

Using GIS to Examine Biogeographic and Macroevolutionary patterns in some Late Paleozoic Cephalopods from the North American Midcontinent Sea

Kayla M. Kolis¹ and Bruce S. Lieberman^{1,2}

¹Biodiversity Institute, University of Kansas, Lawrence, Kansas 66045, U. S. A.

²Department of Ecology & Evolutionary Biology, University of Kansas, Lawrence, Kansas 66045, U. S. A.

Corresponding author:

Bruce S. Lieberman

E-mail address: blieber@ku.edu

Abstract

Geographic range is an important macroevolutionary parameter frequently considered in paleontological studies as species' distributions and range sizes are determined by a variety of biotic and abiotic factors well known to affect the differential birth and death of species. Thus, considering how distributions and range sizes fluctuate over time can provide important insight into evolutionary dynamics. This study uses Geographic Information Systems (GIS) and analyses of evolutionary rates to examine how in some species within the Cephalopoda, an important pelagic clade, geographic range size and rates of speciation and extinction changed throughout the Pennsylvanian and early Permian in the North American Midcontinent Sea. This period is particularly interesting for biogeographic and evolutionary studies because it is characterized by repetitive interglacial-glacial cycles, a global transition from an icehouse to a greenhouse climate during the Late Paleozoic Ice Age, and decelerated macroevolutionary dynamics, i.e. low speciation and extinction rates.

The analyses presented herein indicate that cephalopod species diversity was not completely static and actually fluctuated throughout the Pennsylvanian and early Permian, matching findings from other studies. However, contrary to some other studies, the mean geographic ranges of cephalopod species did not change significantly through time, despite numerous climate oscillations; further, geographic range size did not correlate with rates of speciation and extinction. These results suggest that pelagic organisms may have responded differently to late Paleozoic climate changes than benthic organisms, although additional consideration of this issue is needed. Finally, these results indicate that, at least in the case of cephalopods, macroevolution

23 during the late Paleozoic was more dynamic than previously characterized, and patterns may
24 have varied across different clades during this interval.

25 **Introduction**

26
27 Much work has focused on the relationship between geographic range size and rates of
28 speciation and extinction (e.g., Vrba, 1980; Jablonski, 1986; Eldredge, 1989; Stanley, 1990;
29 Lieberman, 2000; Jablonski & Roy, 2003; Rode & Lieberman, 2004, 2005; Kiessling &
30 Aberhan, 2007; Liow, 2007; Payne & Finnegan, 2007; Abe & Lieberman, 2009; Stigall, 2010;
31 Myers & Saupe, 2013; Myers, MacKenzie, & Lieberman, 2013; Dunhill & Wills, 2015;
32 Jablonski & Hunt, 2015; Orzechowski et al., 2015; Saupe et al., 2015; Castiglione et al., 2017;
33 Pie & Meyer, 2017; Simões et al., 2016; Lam, Stigall, & Matzke, 2018; Schneider, 2018).
34 Furthermore, the use of Geographic Information Systems (GIS) has greatly facilitated
35 investigations into this macroevolutionary relationship (Stigall & Lieberman, 2006; Hendricks,
36 Lieberman, & Stigall, 2008; Dunhill, 2012; Myers, MacKenzie, & Lieberman, 2013; Dunhill &
37 Wills, 2015; Lieberman & Kimmig, 2018). Here, we focus on how geographic range size and
38 rates of speciation and extinction changed throughout the Pennsylvanian and early Permian in
39 the North American Midcontinent Sea in the Cephalopoda, an important clade of pelagic
40 invertebrates (Kullmann, 1983, 1985; House, 1985; Becker & Kullman, 1996; Landman, Tanabe,
41 & Davis, 1996; Wiedmann & Kullmann, 1996; Monnet, De Baets, & C. Klug, 2011; Korn &
42 Klug, 2012; Klug et al., 2015; Korn, Klug, & Walton, 2015), using GIS. This time interval is
43 particularly interesting for biogeographic and evolutionary analysis because it is characterized by
44 repetitive glacial-interglacial cycles, a global transition from an icehouse to greenhouse climate
45 during the Late Paleozoic Ice Age (LPIA) (Montañez & Poulsen, 2013). Further, it is generally

46 considered a time of sluggish macroevolutionary dynamics, i.e. low speciation and extinction
47 rates and low degrees of faunal turnover, that have been demonstrated in studies of other marine
48 invertebrate taxa (Sepkoski, 1998; Stanley & Powell, 2003; Bonelli & Patzkowsky, 2011).
49 However, Ramsbottom (1981), Kullmann (1985), Becker & Kullmann (1996), and Wiedmann &
50 Kullmann (1996) did cogently argue that this was not the case for cephalopods. More recently,
51 Balseiro (2016) did document the existence of some profound evolutionary turnover in bivalves
52 and brachiopods over the course of this interval in regions closer to the ice sheets, such as
53 present-day western Argentina. Furthermore, Segessenman & Kammer (2018) showed that
54 advanced cladid crinoids do display elevated rates of evolution and turnover during this time
55 interval (although three other subclasses of crinoids do show subdued evolutionary rates), and
56 fusulinid foraminifera also fit the pattern shown in the advanced cladids (Groves & Lee, 2008;
57 Groves & Yue, 2009; Segessenman & Kammer, 2018).

58

59 There have been a variety of hypotheses proposed for the postulated decelerated
60 macroevolutionary dynamics (albeit not necessarily in cephalopods) of the LPIA. Some studies
61 contend that this pattern is a result of environmental changes linked to glacial cycling while
62 others point to tectonic activity (Stanley & Powell, 2003; Powell, 2005; Fielding, Frank, &
63 Isbell, 2008; DiMichele et al., 2009; Falcon-Lang & DiMichele, 2010; Bonelli and Patzkowsky,
64 2011; Cecil, DiMichele, & Elrick, 2014; Segessenman & Kammer, 2018). To date, many of the
65 more recent studies focusing on the macroevolutionary dynamics of the LPIA have concentrated
66 on benthic marine invertebrates (e.g., Stanley & Powell, 2003; Powell, 2007; Bonelli &
67 Patzkowsky, 2011; Balseiro, 2016; Segessenman & Kammer, 2018) as they are highly diverse
68 and very abundant. However, it is valuable to also investigate evolutionary patterns in pelagic

69 marine invertebrates as these are also diverse and abundant organisms in late Paleozoic marine
70 | ecosystems (Landman, Tanabe, & Davis, 1996; Monnet, De Baets, & Klug, 2011; Klug et al.,
71 | 2015; Korn, Klug, & Walton, 2015). In particular, given the significant role that geographic
72 factors play in speciation (Mayr, 1942; Eldredge & Gould, 1972; Jablonski, 1986; Brooks &
73 McLennan, 1991; Wiley & Lieberman, 2011; Jablonski & Hunt, 2015; Pie & Meyer, 2017), we
74 might expect that pelagic organisms, because of their innately greater dispersal ability (at least as
75 adults), might show different patterns relative to taxa that were benthic (Rojas et al., 2017;
76 Yacobucci, 2017). This greater dispersal ability might allow pelagic organisms to more fully
77 occupy potentially available habitats than benthic organisms, which could lead to larger
78 geographic ranges and also less change in geographic ranges through time. (In addition, there
79 are certain paleoecological constraints that reduce the dispersal potential of cephalopods, such as
80 minimum water depth required for vertical migration, Ward and Westermann, 1985; Ritterbush
81 et al., 2014; R. T. Becker, 2019, pers. comm.) It also could potentially influence patterns of
82 speciation and extinction by dampening opportunities for geographic isolation and creating
83 larger effective population sizes. Further, sea-level fall is known to cause regular and repeated
84 patterns of extinction in ammonoids (Kullmann, 1983, 1985; House, 1985; Hallam, 1987; Becker
85 & Kullmann, 1996; Wiedmann & Kullmann, 1996; Kaiser et al., 2011; Zhang et al., 2019; and R.
86 T. Becker, 2019, pers. comm.).

87
88 This study focuses on cephalopods from the Pennsylvanian-early Permian (Morrowan, Atokan,
89 Desmoinesian, Missourian, Virgilian, and Wolfcampian) in the Midcontinent Sea of the United
90 States as knowledge of the systematic affinities, geographic distribution and overall diversity of
91 | [these](#) is relatively well understood (Miller, Dunbar, & Condra, 1933; Newell, 1936; Plummer &

92 Scott, 1937; Miller & Youngquist, 1949; Nassichuk, 1975; Boardman et al., 1994; Landman,
93 Tanabe, & Davis, 1996; Kröger, 2005; Klug et al., 2015; Korn, Klug, & Walton, 2015), the
94 stratigraphy of the region is well constrained (Heckel, 2008, 2013), and there are extensive
95 exposures of fossiliferous units in the region. Moreover, at this time the Midcontinent Sea was
96 bordered by the Antler Orogeny to the north, the Ancestral Rocky Mountain Orogeny to the
97 west/northwest and the Ouachita Mountain belt to the south/southeast (as well as various
98 structural arches), such that it constituted a distinct biogeographic region for marine invertebrates
99 (Wells et al., 2007; Nelson & Lucas, 2011; Joachimski & Lambert, 2015).

100

101 The Late Paleozoic Ice Age (LPIA) was the longest lived glacial period of the Phanerozoic and is
102 relatively well understood due to numerous stratigraphic, sedimentologic, paleontologic, and
103 isotopic studies (e.g., Mii, Grossman, & Yancey, 1999; Isbell, 2003; Stanley & Powell, 2003;
104 Raymond & Metz, 2004; Montañez, 2007; Powell, 2007; Tabor & Poulsen, 2007; Fielding,
105 Frank, & Isbell, 2008; Heckel, 2008; DiMichele et al., 2009; Bonelli & Patzkowsky, 2011;
106 Montañez & Poulsen, 2013; Balseiro, 2016; Roark et al., 2017; Segessenman & Kammer, 2018).
107 Glacial cycling in the North American midcontinent region has received much study (e.g., Isbell,
108 2003; Heckel, 2008, 2013). Modern synthesis of the glacial history indicates that the Morrowan
109 to early Desmoinesian represented a localized glacial period, the late Desmoinesian to early
110 Virgilian represented a widespread interglacial period with minor glaciation, and the late
111 Virgilian to early Wolfcampian represented the apex of widespread glaciation (Montañez &
112 Poulsen, 2013). Modeling predicts that sea-level oscillations in the late Pennsylvanian were
113 between 50-100 meters depending upon the number and volume of melting ice sheets, and that
114 water temperatures are estimated to have been between 4-7°C cooler during glacial maxima than

115 inter-glacial periods (Heckel, 1986; Isbell, 2003; Montañez, 2007; Tabor, 2007; Heckel, 2008;
116 Cecil, DiMichele, & Elrick, 2014). The sea-level and temperature changes were likely to have
117 had an important influence on species distribution and geographic range size during this time
118 (Waterhouse & Shi, 2010). Perhaps cephalopod taxa would be less influenced by glacial sea-
119 level cycles than benthic taxa, as these cycles are also known to cause variation in seafloor
120 ventilation, with concomitant dysoxia/anoxia that is more severe for benthic taxa (A. Dunhill,
121 pers. comm., 2018). By contrast, sea-level fall is known to have caused ammonoid extinctions
122 and Paleozoic cephalopods were sensitive to water temperature (R. T. Becker, pers. comm.,
123 2019).

