  $34.02\pm16.54$  $39.13 \pm 14.76$  $11.86 \pm 2.75$ S. fluitans  $12.39 \pm 8.33$ S. fluitans S. fluitans  $13.00 \pm 6.99$ S. natans S. natans  $4.92\pm3.14$ S. natans  $7.07\pm3.26$  $10.03\pm7.94$ Sargassum's  $3.10 \pm 1.21$ Sargassum's  $3.23 \pm 1.28$ Sargassum's  $1.98 \pm 1.29$ associated associated associated turf turf turf

Table 1Algal biomass values. Mean  $\pm$  standard deviation values of algal biomass (grams dry weight m<sup>-2</sup>) at Mahahual, Xahuayxol and Xcalak. Genus considered for<br/>the mixing models analysis. Data below the grey line belongs to pelagic taxa.

Mahahual			Xahuayxol			Xcalak		
Genus	$\delta^{13}C$	$\delta^{15}N$	Genus	$\delta^{13}C$	$\delta^{15}N$	Genus	$\delta^{13}C$	$\delta^{15}$ N
Caulerpa	$-9.89\pm0.15$	$2.22\pm0.01$	Caulerpa	$-8.86 \pm 0.19$	$5.68\pm0.01$	Caulerpa	$-12.60 \pm 0.04$	$1.00\pm0.10$
Caulerpa*	$-16.22\pm0.55$	$\textbf{0.93} \pm \textbf{0.08}$	Dictyota	$-15.71 \pm 0.90$	$2.29\pm0.41$	Caulerpa*	$-9.63\pm0.02$	$2.02\pm0.08$
Dictyota	$-16.38\pm1.23$	1.56 ±1.37	Dictyota*	$-16.31\pm0.95$	$\textit{0.71}\pm\textit{0.02}$	Codium	$-12.17\pm0.07$	$1.25\pm0.07$
Dictyota*	$-15.95\pm0.04$	$0.82\pm0.04$	Halimeda*	$-12.61\pm1.70$	$0.88\pm0.01$	Dictyota	$-15.47\pm0.68$	$0.67\pm0.03$
Halimeda	$-7.01\pm1.25$	$0.29\pm0.43$	Laurencia	$-14.81\pm0.23$	$1.36\pm0.71$	Dictyota*	$-15.69\pm0.20$	$0.04\pm0.06$
Halimeda*	$-8.39\pm0.69$	$0.68\pm0.12$	Lobophora*	$-10.49\pm1.35$	$0.33\pm0.64$	Lobophora	$-14.15\pm0.53$	$0.77\pm0.33$
Laurencia	$-16.16\pm0.90$	$2.61 \pm 1.41$	Penicillus	$-11.51\pm8.28$	$1.84\pm0.30$	Padina	$-10.18\pm0.18$	$0.25\pm0.19$
Stypopodium	$-11.33\pm0.52$	$0.67\pm0.05$	Sargassum	$-14.65\pm1.82$	$3.21\pm0.23$	Penicillus	$-14.50\pm0.08$	$0.06\pm0.08$
Turf	$-13.44\pm0.00$	$3.03\pm0.02$	Stypopodium	$-16.80\pm1.40$	$1.47\pm0.56$	Penicillus*	$-9.75\pm0.14$	$1.98\pm0.04$
Turf*	$-16.54\pm0.22$	$-0.51\pm0.02$	Turf	$-16.43\pm1.32$	$1.84\pm0.30$	Sargassum*	$-14.76\pm0.87$	$0.37\pm0.08$
Udotea	$-12.86\pm0.42$	$2.19\pm0.03$	Turf*	$-18.56\pm0.04$	$-0.89\pm0.11$	Turf	$-17.44\pm0.48$	$4.59\pm0.64$
			Udotea	$-11.62\pm1.34$	$2.42\pm1.12$	Turf*	$-21.98\pm0.10$	$0.41\pm0.01$
			Udotea *	$-12.65\pm0.20$	$2.65\pm0.77$			
S. fluitans	$-16.03\pm0.99$	$-0.53\pm0.26$	S. fluitans	$-16.36\pm0.15$	$-1.74\pm0.38$	S. fluitans	$-16.26\pm0.17$	$-2.51\pm0.52$
S. natans	$-17.44\pm0.71$	$-1.59\pm0.70$	S. natans	$-16.82\pm0.73$	$-1.49\pm0.42$	S. natans	$-17.28\pm0.81$	$-1.62\pm0.55$
Sargassum's associated turf	$-18.29\pm0.51$	$-1.13 \pm 0.05$	Sargassum's associated turf	$-15.93 \pm 0.79$	$-0.47 \pm 0.07$	<i>Sargassum</i> 's associated turf	$-16.27 \pm 0.63$	$-0.96\pm0.01$

 Table 2
 Mean ± standard deviation values of  $\delta^{13}$ C and  $\delta^{15}$ N of algal genus considered in the mixing model analysis taken from Mahahual, Xahuayxol and Xcalak, the asterisks represent the sources under Sargassum effect.

the lowest mean values of  $\delta^{15}N$  (-2.51 ± 0.52‰). Turf associated to floating *Sargassum*  $\delta^{15}N$  values fluctuated from -0.42‰ to -1.17‰. The most depleted  $\delta^{15}N$  was exhibited at Mahahual (-1.13 ± 0.05‰) and the less negative mean value was displayed in Xahuayxol (-0.47 ± 0.07‰).

#### Sea urchins

There were significant differences  $\delta^{15}$ N among sites (ANOVA df = 2, F = 6.473, p = 0.005) and the interaction between site\*effect (USE and WSE) showed significant differences (ANOVA, df = 2, F = 7.321, p = 0.003).

*D. antillarum* exhibited no differences among sites for  $\delta^{13}$ C values p > 0.05. However, we found significant differences analysing the USE and WSE effect (ANOVA df = 1, F = 5.301, p = 0.03). The isotopic ratios of *D. antillarum* (USE) varied from 3.83% to 6.13% for  $\delta^{15}$ N, while  $\delta^{13}$ C ranged from -9.41% to -13.62%. Mahahual was the site with the highest average values for  $\delta^{15}$ N  $5.80 \pm 0.30\%$ , while Xcalak displayed the lowest average value  $4.38 \pm 0.29\%$ . The isotopic ratios of *D. antillarum* (WSE) ranged from 4.69% to 6.16 for  $\delta^{15}$ N, while  $\delta^{13}$ C fluctuated from -8.83% to -13.42%. We found significant differences for  $\delta^{15}$ N for sea urchins between sites (USE, ANOVA, df = 2, F = 6.47, p < 0.005).-Xcalak showed particularly low values under *Sargassum* effect (average value  $4.38 \pm 0.29\%$  *versus* WSE average value  $5.44 \pm 0.36\%$ ). Nevertheless,  $\delta^{13}$ C under *Sargassum* effect (USE).

#### Algal source contributions (SIAR)

Mixing models provided evidence for the contribution of different algal resources for three sites USE and WSE (Table 3). SIAR analysis showed that D. antillarum behaved as an opportunistic grazer under the Sargassum effect, it is important to note that pelagic Sargassum, despite being one of the most abundant available resources, was not the most assimilated resource (Fig. 4). Relatedely, there was a reduction in benthic food sources USE (Fig. 4). Without Sargassum effect D. antillarum consumed, Laurencia, Stypopodium and Udotea (12-15% in average) at Mahahual; Caulerpa, Laurencia, Penicillus, Sargassum and Stypopodium (8-14% in average) at Xahuayxol; and Codium, Lobophora and Padina (13-15% in average) at Xcalak. Nevertheless, those resources were absent in the diet of D. antillarum under Sargassum effect (Table 3). Hence, the species displayed differential resource assimilation and Caulerpa was the most important resource for D. antillarum in Mahahual WSE (up to 37%), followed by Turf (up to 34%) and Halimeda and Udotea (up to 29% for both). USE the most important resource was Halimeda (up to 44%) followed by Caulerpa and Dictyota (both up to 31% of contribution). S. fluitans and S. natan s were no important sources (0-28% and 0-23% respectively), and turf associated to Sargassum blooms was the lesser assimilated resources by *D. antillarum* from 0 up to 22% (Table 3).

