

1Manuscript Title

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3A comprehensive molecular phylogeny of Geometridae (Lepidoptera) with a focus on enigmatic
4small subfamilies

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32Abstract

33Our study aims to investigate the relationships of the major lineages within the moth family
34Geometridae, with a focus on the poorly studied Oenochrominae-Desmobathrinae complex, and
35to translate some [of](#) the results into a coherent subfamily and tribal level classification for the
36family. We analyzed a molecular dataset of 1206 Geometridae terminal taxa from all
37biogeographical regions comprising up to 11 molecular markers that included one mitochondrial
38(COI) and 10 protein-coding nuclear gene regions (Wingless, ArgK, MDH, RpS5, GAPDH, IDH,
39Ca-ATPase, Nex9, EF-1alpha, CAD). The molecular data set was analyzed using maximum
40likelihood as implemented in IQ-TREE and RAxML. We found high support for the traditional
41subfamilies Larentiinae, Geometrinae and Ennominae in their traditional scopes. Sterrhinae is
42monophyletic only if *Ergavia*, *Ametris* and *Macrotres*, which are currently placed in
43Oenochrominae, are formally transferred to Sterrhinae. Desmobathrinae and Oenochrominae are
44found to be polyphyletic. The concepts of Oenochrominae and Desmobathrinae required major
45revision and, after appropriate rearrangements, these groups also form monophyletic subfamily-
46level entities. Oenochrominae *s.str.* as originally conceived by Guenée is phylogenetically distant
47from *Epidesmia*. The latter is hereby described as the subfamily Epidesmiinae Murillo-Ramos,
48Sihvonen & Brehm, **subfam. nov.** Epidesmiinae are a lineage of “slender bodied
49Oenochrominae” that include the genera *Ecphyas* Turner, *Systatica* Turner, *Adeixis* Warren,
50*Dichromodes* Guenée, *Phrixocomes* Turner, *Abraxaphantes* Warren, *Epidesmia* Duncan [&
51Westwood] and *Phrataria* Walker. Archiearinae are monophyletic [whenif](#) *Dirce* and *Acalyphes*
52are formally transferred to Ennominae. We also found that many tribes were para- or polyphyletic
53and therefore propose tens of taxonomic changes at the tribe and subfamily levels.
54Archaeobalbini Viidalepp (Geometrinae) is raised from synonymy of Pseudoterpnini Warren to
55[the-tribetribal](#) rank. Chlorodontoperini Murillo-Ramos, Sihvonen & Brehm, **trib. nov.** and
56Drepanogynini Murillo-Ramos, Sihvonen & Brehm, **trib. nov.** are described as new tribes in
57Geometrinae and Ennominae, respectively.

58

59**Keywords:** Phylogeny, new subfamily, moths, Epidesmiinae, taxonomy.

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63 Introduction

64 Geometridae are the second most species-rich family of Lepidoptera, with approximately 24,000
65 described species (Nieukerken et al., 2011, updated  found in all regions except Antarctica. The
66 monophyly of Geometridae is well supported based on distinctive morphological characters
67 (Cook & Scoble, 1992; Scoble, 1992; Minet & Scoble, 1999). In particular, adult members of the
68 family possess paired tympanal organs at the base of the abdomen, while in ~~the~~ larvae, the ventral
69 prolegs are reduced to two pairs in almost all species, which causes the larvae to move in a
70 looping manner (Minet & Scoble, 1999).

71 The phylogenetic relationships of the major subdivisions of Geometridae have been
72 studied based on molecular data, which have contributed to the understanding of the evolutionary
73 relationships within the family (Abraham et al., 2001; Yamamoto & Sota, 2007; Sihvonen et al.,
74 2011). At ~~the~~ present, eight subfamilies are recognized in Geometridae (Sihvonen et al., 2011).
75 Several recent studies have attempted to confirm the monophyly or clarify the taxonomy of most
76 of these groups, for instance: Sterrhinae (Holloway, 1997; Hausmann, 2004; Sihvonen & Kaila,
77 2004; Öunap et al., 2008), Larentiinae (Holloway, 1997; Mironov, 2003; Viidalepp, 2006, 2011;
78 Hausmann & Viidalepp, 2012; Öunap et al., 2016), Desmobathrinae (Holloway, 1996;
79 Hausmann, 2001), Archiearinae (Hausmann, 2001; Young, 2006), Oenochrominae (Holloway,
80 1996; Scoble & Edwards, 1990; Cook & Scoble, 1992; Hausmann, 2001; Young, 2006),
81 Geometrinae (Cook, 1993; Pitkin, 1996; Hausmann, 2001; Ban et al., 2018), Orthostixinae
82 (Holloway, 1997) and Ennominae (Holloway, 1994; Pitkin, 2002; Beljaev, 2006; Young, 2006;
83 Wahlberg et al., 2010; Öunap et al., 2011; Skou & Sihvonen, 2015; Sihvonen et al., 2015). An
84 important shortcoming is that our understanding of geometrid systematics is biased towards the
85 long-~~studied~~ European fauna, whereas the highest diversity of this family is in the tropics, which
86 is still largely unexplored (Brehm et al., 2016). Many species remain undescribed and there are
87 many uncertainties in tribe and genus level classifications.

88 One of the most ~~complete~~comprehensive phylogenetic studies on Geometridae to date
89 was published by Sihvonen et al. (2011). They analyzed a data set of 164 taxa and eight genetic
90 markers, and the most species-rich subfamilies were recovered as monophyletic. However, the
91 systematic positions of Oenochrominae and Desmobathrinae remained uncertain due to low taxon
92 sampling, and the groups were suggested to be polyphyletic. Moreover, many geometrid genera
93 remained unassigned to tribe .

94 This study is the first in a series of papers, which investigate the phylogenetic
95 relationships of Geometridae on the basis of a sample with global coverage. Our dataset
96 comprises 1206 terminal taxa of Geometridae with samples from all major biomes, using up to 11
97 molecular markers. Our paper includes an overview of the relationships of the major lineages
98 within the family, with **particular focus** on defining the limits and finding the phylogenetic
99 affinities of the subfamilies, **with a focus** on Oenochrominae and Desmobathrinae. Further
100 papers in the series will focus on particular subfamilies and regions, and they will propose further
101 formal taxonomic changes beyond those suggested in the present article: tribe and genus level
102 relationships in Sterrhinae (Sihvonen et al., in prep), New World taxa (Brehm et al., in prep),
103 Larentiinae (Öunap et al., in prep) and the Ennominae. The Boarmiini (Murillo-Ramos et al., in
104 prep).

105 A close relationship of Oenochrominae and Desmobathrinae has been proposed both in
106 morphological (Meyrick, 1889; Cook & Scoble, 1992; Holloway, 1996) and in molecular studies
107 (Sihvonen et al., 2011; Ban et al., 2018). In the first early classifications, species of
108 Desmobathrinae and Oenochrominae were included in the former family Monoctenidae. Meyrick
109 (1889) diagnosed them on the basis of the position of the Rs veins in the hindwing veins and vein
110 Sc+R1 in the forewing, which approximate to the upper margin of the cell from near its base to
111 its middle-cell or beyond (Scoble & Edwards, 1990). However, the classification proposed by
112 Meyrick was not fully supported by subsequent taxonomic work (Scoble & Edwards, 1990; Cook
113 & Scoble, 1992; Holloway, 1996). Unfortunately, Oenochrominae became a “trash bin”
114 of geometrids that could not be placed in other subfamilies, including even Lyalidae, a family of
115 moth-like butterflies (Scoble, 1992). Unsurprisingly, many taxa traditionally classified in
116 Oenochrominae have recently been shown to be misplaced (Holloway, 1997; Staude, 2001;
117 Sihvonen & Staude, 2011; Staude & Sihvonen, 2014). In Scoble & Edwards (1990), the family
118 concept of Oenochrominae was restricted to the robust-bodied Australian genera, with one
119 representative from the Oriental region. These authors were not able to find synapomorphies to
120 define Monoctenidae *sensu* Meyrick, and referred back to the original grouping proposed by
121 Guenée (1858). Scoble & Edwards (1990) defined a narrower group for Oenochrominae based on
122 the male genitalia: The sclerotisation of the diaphragm dorsal to the anellus fuses with the
123 transtilla to form a rigid plate. Cook & Scoble (1992) suggested that the circular form of the

124 lacinia and its orientation parallel to the tympanum was apomorphic for these robust-
125 bodied Oenochrominae.

126 In an extensive morphological study, Holloway (1996) revived  subfamily
127 Desmobathrinae to include species with appendages and slender bodies previously assigned to
128 Oenochrominae. According to Holloway (1996), Desmobathrinae comprises two tribes:
129 Eumeleini and Desmobathrini. However, no synapomorphies were found to link Eumeleini and
130 Desmobathrini. Holloway (1996) highlighted that the modification of the tegumen of the male
131 genitalia is was variable in both groups but that the reduction of cremastral spines in the pupa
132 from eight to four in *Ozola* Walker, 1861 and *Eumelea* Duncan [& Westwood], 1841 provided
133 evidence of a closer relationship between Eumeleini and Desmobathrini. The proposed
134 classification is included in the “World list of family group names in Geometridae” (Forum
135 Herbulot, 2007). Currently, 328 species (76 genera) are included in Oenochrominae, and 248
136 species (19 genera) are assigned to Desmobathrinae (Beccaloni et al., 2003; Sihvonen et al.,
137 2011, 2015).

138 Most recent molecular phylogenies have shown Oenochrominae and Desmobathrinae taxa 
139 to be intermingled (Sihvonen et al., 2011; Ban et al., 2018), but taxon sampling was limited to
140 eight and four species, respectively. The poor taxon sampling and the obviously unresolved
141 relationships around the Oenochrominae and Desmobathrinae complex called for a sound
142 phylogenetic study that clarifies the relationships of these poorly known taxa within
143 Geometridae. We hypothesize that both Oenochrominae and Desmobathrinae are para- or
144 polyphyletic assemblages, and our paper aims to establish a new concept in which all subfamilies
145 of the Geometridae represent monophyletic entities. Our new study comprises 29 terminal taxa of
146 Oenochrominae and 11 representatives of Desmobathrinae. Most species are distributed in the
147 Australian and Oriental Region, but some also occur in other parts of the world.

148

149 **Materials & Methods**

150 The electronic version of this article in Portable Document Format (PDF) will represent a
151 published work according to the International Commission on Zoological Nomenclature (ICZN),
152 and hence the new names contained in the electronic version are effectively published under that
153 Code from the electronic edition alone. This published work and the nomenclatural acts it
154 contains have been registered in ZooBank,  online registration system for the ICZN. The

ZooBank LSIDs (Life Science Identifiers) can be resolved and the associated information viewed through any standard web browser by appending the LSID to the prefix <http://zoobank.org/>. The LSID for this publication is: Epidesmiinae subfam.nov.

