

1 **Manuscript Title**

2

3 A comprehensive molecular phylogeny of Geometridae (Lepidoptera) with a focus on enigmatic  
4 small subfamilies

5

6 Leidys Murillo-Ramos<sup>1,2</sup>, Gunnar Brehm<sup>3</sup>, Pasi Sihvonen<sup>4</sup>, Axel Hausmann<sup>5</sup>, Sille Holm<sup>6</sup>, Hamid  
7 Ghanavi<sup>2</sup>, Erki Õunap<sup>6,7</sup>, Andro Truuverk<sup>6,8</sup>, Hermann Staude<sup>9</sup>, Egbert Friedrich<sup>10</sup>, Toomas  
8 Tammaru<sup>6</sup>, Niklas Wahlberg<sup>2</sup>.

9

10 <sup>1</sup>Grupo Biología Evolutiva, Department of Biology, Universidad de Sucre, Puerta Roja,  
11 Sincelejo, Sucre, Colombia.

12 <sup>2</sup>Systematic Biology group, Department of Biology, Lund University, Lund, Sweden.

13 <sup>3</sup>Institut für Zoologie und Evolutionsbiologie, Phyletisches Museum, Jena, Germany.

14 <sup>4</sup>Finnish Museum of Natural History, Helsinki, Finland.

15 <sup>5</sup>Staatliche Naturwissenschaftliche Sammlungen Bayerns, München, Germany

16 <sup>6</sup>Department of Zoology, Institute of Ecology and Earth Sciences, University of Tartu,  
17 Vanemuise 46, 51014 Tartu, Estonia.

18 <sup>7</sup>Institute of Agricultural and Environmental Sciences, Estonian University of Life Sciences,  
19 Kreutzwaldi 5, 51014 Tartu, Estonia.

20 <sup>8</sup>Natural History Museum, University of Tartu, Vanemuise 46, 51014 Tartu, Estonia

21 <sup>9</sup>LepsocAfrica, Magaliesburg, South Africa

22 <sup>10</sup>Berghoffsweg 5, 07743 Jena, Germany.

23

24 Corresponding Authors:

25 <sup>1</sup>Leidys Murillo-Ramos

26 and <sup>2</sup>Niklas Wahlberg

27 Email address: leidys.murillo@unisucra.edu.co, niklas.wahlberg@biol.lu.se

28

29

30

31

Formatted: Right: 0.25"

32

### 33 Abstract

34 Our study aims to investigate the relationships of the major lineages within the moth family  
35 Geometridae, with a focus on the poorly studied Oenochrominae-Desmobathrinae complex, and  
36 to translate some of the results into a coherent subfamilial and tribal level classification for the  
37 family. We analyzed a molecular dataset of 1206 Geometridae terminal taxa from all  
38 biogeographical regions comprising up to 11 molecular markers that includes one mitochondrial  
39 (COI) and 10 protein-coding nuclear gene regions (Wingless, *ArgK*, MDH, RpS5, GAPDH, IDH,  
40 Ca-ATPase, Nex9, EF-1alpha, CAD). The molecular data set was analyzed using maximum  
41 likelihood as implemented in IQ-TREE and RAxML. We found high support for the subfamilies  
42 Larentiinae, Geometrinae and Ennominae in their traditional scopes. Sterrhinae **becomes**  
43 monophyletic only if *Ergavia* Walker, *Ametris* Hübner and *Macrotres* Westwood, which are  
44 currently placed in Oenochrominae, are formally transferred to Sterrhinae. Desmobathrinae and  
45 Oenochrominae are found to be polyphyletic. The concepts of Oenochrominae and  
46 Desmobathrinae required major revision and, after appropriate rearrangements, these groups also  
47 form monophyletic subfamily-level entities. Oenochrominae *s.str.* as originally conceived by  
48 Guenée is phylogenetically distant from *Epidesmia*. The latter is hereby described as the  
49 subfamily Epidesmiinae Murillo-Ramos, Sihvonen & Brehm, **subfam. nov.** Epidesmiinae are a  
50 lineage of “slender-bodied Oenochrominae” that include the genera *Ecphyas* Turner, *Systatica*  
51 Turner, *Adeixis* Warren, *Dichromodes* Guenée, *Phrixocomes* Turner, *Abraxaphantes* Warren,  
52 *Epidesmia* Duncan [ & Westwood] and *Phrataria* Walker. Archiearinae are monophyletic **when**  
53 *Dirce* and *Acalyphes* are formally transferred to Ennominae. We also found that many tribes were  
54 para- or polyphyletic and therefore propose tens of taxonomic changes at the tribe and subfamily  
55 levels. Archaeobalbini **stat. rev.** Viidalepp (Geometrinae) is raised from synonymy with  
56 Pseudoterpnini Warren to tribal rank. Chlorodontoperini Murillo-Ramos, Sihvonen & Brehm,  
57 **trib. nov.** and Drepanogynini Murillo-Ramos, Sihvonen & Brehm, **trib. nov.** are described as  
58 new tribes in Geometrinae and Ennominae, respectively.

59

60 **Keywords:** Phylogeny, new subfamily, moths, Epidesmiinae, taxonomy, loopers.

61

62

Formatted: Font: Italic

Deleted: is

Deleted: if

Formatted: Right: 0.25"

## 65 Introduction

66 Geometridae are the second most species-rich family of Lepidoptera, with approximately 24,000  
67 described species ([number](#) updated by the authors from Nieukerken et al., 2011) found in all  
68 regions except Antarctica. The monophyly of Geometridae is well supported based on distinctive  
69 morphological characters (Cook & Scoble, 1992; Scoble, 1992; Minet & Scoble, 1999). In  
70 particular, adult members of the family possess paired tympanal organs at the base of the  
71 abdomen, while in larvae the prolegs are reduced to two pairs in almost all species, which causes  
72 the larvae to move in a looping manner (Minet & Scoble, 1999).

73 The phylogenetic relationships of the major subdivisions of Geometridae have been  
74 studied based on molecular data, which have contributed to the understanding of the evolutionary  
75 relationships within the family (Abraham et al., 2001; Yamamoto & Sota, 2007; Sihvonen et al.,  
76 2011). Eight subfamilies are recognized in Geometridae (Sihvonen et al., 2011). Several recent  
77 molecular and morphological studies have attempted to confirm the monophyly or clarify the  
78 taxonomy of most of these groups, for instance: Sterrhinae (Holloway, 1997; Hausmann, 2004;  
79 Sihvonen & Kaila, 2004; Öunap et al., 2008), Larentiinae (Holloway, 1997; Mironov, 2003;  
80 Viidalepp, 2006, 2011; Hausmann & Viidalepp, 2012; Öunap et al., 2016), Desmobathrinae  
81 (Holloway, 1996; Hausmann, 2001), Archiearinae (Hausmann, 2001; Young, 2006),  
82 Oenochrominae (Holloway, 1996; Scoble & Edwards, 1990; Cook & Scoble, 1992; Hausmann,  
83 2001; Young, 2006), Geometrinae (Cook, 1993; Pitkin, 1996; Hausmann, 2001; Ban et al., 2018),  
84 Orthostixinae (Holloway, 1997) and Ennominae (Holloway, 1994; Pitkin, 2002; Beljaev, 2006;  
85 Young, 2006; Wahlberg et al., 2010; Öunap et al., 2011; Skou & Sihvonen, 2015; Sihvonen et al.,  
86 2015), [but questions remain](#). An important shortcoming is that our understanding of geometrid  
87 systematics is biased towards the long-studied European fauna, whereas the highest diversity of  
88 this family is in the tropics, which [are](#) still largely unexplored (Brehm et al., 2016). Many species  
89 remain undescribed and there are many uncertainties in [the classification of tropical taxa](#).

90 One of the most comprehensive phylogenetic studies on Geometridae to date was  
91 published by Sihvonen et al. (2011). They analyzed a data set of 164 taxa and up to eight genetic  
92 markers, and the most species-rich subfamilies were confirmed as monophyletic. However, the  
93 systematic positions of Oenochrominae and Desmobathrinae remained uncertain due to low taxon  
94 [sampling](#), and both subfamilies were suspected to be polyphyletic. Moreover, [because of](#)

Deleted: is

Deleted: speciess

Commented [WD1]: But also the numbers of genes/markers, right?

Formatted: Right: 0.25"

97 taxonomic uncertainty, many geometrid genera, especially among tropical taxa, remained  
98 unassigned to any tribe.

99 This study is the first in a series of papers that investigate the phylogenetic relationships  
100 of Geometridae on the basis of global sampling. Our dataset comprises 1192 terminal taxa of  
101 Geometridae and 14 outgroup taxa, with samples from all major biomes, using up to 11  
102 molecular markers. Our paper includes an overview of the relationships of the major lineages  
103 within the family, with the particular aim of defining the limits and finding the phylogenetic  
104 affinities of the subfamilies, with a focus on Oenochrominae and Desmobathrinae. Further papers  
105 in the series will focus on particular subfamilies and regions, and will build upon the taxonomic  
106 changes proposed in the present article: e.g., relationships in Sterrhinae (Sihvonen et al., in prep),  
107 New World taxa (Brehm et al., submitted), Larentiinae (Öunap et al., in prep) and the ennomine  
108 tribe Boarmiini (Murillo-Ramos et al., in prep).

109 Oenochrominae and Desmobathrinae are considered the most controversial subfamilies in  
110 Geometridae. A close relationship of these subfamilies has been proposed both in morphological  
111 (Meyrick, 1889; Cook & Scoble, 1992; Holloway, 1996) and in molecular studies (Sihvonen et  
112 al., 2011; Ban et al., 2018). In early classifications, species of Desmobathrinae and  
113 Oenochrominae were included in the former family Monoctenidae (Meyrick, 1889). Meyrick  
114 diagnosed them on the basis of the position of the R veins in the hindwing and Sc+R1 in the  
115 forewing (Scoble & Edwards, 1990). However, the classification proposed by Meyrick was not  
116 fully supported by subsequent taxonomic work (Scoble & Edwards, 1990; Cook & Scoble, 1992;  
117 Holloway, 1996). Too often, Oenochrominae was used for geometrids that could not be placed in  
118 other subfamilies, and at some point, even included Hedylidae, the moth-butterflies (Scoble,  
119 1992). Unsurprisingly, many taxa formerly classified in Oenochrominae have recently been  
120 shown to be misplaced (Holloway, 1997; Staude, 2001; Sihvonen & Staude, 2011; Staude &  
121 Sihvonen, 2014). In Scoble & Edwards (1990), the family concept of Oenochrominae was  
122 restricted to the robust-bodied Australian genera, with one representative from the Oriental  
123 region. Scoble & Edwards were not able to find synapomorphies to define Monoctenidae sensu  
124 Meyrick, and referred back to the original grouping proposed by Guenée (1858). They restricted  
125 Oenochrominae to a core clade based on male genitalia: the diaphragm dorsal to the anellus is  
126 fused with the transtilla to form a rigid plate. Additionally, Cook & Scoble (1992) suggested that

Deleted: because they have not been sampled or because of their uncertain placement.

Deleted: ,

Deleted: which

Deleted: they

Deleted: propose further

Deleted: formal

Deleted: beyond those suggested

Deleted: i

Deleted: Oenochrominae and Desmobathrinae

Deleted: Unfortunately

Deleted: as the principle subfamily

Deleted: a family of

Deleted: traditionally

Deleted: These authors

Deleted: Scoble & Edwards

Deleted: (1990)

Deleted: The sclerotisation of

Deleted: fuses

Formatted: Right: 0.25"

146 the circular form of the lacinia and its orientation parallel to the tympanum was apomorphic for  
147 these robust-bodied Oenochrominae.

148 In an extensive morphological study, Holloway (1996) re-established the subfamily  
149 Desmobathrinae to include species with appendages and slender bodies previously assigned to  
150 Oenochrominae. According to Holloway (1996), Desmobathrinae comprises two tribes:  
151 Eumeleini and Desmobathrini. However, no synapomorphies were found to link the two tribes.  
152 Holloway (1996) noted that the modification of the tegumen of the male genitalia was variable in  
153 both groups but that the reduction of cremastral spines in the pupa from eight to four in *Ozola*  
154 Walker, 1861 and *Eumelea* Duncan [ & Westwood], 1841 provided evidence of a close  
155 relationship between Eumeleini and Desmobathrini. A proposed classification for ??? is  
156 included in the *World list of family group names in Geometridae* (Forum Herbulot, 2007).  
157 Currently, 328 species (76 genera) are included in Oenochrominae, and 248 species (19 genera)  
158 are assigned to Desmobathrinae (Beccaloni et al., 2003; Sihvonen et al., 2011, 2015).

159 Most recent molecular phylogenies have shown Oenochrominae and Desmobathrinae to  
160 be intermingled (Sihvonen et al., 2011; Ban et al., 2018), but previous taxon sampling was  
161 limited to eight and four species, respectively. The poor taxon sampling and unresolved  
162 relationships around the oenochromine and desmobathrine complex called for additional  
163 phylogenetic study to clarify the relationships of these poorly known taxa within Geometridae.  
164 We hypothesize that both Oenochrominae and Desmobathrinae are para- or polyphyletic  
165 assemblages, and our paper aims to establish a new concept in which all subfamilies of the  
166 Geometridae represent monophyletic entities. Our study includes 29 terminal taxa of  
167 Oenochrominae and 11 Desmobathrinae, mostly from the Australian and Oriental Regions.

## 169 Materials & Methods

### 172 Material acquisition, taxon sampling and species identification

173 In addition to 461 terminal taxa with published sequences (see Supplemental data S1), we  
174 included sequences from 745 terminal taxa in our study (Supplemental data S1). Representative  
175 taxa of all subfamilies recognized in Geometridae were included, except for the small subfamily  
176 Orthostixinae for which most molecular markers could not be amplified successfully. A total of

Commented [LM3]: once again, this gives the impression that it was lost for a while, this is not said above

Deleted: Eumeleini and Desmobathrini

Deleted: highlighted

Commented [WD4]: Closer than what? Close is best.

Deleted: r

Commented [WD5]: "The" proposed classification for what taxon? Desmobathrinae or Geometridae?

Deleted: The

Deleted: "

Formatted: Font: Italic

Deleted: "

Deleted: the obviously

Deleted: a sound

Deleted: that

Deleted: ies

Deleted: new

Deleted: representatives of

Deleted: .

Deleted: Most species are distributed

Deleted: in

Deleted: , but some also occur in other parts of the world.

Moved down [1]: The electronic version of this article in Portable Document Format (PDF) will represent a published work according to the International Commission on Zoological Nomenclature (ICZN), and hence the new names contained in the electronic version are effectively published under that Code from the electronic edition alone. This published work and the nomenclatural acts it contains have been registered in ZooBank, the online registration system for the ICZN. The ZooBank LSIDs (Life Science Identifiers) can be resolved and the associated information viewed through any standard web browser by appending the LSID to the prefix <http://zoobank.org/>. For this publication: Epidesmiinae subfam.nov. LSIDurn:lsid:zoobank.org:act:34D1E8F7-99F1-4914-8E12-0110459C2040, Chlorodontoperini trib.nov. LSIDurn:lsid:zoobank.org:act:0833860E-A092-43D6-B2A1-FB57D9F7988D, and Drepanogynini trib.nov., LSIDurn:lsid:zoobank.org:act:AA384988-009F-4175-B98C-6209C8868B93. The online version of this work is archived and available from the following digital repositories: PeerJ, PubMed Central and CLOCKSS.

Formatted: Right: 0.25"

215 93 tribes are represented in this study following recent phylogenetic hypotheses and  
216 classifications (Sihvonen et al., 2011; Wahlberg et al., 2010; Sihvonen et al., 2015; Öunap et al.,  
217 2016; Ban et al., 2018). In addition, 14 non-geometrid species belonging to other families of  
218 Geometroidea were included as outgroups based on the hypothesis proposed by Regier et al.  
219 (2009; 2013). Where possible, two or more samples were included per tribe and genus, especially  
220 for species-rich groups that are widely distributed and in cases where genera were suspected to be  
221 poly- or paraphyletic. We emphasized type species or species similar to type species, judged by  
222 morphological characters and/or genetic similarity of DNA barcodes in order to better inform  
223 subsequent taxonomic work, to favor nomenclatorial stability and to establish the phylogenetic  
224 position of genera unassigned to tribes.

