

Quantitative heterodonty in Crocodylia: variability and decoupling in size and shape across modern and extinct taxa. (#29741)

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Quantitative heterodonty in Crocodylia: variability and decoupling in size and shape across modern and extinct taxa.

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Heterodonty in Crocodylia has not been defined quantitatively, as the teeth rarely have been measured. This has resulted in a range of qualitative descriptors, with little consensus on the state of dental morphology in the clade. The purpose of this study was to quantify both shape and size morphology along the tooth row in a multispecific sample of both extant and extinct members Crocodylia using geometric morphometrics. Data were collected from several crown crocodylian (and one peirosaurid) dry skeletal and fossil specimens. Digital photographs were taken of each tooth, and the margins of the tooth were converted into landmarks/semilandmarks. Heterodonty was expressed as Foote's morphological disparity, and a principal components analysis visualized shape variance. All living crocodylians are heterodont to varying degrees. The majority of the shape variance was represented by a 'caniniform' to 'molariform' transition. Heterodonty varied significantly between positions; size undulated whereas shape was significantly linear from mesial to distal, suggesting they are developmentally decoupled. Snout shape correlated to tooth shape but not size, with longirostrine taxa having more caniniform teeth and brevirostrine having more molariform. Caimanines and *Crocodylus porosus* had high size-heterodonty with large pseudocanines, reflecting a prioritization of securing struggling, compliant prey. *Alligator sinensis* and *Osteolaemus tetraspis* had large, highly molariform, distal teeth: a consequence of high-frequency durophagy combined with prey size. *Gavialis gengeticus*, *Mecistops cataphractus* and *Tomistoma schlegelii* had a caniniform arcade with low heterodonty, almost exclusively for capturing small underwater prey with minimal processing. *Alligator prenasalis*, *Brachychampsa sp.*, and "*Crocodylus*" *affinis* were primarily molariform. *Leidyosuchus* and *Borealosuchus* tooth sizes and shapes typical of modern *Alligator* and *Crocodylus* respectively, although the former may have been less durophagous. *Hamadasuchus rebouli* similarities to *C. porosus* indicate it may have dealt with vertebrate prey, but prey processing may have been different due to its terrestrial

habitat. Although Foote's disparity is a reliable method for assessing heterodonty, incomplete tooth rows may give inflated/deflated values. Our methods may be very useful for predicting the shape of missing teeth in fossil taxa, as well as inferring dietary behaviors and other life history characters.

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13 ABSTRACT

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 26 developmentally decoupled. Snout shape correlated to tooth shape but not size, with
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Keywords: Caniniform, Crown, Dentition, Diet, Geometric morphometrics, Molariform, Semilandmarks

INTRODUCTION

What constitutes heterodonty often seems like a moving target, with different qualitative definitions in place depending on the clade being studied (Shimada, 2001). In 1993, Kieser et al. referred to the exact definition of the term heterodont as "a bone of contention." Since that time, the issue never has been fully resolved. Nowhere is this lack of clarity more pronounced than within the non-mammalian vertebrate literature, especially the members of Crocodylia. Traditionally crocodylians were considered homodont, due to a lack of discrete dental categories or variability in cusp number as in mammals (Peyer, 1968; Langston, 1973; Osborn, 1998; Larsson & Sidor, 1999; Zahradnicek et al., 2014). Ferguson (1981) refers to *Alligator mississippiensis* as "pseudoheterodont," because it showed a gradual, as opposed to punctuated, change in tooth shape along the tooth row (see also Grigg and Gans, 1993; Kieser et al., 1993; Hendrickx, Mateus, & Araújo, 2015b). Size changes have motivated the term

56 “heterometric homodonty” for *Crocodylus niloticus* (Fruchard, 2012). To add to the ambiguity,
 57 certain fossil crocodylians, especially those taxa that are interpreted as herbivores or
 58 omnivores, are called “heterodont” in order to distinguish their dentition from their modern
 59 relatives (e.g. Martin, 2007; Ősi, Clark, & Weishampel, 2007; Novas et al., 2009). 

60 Semantics aside, one reason for the lack of resolution concerning crocodylian
 61 heterodonty is that their teeth have rarely been measured. There have been few studies
 62 looking at morphometrics of crocodylian teeth, and, of those studies, measurements have been
 63 Euclidean distances taken for determining replacement rates (Frey & Monninger, 2010;
 64 Bennett, 2012) or for biomechanical analyses (Monfroy, 2017). Aside from a single study
 65 measuring Euclidean distances in a fossil notosuchian (Lecuona & Pol, 2008), little research has
 66 quantitatively investigated heterodonty either within or between species. Typically, crocodylian
 67 dentition is described qualitatively, with the goal of fossil identification, characterization for
 68 phylogenetic analysis, or paleoecological inference (e.g. Schwarz-Wings, Rees, & Lindgren,
 69 2009; Young et al., 2012; Salas-Gismondi et al., 2015; Adams, Noto, & Drumheller, 2017).