LSIDurn:lsid:zoobank.org:act:34D1E8F7-99F1-4914-8E12-0110459C2040, Chlorodontoperini trib.nov. LSIDurn:lsid:zoobank.org:act:0833860E-A092-43D6-B2A1-FB57D9F7988D, and Drepanogynini trib.nov., LSIDurn:lsid:zoobank.org:act:AA384988-009F-4175-B98C-6209C8868B93. The online version of this work is archived and available from the following digital repositories: PeerJ, PubMed Central and CLOCKSS.

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Material acquisition, taxon sampling and species identification

In addition to 461 terminal taxa with published sequences (see Supplemental data S1), we included sequences from 745 new terminal taxa in our study. They were gathered from several museum collections and collectors, including most of the authors (Supplemental data S1). Representative taxa of all subfamilies recognized in Geometridae were included, except for the small subfamily Orthostixinae for which most molecular markers could not successfully amplified. A total of 93 tribes are represented in this study following recent phylogenetic hypotheses and classifications (Sihvonen et al., 2011; Wahlberg et al., 2010; Sihvonen et al., 2015; Öunap et al., 2016; Ban et al., 2018). In addition, 14 non-Geometridae species belonging to other families of Geometroidea were included as outgroups based on the hypothesis proposed by Regier et al. (2009; 2013). Where possible, two or more samples were included per tribe and genus, especially for species-rich groups that are widely distributed and in cases where genera were suspected to be poly- or paraphyletic. We preferred type species or species phylogenetically close to type species in order to ease subsequent taxonomic work, favor nomenclatorial stability and to establish the phylogenetic position of genera unassigned to tribes.

Sampled individuals were identified by the authors using their complementary expertise and appropriate literature, and by comparing type material from different collections and museums. Moreover, we compiled an illustrated catalogue of all Archiearinae, Desmobathrinae and Oenochrominae taxa included in this study, to display the external diversity to allow facilitate subsequent verification of our identifications. This catalogue contains images of all analysed specimens as well as photographs of the respective type material (Supplemental data S2). Many further specimens will be illustrated in other papers (Brehm et al. in prep., Sihvonen et

186al. in prep., Öunap et al. in prep.) Some of the studied individual specimens could not yet be
187assigned to species, and their identifications are preliminary because of a lack of modern
188identification tools, particularly for (potentially undescribed) tropical species. Taxonomic data,
189voucher ID, number of genes, current systematic placement, and references to relevant literature
190where the tribal association is used, are shown in Supplemental data S1.

191

192*Molecular techniques*

193

194DNA was extracted from 1–3 legs preserved either in ethanol or dry. In a few cases, other sources
195of tissue, such as parts of larvae, were used. The remaining parts of specimens were preserved as
196vouchers and will ~~be~~ eventually be deposited in public museum collections. Genomic DNA was
197extracted and purified using a NucleoSpin® Tissue Kit (MACHERY-NAGEL),

198~~according to~~ the manufacturer's protocol. DNA amplification and sequencing were
199carried out following protocols proposed by Wahlberg & Wheat (2008) and Wahlberg et al.

200(2016). PCR products were visualized on agarose gels. PCR products were cleaned enzymatically
201and sent to Macrogen Europe (Amsterdam) for Sanger sequencing. One mitochondrial (COI) and
20210 protein-coding nuclear gene regions (Wingless, ArgK, MDH, RpS5, GAPDH, IDH, Ca-

203ATPase, Nex9, EF-1alpha, CAD) were sequenced. The final dataset had a concatenated length of
2047665 bp with gaps. ~~We~~ check for potential misidentifications, DNA barcode sequences were
205compared to those in BOLD (Barcode of Life Data Systems,

206(<http://www.barcodinglife.org/views/login.php>) where references of more than 21,000 geometrid
207species are available, some 10,000 of them being reliably identified to Linnean species names

208(Ratnasingham & Hebert, 2007). GenBank accession numbers for sequences used in this study
209are provided in Supplemental data S1.

210

211*Alignment and cleaning sequences*

212

213Multiple sequence alignments were ~~done~~ carried out in MAFFT as implemented in Geneious

214v.11.0.2 (Biomatters, <http://www.geneious.com/>) for each gene based on a reference sequence of
215Geometridae downloaded from the database VoSeq (Peña & Malm, 2012). ~~We used MAFFT~~

216algorithm as implemented in Geneious v.11.0.2 (Biomatters, <http://www.geneious.com/>). The

alignments ~~per of each~~ gene ~~were as~~ carefully checked by eye ~~relative to the reference sequence~~,
taking into ~~consideration relevant~~ ~~account the respective~~ genetic codes and reading frames,
~~relative to the reference sequence~~. Heterozygous positions were coded with IUPAC
codes. Sequences with bad quality and ambiguities ~~were~~ removed from the alignments. Finally,
aligned sequences were uploaded to VoSeq (Peña & Malm, 2012) and then assembled in a
dataset comprising 1206 taxa. To check for possible errors in alignments ~~and~~, potentially
contaminated ~~or identical~~ sequences ~~and misidentifications~~, we constructed maximum likelihood
trees for each gene. ~~With these trials, we also looked for identical sequences or~~
~~misidentifications~~. These ~~trial~~ ~~preliminary~~ analyses were conducted using RAXML-HPC2
V.8.2.10 (Stamatakis, 2014) on the web-server CIPRES Science Gateway (Miller et al., 2010).
After cleaning, ~~the~~ final data set included at least three genes per taxon except for *Oenochroma*
vinaria (Guenée, 1858), *Acalyphes philorites* Turner, 1925, *Dirce lunaris* (Meyrick, 1890), *D.*
aesiodora Turner, 1922, *Furcatrox australis* (Rosenstock, 1885), *Chlorodontopera mandarinata*
(Leech, 1889), *Chlorozancla falcatus* (Hampson, 1895), *Pamphlebia rubrolimbraria* (Guenée,
1858) and *Thetidia albocostaria* (Bremer, 1864). For these taxa, included in studies by Young
(2006) and Ban et al. (2018), only two markers were available.

233

234 Tree search strategies and model selection

235 We ran maximum likelihood analyses with a data set partitioned by gene and codon position
236 using IQ-TREE V1.6.6 (Nguyen et al., 2015) and data partitioned by codon in RAXML
237 (Stamatakis et al 2014). IQ-TREE is a stochastic algorithm suitable for analyzing big datasets
238 (Nguyen et al., 2015). ~~Different~~ ~~Best-fitting~~ substitution models were ~~determined~~
239 ~~implementing~~ ~~selected by~~ ModelFinder, which is a model-selection method that incorporates a
240 model of ~~free~~ ~~flexible~~ rate heterogeneity across sites (Kalyaanamoorthy et al., 2017). ModelFinder
241 implements a greedy strategy as implemented in PartitionFinder that starts with the full
242 partitioned model and consequentially merges two partitions (TESTNEWMERGE option) until
243 the model fit does not increase (Lanfear et al., 2012). After the best model ~~is has been~~ found, IQ-
244 TREE starts the tree reconstruction under the best model scheme. The phylogenetic analyses
245 were carried out with ~~the~~ ~~-spp~~ option that allowed each partition to have its own evolutionary
246 rate. The RAXML analysis was ~~implemented~~ ~~carried out~~ on CIPRES using the GTR+GAMMA
247 option with a data set partitioned by gene and codon position.

248 Support for nodes ~~were~~was evaluated with 1000 ultrafast bootstrap (UFBoot2)
249 approximations (Hoang et al., 2017) in IQ-TREE, and rapid bootstrap (RBS) in RAxML
250 (Stamatakis, 2008). Additionally, we implemented SH-like approximate likelihood ratio test
251 (Guindon et al., 2010), which is considered to be a useful complement to bootstrap analysis. To
252 reduce the risk of overestimating branch supports with UFBoot2 test, we implemented *-bnni*
253 option, which optimizes each bootstrap tree using a hill-climbing nearest neighbor interchange
254 (NNI) search. Trees were visualized and edited in FigTree v1.4.3 software (Rambaut, 2012). The
255 final trees were rooted with species of the families Sematuridae, Epicopeiidae, Pseudobistonidae
256 and Uraniidae following previous hypotheses proposed in Regier et al. (2009; 2013), Rajaei et al.
257 (2015) and Heikkilä et al. (2015).

258

259 Results

260

261 *Searching strategies and model selection*

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263 The results from ModelFinder suggested that each gene and codon position kept their own
264 evolutionary model, i.e. no partitions were combined. Similarly, Akaike information criterion
265 (AIC) and Bayesian information criterion (BIC) values showed best partition schemes for the
266 data partitioned by codon position, with 33 partitions in total (evolutionary models are listed in
267 Supplemental data S3). Topologies recovered by IQ-TREE and RAxML analyses resulted in
268 trees with nearly identical patterns of relationships. Also, node support methods tended to agree
269 on the support of nodes with strong phylogenetic signal. However, in most of the cases UFBoot2
270 from IQ-TREE showed higher support values compared to RBS in RAxML (RAxML tree with
271 support values is showed in Supplemental data S4). UFBoot2 and SH-like performed similarly,
272 with UFBoot2 showing slightly higher values, and both tend to show high support for the same
273 nodes (Fig. 1). As noted by the authors of IQ-TREE, values of UF ≥ 95 and SH ≥ 80 indicate
274 well-supported clades (Trifinopoulos & Minh, 2018).

275

276 *General patterns in the phylogeny of Geometridae*

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Analyses of the dataset of 1206 terminal taxa, comprising up to 11 markers and an alignment length of 7665 bp recovered topologies with many well supported clades. About 20 terminal taxa were recovered as very similar genetically and they are likely to represent closely related species, subspecies or specimens of a single species. The examination of their taxonomic status is not the focus of this study, so the number of unique species in the analysis is slightly less than 1200. Our findings confirm the monophyly of Geometridae (values of UFBoot2, SH-like = 100) (Fig. 1). The general patterns in our phylogenetic hypotheses suggest that Sterrhinae are the sister group to the rest of Geometridae. This subfamily is recovered as monophyletic when three genera traditionally included in Oenochrominae are considered as belonging to Sterrhinae. Tribes in Sterrhinae, such as Cosymbiini and Timandriini were not recovered as monophyletic (Fig- 2). A detailed analysis, including formal changes to the classification of Sterrhinae, will be provided by Sihvonen et al. (in prep).

