

Supplemental Table S3:

Geographic range values through time in km² by stage (youngest to oldest from left to right) for the species considered in the analysis.

Genus	species	Wolfcampian	Virgilian	Missourian	Desmoinesian	Atokan	Morrowan
<i>Brachycycloeras</i>	<i>bransonii</i>	0	78.539816	78.539816	0.250798	0	0
<i>Brachycycloeras</i>	<i>crebrincinctum</i>	0	0.523798	78.539816	1176.718476	0	0
<i>Brachycycloeras</i>	<i>curtum</i>	0	377.412261	78.539816	78.539816	0	0
<i>Brachycycloeras</i>	<i>longulum</i>	0	1065.654683	78.539816	0.833697	0	0
<i>Brachycycloeras</i>	<i>normale</i>	0	0.494043	78.539816	8757.000000	78.539816	0
<i>Domatoceras</i>	<i>bradyi</i>	0	78.539816	78.539816	0	0	0
<i>Domatoceras</i>	<i>kleihegei</i>	0	0	0.220338	78.539816	0	0
<i>Domatoceras</i>	<i>moorei</i>	0	0	117036.000000	78.539816	0	0
<i>Domatoceras</i>	<i>sculptile</i>	0	78.539816	78.539816	78.539816	0	0
<i>Domatoceras</i>	<i>umbilicatum</i>	78.539816	12010.000000	3440.000000	12489.577104	0	0
<i>Domatoceras</i>	<i>williamsi</i>	78.539816	78.539816	0.729789	3762.000000	0	0
<i>Ephippioceras</i>	<i>ferratum</i>	0	2810.000000	127692.000000	3269.000000	0	0
<i>Euloxoceras</i>	<i>greenii</i>	0	7594.000000	3827.000000	78.539816	0	0
<i>Gonioloboceras</i>	<i>bridgeportensis</i>	0	0	78.539816	0	0	0
<i>Mescalites</i>	<i>discoidalis</i>	78.539816	0	0	0	0	0
<i>Gonioloboceras</i>	<i>elaisi</i>	0	0	0	0.466069	0	0
<i>Gonioloboceras</i>	<i>goniolobum</i>	0	15174.000000	78.539816	78.539816	0	0
<i>Gonioloboceras</i>	<i>gracellenae</i>	0	78.539816	0	0	0	0
<i>Gonioloboceras</i>	<i>welleri</i>	0	115412.000000	78.539816	78.539816	0	0
<i>Hebertothoceras</i>	<i>unicamera</i>	0	0	0	0.312085	0	0
<i>Knightoceras</i>	<i>abundum</i>	0	0	0.149475	0	0	0
<i>Knightoceras</i>	<i>missouriense</i>	0	0	0	78.539816	0	0
<i>Liroceras</i>	<i>liratum</i>	0	0	119598.000000	78.539816	78.539816	6791.691877
<i>Liroceras</i>	<i>milleri</i>	0	0	4731.000000	78.539816	78.539816	0
<i>Megapronotites</i>	<i>baconi</i>	0	0	0	0	78.539816	78.539816
<i>Metacoceras</i>	<i>angulatum</i>	78.539816	13702.448632	9458.490734	0.130773	0	0
<i>Metacoceras</i>	<i>bituberculatum</i>	0	0	0	78.539816	0	0
<i>Metacoceras</i>	<i>bowmani</i>	0	0	9231.47593	0	0	0
<i>Metacoceras</i>	<i>cheneyi</i>	0	78.539816	0.639810	78.539816	0	0
<i>Metacoceras</i>	<i>cornutum</i>	0	85771.000000	0.793486	670.592743	0	0
<i>Metacoceras</i>	<i>dubium</i>	5745.847979	78.539816	0.101843	0	0	0

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<i>Metacoceras</i>	<i>inconspicuim</i>	0	0	78.539816	0	0	0
<i>Metacoceras</i>	<i>jacksonense</i>	0	78.539816	2214.000000	0	0	0
<i>Metacoceras</i>	<i>knighti</i>	0	0	0.497333	0	0	0
<i>Metacoceras</i>	<i>mutabile</i>	0	0	4982.000000	0.175450	0	0
<i>Metacoceras</i>	<i>nodosum</i>	78.539816	78.539816	4673.000000	78.539816	0	0
<i>Metacoceras</i>	<i>perelegans</i>	0	0	0.793486	0	0	0
<i>Metacoceras</i>	<i>sulciferum</i>	0	78.539816	78.539816	0	0	0
<i>Metacoceras</i>	<i>sublaeve</i>	78.539816	0	0	0	0	0
<i>Milikoninckioeras</i>	<i>elaisi</i>	0	0	1022.815493	0	0	0
<i>Milikoninckioeras</i>	<i>jewetti</i>	0	0	78.539816	0	0	0
<i>Milikoninckioeras</i>	<i>wyandottense</i>	0	0	78.539816	0	0	0
<i>Mooreoceras</i>	<i>bakeri</i>	0	4981.349200	4915.000000	0	0	0
<i>Mooreoceras</i>	<i>condrai</i>	0	78.539816	3676.000000	78.539816	78.539816	0
<i>Mooreoceras</i>	<i>conicum</i>	0	0	4467.000000	78.539816	0	0
<i>Mooreoceras</i>	<i>giganteum</i>	0	78.539816	0	0	0	0
<i>Mooreoceras</i>	<i>normale</i>	0	0	4979.000000	8202.000000	78.539816	78.539816
<i>Mooreoceras</i>	<i>ovale</i>	0	78.539816	2339.000000	0	0	0
<i>Mooreoceras</i>	<i>tuba</i>	0	0	78.539816	0	0	0
<i>Mooreoceras</i>	<i>wedingtonianum</i>	0	0	3758.113739	0	0	0
<i>Orthoceras</i>	<i>dunbari</i>	0	0	78.539816	78.539816	0	0
<i>Orthoceras</i>	<i>kansasense</i>	0	17195728778.000000	610.050914	78.539816	0	0
<i>Orthoceras</i>	<i>longissimicameratum</i>	0	78.539816	78.539816	78.539816	0	0
<i>Orthoceras</i>	<i>occidentale</i>	0	78.539816	78.539816	78.539816	0	0
<i>Parashumardites</i>	<i>senex</i>	0	78.539816	0	0	0	0
<i>Phaneroceeras</i>	<i>compressum</i>	0	0	0	0	24924.000000	78.539816
<i>Phaneroceeras</i>	<i>kesslerense</i>	0	0	0	0	0.368368	0
<i>Pseudoparategoceras</i>	<i>brazoense</i>	0	0	0	0.505825	0.001144	0
<i>Pseudorthoceras</i>	<i>knoxense</i>	78.539816	71899.000000	7577.000000	64660.000000	30599.000000	0
<i>Pseudopronorites</i>	<i>arkansasensis</i>	0	0	0	0	0	8916.063243
<i>Pseudopronorites</i>	<i>kansasensis</i>	0	0	78.539816	0	0	0
<i>Pronorites</i>	<i>pseudotimorensis</i>	0	78.539816	78.539816	78.539816	78.539816	78.539816
<i>Properrinites</i>	<i>boesei</i>	0.559262	0	0	0	0	0
<i>Properrinites</i>	<i>cumminsi</i>	78.539816	0	0	0	0	0
<i>Properrinites</i>	<i>plummeri</i>	78.539816	0	0	0	0	0
<i>Schistoceras</i>	<i>hildrethi</i>	0	139876.000000	23915.000000	78.539816	78.539816	78.539816
<i>Schistoceras</i>	<i>missouriense</i>	78.539816	145678.147758	145678.147758	78.539816	78.539816	0
<i>Schistoceras</i>	<i>unicum</i>	0	0.275353	78.539816	78.539816	78.539816	0

Kommentiert [Office3]: Missourian

Kommentiert [Office4]: Metapronorites; Kasimovian

Kommentiert [Office5]: Sakmarian

Kommentiert [Office6]: Paraschistoceras; Miss, Virg

Kommentiert [Office7]: Miss, Virg

Kommentiert [Office8]: Eoschistoceras; Missourian

Shumardites	cuyleri	0	0.040084	0	0	0	0
Solenochilus	brammeri	0	0	25888.015389	0	0	0
Solenochilus	kempae	0	0	3067.000000	0	0	0
Solenochilus	kerefordensis	0	2277.109781	78.539816	0	0	0
Solenochilus	missouriense	0	0	1321.366476	0	0	0
Solenochilus	newloni	0	78.539816	0	0	0	0
Solenochilus	peculiare	0	0	0	78.539816	0	0
Shumarites	simonsi	0	0.103678	0	0	0	0
Solenochilus	springeri	0	0	0	9839.217950	78.539816	78.539816
Solenochilus	syracusense	0	0	0	78.539816	0	0
Vidriocerus	uddeni	78.539816	0	0	0	0	0

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Using GIS to examine biogeographic and macroevolutionary patterns in late Paleozoic cephalopods from the North American Midcontinent Sea (#31237)

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Using GIS to examine biogeographic and macroevolutionary patterns in late Paleozoic cephalopods from the North American Midcontinent Sea

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Geographic range is an important macroevolutionary parameter frequently considered in paleontological studies as species' distributions and range sizes are determined by a variety of biotic and abiotic factors well known to affect the differential birth and death of species. Thus, considering how distributions and range sizes fluctuate over time can provide important insight into evolutionary dynamics. This study uses Geographic Information Systems (GIS) and analyses of evolutionary rates to examine how in the Cephalopoda, an important pelagic clade, geographic range size and rates of speciation and extinction changed throughout the Pennsylvanian and Early Permian in the North American Midcontinent Sea. This period is particularly interesting for biogeographic and evolutionary studies because it is characterized by repetitive interglacial-glacial cycles, a global transition from an icehouse to a greenhouse climate during the Late Paleozoic Ice Age, and sluggish macroevolutionary dynamics, i.e. low speciation and extinction rates. The analyses presented herein indicate that cephalopod species diversity was not completely static and actually fluctuated throughout the Pennsylvanian and Early Permian, matching findings from other studies. However, contrary to some other studies, the mean geographic ranges of cephalopod species did not change significantly through time, despite numerous climate oscillations; further, geographic range size did not correlate with rates of speciation and extinction. These results suggest that pelagic organisms may have responded differently to Late Paleozoic climate changes than benthic organisms, although additional consideration of this issue is needed. Finally, these results indicate that, at least in the case of cephalopods, macroevolution during the Late Paleozoic was more dynamic than previously characterized, and patterns may have varied across different clades during this interval.