 Qualitative descriptors of crocodylian tooth morphology are numerous, and include terms such
 71 as blunt, bulbous, broadened, button-shaped, conical, globular, fang, kidney-shaped,
 72 lanceolate, needle-like, robust, short, slender, spike-like, and thick (e.g. Brazaitis, 1973;
 73 Groombridge, 1982; Aoki, 1989; Brochu, 1999; Erickson, Lappin, & Vliet, 2003; Ősi, Clark, &
 74 Weishampel, 2007; Schwarz-Wings, Rees, & Lindgren, 2009; Fruchard, 2012; Salas-Gismondi et
 75 al., 2015; Berkovitz and Shellis, 2017).

Research typically has focused on crocodylian tooth development and function as opposed to quantitative morphology, including numerous studies on material characteristics (Shimada, Sato, & Moriyama, 1992; Erickson, 1996; Enax et al., 2013), implantation and replacement (Edmund, 1962; Westergaard and Ferguson, 1986, 1987, 1990; LeBlanc et al., 2017), dental wear (Ősi & Barrett, 2011), jaw musculature and kinematics (Iordansky, 1964; Van Drongelen & Dullemeijer, 1982; Busbey, 1989; Cleuren & De Vree, 1992; Endo et al., 2002), bite force (Cleuren, Aerts, & Vree, 1995; Erickson, Lappin, & Vliet, 2003; Erickson et al., 2012, 2014), death rolling (Fish et al., 2007; SK Drumheller, unpublished data), and taphonomic traces on bone surfaces (Njau ~~and~~ Blumenschine, 2006, 2012; Drumheller ~~and~~ Brochu, 2014; 2016).

Non-mammalian dental morphometrics has seen a burst of research in the past decade. Dinosaur teeth have probably received the most attention, with multiple studies using Euclidean distances for the identification of loose fossil crowns or to infer functional paleoecology (D'Amore, 2009; Larson & Currie, 2013; Buckley and Currie, 2014; Hendrickx and Mateus, 2014; Torices, Reichel, & Currie, 2014; Hendrickx, Mateus, & Araújo, 2015a, Gerke and Wings, 2016; Larson, Brown, & Evans, 2016). Extant reptiles have been investigated quantitatively as well, including colubrid snakes (Britt, Clark, & Bennett, 2009) and varanid lizards (D'Amore, 2015). Prior to this, lamniform sharks were investigated heavily (Shimada, 2002b, 2004; Shimada and Seigel, 2005). These morphometric analyses have shed light on the nature of heterodonty, dental allometry, and ecomorphology in non-mammalian vertebrates, and these methods may be applied to Crocodylia in the hopes to elaborate upon the state of heterodonty in this taxon.

The purpose of this study **was** to quantify both shape and size morphology along the tooth row in a multispecific sample of both extant and extinct members Crocodylia using geometric morphometrics. Our goals were to  introduce a multifaceted method for assessing **heterodonty** in a given crocodylian specimen; 2) document any developmental consistencies found within the clade as a whole; 3) distinguish crocodylian dental morphotypes between individuals; and 4) outline the advantages, limitations, and potential future uses of the method when analyzing crocodylian heterodonty.

MATERIALS AND METHODS

Nomenclature

 Crocodylian teeth have very few discrete homologous anatomical loci, but, because they exhibit thecodont dentition (Edmund, 1969), we defined them as having a crown with an apex, a neck, and a root within an alveolus. Nomenclature for tooth morphology used here was proposed by Smith & Dodson (2003): **mesial, towards the central premaxilla and mandibular symphysis; distal, away from the central premaxilla and mandibular symphysis; lingual, towards the tongue; labial, towards the lips; basal, towards the base of the tooth/alveolus; apical, away from the alveolus/towards the apex.** For simplicity the mesial-most teeth are referred to as ‘mesials’ and the distal-most as ‘distals.’