The monophyly of Larentiinae was established in previous studies (Sihvonen et al., 2011; Öunap et al., 2016) and our results are in full agreement with their hypotheses. However, our results do not support the sister relationship between Sterrhinae and Larentiinae found in the previous studies. In concordance with recent findings (Sihvonen et al., 2011; Öunap et al. 2016; Strutzenberger et al., 2017), we recover Dyspteridini as the sister group to the remaining Larentiinae (Fig. 3). Phylogenetic relationships within Larentiinae were treated in detail by Öunap et al. (2016). Further details of the analyses and changes to the classification of Larentiinae will be discussed by Brehm et al. (in prep) and Öunap et al. (in prep).

Archiearinae are represented by more taxa than in a previous study (Sihvonen et al., 2011), and ~~the subfamily~~ ~~is sister of~~ Oenochrominae + Desmobathrinae complex + Geometrinae and Ennominae (Fig. 4). The monophyly of this subfamily is well supported (values of SH-like, UFBoot2 = 100). However, as in the previous study (Sihvonen et al. 2011), the Australian genera *Dirce* Prout, 1910 and *Acalyphes* Turner, 1926 are not part of Archiearinae but can clearly be assigned to Ennominae. Unlike previously assumed (e.g., McQuillan & Edwards 1994), the subfamily Archiearinae doesn't occur in Australia, despite superficial similarities and the shared high-altitude distribution of *Dirce*, *Acalyphes* and Archiearinae.

Desmobathrinae were shown ~~asto be~~ paraphyletic by Sihvonen et al. (2011). In our analysis, the monophyly of this subfamily is not recovered either, as we find three taxa traditionally placed in Oenochrominae; (i.e. *Zanclopteryx* Herrich-Schäffer, [1855], *Nearcha*

309 Guest, 1887 and *Racasta* Walker, 1861) nested within Desmobathrinae (Fig. 4). We formally
310 transfer these taxa to Desmobathrinae. In the revised sense, Desmobathrinae ~~are~~reform a well-
311 supported group with two main lineages. One of them comprises the genera *Ozola* Walker, 1861,
312 *Derambila* Walker, [1863] and *Zanclopteryx*. This lineage is sister to a well-supported clade
313 comprising *Conolophia* Warren, 1894, *Noreia* Walker, 1861, *Leptoctenopsis*, *Racasta*,
314 *Ophiogramma* Hübner, [1831], *Pycnoneura* Warren, 1894 and *Dolichoneura* Warren, 1894. The
315 genus *Eumelea* Duncan [& Westwood], 1841 has an unclear phylogenetic position in our
316 analyses. The IQ-TREE result suggested  genus to be sister to the subfamily Geometrinae,
317 whereas RAxML recovered *Eumelea* in Ennominae as the sister of *Plutodes* Guenée, [1858].

318 Oenochrominae in the broad sense are not a monophyletic group. However,
319 Oenochrominae *sensu stricto* (Scoble & Edwards, 1990) form a well-supported lineage
320 comprising two clades. One of them contains a polyphyletic *Oenochroma* with *O. infantilis*
321 Prout, 1910 being sister to *Dinophalus* Prout, 1910, *Hypographa* Guenée, [1858], *Lissomma*
322 Warren, 1905, *Sarcinodes* Guenée, [1858] and two further species of *Oenochroma*, including the
323 type species *O. vinaria* Guenée, [1858]. The other clade comprises the genera *Monoctenia*
324 Guenée, [1858], *Onycodes* Guenée, [1858], *Parepisparis* Bethune-Baker, 1906, *Antictenia* Prout,
325 1910, *Arthodia* Guenée, [1858], *Gastrophora* Guenée, [1858] and *Homospora* Turner, 1904 (Fig.
326 4). Most of the remaining genera traditionally placed in Oenochrominae, including e.g.
327 *Epidesmia* Duncan [& Westwood], 1841, form a well-supported monophyletic clade that is sister
328 to Oenochrominae *s. str.* + *Eumelea ludovicata* + Geometrinae + Ennominae assemblage.
329 *Ergavia* Walker, 1866, *Ametris* Guenée, [1858] and *Macrotres* Westwood, 1841 form a
330 monophyletic group within Sterrhinae (see also Sihvonen et al., 2011).

331 The monophyly of Geometrinae is well supported (Fig. 5) and it was recovered as the
332 sister-taxon of *Eumelea*. The *Eumelea* + Geometrinae clade is sister to Oenochrominae *s. str.*
333 Although a recent phylogenetic study proposed several taxonomic changes (Ban et al., 2018), the
334 tribal composition in this subfamily is still problematic. Many tribes were recovered as
335 paraphyletic, because their constituent genera were intermingled in the phylogenetic tree.
336 Hemitheini *sensu* Ban et al. (2018) were recovered as a well-supported clade, which is sister to
337 the rest of Geometrinae. In turn, the African genus *Lophostola* Prout, 1912 was resolved as sister
338 to all other Hemitheini. The monophyly of Pseudoterpnini could not be recovered, instead this
339 tribe splits up into three well-defined groups. *Crypsiphona ocularia* Meyrick, 1888 is recovered

340 as an isolated lineage, *Xenozancla* Warren, 1893 is sister to a clade comprising Dysphaniini and
341 *Pseudoterpnini* s.str. In addition, several genera currently placed in *Pseudoterpnini* s.l. were
342 recovered as an independent lineage clearly separate from *Pseudoterpnini* s.str. (SH-like = 86.3,
343 UFBoot2 = 96). *Ornithospilini* and *Agathiini* clustered together but they were not sister to all
344 *Geometrinae* as shown by Ban et al. (2018). Although there are no phylogenetic studies which
345 investigate the relationship between *Ornithospila* Warren, 1894 and *Agathia* Guenée, [1858], our
346 results suggested that these genera are sister clades. *Aracimini*, *Neohipparchini*,
347 *Timandromorphini*, *Geometrini* and *Comibaenini* were recovered as monophyletic groups.
348 *Synchlorini* were nested within *Nemoriini* in a well-supported clade (support branch SH-like =
349 99.8, UFBoot2 = 100, RBS = 93).
350 *Ennominae* are strongly supported as monophyletic in IQ-TREE analyses (UFBoot2, and
351 SH-like = 100) whereas in RAxML the monophyly is weakly supported (RBS = 63). Detailed
352 results concerning the classification, especially for the Neotropical taxa, will be presented by
353 Brehm et al. (in prep.), but the main results are summarized here (Fig. 6). Very few tribes are
354 monophyletic according to the results of the present study. One group of Neotropical taxa
355 currently assigned to *Gonodontini*, *Gnophini*, *Odontoperini*, and *Bryoptera* Guenée, [1858] +
356 *Ectropis* Hübner, [1825], *Nacophorini*, and *Ennomini* (sensu Beljaev, 2008) grouped together in a
357 large well-supported clade (SH-like = 96.6, UFBoot2 = 97). *Ennomini* were sister to the
358 whole is entire group. The New Zealand genus *Declana* Walker, 1858 appeared as an isolated
359 lineage sister to *Campaeini*, which in turn is sister to *Alsophilini* + *Wilemaniini* + *Colotoini*.
360 These groups are in turn the sister to *Grabiola* Taylor, 1904 + *Acalyphes* Turner, 1926 and a large
361 complex including *Lithinini*, intermixed with some genera placed currently in *Nacophorini* and
362 *Diptychini*. *Theriini* were recovered close to the genera *Erastria* Hübner, [1813] + *Metarranthia*
363 Warren, 1894 and *Palyadini* + *Plutodes* Guenée, [1858]. The IQ-TREE analyses show *Palyadini*
364 as a well-defined lineage, sister to *Plutodes*. However, in RAxML analyses *Eumelea* and
365 *Plutodes* grouped together and *Palyadini* clustered with a group of *Caberini* species. The genera
366 *Neobapta* Warren, 1904 and *Oenoptila* Warren, 1895 formed an independent lineage.
367 *Hypochrosini* formed a lineage with *Apeirini*, *Epionini*, *Sericosema* Warren, 1895 and *Ithysia*
368 Hübner, [1825]. This lineage is in turn the sister of the African *Drepanogynis* Guenée, [1858]
369 which grouped together with the genera *Sphingomima* Warren, 1899, *Thenopa* Walker, 1855 and
370 *Hebdomophruda* Warren, 1897. *Caberini* came out was placed as the sister of an unnamed clade

371 composed of *Trogonia* Warren, 1905, *Acrotomodes* Warren, 1895, *Acrotomia* Herrich-Schäffer,
372 [1855] and *Pyrinia* Hübner, 1818. Finally, our analyses recovered a very large well-supported
373 clade comprising the tribes Macariini, Cassymini, Abraxini, Eutoeini and Boarmiini (SH-like and
374 UFB_{boot2} = 100). This large clade has previously been referred to informally as the “boarmiines”
375 by Forbes (1948) and Wahlberg et al. (2010). The tribe Cassymini is clearly paraphyletic: genera
376 such as *Cirrhosoma* Warren, 1905, *Berberodes* Guenée, 1858, *Hemiphricta* Warren, 1906 and
377 *Ballantiophora* Butler, 1881 currently included in Cassymini, clustered in their own clade
378 together with *Dorsifulcrum* Herbulot, 1979 and *Odontognophos* Wehrli, 1951, as sister to the
379 Abraxini and Eutoeini complex. We were unable to include Orthostixinae in the analyses, so we
380 could not clarify the taxonomic position of this subfamily with regard to the possible synonymy
381 with Ennominae (Sihvonen et al., 2011).

382

383 Discussion

384

385 Optimal partitioning scheme and support values

386 The greedy algorithm implemented in ModelFinder to select the best-fitting partitioning scheme
387 treated the partitions independently and failed to merge any data subsets. The results recovered
388 highest values (AIC and BIC) for data partitioned by codon position. These results are not
389 different from previous studies that tested the performance of different data partitioning schemes
390 and found that in some cases partitioning by gene can result in suboptimal partitioning schemes
391 and may limit the accuracy of phylogenetic analyses (Lanfear et al., 2012). However, we
392 highlight that although the AIC and BIC values were lower in data partitioned by gene, the tree
393 topology recovered was nevertheless almost the same as when data were partitioned by codon,
394 suggesting that the phylogenetic signal in the data is robust to partitioning schemes. The analyses
395 found some disagreements ~~in the methods implemented to evaluate~~ between the different
396 measures of node support. Ultrafast bootstrap gave the highest support values, followed by SH-
397 like and finally standard bootstrap as implemented in RAxML gave the lowest. Although support
398 indices obtained by these methods are not directly comparable,  differences in node support of
399 some clades can be attributed to the small number of markers, insufficient or saturated divergence
400 levels (Guindon et al., 2010).