At Xahuayxol WSE *D. antillarum* showed *Caulerpa* was the most important resource for *D. antillarum* (from 2 up to 25%) and for the rest of algae there were very similar algal contribution (from 0 up to 23%). The main macroalgal contributor of USE was *Udotea* with up to 61%, followed by *Halimeda* and *Lobophora* (with up to 35% and 38% respectively) as secondary resources. *Sargassum's* associated turf showed evidence of low

Mahahual			Xahuayxol			Xcalak		
Genus	WSE	USE	Genus	WSE	USE	Genus	WSE	USE
Caulerpa	19 (1–37)	14 (0-31)	Caulerpa	14 (2–25 )	_	Caulerpa	14 (0–29)	22 (0-40)
Dictyota	9 (0-22)	14 (0-31)	Dictyota	11 (0-22)	9 (0–23)	Codium	15 (0-30)	-
Halimeda	16 (1–29)	31 (17–44)	Halimeda	10 (0–19)	17 (0-35)	Dictyota	12 (0–26)	11 (0–26)
Laurencia	12 (0-25)	_	Laurencia	11 (0–21)	_	Lobophora	13 (0-27)	-
Stypopodium	12 (0-25)	_	Lobophora	_	20 (0-38)	Padina	15 (0–29)	-
Turf	17 (0-34)	11(0-26)	Penicillus	8 (0–18)	_	Penicillus	12 (0–26)	22 (4–39)
Udotea	15 (0-29)	_	Sargassum	12 (0–23)	_	Turf	19 (0-45)	8 (0–19)
S. fluitans	_	12(0-28)	Stypopodium	11 (0–21)	_	Sargassum	_	13 (0–29)
S. natans	_	9(0-23)	Turf	11 (0-22)	6 (0–16)	S. fluitans	_	7 (0–18)
Sargassum's associated turf	-	9 (0–22)	Udotea	12 (0–23)	28 (2–61)	S. natans	_	8 (0–19)
			S. fluitans	_	6 (0–17)	<i>Sargassum</i> 's associated turf	-	9 (0–23)
			S. natans	_	6 (0–17)			
			<i>Sargassum</i> 's associated turf	-	8 (0–21)			

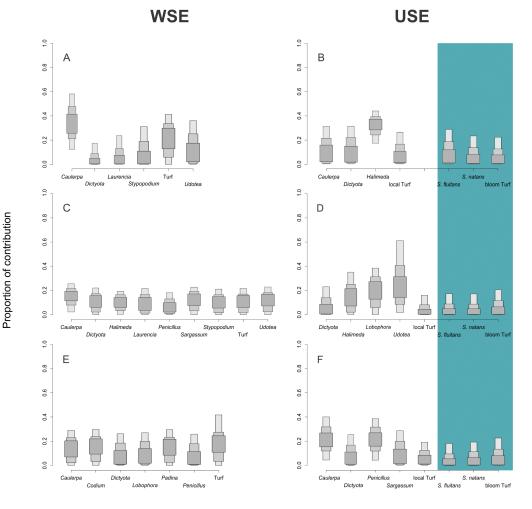
Table 3Average percentage (%) contribution of algal genus to the diet of the sea urchins D. antillarum considering the effect of Sargassum:without Sargassum effect (WSE) and under Sargassum effect (USE) at Mahahual, Xahyayxol and Xcalak produced by the SIAR model using iso-tope values from algae.Minimum and maximum values for each algae are shown in parentheses.

contribution (from 0 up to 21%) and *S. fluitans*, *S. natan* s had negligible contribution to *D. antillarum* diet with a maximum of 17% of the proportional contribution (Table 3).

Turf was the main algal resources for *D. antillarum* in Xcalak WSE (up to 45%) followed by *Caulerpa*, *Codium* and *Padina* as secondary resources (close to 30% maximum of contribution); contrasting USE the main macroalgal contributors in Xcalak were *Penicillus* and *Caulerpa* with up to 39% and 40% respectively. Likewise *Dictyota* and *Sargassum polyceratium* ( benthic *Sargassum*) were secondary resources up to 26% and 29%, respectively. The pelagic components in the other reef lagoons were negligible contributors for *D. antillarum* diet with just 18–23% of maximum contribution (Table 3, Fig. 4).

# **Trophic Levels**

The overall trophic level data for *D. antillarum* (TL) ranged from 1.97 to 3.22. The species exhibited significant differences among sites (ANOVA df = 2, F = 10.63, p = 0.0004), and exhibited significant differences between WSE and USE (ANOVA, df = 1, F = 17.7, p = 0.0003). Likewise, calculating the interaction between site\*effect (USE and WSE) revealed significant differences (ANOVA, df = 2, F = 12.65, p = 0.0001). The highest TL values were reported for Mahahual USE, while the lowest one was recorded in Xahauayxol WSE. At Mahahual, the TL mean value of *D. antillarum* was  $2.35 \pm 0.18$  WSE and  $3.08 \pm 0.13$  USE; at Xahuayxol, the TL mean value was  $2.13 \pm 0.30$  WSE and  $2.49 \pm 0.27$  USE, and at Xcalak TL mean value was  $2.62 \pm 0.15$  WSE and  $2.45 \pm 0.12$  USE (Table 4).



Source (algae)

**Figure 4** Algal resources proportions consumed by *Diadema antillarum*. Contribution rates of algae to the diet of *Diadema antillarum* in the two scenarios (WSE and USE). Results are shown as 25% (light error bars), 75% (grey error bars) and 95% (dark error bars) of credibility intervals. (A) Represents the contribution for *D. antillarum* at Mahahual without *Sargassum* effect (WSE), (B) represents *D. antillarum* at Mahahual without *Sargassum* effect (WSE), (B) represents *D. antillarum* at Mahahual under *Sargassum* effect (USE); (C) represents *D. antillarum* in Xahuayxol WSE, (D) represents *D. antillarum* in Xahuayxol USE; (E) represents *D. antillarum* in Xcalak WSE and (F) represents *D. antillarum* in Xcalak USE. Bloom turf is the *Sargassum*'s associated turf. The blue bar represents the pelagic sources USE.

#### Full-size DOI: 10.7717/peerj.7589/fig-4

 Table 4
 Trophic level of *D. antillarum*. Mean Trophic level (TL), and  $\delta^{15}N$  and  $\delta^{13}C \pm$  standard deviation of *D. antillarum* without Sargassum effect (WSE) and under Sargassum effect (USE) at Mahahual, Xahuayxol and Xcalak.

Site	TL WSE	TL USE	$\delta^{15}$ N WSE	$\delta^{15}$ N USE	$\delta^{13}$ C WSE	$\delta^{13}$ C USE
Mahahual	$2.35\pm0.18$	$3.08\pm0.13$	$5.22\pm0.43$	$5.8\pm0.3$	$-10.46\pm0.6$	$-12.32\pm0.95$
Xahuayxol	$2.13\pm0.3$	$2.49\pm0.27$	$5.09\pm0.71$	$4.9\pm0.24$	$-11.5\pm0.81$	$-11.21\pm1.48$
Xcalak	$2.62\pm0.15$	$2.45\pm0.12$	$5.44\pm0.18$	$4.38\pm0.29$	$-10.58\pm2.01$	$-12.02\pm0.89$

Table 5Trophic niche breadth of sea urchins without Sargassum effect (WSE) and under Sargassumeffect (USE) at Mahahual, Xahuayxol and Xcalak calculated by SIBER analysis of muscle values.SEAc,corrected standard ellipse area.

Niche breadth	Mahahual		Xahu	Xahuayxol		Xcalak	
	WSE	USE	WSE	USE	WSE	USE	
SEA	0.62	0.71	1.79	2.97	2.32	2.32	
SEAc	0.83	0.89	2.68	3.57	3.48	0.14	

#### **Isotopic Niches**

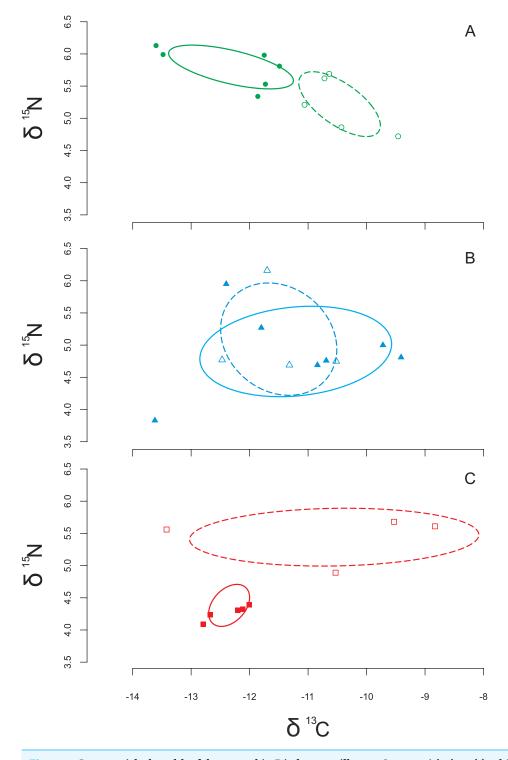
Table 5 shows data on isotopic niche breadth as measured by the corrected standard ellipse area (SEAc). The Stable Isotope Bayesian Ellipses in R (SIBER) analysis suggested a reduction in trophic niche particularly in Xcalak. This site showed the main difference in the trophic niche breadth with SEAc of 3.48 and 0.14 (WSE and USE respectively). An overlap of isotopic niches between WSE and USE was only found in Xahuayxol (Fig. 5). SEAc was higher USE in this site with 3.57 versus 2.68 SEAc WSE (Fig. 5).

