

Effects of root phenotypic changes on the deep rooting of *Populus euphratica* seedlings under drought stresses

Zi-qi Ye^{1,2}, Jian-ming Wang¹, Wen-juan Wang¹, Tian-han Zhang¹, Jing-wen Li^{Corresp. 1}

¹ The College of Forestry, Beijing Forestry University, Beijing, China

² Institute of Forest Ecology, Environment and Protection, Chinese Academy of Forestry, Beijing, Beijing, China

Corresponding Author: Jing-wen Li

Email address: lijingwenhy@bjfu.edu.cn

Background. Deep roots are critical for the survival of *Populus euphratica* seedlings on the floodplains of arid regions where they easily suffer drought stress. Drought typically suppresses root growth, but *P. euphratica* seedlings can adjust phenotypically in terms of root-shoot allocation and root architecture and morphology, thus promoting deep rooting. However, the root phenotypic changes undertaken by *P. euphratica* seedlings as a deep rooting strategy under drought conditions remain unknown. **Methods.** We quantified deep rooting capacity by the relative root depth (RRD), which represents the ratio of taproot length to plant biomass and is controlled by root mass fraction (RMF), taproot mass fraction (TRMF), and specific taproot length (STRL). We recorded phenotypic changes in one-year-old *P. euphratica* seedlings under control, moderate and severe drought stress treatments and assessed the effects of RMF, TRMF, and STRL on RRD. **Results.** Drought significantly decreased absolute root depth but substantially increased RRD via exerting positive effects on TRMF, RMF, and STRL. Under moderate drought, TRMF contributed 55%, RMF 27% and STRL 18% to RRD variation. Under severe drought, the contribution of RMF to RRD variation increased to 37%, which was similar to the 41% for TRMF. The contribution of STRL slightly increased to 22%. **Conclusion.** These results suggest that the adjustments in root architecture and root-shoot allocation were predominantly responsible for deep rooting in *P. euphratica* seedlings under drought conditions, while morphological changes played a minor role. Moreover, *P. euphratica* seedlings rely mostly on adjusting their root architecture to maintain root depth under moderate drought conditions, whereas root-shoot allocation responds more strongly under severe drought conditions, to the point where it plays a role as important as root architecture does on deep rooting.

1 **Effects of root phenotypic changes on the deep**
2 **rooting of *Populus euphratica* seedlings under**
3 **drought stresses**

4

5 Zi-qi Ye^{1,2}, Jian-ming Wang¹, Wen-juan Wang¹, Tian-han Zhang¹, Jing-wen Li^{1*}

6

7 1 The College of Forestry, Beijing Forestry University, No. 35 Qinghua East Road, Haidian
8 District, Beijing 100083, China.

9 2 Institute of Forest Ecology, Environment and Protection, Chinese Academy of Forestry, No.2
10 Dongxiaofu, Xiangshan Road, Haidian District, Beijing 100091, China

11 *Corresponding Author: Jing-wen Li

12 Email address: lijingwenhy@bjfu.edu.cn

13

14 **ABSTRACT**

15 **Background.** Deep roots are critical for the survival of *Populus euphratica* seedlings on the
16 floodplains of arid regions where they easily suffer drought stress. Drought typically suppresses
17 root growth, but *P. euphratica* seedlings can adjust phenotypically in terms of root-shoot allocation
18 and root architecture and morphology, thus promoting deep rooting. However, the root phenotypic
19 changes undertaken by *P. euphratica* seedlings as a deep rooting strategy under drought conditions
20 remain unknown.

21 **Methods.** We quantified deep rooting capacity by the relative root depth (RRD), which represents
22 the ratio of taproot length to plant biomass and is controlled by root mass fraction (RMF), taproot
23 mass fraction (TRMF), and specific taproot length (STRL). We recorded phenotypic changes in
24 one-year-old *P. euphratica* seedlings under control, moderate and severe drought stress treatments
25 and assessed the effects of RMF, TRMF, and STRL on RRD.

26 **Results.** Drought significantly decreased absolute root depth but substantially increased RRD via
27 exerting positive effects on TRMF, RMF, and STRL. Under moderate drought, TRMF contributed
28 55%, RMF 27% and STRL 18% to RRD variation. Under severe drought, the contribution of RMF
29 to RRD variation increased to 37%, which was similar to the 41% for TRMF. The contribution of
30 STRL slightly increased to 22%.

31 **Conclusion.** These results suggest that the adjustments in root architecture and root-shoot
32 allocation were predominantly responsible for deep rooting in *P. euphratica* seedlings under
33 drought conditions, while morphological changes played a minor role. Moreover, *P. euphratica*
34 seedlings rely mostly on adjusting their root architecture to maintain root depth under moderate
35 drought conditions, whereas root-shoot allocation responds more strongly under severe drought
36 conditions, to the point where it plays a role as important as root architecture does on deep rooting.

37

38 INTRODUCTION

39 The root system is the main plant organ acquireing below-ground resources. To adapt to the
40 inherent heterogeneity of soil resources, plants can adjust root phenotype on different integrated
41 levels (Chapin, 1991; Nicotra et al., 2010; Poorter et al., 2012): they can change their relative
42 investment of biomass in shoots and roots on an individual-level, they can adjust root system
43 architecture on an organ-level, or alter root morphology on a module-level (Fig. 1). Most likely,
44 plants adjust their roots on all three levels. However, different environmental stresses may result
45 in different degrees of root phenotypic adjustments, resulting in the different magnitude in
46 functional contributions for plants under environmental stress. For instance, Freschet, Swart &
47 Cornelissen (2015) found that root mass fraction (RMF; the proportion of total plant mass allocated
48 to roots) responded more strongly than specific root length (SRL; root length for a given unit of
49 plant mass) to nutrient deficiency, thus suggesting that with nutrient stress, the increased allocation
50 to roots seems more important than the root morphological change for the plant to achieve an
51 increase in root length. Nevertheless, root architectural change is a basic way to improve fine-root
52 function efficacy (Lynch, 1995; McCormack et al., 2017). Increasing the density and length of the
53 distal roots can increase root absorption area (Kong et al., 2014). Additionally, different root
54 patterns can influence plant uptake efficiency in heterogeneous resource environments (Lynch,
55 2005) and can affect the ability of root systems to capture relatively immobile versus mobile soil
56 resources (Fitter, 1987). However, few studies have focused on the relative functional importance
57 of the different phenotypic changes in stressed plants, especially the relative role of root
58 architectural changes (Weemstra et al., 2016; except Kramer-walter & Laughlin, 2017; Freschet
59 et al., 2018).

60 *Populus euphratica* Oliv. (Salicaceae) is a dioecious riparian tree species found
61 discontinuously within the continental-arid climate region of Central Asia (Browicz, 1977; Wang,
62 1996), which forms monospecific stands along continental rivers. This poplar is an obligate
63 phreatophyte with a root system that continuously contacts the groundwater or the soil water-
64 saturated zone (Zhu et al., 2009), meaning that its growth and survival depends highly on locating

65 and acquiring groundwater (Gries et al., 2003). However, its seedlings cannot reach groundwater
66 during the early stages, meaning that under the harsh environments in arid regions, *P. euphratica*
67 can only propagate generatively in the freshly deposited floodplain soils from May to August (Cao
68 et al., 2012). Even so, the optimal soil conditions in such floodplains for seedling germination and
69 growth only exist for a short time, as the surface soil post-flood rapidly becomes increasingly dry
70 and salty due to evaporation. The rapid onset of winter also shortens the growth period for the
71 seedlings. As a result, *P. euphratica* seedlings must establish a deep root system during the early
72 stage, as only seedlings with deep roots can secure water uptake during the dry period and survive
73 the following year (Zerbe & Thevs, 2011). According to Thevs et al. (2008), Wiehle et al. (2009),
74 and our previous field investigations, *P. euphratica* seedlings established in floodplains possess
75 deep root systems very asymmetrical in size to the shoots. Therefore, deep rooting is considered a
76 key process for survival and establishment of early *P. euphratica* seedlings in an inconsistent
77 riparian environment (Hukin et al., 2005; Wang et al., 2015).

78 *Populus euphratica* seedlings are easily subjected to drought stress in semi-arid regions,
79 (Thevs et al., 2008; Stella et al., 2010), which decreases biomass accumulation and limits root
80 development into the deep soil. Accordingly, they take some phenotypic adjustments in order to
81 deal with drought stress, such as increased root biomass allocation (Wang et al., 2015), root
82 architecture adjustment and root morphology regulation (i.e., increased SRL; Lü et al., 2015). Each
83 of these changes was believed to cause increased seedling root depth. However, as previously
84 outlined, different levels of phenotypic changes may have different functional contributions, and
85 thus the knowledge is still incipient regarding the exact effects of root phenotypic changes on the
86 root depth of *P. euphratica* seedlings under drought stress. Further research on this issue would
87 aid in our understanding of the drought adaptive strategy of *P. euphratica* and other similar deep-
88 rooted plants in the arid regions.

