Properties of a cryptic lysyl oxidase from haloarchaeon *Haloterrigena turkmenica*

Nikolay B Pestov Corresp., [1] , **Daniel V Kalinovsky** [1] , **Tatyana D Larionova** [1] , **Alia Z Zakirova** [1] , **Nikolai N Modyanov** [2] , **Irina A Okkelman** [1] , **Tatyana V Korneenko** [1]

[1] Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry, Moscow, Russia

[2] Department of Physiology and Pharmacology, University of Toledo, Toledo, OH, United States of America

Corresponding Author: Nikolay B Pestov
Email address: korn@mail.ibch.ru

B ackground: Lysyl oxidases (LOX) have been extensively studied in mammals, whereas properties and functions of recently found homologues in prokaryotic genomes remain enigmatic. **Methods:** LOX open reading frame was cloned from *Haloterrigena turkmenica* in an *E. coli* expression vector. Recombinant *Haloterrigena turkmenica* lysyl oxidase *(*HTU-LOX) proteins were purified using metal affinity chromatography under denaturing conditions followed by refolding. Amine oxidase activity has been measured fluorometrically as hydrogen peroxide release coupled with the oxidation of 10-acetyl-3,7-dihydroxyphenoxazine in the presence of horseradish peroxidase. Rabbit polyclonal antibodies were obtained and used in western blotting. **Results:** Cultured *H. turkmenica* has no detectable amine oxidase activity. HTU-LOX may be expressed in *E. coli* with a high protein yield. The full-length protein gives no catalytic activity. For this reason, we hypothesized that the hydrophobic N-terminal region may interfere with proper folding and its removal may be beneficial. Indeed, truncated His-tagged HTU-LOX lacking the N-terminal hydrophobic signal peptide purified under denaturing conditions can be successfully refolded into an active enzyme, and a larger N-terminal truncation further increases the amine oxidase activity. Refolding is optimal in the presence of Cu^{2+} at pH 6.2 and is not sensitive to salt. HTU-LOX is sensitive to LOX inhibitor 3-aminopropionitrile. HTU-LOX deaminates usual substrates of mammalian LOX such as lysine-containing polypeptides and polymers. The major difference between HTU-LOX and mammalian LOX is a relaxed specificity of the former. HTU-LOX readily oxidizes various primary amines including such compounds as taurine and glycine, benzylamine being a poor substrate. Of note, HTU-LOX is also active towards several aminoglycoside antibiotics and polymyxin. Western blotting indicates that epitopes for the anti-HTU-LOX polyclonal antibodies coincide with a high molecular weight protein in *H. turkmenica* cells. **Conclusion:** *H. turkmenica* contains a lysyl oxidase gene that was heterologously expressed yielding an

active recombinant enzyme with important biochemical features conserved between all known LOXes, for example, the sensitivity to 3-aminopropionitrile. However, the native function in the host appears to be cryptic. **Significance:** This is the first report on some properties of a lysyl oxidase from Archaea and an interesting example of evolution of enzymatic properties after hypothetical horizontal transfers between distant taxa.

Properties of a cryptic lysyl oxidase from haloarchaeon *Haloterrigena turkmenica*

Nikolay B. Pestov[1,*], Daniel V. Kalinovsky[1], Tatyana D. Larionova[1], Alia Z. Zakirova[1], Nikolay N. Modyanov[2], Irina A. Okkelman[1], Tatyana V. Korneenko[1]

[1] Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry, Moscow, Russia

[2] Department of Physiology and Pharmacology, University of Toledo, Toledo, OH, United States of America

Short title: *Archaeal Lysyl Oxidase*

* To whom correspondence should be addressed: Nikolay B. Pestov, Shemyakin and Ovchinnikov Institute of Bioorganic Chemistry, Miklukho-Maklaya 16/10, Moscow, 117997, Russia, Tel: +7(495) 330-6556; Fax: +7(495) 330-6556; E-mail: korn@mail.ibch.ru

ABSTRACT

Background: Lysyl oxidases (LOX) have been extensively studied in mammals, whereas properties and functions of recently found homologues in prokaryotic genomes remain enigmatic. **Methods:** LOX open reading frame was cloned from *Haloterrigena turkmenica* in an *E. coli* expression vector. Recombinant *Haloterrigena turkmenica* lysyl oxidase *(*HTU-LOX) proteins were purified using metal affinity chromatography under denaturing conditions followed by refolding. Amine oxidase activity has been measured fluorometrically as hydrogen peroxide release coupled with the oxidation of 10-acetyl-3,7-dihydroxyphenoxazine in the presence of horseradish peroxidase. Rabbit polyclonal antibodies were obtained and used in western blotting. **Results:** Cultured *H. turkmenica* has no detectable amine oxidase activity. HTU-LOX may be expressed in *E. coli* with a high protein yield. The full-length protein gives no catalytic activity. For this reason, we hypothesized that the hydrophobic N-terminal region may interfere with proper folding and its removal may be beneficial. Indeed, truncated His-tagged HTU-LOX lacking the N-terminal hydrophobic signal peptide purified under denaturing conditions can be

successfully refolded into an active enzyme, and a larger N-terminal truncation further increases the amine oxidase activity. Refolding is optimal in the presence of Cu^{2+} at pH 6.2 and is not sensitive to salt. HTU-LOX is sensitive to LOX inhibitor 3-aminopropionitrile. HTU-LOX deaminates usual substrates of mammalian LOX such as lysine-containing polypeptides and polymers. The major difference between HTU-LOX and mammalian LOX is a relaxed specificity of the former. HTU-LOX readily oxidizes various primary amines including such compounds as taurine and glycine, benzylamine being a poor substrate. Of note, HTU-LOX is also active towards several aminoglycoside antibiotics and polymyxin. Western blotting indicates that epitopes for the anti-HTU-LOX polyclonal antibodies coincide with a high molecular weight protein in *H. turkmenica* cells. **Conclusion:** *H. turkmenica* contains a lysyl oxidase gene that was heterologously expressed yielding an active recombinant enzyme with important biochemical features conserved between all known LOXes, for example, the sensitivity to 3-aminopropionitrile. However, the native function in the host appears to be cryptic. **Significance:** This is the first report on some properties of a lysyl oxidase from Archaea and an interesting example of evolution of enzymatic properties after hypothetical horizontal transfers between distant taxa.

INTRODUCTION

Lysyl oxidase is an amine oxidase that is well characterized in mammals. The human genome contains five lysyl oxidase isoforms (LOX and LOXL1-4), all of them possess the highly conserved C-terminal catalytic domain, the N-terminal signal peptide, and the accessory segments in between. Catalytic domain of LOX is unique among other mammalian amine oxidases because of its ability to oxidatively deaminate various amines including ε-amino groups of lysine residues in peptides and proteins. LOX activity initiates cross-link formation between certain proteins, including elastin, collagen, and fibronectin, and this process is important for maturation and remodeling of the extracellular matrix (*Lucero and Kagan, 2006).* Most animal genomes sequenced to date contain from one to five LOX genes, only placozoans, nematodes and ctenophores seem to lack any LOX genes. (*Grau-Bove et al, 2015*). There are LOX genes in some fungal genomes, whereas plants are unknown to possess it. LOX genes are also absent from the vast majority of prokaryotic genomes. Therefore, the presence of true homologues of

animal LOX in just several species of Eubacteria and Archaea is of significant interest; it reflects the unique history of this enzyme, that most parsimoniously can be explained by multiple horizontal transfer events (HGT) (*Grau-Bove et al, 2015*). Among Eubacteria, LOX genes are frequent in Actinomycetes (especially Streptomyces), some Deltaproteobacteria, occasionally – in other eubacteria, and very rarely – among Archaea. This aspect is exciting not only from the phylogenetic point of view, but also because of potential biotechnological applications, i.e. the fact that distantly related enzymes may have useful properties *(Noda-García et al, 2013)*.
It is interesting to note that, in contrast to eukaryotic lysyl oxidases, several LOX homologues identified in prokaryotes exhibit a simple architecture even without a signal peptide (*Grau-Bove et al, 2015*). On the other hand, some prokaryotic LOXes are more complex. Specifically, LOX from *Sorangium cellulosum (*WP_012233967.1*)* possesses a unique Cys-rich C-terminal non-catalytic domain, which is presumably highly disulfide cross-linked.
The few lysyl oxidase homologues from Archaea that have been sequenced are clustered in two independent groups. This suggests that the two major phyla, Thaumarchaeotes and Euryarchaeotes, may had acquired LOX genes in two independent HGT events (*Grau-Bove et al, 2015*). Indeed, HGT is widespread in Archaea (*Papke et al, 2004, Papke et al, 2015)*.
Haloterrigena turkmenica was isolated from Turkmenistani sulfate saline soil by Zvyagintseva and Tarasov and described in 1987 as *Halococcus turkmenicus* (*Zvyagintseva and Tarasov, 1987*). In 1999 it was proposed to rename it to *Haloterrigena* (*Ventosa et al, 1999*). *H. turkmenica* belongs to the family Halobacteriaceae typus Euryarchaeota and is a fairly fast growing chemoorganotrophic extreme halophile that requires at least 2 M NaCl with optimal temperature around 45°C. The complete genome of this archaeon has been sequenced. It consists of 5,440 kbp (including plasmid 6), and it was annotated as encoding 5,287 proteins and 63 ncRNAs *(Saunders et al, 2010*).