Tooth position was indicated by either the presence of a tooth or an empty alveolus in the host bone. Cranial teeth included premaxillary and maxillary teeth, and dentary teeth only **occurred** on the dentary bone. Teeth were numbered in ascending order from mesial to distal positions. Not all crocodylian specimens had the same **number of tooth positions**. Maxillary

118 teeth ranged from M1 to M11-M24, and dentary teeth ranged from D1 to D14-D26. All
 119 specimens were assumed to have 5 premaxillary positions (P1–P5). Members of *Paleosuchus*
 120 and *Osteolaemus* have only 4 premaxillary teeth during early stages of ontogeny (Brochu and
 121 Storrs, 2012; Narvaez et al., 2015), whereas several other species atrophy an alveolus (usually
 122 P2) as they grow (Webb & Messel, 1978; Brown et al., 2015; personal observation). In order to
 123 standardize, the procumbent teeth were always assigned P4 and the other teeth and atrophied
 124 positions were numbered based off of it.

125 Specimens

126  Data were collected from 25 extant, and 14 extinct, crown crocodylian specimens. In
 127 addition, we added the peirosaurid crocodylomorph *Hamadasuchus rebouli*. This resulting in a
 128 total of 24 species. From these we measured 1,224 teeth in total. For extant crocodylian taxa,
 129 data was collected from dry skeletal specimens from the American Museum of Natural History
 130 (AMNH). At least 40% of the tooth positions had to be represented by measureable teeth on
 131 the left and/or right for the specimen to be considered.  Specimens where the only teeth
 132 available biased towards the upper or lower size extremes were excluded. As ontogeny is
 133 beyond the scope of our present study, juveniles were avoided. Fossil taxa were sampled from
 134  the Royal Ontario Museum (ROM), the University of California Museum of Paleontology
 135 (UCMP), and the Smithsonian National Museum of Natural History (USNM). Completeness of
 136 tooth row was not considered in the inclusion of fossils. Although *Caiman crocodilus* is an
 137 extant species, a fossil specimen was  luded.

138 Data collection



Methods were similar to those proposed in D'Amore (2015). Teeth were photographed using either an Olympus Stylus or a Canon Rebel T3 EOS, alternating between a macro and an 18-55 mm lens depending on the size of the tooth, against a dark background with a scale (Figure 1A). Digital photographs were taken from the labial perspective, and a separate picture was taken for each tooth. Only fully erupted teeth with the neck visible were included. Tooth quality was variable in extant specimens. We included teeth with slightly worn apices, but excluded teeth that had large wear facets and chips as long as they interfered noticeably with the outline. Cracks down the long axis of the teeth were common, and were omitted if the crack distorted the shape of the tooth or resulted in a space where light could be seen from the other side.

We used a sliding semilandmark analysis (Bookstein 1997; Sheets, Kim, & Mitchell 2004; Zelditch et al., 2004; Mitteroecker et al., 2013) to derive shape measurements from the tooth's outline. Photographs were entered in TpsDig 2.16, and the margin of the tooth was traced using the curve drawing tool (Rohlf, 2010) (Figure 1B). Because the enamel margin was not always clear, each tooth was traced from apex to the point where the tooth ceases to taper on the neck for both the mesial and distal side. The two traced margins were then each transformed into 30 equidistant coordinates, and the apical coordinates were combined into one. This resulted in 3 landmarks and 56 semilandmarks (Figure 1C), the latter of which were then slid to minimize the bending energy (Perez, Bernal, & Gonzalez, 2006; Gunz & Mitteroecker, 2013) using TpsRelw 1.53 (Rohlf, 2013). This program also performed generalized least squares Procrustes superimposition on the data, and calculated centroid size (CS). Bilateral symmetry of



the jaw was assumed, and the superimposed coordinates and CS were averaged between teeth
from the same position on left and right sides.



Rostrum shape has long been considered both an important phylogenetic and ecomorphological feature in crocodylians (Busbey, 1995; Daniel & McHenry, 2000; Brochu, 2001; Sadleir & Makovicky, 2008; Salas-Gismondi et al., 2016). All specimens' skulls were photographed with a scale from the dorsal perspective. Snout shape data were derived following Drumheller et al., (2016) and Wilberg (2017), using a modified version of our technique for tooth outlines. We traced the skull margin from the rostral-most point of contact between the premaxillae to the caudal-most quadratojugal on each side in TpsDig, broke each margin into 50 equidistant coordinates, combined the anterior-most coordinate (resulting in 3 landmarks and 97 semilandmarks total), and slid them with TpsRelw again. Head length was derived from these landmarks, which began at the rostral-most landmark and ended in-between the posterior-most landmarks along the mid-sagittal plane. In specimens with damaged or missing bones on one side, bilateral symmetry was assumed and the intact side was mirrored. Head length was used as a measure of body size in lieu of snout-vent length, and all CS values were divided by it to normalize relative tooth size. *Allognathosuchus* sp. (UCMP 150180), *Argochampsa rebouli* (ROM 73872), and *Boverisuchus vorax* (UCMP 170767) did not have intact skulls associated with their dentition. Photographs of intact skulls of conspecifics



were used instead to derive head shape and length (substituted specimens included: Yale Peabody Museum 17111, Field Museum of Natural History PR399, and Office Chérifien des Phosphates DEK-GE 1201)

