Peer

Differences in biomass and silica content in typical plant communities with ecotones in the Min River estuary of southeast China

Hui Gao¹, Shuijing Zhai¹, Zhigao Sun¹, Juan Liu² and Chuan Tong¹

¹ School of Geographical Sciences, Key Laboratory of Humid Subtropical Eco-geographical Process, Ministry of Education, Fujian Normal University, Fuzhou, China

² Innovation Center and Key Laboratory of Water Quality and Reservation in the Pearl River Delta, Institute of Environmental Research At Greater Bay, School of Environmental Science and Engineering, Guangzhou University, Guangzhou, China

ABSTRACT

Silica (Si) is a basic nutrient requirement for many aquatic organisms and its biogeochemical cycle plays an important role in estuarine coastal ecosystems. However, little is known about the role Si plays during plant-plant interactive processes in the marsh ecosystems. Here, variations in biomass, biogenic silica (BSi) content, and available Si content of Cyperus malaccensis-dominated marshes, Phragmites australis-dominated marshes, and their ecotonal marshes were studied in the Shanyutan marsh in the Min River estuary, China. Results showed that C. malaccensis and P. australis biomass in ecotones was lower than those in typical communities by 46.4% and 46.3%, respectively. BSi content in aboveground organs of C. malaccensis and culms and roots of P. australis was lower in ecotones than in typical communities, whereas BSi content in other organs showed the opposite trend. Biomass allocation in C. malaccensis and P. australis roots in ecotones was higher by 56.9% and 19.5%, respectively, and BSi stock in C. malaccensis and P. australis roots was higher than that in typical communities by 120.9% and 18.9%, respectively. Available Si content in ecotonal marsh soils was 12.6% greater than that in typical communities. Thus, the two plant species may use different strategies for Si accumulation and allocation in ecotones to adapt to the competitive environment. P. australis may expand primarily via occupation of wider aboveground space, thereby increasing the Si accumulation capacity in aboveground organs. Meanwhile, C. malaccensis may increase the Si allocation capacity of its roots to withstand the pressure from *P. australis*. This study will provide new insights into marsh plant competition from the perspective of Si, which can also benefit plant management in marsh ecosystems.

Subjects Soil Science, Climate Change Biology, Natural Resource Management, Biogeochemistry, Environmental Impacts

Keywords Variation, Silica, Ecotone, Competition, Marsh, Min River estuary

INTRODUCTION

Silica (Si) is considered as a beneficial element for plants (*Ma & Yamaji, 2015; Klotzbücher et al., 2018; Coskun et al., 2019*). Many terrestrial and aquatic plants accumulate high amounts of Si in their tissues (*Struyf et al., 2007; Ma & Yamaji, 2015; Carey et al., 2017*).

Submitted 7 January 2019 Accepted 31 May 2019 Published 22 July 2019

Corresponding authors Shuijing Zhai, s2008shuijing@163.com Juan Liu, liujuan@gzhu.edu.cn

Academic editor Jianhua Xu

Additional Information and Declarations can be found on page 13

DOI 10.7717/peerj.7218

Copyright 2019 Gao et al.

Distributed under Creative Commons CC-BY 4.0

OPEN ACCESS

Si is still not recognized as an essential element for plant growth; however, its beneficial effects on the development, yield, mineral nutrition, health, and survival have been observed in a variety of plant species for decades in agricultural ecosystems (*Epstein*, 1994; Datnoff, Deren & Snyder, 1997; Savant et al., 1999; Datnoff, Snyder & Korndörfer, 2001; Ma & Yamaji, 2006). During the last decade, Si has received increased attention in natural ecosystems owing to its importance in plant growth, nutrient content (Schaller et al., 2012b, 2013; Brackhage et al., 2013; Coskun et al., 2016), stoichiometry (Schaller & Struyf, 2013; Schaller et al., 2016; Schaller, Hines & Brackhage, 2017), providing rigidity to plant structures, and enhancing resistance to abiotic and biotic stresses (Epstein, 1994, 2009; Querné, Ragueneau & Poupart, 2012; Schoelynck et al., 2014). Si availability has a great influence on other nutrient content and the stoichiometry of marsh grasses during plant growth (Schaller et al., 2012b; Brackhage et al., 2013), and can influence element cycling during litter decomposition (Schaller, Hines & Brackhage, 2017). Furthermore, Si may affect the nutrient element turnover via altered plant nutrient and litter decomposition, and the trade-off between resource efficiency and plant stabilization/defense (Schaller, Brackhage & Dudel, 2012a; Schaller & Struyf, 2013; Schoelynck et al., 2014; Schaller et al., 2016). Si could play a significant role in vegetation dynamics, and the processing and functioning of ecosystems (Struyf & Conley, 2009; Schaller et al., 2012b; Schoelynck et al., 2014; Struyf et al., 2015; Carey et al., 2017; Coskun et al., 2019).

Estuarine marshes, located in the land-ocean transition zone, are some of the most sensitive ecosystems to global climate change (Simas, Nunes & Ferreira, 2001), and are considered as important areas for studying Si transport and processing (Struyf et al., 2007; Struyf & Conley, 2009; Carey & Fulweiler, 2013). The efficient use of Si by different plant species potentially influences competitiveness in dynamic environments (Struyf & *Conley*, 2009). Plants that are able to utilize Si to their benefit are favored under changing environmental conditions in estuaries at the community level (Querné, Ragueneau & *Poupart, 2012*). Si is important for plant decomposition processes, plant competitiveness, and stress tolerance (Schoelynck et al., 2014). Previous studies have been conducted to explore Si content variations of typical plant communities in estuarine marsh ecosystems (Struyf et al., 2005; Hou et al., 2010; Querné, Ragueneau & Poupart, 2012; Jacobs et al., 2013), whereas those in ecotones (i.e., the transitional zone between two plant species) remain neglected. The competition among different plant species and the ecological edge effect in ecotones are clearly obvious (Cadenasso et al., 2003; Theriot et al., 2013). However, Si cycling in ecotones and its role for plant competition in estuarine marshes has not been explained in recent studies (Schoelynck et al., 2014). As a beneficial nutrient, Si content might show more variability in ecotones compared to typical communities, and thus may provide new insight into marsh plant competition.

