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Abstract

Emus (*Dromaius novaehollandiae*) are exclusively terrestrial, bipedal and cursorial ratites with some similar biomechanical characteristics to humans. Their growth rates are impressive as their body mass increases eighty-fold from hatching to adulthood whilst maintaining the same mode of locomotion throughout life. These ontogenetic characteristics stimulate biomechanical questions about the strategies that allow them to cope through these changes. To answer such questions, in this study we have collected pelvic limb anatomical data (muscle architecture, tendon length, tendon mass and bone lengths) and calculated muscle physiological cross sectional area (PCSA) and average tendon cross sectional area from emus across an ontogenetic series (n=17, body masses from 3.6 to 42 kg). The data were analysed by reduced major axis regression to determine scaling relationships with body mass. Muscle mass and PCSA showed a marked trend towards positive allometry (26 and 27 out of 34 muscles respectively) and fascicle length showed a more mixed scaling pattern. The long tendons of the main digital flexors scaled with positive allometry for all characteristics whilst other tendons demonstrated a less clear scaling pattern. Finally, the two longer bones of the limb (tibiotarsus and tarsometatarsus) also exhibited positive allometry for length and the other two (femur and first phalanx of the pes) had trends towards isometry. These results indicate that emus increase their muscle force-generating capacities, as well as potentially increasing the force-sustaining capacities of their tendons, as they grow. Furthermore, we have clarified anatomical descriptions and provided illustrations of the pelvic limb muscle-tendon units in emus.

Comment [TW1]: What does 'them' refer to? Is it characteristics or questions or 'emus'

Comment [TW2]: There are quite a few other bones in leg, (fibula, and 11 phalanges)

Comment [TW3]: There are 3 digits, each has a first phalanx

Comment [TW4]: Would this not be obvious – if they didn't as they grew mass would exceed ability of the muscle to move it

Introduction

Scaling studies (relating animal body mass to other biological parameters) have broadly elucidated locomotor adaptations across a wide range of body sizes. These studies have also described important size-related biomechanical (Alexander et al. 1979; Bertram & Biewener 1990; Biewener 1982; Gatesy & Biewener 1991; LaBarbera 1989; Maloiy et al. 1979; McMahon 1975) and metabolic constraints (Gillooly et al. 2001; Hemmingsen 1960; Hokkanen 1986; Kleiber 1932; Schmidt-Nielsen 1984; Taylor et al. 1981). Intraspecific scaling studies (especially in species with adult size variation or sexual dimorphism) are less common (Allen et al. 2010,2014; Carrier & Leon 1990; Carrier 1983; Dial & Jackson 2011; Main & Biewener 2007; Miller et al. 2008; Picasso 2012a; Smith & Wilson 2013; Young 2009). An ontogenetic approach can yield valuable insights into musculoskeletal adaptations during growth and size-related constraints, often with fewer parameters changing (e.g., similar locomotor strategies and basic anatomy preserved across ontogeny). Therefore, when interpreted together with functional information it is possible to infer life history strategies and trade-offs that might occur during growth. Such information is also useful to understand developmental abnormalities and study intervention strategies to manage them.

Ratites are large flightless birds with cursorial morphology e.g., (Smith et al. 2010; Smith & Wilson 2013) that makes them attractive subjects for studies of terrestrial locomotion and bipedalism. Certain characteristics make emus (*Dromaius novaehollandiae*) particularly useful: they have some anatomical and functional similarities to other bipedal animals, including purportedly humans (Goetz et al. 2008). Compared to ostriches, they are generally easier to handle and train in experimental settings, due to their smaller size and calmer temperament. Finally, their growth rate is impressive, as they multiply their body weight ~80 times in their first 18 months of life (Minnaar & Minnaar 1998), making them useful subjects for ontogenetic scaling studies. Despite this interest, there are still some discrepancies in published anatomical descriptions and depictions of the pelvic limb musculature of emus (Haughton 1867; Patak & Baldwin 1998; Vanden Berge & Zweers 1993), and clear visual anatomical aids are lacking in the literature.

Some of the biomechanical changes occurring during the growth of emus have been studied (Main & Biewener 2007). This study described the skeletal strain patterns on the surfaces of the femur and the tibiotarsus (TBT) in running birds, demonstrating a significant increase in the magnitude of cranial and caudal femoral and caudal tibiotarsal strains during ontogeny, despite the enlargement and strengthening of those bones via positive allometric scaling of the second moment of area. Muscles have been shown to influence the pattern of strain in bones (Yoshikawa et al. 1994), and although other factors are likely to be involved in the strain pattern changes reported across ontogeny in emus (Main & Biewener 2007), allometric scaling of the musculature could also play a role in these changes in bone tissue loading. The strains induced by muscle contraction will be proportional to the muscle forces acting on the bone; therefore by estimating muscle forces (e.g., maximal force capacity based upon anatomy), associations between these two findings would be possible.

In order to build on already available data for emus (Goetz et al. 2008; Main & Biewener 2007) and improve our understanding of their developmental and biomechanical scaling strategies, we aim here to quantify the ontogenetic scaling patterns of the long bones, pelvic muscles and their tendons and in the process describe and compare the functional and descriptive anatomy of the pelvic limb musculature of emus. We use regression analysis to determine the relationship of muscle architectural properties with body mass in an ontogenetic series of emus and then examine the implications of these findings for their locomotor ontogeny.

Comment [TW5]: Usually it would be discrepancy between x and y, so do the authors mean errors?

Comment [TW6]: So does this mean Main & Biewener showed that only some of the biomechanical changes occurring during ontogeny have been studied, or did they make one such study and should be cited'e.g. Main.."

Materials and methods

Animal subjects and care: UK group

We dissected 17 emus for this study, obtained from our ongoing research examining emu ontogenetic biomechanics (conducted with ethical approval under a UK Home Office license). These emus were divided in three groups of animals according to their age: Group 1: Three individuals at 4-6 weeks old; Group 2: One 24-28 weeks (6 months) old individual; and Group 3: Six 64-68 weeks (16 months) old individuals. All birds had been used as experimental animals and kept in a small pen (7x7m) for the first six weeks of life, after which they were moved to an outdoor larger enclosure with grass footing (40mx15m) until they were six months old; after this they were moved to a large (1.6 hectares) grass field (maximal animal density at one time was 8 birds/ha). The birds were all born in three consecutive yearly breeding seasons. Only the birds in Group 3 were from the same breeding season but not necessarily the same progenitors; birds from the other two Groups were from two different seasons.

All animals were hatched at a commercial breeding farm in the UK and raised from four weeks of age at the Royal Veterinary College. They were fed a commercial ostrich pelleted diet supplemented with grass and from six weeks of age were kept with free access to commercial food and grass. At 24 weeks, their diet changed from an ostrich grower diet to adult ostrich pelleted food (Dodson and Horrel Ltd., Kettering, Northamptonshire, UK). There were no restrictions or enforcements on the animals' regular exercise regime. All animals were euthanized after other experimental procedures were completed, by lethal intravenous injection of a barbiturate following induction of deep terminal general anaesthesia by intramuscular injection of ketamine and xylazine. Carcasses were kept frozen in a -20°C freezer for up to 2 years before dissection. Thawing was allowed at variable ambient temperatures and for variable amounts of time depending on the size of the animal, and dissection started no longer than 4 days after removal from the freezer. All dissections were performed within a six week period and led by the same individual (L.P.L.).

USA group of emus

Unpublished raw data of muscle masses from a different group of 29 emus (0.74 to 51.7 kg body mass) used for similar purposes as those described for the UK group were also included in this study. This group was bred and reared in the USA (Concord Field Station, Harvard University) under the care of another investigator (R.P.M.) who led all dissections for this group. The size and age composition for this group was more heterogeneous, and only body masses and muscle masses were available for analysis. Because the purpose of the dissections in the group was not a systematic ontogenetic musculoskeletal scaling study, the number of muscles dissected per animal varied.

Bone measurements

Maximal interarticular lengths of the femur, tibiotarsus (TBT), tarsometatarsus (TMT) and first phalanx of the middle (third) digit were measured using an ordinary flexible measuring tape (±1 mm) once they were cleared of all soft tissues.

Myology and muscle architecture

We identified muscles of emus using four separate literature sources (Haughton 1867; Patak & Baldwin 1998; Smith et al. 2007; Vanden Berge & Zweers 1993); when our observations differed from these, we described the anatomical landmarks and attachments in detail according to our observations. General main actions of the muscle were defined based on these publications and confirmed by identifying the muscle attachments and paths and then mimicking the muscle action by applying tension on the muscle during dissection. We used additional reference to a biomechanical model of an ostrich (Hutchinson et al. 2014) to refine the three-dimensional actions of the hip muscles, as those actions are difficult to accurately ascertain from visual inspection and

manipulation. Table 1 shows our simplified description of the anatomy, abbreviations used throughout this study, and inferred muscle actions. Figures 1 to 3 show schematic anatomical representations of the muscle anatomy.

To avoid freeze drying of the carcasses, we ensured all animals were frozen soon after euthanasia kept in sealed bags, and were not thawed and refrozen before dissection. Although this is not essential and often not possible in this type of work, the carcasses showed minimal autolysis and therefore an easier and better dissection during which muscle actions could be approximated without damaging their structure and attachments was possible.

Dissection of the right pelvic limb muscles was performed in all specimens apart from the first two subjects, in which the muscles of the left limb were dissected first to standardise the technique. Measurements taken from the muscles of the left limb were not used (avoiding duplication of information), with the exception of when there were unidentified/damaged muscles from the right limb of the same specimen, in order to create a complete set of muscles for each specimen.