# DISCUSSION

Our results provide evidence of the detrimental effect of *Sargassum* blooms on the physicochemical water properties and ecological processes in near-shore coral reef communities as recently has been identified in our study area (*Rodríguez-Martínez, van Tussenbroek & Jordán-Dahlgren, 2016; van Tussenbroek et al., 2017; Cuevas, Uribe-Martínez & Liceaga-Correa, 2018*). Particularly, the results provide evidence for the input of external carbon and nitrogen resulting from *Sargassum* blooms on benthic communities that alter the nutrient inputs and trophic niche for *D. antillarum*. These findings contribute to the growing recognition of the role of exogenous nutrient enrichment in modifying natural sources in a food web. Hence the organic matter inputs from *Sargassum* coupled with hypoxia leads to modification of natural algal resources for *D. antillarum*. Considering the detrimental effects this likely represents a nutrient limitation to sea urchin herbivory.

Onshore *Sargassum* exhibits physical processes of fragmentation, decomposition and remineralization by bacteria, meiofauna and grazers (*Colombini & Chelazzi, 2003*). The algae-derived organic matter, product of that decomposition, has an effect on *in situ* oxygen availability (*Haas et al., 2010*). *Sargassum* blooms clearly showed a negative impact hypoxic conditions found at the three studied reef lagoons (Fig. 3). This could ultimately drive the success of the communities' nitrogen fixation, evidenced by depleted values of  $\delta^{15}$ N as reported by *Dorado et al. (2012)* and *France (1995*).

The dissolved oxygen values in the back reefs of our study areas were lower than the standard values for coral reefs dominated by algae  $(7.9 \pm 0.5 \text{ mg l}^{-1})$  according to *Haas et al.* (2010) and values reported by *Camacho-Cruz et al.* (2019) for Xahuayxol and Mahahual. This supports ideas from *Kendrick et al.* (2000) and *Haas et al.* (2010), who argue that benthic communities linked to reef lagoons are very susceptible to environmental degradation. Some benthic algae play an important play in the transfer of energy and can be catalyzers of oxygen dynamics in reefs due to coral reef associated algae-derived organic matter (*Wild et al.*, 2010).



**Figure 5** Isotope niche breadth of the sea urchin *Diadema antillarum*. Isotope niche breadth of the sea urchin *Diadema antillarum* at Mahahual (A), Xahuayxol (B) and Xcalak (C). Dotted lines are without *Sargassum* effect (WSE) and solid lines under *Sargassum* effect (USE).

Full-size DOI: 10.7717/peerj.7589/fig-5

#### Isotopic variations in the algal resources

We found that the composition of benthic macroalgae assemblages were different under *Sargassum* and without *Sargassum* effect. USE showed a reduction in the taxonomic diversity of macroalgal food sources available to *D. antillarum* and isotope values presented substantially lower  $\delta^{15}$ N values (Table 2). The fact that there were fewer available algal sources in the USE condition implies that the trophic chain becomes less complex as the interaction of primary consumers with their resources is reduced (*Phillips & Gregg, 2003*).

Overall  $\delta^{13}$ C values ranged from -21.98 to -5.65% are similar to ranges reported by *Fry & Sherr (1984)* and *Morillo-Velarde, Briones-Fourzán & Álvarez Filip (2018)*. Those authors reviewed the  $\delta^{13}$ C data of benthic algae, noting that values ranged between -30and 5‰.  $\delta^{15}$ N overall algae values fluctuated from 0.02 to 5.68‰. Despite these values agree with the variation reported in other studies like *Owens (1987)* and *France (1995)*, we found USE very low, ergo according to *Lapointe et al. (2005)* and *France et al. (1998)*. These low  $^{15}$ N:<sup>14</sup>N ratios can be indicative of macroalgae living in oligotrophic reefs which experience nitrogen fixation (*Montoya, Carpenter & Capone, 2002*). In the presence of the leachates of decomposing *Sargassum*, it is possible that anaerobic bacteria gained significance over other benthic groups (Table 2), (*Carpenter & Cox, 1974; Rooker, Turner & Holt, 2006*), and could be the cause of the low macroalgal isotopic signatures. On the other hand, high values of  $\delta^{15}$ N in macroalgae are linked to land-based N enrichment sources, being a good indicator of anthropogenic nitrogen inputs (*Umezawa et al., 2002*) such as sewage discharges (*Risk et al., 2009; Lapointe et al., 2011*).

*France (1995)* reported nitrogen ranges of marine macroalgae from -3 to 18‰. The inconsistencies in this pattern with values of  $\delta^{15}$ N close to atmospheric signature of 0% suggest a fixation of nitrogen. *Dorado et al. (2012)* associated the depleted values of  $\delta^{15}$ N with nitrogen fixation and its impact on the trophic position of consumers. So, temporal difference between values in this study WSE and USE might be explained by the influence of organic input derived from floating *Sargassum* dragged components. We considered that it is likely that the *Sargassum* effect modifies organic matter dynamics. These modifications stem from changes in the oxygen levels, which were consistently reflected in the low  $\delta^{15}$ N values we recorded of for the primary producers.

#### Status of Diadema antillarum in the Mexican Caribbean

It is important to note that we focused our study on the most abundant species at the three localities and the most important shallow-bottom herbivore on Caribbean reefs (*Carpenter, 1981; Hughes, 1994; Aronson & Precht, 2006; Kissling et al., 2014*). For the Mexican Caribbean, there has been considerable variation in *D. antillarum* population data. *Jordán-Garza et al. (2008)* showed a high presence of *D. antillarum* with densities of more than 7 ind m<sup>-2</sup> in several areas, including our study area. *Jorgensen, Espinoza-Ávalos & Bahena-Basave (2008)* reported densities of 12.6 ind m<sup>-2</sup> after hurricane Dean. According to *Maldonado-Sánchez (2018)* population density of *D. antillarum* displayed <1 ind m<sup>-2</sup> for five different habitats of the Parque Nacional Arrecifes de Xcalak (PNAX) reef lagoon (back reef, seagrasses, sandy bottoms and reef patches) and the fore reef. The back reef exhibited the highest abundance with an average of 0.5 ind m<sup>-2</sup>. However for Mahahual,

we registered an average density of 0.6 ind  $m^{-2}$  (N Cabanillas-Terán, pers. obs., 2017), because of the broad variability exhibited in *D. antillarum* populations from the back reef.

#### Trophic parameters of *D. antillarum*

Our results support the evidence that *Sargassum* blooms impacted  $\delta^{15}$ N differentially among sites, as the ratios of  $\delta^{15}$ N and  $\delta^{13}$ C are determined by their resources (*Phillips* & *Gregg*, 2003). It was conspicuous that *D. antillarum* showed higher  $\delta^{15}$ N values USE at Mahahual.

Although some available resources (e.g., *Dictyota* and turf) were present in both conditions (WSE and USE), measuring the contribution of algae to the sea urchin tissues can display key information about how consumers assimilate habitat resources and this could reveal information on the degree of disturbance (*Layman et al., 2007b*). Therefore, it is possible that the ecological role of *D. antillarum* was different in each site and could be explained by the variation in the number of available resources and a differential assimilation (Table 3). The higher  $\delta^{15}$ N values USE in the muscle of *D. antillarum* were a result of the synergistic effect determined by resource availability and disturbance condition.