Here, we attempted for the first time a study on the properties of lysyl oxidase from this haloarchaeon.

EXPERIMENTAL PROCEDURES

Materials and strains. A fresh stock of *Haloterrigena turkmenica VKMB-1734* was purchased from the All-Russian Collection of Microorganisms (G.K. Skryabin Institute of Biochemistry and Physiology of Microorganisms, Pushchino, Moscow Region, Russia). Capreomycin was from S.P. Incomed (Moscow, Russia), amikacin from OAO Sintez (Kurgan, Russia), substance P was custom synthesized at Syneuro (Moscow, Russia), hexylamine and 3-aminopropionitrile fumarate from Alfa Aesar (USA), all other amine substrates were from Sigma (USA).

Cultivation of H. turkmenica. Various haloarchaeal media with NaCl around 200 g/l such as INMI medium-3, DSMZ-372 are suitable. Care should be taken to adjust pH since *H. turkmenica* does not grow in acidic media. We found that a simpler medium (hereafter referred to as **IAO**) is a better choice: casamino acids, 5 g/l; yeast extract, 5 g/l; NaCl, 220 g/l; pH 7.6 – autoclaved and supplemented with $MgSO_4$, 5 mM; $CuCl_2$,10 μM. Solid IAO medium may be used for growing single colonies, however, only with high quality agar (some batches inhibit growth). Also, *H. turkmenica* can be easily adapted to a defined medium, hereafter referred to as **MHTU,** an enriched version of HMM (*Mosin and Ignatov, 2014*)): L-alanine, 0.4 g/l; L-arginine, 0.4 g/l; D-asparagine, 0.2 g/l; L-aspartic acid, 0.4 g/l; L-cysteine, 0.1 g/l; L-glutamic acid, 1.5 g/l; L-histidine, 0.7 g/l; L-isoleucine, 0.5 g/l; L-leucine, 0.8 g/l; D,L-lysine, 2 g/l; D,L-methionine, 0.4 g/l; L-phenylalanine, 0.3 g/l; L-proline, 0.4 g/l; D,L-serine, 0.6 g/l; L-threonine, 1 g/l; L-tyrosine, 0.2 g/l; D,.L-tryptophan, 0.5 g/l; L-valine, 1 g/l; AMP, 0.1 g/l; NaCl, 220 g/l; $MgSO_4 \cdot 7H_2O$, 20 g/l; KCl, 2 g/l; NH_4Cl, 0.5 g/l; KNO_3, 0.1 g/l; KH_2PO_4,0.1 g/l; K_2HPO_4, 0.1 g/l; Na_3·citrate, 0.8 g/l; $MnSO_4 \cdot 2H_2O$, 0.0003 g/l; $CaCl_2 \cdot 6H_2O$, 0.1 g/l; $ZnSO_4 \cdot 7H_2O$, 0.05 mg/l; $FeSO_4 \cdot 7H_2O$, 0.05 g/l; $CuCl_2$, 10 μM; glycerol, 1 g/l; D-leucine-OH, 0.1 g/l; norleucine, 0.1 g/l; thymine, 0.1 g/l; uracil, 0.1 g/l; pH 7.5.

Isolation of Halorubrum sp. VKK1262. Commercial salt from Upper-Kama deposit (Uralmedprom, Berezniki, Russia) was mixed with IAO medium without NaCl (200 g/l) and filtered through a 0.2 μm GSWP filter (Millipore, USA). Filters were incubated on IAO medium plates prepared with Noble agar (Difco, USA) at 37°C for one week and colored colonies were restreaked several times on fresh plates. 16S RNA sequence was analyzed using PCR with primers Arch16S-f2 and Arch16S-r934 and Sanger sequencing of the amplicons in both directions.

Gene cloning. The DNA used as a template for PCR was isolated from the cell culture using a ZR Fungal / Bacterial DNA MicroPrep kit (Zymo Research, USA) according to the manufacturer's

instructions. For PCR, in an equal volume of 25 µl, primers (sequences in Supplement) at a concentration of 0.8 µM, PCR buffer 5x Phusion GC reaction buffer, 2 µl 2.5 mM deoxyribonucleotide solution, 0.2 µl of Phusion DNA polymerase, and *H. turkmenica* genomic DNA as a template were used. The cycling parameters were as follows: 1. Hot start 98ºC for 2 min; 2. Denaturation at 98ºC, 30 s; 3. Annealing at 55ºC, 1 min; 4. Elongation at 72ºC, 2 min. 30 cycles between steps 4 and 2. 5. Final elongation at 72ºC for 7 min. Purified polynucleotide fragments HTU-AA and HTU-QV (AA and QV stand for corresponding dipeptides in the HTU-LOX sequence) were digested with *Bam*H I and *Hin*d III restriction enzymes and ligated into the corresponding sites of the pQE-30 vector (Qiagen, USA), followed by transformation of the *E. coli* strain XL1-Blue by electroporation. Colonies screening was performed by PCR, and the sequence was confirmed by Sanger sequencing.

Protein expression. The XL-1 Blue transformants HTU-AA and HTU-QV were grown in LB medium containing ampicillin on an orbital shaker at 37ºC until OD_{600} = 0.7 was reached, followed by induction of expression with 0.5 mM IPTG for 3 hours. The cells were then harvested by centrifugation and stored at -70ºC. His-tagged proteins were purified under denaturing conditions (8 M urea) on the metal-chelating sorbent Ni-NTA agarose (*Korneenko and Pestov, 1997*). Typical yields of the purified proteins HTU-QV and HTU-AA were around 25-27 mg per liter of culture. The resulting proteins in 8 M urea pH 6.3 buffered with 0.5 M imidazole, 0.1 M sodium phosphate, and 20 mM Tris were dialyzed against different buffers (optimization briefly described in Results).

Activity assays. Determination of substrate specificity was performed using a fluorometric method suitable for various amine oxidases as the release of hydrogen peroxide coupled to the oxidation of 10-acetyl-3,7-dihydroxyphenoxazine (Biotium, Germany), also known as Amplex red, in the presence of horseradish peroxidase (*Palamakumbura and Trackman, 2002*). The fluorescence of the reaction product (resorufin) was assayed with a Microplate analyzer "Fusion" (Perkin Elmer, USA) at excitation and emission of 535 and 620 nm, respectively. More specifically, the reaction was carried out in 0.1 M borate buffer pH 8.3 in the presence of 1 U / ml horseradish peroxidase at 37°C. For the negative control, 0.1 mM 3-aminopropionitrile was added to block any lysyl oxidase activity. Calibration has been done with known amount of hydrogen peroxide, and data were fitted to Michaelis-Menten equation using Prism software package (GraphPad, USA). Sheep LOX was

isolated from aorta as described before for the purpose of comparison with HTU-LOX (*Pestov et al, 2014*).