89 To understand the changes to plant root depth in response to environmental conditions, it is
90 important to distinguishing between absolute and relative plant dimensions because water deficit
91 may strongly influences the overall plant size in a negative manner (Fig. 1), which often obscures

92 the underlying deep-rooting process (Schenk & Jackson, 2002). Therefore, we adopted the
93 absolute root depth to total plant biomass ratio to quantify relative root depth (RRD), i.e., deep-
94 rooting capacity, which follows the method of Ryser & Lambers (1995) who used root length per
95 total plant biomass to express the relative length of fine roots, i.e., root uptake capacity. Here, we
96 define deep-rooting capacity as the capacity of the plant use all available resources to achieve an
97 increase in root depth via all possible ways. As so, deep-rooting capacity is regarded as an overall
98 capacity that could be depicted by several facets of drought responses (this definition is similar to
99 that of root nutrient acquisition capacity used by Freschet et al. (2018)). Thus, RRD
100 mathematically follows from the increases in RMF and/or the proportion of root system mass
101 invested in taproot (i.e., taproot mass fraction (TRMF)) and/or the taproot length achieved per unit
102 taproot mass (i.e., specific taproot length (STRL)) (see Eq. 1).

103 The present study aimed to answer the following questions: (1) How do *P. euphratica*
104 seedling roots change phenotypically on the individual-, organ- and modular- levels under drought
105 stress? (2) Which level of root phenotypic adjustment (root-shoot allocation, root architecture, or
106 morphology) plays a leading role in facilitating the deep rooting of the drought-stressed seedlings?
107 and (3) Does the relative contributions of these different root phenotypic adjustments on deep
108 rooting vary with drought intensity?

110 MATERIALS & METHODS

111 Experimental method

112 *Nursery phase*

113 This study was conducted at the State Forest Farm located in Ejin Banner, Inner Mongolia, China.
114 All seeds were randomly collected from a mature natural forest dominated by *P. euphratica* (41°57
115 51.3" N, 101°05 06.0" E) along the Ejin River. Planted pots were 20 L in volume, 40 cm in depth,
116 and filled with 16 kg (dry weight) of substrate – a 4/6 (v/v) mixture of peat and sand. A slow-
117 release fertilizer (4 g L⁻¹ Osmocote 16:9:12 NPK and trace elements, product code: 8840) was pre-
118 mixed within the substrate. From May to June 2016, *P. euphratica* seeds were sown and
119 germinated in 90 plastic pots placed in a greenhouse. After a month of growth with normal
120 management, three seedlings per pot, averaging 2.0 ± 0.5 cm in height and bearing four to six
121 leaves, were selected and transferred to the open-air nursery. After two weeks of acclimation, only
122 a single healthy seedling per pot, averaging 7.0 ± 1.5 cm in height and 0.9 ± 0.2 mm in ground
123 diameter, was kept. This nursery phase took about 50 days.

124 *Experimental treatments*

125 This experiment was conducted over a period of 60 days from July to September 2016, during
126 which the average of daily minimum and maximum temperatures were 21.7 °C and 34.3 °C
127 respectively. The daily maximum temperature ranged from 25 °C to 44 °C. The experiment was
128 conducted in a completely randomized design, including three water treatments: 70–80% of field
129 capacity [(optimal water content (OW)], 50–60% of field capacity [moderate drought stress (MD)],
130 and 30–40% of field capacity [(severe drought stress (SD)]. Each treatment was replicated 30
131 times, and each replication consisted of s a seedling planted in an independent pot.

132 The seedlings were randomly assigned to one of the three treatments to avoid the variance in
133 individual growth. Pot body was completely hidden into ground to avoid pot warming and just
134 exposed surface soil to the arid climate, by which the soil in pot could have the relative real
135 underground temperature and moisture in vertical gradient. Because the evapotranspiration rate in
136 Ejin is high during summer days, soil water content was supplemented every day using the weight

137 method to keep it at a certain range. After water supplementation, the pots were rearranged
138 randomly to neutralize the influence of potential environmental heterogeneity. Plastic film was
139 used to cover the nursery to avoid rainfall. Owing to the death of five seedlings subject to the SD
140 treatment, and to the destruction of two seedlings from OW treatment and two seedlings from MD
141 treatment during destructive sampling and root measurements, only 81 seedlings were successfully
142 measured and accessed to analyses.

143 *Growth measurements and destructive sampling*

144 Intact root systems were cleaned of soil with a gentle water jet while a sieve was used to collect
145 any root fragments detached from the system during this process. The cleaned root systems were
146 then floated on water in a transparent tray and imaged using an Epson Expression Perfection V850
147 Pro Scanner with 800 dpi resolution (Fig. 2). Adobe Photoshop CS6 software was used to reduce
148 image noise and black margins. The image analysis software (Win-RhIZO 2013a Pro Instruments
149 Inc., Québec, Canada) was used to analyze images and to estimate the length of total, lateral, and
150 distal roots, the average diameter of the total roots, the number of lateral roots, as well as the
151 external path length (p_e), magnitude (μ), and altitude (a) (Table 1). The taproot was then detached
152 from the root system with scissors, and its length and dry mass were determined. Lateral roots
153 were defined as the root segments connected to the taproot with a root order > 3 , so as to guarantee
154 that such roots perform conducting and foraging functions. Three intact lateral root branch
155 segments with over three levels of root order were selected randomly and scanned to estimate the
156 average length and diameter of the distal roots. Using a scalpel, all distal fine roots were dissected
157 and was determined with its dry mass. If it was impossible to meet these criteria, the whole root
158 system was measured. The dry weights of the taproot, sampled distal roots, and the other root parts
159 were obtained by air drying plant matter in an oven at 72 °C for 60 h. Total root biomass, specific
160 root length, biomass-related variables, and two topological indices were calculated using these dry
161 weight values (Table 1).

162

163 **How to represent root phenotypic adjustments and deep-rooting capacity**

164 As in many other studies, root-shoot allocation was quantified as the fraction of plant biomass
165 invested in the roots (RMF; see Table 1 for definition), and taproot morphological change could
166 be expressed as specific taproot length (STRL; see Table 1 for definition). These two aspects of
167 root phenotypic change both have independent effects in the adjustment of root length (i.e., root
168 depth in this study) (Fig. 1).

169 As it is difficult to fully measure root architecture, previous studies commonly used topology
170 to describe the altered branching patterns indicative of a facet of the root architectural change
171 (Fitter, 1987; Harper, Jones & Sackville, 1991; Lynch, 1995). Here, we not only used the
172 topological index (Fitter, 1987; Glimskär, 2000) and the dichotomous branching index
173 (Šmilauerová & Šmilauer, 2002), both commonly applied to characterize root topology (i.e., TI
174 and DBI, respectively; see Table 1 for definitions), but also used the taproot mass fraction (TRMF;
175 see Table 1 for definition) to characterize root architectural change.

176 It is logical to characterize root architecture as the biomass proportion of a certain functional
177 root module in relation to the whole root system (e.g. TRMF) because the root branching pattern
178 just refers to the coordinated growing relation among different functional root modules, and this
179 relation could be represented with biomass proportion as a mass proxy. In particular, an extreme
180 herringbone-like branching pattern has been found to be primarily confined to the main axis (Fitter
181 et al., 1991) – that is, possessing the largest proportion of taproot biomass. Therefore, considering
182 taproot as a key root module functioning in deep rooting, in this study TRMF was specially used
183 to trace the facet of root architectural change that contribute to deep rooting. In addition, the
184 correlation between TRMF and the commonly used TI and DBI has been examined to determine
185 the availability of TRMF to representing root architecture ($R^2=0.643$ and 0.698 respectively, both
186 $P < 0.001$); Fig. S1), and the results indicated that TRMF could be feasibly used in this study.

187 *Populus euphratica* has an obvious taproot that determines root system depth, and thus
188 absolute root depth can be reflected by taproot length. Accordingly, relative root depth (RRD; see
189 Table 1) was calculated as the absolute taproot length to taproot biomass ratio. Finally, RRD can
190 be factored into RMF, TRMF, and STRL, as follows:

191

192

$$RRD = RMF \times TRMF \times STRL \quad (\text{Eq. 1})$$

193

194 Data analyses

195 Differences observed in biomass allocation as well as in root architecture and morphology among
 196 the three drought treatments were tested using one-way analysis of variance (ANOVA, Welch's F
 197 test). After that, variations of statistical significance were further subjected to *post hoc* pairwise
 198 analysis by applying *t*-tests with Bonferroni corrections, or Games-Howell tests if the homogeneity
 199 of variances was not assumed, considering $P < 0.05$ as significant. The dependence between TRMF
 200 and TI or TRMF and DBI was determined by Pearson's correlation analysis. Statistical analyses
 201 were performed in SPSS (version 19, SPSS Inc., Chicago, IL, USA).