The Shanyutan marsh is the largest wetland in the Min River estuary in the transitional subtropical zone of southeast China. The main vegetation includes native species *Cyperus malaccensis, Phragmites australis,* and *C. compressus,* and invasive species *Spartina alterniflora,* which started to invade this marsh in 2002 (*Tong et al., 2011*). Different types of typical plant communities are distributed zonally from land to sea. The ecotones of two plant communities, that is, the transition regions gradually formed during the process

of plant expansion, are clearly distinct in this zone. *C. malaccensis* and *P. australis* in ecotones grow as vigorously as those in typical communities. Therefore, this provides a suitable area to study the differences between ecotones and typical community zones in the Min River estuary. Studies regarding Si in the Min River estuary marshes have mainly focused on soil and porewater in typical vegetation areas (*Zhai & Xue, 2016*; *Hou et al., 2017*; *Gao et al., 2018*), whereas Si cycling in transitional systems has been neglected. Previous studies have shown that biomass allocation and biogenic silica (BSi) stock in roots was higher in ecotones than in typical communities (*Gao et al., 2017*). This has important implications for understanding the mechanisms behind habitat competition of different plant species. However, the accumulation and allocation of biomass and Si content among different plant species in ecotones of plant–plant interactive processes have been overlooked.

In the present study, the differences in biomass and BSi content of *P. australis* and *C. malaccensis* during their competition were investigated, as was the available Si content of soils in the Shanyutan marsh. The objectives were to: (i) compare the variation in Si content and biomass of typical communities with that of ecotones; and (ii) discuss Si allocation variations among plant organs and between typical communities and ecotones. This would not only enhance the knowledge regarding the variations in Si content, but also help understand the potential competitiveness mechanism of native plant species from Si cycling in the estuarine marsh ecosystems.

MATERIALS AND METHODS

Study area

The present study was undertaken in the Shanyutan marsh (119°34'12"–119°40'40"E, $26^{\circ}00'36''-26^{\circ}03'42''N$), which is the largest wetland (approximately 3,120 ha) in the Min River estuary, southeast China (Liu, Zeng & Chen, 2006). The climate is relatively warm and wet, with a mean annual temperature of 19.6 °C and a mean annual precipitation of 1,300 mm. This area is featured by typical semi-diurnal tide with a mean tidal range of 4.37-4.46 m (Zhang, 1991; Dai, 2004). The marsh sediment is dominated by saline soil with low pH. The width of the Shanyutan coastal marsh (gradient $1-2^{\circ}$) is generally larger than one km (Liu, Zeng & Chen, 2006; Chen, Tang & Li, 2015). The main marsh vegetation includes three native species (P. australis, C. malaccensis, and C. compressus) and one invasive species S. alterniflora (Tong et al., 2010). In the western Shanyutan marsh, there are five typical plant communities distributed in a belt-like pattern from land to sea (Fig. 1): C. malaccensis community, P. australis community, C. malaccensis community, S. alterniflora community, and C. compressus community, with the latter two communities excluded from the present study. In this region, P. australis and C. malaccensis coexist without other species in the transitional zone, and are distributed from both C. malaccensis community to P. australis community and also from *P. australis* community to *C. malaccensis* community, with a width of approximately 50-100 m. Therefore, there were three typical communities and two distinct ecotones in our study.



Figure 1 The study area and location of the sampling sites.W1, Cyperus malaccensis marsh;W2, C. malaccensis-Phragmites australis marsh;W3, P. australis marsh;W4, P. australis-C. malaccensismarsh;W5, C. malaccensis marsh.Full-size DOI: 10.7717/peerj.7218/fig-1

Sample collection and pretreatment

In July 2015, two parallel transects perpendicular to the coastline were laid from land to sea in the high-middle flat tidal zone of the western Shanyutan marsh. The distance between the two transects was approximately 100 m, and on each transect five sampling sites were selected that included three typical community marshes and two ecotonal marshes, that is, *C. malaccensis* community (closed to land and more than 50 m away from the dam, W1), *C. malaccensis–P. australis* community (*C. malaccensis* was dominant with a relative culms density of 82.7% per square meter, W2), *P. australis* community (W3), *P. australis–C. malaccensis* community (*C. malaccensis* was dominant with a relative culms density of 66.1% per square meter, W4), and *C. malaccensis* community (closed to sea, W5). To better distinguish the same plant species among the two ecotones, we recorded *C. malaccensis* in W2 as W2-C and *P. australis* as W2-P whereas *C. malaccensis* in W4 was recorded as W4-C and *P. australis* was W4-P.

Three replicate plots (50×50 cm) were randomly chosen from each of the five sites for the collection of soil and plant samples. Soil profiles (0–40 cm) were taken with a columnar sampler (diameter 10 cm and height 80 cm) and samples were collected at 10 cm intervals (0–10, 10–20, 20–30, and 30–40 cm) from each plot. Soil electrical conductivity and pH were measured simultaneously in situ using a portable instrument (Spectrum Technologies Inc., Chicago, IL, USA, and HACH-sensION3, Loveland, CO, USA, respectively). A total of 120 soil samples were collected and air-dried, ground,

Table 1 Physical and chemical properties of soil profiles from five sampling sites.									
Sites	EC (mS/cm)	рН	Moisture (%)	SOM (%)	BD (g/cm ³)	TN (mg/g)	TC (mg/g)		
W1	3.21 ± 0.86^{ab}	6.43 ± 0.28^{a}	39.78 ± 4.37^{a}	4.38 ± 1.37^{ab}	1.12 ± 0.11^{a}	2.22 ± 1.06^{a}	28.28 ± 11.63^{a}		
W2	3.77 ± 0.85^{a}	6.28 ± 0.38^{a}	$42.23 \pm 8.53^{\rm ac}$	4.89 ± 0.92^{ab}	$1.06 \pm 0.16^{\rm ac}$	$1.74 \pm 0.70^{\rm ac}$	$22.12 \pm 7.95^{\rm ac}$		
W3	3.55 ± 1.29^{ab}	$6.17 \pm 0.27^{\rm ac}$	50.98 ± 4.41^{bd}	4.52 ± 1.03^{ab}	0.94 ± 0.08^{bd}	2.15 ± 0.64^{ac}	26.46 ± 7.37^{a}		
W4	3.29 ± 0.90^{ab}	5.96 ± 0.15^{bc}	52.39 ± 3.15^{b}	5.21 ± 1.33^{a}	$0.89\pm0.05^{\rm b}$	$1.82 \pm 0.09^{\rm ac}$	22.12 ± 1.29^{ac}		
W5	$2.77 \pm 0.95^{\rm bc}$	6.17 ± 0.32^{ab}	46.99 ± 1.83^{cd}	3.77 ± 1.09^{b}	1.00 ± 0.03^{cd}	$1.52 \pm 0.24^{\rm bc}$	$18.88 \pm 2.67^{\rm bc}$		