Pelagic sources may provide new sources of food and the possible nitrogen fixation carried out by turf attached to pelagic *Sargassum* undoubtedly brought a new source of organic matter to basal trophic levels (*Rooker, Turner & Holt, 2006*). However, those sources were not major contributors for *D. antillarum* and appear to avoid the invasive pelagic macroalgae. This is consistent with the feeding ecology by marine generalist herbivores (*Boudouresque & Verlaque, 2001*) and such feeding response is in line with evidence from other sea urchin species in the face of other invasive resources. The experiments carried out by *Tomas, Box & Terrados (2011)* provide evidence that some seaweed invaders were strongly avoided by *Paracentrotus lividus* and therefore escape enemy control by reducing herbivore preference.

The trophic level metric is very useful because the classical discrete trophic level definitions ignore the value of food web connections, omnivory, and diet changes (*Polis & Strong, 1996*; *Vanderklift, Kendrick & Smit, 2006*). Generally the sea urchin *D. antillarum* has been considered as a generalist herbivore (*Ogden & Lobel, 1978*; *Sammarco, 1980*; *Solandt & Campbell, 2001*; *Weil, Torres & Ashton, 2005*). *Morillo-Velarde, Briones-Fourzán & Álvarez Filip (2018)* found that for the North of Quintana Roo Mexico *D. antillarum* occupied an herbivorous trophic position. However, invertebrate samples have been found in the stomach contents this species in the Caribbean, suggesting omnivorous behaviour (Rotjan & Lewis, 2008; Rodríguez-Barreras et al., 2015; Rodríguez-Barreras et al., 2016).

The mean trophic level for *D. antillarum* exhibited at Mahahual was  $2.35 \pm 0.18$  WSE up to  $3.08 \pm 0.13$  USE. Hence, WSE supported the idea that this species occupies an herbivorous position. However USE *D. antillarum* revealed that the species can occupy different trophic niches when faced with resource limitation. Under *Sargassum* blooms, *D. antillarum* displayed a position more in line with omnivorous conditions, suggesting trophic level indicative of herbivorous behaviour tending towards omnivory, according to *Vander Zanden & Rasmussen (1999)*. These authors stated that primary consumers

have a trophic position of 2.0 (strictly herbivorous); but if organisms assimilate primary consumers, they are considered to be a trophic level of 3.0. The results for Mahahual are consistent with *Andrew (1989)* who argued that sea urchins could take advantage of ecosystem changes through omnivory if variation exists in the availability of resources. Our results suggest that *D. antillarum* behave as a facultative omnivore depending on patterns of nutrient availability.  $\delta^{15}$ N signatures for *D. antillarum* in Mahahual suggest a different carbon source USE. These signatures are also likely the result of anthropogenic nitrogen inputs, as this site has a high eutrophication, being an area with elevated touristic demand (*Martínez-Rendis et al., 2016; Arias-González et al., 2017*). Furthermore, possible nitrogen fixation by anaerobic bacteria as an important factor in the variation of available sources of food.

Regarding the TL values exhibited for *D. antillarum* in Mahahual USE  $3.08 \pm 0.13$  versus  $2.35 \pm 0.18$  for WSE would place *D. antillarum* in an omnivorous position tending towards carnivory. Similar values were obtained from Mediterranean sea urchins as a strategy to avoid exclusion by sympatric species (*Wangensteen et al., 2011*). However, we cannot state that *D. antillarum* is carnivorous in Mahahual. This would require a more complete temporal study, and an adjustment of a new  $\delta^{15}$ N baseline for primary producers, considering that  ${}^{15}$ N/ ${}^{14}$ N ratios can vary spatially and temporally (*Jennings et al., 1997; Vanderklift, Kendrick & Smit, 2006*).

The results for Xahuayxol showed also a trend towards higher  $\delta^{15}$ N. However by analyzing the condition of *D. antillarum* in Xahuayxol no significant differences were observed. We can assume that this locality was least changed in its foraging behavior position against the nutrients modification and the species occupied a lower trophic level WSE. Meanwhile, Xcalak displayed the opposite trend compared to Mahahual and Xahuayxol and USE *D. antillarum* trophic level was lower than WSE. Our results suggest that for Xcalak the effect of *Sargassum* blooms completely modified and reduced the possibility for finding available resources, displaying a trophic level around 2.5 between the two scenarios of *Sargassum* blooms. This corresponds to a predominantly herbivorous to omnivorous condition. Moreover this was confirmed with the isotopic niche breadth data where a reduced niche was observed for Xcalak (Fig. 3).

The rank found for *D. antillarum* in this study is consistent with the study conducted by *Rodríguez-Barreras et al.* (2015) in Puerto Rico where microinvertebrates were used as source of organic matter by the sea urchin. Finally, TL values support the premise that echinoids are able to modify their foraging behaviour depending on the availability of resources (*Randall, Schroeder & Starck, 1964; Muthiga & McClanahan, 2007*), and in this case under *Sargassum* blooms condition was not only determined by macroalgae availability, but for unusual conditions that caused a shift in the ecosystem (*Cabanillas-Terán et al., 2016*).

#### Isotopic niche breadth

The ellipses provide integrated information on the relationship between the availability of sources and the niche width. The results of Mahahual indicated that in USE. *D. antillarum* consumes different carbon and nitrogen sources (Fig. 4).

Several studies (*Lawrence, 1975; Carpenter, 1981; Sammarco, 1982; Hay & Fenical, 1988*) noted that echinoids have the ability to adapt their foraging behavior depending on algae availability as well as their population density and site characteristics (*Bak, Carpay & De Ruyter Van Steveninck, 1984; Bak, 1994; Alvarado et al., 2016*). We observed at Mahahual that USE *D. antillarum* exhibited a broader trophic niche than WSE. Despite the limited resources this could lead to trophic overlap and stronger habitat degradation. SIAR results showed a resource shift and this could be explained in terms of omnivory as stated by *France et al. (1998*) "omnivory is a prevalent attribute of aquatic food webs".

The trophic niche of Xahuayxol reflects that there was no difference in the use of carbon and nitrogen sources. It is noteworthy that for the case of Xcalak, the resulting isotopic niche of *D. antillarum* was significantly smaller under *Sargassum* effect. This is consistent with the metric that associates smaller niche amplitude with disturbed ecosystems (*Layman et al., 2007b*).

#### Limitations of the study

To assess the effect of differential management of *Sargassum* and to effectively evaluate the effect of disposal management, quantitative information on beach disposal would be necessary.

From our results, it is clear that algae communities were modified due to *Sargassum*. However, due to the structuring role of sea urchins, and, considering that algae respond to temporal variability naturally, it would be necessary to study changing gradients at different time scales. Such a temporal study would provide more conclusive information about the effect of *Sargassum* spp. on benthic communities.

It is necessary to strengthen the sampling effort to evaluate current population status. A more comprehensive discussion would need to include the interactions with other herbivorous/omnivorous species, that coexist at each site and whether, or how they carry out resource partitioning.

The metrics used in this study allowed us to evaluate the variation of the isotopic signatures that formed the trophic spectrum of *D. antillarum* under two different scenarios. Metric values based on an instantaneous characterization of a single food web provide a limited view of the food web. Therefore, to evaluate the trophic structure and consequently its functional structure, the most promising evaluations would have to include a comparison of multiple gradients, and, to examine the same food web on a longer temporal perspective.

The deposited biomass regarding to *S. fluitans* and *S. natans* did not include a measurement of the total arrived *Sargassum* blooms. However, our results established a baseline for the amounts that were more available for the echinoids that inhabit the back section of the Caribbean shallow reefs.

It would be challenging to evaluate the ecological role of other coexisting species (*Echinometra viridis*, *E. lucunter* and *Eucidaris tribuloides*), and to include samples of micro-invertebrates. However, this could offer new clues to the connectivity between sympatric species, including trophic loops and successional states of algal communities (*Camus, Daroch & Opazo, 2008*) within the benthic communities of coral reefs.

# **CONCLUSIONS**

The present study provides an initial review of how trophic parameters of *D. antillarum* were modified by the impact of pelagic *Sargassum* blooms in the Mexican Caribbean. The results indicated that the effects of the leachates generated by the decomposition process, the input of organic material and deposition in its vegetal structures modify the organic matter in the environment and hence the isotopic signatures. This has negative consequences in the benthic trophic structure, limiting the natural herbivory of *D. antillarum*. The source of available carbon and nitrogen was modified, and the isotopic signatures of macroalgae associated with the reef sites exhibited significantly lower values of  $\delta^{15}$ N. Consequently, the trophic niches were changed and in the case of Xcalak, significantly reduced.