202 We calculated the relative contributions of the variance in RMF, TRMF, and STRL to RRD,
 203 referencing the variance partitioning method of Rees et al. (2010) and Freschet et al. (2015). Given
 204 that $RRD = RMF \times TRMF \times STRL$, our calculation can be expressed as:

$$205 \quad rrd = rmf + trmf + strl \quad (\text{Eq. 2})$$

206 , where the lowercase acronyms indicate \log_e -transformed variables (e.g., $rrd = \ln(RRD)$). Thus,
 207 the variance decomposition of *rrd*, for instance, can be expressed as follows:

$$208 \quad \text{Var}(rrd) = \text{Var}(rmf) + \text{Cov}(rmf, trmf) + \text{Cov}(rmf, strl) + \text{Var}(trmf) + \text{Cov}(rmf, trmf) + \text{Cov}(trmf, strl) + \text{Var}(strl) + \text{Cov}(rmf, strl) + \text{Cov}(trmf, strl) \quad (\text{Eq. 3})$$

209 Following Eq. 3, as a sample, the contribution of variation in *trmf* to the variation in *rrd* can be
 210 written as:

$$212 \quad \text{Cont}(trmf) = [\text{Var}(trmf) + \text{Cov}(rmf, trmf) + \text{Cov}(trmf, strl)] / \text{Var}(rrd) \quad (\text{Eq. 4})$$

213 , where *Var* is the variance and *Cov* is the covariance. This variance partitioning was only
 214 performed when substantial variation (i.e., $\geq 15\%$) in *rrd* was observed across treatments, so as to
 215 avoid meaningless results.

216

217 RESULTS

218 Drought-induced phenotypic changes in biomass, root morphology, and root architecture

219 Drought stress inhibited *P. euphratica* seedlings' growth in both biomass and root morphology
220 (Table. 2). Drought conditions caused a dramatic decrease in the biomass of different plant parts,
221 total root length, and the average length of taproot, lateral roots, and distal roots. However, when
222 examining responses to differing drought intensities, taproot length and taproot biomass did not
223 display a consistent declining trend with increasing drought intensity, and there was no difference
224 in taproot length and taproot biomass between MD and SD treatments (Table 2). Moreover, the
225 diameter of different root classifications had different responses to drought (Table 2). Root
226 diameter (calculated with total roots) did not change significantly under drought conditions, but
227 distal root diameter increased and lateral root diameter decreased significantly (Table 2). Taproot
228 diameter decreased with drought, but not significantly. Furthermore, while the SRL (calculated
229 with total roots) did not change by drought, the SRL of the distal roots (i.e., SDRL) – the root
230 components more actively involved in water uptake – showed a significant decrease in SD in
231 relation to OW (Table 2).

232 *Populus euphratica* seedlings showed a high plasticity in root branching patterns under
233 different drought stresses. DBI and TI measures the degree to which a root is perfectly herringbone
234 (DBI or TI equal to 1) or dichotomous branching (DBI equal to 0, or TI close to 0.5). Values of
235 DBI and TI differed significantly among the three drought treatments, with the highest values in
236 the SD and the lowest in the OW (Table 2). The ranges of value presented by the total samples
237 were 0.650 to 0.844 for TI and 0.032 to 0.392 for DBI. Lateral root branching density, a simple
238 but direct trait reflecting root branching, showed a markedly decreased trend from OW to SD
239 treatments (Table 2).

240

**241 Different levels of root phenotypic variables related to deep rooting changed differently
242 under drought**

243 Drought had a significant positive effect on RRD, but this increase was not proportional to the
244 differences in RRD between SD and MD, it was almost three times greater than the difference in

245 RRD between MD and OW (Table 3). Regarding the indices representing different levels of root
246 phenotypic changes, RMF and TRMF increased markedly with increasing drought severity, while
247 STRL did not significantly increase under drought conditions. In addition, the TRMF under SD
248 was not significantly different from that under MD, but RMF markedly increased under SD in
249 relation to that under MD (Table 3).

250

251 **Relative contributions of different root phenotypic changes to deep rooting**

252 These results indicated that drought stress had a positive effect on RRD mainly via significant
253 positive effects on TRMF and RMF. Generally, TRMF and RMF contributed to over 75% of the
254 variation in RRD, while STRL contributed to only 20% (Fig. 3). Moreover, TRMF always had the
255 largest relative contribution to the variation of RRD both under MD (55%) and under SD (41%)
256 in relation to that under OW (Fig. 3). Furthermore, the relative contributions of RMF, TRMF, and
257 STRL to RRD variation changed with drought intensity (Fig. 3). TRMF contributed 55% of RRD
258 variation between MD and OW, which was larger than the 27% contributed by RMF and 18% by
259 STRL. However, under severe drought stress, RMF contributed to 37% of the variation in RRD,
260 nearly equal with 41% for TRMF. The contribution of STRL on RRD variation also increased
261 slightly to 22%. In addition, the variation of RRD from MD to SD treatments was even more
262 attributable to RMF changes (41%) than to TRMF changes (36%) (Fig. 3).

263

264 **DISCUSSION**

265 **Root adaptive responses to drought stress in terms of allocation, morphology, and** 266 **architecture**

267 One of the basic ways for plants to adapt to a shortage of below-ground resources is to maximize
268 fine root area (e.g., decreased root diameter (RD) and increased SRL under drought stress) (Fitter,
269 1985; Cortina et al., 2008; Olmo, Lopez-Iglesias & Villar, 2014). However, in the present study,
270 the RD and SRL of whole roots exhibited no significant change, but the RD of distal roots
271 increased and SDRL decreased under drought conditions (Table 2). Commonly, roots with a

272 smaller RD and higher SRL is advantageous under drought stress, as the smaller RD conveys
273 higher resistance to root embolism (Alameda & Villar, 2012) through the smaller xylem vessel
274 diameter (Fitter, 1987), and a higher SRL probably is meant to commit limited carbohydrate supply
275 for extensional growth (Trubat, Cortin & Vilagrosa, 2006). The unusual results (increased RD of
276 distal root and decreased SDRL under drought) in this study were likely due to the smaller SRL,
277 which is positively correlated with root life span and respiration rate (McCormack et al., 2012),
278 allowing *P. euphratica* seedlings to save considerable energy in dry soil through a low root
279 turnover rate in distal roots. Moreover, thicker fine roots would be able to penetrate into the more
280 compacted soils imposed by soil drying (Bengough et al., 2005). In addition, the thicker distal
281 roots of *P. euphratica* seedlings show higher drought resistance to preserve their vitality in the
282 topsoil of arid-region floodplains where soil water availability is widely fluctuating with the
283 regime of flood and dry seasons (Leon et al., 2011).

284 Plants mostly elongate root internode lengths to adapt to drought environments (Nicotra &
285 Westoby, 2002) as this also extends the volume of root exploring (Fitter & Stickland, 1992).
286 However, in our results, average root length did not significantly change and lateral root length
287 clearly decreased under drought stress (Table 2). Ecologically, floodplain soil possesses a high
288 infiltration rate, which is due to sandy soil layer (several meters in depth) deposited under the 10–
289 50 cm deep surface clay soil layer formed by floodwater sedimentation (Thevs et al., 2008). Thus,
290 rooting downward into deep soil is more critical for drought-stressed *P. euphratica* seedlings than
291 rooting in other directions and shortening most of their lateral roots can avoid inefficient
292 investments in the horizontal exploration of soil zone (Padilla & Pugnaire, 2007; Bauerle et al.,
293 2008).

294 Root architecture plays a major role in determining root resource-uptake efficiency (Fitter,
295 1987; Lynch, 2005). This multidimensional root feature is generally described by measuring root
296 topology (Harper et al., 1991) or branching density/intensity (Kong et al., 2014). In the present
297 study, the lateral root branching density was reduced under drought conditions (Table 2). From a
298 functional point of view, the sparse lateral root branching of *P. euphratica* seedlings seems to

299 conserve the high metabolic cost of root construction and maintenance, which can commonly
300 exceed 50% of daily photosynthesis (Lambers, Atkin & Millenaar, 2002). Besides, the sparse
301 lateral root branching may reduce the competition for water among the roots of an individual plant
302 (Fitter et al., 1991; Taub & Goldberg, 1996), which effectively increases the uptake efficiency per
303 unit of lateral root length (Postma, Dathe & Lynch, 2014).