Eliminado: Though p

124 **Materials and methods**

125 126 127 **Taxa considered, stratigraphic correlation, specimens examined, and georeferencing: 79**

128 species belonging to 26 genera (13 nautiloids and 13 ammonoids) of cephalopods in the
129 Pennsylvanian-Permian North American Midcontinent Sea were considered (Table S1). These
130 represent abundant, well preserved, and taxonomically well understood species for which we
131 were able to obtain type material and collections material of sufficient quality to enable
132 taxonomic assignments on a breadth of material. Other species from the mid-continent of North
133 America certainly exist and adding these to our analyses could change our results. However, at
134 this time it was not possible to consider these via obtaining type and other material for them and
135 pursuing the significant additional taxonomic work this would entail. Therefore, results are
136 based on consideration of what is essentially a random selection of some of the (albeit well
137 known) species in the region and this analysis is best viewed as an initial approach to considering
138 paleobiogeographic dynamics in the region. Range reconstructions relied on the occurrence

140 records of specimens derived from a comprehensive consideration of the entire taxonomic
141 literature on the taxa studied. In particular, the following publications were utilized: Cox (1857),
142 Swallow (1858), McChesney (1860), Meek & Worthen (1860, 1870), White & St. John (1867),
143 White (1889), Hyatt (1891, 1893), Keyes (1894), Miller (1892), Smith (1896, 1903), Girty
144 (1911, 1915), Mather (1915) Böse (1919, 1920), Miller (1930), Sayre (1930), Miller, Dunbar, &
145 Condra (1933), Miller & Cline (1934), Miller & Owen (1934, 1937, 1939), Foerste (1936),
146 Miller & Thomas (1936), Newell (1936), Plummer & Scott (1937), Elias (1938a, b), Miller &
147 Moore (1938), Smith (1938), Miller & Furnish (1940a, b, 1957), Teichert (1940), Clifton (1942),
148 Miller & Unklesbay (1942), Young (1942), Sturgeon (1946), Miller, Lane, & Unklesbay (1947),
149 Miller & Downs (1948, 1950), Miller & Youngquist (1947, 1949), Miller, Youngquist, &
150 Nielsen (1952), Kummel (1953, 1963), Ruzhentsev & Shimanskiy (1954), Unklesbay (1954),
151 Arkell et al. (1957), Unklesbay & Palmer (1958), Hoare (1961), Furnish, Glenister, & Hansman
152 (1962), McCaleb (1963), Gordon (1964), Miller & Breed (1964), Teichert et al. (1964), Furnish
153 & Glennister (1971), Ruzhentsev & Bogoslovskaya (1971), Nassichuk (1975), Sturgeon et al.
154 (1982), Hewitt et al. (1989), Boardman et al. (1994), Kues (1995), White & Skorina (1999),
155 Kröger & Mapes (2005), Furnish et al. (2009), and Niko & Mapes (2009) as well as from
156 examination of all specimens, including types, housed in: the Division of Invertebrate
157 Paleontology, Biodiversity Institute, University of Kansas (KUMIP); the University of Iowa
158 Paleontology Repository (UI); and the Yale University Peabody Museum of Natural History
159 (YPM). These institutions are among the most complete repositories of cephalopod diversity
160 from this region and time and contain many of the type specimens of the species examined.
161 Moreover, all specimens used in the analysis were personally examined and taxonomically-
162 vetted via consideration of the literature, relevant type specimens, and other material, with

species assignments and determinations made by the first author. Over 1,100 specimens were identified to species level in this study (Kolis, 2017). We chose to focus on the particular species considered, rather than downloading data from the Paleobiology Data Base (PBDB), as we wanted to be able to personally validate the taxonomic identity of specimens using collections data in conjunction with the literature in order to present more rigorously corroborated hypotheses about the geographic distributions of species. We consider this approach to be complementary to those approaches that utilize the PBDB in paleobiogeographic studies. On the one hand, our approach did limit the number of species we were able to consider. On the other hand, we believe it is quite important to evaluate hypotheses about systematic affinities of fossil specimens, the actual data of the fossil record themselves, in detail and thereby accurately define the taxonomic units considered. Given that species represent key macroevolutionary units in nature (Eldredge, 1989; Wiley & Lieberman, 2011; Hendricks et al., 2014), correctly characterizing them taxonomically, and thus validating the scope of their geographic distributions, is critical. Moreover, it has recently been shown by Marshall et al. (2018) that incorporating museum specimen data in the manner that our study has can greatly expand, enhance, and improve knowledge of geographic distributions of fossil species, relative to studies that only utilize data from the PBDB. In the case of some species, ~ 30% of the total considered, our analyses indicated moderate changes in stratigraphic range (addition of a stage, etc.) relative to what is presented in the PBDB. This happened primarily because via this study we were able to identify specimens to species that previously had been treated as indeterminate at the species level, or we were able to determine that specimens had previously been mis-identified to species.

184

185 Specimens were assigned to the Virgilian, Missourian, Desmoinesian, Atokan, Morrowan, or
186 Wolfcampian stages using the USGS National Geologic Map Database (U.S. Geological Survey,
187 2017), Sawin et al. (2006, 2008, 2009), Zeller (1968), Pope (2012), and Heckel (2013). The
188 temporal boundaries of stages were derived from Davydov, Korn, & Schmitz (2012) (Table S2).

189 It is important to note that the boundaries of international stages are based on few
190 geochronological tie points and the correlation of the North American stage boundaries with
191 these is arbitrary; also, some of the boundaries used are still being researched (R. T. Becker,
192 pers. comm., 2019.) In addition, while more resolved stratigraphic assignment to biostratigraphic
193 zone is possible for units in Europe (e.g., Davydov & Leven, 2003), the northern Appalachian
194 Basin of North America (e.g., Heckel et al., 2011), and parts of the North American
195 midcontinent (e.g., Boardman et al., 1994; Heckel et al., 2011), it is less tractable to associate the
196 boundaries of the biostratigraphic zones from the North American midcontinent with radiometric
197 dates for the stratigraphic units and regions considered herein. Furthermore, the museum
198 specimens considered herein lacked the information needed to make it possible to constrain them
199 to biostratigraphic zone, only stage. For this reason, it was unfortunately not possible to consider
200 changes in geographic range, nor rates of speciation and extinction, at a temporal scale more
201 resolved than stage. Although this is often the standard degree of temporal resolution used in a
202 variety of paleobiogeographic studies, it does entail that we were not able to discern events
203 transpiring more rapidly than the time scale of stage. This means that we will be missing
204 important patterns; although speciation and extinction does not appear to frequently be
205 transpiring within stage boundaries in this region, at least sometimes it is, and moreover
206 geographic range shifts by species were certainly happening within these boundaries.
207

Eliminado: (

Eliminado: ,

210 | All specimen localities were georeferenced during the course of the study. *GEOLocate* (Rios &
211 | Bart, 2018) and the *MaNIS Georeferencing Calculator* (Wieczorek, 2015) were used to obtain
212 | coordinates and uncertainty radii. All points were calculated in decimal degrees within the
213 | WGS84 model in the *GEOLocate* (Rios & Bart, 2018) world topo layer to ensure consistency
214 | and accuracy in determinations. Most uncertainty radii were less than 10 kms. Any specimens
215 | with questionable locality information were excluded from analyses, as were specimens with an
216 | uncertainty radius larger than the county they were contained within. This left 950 specimens
217 | (Table S1) to use in range reconstruction and statistical analysis of geographic range through
218 | geologic time. All statistical analyses were performed using Minitab® Statistical Software
219 | *Minitab v. 17* (Minitab, 2016) and *R-Studio Version 3.4.0* (2017).

220 |
221 | **Range reconstruction using GIS:** Methods for range reconstruction follow Rode & Lieberman
222 | (2004, 2005), Stigall & Lieberman (2006), Hendricks, Lieberman, & Stigall (2008), Myers &
223 | Lieberman (2011), Myers, MacKenzie, and Lieberman (2013), and Dunhill & Wills (2015). In
224 | particular, after specimen occurrence data were georeferenced and assigned to temporal bins,
225 | *Excel* CSV files were compiled for the occurrence points for all specimens within species. CSV
226 | files were imported into *ArcGIS v. 10.3* (ESRI, 2014) and layers were created using geographic
227 | coordinate system ‘WGS 1984’ and projected coordinate system ‘WGS 1984 World Mercator’
228 | (Fig. 1). These layers were input into *PaleoWeb* (The Rothwell Group LP, 2016) to rotate
229 | coordinates into continental configuration and geographic position of the midcontinent region
230 | during the Pennsylvanian-early Permian (Fig. 2). These paleo-coordinate layers were then re-
231 | projected into *ArcMap* (ESRI, 2014).

232 | Geographic range values were calculated for each species (Table S3) using minimum bounding
233 | geometry. This method has been shown to provide the most accurate procedure for

234 reconstructing changes in geographic range, especially for fossil taxa (Darroch & Saupe, 2018).
235 Convex hulls or buffers were given to every specimen occurrence point in each species and these
236 shapefiles were re-projected in 'South America-Albers Equal Area Conic'. This model was used
237 to accommodate the rotation of species occurrence coordinates into the southern hemisphere
238 during the late Paleozoic. Species with three or more occurrence points were given a convex hull
239 that spanned the entire area between occurrences (see Rode & Lieberman, 2004; Hendricks,
240 Lieberman, & Stigall, 2008; Myers & Lieberman, 2011; and Darroch & Saupe, 2018). In this
241 way, multiple occurrence points were combined to recreate the geographic range of a single
242 species. Species with only one occurrence point were given a 10km² buffer; species with just two
243 occurrence points were given a 10km² wide buffer which was used, in conjunction with their
244 distance, to derive an area value (following Rode & Lieberman, 2004, 2005; Hendricks,
245 Lieberman, & Stigall, 2008; Myers & Lieberman, 2011; and Myers, MacKenzie, and Lieberman,
246 2013). Species geographic range size data were tested for normality within each temporal stage
247 using the Anderson-Darling normality test (this is a commonly used test to assess normality, see
248 Sokal & Rohlf, 1994).

249

250 **Assessing fossil record bias:** A common concern when studying the fossil record is that there
251 might be biases that could lead to inaccurate or artifactual findings. This concern can be
252 manifold, but the two most pertinent issues here involve incomplete sampling and/or issues of
253 stratigraphic bias. While it is important to be aware of the fact that the fossil record is
254 incomplete, it is worth recognizing that there is a large body of research that demonstrates that
255 many of the biogeographic patterns preserved in the fossil record, particularly in marine settings,
256 represent real biological phenomena, rather than taphonomic artifacts (Myers & Lieberman,

257 2011; Rook, Heim, & Marcot, 2013; Dunhill & Wills, 2015), although that does not mean that
258 such artifacts played no role in this study. Further, it is also prudent to realize that sampling bias
259 is a common issue in studies of extant biodiversity and species distribution, and much work
260 needs to be done in this area to alleviate the biases of the extant biota (Lieberman, 2002;
261 Carrasco, 2013).