Immunization of rabbits was carried out with purified folded protein HTU-QV. Initially, rabbits were subcutaneously injected with 100 μg protein as an emulsion in Freund's complete adjuvant. The first booster injection was made with the same quantity of the antigen in incomplete Freund's adjuvant 5 weeks after the first immunization, and the second booster injection – with 250 μg antigen and no adjuvants 6 weeks later. One week after the second booster injection sera were collected and stored with the preservative sodium azide at 4ºC. Immunization of rabbits has been approved by Animal Care and Use Review Board of Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry, protocol No 15/2011.

Affinity purification of antibodies has been performed on a small-scale essentially as before (*Pestov et al, 2004*) using purified HTU-QV protein electrophoresed using SDS-PAGE and blotted on a PVDF membrane: the HTU-QV band on PVDF was blocked in TBST buffer containing 5% bovine serum albumin and 5% non-fat milk, then serum was added and incubated for 3 h. After several washes with TBST bound antibodies were eluted with 0.1 M sodium citrate (pH 2.0) for 10 min at followed by immediate neutralization with unbuffered tris and addition of 0.1% bovine serum albumin and 0.02% sodium azide.

Western blotting. H. turkmenica cells were centrifuged and the pellets were lyzed in 10 mM tris-HCl, 1 mM $MgCl_2$, pH 7.5 containing 0.5 mM tris(2-carboxyethyl)phosphine (Sigma, USA), Complete protease inhibitor cocktail (Roche, Switzerland) and 1 u/ml Benzonase (EMD Millipore, USA) for 15 min at 37°C followed by centrifugation for 10 min at 15000 g. The supernatants were mixed with Laemmli sample loading buffer without mercaptoethanol and analyzed by electrophoresis in 8% SDS PAGE gels. Following electrophoresis, the protein samples and colored protein weight markers (Spectra Multicolor Brad Range, Thermo, USA) were transferred from polyacrylamide gel onto a PVDF membrane (GE Healthcare, USA). The membrane was washed for 5 min with 2% SDS, then blocked in TBST buffer containing 5% non-fat milk, 0.02% sodium azide and 10% w/w Bløk blocker (EMD Millipore, USA), overnight at 4ºC. On the next stage, the membrane was incubated in 10 ml of TBST buffer solution with 0.1% non-fat dry milk and primary rabbit antibodies (1:10000) for one hour at room temperature, rinsed out with TBST buffer 10 times for 5 minutes each, followed by incubation with secondary antibodies (HRP-conjugated anti-rabbit

antibodies, Biotium, Germany, 1:50000) in 10 ml TBST buffer with 0.1% non-fat dry milk for one hour, and rinsed out again in the same way. Chemiluminescence was recorded using Femto Maximum Sensitivity Western Blotting Detection Reagent (Thermo, USA) and Carestream Kodak Biomax Light film (Sigma, USA).

RESULTS

We initially attempted to produce the full-length HTU-LOX protein in *E. coli* but found that it precipitates as inclusion bodies without any detectable amine oxidase activity, and all attempts at its refolding were unsuccessful (results not shown). For this reason, we proceeded to deletion mutants without the N-terminal peptide (hydrophobic segments are common sources of problematic expression in *E. coli*) with subsequent purification under denaturing conditions and refolding. The purity of the resulting eluate was checked by SDS PAGE. Fig. 1 illustrates the expression and purification of HTU-LOX exemplified by HTU-QV variant. Of note is the fact of its anomalously slow electrophoretic mobility that corresponds to an apparent molecular weight of 34 kDa, whereas the theoretical value of the His-tagged HTU-QV is 24.3 kDa. Since HTU-LOX is a rather acidic protein (theoretical pI 4.58 for the His-tagged HTU-QV), this peculiarity is common among acidic proteins (*García-Ortega et al, 2005*).

Refolding of the purified proteins HTU-AA and HTU-QV was achieved using dialysis against different buffers and results in good amine oxidase activity. We investigated a variety of factors that may improve the formation of catalytically active proteins HTU-AA and HTU-QV: buffer type and concentration (Tris, phosphate buffered saline, acetate, etc.), the ionic strength of the solution (concentration of NaCl), temperature, metal ions (Cu, Fe, Zn, Ni, Co, Mn) in different concentrations, pH of the solution (5.0 – 8.0), as well as the dialysis with a gradual decrease in the concentration of the denaturing agent (urea). Optimal pH is around 6.2 (Fig. 2A). Since it is known that mammalian LOX requires the presence of a copper ion in the catalytic domain in order to achieve the formation of the lysyl-tyrosine quinone (LTQ) in the catalytic center, we expected similar results of for HTU-LOX. Indeed, only Cu^{2+} increases activity (Fig. 2B,C), whereas a mixture of different ions gives an inhibition (Fig. 2C). It is interesting to note that refolding efficiency is only slightly affected by NaCl concentration, contrary to the expectations from the fact that *H. turkmenica* is an extreme halophile that requires at least 2 M NaCl (Fig. 2B). In the case of NaCl similar results were obtained in folding by dilution experiments,

210 demonstrating also that 1 M and 2 M NaCl cannot improve activity any further (results not
211 shown).

212 A slow decrease of the denaturant (urea) concentration was found to lack any advantages over
213 the stepwise approach with immediate transfer into a buffer without urea. This was confirmed by
214 refolding by dilution (results not shown). Ultimately, a simple refolding procedure may be
215 considered as optimal:

216 • Dialysis against 40 mM sodium acetate, pH 6.2 with 1 mM $CuSO_4$ at 4ºC for 3 hours;
217 • Dialysis against 40 mM sodium acetate, pH 6.2 without copper at 4ºC overnight.
218
219 Interestingly, under any conditions used, the amine oxidase activity of the protein HTU-QV AA
220 (typical activity for HTU-QV with 1 mM taurine at pH 8.3 was approximately 0.014 μmole/min
221 hydrogen peroxide per·mg protein) was about fifteen times higher than that of HTU-. Therefore,
222 the segment of HTU-LOX sequence from Ala^{39} to Gln^{92} may function as an inhibitory
223 (pro)peptide/

224 Refolded proteins HTU-QV and HTU-AA exhibit activity against a wide variety of primary
225 amines (Table 1): histamine, methylamine, lysine, cadaverine, tyramine, etc. Even glycine, β-
226 alanine are efficiently oxidized, in contrast with mammalian LOX. HTU-LOX readily oxidizes
227 some amine-containing antibiotics: polymyxin and aminoglycosides such as capreomycin and
228 amikacin. This is a unique property of lysyl oxidases, since other amine oxidases either do not
229 deaminate aminoglycosides or even are inhibited by them, as in the case of *E. coli* amine oxidase
230 (*Elovaara et al, 2015*). Regarding various proteins, lysine-containing peptides, and polymers
231 (e.g., poly-L-lysine, poly-allylamine, lysozyme, and substance P as an example of a Lys-
232 containing peptide), the HTU-LOX behaves almost like LOX from the aorta. Taurine is one of
233 the best substrates for HTU-LOX. It is also capable of oxidizing glycine, β-alanine, and γ-
234 aminobutyric acid. The only amine that HTU-LOX oxidizes much worse than mammalian LOX
235 is benzylamine. Importantly, the HTU-LOX demonstrated good sensitivity to the classical
236 inhibitor of all LOXes – 3-aminopropionitrile (BAPN). Also, HTU-LOX is somewhat different
237 from the mammalian enzyme in terms of pH dependence. In contrast to the latter, HTU-LOX
238 activity does not exhibit a steep decline from its maximum around 8.3, and even displays a
239 certain degree of bimodality retaining some activity even below 7 (Fig. 3).

We also attempted to study the HTU-LOX protein in the host – the archaeal halophile *H. turkmenica*. For this purpose, we raised polyclonal antibodies against the truncated HTU-LOX (variant QV). The full-size HTU-LOX theoretically contains 308 amino acids with a molecular weight of 33829 Da, whereas the full-length protein expressed in *E. coli* has electrophoretic mobility corresponding to 52 kDa (Suppl. Fig.). However, this apparently large discrepancy should be regarded as normal, since the anomalous mobility has been observed for purified recombinant HTU-LOX (Fig. 1). Western blotting (Fig. 4, lanes 1-2, compare to negative controls in lanes 4-6) showed the presence of HTU-LOX in *H. turkmenica* cells. It should be emphasized that specific bands were reliably detected only at a high sensitivity, meaning that the normal expression level of the protein in cultured *H. turkmenica* is quite low, and detection of the full-length, unprocessed HTU-LOX was obscured by non-specific bands (Fig. 4, lanes 4-6). Most interestingly, only a very high molecular weight band specific to anti-HTU-LOX antibodies of about 210 kDa has been reliably detected. Long incubation of the cells in saturated salt in the medium results in a marked decrease in the intensity of this band (Suppl. Fig.). Therefore, we can hypothesize that HTU-LOX in *H. turkmenica* predominantly exists in a moderately stable homo- or heterooligomeric form, which migration in SDS-PAGE corresponds to an apparent molecular weight of 210 kDa, however, this anomaly awaits further studies.