Notes:

Values are means \pm S.E. Different lowercase letters indicate statistical significant differences between sampling sites within the same column at the level of p < 0.05. EC, electrical conductivity; Moisture, soil moisture; SOM, soil organic matter; BD, bulk density; TN, total nitrogen; TC, total carbon.

and sieved through an 80-mesh nylon sieve. Aboveground parts were clipped at the soil surface and separated into culms, leaves, and litter (withered parts). Underground parts were also collected. Plant samples of *C. malaccensis* or *P. australis* collected from typical communities or ecotones were wholly divided and pretreated based on the different organs of plant species in the ecotones. All plant organs were washed thoroughly with deionized water and dried in an oven (GZX-9140; MBE, Beijing, China) at 65 °C until a constant weight. Then, the samples were ground into a fine powder (<0.25 mm) for Si analysis.

Sample analysis

To extract BSi, 30 mg of plant powder was digested for 5 h with Na₂CO₃ (0.1 mol/L) at 85 °C (*Demaster*, 1981; Struyf et al., 2005). BSi and available Si content of the soil were extracted with alkaline Na₂CO₃ (*Demaster*, 1981) and citric acid (*Acquaye & Tinsley*, 1965; Lu, 1999; Narayanaswamy & Prakash, 2010; Babu et al., 2016), respectively. The dissolved Si content in the extractions was measured using the molybdate blue spectrophotometric method. Total carbon and total nitrogen in soil were determined with a Vario EL Elemental Analyser (Elementar Scientific Instruments, Langenselbold, Germany). Soil grain size was measured with a Master Sizer 2000 Laser Particle Size Analyzer (Master Scientific Instruments, Malvern, UK). Soil organic matter (SOM) was analyzed by the K₂Cr₂O₇ oxidation method. Soil bulk density (BD) was measured using the cutting ring method. Soil moisture content was the weight difference between fresh soils before and after drying in an oven at 105 °C for 24 h. The physical and chemical properties of soil profiles from the five sampling sites are shown in Table 1.

Calculations

The biomass or BSi stock allocation proportions (AP, %) in different plant tissues were calculated by the following equation:

$$AP = \frac{Bi}{Bn} \times 100\%$$
 (1)

where, Bi (g/m^2) is the biomass or BSi stock in different plant organs (including roots, culms, leaves, and litter); and Bn (g/m^2) is the total biomass or total BSi stock. BSi stock is BSi content multiplied by relevant biomass (mg/m^2) .

Peer

The relative competition coefficient of *C. malaccensis* to *P. australis* (Rcp) was calculated based on *De Wit's* (1960) model of resource competition to represent the interspecific competition ability as follows:

$$Rcp = \frac{Yce/Ype}{Yct/Ypt}$$
(2)

where, Yce (g/m^2) and Yct (g/m^2) are the biomass of *C. malaccensis* in ecotones and typical communities, respectively; and Ype (g/m^2) and Ypt (g/m^2) are the biomass of *P. australis* in ecotones and typical communities, respectively. Rcp > 1 means that *C. malaccensis* has a greater competition ability than that of *P. australis*.

For each plant species, we used the data regarding BSi content to calculate the values of roots/culms (B_R/B_C), roots/leaves (B_R/B_L), culms/leaves (B_C/B_L), and shoots/roots (B_S/B_R), which could represent the ability of relative transportation or immobilization of BSi under different situations, according to the methodology described by *Dahmani-Muller et al.* (2000). For example, if the value of B_R/B_C was lower, especially less than 1, more BSi was transported from roots to culms or more BSi was immobilized in culms.

Accumulation factors (AF) in different organs were calculated using the ratio of $(\text{Element})_{\text{plant}}$ to $(\text{Element})_{\text{soil}}$ (*Duman, Cicek & Sezen, 2007*) (Eq. (3)) to evaluate the ability of plants to absorb Si from marsh soils:

$$(AF)_{plant} = \frac{C_{plant}}{C_{soil}}$$
(3)

where, C_{plant} and C_{soil} (mg/g) were the average BSi content in different plant organs (including roots, culms, leaves, and litter) and in surface soils, respectively.

Statistical analyses

Statistical analyses were performed with the SPSS Version 19.0 Statistical Software Package (SPSS Inc., Chicago, IL, USA). One-way analysis of variance was used to analyze significant differences in Si content of marsh plants and soils between typical communities and ecotones. Correlations between available Si content and physical and chemical properties were evaluated using Pearson correlation coefficients with a significance level of p = 0.05. Principal component analysis and stepwise linear regression analysis were used to test the major influencing factors of available Si content in marsh soils.

RESULTS

Variation of biomass allocation of marsh plants in typical communities and ecotones

Biomass allocation was widely different in typical communities compared to ecotones for the same plant species (Fig. 2A; Table 2A). Each organ biomass of *C. malaccensis* and *P. australis* in typical communities was higher than that in ecotones. The total biomass of *C. malaccensis* and *P. australis* in typical communities was higher by 46.4% (p < 0.01) and 46.3% (p < 0.01) than those in ecotones, respectively.



Figure 2 Biomass (A) and allocation proportion of biomass (B) to different organs of Cyperus malaccensis and Phragmites australis in typicalcommunities and ecotones. The species C. malaccensis and P. australis in W2 were recorded as W2-C and W2-P while those in W4 were recorded asW4-C and W4-P respectively. Error bars represent standard error of the mean.Full-size Image: Full-size Image: Full

Table 2 Biomass (A) and allocation proportion (B) to different organs of *Cyperus malaccensis* and *Phragmites australis* in typical communities and ecotones.