262 The possibility that biases in the fossil record might lead to artifactual results was assessed in a
263 few different ways. First, the relationship between outcrop availability and the geographic range
264 of Pennsylvanian and Permian cephalopods was determined (see Myers & Lieberman, 2011). A
265 percent coverage table of the range size of species overlaid against temporal outcrop availability
266 was created using *ArcGIS v. 10.3* (ESRI, 2014). A low percentage of overlap between range size
267 and outcrop area would suggest species distributions are more likely to reflect ‘real’
268 biogeographic patterns while a high percentage of overlap would suggest the presence or absence
269 of outcrop was significantly influencing results (Myers & Lieberman, 2011; Myers, MacKenzie,
270 & Lieberman, 2013; however, see also Dunhill, 2012 for an alternative viewpoint). The second
271 test used was an “n-1” jackknifing analysis (see Myers & Lieberman, 2011; Myers, MacKenzie,
272 & Lieberman, 2013). This procedure sub-sampled species range size within each temporal bin to
273 test the resilience of data to outliers. Mean range size estimations were generated for each
274 temporal bin; these were input into a one-way ANOVA to compare jackknife estimates with the
275 initial geographic range size estimates (Myers & Lieberman, 2011; Myers, MacKenzie, &
276 Lieberman, 2013). Finally, a Pearson rank correlation test was performed to test the association
277 of occurrence points and geographic range size; a close correlation would indicate that
278 reconstructed ranges were very much dependent on sampling and suggest that reconstructed
279 biogeographic patterns might be an artifact of a biased fossil record (Myers, MacKenzie, &
280

281 Lieberman, 2013).

282

283 **Speciation and extinction rate calculations:** Speciation and extinction rates were calculated in
284 order to consider macroevolutionary dynamics in cephalopods from the Late Paleozoic
285 Midcontinent Sea. Macroevolutionary rates were calculated using the following equation,
286 presented in Foote (2000) and Rode & Lieberman (2005):

287
288
$$N_f = N_0 e^{rt}$$

289

290 where N_0 is the species richness at the beginning of a temporal bin, N_f is the species richness at
291 the end of a temporal bin, t is the duration of a temporal bin, and r is the total rate of diversity
292 change. The temporal bins used were North American stages (Table S2). Species richness values
293 (N_f) were determined for each temporal bin and were parsed into ‘carry-over’ (N_0) and ‘new’
294 species richness values to ensure the accuracy of speciation and extinction rate calculation. In
295 this way, it was possible to calculate the rate of diversity change between bins. For example, r
296 $_{Atokan} = (\ln N_{0-Desmoinesian} - \ln N_{0-Atokan}) / t_{Atokan}$. Speciation rate within each temporal bin was
297 calculated using the equation $S_{Atokan} = (\ln N_{f-Atokan} - \ln N_{0-Atokan}) / t_{Atokan}$, and extinction rate within
298 each temporal bin was calculated using the equation $E_{Atokan} = S_{Atokan} - r_{Atokan}$ for each temporal
299 stage (Foote, 2000; Rode & Lieberman, 2005).

300

301 Results

302

303 **Paleobiogeographic patterns:** Geographic range data were analyzed separately across all
304 cephalopods and individually for both nautiloids and ammonoids. As mentioned above, species

geographic range size data were tested for normality within each temporal stage using the Anderson-Darling normality test (see Sokal & Rohlf, 1994). Range size data within each temporal stage were not normally distributed for any data combination ($P < 0.005$). Instead, distributions were left skewed across all temporal stages for every data grouping. Data were subsequently log-transformed to normalize data, and statistical analyses were performed on both original and transformed data.

311

In general, geographic range size (either mean of transformed data or median of original) of ammonoids and nautiloids increases during the Missourian and Virgilian stages (Fig. 3), which was a time of sea-level rise due to warming during an interglacial (Isbell, 2003; Montañez & Poulsen, 2013), such that there may be an association between the sea-level rise and the increase in geographic range. Another possibility is that there was some change in taphonomic or collecting conditions that occurred during the Virgilian that made it easier to discern the actual biogeographic distributions of species at this time, relative to other time intervals (G. Piñero, pers. comm., 2018). However, none of the changes in geographic range were statistically significant, so it is not possible to infer strong correlation between the sea-level rise, or possible taphonomic factors, and the range expansion. For instance, Mann-Whitney U tests, a non-parametric test used to compare two sample medians (see Sokal & Rohlf, 1994), found no statistically significant changes (at $P \leq 0.05$) in median geographic range size for any temporal stages separately across all the studied cephalopods, as well as individually for nautiloids and ammonoids, even prior to correction for multiple comparisons. This is because with the Mann-Whitney U test median range values are considered, and for all cephalopods the median range values are constant through time (79km²). Mean values do show more change through time in

Con formato: Resaltar

Comentario [GP1]: This sentence has not sense, please, be more explicit.

Comentario [GP2]: Well, we have to say that mean and median are *per se* statistically valuable data!.

Con formato: Resaltar

Con formato: Resaltar

our data than the corresponding median values, as might be expected, and median values are better to focus on for statistical purposes when the data are not normally distributed.

The same was true for two-sample t-tests (see Sokal & Rohlf, 1994) performed on log-transformed data which again found no statistically significant changes (at $P \leq 0.05$) in mean geographic range size through time, even prior to employing a statistical correction needed in the case when there are multiple comparisons. Again, recall that *mean* range size data are shown in Figure 3, and the differences among log-transformed data through time are far less substantial (and ultimately not significant). Furthermore, a one-way ANOVA, either with or without the assumption of equal variance, failed to find any significant differences (at $P \leq 0.05$) between stages for log-transformed mean geographic range size across all cephalopods as well as individually for nautiloids and ammonoids. Still, it is worth noting that changes in range size are occurring through time, most notably in the Virgilian, and these could be related to climatic changes that occurred then, and also changes in the paleogeography of the region, although in the absence of statistical evidence we could not convincingly document such a link in the present study. However, it is important to note that previous studies (e.g., Ramsbottom, 1981) have documented such a link.

Analysis of macroevolutionary rates: Speciation rate (S) and extinction rate (E) were calculated for the Atokan, Desmoinesian, Missourian, and Virgilian stages across all selected cephalopods and within selected nautiloids and ammonoids, respectively. The S and E presented across all selected cephalopods are comprised of two calculations; one calculation included taxa that only occurred in a single temporal stage (singletons) (Table 1; Fig. 4), while the other

351 calculation excluded taxa that occurred in a single temporal stage (Table S4). S and E was also
 352 calculated for ammonoids and for nautiloids including (Tables S5, S6) and excluding taxa that
 353 occurred in a single stage (Tables S7, S8). Note, due to the dependence of calculations on
 354 diversity metrics from both adjacent stages, it is not possible to accurately calculate the rate of
 355 biodiversity change (R), or S and E for the first stage considered, the Morrowan, nor R or E for
 356 the last stage considered, the Wolfcampian (these are thus left blank in Table 1 and Tables S4-
 357 S8). While it might have been possible to infer S and E using other methods, to do so would
 358 exaggerate the significance of edge effects and thus be problematic (Foote, 2000). A problem
 359 with including singleton taxa, which occur in just a single stratigraphic stage, and why it is
 360 sometimes recommended that these be excluded, is that perforce for these there will be a direct
 361 correlation between S and E (Foote, 2000); however, when singletons are not included, a higher
 362 proportion of ammonoids cannot be considered, as many of these have short biostratigraphic
 363 ranges (R. T. Becker, pers. comm., 2019).

Comentario [GP3]: The text below was removed because it is redundant.

Eliminado:

Eliminado: such that analyses were performed both with and without singletons

Con formato: Resaltar

364
 365 Across all cephalopods studied, S was high in the Atokan and Desmoinesian, fell in the
 366 Missourian, and reached very low levels in the Virgilian and Wolfcampian (Fig. 4). By contrast,
 367 E was low in the Atokan and Desmoinesian, began to rise in the Missourian, and reached even
 368 higher levels in the Virgilian (Fig. 4). Essentially, across all cephalopods examined, when S is
 369 high, E is low, and when S is low, E is high. This is potentially contrary to the pattern expected
 370 with an ecological opportunity model of speciation (Simões et al., 2016), although the specific
 371 processes driving the diversification could not be determined at this time. However, it is
 372 possible that when S was high there may have been many short-lived species that could not be
 373 sampled that were actually going extinct, and this phenomenon would artificially depress E. To

Comentario [GP4]: This paragraph and in general the explanation about the reasons to avoid the use of singletons is too much confusing. Please, reword because it is obvious that if you exclude singletons a higher proportion of ammonoids cannot be considered. That is indeed a problem that Dr. Becker remarked. By the way, who recommended sometimes that singletons must be excluded? In which situations?

Con formato: Resaltar

378 consider this in more detail, what is truly needed is a zone by zone analysis of all cephalopod
379 species known from the North American midcontinent (R. T. Becker, 2019, pers. comm.).

380

381 As expected, S and E are lower when singletons are excluded (see Tables 1, S4). Segesseman &
382 Kammer (2018) found in their macroevolutionary study on crinoids during this interval that

383 including or excluding singletons substantially influenced their results, but in our study including

384 or excluding these did not produce such a substantial change in results. Notably, S and E patterns

385 diverge somewhat between ammonoids and nautiloids when considered individually (and the

386 patterns in nautiloids better match the overall patterns across all the studied cephalopods). For

387 instance, in nautiloids S is high in the Atokan and Desmoinesian, then declines to moderate in

388 the Missourian, and is at its lowest in the Missourian and Wolfcampian (Table S6), whereas in

389 ammonoids S is only high in the Atokan, declines to moderate in the Desmoinesian, declines

390 somewhat more in the Missourian and then remains essentially constant through the

391 Wolfcampian (Table S5). In addition, E is low in ammonoids during the Desmoinesian and

392 Missourian but high in the Atokan and Wolfcampian (Table S5), whereas in nautiloids there are

393 no observed extinctions during the Atokan; values remain quite low for nautiloids in the

394 Desmoinesian, rise somewhat in the Missourian, and then rise again in the Virgilian (Table S6).

395

396 An important caveat regarding the calculation of S is that many of the species analyzed belong to

397 genera that were widely distributed beyond the Midcontinent Sea during the Late Paleozoic.

398 Thus, although none of the species considered in these analyses occurred outside of the

399 Midcontinent Sea, their close relatives did. It is conceivable that while speciation events and

400 rates by necessity are herein treated as occurring *in situ*, this might not always have been the

Eliminado: (

Eliminado: [

Eliminado:]

Eliminado:)

case. Instead, some speciation events could have occurred outside of the Midcontinent Sea with subsequent invasion events into that region. These invasions would appear as *in situ* speciation events in this analysis, although they actually were not. In the absence of phylogenetic hypotheses for the genera considered it is not currently possible to consider how much of the pattern pertaining to speciation rate shown in Fig. 4 is due to invasion instead of speciation such that both might be playing a role. (*Metacoceras* is one example where the genus occurs well outside of the North American mid-continent, it is known to occur in beds ~ 100kms southeast of Moscow, Russia, such that some of the cladogenetic events involving this genus might comprise instances of invasion). Further, a related phenomenon could affect the calculation of E: at times what were treated as extinction events might have simply been local extinctions in the Midcontinent Sea which could have included emigration to other regions. As mentioned previously, it does not appear that any of the species considered occur outside of the Midcontinent Sea, but a phylogenetic hypothesis for these groups would be valuable for considering this issue in greater detail.

Eliminado: .

Con formato: Fuente: Cursiva

Eliminado: .

