Recognizing two new *Hippolyte* species (Decapoda, Caridea, Hippolytidae) form from the South China Sea based on integrative taxonomy

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Abstract

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38 39 Shrimps of Hippolyte shrimps exhibit abundant biological diversity and display great ecologically significant significance in the ecosystem of seaweed bed ecosystems. Dozens of Hippolyte specimens were collected from Hainan Island and the Xisha Islands in the South China Sea. Detailed examination indicates that they one of these specimens represent some new Hippolyte species to the genus. Based on morphological, genetic, and ecological data, Hippolyte H. chacei sp. nov. and Hippolyte-H. nanhaiensis sp. nov. are described. Hippolyte-H. chacei sp. nov. was collected from the biotope of Sargassum sp. in Hainan Island, it and is distinguishes distinguished from the its congeners by its the unique mandible and particular dactyli of the third to fifth pereiopods, which corresponding corresponds to its basal position in the Indo-West pacific Pacific species clade of the phylogenetic tree. Hippolyte-H. nanhaiensis sp. nov. was collected from the biotopes of Galaxaura sp. and Halimeda sp. in the Xisha Islands, it-This new species differs from the its congeners by the combined based on a combination of features of the rostrum, scaphocerite, antennular peduncle, and the spines in on the dactyli of the third to fifth pereiopods, ; and additionally, it isforms sister group withto Hippolyte-H. australiensis in the phylogenetic tree. An identification key to mature female Hippolyte of the Indo-West Pacific and neighboring seas is provided.

Key words: Caridea, *Hippolyte*, Integrative taxonomy, Marine biodiversity, New species, South China Sea

Introduction

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Shrimps belonging toof the genus Hippolyte Leach, 1814, which display incrediblehigh diversity in morphology, coloration, and ecological habits, are mainly occur in tropical and temperate oceans, but can also occur in the polar region. For example, H. varians Leach, 1814 were recorded from the Arctic Circle in Norway (d'Udekem d'Acoz, 2007). Most Hippolyte species eommonly perching inhabit in the seaweed of the euphotic layer of tropical and subtropical oceans around the planet, and a few are obligatory or facultative symbionts of other organisms, such as gorgonians, and crinoids (d'Udekem d'Acoz, 2007; Marin et al., 2011). and display wondrously bio diverse in morphology, coloration, and ecological habit. In the last decade, the taxonomy, phylogeny, and biology of Hippolytegenus has have attracted considerable attention in the research of taxonomy, phylogeny and biology (Manjón-Cabeza et al., 2011; Marin et al., 2011; Terossi & Mantelatto, 2012; Liasko et al., 2015; Duarte & Flores, 2017; Duarte et al., 2017; Gan & Li, 2017a; 2017b; Liasko et al., 2017; Terossi et al., 2017). Currently, a total of 35 valid species are recognized worldwide (d'Udekem d'Acoz, 1996; 2007; De Grave & Fransen, 2011; Marin et al., 2011; Gan & Li, 2017a; 2017b; Terossi et al., 2017). Among these species, no more than half of themfewer than half (about approximately 12 species) are occurring in the Indo-West Pacific region. In the meantime Moreover, some unnamed species or and cryptic species are were also documented in the previous publications (Hayashi, 1986; d'Udekem d'Acoz, 1996; 2007; Terossi et al., 2017) waiting to be revised.

Because of its extensive morphological diversity and morphic overlap, as well as complex information described in previously published literatureas well as the confusions appeared in the literatures, the taxonomical research of *Hippolyte* is has always been considered to be difficult based on the morphological methodmorphology (d'Udekem d'Acoz, 1996; Gan & Li, 2017a). The situation is even more serious referring to complicated for the 'Hippolyte H. ventricosa H. Milne Edwards, 1837' species complex, which includes H. acuta (Stimpson, 1860), H. australiensis (Stimpson, 1860), H. ngi Gan & Li, 2017; H. singaporensis Gan & Li, 2017, H. ventricosa H. Milne Edwards, 1837, Hippolyte sp. A from Australia, Hippolyte sp. B from Hawaii, Hippolyte sp. C from the Malay Archipelago, and Hippolyte sp. D from Madagascar (d'Udekem d'Acoz, 1996; Gan & Li, 2017a; 2017b). More recently, Terossi et al. (2017) further recognized four cryptic or pseudocryptic species (H. ventricosa group-sp. 1 and sp. 2 from Indonesia, H. ventricosa group-sp. 3 from Fiji, and H. ventricosa group-sp. 4 from Taiwan) based on genetic analysis, but-however, their morphological features are greatly-very similar to those of H. ventricosa.

During recent biodiversity surveys of islands (2014_2018) of the South China Sea, dozens of *Hippolyte* specimens were collected from Hainan Island and the Xisha Islands by snorkeling. After detailed examination and multiple analysisanalyses, we described two new species of the

Comentário [AdS1]: Rephrase this sentence. E.g. inhabit the euphotic layer on seaweed beds?