1 **Using GIS to Examine Biogeographic and Macroevolutionary**
2 **patterns in Late Paleozoic Cephalopods from the North American**
3 **Midcontinent Sea**

4

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23 **Abstract**

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25 Geographic range is an important macroevolutionary parameter frequently considered in
26 paleontological studies as species' distributions and range sizes are determined by a variety of
27 biotic and abiotic factors well known to affect the differential birth and death of species. Thus,
28 considering how distributions and range sizes fluctuate over time can provide important insight
29 into evolutionary dynamics. This study uses Geographic Information Systems (GIS) and analyses
30 of evolutionary rates to examine how in the Cephalopoda, an important pelagic clade, geographic
31 range size and rates of speciation and extinction changed throughout the Pennsylvanian and
32 Early Permian in the North American Midcontinent Sea. This period is particularly interesting
33 for biogeographic and evolutionary studies because it is characterized by repetitive interglacial-
34 glacial cycles, a global transition from an icehouse to a greenhouse climate during the Late
35 Paleozoic Ice Age, and sluggish macroevolutionary dynamics, i.e. low speciation and extinction
36 rates.

37 The analyses presented herein indicate that cephalopod species diversity was not completely
38 static and actually fluctuated throughout the Pennsylvanian and Early Permian, matching
39 findings from other studies. However, contrary to some other studies, the mean geographic
40 ranges of cephalopod species did not change significantly through time, despite numerous
41 climate oscillations; further, geographic range size did not correlate with rates of speciation and
42 extinction. These results suggest that pelagic organisms may have responded differently to Late
43 Paleozoic climate changes than benthic organisms, although additional consideration of this issue
44 is needed. Finally, these results indicate that, at least in the case of cephalopods, macroevolution

45 during the Late Paleozoic was more dynamic than previously characterized, and patterns may
46 have varied across different clades during this interval.

47 **Introduction**

48

49 Much work has focused on the relationship between geographic range size and rates of
50 speciation and extinction (e.g., Vrba, 1980; Jablonski, 1986; Eldredge, 1989; Stanley, 1990;
51 Lieberman, 2000; Jablonski & Roy, 2003; Rode & Lieberman, 2004, 2005; Kiessling &
52 Aberhan, 2007; Liow, 2007; Payne & Finnegan, 2007; Abe & Lieberman, 2009; Stigall, 2010;
53 Myers & Saupe, 2013; Myers, MacKenzie, & Lieberman, 2013; Dunhill & Wills, 2015;
54 Jablonski & Hunt, 2015; Orzechowski et al., 2015; Saupe et al., 2015; Castiglione et al., 2017;
55 Pie & Meyer, 2017; Simoes et al., 2016; Lam, Stigall, & Matzke, 2018; Schneider, 2018).
56 Furthermore, the use of Geographic Information Systems (GIS) has greatly facilitated
57 investigations into this macroevolutionary relationship (Stigall & Lieberman, 2006; Hendricks,
58 Lieberman, & Stigall, 2008; Dunhill, 2012; Myers, MacKenzie, & Lieberman, 2013; Dunhill &
59 Wills, 2015; Lieberman & Kimmig, 2018). Here, we focus on how geographic range size and
60 rates of speciation and extinction changed throughout the Pennsylvanian and Early Permian in
61 the North American Midcontinent Sea in the Cephalopoda, an important clade of pe[REDACTED]
62 invertebrates (Landman, Tanah[REDACTED] & Davis, 1996; Monnet, De Baets, & C. Klug, 2011; Körn et
63 al., 2015), using GIS. This period is particularly interesting for biogeographic and evolutionary
64 analysis because it is characterized by repetitive glacial-interglacial cycles, a glob[REDACTED] transition
65 from an icehouse to greenhouse climate during the Late Paleozoic Ice Age (LPIA) (Montañez &
66 Poulsen, 2013). Further, it is generally considered a time of sluggish macroevolutionary
67 dynamics, i.e. low speciation and extinction rates and low degrees of faunal turnover, that have

68 been demonstrated in studies of other marine invertebrate taxa (Sepkoski, 1998; Stanley &
69 Powell, 2003; Bonelli & Patzkowsky, 2011). However, Balseiro (2016) did document the
70 existence of some profound **evolutionary turnover** over the course of this interval in regions
71 closer to the ice sheets, such as present day western Argentina. Furthermore, recently
72 Segessenman & Kammer (2018) showed that advanced cladid crinoids do display elevated rates
73 of evolution and turnover during this time interval, although three other subclasses of crinoids do
74 show subdued evolutionary rates. It is worth noting that fusulinid foraminifera also fit the pattern
75 shown in the advanced cladids (Groves & Lee, 2008; Groves & Yue, 2009; Segessenman &
76 Kammer, 2018).

77
78 There have been a variety of hypotheses proposed for the postulated **sluggish** macroevolutionary
79 dynamics of the LPIA. Some studies contend that this pattern is a result of environmental
80 changes linked to glacial cycling while others point to tectonic activity (Stanley and Powell,
81 2003; Powell, 2005; Fielding, Frank, & Isbell, 2008; DiMichele et al., 2009; Falcon-Lang &
82 DiMichele, 2010; Bonelli and Patzkowsky, 2011; Cecil, DiMichele, & Elrick, 2014;
83 Segessenman & Kammer, 2018). To date, many of the studies focusing on the
84 macroevolutionary dynamics of the LPIA have concentrated on benthic marine invertebrates
85 (e.g., Stanley & Powell, 2003; Powell, 2007; Bonelli & Patzkowsky, 2011; Balseiro, 2016;
86 Segessenman & Kammer, 2018) as they are highly diverse and very abundant. However, it is
87 valuable to explicitly investigate evolutionary patterns in pelagic marine invertebrates as these
88 are also diverse and abundant organisms in **Late** Paleozoic marine ecosystems (Landman,
89 Tanabe, & Davis, 1996; Monnet, De Baets, & Klug, 2011; **Körn** et al., 2015). In particular, given
90 the significant role that geographic factors play in speciation (Mayr, 1942; Eldredge & Gould,

91 1972; Jablonski, 1986; Brooks & McLennan, 1991; Wiley & Lieberman, 2011; Jablonski &
92 Hunt, 2015; Pie & Meyer, 2017), we might expect that pelagic organisms, because of their
93 innately greater dispersal ability (at least as adults), might show different patterns relative to taxa
94 that were benthic (Rojas et al., 2017; Yacobucci, 2017).

95

96 This study focuses on cephalopods from the Pennsylvanian-Early Permian (Morrowan, Atokan,
97 Desmoinesian, Missourian, Virgilian, and Wolfcampian) in the Midcontinent Sea of the United
98 States as knowledge of the systematic affinities, geographic distribution and overall diversity of
99 cephalopods during this interval is relatively well understood (Miller, Dunbar, & Condra, 1933;
100 Newell, 1936; Plummer & Scott, 1937; Miller & Youngquist, 1949; Landman, Tanabe, & Davis,
101 1996; Kröger, 2005; **Körn** et al., 2015), the stratigraphy of the region is well constrained (Heckel,
102 2008, 2013), and there are extensive exposures of fossiliferous units in the region. Moreover, at
103 this time the Midcontinent Sea was bordered by the Antler Orogeny to the north, the Ancestral
104 Rocky Mountain Orogeny to the west/northwest and the Ouachita Mountain belt to the
105 south/southeast (as well as various structural arches), such that it constituted a distinct
106 biogeographic region for marine invertebrates (Wells et al., 2007; Nelson & Lucas, 2011;
107 Joachimski & Lambert, 2015).