Species	Locations	Biomass (g/m ² , A)						Allocation proportion (%, B)			
		Culm	Litter	Leaf	Root	Total	Culm	Litter	Leaf	Root	
Cyperus	Typical	1,023.90 ± 97.89***	57.58 ± 6.17***	8.37 ± 1.55***	796.58 ± 32.52*	1,886.42 ± 73.10***	54.16***	3.04**	0.44*	42.36***	
malaccensis	Ecotone	317.97 ± 11.69***	17.44 ± 1.05***	$2.74 \pm 0.07^{***}$	$672.88 \pm 47.17^*$	1,011.02 ± 34.36***	31.53***	1.73**	0.27*	66.47***	
Phragmites	Typical	684.83 ± 46.08***	$218.5 \pm 70.64^*$	376.51 ± 39.78***	1,034.77 ± 168.95**	2,311.63 ± 219.01***	29.50*	9.45*	16.29**	44.76**	
australis	Ecotone	346.49 ± 14.14***	99.39 ± 4.07*	131.23 ± 17.93***	663.58 ± 37.50**	1,240.69 ± 65.50***	27.95*	8.05*	10.53**	53.47**	

Note:

Asterisks indicate significant differences between typical communities and ecotones within *Cyperus malaccensis* or *Phragmites australis* communities. p > 0.05.

p < 0.05.

p < 0.01.

Generally, the allocation proportion to *C. malaccensis* root biomass was similar to that of *P. australis* in typical communities (p > 0.05); however, it was obviously higher in *C. malaccensis* than that in *P. australis* in the ecotones (p < 0.05; Fig. 2B; Table 2B). For the same plant species, root biomass allocation proportions in ecotones were higher than those in typical communities by 56.9% (*C. malaccensis*, p < 0.01) and 19.5% (*P. australis*, p < 0.05). However, biomass allocation proportions of culms (p < 0.01) and litter (p < 0.05) of *C. malaccensis* in ecotones were lower than those in typical communities by 41.8% and 43.1%, respectively, with no obvious discrepancy with *P. australis* (p > 0.05). The leaf biomass allocation proportion of *P. australis* in ecotones was significantly lower than that in typical communities (p < 0.05) by 35.4%, whereas no difference was observed for *C. malaccensis* (p > 0.05; Table 2B).





BSi variation of marsh plants in typical communities and ecotones variation in BSi content

BSi content in different organs of *C. malaccensis* in typical communities and ecotones was lower than that of *P. australis* (p < 0.05; Fig. 3A; Table 3A). For *C. malaccensis*, the mean BSi content of different organs in W1, W2, W4, and W5 was 12.64, 9.90, 6.35, and 7.41 mg/g, respectively. The mean BSi content in *C. malaccensis* culms (p > 0.05), litter (p > 0.05), and leaves (p < 0.05) in ecotones was lower than that in typical communities, whereas the mean BSi content in roots was greater (by 2.27%, p > 0.05; Table 3A). The average BSi content of different organs in *P. australis* was highest in W2, followed by W3, and was lowest in W4 (Fig. 3A). The average BSi content of *P. australis* culms and roots in ecotones was lower than that in typical communities (p > 0.05), whereas the average BSi content in litter and leaves showed the opposite trend (p > 0.05; Table 3A). Thus, the same plant species had approximately more BSi near the shore and less BSi near the sea, except for *C. malaccensis* in W4.

Allocation proportion of BSi stock

In ecotones, the allocation proportion of *P. australis* BSi stock was higher in roots and litter, and lower in leaves and culms than those in typical communities. The allocation proportion of BSi stock in *C. malaccensis* roots was higher in ecotones than in typical communities, whereas other organs showed the opposite trend (Fig. 3B). In contrast to BSi stocks in culms and leaves, the average allocation proportion in the roots of *C. malaccensis* and *P. australis* in ecotones was higher than that in typical communities by 120.9%

Species Locations BSi content (mg/g, A) Allocation proportion (%, B) Culm Litter Leaf Root Culm Litter Leaf Root 71.54* Cyperus Typical 11.06 ± 3.12 10.40 ± 2.15 13.55 ± 3.85* 4.84 ± 1.09 3.89*** 0.71* 23.86 malaccensis Ecotone 9.35 ± 2.38* $8.44 \pm 1.87^*$ 8.21 ± 1.50** $4.95 \pm 0.21^{*}$ 44.71* 2.22*** 0.35* 52.71* 15.19 ± 1.89 Phragmites Typical $17.86 \pm 1.89^{*}$ 18.43 ± 1.52 $9.98 \pm 0.79^{*}$ 38.54* 11.94* 17.33 32.19* 17.17 ± 4.45 australis Ecotone 19.95 ± 2.53 15.43 ± 0.99 9.87 ± 3.09 36.56 13.70* 11.49 38.26*

Table 3 BSi content (A) and stock allocation proportion (B) to different organs of Cyperus malaccensis and Phragmites australis in typical communities and ecotones.

Note:

Asterisks indicate significant differences between typical communities and ecotones within Cyperus malaccensis or Phragmites australis communities. * *p* > 0.05.

Table 4 BSi transfer coefficients (A) and accumulation factors (B) in Cyperus malaccensis and Phragmites australis of typical communities and ecotones.

Species	Locations	Transfer coefficients (A)				Accumulation factors (B)			
		B_R/B_C	B_R/B_L	B_C/B_L	B_S/B_R	Culm	Litter	Leaf	Root
Cyperus	Typical	0.44	0.36	0.82	2.46	1.59	1.59	1.95	0.70
malaccensis	Ecotone	0.56	0.63	1.13	1.71	1.44	1.43	1.27	0.81
Phragmites	Typical	0.44	0.45	1.01	2.26	2.88	2.97	2.45	1.61
australis	Ecotone	0.54	0.61	1.11	1.92	2.48	3.07	2.23	1.35

and 18.9%, respectively (p > 0.05; Table 3B). For the different plant species, the allocation proportion of BSi stock in C. malaccensis roots (23.9%) was lower than that of P. australis (32.2%) in typical communities; however, it was higher in ecotones, with an allocation of 52.7% in C. malaccensis and 38.3% in P. australis (Table 3B). Therefore, both species might regulate BSi allocation in culms and leaves to be habituated to the homogeneous environment in typical communities while increasing the root BSi allocation to adapt to the competitive environmental conditions.