401

403

404 **Geometridae Leach, 1815**

405 The phylogenetic hypothesis presented in this study is by far the most comprehensive to date in
406 terms of the number of markers, sampled taxa, and geographical coverage. In total our sample
407 includes 814 genera, thus representing 41% of the currently recognised Geometridae genera
408 (Scoble & Hausmann, 2007). Previous phylogenetic hypotheses were based mainly on the
409 European fauna and many clades were not unambiguously supported due to low taxon sampling.
410 The general patterns of the phylogenetic relationships between the subfamilies recovered in this
411 article largely agrees with previous hypotheses based on morphological characters and different
412 sets of molecular markers (Holloway, 1997; Abraham, 2001; Yamamoto & Sota, 2007; Sihvonen
413 et al., 2011). However, the results of our larger dataset differ in many details and sheds light on
414 the phylogenetic relationships of especially the poorly resolved small subfamilies.

415 Sterrhinae are recovered as the sister subfamily to the remaining Geometridae. This result
416 is not in concordance with Sihvonen et al. (2011), Yamamoto & Sota (2007) and Regier et al.
417 (2009), who found a sister group relationship between Sterrhinae and Larentiinae which in turn
418 were sister to the rest of Geometridae. Sihvonen et al. (2011) showed these relationships with low
419 support, while Yamamoto & Sota (2007) and Regier et al. (2009) included only a few samples in
420 their analyses, which could have had an influence on the results. Our analyses include
421 representatives from almost all known tribes currently included in Sterrhinae and Larentiinae.
422 The higher number of markers, improved methods of analysis, the broader taxon sampling as
423 well as the stability of our results suggests that Sterrhinae are indeed the sister group to the
424 remaining Geometridae Sterrhinae (after transfer of *Ergavia*, *Ametris* and *Macrotis*, see details
425 below), Larentiinae, Archiearinae, Geometrinae and Ennominae were highly supported as
426 monophyletic. Oenochrominae and Desmobathrinae formed polyphyletic and paraphyletic
427 assemblages respectively. The monophylies of Oenochrominae and Desmobathrinae have always
428 been questioned. Morphological studies addressing Oenochrominae or Desmobathrinae have
429 been very limited and the majority of genera have never been examined in depth. In addition, it
430 has been very difficult to establish the boundaries of these subfamilies only on the basis of
431 morphological examination (Scoble & Edwards, 1990). Sihvonen et al. (2011) showed that
432 neither Oenochrominae nor Desmobathrinae were monophyletic, but these results were

433 considered preliminary due to the limited number of sampled taxa, and no formal transfers were
434 proposed. To date, the phylogenetic positions of these subfamilies are not clear. The systematic
435 status of Orthostixinae remains unclear because it was not included in our study. Sihvonen et al.
436 (2011) included the genus *Naxa* Walker, 1856, formally placed in Orthostixinae, and found it to
437 be nested within Ennominae. However, only three genes were successfully sequenced from this
438 taxon, and its position in the phylogenetic tree turned out to be a highly unstable taxon in our
439 analyses. It was thus excluded from our dataset. Without a doubt, *Orthostixis* Hübner, [1823], the
440 type genus of the subfamily, needs to be included in future analyses.

441

442 **Sterrhinae Meyrick, 1892**

443 We included 74 Sterrhinae taxa in our analyses, with all tribes recognized in Forum Herbulot
444 (2007) being represented. The recovered patterns generally agree with previous phylogenetic
445 hypotheses of the subfamily (Sihvonen, 2004, Sihvonen et al., 2011). The genera *Ergavia*,
446 *Ametris* and *Macrotis*, which currently are placed in Oenochrominae were found to form a well-
447 defined lineage within Sterrhinae with strong support (SH-Like = 99 UFBoot2 = 100). These
448 genera are distributed in the New World, whereas the range of true Oenochrominae is restricted
449 to the Australian and Oriental region. Sihvonen et al. (2011) already found that *Ergavia* and
450 *Afrophyla* Warren, 1895 belong to Sterrhinae and suggested more extensive analyses to clarify
451 the position of these genera, which we did. *Afrophyla* was already transferred to Sterrhinae
452 (Sihvonen & Staude, 2011) and *Ergavia*, *Ametris* and *Macrotis* (plus *Almodes* Guenée, [1858])
453 will be transferred by Sihvonen et al. (in prep.).

454 Cosymbiini, Timandrini, Rhodometrini and Lythriini are closely related as shown
455 previously (Sihvonen & Kaila, 2004; Öunap et al., 2008; Sihvonen et al., 2011). Cosymbiini
456 appear as sister to the Timandrini + Rhodometrini + Lythriini clade. Lythriini are closely related
457 to Rhodometrini as shown by Öunap et al. (2008) with both molecular and morphological data.
458 However, Timandrini was not the closest to Rhodometrini + Lythriini clade due to the
459 phylogenetic position of *Traminda* Saalmüller, 1891 (Timandrini) and *Pseudosterrha* Warren,
460 1888 (Cosymbiini). These taxa grouped together forming a different lineage which is sister to
461 Rhodometrini + Lythriini clade (Fig. 2).

462 Rhodostropiini and Cyllopodini were recovered polyphyletic with species of Cyllopodini
463 clustering within Rhodostropiini. Similar results were recovered before (Sihvonen & Kaila,

2004; Sihvonen et al., 2011), suggesting that further work needs to be done to clarify the status and systematic position of these tribes. On the other hand, Sterrhini and Scopulini were recovered as sister taxa as proposed by Sihvonen & Kaila (2004); Hausmann (2004); Öunap et al. (2008) and Sihvonen et al. (2011). Our new phylogenetic hypothesis constitutes a large step towards understanding the evolutionary relationships of the major lineages of Sterrhinae. Further taxonomic changes and more detailed interpretation of the clades will be dealt with by Sihvonen et al. (in prep.).

Larentiinae Duponchel, 1845

Larentiinae are a monophyletic entity (Fig. 3). In concordance with the results of Sihvonen et al. (2011), Viidalepp (2011), Öunap et al. (2016) and Strutzenberger et al. (2017), Dyspteridini are placed as sister to all other larentiines. Such a systematic position is furthermore supported by the green coloration of the wings and the reduced size of the hindwings. Remarkably, *Brabiroides* Warren, 1904 forms an independent lineage. Chesiadini are monophyletic and sister to all larentiines except Dyspteridini, *Brabiroides* and Trichopterygini. These results do not support the suggestion by Viidalepp (2006) and Sihvonen et al. (2011) that Chesiadini are sister to Trichopterygini.

In our phylogenetic hypothesis, Asthenini are sister to the Perizomini + Melanthiini + Eupitheciini clade. These results do not fully agree with Öunap et al. (2016) who found Asthenini to be sister to all Larentiinae except Dyspteridini, Chesiadini, Trichopterygini and Eudulini. However, our results do support Melanthiini + Eupitheciini complex as a sister lineage to Perizomini. Sihvonen et al. (2011) recovered Phileremini and Rheumapterini as well-supported sister taxa. Our results suggest *Triphosa dubitata* Linnaeus 1758 as sister of Phileremini while Rheumapterini is the sister to this clade. Cidariini were recovered as polyphyletic, as the genera *Coenotephria* Prout, 1914 and *Lampropteryx* Stephens, 1831 cluster in a different clade apart from the lineage comprising the type genus of the tribe, *Cidaria* Treitschke, 1825. Also, *Ceratotalia* Packard, 1876, currently placed in Hydriomenini and *Trichodezia* Warren, 1895 were mixed in Cidariini. This result is not in concordance with Öunap et al. (2016), who found this tribe monophyletic. Scotopterygini were sister to a lineage comprising *Ptychorrhoe blosyrata* Guenée [1858], *Disclioprocta* sp, Euphyiini, an unnamed clade, Xanthorhoini and Cataclysmiini. Euphyiini are monophyletic, but Xanthorhoini were recovered as mixed with

495Cataclysmiini. The same findings were shown by Õunap et al. (2016), but no taxonomic
496rearrangements were proposed. Larentiini are monophyletic and sister of Hererusiini,
497Hydriomenini, Erateinini, Stamnodini and some unnamed clades. Heterusiini are recovered as a
498polyphyletic group, while Erateinini are close to Stamnodini as proposed by Sihvonen et al.
499(2011). Although with some differences, our results support the major phylogenetic patterns of
500Õunap et al. (2016).

501 Despite substantial progress, the tribal classification and phylogenetic relationships of
502Larentiinae are far from being sufficiently resolved (Õunap et al. 2016). Forbes (1948) proposed
503eight tribes based on morphological information, Viidalepp (2011) raised the number to 23 and
504Õunap et al. (2016) recovered 25 tribes studying 58 genera. Our study includes 23 tribes and 125
505genera (with a focus on Neotropical taxa). However, the phylogenetic position of many taxa
506remains unclear, and many tropical genera have not yet been formally assigned to any tribe.
507Formal descriptions of these groups will be treated in detail by Brehm et al. (in prep) and Õunap
508et al (in prep).

509

510**Archiearinae Fletcher, 1953**

511The hypothesis presented in this study recovered Archiearinae as a monophyletic entity if some
512taxonomic rearrangements are done. This subfamily was previously considered as sister to
513Geometrinae + Ennominae (Abraham et al., 2001), whereas Yamamoto & Sota (2007) proposed
514them as the sister-taxon to Orthostixinae + Desmobathrinae. Our findings agree with Sihvonen et
515al. (2011) who recovered Archiearinae as the sister-taxon to the rest of Geometridae excluding
516Sterrhinae and Larentiinae, although only one species was included in their study. *Archiearis*
517Hübner, [1823] is sister to *Boudinotiana* Esper, 1787 and these taxa in turn are sister to
518*Leucobrephe* Grote, 1874 (Fig. 4). The southern hemisphere Archiearinae require more
519attention. Young (2006) suggested that two Australian Archiearinae genera, *Dirce* and *Acalyphes*,
520actually belong to Ennominae. Our analyses clearly support this view and we therefore propose to
521formally transfer *Dirce* and *Acalyphes* to Ennominae (all formal taxonomic changes are provided
522in Table 1). Unfortunately, the South American Archiearinae genera *Archiearides* Fletcher, 1953
523and *Lachnocephala* Fletcher, 1953, and Mexican *Caenosyntes* Dyar, 1912 (Pitkin & Jenkins
5242004), could not be included in our analyses. The position in Archiearinae requires further study.

525 These presumably diurnal taxa may only be superficially similar to northern hemisphere
526 Archiearinae as was the case with Australian *Dirce* and *Acalyphes*.