225       Sampled individuals were identified by the authors using appropriate literature, by  
226 comparing them with type material from different collections, museums and DNA barcode  
227 sequences. Moreover, we compiled an illustrated catalogue of all Archiearinae, Desmobathrinae  
228 and Oenochrominae taxa included in this study, to demonstrate their morphological diversity and  
229 to facilitate subsequent verification of our identifications. This catalogue contains images of all  
230 analysed specimens of the above-mentioned taxa as well as photographs of the respective type  
231 material (Supplemental data S2). Further taxa from other subfamilies will be illustrated in other  
232 papers (Brehm et al. in prep., Sihvonen et al. in prep., Öunap et al. in prep.). Some of the studied  
233 specimens could not yet be assigned to species, and their identifications are preliminary,  
234 particularly for (potentially undescribed) tropical species. Taxonomic data, voucher IDs, number  
235 of genes, current systematic placement, and references to relevant literature with regard to tribal  
236 assignment, are shown in Supplemental data S1.

#### 237 *Molecular techniques*

238  
239  
240 DNA was extracted from 1–3 legs of specimens either preserved in ethanol or dry. In a few cases,  
241 other sources of tissue were used, such as parts of larvae. The remaining parts of specimens were  
242 preserved as vouchers deposited in the collections of origin, both public and private (eventually  
243 private material will be deposited in public museum collections). Genomic DNA was extracted  
244 and purified using a NucleoSpin® Tissue Kit (MACHEREY-NAGEL), following the  
245 manufacturer's protocol. DNA amplification and sequencing were carried out following protocols

Deleted: and will eventually be deposited in public museum collections...

Formatted: Right: 0.25"

248 proposed by Wahlberg & Wheat (2008) and Wahlberg et al. (2016). PCR products were  
249 visualized on agarose gels. PCR products were cleaned enzymatically with Exonuclease I and  
250 FastAP Thermosensitive Alkaline Phosphatase (ThermoFisher Scientific) and sent to Macrogen  
251 Europe (Amsterdam) for Sanger sequencing. One mitochondrial (COI) and 10 protein-coding  
252 nuclear gene regions (Wingless, ArgK, MDH, RpS5, GAPDH, IDH, Ca-ATPase, Nex9, EF-  
253 1alpha and CAD) were sequenced. To check for potential misidentifications, DNA barcode  
254 sequences were compared to those in BOLD (Ratnasingham & Hebert 2007) where references of  
255 more than 21,000 geometrid species are available, some 10,000 of them being reliably identified  
256 to Linnean species names (Ratnasingham & Hebert, 2007). GenBank accession numbers for  
257 sequences used in this study are provided in Supplemental data S1.

#### 259 *Alignment and cleaning sequences*

260  
261 Multiple sequence alignments were carried out in MAFFT as implemented in Geneious v.11.0.2  
262 (Biomatters, <http://www.geneious.com/>) for each gene based on a reference sequence of  
263 Geometridae downloaded from the database VoSeq (Peña & Malm, 2012). The alignment of each  
264 gene was carefully checked by eye relative to the reference sequence, taking into account the  
265 respective genetic codes and reading frames. Heterozygous positions were coded with IUPAC  
266 codes. Sequences with bad quality were removed from the alignments. Aligned sequences were  
267 uploaded to VoSeq (Peña & Malm, 2012) and then assembled into a dataset comprising 1206  
268 taxa. The final dataset had a concatenated length of 7665 bp including gaps. To check for  
269 possible errors in alignments, potentially contaminated or identical sequences and  
270 misidentifications, we constructed maximum likelihood trees for each gene. These preliminary  
271 analyses were conducted using RAxML-HPC2 V.8.2.10 (Stamatakis, 2014) on the web-server  
272 CIPRES Science Gateway (Miller et al., 2010). The final data set included at least three genes per  
273 taxon except for *Oenochroma vinaria* (Guenée, 1858), *Acalyphes philorites* Turner, 1925, *Dirce*  
274 *lunaris* (Meyrick, 1890), *D. aesiodora* Turner, 1922, *Furcatrox australis* (Rosenstock, 1885),  
275 *Chlorodontopera mandarinata* (Leech, 1889), *Chlorozancla falcatus* (Hampson, 1895),  
276 *Pamphlebia rubrolimbraria* (Guenée, 1858) and *Thetidia albocostaria* (Bremer, 1864). For these  
277 taxa, included in studies by Young (2006) and Ban et al. (2018), only two markers were  
278 available. The final data matrix included 32% missing data.

Deleted: -

Formatted: Right: 0.25"

280

### 281 *Tree search strategies and model selection*

282 We ran maximum likelihood analyses with a data set partitioned by gene and codon position  
283 using IQ-TREE V1.6.10 (Nguyen et al., 2015) and data partitioned by codon in RAxML  
284 (Stamatakis et al 2014). Best-fitting substitution models were selected by ModelFinder, which is  
285 a model-selection method that incorporates a model of flexible rate heterogeneity across sites  
286 (Kalyaanamoorthy et al., 2017). ModelFinder implements a greedy strategy as implemented in  
287 PartitionFinder that starts with the full partitioned model and consequentially merges two  
288 partitions (MFP+MERGE option) until the model fit does not increase (Lanfear et al., 2012).  
289 After the best model has been found, IQ-TREE starts the tree reconstruction under the best model  
290 scheme. The phylogenetic analyses were carried out with the *-spp* option that allowed each  
291 partition to have its own evolutionary rate. The RAxML-HPC2 V.8.2.10 analysis was carried out  
292 on CIPRES using the GTR+CAT option.

293 Support for nodes was evaluated with 1000 ultrafast bootstrap (UFBoot2) approximations  
294 (Hoang et al., 2017) in IQ-TREE, and SH-like approximate likelihood ratio test (Guindon et al.,  
295 2010). Additionally, we implemented rapid bootstrap (RBS) in RAxML (Stamatakis, 2008). To  
296 reduce the risk of overestimating branch supports in UFBoot2 test, we implemented *-bnni* option,  
297 which optimizes each bootstrap tree using a hill-climbing nearest neighbor interchange (NNI)  
298 search. Trees were visualized and edited in FigTree v1.4.3 software (Rambaut, 2012). The final  
299 trees were rooted with species of the families Sematuridae, Epicopeiidae, Pseudobistonidae and  
300 Uraniidae following previous hypotheses proposed in Regier et al. (2009; 2013), Rajaei et al.  
301 (2015) and Heikkilä et al. (2015).

### 302 *Taxonomic decisions*

303 The electronic version of this article in Portable Document Format (PDF) will represent a  
304 published work according to the International Commission on Zoological Nomenclature (ICZN),  
305 and hence the new names contained in the electronic version are effectively published under that  
306 Code from the electronic edition alone. This published work and the nomenclatural acts it  
307 contains have been registered in ZooBank. The ZooBank LSIDs (Life Science Identifiers) can be  
308 resolved and the associated information viewed through any standard web browser by appending

Deleted: 6

Deleted: TESTNEWMERGE

Deleted: RAxML

Commented [LM8]: If we want the last tree.

Deleted: GAMMA

Moved (insertion) [1]

Deleted: , the online registration system for the ICZN

Formatted: Right: 0.25"



the LSID to the prefix <http://zoobank.org/>. For this publication: *Epidesmiinae* subfam.nov.  
 LSIDurn:lsid:zoobank.org:act:34D1E8F7-99F1-4914-8E12-0110459C2040, *Chlorodontoperini*  
 trib.nov. LSIDurn:lsid:zoobank.org:act:0833860E-A092-43D6-B2A1-FB57D9F7988D, and  
*Drepanogynini* trib.nov., LSIDurn:lsid:zoobank.org:act:AA384988-009F-4175-B98C-  
 6209C8868B93. The online version of this work is archived and available from the following  
 digital repositories: PeerJ, PubMed Central and CLOCKSS.

Deleted: ¶

## Results

### *Searching strategies and model selection*

The best-fit partition model in ModelFinder resulted in merging the codon positions into 26  
 partitions. Similarly, Akaike information criterion (AIC) and Bayesian information criterion  
 (BIC) values show best partition schemes for the data partitioned by codon position (evolutionary  
 models are listed in Supplemental data S3). Topologies recovered by IQ-TREE and RAxML  
 analyses result in trees with nearly identical topology. Also, the different support methods tend to  
 agree in recovering same nodes with strong phylogenetic signal. However, in most of the cases  
 UFBoot2 from IQ-TREE show higher support values compared to RBS in RAxML (RAxML tree  
 with support values is showed in Supplemental data S4). *SH-like* and UFBoot2 performed  
 similarly, with UFBoot2 showing slightly higher values, and both tend to show high support for  
 the same nodes (Fig. 1). As noted by the authors of IQ-TREE, values of *SH*  $\geq 80$  UFBoot2  $\geq$   
 95 and indicate well-supported clades (Trifinopoulos & Minh, 2018).

Deleted: The results from

Deleted: suggest

Deleted: that each gene and

Deleted: keep their own evolutionary model, i.e. no partitions were combined in

Deleted: , with 33 partitions in total

Deleted: and *SH-like*

Deleted: *SH*  $\geq 80$

### *General patterns in the phylogeny of Geometridae*

Analyses of the dataset of 1206 terminal taxa, comprising up to 11 markers and an  
 alignment length of 7665 bp recovered topologies with many well-supported clades. About 20  
 terminal taxa are recovered as very similar genetically and they are likely to represent closely  
 related species, subspecies or specimens of a single species. The examination of their taxonomic  
 status is not the focus of this study, so the number of unique species in the analysis is slightly less  
 than 1200. Our findings confirm the monophyly of Geometridae (values of *SH-like*, UFBoot2 =

Deleted: ,

Deleted: *SH-like*

Formatted: Right: 0.25"

100) (Fig. 1). The general patterns in our phylogenetic hypotheses suggest that Sterrhinae are the sister group to the rest of Geometridae. This subfamily is recovered as monophyletic when three genera traditionally included in Oenochrominae are considered to belong to Sterrhinae (see details below). Tribes in Sterrhinae, such as Timandriini, Rhodometrini, Lythriini, Rhodostrophiini and Cylopodini, are not recovered as monophyletic (Fig. 2). A detailed analysis, including formal changes to the classification of Sterrhinae, will be provided by Sihvonen et al. (in prep).

The monophyly of Larentiinae is established in previous studies (Sihvonen et al., 2011; Öunap et al., 2016) and our results are largely in agreement with their hypotheses. However, our results do not support the sister relationship between Sterrhinae and Larentiinae found in previous studies. Rather, we find that Sterrhinae are the sister to the rest of Geometridae. Within Larentiinae, in concordance with recent findings (Sihvonen et al., 2011; Öunap et al. 2016; Strutzenberger et al., 2017), we find Dyspteridini as the sister group to the remaining Larentiinae (Fig. 3). Phylogenetic relationships within Larentiinae were treated in detail by Öunap et al. (2016). Further details of the analyses and changes to the classification of Larentiinae will be discussed by Brehm et al. (submitted) and Öunap et al. (in prep).

Archiearinae are represented by more taxa than in a previous study (Sihvonen et al., 2011). Archiearinae grouped as sister to Oenochrominae + Desmobathrinae complex+ *Eumelea* + Geometrinae and Ennominae (Fig. 4). The monophyly of this subfamily is well supported (values of SH-like, UFBoot2 = 100). However, as in the previous study (Sihvonen et al. 2011), the Australian genera *Dirce* Prout, 1910 and *Acalyphes* Turner, 1926 are not part of Archiearinae but can clearly be assigned to Ennominae. Unlike previously assumed (e.g., McQuillan & Edwards 1994), the subfamily Archiearinae does probably not occur in Australia, despite superficial similarities of *Dirce*, *Acalyphes* and Archiearinae.

Desmobathrinae were shown to be paraphyletic by Sihvonen et al. (2011). In our analysis, the monophyly of this subfamily is not recovered either, as we find three taxa traditionally placed in Oenochrominae (i.e. *Zancloteryx* Herrich-Schäffer, [1855] and *Racasta* Walker, 1861) nested within Desmobathrinae (Fig. 4). We formally transfer these taxa to Desmobathrinae. In the revised sense, Desmobathrinae form a well-supported group with two main lineages. One of them comprises *Ozola* Walker, 1861, *Derambila* Walker, [1863] and *Zancloteryx*. This lineage is sister to a well-supported clade comprising *Conolophia* Warren, 1894, *Noreia* Walker, 1861,

Deleted: as

Deleted: ing

Deleted: Cosymbiini and

Deleted: recover

Deleted: + Trichopterygini

Deleted: , *Nearcha* Guest, 1887

Deleted: the genera

Formatted: Right: 0.25"

394 *Leptoctenopsis*, *Racasta*, *Ophiogramma* Hübner, [1831], *Pycnoneura* Warren, 1894 and  
 395 *Dolichoneura* Warren, 1894.  
 396 Oenochrominae in the broad sense are not a monophyletic group. However,  
 397 Oenochrominae *sensu stricto* (Scoble & Edwards, 1990) form a well-supported lineage  
 398 comprising two clades. One of them contains a polyphyletic *Oenochroma* with *O. infantilis*  
 399 Prout, 1910 being sister to *Dinophalus* Prout, 1910, *Hypographa* Guenée, [1858], *Lissomma*  
 400 Warren, 1905, *Sarcinodes* Guenée, [1858] and two further species of *Oenochroma*, including the  
 401 type species *O. vinaria* Guenée, [1858]. The other clade comprises *Monoctenia* Guenée, [1858],  
 402 *Onycodes* Guenée, [1858], *Parepisparis* Bethune-Baker, 1906, *Antictenia* Prout, 1910, *Arthodia*  
 403 Guenée, [1858], *Gastrophora* Guenée, [1858] and *Homospira* Turner, 1904 (Fig. 4). Most of the  
 404 remaining genera traditionally placed in Oenochrominae, including e.g. *Epidesmia* Duncan [&  
 405 Westwood], 1841, form a well-supported monophyletic clade that is sister to Oenochrominae *s.*  
 406 *str.* + *Eumelea ludovicata* + Geometrinae + Ennominae assemblage.

407 The genus *Eumelea* Duncan [& Westwood], 1841 has an unclear phylogenetic position in our  
 408 analyses. The IQ-TREE result suggests *Eumelea* to be sister to the subfamily Geometrinae (SH-  
 409 like = 93.6, UFBoot2 = 71, whereas RAXML recovered *Eumelea* in Ennominae as sister of  
 410 *Plutodes* Guenée, [1858] (RBS = 60).

412 The monophyly of Geometrinae is well supported (Fig. 5) and in IQ-TREE results  
 413 Geometrinae are recovered as the sister-taxon of *Eumelea*. The *Eumelea* + Geometrinae clade is  
 414 sister to Oenochrominae *s. str.* Although a recent phylogenetic study proposed several taxonomic  
 415 changes (Ban et al., 2018), the tribal composition in Geometrinae is still problematic. Many tribes  
 416 are recovered as paraphyletic. *Ornithospila* Warren, 1894 and *Agathia* Guenée, [1858], our  
 417 results suggest are the sisters to the rest of Geometrinae. *Chlorodontopera* is placed as an isolated  
 418 lineage sister of *Aracimini*, *Neohipparchini*, *Timandromorphini*, *Geometrini* and *Comibaenini*  
 419 which are recovered as monophyletic groups. *Synchlorini* are nested within *Nemoriini* in a well-  
 420 supported clade (support branch SH-like = 98.3, UFBoot2 = 91, RBS = 93). The monophyly of  
 421 *Pseudoterpnini* could not be recovered, instead this tribe splits up into three well-defined groups.  
 422 Several genera currently placed in *Pseudoterpnini s.l.* are recovered as an independent lineage  
 423 clearly separate from *Pseudoterpnini s.str.* (SH-like, UFBoot2 = 100). *Xenozancla* Warren, 1893  
 424 is sister to a clade comprising *Dysphaniini* and *Pseudoterpnini s.str.* *Hemitheini sensu* Ban et al.