Relationship between biogeography and macroevolutionary rates: Across all the studied cephalopods, mean geographic range size increased during the Virgilian (and in ammonoids first in the Missourian but then more prominently in the Virgilian) and declined in the Wolfcampian (Fig. 3); speciation rates were generally high in the Atokan and Desmoinesian and fell in the Virgilian (Fig. 4); extinction rates were generally low in the Atokan and Desmoinesian and rose in the Virgilian (Fig. 4). The Pearson correlation test in *Minitab 17* (Minitab, 2016) was used to examine the association between geographic range and either speciation rate extinction rate in greater detail. No significant (at $P \leq 0.05$) correlation between speciation or extinction rate and

range size was found across all cephalopods or within ammonoids or nautiloids individually (Table 2). However, in cases the values approach $P = 0.05$. For instance, the association between decreasing geographic range size and increasing extinction for all cephalopods and for ammonoids alone, so it is clear that generally there is some association between the two, but unfortunately significant support at the .05 level is lacking. We note that numerous previous studies have documented an association between decreasing geographic range size and increasing extinction rate (e.g. Vrba, 1980; Jablonski, 1986; Eldredge, 1989; Stanley, 1990; Jablonski & Roy, 2003; Rode & Lieberman, 2004, 2005; Kiessling & Aberhan, 2007; Payne & Finnegan, 2007; Stigall, 2010; Dunhill & Wills, 2015; Jablonski & Hunt, 2015; Orzechowski et al., 2015; Saupe et al., 2015; Castiglione et al., 2017; Pie & Meyer, 2017; Lam, Stigall, & Matzke, 2018; Schneider, 2018) and thus this a very robust phenomenon in general and likely to be operating to some extent herein. However, over this time interval and for this particular group of species the association is not statistically significant (Table 2), probably because sample sizes are not large, and further this is likely because many taxa were culled by the late Mississippian extinction (M. Powell, pers. comm., 2018). Further, sample size could also be influencing the results pertaining to changes in geographic range size through time (G. Piñeiro, pers. comm., 2019).

447

Analysis of fossil record bias: The low percentage of overlap between cephalopod species geographic ranges and the availability of outcrop, less than 1% in 29 out of 30 species (Table S9; the one species with a larger percentage value, “*Orthoceras*” *kansasense*, occurs throughout the Midcontinent Sea), suggests the results are not simply an artifact of an incomplete fossil record, at least pertaining to outcrop availability or changes in the paleogeography of the region. The

453 “n-1” jackknifing analysis also supports the robustness of the reconstructed ranges, as no
454 statistically significant differences were found between the mean of the reconstructed and
455 subsampled range values for any time interval (all P-values > 0.9), suggesting that one or a few
456 occurrence records are not having a major influence on biogeographic patterns. Similar results
457 were found in other taxa and time periods by Hunt, Roy, & Jablonski (2005), Myers &
458 Lieberman (2011), and Myers, MacKenzie, & Lieberman (2013), although Dunhill, Hannisdal,
459 & Benton (2014) did find some association between outcrop area and diversity in the case of the
460 marine fossil record of Great Britain. Finally, the Pearson correlation test shows no correlation (-
461 0.055, P-Value = 0.789) between the number of occurrence points and geographic range size;
462 this provides further evidence that the biogeographic signatures of Late Paleozoic cephalopods
463 are unlikely to be simply an artifact of the fossil record.

464

465 **Diversity patterns:** Across all cephalopods, species richness increased from the Morrowan to
466 the Atokan, peaked in the Desmoinesian, and decreased through the Wolfcampian (Fig. S1). A
467 similar pattern is seen in the nautiloids (Fig. S2). However, the ammonoids (Fig. S3) demonstrate
468 an earlier peak in the Atokan, followed by a Desmoinesian to Virgilian plateau, with a decrease
469 in the Wolfcampian. This indicates that the data from nautiloids are most influencing the
470 recovered patterns (G. Piñeiro, pers. comm., 2019). Notably, previous studies of late Paleozoic
471 brachiopod communities in Bolivia showed a consistent trend between diversity and glacial
472 cycling with increased diversity during glacial periods and decreased diversity during inter-
473 glacial periods (Badyrka, Clapham, & Lopez, 2013). However, there seems to be less
474 consistency between species richness trends and glacial cycling in the Midcontinent Sea. For
475 instance, there is an increase in cephalopod species richness throughout the Morrowan to

Desmoinesian associated with localized glaciation, and an interglacial period with generally minor glaciation is associated with a decrease in cephalopod species richness from the Desmoinesian to Virgilian, yet by contrast widespread glaciation is associated with a decrease in species richness from the Virgilian to the Wolfcampian. Important points, however, are that these are raw diversity patterns, and sample standardized diversity patterns show a different result (M. Powell, pers. comm., 2018), and further that brachiopods and cephalopods can show different behaviors in response to climatic changes (G. Piñeiro, pers. comm., 2019).

Discussion

Geographic range shifts through time are one of the pervasive phenomena in the history of life; these are manifest both within species and higher-level clades, occur at a number of different time scales, and are frequently linked to climatic change (Wiley & Lieberman, 2011). Specific examples do come from the late Paleozoic, a time of extensive climate change including profound glaciation along with numerous glacial and interglacial cycles and associated cycles of sea-level rise and fall (Montañez and Poulsen, 2013). (Previous studies of ammonoids have shown that these changes in sea-level may have caused more significant changes in biogeographic ranges of taxa than temperature changes during this time period, and other time periods as well [Hallam, 1987; Hartenfels & Becker, 2016; Zhang et al., 2019]). Those changes impacted patterns of geographic range in both terrestrial plant (e.g., DiMichele et al., 2009; Falcon-Lang & DiMichele, 2010) and marine invertebrate ecosystems (e.g., Ramsbottom, 1981; Leighton, 2005; Powell, 2007; Waterhouse & Shi, 2010; Balseiro & Halpern, 2019). When it comes to marine invertebrates from this time interval, most of the focus has been on the highly diverse benthic faunas (e.g., Stanely & Powell, 2003; Powell, 2007; Bonelli & Patzkowsky,

2011; Balseiro, 2016; Segessenman & Kammer, 2018; Balseiro & Halpern, 2019); however, taxa that have a pelagic life style are also worth examining. Herein, 79 pelagic species of cephalopods were examined for patterns of range size change using GIS and although in general these species exhibit some evidence for changes in geographic range size (Fig. 3) especially in the Virgilian, and to a lesser extent in the Missourian, those changes were not statistically significant, making it hard to directly tie them to climate changes. However, there is strong evidence that climate change played a prominent role in influencing geographic range of cephalopods from other regions during this time period (e.g., Ramsbottom, 1981) and indeed in cephalopods from other time periods (e.g., Hallam, 1987; Jacobs et al., 1994; Kaiser et al., 2011; Hartenfels & Becker, 2016; Zhang et al., 2019). In a similar vein, many paleontological studies have demonstrated that species with larger geographic ranges tend to have lower extinction rates than species with narrower geographic range sizes (e.g., Vrba, 1980; Jablonski, 1986; Eldredge, 1989; Stanley, 1990; Rode & Lieberman, 2004; Stigall & Lieberman, 2006; Payne & Finnegan, 2007; Stigall, 2010; Hopkins, 2011; Dunhill & Wills, 2015). Again, this phenomenon is not found to be statistically significant in the case of the late Paleozoic cephalopod species considered herein (Table 2), but there is some general quantitative evidence for the phenomenon.

There may be a few different explanations for these findings. First, it may be that some cephalopod species were not significantly affected by the glacial-interglacial climatic cycles transpiring within the Late Paleozoic Midcontinent Sea. A second possible explanation, perhaps coupled to the first, is that since cephalopods are highly mobile relative to benthic marine invertebrates such as gastropods, bivalves, brachiopods, etc., they can more easily occupy a greater portion of their potential range. Further, perhaps the available potential range of

523 cephalopod species does not change much in glacial relative to interglacial regimes. This may
524 seem unlikely given the vast fluctuations in sea level occurring at the time, but pelagic marine
525 organisms, because of their ease of dispersal, may more easily maintain consistent geographic
526 ranges relative to benthic counterparts. Another possible explanation for the pattern retrieved is
527 that, given the limits of stratigraphic correlation, sample size, and the completeness of the fossil
528 record, it was necessary for the analyses of species distribution conducted herein to focus on the
529 time scale of geological stages, whereas in actuality there were climatic changes occurring within
530 stages (Heckel, 2008, 2013); these certainly did cause fluctuations in species' geographic ranges
531 within stages, but simply could not be observed in the present study. The inability to observe
532 changes in geographic range size of species at a scale more resolved than stage, in particular,
533 likely played an important limiting role in the conclusions that could be derived. For instance,
534 other studies such as Ramsbottom (1981) have looked at European taxa from the same time
535 period, but focused at the level of zones, and did find a strong association between climate, sea-
536 level, and geographic distribution. A final set of explanations are related to the issue of sampling.
537 For instance, it was more difficult for the analyses presented herein to detect a relationship
538 between geographic range size and macroevolutionary rate because speciation and extinction
539 rates could only be calculated for four stages. Although we did not observe a substantial amount
540 of speciation and extinction occurring within stage boundaries, certainly being able to consider
541 more stages would have enhanced our ability to retrieve patterns. We suspect that another
542 important explanation for our results is the relatively limited number of species that could be
543 considered herein. An expansion in the number of taxa considered could absolutely change our
544 results in various ways, including via increasing statistical power. Thus, what is presented herein
545 should only be treated as preliminary results that require further data and additional testing. We

546 would note, though, that detailed taxonomic vetting of specimens, including through comparison
547 of type material, especially involving taxonomic studies conducted in some cases more than 70
548 years ago, requires a significant amount of time investment. Thus, dramatic expansions to this
549 dataset would require concomitant investments of time. However, other datasets such as
550 AMMON and the PBDB could be used if one did not feel it was necessary to spend time vetting
551 taxonomic assignments. Although we posit that it is important to vet taxonomic assignments that
552 may be outdated, we would assert that our approach should be viewed as complementary to
553 approaches that rely on mining currently existing paleontologically oriented databases, and that
554 both types of approaches have value. As a final possible explanation for our results, we further
555 note that a common concern when studying the fossil record is the potential role biases can play.
556 This concern can be manifold. It is somewhat obviated by the results presented herein regarding
557 the apparent quality of the fossil record, but that does not mean that there are no inherent
558 problems with the cephalopod record that are at present difficult to ascertain; these could be
559 influencing the results retrieved in some at present unspecified way.