527

528 **Desmobathrinae Meyrick, 1886**

529 Taxa placed in Desmobathrinae were formerly recognized as Oenochrominae genera with slender
530 appendages. Holloway (1996) revived this subfamily from synonymy with Oenochrominae and
531 divided it into the tribes Eumeleini and Desmobathrini. Desmobathrinae species have a
532 pantropical distribution and they apparently (still) lack recognized morphological apomorphies
533 (Holloway, 1996). Our phylogenetic analysis has questioned the monophyly of Desmobathrinae
534 *sensu* Holloway because some species currently placed in Oenochrominae were embedded within
535 the group (see also Sihvonen et al., 2011), and also the phylogenetic position of the tribe
536 Eumeleini is unstable (see below). Desmobathrinae can be regarded as a monophyletic group in
537 our study, after the transfer of *Zanclopteryx*, *Nearcha* and *Racasta* from Oenochrominae to
538 Desmobathrinae, and the removal of Eumeleini (Table 1). Desmobathrinae as circumscribed here
539 are an independent lineage that is sister to all Geometridae except Sterrhinae, Larentiinae and
540 Archiearinae.

541 The monobasic Eumeleini (comprising only the genus *Eumelea*) has had a dynamic
542 taxonomic history: *Eumelea* was transferred from Oenochrominae *s.l.* to Desmobathrinae based
543 on the pupal cremaster (Holloway, 1996), whereas Beljaev (2008) pointed out that *Eumelea*
544 could be a member of Geometrinae based on the skeleto-muscular structure of the male genitalia.
545 Molecular studies (Sihvonen et al., 2011, Ban et al., 2018) suggested that *Eumelea* was part of
546 Oenochrominae *s.str.*, but these findings were not well-supported and no formal taxonomic
547 changes were proposed. Our analyses with IQTREE and RAxML recovered Eumeleini in two
548 very different positions,  either as sister to Geometrinae (SH-like = 92, UFBoot2 = 98) rather than
549 belonging to Desmobathrinae (figs 4, 5), or as sister of *Plutodes* in Ennominae (RBS = 60)
550 (Supplemental data S4). The examination of morphological details suggests that the position as
551 sister to Geometrinae is more plausible: hindwing vein M2 is present and tubular; anal margin of
552 the hindwing is elongated; and large coremata originate from saccus (Holloway 1994, our
553 observations). The morphology of *Eumelea* is partly unusual, and for that reason we illustrate
554 selected structures (Supplemental data S5), which include for instance the following: antennae
555 and legs of both sexes are very long; forewing vein Sc (homology unclear) reaches wing margin;

556 in male genitalia coremata are extremely large and branched; uncus is cross-shaped (cruciform);
557 tegumen is narrow and it extends ventrally beyond the point of articulation with vinculum; saccus
558 arms are extremely long, looped; and vesica is with lateral rows of cornuti. However, the green
559 geoverdin pigment concentration of *Eumelea* is low in comparison to Geometrinae (Cook et al.,
560 1994). We tentatively conclude that *Eumelea* is probably indeed associated with Geometrinae.
561 However, since eleven genetic markers were not sufficient to clarify the phylogenetic affinities of
562 *Eumelea*, we provisionally place the genus as *incertae sedis* (Table 1).

563

564 **Oenochrominae Guenée, [1858]**

565 Oenochrominae has obviously been the group comprising taxa that could not easily be assigned
566 to other subfamilies. Out of the 76 genera currently assigned to Oenochrominae, our study
567 includes 25 genera (28 species). Three of these genera will be formally transferred to Sterrhinae
568 (Sihvonen et al. in prep.), two are here transferred to Desmobathrinae (see above, Table 1), and
569 eight are transferred to Epidesmiinae (see below). In agreement with Sihvonen et al. (2011),
570 Oenochrominae s. str. grouped together in a well-supported lineage. Genera of this clade can be
571 characterized as having robust bodies, and their male genitalia have a well-developed uncus and
572 gnathos, broad valvae and a well-developed anellus (Scoble & Edwards, 1990). Common host
573 plants are members of Proteaceae and Myrtaceae (Holloway, 1996). Our results strongly suggest
574 that the genus *Oenochroma* is polyphyletic: *O. infantilis* is sister to a clade including *Dinophalus*,
575 *Hypographa*, *Lissomma*, *Sarcinodes* and (at least) two species of *Oenochroma*. To date, 20
576 species have been assigned to *Oenochroma* by Scoble (1999), and one additional species was
577 described by Hausmann et al. (2009), who suggested that *O. vinaria* is a species complex. We
578 agree with Hausmann et al. (2009), who pointed out the need of major revision and taxonomic
579 definition of *Oenochroma*.

580 In our phylogenetic hypothesis, *Sarcinodes* is sister to *O. orthodesma* and *O. vinaria*, [the](#)
581 [type species of *Oenochroma*](#). Although *Sarcinodes* and *Oenochroma* resemble each other in
582 external morphology, a sister-group relationship between these genera has not been hypothesized
583 before. The inclusion of *Sarcinodes* in Oenochrominae is mainly based on shared tympanal
584 characters (Scoble & Edwards, 1990). However, the circular form of the lacinia, which is an
585 apomorphy of Oenochrominae s.str. is missing or not apparent in *Sarcinodes* (Holloway, 1996).
586 In addition, *Sarcinodes* is found in the Oriental rather than in the Australian region, where all

587 *Oenochroma* species are distributed. A second clade of Oenochrominae s.str. comprises of the
588 genera *Monoctenia*, *Onycodes*, *Parepisparis*, *Antictenia*, *Arhodia*, *Gastrophora* and *Homospora*,
589 which clustered together as the sisters of *Oenochroma* and its relatives. These genera are widely
590 recognized in sharing similar structure of male genitalia (Scoble & Edwards, 1990), yet their
591 phylogenetic relationships have never been tested. Young (2006) suggested the monophyly of
592 Oenochrominae s.str., however, with a poorly resolved topology and low branch support. In her
593 study, *Parepisparis*, *Phallaria* and *Monoctenia* shared a bifid head, while in *Parepisparis* and
594 *Onychodes*, the aedeagus was lacking caecum and cornuti. Our analysis supports these
595 morphological similarities. ***Monoctenia*, *Onycodes* and *Parepisparis* clustered together**
596 However, a close relationship of the genera *Antictenia*, *Arhodia*, *Gastrophora* and *Homospora*
597 has not been suggested before. Our analysis thus strongly supports the earliest definition of
598 Oenochrominae proposed by Guenée (1858), and reinforced by Cook & Scoble (1992).
599 Oenochrominae should be restricted to *Oenochroma* and related genera such as *Dinophalus*,
600 *Hypographa*, *Lissomma*, *Sarcinodes*, *Monoctenia*, *Onycodes*, *Parepisparis*, *Antictenia*, *Arhodia*,
601 *Gastrophora*, *Homospora*, *Phallaria* and *Palaeodoxa*. We consider that genera included to in
602 Oenochrominae by (Scoble & Edwards, (1990), but recovered in a separate lineage apart separate
603 from *Oenochroma* and its close relatives in our study, belong to a hitherto unknown subfamily,
604 which is described below.

605

606 **Epidesmiinae** Murillo-Ramos, Brehm & Sihvonen **new subfamily**

607

608 Type genus: *Epidesmia* Duncan [& Westwood], 1841.

609 Material examined: Taxa included in the molecular phylogeny: *Ecphyas* Turner, 1929, *Systatica*
610 Turner, 1904, *Adeixis* Warren, 1987, *Dichromodes* Guenée, 1858, *Phrixocomes* Turner, 1930,
611 *Abraxaphantes* Warren, 1894, *Epidesmia* Duncan [& Westwood], 1841, and *Phrataria* Walker,
612 [1863]. 

613 Most of the slender-bodied Oenochrominae, excluded from Oenochrominae s. str. by Holloway
614 (1996), were recovered as an independent lineage (Fig. 4) that consists of two clades: *Ecphyas* +
615 *Systatica* and *Epidesmia* + five other genera. Branch support values in the from IQ-TREE strongly
616 support the monophyly of this clade (UFBoot2; and SH-like = 100), while in RAxML the clade
617 is moderately supported (RBS = 89). These genera have earlier been assigned to Oenochrominae

618*s.l.* (Scoble & Edwards, 1990). However, we recovered the group as a well-supported lineage
619independent from Oenochrominae *s. str.* and transfer them to Epidesmiinae, subfam. n. (Table 1).
620Phylogenetic position: Epidesmiinae is sister to Oenochrominae *s. str.* + *Eumelea* + Geometrinae
621+ Ennominae.

622Short description of Epidesmiinae: Antennae in males unipectinate (exception: *Adeixis*), towards
623apex shorter towards the apex. Pectination moderate or long. Thorax and abdomen slender
624(unlike in Oenochrominae). Forewings with sinuous postmedial line and areole present.
625Forewings planiform (with wings lying flat on the substrate) in resting position, held like a
626triangle, and cover the hindwings.

627Diagnosis of Epidesmiinae: The genera included in this subfamily form a strongly supported
628clade with DNA sequence data from the following gene regions (exemplar *Epidesmia chilonaria*
629Herrich-Schäffer, [1855]) ArgK (GB Accession number), Ca-ATPase (GB Accession number),
630CAD (GB Accession number), COI (GB Accession number), EF1a (GB Accession number),
631GAPDH (GB Accession number), MDH (GB Accession number) and Nex9 (GB Accession
632number). **(note to the editor: GB accession numbers will be provided on acceptance).** A
633thorough morphological diagnosis requires further research.

634Distribution: Most genera are distributed in the Australian region, with range of some extending
635to the Orient as well, and *Apraxaphantes* is the only genus that occurs exclusively in the Oriental
636region

637

638Geometrinae Stephens, 1829

639The monophyly of Geometrinae is strongly supported, but the number of tribes included in this
640subfamily is still unclear. Sihvonen et al. (2011) analyzed 27 species assigned to 11 tribes,
641followed by Ban et al. (2018) with 116 species in 12 tribes. Ban et al. (2018) synonymized nine
642tribes, and validated the monophyly of 12 tribes, with two new tribes Ornithospilini and Agathiini
643being the first two clades branching off the main lineage of Geometrinae. Our study (168 species)
644validates the monophyly of 13 tribes, eleven of which were defined in previous studies:
645Hemitheini, Dysphaniini, Pseudoterpnini *s.str.*, Ornithospilini, Agathiini, Aracimini,
646Neohipparchini, Timandromorphini, Geometrini, Comibaeini, Nemoriini. One synonymization is
647proposed: Synchronini Ferguson, 1969 **syn. nov.** is synonymized with Nemoriini.

tribe is proposed as new: Chlorodontoperini **trib. nov.**, and one tribe (Archaeobalbini Viidalepp, 1981, **stat. rev.**) is raised from synonymy of Pseudoterpnini to tribe status.