Comentário [AdS2]: This sentence should be revised. It is not clear what H. ventricosa are you exactly mentioning in the end. 'H. ventricosa H. Milne Edwards, 1837' species complex based on integrative methods, namely, the validity of the new species is supported by the morphological, genetic and ecological data.

Materials & Methods

Sample collection and morphological examination. All the specimens were collected by a handheld net when snorkeling among the seaweed. After photographing, the specimens were preserved in 95% ethanol. Dissection and illustrations were made using a stereomicroscope (Nikon SMZ 1500, Japan) and a microscope (Nikon AZ100, Japan). Measurements and length ratios were calculated according to the method proposed by d'Udekem d'Acoz (1996). All the specimens are depositing deposited in the Marine Biological Museum of Chinese Academy of Sciences (MBM) in the Institute of Oceanology of Chinese Academy of Sciences, Qingdao, China (IOCAS)

Molecular data and analysis. Total genomic DNA was extracted from the pleopods of the specimens using the QIAamp DNA Mini Kit (QIAGEN, Germany) following the manufacturer's instructions. The extracted DNA was eluted in 100 μl of double-distilled H₂O (ddH₂O). Partial sequences of the 16S rRNA genes were amplified from the diluted DNA via the polymerase chain reaction (PCR). The reactions were carried out in a 50-μl volume containing the following reagents: 25 μl Premix Taq (TaKaRa TaqTM Version 2.0 plus dye, Japan), 1 μl forward and reverse primers (10 μM) respectively, 3 μl DNA template, and 20 μl ddH2O. The primers 16S-AR/1472, 5'-CGCCTGTTTATCAAAAACAT-3'/5'-AGATAGAAACCAACCTGG-3', were used (Crandall & Fitzpatrick, 1996). The PCR profile was as follows: 3 min at 94 °C for initial denaturation, followed by 35 cycles of denaturation at 94 °C for 30 s, annealing at 52 °C for 40 s and elongation at 72 °C for 50 s, with a final extension at 72 °C for 10 min. The PCR products were purified using the QIAquick Gel Extraction Kit (QIAGEN, Germany), and then bidirectionally sequenced using the same primers with an ABI 3730xl Analyzer (Applied Biosystems, USA). The obtained sequences were checked and proofread by ContigExpess 6.0 (a component of the Vector NTI Suite 6.0).

Besides ofIn addition to the sequences obtained by PCR (Table 1, Dataset S1), we—also downloaded some other sequences of Hippolyte sequences species from Genbank—with caution, including the previously reported cryptic or pseudocryptic species, namely H. ventricosa group-sp. 1 (KX588914), H. ventricosa group-sp. 2 (KX588915), H. ventricosa group-sp. 3 (KX588915), and H. ventricosa group-sp. 4 (KX588915) reported by Terossi et al. (2017) and H. ventricosa group-sp. 5 (KF023090) reported by De Grave et al. (2014). The molecular data, including 37 sequences of 16S rRNA genes, were aligned using MUSCLE 3.8 (Edgar, 2004). The highly divergent and poorly aligned sites were omitted from the alignment according to GBlocks 0.91b (Castresana, 2000). The best-fitting nucleotide base substitution model (GTR+I+G) for the alignment data was determined by ModelTest 3.7 (Posada & Crandall, 1998). Then this model was subsequently applied to phylogenetic analysis using the maximum likelihood (ML) method by PhyML 3.1 (Guindon & Gascuel, 2003) with 1000 bootstrap reiterations. The Bayesian inference (BI) tree was constructed using MrBayes 3.2 (Huelsenbeck

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& Ronquist, 2001), the Markov chains were run for 2000000 generations, with sampling every 2000 generations, after the first 25% trees were discarded as burn-in, the remaining trees were used to construct the 50% majority-rule consensus tree and to estimate posterior probabilities. The genetic distances were calculated under the Kimura 2-parameter (K2P) model in MEGA 7.0 (Kumar et al., 2016).

Ecological data. When the shrimp specimens were captured, their biotopes (mainly the algal colony where in which the shrimp living ined) were documented.

The following abbreviations are used: CL, carapace length, the length from the posterior orbital margin to the posterior dorsal border of the carapace; Coll., collector (s).

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Results

141 Taxonomy

- 142 Order Decapoda Latreille, 1802
- 143 Family Hippolytidae Spence Bate, 1888
- 144 Genus Hippolyte Leach, 1814
- 145 Hippolyte chacei sp. nov.
- 146 (Figs. 1–<u>54</u>, <u>10A5A</u>)

Material examined. *Holotype*: MBM285015, non-ovigerous female, 3.3 mm CL, Hongtang bay, Hainan Island, northern South China Sea, 1–3 m, Coll. Z B, Gan, 25 March 2018 (GenBank accession number of 16S rRNA gene: MK231007). *Paratypes*: MBM285016, 1 male, 2.3 mm CL, same collection data as holotype (GenBank accession number of 16S rRNA gene: MK231008); MBM285017, 2 non-ovigerous female, 2.7–3.0 mm CL, Houhai bay, Hainan Island, northern South China Sea, 2–3 m, Coll. Z B. Gan, 22 March 2018.