Variation in BSi transport was observed in C. malaccensis and P. australis as well as in typical communities and ecotones (Table 4A). The values of B_R/B_C , B_R/B_L , and B_C/B_L of C. malaccensis and P. australis in ecotones were higher than those in typical communities, whereas B_S/B_R ratios showed the opposite trend. Therefore, the mobility of BSi from roots to aboveground parts in ecotones was relatively weaker than that of typical communities.

The accumulation of BSi in different organs of the two plant species is shown in Table 4B. Apart from the roots, the AF of other organs of C. malaccensis in typical communities was generally higher than those in the ecotones. However, the AF in the roots of *P. australis* in the ecotones was lower than that in the typical communities by 16.2%.

Variation in available Si content of marsh soils in typical communities and ecotones

Generally, available Si content of marsh soils gradually increased from land to sea except for W5, and the mean value of available Si content in ecotones was significantly higher

p < 0.05.p < 0.01.



Figure 4 Available Si content of marsh soils in typical communities and ecotones. Error bars represent standard error of the mean. Uppercase letters represent significant differences (p < 0.05) in available Si content in the same soil layer between different sampling sites. Lowercase letters represent significant differences (p < 0.05) between different soil depths in the same site. Full-size \square DOI: 10.7717/peerj.7218/fig-4

than that in typical communities by 12.6% (p < 0.05; Fig. 4). Significant differences were observed in available Si content of soil layers from 0 to 30 cm (p < 0.05) in the five sampling sites, with no significant difference in the 30–40 cm soil layer (p > 0.05). In general, available Si content decreased with increasing depth in different marshes (p < 0.01). Therefore, the available Si content in surface soil may be influenced by plant rooting, tides, and other factors, whereas the available Si content in deeper soils was relatively stable.

DISCUSSION

Biomass allocation of marsh plants in ecotones and its implications for plant competition

Plasticity in biomass allocation determines the ability of plants to obtain resources in heterogeneous environments (*Poorter, Remkes & Lambers, 1990*), thereby influencing their competitiveness in different plant communities. Plants may improve their competitive ability by increasing biomass (such as roots, culms, etc.) and by allocating more resources to root systems under low nutrient conditions (*Weiner, 2004; Wu, Chen & Du, 2010*). In the Shanyutan marsh, the biomass of *C. malaccensis* and *P. australis* in ecotones was lower than that in typical communities (Table 2A), indicating that competition might exist between *C. malaccensis* and *P. australis* in ecotones, and plant growth was inhibited to some extent. Additionally, biomass allocation to *C. malaccensis* and *P. australis* roots in

ecotones was higher than that in typical communities (Table 2B). In ecotones, the biomass allocation of *C. malaccensis* culms and *P. australis* leaves was significantly decreased. Therefore, plants might allocate more resources to roots, thus promoting root growth. To improve survival ability and competitiveness, plants may take different strategies for biomass allocation to make better use of resources under different environmental conditions (*Lloret, Casanovas & Penuelas, 2010*). In this region, both plant species might increase their ability to absorb soil nutrients, resulting in lower average C and N content of soils in ecotonal marshes (Table 1). Under this condition, plants in ecotones may increase the biomass allocation proportion of roots to improve their competitiveness and resist nutrient stress.

In ecotones, *P. australis* occupied more aboveground space with greater density and higher height, whereas the density of *C. malaccensis* was greatly reduced (Table S1). Thus, *C. malaccensis* had to promote a high biomass allocation proportion of roots to absorb more nutrients from subsurface layers and better withstand competition from *P. australis* in ecotones. In addition, the relative competitive stress of *C. malaccensis* was higher than that of *P. australis* (Rcp > 1), especially for underground resources (Table S2), which also showed that the underground parts of *C. malaccensis* had a stronger competition advantage in ecotones.

Changed BSi stock allocation of marsh plants in ecotones and its implications for plant competition

Biomass of each organ in plants determines the amount of accumulated nutrients and the distribution of nutrients within plants (*Zeng, Zhang & Tong, 2009; Li, 2013*). Several studies have indicated that the differences of plant BSi storage were attributable to the discrepancies of biomass (*Schoelynck et al., 2010; Struyf et al., 2015; Carey et al., 2017*). In the present study, BSi stock allocation in *C. malaccensis* and *P. australis* roots was obviously higher in ecotones than that in typical communities, principally owing to the increased biomass allocation proportion (Fig. 2B; Table 2B). Especially for *C. malaccensis*, BSi allocation proportion of roots in ecotones almost doubled in typical communities, whereas the BSi allocation proportions in leaves and culms were nearly decreased by half. However, BSi allocation proportion in *P. australis* roots only increased by 18.9% in ecotones. Therefore, this might illustrate that *C. malaccensis* roots reserving more BSi may help it to compete with *P. australis* in ecotones.

Usually plants allocate more resources to roots to obtain more nutrients to improve their competitiveness under low nutrient conditions (*Weiner, 2004*). Regarding Si, active plant Si uptake has been suggested for native species exhibiting strong environmental stress conditions (*Schoelynck & Struyf, 2016*). The level of BSi accumulation in marsh plants could be automatically adaptive, that is, Si accumulation in the vegetation would be increased under stressful conditions (*Schoelynck & Struyf, 2016*). In the Shanyutan marsh, *P. australis* flourished easier to gain light and space on the ground than *C. malaccensis*. Therefore, *C. malaccensis* had to promote the Si enrichment ability of its roots to resist the spread of *P. australis* in ecotones, and adjust the absorption and allocation of Si to adapt to this competitive environment (*Gao et al., 2017*). In the present study, the allocation proportion of BSi stock in *C. malaccensis* and *P. australis* was different among ecotones, compared with that in the typical communities. In ecotones, BSi allocation proportion of each organ in *C. malaccensis* decreased except for the roots, whereas the increased value of BSi allocation proportion in *P. australis* was focused on the leaves and litter. Generally, Si has distinct effects in Si-accumulators (such as marsh plants) that are exposed to abiotic and biotic stresses (*Ma & Yamaji, 2015; Coskun et al., 2019*). Therefore, for superficial estimates, more BSi was focused on the roots in *C. malaccensis* rather than on the leaves in *P. australis* in ecotones to resist biotic pressure.

