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Marine biodiversity from zero to a thousand meters at Clipperton Atoll (Île de La Passion), Tropical Eastern Pacific

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ABSTRACT

Clipperton Atoll (Île de La Passion) is the only atoll in the Tropical Eastern Pacific (TEP) ecoregion and, owing to its isolation, possesses several endemic species and is likely an important stepping stone between Oceania, the remainder of the TEP, including other oceanic islands and the west coast of Central America. We describe the biodiversity at this remote atoll from shallow water to depths greater than one thousand meters using a mixture of technologies (SCUBA, stereo baited remote underwater video stations, manned submersible, and deep-sea drop cameras). Seventy-four unique taxa of invertebrates were identified during our expedition. The majority (70%) of these taxa were confined to the top 400 m and consisted mostly of sessile organisms. Decapod crustaceans and black corals (Antipatharia) had the broadest depth ranges, 100-1,497 m and 58–967 m, respectively. Decapods were correlated with the deepest depths, while hard corals were correlated with the shallow depths. There were 96 different fish taxa from 41 families and 15 orders, of which 70% were restricted to depths < 200 m. While there was a decreasing trend in richness for both fish and invertebrate taxa with depth, these declines were not linear across the depth gradient. Instead, peaks in richness at \sim 200 m and \sim 750 m coincided with high turnover due to the appearance of new taxa and disappearance of other taxa within the community and is likely associated with the strong oxygen minimum zone that occurs within the region. The overall depth effect was stronger for fishes compared with invertebrates, which may reflect ecological preferences or differences in taxonomic resolution among groups. The creation of a no-take marine reserve 12 nautical miles around the atoll in 2016 will help conserve this unique and relatively intact ecosystem, which possesses high predator abundance.

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INTRODUCTION

Clipperton Atoll (Île de la Passion) is an uninhabited French island in the Tropical Eastern Pacific (TEP) ecoregion (*Spalding et al., 2007*), located 1,080 km south-west of Mexico (*Jost & Andréfouët, 2006*; Fig. 1). It is the only atoll in the TEP, and with 3.7 km² of coral-reef habitat, is the largest single coral reef in the region (*Glynn, Veron & Wellington, 1996*). The atoll has a highly eutrophic brackish-water lagoon with no outlet and a novel microbial community (*Galand et al., 2012*). Clipperton is an important location for seabirds, with the world's largest breeding colony of masked boobies (*Sula dactylatra*) and the second largest brown booby (*S. leucogaster*) colony (*Pitman, Ballance & Bost, 2005*).

The atoll has been occupied at various times by guano miners, would-be settlers, or military personnel, mostly from Mexico, which claimed it until international arbitration awarded it to France in 1931 (*Jost & Andréfouët, 2006*). It was later occupied by the US military during WWII and is now under direct authority of the Ministry of Overseas France (Ministère des Outre-mer). Despite its remoteness, Clipperton is frequented by sport fishermen, as well as tuna and shark fishermen from various nations (*Jost et al., 2015*; *Kroodsma et al., 2018*).

The TEP is separated from the central and western Pacific by the East Pacific Rise, a divergent tectonic plate boundary on the Pacific floor running north to south that coincides with the boundaries of a unique biogeographic province with a high proportion of endemic species (*Ekman, 1953; Allen, 2008*). Owing to its isolation and western location in the TEP, Clipperton is likely an important stepping stone for dispersal of marine organisms between Oceania, the TEP, including other oceanic islands and the west coast of Central America (*Emerson, 1994; Glynn, Veron & Wellington, 1996; Robertson & Allen, 1996; Lessios & Robertson, 2006*). Within the TEP, Clipperton lies within the Ocean Island Province, which consists of five oceanic islands/archipelagos and possesses more transpacific species and more highly localized endemics compared to the mainland Cortez and Panamic provinces (*Robertson & Cramer, 2009*).

Several studies have been conducted on the shallow-water marine biodiversity of Clipperton, which have revealed a relatively depauperate flora and fauna (*Allen & Robertson*, 1997; *Jost et al.*, 2015; *Jost et al.*, 2016). In 2004–2005 a French-led expedition to Clipperton documented species from a wide range of taxonomic groups including: algae (83 species), corals (23), gastropods (70), bivalves (22), decapods (95), echinoderms (28), and fishes (163) (*Salvat, Adjeroud & Charpy, 2008*). A more recent and comprehensive checklist of cartilaginous and bony fishes from Clipperton yielded 197 species from 62 families (*Fourriére et al., 2014*). Most of these shallow-water organisms have affinities with the Indo-Pacific and central Pacific regions. Few efforts, however, have been made to describe the mesophotic coral ecosystems (MCEs) and deep-water habitats of this unique atoll, or across the TEP in general.

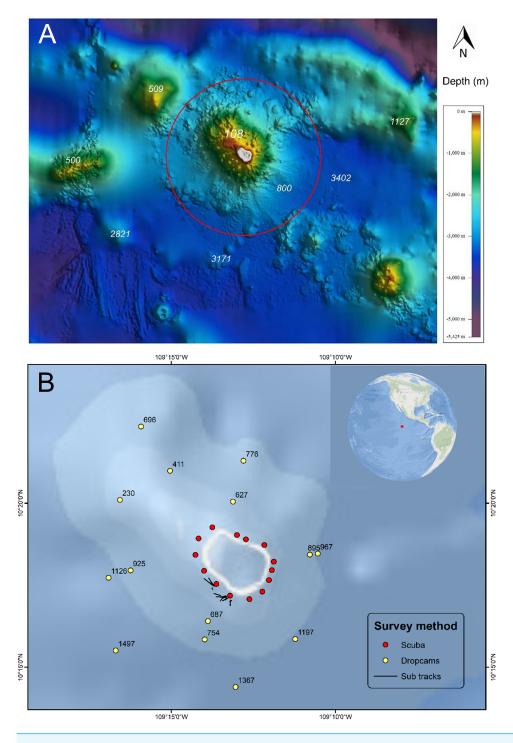


Figure 1 Bathymetry and sampling locations around Clipperton Atoll. (A) Bathymetry surrounding Clipperton Atoll. Data from French Naval Hydrographic and Oceanographic Service (SHOM: Service hydrographique et océanographique de la Marine). Red circle is 12 nautical miles no-take reserve around the atoll. (B) Locations of underwater surveys. Numbers next to dropcam points are depths (m).

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MCEs (30–150 m) are considered extensions of shallow reef communities and may greatly increase the available habitats for reef organisms, yet they have received relatively little attention owing to the difficulty in studying these habitats (*Lesser, Slattery & Leichter, 2009; Puglise et al., 2009; Hinderstein et al., 2010; Rocha et al., 2018*). Little is known about the deep-water communities within the region except for a few studies on slopes and seamounts around Isla del Coco (Costa Rica) to depths of 450 m using manned submersibles (*Cortés & Blum, 2008; Starr, Green & Sala, 2012; Starr et al., 2012*), and recent explorations at the Galápagos using Remote Operated Vehicles and manned submersibles to depths of 4,000 m (*Carey et al., 2016*).

The objective of this research was to describe the biodiversity at this remote atoll from shallow water to depths of over one thousand meters using a mixture of technologies to obtain a better understanding of the entire ecosystem. In addition, this work is intended to establish a baseline of ecosystem biodiversity and raise awareness of this unique location.

MATERIALS & METHODS

Site description

Clipperton Atoll is small (8.9 km² including the lagoon), with an emergent land surface of only 1.7 km², and a circumference of 11.8 km (*Jost, 2003*). It is surrounded by an almost continuous coral-rich fringing reef (ca. 50% live coral cover) and possesses high fish biomass (>3 t ha) (*Ricart et al., 2016*). The 50-m isobath is, on average, <500 m from shore (*Glynn, Veron & Wellington, 1996*).