After identification of each muscle, we performed complete dissection and removal of it by transection at its origin and insertion(s). Next, the muscle was laid flat on a table and we took four muscle architectural measurements in a standard protocol: muscle mass (M_m), fascicle length (L_f), muscle belly length and pennation angle (Θ). Muscle mass was measured on an electronic scale (± 0.01 g) after removal of tendons, fat and aponeuroses, fascicle length was measured from at least five random sites within the muscle belly using digital callipers (± 0.1 mm), muscle belly length was measured as the length (± 1 mm) from the origin of the most proximal muscle fascicles to the insertion of the most distal fascicles into the distal tendon or aponeurosis, and the pennation angle was measured at least five times using a goniometer (±5°); the mean of the latter measurements was taken as the pennation angle for the muscle. The repeated measurements were taken from multiple cuts into the muscle to expose different anatomical orientations of the fascicles with the same muscle. This methodology minimises the differences that may be seen across an individual muscle and ensures mean values used for further calculations are representative of the overall architecture of the muscle. We calculated total limb muscle mass by adding the individual masses of the muscle bellies. Our approach was straightforward for most muscles, apart from three smaller muscles of the limb: IFI, ISF and FPPDII (Table 1), where minor dissection mistakes might have impaired estimates of their masses and architectural properties.

Muscle volume was calculated by dividing muscle mass by estimated muscle density of vertebrates (1.06 g cm^{-3}; (Brown et al. 2003; Hutchinson et al. 2014; Mendez & Keys 1960)). From these data we calculated physiological cross-sectional area (PCSA) for each muscle via the standard formula (Powell et al. 1984; Sacks & Roy 1982) (Equation 1):

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$$)$$

When a tendon was present it was dissected down to its insertion onto the bone together with the muscle. The tendon was then transected at the musculotendinous junction when a clear separation became apparent and stretched on a flat surface. We then measured lengths with a standard ruler or flexible measuring tape (±1mm), and tendon mass was also calculated using the same instrumentation as for the muscles.

Tendon cross-sectional area (TCSA) was calculated using tendon length (L_{ten}); from muscle origin to bony insertion; and tendon mass (M_{ten}) as follows (Equation 2):

Where 1120 kg m^{-3} is assumed as the density of tendon (Hutchinson et al. 2014; Ker 1981).

Statistical analysis

Ontogenetic scaling relationships of (non-normalized) muscle properties were analysed using reduced major axis ("Model II") regression for $\log_{10}$ of each property vs $\log_{10}$ body mass using custom-designed R software code (R Development Core Team 2010) ~~code~~. A Shapiro-Wilk test was performed to assess normality of distribution of the residuals, and the *p* value for significance was set to <0.05. The inclusion criteria for data presented were: Datasets first had to have a *p* value <0.05 in the above described Shapiro-Wilk test. If this *p* was >0.05, the data were then tested for the presence of outliers (which were set at ±2 standard deviations [SD] from the mean) and outliers removed. The RMA linear regression was performed again using this dataset and again, data were only presented if the *p* value for distribution of residuals was <0.05. Once the datasets were defined, R^2 correlation values and upper and lower bounds of the 95% confidence interval (CI) were calculated to assess the spread of data points around each regression line.

In order to obtain relative values to compare results from individuals of different size, muscle mass, PCSA and F_{length} were normalized to body mass (BM) by dividing each value by the subject's BM, $BM^{0.67}$ and $BM^{0.33}$ respectively. We used body mass (BM) as our independent variable and the target architectural parameter as our dependant variable. Overall, we followed a similar approach as that described by Allen et al. (2010,2014).

Briefly, for two objects to be considered geometrically similar (and thus for an isometric scaling pattern to be inferred), areas should scale to the square product of lengths and volumes to the cube of lengths. Because mass is a volumetric property, the dependant variable is considered to scale isometrically if the mass of the structure scales with BM^1, areal properties (PCSA, TCSA) scale to $BM^{0.67}$ and lengths scale to $BM^{0.33}$, whereas angles and other non-dimensional variables should scale as BM^0.

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Results

We obtained 6524 measurements of seven different muscle-tendon architectural parameters from 34 pelvic limb muscles and four pelvic limb bones in 17 emus from 3.6 to 42 kg of body mass. We found strong evidence for positive allometric scaling of many of these architectural parameters, as described below. To aid interpretation of our results, we have divided the muscles of the limb into proximal (those acting mostly ~~acting~~ on the hip and knee joints) and distal (those acting on the ankle, foot and digits) ~~muscles~~ groups and have used this division to compare trends between the two regions.

Bone lengths

The lengths of the four bones scaled with moderate positive allometry (expected slope representing isometry would be 0.33). The femur had the least marked allometric exponent (0.38), whilst the tarsometatarsus the greatest (0.44), the tibiotarsus had a slope value of 0.41 and for the 1st phalanx of the second digit (P1) the value was 0.39. The lower CIs for all bones were greater than isometry, except for P1 where the lower CI was 0.33. Finally, the R^2 values were >0.95 for all bones, indicating good correlation of the data (for full results see Table 2).

Myology, architectural characteristics of muscles and functional interpretation

We classified a total of 34 muscles in Table 1. As noted by Regnault et al. (2014), there is no patellar ossification in the knee joint of emus, unlike ostriches and some other palaeognaths as well as most extant birds. Although muscle origins, insertions and paths were generally found to agree with previous publications (Haughton 1867; Patak & Baldwin 1998; Vanden Berge & Zweers 1993) and

Comment [TW7]: It would improve the utility of the ms if an additional figure was introduced that depicted the areas of insertions of all these muscles because the table and its descriptions are very confusing. Eg especially with regard to lateral femur

hence detailed redescription is unnecessary, there were a few muscles for which we have found some differences worth noting, or for which we needed to use methodological simplifications:

M. iliotibialis lateralis pars post acetabularis (IL): The distal fusion and similar actions of both parts of the IL muscle meant that, in order to avoid dissection errors when finding the division between the cranial and caudal parts of the muscle, we measured and presented them together.

M. iliotrochantericus cranialis (ITCR): Although this was a clear, separate muscle in most specimens, it was found to be fused with the ITM in two specimens of body mass ~20 kg, which is a common finding in birds (Gangl et al. 2004)

M. ischiofemoralis (ISF): This small muscle is difficult to detect and dissect, which is likely to have affected the accuracy of the data obtained from it (leading to lower R^2 values and wider 95% CI ranges). Its action is likely to involve fine motor control, proprioception and stabilisation of the hip joint, given its very small size. Some studies have considered this muscle to be absent (or fused with other muscles; e.g. CFP) in emus (Haughton 1867; Patak & Baldwin 1998), which would be unusual for any birds. The origin and insertion of the muscle that we label the ISF is best interpreted as a reduced -- but still present – muscle, similar to that in ostriches (Gangl et al. 2004; Zinoviev 2006).

M. caudofemoralis pars pelvica (CFP): We consider, contrary to other reports (Haughton 1867; Patak & Baldwin 1998), that this muscle is present in emus. Prior studies classified this muscle as the "iliofemoralis" but we agree with the *Nomina Anatomica Avium* (Vanden Berge & Zweers 1993) and other reports (Gangl et al. 2004; Hutchinson 2004a; Hutchinson et al. 2014; Zinoviev 2006) that it is present in ratites, related to a reduced portion of the large caudofemoralis muscle that is ancestrally present in tailed reptiles (Gatesy 1999). There is no evidence of a caudalis part to the M. caudofemoralis in emus, unlike in ostriches (Gangl et al. 2004) and some other ratites, so that is either fused to the CFP or lost.

M. ambiens (AMB): We found this muscle to have two insertions, previously unnoticed: a tendinous one onto the tibia and a fleshy one onto the distal femur. Unusual modifications of this muscle seem common in ratite birds (Hutchinson et al. 2014)

M. popliteus: This is a short, deeply positioned, fleshy muscle with multiple fibrous planes within it, originating on the caudolateral, proximal aspect of the tibiotarsus and inserting onto the medial side of the proximal fibula. It is likely a stabiliser or pronator/supinator of the fibula, as in ostriches (Fuss 1996), and may act a proprioceptive or ligament-like structure.

Normalized data for individual muscles

Normalized data allowing relative comparisons between muscles for mass, fascicle length and PCSA are presented in Figure 4. The largest relative muscles with regards to mass were three proximal (ILPO, ILFB and IC) and three distal muscles (GM, FL and GL). This order changes when muscles are ranked according to PCSA because parallel-fibred muscles tend to drop down the list, with the large ILPO being the only parallel muscle seen in the top 10 of a list that is otherwise dominated by distal muscles (FL, GM, FPDIII, TC and GL). On the other hand, when fascicle length is compared, the three parts of the gastrocnemius (GIM, GM and GL) are the only distal muscles listed amongst the 10 longest fibred muscles of a ranking that is topped by the FCLP, IC, ILFB and ILPO.

Limb muscle masses

~~When~~ The sum of all mass values of the limb musculature ~~were added, these~~ represented a mean of 13.4±3% of BM, with the proximal limb musculature (PLM) representing 61±2% of limb muscle mass and the distal limb muscles (DLM) accounting for the remaining 39±2%. However, if only values for the six larger birds (adults) are analysed, limb muscle mass accounts for 14.8±1% of BM, but the limb muscle mass is only 10.8%±3 of body mass in the five birds that were 4-6 weeks old.

Scaling regression analysis

The slopes of the reduced major axis regression lines for muscle properties vs. body mass are shown in Tables 3 and 4, with R^2 and 95% CIs, as well as represented in Figures 5 (A and B) and 6. Ranges of the slope and amplitudes of the CIs referred to below are the upper and lower bounds of the 95% CIs for the regression slopes. Scaling exponents and CIs are presented in Table 3. Scaling exponents and isometry border lines are plotted in Figure 5A and 5B (M_m, L_f and PCSA) and Figure 7 (M_{ten}, L_{ten} and TCSA). In summary, there was strong positive allometry of muscle mass and mild positive allometry or isometry of fascicle length, leading to a marked positive allometry of PCSA.

Scaling of limb muscle masses

We found limb muscle mass as well as the masses of proximal (PLM) and distal limb muscles (DLM) to be tightly correlated with body mass across all three groups. The regression slope of limb muscle mass vs. BM was 1.16 (1.05<CI<1.29, R^2=0.96), whilst PLM had a value of 1.14 (1.02<CI<1.27, R^2=0.96) and DLM exhibited a slope of 1.20 (1.09<CI<1.32, R^2=0.97) (Table 3).