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182

183 Ordination and statistics

184 All analyses were conducted in MorphoJ v. 106d (Klingenberg 2011) and SPSS Version

 185 19.0 [IBM Corp, Armonk, NY]. A 10,000 permutations test was run on the Procrustes distance

186 between cranial and dentary teeth for extant specimens, and showed no significant differences

187 ($P = 0.2000$). Therefore, all subsequent analyses were conducted with both arcades combined

188 (unless position is being considered). Interspecific differences in mean tooth size and shape

189 were analyzed using an analyses of variance (ANOVA). Size was heteroscedastic according to

190 Levene's test ($P = 0.0002$), so we ran a Welch's ANOVA of  normalized CS values for all members

191 of each species in SPSS. For shape, Procrustes ANOVA was run in MorphoJ.

192 A singular measure of heterodonty was derived for each specimen in the form of

193 Foote's morphological disparity [$MD = (\sum_{i=1}^m D_i^2) / (m - 1)$] (Foote, 1993; Zelditch, Sheets, &

194 Fink, 2003; Sheets & Zelditch, 2013). Disparity (MD) is the sum of the differences of the values

195 of a given tooth (i) from the mean for all teeth from that specimen (Di) squared, with the

196 number of tooth positions (m) factored in. We calculated disparity for both cranial and dentary

197 teeth together from each specimen. For size heterodonty, Di is simply the difference in CS of a

198 tooth from the mean of the individual (Zelditch et al., 2004). Only specimens with intact skulls

199 for normalization were included  or shape Di is the Procrustes distance between the tooth and

200 the mean, and was calculated using DisparityBox7 (Sheets, 2012).

Although Foote's MD is an inclusive measure of heterodonty, it is not descriptive of multivariate measures such as shape. Therefore, principal components analyses (PCA) were conducted to visualize the degree of shape variance within all cranial and dentary teeth separately. All PCs representing over 5% of the variance were considered, and shape variance was visualized in vector diagrams (Figure 1D). Both CS values and PC scores were graphed as box plots for all members of each species, to visualize the range of shape and size that contribute to MD. Similar to teeth, a PCA was run for skull shape. Skull shape PCs were compared to the average tooth PC for each extant individual in a bivariate plot, and regressed to determine if skull shape and tooth shape are correlated.

Shape and size were plotted against tooth position to visualize variability along the tooth row. For size, CS was plotted against tooth position. For shape, the aforementioned tooth PC scores were plotted against tooth position. Data were clustered into box plots to depict trends along the dental arcade for each family within our extant sample. Each box represented a position. Note that these positions were not normalized, so specimens with more tooth positions will be the only occupants of the distal-most categories. As positional data were often heteroscedastic, Welch's ANOVAs were conducted to determine if shape and/or size were significantly different by position for each arcade in each family. Shape data were also regressed to determine if serial homology is linear along the tooth row. To standardize these regressions, tooth position was normalized into a percentage. (We numbered the positions along the tooth row starting with 1 at the mesial-most position, divided each by the total number of positions along the arcade, and then subtracted 0.5. This placed the y-intercept

222 halfway along the arcade). Included PCs were then regressed against position-percentage, and
 223 regression statistics were tabulated.

224 Results:

225 Shape variability in the sample

226  The majority of the shape variance was represented by the  principal component
227 (PC1). It accounted for over 91.26% of the variance, and is the only PC considered further.

228 When visualized, the axes indicated a 'caniniform' to 'molariform' transition with increased
229 values. The negative-most condition was an elongate, narrow crown coupled with a gentle
230 concavity on the distal margin. The positive-most values depicted an apical-basal shortening
231 and mesial-distal broadening (Figure 2), giving the tooth a stout, rounded crown with a
232 relatively narrow neck. Although PC1 was treated as a continuous variable in all subsequent

 analyses, we assigned names to ranges of PC scores for the purposes of description. These
234 descriptors include; high-caniniform (<-0.25); mid-caniniform (-0.25 to -0.15); low-caniniform ($-$
235 0.15 to -0.05); average (-0.05 to 0.05); low-molariform (0.05 to 0.15); mid-molariform ($0.15-$
236 0.25); and high-molariform (> 0.25).