Description. Large sized shrimp of *Hippolyte*, oOutline robust (Fig. 1,2). Ratio lateral length/height of carapace 1.56–1.72. Rostrum long, slightly shorter than carapace, distinctly overreaching antennular peduncle, nearly reaching to the end of scaphocerite, without lateral carina, superior border slightly concave, unarmed in the female specimens (Fig. 3A2A,B) and armed with only one proximal tooth in the male specimen (Fig. 3C2C), inferior border slightly convex, armed with 4 teeth in the distal half length. Carapace smooth and glabrous, with robust

supraorbital-spine, antennal spine and hepatic spines (Fig. 3A2A,B). Base of supraorbital spine posterior to the posterior orbital margin. Tip of antennal spine slightly overreaching inferior orbital angle. Tip of hepatic spine falling short of anterior edge of carapace. Inferior orbital angle strongly produced, knob_like (Fig. 3B2B,D). Branchiostegal margin with a_distinct notch. Pterygostomian region rounded, strongly produced (Fig. 1,3B2B).

Abdominal segments smooth (Fig. 1). Third abdominal segment geniculately curved. Ratio dorsal length/height of the sixth abdominal segment 1.95–2.10. Telson (Fig. 3E2E) longer than the sixth abdominal segment; posterior margin rounded, armed with eight strong spines, outer spines smallest, medial two longest, without intermediate spinules or seta; Dorsal dorsal surface armed with two pairs of spines situated on distal 0.31–0.35 and 0.59–0.63 telson length.

Eye (Fig. 3A2A) well developed; tip of cornea nearly reaching to the end of first segment of antennular peduncle when extended forward; unpigmented part of eyestalk longer than broad; cornea semispherical, distinctly shorter than unpigmented part of eyestalk.

Antennular peduncle (Fig. 3F2F) slightly overreaching middle_length of scaphocerite. First segment of antennular peduncle with one well developed distolateral tooth; inner ventral tooth (Fig. 3G2G) on-0.47-050 of first segment (excluding distolateral tooth). Stylocerite large, reaching 0.56-0.62 (distolateral tooth included), or 0.69-0.75 (distolateral tooth excluded) of first segment. Second segment of antennular peduncle 0.81-0.86 times as long as broad in dorsal view, approximately 0.86-0.98 times as long as third segment in dorsal view. Outer antennular flagellum shorter than inner flagellum—one and proximal 6-8 segments thicker than orthersothersdistal ones. Scaphocerite (Fig. 2H2H) 3.06-3.18 times as long as wide, distolateral spine of scaphocerite far from reaching distal margin of blade, distolateral spine and blade separated by a distinct notch.

Mandible (Fig. 4A3A,B) without palp, incisor process unique in the genus *Hippolyte*, with 15–17 acute teeth. Maxillula (Fig. 4C3C) with broad curved palp, distal margin of upper lacinia armed with 14–18 spines and scattered simple long setae. Maxilla (Fig. 4D3D) with short palp; scaphognathite broad and long, lateral border nearly straight; inner lacinia bilobed, distal margin furnished with row of spines and long plumose setae; proximal endite round, with long setae on distal margin. Epipod of first maxilliped (Fig. 4E3E) slightly bilobed, endopod broad with distal long setae, exopod—with well-developed, caridean lobe broad. Second maxilliped (Fig. 4F3F) with well-developed exopod, flagelliform; endopod normal, dactylar segment oval, terminal margin furnished with simple and spinous setae; propodal segment with anteromedial margin round, bearing simple setae; carpus broader than long, and shorter than merus; ischium and basis fused. Third maxilliped (Fig. 4G3G) reaching to 0.32–0.39 of the scaphocerite when extended forward—, Exopod—exopod relatively short, only reaching to the mid-length of antepenultimate segment—of endopod; Ultimate—ultimate segment (excluding apical spine) of endopod 1.23–1.32 times as long as penultimate segment, distal half armed with 7–9 strong spines; antepenultimate segment nearly equal length to the last two segments combined.

First pereiopod (Fig. <u>5A4A</u>) shortest among pereiopods, robust and oblique, reaching to the end of basicerite when extended forward. Ventral margin of ischium, merus and carpus furnished

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with long simple setae. Terminal margin of carpus cotyloid. Cutting edges of chela non-denticulate, with tiny setula and long simple <u>distal</u> setae; tip of fingers armed with 3 acute spines respectively (Fig. <u>5B4B</u>).