560

561 There is, however, another finding contrary to what might typically be expected for the late
562 Paleozoic that is worth mentioning. That is the fact that there seems to have been at least some
563 moderate degree of evolutionary diversification and turnover within cephalopods, such that
564 species diversity did fluctuate throughout the Pennsylvanian and early Permian. Pennsylvanian
565 rates of macroevolution are typically classified as ‘sluggish’ or ‘stolid’ across all marine animals,
566 and Sepkoski (1998) formalized the notion that there was a marked decline in evolutionary rates
567 of Carboniferous and Permian marine faunas. Stanley & Powell (2003) reiterated this result and
568 identified low mean macroevolutionary rates for marine invertebrate taxa. Bonelli &

Eliminado: macroevolution

Patzkowsky (2011) also documented a pattern of low turnover in the face of major episodes of sea-level rise and fall due to climatic change. The results from the analyses presented herein could indicate that macroevolutionary rate in the case of late Paleozoic cephalopods was more dynamic than often thought, supporting the conclusions of a variety of other important studies considering late Paleozoic ammonoid diversity including Kullmann (1983, 1985), House (1985), Becker & Kullman (1996), Wiedmann & Kullmann (1996), and Kullmann, Wagner, & Winkler Prins (2007). One possible reason why cephalopods may show a higher rate of diversification than other groups is that they were a fairly evolutionarily volatile group (Lieberman & Melott, 2013); thus, relative to many other marine invertebrate groups, they had relatively high rates of speciation and extinction (Stanley, 1979; Jacobs et al., 1994; Landman, Tanabe, & Davis, 1996; Monnet, De Baets, & Klug, 2011; Klug et al., 2015; Korn, Klug, & Walton, 2015). However, this may not be the entire explanation, as some other groups also show elevated rates of speciation and extinction during this time interval. For instance, Balseiro (2016) and Balseiro and Halpern (2019) did document evolutionary turnover at high latitudes, and elevated evolutionary rates have also been found in fusulinid foraminifera (Groves & Lee, 2008; Groves & Yue, 2009) and advanced cladid crinoids (Segessenman & Kammer, 2018). Ultimately, we support the contention raised by Segessenman & Kammer (2018) that patterns from a few individual groups do not refute the general pattern of sluggish macroevolution postulated for this time period in the history of life. The results may lend credence to the notion that macroevolutionary patterns across all marine animals are rarely unitary for any one time period in the history of life, and instead often tend to be variegated.

Conclusions

593

594 Patterns of range size change in late Paleozoic cephalopods from the North American
595 Midcontinent Sea were investigated using GIS. These species do exhibit some evidence for
596 changes in geographic range size through time, but the changes were not statistically significant
597 nor could they be directly tied to climate change. Further, in contradistinction to what is usually
598 found in the fossil record, cephalopod species with larger geographic ranges were not found to
599 have lower extinction rates than species with narrower geographic ranges. These distinctive
600 patterns may perhaps be related to the fact that cephalopods are pelagic and highly mobile, at
601 least relative to many benthic marine invertebrates, but it may also be due to the fact that only 79
602 species could be considered in our study, or to the fact that we were constrained to analyze
603 patterns at the temporal level of stage. Finally, the group shows more evolutionary
604 diversification and turnover during the Pennsylvanian and early Permian than is typical of other
605 marine invertebrate groups and this could be related to the fact that cephalopods are an
606 evolutionarily volatile group.

607

608 **Acknowledgements**

609

610 Thanks to Chris Beard, Kirsten Jensen, Julien Kimmig, and Luke Strotz for very helpful
611 discussions on this work and thanks to them and Matthew Powell, Alexander Dunhill, Ralph
612 Thomas Becker, Dieter Korn, Graciela Piñeiro, Thomas Algeo, and Wolfgang Kiessling for
613 comments on previous versions of the manuscript. Thanks to Julien Kimmig for assistance with
614 collections related matters and for providing access to specimens in the KUMIP; thanks to
615 Tiffany Adrain for assistance with collections related matters and for providing access to

specimens in the UI; and thanks to Susan Butts for assistance with collections related matters and for providing access to specimens in the YPM. Thanks to Michelle Casey and Erin Saupe for assistance with stratigraphic correlations and use of GIS.

References

- Abe FR, Lieberman BS. 2009.** The nature of evolutionary radiations: a case study involving Devonian trilobites. *Evolutionary Biology* **36**:225-234.
- Arkell WJ, Furnish WM, Kummel B, Miller AK, Moore RC, Schindewolf OH, Sylvester-Brady PC, Wright CW. 1957.** Part L. Mollusca Cephalopoda Ammonoidea. *Treatise on Invertebrate Palaeontology*. Lawrence, Kansas: Geological Society of America.
- Badyrka K, Clapham ME, Lopez S. 2013.** Paleoecology of brachiopod communities during the late Paleozoic ice age in Bolivia (Copacabana Formation, Pennsylvanian-Early Permian). *Palaeogeography, Palaeoclimatology, Palaeoecology* **387**:56-65.
- Balseiro D. 2016.** Compositional turnover and ecological changes related to the waxing and waning of glaciers during the Late Paleozoic ice age in ice-proximal regions (Pennsylvanian, western Argentina). *Paleobiology* **42**:335-357.
- Balseiro D, Halpern K. 2019.** Immigration and extirpation selectivity patterns of brachiopods and bivalves across the Carboniferous glacial to non-glacial transition (Pennsylvanian, central western Argentina) and their influence in building the biotic bathymetric gradient. *Palaeogeography, Palaeoclimatology, Palaeoecology* . **In press**.
<https://doi.org/10.1016/j.palaeo.2018.11.031> .

638 **Becker RT, Kullmann J. 1996.** Paleozoic ammonoids in space and time. In: Landman NH,
639 Tanabe K, Davis RA, eds. *Ammonoid Paleobiology*. Boston: Springer, 711-753.

640 **Boardman DR II, Work DM, Mapes RH, Barrick JE. 1994.** Biostratigraphy of middle and late
641 Pennsylvanian (Desmoinesian-Virgilian) ammonoids. *Kansas Geological Survey Bulletin*
642 **232**:1-121.

643 **Bonelli JR, Patzkowsky ME. 2011.** Taxonomic and ecologic persistence across the onset of the
644 Late Paleozoic Ice Age: evidence from the upper Mississippian (Chesterian series),
645 Illinois basin, United States. *Palaios* **26**:5-17.

646 **Böse E. 1919.** The Permo-Carboniferous ammonoids of the Glass Mountains, West Texas, and their
647 stratigraphical significance. *University of Texas Bulletin* **1762**:1-241.

648 **Böse E. 1920.** On ammonoids from the Abo Sandstone of New Mexico and the age of the beds
649 which contain them. *American Journal of Science* **49**:51-60.

650 **Brooks DR, McLennan DA. 1991.** *Phylogeny, Ecology, and Behavior: a Research Program in*
651 *Comparative Biology*. Chicago: University of Chicago Press.

652 **Carrasco MA. 2013.** The impact of taxonomic bias when comparing past and present species
653 diversity. *Palaeogeography, Palaeoclimatology, Palaeoecology* **372**:130-137.

654 **Castiglione S, Mondanaro A, Melchionna M, Serio C, Di Febbraro M, Carotenuto F, Raia**
655 **P. 2017.** Diversification rates and the evolution of species range size frequency
656 distribution. *Frontiers in Ecology and Evolution* **5**:147, 1-10.

657 **Cecil CB, DiMichele WA, Elrick SD. 2014.** Middle and Late Pennsylvanian cyclothems,
658 American Midcontinent: Ice-Age environmental changes and terrestrial biotic dynamics.
659 *Comptes Rendus Geoscience* **346**:159-168.

660 **Clifton RL. 1942.** Invertebrate faunas from the Blaine and the Dog Creek formations of the Permian
661 Leonard Series. *Journal of Paleontology* **16**:685-699.

662 **Cox DD. 1857.** First report of a geological reconnaissance of the northern countries of Arkansas,
663 made during the years 1857 and 1858. *Arkansas Geological Survey*.

664 **Darroch SAF, Saupe EE. 2018.** Reconstructing geographic range-size dynamics from fossil
665 data. *Paleobiology* **44**:25-39.

666 **Davydov VI, Leven EJ. 2003.** Reconstructing geographic range-size dynamics from fossil data.
667 *Palaeobiogeography, Palaeoclimatology, Palaeoecology* **196**:39-57.

668 **Davydov VI, Korn D, Schmitz MD. 2012.** Chapter 23 - The Carboniferous Period. In:
669 Gradstein FM, Ogg JG, Schmitz MD, Ogg GM, eds. *The Geologic Time Scale*.
670 Amsterdam: Elsevier, 603 - 651.

671 **DiMichele WA, Montañez IP, Poulsen CJ, Tabor NJ. 2009.** Climate and vegetational regime
672 shifts in the late Paleozoic ice age earth. *Geobiology* **7**:200-226.

673 **Dunhill AM. 2012.** Problems with using rock outcrop area as a paleontological sampling proxy:
674 rock outcrop and exposure area compared with coastal proximity, topography, land use,
675 and lithology. *Paleobiology* **38**:126-143.

676 **Dunhill AM, Wills MA. 2015.** Geographic range did not confer resilience to extinction in
677 terrestrial vertebrates at the end-Triassic crisis. *Nature Communications* **6**:7980.

678 **Dunhill AM, Hannisdal B, Benton MJ. 2014.** Disentangling rock record bias and common-
679 cause from redundancy in the British fossil record. *Nature Communications* **5**:4818.

680 **Eldredge N. 1989.** *Macroevolutionary Dynamics: Species, Niches, and Adaptive Peaks*. New

681 York: McGraw-Hill.

682

683 **Eldredge N, Gould SJ. 1972.** Punctuated equilibria: an alternative to phyletic gradualism. In:

684 Schopf TJM, ed. *Models in Paleobiology*. San Francisco: Freeman, Cooper, 82-115.

685 **Elias MK. 1938a.** *Properrinites plummeri* Elias, n. gen and sp., from late Paleozoic rocks of Kansas.

686 *Journal of Paleontology* **12**:101-105.

687 **Elias MK. 1938b.** Revision of *Gonioloboceras* from late Paleozoic rocks of the midcontinent region.

688 *Journal of Paleontology* **12**:91100.

689 **ESRI. 2014.** *ArcGIS Desktop: Release 10.3*. Redlands, CA: Environmental Systems Research

690 Institute.

691 **Falcon-Lang HJ, DiMichele WA. 2010.** What happened to the coal forests during

692 Pennsylvanian glacial phases? *Palaaios* **25**:611-617.

693 **Fielding CR, Frank TD, Isbell JL. 2008.** The Late Paleozoic Ice Age: a review of current

694 understanding and synthesis of global climate patterns. *Geological Society of America*

695 *Special Paper* **441**:343-354.

696 **Foerste AF. 1936.** Silurian cephalopods of the Port Daniel area on Gaspé Peninsula, in eastern

697 Canada. *Bulletin of Denison University, Journal of the Scientific Laboratories* **31**:21-92.

698 **Foote M. 2000.** Origination and extinction components of taxonomic diversity: general

699 problems. *Paleobiology* **26**:74-102.

700 **Furnish WM, Glenister BF. 1971.** Permian Gonioloboceratidae (Ammonoidea). *Smithsonian*

701 *Contributions to Paleobiology* **3**:301-312.

702 **Furnish WM, Glenister BF, Hansman RH. 1962.** Brachycycloceratidae, novum, deciduous
703 Pennsylvanian nautiloids. *Journal of Paleontology* **36**:1341-1356.

704 **Furnish WM, Glenister BF, Kullmann J, Zhou Z. 2009.** Part L. Mollusca 4, Revised, Vol. 2:
705 Carboniferous and Permian Ammonoidea (Goniatitida and Prolecanitida), pg. 136-144.
706 *Treatise on Invertebrate Palaeontology*. Lawrence: The University of Kansas Paleontological
707 Institute.

708 **Girty GH. 1911.** On some new genera and species of Pennsylvanian fossils from the Wewoka
709 Formation of Oklahoma. *Annals of the New York Academy of Sciences* **21**:119-156.

710 **Girty GH. 1915.** Fauna of the Wewoka Formation of Oklahoma. *United States Geological Survey*
711 *Bulletin* **544**:1-353.

712 **Gordon M. 1964.** Carboniferous cephalopods of Arkansas. *United States Geological Survey*
713 *Professional Paper* **460**:1-322.