108

109 The **Late** Paleozoic Ice Age (LPIA) was the longest lived glacial period of the Phanerozoic and is
110 relatively well understood due to numerous stratigraphic, sedimentologic, paleontologic, and
111 isotopic studies (e.g., Mii, Grossman, & Yancey, 1999; Isbell, 2003; Stanley & Powell, 2003;
112 Raymond & Metz, 2004; Montañez, 2007; Powell, 2007; Tabor & Poulsen, 2007; Fielding,
113 Frank, & Isbell, 2008; Heckel, 2008; DiMichele et al., 2009; Bonelli & Patzkowsky, 2011;

114 Montañez & Poulsen, 2013; Balseiro, 2016; Roark et al., 2017; Segessenman & Kammer, 2018).
115 Glacial cycling in the midcontinent region has received much study (e.g., Isbell, 2003; Heckel,
116 2008, 2013). Modern synthesis of the glacial history indicates that the Morrowan to **Early**
117 Desmoinesian represented a localized glacial period, the **Late** Desmoinesian to **Early** Virgilian
118 represented a widespread interglacial period with minor glaciation, and the **Late** Virgilian to
119 Early Wolfcampian represented the apex of widespread glaciation (Montañez & Poulsen, 2013).
120 Modeling predicts that sea-level oscillations in the Late Pennsylvanian were between 50-100
121 meters depending upon the number and volume of melting ice sheets, and that water
122 temperatures are estimated to have been between 4-7°C cooler during glacial maxima than inter-
123 glacial periods (Heckel, 1986; Isbell, 2003; Montañez, 2007; Tabor, 2007; Heckel, 2008; Cecil,
124 DiMichele, & Elrick, 2014). The sea-level and temperature changes were likely to have had an
125 important influence on species distribution and geographic range size during this time
126 (Waterhouse & Shi, 2010).

127 **Materials and methods**

128
129

130 **Taxa considered, stratigraphic correlation, specimens examined, and georeferencing:** 79

131 species belonging to 26 genera (13 nautiloids and **13 ammonoids**) of cephalopods in the
132 Pennsylvanian-Permian North American Midcontinent Sea were considered (Table S1). These
133 represent the most abundant, well preserved, and taxonomically well understood species. Range
134 reconstructions relied on the occurrence records of specimens from: the Division of Invertebrate
135 Paleontology, Biodiversity Institute, University of Kansas (KUMIP); the University of Iowa
136 Paleontology Repository (UI); and the Yale University Peabody Museum of Natural History
137 (YPM). These institutions house the most complete repository of cephalopod diversity from this

138 region and time as well as contain many of the type specimens of the species examined.

139 Moreover, all specimens used in the analysis were personally examined and taxonomically-

140 vetted, with species assignments and determinations made by the first author. Over 1,100

141 specimens were identified to species level in this study (Kolis, 2017).

142

143 Specimens were assigned to the Virgilian, Missourian, Desmoinesian, Atokan, Morrowan, or

144 Wolfcampian Stages using the USGS National Geologic Map Database (U.S. Geological Survey,

145 2017), Sawin et al. (2006, 2008, 2009), Zeller (1968), Pope (2012), and Heckel (2013). The

146 temporal boundaries of stages were derived from Davydov, Korn, and Schmitz (2012) (Table

147 S2).

148 All specimen localities were georeferenced during the course of the study. *GEOLocate* (Rios and

149 Bart, 2018) and the *MaNIS Georeferencing Calculator* (Wieczorek, 2015) were used to obtain

150 coordinates and uncertainty radii. All points were calculated in decimal degrees within the

151 WGS84 model in the *GEOLocate* (Rios & Bart, 2018) world topo layer to ensure consistency

152 and accuracy in determinations. Most uncertainty radii were less than 10 kms. Any specimens

153 with questionable locality information were excluded from analyses, as were specimens with an

154 uncertainty radius larger than the county they were contained within. This left 950 specimens

155 (Table S1) to use in range reconstruction and statistical analysis of geographic range through

156 geologic time. All statistical analyses were performed using Minitab® Statistical Software

157 *Minitab v. 17* (Minitab, 2016) and *R-Studio Version 3.4.0*.

158

159 **Range reconstruction using GIS:** Methods for range reconstruction follow Rode & Lieberman

160 (2004, 2005), Stigall & Lieberman (2006), Hendricks, Lieberman, & Stigall (2008), Myers &

161 Lieberman (2011), Myers, MacKenzie, and Lieberman (2013), and Dunhill & Wills (2015). In
162 particular, after specimen occurrence data were georeferenced and assigned to temporal bins,
163 *Excel* CSV files were compiled for the occurrence points for all specimens within species. CSV
164 files were imported into *ArcGIS v. 10.3* (ESRI, 2014) and layers were created using geographic
165 coordinate system ‘WGS 1984’ and projected coordinate system ‘WGS 1984 World Mercator’
166 (Fig. 1). These layers were input into *PaleoWeb* (The Rothwell Group LP, 2016) to rotate
167 coordinates into continental configuration and geographic position of the midcontinent region
168 during the Pennsylvanian-Early Permian (Fig. 2). These paleo-coordinate layers were then re-
169 projected into *ArcMap* (ESRI, 2014).

170

171 Geographic range values were calculated for each species (Table S3) using minimum bounding
172 geometry. This method has been shown to provide the most accurate method for reconstructing
173 changes in geographic range, especially for fossil taxa (Darroch & Saupe, 2018). Convex hulls or
174 buffers were given to every specimen occurrence point in each species and these shapefiles were
175 re-projected in ‘South America-Albers Equal Area Conic’. This model was used to accommodate
176 the rotation of species occurrence coordinates into the southern hemisphere during the Late
177 Paleozoic. Species with three or more occurrence points were given a convex hull that spanned
178 the entire area between occurrences. In this way, multiple occurrence points were combined to
179 recreate the geographic range of a single species. Species with only one occurrence point were
180 given a 10km² buffer; species with just two occurrence points were given a 10km² wide buffer
181 which was used, in conjunction with their distance, to derive an area value (following Rode &
182 Lieberman [2004, 2005], Hendricks, Lieberman, & Stigall [2008], Myers & Lieberman [2011],
183 and Myers, MacKenzie, and Lieberman [2013]).

184

185 **Assessing fossil record bias:** A common concern when studying the fossil record is that there
186 might be biases that could lead to inaccurate or artifactual findings. This concern can be
187 manifold, but the two most pertinent issues here involve incomplete sampling and/or issues of
188 stratigraphic bias. While it is important to be cognizant of the fact that the fossil record is
189 incomplete, it is worth recognizing that there is a large body of research that demonstrates many
190 of the biogeographic patterns preserved in the fossil record, particularly in marine settings,
191 represent real biological phenomena, rather than taphonomic artifacts (Myers & Lieberman,
192 2011; Rook, Heim, & Marcot, 2013; Dunhill & Wills, 2015). Further, it is also prudent to realize
193 that sampling bias is a common issue in studies of extant biodiversity and species distribution,
194 and much work needs to be done in this area to alleviate the biases of the extant biota
195 (Lieberman, 2002; Carrasco, 2003).

196

197 The possibility that biases in the fossil record might lead to artifactual results was assessed in a
198 few different ways. First, the relationship between outcrop availability and the geographic range
199 of Pennsylvanian and Permian cephalopods was determined (see Myers & Lieberman, 2011). A
200 percent coverage table of the range size of species overlaid against temporal outcrop availability
201 was created using *ArcGIS v. 10.3* (ESRI, 2014). A low percentage of overlap between range size
202 and outcrop area would suggest species distributions are more likely to reflect ‘real’
203 biogeographic patterns while a high percentage of overlap would suggest the presence or absence
204 of outcrop was significantly influencing results (Myers & Lieberman, 2011; Myers, MacKenzie,
205 & Lieberman, 2013; however, see also Dunhill, 2012 for an alternative viewpoint). The second
206 test used was an “n-1” jackknifing analysis. This procedure sub-sampled species range size
207 within each temporal bin to test the resilience of data to outliers. Mean range size estimations

208 were generated for each temporal bin; these were input into a one-way ANOVA to compare
 209 jackknife estimates with the initial geographic range size estimates (Myers & Lieberman, 2011;
 210 Myers, MacKenzie, & Lieberman, 2013). Finally, a Pearson rank correlation test was performed
 211 to test the association of occurrence points and geographic range size; a close correlation would
 212 indicate that reconstructed ranges were very much dependent on sampling and suggest that
 213 reconstructed biogeographic patterns might be an artifact of a biased fossil record (Myers,
 214 MacKenzie, & Lieberman, 2013).

215

216 **Speciation and extinction rate calculations:** Speciation and extinction rates were calculated in
 217 order to consider macroevolutionary dynamics in cephalopods from the Late Paleozoic
 218 Midcontinent Sea. Macroevolutionary rates were calculated using the following equation,
 219 presented in Foote (2000) and Rode & Lieberman (2005):

220

221

$$N_f = N_0 e^{rt}$$

222

223 where N_0 is the species richness at the beginning of a temporal bin, N_f is the species richness at
 224 the end of a temporal bin, t is the duration of a temporal bin, and r is the total rate of diversity
 225 change. The temporal bins used were North American stages. Species richness values (N_f) were
 226 determined for each temporal bin and were parsed into ‘carry-over’ (N_0) and ‘new’ species
 227 richness values to ensure the accuracy of speciation and extinction rate calculation. In this way,
 228 it was possible to calculate the rate of diversity change between bins. For example, $r_{\text{Atokan}} = (\ln$
 229 $N_{0\text{-Desmoinesian}} - \ln N_{0\text{-Atokan}}) / t_{\text{Atokan}}$. Speciation rate within each temporal bin was calculated
 230 using the equation $S_{\text{Atokan}} = (\ln N_{f\text{-Atokan}} - \ln N_{0\text{-Atokan}}) / t_{\text{Atokan}}$, and extinction rate within each

231 temporal bin was calculated using the equation $E_{Atokan} = S_{Atokan} - r_{Atokan}$ for each temporal stage
232 (Foote, 2000; Rode & Lieberman, 2005).