Also, we found that BAPN (even at a rather high concentration of 1 mM) had no significant effect on sensitivity of fresh cells to osmotic stress, on formation of hypotonically-resistant cysts, or on growth rate in both conventional (IAO) and defined (MHTU) media.

DISCUSSION

Amino acid sequence alignments (Fig. 5-6) of LOX proteins demonstrate poor overall conservation (for example, high variability in the number of disulfide bonds) with only a few hyperconserved amino acid residues like Cu-binding His and LTQ formation (*Zhang et al, 2018).* A fundamental aspect that needs to be emphasized is the fact that relatively little research has been carried out on the influence of HGT with the subsequent adaptation of the catalytic properties of the enzymes to a new host.

Refolding efficiency is not significantly affected by NaCl concentration. This surprising fact could reflect the history of prokaryote LOX genes: halophile archaea may have acquired these

genes from microorganisms with a rather different requirement for salt. The ancient HGT event may had even originated from a halophobic organism, followed by "domestication" that suppressed the formation of misfolded protein. Besides, LOX may had served as an antibiotic resistance enzyme under aerobic conditions. This, however, is unlikely in extant *H. turkmenica*, since Archaea are usually highly resistant to both polymyxin and common aminoglycosides. Also, HTU-LOX oxidizes some peptide antibiotics and theoretically this feature may be useful for competition with other species of haloarchaea (*Besse et al, 2015*) in the natural habitat of *H. turkmenica*. The low expression level of the enzyme suggests that HTU-LOX plays a modest functional role in increasing availability of nitrogen from non-typical amines. Its promiscuous substrate specificity and negligible enzymatic activity in *H. turkmenica* cells make it difficult to demonstrate this fact experimentally.

HTU-LOX accepts glycine, β-alanine, and γ-aminobutyric acid as substrates. This observation is unusual, because the presence of any acidic groups in vicinity of the amino group almost completely prevents oxidation by most amine oxidases. Thus, it is safe to conclude that the HTU-LOX has a relaxed substrate specificity in comparison with its mammalian homologue (*Shah et al, 1993*). Perhaps a low selection pressure on the lysyl oxidase gene allowed it to lose substrate specificity. This, however, may be useful for biotechnological purposes as a starting point for molecular evolution in any direction.

Another interesting fact is that the amine oxidase activity of the truncated protein HTU-QV is much higher than that of the longer one, HTU-AA. This observation is in line with the general view that LOX catalytic domain is usually (except for animal LOXL2-4 and homologs) preceded by an autoinhibitory sequence, together forming a propeptide. In the case of HTU-LOX, the autoinhibitory sequence corresponds to the stretch from Ala^{39} to Gln^{92}. However, the inhibition is relatively inefficient, and this may also reflect the evolution of HTU-LOX gene after the in-Archaea HGT that resulted in a partial degradation of the autoinhibitory function of the propeptide. LOX genes in Archaea underwent at least two independent HGTs (*Grau-Bove et al, 2015*) and this is just an example of HGT in Archaea (*Papke et al, 2004, Papke et al, 2015).* The widespread occurrence of these HGT events may also indicate that the transferred genes not necessarily possess indispensable functions in every species.

What is the origin of the animal lysyl oxidase? Has it emerged in primitive animals at the beginning of their evolution through HGT from Eubacteria? Or, conversely, LOX genes, which have important functions in animals, made their way several times into the world of prokaryotes? The second option seems highly unlikely due to splitting of animal ORFs into exons and molecular phylogeny (*Grau-Bové et al, 2015*) but cannot be excluded completely. In any respect, the most parsimonious explanation of the evolution of the catalytic LOX domain is that inter-kingdom saltations of LOX genes between distant branches of Life occurred more than once.

CONCLUSIONS

- *H. turkmenica* LOX (HTU-LOX) was successfully expressed in *E. coli*
- Optimal refolding conditions are different from those for the growth of the host cells
- Sensitivity to 3-aminopropionitrile is conserved in HTU-LOX
- HTU-LOX has a relaxed substrate specificity in comparison with mammalian LOX
- Benzylamine is a poor substrate for HTU-LOX
- N-terminal truncation of HTU-LOX increases activity
- Cultured *H. turkmenica* does not exhibit any detectable amine oxidase activity
- In *H. turkmenica,* HTU-LOX expression level is low, polyclonal anti-HTU-LOX antibodies detect a band protein with apparent molecular weight of 210 kDa
- Native function of *H. turkmenica* lysyl oxidase may be cryptic

ACKNOWLEDGMENTS

REFERENCES

Besse A, Peduzzi J, Rebuffat S, Carré-Mlouka A. 2015. Antimicrobial peptides and proteins in the face of extremes: Lessons from archaeocins. *Biochimie* **118: 344-355.**

Dmitriev RI, Okkelman IA, Abdulin RA, Shakhparonov MI, Pestov NB. 2009. Nuclear transport of protein TTC4 depends on the cell cycle. *Cell Tissue Res* **336: 521-527**.

García-Ortega L, De los Ríos V, Martínez-Ruiz A, Oñaderra M, Lacadena J, Martínez del Pozo A, Gavilanes JG. 2005. Anomalous electrophoretic behavior of a very acidic protein: ribonuclease U2. *Electrophoresis*. 26: 3407

Grau-Bové X, Ruiz-Trillo I, Rodriguez-Pascual F. 2015. Origin and evolution of lysyl oxidases. *Scientific Reports* **5:** 10568

Elovaara H, Huusko T, Maksimow M, Elima K, Yegutkin GG, Skurnik M, Dobrindt U, Siitonen A, McPherson MJ, Salmi M, Jalkanen S. 2015. Primary amine oxidase of *Escherichia coli* is a metabolic enzyme that can use a human leukocyte molecule as a substrate. *PLOS One* **10:** e0142367

Lucero HA, Kagan HM. 2006. Lysyl oxidase: an oxidative enzyme and effector of cell function. *Cellular and Molecular Life Sciences* 63: 2304-2316

Korneenko TV, Pestov NB, Egorov MB, Ivanova MV, Kostina MB, Shakhparonov MI. 1997. Monoclonal antibodies to the α-subunit of the putative human H^+,K^+-ATPase encoded by the atp1al1 gene. *Bioorganicheskaya khimiya* 23: 800-8004.

Mosin O, Ignatov, I. 2014. Photochrome Transmembrane Protein Bacteriorhodopsin from Purple Membranes of *Halobacterium halobium*. Applications in Bio- and Nanotechnologies. *Journal of Medicine, Physiology and Biophysics* **6:** 42-60

Noda-García L, Camacho-Zarco AR, Medina-Ruíz S, Gaytán P, Carrillo-Tripp M, Fülöp V, Barona-Gómez F. 2013. Evolution of substrate specificity in a recipient's enzyme following horizontal gene transfer. *Molecular Biology and Evolution* **30:** 2024-2034

Palamakumbura AH, Trackman PC. 2002. A fluorometric assay for detection of lysyl oxidase enzyme activity in biological samples. *Analytical Biochemistry* **300:** 245-251.

Papke RT, Corral P, Ram-Mohan N, Haba RR, Sánchez-Porro C, Makkay A, Ventosa A. 2015. Horizontal gene transfer, dispersal and haloarchaeal speciation. *Life (Basel)* 5: 1405-1426.

Papke RT, Koenig JE, Rodríguez-Valera F, Doolittle WF. 2004. Frequent recombination in a saltern population of Halorubrum. *Science* 306: 1928-1929.