304 Our results indicated that *P. euphratica* seedlings tended to create a herringbone-like
305 branching pattern under drought conditions, as revealed by the increased TI and DBI values under
306 increased drought treatments (Table 2). This finding is in line with most model-based and
307 empirical studies conducted on plants under drought conditions (Fitter, 1991; Taub & Goldberg,
308 1996). Herringbone-like root systems possess higher exploration efficiency (Ho et al., 2005; Paula
309 & Pausas, 2011), thereby allowing for *P. euphratica* seedlings to reach water-rich deep soils
310 quickly. Additionally, it should be noticed that although our experiment exists an obvious vertical
311 gradient of soil moisture in pot, stimulating more actual ground water condition and different soil
312 texture layers would be better for determining deep-rooting strategy of *P. euphratica* seedlings in
313 further studies.

314

315 **Changes to absolute and relative root depth under drought stress**

316 The four-month-old *P. euphratica* seedlings grown in our experiment ultimately presented total
317 biomasses ranging from about 0.2–0.5 g (Table 2), and, surprisingly, they developed taproots with
318 approximately 26 to 35 cm long (Table 2). This indicates that, despite their low biomass
319 accumulation rate, *P. euphratica* seedlings have a great capability to root deeply at their early
320 stages. The miniscule biomass accumulation by the first-year seedlings might be due to their
321 extremely small and light seeds (0.1–0.2 g per thousand seeds). A similar ontogeny was also found
322 in other studies concerning first-year riparian seedlings. An experiment under drought conditions
323 conducted by Wang et al. (2015) showed that *P. euphratica* seedlings sown in April had about 1 g
324 dry mass and over 22 cm taproot length by the end of July. Likewise, a study of riparian tree
325 seedlings (family Salicaceae) conducted by Stella et al. (2010) demonstrated that three-month-old

326 cottonwood seedlings (*P. fremontii*) had 0.3 g dry weight with roots over 20 cm in depth, and that
327 three-month-old *Salix exigua* and *S. gooddingii* seedlings developed root depths exceeding 25 cm
328 and 40 cm, respectively, despite exhibiting dry weights of only 0.22 g and 0.4 g, respectively.
329 Clearly, the tiny but deep-rooting seedling phenotype seems common in riparian tree species
330 growing in arid regions.

331 Changes in plant root depth under drought stress are controlled by two processes (Fig. 1)
332 (Sultan, 2000). On the one hand, drought stress weakens photosynthesis, dwindles the accumulated
333 biomass and body size, and thereby shortening the root depth. On the other hand, the root depth is
334 influenced by different drought-induced root phenotypic responses. In this study, the absolute root
335 depth of *P. euphratica* seedlings decreased significantly (Table 2) while the RRD increased
336 significantly under drought conditions (Table 3), indicating that the limitation in growth caused by
337 drought had an overwhelming effect on root depth, but which was compensated by enhancing
338 deep-rooting capacity of *P. euphratica* seedlings under drought.

339

340 **Relative contributions of root phenotypic changes to the increase of relative root depth**

341 Our results indicated that changes to root architecture and root-shoot allocation in *P. euphratica*
342 seedlings dominated for achieving deep rooting under drought conditions, while the role of
343 morphological changes was minor (Fig. 3). This supports the perspective of Freschet et al. (2015)
344 that root-shoot allocation was more important than root morphological changes for plant
345 adaptations to changing environmental conditions. Besides, STRL in this study did not
346 significantly increase under drought stress (table 3) and maintained relatively slight contributions
347 to deep rooting regardless of drought intensity (Fig. 3), although taproot stretching seems to be an
348 efficient way to increase root depth. This imply that phenotypic changes in module-level
349 (morphology) seems not much important for plants to respond belowground resources, given that
350 the role of SRL of fine roots (absorptive roots) are also marginal for plant phenotypic adaption to
351 nutrient limitation (Kramer-walter & Laughlin, 2017). The potential negative effects of increased
352 STRL on taproot function may account for this result. Because SRL is determined negatively by

353 the root diameter and root tissue mass density (Nicotra & Westoby, 2002), increased STRL means
354 decreased taproot diameter or tissue density. This negatively affects taproot conduction,
355 anchorage, and penetration, which are all essential functions for drought-suffered seedlings.

356 The relative contributions of changes in root-shoot allocation and root architecture were
357 altered obviously under different drought intensities (Fig. 3). Under MD, root architectural changes
358 played a decisive role (contribution over 50%) for increasing RRD. However, under SD, the root-
359 shoot allocation response was stronger than that under MD (Table 3), and its relative contribution
360 to deep rooting became nearly as important as that of root architectural changes (Fig. 3). This is
361 likely because increased root-shoot allocation would have decreased photosynthetic capacity and
362 accumulation of photosynthates (Muller et al., 2011), while root architecture changes seems more
363 carbon economical under drought conditions, by which plant would root deeply only at the cost of
364 weakening exploration capacity to the horizontal and surface soil (Thevs et al., 2008). Moreover,
365 the variation of RRD from SD to MD is about three times that of the variation from OW to MD
366 (Table 3), which indicates that keeping deep root is more important when drought becomes severe.
367 Therefore, adjustment in root architecture seems to be insufficient for *P. euphratica* seedlings to
368 root deeply under severe drought conditions. From a functional view, the seedlings possibly cannot
369 easily acquire the adequate amount of water to maintain metabolic processes with increased
370 drought stress. Accordingly, the seedlings would rely more on allocating biomass to roots, which
371 have an advantage in minimizing water loss of shoot transpiration and in enhancing deep rooting
372 potential (Brunner et al., 2015). In addition, our study revealed a practical implication for breeding
373 *P. euphratica* seedlings. To promote the survival of container seedlings transplanted in the field,
374 it is a good idea to breed the seedlings under moderate drought stress before transplantation, in
375 order to promote deeper and steeper root systems while minimally affecting their sizes.

376 It has been widely reported for many plants, including *P. euphratica* seedlings (Bogeat-Triboulot
377 et al., 2007), that root-shoot allocation responds significantly only to severe environmental stresses
378 (Poorter et al., 2012). A general explanation is that plants maintain their above-ground growth for
379 as long as possible under moderate soil environmental stresses to keep aboveground

380 competitiveness (Padilla et al., 2009; Poorter et al., 2012), but how plants tackle moderate
381 belowground resources stress to maintain shoot growth is still unclear. Our results indicate that *P.*
382 *euphratica* seedlings are able to adapt to moderate drought stress mainly via phenotypic
383 adjustments within their root systems. Here, we could raise an assumption that before root-shoot
384 allocation strongly responds to severe drought (Poorter et al., 2012), change in root architecture
385 mainly shoulder the absent role of root-shoot allocation for drought-suffered plant. This hypothesis
386 is based on a premise that plant phenotypic changes at the organ-level are more carbon economical
387 than that at the individual-level, while the latter should be more water economical than the former
388 for drought adaptation. To confirm this hypothesis, more functionally-different species need to be
389 further studied.

390

391 **CONCLUSIONS**

392 (1) *Populus euphratica* seedlings showed a conservative resource-use strategy in response to
393 drought stress, evidenced by thicker and shorter distal roots with lower SRL, sparser lateral
394 root branching, and herringbone-like root architecture under drought.

395 (2) The absolute root depth of *P. euphratica* seedlings was strongly constrained by drought, but
396 this negative effect was alleviated by changes in root-shoot allocation, root architecture, and
397 taproot morphology, resulting in a significant increase in relative root depth.

398 (3) Root architectural changes and root-shoot allocation dominated in order to achieve deep
399 rooting under drought conditions, while the role of taproot morphological changes was
400 relatively minor. Interestingly, their relative contributions to deep rooting varied with drought
401 intensity. Under moderate drought conditions, root architectural changes exerted a
402 predominant effect on increased relative root depth, but under severe drought, root-shoot
403 allocation and root architecture played equally important roles.

404 ACKNOWLEDGEMENTS

405 We are profoundly thankful to Ejin Banner State Forest Farm for support to conduct this
406 experiment, and to our colleague Liu W., Dong. F.Y. and Zhong Y.M. for their assistance in the
407 field and indoors. We also thank the public infrastructure laboratory in the College of Forestry of
408 Beijing Forestry University for their support of instruments.