Second pereiopod (Fig. 5C4C) slightly overreaching the end of third maxilliped when extended forward. Carpus with three subsegments, first subsegment 1.70–1.85 times as long as second subsegment, third subsegment slightly longer than or subequal to first subsegment; first subsegment 2.45–2.56 times as long as wide, second subsegment 1.08–1.12 times as long as wide, third subsegment 2.06–2.12 times as long as wide. Cutting edges of chela not denticulate, with tiny setula and long simple setae; tip of fixed finger and dactylus armed with 3 acute spines respectively (Fig. 5D4D).

Third to fifth pereiopods long and robust. Third pereiopod (Fig. <u>5E4E</u>) of female specimen reaching to the distolateral spine of scaphocerite when extended forward...; Daetylus dactylus of third pereiopod with 13–16 spines, the last 2–3 subdorsal spines distinctly shorter than the other terminal spines (Fig. 5F4F)...); Propodus propodus 5.56-5.62 times as long as wide, armed with 6-7 pairs of spines on ventral margin.; Carpus carpus 2.66-2.73 times as long as wide, armed with one proximal lateral spine...; Merus merus 5.58-5.62 times as long as wide, armed with 3 lateral spines. Ratio length of third pereiopod dactylus with longest apical spine/length of propodus 0.45-0.49; ratio length of third pereiopod dactylus with longest apical spine/length of carpus 0.79-0.83; ratio length of dactylus without spines/breadth of dactylus without spines 2.61–2.69; ratio length of dactylus with longest spines/breadth of dactylus without spines 2.95– 3.10; ratio length of longest spine of dactylus/breadth of dactylus without spines 0.62–0.71; ratio length of longest spine of dactylus/length of dactylus without spines 0.22-0.28. Third pereiopod (Fig. 5G4G,H) of male specimen with propodus and dactylus forming a prehensile apparatus. Fourth and fifth pereiopods (Fig. 5141,J,K,L) similar in shape to third pereiopod of female specimen, but slightly decreasing in size. Merus of fourth pereiopod armed with 2 lateral spine; merus of fifth pereiopod without lateral spine.

First pleopod (Fig. <u>5M4M</u>) of female specimen normal, endopod about 0.54–0.62 times as long as exopod. First pleopod (Fig. <u>5N4N</u>) of male specimen with endopod about 0.41–0.46 times as long as exopod. Second pleopod (Fig. <u>5O4O</u>) of male specimen with endopod about 0.81–0.89 times as long as exopod, appendix masculina with 9 apical setae, about 0.39–0.43 times as long as appendix interna (Fig. <u>5P4P</u>).

Coloration. Generally light brown over body (Fig. 25A), with few tawny stripes on carapace, and with faintpindling tawny spots on abdomen.

Biotope. All specimens were captured among gulfweed (*Sargassum* sp.) under the at depths of 1–3 m. Plenty of *Hippolyte* cf. ventricosa were co-captured at the same time simultaneously, the ratio of prisal nearly reached to 1:8.

Distribution. Hongtang bay and Houhai bay, Hainan Island, northern South China Sea; presumably Presumably, also distribute in Malayan Archipelago and Madagascar (see discussion).

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Etymology. The new species is named after Dr. Fenner A. Chace, Jr. in recognition of his great contribution to the crustacean taxonomy.

Hippolyte nanhaiensis sp. nov.

(Figs. 6–9, 10B<u>5B</u>–E)

Material examined. *Holotype:* MBM285018, ovigerous female, 1.6 mm CL, Ganquan Island, Xisha Islands, the South China Sea, 1–3 m, Coll. Z B, Gan, 15 May 2015 (GenBank accession number of 16S rRNA gene: MK231005). *Paratypes:* MBM285019, 1 male, 1.1 mm CL, same collection data as holotype (GenBank accession number of 16S rRNA gene: MK231006); MBM189210, 4 ovigerous female, 1.3–1.6 mm CL, 2 female, 1.2–1.3 mm CL, 2 male, 0.9–1.1 mm CL, 1 juvenile 0.6 mm CL, same collection data as holotype; MBM189211, 19 ovigerous female, 1.3–1.9 mm CL, 6 female, 1.0–1.4 mm CL, 5 male, 0.8–1.1 mm CL, 5 juvenile 0.6-0.8 mm CL, Bei Island, Xisha Islands, the South China Sea, 1–3 m, Coll. Z B. Gan, 13 May 2015.

Description. Middle sized shrimp of *Hippolyte*, eQutline stoutsoft (Fig. 6, 10B E). Ratio lateral length/height of carapace 1.49–1.58. Rostrum distinctly shorter than carapace, reaching to or slightly overreaching the end of antennular peduncle, without lateral carina; superior border straight, armed with 1–2 tooth in proximal position (Fig. 7A–D); inferior border armed with 1 subdistal toothdistal teeth in female specimens (Fig. 7C), and unarmed or only with 1 tiny distal notch in male specimens (Fig. 7D). Carapace smooth and glabrous, with supraorbital—spine, antennal spine—and hepatic spines (Fig. 7B,C). Base of supraorbital spine posterior to posterior orbital margin. Antennal spine small, slightly overreaching inferior orbital angle. Hepatic spine reaching to or slightly overreaching anterior edge of carapace. Inferior orbital angle produced, knob_like (Fig. 7B,C). Branchiostegal margin sinuous. Pterygostomian region rounded, strongly produced (Fig. 7C).