Pestov NB, Korneenko TV, Radkov R, Zhao H, Shakhparonov MI, Modyanov NN. 2004. Identification of the β-subunit for nongastric H-K-ATPase in rat anterior prostate. *American Journal of Physiology. Cell Physiology*. 286: C1229-C1237.

Pestov NB, Okkelman IA, Shmanai VV, Hurski AL, Giaccia AJ, Shchepinov MS. 2011. Control of lysyl oxidase activity through site-specific deuteration of lysine. *Bioorganic & Medicinal Chemistry Letters* **21:** 255-258.

Saunders E, Tindall BJ, Fähnrich R, Lapidus A, Copeland A, Del Rio TG, Lucas S, Chen F, Tice H, Cheng JF, Han C, Detter JC, Bruce D, Goodwin L, Chain P, Pitluck S, Pati A,

Ivanova N, Mavromatis K, Chen A, Palaniappan K, Land M, Hauser L, Chang YJ, Jeffries CD, Brettin T, Rohde M, Göker M, Bristow J, Eisen JA, Markowitz V, Hugenholtz P, Klenk HP, Kyrpides NC. 2010. Complete genome sequence of *Haloterrigena turkmenica* type strain (4k). *Standards in Genomic Sciences* **2:** 107-116.

Shah MA, Scaman CH, Palcic MM, Kagan HM. 1993. Kinetics and stereospecificity of the lysyl oxidase reaction. *Journal of Biological Chemistry* 268: 11573-11579.

Ventosa A, Carmen Gutierrez M, Kamekura M, Dyall-Smith M L. 1999. Proposal to transfer *Halococcus turkmenicus*, *Halobacterium trapanicum* JCM 9743 and strain GSL-11 to *Haloterrigena turkmenica* gen. nov., comb. nov. *International Journal of Systematic Bacteriology* **49:** 131-136.

Zhang X, Wang Q, Wu J, Wang J, Shi Y, Liu M. 2018. Crystal structure of human lysyl oxidase-like 2 (hLOXL2) in a precursor state. Proceedings of the National Academy of Sciences of the United States of America. **115:** 3828-3833.

Zvyagintseva IS, Tarasov AL. 1987. Extreme halophilic bacteria from alkaline soils. *Mikrobiologiya* **56:** 839-844.

FIGURE LEGENDS

Table

Substrate efficiencies of recombinant *Haloterrigena* lysyl oxidase (HTU-QV variant) in comparison with mammalian enzymes from aorta. V_{max}/K_m ratios normalized with respect to tyramine. Values for polymers and lysozyme calculated per amine groups. Data for LOX from bovine aorta from (*Shah et al, 1993*) (note that conditions used were significantly different from this study). For HTU-LOX the short variant QV has been used. ND – no data. Z – rate too low for accurate determination. CK – complex kinetics with inhibition by substrate at high concentrations.

Figure 1. Electrophoretic analysis of expression and purification of recombinant N-terminally truncated *H. turkmenica* LOX (HTU-QV). 1 – molecular weight marker proteins; 2 – *E. coli* proteins before addition of IPTG; 3 – expression induced with IPTG; 4 – purified HTU-QV protein.

Fig. 2. Folding of recombinant *H. turkmenica* lysyl oxidase (HTU-QV). Folding by dialysis. A – Influence of pH; B – effect of NaCl; C – 1 mM salts of various metals added to the dialysis buffer; D – different concentrations of $CuSO_4$.

Fig. 3. pH dependence of recombinant *H. turkmenica* lysyl oxidase. Amine oxidase reaction rates for HTU-QV protein in comparison with LOX from sheep aorta were measured in universal borate-phosphate-acetate buffer with histamine as the substrate.

Figure 4. Immunoblotting with anti-HTU-LOX antibodies and proteins from *H. turkmenica* cells. Chemiluminescence of bound HRP-labeled antibodies; positions of molecular weight markers on the left and in the middle. 1-3 – affinity purified antibodies against HTU-QV, the catalytic domain of LOX from *H. turkmenica*. 4-6 – negative control with rabbit serum against an unrelated antigen (*Dmitriev et al, 2009*). 1,2,4,5 – proteins of *H. turkmenica* grown to log-phase (lanes 1 and 4 were loaded with three times more protein than lanes 2 and 5). 3,6 – proteins of *Halorubrum sp. VKK1262* (loading equal to lanes 2 and 5).

Fig. 5. Multiple alignment of archaeal lysyl oxidases. Obtained using Muscle algorithm. (HaloterrigenalimiCola – *Haloterrigena limicola*, HaloterrigenaturCk – *Haloterrigena turkmenica*, NatronoCoCCusjeotgali – *Natronococcus jeotgali,* NitrosopumilussalariaDB31 and Nitrosopimulus – *Nitrosopumilus sequnces.* Yellow – cysteine residues marked in yellow; red – LTQ-forming lysine and tyrosine; purple – three hyperconserved histidine residues necessary for the binding of Cu^{2+}.

Fig. 6. Multiple alignment of the conserved segments of catalytic domains from all lysyl oxidases representing different kingdoms. Obtained using Muscle algorithm from consensus sequences of different taxa. ARCHAE – Archaeal LOX sequences, DELTA – *Deltaproteobacteria*, FUNGI – fungal LOXes, ANIMA – various animal LOXes, LOW – *Mesomycetozoa* and *Orthonectida*, short – LOX from *Truepera radiovitrix, Deinococcus pimensis, Nitrospira nitrosa,* and a few samples from Parcubacteria, BETA – *Betaproteobacteria*, Actinshort – *Amycolatopsis mediterranei* LOX and closest homologues,

416 STREPTOM – LOX from *Streptomyces*, ACTINvar – other actinomycetal LOXes, BACIES – all
417 other eubacterial LOXes. Yellow – cysteine residues marked in yellow; red – LTQ-forming
418 lysine and tyrosine; purple – three hyperconserved histidine residues necessary for the binding of
419 copper.
420

Table 1 (on next page)

Substrate efficiencies of recombinant *Haloterrigena* lysyl oxidase (HTU-QV variant) in comparison with mammalian enzymes from aorta.

V_{max}/K_m ratios normalized with respect to tyramine. Values for polymers and lysozyme calculated per amine groups. Data for LOX from bovine aorta from (*Shah et al, 1993*) (note that conditions used were significantly different from this study). For HTU-LOX the short variant QV has been used. ND – no data. Z – rate too low for accurate determination. CK – complex kinetics with inhibition by substrate at high concentrations.

1

substrate	HTU-LOX	Sheep	Bovine
L-lysine	0.058	0.028	ND
cadaverine	0.370	1.070	1.09
histamine	0.550	0.920	ND
taurine	1.120	0.120	ND
glycine	0.020	z	ND
β-alanine	0.005	z	ND
GABA	0.015	z	ND
methylamine	0.020	ND	ND
substance P	0.068	ND	ND
lysozyme	CK	0.080	ND
polyallylamine	CK	0.080	ND
amikacin	0.260	ND	ND
capreomycin	0.120	0.190	ND
polymyxin	0.780	ND	ND
benzylamine	z	0.170	0.52
hexylamine	0.28	1.140	0.14

2

3

4 **Substrate efficiencies of recombinant *Haloterrigena* lysyl oxidase (HTU-QV variant) in**
5 **comparison with mammalian enzymes from aorta.** V_{max}/K_m ratios normalized with respect to
6 tyramine. Values for polymers and lysozyme calculated as for molar amine groups. Data for LOX
7 from bovine aorta from (*Shah et al, 1993*) (note that conditions used were significantly different
8 from this study). For HTU-LOX the short variant QV has been used. ND – no data. Z – rate too
9 low for accurate determination. CK – complex kinetics with inhibition by substrate at high
10 concentrations.

Figure 1

Electrophoretic analysis of recombinant N-terminally truncated *H. turkmenica* LOX (HTU-QV) expression and purification.

1 - molecular weight marker proteins; 2 - *E. coli* proteins before addition of IPTG; 3 – Expression induced with IPTG; 4 – purified HTU-QV protein.