409

410 **REFERENCES**

- 411 **Alameda D, Villar R. 2012.** Linking root traits to plant physiology and growth in *Fraxinus*
412 *angustifolia*, vahl. seedlings under soil compaction conditions. *Environmental & Experimental*
413 *Botany* **79**:49-57 DOI 10.1016/j.envexpbot.2012.01.004.
- 414 **Bauerle TL, Smart DR, Bauerle WL, Stockert C, Eissenstat DM. 2008.** Root foraging in
415 response to heterogeneous soil moisture in two grapevines that differ in potential growth rate.
416 *New Phytologist* **179**:857-866 DOI 10.1111/j.1469-8137.2008.02489.x.
- 417 **Bengough AG, Bransby MF, Hans J, McKenna SJ, Roberts TJ, Valentine TA. 2005.** Root
418 responses to soil physical conditions: growth dynamics from field to cell. *Journal of*
419 *Experimental Botany* **57**:437-447 DOI 10.1093/jxb/erj003.
- 420 **Bogeat-Triboulot MB, Brosche M, Renaut J, Jouve L, Le Thiec D, Fayyaz P, Vinocur B,**
421 **Witters E, Laukens K, Teichmann T, Altman A, Hausman JF, Polle A, Kangasjärvi J,**
422 **Dreyer E. 2007.** Gradual soil water depletion results in reversible changes of gene expression,
423 protein profiles, ecophysiology, and growth performance in *Populus euphratica*, a poplar
424 growing in arid regions. *Plant Physiology* **143**:876-892 DOI 10.1104/pp.106.088708.
- 425 **Browicz K. 1977.** Chorology of *Populus euphratica* Olivier. *Arboretum Kornickie* **22**:5-27.
- 426 **Brunner I, Herzog C, Dawes MA, Arend M, Sperisen C. 2015.** How tree roots respond to
427 drought. *Frontiers in Plant Science* **6**:547 DOI 10.3389/fpls.2015.00547.
- 428 **Cao D, Li J, Huang Z, Baskin CC, Baskin JM, Hao P, Zhou W, Li J. 2012.** Reproductive
429 characteristics of a *Populus euphratica* population and prospects for its restoration in China.
430 *Plos One* **7(7)**: e39121 DOI 10.1371/journal.pone.0039121.
- 431 **Chapin FS III. 1991.** Integrated responses of plants to stress. *BioScience* **41**: 29-36 DOI:
432 10.2307/1311538
- 433 **Cortina J, Green JJ, Baddeley JA, Watson CA. 2008.** Root morphology and water transport of
434 *Pistacia lentiscus* seedlings under contrasting water supply: a test of the pipe stem theory.
435 *Environmental & Experimental Botany* **62**:343-350 DOI 10.1016/j.envexpbot.2007.10.007.
- 436 **Fitter AH. 1985.** Functional significance of root morphology and root system architecture. In:

- 437 Fitter AH, Read DJ, Atkinson D, Usher MB, eds. *Ecological Interactions in Soil*. Oxford, UK:
438 Blackwell Scientific Publications, 87-106.
- 439 **Fitter AH. 1987.** An architectural approach to the comparative ecology of plant root systems. *New*
440 *Phytologist* **106**:61-77 DOI 10.1111/j.1469-8137.1987.tb04683.x.
- 441 **Fitter AH, Stickland TR, Harvey ML, Wilson GW. 1991.** Architectural analysis of plant root
442 systems 1. Architectural correlates of exploitation efficiency. *New Phytologist* **118**:375-382.
443 DOI 10.1111/j.1469-8137.1992.tb01110.x.
- 444 **Fitter AH, Stickland TR. 1992.** Fractal characterization of root system architecture. *Functional*
445 *Ecology* **6**:632-635 DOI 10.2307/2389956.
- 446 **Freschet GT, Swart EM, Cornelissen JHC. 2015.** Integrated plant phenotypic responses to
447 contrasting above- and below-ground resources: key roles of specific leaf area and root mass
448 fraction. *New Phytologist* **206(4)**:1247-1260 DOI 10.1111/nph.13352.
- 449 **Freschet GT, Violle C, Bourget MY, Scherer-Lorenzen M, Fort F. 2018.** Allocation,
450 morphology, physiology, architecture: the multiple facets of plant above- and below-ground
451 responses to resource stress. *New Phytologist*. **219**:4 DOI 10.1111/nph.15225.
- 452 **Glimskär A. 2000.** Estimates of root system topology of five plant species grown at steady-state
453 nutrition. *Plant & Soil* **227**:249-256 DOI 10.1023/a: 1026531200864.
- 454 **Gries D, Zeng F, Foetzki A, Arndt SK, Bruelheide H, Thomas FM, Zhang X, Runge M. 2003.**
455 Growth and water relations of *Tamarix ramosissima*, and *Populus euphratica*, on Taklamakan
456 desert dunes in relation to depth to a permanent water table. *Plant, Cell & Environment* **26**:725-
457 736 DOI 10.1046/j.1365-3040.2003.01009.x.
- 458 **Harper J, Jones M, Sackville HN. 1991.** The evolution of roots and the problems of analysing
459 their behaviour. In: Atkinson D, ed. *Plant Root Growth: An Ecological Perspective*. Oxford,
460 UK: Blackwell Scientific Publications, 3-22.
- 461 **Ho MD, Rosas JC, Brown KM, Lynch JP. 2005.** Root architectural tradeoffs for water and
462 phosphorus acquisition. *Functional Plant Biology* **32**:737-748 DOI 10.1071/FP05043.
- 463 **Hukin D, Cochard H, Dreyer E, Thiec DL, Bogeat-Triboulot MB. 2005.** Cavitation

- 464 vulnerability in roots and shoots: Does *Populus euphratica* Oliv., a poplar from arid areas of
465 Central Asia, differ from other poplar species? *Journal of Experimental Botany* **56(418)**:2003-
466 2010 DOI 10.1093/jxb/eri198.
- 467 **Kramer-walter KR, Laughlin DC. 2017.** Root nutrient concentration and biomass allocation are
468 more plastic than morphological traits in response to nutrient limitation. *Plant & Soil* **416(1-**
469 **2)**:1-12 DOI 10.1007/s11104-017-3234-9
- 470 **Kong D, Ma C, Zhang Q, Li L, Chen X, Zeng H, Guo D. 2014.** Leading dimensions in
471 absorptive root trait variation across 96 subtropical forest species. *New Phytologist* **203**:863-
472 72 DOI 10.1111/nph.12842.
- 473 **Lambers H, Atkin O, Millenaar FF. 2002.** Respiratory patterns in roots in relation to their
474 functioning. In: Waisel Y, Eshel A, Kafkaki K. eds. *Plant roots: the Hidden Half*. New York,
475 USA: Marcel Dekker, 521-552.
- 476 **Leon MF, Squeo FA, Gutierrez JR, Holmgren M. 2011.** Rapid root extension during water
477 pulses enhances establishment of shrub seedlings in the Atacama Desert. *Journal of Vegetation*
478 *Science* **22(1)**:120-129 DOI 10.1111/j.1654-1103.2010.01224.x.
- 479 **Lü S, Zhang X, Zhang N, Xia Y, Jing L, Li J. 2015.** Response of root growth and architecture
480 of *Populus euphratica* seedling to changes of soil moisture. (in Chinese with English abstract)
481 *Acta Botanica Boreali-Occidentalia Sinica* **35**:1005-1012.
- 482 **Lynch JP. 1995.** Root architecture and plant productivity. *Plant Physiology* **109**:7-13 DOI
483 10.1104/pp.109.1.7.
- 484 **Lynch JP. 2005.** Root architecture and nutrient acquisition. In: Bassirirad H, ed. *Nutrient*
485 *Acquisition by Plants: An Ecological Perspective*. Berlin, Germany: Springer, 147-183.
- 486 **Markestijn L, Poorter L. 2009.** Seedling root morphology and biomass allocation of 62 tropical
487 tree species in relation to drought- and shade-tolerance. *Journal of Ecology* **97**:311-325 DOI
488 10.1111/j.1365-2745.2008.01466.x.
- 489 **Muller B, Pantin F, Génard M, Turc O, Freixes S, Piques M, Gibon Y. 2011.** Water deficits
490 uncouple growth from photosynthesis, increase C content, and modify the relationships

- 491 between C and growth in sink organs. *Journal of Experimental Botany* **62**:1715-1729 DOI
492 10.1093/jxb/erq438.
- 493 **McCormack ML, Adams TS, Smithwick EH, Eissenstat DM. 2012.** Predicting fine root
494 lifespan from plant functional traits in temperate trees. *New Phytologist* **195**:823-831. DOI
495 10.1111/j.1469-8137.2012.04198.x.
- 496 **McCormack ML, Guo D, Iversen CM, Chen WL, Eissenstat DM, Fernandez CW, Li L, Ma
497 C, Ma Z, Poorter H, Reich PB, Zadworny M, Zanne A. 2017.** Building a better foundation:
498 improving root-trait measurements to understand and model plant and ecosystem processes.
499 *New Phytologist* **215(1)**:27-37 DOI 10.1111/nph.14459.
- 500 **Nicotra AB, Westoby M. 2002.** Seedling root anatomy and morphology: An examination of
501 ecological differentiation with rainfall using phylogenetically independent contrasts.
502 *Oecologia* **130(1)**:136-145 DOI 10.1007/s004420100788.
- 503 **Nicotra AB, Atkin OK, Bonser SP, Davidson AM, Finnegan EJ, Mathesius U, Poot P,
504 Purugganan MD, Richards CL, Valladares F, van Kleunen M. 2010.** Plant phenotypic
505 plasticity in a changing climate. *Trends in Plant Science* **15**: 684-692 DOI
506 10.1016/j.tplants.2010.09.008
- 507 **Olmo M, Lopez-Iglesias B, Villar R. 2014.** Drought changes the structure and elemental
508 composition of very fine roots in seedlings of ten woody tree species, implications for a drier
509 climate. *Plant & Soil* **384**:113-129 DOI 10.1007/s11104-014-2178-6.
- 510 **Padilla FM, Pugnaire FI. 2007.** Rooting depth and soil moisture control Mediterranean woody
511 seedling survival during drought. *Functional Ecology* **21**:489-495 DOI 10.1111/j.1365-
512 2435.2007.01267.x.
- 513 **Padilla FM, Miranda JD, Jorquera MJ, Pugnaire FI. 2009.** Variability in amount and
514 frequency of water supply affects roots but not growth of arid shrubs. *Plant Ecology* **204**:261-
515 270 DOI 10.1007/s11258-009-9589-0.
- 516 **Paula S, Pausas JG. 2011.** Root traits explain different foraging strategies between resprouting
517 life histories. *Oecologia* **165(2)**: 321-331 DOI 10.1007/s00442-010-1806-y.