233

234 **Results**

235

236 **Paleobiogeographic patterns:** Geographic range data were analyzed separately across all
237 cephalopods and individually for both nautiloids and ammonoids. Species geographic range size
238 data were tested for normality within each temporal stage using the Anderson-Darling normality
239 test. Range size data within each temporal stage were not normally distributed for any data
240 combination ($P < 0.005$). Instead, distributions were left skewed across all temporal stages for
241 every data grouping. Data were subsequently log-transformed to normalize data, and statistical
242 analyses were performed on both original and transformed data.

243

244 In general, geographic range size (either mean of transformed data or median of original) of
245 ammonoids and nautiloids increases during the Missourian and Virgilian stages (Fig. 3), which
246 loosely correlates with a time of sea-level rise due to warming during an interglacial (Isbell,
247 2003; Montañez & Poulsen, 2013). However, none of the changes were statistically significant.
248 For instance, Mann-Whitney U Tests found no statistically significant changes (at $P \leq 0.05$) in
249 median geographic range size for any temporal stages separately across all cephalopods, as well
250 as individually for nautiloids and ammonoids, even prior to correction for multiple comparisons.

251

252 The same was true for two-sample t-tests performed on log-transformed data which again found
253 no statistically significant changes (at $P \leq 0.05$) in mean geographic range size though time, even

254 prior to correction for multiple comparisons. Furthermore, a one-way ANOVA, either with or
255 without the assumption of equal variance, failed to find any significant differences (at $P \leq 0.05$)
256 between stages for log-transformed mean geographic range size across all cephalopods as well as
257 individually for nautiloids and ammonoids.

258

259 **Analysis of macroevolutionary rates:** Speciation rate (S) and extinction rate (E) were
260 calculated for the Atokan, Desmoinesian, Missourian, and Virgilian stages across all
261 cephalopods and within nautiloids and ammonoids, respectively. The S and E presented across
262 all cephalopods are comprised of two calculations; one calculation included taxa that only
263 occurred in a single temporal stage (Table 1; Fig. 4), while the other calculation excluded taxa
264 that occurred in a single temporal stage (Table S4). S and E was also calculated for ammonoids
265 and for nautiloids including (Tables S5, S6) and excluding taxa that occurred in a single stage
266 (Tables S7, S8). Note, due to the dependence of calculations on diversity metrics from both
267 adjacent stages, it is not possible to accurately calculate the rate of biodiversity change (R), or S
268 and E for the first stage considered, the Morrowan, nor R or E for the last stage considered, the
269 Wolfcampian (these are thus left blank in Table 1 and Tables S4-S8). While it might have been
270 possible to infer S and E using other methods, to do so would exaggerate the significance of edge
271 effects and thus be problematic (Foote, 2000).

272

273 Across all cephalopods, S was high in the Atokan and Desmoinesian, fell in the Missourian, and
274 reached very low levels in the Virgilian and Wolfcampian (Fig. 4). By contrast, E was low in the
275 Atokan and Desmoinesian, began to rise in the Missourian, and reached even higher levels in the
276 Virgilian (Fig. 4). Essentially, across all cephalopods examined, when S is high, E is low, and

277 when S is low, E is high. This is potentially contrary to the pattern expected with an ecological
278 opportunity model of speciation (Simões et al., 2016), although the specific processes driving the
279 diversification could not be determined at this time. However, it is possible that when S was
280 high there may have been many short-lived species that could not be sampled that were actually
281 going extinct, and this phenomenon would artificially depress E.

282

283 As expected, S and E are lower when singletons are excluded (see Tables 1, S4). (See
284 Segesseman & Kammer [2018] for a recent discussion of how singletons can affect manifest
285 patterns in these types of studies.) Notably though, S and E patterns diverge somewhat between
286 ammonoids and nautiloids when considered individually. For instance, in nautiloids S is high in
287 the Atokan and Desmoinesian, then declines to moderate in the Missourian, and is at its lowest in
288 the Missourian and Wolfcampian (Table S6), whereas in ammonoids S is only high in the
289 Atokan, declines to moderate in the Desmoinesian, declines somewhat more in the Missourian
290 and then remains essentially constant through the Wolfcampian (Table S5). In addition, E is low
291 in ammonoids during the Desmoinesian and Missourian but high in the Atokan and
292 Wolfcampian (Table S5), whereas in nautiloids there are no observed extinctions during the
293 Atokan; values remain quite low in the Desmoinesian, rise somewhat in the Missourian, and then
294 rise again in the Virgilian (Table S6).

295

296 An important caveat regarding the calculation of S is that many of the species analyzed belong to
297 genera that were widely distributed beyond the Midcontinent Sea during the Late Paleozoic.
298 Thus, although none of the species considered in these analyses occurred outside of the
299 Midcontinent Sea, their close relatives did. It is conceivable that while speciation events and

300 rates by necessity are treated as occurring *in situ* herein, this might not always have been the
301 case. Instead, some speciation events could have occurred outside of the Midcontinent Sea with
302 subsequent invasion events into that region. These invasions would appear as *in situ* speciation
303 events in this analysis, although they actually were not. In the absence of phylogenetic
304 hypotheses for the genera considered it is not currently possible to consider how much of the
305 pattern pertaining to speciation rate shown in Fig. 4 is due to invasion instead of speciation.
306 Further, a related phenomenon could affect the calculation of E: at times what were treated as
307 extinction events might have simply been local extinctions in the Midcontinent Sea which could
308 have included emigration to other regions. As mentioned previously, it does not appear that any
309 of the species considered occur outside of the Midcontinent Sea, but a phylogenetic hypothesis
310 for these groups would be valuable for considering this issue in greater detail.

311

312 **Relationship between biogeography and macroevolutionary rates:** Mean geographic range
313 size increased during the Missourian and Virgilian and declined in the Wolfcampian (Fig. 3);
314 speciation rates were high in the Atokan and Desmoinesian and fell in the Virgilian (Fig. 4);
315 extinction rates were low in the Atokan and Desmoinesian and rose in the Virgilian (Fig. 4). The
316 Pearson correlation test in *Minitab 17* (Minitab, 2016) was used to examine the association
317 between geographic range and either speciation rate extinction rate in greater detail. No
318 significant (at $P \leq 0.05$) correlation between speciation or extinction rate and range size was
319 found across all cephalopods or within ammonoids or nautiloids individually (Table 2).
320 However, in cases the values approach $P = 0.05$: for instance, the association between
321 geographic range size and extinction for all cephalopods and for ammonoids alone. Notably, an
322 association between geographic range size and extinction has been documented by numerous

323 studies (e.g. Vrba, 1980; Jablonski, 1986; Eldredge, 1989; Stanley, 1990; Jablonski & Roy,
324 2003; Rode & Lieberman, 2004, 2005; Kiessling & Aberhan, 2007; Payne & Finnegan, 2007;
325 Stigall, 2010; Dunhill & Wills, 2015; Jablonski & Hunt, 2015; Orzechowski et al., 2015; Saupe
326 et al., 2015; Castiglione et al., 2017; Pie & Meyer, 2017; Lam, Stigall, & Matzke, 2018;
327 Schneider, 2018) and thus is a very robust phenomenon in general and likely to be operating to
328 some extent herein. However, over this time interval and for this particular group of species the
329 association is not statistically significant (Table 2).

330

331 **Analysis of fossil record bias:** The low percentage of overlap between cephalopod species
332 geographic ranges and available outcrop, less than 1% in 29 out of 30 species (Table S9; the one
333 species with a larger percentage value, *Orthoceras kansasense*, occurs throughout the
334 Midcontinent Sea), suggests the results are not simply an artifact of an incomplete fossil record,
335 at least pertaining to outcrop availability. The “n-1” jackknifing analysis also supports the
336 robustness of the reconstructed ranges, as no statistically significant differences were found
337 between the mean of the reconstructed and subsampled range values for any time interval (all P-
338 values > 0.9), suggesting that one or a few occurrence records are not having a major influence
339 on biogeographic patterns. Similar results were found in other taxa and time periods by Hunt,
340 Roy, & Jablonski (2005), Myers & Lieberman (2011), and Myers, MacKenzie, & Lieberman
341 (2013). Finally, the Pearson correlation test shows no correlation (-0.055, P-Value = 0.789)
342 between the number of occurrence points and geographic range size; this provides further
343 evidence that the biogeographic signatures of Late Paleozoic cephalopods are unlikely to be
344 simply an artifact of the fossil record.

345

346 **Diversity patterns:** Across all cephalopods, species richness increased from the Morrowan to
347 the Atokan, peaked in the Desmoinesian, and decreased through the Wolfcampian (Fig. S1). A
348 similar pattern is seen in the nautiloids (Fig. S2). However, the ammonoids (Fig. S3) demonstrate
349 an earlier peak in the Atokan, followed by a Desmoinesian to Virgilian plateau, with a decrease
350 in the Wolfcampian. Notably, previous studies of Late Paleozoic brachiopod communities in
351 Bolivia showed a consistent trend between diversity and glacial cycling with increased diversity
352 during glacial periods and decreased diversity during inter-glacial periods (Badyrka, Clapham, &
353 Lopez, 2013). However, there seems to be less consistency between species richness trends and
354 glacial cycling in the Midcontinent Sea. For instance, there is an increase in cephalopod species
355 richness throughout the Morrowan to Desmoinesian associated with localized glaciation, and an
356 interglacial period with generally minor glaciation is associated with a decrease in cephalopod
357 species richness from the Desmoinesian to Virgilian, yet by contrast widespread glaciation is
358 associated with a decrease in species richness from the Virgilian to the Wolfcampian.