Deleted: The genus *Eumelea* Duncan [& Westwood], 1841 has an unclear phylogenetic position in our analyses. The IQ-TREE result suggests *Eumelea* to be sister to the subfamily Geometrinae (SH-like = 92.5, UFBoot2 = 98, whereas RAXML recovered *Eumelea* in Ennominae as sister of *Plutodes* Guenée, [1858] (RBS = 60)....

Deleted: the genera

Deleted: *Ergavia* Walker, 1866, *Ametris* Guenée, [1858] and *Macrotis* Westwood, 1841 form a monophyletic group within Sterrhinae (see also Sihvonen et al., 2011), ...

Formatted: Highlight

Moved (insertion) [2]

Deleted: Although there are no phylogenetic studies which investigate the relationship between ...

Deleted: that these genera are sister clades

Deleted: .

Formatted: Font: Italic

Moved (insertion) [5]

Deleted: 100

Deleted: SH-like = 99.8,

Formatted: Right: 0.25"

(2018) are recovered as a well-supported clade. The African *Lophostola* Prout, 1912 and *Crypsiphona ocularia* Meyrick, 1888 were resolved as a single lineages, close to Hemitheini. Ennominae are strongly supported as monophyletic in IQ-TREE analyses (SH-like = 100, and UFBoot2) whereas in RAxML the monophyly is weakly supported (RBS = 63). Detailed results concerning the classification, especially for the Neotropical taxa, will be presented by Brehm et al. (submitted), but the main results are summarized here (Fig. 6). Very few tribes are monophyletic according to the results of the present study. One group of Neotropical taxa currently assigned to Gonodontini (unnamed E1), *Idialcis* Warren 1906 (unnamed clade E2), *Gonodontini* s.str., Gnophini, Odontoperini, unnamed clade E3, Nacophorini, and Ennomini (*sensu* Beljaev, 2008) group together (SH-like = 90.3, UFBoot2 = 87). Ennomini were sister to this entire group. *Campaeni* is recovered as sister of *Alsophilini* + *Wilemaniini* and *Colotoini*. In turn, they are sister to the New Zealand genus *Declana* Walker, 1858 (unnamed E4), which appeared as an isolated lineage sister to *Acalyphes* Turner, 1926 + *Dirce* Prout, 1910 and a large complex including Lithinini, intermixed with some genera currently placed in Nacophorini and Diptychini. *Neobapta* Warren, 1904 and *Oenoptila* Warren, 1895 formed an independent lineage (unnamed E5) sister to Theriini, which form a supported clade with *Lomographa* (Baptini). Likewise, we recovered *Erastria* Hübner, [1813] + *Metarranthia* Warren, 1894 (unnamed E6) and *Plutodes* Guenée, [1858] + *Palyadini*. The IQ-TREE analyses show *Palyadini* as a well-defined lineage, sister to *Plutodes*. However, in RAxML analyses, *Eumelea* and *Plutodes* group together and *Palyadini* cluster with a group of Caberini species. *Apeirini* formed a lineage with *Hypochrosini*, Epionini, *Sericosema* Warren, 1895 and *Ithysia* Hübner, [1825]. This lineage is in turn sister of African *Drepanogynis* Guenée, [1858] which groups together with *Sphingomima* Warren, 1899, *Thenopa* Walker, 1855 and *Hebdomophruda* Warren, 1897. Caberini are sister to an unnamed clade composed of *Trotogonia* Warren, 1905, *Acrotomodes* Warren, 1895, *Acrotomia* Herrich-Schäffer, [1855] and *Pyrinia* Hübner, 1818. Finally, our analyses recover a very large, well-supported clade comprising the tribes Macariini, Cassymini, Abraxini, Eutoeini and Boarmiini (SH-like = 100 and UFBoot2 = 99). This large clade has previously been referred to informally as the “boarmiines” by Forbes (1948) and Wahlberg et al. (2010). The tribe Cassymini is clearly paraphyletic: genera such as *Cirrhosoma* Warren, 1905, *Berberodes* Guenée, 1858, *Hemiphricta* Warren, 1906 and *Ballantiophora* Butler, 1881 currently included in Cassymini, cluster in their

- Deleted: , which is sister to the rest of Geometrinae
- Deleted: In turn, t
- Deleted: genus
- Moved (insertion) [3]
- Deleted: was
- Deleted: as sister to all other Hemitheini
- Deleted: The monophyly of Pseudoterpnini could not be recovered, instead this tribe splits up into three well-defined groups. In addition, several genera currently placed in Pseudoterpnini s.l. are recovered as an independent lineage clearly separate from Pseudoterpnini s.str. (SH-like = 86.3, UFBoot2 = 96). The monophyly of Pseudoterpnini could not
- Moved up [4]: The monophyly of Pseudoterpnini could not be recovered, instead this tribe splits up into three well-
- Moved up [3]: In addition, several genera currently placed
- Moved up [2]: Although there are no phylogenetic studies
- Moved up [5]: Aracimini, Neohipparchini,
- Moved (insertion) [4]
- Deleted: *Crypsiphona ocularia* Meyrick, 1888 is recovered...
- Formatted: Pattern: Clear (White), Not Highlight
- Deleted: , and SH-like = 100
- Deleted: and *Bryoptera* Guenée, [1858] + *Ectropis* Hübner...
- Deleted: in a large well-supported clade
- Deleted: 96.6
- Deleted: 9
- Deleted: 7
- Formatted: Font: Italic
- Deleted: The New Zealand genus *Declana* Walker, 1858... [3]
- Commented [WD9]: How supported? Weakly? Strongly? [4]
- Deleted: con
- Formatted: Font: Italic
- Deleted: re
- Deleted: close to
- Deleted: *Palyadini* +
- Deleted: .
- Deleted: *Neobapta* Warren, 1904 and *Oenoptila* Warren... [5]
- Deleted: *Hypochrosini*
- Deleted: *Apeirini*
- Deleted: of
- Commented [WD10]: Elements of a series that contain... [6]
- Deleted:
- Deleted: = 100
- Formatted: Right: 0.25"

own clade together with *Dorsifulcrum* Herbulot, 1979 and *Odontognophos* Wehrli, 1951. We were unable to include Orthostixinae in the analyses, so we could not clarify the taxonomic position of this subfamily with regard to *its* possible synonymy with Ennominae (Sihvonen et al., 2011).

Deleted: , as sister to the Abraxini-Eutocini complex

Deleted: the

Formatted: Font: (Default) Times, Font color: Text 1, Pattern: Clear (White)

## Discussion

### *Optimal partitioning scheme and support values*

The greedy algorithm implemented in ModelFinder to select the best-fitting partitioning scheme parsed the codon positions into 26 subsets. The results recovered highest values (AIC and BIC) for data partitioned by codon position. These results are not different from previous studies that tested the performance of different data partitioning schemes and found that in some cases partitioning by gene can result in suboptimal partitioning schemes and may limit the accuracy of phylogenetic analyses (Lanfear et al., 2012). However, we note that although the AIC and BIC values were lower when the data were partitioned by gene, the tree topology recovered was almost the same as when data were partitioned by codon, suggesting that much of the phylogenetic signal in the data is robust to partitioning schemes. As would be expected, the analyses yielded different measures of node support. Ultrafast bootstrap gave the highest support values, followed by SH-like and finally standard bootstrap as implemented in RAxML gave the lowest. Although support indices obtained by these methods are not directly comparable, differences in node support of some clades can be attributed to the small number of markers, insufficient or saturated divergence levels (Guindon et al., 2010).

Deleted: treated

Deleted: partitions

Deleted: independently and failed to merge any

Deleted: data

Commented [WD11]: Awkward. Can this sentence be combined with previous for clarity. Otherwise re-write.

Deleted: highlight

Deleted: in

Deleted: nevertheless

Deleted: The

Deleted: resulted

Deleted: in some disagreements between the

Commented [WD12]: Why is this word inserted here? Sounds redundant with "small numbers of markers."

### *Current understanding of Geometridae phylogeny and taxonomic implications*

#### **Geometridae Leach, 1815**

The phylogenetic data presented in this study are by far the most comprehensive to date in terms of the number of markers, sampled taxa, and geographical coverage. In total, our sample includes 814 genera, thus representing 41% of the currently recognised Geometridae genera (Scoble & Hausmann, 2007). Previous phylogenetic hypotheses were based mainly on the European fauna and many clades were not unambiguously supported due to low taxon sampling. The general

Deleted: hypothesis

Deleted: is

Formatted: Right: 0.25"

593 patterns of the phylogenetic relationships among the subfamilies recovered in our study largely  
 594 agrees with previous hypotheses based on morphological characters and different sets of  
 595 molecular markers (Holloway, 1997; Abraham, 2001; Yamamoto & Sota, 2007; Sihvonen et al.,  
 596 2011). However, the results of our larger dataset differ in many details and shed light on the  
 597 phylogenetic relationships of several, poorly resolved, small subfamilies.

Deleted: between

598 Sterrhinae are recovered as the sister subfamily to the remaining Geometridae. This result  
 599 is not in concordance with Sihvonen et al. (2011), Yamamoto & Sota (2007) and Regier et al.  
 600 (2009), who found a sister group relationship between Sterrhinae and Larentiinae which in turn  
 601 were sister to the rest of Geometridae. Sihvonen et al. (2011) showed these relationships with low  
 602 support, while Yamamoto & Sota (2007) and Regier et al. (2009) included only a few samples in  
 603 their analyses. Our analyses include representatives from almost all known tribes currently  
 604 included in Sterrhinae and Larentiinae. The higher number of markers, improved methods of  
 605 analysis, the broader taxon sampling as well as the stability of our results suggests that Sterrhinae  
 606 are indeed the sister group to the remaining Geometridae. Sterrhinae (after transfer of *Ergavia*,  
 607 *Ametris* and *Macrotis*, see details below), Larentiinae, Archiearinae, Geometrinae and  
 608 Ennominae were highly supported as monophyletic. Oenochrominae and Desmobathrinae formed  
 609 polyphyletic and paraphyletic assemblages respectively. The monophylies of Oenochrominae and  
 610 Desmobathrinae have long been questioned. Morphological studies addressing Oenochrominae or  
 611 Desmobathrinae have been limited and the majority of genera have never been examined in  
 612 depth. In addition, it has been very difficult to establish the boundaries of these subfamilies on  
 613 the basis of morphological structures (Scoble & Edwards, 1990). Sihvonen et al. (2011) showed  
 614 that neither Oenochrominae nor Desmobathrinae were monophyletic, but these results were  
 615 considered preliminary due to the limited number of sampled taxa, and as a consequence no  
 616 formal transfers were proposed. To date, the phylogenetic positions of these subfamilies have  
 617 been unclear.

Deleted: especially the

Deleted: previously

Commented [WD13]: Not clear what these refers to here. Your findings or those of Yamamoto & Sota (2007) + Regier et al. (2009)

Formatted: Highlight

Formatted: Highlight

Deleted: , which could have had an influence on the results

Deleted: always

Deleted: only

Deleted: examination

Formatted: Strikethrough

618 The systematic status of Orthostixinae remains uncertain because it was not included in  
 619 our study. Sihvonen et al. (2011) included the genus *Naxa* Walker, 1856, formally placed in  
 620 Orthostixinae, and found it to be nested within Ennominae. However, only three genes were  
 621 successfully sequenced from this taxon, and its position in the phylogenetic tree turned out to be  
 622 highly unstable in our analyses. It was thus excluded from our dataset. *Orthostixis*  
 623 Hübner, [1823], the type genus of the subfamily, needs to be included in future analyses.

Deleted: Without a doubt,

Formatted: Right: 0.25"

632

633 **Sterrhinae Meyrick, 1892**

634 We included 74 Sterrhinae taxa in our analyses, with all tribes recognized in Forum Herbulot  
 635 (2007) being represented. The recovered patterns generally agree with previous phylogenetic  
 636 hypotheses of the subfamily (Sihvonen, 2004, Sihvonen et al., 2011). The genera *Ergavia*  
 637 *Walker, 1866*, *Ametris* *Guenée, [1858]* and *Macrotis* *Westwood, 1841*, which currently are  
 638 placed in Oenochrominae were found to form a well-defined lineage within Sterrhinae with  
 639 strong support (SH-Like = 99 UFBot2 = 100). These genera are distributed in the New World,  
 640 whereas the range of true Oenochrominae is restricted to the Australian and Oriental *Regions*.  
 641 Sihvonen et al. (2011) already found that *Ergavia* and *Afrophylla* Warren, 1895 belong to  
 642 Sterrhinae and suggested more extensive analyses to clarify the position of these genera, which  
 643 we did. *Afrophylla* was transferred to Sterrhinae by Sihvonen & Staude (2011) and *Ergavia*,  
 644 *Ametris* and *Macrotis* (plus *Almodes* Guenée, [1858]) will be transferred by Sihvonen et al. (in  
 645 prep.).

646 Cosymbiini, Timandrini, Rhodometrini and Lythriini are closely related as shown  
 647 previously (Sihvonen & Kaila, 2004; Öunap et al., 2008; Sihvonen et al., 2011). Cosymbiini  
 648 appear as sister to the Timandrini + *Traminda* *Saalmüller, 1891* + *Pseudosterrha* *Warren, 1888*  
 649 and Rhodometrini + Lythriini clade. Lythriini are closely related to Rhodometrini as shown by  
 650 Öunap et al. (2008) with both molecular and morphological data. *While* *Traminda* (Timandrini)  
 651 and *Pseudosterrha* (Cosymbiini) grouped together forming a different lineage *that* is sister to  
 652 Rhodometrini + Lythriini clade (Fig. 2).

653 Rhodostrophiiini and Cyllopodini were recovered *as* polyphyletic with species of  
 654 Cyllopodini clustering within Rhodostrophiiini. Similar results were recovered *previously*  
 655 (Sihvonen & Kaila, 2004; Sihvonen et al., 2011), suggesting that *additional* work *is needed* to  
 656 *clarify* the status and systematic positions of these tribes. Sterrhini and Scopulini were recovered  
 657 as sister taxa as proposed by Sihvonen & Kaila (2004), Hausmann (2004), Öunap et al. (2008)  
 658 and Sihvonen et al. (2011). Our new phylogenetic hypothesis constitutes a large step towards  
 659 understanding the evolutionary relationships of the major lineages of Sterrhinae. Further  
 660 taxonomic changes and more detailed interpretation of the clades *are ongoing*.

661

662 **Larentiinae Duponchel, 1845**

Formatted: Font: 12 pt, Not Highlight

Formatted: Font: 12 pt, Not Highlight

Formatted: Font: 12 pt, Not Highlight

Deleted: region

Deleted: However, Timandrini was not the closest to Rhodometrini + Lythriini clade due to the phylogenetic position of ...

Deleted: Saalmüller, 1891

Deleted: Warren, 1888

Deleted: . These taxa

Deleted: which

Deleted: before

Deleted: further

Deleted: needs

Deleted: be done to

Deleted: On the other hand,

Deleted: );

Deleted: );

Deleted: will be dealt with

Deleted: by Sihvonen et al. (in prep.).

Formatted: Right: 0.25"



Larentiinae are a monophyletic entity (Fig. 3). **Partly** in concordance with the results of Sihvonen et al. (2011), Viidalepp (2011), Õunap et al. (2016) and Strutzenberger et al. (2017), Dyspteridini are **supported as** sister to all other larentiines. Remarkably, *Brabiroides* Warren, 1904 forms an independent lineage. Chesiadini are monophyletic and sister to all larentiines except Dyspteridini, *Brabiroides* and Trichopterygini. These results do not support the suggestion by Viidalepp (2006) and Sihvonen et al. (2011) that Chesiadini are sister to Trichopterygini.