Data collection

In March 2016, we used a mixture of visual systems including underwater visual censuses by SCUBA, stereo baited remote underwater video stations (BRUVS), manned submersibles, and deep-sea drop cameras to assess the biodiversity of Clipperton Atoll to depths greater than one thousand meters. We did not survey cryptic biodiversity and limited collecting to a few samples taken with the manipulator arm of the submersible. All sampling was conducted during daylight hours (\sim 7:00 to 17:00). Permission to conduct this research was granted by the Haut-Commissariat de la République en Polynésie française (NO-HC/167/CAB/BSIRI/MG).

Underwater visual census

Quantitative fish and benthic surveys were conducted at 14 sites around the atoll. Each site was surveyed at two depth strata (~10 and 20 m), amounting to 28 site ×depth combinations. Three 25-m long transects were laid along isobaths within a homogeneous habitat, with 5 m separating each fish transect. At each survey site, a scuba diver counted and sized all fishes of total length (TL) \geq 20 cm within a 4 m wide strip on an initial "swim out" while the transect line was laid (transect area = 100 m²), and all fishes of TL <20 cm were counted in a 2 m wide strip along the transect line on the way back (transect area = 50 m²). Total fish lengths were estimated to the nearest cm. All fishes were identified to the species level based on *Robertson & Allen (2015)*.

Characterization of the benthos was carried out by divers on 50 m long transects laid parallel to the coast, with two depth strata (\sim 10 and 20 m). One 50-m transect was

sampled at each depth strata. For algae, corals and other sessile macroinvertebrates, a point-intercept method was used along each transect. Species or taxa found every 20 cm on the tape measure were recorded. Echinoderms were quantified in 50×50 cm quadrats (25 random quadrats per depth stratum). Qualitative surveys of fishes and benthos were conducted at each survey site down to 40.8 m.

Stereo Baited Remote Underwater Video Stations (s-BRUVS)

A stereo-video system consisting of two high definition cameras (GoPro Hero 4+) in waterproof housings (SeaGIS, Australia) was mounted on an aluminum A-frame and deployed in 20 m (*Acuña Marrero et al., 2018*). The frame was attached to a 20 kg weight with a buoy designed to keep the video system 1–1.5 m above the seabed. A bait bag filled with \sim 1 kg of crushed tuna was attached to a 1.3 m long PVC pipe located in front of the cameras. A total of 24 BRUVS were deployed for a period of 90 min each around the perimeter of the atoll. BRUVS sampling was conducted over consecutive days from 14 to 20 March 2016, with an average number of deployments of 3.7 (±0.5 sd) per day. BRUVS were spaced at least 500 m apart to avoid overlap of individual camera bait plumes.

Manned submersible dives

The *DeepSee* is a custom-built one-atmosphere submarine, capable of carrying a pilot and two passengers to a depth of 450 m (*Cortés & Blum, 2008*). The pilot and passengers are housed in a spherical acrylic compartment that provides an unrestricted 360-degree view. The submersible is equipped with a High-Definition digital video camera and a high-intensity-discharge lighting system. Invertebrates and fishes were identified to the lowest possible taxa and attributed to their observed depths using visual observations and video recordings during each submersible dive. A few samples of gorgonians were taken between 100 and 300 m with the manipulator arm of the submersible.

Deep drop cameras

Deep Ocean Dropcams are high definition Sony Handycam HDR-XR520V 12-megapixel cameras encased in a borosilicate glass sphere and rated to a depth of 10,000 m (*Turchik et al., 2015*). Cameras were baited with \sim 1 kg of frozen fish. The viewing area per frame was between 2–6 m², depending on the slope of the substrate. Each individual organism documented within the video frame was identified to the lowest possible taxa.

Statistical methods

Taxa were classified to the lowest possible taxonomic resolution across all taxonomic groups and methodologies. For invertebrates, this was mostly at the level of order or class, although some were only identifiable to phyla. Higher taxonomic levels included those organisms of the group that we were unable to assign to a lower level taxonomic rank. Shallow-water fishes were identified to species level, while deeper water species were only identifiable to family in many cases (*McCosker & Rosenblatt, 2010; Robertson & Allen, 2015*). All fishes were reclassified to family for analyses. All data were converted to presence at each depth observed. All observations were grouped into 20-m depth bins from 10 to 320 m, followed by 20-m bins at 420, 680, 740, 760, 880, 920, 960, 1120, 1180, 1360, and 1480 m.

We used Generalized Linear Mixed Effects Models (GLMMs) with a logit link to investigate fish family and invertebrate taxa probability of occurrence according to depth. Taxa were first grouped into 20 m depth bins, and then modeled as a binomial distribution with the statistical package lme4 (*Bates et al., 2014*). Models included a random effect to allow the relationship of the response variable to vary by depth. Therefore, the magnitude of the random effect quantified how community composition varied with depth. The significance of depth as a predictor of occurrence, and the significance of the relationship varying by taxon was assessed using a Likelihood Ratio Test (LRT) with the statistical package LmerTest (*Bates et al., 2014*). All data analyses were performed in the R programing environment, Version 1.0.136 (*R Core Team, 2016*).

Principal Coordinate Analysis (PCO) was used to investigate patterns in assemblage composition by depth with biplot vectors defining correlations overlaid on the ordinations. Bray Curtis similarity matrices of presence-absence data of invertebrates and fishes grouped into 20 m depth bins were used to conduct the PCO. Vectors defined correlations between invertebrates or fishes and the ordination and were calculated based on Spearman's rank order correlation (ρ).

To investigate the change in community composition with depth, richness (number of taxa) and turnover (change in taxa) were calculated using the statistical package "codyn" in the R programing environment (*Hallett et al., 2016*). Taxon frequency of occurrence was grouped in 20 m depth bins, and three metrics of species turnover were calculated for the spatial gradient from 0–1,500 m depth: total turnover, appearances, and disappearances. Total turnover calculates the proportion of species that differ between depth points, while appearance and disappearance differentiate depths in which species appear vs. disappear, respectively (*Hallett et al., 2016*).

RESULTS

Sample allocation

There were 14 sites sampled by four divers on open-circuit scuba to a maximum depth of 40.8 m (Table 1). BRUVS were performed at 24 locations around the atoll at a depth of 20 m for a total of 36 recording hours. We conducted 16 submersible dives ranging in duration from 2:00 to 2:48 hrs (total = 33.2 hrs). Maximum depths of each submersible dive ranged from 150 m to 330 m, with an average of 251 m (sd \pm 59). Dropcams were deployed at 14 locations for 4 to 6.5 hrs (165 to 240 min of record-time), with an average record-time of 3 hrs. Deployment depths ranged from 230 m to 1,497 m ($\bar{X} = 868.2 \pm$ sd 349.8). Eight of the fourteen dropcam deployments occurred on soft bottom communities.

Invertebrates

Seventy-four different taxa of invertebrates were identified during surveys around Clipperton (Table S1). Taxa were grouped into order or the lowest possible taxonomic resolution for analyses, resulting in thirty high-level taxonomic categories. More than half of these higher-level taxa (52%) had a maximum depth of 310 m or less (Fig. 2). Most of these shallow groups had narrow depth ranges ($\bar{X} = 148.7 \text{ m} \pm 242.2 \text{ sd}$), excluding Hydrozoa, Porifera, and Alcyonacea, which had maximum depths of 776, 895, and 1,497 m,

Method	Ν	Average depth m (± sd)	Min. depth (m)	Max. depth (m)	Total hrs
Open-circuit scuba	14	19.2 (7.7)	12.2	40.8	70.0 ^a
BRUVS ^b	24	20.0	20.0	20.0	36.0
Submersible	16	250.6 (58.7)	0.0 ^c	330.0	31.1
Dropcams	14	868.2 (349.8)	230.0	1497.0	40.0

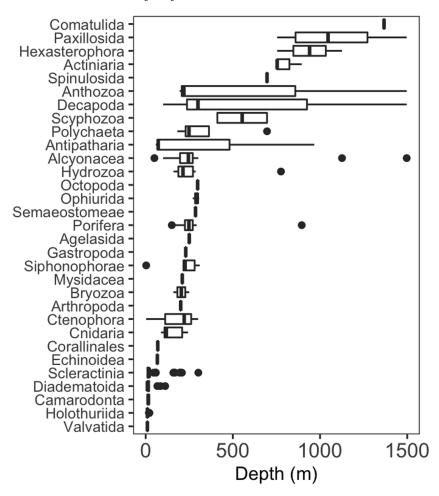
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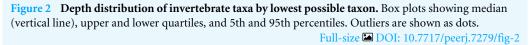
^aFour divers per station—17.5 hrs total \times 4 divers.