Abdominal segments smooth (Fig. 6), without or with few long plumose setae on tergum. Third abdominal segment geniculately curved. Ratio dorsal length/height of the sixth abdominal segment 1.91–2.08. Telson (Fig. 7E) longer than sixth abdominal segment;—, posterior margin rounded, armed with eight strong spines, outer spines smallest, medial two longest, without or with two intermediate long plumose setae—; Dorsal dorsal surface armed with two pairs of spines situated on distal 0.21–0.26 and 0.43–0.49 telson length.

Eye (Fig. 7A) well developed, tip of cornea falling short of the first segment of antennular peduncle when extended forward,—; unpigmented part of eyestalk slightly longer than broad,—; cornea semispherical, slightly shorter than unpigmented part of eyestalk.

Antennular peduncle (Fig. 7F) distinctly overreaching mid_dle_length of scaphocerite. First segment of antennular peduncle with one distolateral tooth,—; inner ventral tooth (Fig. 7G) on 0.59–0.66 of first segment (excluding distolateral tooth), smallsamll. Stylocerite large, reaching 0.86–0.92 (distolateral tooth included), or 0.76–0.81 (distolateral tooth excluded) of first segment. Second segment of antennular peduncle 0.88–0.96 times as long as broad in dorsal view, 0.83–0.95 times as long as third segment. Outer antennular flagellum shorter than inner flagellum and proximal 7–9 segments thicker than orthersothers distal ones. Scaphocerite (Fig.

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Comentário [AdS10]: Revise this sentence to clarify the meaning.

Comentário [AdS111: ?

7H) 2.19–2.38 times as long as wide, distolateral spine of scaphocerite far from reaching distal margin of blade, distolateral spine and blade separated by a notch.

Mouthparts typical for genus. Mandible (Fig. 8A) without palp, incisor process with 5 acute teeth. Maxillula (Fig. 8B) with curved palp, distal margin of upper lacinia armed with 8–10 spines and two long plumose setae. Maxilla (Fig. 8C) with pudgy short palp; scaphognathite broad in upper half and narrow in lower half, lateral border nearly straight; inner lacinia bilobed, distal margin furnished with spinous setae; proximal endite round, with long plumose setae on distal margin. Endopod of first maxilliped (Fig. 8D) slender, with long plumose setae; exopod with feeble caridean lobe on base. Second maxilliped (Fig. 8E) with well-developed exopod, flagelliform; endopod normal, dactylar segment arched, terminal margin armed with row of long spines; propodal segment bearing few long plumose setae; carpus longer than broad, shorter than merus. Third maxilliped (Fig. 8F) reaching to mid-lenghtlength of the scaphocerite when extended forward; exopod reaching to 0.72–0.79 of antepenultimate segment; ultimate segment (excluding apical spine) of endopod 1.61–1.78 times as long as penultimate segment, distal half armed with 6–9 strong spines; antepenultimate segment slightly shorter than the last two segments combined.

First pereiopod (Fig. 9A) shortest among pereiopods, oblique, nearly reaching to midlenghtlength of the scaphocerite when extended forward. Ventral margin of basis, ischium, and merus furnished with long plumose setae. Terminal margin of carpus cotyloid. Cutting edges of chela non-denticulate, with tiny setula and long simple setae; tip of fixed finger with 3 acute spines, tip of dactylus with 4 acute spines (Fig. 9B).

Second pereiopod (Fig. 9C) slightly overreaching to the distolateral spine of scaphocerite when extended forward. Carpus with three subsegments, first subsegment 2.13–2.26 times as long as second subsegment, third subsegment slightly shorter than first subsegment; first subsegment 2.65–2.76 times as long as wide, second subsegment 1.08–1.16 times as long as wide, third subsegment 1.76–1.83 times as long as wide. Cutting edges of chela not denticulate, with tiny setula and long simple distal setae. Tip of fixed finger with 3 acute spines, tip of dactylus with 4 acute spines (Fig. 9D).