- 518 **Poorter H, Niklas KJ, Reich PB, Oleksyn J, Poot P, Mommer L. 2012.** Biomass allocation to
519 leaves, stems and roots: Meta-analyses of interspecific variation and environmental control.
520 *New Phytologist* **193**:30-50 DOI 10.1111/j.1469-8137.2011.03952.x.
- 521 **Postma JA, Dathe A, Lynch JP. 2014.** The optimal lateral root branching density for maize
522 depends on nitrogen and phosphorus availability. *Plant Physiology* **166(2)**:590-602 DOI
523 10.1104/pp.113.233916.
- 524 **Rees M, Osborne CP, Woodward FI, Hulme SP, Turnbull LA, Taylor SH. 2010.** Partitioning
525 the components of relative growth rate: how important is plant size variation. *American*
526 *Naturalist* **176(6)**:E152 DOI 10.1086/657037.
- 527 **Ryser P, Lambers H. 1995.** Root and leaf attributes accounting for the performance of fast- and
528 slow-growing grasses at different nutrient supply. *Plant & Soil* **170**:251-265 DOI
529 10.1007/BF00010478.
- 530 **Sultan SE. 2000.** Phenotypic plasticity for plant development, function and life history. *Trends in*
531 *Plant Science* **5(12)**:537-542 DOI 10.1016/S1360-1385(00)01797-0.
- 532 **Schenk HJ, Jackson RB. 2002.** Rooting depths, lateral root spreads and
533 below-ground/above-ground allometries of plants in water-limited ecosystems. *Journal of*
534 *Ecology* **90**:480-494 DOI 10.1046/j.1365-2745.2002.00682.x.
- 535 **Stella JC, Battles JJ, McBride JR, Orr BK. 2010.** Riparian seedling mortality from simulated
536 water table recession, and the design of sustainable flow regimes on regulated rivers.
537 *Restoration Ecology* **18**:284-294 DOI 10.1111/j.1526-100X.2010.00651.x.
- 538 **Šmilauerová M, Šmilauer P. 2002.** Morphological responses of plant roots to heterogeneity of
539 soil resources. *New Phytologist* **154(3)**:703-715 DOI 10.1046/j.1469-8137.2002.00416.x.
- 540 **Taub DR, Goldberg D. 1996.** Root system topology of plants from habitats differing in soil
541 resource availability. *Functional Ecology* **10**:258-264 DOI 10.2307/2389851.
- 542 **Thevs N, Zerbe S, Peper J, Succow M. 2008.** Vegetation and vegetation dynamics in the Tarim
543 River floodplain of continental-arid Xinjiang, NW China. *Phytocoenologia* **38(2)**:65-84 DOI
544 10.1127/0340-269X/2008/0038-0065.

- 545 **Trubat R, Cortina J, Vilagrosa A. 2006.** Plant morphology and root hydraulics are altered by
546 nutrient deficiency in *Pistacia lentiscus* (L.). *Trees* **20**:334-339 DOI 10.1007/s00468-005-
547 0045-z.
- 548 **Wang L, Zhao C, Li J, Liu Z, Wang J. 2015.** Root plasticity of *Populus euphratica* seedlings in
549 response to different water table depths and contrasting sediment types. *PloS One* **10**:e0118691
550 DOI 10.1371/journal.pone.0118691.
- 551 **Wang S. 1996.** The Status, conservation and recovery of global resources of *Populus euphratica*.
552 (in Chinese). *World Forestry Research* **6**:37-44 DOI 10.13348/j.cnki.sjlyyj. 1996.06.005.
- 553 **Weemstra M, Mommer L, Visser EJW, Ruijven J, Kuyper TW, Mohren GMJ, Sterck FJ.**
554 **2016.** Towards a multidimensional root trait framework: A tree root review. *New Phytologist*
555 **211(4)**:1159-1169 DOI 10.1111/nph.14003.
- 556 **Wiehle M, Eusemann P, Thevs N, Schnittler M. 2009.** Root suckering patterns in *Populus*
557 *euphratica*, (Euphrates poplar, Salicaceae). *Trees* **23(5)**:991-1001 DOI 10.1007/s00468-009-
558 0341-0.
- 559 **Zerbe S, Thevs N. 2011.** Restoring central Asian floodplain ecosystems as natural capital and
560 cultural heritage in a continental desert environment. In: Hong SK, ed. Landscape Ecology in
561 Asian Cultures. *Ecological Research Monographs*. Tokyo, Japan: Springer 277-297.
- 562 **Zhu Y, Ren L, Skaggs TH, Lü H, Yu Z, Wu Y, Fang X. 2009.** Simulation of *Populus euphratica*
563 root uptake of groundwater in an arid woodland of the Ejina Basin, China. *Hydrology Process*
564 **23**:2460-2469 DOI 10.1002/hyp.7353.

Figure 1

Schematic representation of partitioning the total drought effect into environmental limitation of growth and different levels of root phenotypic responses

(A.) A scheme to increasing a certain root length through three levels of root phenotypic changes respectively. To achieve an increase of certain root length, plant can phenotypically adjust at three levels respectively, but most commonly, plant roots adjust at all three levels when they respond to drought. (B.) Drought exerts considerable influence in metabolism, causing an overall limitation of growth. (C.) Real growth situation is a syndrome from environmental limitation and multiple facets of integrated phenotypic responses