When the mass of each muscle was analysed against body mass, overall the trend was towards positive allometry. Out of 34 muscles, 26 had slopes for M_m vs. BM with their lower CI limit >1 (consistent with positive allometry), and only eight (ITCr, ITM, IFI, ISF, FCLA, FMTL, AMB and FPPDII) had a lower CI boundary for the regression slope lower than 1 (indicating potential negative allometry). Of the 26 muscles showing positive allometry of M_m, we found strong positive allometry (regression slopes with the lower boundary of the CI greater than 1.1) in 18/34. The CI amplitudes were lower than 0.4 for 25 muscles and higher for the remaining nine. The R^2 values demonstrated tight correlations between M_m and BM, with 26/34 muscles having R^2 values >0.9 and only two muscles (ITM=0.73; GIM=0.76) having R^2 values <0.8.

Similarly, scaling patterns of the muscle masses for the USA group of emus (Figure 6), showed similar scaling patterns to the UK group, with only five muscles having a lower CI boundary <1 (POP, ILPO, FPDIV, OBTII and FPDII) and the remaining having their CIs entirely within positive allometry values. R^2 values for the muscles measured in this group were similarly high, with 30/32 muscles having values greater than 0.9.

Scaling of muscle fascicle length

In general, fascicle length (L_f) was only moderately well correlated with body mass due to substantial variation in the data (a combination of inevitable measurement errors, sampling bias and true biological variation, as usual for muscle fascicle measurements (e.g., Allen et al. [2010,2014]) The datasets for four muscles (ISF, PIFLM, FPDII and FPPDII) had a *p* value>0.05, so these are not presented (Table 3). Of the remaining 30 muscles, only 16/30 had R^2 values >0.5. Scaling of L_f vs. BM showed a trend towards positive allometry for 18/30 muscles (lower limit of the slope's CI >0.33), and for the remaining 12 muscles a slope of 0.33 was included in the CIs, so isometry could not be ruled out. The amplitude of the CI was <0.4 for 17 muscles and lower than 0.6 for 4 muscles. Five muscles had a CI amplitude greater than 0.6.

Scaling of muscle PCSA

R^2 values greater than 0.9 were found for only eight muscles (IC, ILPO, ILFB, IFE, FCLP, GL, GM and FL), but only two (ITM and FPPDII) had a value <0.5. The lower boundary of the CIs of the scaling slope was greater than 0.66 for 27 muscles (i.e., exhibiting positive allometry) and a value <0.66 (suggesting a potential negative allometry of muscle PCSA in emus) was seen for eight muscles (ITM, ITC, IFI, FMTL, AMB, TC and FPPDII); however, all of these had a value of the upper CI higher than 0.66, so those results were indistinguishable from isometry. The CI ranges were narrow for 15 muscles (<0.4) and lower than 1.0 for the remaining (Table 3).

Scaling of tendon mass

We recorded tendon characteristics for 28 muscles (Table 4); the six muscles excluded did not have a discrete tendon at either of their attachments (CFP, FCLA, FCLP, IC, PIFLM, POP). We encountered difficulties in achieving a consistent method for tendon dissection and measurement of muscles with thin (IFE, AMB), very short (ISF and IFI) or multiple tendons (FMTM, FMTIM), which lead us to exclude data from these as well. The tendon of the GIM was included with the GM tendon, and the FMTL tendon was not measured because the muscle was transected at the proximal aspect of the large patellar tendon for studies of patellar tendon morphology by Regnault et al. (2014). Thus data are presented for the tendons of 20 muscles. The major gastrocnemius tendon resulting from the fusion of the three gastrocnemius muscles was dissected by transecting the tendon of the GL at the site of insertion of onto common tendon; therefore the GM remained with the extensive common portion of the tendon, which distally was transected at its insertion onto the fibrous scutum at the level of the ankle joint.

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In general, we detected a strong correlation between M_{ten} and BM in 7/20 tendons (FL, FPDIII, FPPDIII, FPDIV, FHL, FDL and EDL), with an R^2 value >0.9. For a further 7/20 tendons we obtained $0.75<R^2<0.9$. The ITM tendon had a low R^2 value of 0.4, and the remaining five muscles had $0.75>R^2>0.5$. The scaling slopes for tendon mass indicate positive allometry in 10 out of 20 tendons (lower CI boundary >1) across emu ontogeny. The remaining 10 had this lower boundary <1, so the kind of scaling exhibited by these tendons is inconclusive because the upper limit of the CI was >1.2. The ranges of CIs for these slopes were narrow for 6/20 muscles (≤0.4), moderate for 8/20 (0.4<CI<0.6) and six tendons had a large CI range (>0.6). Notably, apart from FPPDII, all digital flexors and other muscles with relatively long tendons had higher R^2 values and narrower CI ranges.

Scaling of tendon length

We measured L_{ten} for 20 muscles (Table 4); same as for masses; from the end of the muscle belly to the insertion. Statistical analysis for one muscle (ITCr) excluded the values for this muscle because the p value >0.05. For the other 19 tendons, the general scaling trend was towards strong positive allometry, with 16 muscles having the lower limit of the CI >0.33 and the remaining three (FCM, GM, FL) with an upper limit of the CI that was significantly higher (>0.43) than 0.33. However, exponents indicating negative allometry are included in the CIs for all of the latter tendons, and hence isometry (or negative allometry) could not be conclusively excluded, although we infer a potential trend for positive allometry of tendon length in growing emus. In general, tendon length was poorly correlated with BM, because only nine muscles had R^2 values greater than 0.7 (presumably due to errors and variation, as noted above for L_F). However, similar to the tendon mass and BM correlations, all digital flexors apart from FPDII and the FPPDII had among the best R^2 values (>0.7). The confidence intervals of the scaling slopes had a range of <0.4 for 11 tendon lengths.

Scaling of tendon cross-sectional area

Average TCSA was calculated for the same 20 tendons as above (Table 4). The dataset for ILPO had a *p* value >0.05 and was excluded. Of the 19 remaining tendons, 10 showed a lower CI limit of the slope consistent with positive allometry (>0.66). Of the tendons with a lower CI limit that included values indicating potential negative allometry, the only digital flexor tendon in this group was the FPPDII. However, all tendons had the upper CI limit >0.66, indicating possible isometry or positive allometry as well, so overall our results for tendon CSA were inconclusive. The CI interval was <0.4 for 6 muscles (GM, FL, FPPDIII, FPDIV, FHL and FDL) and >0.6 in 9/19 muscles (ITC, ITCr, ITM, FCM, ILFB, GL, FPDII, FPPDII and EDL). R^2 values were >0.8 for eight tendons (FCM, GM, FL, FPDIII, FPPDIII, FPDIV, FHL and FDL) but <0.5 (indicating poor fit of the data to the regression line) for 6 tendons (ITC, ITM, ILFB, OMII, OMIP and FPPDII), with the latter one being the only distal limb and relatively long tendon in this group.

362
363
364 **Discussion**

365 Our dissections refined the myology of the pelvic limb in emus (Table 1 and Figures 1-3) and found
366 some anatomical aspects that were previously misunderstood. This is important, as functional
367 studies depending on inaccurate anatomical accounts of the myology could obtain unrealistic results
368 from biomechanical models using such data (Goetz et al. 2008; Hutchinson et al. 2014). We have
369 also measured muscle architectural parameters including mass, fascicle length and angle of
370 pennation, in addition to calculating physiological cross sectional areas, for 34 pelvic limb muscles.
371 We also obtained masses and lengths from 20 tendons and calculated their average cross sectional
372 areas. Finally, we measured bone lengths from four bones. All of these data were acquired from an
373 ontogenetic series of birds ranging from 4 weeks to 18 months of age (near sexual maturity). With
374 the use of regression analysis, we estimated the slope of each of these parameters vs body mass and
375 with this estimated the scaling trends for each of them. This dataset is the first of its kind for a large,
376 cursorial bipedal animal and is key to the understanding of ontogenetic adaptation strategies and
377 trade-offs occurring in the locomotor system of this remarkable avian species.

378 Emus, like other ratites and some other birds, must have locomotor independence from hatching
379 (termed precociality) and develop into large, running adult birds within 16-18 months (Davies &
380 Bamford 2002). Taking into consideration their initial development within the egg, their ontogeny
381 poses interesting questions about their locomotor development, related to our study's aims, such
382 as: How do muscle structure and anatomy change to accommodate precocial development of emus?
383 What are the strategies that growing emus use to maintain tissue mechanical safety factors during
384 rapid development of cursorial morphology and high-speed locomotor abilities? Our data suggest
385 some answers to these questions, as follows.
386
387 *Scaling patterns across ontogeny*
388 We found significant and strong positive allometry of emu pelvic limb muscle masses, indicating that
389 most get significantly more powerful (in relative and absolute terms) as the animals grow. However,
390 the functional relevance of this observation is slightly mitigated by a less marked positive allometry
391 of PCSA (and therefore maximal muscle force), driven by a trend for fascicle length that is closer to
392 isometry (i.e., preserving geometric similarity).

393 In the proximal part of the pelvic limb, the developmental and functional mechanism appears to rely
394 on the arrangement of large and metabolically expensive muscles (ILPO, ILFB, IC, FCLP and FMTL) to
395 provide the wide range of motion of the knee joint (and hip, during faster running) in combination
396 with a relatively short femur that scales close to isometry. This arrangement also leads to the
397 proximal to distal gradient of muscle mass previously reported for other birds (Paxton et al. 2010;
398 Smith et al. 2006), which has long been thought to favour energy-saving by keeping the distal end of
399 the limb light and its muscles dependent on springy tendons. The proximal-distal gradient also
400 concentrates large, power-generating muscles in the proximal limb (Alexander 1974; Alexander
401 1991) with large moment arms (Hutchinson et al. 2014; Smith et al. 2007) and thus the ability to
402 produce the considerable joint moments needed for high-speed running (Hutchinson 2004).