^bBRUVS—Baited remote underwater video stations.

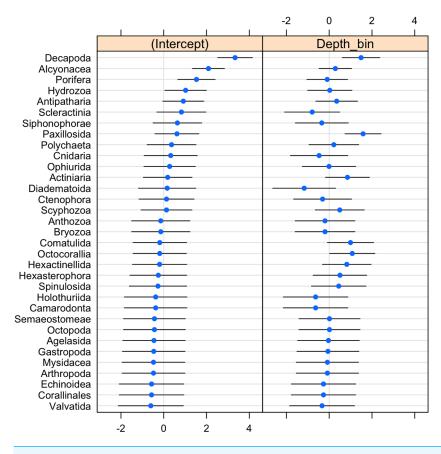
^cTaxa were counted at the beginning of the dive as the sub descended.

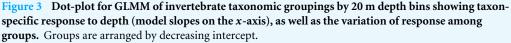
 Table 1
 Depth and total hours (hrs) for each survey method.





respectively. Decapoda and Antipatharia had the largest depth ranges, 100–1497 m and 58–967 m, respectively. Only two taxonomic groups (Hexactinellida and Comatulida) were restricted to depths >1,000 m.





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The probability of invertebrate taxa occurrence declined with increasing depth (Fig. S1, $\beta = -0.68$, se = 0.27, p = 0.01). The significant negative relationship (LRT: $\chi^2(1) = 15.64$, p < 0.001) explained 43% of the variability in these data. Community composition varied significantly with depth (LRT: $\chi^2(2) = 11.5$, p = 0.003). Decapods such as crabs, shrimps, and lobsters along with sea stars (Paxillosida) responded positively to increasing depth, while stony corals (Scleractinia) responded negatively to depth (though the variability in slope overlapped with zero) (Fig. 3).

Invertebrate taxa separated out by depth along both axes in ordination space (Fig. 4). PCO1 explained 25.8% of the variation in assemblages, with Decapoda ($\rho = -0.807$) correlated with the deepest depths. Scyphozoa and Diadematoida were correlated with a mix of depth ranges ($\rho = 0.427$ and 0.350, respectively), while hard corals (Scleractinia, $\rho = 0.366$) and siphonophores (Siphonophorae, $\rho = 0.321$) were correlated with the shallow depths. PCO2 explained 21.9% of the variation in invertebrate taxa structure, with soft corals (Alcyonacea, $\rho = -0.733$), and sponges (Porifera, $\rho = -0.656$) correlated with the intermediate depth ranges from 140 to 300 m.

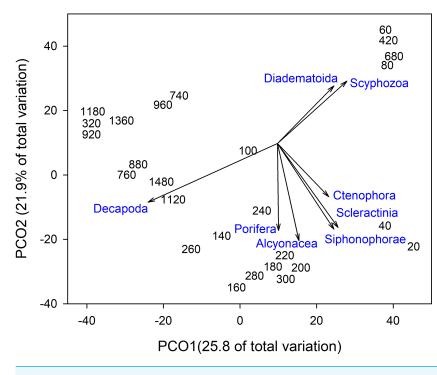


Figure 4 Principal Coordinates Analysis of invertebrate taxa presence-absence data using Bray Curtis similarity matrices of invertebrate taxa grouped into 20 m depth bins. Vectors defined correlations based on Spearman's rank order correlation between invertebrate taxa and the ordination. Full-size DOI: 10.7717/peerj.7279/fig-4

Taxonomic richness (number of taxa) for invertebrates peaked at ~ 200 m, following a relatively high turnover at ~ 100 m (Fig. 5). This turnover was due to the appearance of new taxa observed with increasing depth. Richness continued to decline with depth, with a smaller second peak at ~ 750 m. Again, this increase in richness coincided with a high turnover rate due to appearances of new taxa and the disappearance of others.

Fishes

We identified a total of 96 unique fish taxa from 41 fish families and 15 orders (Fig. 6, Table S2). The most specious families were Carangidae (n = 7), Labridae (n = 7), and Serranidae (n = 7). We report 15 new records for fishes at Clipperton, including eight new families (Scyliorhinidae, Etmopteridae, Echinorhinidae, Chimaeridae, Macrouridae, Moridae, Bythitidae, and Liparidae). Most of these new records are from deep-water species (Fig. 7, Fig. S2). Of the seven endemic fish species known from Clipperton, four were observed during our surveys. Of these, 67% were restricted to depths <200 m. Average depth ranges for fish families found in <200 m depth was 72.5 m (\pm 50.4 sd), and only slightly higher for those families in <400 m ($\bar{X} = 84.4$ m \pm 61.3 sd). Three families (Etmopteridae, and Moridae) were restricted to depths >1,000 m. Bramble sharks (Echinorhinidae) had the broadest depth distribution (230–895 m).

The probability of fish family occurrence declined with increasing depth (Fig. S3, $\beta = -6.26$, *se* = 1.12, *p* < 0.001). This negative relationship with depth was highly significant (LRT: $\chi^2[1] = 34.76$, *p* < 0.001), and explained 95% of the variability in

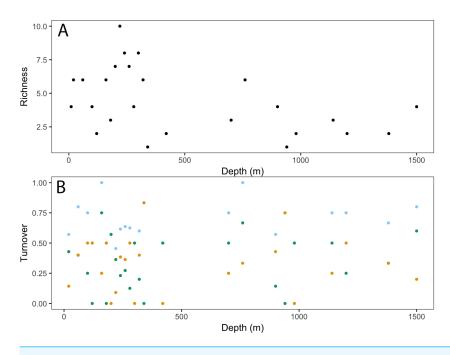


 Figure 5
 Invertebrate community patterns across depth. (A) Taxa richness; (B) turnover (total = light blue, appearances = dark green, disappearances = yellow) from 0–1,500 m depth at Clipperton Atoll.

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the data ($R^2 = 0.95$). Community composition also varied significantly with depth (LRT: $\chi^2[2] = 147.02$, p < 0.001). The magnitude of variation was 18.575 (se = 4.31) on a logit scale, indicating that the community composition changed entirely in terms of which fish families were present depending on depth.

The scorpionfishes (Scorpaenidae), grenadiers (Macrouridae), chimaeras (Chimaeridae), catsharks (Scyliorhinidae), and morids (Moridae) increased significantly in probability of occurrence with depth (Fig. 8). The majority (68%) of the fish families had a negative relationship to increasing depth, with requiem sharks (Carcharhinidae), jacks (Carangidae), groupers (Serranidae), wrasses (labridae), and tunas and mackerels (Scombridae) showing the strongest relationships. However, except for the five deep-water families noted above, the depth distributions of all other families were highly variable as evidenced by the modelled slopes overlapping with zero.