359 **Discussion**

360
361

362 Geographic range shifts through time are one of the pervasive phenomena in the history of
363 life; these are manifest both within species and higher-level clades, occur at a number of
364 different time scales, and are frequently linked to climatic change (Wiley & Lieberman, 2011).
365 Specific examples do come from the Late Paleozoic, a time of extensive climate change
366 including profound glaciation along with numerous glacial and interglacial cycles (Montañez and
367 Poulsen, 2013). Those changes impacted patterns of geographic range in both terrestrial plant
368 (e.g., DiMichele et al., 2009; Falcon-Lang & DiMichele, 2010) and marine invertebrate
369 ecosystems (e.g., Leighton, 2005; Powell, 2007; Waterhouse & Shi, 2010). When it comes to

370 marine invertebrates from this time interval, most of the focus has been on the highly diverse
371 benthic faunas (e.g., Stanely & Powell, 2003; Powell, 2007; Bonelli & Patzkowsky, 2011;
372 Balseiro, 2016; Segessenman & Kammer, 2018); however, taxa that have a pelagic life style (as
373 adults) are also worth examining. Herein, 79 pelagic species of cephalopods were examined for
374 patterns of range size change using GIS and although in general these species exhibit some
375 evidence for changes in geographic range size (Fig. 3), those changes were not statistically
376 significant nor can they be directly tied to climate change. In a similar vein, many
377 paleontological studies have demonstrated that species with larger geographic ranges tend to
378 have lower extinction rates than species with narrower geographic range sizes (e.g., Vrba, 1980;
379 Jablonski, 1986; Eldredge, 1989; Stanley, 1990; Rode & Lieberman, 2004; Stigall & Lieberman,
380 2006; Payne & Finnegan, 2007; Stigall, 2010; Hopkins, 2011; Dunhill & Wills, 2015). Again,
381 this phenomenon was not found to be statistically significant in the case of the Late Paleozoic
382 cephalopod species considered herein (Table 2).

383

384 There may be a few different explanations for these findings. First, it may be that cephalopod
385 species were not significantly affected by the glacial-interglacial climatic cycles transpiring
386 within the Late Paleozoic Midcontinent Sea. A second possible explanation, perhaps coupled to
387 the first, is that since cephalopods are highly mobile, they can more easily occupy a greater
388 portion of their potential range relative to taxa that are benthic (at least as adults), such as
389 brachiopods. Further, perhaps the available potential range of cephalopod species does not
390 change much in glacial relative to interglacial regimes. This may seem unlikely given the vast
391 fluctuations in sea level occurring at the time, but pelagic marine organisms, because of their
392 ease of dispersal, may more easily maintain consistent geographic ranges relative to benthic

393 counterparts. Another possible explanation for the pattern retrieved is that, given the limits of
394 stratigraphic correlation, sample size, and the completeness of the fossil record, it was necessary
395 for the analyses of species distribution conducted herein to focus on the time scale of geological
396 stages, whereas in actuality there were climatic changes occurring within stages (Heckel, 2008,
397 2013); these probably did cause fluctuations in species' geographic ranges within stages, but
398 simply could not be observed in the present study. A final set of explanations are related to the
399 issue of sampling. For instance, it was more difficult for the analyses presented herein to detect
400 a relationship between geographic range size and macroevolutionary rate because speciation and
401 extinction rates could only be calculated for four stages. Further, a common concern when
402 studying the fossil record is that there might be biases that can lead to inaccurate findings. This
403 concern can be manifold, and although it is not entirely obviated by the results presented
404 regarding the apparent quality of the fossil record suggested by the various tests presented, it
405 does become harder to invoke as a specific, primary reason for results retrieved.

406

407 Another finding perhaps contrary to what might typically be expected for the Late Paleozoic is
408 that there was at least some evolutionary diversification and turnover within cephalopods, such
409 that species diversity did fluctuate throughout the Pennsylvanian and Early Permian.

410 Pennsylvanian rates of macroevolution are typically classified as 'sluggish' or 'stolid' across all
411 marine animals, and Sepkoski (1998) formalized the notion that there was a marked decline in
412 evolutionary rates of Carboniferous and Permian marine faunas. Stanley & Powell (2003)
413 reiterated this result and identified low mean macroevolutionary rates for marine invertebrate
414 taxa. Bonelli & Patzkowsky (2011) also documented a pattern of low turnover in the face of
415 major episodes of sea-level rise and fall due to climatic change. The results from the analyses

416 presented herein could indicate that macroevolutionary rate, at least in the case of Late Paleozoic
417 cephalopods, was more dynamic than often thought. One possible reason for this result is that
418 cephalopods are a fairly evolutionarily volatile group (Lieberman & Melott, 2013) relative to
419 many other marine invertebrate groups and have relatively high rates of diversification (Stanley,
420 1979; Jacobs et al., 1994; Landman, Tanabe, & Davis, 1996; Monnet, De Baets, & Klug, 2011;
421 Körn, Klug, & Walton, 2015; Körn et al., 2015); thus, they would generally be expected to have
422 higher rates of speciation and extinction than typical. However, this may not be the entire
423 explanation, as other groups also seem to show elevated rates of speciation and extinction during
424 this time interval. For instance, Balseiro (2016) did document evolutionary turnover at high
425 latitudes, and elevated evolutionary rates have also been found in fusulinid foraminifera (Groves
426 & Lee, 2008; Groves & Yue, 2009) and advanced cladid crinoids (Segessenman & Kammer,
427 2018). Ultimately, we support the contention raised by Segessenman & Kammer (2018) that
428 patterns from a few individual groups do not refute the general pattern of sluggish
429 macroevolution postulated for this time period in the history of life. The results may lend
430 credence to the notion that macroevolutionary patterns across all marine animals are rarely
431 unitary for any one time period in the history of life, and instead often tend to be variegated.
432

433 **Conclusions**

434

435 Patterns of range size change in late Paleozoic cephalopods from the North American
436 Midcontinent Sea were investigated using GIS. These species do exhibit some evidence for
437 changes in geographic range size through time, but the changes were not statistically significant
438 nor could they be directly tied to climate change. Further, in contradistinction to what is usually

439 found in the fossil record, cephalopod species with larger geographic ranges were not found to
440 have lower extinction rates than species with narrower geographic ranges. These distinctive
441 patterns may perhaps be related to the fact that cephalopods are pelagic and highly mobile.
442 Finally, the group shows more evolutionary diversification and turnover during the
443 Pennsylvanian and Early Permian than is typical of other marine invertebrate groups and this
444 could be related to the fact that cephalopods are an evolutionarily volatile group.

445

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447

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457

458 **References**

459

460

- 461 **Abe FR, Lieberman BS. 2009.** The nature of evolutionary radiations: a case study involving
462 Devonian trilobites. *Evolutionary Biology* **36**:225-234.
- 463 **Badyrka K, Clapham ME, Lopez S. 2013.** Paleoeology of brachiopod communities during the
464 late Paleozoic ice age in Bolivia (Copacabana Formation, Pennsylvanian-Early Permian).
465 *Palaeogeography, Palaeoclimatology, Palaeoecology* **387**:56-65.
- 466 **Balseiro D. 2016.** Compositional turnover and ecological changes related to the waxing and
467 waning of glaciers during the Late Paleozoic ice age in ice-proximal regions
468 (Pennsylvanian, western Argentina). *Paleobiology* **42**:335-357.
- 469 **Bonelli JR, Patzkowsky ME. 2011.** Taxonomic and ecologic persistence across the onset of the
470 Late Paleozoic Ice Age: evidence from the upper Mississippian (Chesterian series),
471 Illinois basin, United States. *Palaios* **26**:5-17.
- 472 **Brooks DR, McLennan DA. 1991.** *Phylogeny, Ecology, and Behavior: a Research Program in*
473 *Comparative Biology*. Chicago: University of Chicago Press.
- 474 **Carrasco MA. 2013.** The impact of taxonomic bias when comparing past and present species
475 diversity. *Palaeogeography, Palaeoclimatology, Palaeoecology* **372**:130–137.
- 476 **Castiglione S, Mondanaro A, Melchionna M, Serio C, Di Febbraro M, Carotenuto F, Raia**
477 **P. 2017.** Diversification rates and the evolution of species range size frequency
478 distribution. *Frontiers in Ecology and Evolution* **5**:147, 1-10.
- 479 **Cecil CB, DiMichele WA, Elrick SD. 2014.** Middle and Late Pennsylvanian cyclothems,
480 American Midcontinent: Ice-Age environmental changes and terrestrial biotic dynamics.
481 *Comptes Rendus Geoscience* **346**:159-168.
- 482 **Darroch SAF, Saupe EE. 2018.** Reconstructing geographic range-size dynamics from fossil