403 The distal limb, on the other hand, is heavily dependent on the triad of M. gastrocnemius (GL, GIM
404 and GM) along with M. fibularis longus (FL); both ankle extensors; as well as M. tibialis cranialis (TC)
405 and M. extensor digitorum longus (EDL); both ankle flexors. Combined, these muscles constitute
406 80% of the muscle mass and 60% of the force-generating capacity (PCSA) of this portion of the limb.
407 The unusual proportion of body mass taken up by the ankle extensors has been noted before
408 (Hutchinson 2004a; Hutchinson 2004b) and is characteristic of birds in general (e.g., (Paxton et al.
409 2010) but is taken to an extreme in large ratites (e.g. Smith et al. 2006).

Further distally, the long and slender tarsometatarsus bone lends itself well as a support for the long tendons of the digital flexor muscles which in turn provide essential springs used in support and propulsion of the limbs and body. The relatively small muscles and long tendons of the digital flexors make them likely to operate mainly as energy storage devices at faster speeds, as seen in other species like horses and smaller running birds (Biewener 1998; Daley & Biewener 2011). The positive allometry of many tendon properties is in line with this increase in force-generating capacity seen during ontogeny. As in most other birds, the tendons running along the tarsometatarsus are almost exclusively on the cranial and caudal (dorsal/plantar) side. It would also be interesting to know the effect on bone strains from this "bow and arrow" anatomical arrangement between the tarsometatarsus and the dorsal/plantar tendons to see if it might modify the predominantly torsional loads seen in the proximal two pelvic limb bones (Main & Biewener 2007).

For these spring-like tendons, a trade-off between muscle force and tendon elasticity does not seem to occur in emus. This lack of a trade-off is indicated by the similar scaling patterns of the cross-sectional areas of the digital flexor muscles and tendons, both of which trend towards positive allometry across emu ontogeny. As seen in other species (Ker et al. 1988), the relative increases in the cross-sectional areas of tendons might maintain tendon safety factors (maximal stresses before failure vs. *in vivo*) as the birds increase in size. However, tendons might also change their biomechanical properties (stiffness; Young's modulus) with age, as seen in other species (Shadwick 1990; Thorpe et al. 2014), therefore influencing biomechanical interpretations of the data presented here.

To compliment data from Main et al., who showed the scaling patterns of the cross-sectional areas of the femur and tibiotarsus of emus to be close to isometry (Main & Biewener 2007), here we analysed the scaling patterns of the lengths of the four limb bones and the third toe. Our data indicate positive allometry of the two longer bones, the tibiotarsus (lower CI limit=0.37) and tarsometatarsus (lower CI limit=0.39), but a less marked positively allometric scaling trend for the femur (lower limit of CI=0.34) and for the first phalanx of digit III (lower CI limit=0.33). These results differ from those reported for another ratite, the greater rhea (*Rhea americana*), where only the tarsometatarsus showed positive allometry (Picasso 2012a). Considering our results, if similar cross-sectional geometry is assumed along the length of the bone shafts, this would lead to an increase in strains (at least for bending) at the mid-shaft with increasing body mass. The increase of strain magnitudes in the femur and tibiotarsus across ontogeny in birds (Main & Biewener 2007) can be explained as a change in cross-sectional area geometry whilst relative limb loads are maintained during growth (Doube et al. 2012; Main & Biewener 2007). Although this geometrical change might suffice to explain the increase in strain magnitudes during ontogeny, it leaves unanswered two questions: First, how might internal forces (of soft tissues) influence bone mechanics and therefore adaptation during growth? Second, what is the mechanical environment in areas of bone distant to the commonly measured mid shaft?

Comment [TW9]: Suggest state femur, tibiotarsus, tarsometatarsus and proximal phalanx of digit III – because there are more than 4 limb bones and you did not use all bones of the third toe…

Although there are very limited data on the ontogeny of skeletal muscle physiology, experiments in mice and cats (Close 1964; Close & Hoh 1967) demonstrate that although muscle force: velocity parameters change from newborns to adults, these changes appear to occur in a relatively short period and therefore newborn muscle, after the first few days of life, becomes similar to that of adults. However, mice and cats, like many other mammals, are born with neuromotor immaturity (Muir 2000), in contrast to emus. It is therefore reasonable to speculate that, like other birds (Gaunt & Gans 1990), emus are unlikely to have appreciable changes in muscle physiology during growth. Thus changes in functional (e.g., maximal force generating capacity) and biomechanical parameters should be detectable by anatomical studies such as ours.

Few studies have quantified the ontogenetic scaling patterns of limb musculature in birds (Carrier & Leon 1990; Dial & Carrier 2012; Paxton et al. 2014; 2012b), but positive allometry predominates in

458 the muscle masses involved in the major adult mode of locomotion (flying vs. cursorial). In the
459 Californian gull, the M. gastrocnemius scaled isometrically but the M. pectoralis had strong positive
460 allometry with an inflection point when the fledglings started exercising their wings (Carrier & Leon
461 1990). Paxton et al. (2014; also 2010) recently reported the ontogenetic scaling patterns of the
462 musculature of a highly modified galliform, the broiler chicken. These birds, unsurprisingly due to
463 their selective breeding, were found to have positive allometry of muscle mass of the main pelvic
464 limb muscles but isometry of the fascicle lengths (Paxton et al. 2014), a pattern that is nonetheless
465 similar to our findings. Picasso et al. (2012b) found quite similar scaling patterns across rhea
466 ontogeny: an average 64-fold increase in pelvic limb muscle mass from 1 month of age to adulthood
467 whilst only a 34 fold increase in body mass. Together, these data suggest that positive allometry
468 prevails across ontogeny for leg muscles in all extant birds with precocial development; potentially a
469 homologous feature of their development that is quite unlike the isometry prevailing in their closest
470 extant relatives, Crocodylia (Allen et al., 2010,2014).

Comment [TW10]: Suspect this is a bit of an over intrepretation – surely all these birds have long tmt relative to femora – I expect of those many birds with tmt = femur length or less this pattern would not prevail.

471 In birds, ontogenetic allometric patterns for muscle architecture might be exaggerated for two
472 reasons. Birds have a more limited embryonic space in their eggs and tend to be born relatively
473 smaller than other species. Alternatively, as suggested by Dial and Carrier (2012), birds must
474 optimise their energy consumption to achieve their ultimate functional gait whilst channelling
475 resources to their precocial gait (Dial & Carrier 2012) (running vs. swimming or flying). Ratites are
476 unusual for birds in that they solely have terrestrial gaits throughout their life and, in the case of
477 emus, their wings have atrophied to such an extent that they should not present much metabolic
478 competition to hindlimb development. Considering the approximately isometric scaling of kinematic
479 parameters seen in ratites (Main & Biewener 2007; Smith et al. 2010), it is likely that this increase in
480 muscle masses will lead to limb design that is adapted for power production and manoeuvrability.
481 The former is also supported by metabolic studies which found a predominance of fast fibres in the
482 M. gastrocnemius of emus (Patak 1993), although more studies of muscle physiology in emus and
483 other ratites would be valuable.

484 The need for locomotor independence and high performance in vulnerable, young, precocial and
485 cursorial birds might favour the aforementioned characteristics (Carrier 1996). If so, could adult
486 muscle phenotypes be a reflection of the locomotor needs during early development and therefore
487 be overdesigned for their demands? Alternatively, negative allometric scaling of such features may
488 occur as seen in goats (Main & Biewener 2004) and jackrabbits (Carrier 1983). It is hard to draw an
489 inference from our data, because the overall positive allometry seen in the pelvic limb musculature
490 could indicate a necessity to grow faster and stronger to adulthood to compensate for a juvenile
491 disadvantage or could reflect selective pressures on the locomotor ontogeny of emus in which
492 muscles congenitally primed for fast growth during adolescence could lead to continued growth past
493 an optimum in adulthood. Although direct measurements of maximal performance of complex
494 locomotor systems is problematic, a modelling approach using the data presented here, would be a
495 valid approach to answer this question.

496 *How well are farmed emus representative of the species overall?*
497 Although emu farming is relatively common, its goal is to extract meat, oil and skin and therefore
498 these birds are not bred in captivity for their locomotor behaviour, nor do they suffer strong
499 predatory pressures on it. The diet of captive bred birds as well as their relative sedentary regime
500 when compared to wild animals is likely to influence tissue development and distribution. However,
501 as farming of these birds is a recent activity and it is not a highly specialised or intense process as
502 with other domesticated species (Goonewardene et al. 2003), it is unlikely that heritable traits of the
503 emu musculoskeletal system have been significantly altered. Therefore, we expect the muscle
504 distribution and scaling patterns of our emus to be similar to wild emus.
505

By presenting muscle mass data from two distinct groups of birds (UK and USA groups), we established that these groups at least have similar scaling patterns, ruling out any potential bias imposed by different breeding regimes. With regards to diet, it was apparent that our birds were carrying a significant amount of subcutaneous and peritoneal fat; likely encouraged by their *ad libitum* access to a commercial pelleted diet. Although the emus had no restriction in exercise in their large enclosure, they tended to occupy a small area of the field near the entrance and exercised only a few times a day when stimulated by an event like an unfamiliar bird (e.g., crows) eating their food or animal movements in the adjacent fields (e.g., horses and sheep). At approximately 14 months of age and coinciding with the onset of sexual maturity, the birds became more territorial, and the females more dominant and aggressive towards certain males. This led to an increase in the group's activity from that period onwards, but given that the timing of this increase was so close to the euthanasia of the birds we deem it unlikely that it changed their muscle architecture. Finally, but harder to infer, would be the possibility that weather differences from their natural habitat could lead them to deposit greater amounts of body fat during the winter season when bred in cooler climates like the one in the UK.

Comment [TW11]: Does not seem relevant to muscle mass – so long as all birds had the same 'condition factor' then muscle mass as proportion of body mass is unaffected – remove this from here and put comment in methods that condition was simiular among birds.