Analysis of fish assemblages showed clear separation in ordination space, with three major depth clusters (20–140, 160–420, 760–1,500, Fig. 9). PCO1 explained 35.8% of the variation in assemblages, with grenadiers (Macrouridae, $\rho = -0.698$) and chimeras (Chimaeridae, $\rho = -0.548$) correlated with the deepest depth cluster, and scorpionfishes (Scorpaenidae) correlated with the middle depth stratum ($\rho = 0.872$). PCO2 explained 28.7% of the variation in fish assemblage structure, with jacks (Carangidae, $\rho = 0.815$), groupers (Serranidae, $\rho = 0.771$), sharks (Carcharhinidae, $\rho = 0.766$), and wrasses (Labridae, $\rho = 0.746$) correlated with the shallow depth cluster. This shallow cluster was somewhat disconcordant, with the deeper depth bins separated from the shallower ones.

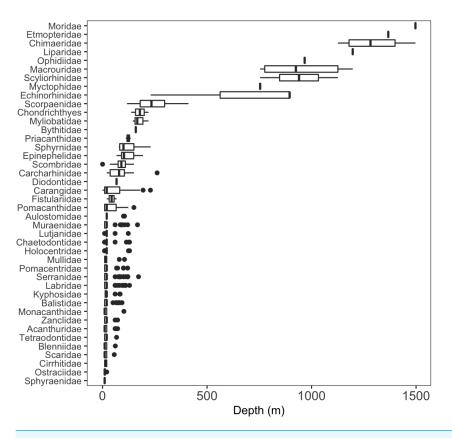


Figure 6 Depth distributions of fish families. A few sharks were only identifiable to class = Chondrichthyes.

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Fish assemblage richness declined rapidly with depth, with slight increases at \sim 100–200 m, \sim 750 m, and at \sim 1,200 m (Fig. 10).

DISCUSSION

This study represents the first examination of the biodiversity at a remote atoll from shallow-water to depths greater than one thousand meters using a mixture of survey methods and technologies. While there is a long history of research conducted at Clipperton (*Snodgrass & Heller, 1904; Hertlein et al., 1957*), most studies have focused on the shallow-water ecosystems (*Glynn, Veron & Wellington, 1996; Jost et al., 2015; Fourriére et al., 2014; Ricart et al., 2016*). As a result, the MCE and deep-water habitats of the atoll had not been well described prior to our expedition. Our integrated approach sheds new light on the distribution of marine communities across a large depth gradient at the only coral atoll in the TEP and represents one of the first studies to integrate multiple methods to characterize an ecosystem from nearshore depths to nearly 1,500 m.

While there is a decreasing trend in taxa richness for both fishes and invertebrates with increasing depth, these declines are not linear across the depth gradient. Instead, peaks in richness were observed at \sim 200 m and 750 m, which also coincided with higher assemblage

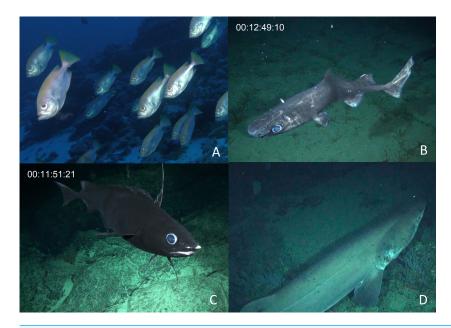
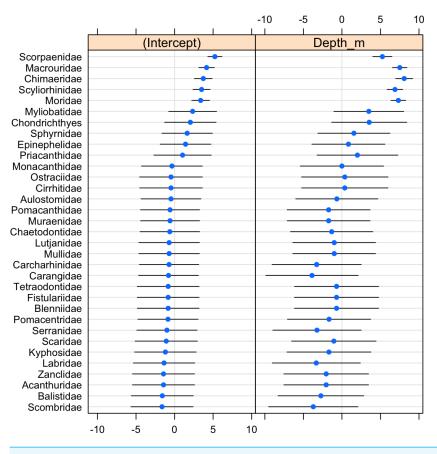
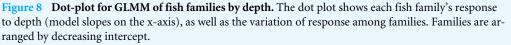


Figure 7 New fish taxa recorded at Clipperton. (A) Cookeolus japonicus (135 m), (B) Etmopterus benchley i (1,367), C. Antimora sp. (1,497 m), and D. Echinorhinus cookei (895 m). Full-size DOI: 10.7717/peerj.7279/fig-7

turnovers at these depths. Fish assemblages showed three major depth clusters (20-140 m, 160-420 m, 760-1,500 m), and these align somewhat with observed patterns of richness.

The overall depth effect was stronger for fish families than for invertebrate taxonomic groups, which may reflect physiological constrains (i.e., the influence of the oxygenminimum zones - OMZ), substrate differences, ecological preferences, or the differences in taxonomic resolution owing to our limited ability to identify taxa from visual methods, particularly for deeper water taxa, where taxonomy is typically poorly resolved (Figs. S1 and S3). Although little is known about the biodiversity of deep-sea ecosystems of the TEP, at Isla del Coco National Park in Costa Rica, sharp faunal changes were reported at relatively shallow depths (~50 m) during quantitative submersible dives to 450 m (*Cortés & Blum*, 2008). Similarly, underwater surveys of reef fish assemblages and coral communities in the Pacific and western Atlantic showed high dissimilarity between shallow (altiphotic) and mesophotic depth strata (Rocha et al., 2018). Olavo et al. (2011) proposed a faunal corridor for species associated with deep reef formations along the shelf-edge zone (40–200 m) of the South American continental margin, connecting the south-western Atlantic and the Caribbean provinces. In the Caribbean, submersible-based surveys revealed a unique reef-fish assemblage below the mesophotic zone between ~130 and 309 m (rariphotic zone) that was taxonomically distinct from shallower faunas (Baldwin, Tornabene & Robertson, 2018). We observed a similar break in community structure between the mesophotic and rariphotic zones at Clipperton. Unlike Baldwin, Tornabene & Robertson (2018), who found that the rariphotic assemblage was dominated by deep representatives of typical shallow taxa, the rariphotic assemblage at Clipperton was dominated almost exclusively by scorpaenids. Our sampling was conducted at a much broader scale than the studies noted





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above, which may have limited our ability to detect differences in community structure in the shallower depths (<140 m).

At Tristan da Cunha in the South Atlantic, *Caselle et al. (2018)* found distinct differences in the deep fish community above and below ~750 m, which was correlated with discontinuities in temperature and salinity at this depth. Similarly, characterization of the demersal fishes on the continental slopes of New Zealand using baited cameras also found that species richness peaked in shallow water, followed by a decrease beyond 100 m to a stable average from 700 to 1,200 m (*Zintzen et al., 2012*). The regional-scale distribution of benthic invertebrate megafauna along the Western Australian continental shelf down to >1,000 m is influenced by transitions in bottom temperature and oxygen concentration, which are associated with the warmer, oxygen-poor tropical waters of the southward flowing Leeuwin Current, the northward-flowing, colder, oxygen-rich Leeuwin undercurrent at 200–400 m, and Antarctic Intermediate Water below 400 m (*Williams et al., 2010*).