In our phylogenetic hypothesis, a large clade including the former tribes Lophochoristini, Heliotheini, Microloxiini, Thalerini, Rhomboristini, Hemistolini, Comostolini, Jodini and Thalassodini is recovered as sister to the rest of Geometrinae. These results are in full agreement with Ban et al. (2018), who synonymized all [of](#) these tribes with Hemitheini. Although the monophyly of Hemitheini is strongly supported, our findings recovered only a few monophyletic subtribes. For example, genera placed in Hemitheina were intermixed with those belonging to Microloxiina, Thalassodina and Jodina. Moreover, many genera which were unassigned to tribe, were recovered as belonging to Hemitheini. Our findings recovered *Lophostola* Prout, 1912 as sister to all Hemitheini. These results are quite different from those found by Ban et al. (2018) who suggested Rhomboristina as being sister to the rest of Hemitheini. In contrast, our results recovered Rhomboristina mingled with Hemistolina. These different results are probably influenced by the presence of African and Madagascan *Lophostola* in our analysis. We feel that the concept of subtribe is not practical at this point in time and thus do not advocate its use in [G](#)geometridae classification.

The Australian genus *Crypsiphona* Meyrick, 1888 is sister to all tribes included in Geometrinae except Hemitheini. *Crypsiphona* has been assigned to Pseudoterpnini (e. g. Pitkin et al. 2007, Öunap & Viidalepp 2009), but is recovered as a separate lineage in our tree. Given the isolated position of *Crypsiphona*, the designation of a new tribe could be considered, but due to low support of branches in our analyses, further information (including morphology) is needed to confirm the phylogenetic position of this genus. *Xenozancla* Warren, 1893 is placed as sister to the clade comprising Dysphaniini and Pseudoterpnini s. str.. Sihvonen et al. (2011) did not include *Xenozancla* in their analyses and suggested the sister relationships of Dysphaniini and Pseudoterpnini but with low support. According to Ban et al. (2018), *Xenozancla* is more closely related to Pseudoterpnini s.str. rather than to Dysphaniini. However, due to low support of clades, Ban et al. (2018) did not propose a taxonomic assignment to *Xenozancla*, which is currently not assigned to a tribe. Although our IQ-TREE results show that *Xenozancla* is sister of clade comprising Dysphaniini and Pseudoterpnini s. str., the RAxML analysis did not recover the same phylogenetic relationships. Instead, Dysphaniini + Pseudoterpnini s.str. are found to be sister to each other, but *Xenozancla* is placed close to *Rhomborista monosticta* (Wehrli, 1924). As in Ban

et al. (2018), due to low support of nodes, we cannot reach ~~to~~ any conclusion about the phylogenetic affinities of these tribes based on our results due to low support of nodes.

The monophyly of Pseudoterpnini *sensu* Pitkin et al. (2007) could not be recovered. Same results were shown by Ban et al. (2018) who recovered Pseudoterpnini *s.l.* including all the genera previously studied by Pitkin et al. (2007) ~~and~~, forming a separate clade from *Pseudoterpna* Hübner, [1823]+ *Pingasa* Moore, 1887. Our results showed the African *Mictoschema* Prout, 1922 falling within Pseudoterpnini *s.str.*, and it is sister to *Pseudoterpna* and *Pingasa*. A second group of Pseudoterpnini *s.l.* was recovered as an independent lineage clearly separate from Pseudoterpnini *s.str.* (SH-like = 86.3, UFBoot2 = 96). Ban et al. (2018) did not introduce a new tribe due to the morphological similarities and difficulty in finding apomorphies of Pseudoterpnini *s.str.* In addition, their results were weakly supported. Considering that two independent studies have demonstrated the paraphyly of Pseudoterpnini *sensu* Pitkin et al (2007), we see no reason for retaining the wide concept of this tribe. Instead we propose the revival of the tribe status of Archaeobalbini and the description of a new tribe Chlorodontoperini, which removes paraphyly from the clades in question.

694

Archaeobalbini Viidalepp, 1981, **status revised**

(original spelling: Archeobalbini, justified emendation in Hausmann (1996))

Type genus: *Archaeobalbis* Prout, 1912 (synonymized with *Herochroma* Swinhoe, 1893 in Holloway (1996))

Material examined: *Herochroma curvata* Han & Xue, 2003, *H. baba* Swinhoe 1893, *Metallolophia inanularia* Han & Xue, 2004, *M. cuneataria* Han & Xue, 2004, *Actenochroma muscicoloraria* (Walker, 1862), *Absala dorcada* Swinhoe, 1893, *Metaterpna batangensis* Hang & Stünig, 2016, *M. thyatiraria* (Oberthür, 1913), *Limbatochlamys rosthorni* Rothschild, 1894, *Pachyodes pictaria* Moore, 1888, *Dindica para* Swinhoe, 1893, *Dindicodes crocina* (Butler, 1880), *Lophophelma erionoma* (Swinhoe, 1893), *L. varicoloraria* (Moore, 1868), *L. iterans* (Prout, 1926) and *Pachyodes amplificata* (Walker, 1862).

706

This lineage splits into four groups: *Herochroma* Swinhoe, 1893 + *Absala* Swinhoe, 1893 + *Actenochroma* Warren, 1893 is the sister lineage of the rest of Archaeobalbini that were recovered as a polytomic bunch of three clades ~~conforming~~comprising the genera

710 *Limbatochlamys* Rothschild, 1894, *Psilotagma* Warren, 1894, *Metallolophia* Warren, 1895,
711 *Metaterpna* Yazaki, 1992, *Dindica* Warren, 1893, *Dindicodes* Prout, 1912, *Lophophelma* Prout,
712 1912 and *Pachyodes* Guenée, 1858. This tribe can be diagnosed by the combination of DNA data
713 from six genetic markers, see for instance *Pachyodes amplificata* (CAD, COI, EF1a, GAPDH,
714 MDH RpS5) shown in supplementary material. Branch support values in IQ-TREE strongly
715 confirm the monophyly of this clade (SH-like = 86.3, UFBoot2 = 96). GenBank accession
716 numbers are shown in supplementary material. A morphological diagnosis requires further
717 research. 

718

719 Chlorodontoperini Murillo-Ramos, Sihvonen & Brehm, **new tribe**

720 Type genus: *Chlorodontopera* Warren, 1893

721 Material examined: Taxa in the molecular phylogeny: *C. discospilata* (Moore, 1867) and *C.*
722 *mandarinata* (Leech, 1889).

723

724 Some studies (Inoue, 1961; Holloway, 1996) suggested the morphological similarities of
725 *Chlorodontopera* Warren, 1893 with members of Aracimini. Moreover Holloway (1996)
726 considered this genus as part of Aracimini. Our results suggest a sister relationship of
727 *Chlorodontopera* with Aracimini rather than the inclusion in the tribe as well as the sister
728 relationship with a large lineage comprising the rest of Geometrinae. Considering that our
729 analysis strongly supports *Chlorodontopera* as an independent lineage (branch support SH-like =
730 99 UFBoot2 = 100, RBS = 99), we introduce the monobasic tribe Chlorodontoperini. This tribe
731 can be diagnosed by the combination of DNA data from six genetic markers (exemplar
732 *Chlorodontopera discospilata*) CAD (MG015448), COI (MG014735), EF1a (MG015329),
733 GAPDH (MG014862), MDH (MG014980) and RpS5 (MG015562). Ban et al. (2018) did not
734 introduce a new tribe because the relationship between *Chlorodontopera* and *Euxena* Warren,
735 1896 was not clear in their study. This relationship was also been proposed by Holloway (1996)
736 based on similar wing patterns. Further analyses are needed to clarify the affinities between
737 *Chlorodontopera* and *Euxena*.

738 The tribe Chlorodontoperini is diagnosed by distinct discal spots with pale margins on the
739 wings, which are larger on the hindwing; a dull reddish-brown patch is present between the discal
740 spot and the costa on the hindwing, and veins M3 and CuA1 are not stalked on the hindwing

741(Ban et al., 2018). In the male genitalia, the socii are stout and setose and the lateral arms of the
742gnathos are developed, not joined. Sternite 3 of the male has setal patches. Formal taxonomic
743changes are listed in Table 1.

744

745Aracimini, Neohipparchini, Timandromorphini, Geometrini and Comibaenini were recovered as
746monophyletic groups. These results are in full agreement with Ban et al. (2018). However, the
747phylogenetic position of *Eucyclodes* Warren, 1894 is ~~not clear~~uncertain. This genus is placed as
748sister of Comibaenini (support branch SH-like = 32.4, UFBoot2 = 100, RBS = 67). The
749monophyly of Nemoriini and Synchronini is not supported. Instead, Synchronini are nested within
750Nemoriini (support branch SH-like = 99.8, UFBoot2 = 100, RBS = 93). Our findings are in
751concordance with Sihvonen et al. (2011) and Ban et al. (2018), but our analyses included a larger
752number of markers and a much higher number of taxa. Thus, we formally synonymize
753Synchronini **syn. nov.** with Nemoriini (Table 1).

754

755Ennominae Duponchel, 1845

756Ennominae are the most species-rich subfamily of geometrids. The loss of vein M2 on the
757hindwing is probably the best apomorphy (Holloway, 1993), although this character does not
758occur in a few ennomine taxa (Staude, 2001; Skou & Sihvonen, 2015). Ennominae are a
759morphologically highly diverse subfamily, and attempts to find further synapomorphies shared by
760all major tribal groups have failed.

761 The number of tribes as well as phylogenetic relationships among tribes are still debatable
762(see Skou & Sihvonen, 2015 for an overview). Moreover, the taxonomic knowledge of this
763subfamily in tropical regions is still poor. Holloway (1993) recognized 21 tribes, Beljaev (2006)
76424 tribes, and Forum Herbulot (2007) 27 tribes. To date, five molecular studies have corroborated
765the monophyly of Ennominae (Young, 2006; Yamamoto & Sota, 2007; Wahlberg et al., 2010;
766Öunap et al., 2011, Sihvonen et al. 2011) with no conflicting evidence ever presented, with
767Young (2006) being the only exception who found a paraphyletic Ennominae. Moreover, three
768large-scale taxonomic revisions (without a phylogenetic hypothesis) were published by Pitkin
769(2002) for the Neotropical region, Skou & Sihvonen (2015) for the Western Palaearctic region,
770and Holloway (1994) for Borneo. More detailed descriptions of taxonomic changes in Ennominae

will be given by Brehm et al. (in prep) and Murillo-Ramos et al. (in prep), here we discuss general patterns and give details for taxonomic acts not covered in the other two papers.