None of the above issues can be tested with available data, but Hutchinson et al. (2014) noted a possible reduction in relative muscle masses in wild vs. captive bred ostriches, which could also apply to emus. Regardless, it is less certain that the scaling patterns for muscle/tendon architecture observed here would differ in wild vs. captive emus.

Conclusions

We have provided a new dataset on the ontogenetic scaling of pelvic limb anatomy and muscle architectural properties of a cursorial bird (the first complete architectural dataset of its kind), and we have done this using a group of 17 emus across a tenfold increase in body mass. A marked trend of positive allometry of muscle masses and PCSAs is accompanied by less marked positive allometry of fascicle lengths. Tendons, specially the long digital flexors, also demonstrate positive allometry of their lengths, as do the two longer limb bones (tibiotarsus and tarsometatarsus). We have also provided anatomical descriptions, illustrations and clarifications of the pelvic limb anatomy of the emu. This work should be a valuable resource for future functional, comparative and evolutionary studies of emus, other birds and extinct related animals, and we have illuminated the ontogenetic adaptation of the musculoskeletal system in an extreme example of size variation during rapid growth.

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548 **Tables and table captions**

549 **Table 1.** Pelvic limb muscles of emus and their apparent actions.

Muscle	Abbreviation	Origin	Insertion	Action
M. iliotibialis cranialis	IC	Dorsal edge of preacetabular ilium	Insertion on the medial aspect of the proximal tibiotarsus	Main: Hip flexion; knee extension/flexion Other: Hip medial rotation, adduction
M. iliotibialis lateralis (cranial and caudal portions)	ILPO	Lateral edge of acetabular ala	Craniolateral proximal tibiotarsus (cranial and lateral cristae cnemiales) via aponeurosis	Main: Hip extension,abduction; knee extension Other: Hip medial/lateral rotation
M. iliotrochantericus cranialis	ITCr	Cranial surface of preacetabular ilium	Lateral aspect of the femoral trochanteric crest (distal to IFE insertion)	Main: Hip flexion, medial rotation Other: Hip abduction/adduction
M. iliotrochantericus medialis	ITM	Craniodorsal surface of preacetabular ilium	Lateral aspect of the femoral trochanteric crest (proximal to IFE insertion)	Main: Hip flexion, medial rotation Other: Hip abduction/adduction
M. iliotrochantericus caudalis	ITC	Ala preacetabularis ilii: fossa iliaca dorsalis	Lateral aspect of the femoral trochanteric crest	Main: Hip flexion, medial rotation Other: Hip abduction/adduction
M. iliofibularis	ILFB	Ala postacetabularis ilii: facies lateralis	Proximal third of the corpus fibulae	Main: Knee flexion;, hip extension Other: Hip abduction
M. iliofemoralis externus	IFE	Crista iliaca dorsalis, dorsal to foramen acetabulum	Lateral side of femoral trochanteric crest (between ITC and ITM insertions)	Main: Hip flexion, abduction Other: Hip medial/lateral rotation
M. iliofemoralis internus	IFI	Ventral preacetabular ilium	Medial side of proximal femoral shaft; tubercle	Main: Hip flexion, adduction Other: Hip medial/lateral rotation
M. ischiofemoralis	ISF	Cranial margin of the foramen ilioischiadicum	Proximal caudal femur under origin of FMTL	Main: Hip abduction, lateral rotation Other: Hip flexion/extension
M. caudofemoralis p. pelvica	CFP	Caudolateral ilium and ischium	Proximal caudomedial femur	Main: Hip extension Other: Hip lateral rotation, abduction
M. flexor cruris lateralis pars pelvica	FCLP	Caudolateral corner of pelvis	Proximal craniomedial tibiotarsus	Main: Hip extension, abduction Other: Medial rotation of hip and knee; knee flexion
M. flexor cruris lateralis pars accessoria	FCLA	By a raphe from the distal third of the FCLP	Caudomedial femoral shaft	Main: Hip extension, abduction Other: Hip medial rotation
M. flexor cruris medialis	FCM	Caudolateral extremes of ischium and pubis	Via split cranial aponeurosis: on the caudal femoral shaft, and on the caudoproximal	Main: Hip extension, abduction; knee flexion Other: Hip medial rotation

Comment [TW12]: It would be greta to have the insertions etc shown on diagrams of the bones!

			tibiotarsus, caudodistally to the insertion of the FCLP.	
M. puboischiofemoralis p. lateralis and p. medialis	PIFLM	Along the length of the lateral ischium	Via thin tendinous insertion onto the caudal aspect of the femoral shaft	Main: Hip extension, abduction Other: Hip lateral rotation
M. femorotibialis lateralis (Cranial, intermediate and caudal portions)	FMTL	Caudolateral surface of femoral shaft. With 3 fused parts: cranial, intermediate and caudal	Crista cnemalis of tibiotarsus via a thick patellar tendon (no ossified patella) with ILPO	Knee extension
M. femorotibialis intermedialis	FMTIM	Cranial surface of the proximal femoral shaft	Medial side of crista cnemalis cranialis of tibiotarsus	Knee extension
M. femorotibialis medialis	FMTM	3 distinct heads originating from the medial surface of the femur, cranial and caudal portions on the proximal third and distal portion on the distal third	Proximo-medial extremity of tibiotarsus	Knee flexion, adduction
M. obturatorius medialis (Ilium – Ischium part)	OMII	Surface of fenestra ilioischium	Long tendon that passes through the foramen ilioischiadicum and inserts onto the lateral side of the femoral trochanteric crest	Main: Hip lateral rotation Other: Hip flexion, adduction
M. obturatorius medialis (Ischium – pubis part)	OMIP	Surface of fenestra ischiopubica	As OMII	Main: Hip lateral rotation Other: Hip flexion, adduction
M. ambiens	AMB	Cranial pubic rim (preacetabular process)	Two insertions on the medial knee ligaments, one tendinous and the other one fleshy	Main: Hip adduction; knee flexion Other: Hip medial rotation
M. gastrocnemius lateralis	GL	Lateral condyle of femur, aponeurosis of M. Iliotibialis and tendon from cranial fibula	Tendons fusing to form a thick fibrous calcaneal pad, onto caudal side of tarsometatarsus (Calcaneal scutum)	Main: Ankle extension; knee flexion
M. gastrocnemius medialis	GM	Aponeurosis of M. Iliotibialis and facies gastrocnemialis, connecting to the medial surface of the proximal tibia	As GL	Main: Ankle extension; knee flexion
M. gastrocnemius Intermedius	GIM	Craniolateral femur, adjacent of the origin of FHL muscle	As GL and GIM	Main: Ankle extension; knee flexion
M. fibularis longus	FL	Proximal origin from medial distal patellar ligament and craniolaterally onto proximal tibiotarsus.	Two tendinous insertions: Plantar calcaneal scutum and joining the tendon of FPDIII	Main: Ankle extension Other: Knee flexion; toe flexion via FPDIII tendon
M. tibialis cranialis c. tibiale and c. femorale	TC	2 heads: A fleshy one onto the proximal cranial tibiotarsus, and via a thick tendon onto the cranial aspect of the lateral trochlear ridge of the femur	Cranial side of proximal tarsometatarsus	Main: Ankle flexion Other: Knee extension (femoral head)
M. popliteus	POP	Medial side of proximal fibula	Caudal side of proximal tibiotarsus	Main: Fibular rotation
M. flexor perforatus digiti II	FPDII	Via origin of FPDIII	Splits into 2 branches at level of proximal phalanx to insert on either side of middle	Main: Digit II flexion Other: Ankle extension

			phalanx, ventrally	
M. flexor perforatus digiti III	FPDIII	2 tendons: Cranial fibula and medial side of the medial condyle of the femur	Proximal phalanx, small portion fused to FPPDII tendon in some specimens, ventrally	Main: Digit III flexion Other: Ankle extension
M. flexor perforans et perforans digiti II	FPPDII	Deep fibular tendon of GL muscle	Middle phalanx of digit II, ventrally	Main: Digit II flexion Other: Ankle extension
M. flexor perforans et perforans digiti III	FPPDIII	Lateral knee ligaments and FPDIV origin	Middle phalanx of digit III, ventrally	Main: Digit III flexion Other: Ankle extension
M. flexor perforatus digiti IV	FPDIV	Superficial side of FPDIII origin	Proximal and middle phalanges of digit IV, ventrally	Main: Digit IV flexion Other: Ankle extension
M. flexor hallucis longus	FHL	2 heads: lateral and caudal aspects of distal femur near condyles	Fuses with FDL tendon	Main: Ankle extension; knee flexion
M. flexor digitorum longus	FDL	2 heads: proximal tibiotarsus and distal third of fibula (3/4 of length)	Splits into 3 parts above MTP joint to insert onto the distal, ventral phalanx of each toe	Main: Digits II, III and IV flexion Other: Ankle extension
M. extensor digitorum longus	EDL	Cranial proximal tibiotarsus	Dorsal surface of each phalanx	Main: Digits II, III and IV extension; ankle flexion

Bone	Scaling exponent	Lower 95% CI	Upper 95% CI	R^2
Femur	0.38	0.34	0.42	0.96
Tibiotarsus	0.41	0.38	0.45	0.97
Tarsometatarsus	0.44	0.39	0.49	0.96
First Phalanx (Dig III)	0.39	0.33	0.46	0.91

551 **Table 2.** Regression analysis results for the lengths of the four limb bones. The lower 95% boundary
552 (>0.33) demonstrates positive allometry of the tibiotarsus and the tarsometatarsus but results
553 are closer to isometry for the femur and first phalanx of digit III.