# ACKNOWLEDGEMENTS

We acknowledge Katie Cramer, Gerald Islebe and two anonymous reviewers for their valuable and helpful comments on the manuscript. We thank Comisión Nacional de Áreas Naturales Protegidas (CONANP) for logistic support and Alberto de Jesus-Navarrete for lending the multi-parameter water quality checker. The first author is grateful to María Alfaro-Padilla, Roberto Herrera-Pavón and James Boon for their logistic help during fieldwork and laboratory work, to Alejandro Aragón for his help in editing Fig. 1 and helping to modify Fig. 2, and Rebecca Friedel is acknowledged for improving the English of the manuscript.

# **ADDITIONAL INFORMATION AND DECLARATIONS**

#### Funding

This work was funded by Secretaría de Medio Ambiente y Recursos Naturales (SEMAR-NAT) through the project: OGRMIS- DAC-UCR #001/2015 ECOSUR/SEMARNAT. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

#### **Grant Disclosures**

The following grant information was disclosed by the authors: OGRMIS- DAC-UCR: #001/2015.

#### **Competing Interests**

The authors declare there are no competing interests.

#### **Author Contributions**

- Nancy Cabanillas-Terán conceived and designed the experiments, performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Héctor A. Hernández-Arana conceived and designed the experiments, performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, authored or reviewed drafts of the paper, approved the final draft.

- Miguel-Ángel Ruiz-Zárate and Alejandro Vega-Zepeda performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, authored or reviewed drafts of the paper, approved the final draft.
- Alberto Sanchez-Gonzalez analyzed the data, contributed reagents/materials/analysis tools, authored or reviewed drafts of the paper, approved the final draft.

## **Field Study Permissions**

The following information was supplied relating to field study approvals (i.e., approving body and any reference numbers):

The collection permit (PPF/DGOPA-002/17) was obtained from the Comisión Nacional de Acuacultura y Pesca (CONAPESCA).

#### **Data Availability**

The following information was supplied regarding data availability: The raw data are available in the Supplemental Files.

#### **Supplemental Information**

Supplemental information for this article can be found online at http://dx.doi.org/10.7717/peerj.7589#supplemental-information.

# REFERENCES

- Alvarado JJ, Cortés J, Guzman H, Reyes-Bonilla H. 2016. Bioerosion by the sea urchin Diadema mexicanum along Eastern Tropical Pacific coral reefs. *Marine Ecology* 37:1088–1102 DOI 10.1111/maec.12372.
- Alvarez-Filip L, Gill JA, Dulvy NK, Perry AL, Watkinson AR, Côté IM. 2011. Drivers of region-wide declines in architectural complexity on Caribbean reefs. *Coral Reefs* 30:1051–1060 DOI 10.1007/s00338-011-0795-6.
- Andrew NL. 1989. Contrasting ecological implications of food limitation in sea urchins and herbivorous gastropods. *Marine Ecology Progress Series* 51:189–193 DOI 10.3354/meps051189.
- Arellano-Verdejo J, Lazcano-Hernandez HE, Cabanillas-Terán N. 2019. ERISNet: deep neural network for Sargassum detection along the coastline of the Mexican Caribbean. *PeerJ* 7:e6842 DOI 10.7717/peerj.6842.
- Arias-González JE, Fung T, Seymour RM, Garza-Pérez JR, Acosta-González G, Bozec Y-M, Johnson CR. 2017. A coral-algal phase shift in Mesoamerica not driven by changes in herbivorous fish abundance. *PLOS ONE* 12(4):e0174855 DOI 10.1371/journal.pone.0174855.
- Aronson RB, Precht WF. 2006. Conservation, precaution, and Caribbean reefs. *Coral Reefs* 25:441–450 DOI 10.1007/s00338-006-0122-9.
- **Bak RPM. 1994.** Sea urchin bioerosion on coral reefs: place in the carbonate budget and relevant variables. *Coral Reefs* **13**:99–103 DOI 10.1007/BF00300768.
- Bak RPM, Carpay M, De Ruyter Van Steveninck ED. 1984. Densities of the sea urchin Diadema antillarum before and after mass mortalities on the coral reefs of Curacao. *Marine Ecology Progress Series* 17:105–108.

- Bauman AG, Burt JA, Feary DA, Marquis E, Usseglio P. 2010. Tropical harmful algal blooms: an emerging threat to coral reef communities? *Marine Pollution Bulletin* 60:2117–2122 DOI 10.1016/j.marpolbul.2010.08.015.
- Bearhop S, Adams CE, Waldron S, Fuller RA, MacLeod H. 2004. Determining trophic niche width: a novel approach using stable isotope analysis. *The Journal of Animal Ecology* 73:1007–1012 DOI 10.1111/j.0021-8790.2004.00861.x.
- **Behmer ST, Joern A. 2008.** Coexisting generalist herbivores occupy unique nutritional feeding niches. *Proceedings of the National Academy of Sciences of the United States of America* **105**:1977–1982 DOI 10.1073/pnas.0711870105.
- **Ben-David M, Schell DM. 2001.** Mixing models in analyses of diet using multiple stable isotopes: a response. *Oecologia* **127**:180–184 DOI 10.1007/s004420000570.
- Boecklen WJ, Yarnes CT, Cook BA, James AC. 2011. On the use of stable isotopes in trophic ecology. *Annual Review of Ecology, Evolution, and Systematics* **42**:411–440 DOI 10.1146/annurev-ecolsys-102209-144726.
- **Boudouresque CF, Verlaque M. 2001.** Ecology of Paracentrotus lividus. *Developments in Aquaculture and Fisheries Science* **32**:177–216 DOI 10.1016/S0167-9309(01)80013-2.
- **Box GEP, Cox DR. 1964.** An analysis of transformations. *Journal of the Royal Statistical Society: Series B (Statistical Methodology)* **26**:211–243.
- Cabanillas-Terán N, Loor-Andrade P, Rodríguez-Barreras R, Cortés J. 2016. Trophic ecology of sea urchins in coral-rocky reef systems, Ecuador. *PeerJ* 4:e1578 DOI 10.7717/peerj.1578.
- Camacho-Cruz KA, Ortiz-Hernández MC, Sánchez A, Carrillo L, De Jesús Navarrete A. 2019. Water quality in the eastern karst region of the Yucatan Peninsula: nutrients and stable nitrogen isotopes in turtle grass, Thalassia testudinum. *Environmental Science and Pollution Research* 1–17 DOI 10.1007/s11356-019-04757-3.
- **Camus PA, Daroch K, Opazo LF. 2008.** Potential for omnivory and apparent intraguild predation in rocky intertidal herbivore assemblages from northern Chile. *Marine Ecology Progress Series* **361**:35–45 DOI 10.3354/meps07421.
- **Carpenter RC. 1981.** Grazing by Diadema antillarum (Philippi) and its effects on the benthic algal community. *Journal of Marine Research* **39**:749–765.
- **Carpenter RC. 1986.** Partitioning herbivory and its effects on coral reef algal communities. *Ecological Monographs* **56**:345–364 DOI 10.2307/1942551.
- **Carpenter EJ, Cox JL. 1974.** Production of pelagic Sargassum and a blue–green epiphyte in the western Sargasso Sea. *Limnology and Oceanography* **19**:429–436 DOI 10.4319/lo.1974.19.3.0429.
- **Colombini I, Chelazzi L. 2003.** Influence of marine allochthonous input on sandy beach communities. *Oceanography and Marine Biology: An Annual Review* **41**:115–159.
- Cramer KL, Jackson JBC, Angioletti CV, Leonard-Pingel J, Guilderson TP. 2012. Anthropogenic mortality on coral reefs in Caribbean Panama predates coral disease and bleaching. *Ecology Letters* 15:561–567 DOI 10.1111/j.1461-0248.2012.01768.x.
- **Cuevas E, Uribe-Martínez A, Liceaga-Correa M De LÁ. 2018.** A satellite remotesensing multi-index approach to discriminate pelagic Sargassum in the waters of the

Yucatan Peninsula, Mexico. *International Journal of Remote Sensing* **39**:3608–3627 DOI 10.1080/01431161.2018.1447162.