In our phylogenetic hypothesis, Asthenini are sister to the Perizomini + Melanthiini + Eupitheciini clade. These results do not fully agree with Õunap et al. (2016) who found Asthenini to be sister to all Larentiinae except Dyspteridini, Chesiadini, Trichopterygini and Eudulini. However, our results do support the Melanthiini + Eupitheciini complex as a sister lineage to Perizomini. Sihvonen et al. (2011) recovered Phileremini and Rheumapterini as well-supported sister taxa. Our results suggest *Triphosa dubitata* (**Triphosini**) Linnaeus 1758 **is** sister **to** Phileremini, **with** Rheumapterini sister to this clade. Cidariini were recovered as **paraphyletic**, as the genera *Coenotephria* Prout, 1914 and *Lampropteryx* Stephens, 1831 cluster in a different clade (**unnamed clade L7**) apart from the lineage comprising the type genus of the tribe, *Cidaria* Treitschke, 1825. *Ceratotalia* Packard, 1876, currently placed in Hydriomenini and *Trichodezia* Warren, 1895 **nested within** Cidariini. This result is not in concordance with Õunap et al. (2016), who **regarded** this tribe **to be** monophyletic. Scotopterygini are sister to a lineage comprising *Ptychorrhoe blosyrata* Guenée [1858], *Disclisioprocta natalata* (Walker, 1862) (**placed in unnamed clade L8**), Euphyiini, **an unnamed clade L9 comprising the genera** *Pterocypha*, *Archirhoe* and *Obila*, Xanthorhoini and Cataclymini. Euphyiini are monophyletic, but Xanthorhoini are recovered as mixed with Cataclymini. The same findings were shown by Õunap et al. (2016), but no taxonomic rearrangements were proposed. Larentiini are monophyletic and sister of **Hydriomenini**, **Heterusiini**, Erateinini, Stamnodini and some unnamed clades (**L11-14**). Although with some differences, our results support the major phylogenetic patterns of Õunap et al. (2016).

Despite substantial progress, the tribal classification and phylogenetic relationships of Larentiinae are far from being resolved (Õunap et al. 2016). Forbes (1948) proposed eight tribes based on morphological information, Viidalepp (2011) raised the number to 23 and Õunap et al. (2016) recovered 25 tribes studying 58 genera. Our study includes 23 **of the currently recognized** tribes and 125 genera (with **an emphasis** on Neotropical taxa). However, the phylogenetic

Deleted: ial

Deleted: I

Deleted: of

Deleted: while

Deleted: is the

Deleted: polyphyletic

Deleted:

Deleted: Also,

Deleted: were mixed

Deleted:

Deleted: found

Deleted: *sp*

Formatted: Font: Italic

Formatted: Font: Italic

Formatted: Font: Italic

Deleted: an unnamed clade

Deleted: r

Deleted: Hydriomenini,

Deleted: Heterusiini are recovered as a polyphyletic group, while Erateinini are close to Stamnodini as proposed by Sihvonen et al. (2011). Although with some differences, our

Deleted: sufficiently

Deleted: a focus

Formatted: Right: 0.25"



731 position of many taxa remains unclear, and many tropical genera have not yet been formally  
732 assigned to a tribe. Formal descriptions of these groups will be treated in detail by Brehm et al.  
733 (submitted) and Öunap et al. (in prep).

Formatted: Highlight

Formatted: Highlight

Deleted: any

734  
735 I STOPPED LINE EDITING HERE. WILL SKIM TO CONCLUSION

#### 736 737 **Archiearinae Fletcher, 1953**

738 The hypothesis presented in this study recovered Archiearinae as a monophyletic entity if some  
739 taxonomic rearrangements are performed. This subfamily was previously considered as sister to  
740 Geometrinae + Ennominae (Abraham et al., 2001), whereas Yamamoto & Sota (2007) proposed  
741 them as the sister-taxon to Orthostixinae + Desmobathrinae. Our findings agree with Sihvonen et  
742 al. (2011) who recovered Archiearinae as the sister-taxon to the rest of Geometridae excluding  
743 Sterrhinae and Larentiinae, although only one species was included in their study. *Archiearis*  
744 Hübner, [1823] is sister to *Boudinotiana* Esper, 1787 and these taxa in turn are sister to  
745 *Leucobrephe* Grote, 1874 (Fig. 4). The southern hemisphere Archiearinae require more  
746 attention. Young (2006) suggested that two Australian Archiearinae genera, *Dirce* and *Acalyphes*,  
747 actually belong to Ennominae. Our analyses clearly support this view and we therefore propose to  
748 formally transfer *Dirce* and *Acalyphes* to Ennominae (all formal taxonomic changes are provided  
749 in Table 1). Unfortunately, the South American Archiearinae genera *Archiearides* Fletcher, 1953  
750 and *Lachnocephala* Fletcher, 1953, and Mexican *Caenosyntes* Dyar, 1912 (Pitkin & Jenkins  
751 2004), could not be included in our analyses. The position in Archiearinae requires further study.  
752 These presumably diurnal taxa may only be superficially similar to northern hemisphere  
753 Archiearinae as was the case with Australian *Dirce* and *Acalyphes*.

#### 754 755 **Desmobathrinae Meyrick, 1886**

756 Taxa placed in Desmobathrinae were formerly recognized as Oenochrominae genera with slender  
757 appendages. Holloway (1996) revived this subfamily from synonymy with Oenochrominae and  
758 divided it into the tribes Eumeleini and Desmobathrini. Desmobathrinae species have a  
759 pantropical distribution and they apparently (still) lack recognized morphological apomorphies  
760 (Holloway, 1996). Our phylogenetic analysis has questioned the monophyly of Desmobathrinae  
761 *sensu* Holloway because some species currently placed in Oenochrominae were embedded within

Formatted: Right: 0.25"

763 the group (see also Sihvonen et al., 2011), and also the phylogenetic position of the tribe  
764 Eumeleini is unstable (see below). Desmobathrinae can be regarded as a monophyletic group in  
765 our study, after the transfer of *Zanclopteryx*, *Nearcha* and *Racasta* from Oenochrominae to  
766 Desmobathrinae, and the removal of Eumeleini (Table 1). Desmobathrinae as circumscribed here  
767 are an independent lineage that is sister to all Geometridae except Sterrhinae, Larentiinae and  
768 Archiearinae.

769 The monobasic Eumeleini has had a dynamic taxonomic history: *Eumelea* was transferred  
770 from Oenochrominae *s.l.* to Desmobathrinae based on the pupal cremaster (Holloway, 1996),  
771 whereas Beljaev (2008) pointed out that *Eumelea* could be a member of Geometrinae based on  
772 the skeleto-muscular structure of the male genitalia. Molecular studies (Sihvonen et al., 2011,  
773 Ban et al., 2018) suggested that *Eumelea* was part of Oenochrominae *s.str.*, but these findings  
774 were not well-supported and no formal taxonomic changes were proposed. Our analyses with  
775 IQTREE and RAxML recovered Eumeleini in two very different positions, either as sister to  
776 Geometrinae (SH-like = 93.6, UFBoot2 = 71) rather than belonging to Desmobathrinae (figs 4,  
777 5), or as sister of *Plutodes* in Ennominae (RBS = 60) (Supplemental data S4). The examination of  
778 morphological details suggests that the position as sister to Geometrinae is more plausible:  
779 hindwing vein M2 is present and tubular; anal margin of the hindwing is elongated; and large  
780 coremata originate from saccus (Holloway 1994, our observations). The morphology of *Eumelea*  
781 is partly unusual, and for that reason we illustrate selected structures (Supplemental data S5),  
782 which include for instance the following: antennae and legs of both sexes are very long; forewing  
783 vein Sc (homology unclear) reaches wing margin; in male genitalia coremata are extremely large  
784 and branched; uncus is cross-shaped (cruciform); tegumen is narrow and it extends ventrally  
785 beyond the point of articulation with vinculum; saccus arms are extremely long, looped; and  
786 vesica is with lateral rows of cornuti. However, the green geoverdin pigment concentration of  
787 *Eumelea* is low in comparison to Geometrinae (Cook et al., 1994). We tentatively conclude that  
788 *Eumelea* is probably indeed associated with Geometrinae. However, since eleven genetic markers  
789 were not sufficient to clarify the phylogenetic affinities of *Eumelea*, we provisionally place the  
790 genus as *incertae sedis* (Table 1).

791

792 **Oenochrominae Guenée, [1858]**

Deleted: 92

Deleted: 98

Formatted: Right: 0.25"

Oenochrominae has obviously been the group comprising taxa that could not easily be assigned to other subfamilies. Out of the 76 genera currently assigned to Oenochrominae, our study includes 25 genera (28 species). Three of these genera will be formally transferred to Sterrhinae (Sihvonen et al. in prep.), two are here transferred to Desmobathrinae (see above, Table 1), and eight are transferred to Epidesmiinae (see below). In agreement with Sihvonen et al. (2011), Oenochrominae *s. str.* grouped together in a well-supported lineage. Genera of this clade can be characterized as having robust bodies, and their male genitalia have a well-developed uncus and gnathos, broad valvae and a well-developed anellus (Scoble & Edwards, 1990). Common host plants are members of Proteaceae and Myrtaceae (Holloway, 1996). Our results strongly suggest that the genus *Oenochroma* is polyphyletic: *O. infantilis* is sister to a clade including *Dinophalus*, *Hypographa*, *Lissomma*, *Sarcinodes* and (at least) two species of *Oenochroma*. To date, 20 species have been assigned to *Oenochroma* by Scoble (1999), and one additional species was described by Hausmann et al. (2009), who suggested that *O. vinaria* is a species complex. We agree with Hausmann et al. (2009), who pointed out the need of major revision and taxonomic definition of *Oenochroma*.

In our phylogenetic hypothesis, *Sarcinodes* is sister to *O. orthodesma* and *O. vinaria*, the type species of *Oenochroma*. Although *Sarcinodes* and *Oenochroma* resemble each other in external morphology, a sister-group relationship between these genera has not been hypothesized before. The inclusion of *Sarcinodes* in Oenochrominae is mainly based on shared tympanal characters (Scoble & Edwards, 1990). However, the circular form of the lacinia, which is an apomorphy of Oenochrominae *s.str.* is missing or not apparent in *Sarcinodes* (Holloway, 1996). In addition, *Sarcinodes* is found in the Oriental rather than in the Australian region, where all *Oenochroma* species are distributed. A second clade of Oenochrominae *s.str.* comprises the genera *Monoctenia*, *Onycodes*, *Parepisparis*, *Antictenia*, *Arhodia*, *Gastrophora* and *Homospora*, which clustered together as the sister of *Oenochroma* and its relatives. These genera are widely recognized in sharing similar structure of male genitalia (Scoble & Edwards, 1990), yet their phylogenetic relationships have never been tested. Young (2006) suggested the monophyly of Oenochrominae *s.str.*, however, with a poorly resolved topology and low branch support. In her study, *Parepisparis*, *Phallaria* and *Monoctenia* shared a bifid head, while in *Parepisparis* and *Onychodes*, the aedeagus was lacking caecum and cornuti. Our analysis supports these morphological similarities. *Monoctenia*, *Onycodes* and *Parepisparis* clustered together. However,

Formatted: Right: 0.25"

826 a close relationship of the genera *Antictenia*, *Arhodia*, *Gastrophora* and *Homospora* has not been  
827 suggested before. Our analysis thus strongly supports the earliest definition of Oenochrominae  
828 proposed by Guenée (1858), and reinforced by Cook & Scoble (1992). Oenochrominae should be  
829 restricted to *Oenochroma* and related genera such as *Dinophalus*, *Hypographa*, *Lissomma*,  
830 *Sarcinodes*, *Monoctenia*, *Onycodes*, *Parepisparis*, *Antictenia*, *Arhodia*, *Gastrophora*,  
831 *Homospora*, *Phallaria* and *Palaeodoxa*. We consider that genera included in Oenochrominae by  
832 Scoble & Edwards (1990), but recovered in a lineage separate from *Oenochroma* and its close  
833 relatives in our study, belong to a hitherto unknown subfamily, which is described below.

834

835 **Epidesmiinae** Murillo-Ramos, Brehm & Sihvonen **new subfamily**

836 [LSIDurn:lsid:zoobank.org:act:34D1E8F7-99F1-4914-8E12-0110459C2040](https://lsid.zoobank.org/act:34D1E8F7-99F1-4914-8E12-0110459C2040)

837 Type genus: *Epidesmia* Duncan [ & Westwood ], 1841.

838 Material examined: Taxa included in the molecular phylogeny: *Ecphyas holopsara* Turner, 1929,  
839 *Systatica xanthastis* Lower, 1894, *Adeixis griseata* Hudson 1903, *Dichromodes indicataria*  
840 Walker 1866, *Phrixocomes* sp. Turner 1930, *Abraxaphantes perampla* Swinhoe 1890, *Epidesmia*  
841 *chilonaria* Herrich-Schäffer [1855], *Phrataria replicataria* Walker 1866.

842 Most of the slender-bodied Oenochrominae, excluded from Oenochrominae *s. str.* by Holloway  
843 (1996), were recovered as an independent lineage (Fig. 4) that consists of two clades: *Ec.*  
844 *holopsara* + *S. xanthastis* and *Ep. chilonaria* + five other genera. Branch support values from IQ-  
845 TREE strongly support the monophyly of this clade (SH-like and UFBoot2 = 100), while in  
846 RAXML the clade is moderately supported (RBS = 89). These genera have earlier been assigned  
847 to Oenochrominae *s.l.* (Scoble & Edwards, 1990). However, we recovered the group as a well-  
848 supported lineage independent from Oenochrominae *s. str.* and transfer them to Epidesmiinae,  
849 subfam. n. (Table 1).

850 Phylogenetic position: Epidesmiinae is sister to Oenochrominae *s. str.* + *Eumelea* + Geometrinae  
851 + Ennominae.

852 Short description of Epidesmiinae: Antennae in males unipectinate (exception: *Adeixis*), towards  
853 apex shorter towards the apex. Pectination moderate or long. Thorax and abdomen slender  
854 (unlike in Oenochrominae). Forewings with sinuous postmedial line and areole present.

Deleted: *Ecphyas*

Deleted: *Systatica*

Deleted: and *Epidesmia*

Deleted: and SH-like

Formatted: Right: 0.25"

859 Forewings planiform (with wings lying flat on the substrate) in resting position, held like a  
860 triangle, and cover the hindwings.

861 Diagnosis of Epidesmiinae: The genera included in this subfamily form a strongly supported  
862 clade with DNA sequence data from the following gene regions (exemplar *Epidesmia chilonaria*  
863 Herrich-Schäffer, [1855]) ArgK (MK738299), Ca-ATPase (MK738690), CAD (MK738960),  
864 COI (MK739187), EF1a (MK740168), GAPDH (MK740402), MDH (MK740974) and Nex9  
865 (MK741433). A thorough morphological investigation of the subfamily, including diagnostic  
866 characters, is under preparation (Murillo-Ramos et al.).

867 Distribution: Most genera are distributed in the Australian region, with some species ranging into  
868 the Oriental region. *Abraxaphantes* occurs exclusively in the Oriental region.

#### 869 **Geometrinae Stephens, 1829**

871 The monophyly of Geometrinae is strongly supported, but the number of tribes included  
872 in this subfamily is still unclear. Sihvonen et al. (2011) analyzed 27 species assigned to 11 tribes,  
873 followed by Ban et al. (2018) with 116 species in 12 tribes. Ban et al. (2018) synonymized nine  
874 tribes, and validated the monophyly of 12 tribes, with two new tribes Ornithospilini and Agathiini  
875 being the first two clades branching off the main lineage of Geometrinae. Our study (168 species)  
876 validates the monophyly of 13 tribes, eleven of which were defined in previous studies:

877 Hemitheini, Dysphaniini, Pseudoterpnini s.str., Ornithospilini, Agathiini, Aracimini,  
878 Neohipparchini, Timandromorphini, Geometrini, Comibacini, Nemoriini. One synonymization is  
879 proposed: Synchlorini Ferguson, 1969 **syn. nov.** is synonymized with Nemoriini Gumpfenberg,  
880 1887. One tribe is proposed as new: Chlorodontoperini **trib. nov.**, and one tribe (Archaeobalbini

881 Viidalepp, 1981, **stat. rev.**) is raised from synonymy with Pseudoterpnini.

883 Although there are no phylogenetic studies which investigate the relationships between  
884 *Ornithospila* Warren, 1894 and *Agathia* Guenée, [1858], our results suggest  
885 Ornithospilini+Agathiini are sister clades. This tribes are the sisters to the rest of Geometrinae.  
886 *Chlorodontopera* is placed as an isolated lineage as shown by Ban et al. (2018). Given that  
887 *Chlorodontopera* is clearly forming an independent and well-supported lineage we propose the  
888 description of a new tribe *Chlorodontoperini*.