554

	M_m vs BM					L_f vs BM					PCSA vs BM				
Muscle	Outliers	Slope	Lower 95% CI	Upper 95% CI	R^2	Outliers	Slope	Lower 95% CI	Upper 95% CI	R^2	Outliers	Slope	Lower 95% CI	Upper 95% CI	R^2
AMB	0	1.08	0.96	1.21	0.96	0	0.42	0.31	0.57	0.67	0	0.81	0.64	1.03	0.81
CFP	0	1.18	1.09	1.28	0.98	0	0.48	0.31	0.73	0.36	0	0.94	0.78	1.13	0.89
EDL	0	1.25	1.10	1.41	0.95	0	0.54	0.39	0.75	0.64	0	0.82	0.67	1.01	0.86
FCLA	1	1.16	0.95	1.43	0.87	1	0.36	0.24	0.53	0.51	1	0.89	0.73	1.09	0.87
FCLP	0	1.26	1.16	1.36	0.98	0	0.33	0.24	0.44	0.69	0	0.99	0.89	1.09	0.97
FCM	1	1.31	1.16	1.48	0.95	1	0.60	0.39	0.91	0.42	1	0.95	0.75	1.20	0.83
FDL	1	1.29	1.15	1.44	0.96	1	0.58	0.37	0.90	0.36	1	0.93	0.76	1.15	0.86
FHL	1	1.22	1.04	1.42	0.93	1	0.66	0.42	1.04	0.34	1	0.98	0.70	1.37	0.64
FL	0	1.32	1.23	1.42	0.98	0	0.44	0.33	0.58	0.73	0	0.98	0.84	1.16	0.91
FMTIM	0	1.24	1.05	1.48	0.90	0	0.64	0.43	0.97	0.40	0	0.99	0.70	1.41	0.57
FMTL	0	1.19	0.95	1.49	0.83	0	0.43	0.31	0.60	0.64	0	0.86	0.65	1.14	0.73
FMTM	0	1.29	1.05	1.59	0.86	0	0.45	0.29	0.70	0.31	0	0.99	0.80	1.22	0.85
FPDII	0	1.45	1.26	1.67	0.93		-	-	-	-	0	1.40	1.06	1.84	0.74
FPDIII	0	1.34	1.19	1.51	0.95	0	0.60	0.41	0.88	0.47	0	1.03	0.78	1.36	0.74
FPDIV	0	1.20	1.09	1.32	0.97	0	0.43	0.28	0.65	0.38	0	0.99	0.80	1.22	0.85
FPPDII	0	0.75	0.59	0.95	0.81	0	0.74	0.49	1.14	0.37	0	0.68	0.44	1.07	0.29
FPPDIII	0	1.29	1.15	1.45	0.96		-	-	-	-	0	0.98	0.72	1.34	0.67
GIM	0	1.32	1.01	1.73	0.75	0	0.46	0.34	0.63	0.69	0	1.03	0.72	1.48	0.54
GL	0	1.30	1.19	1.43	0.97	0	0.51	0.40	0.67	0.77	0	0.88	0.76	1.01	0.93
GM	0	1.24	1.14	1.33	0.98	0	0.34	0.26	0.43	0.77	0	0.93	0.82	1.06	0.95
IC	0	1.27	1.15	1.40	0.97	0	0.31	0.24	0.39	0.81	0	1.00	0.88	1.13	0.95
IFE	0	1.26	1.11	1.42	0.95	0	0.56	0.42	0.75	0.72	0	0.79	0.66	0.93	0.91
IFI	2	1.22	0.97	1.54	0.85	2	0.49	0.33	0.72	0.57	2	0.92	0.66	1.28	0.68
IB	0	1.32	1.22	1.42	0.98	0	0.36	0.30	0.44	0.89	0	0.98	0.89	1.07	0.97
ILPO	0	1.29	1.16	1.43	0.96	0	0.31	0.21	0.46	0.50	0	1.08	0.92	1.26	0.92
ISF	3	1.10	0.93	1.32	0.92	-	-	-	-	--	3	1.06	0.73	1.54	0.63
ITC	2	1.26	1.14	1.39	0.97	2	0.76	0.61	0.95	0.86	2	0.64	0.50	0.81	0.84
ITCr	0	1.16	0.99	1.36	0.92	0	0.37	0.27	0.50	0.68	0	0.89	0.70	1.13	0.80
ITM	2	1.12	0.83	1.51	0.75	2	0.78	0.49	1.23	0.39	2	0.89	0.55	1.45	0.29
OMII	0	1.23	1.10	1.39	0.95	0	0.73	0.46	1.15	0.27	0	1.05	0.76	1.45	0.65
OMIP	0	1.23	1.11	1.36	0.97	0	0.53	0.36	0.77	0.49	0	0.94	0.77	1.15	0.87
PIFLM	0	1.24	1.13	1.36	0.97	-	-	-	-	-	0	1.11	0.89	1.39	0.83
POP	2	1.44	1.17	1.76	0.88	2	0.68	0.41	1.13	0.22	2	1.15	0.88	1.51	0.79
TC	0	1.20	1.08	1.33	0.97	0	0.68	0.50	0.93	0.67	0	0.77	0.55	1.07	0.63

Table 3. Results of RMA linear regression of muscle architecture vs. body mass (BM) for the pelvic limb of *Dromaius novaehollandiae*, across ontogeny. M_m, muscle mass (kg); L_f, fascicle length (m), *PCSA*, physiological cross-sectional area (m^2).

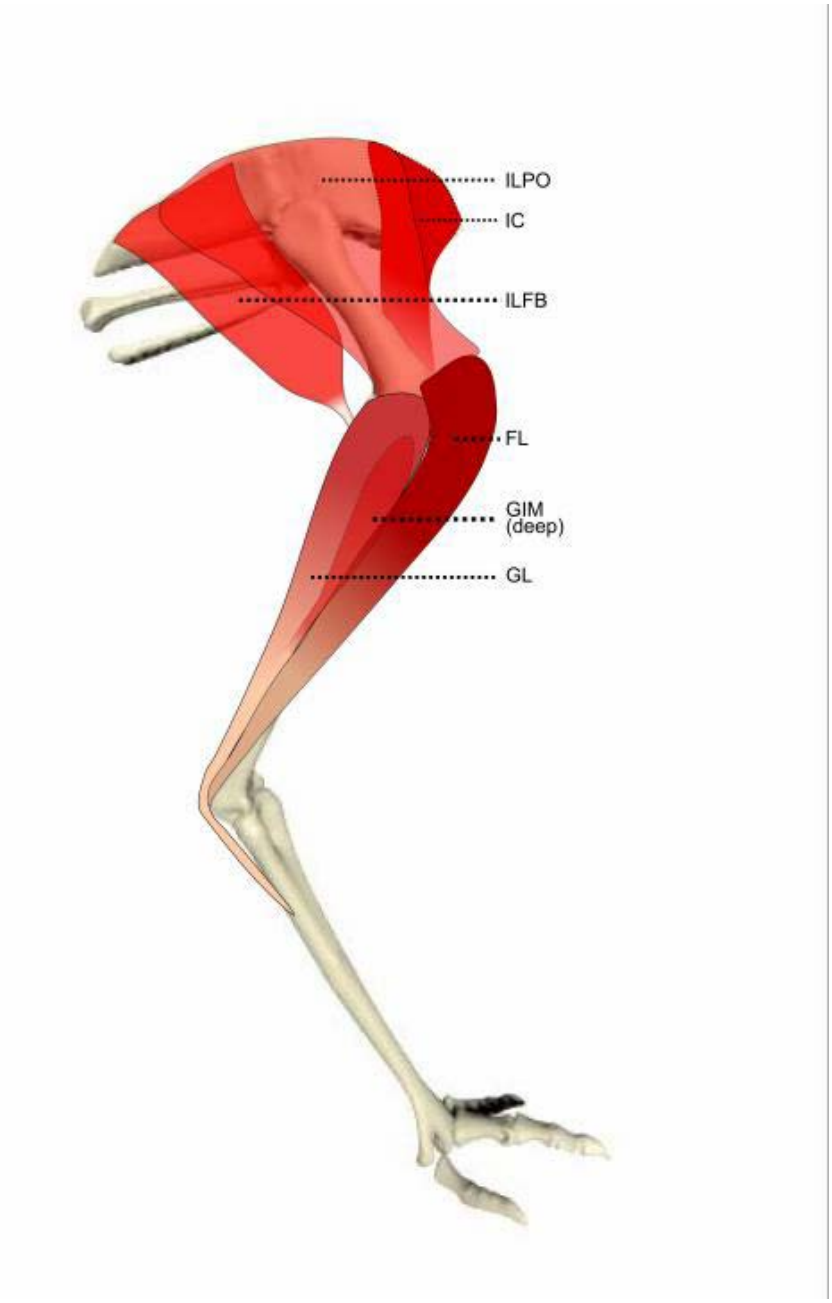
557

	M_{ten} vs BM					L_{ten} vs BM					TCSA vs BM				
Tendon	*Outliers*	*Slope*	*Lower 95% CI*	*Upper 95% CI*	R^2	*Outliers*	*Slope*	*Lower 95% CI*	*Upper 95% CI*	R^2	*Outliers*	*Slope*	*Lower 95% CI*	*Upper 95% CI*	R^2
EDL	0	1.26	1.10	1.44	0.94	1	-0.81	-1.07	-0.61	0.75	0	0.86	0.61	1.22	0.58
FCM	0	1.31	1.01	1.69	0.86	0	0.46	0.27	0.79	0.34	0	1.05	0.78	1.43	0.81
FDL	1	1.22	1.08	1.39	0.95	1	0.43	0.36	0.51	0.91	1	0.81	0.70	0.93	0.94
FHL	1	1.29	1.09	1.53	0.91	1	0.45	0.34	0.60	0.74	1	0.87	0.75	1.01	0.93
FL	0	1.33	1.15	1.52	0.94	0	0.39	0.31	0.50	0.81	0	0.99	0.82	1.20	0.88
FPDII	0	1.26	1.03	1.53	0.87	0	0.63	0.40	0.97	0.32	0	1.09	0.76	1.57	0.56
FPDIII	0	1.38	1.21	1.58	0.94	0	0.43	0.36	0.52	0.88	0	1.01	0.82	1.24	0.86
FPDIV	0	1.17	1.05	1.31	0.96	0	0.42	0.37	0.48	0.95	0	0.76	0.67	0.86	0.95
FPPDII	0	1.34	0.95	1.88	0.60	0	0.78	0.58	1.06	0.69	0	0.80	0.50	1.27	0.24
FPPDIII	0	1.24	1.06	1.44	0.92	0	0.43	0.38	0.49	0.95	0	0.83	0.68	1.03	0.85
GL	0	1.63	1.19	2.23	0.66	0	0.89	0.59	1.36	0.38	0	0.95	0.69	1.30	0.66
GM	0	0.98	0.78	1.23	0.83	0	0.28	0.18	0.43	0.37	0	0.79	0.64	0.97	0.85
IB	1	1.03	0.79	1.33	0.79	1	0.51	0.35	0.73	0.57	1	0.81	0.53	1.23	0.43
ILPO	2	1.38	0.99	1.93	0.68	2	1.04	0.69	1.56	0.51	-	-	-	-	-
ITC	3	1.04	0.81	1.33	0.84	3	0.61	0.44	0.83	0.74	3	0.75	0.46	1.22	0.34
ITCr	1	1.02	0.76	1.36	0.73	-	-	-	-	-	1	1.18	0.80	1.74	0.52
ITM	7	1.37	0.76	2.46	0.43	6	0.72	0.37	1.42	0.09	7	1.19	0.61	2.33	0.21
OMII	0	1.26	0.98	1.62	0.79	0	0.71	0.53	0.94	0.72	0	0.75	0.51	1.10	0.48
OMIP	0	0.99	0.74	1.33	0.70	1	0.48	0.36	0.65	0.71	1	0.67	0.44	1.02	0.43
TC	0	1.06	0.85	1.30	0.85	0	0.50	0.34	0.73	0.47	0	0.75	0.56	1.00	0.71