- **DeNiro MJ, Epstein S. 1981.** Influence of diet on the distribution of nitrogen isotopes in animals. *Geochimica et Cosmochimica Acta* **45**:341–351 DOI 10.1016/0016-7037(81)90244-1.
- De Széchy MTM, Guedes PM, Baeta-Neves MH, Oliveira EN. 2012. Verification of Sargassum natans (Linnaeus) Gaillon (Heterokontophyta: Phaeophyceae) from the Sargasso Sea off the coast of Brazil, western Atlantic Ocean. *Check List* **8**:638–641 DOI 10.15560/8.4.638.
- Dorado S, Rooker JR, Wissel B, Quigg A. 2012. Isotope baseline shifts in pelagic food webs of the Gulf of Mexico. *Marine Ecology Progress Series* 464:37–49 DOI 10.3354/meps09854.
- Eklöf JS, De la Torre-Castro M, Gullström M, Uku J, Muthiga N, Lyimo T, Bandeira SO. 2008. Sea urchin overgrazing of seagrasses: a review of current knowledge on causes, consequences, and management. *Estuarine, Coastal and Shelf Science* 79:569–580 DOI 10.1016/j.ecss.2008.05.005.
- **France RL. 1995.** Differentiation between littoral and pelagic food webs in lakes using stable carbon isotopes. *Limnology and Oceanography* **40**:1310–1313 DOI 10.4319/lo.1995.40.7.1310.
- **France R, Holmquist J, Chandler M, Cattaneo A. 1998.** δ15N evidence for nitrogen fixation associated with macroalgae from a seagrass-mangrove-coral reef system. *Marine Ecology Progress Series* **167**:297–299 DOI 10.3354/meps167297.
- **Franks JS, Johnson DR, Ko DS. 2016.** Pelagic Sargassum in the tropical North Atlantic. *Gulf and Caribbean Research* **27**:SC6–SC11.
- Fry B. 2006. Stable isotope ecology. New York: Springer DOI 10.1007/0-387-33745-8.
- **Fry B, Sherr EB. 1984.** δ13C measurements as indicators of carbon flow in marine and freshwater ecosystems. *Contributions to Marine Science* **27**:13–47.
- Fry B, Sherr EB. 1989. δ13C Measurements as Indicators of Carbon Flow in Marine and Freshwater Ecosystems. In: Rundel PW, Ehleringer JR, Nagy KA, eds. *Stable isotopes in ecological research*. New York: Springer, 196–229 DOI 10.1007/978-1-4612-3498-2\_12.
- Gower J, Young E, King S. 2013. Satellite images suggest a new Sargassum source region in 2011. *Remote Sensing Letters* 4:764–773 DOI 10.1080/2150704X.2013.796433.
- Haas AF, Jantzen C, Naumann MS, Iglesias-Prieto R, Wild C. 2010. Organic matter release by the dominant primary producers in a Caribbean reef lagoon: implication for *in situ* O2 availability. *Marine Ecology Progress Series* 409:27–39 DOI 10.3354/meps08631.
- Hamaoka H, Okuda N, Fukumoto T, Miyasaka H, Omori K. 2010. Seasonal dynamics of a coastal food web: stable isotope analysis of a higher consumer. In: Ohkouchi N, Tayasu I, Koba K, eds. *Earth, life, and isotopes*. Kyoto: Kyoto University Press, 161–181.

- Hay ME, Fenical W. 1988. Marine plant-herbivore interactions: the ecology of chemical defense. *Annual Review of Ecology and Systematics* 19:111–145 DOI 10.1146/annurev.es.19.110188.000551.
- Hobson KA. 1999. Tracing origins and migration of wildlife using stable isotopes: a review. *Oecologia* 120:314–326 DOI 10.1007/s004420050865.
- Hobson KA, Welch HE. 1992. Determination of trophic relationships within a high Arctic marine food web using  $\delta$  13 C and  $\delta$  15 N analysis. *Marine Ecology Progress Series* 84:9–18 DOI 10.3354/meps084009.
- **Hoffman DM. 2009.** Institutional legitimacy and co-management of a marine protected area: implementation lessons from the case of Xcalak Reefs National Park, Mexico. *Human Organization* **68**:39–54 DOI 10.17730/humo.68.1.28gw1106u131143h.
- Hughes TP. 1994. Catastrophes, phase shifts, and large-scale degradation of a Caribbean coral reef. *Science* 265:1547–1551 DOI 10.1126/science.265.5178.1547.
- Hunter MD, Price PW. 1992. Playing chutes and ladders: heterogeneity and the relative roles of bottom-up and top-down forces in natural communities. *Ecology* 73:724–732.
- Jackson AL, Inger R, Parnell AC, Bearhop S. 2011. Comparing isotopic niche widths among and within communities: SIBER–Stable Isotope Bayesian Ellipses in R. *The Journal of Animal Ecology* 80:595–602 DOI 10.1111/j.1365-2656.2011.01806.x.
- Jennings S, Reñones O, Morales-Nin B, Polunin NV, Moranta J, Coll J. 1997. Spatial variation in the 15N and 13C stable isotope composition of plants, invertebrates and fishes on Mediterranean reefs: implications for the study of trophic pathways. *Marine Ecology Progress Series* 146:109–116 DOI 10.3354/meps146109.
- Johnson DR, Ko DS, Franks JS, Moreno P, Sanchez-Rubio G. 2013. The Sargassum Invasion of the Eastern Caribbean and Dynamics of the Equatorial North Atlantic Invasión de Sargazo en el Caribe Oriental y la Dinámica en la Zona Ecuatorial del Atlántico Norte L'Invasion de Sargasse dans les Caraïbes Orientales et leur Dynamique dans la. In: *Proceedings of the 65th gulf and caribbean fisheries institute*. 102–103.
- Jordán-Garza AG, Maldonado MA, Baker DM, Rodríguez-Martínez RE. 2008. High abundance of Diadema antillarum on a Mexican reef. *Coral Reefs* 27:295–295 DOI 10.1007/s00338-007-0338-3.
- Jorgensen P, Espinoza-Ávalos J, Bahena-Basave H. 2008. High population density survival of the sea urchin Diadema antillarum (Philippi 1845) to a category 5 hurricane in southern Mexican Caribbean. *Hidrobiológica* 18:257–260.
- Kendrick GA, Hegge BJ, Wyllie A, Davidson A, Lord DA. 2000. Changes in seagrass cover on success and Parmelia Banks, Western Australia Between 1965 and 1995. *Estuarine, Coastal and Shelf Science* **50**:341–353 DOI 10.1006/ecss.1999.0569.
- Kissling DL, Precht WF, Miller SL, Chiappone M. 2014. Historical reconstruction of population density of the echinoid Diadema antillarum on Florida Keys shallow bank-barrier reefs. *Bulletin of Marine Science* 90:665–679 DOI 10.5343/bms.2013.1022.

- Lapointe BE, Barile PJ, Littler MM, Littler DS. 2005. Macroalgal blooms on southeast Florida coral reefs: II. Cross-shelf discrimination of nitrogen sources indicates widespread assimilation of sewage nitrogen. *Harmful Algae* 4:1106–1122 DOI 10.1016/j.hal.2005.06.002.
- Lapointe BE, Langton R, Bedford BJ, Potts AC, Day O, Hu C. 2010. Land-based nutrient enrichment of the Buccoo Reef Complex and fringing coral reefs of Tobago, West Indies. *Marine Pollution Bulletin* 60:334–343 DOI 10.1016/j.marpolbul.2009.10.020.
- Lapointe BE, Thacker K, Hanson C, Getten L. 2011. Sewage pollution in Negril, Jamaica: effects on nutrition and ecology of coral reef macroalgae. *Chinese Journal* of Oceanology and Limnology 29:775–789 DOI 10.1007/s00343-011-0506-8.
- Lawrence JM. 1975. On the relationships between marine plants and sea urchins. Oceanography and Marine Biology: An Annual Review 13:213–286.
- Layman CA, Araujo MS, Boucek R, Hammerschlag-Peyer CM, Harrison E, Jud ZR, Matich P, Rosenblatt AE, Vaudo JJ, Yeager LA, Post DM, Bearhop S. 2012. Applying stable isotopes to examine food-web structure: an overview of analytical tools. *Biological reviews of the Cambridge Philosophical Society* 87:545–562 DOI 10.1111/j.1469-185X.2011.00208.x.
- Layman CA, Arrington DA, Montaña CG, Post DM. 2007a. Can stable isotope ratios provide for community-wide measures of trophic structure? *Ecology* 88:42–48 DOI 10.1890/0012-9658(2007)88[42:CSIRPF]2.0.CO;2.
- Layman CA, Quattrochi JP, Peyer CM, Allgeier JE. 2007b. Niche width collapse in a resilient top predator following ecosystem fragmentation. *Ecology Letters* **10**:937–944 DOI 10.1111/j.1461-0248.2007.01087.
- Lessios HA, Cubit JD, Robertson DR, Shulman MJ, Parker MR, Garrity SD, Levings SC. 1984. Mass mortality of Diadema antillarum on the Caribbean coast of Panama. *Coral Reefs* 3:173–182 DOI 10.1007/BF00288252.
- Littler DS, Littler MM. 2000. Caribbean reef plants. Washington: OffShore Graphics.
- Louime C, Fortune J, Gervais G. 2017. Sargassum invasion of coastal environments: a growing concern. *American Journal of Environmental Sciences* 13:58–64 DOI 10.3844/ajessp.2017.58.64.
- Maldonado-Sánchez MA. 2018. Influencia ecológica del erizo de mar Diadema antillarum (Philippi, 1845) sobre la estructuración de la comunidad bentónica de la laguna arrecifal de Xcalak, Quintana Roo. PhD thesis, Centro de Investigación y de Estudios avanzados del Instituto Politécnico Nacional Unidad Mérida.
- Maldonado-Sánchez J, Mariño Tapia I, Teresa Herrera-Dorantes M, Ardisson P-L. 2019. Hydrodynamic conditions that favor the settlement of Diadema antillarum to a western Caribbean coral reef. *Bulletin of Marine Science* **95**:251–264 DOI 10.5343/bms.2018.0001.
- Mariño-Tapia I, Silva R, Enriquez C, Mendoza-Baldwin E, Escalante-Mancera E, Ruiz-Rentería F. 2010. Wave transformation and wave-driven circulation on natural reefs under extreme hurricane conditions. *Coastal Engineering Proceedings* **32**:Article 28.
- Martínez-Rendis M, Acosta-González G, Hernández-Stefanoni JL, Arias González JE. 2016. Quantifying the reefscape transformation of a coastal Caribbean coral reef