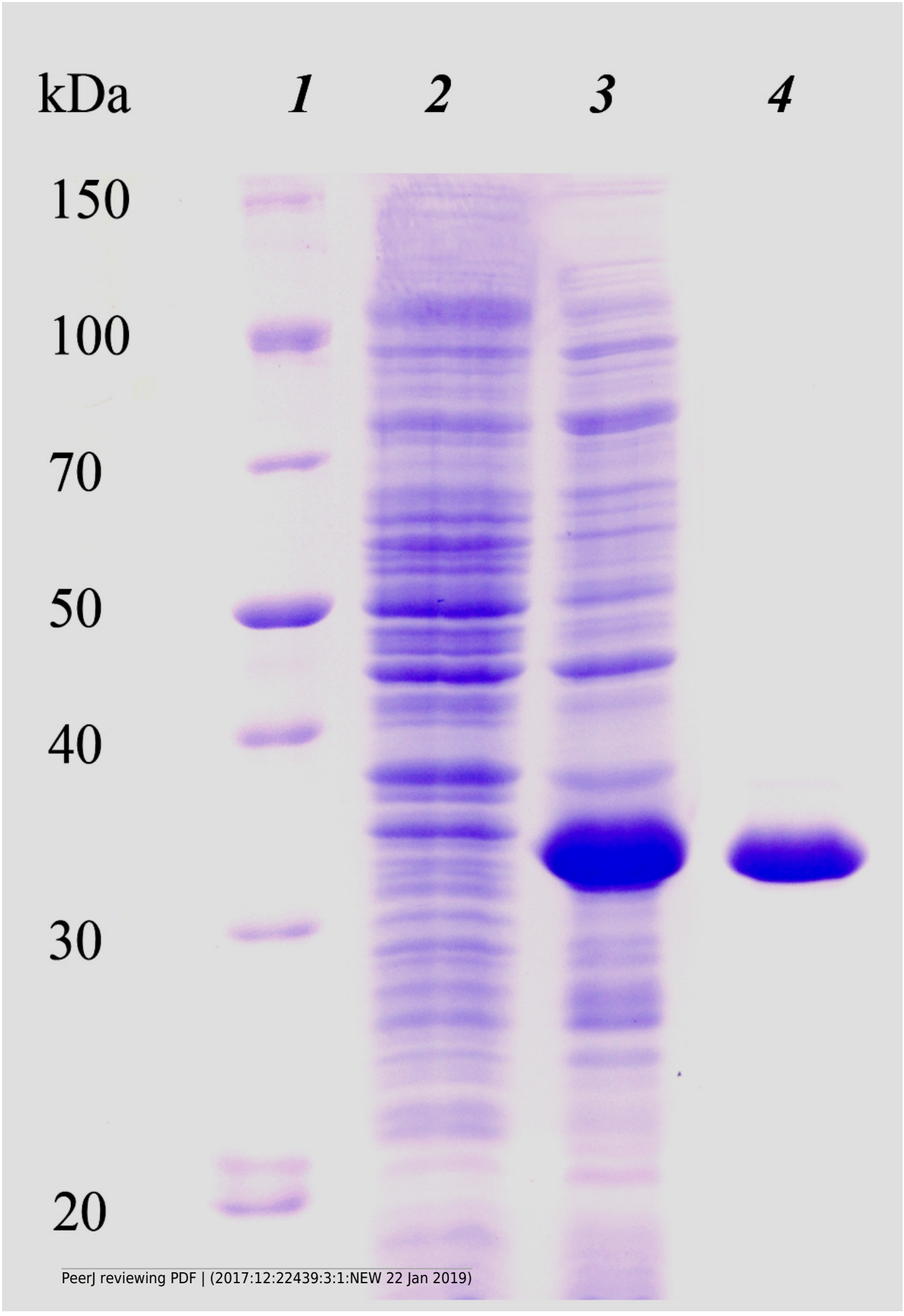

kDa
1
2
3
4
150
100
70
50
40
30
20

Figure 2

Folding of recombinant *H. turkmenica* lysyl oxidase (HTU-QV).

Folding by dialysis. A – Influence of pH; B – effect of NaCl; C – 1 mM salts of various metals added to the dialysis buffer; D – different concentrations of $CuSO_4$.

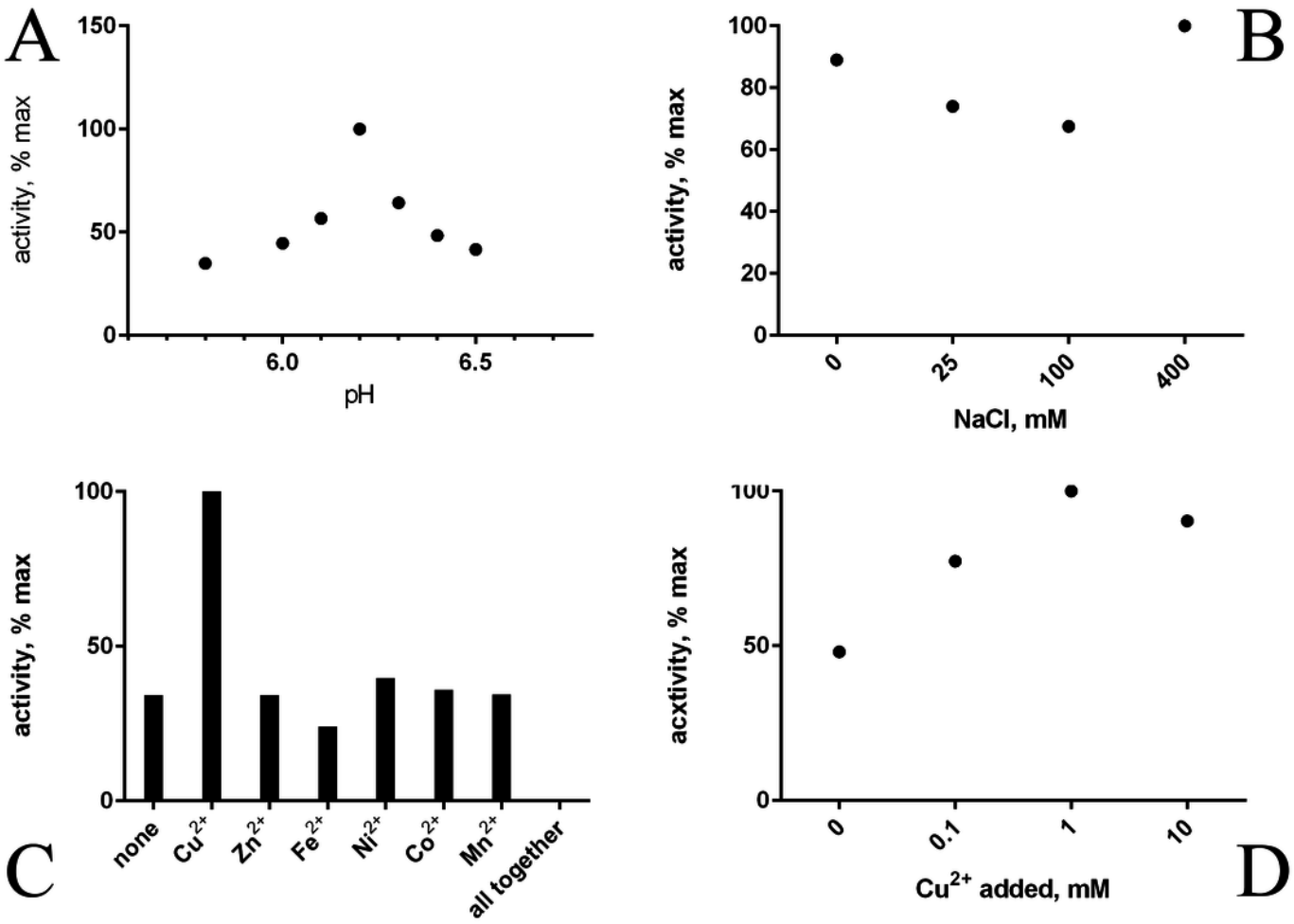


Figure 3

pH dependence of recombinant *H. turkmenica* lysyl oxidase.

In comparison with LOX from sheep aorta amine oxidase reaction rates were measured for HTU-QV protein if universal borate-phosphate-acetate buffer with histamine as the substrate.