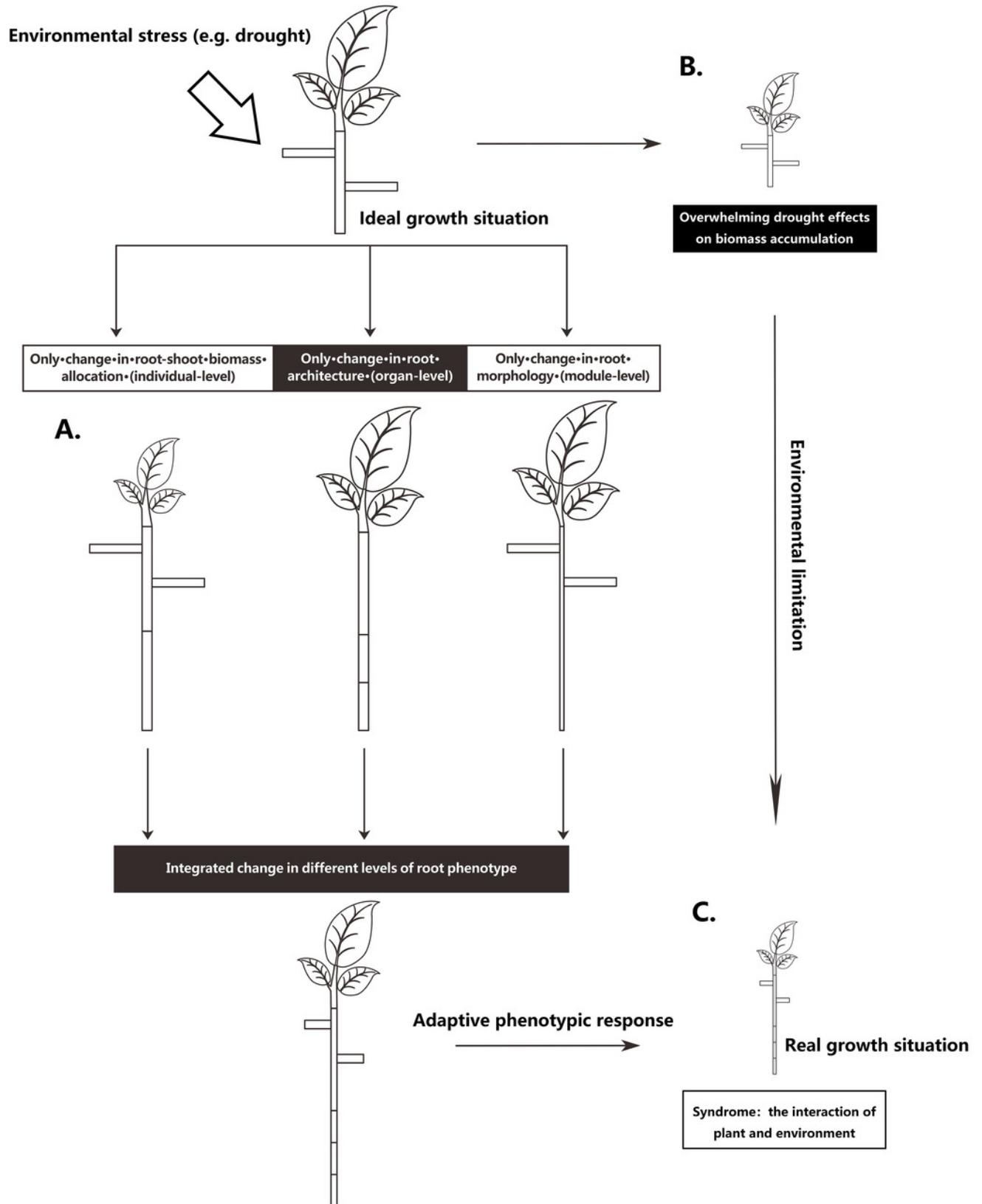


Figure 2

Representative scan images used in the study to illustrate changes in root system of *P. euphratica* seedlings with drought treatments

The representative root images of *P. euphratica* seedlings show the relatively denser branch, deeper root depth and more herringbone-like branching pattern of root with drought increasing. (A) & (B) show the root images from the seedlings under OW, in which their TI=0.697, DBI=0.056, and TI=0.650, DBI= 0.032, respectively. (C) & (D) from the seedlings under OW, in which their TI=0.702, DBI=0.074, and TI=0.702, DBI=0.086, respectively. (E) & (F) from seedlings under SD in which their TI=0.754, DBI=0.185, and TI=0.773, DBI=0.258, respectively. **Note:** OW: optimal water content; MD: moderate drought stress; SD: severe drought stress. DBI is the dichotomous branching index and TI is the topology index, and both of them are used to quantify root architectural changes (for detailed definition see table 1). A DBI of 0 is characteristic of a perfectly dichotomous branching structure and similarly, a TI close to 0.5 means a dichotomous-like branching structure. Contrastly, both a DBI of 1 and a TI of 1 represents a perfectly herringbone branching structure.

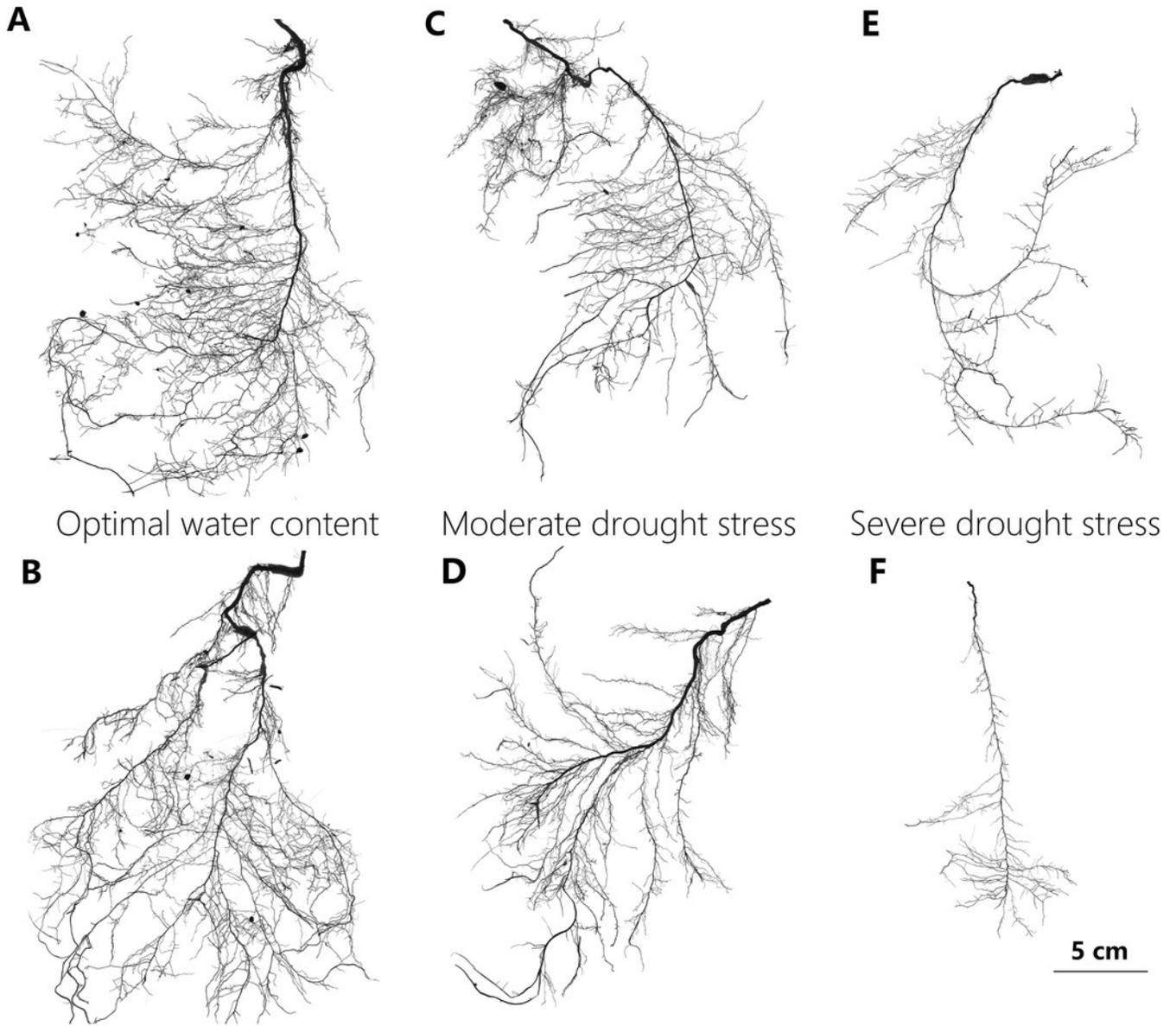


Figure 3

Relative contributions of root-shoot allocation (RMF, light grey bars), morphology (STRL, dark grey bars) and architecture (TRMF, black bars) variables to the variation in relative root depth (RRD).

From left to right, the first bar represents total contributions to the variation in RRD between OW and MD, the second bar represents that between OW and SD, and the third bar represents that between MD and SD. OW: optimal water content (control); MD: moderate drought stress; SD: severe drought stress.