714 **Groves JR, Lee A. 2008.** Accelerated rates of foraminiferal origination and extinction during the
715 late Paleozoic ice age. *Journal of Foraminiferal Research* **38**:74-84.

716 **Groves JR, Yue W. 2009.** Foraminiferal diversification during the late Paleozoic ice age.
717 *Paleobiology* **35**:367-392.

718 **Hallam A. 1987.** Radiations and extinctions in relation to environmental change in the marine
719 Lower Jurassic of northwest Europe. *Paleobiology* **13**:152-168.

720 **Hartenfels S, Becker RT. 2016.** The global *Annulata* events: review and new data from the
721 Rheris Basin (northern Tafilalt) of SE Morocco. *Geological Society of London, Special*
722 *Publications* **423**:291-354.

723 **Heckel PH. 1986.** Sea-level curve for Pennsylvanian eustatic marine transgressive-regressive
724 depositional cycles along midcontinent outcrop belt, North America. *Geology* **14**:330-
725 334.

726 **Heckel PH. 2008.** Pennsylvanian cyclothems in midcontinent North America as far-field effects
727 of waxing and waning of Gondwana ice sheets. *Geological Society of America Special*
728 *Paper* **441**:275-289.

729 **Heckel PH. 2013.** Pennsylvanian cyclothems of northern midcontinent shelf and biostratigraphic
730 correlation of cyclothems. *Stratigraphy* **10**:3-40.

731 **Heckel PH, Barrick JE, Rosscoe SJ. 2011.** Condonot-based correlation of marine units in the
732 lower Conemaugh Group (Late Pennsylvanian) in Northn Appalachian Basin.
733 *Stratigraphy* **8**:253-269.

734 **Hendricks JR, Lieberman BS, Stigall AL. 2008.** Using GIS to study palaeobiogeographic and
735 macroevolutionary patterns in soft-bodied Cambrian arthropods. *Palaeogeography,*
736 *Palaeoclimatology, Palaeoecology* **264**:163-175.

737 **Hendricks JR, Saupe EE, Myers CE, Hermesen EJ, Allmon WD. 2014.** The generification of
738 the fossil record. *Paleobiology* **40**:511-528.

739 **Hewitt RA, Dokainish MA, El Aghoury M, Westermann GE. 1989.** Bathymetric limits of a
740 Carboniferous orthoconic nautiloid deduced by finite element analysis. *Palaaios* **4**:157-167.

741 **Hoare RD. 1961.** Desmoinesian Brachiopoda and Mollusca from southwest Missouri. *Missouri*
742 *University Studies* **36**:1-262.

743 **Hopkins MJ. 2011.** How species longevity, intraspecific morphological variation, and
744 geographic range size are related: a comparison using Late Cambrian trilobites.

745 *Evolution* **65**:3253-3273.

746 **House MR. 1985.** Ammonoid extinction events. *Philosophical Transactions of the Royal*
747 *Society of London Series B* **325**:307-326.

748 **Hunt G, Roy K, Jablonski D. 2005.** Species-level heritability reaffirmed: a comment on 'on the
749 heritability of geographic range sizes'. *American Naturalist* **166**:129-135.

750 **Hyatt A. 1891.** Carboniferous cephalopods. *Geological Survey of Texas Annual Report* **2**:329-356.

751 **Hyatt A. 1893.** Carboniferous cephalopods: Second paper. *Geological Survey of Texas Annual*
752 *Report* **4**:377-474.

753 **Isbell JL. 2003.** Timing of the late Paleozoic glaciation in Gondwana: was glaciation responsible
754 for the development of northern hemisphere cyclothems? *Geological Society of America*
755 *Special Paper* **370**:5-24.

756 **Jablonski D. 1986.** Larval ecology and macroevolution in marine invertebrates. *Bulletin of*
757 *Marine Science* **39**:565-587.

758 **Jablonski D, Hunt G. 2015.** Larval ecology, geographic range, and species survivorship in
759 Cretaceous mollusks: organismic versus species-level explanations. *American Naturalist*
760 **168**:556-564.

761 **Jablonski D, Roy K. 2003.** Geographical range and speciation in fossil and living
762 molluscs. *Proceedings of the Royal Society of London, Series B* **270**:401-406.

763 **Jacobs DK, Landman NH, Chamberlain JA Jr. 1994.** Ammonite shell shape covaries with
764 facies and hydrodynamics: iterative evolution as a response to changes in basinal
765 environment. *Geology* **22**:905-908.

766 **Joachimski MM, Lambert LL. 2015.** Salinity contrast in the US Midcontinent Sea during

767 Pennsylvanian glacio-eustatic highstands: evidence from conodont apatite $\delta^{18}\text{O}$.
 768 *Palaeogeography, Palaeoclimatology, Palaeoecology* **433**:71-80.

769 **Kaiser SI, Becker RT, Steuber T, Aboussalam SZ. 2011.** Climate-controlled mass extinctions,
 770 facies and sea-level changes around the Devonian-Carboniferous boundary in the
 771 eastern Anti-Atlas (SE Morocco). *Palaeogeography, Palaeoclimatology, Palaeoecology*
 772 **310**:340-364.

773 **Keyes CR. 1894.** Paleontology of Missouri. *Missouri Geological Survey* **4**:1-226.

774 **Kiessling W, Aberhan M. 2007.** Geographical distribution and extinction risk: lessons from
 775 Triassic-Jurassic marine benthic organisms. *Journal of Biogeography* **34**:1473-1489.

776 **Klug C, Korn D, De Baets K, Kruta I, Mapes RH, eds. 2015.** *Ammonoid Paleobiology: from*
 777 *Macroevolution to Paleogeography*. Berlin: Springer.

778 **Kolis K. 2017.** The biogeography and macroevolutionary trends of late Paleozoic cephalopods in
 779 the North American Midcontinent Sea: understanding the response of pelagic organisms
 780 to changing climate during the Late Paleozoic Ice Age. Master's Thesis, Department of
 781 Ecology & Evolutionary Biology, University of Kansas.

782 **Korn D, Klug C. 2012.** Paleozoic ammonoids - diversity and development of conch
 783 morphology. In: Talent JA, ed. *Earth and Life, International Year of Planet Earth*.
 784 Berlin: Springer, 491-534.

785 **Korn D, Klug C, Walton SA. 2015.** Taxonomic diversity and morphological disparity of
 786 Paleozoic ammonoids. In: Klug C, Korn D, De Baets K, Kruta I, Mapes RH, eds.
 787 *Ammonoid Paleobiology: from Macroevolution to Paleogeography*. Berlin: Springer,
 788 231-264.

789 **Kröger B. 2005.** Adaptive evolution in Paleozoic coiled cephalopods. *Paleobiology* **31**:253-268.

790 **Kröger B, Mapes RH. 2005.** Revision of some common Carboniferous genera of North American
791 orthocerid nautiloids. *Journal of Paleontology* **79**:1002-1011.

792 **Kullmann J. 1983.** Maxima im tempo der evolution Karbonischer Ammonoideen.
793 *Paläontologische Zeitschrift* **57**:231-240.

794 **Kullmann J. 1985.** Drastic changes in Carboniferous ammonoid rates of evolution. In: Bayer U,
795 Seilacher A, eds. *Sedimentary and Evolutionary Cycles. Lecture Notes in Earth Sciences*,
796 vol. 1. Berlin: Springer, 35-47.

797 **Kullmann J, Wagner RH, Winkler Prins CF. 2007.** Significance for international correlation
798 of the Perapertú Formation in northern Palencia, Cantabrian Mountains.
799 Tectonic/stratigraphic context and description of Mississippian and upper Bashkirian
800 goniatites. *Revista Española de Paleontología* **22**:127-145.

801 **Kues BS. 1995.** Marine fauna of the Early Permian (Wolfcampian) Robledo Mountains Member,
802 Hueco Formation, southern Robledo Mountains, New Mexico. *New Mexico Museum of*
803 *Natural History and Science Bulletin* **6**:63-90.

804 **Kummel B. 1953.** American Triassic coiled nautiloids. *U. S. Geological Survey Professional Paper*
805 **250**:1-149.

806 **Kummel B. 1963.** Miscellaneous nautilid type species of Alpheus Hyatt. *Bulletin of the Museum of*
807 *Comparative Zoology* **128**:325-368.

808 **Lam AR, Stigall AL, Matzke N. 2018.** Dispersal in the Ordovician: speciation patterns and
809 paleobiogeographic analyses of brachiopods and trilobites. *Palaeogeography*,
810 *Palaeoclimatology, Palaeoecology* **489**:147-165.

811 **Landman N, Tanabe K, Davis RA, eds. 1996.** Ammonoid Paleobiology. New York: Plenum.

812 **Leighton LR. 2005.** The latitudinal diversity gradient through deep time: testing the "Age of
813 Tropics" hypothesis using Carboniferous productidine brachiopods. *Evolutionary*
814 *Ecology* **19**:563-581.

815 **Liow LH. 2007.** Does versatility as measured by geographic range, bathymetric range and
816 morphological variability contribute to taxon longevity? *Global Ecology and*
817 *Biogeography* **16**:117-128.

818 **Lieberman BS. 2000.** *Paleobiogeography: Using Fossils to Study Global Change, Plate*
819 *Tectonics, and Evolution*. New York: Kluwer Academic/Plenum.

820 **Lieberman BS. 2002.** Biogeography with and without the fossil record. *Palaeogeography,*
821 *Palaeoclimatology, Palaeoecology* **178**:39-52.

822 **Lieberman BS, Kimmig J. 2018.** Museums, paleontology, and a biodiversity science based
823 approach. In: Rosenberg GD, Clary RM, eds. *Museums at the Forefront of the History of*
824 *Geology: History Made, History in the Making. Geological Society of America Special*
825 *Paper* **535**. In press. [https://doi.org/10.1130/2018.2535\(22\)](https://doi.org/10.1130/2018.2535(22))

826 **Lieberman BS, Melott AL. 2013.** Declining volatility, a general property of disparate systems:
827 from fossils, to stocks, to the stars. *Palaeontology* **56**:1297-1304.

828 **Marshall CR, Finnegan S, Clites EC, Holroyd PA, Bonuso N, Cortez C, Davis E, Dietl GP,**
829 **Druckenmiller PS, Eng RC, Garcia C, Estes-Smargiassi K, Hendy A, Hollis KA,**
830 **Little H, Nesbitt EA, Roopnarine P, Skibinski L, Vendetti J, White LD. 2018.**
831 Quantifying the dark data in museum fossil collections as palaeontology undergoes a
832 second digital revolution. *Biology Letters* **14**:20180431.

833 **Mather KF. 1915.** The fauna of the Morrow group of Arkansas and Oklahoma. *Bulletin of Science*

834 *Laboratories of Denison University* **18**:59-284.

835 **Mayr E. 1942.** *Systematics and the Origin of Species*. Cambridge: Harvard University Press.

836 **McCaleb JA. 1963.** The goniatite fauna from the Pennsylvanian Winslow Formation of northwest
837 Arkansas. *Journal of Paleontology* **37**:110-115.

838 **McChesney AM. 1860.** Descriptions of new species of fossils from the Paleozoic rocks of the
839 western states. *Transactions of the Chicago Academy of Sciences* **1**:1-76.

840 **Meek FB, Worthen AH. 1860.** Descriptions of new Carboniferous fossils from Illinois and other
841 western states. *Proceeding of the Academy of Natural Sciences of Philadelphia* **4**:447-472.