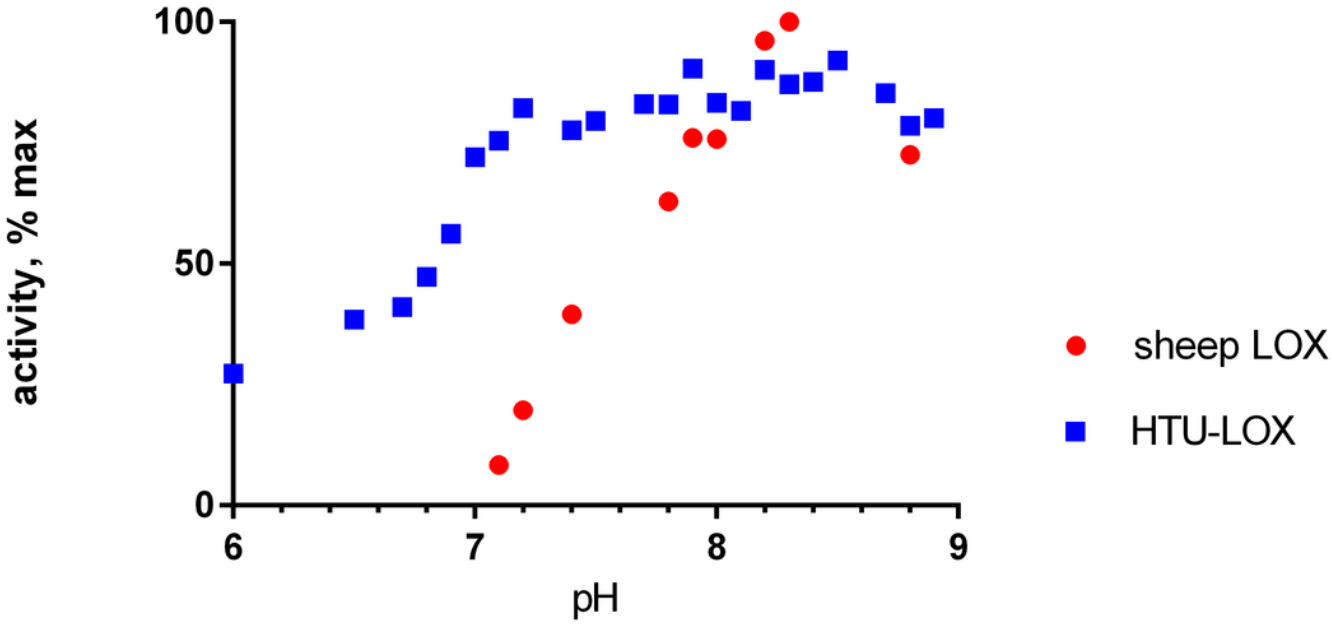


Figure 4

Figure 4. Immunoblotting detection of lysyl oxidase in *H. turkmenica* cells with anti-HTU-LOX antibodies.

Chemiluminescence of bound HRP-labeled antibodies; positions of molecular weight markers on the left and in the middle. 1-3 – affinity purified antibodies against HTU-QV, the catalytic domain of LOX from *H. turkmenica*. 4-6 – negative control with rabbit serum against an unrelated antigen (*Dmitriev et al, 2009*). 1,2,4,5 – proteins of *H. turkmenica* grown to log-phase (lanes 1 and 4 were loaded with three times more protein than lanes 2 and 5). 3,6 – proteins of *Halorubrum sp. VKK1262* (loading equal to lanes 2 and 5).

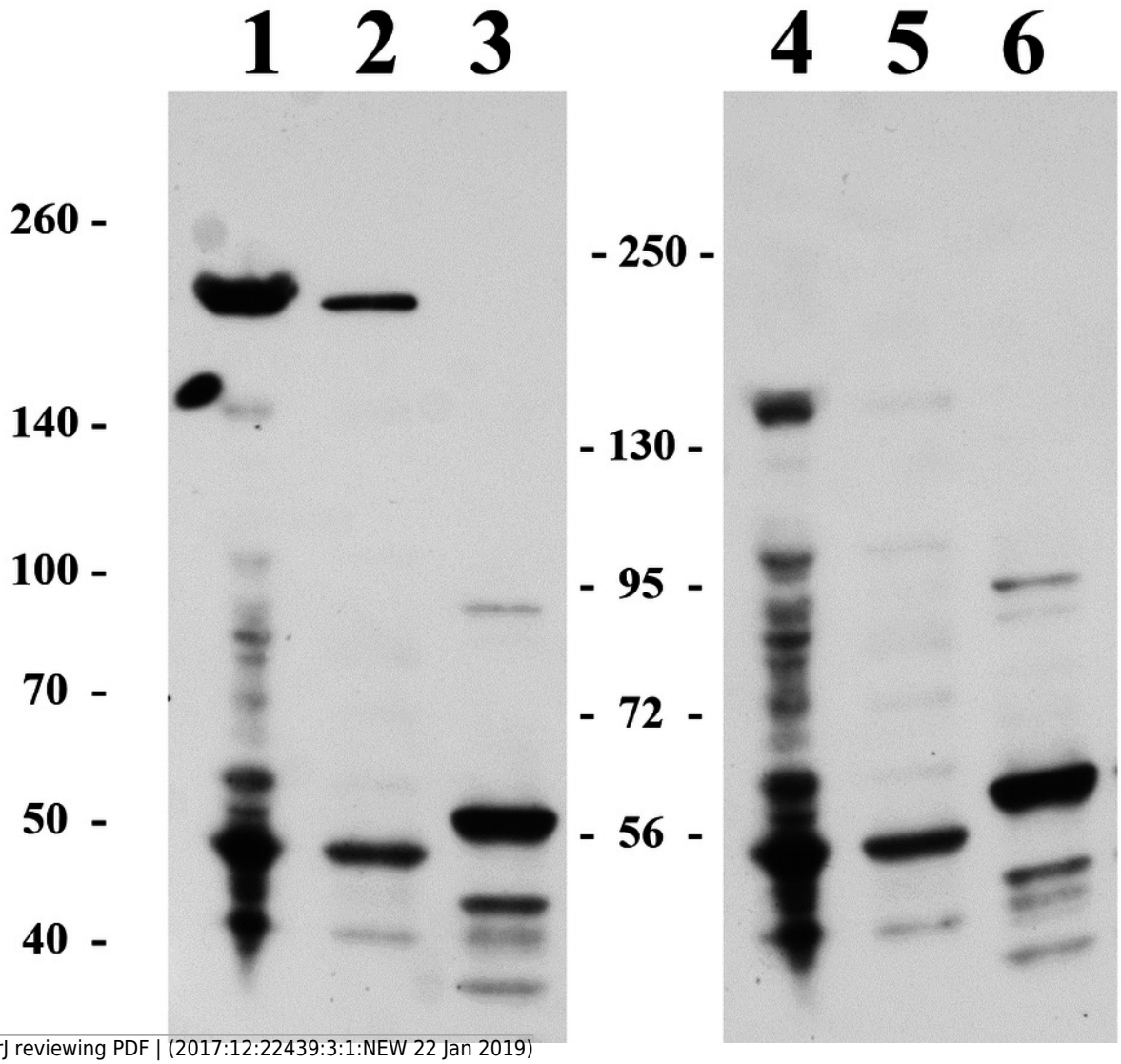


Figure 5

Multiple alignment of archaeal lysyl oxidases.

Obtained using Muscle algorithm. (HaloterrigenalimiCola – *Haloterrigena limicola*, HaloterrigenaturCk – *Haloterrigena turkmenica*, NatronoCoCCusjeotgali – *Natronococcus jeotgali,* NitrosopumilussalariaDB31 and Nitrosopimulus – *Nitrosopumilus sequnces.* Yellow – cysteine residues marked in yellow, red – LTQ-forming lysine and tyrosine, purple – three hyperconserved histidine residues necessary for the binding of Cu^{2+}.