890 *Chlorodontoperini* Murillo-Ramos, Sihvonen & Brehm, **new tribe**

891 LSIDurn:lsid:zoobank.org:act:0833860E-A092-43D6-B2A1-FB57D9F7988D

Formatted: Font: Times New Roman

Formatted: Line spacing: Multiple 1.15 li

Field Code Changed

Deleted: GB Accession number

Formatted: Font: Times New Roman

Deleted: GB Accession number

Deleted: GB Accession number

Formatted: Font: Times New Roman

Field Code Changed

Formatted: Font: Times New Roman

Formatted: Font: Times New Roman

Field Code Changed

Formatted: Font: Times New Roman

Formatted: Font: Times New Roman

Field Code Changed

Formatted: Font: Times New Roman

Deleted: GB Accession number

Formatted: Font: Times New Roman

Field Code Changed

Formatted: Font: Times New Roman

Deleted: GB Accession number

Formatted: Font: Times New Roman

Field Code Changed

Formatted: Font: Times New Roman

Deleted: GB Accession number

Formatted: Font: Times New Roman

Field Code Changed

Formatted: Font: Times New Roman

Deleted: GB Accession number

Formatted: Font: Times New Roman

Field Code Changed

Formatted: Font: Times New Roman

Deleted: GB Accession number

Formatted: Font: Times New Roman

Deleted: (note to the editor: GB accession numbers will be provided on acceptance)...

Formatted: Font: (Default) Times New Roman

Deleted: .

Formatted: Font: (Default) Times New Roman, Font color: Auto, Not Highlight

Formatted: Right: 0.25"

Type genus: *Chlorodontopera* Warren, 1893

Material examined: Taxa in the molecular phylogeny: *C. discospilata* (Moore, 1867) and *C. mandarinata* (Leech, 1889).

Some studies (Inoue, 1961; Holloway, 1996) suggested the morphological similarities of *Chlorodontopera* Warren, 1893 with members of Aracimini. Moreover, Holloway (1996) considered this genus as part of Aracimini. Our results suggest a sister relationship of *Chlorodontopera* with Aracimini rather than the inclusion in the tribe as well as the sister relationship with a large lineage comprising the rest of Geometrinae. Considering that our analysis strongly supports *Chlorodontopera* as an independent lineage (branch support SH-like = 99 UFBoot2 = 100, RBS = 99), we introduce the monobasic tribe Chlorodontoperini. This tribe can be diagnosed by the combination of DNA data from six genetic markers (exemplar *Chlorodontopera discospilata*) CAD (MG015448), COI (MG014735), EF1a (MG015329), GAPDH (MG014862), MDH (MG014980) and RpS5 (MG015562). Ban et al. (2018) did not introduce a new tribe because the relationship between *Chlorodontopera* and *Euxena* Warren, 1896 was not clear in their study. This relationship was also been proposed by Holloway (1996) based on similar wing patterns. Further analyses are needed to clarify the affinities between *Chlorodontopera* and *Euxena*.

The tribe Chlorodontoperini is diagnosed by distinct discal spots with pale margins on the wings, which are larger on the hindwing; a dull reddish-brown patch is present between the discal spot and the costa on the hindwing, and veins M3 and CuA1 are not stalked on the hindwing (Ban et al., 2018). In the male genitalia, the socii are stout and setose and the lateral arms of the gnathos are developed, not joined. Sternite 3 of the male has setal patches. Formal taxonomic changes are listed in Table 1.

Aracimini, Neohipparchini, Timandromorphini, Geometrini and Comibaenini were recovered as monophyletic groups. These results are in full agreement with Ban et al. (2018). However, the phylogenetic position of *Eucyclodes* Warren, 1894 is uncertain (unnamed G2). This genus is placed as sister of Comibaenini (support branch SH-like = 32.4, UFBoot2 = 100, RBS = 67). The monophyly of Nemoriini and Synchronini is not supported. Instead, Synchronini are nested within Nemoriini (support branch SH-like = 98.3, UFBoot2 = 91, RBS = 93). Our findings are in concordance with Sihvonen et al. (2011) and Ban et al. (2018), but our analyses included a larger

Formatted: Right: 0.25"

number of markers and a much higher number of taxa. Thus, we formally synonymize  
 Synchronini **syn. nov.** with Nemoriini (Table 1).

The monophyly of Pseudoterpnini *sensu* Pitkin et al. (2007) could not be recovered.  
 Similar results were shown by Ban et al. (2018) who recovered Pseudoterpnini *s.l.* including all  
 the genera previously studied by Pitkin et al. (2007), forming a separate clade from *Pseudoterpna*  
 Hübner, [1823] + *Pingasa* Moore, 1887. Our results showed African *Mictoschema* Prout, 1922  
 falling within Pseudoterpnini *s.str.*, and it is sister to *Pseudoterpna* and *Pingasa*. A second group  
 of Pseudoterpnini *s.l.* was recovered as an independent lineage clearly separate from  
 Pseudoterpnini *s.str.* (SH-like = 88.3, UFBoot2 = 64). Ban et al. (2018) did not introduce a new  
 tribe due to the morphological similarities and difficulty in finding apomorphies of  
 Pseudoterpnini *s.str.* In addition, their results were weakly supported. Considering that two  
 independent studies have demonstrated the paraphyly of Pseudoterpnini *sensu* Pitkin et al (2007),  
 we see no reason for retaining the wide concept of this tribe. Instead, we propose the revival of  
 the tribe status of Archaeobalbini.

Archaeobalbini Viidalepp, 1981, status revised  
 (original spelling: Archeobalbini, justified emendation in Hausmann (1996))  
 Type genus: *Archaeobalbis* Prout, 1912 (synonymized with *Herochroma* Swinhoe, 1893 in  
 Holloway (1996))  
 Material examined: *Herochroma curvata* Han & Xue, 2003, *H. baba* Swinhoe 1893,  
*Metallophilia inanularia* Han & Xue, 2004, *M. cuneataria* Han & Xue, 2004, *Actenochroma*  
*muscoloraria* (Walker, 1862), *Absala dorcada* Swinhoe, 1893, *Metaterpna batangensis* Hang  
 & Stüning, 2016, *M. thyatiraria* (Oberthür, 1913), *Limbatochlamys rosthorni* Rothschild, 1894,  
*Pachyodes pictaria* Moore, 1888, *Dindica para* Swinhoe, 1893, *Dindicodes crocina* (Butler,  
 1880), *Lophophelma erionoma* (Swinhoe, 1893), *L. varicoloraria* (Moore, 1868), *L. iterans*  
 (Prout, 1926) and *Pachyodes amplificata* (Walker, 1862).

This lineage splits into four groups: *Herochroma* Swinhoe, 1893 + *Absala* Swinhoe, 1893 +  
*Actenochroma* Warren, 1893 is the sister lineage of the rest of Archaeobalbini that were  
 recovered as a polytomic bunch of three clades comprising the genera *Limbatochlamys*  
 Rothschild, 1894, *Psilotagma* Warren, 1894, *Metallophilia* Warren, 1895, *Metaterpna* Yazaki,

Moved (insertion) [6]

Deleted: 6

Deleted: 96

Deleted: and the description of a new tribe  
Chlorodontoperini.

Formatted: Right: 0.25"

969 1992, *Dindica* Warren, 1893, *Dindicodes* Prout, 1912, *Lophophelma* Prout, 1912 and *Pachyodes*  
970 *Guenée*, 1858. This tribe can be diagnosed by the combination of DNA data from six genetic  
971 markers, see for instance *Pachyodes amplificata* (CAD, COI, EF1a, GAPDH, MDH RpS5)  
972 shown in supplementary material. Branch support values in IQ-TREE strongly confirm the  
973 monophyly of this clade (SH-like = 88.3, UFBoot2 = 64). GenBank accession numbers are  
974 shown in supplementary material. A morphological diagnosis requires further research.

Deleted: 6

Deleted: 96

975  
976 *Xenozancla* Warren, 1893 (unnamed G3) is sister to the clade comprising *Dysphaniini*  
977 *and Pseudoterpnini s. str.* Sihvonon et al. (2011) did not include *Xenozancla* in their analyses and  
978 suggested a sister relationship of *Dysphaniini* and *Pseudoterpnini*, but with low support.  
979 According to Ban et al. (2018), *Xenozancla* is more closely related to *Pseudoterpnini s. str.* rather  
980 than to *Dysphaniini*. However, due to low support of clades, Ban et al. (2018) did not propose a  
981 taxonomic assignment to *Xenozancla*, which is currently not assigned to a tribe. Although our IQ-  
982 TREE results show that *Xenozancla* is sister of clade comprising *Dysphaniini* and *Pseudoterpnini*  
983 *s. str.*, the RAxML analysis did not recover the same phylogenetic relationships. Instead,  
984 *Dysphaniini + Pseudoterpnini s. str.* are found to be sister taxa, but *Xenozancla* is placed close to  
985 *Rhomborista monosticta* (Wehrli, 1924). As in Ban et al. (2018), due to low support of nodes, we  
986 cannot reach a conclusion about the phylogenetic affinities of these tribes based on our results.

Formatted: Indent: First line: 0.5"

987 The Australian genus *Crypsiphona* Meyrick, 1888 (unnamed G4) placed close to  
988 *Lophostola* and *Hemitheini*. *Crypsiphona* has been assigned to *Pseudoterpnini* (e. g. Pitkin et al.  
989 2007, Öunap & Viidalepp 2009), but is recovered as a separate lineage in our tree. Given the  
990 isolated position of *Crypsiphona*, the designation of a new tribe could be considered, but due to  
991 low support of branches in our analyses, further information (including morphology) is needed to  
992 confirm the phylogenetic position of this genus. In our phylogenetic hypothesis, a large clade  
993 including the former tribes Lophochoristini, Heliotheini, Microloxiini, Thalerini, Rhomboristini,  
994 Hemistolini, Comostolini, Jodini and Thalassodini is recovered as sister to the rest of  
995 Geometrinae. These results are in full agreement with Ban et al. (2018), who synonymized all of  
996 these tribes with *Hemitheini*. Although the monophyly of *Hemitheini* is strongly supported, our  
997 findings recovered only a few monophyletic subtribes. For example, genera placed in *Hemitheina*  
998 were intermixed with those belonging to *Microloxiina*, *Thalassodina* and *Jodina*. Moreover, many  
999 genera which were unassigned to tribe, were recovered as belonging to *Hemitheini*. Our findings

Formatted: Not Highlight

Formatted: Right: 0.25"



1002 recovered *Lophostola* Prout, 1912 as sister to all Hemitheini. These results are quite different  
1003 from those found by Ban et al. (2018) who suggested Rhomboristina as being sister to the rest of  
1004 Hemitheini. In contrast, our results recovered Rhomboristina mingled with Hemistolina. These  
1005 different results are probably influenced by the presence of African and Madagascan *Lophostola*  
1006 in our analysis. We feel that the subtribe concept is not practical thus do not advocate its use in  
1007 geometrid classification.

#### 1008 **Ennominae Duponchel, 1845**

1010 Ennominae are the most species-rich subfamily of geometrids. The loss of vein M2 on the  
1011 hindwing is probably the best apomorphy (Holloway, 1993), although this character does not  
1012 occur in a few ennomine taxa (Staude, 2001; Skou & Sihvonen, 2015). Ennominae are a  
1013 morphologically highly diverse subfamily, and attempts to find further synapomorphies shared by  
1014 all major tribal groups have failed.

1015 The number of tribes as well as phylogenetic relationships among tribes are still debatable  
1016 (see Skou & Sihvonen, 2015 for an overview). Moreover, the taxonomic knowledge of this  
1017 subfamily in tropical regions is still poor. Holloway (1993) recognized 21 tribes, Beljaev (2006)  
1018 24 tribes, and Forum Herbulot (2007) 27 tribes. To date, four molecular studies have  
1019 corroborated the monophyly of Ennominae (Yamamoto & Sota, 2007; Wahlberg et al., 2010;  
1020 Öunap et al., 2011, Sihvonen et al. 2011), with Young (2006) being the only exception who  
1021 found Ennominae paraphyletic. Moreover, three large-scale taxonomic revisions (without a  
1022 phylogenetic hypothesis) were published by Pitkin (2002) for the Neotropical region, Skou &  
1023 Sihvonen (2015) for the Western Palaearctic region, and Holloway (1994) for Borneo. More  
1024 detailed descriptions of taxonomic changes in Ennominae will be given by Brehm et al.  
1025 (submitted) and Murillo-Ramos et al. (in prep.). We here discuss general patterns and give details  
1026 for taxonomic acts not covered in the other two papers.

1027 Our findings recover Ennominae as a monophyletic entity, but results were not highly  
1028 supported in RAxML (RBS = 67) results compared to IQ-TREE (SH-Like = 100, UFBoot2 = 99).  
1029 The lineage comprising Geometrinae and Oenochrominae is recovered as the sister clade of  
1030 Ennominae. In previous studies, Wahlberg et al. (2010) sampled 49 species of Ennominae,  
1031 Öunap et al. (2011) sampled 33 species, and Sihvonen et al. (2011) 70 species including up to  
1032 eight markers per species. All these studies supported the division of Ennominae into

Deleted: ¶

**Moved up [6]:** The monophyly of Pseudoterpnini *sensu* Pitkin et al. (2007) could not be recovered. Similar results were shown by Ban et al. (2018) who recovered Pseudoterpnini *s.l.* including all the genera previously studied by Pitkin et al. (2007), forming a separate clade from *Pseudoterpna* Hübner, [1823] + *Pingasa* Moore, 1887. Our results showed African *Mictoschema* Prout, 1922 falling within Pseudoterpnini *s.str.*, and it is sister to *Pseudoterpna* and *Pingasa*. A second group of Pseudoterpnini *s.l.* was recovered as an independent lineage clearly separate from Pseudoterpnini *s.str.* (SH-like = 86.3, UFBoot2 = 96). Ban et al. (2018) did not introduce a new tribe due to the morphological similarities and difficulty in finding apomorphies of Pseudoterpnini *s.str.* In addition, their results were weakly supported. Considering that two independent studies have demonstrated the paraphyly of Pseudoterpnini *sensu* Pitkin et al (2007), we see no reason for retaining the wide concept of this tribe. Instead, we propose the revival of the tribe status of Archaeobalbinini and the description of a new tribe Chlorodontoperini. ¶

Archaeobalbinini Viidalepp, 1981, **status revised** ¶

(original spelling: Archeobalbinini, justified emendation in Hausmann (1996)) ¶

Type genus: *Archaeobalbis* Prout, 1912 (synonymized with *Herochroma* Swinhoe, 1893 in Holloway (1996)) ¶  
Material examined: *Herochroma curvata* Han & Xue, 2003, *H. baba* Swinhoe 1893, *Metallolopia inanularia* Han & Xue, 2004, *M. cuneataria* Han & Xue, 2004, *Actenochroma muscicoloraria* (Walker, 1862), *Absala dorcada* Swinhoe, 1893, *Metaterpna batangensis* Hang & Stüning, 2016, *M. thyatiraria* (Oberthür, 1913), *Limbatochlamys rosthorni* Rothschild, 1894, *Pachyodes pictaria* Moore, 1888, *Dindica para* Swinhoe, 1893, *Dindicodes crocina* (Butler, 1880), *Lophophelma erionoma* (Swinhoe, 1893), *L. varicoloraria* (Moore, 1868), *L. iterans* (Prout, 1926) and *Pachyodes amplificata* (Walker, 1862). ¶

This lineage splits into four groups: *Herochroma* Swinhoe, 1893 + *Absala* Swinhoe, 1893 + *Actenochroma* Warren, 1893 is the sister lineage of the rest of Archaeobalbinini that were recovered as a polytomic bunch of three clades comprising the genera *Limbatochlamys* Rothschild, 1894, *Psilotagna* Warren, 1894, *Metallolopia* Warren, 1895, *Metaterpna* Yazaki, 1992, *Dindica* Warren, 1893, *Dindicodes* Prout, 1912, *Lophophelma* Prout, 1912 and

**Deleted:** —The Australian genus *Crypsiphona* Meyrick, 1888 is sister to all tribes included in Geometrinae except Hemitheini. *Crypsiphona* has been assigned to Pseudoterpnini (e. g. Pitkin et al. 2007, Öunap & Viidalepp 2009), but is recovered as a separate lineage in our tree. Given the isolated position of *Crypsiphona*, the designation of a new tribe could be considered, but due to low support of branches in our analyses, further information (including morphology) is needed to confirm the phylogenetic position of this genus. *Xenozancla* Warren, 1893 is sister to the clade comprising Dysphaniini and Pseudoterpnini *s. str.* Sihvonen ¶

**Deleted:** and SH-Like = 100

**Formatted:** Right: 0.25"

1222 “boarmiine” and “ennomine” moths (Holloway, 1994). This grouping was proposed by Forbes  
1223 (1948) and Holloway (1994), who suggested close relationships between the tribes Boarmiini,  
1224 Macariini, Cassymini and Eutoeini based on the bifid pupal cremaster and the possession of a  
1225 fovea in the male forewing. The remaining tribes were defined as “ennomines” based on the loss  
1226 of a setal comb on male sternum A3 and the presence of a strong furca in male genitalia. Both  
1227 Wahlberg et al. (2010) and Sihvonen et al. (2011) found these two informal groupings to be  
1228 reciprocally monophyletic.