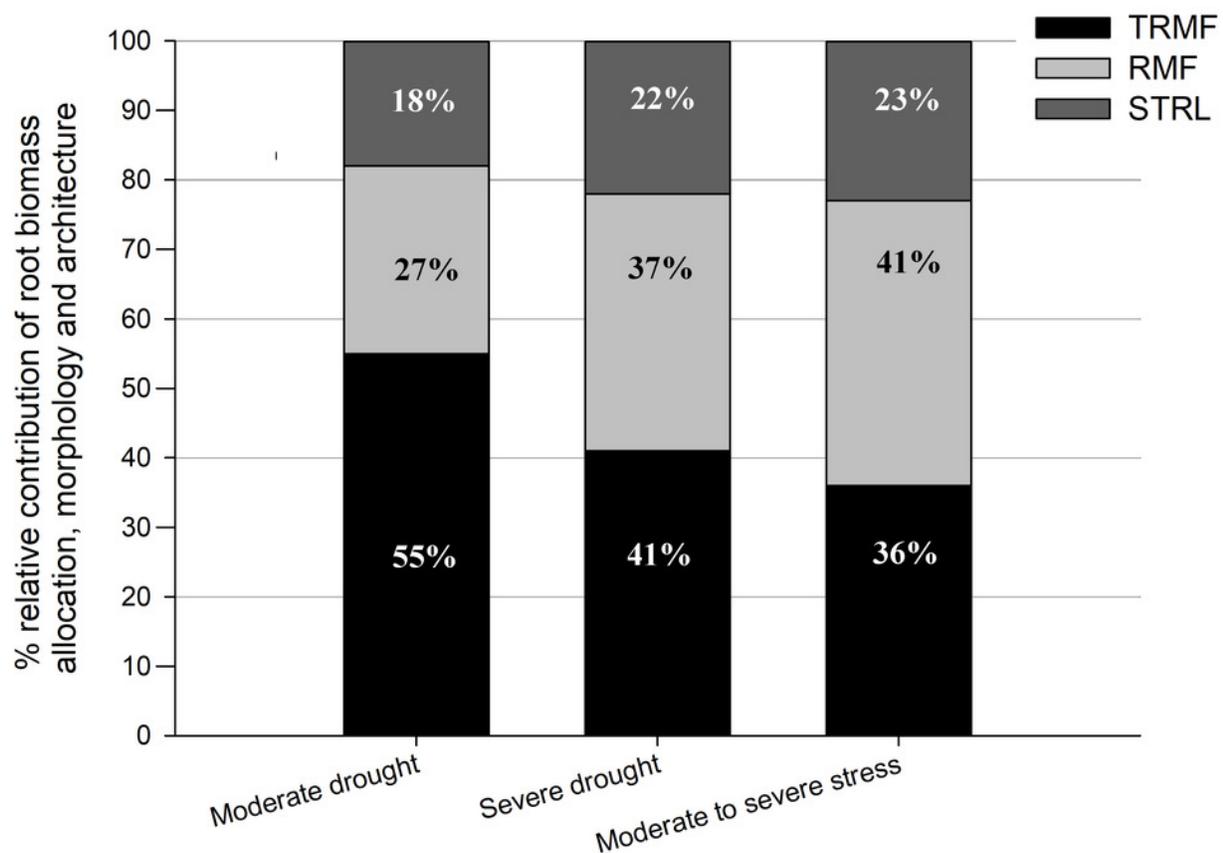


Table 1 (on next page)

Calculated root variables and its abbreviations, units, description and functional role

1 **Table 1 Calculated root variables and its abbreviations, units, description and functional role**

Root variables (Abbrev.) units	Description	Functional role
Relative root depth (<i>RRD</i>) cm taproot [g plant] ⁻¹	Taproot length per total plant dry mass	Capacity for deep rooting.
Root mass fraction (<i>RMF</i>) g root [g plant] ⁻¹	The proportion of total plant mass allocated to roots	Total plant resources allocated to root functions (e.g. absorption, transport, foraging, anchoring) (Markesteyn & Poorter, 2009)
Taproot mass fraction (<i>TRMF</i>) g taproot [g root] ⁻¹	The proportion of total root mass allocated to taproot	Total root resources allocated to the taproot functions.(e.g., anchoring, foraging, transport)
Specific taproot length (<i>STRL</i>) cm taproot[g taproot] ⁻¹	Taproot length per amount of taproot biomass invested	The efficiency of taproot resources used to deep rooting.
Lateral root branching density (<i>LRBD</i>) number lateral root[cm taproot] ⁻¹	Lateral roots branching number per unit of taproot length	Potential capacity of exploration and exploitation to horizontal soil resources
Topology Index (<i>TI</i>)	$TI = \log_{10}(a)/\log_{10}(\mu)$; altitude a is number of links in the longest path from base to tips; magnitude μ is number of external links or the number of root tips (Glimskär, 2000)	TI vary from 1 close to 0.5, and DBI vary between 0 and 1, both with large values indicative of a more herringbone-like root system that are thought to be more efficient at intercepting mobile resources , such as water, by extensive soil exploration, contrasted with dichotomous-like systems that are better at acquiring immobile resources, such as phosphorus by intensive soil exploration (Fitter, 1987)
Dichotomous branching index (<i>DBI</i>)	$DBI = [p_e - \min(p_e)] / [\max(p_e) - \min(p_e)]$; p_e is the sum of the number of links in all paths from each external link to the base link; $\max(p_e)$ and $\min(p_e)$ respectively is the theoretical external path length for a system of given magnitude that has a completely herringbone and dichotomous topology, for detailed calculation see Šmilauerová & Šmilauer (2002)	

2

Table 2 (on next page)

Mean values of biomass variables, root branching variables and root morphological variables of *P. euphratica* seedlings at 110 days under drought treatments

Notes: Differences in variables among treatments were tested using factorial analysis of variance (ANOVA). Values are means \pm standard error (OW: n=25; MD: n=28; SD: n=28). In each row, means followed by different letters are significantly different ($P < 0.05$). OW, MD and SD refers to optimal water content (control), moderate drought stress and severe drought stress separately. SRL=Specific Root Length; SRDL=Specific Distal Root Length; TI=Topology Index; DBI=Dichotomous Branching Index.

1 **Table 2 Mean values of biomass variables, root branching variables and root morphological**
 2 **variables of *P. euphratica* seedlings at 110 days under drought treatments**

	Variables	OW (control)	MD	SD
Biomass	Total plant biomass (g)	0.46±0.02 ^a	0.35±0.02 ^b	0.22±0.02 ^c
	Above-ground biomass (g)	0.30±0.02 ^a	0.23±0.02 ^b	0.13±0.01 ^c
	Below-ground biomass (g)	0.16±0.01 ^a	0.12±0.01 ^b	0.09±0.01 ^c
	Taproot biomass (g)	0.043±0.003 ^a	0.032±0.002 ^b	0.030±0.002 ^b
Morphology	Total root length (cm)	662±37 ^a	516±44 ^b	393±41 ^c
	Average root length (cm)	0.376±0.048 ^a	0.367±0.038 ^a	0.236±0.027 ^a
	Root diameter (mm)	0.348±0.007 ^a	0.357±0.006 ^a	0.359±0.009 ^a
	SRL (cm/g)	4766±290 ^a	4846±357 ^a	5141±343 ^a
	Average distal root length (cm)	0.319±0.122 ^a	0.279±0.191 ^b	0.156±0.057 ^c
	Distal root diameter (mm)	0.278±0.006 ^c	0.302±0.007 ^b	0.337±0.007 ^a
	SDRL(cm/g)	10032±425 ^a	9487±661 ^{ab}	7843±361 ^b
	Average lateral root length (cm)	5.113±0.324 ^a	4.680±0.333 ^{ab}	3.765±0.298 ^c
	Lateral root diameter (mm)	0.881±0.035 ^a	0.757±0.034 ^b	0.653±0.029 ^c
	Taproot length (cm)	35.3±1.1 ^a	27.3±0.8 ^b	26.2±0.7 ^b
Taproot diameter (mm)	1.60±0.091 ^a	1.48±0.093 ^a	1.35±0.082 ^a	
Architecture	TI	0.713±0.007 ^b	0.731±0.008 ^{ab}	0.757±0.010 ^a
	DBI	0.101±0.010 ^b	0.153±0.016 ^{ab}	0.216±0.020 ^a
	Lateral root branching density (n/cm)	1.21±0.09 ^a	1.03±0.05 ^b	0.85±0.04 ^c

3 Notes: Differences in variables among treatments were tested using factorial analysis of variance
 4 (ANOVA). Values are means± standard error (OW: n=25; MD: n=28; SD: n=28). In each row,
 5 means followed by different letters are significantly different (P<0.05). OW, MD and SD refers to
 6 optimal water content (control), moderate drought stress and severe drought stress separately.
 7 SRL=Specific Root Length; SRDL=Specific Distal Root Length; TI=Topology Index;
 8 DBI=Dichotomous Branching Index.

Table 3(on next page)

Representative indices of different root phenotypic changes and deep-rooting capacity of *P. euphratica* seedlings under drought treatments

Notes: Differences in Indices among treatments were tested using factorial analysis of variance (ANOVA). Values are means \pm standard error (OW: n=25; MD: n=28; SD: n=28). Within a row, means followed by different letters are significantly different (P<0.05). OW, MD and SD refers to optimal water content (control), moderate drought stress and severe drought stress respectively. RMF=Root Mass Fraction; TRMF=Taproot Mass Fraction; STRL=Specific Taproot Length; RRD=Relative Root Depth.

1 **Table 3 Representative indices of different root phenotypic changes and deep-rooting**
 2 **capacity of *P. euphratica* seedlings under drought treatments**

Indices	OW (control)	MD	SD
RMF	0.338±0.009 ^b	0.353±0.013 ^b	0.418±0.013 ^a
TRMF	0.304±0.018 ^b	0.333±0.026 ^{ab}	0.393±0.030 ^a
STRL (cm/g)	889±37 ^a	919±45 ^a	1020±67 ^a
RRD (cm/g)	90.8±7.26 ^b	108.0±11.0 ^{ab}	166.0±15.9 ^a

3 Notes: Differences in Indices among treatments were tested using factorial analysis of variance
 4 (ANOVA). Values are means± standard error (OW: n=25; MD: n=28; SD: n=28). Within a row,
 5 means followed by different letters are significantly different (P<0.05). OW, MD and SD refer to
 6 optimal water content (control), moderate drought stress and severe drought stress respectively.
 7 RMF=Root Mass Fraction; TRMF=Taproot Mass Fraction; STRL=Specific Taproot Length;
 8 RRD=Relative Root Depth.