during a phase shift and the associated coastal landscape change. *Marine Ecology* **37**:697–710 DOI 10.1111/maec.12334.

- **Minagawa M, Wada E. 1984.** Stepwise enrichment of 15N along food chains: further evidence and the relation between δ15N and animal age. *Geochimica et Cosmochimica Acta* **48**:1135–1140 DOI 10.1016/0016-7037(84)90204-7.
- Montoya JP, Carpenter EJ, Capone DG. 2002. Nitrogen fixation and nitrogen isotope abundances in zooplankton of the oligotrophic North Atlantic. *Limnology and Oceanography* **47**:1617–1628 DOI 10.4319/lo.2002.47.6.1617.
- Moore JW, Semmens BX. 2008. Incorporating uncertainty and prior information into stable isotope mixing models. *Ecology Letters* 11:470–480 DOI 10.1111/j.1461-0248.2008.01163.x.
- Morillo-Velarde PS, Briones-Fourzán P, Álvarez Filip L. 2018. Habitat degradation alters trophic pathways but not food chain length on shallow Caribbean coral reefs. *Scientific Reports* 8:4109 DOI 10.1038/s41598-018-22463-x.
- Muthiga NA, McClanahan TR. 2007. Ecology of Diadema. In: *Developments in aquaculture and Fisheries Science*. Amsterdam: Elsevier, 205–225.
- Newsome SD, Martinez del Rio C, Bearhop S, Phillips DL. 2007. A niche for isotopic ecology. *Frontiers in Ecology and the Environment* 5:429–436 DOI 10.1890/060150.1.
- **Ogden JC, Lobel PS. 1978.** The role of herbivorous fishes and urchins in coral reef communities. *Environmental Biology of Fishes* **3**:49–63 DOI 10.1007/BF00006308.
- **Owens NJP. 1987.** Natural Variations in 15N in the Marine Environment. In: Blaxter JHS, Southward AJ, eds. *Advances in marine biology*. New York: Academic Press, 389–451 DOI 10.1016/S0065-2881(08)60077-2.
- Oyesiku OO, Egunyomi A. 2014. Identification and chemical studies of pelagic masses of Sargassum natans (Linnaeus) Gaillon and S. fluitans (Borgessen) Borgesen (brown algae), found offshore in Ondo State, Nigeria. *African Journal of Biotechnology* 13:1188–1193 DOI 10.5897/AJB2013.12335.
- Parnell AC, Inger R, Bearhop S, Jackson AL. 2010. Source partitioning using stable isotopes: coping with too much variation. *PLOS ONE* 5(3):e9672 DOI 10.1371/journal.pone.0009672.
- **Parnell A, Jackson A. 2013.** siar: stable isotope analysis in R. R package version 4.2. *Available at http://CRAN.R-project.org/package=siar*(23 March 2014) [Links].
- **Peterson BJ. 1999.** Stable isotopes as tracers of organic matter input and transfer in benthic food webs: a review. *Acta Oecologica* **20**:479–487 DOI 10.1016/S1146-609X(99)00120-4.
- Peterson BJ, Fry B. 1987. Stable isotopes in ecosystem studies. *Annual Review of Ecology and Systematics* 18:293–320 DOI 10.1146/annurev.es.18.110187.001453.
- Phillips DL, Gregg JW. 2003. Source partitioning using stable isotopes: coping with too many sources. *Oecologia* 136:261–269 DOI 10.1007/s00442-003-1218-3.
- Phillips DL, Inger R, Bearhop S, Jackson AL, Moore JW, Parnell AC, Semmens BX, Ward EJ. 2014. Best practices for use of stable isotope mixing models in food-web studies. *Canadian Journal of Zoology* 92:823–835 DOI 10.1139/cjz-2014-0127.

- Phillips DL, Koch PL. 2002. Incorporating concentration dependence in stable isotope mixing models. *Oecologia* 130:114–125 DOI 10.1007/s004420100786.
- **Polis GA, Strong DR. 1996.** Food web complexity and community dynamics. *The American Naturalist* **147**:813–846 DOI 10.1086/285880.
- Polunin NVC, Morales-Nin B, Pawsey WE, Cartes JE, Pinnegar JK, Moranta J. 2001. Feeding relationships in Mediterranean bathyal assemblages elucidated by stable nitrogen and carbon isotope data. *Marine Ecology Progress Series* 220:13–23 DOI 10.3354/meps220013.
- Post DM, Layman CA, Arrington DA, Takimoto G, Quattrochi J, Montaña CG. 2007. Getting to the fat of the matter: models, methods and assumptions for dealing with lipids in stable isotope analyses. *Oecologia* 152:179–189 DOI 10.1007/s00442-006-0630-x.
- Prado P, Alcoverro T, Romero J. 2010. Influence of nutrients in the feeding ecology of seagrass (Posidonia oceanica L.) consumers: a stable isotopes approach. *Marine Biology* 157:715–724 DOI 10.1007/s00227-009-1355-2.
- **Randall JE, Schroeder RE, Starck WA. 1964.** Notes on the biology of the echinoid Diadema antillarum. *Caribbean Journal of Science* **4**:421–433.
- **Risk MJ, Lapointe BE, Sherwood OA, Bedford BJ. 2009.** The use of δ15N in assessing sewage stress on coral reefs. *Marine Pollution Bulletin* **58**:793–802 DOI 10.1016/j.marpolbul.2009.02.008.
- Rodríguez SR. 2003. Consumption of drift kelp by intertidal populations of the sea urchin Tetrapygus niger on the central Chilean coast: possible consequences at different ecological levels. *Marine Ecology Progress Series* 251:141–151 DOI 10.3354/meps251141.
- Rodríguez-Barreras R, Cuevas E, Cabanillas-Terán N, Branoff B. 2016. Understanding trophic relationships among Caribbean sea urchins. *Revista de Biología Tropical* 64:837–848 DOI 10.15517/rbt.v64i2.19366.
- Rodríguez-Barreras R, Cuevas E, Cabanillas-Terán N, Sabat AM. 2015. Potential omnivory in the sea urchin Diadema antillarum? *Regional Studies in Marine Science* 2:11–18 DOI 10.1016/j.rsma.2015.08.005.
- Rodríguez-Martínez RE, Medina-Valmaseda AE, Blanchon P, Monroy-Velázquez LV, Almazán-Becerril A, Delgado-Pech B, Vásquez-Yeomans L, Francisco V, García-Rivas MC. 2019. Faunal mortality associated with massive beaching and decomposition of pelagic Sargassum. *Marine Pollution Bulletin* 146:201–205 DOI 10.1016/j.marpolbul.2019.06.015.
- Rodríguez-Martínez RE, van Tussenbroek B, Jordán-Dahlgren E. 2016. Afluencia masiva de sargazo pelágico a la costa del Caribe mexicano (2014-2015). In: García-Mendoza E, Quijano-Scheggia SI, Olivos-Ortiz A, Núñez-Vázquez EJ, eds. *Florecimientos Algales Nocivos en México*. Ensenada: CICESE, 352–365.
- Rooker JR, Turner JP, Holt SA. 2006. Trophic ecology of Sargassum-associated fishes in the Gulf of Mexico determined from stable isotopes and fatty acids. *Marine Ecology Progress Series* 313:249–259 DOI 10.3354/meps313249.