1229 In our analyses, 653 species with up to 11 markers were sampled, with an emphasis on  
1230 Neotropical taxa which so far had been poorly represented in the molecular phylogenetic  
1231 analyses. Our results recovered the division into two major subclades, a core set of ennomines in  
1232 a well-supported clade, and a poorly supported larger clade that includes the “boarmiines” among  
1233 four other lineages usually thought of as “ennomines”. The traditional “ennomines” are thus not  
1234 found to be monophyletic in our analyses, questioning the utility of such an informal name. Our  
1235 phylogenetic hypothesis supports the validation of numerous tribes earlier proposed, in addition  
1236 to several unnamed clades. We validate 23 tribes (Forum Herbulot, 2007; Skou & Sihvonen,  
1237 2015): Gonodontini, Gnophini, Odontoperini, Nacophorini, Ennomini, Campaeini, Alsophilini,  
1238 Wilemaniini, Prosoplophini, Diptychini, Theriini, Plutodini, Palyadini, Hypochrosini, Apeirini,  
1239 Epionini, Caberini, Macariini, Cassymini, Abraxini, Eutoeini and Boarmiini. We hereby propose  
1240 one new tribe: Drepanogynini **trib. nov.** (Table 1). Except for the new tribe, most of the groups  
1241 recovered in this study are in concordance with previous morphological classifications  
1242 (Holloway, 1993; Beljaev, 2006, 2016; Forum Herbulot, 2007; Skou & Sihvonen, 2015).

1243 Five known tribes and two further unnamed lineages (E1, E2 in figure 6) form the core  
1244 Ennominae: Gonodontini, Gnophini, Odontoperini, Nacophorini and Ennomini. Several  
1245 Neotropical clades that conflict with the current tribal classification of Ennominae will be  
1246 described as new tribes by Brehm et al (submitted). Gonodontini and Gnophini are recovered as  
1247 sister taxa. Gonodontini was defined by Forbes (1948) and studied by Holloway (1994), who  
1248 showed synapomorphies shared by *Gonodontis* Hübner, [1823], *Xylinophylla* Warren, 1898 and  
1249 *Xenimpia* Warren, 1895. Our results recovered the genus *Xylinophylla* as sister of *Xenimpia* and  
1250 *Psilocladia* Warren, 1898. *Psilocladia* is an African genus currently unassigned to tribe (see  
1251 Sihvonen et al., 2015 for details). Considering the strong support and that the facies and  
1252 morphology are somewhat similar to other analysed taxa in Gonodontini, we formally include

Formatted: Right: 0.25"

1253 *Psilocladia* in Gonodontini (Table 1). Gnophini are are monophyletic and we formally transfer  
1254 the African genera *Oedicentra* Warren, 1902 and *Hypotephrina* Janse, 1932, from unassigned to  
1255 Gnophini (Table 1). The total number of species, and number of included genera in Gnophini are  
1256 still uncertain (Skou & Sihvonen, 2015). Based on morphological examination, Beljaev (2007,  
1257 2016) treated Angeronini as a synonym of Gnophini. The costal projection on male valva bearing  
1258 a spine or group of spines was considered as a synapomorphy of the group. Using molecular data,  
1259 Yamamoto & Sota (2007) showed a close phylogenetic relationship between *Angerona*  
1260 Duponchel, 1829 (Angeronini) and *Chariaspilates* Wehrli, 1953 (Gnophini). Similar results were  
1261 shown by Sihvonen et al. (2011) who recovered *Angerona* and *Charissa* Curtis, 1826 as sister  
1262 taxa, and our results also strongly support treating Angeronini as synonym of Gnophini.

Deleted: a well-defined assemblage

1263 Holloway (1993) suggested close affinities among Nacophorini, Azelinini and  
1264 Odontoperini on the basis of larval characters. In a morphology-based phylogenetic study, Skou  
1265 & Sihvonen (2015) suggested multiple setae on the proleg on A6 of the larvae as a  
1266 synapomorphy of the group. Our results also support a close relationship of Nacophorini,  
1267 Azelinini and Odontoperini. These clades will be treated in more detail by Brehm et al.  
1268 (submitted).

1269 Following the ideas of Pitkin (2002), Beljaev (2008) synonymized the tribes  
1270 Ourapterygini and Nephodiini with Ennomini. He considered the divided vinculum in male  
1271 genitalia and the attachment of muscles *m*<sub>3</sub> as apomorphies of the Ennomini, but did not provide  
1272 a phylogenetic analysis. Sihvonen et al. (2011) supported Beljaev's assumptions and recovered  
1273 *Ennomos* Treitschke, 1825 (Ennomini), *Ourapteryx* Leach, 1814 (Ourapterygini) and *Nephodia*  
1274 Hübner, [1823] (Nephodiini) as belonging to the same clade. Our comprehensive analysis  
1275 confirms those previous findings and we agree with Ennomini as the valid tribal name for this  
1276 large clade.

Formatted: Indent: First line: 0.5"

1277 Campaeini, Alsophilini, Wilemaniini and Prosoplophini grouped together in a well-  
1278 supported clade (SH-like = 100 and UFBoot2 = 99). Previous molecular analyses have shown an  
1279 association of Colotoini [= Prosoplophini] and Wilemaniini (Yamamoto & Sota, 2007;  
1280 Sihvonen et al., 2011), although no synapomorphies are known to support synonymization (Skou  
1281 & Sihvonen, 2015). The Palaearctic genera *Compsoptera* Blanchard, 1845, *Apochima* Agassiz,

Deleted: ¶

Moved (insertion) [7]

Deleted: = 100

Formatted: Right: 0.25"

1285 1847, *Dasycorsa* Prout, 1915, *Chondrosoma* Anker, 1854 and *Dorsispina* Nupponen &  
1286 Sihvonen, 2013, are potentially part of the same complex (Skou & Sihvonen, 2015, Sihvonen  
1287 pers. obs.), but they were not included in the current study. Campaeini is a small group including  
1288 four genera with Oriental, Palaearctic and Nearctic distribution, apparently closely related to  
1289 Alsophilini and Prosoplophini, but currently accepted as a tribe (Forum Herbulot, 2007;  
1290 Sihvonen & Skou, 2015). Our results support the close phylogenetic affinities among these tribes,  
1291 but due to the limited number of sampled taxa, we do not propose any formal changes.

1292 The genus *Declana* Walker, 1858 is recovered as an isolated clade sister to **to Diptychini,**  
1293 **including *Acalyphes* and *Dirce*.** This genus is endemic to New Zealand, but to date has not been  
1294 assigned to tribe. According to our results, *Declana* could well be defined as its own tribe.  
1295 However, the delimitation of this tribe is beyond the scope of our paper and more genera from  
1296 Australia and New Zealand should first be examined.

1297 A close relationship between Nacophorini and Lithinini was suggested by Pitkin (2002),  
1298 based on the similar pair of processes of the anellus in the male genitalia. Pitkin also noted a  
1299 morphological similarity in the male genitalia (processes of the juxta) shared by Nacophorini and  
1300 Diptychini. In a study of the Australasian fauna, Young (2008) suggested the synonymization of  
1301 Nacophorini and Lithinini. This was further corroborated by Sihvonen et al. (2015) who found  
1302 that Diptychini were nested within some Nacophorini and Lithinini. However, none of the studies  
1303 proposed formal taxonomic changes because of limited taxon sampling. In contrast, samples in  
1304 our analyses cover all biogeographic regions and the results suggest that true Nacophorini is a  
1305 clade which comprises almost exclusively New World species. This clade is clearly separate from  
1306 Old World “nacophorines” (cf. Young, 2003) that are intermixed with Lithinini and Diptychini.  
1307 We here formally transfer Old World nacophorines to Diptychini and synonymize Lithinini **syn.**  
1308 **nov.** with Diptychini (Table 1). Further formal taxonomic changes in the Nacophorini complex  
1309 are provided by Brehm et al. (submitted).

1310 *Theria* Hübner, [1825], the only representative of Theriini in this study, clustered together  
1311 with *Lomographa* Hübner, [1825] (Baptini in Skou & Sihvonen, 2015), in a well-supported  
1312 clade, agreeing with the molecular results of Sihvonen et al. (2011). The placement of  
1313 *Lomographa* in Caberini (Rindge, 1979; Pitkin, 2002) is not supported by our study nor by that of  
1314 Sihvonen et al. (2011). The monophyly of *Lomographa* has not been tested before, but we show

Deleted: a complex lineage comprising Campaeini, Alsophilini, Wilemaniini and Prosoplophini

Formatted: Font: Italic

Moved up [7]: Campaeini, Alsophilini, Wilemaniini and Prosoplophini grouped together in a well-supported clade (SH-like and UFBoot2 = 100). Previous molecular analyses have shown an association of Colotoini [= Prosoplophini] and Wilemaniini (Yamamoto & Sota, 2007; Sihvonen et al., 2011), although no synapomorphies are known to support synonymization (Skou & Sihvonen, 2015). The Palaearctic genera *Compsoptera* Blanchard, 1845, *Apochima* Agassiz, 1847, *Dasycorsa* Prout, 1915, *Chondrosoma* Anker, 1854 and *Dorsispina* Nupponen & Sihvonen, 2013, are potentially part of the same complex (Skou & Sihvonen, 2015, Sihvonen pers. obs.), but they were not included in the current study. Campaeini is a small group including four genera with Oriental, Palaearctic and Nearctic distribution, apparently closely related to Alsophilini and Prosoplophini, but currently accepted as a tribe (Forum Herbulot, 2007; Sihvonen & Skou, 2015). Our results support the close phylogenetic affinities among these tribes, but due to the limited number of sampled taxa, we do not propose any formal changes.

Formatted: Font: Not Highlight

Formatted: Right: 0.25"

1337 that one Neotropical and one Palaearctic *Lomographa* species indeed group together. Our results  
1338 show that Caberini are not closely related to the Theriini + Baptini clade, unlike in earlier  
1339 morphology-based hypotheses (Rindge, 1979; Pitkin 2002). Morphologically, Theriini and  
1340 Baptini are dissimilar, therefore we recognize them as valid tribes (see description and  
1341 illustrations in Skou & Sihvonen, 2015).

1342 According to our results, 11 molecular markers were not enough to infer phylogenetic  
1343 affinities of Plutodini (represented by one species of *Plutodes*). Similar results were found by  
1344 Sihvonen et al. (2011), who in some analyses recovered *Plutodes* as sister of *Eumelea*. Our  
1345 analyses are congruent with those findings. IQ-TREE results suggest that *Plutodes* is sister to  
1346 Palyadini, but RAxML analyses recovered *Eumelea* as the most probable sister of *Plutodes*.  
1347 Given that our analyses are not in agreement about the sister-group affinities of *Plutodes*, we do  
1348 not make any assumptions about its phylogenetic position. Instead we emphasize that further  
1349 work needs to be done to clarify the phylogenetic positions of *Plutodes* and related groups.

1350 Hypochrosini is only recovered in a well-defined lineage if the genera *Apeira* Gistel, 1848  
1351 (*Apeirini*), *Epione* Duponchel, 1829 (*Epionini*), *Sericosema* (Caberini), *Ithysia* (Theriini),  
1352 *Capasa* Walker, 1866 (unassigned) and *Omisodes* Warren, 1894 (unassigned) were transferred to  
1353 Hypochrosini. Skou & Sihvonen (2015) already suggested a close association of Epionini,  
1354 Apeirini and Hypochrosini. We think that synonymising these tribes is desirable. However, due  
1355 to the limited number of sampled taxa we do not propose any formal changes until more data will  
1356 become available. We do suggest, however, formal taxonomic changes of the genera *Capasa* and  
1357 *Omisodes* from unassigned to Hypochrosini (Table 1).

1358 The southern African genus *Drepanogynis* is paraphyletic and has earlier been classified  
1359 as belonging in Ennomini, and later in Nacophorini (Krüger 2002). In our phylogeny, it is  
1360 intermixed with the genera *Sphingomima* Warren, 1899, and *Thenopa* Walker, 1855.  
1361 *Hebdomophruda errans* Prout, 1917 also clusters together with these taxa, apart from other  
1362 *Hebdomophruda* Warren, 1897 species, which suggests that this genus is polyphyletic. These  
1363 genera form a clade sister to the lineage that comprises several Hypochrosini species.  
1364 Considering that our analysis strongly supports this clade, we place *Thenopa*, *Sphingomima* and  
1365 *Drepanogynis* in a tribe of their own.

1366  
1367 Drepanogynini Murillo-Ramos, Sihvonen & Brehm **new tribe**

Deleted: would be

Formatted: Right: 0.25"

1369 [LSIDurn:lsid:zoobank.org:act:AA384988-009F-4175-B98C-6209C8868B93](https://lsid.zoobank.org/act:AA384988-009F-4175-B98C-6209C8868B93)

Formatted: Spanish

1370 Type genus: *Drepanogynis* Guenée, [1858]

1371

1372 The African genera *Thenopa*, *Sphingomima* and *Drepanogynis* appear as a strongly supported  
1373 lineage (SH-like UFB002, and RBS = 100). Krüger (1997, p. 259) proposed "Boarmiini and  
1374 related tribes as the most likely sister group" for *Drepanogynis*, whereas more recently  
1375 *Drepanogynis* was classified in the putative southern hemisphere Nacophorini (Krüger, 2014;  
1376 Sihvonen et al., 2015). In the current phylogeny, *Drepanogynis* is isolated from Nacophorini  
1377 *sensu stricto* and from other southern African genera that have earlier been considered to be  
1378 closely related to it (Krüger 2014 and references therein). The other southern African genera  
1379 appeared as belonging to Diptychini in our study. The systematic position of *Drepanogynis*  
1380 *tripartita* (Warren, 1898) has earlier been analysed in a molecular study (Sihvonen et al., 2015).  
1381 The taxon grouped together with the Palearctic species of the tribes Apeirini, Theriini, Epionini  
1382 and putative Hypochrosini. Sihvonen et al. (2015) noted that *Argyrophora trofonia* (Cramer,  
1383 [1779]) (representing *Drepanogynis* group III *sensu* Krüger, 1999) and *Drepanogynis tripartita*  
1384 (representing *Drepanogynis* group IV *sensu* Krüger, 2002) did not group together, but no formal  
1385 changes were proposed. Considering that the current analysis strongly supports the placement of  
1386 *Drepanogynis* and related genera in an independent lineage, and the aforementioned taxa in the  
1387 sister lineage (Apeirini, Theriini, Epionini and putative Hypochrosini) have been validated at  
1388 tribe-level, we place *Drepanogynis* and related genera in a tribe of their own.