558 **Table 4.** Results of RMA linear regression of tendon dimensions vs. body mass (BM) for the pelvic limb of *Dromaius novaehollandiae*, across ontogeny. M_{ten},
559 tendon mass (kg); L_{ten}, tendon length (m); *TCSA*, tendon cross-sectional area (m^2).

560

561 **Figures and Figure Captions**



562

563 **Figure 1** Schematic anatomical representation of the most superficial layer of muscles, in lateral
564 view, for the pelvic limb of an adult emu.

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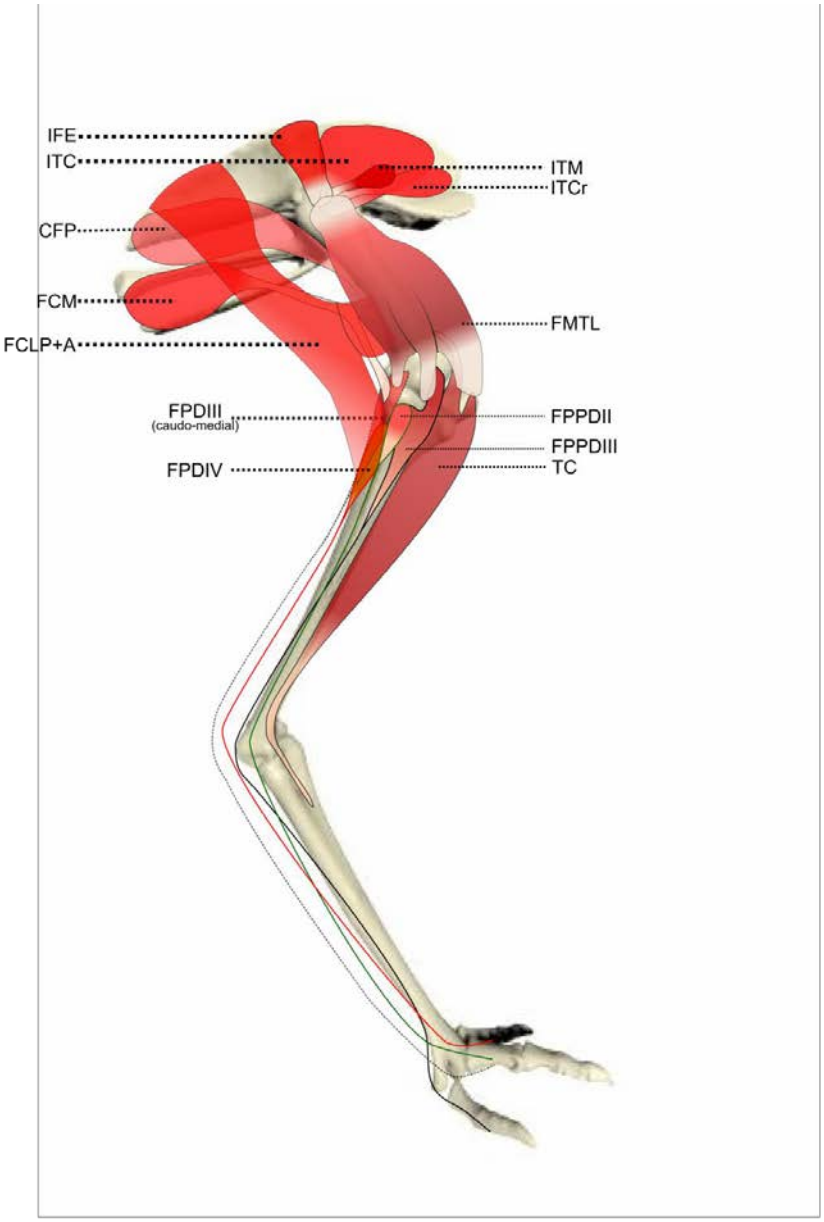
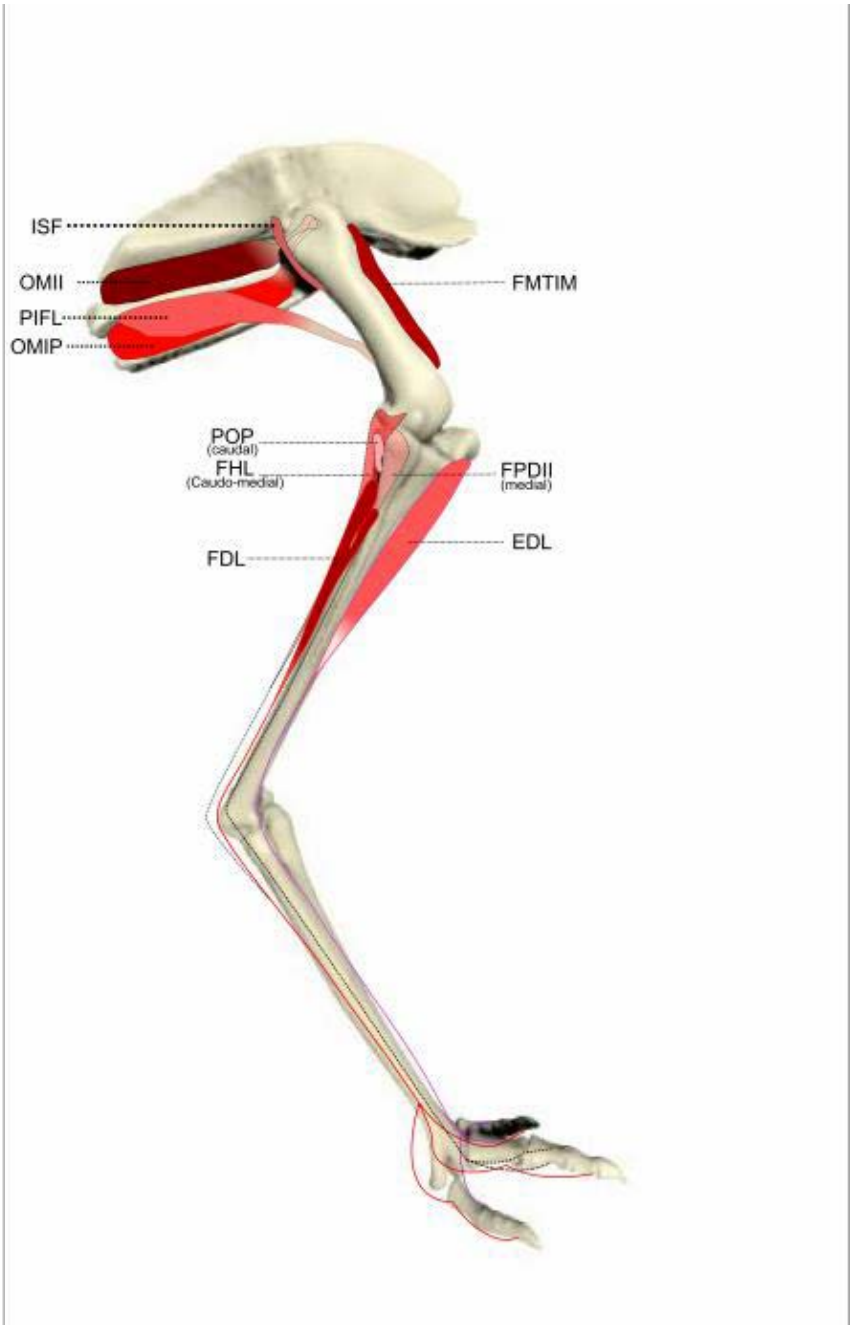