237 Skull morphology:

238  Mean tooth shape showed  significant trends when regressed against head shape, but
239 not size (Figure 3). Head shape yielded a PC1 encompassing 89.81% of shape variance, and
 represented the brevirostrine-to-mesorostrine-to-longirostrine transition (broad-, middle-, and
241 slender-snouted conditions respectively) (Brochu, 2001; Wilberg, 2017). Significant regressions
242 scaled with a slope of -0.67 for extant-only crocodylians. Generally speaking, longirostrine taxa
243 had the most caniniform teeth and brevirostrine taxa had the most molariform teeth. Scatter
244 increases among the broader-snouted specimens below the mean, and the relationship is less

well represented. When extinct taxa were included, the slope decreased to just under -1.0. But the goodness of fit dropped from 0.73 to 0.52. Most of our fossil taxa had broader teeth relative to head shape than the extant sample, which was most likely a consequence of sampling bias (see DISCUSSION).

Footo's disparity and morphotype ranges

When means were compared, ANOVA indicated significant differences in CS between species [$F(13,674) = 6.389; P < 0.0001$]. Heterodonty varied both between and among species (Figure 4). *Alligator* tended to have lower size heterodonty than the caimanines, because the latter had more teeth above the inter-quartile range. Crocodyloids had highly variable size heterodonty. Several members of *Crocodylus* were middle ranged, but both *Crocodylus porosus* specimens rose above the rest. A member *O. tetraspis* was the most size-heterodont, but the others were much lower. This species had the largest teeth relatively speaking. *Mecistops cataphractus* and *Tomistoma schlegelii* were the least heterodont crocodyloids, and had the narrowest size ranges with the lowest maximum tooth sizes. *Gavialis gengeticus* was the least size heterodont in our modern data set, with similar size ranges to the longirostrine crocodyloids.

As with size, shape was also significantly different between species [$F(1482,79572) = 19.61; P < 0.0001$]. The caimanines and most members of *Crocodylus* had similar shape heterodonty, and possessed crowns ranging from mid-caniniform to mid-molariform (Figure 4). *Crocodylus siamensis* was more shape heterodont than its congeners. *Alligator* species had lower shape heterodonty than most caimanines, as their PC1 range was generally more

266 molariform. Unlike size,  *tetraspis* had shape heterodonty similar to members of *Crocodylus*.
 267 *Alligator sinensis*, *C. siamensis*, and *O. tetraspis* were the only modern taxa with teeth in high-
 268 molariform range. Similar to size, the longirostrine taxa have the least shape heterodonty.
 269 *Gavialis gangeticus* teeth all ranged from average to the high-caniniform. *Tomistoma schlegelii*
 270 had teeth ranging from high-caniniform to low-molariform range, whereas *M. cataphractus* was
 271 more constrained between mid-caniniform and low-molariform.

272 Concerning fossil taxa, *C. crocodilus* was similar to modern caimanines concerning
 273 heterodonty, although the maximum tooth size was lower (Figure 4). *Alligator prenasalis*,
 274 *Brachychampsa sp.*, and "*C. affinis*" had some of the lowest shape and size heterodonty in our
 275 sample. Tooth shape was limited from average to high-molariform, as they all lacked any
 276 disparately large teeth. *Leidyosuchus* heterodonty fell within the range of living alligatoroids,
 277 but the majority of the teeth were on the small end. *Borealosuchus sternbergii* had a range of
 278 tooth sizes typical of modern *Crocodylus*. Heterodonty was highly variable in this species.
 279 *Hamadasuchus rebouli* mirrored *C. porosus* in both size-ranges and size-heterodonty, but had
 280 higher shape heterodonty with more molariform teeth. *Allognathosuchus sp.* had the highest
 281 shape heterodonty by a large margin. *Argochampsa krebsi* and *B. vorax* both had low
 282 heterodonty, and were represented by only caniniform and molariform morphs respectively.

283 Heterodonty along the tooth row

284 In both extant Alligatoroidea and Crocodyloidea, size varied significantly between
 285 positions. Size undulated  ee times along the dental arcade resulting in significant differences
 286 between positions for both the cranium and mandible (Figure 5). Each undulation peaked with

a relatively large procumbent tooth (Gignac & Erickson, 2014). The largest procumbent teeth included P4 for both clades, and M4 for alligatoroids and M5 for crocodyloids (sensu Brochu and Storrs, 2012). Members of *Paleosuchus* has very large P3 and M3 teeth as well. A final undulation resulted in a procumbent tooth at M9-11. In alligatoroids this final peak relatively consistent throughout the group, but in crocodyloids these teeth varied greatly in size between specimens concerning both position and degree. Interspersed between these were small teeth, with the distal-most tooth often the smallest. The mandible was similar with procumbent D1 and D4, both followed by a series of small teeth. The third size-peak maximized between D11 and D14, reflecting similar levels of variability to its cranial counterpart. Gavialoids differed markedly by having the two mesial-most teeth enlarged, and the remainder of the teeth show a gradual decrease in size distally.