The geographic and vertical distribution of many species may be restricted by the presence of oxygen-minimum zones (OMZ), which are defined as oxygen concentrations

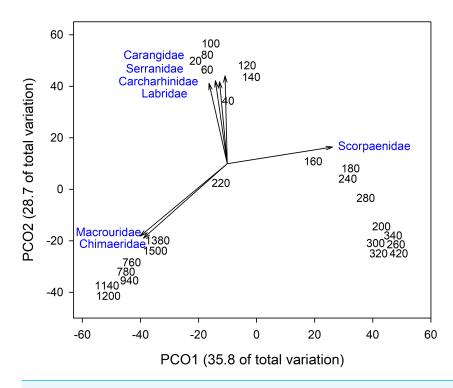


Figure 9 Principal Coordinates Analysis of fish family presence-absence data using Bray Curtis similarity matrices of fish families grouped into 20 m depth bins. Vectors defined correlations based on Spearman's rank order correlation between fish families and the ordination. Full-size DOI: 10.7717/peerj.7279/fig-9

<0.5 ml l⁻¹ (*Rogers, 2000*). Several oceanic regions around the world have large, persistent oxygen minima, including the TEP (Karstensen, Stramma & Visbeck, 2008). The OMZ has its largest westward extension in the tropical North Pacific at around 10° N, with oxygen values of $<50 \ \mu mol \ kg^{-1}$ covering almost the entire area below about 100 m depth (Karstensen, Stramma & Visbeck, 2008). The OMZ has an influence on the abundance and diversity of species, with the upper and lower boundaries of the OMZ being locations of increased biogeochemical activity. Elevated zooplankton abundances and a peak in benthic megafauna and macrofauna abundance was observed at the lower depth range of the OMZ (~740-850 m) elsewhere in the TEP (Levin, 1991; Wishner et al., 1995). There is evidence for increasing dissolved oxygen depletion and vertical expansion of the OMZ worldwide because of global climate change (Stramma et al., 2008; Hofmann & Schellnhuber, 2009; Keeling, Körtzinger & Gruber, 2010), and this could greatly impact the deep-sea biodiversity around Clipperton in the future. This present study can serve as a baseline to monitor changes in benthic biodiversity across depth gradients at Clipperton and elsewhere in the TEP. Studies such as ours are needed because some species are predicted to move deeper with increasing ocean temperatures, along with many unpredictable changes to the ecosystem.

Clipperton's shallow-water ecosystem is biogeographically unique in the TEP (*Ekman*, 1953; *Allen*, 2008). The remoteness and small area of the atoll has resulted in low genetic diversity and apparent kinship for two endemic fish species (*Crane et al.*, 2018). Of the

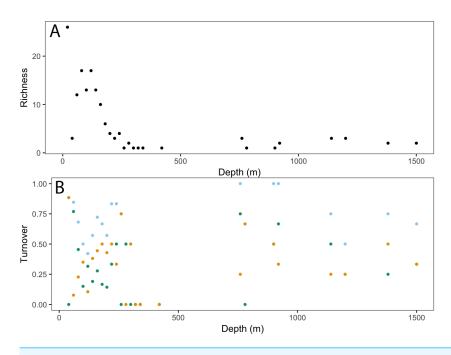


Figure 10Fish community patterns across depth: A: Species richness, B: turnover (total = light blue,
appearances = dark green, disappearances = yellow) from 0–1,500 m depth at Clipperton Atoll.
Full-size DOI: 10.7717/peerj.7279/fig-10

seven endemic fish species known from Clipperton, four were observed during our surveys. The endemic Clipperton angelfish (*Holacanthus limbaughi*) is considered a shallow-water species; however, recent submersible dives have found large aggregations (>100 individuals) at depths of 150 m (*Clua et al., 2019*), thus highlighting the connectivity between shallow and deeper water habitats. Previous studies on the shallow-water marine biodiversity of Clipperton have provided evidence to suggest that it may also be an important stepping-stone between Oceania, the TEP, and the coast of Central America (*Glynn, Veron & Wellington, 1996; Emerson, 1994; Robertson & Allen, 1996*). We report 15 new records for fishes at Clipperton, most of which are deep-water species, which have not previously been well surveyed at Clipperton. While the shallow water ecosystem at Clipperton is unique and is a potential link with other locations within the region, these deeper depths may be an important extension of this distinctive ecosystem as well.

By exploring deeper habitats, this study helps put Clipperton into the larger context of biodiversity value, especially as a link to other isolated coral reef ecosystems in the Eastern Pacific. The olive grouper, *Epinephelus cifuentesi*, was only described in 1993 and is a deep-water species (40–135 m) with a range restricted to the tip of Baja, southern Mexico to Ecuador, Galápagos Islands, Isla del Coco, Revillagigedo, and Alijos rocks (*Craig, Sadovy de Mitcheson & Heemstra, 2011; Rocha et al., 2013*). Olive grouper were once a major component of the deep-water grouper fishery in the Galápagos, but stocks have declined dramatically there due to overfishing and the species is currently rare elsewhere in its range, leading to its listing as Near Threatened by IUCN (*Schiller et al., 2015*). Large olive groupers were relatively abundant and occurred on 44% of our submersible dives at Clipperton. This is the first record for this species at Clipperton and its prevalence at the atoll may suggest an important refuge and potential source population for this overexploited and rare species.

Clipperton is also intriguing from the perspective of the maintenance of biodiversity on a small and remote island. From a conservation perspective, small and remote islands like Clipperton are predicted to suffer a greater loss of biodiversity if immigration is low or reduced, even if niche mechanisms are intact (*Chisholm et al., 2016*). Therefore, changes in oceanographic conditions can increase extinction risks for species with restricted ranges and/or low densities, as is typical for the ecosystems of small isolated islands such as Clipperton. The fire coral (*Millepora exaesa*) was previously reported from Clipperton (*Glynn, Veron & Wellington, 1996*; *Charpy, 2009*) but was absent from our surveys, highlighting the episodic nature of the ecosystem.

The Eastern Pacific Barrier greatly limits connectivity between the TEP and the central Pacific and the long-term persistence of populations depends on local recruitment within the region (*Romero-Torres et al., 2018*). Although bidirectional dispersal pathways exist between Clipperton and the Line Islands, and to a lesser extent Hawai'i, this low frequency and weak connection suggest that populations are more vulnerable to disturbance than previously thought (*Romero-Torres, Acosta & Treml, 2017; Romero-Torres et al., 2018*). The maintenance of stepping stones like Clipperton is critical for the preservation of long-distance connectivity across the TEP and with the Central Pacific (*Romero-Torres et al., 2018*).

Deep-sea mineral companies are particularly interested in the exploration and exploitation of the rich mineral deposits of the Clarion-Clipperton Fracture Zone (CCZ), adjacent to Clipperton's EEZ (*Wedding et al., 2013*; *Wedding et al., 2015*; *Voosen, 2019*). France has not yet exploited the deep minerals in Clipperton's EEZ; however, a French-Mexican oceanographic survey in 1997 found polymetallic nodules within the EEZ (*Jost et al., 2015*). These deep-sea communities have low productivity, delicate biogenic habitat structure, and very low recovery rates from physical disturbance, and as such are particularly vulnerable to human influences (*Smith & Demopoulos, 2003; Smith et al., 2008a; Smith et al., 2011*).

During our expedition, we observed abandoned longline gear on every dive, as well as line, nets, rafts, radio beacons and fish aggregation devices (FADs) scattered along the beach. FADs have commonly washed ashore at Clipperton, with at least 14 in 2015, and 5 in 2016 (*Jost et al., 2016*). Due to absence of a permanent French presence on the island and only intermittent patrols, authorized (48 Mexican ships in 2017) and non-authorized seiners and longliners are frequently seen fishing around Clipperton. The near absence of adult sharks observed during our expedition (*Jost et al., 2016*), which included 36 h of baited camera surveys (BRUVS), is likely the result of both targeted fishing and bycatch interactions. While early reports refer to the abundance of sharks at Clipperton (*Limbaugh, 1963; Skaggs, 1989*), *Allen & Robertson (1997*) reported that they were surprising uncommon in 1994 and noted that intensive fishing had removed \sim 2,000 sharks from the area the previous year.

Another indicator of extensive fishing pressure around Clipperton is the drastic decline in the population of masked boobies in the past decade from 110,000 to 40,000 individuals

(*Jost et al., 2016*). Seabirds depend on tuna to drive baitfish to the surface for them to feed. Local reductions in tuna stocks force seabirds to forage farther from shore, expending additional energy and making them more vulnerable to unregulated high seas fisheries. Masked boobies at Clipperton are now observed to forage more than three times farther offshore (430 nautical miles) (*Jost et al., 2015*), compared with ten years prior (*Weimerskirch et al., 2009*). The impacts of illegal visitation by sport fishers and divers is largely unknown. The endemic Clipperton angelfish (*Holacanthus limbaughi*) is highly prized in the aquarium fish trade and can be illegally sold for several thousand dollars in US markets and up to \$10,000 in Asian markets (*Clua et al., 2019*), making this illegal fishery very lucrative.