Deleted: ed

Deleted: , SH-like and

Deleted: UFB002

1389 Material examined and taxa included: *Drepanogynis mixtaria* Guenée, [1858], *D.*  
1390 *tripartita*, *D. determinata* (Walker, 1860), *D. arcuifera* Prout, 1934, *D. arcuatilinea* Krüger,  
1391 2002, *D. cnephaeogramma* (Prout, 1938), *D. villaria* (Felder & Rogenhofer, 1875),  
1392 "*Sphingomima*" *discolucida* Herbulot, 1995 (genus combination uncertain, see taxonomic notes  
1393 below), *Thenopa diversa* Walker, 1855, "*Hebdomophruda*" *errans* Prout, 1917 (genus  
1394 combination uncertain, see taxonomic notes below).

1395 Taxonomic notes: We choose *Drepanogynis* Guenée, [1858] as the type genus for  
1396 Drepanogynini, although it is not the oldest valid name (ICZN Article 64), because extensive  
1397 literature has been published on *Drepanogynis* (Krüger 1997, 1998, 1999, 2014), but virtually  
1398 nothing exists on *Thenopa*, except the original descriptions of its constituent species. Current  
1399 results show the urgent need for more extensive phylogenetic studies within Drepanogynini.

Formatted: Right: 0.25"

1403 *Thenopa* and *Sphingomima* are embedded within *Drepanogynis*, rendering it paraphyletic, but  
 1404 our taxon coverage is too limited to propose formal changes in this species-rich group.  
 1405 Drepanogynini, as defined here, are distributed in sub-Saharan Africa. *Drepanogynis sensu*  
 1406 Krüger (1997, 1998, 1999, 2014) includes over 150 species and it ranges from southern Africa to  
 1407 Ethiopia (Krüger 2002, Vári et al. 2002), whereas the genera *Sphingomima* (10 species) and  
 1408 *Thenopa* (4 species) occur in Central and West Africa (Scoble 1999). *Sphingomima* and *Thenopa*  
 1409 are externally similar, so the recovered sister-group relationship in the current phylogeny analysis  
 1410 was anticipated. In the current analysis, *Hebdomophruda errans* Prout, 1917 is isolated from  
 1411 other analysed *Hebdomophruda* species (the others are included in Diptychini), highlighting the  
 1412 need for additional research. Krüger (1997, 1998) classified the genus *Hebdomophruda* into  
 1413 seven species groups on the basis of morphological characters, and *H. errans* group is one of  
 1414 them (Krüger 1998). We do not describe a new genus for the taxon *errans*, nor do we combine it  
 1415 with any genus in the Drepanogynini, highlighting its uncertain taxonomic position (*incertae*  
 1416 *sedis*) pending more research. In the current analysis, *Sphingomima discolorida* Herbulot, 1995 is  
 1417 transferred from unassigned tribus combination to Drepanogynini, but as the type species of  
 1418 *Sphingomima* (*S. heterodoxa* Warren, 1899) was not analysed, we do not transfer the entire genus  
 1419 *Sphingomima* into Drepanogynini. We highlight the uncertain taxonomic position of the taxon  
 1420 *discolorida*, acknowledging that it may eventually be included again in *Sphingomima* if the entire  
 1421 genus should get transferred to Drepanogynini.

1422  
 1423 Diagnosis: Drepanogynini can be diagnosed by the combination of DNA data with up to 11  
 1424 genetic markers (exemplar *Drepanogynis mixtaria* Guenée, [1858]) ArgK (MK738841), COI  
 1425 (MK739615), EF1a (MK739960), IDH (MK740862), MDH (MK741181), Nex9  
 1426 (MK741630), RpS5 (MK741991) and Wingless (MK742540). In the light of our  
 1427 phylogenetic results, the *Drepanogynis* group of genera, as classified earlier (Krüger 2014), is  
 1428 split between two unrelated tribes (Drepanogynini and Diptychini). More research is needed to  
 1429 understand how other *Drepanogynis* species and the *Drepanogynis* group of genera *sensu* Krüger  
 1430 (1997, 1998, 1999, 2014) (at least 11 genera), should be classified.

1431 Boarmiini are the sister group to a clade that comprises Macariini, Cassymini, Abraxini  
 1432 and Eutoeini. We found that many species currently assigned to Boarmiini are scattered  
 1433 throughout Ennominae. Boarmiini *s. str.* are strongly supported but are technically not

Deleted: GB Accession number), Ca-ATPase (GB Accession number)

Deleted: CAD (GB Accession number),

Deleted: GB Accession number),

Deleted: GB Accession number

Deleted: GAPDH (GB Accession number),

Deleted: GB Accession number

Deleted: GB Accession number

Deleted: GB Accession number

Deleted: GB Accession number

Deleted: GB Accession number

Deleted: ¶

Formatted: Pattern: Clear (White), Not Highlight

Formatted: Right: 0.25"



monophyletic because of a large number of genera which need to be formally transferred from other tribes to Boarmiini (see Brehm et al., submitted for Neotropical taxa and Murillo-Ramos et al., in prep. for other taxa). The results are principally in concordance with Jiang et al. (2017), who supported the monophyly of Boarmiini but with a smaller number of taxa.

The divided valva in male genitalia was suggested as a synapomorphy of Macariini + Cassymini + Eutoeini by Holloway (1994). In addition, he proposed the inclusion of Abraxini in Cassymini. *Although our findings support a close relationship, this group requires more study and a more extensive sampling effort.* Similar findings were provided by Jiang et al. (2017) who suggested more extensive sampling to study the evolutionary relationships of these tribes.

#### Orthostixinae Meyrick, 1892

Orthostixinae were not included in our study. Sihvonen et al. (2011) showed this subfamily as deeply embedded within Ennominae, but unfortunately it was not represented by the type genus of the tribe. These results agree with Holloway (1996) who examined *Orthostixis* Hübner, [1823] and suggested the inclusion in Ennominae despite the full development of hindwing vein M2, the presence of a forewing areole and the very broad base of the tympanal ansa. We sampled the species *Naxa textilis* (Preyer, 1884) and *Orthostixis cribraria* (Hübner, 1796) but, only three and one marker were successfully sequenced for these samples, respectively. We included these species in the preliminary analyses but results were so unstable that we excluded them from the final analysis. Further research including fresh material and more genetic markers are needed to investigate the position of Orthostixinae conclusively.

#### Conclusions

This study elucidated *important* evolutionary relationships *among* major groups within Geometridae. The monophyly of the subfamilies and the most widely accepted tribes were tested. We found strong support for the *traditional concepts of* Larentiinae, Geometrinae and Ennominae. Sterrhinae also becomes monophyletic when *Ergavia*, *Ametris* and *Macrotis*, currently placed in Oenochrominae, are transferred to Sterrhinae. The concepts of Oenochrominae and Desmobathrinae required major revision and, after appropriate rearrangements, these groups will also form monophyletic subfamily-level entities. Archiearinae

Deleted: Our

Deleted: Holloway's suggestions; Cassymini is recovered as polyphyletic and Abraxini and Eutoeini as sister taxa. Synonymization of Eutoeini and Cassymini with Abraxini should be considered in future studies, but the support values of the basal branches are too low in our hypothesis to draw final conclusions. Similar findings were

Deleted: some of the

Deleted: of

Deleted: the

Deleted: subfamilies

Deleted: in their traditional scopes

Deleted: formally

Formatted: Right: 0.25"



are monophyletic with the transfer of *Dirce* and *Acalyphes* to Ennominae. We treat Epidesmiinae as a new subfamily.

This study proposes the recognition of eight monophyletic geometrid subfamilies. Many geometrid tribes were recovered as para- or polyphyletic. We attempted to address the needed taxonomic changes, in order to favor taxonomic stability of the subfamilies and many tribes, even if in an interim way, to allow other researchers to use an updated higher-taxonomic structure that better reflects our current understanding of geometrid phylogeny. Further papers will be added to this work and will provide a large number of additional taxonomic changes in the Geometridae (see Introduction). Although we included a large number of new taxa in our study, many clades remain poorly represented. This is particularly true for taxa from tropical Africa and Asia. Tribes in special need of reassessment include Eumeleini, Plutodini, Eutoeini, Cassymini and Abraxini. Denser taxon sampling will ensure more robust phylogenetic conclusions towards a natural classification. We hope the phylogenetic hypotheses shared here will open new paths of inquiry across the Geometridae. Morphological synapomorphies have not yet been identified for many of the re- and newly defined higher taxa circumscribed by our 11-gene data set. Likewise, there is great need, across the family, to begin the work of mapping behavioral and life history attributes to the clades identified in this work.

#### Acknowledgements

Harri Sihvonen (Finland) is thanked for preparing extensive African materials for the study. We are grateful to Cathy Byrne (Hobart, Australia), B.C. Schmidt (Canada, Ottawa), Alfred Moser (Curitiba, Brazil), Rolf M\"ortter (Karlsruhe, Germany), Daniel Bolt (Domat /Ems, Switzerland), Florian Bodner (Vienna, Austria), Dominik Rabl (Vienna, Austria), Aare Lindt (Tallinn, Estonia), Luis Parra (Concepci\"on, Chile), Andreas Kopp (St. Margarethen, Switzerland), Stefan Naumann (Berlin, Germany), Jaan Viidalepp (Tartu, Estonia) for providing samples for this study. We thank John Chainey, Geoff Martin and Linda Pitkin at the NHM (London) for providing access to the collections and photographs of Neotropical Ennominae moths. We thank David Wagner, Andreas Zwick and Kevin Keegan for constructive comments.

#### References

Deleted: separated

Deleted: As a result, t

Deleted: a higher-level classification of

Deleted: Geometrid

Deleted: ae comprising eight monophyletic

Deleted: Moreover, we found that m

Formatted: Indent: First line: 0.5"

Deleted: in the different subfamilies were

Deleted: applied

Formatted: Strikethrough

Commented [WD14]: This is not a conclusion of this study and this point has already been made repeatedly.

Formatted: Strikethrough

Deleted: Despite our efforts t

Deleted: o

Deleted: very

Deleted: to be analyzed

Deleted: we acknowledge that

Deleted: are still

Deleted: strongly under

Deleted: ,

Deleted: and more detailed phylogenetic studies are required

Deleted: ing e.g. the tribes

Deleted: A-d

Formatted: Strikethrough

Deleted: in these regions

Commented [WD15]: This sentence already implied by the two previous sentences.

Formatted: Strikethrough

Formatted: None

Deleted: Tartu

Formatted: Right: 0.25"

- 1539 Abraham D, Ryrholm N, Wittzell H, Jeremy DH, Scoble MJ, Löfstedt C. 2001. Molecular  
1540 phylogeny of the subfamilies in Geometridae (Geometroidea: Lepidoptera). *Molecular*  
1541 *Phylogenetics and Evolution* 20: 65–77.
- 1542 Ban X, Jiang N, Cheng R, Xue D, Han H. 2018. Tribal classification and phylogeny of  
1543 Geometrinae (Lepidoptera: Geometridae) inferred from seven gene regions. *Zoological*  
1544 *Journal of the Linnean Society*, 184: 653–672 DOI: [10.1093/zoolinnean/zly013](https://doi.org/10.1093/zoolinnean/zly013).
- 1545 Beccaloni G Scoble M, Kitching I, Simonsen T, Robinson G, Pitkin B, Hine A, Lyal C.  
1546 2003. “Subfamily Oenochrominae” The Global Lepidoptera Names Index. Natural  
1547 History Museum, London. Available at <http://www.nhm.ac.uk/our-science/data/lepindex/>  
1548 (accessed 3 August 2018).
- 1549 Beljaev EA. 2006. A morphological approach to the Ennominae phylogeny (Lepidoptera,  
1550 Geometridae). *Spixiana* 29: 215–216.
- 1551 Beljaev EA. 2007. Tentative tribal system of Ennominae based on current family group names.  
1552 Personal communication (not peer-reviewed), available at  
1553 [http://www.herbulot.de/pdf/Family\\_group\\_names\\_in\\_Geometridae\\_alternative\\_12\\_06\\_20\\_07.pdf](http://www.herbulot.de/pdf/Family_group_names_in_Geometridae_alternative_12_06_20_07.pdf)  
1554 (accessed 3 August 2018).
- 1555 Beljaev EA. 2008. A new concept of the generic composition of the geometrid moth tribe  
1556 Ennomini (Lepidoptera, Geometridae) Based on Functional Morphology of the Male  
1557 Genitalia. *Entomological Review* 88: 50–60.
- 1558 Brehm G, Hebert PDN, Colwell RK Adams MO, Bodner F, Friedemann K, Möckel L, Fiedler K.  
1559 2016. Turning up the heat at a hotspot: DNA barcodes reveal 80% more species of  
1560 geometrid moths along an Andean elevational gradient. *PlosOne* 11: e0150327 DOI:  
1561 [10.1371/journal.pone.0150327](https://doi.org/10.1371/journal.pone.0150327)
- 1562 Cook MA, Scoble MJ. 1992. Tympanal organs of geometrid moths: a review of their  
1563 morphology, function, and systematic importance. *Systematic Entomology* 17: 219–232.
- 1564 Forbes WTM. 1948. Lepidoptera of New York and neighboring states. II. Memoirs of the Cornell  
1565 University Agricultural Experiment Station 274: 1–263.
- 1566 Forum Herbulot. 2007. World list of family-group names in Geometridae. Available at  
1567 <http://www.herbulot.de/famgroup.htm> (accessed 3 August 2018).
- 1568 Guindon S, Dufayard JF, Lefort V, Anisimova M, Hordijk W, Gascuel O. 2010. New  
1569 algorithms and methods to estimate maximum-likelihood phylogenies: assessing the  
1570 performance of PhyML 3.0. *Systematic Biology* 59: 307–321.
- 1571 Hausmann A. 2001. *Geometrid moths of Europe. Vol. 1: introduction to the series. Archiearinae,*  
1572 *Oenochrominae, Geometrinae*. Apollo books, Stenstrup.
- 1573 Hausmann A. 2004 *Geometrid moths of Europe. Vol. 2: Sterrhinae*. Apollo books, Stenstrup.

Formatted: Swedish

Formatted: Right: 0.25"

- 1574 Hausmann A, Hebert PDN, Mitchell A, Rougerie R, Sommerer M, Edwards T, Young K. 2009.  
1575 Revision of the Australian *Oenochroma vinaria* Guenée, 1858 species-complex  
1576 (Lepidoptera: Geometridae, Oenochrominae): DNA barcoding reveals cryptic diversity  
1577 and assesses status of type specimen without dissection. *Zootaxa* 2239: 1–21.
- 1578 Hausmann A, Viidalepp J. 2012. *Geometrid moths of Europe. Vol. 3: Larentiinae I*. Apollo  
1579 books, Stenstrup.
- 1580 Heikkilä M, Mutanen M, Wahlberg N, Sihvonen P, Kaila L. 2015. Elusive ditrysian phylogeny:  
1581 an account of combining systematized morphology with molecular data (Lepidoptera).  
1582 *BMC Evolutionary Biology* 15: 1–27 DOI: 10.1186/s12862-015-0520-0.
- 1583 Hoang DT, Chernomor O, Haeseler A, Minh BQ, Vinh LS. 2018. UFBoot2: Improving the  
1584 ultrafast bootstrap approximation. *Molecular Biology and Evolution* 35: 518–522 DOI:  
1585 10.1093/molbev/msx281.
- 1586 Holloway J. 1994. The moths of Borneo, part 11: Family Geometridae, subfamily Ennominae.  
1587 *Malayan Nature Journal* 47: 1–309.
- 1588 Holloway J. 1996. The moths of Borneo, part 9: Geometridae (incl. Orthostixini),  
1589 Oenochrominae, Desmobathrinae, Geometrinae, Ennominae. *Malayan Nature Journal* 49:  
1590 147–326.
- 1591 Holloway J. 1997. The moths of Borneo, part 10: family Geometridae, subfamilies Sterrhinae and  
1592 Larentiinae. *Malayan Nature Journal* 51: 1–242.
- 1593 Inoue H. 1961. Lepidoptera: Geometridae. *Insecta Japonica* 4: 1–106.
- 1594 International Commission on Zoological Nomenclature 2012: International Code of Zoological  
1595 Nomenclature, fourth edition. Available online <http://iczn.org/iczn/index.jsp>
- 1596 Jiang N, Li XX, Hausmann A, Cheng R, Xue DY, Han HX. 2017. A molecular phylogeny of the  
1597 Palaearctic and Oriental members of the tribe Boarmiini (Lepidoptera: Geometridae:  
1598 Ennominae). *Invertebrate Systematics* 31: 427–441.
- 1599 Kalyaanamoorthy S, Minh BQ, Wong TKF, Haeseler A von, Jermin LS. 2017. ModelFinder:  
1600 Fast Model Selection for Accurate Phylogenetic Estimates *Nature Methods* 14: 587–589  
1601 DOI: 10.1038/nmeth.4285.
- 1602 Krüger M. 1997. Revision of Afrotropical Ennominae of the *Drepanogynis* group I: the genus  
1603 *Hebdomophruda* Warren, Part 1. *Annals of the Transvaal Museum* 36: 257–291.  
1604
- 1605 Krüger M. 1998. Revision of Afrotropical Ennominae of the *Drepanogynis* group II: the genus  
1606 *Hebdomophruda* Warren, Part 2. *Annals of the Transvaal Museum* 36: 333–349.  
1607
- 1608 Krüger M. 1999. Revision of Afrotropical Ennominae of the *Drepanogynis* group III: the genera  
1609 *Argyrophora* Guenée, *Pseudomaenas* Prout and *Microligia* Warren. *Annals of the*  
1610 *Transvaal Museum* 36: 427–496.