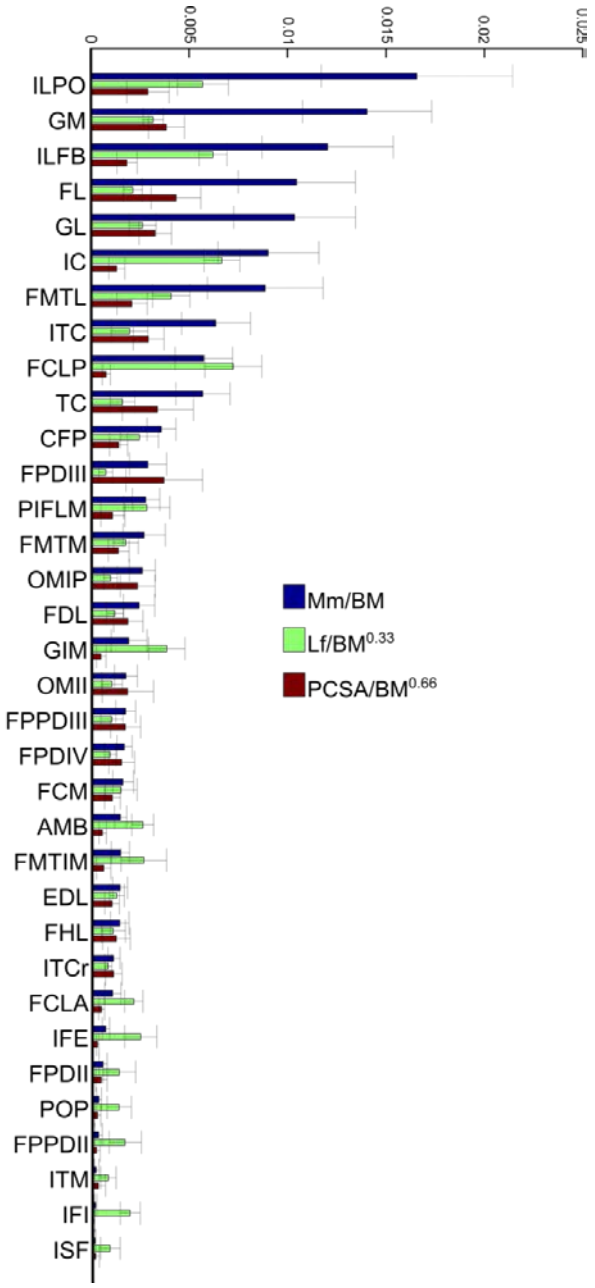
569

570 **Figure 2.** Schematic anatomical representation of the intermediate layer of muscles, from a lateral
571 view, of the pelvic limb of an adult emu.



572

573 **Figure 3.** Schematic anatomical representation of the deeper layer of muscles, from a lateral view, of
574 the pelvic limb of an adult emu.

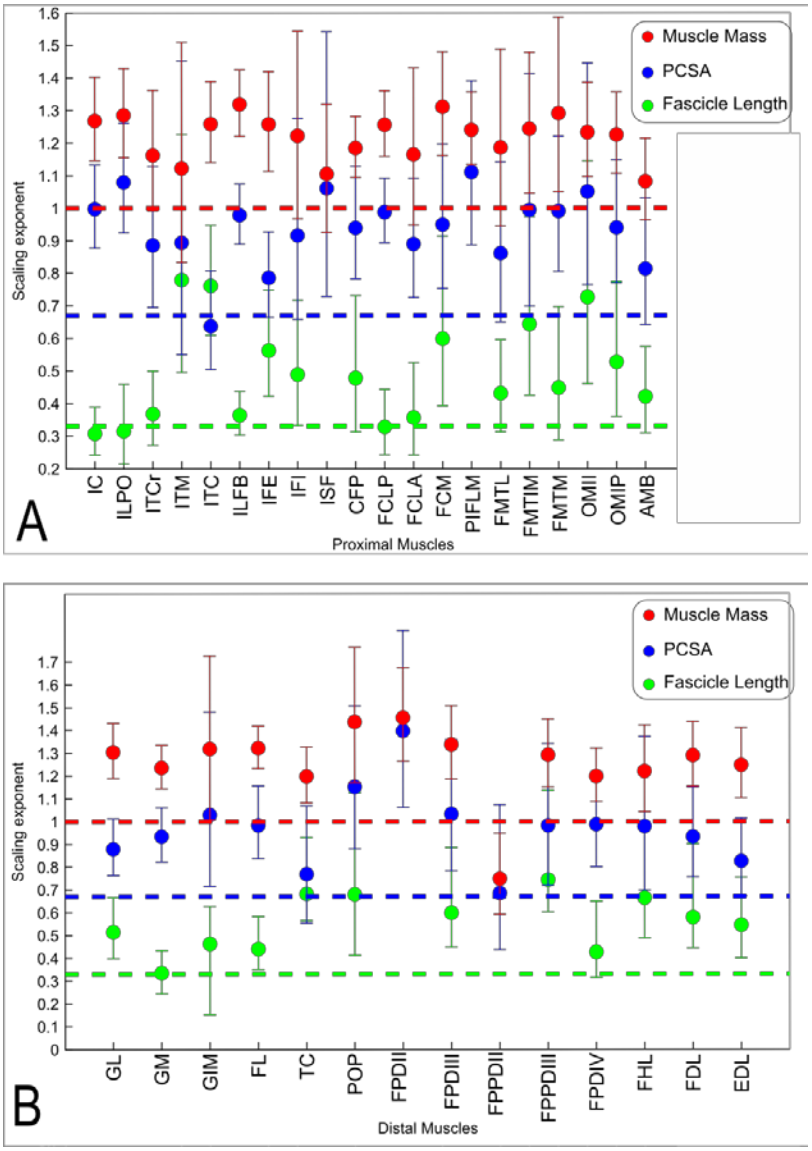


575

576 **Figure 4**. Normalized relative muscle parameters for individual muscles in emu pelvic limbs; mean
577 values (error bars showing ±1 S.D.) are shown. Abbreviations for muscles are in Table 1. The key on
578 the right side of the figure shows how muscle mass (M_m), physiological cross-sectional area (PCSA),
579 and fascicle length (L_f) were normalized. L_f values were adjusted to be 1/10 of the actual results in
580 order to be of similar magnitude to the others. Muscles are organised from top to bottom in
581 decreasing order of muscle mass.

582

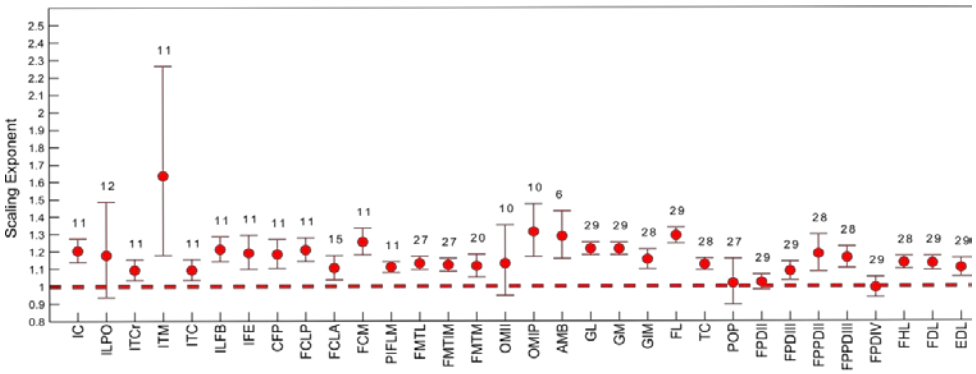
583



584

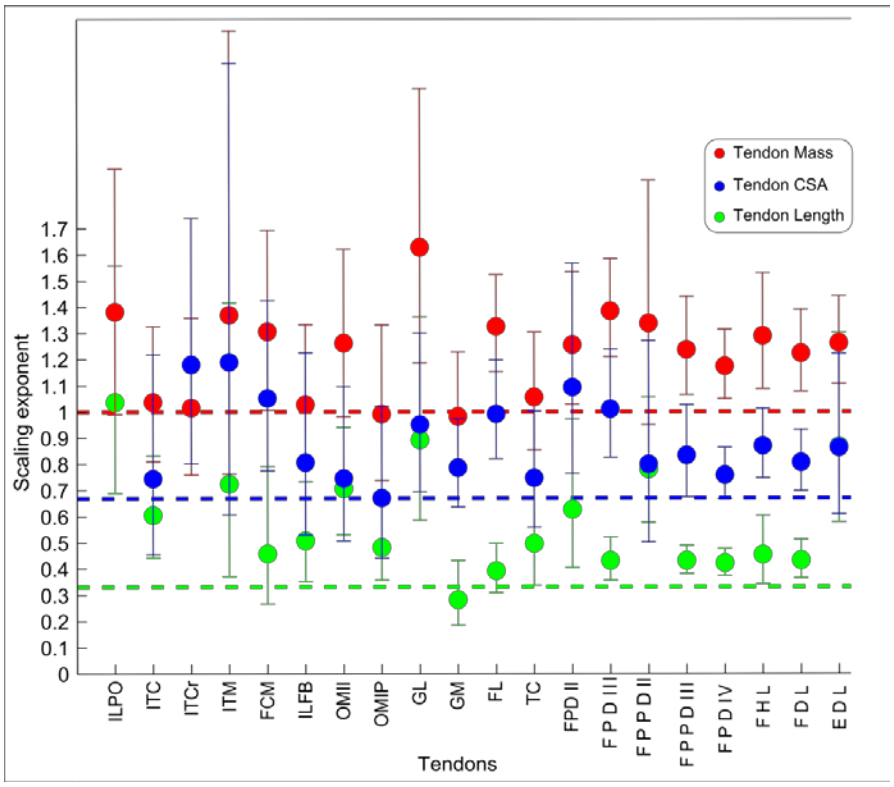
585

586 **Figure 5.** Ontogenetic scaling exponents and 95% confidence intervals (shown as error bars around
587 mean exponent) for muscle mass (red), PCSA (blue) and fascicle length (green) for individual muscles
588 in emu pelvic limbs. Abbreviations for muscles are in Table 1. Dashed lines indicate the expected
589 isometric scaling exponent for each parameter. Data for **A)** proximal limb muscles and **B)** distal limb
590 muscles.



591
592 **Figure 6.** Ontogenetic scaling exponents and 95% confidence intervals for masses of individual
593 muscles in emu pelvic limbs, from the USA group. Abbreviations for muscles are in Table 1. Dashed
594 line indicates the expected isometric scaling exponent (1.0), and the number above each parameter
595 indicates the number of muscles included in each regression analysis.

596



597

598 **Figure 7.** Ontogenetic scaling exponents and 95% confidence intervals for tendon mass (red),
599 average cross sectional area (blue) and length (green) for 20 individual muscles in emu pelvic limbs.
600 Abbreviations for muscles are in Table 1. Dashed lines indicate the expected isometric scaling
601 exponent for each parameter.

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