Third to fifth pereiopods long and robust. Third pereiopod (Fig. 9E) of female specimen reaching beyond terminal blade of scaphocerite by dactylus when extended forward.—; Dactylus dactylus of third pereiopod with 8–10 spines, all spines in ventral and apical positions (none in dorsal or subdorsal positions), with two large apical spines larger than others (the ultimate longer but thinner than the penultimate) (Fig. 9F).—); Propodus propodus 6.98–7.12 times as long as wide, armed with 4–6 pairs of spines on ventral margin.—; Carpus carpus 2.96–3.14 times as long as wide, armed with one proximal lateral spine.—; Merus merus 6.45–6.63 times as long as wide, armed with 2 lateral spines. Ratio length of third pereiopod dactylus with longest apical spine/length of carpus 0.86–0.92; ratio length of third pereiopod dactylus with longest apical spine/length of carpus 0.86–0.92; ratio length of dactylus without spines/breadth of dactylus without spines 2.86–2.93; ratio length of dactylus with longest spines/breadth of dactylus without spines 4.35–4.43; ratio length of longest spine of dactylus/breadth of dactylus without spines

1.50–1.55; ratio length of longest spine of dactylus/length of dactylus without spines 0.53–0.58. Third pereiopod (Fig. 9G,H) of male specimen with propodus and dactylus forming a prehensile apparatus (Fig. 9G,H). Fourth and fifth pereiopods (Fig. 9I,J) similar in shape to third pereiopod of female specimen, but slightly decreasing in size. Merus of fourth pereiopod armed with 0–1 lateral spine; merus of fifth pereiopod without lateral spine.

First pleopod (Fig. 9K) of female specimen normal, endopod about 0.72–0.78 times as long as exopod. First pleopod (Fig. 9L) of male specimen with endopod about 0.25–0.29 times as long as exopod. Second pleopod (Fig. 9M) of male specimen with endopod about 0.79–0.86 times as long as exopod,—; appendix masculina with 8 apical setae, about 0.41–0.47 times as long as appendix interna (Fig. 9N).

Coloration and Biotopes. The specimens collected in-from different biotopes exhibiting exhibited different body eclourscolors. Specimens (Fig. 10B5B,C) captured among *Galaxaura* sp. are generally pink over the body with numerous white spots; specimens (Fig. 10D5D,E) captured among *Halimeda* sp. are generally light green over the body with white or pink stains on the carapace, abdomen, and telson. All specimens were captured under theat depths of 1–3 m, without other *Hippolyte* spp. was eo captured.

Distribution. Xisha Islands, the South China Sea; <u>presumably Presumably</u>, also distribute in Taiwan Islands (see discussion).

Etymology. 'Nanhai' means the South China Sea; the new species is named after its type locality.

Discussion

Hippolyte chacei sp. nov. can be distinguished from all of the valid Hippolyte species of Hippolyte by its the particular unique dactyli of the third to fifth pereiopods. This kind of dactyli was seldom recorded from in previous literatures, and all the specimens with this kind of dactyli were under the name previously considered as H. ventricosa once upon a time, namely such as specimens the recorded from Malayan Archipelago (of Holthuis (, 1947) from Malayan Archipelago, the record of Ledoyer (1970) from Madagascar (Ledoyer, 1970), and also the record of Hayashi (1981) from Hawai (Hayashi, 1981). d'Udekem D'Udekem d'Acoz (1996) considered that all these specimens previous descriptions were not real H. ventricosa, and might represent some undescribed species-; The the present work, based on molecular data, confirmed the this suspicionjudgment of d'Udekem d'Acoz (1996) based on molecular data. In the 16S rRNA phylogenetic tree (Fig. 1110) of 16S rRNA gene segments, H. chacei sp. nov. (two specimens) formed an lonely-isolated branch clustered in the subbasal position of the Indo-West pacific Pacific clade (Terossi et al., 2017). and Additionally, the average genetic divergence between H. chacei sp. nov. and other Hippolyte spp. is was 20.8%, which is slightly larger than the average interspecific genetic divergence, 20.5% (calculated from the 30 Hippolyte species of Hippolyte in the present study). According to Based on the oversimplified and inadequacy inadequate descriptions of by Holthuis (1947) and Ledoyer (1970), their specimens are were morphologically very similar to H. chacei sp. nov. in morphology, so Therefore, it is we speculated that their specimens may belong to H. chacei sp. nov.,...; however, the validation

should be supportthis should be tested by a-further detailed examination of the specimens of Holthuis (1947) and Ledoyer (1970) specimens. Hayashi (1981) stated that the mouthparts of his Hawaiian specimens were similar to those of H. edmondsoni and H. jarvisensis, and but this kind of the seir mouthparts is weare distinctly different from that those of H. chacei sp. nov., and the difference is also shown in moreover, differences were also observed in the position of hepatic spine. Presumably, the specimens recorded by Hayashi (1981) may represent a different species from H. chacei sp. nov..

Our specimens of *Hippolyte chacei* sp. nov. were captured among *Sargassum* sp. together with *Hippolyte of ventricosa*. This may indicate that the two species occupy similar ecological niche, but the ratio of prisal nearly reached to 1:8. We speculate that *H. chacei* sp. nov. probably remain a predicament in the interspecific competition.