- Rotjan RD, Lewis SM. 2008. Impact of coral predators on tropical reefs. *Marine Ecology Progress Series* 367:73–91 DOI 10.3354/meps07531.
- Sala E, Ribes M, Hereu B, Zabala M, Alvà V, Coma R, Garrabou J. 1998. Temporal variability in abundance of the sea urchins Paracentrotus lividus and Arbacia lixula in the northwestern Mediterranean: comparison between a marine reserve and an unprotected area. *Marine Ecology Progress Series* 168:135–145 DOI 10.3354/meps168135.
- Sammarco PW. 1980. Diadema and its relationship to coral spat mortality: grazing, competition, and biological disturbance. *Journal of Experimental Marine Biology and Ecology* **45**:245–272 DOI 10.1016/0022-0981(80)90061-1.
- Sammarco PW. 1982. Echinoid grazing as a structuring force in coral communities: whole reef manipulations. *Journal of Experimental Marine Biology and Ecology* 61:31–55 DOI 10.1016/0022-0981(82)90020-X.
- **Schell JM, Goodwin DS, Siuda ANS. 2015.** Recent Sargassum inundation events in the Caribbean: shipboard observations reveal dominance of a previously rare form. *Oceanography* **28**:8–11.
- Schmitter-Soto JJ, Aguilar-Perera A, Cruz-Martínez A, Herrera-Pavón RL, Morales-Aranda AA, Cobián-Rojas D. 2018. Interdecadal trends in composition, density, size, and mean trophic level of fish species and guilds before and after coastal development in the Mexican Caribbean. *Biodiversity and Conservation* 27:459–474 DOI 10.1007/s10531-017-1446-1.
- Sissini MN, De Barros Barreto MBB, Széchy MTM, De Lucena MB, Oliveira MC, Gower J, Liu G, De Oliveira Bastos E, Milstein D, Gusmão F, Martinelli-Filho JE, Alves-Lima C, Colepicolo P, Ameka G, De Graft-Johnson K, Gouvea L, Torrano-Silva B, Nauer F, Marcos de Castro Nunes J, Barufi JB, Rörig L, Riosmena-Rodríguez R, Mello TJ, Lotufo LVC, Horta PA. 2017. The floating Sargassum (Phaeophyceae) of the South Atlantic Ocean—likely scenarios. *Phycologia* 56:321–328 DOI 10.2216/16-92.1.
- Smetacek V, Zingone A. 2013. Green and golden seaweed tides on the rise. *Nature* 504:84–88 DOI 10.1038/nature12860.
- **Solandt JL, Campbell AC. 2001.** Macroalgal feeding characteristics of the sea urchin Diadema antillarum Philippi at Discovery Bay, Jamaica. *Caribbean Journal of Science* **37**:227–238.
- Solarin BB, Bolaji DA, Fakayode OS, Akinnigbagbe RO. 2014. Impacts of an invasive seaweed Sargassum hystrix var. fluitans (borgesen 1914) on the fisheries and other economic implications for the nigerian coastal waters. *IOSR Journal of Agriculture and Veterinary Science* 7:1–6.
- Steneck RS, Lang JC. 2003. Mexico. Rapid assessment of Mexico's Yucatan reef in 1997 and 1999: pre-and post-1998 mass bleaching and Hurricane Mitch (Stony Corals, Algae and Fishes). *Atoll Research Bulletin* 496(17):294–317 DOI 10.5479/si.00775630.496-17.294.

- Sweatman JL, Layman CA, Fourqurean JW. 2017. Habitat fragmentation has some impacts on aspects of ecosystem functioning in a sub-tropical seagrass bed. *Marine Environmental Research* 126:95–108 DOI 10.1016/j.marenvres.2017.02.003.
- Tomas F, Alvarez-Cascos D, Turon X, Romero J. 2006. Differential element assimilation by sea urchins Paracentrotus lividus in seagrass beds: implications for trophic interactions. *Marine Ecology Progress Series* **306**:125–131 DOI 10.3354/meps306125.
- Tomas F, Box A, Terrados J. 2011. Effects of invasive seaweeds on feeding preference and performance of a keystone Mediterranean herbivore. *Biological Invasions* 13:1559–1570 DOI 10.1007/s10530-010-9913-6.
- **Umezawa Y, Miyajima T, Yamamuro M, Kayanne H, Koike I. 2002.** Fine-scale mapping of land-derived nitrogen in coral reefs by δ15N in macroalgae. *Limnology and Oceanography* **47**:1405–1416 DOI 10.4319/lo.2002.47.5.1405.
- van Tussenbroek BI, Hernández-Arana HA, Rodríguez-Martínez RE, Espinoza-Avalos J, Canizales-Flores HM, González-Godoy CE, Barba-Santos MG, Vega-Zepeda A, Collado-Vides L. 2017. Severe impacts of brown tides caused by Sargassum spp. on near-shore Caribbean seagrass communities. *Marine Pollution Bulletin* 122:272–281 DOI 10.1016/j.marpolbul.2017.06.057.
- Vander Zanden MJ, Rasmussen JB. 1996. A trophic position model of pelagic food webs: impact on contaminant bioaccumulation in Lake Trout. *Ecological Monographs* 66:451–477 DOI 10.2307/2963490.
- Vander Zanden M, Rasmussen JB. 1999. Primary consumer δ13C and δ15N and the trophic position of aquatic consumers. *Ecology* 80:1395–1404
   DOI 10.1890/0012-9658(1999)080[1395:PCCANA]2.0.CO;2.
- Vander Zanden M, Rasmussen JB. 2001. Variation in δ15N and δ13C trophic fractionation: implications for aquatic food web studies. *Limnology and Oceanography* 46:2061–2066 DOI 10.4319/lo.2001.46.8.2061.
- Vanderklift MA, Kendrick GA, Smit AJ. 2006. Differences in trophic position among sympatric sea urchin species. *Estuarine, Coastal and Shelf Science* 66:291–297 DOI 10.1016/j.ecss.2005.09.004.
- Wang M, Hu C, Barnes BB, Mitchum G, Lapointe B, Montoya JP. 2019. The great Atlantic Sargassum belt. *Science* **365**:83–87 DOI 10.1126/science.aaw7912.
- Wang M, Hu C, Cannizzaro J, English D, Han X, Naar D, Lapointe B, Brewton R, Hernandez F. 2018. Remote sensing of Sargassum biomass, nutrients, and pigments. *Geophysical Research Letters* 45:12–359 DOI 10.1029/2018GL078858.
- Wangensteen OS, Turon X, García-Cisneros A, Recasens M, Romero J, Palacín C. 2011. A wolf in sheep's clothing: carnivory in dominant sea urchins in the Mediterranean. *Marine Ecology Progress Series* 441:117–128.
- Weil E, Torres JL, Ashton M. 2005. Population characteristics of the sea urchin Diadema antillarum in La Parguera, Puerto Rico, 17 years after the mass mortality event. *Revista de Biologia Tropical* 53(Suppl 3):219–231.
- Wild C, Haas A, Naumann M, Mayr C, El-Zibdah M. 2010. Comparative investigation of organic matter release by corals and benthic reef algae–implications for pelagic and benthic microbial metabolism. Coral reefs in a time of change–case studies to

understand potential biogeochemical consequences of phase shifts from corals to benthic algae. In: *Proceedings of the 11th international coral reef symposium, vol. 121.* 1319–1323.

- Wing SR, McLeod RJ, Clark KL, Frew RD. 2008. Plasticity in the diet of two echinoderm species across an ecotone: microbial recycling of forest litter and bottomup forcing of population structure. *Marine Ecology Progress Series* 360:115–123 DOI 10.3354/meps07434.
- Zar JH. 1999. Biostatistical analysis. London: Pearson Education.