842 **Meek FB, Worthen AH. 1870.** Descriptions of new species and genera of fossils from the
843 Palaeozoic rocks of the western states. *Proceedings of the Academy of Natural Sciences of*
844 *Philadelphia* **22**:22-56.

845 **Mii HS, Grossman EL, Yancey TE. 1999.** Carboniferous isotope stratigraphies of North
846 America: implications for Carboniferous paleoceanography and Mississippian glaciation.
847 *Geological Society of America Bulletin* **111**:960-973.

848 **Miller AK. 1930.** A new ammonoid fauna of Late Paleozoic age from western Texas. *Journal of*
849 *Paleontology* **4**:383-412.

850 **Miller AK, Cline LM. 1934.** The cephalopod fauna of the Pennsylvanian Nellie Bly Formation of
851 Oklahoma. *Journal of Paleontology* **8**:171-185.

852 **Miller AK, Downs R. 1948.** A cephalopod fauna from the type section of the Pennsylvanian
853 "Winslow Formation" of Arkansas. *Journal of Paleontology* **22**:672-680.

854 **Miller AK, Downs RH. 1950.** Ammonoids of the Pennsylvanian Finis Shale of Texas. *Journal of*

855 *Paleontology* **24**:185-218.

856 **Miller AK, Furnish WM. 1940a.** Permian Ammonoids of the Guadalupe Mountain region and
857 adjacent areas. *Geological Society of America, Special Papers* **26**:1-238.

858 **Miller AK, Furnish WM. 1940b.** Studies of Carboniferous Ammonoids, Parts 5-7. *Journal of*
859 *Paleontology* **14**:521-543.

860 **Miller AK, Furnish WM. 1957.** Introduction to Ammonoidea. Pp. L1-L6 in W. J. Arkell et al., *Part*
861 *L Mollusca 4 Cephalopoda Ammonoidea. Treatise on Invertebrate Paleontology*. Lawrence,
862 Kansas: Geological Society of America.

863 **Miller AK, Moore A, 1938.** Cephalopods from the Carboniferous Morrow group of northern
864 Arkansas and Oklahoma. *Journal of Paleontology* **22**:341-354.

865 **Miller AK, Owen JB. 1934.** Cherokee nautiloids of the northern Mid-Continent region. *University*
866 *of Iowa Studies in Natural History* **16**:185-272.

867 **Miller AK, Owen JB. 1937.** A new Pennsylvanian cephalopod fauna from Oklahoma. *Journal of*
868 *Paleontology* **11**:403-422.

869 **Miller AK, Owen JB. 1939.** An ammonoid fauna from the lower Pennsylvanian Cherokee
870 Formation of Missouri. *Journal of Paleontology* **13**:141-162.

871 **Miller AK, Thomas HD. 1936.** The Casper Formation (Pennsylvanian) of Wyoming and its
872 cephalopod fauna. *Journal of Paleontology* **10**:715-738.

873 **Miller AK, Unklesbay AG. 1942.** Permian nautiloids from western United States. *Journal of*
874 *Paleontology* **16**:719-738.

875 **Miller AK, Youngquist W. 1947.** Lower Permian Cephalopods from the Texas Colorado River

876 Valley. *The University of Kansas Paleontological Contributions* 2:1-15.

877 **Miller AK, Youngquist WL. 1949.** American Permian nautiloids. *Geological Society of*
878 *America Memoir* 41:1-217.

879 **Miller AK, Dunbar CO, Condra GE. 1933.** The nautiloid cephalopods of the Pennsylvanian
880 System in the Mid-continent region. *Nebraska Geological Survey Bulletin* 9:1-240.

881 **Miller AK, Lane JH, Unklesbay AG. 1947.** A nautiloid cephalopod fauna from the Pennsylvanian
882 Winterset Limestone of Jackson Country, Missouri. *University of Kansas Paleontological*
883 *Contributions* 2:1-11.

884 **Miller AK, Youngquist W, Nielsen ML. 1952.** Mississippian cephalopods from western Utah.
885 *Journal of Paleontology* 26:148-161.

886 **Miller HW, Breed WJ. 1964.** *Metacoceras bowmani*, a new species of nautiloid from the Toroweap
887 Formation (Permian) of Arizona. *Journal of Paleontology* 38:877-880.

888 **Miller SA. 1892.** Palaeontology. *Geological Survey of Indiana Annual Report Advance Sheets* 18.

889 **Minitab 17 Statistical Software. 2016.** State College, PA: Minitab, Inc.

890 **Monnet C, De Baets K, Klug C. 2011.** Parallel evolution controlled by adaptation and
891 covariation in ammonoid cephalopods. *BMC Evolutionary Biology* 11:115, 1-21.

892 **Montañez IP. 2007.** CO₂-forced climate and vegetational instability during Late Paleozoic
893 deglaciation. *Science* 315:87-91.

894 **Montañez IP, Poulsen CJ. 2013.** The late Paleozoic Ice Age: an evolving paradigm. *Annual*
895 *Review of Earth Planet Science* 41:629-656.

896 **Myers CE, Lieberman BS. 2011.** Sharks that pass in the night: using Geographical Information

897 Systems to investigate competition in the Cretaceous Interior Seaway. *Proceedings of the*
898 *Royal Society of London, Series B* **278**:681-689.

899 **Myers CE, Saupe EE. 2013.** A macroevolutionary expansion of the modern synthesis and the
900 importance of extrinsic abiotic factors. *Palaeontology* **56**:1179-1198.

901 **Myers CE, MacKenzie RA, Lieberman BS. 2013.** Greenhouse biogeography: the relationship
902 of geographic range to invasion and extinction in the Western Interior Seaway.
903 *Paleobiology* **39**:135-148.

904 **Nassichuk WW. 1975.** Carboniferous ammonoids and stratigraphy in the Canadian Arctic
905 archipelago. *Geological Survey of Canada Bulletin* **237**:1-240.

906 **Nelson WJ, Lucas SG. 2011.** Carboniferous geologic history of the Rocky Mountain region.
907 *New Mexico Museum of Natural History and Science Bulletin* **53**:115-142.

908 **Newell ND. 1936.** Some mid-Pennsylvanian invertebrates from Kansas and Oklahoma: III.
909 Cephalopoda. *Journal of Paleontology* **10**:481-489.

910 **Niko S, Mapes RH. 2009.** Redescription and new information on the Carboniferous cephalopod
911 *Brachycycloceras normale* Miller, Dunbar and Condra, 1933. *Paleontological Research*
912 **13**:337-343.

913 **Orzechowski EA, Lockwood R, Byrnes JEK, Anderson SC, Finnegan S, Finkel ZV, Harnik**
914 **PG, Lindberg DR, Liow LH, Lotze HK, McClain CR, McGuire JL, O'Dea A,**
915 **Pandolfi JM, Simpson C, Tittensor D. 2015.** Marine extinction risk shaped by
916 trait-environment interactions over 500 million years. *Global Change Biology* **21**:3595-
917 3607.

918 **Payne JL, Finnegan S. 2007.** The effect of geographic range on extinction risk during

919 background and mass extinction. *Proceedings of the National Academy of Sciences, USA*
920 **104**:10506-10511.

921 **Pie MR, Meyer ALS. 2017.** The evolution of range sizes in mammals and squamates:
922 heritability and differential evolutionary rates for low- and high-latitude limits.
923 *Evolutionary Biology* **44**:347-355.

924 **Plummer FB, Scott G. 1937.** Upper Paleozoic ammonites of Texas: the geology of Texas.
925 *University of Texas Bulletin* **3701, vol 3, part 1**:1-516.

926 **Pope JP. 2012.** Description of Pennsylvanian units, revision of stratigraphic nomenclature and
927 reclassification of the Morrowan, Atokan, Desmoinesian, Missourian, and Virgilian
928 stages in Iowa. *Iowa Department of Natural Resources Special Report Series* **5**:1-140.

929 **Powell MG. 2005.** Climatic basis for sluggish macroevolution during the late Paleozoic ice age.
930 *Geology* **33**:381-384.

931 **Powell MG. 2007.** Latitudinal diversity gradients for brachiopod genera during late Paleozoic
932 time: links between climate, biogeography and evolutionary rates. *Global Ecology and*
933 *Biogeography* **16**:519-528.

934 **R version 3.4.0. 2017.** *You Stupid Darkness*. Vienna, Austria: R foundation for statistical
935 computing.

936 **Ramsbottom WHC. 1981.** Eustatic control in Carboniferous ammonoid biostratigraphy. In:
937 House MR, Senior JR, eds. *The Ammonoidea, Systematics Association Special Volume*
938 **18**. London: Academic Press, 369-398.

939 **Raymond A, Metz C. 2004.** Ice and its consequences: glaciation in the Late Ordovician, Late

940 Devonian, Pennsylvanian-Permian and Cenozoic compared. *Journal of Geology* **112**:655-
941 670.

942 **Rios NE, Bart HL Jr. 2018.** GEOlocate: a platform for georeferencing natural history
943 collections data. <https://www.geo-locate.org> .

944 **Ritterbush KA, Hoffmann R, Lukender A, De Baets K. 2014.** Pelagic palaeoecology: the
945 importance of recent constraints on ammonoid palaeobiology and life history. *Journal of*
946 *Zoology* **292**:229-241.

947 **Roark A, Flake R, Grossman EL, Olszewski T, Lebold J, Thomas D, Marcantonio F, Miller**
948 **B, Raymond A, Yancey T. 2017.** Brachiopod geochemical records from across the
949 Carboniferous seas of North America: evidence for salinity gradients, stratification, and
950 circulation patterns. *Palaeogeography, Palaeoclimatology Palaeoecology* **485**:136-153.

951 **Rode AL, Lieberman BS. 2004.** Using GIS to unlock the interactions between biogeography,
952 environment, and evolution in Middle and Late Devonian brachiopods and bivalves.
953 *Palaeogeography, Palaeoclimatology, Palaeoecology* **211**:345-359.

954 **Rode AL, Lieberman BS. 2005.** Intergrating evolution and biogeography: a case study
955 involving Devonian crustaceans. *Journal of Paleontology* **79**:267-276.

956 **Rojas A, Patarroyo P, Mao L, Bengtson P, Kowalewski M. 2017.** Global biogeography of
957 Albian ammonoids: a network-based approach. *Geology* **45**:659-662.

958 **Rook DL, Heim NA, Marcot J. 2013.** Contrasting patterns and connections of rock and biotic
959 diversity in the marine and non-marine fossil records of North America.
960 *Palaeogeography, Palaeoclimatology, Palaeoecology* **372**:123-129.

961 **Rothwell Group LP. 2016.** *PaleoWeb: Free Plate Tectonics Software*. Lakewood, CO.