Alligatoroids and crocodyloids both showed a general linear trend concerning tooth shape, making positions significantly different (Figure 5). Mesial teeth ranged from mid-caniniform to average, with distals approaching high-molariform. In cranial teeth there was an apparent plateau from P1-M4, followed by a gradual increase in molariformy distally. Dentary teeth represented a more linear caniniform-to-molariform transition. Crocodyloids were much more varied than alligatoroids, with wider ranges and a series of low outliers (most of which are *T. schlegii*). In *G. gangeticus*, most of the teeth are high-caniniform with a steep increase towards average in the distal-most fifth of the arcade.

When each modern individual's PC1 values were regressed against position, the resultant linear trends represented individual shape heterodonty very well (Table 1). All regressions were significant, and all except *G. gangeticus* had goodnesses of fit greater than 70%. Both cranial and dentary tooth rows typically had slopes between 0.25-0.55. More shape heterodont taxa typically had greater slopes, where *C. siamensis* had the highest slopes of the sample. All the longirostrine taxa had the lowest intercepts indicating high caniniformy, and members of *Alligator* had high y-intercepts indicating general molariformy.

Fossil taxa displayed a much more variable range of size and shape trends along their arcades (Table 2). All fossil specimens with intact teeth in the position showed a procumbent P4, but "*C.*" *affinis* also had a similarly sized P3. Unlike other alligatoroids *L. canadensis* had both procumbent M4 and M5. Some of the largest teeth of *A. prenasalis*, *Brachychampsa*, and "*C.*" *affinis*, were the molariform distals. These teeth typically approached the size of the largest mesial crowns. Regressions for shape were significant for all fossil taxa except the "*C.*" *affinis* mandible (USNM 4048). The teeth of the fossil *Caiman crocodilus* had very similar sizes and shapes to those of modern *Caiman yacare* at the same positions. They only differed in that the coefficients of its cranial regression were dissimilar. All fossil specimens with intact teeth in the position showed a procumbent P4, but "*C.*" *affinis* also has a similarly sized P3. Unlike other alligatoroids *L. canadensis* has both procumbent M4 and M5. Some of the largest teeth of *A. prenasalis*, *Brachychampsa*, and "*C.*" *affinis*, were the molariform distals. These teeth typically approached the size of the largest mesial crowns, which themselves had very high PC1 values. This resulted in  species having the shallowest slopes and the greatest y-intercepts. Although *Allognathosuchus* has similar distal teeth, a mesial crown steepened the regression. Both *B.*

329 *sternbergii* specimens' cranial teeth have similar slopes,  intercepts differed by ~0.15.

330 *Hamadasuchus rebouli* had the largest teeth for its skull length with a dramatic  undulation in

331 size, and the greatest slope of any fossil cranial series. *Boverisuchus vorax* and *A. krebsi* both

332 formed significant trends with very high slopes. It should be noted that these two species have

333 less than a third of their teeth accounted for, and are all located in a narrow region of the

334 dentary 

335 DISCUSSION

336 Developmental trends in crocodylian heterodonty

337  Multiple measures of heterodonty allow for us to describe morphological conditions

338 across the crocodylian specimens sampled in a thorough manner, through a combination of

339 disparity, morphotypes ranges, and serial homology along the tooth row. All living crocodylians

340 are heterodont to varying degrees, and these data showed significant variability of

341 morphotypes along the dental arcade for all specimens. Although dentition varies between

342 species, certain consistencies were seen throughout the extant members of the clade:

- 343 1. Similar teeth occur on both the cranial and dentary dental arcades.
- 344 2. The vast majority of shape variance from the labial perspective ranges from high-
- 345 caniniform to high molariform. Only minor distal curvature is apparent in more
- 346 caniniform crowns.
- 347 3. There is serial homology in tooth shape from-mesial-to-distal along the tooth row, and
- 348 molariformy increases in this direction.  s is significantly linear for both dental
- 349 arcades.

4. Size variability consists of an undulating pattern with three peaks, with large procumbent crowns interspersed within smaller crowns.