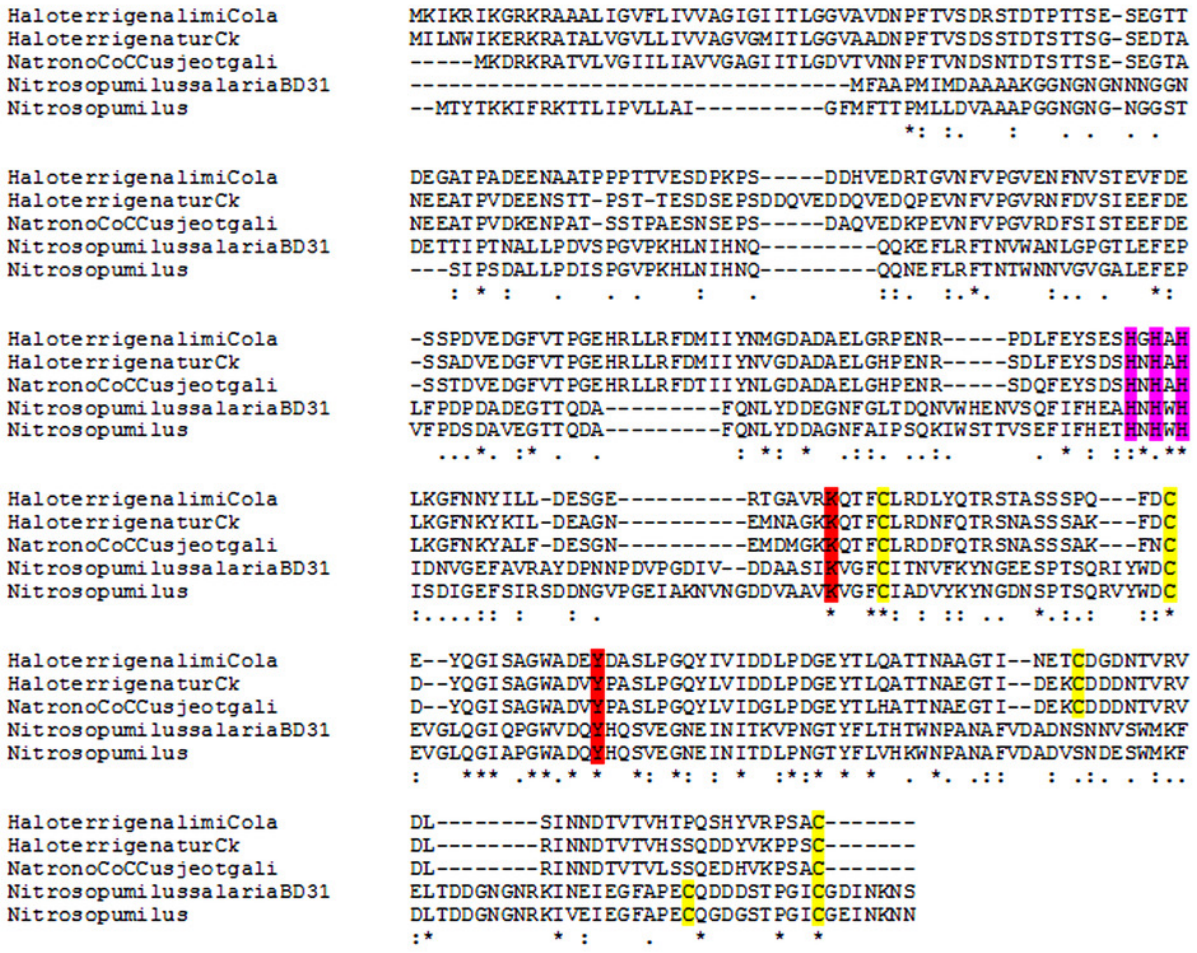
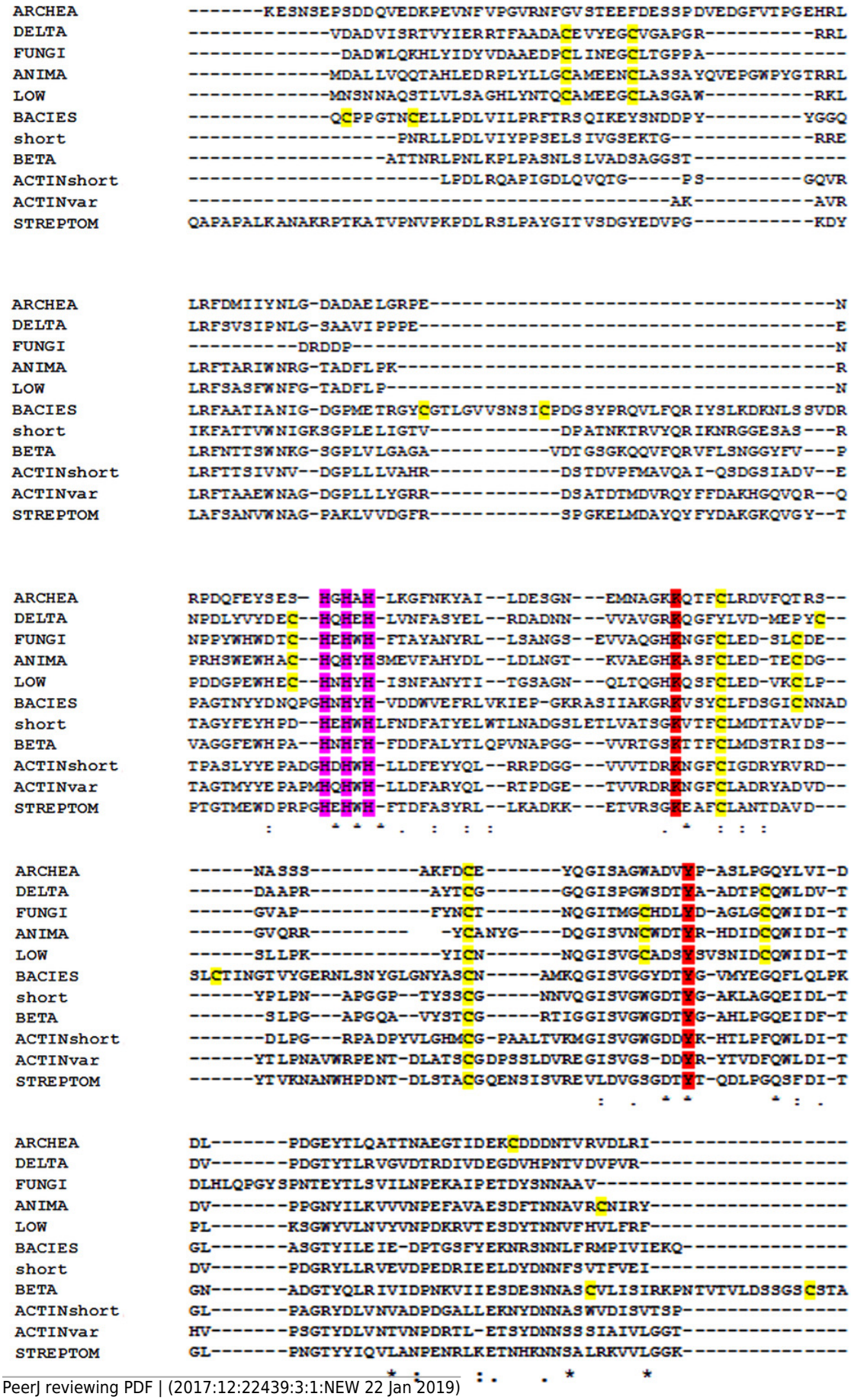


Figure 6

Multiple alignment of the conserved segments of catalytic domains from all lysyl oxidases representing different kingdoms.

Obtained using Muscle algorithm from consensus sequences of different taxa. ARCHAE – Archaeal LOX sequences, DELTA – *Deltaproteobacteria*, FUNGI – fungal LOXes, ANIMA – various animal LOXes, LOW – *Mesomycetozoa* and *Orthonectida*, short – LOX from *Truepera radiovitrix, Deinococcus pimensis, Nitrospira nitrosa,* and a few samples from Parcubacteria*,* BETA – *Betaproteobacteria*, Actinshort – *Amycolatopsis mediterranei* LOX and closest homologues, STREPTOM – LOX from *Streptomyces*, ACTINvar – other actinomycetal LOXes, BACIES – all other eubacterial LOXes. Yellow – cysteine residues marked in yellow; red – LTQ-forming lysine and tyrosine; purple – three hyperconserved histidine residues necessary for the binding of copper.


ARCHEA
DELTA
FUNGI
ANIMA
LOW
BACIES
short
BETA
ACTINshort
ACTINvar
STREPTOM