Our findings recover Ennominae as a monophyletic entity, but results were not highly supported in RAxML (RBS = 67) results compared to IQ-TREE (UFBoot2 and SH-Like = 100). The lineage comprising Geometrinae and Oenochrominae is recovered as the sister clade of Ennominae. In previous studies, Wahlberg et al. (2010) sampled 49 species of Ennominae, Öunap et al. (2011) sampled 33 species, and Sihvonen et al. (2011) 70 species including up to eight markers per species. All these studies supported the division of Ennominae into “boarmiine” and “ennomine” moths (Holloway, 1994). This grouping was proposed by Forbes (1948) and Holloway (1994), who suggested close relationships between the tribes Boarmiini, Macariini, Cassymini and Eutoeini based on the bifid pupal cremaster and the possession of a fovea in the male forewing. The remaining tribes were defined as “ennomines” based on the loss of a setal comb on male sternum A3 and the presence of a strong furca in male genitalia. Both Wahlberg et al. (2010) and Sihvonen et al. (2011) found these two informal groupings to be reciprocally monophyletic.

In our analyses, 653 species with up to 11 markers were sampled, with an emphasis on Neotropical taxa which so far had been poorly represented in the molecular phylogenetic analyses. Our results recovered the division into two major subclades, a core set of ennomines in a well-supported clade, and a poorly supported larger clade that includes the “boarmiines” among four other lineages usually thought of as “ennomines”. The traditional “ennomines” are thus not found to be monophyletic in our analyses, questioning the utility of such an informal name. Our phylogenetic hypothesis supports the validation of numerous tribes earlier proposed, in addition to several unnamed clades. We validate 23 tribes (Forum Herbulot, 2007; Skou & Sihvonen, 2015): Gonodontini, Gnophini, Odontoperini, Nacophorini, Ennomini, Campaeini, Alsophilini, Wilemaniini, Prosopolophini, Diptychini, Theriini, Plutodini, Palyadini, Hypochrosini, Apeirini, Epionini, Caberini, Macariini, Cassymini, Abraxini, Eutoeini and Boarmiini. We hereby propose one new tribe: Drepanogynini **trib. nov.** (Table 1). Except for the new tribe, most of the groups recovered in this study are in concordance with previous morphological classifications (Holloway, 1993; Beljaev, 2006, 2016; Forum Herbulot, 2007; Skou & Sihvonen, 2015).

Five known tribes and two further unnamed lineages form the core Ennominae:

Gonodontini, Gnophini, Odontoperini, Nacophorini and Ennomini. Several Neotropical

clades that conflict with the current tribal classification of Ennominae will be described as new tribes by Brehm et al (in prep). Gonodontini and Gnophini are recovered as sister taxa. Gonodontini was defined by Forbes (1948) and studied by Holloway (1994), who showed synapomorphies shared by *Gonodontis* Hübner, [1823], *Xylinophylla* Warren, 1898 and *Xenimpia* Warren, 1895. Our results recovered the genus *Xylinophylla* as sister of *Xenimpia* and *Psilocladia* Warren, 1898. *Psilocladia* is an African genus currently unassigned to tribe (see Sihvonen et al., 2015 for details). Considering the strong support and that the facies and morphology are somewhat similar to other analysed taxa in Gonodontini, we formally include *Psilocladia* in Gonodontini (Table 1). Gnophini are a well-defined assemblage and we formally transfer the African genera *Oedicentra* Warren, 1902 and *Hypotephrina* Janse, 1932, from unassigned to Gnophini (Table 1). The total number of species, and number of included genera in Gnophini are still uncertain (Skou & Sihvonen, 2015). Based on morphological examination, Beljaev (2007, 2016) treated Angeronini as a synonym of Gnophini. The costal projection on male valva bearing a spine or group of spines was considered as a synapomorphy of the group. Using molecular data, Yamamoto & Sota (2007) showed [thea](#) close phylogenetic relationship between *Angerona* Duponchel, 1829 (Angeronini) and *Chariaspilates* Wehrli, 1953 (Gnophini). Similar results were shown by Sihvonen et al. (2011) who recovered *Angerona* and *Charissa* Curtis, 1826 as sister taxa, and our results also strongly support treating Angeronini as synonym of Gnophini.

Holloway (1993) suggested close affinities among Nacophorini, Azelinini and Odontoperini on the basis of larval characters. In a morphology-based phylogenetic study, Skou & Sihvonen (2015) suggested multiple setae on the proleg on A6 of the larvae as a synapomorphy of the group. Our results also supported a close relationship of Nacophorini, Azelinini and Odontoperini. These clades will be treated in more detail by Brehm et al. (in prep.).

Following the ideas of Pitkin (2002), Beljaev (2008) synonymized the tribes Ourapterygini and Nephodiini with Ennomini. He considered the divided vinculum in male genitalia and the attachment of muscles *m*₃ as apomorphies of the Ennomini, but did not provide a phylogenetic analysis. Sihvonen et al. (2011) supported Beljaev's assumptions and recovered *Ennomos* Treitschke, 1825 (Ennomini), *Ourapteryx* Leach, 1814 (Ourapterygini) and *Nephodia* Hübner, [1823] (Nephodiini) as belonging to the same clade. Our comprehensive analysis

832 confirms those previous findings and we agree with Ennomini as valid tribal name for this large
833 clade.

834

835 The genus *Declana* Walker, 1858 is recovered as an isolated clade sister to a complex
836 lineage comprising Campaeini, Alsophilini, Wilemaniini and Prosopopolophini. This genus is
837 endemic to New Zealand, but to date has not been assigned to any tribe. According to our results,
838 *Declana* could well be defined as its own tribe. However, the delimitation of this tribe is beyond
839 the scope of our paper and more genera from Australia and New Zealand should first be
840 examined.

841 Campaeini, Alsophilini, Wilemaniini and Prosopopolophini grouped together in a well-
842 supported clade (SH-like and UFBoot2 = 100). Previous molecular analyses have shown an
843 association of Colotoini [= Prosopopolophini] and Wilemaniini (Yamamoto & Sota, 2007;
844 Sihvonen et al., 2011), although no synapomorphies are known to support synonymization (Skou
845 & Sihvonen, 2015). The Palaearctic genera *Compsoptera* Blanchard, 1845, *Apochima* Agassiz,
846 1847, *Dasycorsa* Prout, 1915, *Chondrosoma* Anker, 1854 and *Dorsispina* Nupponen &
847 Sihvonen, 2013, are potentially part of the same complex (Skou & Sihvonen, 2015, Sihvonen
848 pers. obs.), but they were not included in the current study. Campaeini is a small group including
849 four genera with Oriental, Palaearctic and Nearctic distribution, apparently closely related to
850 Alsophilini and Prosopopolophini, but currently accepted as a tribe (Forum Herbulot, 2007;
851 Sihvonen & Skou, 2015). Our results support the close phylogenetic affinities among these tribes,
852 but due to the limited number of sampled taxa, we do not propose any formal changes.

853 A close relationship between Nacophorini and Lithinini was suggested by Pitkin (2002),
854 based on the similar pair of processes of the anellus in the male genitalia. Pitkin also noted a
855 morphological similarity in the male genitalia (processes of the juxta) shared by Nacophorini and
856 Diptychini. In a study of the Australasian fauna, Young (2008) suggested the synonymization of
857 Nacophorini and Lithinini. This was further corroborated by Sihvonen et al. (2015) who found
858 that Diptychini were nested within some Nacophorini and Lithinini. However, none of the studies
859 proposed formal taxonomic changes because of limited taxon sampling. In contrast, samples in
860 our analyses cover all biogeographic regions and the results suggest that the true Nacophorini is a
861 clade which comprises almost exclusively New World species. This clade is clearly separate from

862 Old World “nacophorines” (cf. Young, 2003) that are intermixed with Lithinini and Diptychini.
863 We here formally transfer Old World nacophorines to Diptychini and synonymize Lithinini **syn.**
864 **nov.** with Diptychini (Table 1). Further formal taxonomic changes in the Nacophorini complex
865 are provided by Brehm et al. (in prep.).

866 *Theria* Hübner, [1825], the only representative of Theriini in this study, clustered together
867 with *Lomographa* Hübner, [1825] (Baptini in Skou & Sihvonen, 2015), in a well-supported
868 clade, agreeing with the molecular results of Sihvonen et al. (2011). The placement of
869 *Lomographa* in Caberini (Rindge, 1979; Pitkin, 2002) is not supported by our study nor by that of
870 ~~by~~ Sihvonen et al. (2011). The monophyly of *Lomographa* has not been tested before, but we
871 show that the Neotropical and Palaearctic *Lomographa* species indeed group together. Our results
872 show that Caberini are not closely related to the Theriini + Baptini clade, unlike in the earlier
873 morphology-based hypotheses (Rindge, 1979; Pitkin 2002). Morphologically, Theriini and
874 Baptini are dissimilar, therefore we recognize them as valid tribes (see description and
875 illustrations in Skou & Sihvonen, 2015).

876 According to our results, 11 molecular markers were not enough to infer phylogenetic
877 affinities of Plutodini (represented by one species of *Plutodes*). Similar results were found by
878 Sihvonen et al. (2011), who in some analyses recovered *Plutodes* as sister of *Eumelea*. Our
879 analyses are ~~in-concordance~~ **congruent** with those findings, IQ-TREE results suggested that
880 *Plutodes* ~~is~~ **is** sister to Palyadini, but RAxML analyses recovered *Eumelea* as the most probable
881 sister of *Plutodes*. Given that our analyses were not in agreement about the sister-group affinities
882 of *Plutodes*, we do not make any assumptions ~~to about~~ its ~~the~~ phylogenetic position. Instead we
883 emphasize that further works ~~needs~~ **need**s to be done to clarify the phylogenetic positions of *Plutodes*
884 and related groups.

885 Hypochrosini is **only** recovered in a well-defined lineage ~~only~~ if the genera *Apeira* Gistel,
886 1848 (Apeirini), *Epione* Duponchel, 1829 (Epionini), *Sericosema* (Caberini), *Ithysia* (Theriini),
887 *Capasa* Walker, 1866 (unassigned) ~~and~~ **and**; *Omizodes* Warren, 1894 (unassigned) would be
888 transferred to Hypochrosini. Skou & Sihvonen (2015) already suggested a close association of
889 Epionini, Apeirini and Hypochrosini. We think that ~~the~~ **the** synonymizati**ons** ~~ing~~ **ing** of these tribes is
890 desirable. However, due to the limited number of sampled taxa we do not propose any formal
891 changes until more data **will** become available. We do suggest, however, formal taxonomic
892 changes of the genera *Capasa* and *Omizodes* from unassigned to Hypochrosini (Table 1).