Formatted: Right: 0.25"

- 1611  
1612 Krüger M. 2002. Revision of Afrotropical Ennominae of the *Drepanogynis* group IV: the genus  
1613 *Drepanogynis* Guenée (Lepidoptera: Geometridae). *Transvaal Museum Monograph* 13:  
1614 1–220 incl. 442 figs.  
1615
- 1616 Krüger M. 2014: A revision of the *Mauna* Walker, 1865 and *Illa* Warren, 1914 group of genera  
1617 (Lepidoptera: Geometridae: Ennominae: Nacophorini). *Annals of the Ditsong National*  
1618 *Museum of Natural History* 4: 77–173.
- 1619 Lanfear R , Calcott B, Ho SYW, Guindon S. 2012. Partitionfinder: Combined selection of  
1620 partitioning schemes and substitution models for phylogenetic analyses. *Molecular*  
1621 *Biology and Evolution* 29: 1695–1701 DOI:10.1093/molbev/mss020 pmid:22319168.
- 1622 Meyrick E. 1889. Revision of Australian Lepidoptera. *Proceedings of the Linnean Society of New*  
1623 *South Wales* 41: 117–1216.
- 1624 Miller MA, Pfeiffer W, Schwartz T. 2010. Creating the CIPRES science gateway for inference of  
1625 large phylogenetic trees. In Proceedings of the Gateway Computing Environments  
1626 Workshop (GCE), New Orleans, LA pp 1 - 8. Available at <http://www.phylo.org>.  
1627 (accessed June, 2018).
- 1628 Minet J, Scoble MJ. 1999. The Drepanoid/Geometroid assemblage. In: Kristensen NP, ed.  
1629 *Handbook of Zoology, part 35, Lepidoptera, Moths and Butterflies, Vol. 1, Evolution,*  
1630 *Systematics, and Biogeography*: De Gruyter, Berlin, 301–320.
- 1631 Mironov V. 2003. Larentiinae II (Perizomini and Eupitheciini). In: Hausmann A, ed. *The*  
1632 *Geometrid Moths of Europe* 4. Apollo Books, Stenstrup, 1–463.
- 1633 Nguyen LT, Schmidt HA, Haeseler A von, Minh BQ. 2015. IQ- TREE: A fast and effective  
1634 stochastic algorithm for estimating maximum likelihood phylogenies. *Molecular Biology*  
1635 *and Evolution* 32: 268–274 DOI: 10.1093/molbev/msu300.
- 1636 Nieuwerkerken EJ van, Kaila L, Kitching IJ, Kristensen NP, Lees DC, Minet J, Mitter C, Mutanen  
1637 M, Regier JC, Simonsen TJ, Wahlberg N, Yen S, Zahir R, Adamski D, Baixeras J,  
1638 Bartsch D, Bengtsson BÅ, Brown JW, Bucheli SR, Davis DR, Prins J de, Prins W de,  
1639 Epstein ME, Gentili-Poole P, Gielis C, Hättenschwiler P, Hausmann A, Holloway JD,  
1640 Kallies A, Karsholt O, Kawahara AY, Koster J, Kozlov M, Lafontaine JD, Lamas G,  
1641 Landry J, Lee S, Nuss M, Park K, Penz C, Rota J, Schintlmeister A, Schmidt BC, Sohn J,  
1642 Solis MA, Tarmann GM, Warren AD, Weller S, Yakovlev RV, Zolotuhin VV, Zwick A.  
1643 2011. Order Lepidoptera Linnaeus, 1758. In: Zhang, Z.-Q. (Ed.), *Animal biodiversity: An*  
1644 *outline of higher-level classification and survey of taxonomic richness. Zootaxa* 3148:  
1645 212–221.
- 1646 Õunap E, Viidalepp J, Saarma U. 2008. Systematic position of Lythriini revised: transferred from  
1647 Larentiinae to Sterrhinae (Lepidoptera, Geometridae). *Zoologica Scripta* 37: 405–413.

Formatted: Right: 0.25"

- 1648 Õunap E, Viidalepp J. 2009. Description of *Crypsiphona tasmanica* sp. nov. (Lepidoptera:  
1649 Geometridae: Geometrinae), with notes on limitations in using DNA barcodes for  
1650 delimiting species. *Australian Journal of Entomology* 48: 113–124.
- 1651 Õunap E, Javoiš J, Viidalepp J, Tammaru T. 2011. Phylogenetic relationships of selected  
1652 European Ennominae (Lepidoptera: Geometridae). *European Journal of Entomology* 108:  
1653 267–273.
- 1654 Õunap E, Viidalepp J, Truuverk A. 2016. Phylogeny of the subfamily Larentiinae (Lepidoptera:  
1655 Geometridae): integrating molecular data and traditional classifications. *Systematic  
1656 Entomology* 21: 824–843 DOI:10.1111/syen.12195.
- 1657 Peña C, Malm T. 2012. VoSeq: a voucher and DNA sequence web application. *PLoS ONE* 7:  
1658 e39071.
- 1659 Pitkin L. 1996. Neotropical emerald moths: a review of the genera (Lepidoptera: Geometridae,  
1660 Geometrinae). *Zoological Journal of the Linnean Society* 118: 309–440.
- 1661 Pitkin L. 2002. Neotropical Ennomine moths: a review of the genera (Lepidoptera: Geometridae).  
1662 *Zoological Journal of the Linnean Society* 135: 121–401.
- 1663 Pitkin B, Jenkins P. 2004. Butterflies and moths of the world, generic names and their type-  
1664 species. Available at <http://www.nhm.ac.uk/our-science/data/butmoth/> (accessed 29  
1665 August, 2018).
- 1666 Pitkin L, Han H, James S. 2007. Moths of the tribe Pseudoterpnini (Geometridae: Geometrinae):  
1667 a review of the genera. *Zoological Journal of the Linnean Society* 150:343–412.
- 1668 Rajaei H, Greve C, Letsch H, Stüning D, Wahlberg N, Minet J, Misof B. 2015. Advances in  
1669 Geometroidea phylogeny, with characterization of a new family based on *Pseudobiston  
1670 pinratanaei* (Lepidoptera, Glossata). *Zoologica Scripta* 44: 418–436.  
1671 DOI:10.1111/zsc.12108.
- 1672 Rambaut A. 2012. Figtree 1.4.0. Available at <http://tree.bio.ed.ac.uk/software/figtree/> (accessed  
1673 3 August 2018).
- 1674 Ratnasingham S, Hebert PD. 2007. BOLD: the barcode of life data systems. *Molecular Ecology  
1675 Notes* 7: 355–364 DOI:10.1111/j.1471-8286.2007.01678.x.
- 1676 Regier JC, Zwick A, Cummings MP, Kawahara AY, Cho S, Weller S, Roe A, Baixeras J, Brown  
1677 JW, Parr C, Davis DR, Epstein M, Hallwachs W, Hausmann A, Janzen DH, Kitching IJ,  
1678 Solis MA, Yen SH, Bazinet AL, Mitter C. 2009. Toward reconstructing the evolution of  
1679 advanced moths and butterflies (Lepidoptera: Ditrysia): an initial molecular study. *BMC  
1680 Evolutionary Biology* 9: 280 DOI:10.1186/1471-2148-9-280.
- 1681 Regier JC, Mitter C, Zwick A, Bazinet AL, Cummings MP, Kawahara AY, Sohn JC, Zwickl  
1682 DJ, Cho S, Davis DR, Baixeras J, Brown J., Parr C, Weller S, Lees DC, Mitter

Formatted: Finnish

Field Code Changed

Formatted: Right: 0.25"

- 1683 KT. 2013. A large-scale, higher-level, molecular phylogenetic study of the insect order  
1684 Lepidoptera (moths and butterflies). *PLoS ONE* 8: e58568.
- 1685 Rindge FH. 1979. A revision of the North American moths of the  
1686 genus *Lomographa* (Lepidoptera, Geometridae). *American Museum Novitates* 2673: 1–  
1687 18.
- 1688 Scoble MJ, Edwards ED. 1990. *Parepisparis* Bethune-Baker and the composition of the  
1689 Oenochrominae (Lepidoptera: Geometridae). *Entomologica Scandinavica* 20: 371–399.
- 1690 Scoble MJ. 1992. *Lepidoptera: Form Function and Diversity*. Oxford: Oxford University Press.
- 1691 Scoble, M.J. 1999. Geometrid Moths of the World: A catalogue (Lepidoptera, Geometridae). vols.  
1692 1 & 2. Collingwood, CSIRO.
- 1693 Sihvonen P, Kaila L. 2004. Phylogeny and tribal classification of Sterrhinae with emphasis on  
1694 delimiting Scopulini (Lepidoptera: Geometridae). *Systematic Entomology* 29: 324–358.
- 1695 Sihvonen P, Mutanen M, Kaila L, Brehm G, Hausmann A, Staude HS. 2011. Comprehensive  
1696 molecular sampling yields a robust phylogeny for geometrid moths (Lepidoptera:  
1697 Geometridae). *PLoS ONE* 6: e20356 DOI: doi:10.1371/journal.pone.0020356.
- 1698 Sihvonen P, Staude H. 2011. Geometrid moth *Afrophylla vethi* (Snellen, 1886) transferred from  
1699 Oenochrominae to Sterrhinae (Lepidoptera: Geometridae). *Metamorphosis* 22: 102–113.
- 1700 Sihvonen P, Staude HS, Mutanen M. 2015. Systematic position of the enigmatic African cycad  
1701 moths: an integrative approach to a nearly century old problem (Lepidoptera:  
1702 Geometridae, Diptychini). *Systematic Entomology* 40: 606–627.
- 1703 Skou P, Sihvonen P. 2015. *The Geometrid Moths Of Europe. Vol. 5: Ennominae I*. Apollo books,  
1704 Stenstrup.
- 1705 Stamatakis A, Hoover P, Rougemont J. 2008. A rapid bootstrap algorithm for the RAxML Web  
1706 servers. *Systematic Biology* 57: 758–771.
- 1707 Stamatakis, A. (2014) RAxML Version 8: A tool for Phylogenetic Analysis and Post-Analysis of  
1708 Large Phylogenies. *Bioinformatics* 30 (9):13121313 DOI: [10.1093/bioinformatics/btu033](https://doi.org/10.1093/bioinformatics/btu033).
- 1709 Staude HS. 2001. A revision of the genus *Callioratis* Felder (Lepidoptera: Geometridae:  
1710 Diptychinae). *Metamorphosis* 12: 125–156.
- 1711 Staude H, Sihvonen P. 2014. Revision of the African geometrid genus *Zerenopsis* C. & R.  
1712 Felder-moths with peculiar life histories and mating behaviors (Geometridae: Ennominae:  
1713 Diptychini). *Metamorphosis* 25: 11–55.
- 1714 Strutzenberger P, Brehm G, Gottsberger B, Bodner F, Seifert, CL, Fiedler, K. 2017.  
1715 Diversification rates, host plant shifts and an updated molecular phylogeny of Andean

Formatted: Swedish

Formatted: Right: 0.25"

- 1716 *Eois* moths (Lepidoptera: Geometridae). PlosOne 12: [e018843](https://doi.org/10.1371/journal.pone.018843)  
1717
- 1718 Trifinopoulos J, Minh, B. 2018. IQ-TREE Manual: Frequently Asked Questions. Available at  
1719 <http://www.iqtree.org/doc/Frequently-Asked-Questions> (accessed 13 August 2018).
- 1720 Vári L, Kroon DM, Krüger M. 2002. *Classification and Checklist of the Species of Lepidoptera*  
1721 *Recorded in Southern Africa*. Australia: Simple Solutions, Chatswood.
- 1722  
1723 Viidalepp J. 2006. Cladistic analysis of the subfamily Larentiinae. In: Hausmann A, McQuillan  
1724 P, eds. Proceedings of the Forum Herbulot 2006. Integration of molecular, ecological and  
1725 morphological data: Recent progress towards the higher classification of the Geometridae  
1726 (Hobart, 19–20 January 2006). *Spixiana* 29: 202–203.
- 1727 Viidalepp J. 2011. A morphological review of tribes in Larentiinae (Lepidoptera: Geometridae).  
1728 *Zootaxa* 3136: 1–44.
- 1729 Yamamoto S, Sota T. 2007. Phylogeny of the Geometridae and the evolution of winter moths  
1730 inferred from a simultaneous analysis of mitochondrial and nuclear genes. *Molecular*  
1731 *Phylogenetics and Evolution* 44: 711–723.
- 1732 Young CJ. 2003. The Place of the Australian Nacophorini in the Geometridae. *Spixiana* 26: 199-  
1733 200.
- 1734 Young CJ. 2006. Molecular relationships of the Australian Ennominae (Lepidoptera:  
1735 Geometridae) and implications for the phylogeny of the Geometridae from molecular and  
1736 morphological data. *Zootaxa* 1264: 1–147.
- 1737 Young CJ. 2008. Characterization of the Australian Nacophorini using adult morphology, and  
1738 phylogeny of the Geometridae based on morphological characters. *Zootaxa* 1736: 1–141.
- 1739 Wahlberg N, Wheat CW. 2008. Genomic outposts serve the phylogenomic pioneers: designing  
1740 novel nuclear markers for genomic DNA extractions of Lepidoptera. *Systematic Biology*  
1741 57: 231–242.
- 1742 Wahlberg N, Snäll N, Viidalepp J, Ruohomäki K, Tammaru T. 2010. The evolution of female  
1743 flightlessness among Ennominae of the Holarctic forest zone (Lepidoptera, Geometridae).  
1744 *Molecular Phylogenetics and Evolution* 55: 929–938.
- 1745 Wahlberg N., Peña, C., Ahola, M., Wheat C.W., Rota J. 2016. PCR primers for 30 novel gene  
1746 regions in the nuclear genomes of Lepidoptera. *ZooKeys* 596: 129–141.  
1747 <https://doi.org/10.3897/zookeys.596.8399>

Page 12: [1] Deleted Leidys Murillo 4/11/19 4:13:00 PM

▼

Page 12: [2] Deleted Leidys Murillo 4/11/19 4:37:00 PM

▼

Page 12: [3] Deleted Leidys Murillo 4/11/19 4:47:00 PM

▼

Page 12: [4] Commented [WD9] Wagner, David 5/22/19 3:49:00 AM

How supported? Weakly? Strongly? Or just say ,” “formed a clade,” if the support is of average strength (deleting the word supported).

Page 12: [5] Deleted Leidys Murillo 4/12/19 8:49:00 AM

▼

Page 12: [6] Commented [WD10] Wagner, David 5/22/19 3:52:00 AM

Elements of a series that contain commas should be separated with semi-colons.

Page 25: [7] Deleted Leidys Murillo 4/12/19 10:43:00 AM

▼