- 962 **Ruzhentsev VE, Bogoslovskaya MF. 1971.** Namurskiy etap v evolyutsii ammonoidey.
 963 Rannenamyurskie ammonoidei. *Akademiya Nauk SSSR, Trudy Paleontologicheskogo*
 964 *Instituta* **133**:1-382.
- 965 **Ruzhentsev VE, Shimanskiy VN. 1954.** Nizhnepermskie svernutye i sognutye Nautiloidei
 966 yuzhnogo Urala. *Akademiya Nauk SSSR, Trudy Paleontologicheskogo Instituta* **50**:1-150.
- 967 **Saupe EE, Qiao H, Hendricks JR, Portell RW, Hunter SJ, Soberón J, Lieberman BS. 2015.**
 968 Niche breadth and geographic range size as determinants of species survival on
 969 geological time scales. *Global Ecology and Biogeography* **24**:1159-1169.
- 970 **Sawin RS, West RR, Franseen EK, Watney WL, McCauley JR. 2006.** Carboniferous-
 971 Permian boundary in Kansas, midcontinent, USA: current research in earth sciences.
 972 *Kansas Geological Survey, Bulletin* **252, Part 2**:1-13.
- 973 **Sawin RS, Franseen EK, West RR, Ludvigson GA, Watney WL. 2008.** Clarification and
 974 changes in Permian stratigraphic nomenclature in Kansas: current research in earth
 975 sciences. *Kansas Geological Survey, Bulletin* **254, Part 2**:1-4.
- 976 **Sawin RS, Franseen EK, Watney WL, West RR, Ludvigson GA. 2009.** New stratigraphic
 977 rank for the Carboniferous, Mississippian, and Pennsylvanian in Kansas: current research
 978 in earth sciences. *Kansas Geological Survey, Bulletin* **256, Part 1**:1-4.
- 979 **Sayre AN. 1930.** The fauna of the Drum Limestone of Kansas and western Missouri. *University of*
 980 *Kansas Science Bulletin* **19**:1-203
- 981 **Schneider CL. 2018.** Marine refugia past, present, and future: lessons from ancient geologic
 982 crises for modern marine ecosystem conservation. In: Tyler CL, Schneider CL, eds..
 983 *Marine Conservation Paleobiology*. Berlin: Springer, 163-208.

984 **Segessenman DC, Kammer TW. 2018.** Testing evolutionary rates during the late Paleozoic ice
 985 age using the crinoid fossil record. *Lethaia* **51**:330-343.

986 **Sepkoski JJ Jr. 1998.** Rates of speciation in the fossil record. *Philosophical Transactions of the*
 987 *Royal Society of London* **353**:315-326.

988 **Simões M, Breitzkreuz L, Alvarado M, Baca S, Cooper JC, Heins L, Herzog K, Lieberman**
 989 **BS. 2016.** The evolving theory of evolutionary radiations. *Trends in Ecology and*
 990 *Evolution (TREE)* **31**:27-34.

991 **Smith HJ. 1938.** *The Cephalopod Fauna of the Buckhorn Asphalt*. Chicago: University of Chicago
 992 Libraries.

993 **Smith JP. 1896.** Marine fossils from the Coal Measures of Arkansas. *Proceedings of the American*
 994 *Philosophical Society* **35**:213-285.

995 **Smith JP. 1903.** The Carboniferous ammonoids of America. *Monographs of the United States*
 996 *Geological Survey* **52**:1-211.

997 **Sokal RR, Rohlf FJ. 1994.** *Biometry: the Principles and Practices of Statistics in Biological*
 998 *Research*. San Francisco: W. H. Freeman.

999 **Stanley SM. 1979.** *Macroevolution*. San Francisco: W. H. Freeman.

1000 **Stanley SM. 1990.** The general correlation between rate of speciation and rate of extinction:
 1001 fortuitous causal linkages. In: Ross RM, Allmon WD, eds. *Causes of Evolution: a*
 1002 *Paleontological Perspective*. Chicago: University of Chicago Press, 103-127.

1003 **Stanley SM, Powell MG. 2003.** Depressed rates of origination and extinction during the late
 1004 Paleozoic ice age: a new state for the global marine ecosystem. *Geology* **31**:877-880.

1005 **Stigall AL. 2010.** Using GIS to assess the biogeographic impact of species invasions on native
 1006 brachiopods during the Richmondian Invasion in the type-Cincinnatian (Late Ordovician,
 1007 Cincinnati region). *Palaeontologia Electronica* **13(1)**, 5A:1-19.

1008 **Stigall AL, Lieberman BS. 2006.** Quantitative palaeobiogeography: GIS, phylogenetic
 1009 biogeographical analysis and conservation insights. *Journal of Biogeography* **33**:2051-
 1010 2060.

1011 **Sturgeon MT. 1946.** Allegheny fossil invertebrates from eastern Ohio-Nautiloidea. *Journal of*
 1012 *Paleontology* **20**:8-37.

1013 **Sturgeon MT, Windle DL, Mapes RH, Hoare RD. 1982.** New and revised taxa of Pennsylvanian
 1014 cephalopods in Ohio and West Virginia. *Journal of Paleontology* **56**:1453-1479.

1015 **Swallow GC. 1858.** Rocks of Kansas with descriptions of new Permian fossils. *Transactions of the*
 1016 *Academy of Sciences St. Louis* **1**:1-27.

1017 **Tabor NJ. 2007.** Permo-Pennsylvanian palaeotemperatures from Fe-Oxide and phyllosilicate
 1018 δO_{18} values. *Earth and Planetary Science Letters* **253**:159-171.

1019 **Tabor NJ, Poulsen CJ. 2007.** Paleoclimate across the late Pennsylvanian-early Permian tropical
 1020 paleolatitudes: a review of climate indicators, their distribution, and relation to
 1021 palaeophysiographic climate factors. *Palaeogeography, Palaeoclimatology,*
 1022 *Palaeoecology* **268**:293-310.

1023 **Teichert C. 1940.** Contributions to nautiloid nomenclature. *Journal of Paleontology* **14**:590-597.

1024 **Teichert C, Kummel B, Sweet WC, Stenzel HB, Furnish WM, Glenister BF, Erben HK, Moore**
 1025 **RC, Zeller D. 1964.** Part K Mollusca 3 Cephalopoda—General Features, Endoceratoidea—
 1026 Actinoceratoidea—Nautiloidea—Bacritoidea. *Treatise on Invertebrate Paleontology*.

1027 Lawrence, Kansas: The Geological Society of America.

1028 **Unklesbay AG. 1954.** Distribution of American Pennsylvanian cephalopods. *Journal of*
1029 *Paleontology* **28**:84-95.

1030 **Unklesbay AG, Palmer EJ. 1958.** Cephalopods from the Burgner Formation in Missouri. *Journal of*
1031 *Paleontology* **32**:1071-1076.

1032 **US Geological Survey. 2017.** U.S. Geologic Names Lexicon ("Geolex"). Retrieved from the
1033 National Geologic Map Database: <https://ngmdb.usgs.gov/Geolex/search> .

1034 **Vrba ES. 1980.** Evolution, species, and fossils: how does life evolve? *South African Journal of*
1035 *Science* **76**:61-84.

1036 **Ward PD, Westermann GEG. 1985.** Cephalopod paleoecology. *Studies in Geology, Notes for a*
1037 *Short Course (Mollusks)* **13**:215-229.

1038 **Waterhouse JB, Shi GR. 2010.** Late Palaeozoic global changes affecting high-latitude
1039 environments and biotas: an introduction. *Palaeogeography, Palaeoclimatology,*
1040 *Palaeoecology* **298**:1-16.

1041 **Wells MR, Allison PA, Piggott MD, Gorman GJ, Hampson GJ, Pain CC, Fang F. 2007.**
1042 Numerical modeling of tides in the late Pennsylvanian Midcontinent Seaway of North
1043 America with implications for hydrography and sedimentation. *Journal of Sedimentary*
1044 *Research* **77**:843-865.

1045 **White CA. 1889.** On the Permian formation of Texas. *The American Naturalist* **23**:109-128.

1046 **White CA, St. John OH. 1867.** Descriptions of new Subcarboniferous and Coal Measure fossils
1047 collected upon the Geological Survey of Iowa; together with a notice of new generic

1048 characters observed in two species of brachiopods. *Transactions of the Chicago Academy of*
 1049 *Sciences* **1**:115-127.

1050 **White RD, Skorina LK. 1999.** A type catalog of fossil invertebrates (Mollusca: Actinoceratoidea,
 1051 Bactritoidea, Endoceratoidea, and Nautiloidea) in the Yale Peabody Museum. *Postilla* **219**:1-
 1052 39.

1053 **Wieczorek C, Wieczorek J. 2015.** Georeferencing calculator (version 20160920). Museum of
 1054 Vertebrate Zoology, University of California, Berkeley. <http://manisnet.org/gci2.html> .

1055 **Wiedmann J, Kullmann J. 1996.** Crises in ammonoid evolution. In: Landman NH, Tanabe K,
 1056 Davis RA, eds. *Ammonoid Paleobiology*. Boston: Springer, 795-813.

1057 **Wiley EO, Lieberman BS. 2011.** *Phylogenetics, 2nd edition*. New York: J. Wiley & Sons.

1058 **Yacobucci MM. 2017.** Marine life in a greenhouse world: cephalopod biodiversity and
 1059 biogeography during the early Late Cretaceous. *Paleobiology* **43**:587-619.

1060 **Young JA. 1942.** Pennsylvanian Scaphopoda and Cephalopoda from New Mexico. *Journal of*
 1061 *Paleontology* **16**:120-125.

1062 **Zeller DE, ed. 1968.** The stratigraphic succession in Kansas. *Kansas Geological Survey, Bulletin*
 1063 **189**:1-81.

1064 **Zhang M, Becker RT, Ma X, Zhang Y, Zong. 2019.** Hangenberg Black Shale with
 1065 cymaclymeniid ammonoids in the terminal Devonian of South China. *Palaeobiodiversity*
 1066 *and Palaeoenvironments*. **In press**. <https://doi.org/10.1007/s12549-018-0348-x>
 1067
 1068
 1069

1070

1071 **Figure Captions**

1072

1073 **Figure 1: Distribution of Pennsylvanian and early Permian cephalopods.**

1074 A) Distribution of Pennsylvanian nautiloid and ammonoid data points (red) and B) early Permian
1075 nautiloid and ammonoid data points (blue) across the midcontinent region of North America.
1076 Plotted using *ArcGIS* v. 10.3 (ESRI, 2014) software at 1: 20,000,000.

1077

1078 **Figure 2: Occurrence points of *Metacoceras* sp. and *Mooreoceras* sp.**

1079 For the Virgilian, shown on possible paleogeography of that stage, at 1:1,000,000,000 scale;
1080 plotted using *PaleoWeb* (The Rothwell Group LP, 2016).

1081

1082 **Figure 3: Mean geographic range size in km² of cephalopods through time.**

1083 Nautiloid species (A) and ammonoid species (B) range changes occur but are not statistically
1084 significant when analyzed using non-parametric tests (note, median range size data not graphed
1085 but for all cephalopods they are 79km² for all time intervals, for ammonoids they are 78.5km² for
1086 the Desmoinesian and Wolfcampian and 79km² for all other time intervals, and for nautiloids
1087 they are 79km² for all time intervals) or when log transformed data are analyzed using
1088 parametric tests (note log transformed data not graphed but mean transformed values for all
1089 cephalopods are 5.51 [standard error 0.75] for the Morrowan, 4.05 [standard error 1.02] for the
1090 Atokan, 4.36 [standard error 0.49] for the Desmoinesian, 5.65 [standard error 0.49] for the
1091 Missourian, 5.96 [standard error 0.79] for the Virgilian, and 4.31 [standard error 0.52] for the

Eliminado: ¶

¶
¶
¶
¶
¶
¶
¶

1101 Wolfcampian).

1102

1103 **Figure 4: Speciation and extinction rates through time.**

1104 Values given in per Myr and derived from Table 1.