These results might indicate that the two plant species use different strategies for Si accumulation and allocation in ecotones to adapt to the competitive environment. *P. australis* expanded primarily by occupying a wider aboveground space and by increasing its Si accumulation capacity of aboveground organs, whereas *C. malaccensis* was able to resist the competitive pressure of *P. australis* by expanding the Si allocation of its roots.

Differences of available Si content in marsh soils between typical communities and ecotones

Available Si content is important for measuring soil nutrient supply to plants (*Babu et al., 2016*; *Klotzbücher et al., 2018*). Pearson correlation analyses showed that significantly positive correlations occurred between available Si content and soil moisture and SOM (p < 0.01 or p < 0.05; Table S3). Available Si content was relatively higher in soils with more water content (*Gao et al., 2015*), which might be the reason for higher available Si content in ecotones (W2 and W4, Table 1). In addition, a previous study demonstrated that the decomposition of SOM could release Si, and soil organic acids and reductive conditions formed by organic matter degradation may damage iron-Si complexes, which are also conducive to Si dissolution (*Ma, Chen & Chu, 2016*). The higher available Si content observed in ecotonal marsh soils might also be related to the higher SOM (Table 1) compared to typical communities. Thus, soil moisture and SOM had important effects on the distribution of soil available Si in the Shanyutan marsh.

Principal component analyses showed that the major factors influencing available Si content were soil BD, grain composition, pH, and organic components (a variance contribution rate of 81.0%; Table S4). Furthermore, stepwise linear regression analysis revealed that soil BD was the crucial factor influencing available Si content (available Si = $-301.691 \times BD + 793.286$, p = 0.006). In the present study, available Si content was relatively higher in ecotones than in typical communities and had a significantly negative correlation with BD (p < 0.01; Table S3), which might be due to the root systems of the vegetation. Previous studies indicated that root growth was limited because of the hardness and lack of oxygen supply in soils with higher BD, thus decreasing root dry weight (*Grzesiak, 2009; Singh, Salaria & Kaul, 2015*). In the present study, the mean biomass allocation to root systems in ecotones (60.0%) was higher than that in typical communities (43.56%, p < 0.05; Table 2B), whereas soil BD showed the opposite trend in ecotones (0.98 g/cm³) and typical communities (1.02 g/cm³, p > 0.05). Therefore, this might result in the higher available Si content in the marsh soils of ecotones.

CONCLUSIONS

The biomass and BSi stock allocation proportion of *C. malaccensis* and *P. australis* roots were higher in ecotones than those in typical communities in the Shanyutan marsh. In ecotones, *P. australis* might expand primarily by occupying a wider aboveground space and increasing the Si accumulation capacity of aboveground organs, whereas *C. malaccensis* might resist the competition pressure of *P. australis* by increasing the Si allocation capacity of its roots. Available Si content in ecotone soils was also higher than that in typical community soils. Soil moisture, SOM, BD, and plant assimilation were the major factors influencing available Si content in the Shanyutan marsh.

ACKNOWLEDGEMENTS

We would like to thank Tao He, Jiabing Li, Peng Ren and He Zhu for their assistance with field sampling and thank Guanyun Hou for her help with the data analysis. We sincerely thank the reviewers and editor for their valuable comments on the manuscript.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

The work was supported by the Natural Science Foundation of China (Nos. 41401114, 41573008, 41873015 and U1612442) and the Guangzhou University's 2017 training program for young top-notch personnel (BJ201709). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors: Natural Science Foundation of China: 41401114, 41573008, 41873015 and U1612442. Guangzhou University's 2017 training program for young top-notch personnel: BJ201709.

Competing Interests

The authors declare that they have no competing interests.

Author Contributions

- Hui Gao performed the experiments, analyzed the data, prepared figures and/or tables.
- Shuijing Zhai conceived and designed the experiments, contributed reagents/materials/ analysis tools, authored or reviewed drafts of the paper, approved the final draft.
- Zhigao Sun performed the experiments, analyzed the data, prepared figures and/or tables.
- Juan Liu conceived and designed the experiments, authored or reviewed drafts of the paper, approved the final draft.
- Chuan Tong performed the experiments, analyzed the data, prepared figures and/or tables.

Data Availability

The following information was supplied regarding data availability: Data are available as a Supplemental File.

Supplemental Information

Supplemental information for this article can be found online at http://dx.doi.org/10.7717/ peerj.7218#supplemental-information.