- 483 data. *Paleobiology* **44**:25-39.
- 484 **Davydov VI, Körn D, Schmitz MD. 2012.** Chapter 23 - The Carboniferous Period. In:
485 Gradstein FM, Ogg JG, Schmitz MD, Ogg GM, eds. *The Geologic Time Scale*.
486 Amsterdam: Elsevier, 603 - 651.
- 487 **DiMichele WA, Montañez IP, Poulsen CJ, Tabor NJ. 2009.** Climate and vegetational regime
488 shifts in the late Paleozoic ice age earth. *Geobiology* **7**:200-226.
- 489 **Dunhill AM. 2012.** Problems with using rock outcrop area as a paleontological sampling proxy:
490 rock outcrop and exposure area compared with coastal proximity, topography, land use,
491 and lithology. *Paleobiology* **38**:126-143.
- 492 **Dunhill AM, Wills MA. 2015.** Geographic range did not confer resilience to extinction in
493 terrestrial vertebrates at the end-Triassic crisis. *Nature Communications* **6**:1-8.
- 494 **Eldredge N. 1989.** *Macroevolutionary Dynamics: Species, Niches, and Adaptive Peaks*. New
495 York: McGraw-Hill.
- 496
497 **Eldredge N, Gould SJ. 1972.** Punctuated equilibria: an alternative to phyletic gradualism. In:
498 Schopf TJM, ed. *Models in Paleobiology*. San Francisco: Freeman, Cooper, 82-115.
- 499 **ESRI. 2014.** *ArcGIS Desktop: Release 10.3*. Redlands, CA: Environmental Systems Research
500 Institute.
- 501 **Falcon-Lang HJ, DiMichele WA. 2010.** What happened to the coal forests during
502 Pennsylvanian glacial phases? *Palaios* **25**:611-617.
- 503 **Fielding CR, Frank TD, Isbell JL. 2008.** The Late Paleozoic Ice Age: a review of current

- 504 understanding and synthesis of global climate patterns. *Geological Society of America*
505 *Special Paper* **441**:343-354.
- 506 **Foote M. 2000.** Origination and extinction components of taxonomic diversity: general
507 problems. *Paleobiology* **26**:74-102.
- 508 **Groves JR, Lee A. 2008.** Accelerated rates of foraminiferal origination and extinction during the
509 late Paleozoic ice age. *Journal of Foraminiferal Research* **38**:74-84.
- 510 **Groves JR, Yue W. 2009.** Foraminiferal diversification during the late Paleozoic ice age.
511 *Paleobiology* **35**:367-392.
- 512 **Heckel PH. 1986.** Sea-level curve for Pennsylvanian eustatic marine transgressive-regressive
513 depositional cycles along midcontinent outcrop belt, North America. *Geology* **14**:330-
514 334.
- 515 **Heckel PH. 2008.** Pennsylvanian cyclothems in midcontinent North America as far-field effects
516 of waxing and waning of Gondwana ice sheets. *Geological Society of America Special*
517 *Paper* **441**:275-289.
- 518 **Heckel PH. 2013.** Pennsylvanian cyclothems of northern midcontinent shelf and biostratigraphic
519 correlation of cyclothems. *Stratigraphy* **10**:3-40.
- 520 **Hendricks JR, Lieberman BS, Stigall AL. 2008.** Using GIS to study palaeobiogeographic and
521 macroevolutionary patterns in soft-bodied Cambrian arthropods. *Palaeogeography,*
522 *Palaeoclimatology, Palaeoecology* **264**:163-175.
- 523 **Hopkins MJ. 2011.** How species longevity, intraspecific morphological variation, and
524 geographic range size are related: a comparison using Late Cambrian trilobites.

525 *Evolution* **65**:3253-3273.

526 **Hunt G, Roy K, Jablonski D. 2005.** Species-level heritability reaffirmed: a comment on 'on the
527 heritability of geographic range sizes'. *American Naturalist* **166**:129-135.

528 **Isbell JL. 2003.** Timing of the late Paleozoic glaciation in Gondwana: was glaciation responsible
529 for the development of northern hemisphere cyclothem? *Geological Society of America*
530 *Special Paper* **370**:5-24.

531 **Jablonski D. 1986.** Larval ecology and macroevolution in marine invertebrates. *Bulletin of*
532 *Marine Science* **39**:565-587.

533 **Jablonski D, Hunt G. 2015.** Larval ecology, geographic range, and species survivorship in
534 Cretaceous mollusks: organismic versus species-level explanations. *American Naturalist*
535 **168**:556-564.

536 **Jablonski D, Roy K. 2003.** Geographical range and speciation in fossil and living
537 molluscs. *Proceedings of the Royal Society of London, Series B* **270**:401-406.

538 **Joachimski MM, Lambert LL. 2015.** Salinity contrast in the US Midcontinent Sea during
539 Pennsylvanian glacio-eustatic highstands: evidence from conodont apatite $\delta^{18}O$.
540 *Palaeogeography, Palaeoclimatology, Palaeoecology* **433**:71-80.

541 **Kiessling W, Aberhan M. 2007.** Geographical distribution and extinction risk: lessons from
542 Triassic-Jurassic marine benthic organisms. *Journal of Biogeography* **34**:1473-1489.

543 **Kolis K. 2017.** The biogeography and macroevolutionary trends of late Paleozoic cephalopods in
544 the North American Midcontinent Sea: understanding the response of pelagic organisms
545 to changing climate during the Late Paleozoic Ice Age. Master's Thesis, Department of
546 Ecology & Evolutionary Biology, University of Kansas.

- 547 **Körn D, Klug C, Walton SA. 2015.** Taxonomic diversity and morphological disparity of
548 Paleozoic ammonoids. In: Klug C, Körn D, De Baets K, Kruta I, Mapes RH, eds.
549 *Ammonoid Paleobiology: from Macroevolution to Paleogeography*. Berlin: Springer,
550 231-264.
- 551 **Klug C, Körn D, De Baets K, Kruta I, Mapes RH, eds. 2015.** *Ammonoid Paleobiology: from*
552 *Macroevolution to Paleogeography*. Berlin: Springer
- 553 **Kröger B. 2005.** Adaptive evolution in Paleozoic coiled cephalopods. *Paleobiology* **31**:253-268.
- 554 **Lam AR, Stigall AL, Matzke N. 2018.** Dispersal in the Ordovician: speciation patterns and
555 paleobiogeographic analyses of brachiopods and trilobites. *Palaeogeography,*
556 *Palaeoclimatology, Palaeoecology* **489**:147-165.
- 557 **Landman N, Tanabe K, Davis RA, eds. 1996.** Ammonoid Paleobiology. New York: Plenum.
- 558 **Leighton LR. 2005.** The latitudinal diversity gradient through deep time: testing the "Age of
559 Tropics" hypothesis using Carboniferous productidine brachiopods. *Evolutionary*
560 *Ecology* **19**:563-581.
- 561 **Liow LH. 2007.** Does versatility as measured by geographic range, bathymetric range and
562 morphological variability contribute to taxon longevity? *Global Ecology and*
563 *Biogeography* **16**:117-128.
- 564 **Lieberman BS. 2000.** *Paleobiogeography: Using Fossils to Study Global Change, Plate*
565 *Tectonics, and Evolution*. New York: Kluwer Academic/Plenum.
- 566 **Lieberman BS. 2002.** Biogeography with and without the fossil record. *Palaeogeography,*
567 *Palaeoclimatology, Palaeoecology* **178**:39-52.
- 568 **Lieberman BS, Kimmig J. 2018.** Museums, paleontology, and a biodiversity science based
569 approach. In: Rosenberg GD, Clary RM, eds. *Museums at the Forefront of the History of*

- 570 *Geology: History Made, History in the Making. Geological Society of America Special*
571 *Paper 535*. In press. [https://doi.org/10.1130/2018.2535\(22\)](https://doi.org/10.1130/2018.2535(22))
- 572 **Lieberman BS, Melott AL. 2013.** Declining volatility, a general property of disparate systems:
573 from fossils, to stocks, to the stars. *Palaeontology* **56**:1297-1304.
- 574 **Mayr E. 1942.** *Systematics and the Origin of Species*. Cambridge: Harvard University Press.
- 575 **Myers CE, Saupe EE. 2013.** A macroevolutionary expansion of the modern synthesis and the
576 importance of extrinsic abiotic factors. *Palaeontology* **56**:1179-1198.
- 577 **Mii HS, Grossman EL, Yancey TE. 1999.** Carboniferous isotope stratigraphies of North
578 America: implications for Carboniferous paleoceanography and Mississippian glaciation.
579 *Geological Society of America Bulletin* **111**:960-973.
- 580 **Miller AK, Youngquist WL. 1949.** American Permian nautiloids. *Geological Society of*
581 *America Memoir* **41**:1-217.
- 582 **Miller AK, Dunbar CO, Condra GE. 1933.** The nautiloid cephalopods of the Pennsylvanian
583 System in the Mid-continent region. *Nebraska Geological Survey Bulletin* **9**:1-240.
- 584 **Minitab 17 Statistical Software. 2016.** State College, PA: Minitab, Inc.
- 585 **Monnet C, De Baets K, Klug C. 2011.** Parallel evolution controlled by adaptation and
586 covariation in ammonoid cephalopods. *BMC Evolutionary Biology* **11**:115, 1-21.
- 587 **Montañez IP. 2007.** CO₂-forced climate and vegetational instability during Late Paleozoic
588 deglaciation. *Science* **315**:87-91.
- 589 **Montañez IP, Poulsen CJ. 2013.** The late Paleozoic Ice Age: an evolving paradigm. *Annual*
590 *Review of Earth Planet Science* **41**:629-656.