Clipperton is important in the pathways of numerous highly migratory species (e.g., tuna, sharks, seabirds, billfish, marine mammals) (*Jost, 2005; Jost et al., 2015*). Despite its isolation, small size, and location relative to the great East Pacific Barrier, there is connectivity reported across these oceanographic barriers, which makes Clipperton of high scientific interest as a likely important stepping-stone of connectivity between the east and west tropical Pacific for some species (*Lessios & Robertson, 2006*). The presence of widely distributed Indo-Pacific zooxanthellate corals at Clipperton indicates that these NE Pacific Islands probably serve as a stepping stone for dispersal into the far eastern Pacific region (*Glynn, Veron & Wellington, 1996*).

The deeper depths at Clipperton Atoll may serve as a refuge from increasing water temperatures associated with climate change. *Glynn, Veron & Wellington (1996)* noted that that the coral reef community at Clipperton extends down to ca. 70 m and these deeper depths may allow thermally sensitive coral species to endure increasing shallow sea temperatures and help to recolonize shallow areas once environmental conditions become more favorable (*Bridge et al., 2013; Smith et al., 2014*). Since top predators (e.g., sharks, groupers, snappers, jacks) utilize a range of depths and habitats, deep-water refugia only provide partial protection for these species (*Rocha et al., 2018*). In addition, signs of heavy fishing pressure, sedimentation, coral bleaching, and invasive species have been observed on MCEs in the Pacific and Caribbean (*Rocha et al., 2018*). While species composition can be distinct among depth zones, the overall connectivity of the ecosystem from shallow to deep necessitates a whole-ecosystem approach to management (*Baldwin, Tornabene & Robertson, 2018*).

The recent creation of a no-take reserve 12 nautical miles around the atoll in November 2016 will help conserve the entire ecosystem from fishing, landing, and other external threats. There are currently 57 Mexican fishing boats (47 purse seiners and 10 longliners) authorized to fish within the French EEZ surrounding Clipperton, but outside of the recently created 12 nautical mile reserve.

CONCLUSIONS

The results of this study highlight the unique marine ecosystem of Clipperton Atoll, particularly the deep-sea environments, which have been poorly studied prior to our expedition. The mixture of technologies, spanning a broad depth range, provided for an integrated examination of the biodiversity of this isolated atoll in the TEP. The relatively

low diversity, high endemism and unique biogeography of Clipperton is of high scientific interest as it is likely an important stepping-stone of connectivity between the east and west tropical Pacific for some species. Its remoteness has also resulted in substantial illegal, unreported and under-reported fishing, which appears to have negative impacts on the overall health of the ecosystem. To help restore populations of sharks, tunas, and other pelagic species, there is an urgent need for better management of the entire EEZ, which could include a better vessel monitoring system, satellite-based vessel monitoring and surveillance, expansion of the existing 12 nautical mile protected area, and a greater research presence.

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ADDITIONAL INFORMATION AND DECLARATIONS

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Competing Interests

Shmulik Blum is employed by DeepSee, UnderSea Hunter Group.

Author Contributions

- Alan M. Friedlander and Enric Ballesteros conceived and designed the experiments, performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Jonatha Giddens performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Shmulik Blum performed the experiments, contributed reagents/materials/analysis tools, authored or reviewed drafts of the paper, approved the final draft.
- Eric K. Brown, Jennifer E. Caselle, Christian Jost and Pelayo Salinas-de-León conceived and designed the experiments, performed the experiments, contributed reagents/materials/analysis tools, authored or reviewed drafts of the paper, approved the final draft.
- Bradley Henning performed the experiments, contributed reagents/materials/analysis tools, authored or reviewed drafts of the paper, approved the final draft.
- Enric Sala conceived and designed the experiments, contributed reagents/materials/analysis tools, authored or reviewed drafts of the paper, approved the final draft.

Field Study Permissions

The following information was supplied relating to field study approvals (i.e., approving body and any reference numbers):

Permission to conduct this work was provided by the Haut-Commissariat de la Republique en Polynesie Francaise (HC167CABBSIRIMG).

Data Availability

The following information was supplied regarding data availability:

The raw data is available as a Supplemental File. The raw data provides density estimates (number per meter squared) for all taxa observed using underwater surveys, submersibles, and deep drop cameras. Data include date, latitude and longitude.

Supplemental Information

Supplemental information for this article can be found online at http://dx.doi.org/10.7717/ peerj.7279#supplemental-information.

REFERENCES

Acuña Marrero D, Smith AN, Salinas-de León P, Harvey ES, Pawley MD, Anderson MJ. 2018. Spatial patterns of distribution and relative abundance of coastal shark species in the Galapagos Marine Reserve. *Marine Ecology Progress Series* 593:73–95 DOI 10.3354/meps12505.

- Allen GR. 2008. Conservation hotspots of biodiversity and endemism for Indo-Pacific coral reef fishes. *Aquatic Conservation: Marine and Freshwater Ecosystems* 18:541–556 DOI 10.1002/aqc.880.
- Allen GR, Robertson DR. 1997. An annotated checklist of the fishes of Clipperton Atoll, tropical eastern Pacific. *Revista de Biología Tropical* 45:813–843.
- Baldwin CC, Tornabene L, Robertson DR. 2018. Below the mesophotic. *Scientific Reports* 8:4920 DOI 10.1038/s41598-018-23067-1.
- **Bates D, Maechler M, Bolker B, Walker S. 2014.** Fitting linear mixed-effects models using lme4. *Journal of Statistical Software* **67**:1–48.
- Bridge TC, Hughes TP, Guinotte JM, Bongaerts P. 2013. Call to protect all coral reefs. *Nature Climate Change* 3:528–530 DOI 10.1038/nclimate1879.
- Carey S, Fisher CR, Salinas de León P, Roman C, Raineault NA, Suarez J, Smart C, Kane R, Tuzun S, Balcanoff J, Lubetkin M, Jones M, Schwartz D, Fornari D, Soule A, Wanless D, Watling L, Ballard RD. 2016. Exploring the undersea world of the Galápagos Islands. *Oceanography Magazine* 29:32–34 DOI 10.5670/oceanog.2016.07.
- Caselle JE, Hamilton SL, Davis K, Thompson CD, Turchik A, Jenkinson R, Simpson D, Sala E. 2018. First quantification of subtidal community structure at Tristan da Cunha Islands in the remote South Atlantic: from kelp forests to the deep sea. *PLOS ONE* 13(3):e0195167 DOI 10.1371/journal.pone.0195167.
- **Charpy L. 2009.** *Clipperton, environnement et biodiversité d'un microcosme océanique.* Paris: Museum National d'Histoire Naturelle.
- Chisholm RA, Fung T, Chimalakonda D, O'Dwyer JP. 2016. Maintenance of biodiversity on islands. *Proceedings of the Royal Society B: Biological Sciences* 283:20160102 DOI 10.1098/rspb.2016.0102.
- **Clua E, Bessudo S, Caselle JE, Chauvet C, Jost C, Friedlander AM. 2019.** Population assessment of the endemic angelfish *Holacanthus limbaughi* from Île de La Passion-Clipperton Atoll (Tropical Eastern Pacific). *Endangered Species Research* **38**:171–176 DOI 10.3354/esr00947.
- **Cortés J, Blum S. 2008.** Life to 450 m depth at Isla del Coco, Costa Rica. *Revista de Biología Tropical* **56**:189–206.
- **Craig MT, Sadovy de Mitcheson YJ, Heemstra PC. 2011.** *Groupers of the world*. Grahamstown: NISC.
- Crane NL, Tariel J, Caselle JE, Friedlander AM, Robertson DR, Bernardi G. 2018. Clipperton Atoll as a model to study small marine populations: endemism and the genomic consequences of small population size. *PLOS ONE* **13(6)**:e0198901 DOI 10.1371/journal.pone.0198901.
- Ekman S. 1953. Zoogeography of the sea. London: Sidgwick and Jackson Ltd.
- **Emerson WK. 1994.** A zoogeographic summary of the marine mollusks of Clipperton Island (tropical eastern Pacific Ocean). *The Festivus* **26**:62–71.
- **Fourriére M, Reyes-Bonilla H, Rodríguez-Zaragoza FA, Crane N. 2014.** Fishes of Clipperton Atoll, Eastern Pacific: checklist, endemism, and analysis of completeness of the inventory. *Pacific Science* **68**:375–395 DOI 10.2984/68.3.7.