Shape- and size-heterodonty are clearly decoupled, as they change in dramatically different, and seemingly independent, fashions along the arcade. This begs the question; do developmental agents influence size and shape separately? Although quite a bit of research has looked at how crocodylian teeth grow and replace themselves (INTRODUCTION), surprisingly little has been done on what developmental influences affect tooth size and shape. Crocodylian teeth are replaced in waves, or Zahnreihe (Edmund, 1962; Westergaard and Ferguson, 1990; Osborn, 1998), but these waves appear to be unrelated to the morphological variables investigated here. Keiser et al., (1993) compartmentalized the dentition along the tooth row for *C. niloticus*, grouping teeth into incisor, 'premolar,' and 'molar' regions with each reflecting a procumbent tooth. They did not offer a developmental mechanism that differentiates these categories though. Fruchard (2012) suggested that the only difference between procumbent teeth and their smaller counterparts was that the former was "programmed to be bigger," suggesting some sort of additional developmental signaling to enlarge teeth. More research is needed on how tooth shape and size are established developmentally in order to truly understand what generates heterodonty.

Although we were able to define heterodonty successfully, the task of assigning a singular dental morphotype to any one species of crocodylian is much more difficult. Heterodonty seems to vary within species, making the assignment of a singular heterodonty measure to an entire species dubious. As far as biological explanations for this, tooth form is

almost certainly influenced by allometry. Ontogenetic shifts in feeding niche have been documented in most crocodylian species (e.g. Groombridge, 1982; Webb, Manolis, & Buckworth, 1982; Pooley & Gans, 1976; Pooley, 1989; Delany, 1990; Santos et al., 1996; Da Silveira  Magnusson, 1999; Subalusky, A. L., Fitzgerald, L. A., & Smith, 2009 Wallace ~~and~~ Leslie, 2008; Borteiro et al.  2009; Hanson et al., 2014), and allometric changes in the feeding apparatus with size are often explained as a structural consequence of this (e.g. Dodson, 1975; Webb ~~and~~ Messel, 1978; Hutton, 1987; Erickson, Lappin, & Vliet, 2003; Verdade, 2000; Wu et al., 2006; Gignac  Erickson, 2016; Gignac  O'Brien, 2016). Concerning teeth, a qualitative increase in overall molariformy was observed in *A. mississippiensis*, and functioned to meet the mechanical demands of increased durophagy (Erickson, Lappin, & Vliet, 2003; Gignac & Erickson, 2014). Although our sample size is too low to confidently assess dental ontogeny within each species, we did see a similar general trend in conspecifics of different sizes. In particular, the larger of our two *C. porosus* had a greater y-intercepts indicating greater molariformy. This trend was also seen in the *B. sternbergii* cranial specimens. In addition to allometry, phenotypic changes due to environmental factors may also influence teeth. Skull shape and tooth orientation are irregularly influenced by the captive rearing (Erickson et al., 2004; Drumheller et al., 2016), and how this may also influence tooth shape has yet to be determined. Many of our specimens had 'no data' concerning their rearing, so we do not know if captivity influenced either tooth or skull morphology.

Adaptive explanations for interspecific variability in modern taxa

The method presented here indicates that significant differences in tooth morphology exist between modern members of Crocodylia. Bite force in crocodylians is primarily influenced by size (Erickson et al., 2012), and our data set shows that similarly sized crocodylians may have very different tooth dimensions. This rules out maximum bite force as the sole limiting factor dictating tooth form. Although we are reluctant to associate specific prey items with a specific tooth forms, shape and size will influence how a tooth interacts with a particular substrate. We therefore suggest that a biomechanical link should exist between the structural limits imposed by tooth form and the material properties of the substrates it interacts with.

As with all jawed vertebrates, crocodylian teeth must cope with different stresses based on their respective position along the arcade. Distal teeth will endure more force during a bite due to their close proximity to the hinge (Cleuren, Aerts, & Vree, 1995; Erickson, Lappin, & Vliet, 2003; Erickson et al., 2012; McHenry et al., 2006). This explains why these teeth are always on the molariform half of the shape spectrum; the larger base-to-height ratio gives them greater relative bending strengths (Van Valkenburgh and Ruff, 1987; Gignac & Erickson, 2014; Monfroy, 2017). Because force is highest in this region, food processing typically occurs here (Busbey, 1989; Davenport et al., 1990; Cleurens and de Vree, 2000). Their short size also ensures they do not impede jaw closure. This necessity is very apparent in *G. gangeticus*, and explains the poor linear shape relationship along the tooth row. Having all the teeth be high- to mid-caniniform except for the distal-most region in attempt to reduce heterodonty as much

as possible (Grigg and Gans, 1993), while ensuring the distals do not impede jaw closure or break when processing food.