- 591 **Myers CE, Lieberman BS. 2011.** Sharks that pass in the night: using Geographical Information
592 Systems to investigate competition in the Cretaceous Interior Seaway. *Proceedings of the*
593 *Royal Society of London, Series B* **278**:681-689.
- 594 **Myers CE, MacKenzie RA, Lieberman BS. 2013.** Greenhouse biogeography: the relationship
595 of geographic range to invasion and extinction in the Western Interior Seaway.
596 *Paleobiology* **39**:135-148.
- 597 **Nelson WJ, Lucas SG. 2011.** Carboniferous geologic history of the Rocky Mountain region.
598 *New Mexico Museum of Natural History and Science Bulletin* **53**:115-142.
- 599 **Newell ND. 1936.** Some mid-Pennsylvanian invertebrates from Kansas and Oklahoma: III.
600 Cephalopoda. *Journal of Paleontology* **10**:481-489.
- 601 **Orzechowski EA, Lockwood R, Byrnes JEK, Anderson SC, Finnegan S, Finkel ZV, Harnik**
602 **PG, Lindberg DR, Liow LH, Lotze HK, et al. 2015.** Marine extinction risk shaped by
603 trait-environment interactions over 500 million years. *Global Change Biology* **21**:3595-
604 3607.
- 605 **Payne JL, Finnegan S. 2007.** The effect of geographic range on extinction risk during
606 background and mass extinction. *Proceedings of the National Academy of Sciences, USA*
607 **104**:10506-10511.
- 608 **Pie MR, Meyer ALS. 2017.** The evolution of range sizes in mammals and squamates:
609 heritability and differential evolutionary rates for low- and high-latitude limits.
610 *Evolutionary Biology* **44**:347-355.
- 611 **Plummer FB, Scott G. 1937.** Upper Paleozoic ammonites of Texas: the geology of Texas.
612 *University of Texas Bulletin* **3701, vol 3, part 1**:1-516.

- 613 **Pope JP. 2012.** Description of Pennsylvanian units, revision of stratigraphic nomenclature and
614 reclassification of the Morrowan, Atokan, Desmoinesian, Missourian, and Virgilian
615 stages in Iowa. *Iowa Department of Natural Resources Special Report Series* **5**:1-140.
- 616 **Powell MG. 2005.** Climatic basis for sluggish macroevolution during the late Paleozoic ice age.
617 *Geology* **33**:381-384.
- 618 **Powell MG. 2007.** Latitudinal diversity gradients for brachiopod genera during late Paleozoic
619 time: links between climate, biogeography and evolutionary rates. *Global Ecology and*
620 *Biogeography* **16**:519-528.
- 621 **Raymond A, Metz C. 2004.** Ice and its consequences: glaciation in the Late Ordovician, Late
622 Devonian, Pennsylvanian-Permian and Cenozoic compared. *Journal of Geology* **112**:655-
623 670.
- 624 **Rios NE, Bart HL Jr. 2018.** GEOlocate: a platform for georeferencing natural history
625 collections data. <https://www.geo-locate.org> .
- 626 **Roark A, Flake R, Grossman EL, Olszewski T, Lebold J, Thomas D, Marcantonio F, Miller**
627 **B, Raymond A, Yancey T. 2017.** Brachiopod geochemical records from across the
628 Carboniferous seas of North America: evidence for salinity gradients, stratification, and
629 circulation patterns. *Palaeogeography, Palaeoclimatology Palaeoecology* **485**:136-153.
- 630 **Rode AL, Lieberman BS. 2004.** Using GIS to unlock the interactions between biogeography,
631 environment, and evolution in Middle and Late Devonian brachiopods and bivalves.
632 *Palaeogeography, Palaeoclimatology, Palaeoecology* **211**:345-359.
- 633 **Rode AL, Lieberman BS. 2005.** Intergrating evolution and biogeography: a case study

- 634 involving Devonian crustaceans. *Journal of Paleontology* **79**:267-276.
- 635 **Rojas A, Patarroyo P, Mao L, Bengtson P, Kowalewski M. 2017.** Global biogeography of
636 Albian ammonoids: a network-based approach. *Geology* **45**:659-662.
- 637 **Rook DL, Heim NA, Marcot J. 2013.** Contrasting patterns and connections of rock and biotic
638 diversity in the marine and non-marine fossil records of North America.
639 *Palaeogeography, Palaeoclimatology, Palaeoecology* **372**:123-129.
- 640 **Rothwell Group LP. 2016.** *PaleoWeb: Free Plate Tectonics Software*. Lakewood, CO.
- 641 **Saupe EE, Qiao H, Hendricks JR, Portell RW, Hunter SJ, Soberón J, Lieberman BS. 2015.**
642 Niche breadth and geographic range size as determinants of species survival on
643 geological time scales. *Global Ecology and Biogeography* **24**:1159-1169.
- 644 **Sawin RS, West RR, Franseen EK, Watney WL, McCauley JR. 2006.** Carboniferous-
645 Permian boundary in Kansas, midcontinent, USA: current research in earth sciences.
646 *Kansas Geological Survey, Bulletin* **252, Part 2**:1-13.
- 647 **Sawin RS, Franseen EK, West RR, Ludvigson GA, Watney WL. 2008.** Clarification and
648 changes in Permian stratigraphic nomenclature in Kansas: current research in earth
649 sciences. *Kansas Geological Survey, Bulletin* **254, Part 2**:1-4.
- 650 **Sawin RS, Franseen EK, Watney WL, West RR, Ludvigson GA. 2009.** New stratigraphic
651 rank for the Carboniferous, Mississippian, and Pennsylvanian in Kansas: current research
652 in earth sciences. *Kansas Geological Survey, Bulletin* **256, Part 1**:1-4.
- 653 **Schneider CL. 2018.** Marine refugia past, present, and future: lessons from ancient geologic
654 crises for modern marine ecosystem conservation. In: Tyler CL, Schneider CL, eds..
655 *Marine Conservation Paleobiology*. Berlin: Springer, 163-208.

- 656 **Segessenman DC, Kammer TW. 2018.** Testing evolutionary rates during the late Paleozoic ice
657 age using the crinoid fossil record. *Lethaia* **51**:330-343.
- 658 **Sepkoski JJ Jr. 1998.** Rates of speciation in the fossil record. *Philosophical Transactions of the*
659 *Royal Society of London* **353**:315-326.
- 660 **Simões M, Breitzkreuz L, Alvarado M, Baca S, Cooper JC, Heins L, Herzog K, Lieberman**
661 **BS. 2016.** The evolving theory of evolutionary radiations. *Trends in Ecology and*
662 *Evolution (TREE)* **31**:27-34.
- 663 **Stanley SM. 1979.** *Macroevolution*. San Francisco: W. H. Freeman.
- 664 **Stanley SM. 1990.** The general correlation between rate of speciation and rate of extinction:
665 fortuitous causal linkages. In: Ross RM, Allmon WD, eds. *Causes of Evolution: a*
666 *Paleontological Perspective*. Chicago: University of Chicago Press, 103-127.
- 667 **Stanley SM, Powell MG. 2003.** Depressed rates of origination and extinction during the late
668 Paleozoic ice age: a new state for the global marine ecosystem. *Geology* **31**:877-880.
- 669 **Stigall AL. 2010.** Using GIS to assess the biogeographic impact of species invasions on native
670 brachiopods during the Richmondian Invasion in the type-Cincinnatian (Late Ordovician,
671 Cincinnati region). *Palaeontologia Electronica* **13(1)**, **5A**:1-19.
- 672 **Stigall AL, Lieberman BS. 2006.** Quantitative palaeobiogeography: GIS, phylogenetic
673 biogeographical analysis and conservation insights. *Journal of Biogeography* **33**:2051-
674 2060.
- 675 **Tabor NJ. 2007.** Permo-Pennsylvanian palaeotemperatures from Fe-Oxide and phyllosilicate
676 δO_{18} values. *Earth and Planetary Science Letters* **253**:159-171.

- 677 **Tabor NJ, Poulsen CJ. 2007.** Paleoclimate across the late Pennsylvanian-early Permian tropical
678 paleolatitudes: a review of climate indicators, their distribution, and relation to
679 palaeophysiographic climate factors. *Palaeogeography, Palaeoclimatology,*
680 *Palaeoecology* **268**:293-310.
- 681 **US Geological Survey. 2017.** U.S. Geologic Names Lexicon ("Geolex"). Retrieved from the
682 National Geologic Map Database: <https://ngmdb.usgs.gov/Geolex/search> .
- 683 **Vrba ES. 1980.** Evolution, species, and fossils: how does life evolve? *South African Journal of*
684 *Science* **76**:61-84.
- 685 **Waterhouse JB, Shi GR. 2010.** Late Palaeozoic global changes affecting high-latitude
686 environments and biotas: an introduction. *Palaeogeography, Palaeoclimatology,*
687 *Palaeoecology* **298**:1-16.
- 688 **Wells MR, Allison PA, Piggott MD, Gorman GJ, Hampson GJ, Pain CC, Fang F. 2007.**
689 Numerical modeling of tides in the late Pennsylvanian Midcontinent Seaway of North
690 America with implications for hydrography and sedimentation. *Journal of Sedimentary*
691 *Research* **77**:843-865.
- 692 **Wieczorek C, Wieczorek J. 2015.** Georeferencing calculator (version 20160920). Museum of
693 Vertebrate Zoology, University of California, Berkeley. <http://manisnet.org/gci2.html> .
- 694 **Wiley EO, Lieberman BS. 2011.** *Phylogenetics, 2nd edition*. New York: J. Wiley & Sons.
- 695 **Yacobucci MM. 2017.** Marine life in a greenhouse world: cephalopod biodiversity and
696 biogeography during the early Late Cretaceous. *Paleobiology* **43**:587-619.
- 697 **Zeller DE, ed. 1968.** The stratigraphic succession in Kansas. *Kansas Geological Survey, Bulletin*
698 **189**:1-81.