In morphologyMorphologically, Hippolyte-H. nanhaiensis sp. nov. is closely related to H. acuta, H. australiensis, H. ngi; H. singaporensis, and H. ventricosa (redescribed by d'Udekem d'Acoz, 1999.); They they all have the features of first article of the antennular peduncle with one distolateral tooth, fifth pleonite without dorsolateral toothteeth, and third to fifth pereiopods with normal dactyli. H. nanhaiensis sp. nov. differs from H. acuta, H. australiensis, and H. ngi by the its shorter rostrum (reaching to or slightly overreaching the end of the antennular peduncle vs. distinctly overreaching the end of the antennular peduncle). Furthermore, H. acuta is distinguished from H. nanhaiensis sp. nov. also-by a its particularly long eyestalk (Stimpson, 1860; Hayashi & Miyake, 1968; Yanagawa & Watanabe, 1988; d'Udekem d'Acoz, 1996), and H. australiensis is distinguished from H. nanhaiensis sp. nov. by the its rostrum, with which has a sharp lateral carina, and the also by its dactyli of the third to fifth pereiopods, which with have 4 large apical spines (d'Udekem d'Acoz, 2001). H. ngi differs from H. nanhaiensis sp. nov. by the its hepatic, which overreaches the anterior edge of carapace by distal half of hepatic spine overreaches the anterior edge of carapace half length, and also by its the dactyli of the third to fifth pereiopods with, which have 3 large apical spines (Gan & Li, 2017b).

According to the <u>H. ventricosa</u> redescription of <u>Hippolyte ventricosa</u> by d'Udekem d'Acoz (1999) based on type specimens, <u>H. ventricosa</u> and <u>H. nanhaiensis</u> sp. nov. both have <u>2 large apical spines on</u> the dactyli of <u>the</u> third to fifth pereiopods with <u>2 large apical spines</u>, however<u>However</u>, the latter has more longer apical spines. The ratio <u>length</u> of <u>the</u> longest spine of <u>the</u> dactylus/length of <u>the</u> dactylus without spines is 0.53–0.58 in <u>H. nanhaiensis</u> sp. nov., but it is only 0.35 in <u>H. ventricosa</u>. The <u>rostrum of H. ventricosa rostrum</u> distinctly overreaches the end of <u>the</u> antennular peduncle, but it—only reaches to or slightly overreaches the end of <u>the</u> antennular peduncle in <u>H. nanhaiensis</u> sp. nov.. The <u>seaphocerite of H. ventricosa scaphocerite</u> is 3.10 times as long as wide, <u>while itbut</u> is 2.19–2.38 times as long as wide in <u>H. nanhaiensis</u> sp. nov.. According to d'Udekem d'Acoz (1999), the total length <u>of syntype</u> of <u>the H. ventricosa syntypes</u> is up to 17 mm, <u>which is</u> nearly two times larger than the largest <u>specimen of H. nanhaiensis</u> sp. nov. <u>specimen</u>. Furthermore, the two species <u>take upinhabit</u> different ecological niches, <u>.; H. ventricosa living lives</u> among <u>Zostera</u> sp. and <u>Padina</u> sp., <u>maybe and may</u> also <u>be found</u> among <u>Sargassum</u> sp. <u>eteand other organisms</u>. Nevertheless, <u>H. nanhaiensis</u> sp. nov. was

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not found from <u>any of</u> these biotopes, it was only captured from *Galaxaura* sp. and *Halimeda* sp., among which are currently found in higher temperatures at present, and no other species of *Hippolyte* were co captured no congeneric species were found from these biotopes either. As and prefer different species possibly presentmay have different ecological requirements, and prefer different biotopes (d'Udekem d'Acoz, 1996).

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435 436 In the 16S rRNA phylogenetic tree (Fig. 1110) of 16S rRNA gene segments, Hippolyte H. nanhaiensis sp. nov. (two specimens); formed a clade together with H. ventricosa group—sp. 4 (Terossi et al., 2017), form a monophyletic clade being sister group withand this clade is sister to H. australiensis. The average genetic divergence between H. nanhaiensis sp. nov. and other Hippolyte spp. is 22.5%, which is larger than the average interspecific genetic divergence; (20.5%) (calculated from the 30 species of Hippolyte in the present study). The result of 16S rRNA sequences alignment of 16S rRNA gene segments showshowed that the H. nanhaiensis sp. nov. sequences were of H. nanhaiensis sp. nov. is identical withto, or only has had one nucleotide base different difference from, that of H. ventricosa group—sp. 4 (KX588916). In this ease Therefore, H. ventricosa group—sp. 4 and H. nanhaiensis sp. nov. may represent the same species.