- Galand PE, Bourrain M, De Maistre E, Catala P, Desdevises Y, Elifantz H, Kirchman DL, Lebaron P. 2012. Phylogenetic and functional diversity of Bacteria and Archaea in a unique stratified lagoon, the Clipperton atoll (N Pacific). *FEMS Microbiology Ecology* **79**:203–217 DOI 10.1111/j.1574-6941.2011.01209.x.
- Glynn PW, Veron JE, Wellington GM. 1996. Clipperton Atoll (eastern Pacific): oceanography, geomorphology, reef-building coral ecology and biogeography. *Coral Reefs* 15:71–99 DOI 10.1007/BF01771897.
- Hallett LM, Jones SK, MacDonald AA, Jones MB, Flynn DF, Ripplinger J, Slaughter P, Gries C, Collins SL. 2016. Codyn: an R package of community dynamics metrics. *Methods in Ecology and Evolution* 7:1146–1151 DOI 10.1111/2041-210X.12569.
- Hertlein LG, Emerson WK, Fisher RL, Jordan JB, O'Neil S. 1957. Additional notes on the invertebrate fauna of Clipperton Island. *American Museum Novitates* 1859:1–9.
- Hinderstein LM, Marr JC, Martinez FA, Dowgiallo MJ, Puglise KA, Pyle RL, Zawada DG, Appeldoorn R. 2010. Theme section on Mesophotic coral ecosystems: characterization, ecology, and management. *Coral Reefs* 29:247–251 DOI 10.1007/s00338-010-0614-5.
- Hofmann M, Schellnhuber HJ. 2009. Oceanic acidification affects marine carbon pump and triggers extended marine oxygen holes. *Proceedings of the National Academy of Sciences of the United States of America* 106:3017–3022 DOI 10.1073/pnas.0813384106.
- Jost C. 2003. Clipperton Ile de la Passion: une aire française du Pacifique à protéger. In: Lebigre J-M, Decoudras P-M, eds. *Les aires protégées insulaires et littorales tropicales Actes du Colloque <Aires protégées insulaires et littorales tropicales>, Nouméa (Nouvelle-Calédonie)*. Bordeaux: Presses Univ de Bordeaux, 223–243.
- Jost C. 2005. Risques environnementaux et enjeux à Clipperton (Pacifique français). *Cybergeo: European Journal of Geography* 1:314.
- Jost CH, Andréfouët S. 2006. Long-term natural and human perturbations and current status of Clipperton Atoll, a remote island of the Eastern Pacific. *Pacific Conservation Biology* 12:207–217 DOI 10.1071/PC060207.
- Jost C, Friedlander A, Arribas J, Ballesteros E, Betz J, Brown E, Caselle J, Henning B, Hoyos M, Salinas de Leon P, Rose P, Thompson C, Sala E. 2016. L'atoll de Clipperton (île de La Passion): Biodiversité, menaces et recommandations pour sa conservation. Rapport au Gouvernement de la France. Washington D.C./UPF: Août, National Geographic Society—Pristine Seas Tahiti, Polynésie française. 98 pages.
- Jost C, Solis-Weiss V, Dumas P, Sanvicente Añorve L, Gravier-Bonnet N, Jost Xenia , Grellier M, Arakino M, Duchauchoy A, Tchékémian A, Lemus E. 2015. Clipperton Island protection and valorisation. Rapport de l'expédition scientifique internationale PASSION 2015 à Monsieur le Président de la république française, UPF/UNC/ULR/UNAM/ AFD/MAEDI/MOM, nov. 2015. Paris, Tahiti, 90 pages.
- Karstensen J, Stramma L, Visbeck M. 2008. Oxygen minimum zones in the eastern tropical Atlantic and Pacific oceans. *Progress in Oceanography* 77:331–350 DOI 10.1016/j.pocean.2007.05.009.

- Keeling RF, Körtzinger A, Gruber N. 2010. Ocean deoxygenation in a warming world. *Annual Reviews in Marine Science* 2:199–229 DOI 10.1146/annurev.marine.010908.163855.
- Kroodsma DA, Mayorga J, Hochberg T, Miller NA, Boerder K, Ferretti F, Wilson A, Bergman B, White TD, Block BA, Woods P, Sullivan B, Costello C, Worm B. 2018. Tracking the global footprint of fisheries. *Science* 359(6378):904–908 DOI 10.1126/science.aao5646.
- Lesser MP, Slattery M, Leichter JJ. 2009. Ecology of mesophotic coral reefs. *Journal of Experimental Marine Biology and Ecology* 375:1–8 DOI 10.1016/j.jembe.2009.05.009.
- Lessios HA, Robertson DR. 2006. Crossing the impassable: genetic connections in 20 reef fishes across the eastern Pacific barrier. *Proceedings of the Royal Society B: Biological Sciences* 273:2201–2208 DOI 10.1098/rspb.2006.3543.
- Levin LA. 1991. Interactions between metazoans and large, agglutinating protozoans: implications for the community structure of deep-sea benthos. *American Zoologist* 31:886–900 DOI 10.1093/icb/31.6.886.
- Limbaugh C. 1963. Field notes on sharks. In: Gilbert PW, ed. *Sharks and survival*. Boston: D.C. Heath and Co, 63–94.
- McCosker JE, Rosenblatt RH. 2010. The fishes of the Galápagos Archipelago: an update. *Proceedings of the California Academy of Sciences* 61:167–195.
- Miljutin DM, Miljutina MA, Arbizu PM, Galéron J. 2011. Deep-sea nematode assemblage has not recovered 26 years after experimental mining of polymetallic nodules (Clarion-Clipperton Fracture Zone, Tropical Eastern Pacific). *Deep Sea Research Part I: Oceanographic Research Papers* 58:885–897 DOI 10.1016/j.dsr.2011.06.003.
- **Olavo G, Costa PA, Martins AS, Ferreira BP. 2011.** Shelf-edge reefs as priority areas for conservation of reef fish diversity in the tropical Atlantic. *Aquatic Conservation: Marine and Freshwater Ecosystems* **21**:199–209.
- Pitman RL, Ballance LT, Bost C. 2005. Clipperton Island: pig sty, rat hole and booby prize. *Marine Ornithology* 33:193–194.
- Puglise K, Hinderstein L, Marr JCA, Dowgiallo MJ, Martinez FA. 2009. Mesophotic coral eco-systems research strategy: international workshop to prioritize research and management needs for mesophotic coral ecosystems, Jupiter, Florida, 12-15 2008. NOAA Technical Memorandum NOS NCCOS 98 and NOAA OER 2. NOAA/National Centers for Coastal Ocean Science, Silver Spring.
- **R Core Team. 2016.** R: a language and environment for statistical computing. Vienna: R Foundation for Statistical Computing. *Available at https://www.R-project.org/*.
- Ricart AM, Rodríguez-Zaragoza FA, González-Salas C, Ortiz M, Cupul-Magaña AL, Adjeroud M. 2016. Coral reef fish assemblages at Clipperton Atoll (Eastern Tropical Pacific) and their relationship with coral cover. *Scientia Marina* 80:479–486 DOI 10.3989/scimar.04301.12B.
- **Robertson DR, Allen GR. 1996.** Zoogeography of the shorefish fauna of Clipperton Atoll. *Coral Reefs* **15**:121–131 DOI 10.1007/BF01771902.
- **Robertson DR, Allen GR. 2015.** Shorefishes of the tropical Eastern Pacific: online information system. Version 2.0. Balboa: Smithsonian Tropical Research Institute.