699

700 **Figure Captions**

701

702 **Figure 1: Distribution of Pennsylvanian cephalopods.**

703 Distribution of Pennsylvanian nautiloid and ammonoid data points (red, left) and Permian
704 nautiloid and ammonoid data points (blue, right) across the midcontinent region of North
705 America. Plotted using *ArcGIS v. 10.3* (ESRI, 2014) software at 1: 20,000,000.

706

707 **Figure 2: Occurrence points of *Metacoceras* sp. and *Mooreoceras* sp.**

708 For the Virgilian, shown on possible paleogeography of that stage, at 1:1,000,000,000 scale;
709 plotted using *PaleoWeb* (The Rothwell Group LP, 2016).

710

711 **Figure 3: Mean geographic range size in km² of cephalopods through time.**

712 Nautiloid species (left) and ammonoid species (right) range changes occur but are not
713 statistically significant when analyzed using non-parametric tests or when log transformed data
714 are analyzed using parametric tests.

715

716 **Figure 4: Speciation and extinction rates through time.**

717 Values given in per Myr and derived from Table 1.

Table 1 (on next page)

Speciation rates (S) per millions of years (Myr), extinction rates (E) per Myr, and rate of turnover (R) per Myr, for each stage across all cephalopods.

Species richness values, species carryover from the previous stage, new species originating in the stage, N_o , N_f , and duration (in Myr) also given.

1 **Table 1:**
 2 **Speciation rates (S) per millions of years (Myr), extinction rates (E) per Myr, and rate of**
 3 **turnover (R) per Myr, for each stage across all cephalopods, with species richness values,**
 4 **species carryover from the previous stage, new species originating in the stage, N_o , N_f , and**
 5 **duration (in Myr) also given.**

6
7

Stage	Species Richness	Species Carryover	New Species	N_o	N_f	Duration	R	S	E
Wolfcampian	13	7	6	7	13	14		0.0442	
Virgilian	38	32	6	32	38	5	-0.3040	0.0343	0.3383
Missourian	55	33	22	33	55	3	-0.0103	0.1703	0.1805
Desmoinesian	41	12	29	12	41	3	0.3372	0.4096	0.0724
Atokan	15	7	8	7	15	2	0.2694	0.3811	0.1116
Morrowan	8	0	8	0	8	6			

8

Table 2 (on next page)

Pearson correlation test for association between S and geographic range and E and geographic range across all cephalopods and for ammonoids and nautiloids individually.

Pearson's r and P-values given.

1 **Table 2:**

2 **Pearson correlation test for association between S and geographic range and E and**
3 **geographic range across all cephalopods and for ammonoids and nautiloids individually,**
4 **with Pearson's r and P-values given.**

5

Taxon – Speciation	Pearson's r	P-value	Taxon – Extinction	Pearson's r	P-value
All Cephalopods – S	-0.541	0.347	All Cephalopods – E	0.925	0.075
Nautiloids– S	-0.463	0.432	Nautiloids – E	0.913	0.087
Ammonoids – S	-0.519	0.370	Ammonoids – E	0.803	0.197

6

Figure 1(on next page)

Distribution of Pennsylvanian cephalopods.

Distribution of Pennsylvanian nautiloid and ammonoid data points (red, left) and Permian nautiloid and ammonoid data points (blue, right) across the midcontinent region of North America. Plotted using ArcGIS v. 10.3 (ESRI, 2014) software at 1: 20,000,000.

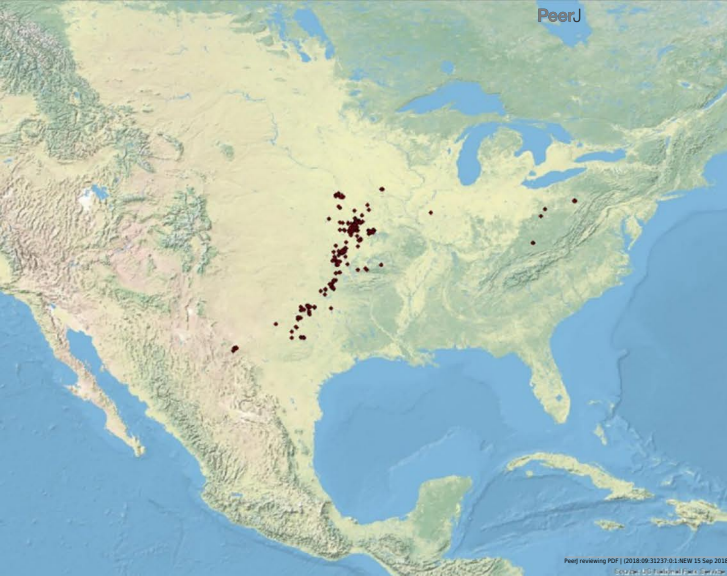


Figure 2 (on next page)

Occurrence points of *Metacoceras* sp. and *Mooreoceras* sp.

For the Virgilian, shown on possible paleogeography of that stage, at 1:1,000,000,000 scale; plotted using PaleoWeb (The Rothwell Group LP, 2016).

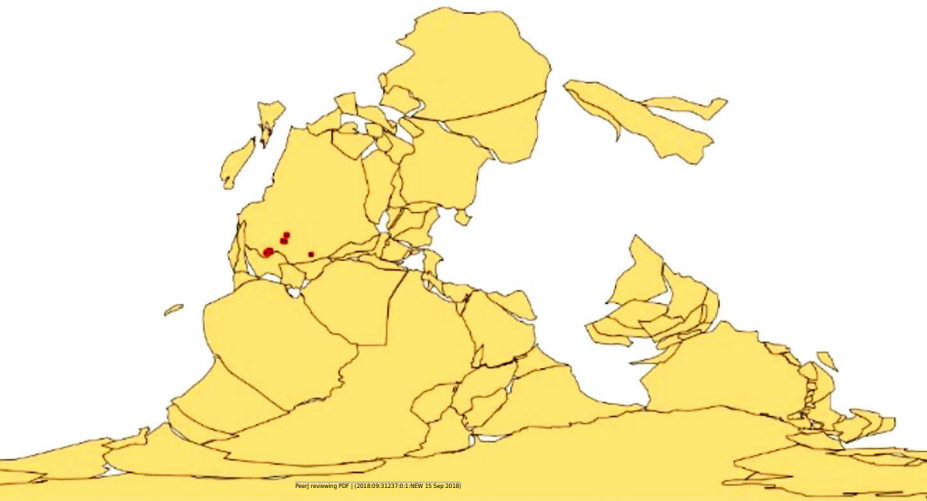


Figure 3(on next page)

Mean geographic range size in km² of cephalopods through time.

Nautiloid species (left) and ammonoid species (right) range changes occur but are not statistically significant when analyzed using non-parametric tests or when log transformed data are analyzed using parametric tests.

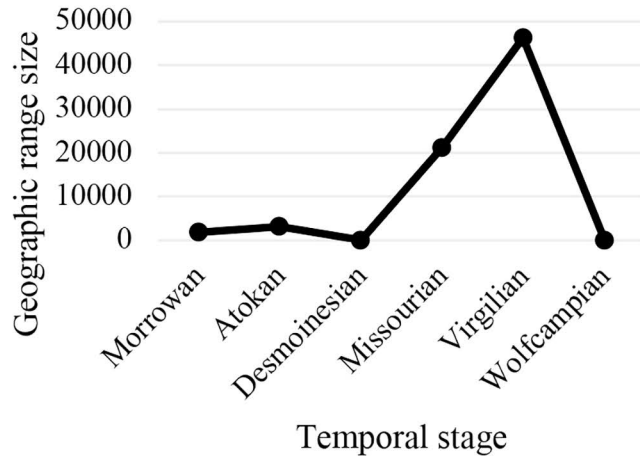
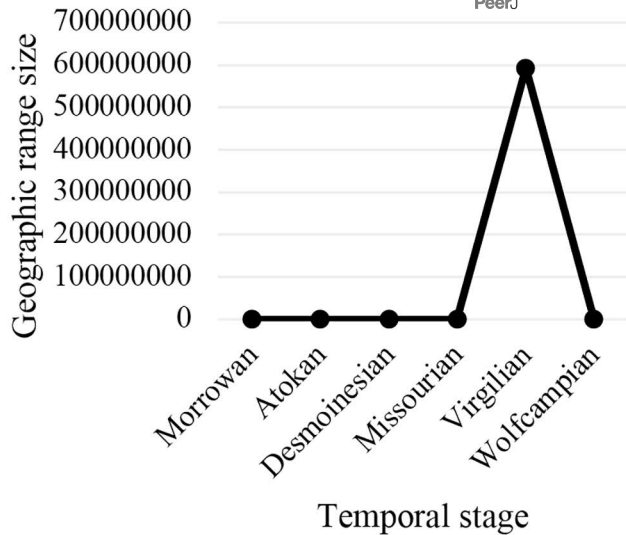


Figure 4(on next page)

Speciation and extinction rates through time.

Values given in per Myr and derived from Table 1.