REFERENCES

- Acquaye D, Tinsley J. 1965. Soluble silica in soils. In: Hallsworth EG, Crawford DV, eds. Experimental Pedology. In: Hallsworth EG, Crawford DV, eds. Proceedings of the Eleventh Easter School in Agricultural Science. London: Butterworths, 126–148.
- Babu T, Tubana B, Datnoff L, Dupree P, White B. 2016. Survey of the plant-available silicon status of agricultural soils in Louisiana. *Journal of Plant Nutrition* 41(3):273–287 DOI 10.1080/01904167.2017.1346668.
- Brackhage C, Schaller J, Bäucker E, Dudel EG. 2013. Silicon availability affects the stoichiometry and content of calcium and micro nutrients in the leaves of common reed. *Silicon* 5(3):199–204 DOI 10.1007/s12633-013-9145-3.
- Cadenasso ML, Pickett STA, Weathers KC, Jones CG. 2003. A framework for a theory of ecological boundaries. *Bioscience* 53(8):750–758 DOI 10.1641/0006-3568(2003)053[0750:affato]2.0.co;2.
- **Carey JC, Fulweiler RW. 2013.** Nitrogen enrichment increases net silica accumulation in a temperate salt marsh. *Limnology and Oceanography* **58(1)**:99–111 DOI 10.4319/lo.2013.58.1.0099.
- Carey JC, Parker TC, Fetcher N, Tang JW. 2017. Biogenic silica accumulation varies across tussock tundra plant functional type. *Functional Ecology* 31(11):2177–2187 DOI 10.1111/1365-2435.12912.
- Chen J, Tang JJ, Li DY. 2015. Influence of outflows from the Minjiang River on the estuary and adjacent coastal areas. Beijing: Science Press.
- Coskun D, Britto DT, Huynh WQ, Kronzucker HJ. 2016. The role of silicon in higher plants under salinity and drought stress. *Frontiers in Plant Science* 7:1072 DOI 10.3389/fpls.2016.01072.
- Coskun D, Deshmukh R, Sonah H, Menzies JG, Reynolds O, Ma JF, Kronzucker HJ, Bélanger RR. 2019. The controversies of silicon's role in plant biology. *New Phytologist* 221(1):67–85 DOI 10.1111/nph.15343.
- **Dahmani-Muller H, Van Oort F, Gélie B, Balabane M. 2000.** Strategies of heavy metal uptake by three plant species growing near a metal smelter. *Environmental Pollution* **109(2)**:231–238 DOI 10.1016/S0269-7491(99)00262-6.
- **Dai ZZ. 2004.** Analysis of hydrologic features in the Fuzhou district. *Hydraulic Science and Technology* **3**:9–11.
- Datnoff LE, Deren CW, Snyder GH. 1997. Silicon fertilization for disease management of rice in Florida. *Crop Protection* 16(6):525–531 DOI 10.1016/S0261-2194(97)00033-1.
- Datnoff LE, Snyder GH, Korndörfer GH. 2001. Silica in agriculture. Amsterdam: Elsevier.
- De Wit CT. 1960. On competition. Versl Landbouwk Onde 66(1):36-39.
- **Demaster DJ. 1981.** The supply and accumulation of silica in the marine environment. *Geochimica Et Cosmochimica Acta* **45(10)**:1715–1732 DOI 10.1016/0016-7037(81)90006-5.
- Duman F, Cicek M, Sezen G. 2007. Seasonal changes of metal accumulation and distribution in common club rush (*Schoenoplectus lacustris*) and common reed (*Phragmites australis*). *Ecotoxicology* 16(6):457 DOI 10.1007/s10646-007-0150-4.
- **Epstein E. 1994.** The anomaly of silicon in plant biology. *Proceedings of the National Academy of Sciences of the United States of America* **91(1)**:11–17 DOI 10.1073/pnas.91.1.11.

- **Epstein E. 2009.** Silicon: its manifold roles in plants. *Annals of Applied Biology* **155(2)**:155–160 DOI 10.1111/j.1744-7348.2009.00343.x.
- Gao Z, Jie DM, Liu LD, Liu HY, Lu MJ, Gao GZ, Liu HT, Gao YS. 2015. Temporal and spatial variation of available soil silicon and its influence on the formation of phytoliths in *Phragmites communis* in Northeast China. *Quaternary Sciences* **35**(4):967–976.
- Gao H, Li JB, He T, Sun ZG, Fan AL, Zhu H, Ren P, Zhai SJ. 2017. Silica distribution characteristics in plant-soil systems of typical vegetation communities and ecotones in Min River estuary wetland. *Journal of Soil and Water Conservation* 31(1):279–285.
- Gao H, Zhai SJ, Sun ZG, He T, Tian LP, Hu XY. 2018. Spatial and temporal variations of available silica content in marsh soils under the *Spartina alterniflora* invasion in the Min River estuary. *Acta Ecologica Sinica* 38(17):6136–6142 DOI 10.5846/stxb201709041593.
- Grzesiak MT. 2009. Impact of soil compaction on root architecture, leaf water status, gas exchange and growth of maize and triticale seedlings. *Plant Root* 3:10–16 DOI 10.3117/plantroot.3.10.
- Hou LJ, Liu M, Yang Y, Ou DN, Lin X, Chen H. 2010. Biogenic silica in intertidal marsh plants and associated sediments of the Yangtze Estuary. *Journal of Environmental Sciences* 22(3):374–380 DOI 10.1016/s1001-0742(09)60118-2.
- Hou GY, Zhai SJ, Gao H, Le XQ. 2017. Effect of salinity on silicon, carbon, and nitrogen during decomposition of *Spartina alterniflora* litter. *Acta Ecologica Sinica* 37(1):184–191 DOI 10.5846/stxb201607261527.
- Jacobs S, Müller F, Teuchies J, Oosterlee L, Struyf E, Meire P. 2013. The vegetation silica pool in a developing tidal freshwater marsh. *Silicon* 5(1):91–100 DOI 10.1007/s12633-012-9136-9.
- Klotzbücher T, Klotzbücher A, Kaiser K, Merbach I, Mikutta R. 2018. Impact of agricultural practices on plant-available silicon. *Geoderma* 331:15–17 DOI 10.1016/j.geoderma.2018.06.011.
- Li XW. 2013. Study on the biomass, nutrient content and C, N reserves of *Betula alnoides* plantation. Master Degree. Guangxi University.
- Liu JQ, Zeng CS, Chen N. 2006. Research of Minjiang River estuary wetland. Beijing: Science Press.
- Lloret F, Casanovas C, Penuelas J. 2010. Seedling survival of Mediterranean shrubland species in relation to root: shoot ratio, seed size and water and nitrogen use. *Functional Ecology* 13(2):210–216 DOI 10.1046/j.1365-2435.1999.00309.x.
- Lu RK. 1999. *Methods of soil science and agricultural chemistry analysis*. Beijing: Agriculture Science and Technology Press.
- Ma X, Chen JJ, Chu GX. 2016. Spatial distribution and affecting factors of soil available Si in Shihezi region. *Acat Agriculturae Boreali-Occidentalis Sinica* 25(2):276–282.
- Ma JF, Yamaji N. 2006. Silicon uptake and accumulation in higher plants. *Trends in Plant Science* 11(8):392–397 DOI 10.1016/j.tplants.2006.06.007.
- Ma JF, Yamaji N. 2015. A cooperative system of silicon transport in plants. *Trends in Plant Science* 20(7):435-442 DOI 10.1016/j.tplants.2015.04.007.
- Narayanaswamy C, Prakash NB. 2010. Evaluation of selected extractants for plant-available silicon in rice soils of southern India. *Communications in Soil Science and Plant Analysis* 41(8):977–989 DOI 10.1080/00103621003646063.
- Poorter H, Remkes C, Lambers H. 1990. Carbon and nitrogen economy of 24 wild species differing in relative growth rate. *Plant Physiology* 94(2):621–627 DOI 10.1104/pp.94.2.621.
- Querné J, Ragueneau O, Poupart N. 2012. In situ biogenic silica variations in the invasive salt marsh plant, *Spartina alterniflora*: a possible link with environmental stress. *Plant and Soil* 352(1-2):157-171 DOI 10.1007/s11104-011-0986-5.