Available at http://biogeodb.stri.si.edu/sftep/en/pages (accessed on 21 December 2018).

- Robertson DR, Cramer KL. 2009. Shore fishes and biogeographic subdivisions of the Tropical Eastern Pacific. *Marine Ecology Progress Series* 380:1–7 DOI 10.3354/meps07925.
- Rocha L, Ferreira B, Choat JH, Craig M, Sadovy Y. 2013. *Epinephelus cifuentesi*. IUCN Red List of Threatened Species, Version 2013.2. *Available at http://www.iucnredlist. org* (accessed on 12 August 2018).
- Rocha LA, Pinheiro HT, Shepherd B, Papastamatiou YP, Luiz OJ, Pyle RL, Bongaerts P. 2018. Mesophotic coral ecosystems are threatened and ecologically distinct from shallow water reefs. *Science* 361:281–284 DOI 10.1126/science.aaq1614.
- **Rogers AD. 2000.** The role of the oceanic oxygen minima in generating biodiversity in the deep sea. *Deep Sea Research Part II: Topical Studies in Oceanography* **47**:119–148 DOI 10.1016/S0967-0645(99)00107-1.
- Romero-Torres M, Acosta A, Treml EA. 2017. The regional structure of spawning phenology and the potential consequences for connectivity of coral assemblages across the Eastern Tropical Pacific. *ICES Journal of Marine Science* 74:613–624 DOI 10.1093/icesjms/fsw218.
- **Romero-Torres M, Treml EA, Acosta A, Paz-García DA. 2018.** The Eastern Tropical Pacific coral population connectivity and the role of the Eastern Pacific Barrier. *Scientific Reports* **8**:9354 DOI 10.1038/s41598-018-27644-2.
- Salvat B, Adjeroud M, Charpy L. 2008. Les récifs coralliens de Clipperton. *Revue d'écologie*, 2008, 1-2 "Les récifs coralliens de l'outre-mer français: suivi et état des lieux" 63(1–2):179–187.
- Schiller L, Alava JJ, Grove J, Reck G, Pauly D. 2015. The demise of Darwin's fishes: evidence of fishing down and illegal shark finning in the Galápagos Islands. *Aquatic Conservation: Marine and Freshwater Ecosystems* 25:431–446 DOI 10.1002/aqc.2458.
- **Skaggs JM. 1989.** *Clipperton, a history of the island the world forgot.* New York: Walker and Company.
- Smith CR, De Leo FC, Bernardino AF, Sweetman AK, Arbizu PM. 2008a. Abyssal food limitation, ecosystem structure and climate change. *Trends in Ecology and Evolution* 23:518–528 DOI 10.1016/j.tree.2008.05.002.
- **Smith CR, Demopoulos AWJ. 2003.** The deep Pacific Ocean floor. In: Tyler PA, ed. *Ecosystems of the world: ecosystems of the deep ocean*. Vol. 28. Amsterdam: Elsevier, 179–218.
- Smith TB, Glynn PW, Maté JL, Toth LT, Gyory J. 2014. A depth refugium from catastrophic coral bleaching prevents regional extinction. *Ecology* 95:1663–1673 DOI 10.1890/13-0468.1.
- Smith CR, Levin LA, Koslow A, Tyler PA, Glover AG. 2008b. The near future of deep seafloor ecosystems. In: Polunin NVC, ed. *Aquatic ecosystems: trends and global prospects*. Cambridge: Cambridge University Press, 334–351.

- Snodgrass RE, Heller E. 1904. Papers from the Hopkins-Stanford Galapagos Expedition, 1898-1899. XVII. Shore fishes of the Revillagigedo, Clipperton, Cocos and Galapagos Islands. *Proceedings of the Washington Academy of Sciences* 6:333–427.
- Spalding MD, Fox HE, Allen GR, Davidson N, Ferdaña ZA, Finlayson MA, Halpern B, Jorge MA, Lombana A, Lourine SA, Martin KD, McManus E, Molnar J, Recchia CA, Robertson J. 2007. Marine ecoregions of the world: a bioregionalization of coastal and shelf areas. *Bioscience* 57:573–583 DOI 10.1641/B570707.
- **Starr RM, Cortés J, Barnes CL, Green K, Breedy O. 2012.** Characterization of deepwater invertebrates at Isla del Coco National Park and Las Gemelas Seamount, Costa Rica. *Revista de Biología Tropical* **Suppl. 3**:303–319.
- Starr RM, Green K, Sala E. 2012. Deepwater fish assemblages at Isla del Coco National Park and Las Gemelas Seamount, Costa Rica. *Revista de Biología Tropical* Suppl. 3:347–362.
- Stramma L, Johnson GC, Sprintall J, Mohrholz V. 2008. Expanding oxygen-minimum zones in the tropical oceans. *Science* **320**:655–658 DOI 10.1126/science.1153847.
- Turchik AJ, Berkenpas EJ, Henning BS, Shepard CM. 2015. *The Deep Ocean Dropcam: a highly deployable benthic survey tool*. Washington, D.C.: OCEANS 15 MTS/IEEE Washington, 1–8 DOI 10.23919/OCEANS.2015.7401978.
- Voosen P. 2019. Scheme to mine the abyss gets sea trial. *Science* 363:1129–1130 DOI 10.1126/science.363.6432.1129.
- Wedding LM, Friedlander AM, Kittinger JN, Watling L, Gaines SD, Bennett M, Hardy SM, Smith CR. 2013. From principles to practice: a spatial approach to systematic conservation planning in the deep sea. *Proceedings of the Royal Society B: Biological Sciences* 280:20131684 DOI 10.1098/rspb.2013.1684.
- Wedding LM, Reiter SM, Smith CR, Gjerde KM, Kittinger JN, Friedlander AM, Gaines SD, Clark MR, Thurnherr AM, Hardy SM, Crowder LB. 2015. Managing mining of the deep seabed. *Science* 349:144–145 DOI 10.1126/science.aac6647.
- Weimerskirch H, Le Corre M, Bost C-A, Ballance LT, Pitman RL. 2009. L'avifaune et l'écologie des oiseaux marins. In: Charpy L, ed. *Clipperton, environnement et biodiversité d'un microcosme océanique*. Vol. 68. Paris: Muséum National d'Histoire Naturelle; IRD, Marseille, Patrimoines naturels, 381–392.
- Williams A, Althaus F, Dunstan PK, Poore GC, Bax NJ, Kloser RJ, McEnnulty FR. 2010. Scales of habitat heterogeneity and megabenthos biodiversity on an extensive Australian continental margin (100–1100 m depths). *Marine Ecology* 31:222–236 DOI 10.1111/j.1439-0485.2009.00355.x.
- Wishner KF, Ashjian CJ, Gelfman C, Gowing MM, Kann L, Levin LA, Mullineaux LS, Saltzman J. 1995. Pelagic and benthic ecology of the lower interface of the Eastern Tropical Pacific oxygen minimum zone. *Deep Sea Research Part I: Oceanographic Research Papers* 42:93–115 DOI 10.1016/0967-0637(94)00021-J.
- Zintzen V, Anderson MJ, Roberts CD, Harvey ES, Stewart AL, Struthers CD. 2012. Diversity and composition of demersal fishes along a depth gradient assessed by baited remote underwater stereo-video. *PLOS ONE* 7(10):e48522 DOI 10.1371/journal.pone.0048522.