893 The southern African genus *Drepanogynis* is paraphyletic and has earlier been classified
894 as belonging in Ennomini, and later in Nacophorini (Krüger 2002). In our phylogeny, it is
895 intermixed with the genera *Sphingomima* Warren, 1899, and *Thenopa* Walker, 1855.
896 *Hebdomophruda errans* Prout, 1917 **also** clustered together with these taxa **also**, apart from other
897 *Hebdomophruda* Warren, 1897 species, which suggests that this genus is polyphyletic. These
898 genera form a clade sister to the lineage that comprises several Hypochrosini species.
899 Considering that our analysis strongly supports this clade, we place *Thenopa*, *Sphingomima* and
900 *Drepanogynis* in a tribe of their own.

901

902 Drepanogynini Murillo-Ramos, Sihvonen & Brehm **new tribe**

903

904 Type genus: *Drepanogynis* Guenée, [1858]

905

906 The African genera *Thenopa*, *Sphingomima* and *Drepanogynis* appeared as a strongly supported
907 lineage (RBS, SH-like and UFBoot2 = 100). Krüger (1997, p. 259) proposed "Boarmiini and
908 related tribes as the most likely sister group" for *Drepanogynis*, whereas more recently
909 *Drepanogynis* was classified in the putative southern hemisphere Nacophorini (Krüger, 2014;
910 Sihvonen et al., 2015). In the current phylogeny, *Drepanogynis* is isolated from Nacophorini
911 *sensu stricto* and from other southern African genera that have earlier been considered to be
912 closely related to it (Krüger 2014 and references therein). The other southern African genera
913 appeared as belonging to Diptychini in our study. The systematic position of *Drepanogynis*
914 *tripartita* (Warren, 1898) has earlier been analysed in a molecular study (Sihvonen et al., 2015).
915 The taxon grouped together with the Palearctic species of the tribes Apeirini, Theriini, Epionini
916 and putative Hypochrosini. Sihvonen et al. (2015) noted that *Argyrophora trofonia* (Cramer,
917 [1779]) (representing *Drepanogynis* group III *sensu* Krüger, 1999) and *Drepanogynis tripartita*
918 (representing *Drepanogynis* group IV *sensu* Krüger, 2002) did not group together, but no formal
919 changes were proposed. Considering that the current analysis strongly supports the placement of
920 *Drepanogynis* and related genera in an independent lineage, and the aforementioned taxa in the
921 sister lineage (Apeirini, Theriini, Epionini and putative Hypochrosini) have been validated at
922 tribe-level, we place *Drepanogynis* and related genera in a tribe of their own.

923 Material examined and taxa included: *Drepanogynis mixtaria* Guenée, [1858], *D.*
924 *tripartita*, *D. determinata* (Walker, 1860), *D. arcuifera* Prout, 1934, *D. arcuatilinea* Krüger,
925 2002, *D. cnephaeogramma* (Prout, 1938), *D. villaria* (Felder & Rogenhofer, 1875),
926 "*Sphingomima*" *discolucida* Herbulot, 1995 (genus combination uncertain, see taxonomic notes
927 below), *Thenopa diversa* Walker, 1855, "*Hebdomophruda*" *errans* Prout, 1917 (genus
928 combination uncertain, see taxonomic notes below).

929 Taxonomic notes: We choose *Drepanogynis* Guenée, [1858] as the type genus for
930 Drepanogynini, although it is not the oldest valid name (ICZN Article 64), because extensive
931 literature has been published on *Drepanogynis* (Krüger 1997, 1998, 1999, 2014), but virtually
932 nothing exists on *Thenopa*, except the original descriptions of its constituent species. Current
933 results show the urgent need for more extensive phylogenetic studies within Drepanogynini.
934 *Thenopa* and *Sphingomima* are embedded within *Drepanogynis*, ~~making rendering~~ it paraphyletic,
935 but our taxon coverage is too limited to propose formal changes in this species-rich group.
936 Drepanogynini, as defined here, are distributed in sub-Saharan Africa. *Drepanogynis sensu*
937 Krüger (1997, 1998, 1999, 2014) includes over 150 species and it ranges from southern Africa to
938 Ethiopia (Krüger 2002, Vári et al. 2002), whereas the genera *Sphingomima* (10 species) and
939 *Thenopa* (4 species) occur in Central and West Africa (Scoble 1999). *Sphingomima* and *Thenopa*
940 are externally similar, so the recovered sister-group relationship in the current phylogeny analysis
941 ~~is was~~ anticipated. In the current analysis *Hebdomophruda errans* Prout, 1917 is isolated from
942 other analysed *Hebdomophruda* species (the others are included in Diptychini), highlighting the
943 need for additional research. Krüger (1997, 1998) classified the genus *Hebdomophruda* into
944 seven species groups on the basis of morphological characters, and *H. errans* group is one of
945 them (Krüger 1998). We do not describe a new genus for the taxon *errans*, nor do we combine it
946 with any genus in the Drepanogynini, highlighting its uncertain taxonomic position (*incertae*
947 *sedis*) ~~waiting for pending~~ more research. In the current analysis *Sphingomima discolucida*
948 Herbulot, 1995 is transferred from unassigned tribus combination to Drepanogynini, but
949 ~~because as~~ the type species of *Sphingomima* (*S. heterodoxa* Warren, 1899) was not analysed, we
950 do not transfer the entire genus *Sphingomima* into Drepanogynini. We highlight the uncertain
951 taxonomic position of the taxon *discolucida*, acknowledging that it may eventually be ~~combined-~~
952 ~~back to included again in~~ *Sphingomima* if the entire genus ~~is should get~~ transferred ~~into~~
953 Drepanogynini.

954

955 Diagnosis  Drepanogynini can be diagnosed by the combination of DNA data with up to 11
956 genetic markers (exemplar *Drepanogynis mixtaria* Guenée, [1858]) ArgK (GB Accession
957 number), Ca-ATPase (GB Accession number), CAD (GB Accession number), COI (GB
958 Accession number), EF1a (GB Accession number), GAPDH (GB Accession number), IDH (GB
959 Accession number), MDH (GB Accession number), Nex9 (GB Accession number), RpS5 (GB
960 Accession number) and Wingless (GB Accession number). In the light of our phylogenetic
961 results, the *Drepanogynis* group of genera, as classified earlier (Krüger 2014), is split between
962 two unrelated tribes (Drepanogynini and Diptychini). More research is needed to understand how
963 other *Drepanogynis* species and the *Drepanogynis* group of genera *sensu* Krüger (1997, 1998,
964 1999, 2014) (at least 11 genera), should be classified.

965

966 Boarmiini are the sister group to a clade that comprises Macariini, Cassymini, Abraxini
967 and Eutoeini. We found that many species currently assigned to Boarmiini are scattered
968 throughout Ennominae. Boarmiini *s. str.* are strongly supported but [are](#) technically ~~is~~-not
969 monophyletic because of a large number of genera which need to be formally transferred from
970 other tribes to Boarmiini (see Brehm et al., in prep. for Neotropical taxa and Murillo-Ramos et
971 al., in prep. for other taxa). The results are principally in concordance with Jiang et al. (2017),
972 who supported the monophyly of Boarmiini but with a smaller number of taxa.

973 The divided valva in male genitalia was suggested as a synapomorphy of Macariini +
974 Cassymini + Eutoeini by Holloway (1994). In addition, he proposed the inclusion of Abraxini in
975 Cassymini. Our findings support Holloway's suggestions; Cassymini is recovered as polyphyletic
976 and Abraxini and Eutoeini were found to be sister taxa. Synonymization of Eutoeini and
977 Cassymini with Abraxini should be considered in future studies, but the support [indices/values](#) of
978 the basal branches are too low in our hypothesis to draw final conclusions. Similar findings were
979 provided by Jiang et al. (2017) who suggested more extensive sampling to study the evolutionary
980 relationships of these tribes.

981

982 **Orthostixinae Meyrick, 1892**

983 Orthostixinae were not included in our study. Sihvonen et al. (2011) showed this
984 subfamily as deeply embedded within Ennominae, but unfortunately it was not represented by the

985type genus of the tribe. These results agree with Holloway (1996) who examined *Orthostixis*
986Hübner, [1823] and suggested the inclusion in Ennominae despite the full development of
987hindwing vein M2, the presence of a forewing areole and the very broad base of the tympanal
988ansa. We sampled the species *Naxa textilis* (Preyer, 1884) and *Orthostixis cribraria* (Hübner,
9891796) but, only three and one marker were successfully sequenced ~~from~~ for these samples,
990respectively. We included these species in the preliminary analyses but results were so unstable
991that we excluded them from the final analysis. Further research including fresh material and more
992genetic markers are needed to investigate the position of Orthostixinae conclusively.

993

994Conclusions

995This study elucidated some of the evolutionary relationships of the major groups within
996Geometridae. The monophyly of the subfamilies and the most widely accepted tribes was tested.
997We found high support for the subfamilies Larentiinae, Geometrinae and Ennominae in their
998traditional scopes. Sterrhinae also becomes monophyletic when *Ergavia*, *Ametris* and *Macrotis*,
999currently placed in Oenochrominae, are formally transferred to Sterrhinae. The concepts of
1000Oenochrominae and Desmobathrinae required major revision and, after appropriate
1001rearrangements, these groups will also form monophyletic subfamily-level entities. Archigeraeinae
1002are monophyletic with the transfer of *Dirce* and *Acalyphes* to Ennominae. We separated
1003Epidesmiinae as a new subfamily. As a result, this study proposes a higher level classification of
1004Geometridae comprising 8 monophyletic subfamilies. Moreover, we found that many tribes in the
1005different subfamilies were para- or polyphyletic. We attempted to address the needed taxonomic
1006changes, in order to favor taxonomic stability of the subfamilies and many tribes, even if in an
1007interim way, to allow applied researchers to use an updated higher taxonomic structure that better
1008reflects our current understanding of geometrid phylogeny. Further papers will be added to this
1009work and will provide a large number of furtheradditional taxonomic changes in the Geometridae
1010(see Introduction). Despite our efforts to include a very large number of new taxa to be analyzed
1011in our study, we acknowledge that many clades are still strongly under-represented. This is
1012particularly true for taxa from tropical Africa and Asia, and more detailed phylogenetic studies
1013are required including e.g. the tribes Eumeleini, Plutodini, Eutoeini, Cassymini and Abraxini. A
1014better taxon sampling in these regions will allow to draw better conclusions about phylogeny and
1015subsequent classification to reflect it. For ~~thisese~~ taxona and many tribes – old and new – we

1016 encourage morphological studies that attempt to find more apomorphies and that include a
1017 broader range of taxa.

1018

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1028

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