- Savant NK, Korndörfer GH, Datnoff LE, Snyder GH. 1999. Silicon nutrition and sugarcane production: a review. *Journal of Plant Nutrition* 22(12):1853–1903 DOI 10.1080/01904169909365761.
- Schaller J, Brackhage C, Dudel EG. 2012a. Silicon availability changes structural carbon ratio and phenol content of grasses. *Environmental and Experimental Botany* 77:283–287 DOI 10.1016/j.envexpbot.2011.12.009.
- Schaller J, Brackhage C, Gessner MO, Bäucker E, Dudel EG. 2012b. Silicon supply modifies C:N:P stoichiometry and growth of *Phragmites australis*. *Plant Biology* 14(2):392–396 DOI 10.1111/j.1438-8677.2011.00537.x.
- Schaller J, Brackhage C, Paasch S, Brunner E, Bäucker E, Dudel EG. 2013. Silica uptake from nanoparticles and silica condensation state in different tissues of *Phragmites australis*. Science of the Total Environment 442(1):6–9 DOI 10.1016/j.scitotenv.2012.10.016.
- Schaller J, Hines J, Brackhage C. 2017. Is initial Si concentration determining the influence of warming and N-supply on stoichiometric changes during litter decomposition? *Aquatic Botany* 138:1–8 DOI 10.1016/j.aquabot.2016.11.013.
- Schaller J, Schoelynck J, Struyf E, Meire P. 2016. Silicon affects nutrient content and ratios of wetland plants. *Silicon* 8(4):479–485 DOI 10.1007/s12633-015-9302-y.
- Schaller J, Struyf E. 2013. Silicon controls microbial decay and nutrient release of grass litter during aquatic decomposition. *Hydrobiologia* 709(1):201–212 DOI 10.1007/s10750-013-1449-1.
- Schoelynck J, Bal K, Backx H, Okruszko T, Meire P, Struyf E. 2010. Silica uptake in aquatic and wetland macrophytes: a strategic choice between silica, lignin and cellulose? *New Phytologist* 186(2):385–391 DOI 10.1111/j.1469-8137.2009.03176.x.
- Schoelynck J, Müller F, Vandevenne F, Bal K, Barão L, Smis A, Opdekamp W, Meire P, Struyf E. 2014. Silicon-vegetation interaction in multiple ecosystems: a review. *Journal of Vegetation Science* 25(1):301–313 DOI 10.1111/jvs.12055.
- Schoelynck J, Struyf E. 2016. Silicon in aquatic vegetation. *Functional Ecology* 30(8):1323–1330 DOI 10.1111/1365-2435.12614.
- Simas T, Nunes JP, Ferreira JG. 2001. Effects of global climate change on coastal salt marshes. *Ecological Modelling* 139(1):1–15 DOI 10.1016/s0304-3800(01)00226-5.
- Singh J, Salaria A, Kaul A. 2015. Impact of soil compaction on soil physical properties and root growth: a review. *International Journal of Food, Agriculture and Veterinary Sciences* 5(1):23–32.
- Struyf E, Conley DJ. 2009. Silica: an essential nutrient in wetland biogeochemistry. *Frontiers in Ecology and the Environment* 7(2):88–94 DOI 10.1890/070126.
- Struyf E, Mosimane K, Van Pelt D, Murray-Hudson M, Meire P, Frings P, Wolski P, Schaller J, Gondwe MJ, Schoelynck J, Conley DJ. 2015. The role of vegetation in the Okavango Delta silica sink. *Wetlands* 35(1):171–181 DOI 10.1007/s13157-014-0607-1.
- Struyf E, Van Damme S, Gribsholt B, Bal K, Beauchard O, Middelburg JJ, Meire P. 2007. *Phragmites australis* and silica cycling in tidal wetlands. *Aquatic Botany* **87**(2):134–140 DOI 10.1016/j.aquabot.2007.05.002.
- Struyf E, Van Damme S, Gribsholt B, Middelbulg JJ, Meire P. 2005. Biogenic silica in tidal freshwater marsh sediments and vegetation (Schelde estuary, Belgium). *Marine Ecology Progress Series* 303:51–60 DOI 10.3354/meps303051.
- Theriot JM, Conkle JL, Pezeshki SR, Delaune RD, White JR. 2013. Will hydrologic restoration of Mississippi River riparian wetlands improve their critical biogeochemical functions? *Ecological Engineering* **60(6)**:192–198 DOI 10.1016/j.ecoleng.2013.07.021.

- Tong C, Jia RX, Wang WQ, Zeng CS. 2010. Spatial variations of carbon, nitrogen and phosphorous in tidal salt marsh soils of the Minjiang River estuary. *Geographical Research* 29(7):1203–1213.
- Tong C, Zhang LH, Wang WQ, Gauci V, Marrs R, Liu BG, Jia RX, Zeng CS. 2011. Contrasting nutrient stocks and litter decomposition in stands of native and invasive species in a sub-tropical estuarine marsh. *Environmental Research* 111(7):909–916 DOI 10.1016/j.envres.2011.05.023.
- Weiner J. 2004. Allocation, plasticity and allometry in plants. *Perspectives in Plant Ecology Evolution and Systematics* 6(4):207–215 DOI 10.1078/1433-8319-00083.
- Wu GL, Chen M, Du GZ. 2010. Response of biomass allocation and morphological characteristics to light and nutrient resources for seedlings of three alpine species. *Acta Ecologica Sinica* 30(1):60–66.
- Zeng CS, Zhang LH, Tong C. 2009. Seasonal dynamics of nitrogen and phosphorus in *Phragmites australis* and *Spartina alterniflora* in the wetlands of Min River estuary. Wetland Science 7(1):16–24.
- Zhai SJ, Xue LL. 2016. Changes in the distribution of silica in the porewaters and sediments of the intertidal zone with different plant communities in the Min River estuary. *Acta Ecologica Sinica* 36(21):6766–6776 DOI 10.5846/stxb201501140109.
- Zhang WK. 1991. The analysis of the developments and evolution of the shoal patch outer Minjiang estuary. *Journal of Fujian Normal University (Natural Science Edition)* 7(3):94–100.