During the biodiversity surveys (Hainai Island biodiversity surveys were conducted in 2014, 2015, 2016, 2017, and 2018, the Xisha Islands biodiversity surveys were conducted in 2015. 2016), dozens of Hippolyte-H. cf. ventricosa (GenBank accession number of 16S rRNA gene: MK231003, MK231004, MK231009) were collected among biotopes of Sargassum sp. and Thalassia sp. from the nearshore waters of Hannai Hainan Island (no specimens were found from the Xisha Islands). These specimens have had the following features: (1) first article of the antennular peduncle has with one distolateral tooth, and fifth pleonite has no dorsolateral tooth; (2) carapace length of mature females is among 1.8-3.3 mm, and total length among is 13-24 mm; (3) rostrum distinctly overreaches overreaching the end of the antennular peduncle but falling short of scaphocerite apex, superior border with 1-2 tooth-teeth and inferior border with 1-5 toothteeth; (4) incisor process of mandible with 5-6 teeth; (5) scaphocerite 2.79-3.38 times as long as wide; (46) the dactyli of the third to fifth pereiopods with 2 large apical spines, but the longest apical spines never exceeding the half-length of dactyli properly, the ratio length of the longest spine of dactylus/length of dactylus without spines is among 0.33-0.41; (57) the specimens displaying various colorations (Fig. 10F5F-G). All of these features differed from that those of H. acuta, H. australiensis, H. ngi; H. singaporensis, and H. nanhaiensis sp. nov., but indicate these specimens belong to are similar to those of H. ventricosa (based on the redescription by d'Udekem d'Acoz, 1999). HoweverRecently, more than four cryptic or pseudocryptic species of *H. ventricosa* were detected through with molecular data (De Grave et al., 2014; Terossi et al., 2017), although d'Udekem d'Acoz (1999) detailedly redescribed H. ventricosa based on type specimens which were also morphologically very similar to H. ventricosa (De Grave et al., 2014; Terossi et al., 2017). Therefore, It it is currently unclear which specimens represent confused to determine which one is true H. ventricosa-; the 16S rRNA gene

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segment or other genetic data derived from the *H. ventricosa* topotype of *H. ventricosa* is expected are expected to clarify this issue.

Conclusions

As pointed noted by d'Udekem d'Acoz (1999; 2001), the systematics of the Indo-West pacific Pacific genus Hippolyte is extremely hysteretiecomplicated, despite even though this region was is considered to be the origin center of the genus (Terossi et al., 2017). The taxonomic troubles confusion eome from both the lack of knowledge on several species, such as H. proteus, H. kraussiana, and H. acuta, the poorly known species and the vast continuous variation of morphological characteristies. Our research indicatestudy revealed that the proportion of the rostrum, the scale of scaphocerite scale, the position of the hepatic spine, and the features of the dactyli of the third to fifth perciopods are more significant extremely important in Hippolyte morphological taxonomy of Hippolyte. And in the future, a new taxon established based on integrative datum, eg morphological data, genetic data, and ecological data and so on, will be more valuable and credible. A preliminary key tofor the identification of mature female of the genus Hippolyte of the Indo-West Pacific and neighboring seas is provided. This key only contains the valid species listed in WoRMS (http://www.marinespecies.org), and the cryptic or pseudocryptic species of H. ventricosa are temporarily pooled together as 'H. ventricosa' sensu lato.

Comentário [AdS18]: Relatively to exactly what?

Comentário [AdS19]: What exactly are the relevant morphological features of the scaphocerite? And it is the scaphocerite scale is exactly?

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4-Carapace with dorsal surface fabulously gibbous. H. dossena

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477	6-Rostrum longer than the half length of carapace, reaching to the end of antennular	
478	peduncle	
479	7-Rostrum without dorsal tooth, base of hepatic spine nearly situating at anterior edge of	
480	carapace	
481	7-Rostrum with 1-2 dorsal teeth, base of hepatic spine situating at posterior to the anterior edge	
482	of carapace8	
483	8-Dactyli of third to fifth pereiopods with three long terminal teeth, distal half of hepatic spine	
484	overreaching anterior edge of carapace	
485	8-Dactyli of third to fifth pereiopods with two terminal teeth, hepatic spine slightly overreaching	
486	anterior edge of carapace 9	
487	9-Distal spine of dactylus of third pereiopod longer than the half length of dactylus proper	
488	(excluding spines)	
489	9-Distal spine of dactylus of third pereiopod shorter than the half length of dactylus proper	
490	(excluding spines). H. jarvinensis	
491	10-Rostrum with postrostral spine, situating at just above the orbit	
492	10-Rostrum without postrostral spine, all dorsal spines situating at prior to the	
493	orbit. H. nanhaiensis sp. nov.	
494	11-Incisor process of mandible with no more than 8 acute teeth	
495	11-Incisor process of mandible with 15–17 acute teeth, dactyli of third to fifth pereiopods with 2–	
496	3 subdorsal spines	
497	12-Unpigmented part of eyestalk 3 times as long as cornea	
498	12-Unpigmented part of eyestalk no more than 3 times as long as cornea	
499	<u>13-Rostrum</u> without dorsal	
500	<u>spine</u> 14	
501	13-Rostrum with dorsal spine	
502	14-Apex of the rRostrum with-trifid apex. H.	
503	<u>multicolorata</u>	
504	14-Apex of rostrum normal, non-trifidsimple	
505	<u>australiensis</u>	
506	15-Apex of the Rrostrum with bifid apex	
507	apexH. bifidirostris	
508	15-Apex of rostrum normal, non-bifidsimple	Format Unidos
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