Particular attention should be paid to the distals in modern alligatoroids and crocodyloids, as their relative size varies noticeably between species (Figure 6a). Most members of these clades have a single procumbent tooth followed distally by several smaller teeth. These are typically mid-molariform, but even the high-molariform distals of *C. siamensis* were some of the smallest teeth in its arcade. *Alligator sinensis* differed from this, in that it had a row of 4-5 relatively large, high-molariform, crowns followed by only one crown reduced in size. Probably the most extreme condition, *O. tetraspis* had distals that were exceptionally large; the largest crowns at positions M10-12 and D11-13 all belonged to members of this species. Aoki (1989) qualitatively noted these unique conditions, and suggested they facilitated durophagy. All alligatoroids and crocodyloids sampled here have been recorded to consume at least some hard prey items though (e.g. Brazaitis, 1973; McIlhenny, 1976; Taylor, 1979; Groombridge, 1982; Ross and Magnusson, 1989; Santos et al., 1996; Selvaraj, 2012; Nifong & Silliman, 2013), so it is unclear what selection pressure resulted in these particular morphologies. It may be a result of body size. Bite force tests of *A. mississippiensis* showed the pressure produced at its procumbent distal (M11) to be adequate to crush its harder prey items (Erickson, Lappin, & Vliet, 2003; Gignac & Erickson, 2014). If this is the case in most of the large crocodylians, enlarging the distal-most crowns would be unnecessary. *Alligator sinensis* and *O. tetraspis*, on the other hand, may need more extreme dentition closer to the hinge; their smaller size would make it more difficult to process foods with similar mechanical properties. Another explanation for this is the frequency of consuming hard prey. Although both *A. sinensis* and *O. tetraspis*

have broad diets, studies have shown certain populations to consume disproportionately large numbers of shelled mollusks and crustaceans (Cheng et al., 1957; Groombridge, 1982; Ross Magnusson, 1989; Luiselli, Akani, & Capizzi, 1999; Pauwels et al., 2007).

Caniniform mesials are ideal for the acquisition of prey, as pointed apices reduce surface area to puncture compliant substrate (Frazetta, 1988). Being farther from the hinge, these teeth move faster during a strike and are more likely to contact prey trying to escape (Busbey, 1989). All taxa measured here also have two sets of large mesial teeth on both arcades, often referred to as 'pseudocanines' (Brochu, 1999). These procumbent teeth are well built for puncturing, likely make first contact with prey during jaw closure, and resilient against struggling prey (Iordansky, 1964). High size heterodonty in caimanines was typically a consequence of large pseudocanines and relatively small remaining crowns, (Figure 6b). Their pseudocanines can become so large that dentary crowns may grow entirely through the cranial rostrum in adults (Brazaitis, 1973). The molariform distals were rather small by comparison. This suggests these taxa prioritize securing prey over processing it. This may be a specialization for hunting mobile and/or compliant prey (Sampaio et al., 2013), as insects and fish can make up a large portion of their diet (Santos et al., 1996). The exceptionally large pseudocanines of *C. porosus* also show a prioritization for puncturing and securing soft-bodied prey in a larger context, as this species is notorious for actively hunting large vertebrates such as sharks, cattle, horses, and humans (e.g. Taylor, 1979; Kar & Bustard, 1981; Groombridge, 1982; Doody, 2009; Hanson et al., 2015). Similar to caimanines, this species atrophies position P2 to make room for enlarged D1 pseudocanines (Brown et al., 2015)

456 The longirostrine species have a reputation for eating small, aquatic prey with a focus
 457 on fish (Peyer, 1968; Webb, Manolis, & Buckworth, 1982), and multiple lines of evidence
 458 suggest the feeding apparatus is well designed for this function. The longirostrine shape
 459 reduces resistance during both lateral motion and jaw adduction when feeding underwater,
 460 and the increased snout length allows for a faster strike (Pooley, 1989; Thorbjarnarson, 1990;
 461 McHenry et al., 2006; Pierce et al., 2008). Highly caniniform teeth can quickly puncture fast-
 462 moving, compliant prey, and their elongate shape may also lower resistance. The longirostrine
 463 cranio-dental morphotype may be prey-size inhibitive, as larger prey could damage it while
 464 struggling. The elongate mandibular symphysis of longirostrines results in a mechanical
 465 disadvantage against forces produced by shaking/twisting prey as well (Walmsley et al., 2013),
 466 and there is a reduction in masticatory musculature relative to other crocodylians (Endo et al.,
 467 2002). The gracile nature of the dentition means a lower bending strength, also making them
 468 more susceptible to breakage by more powerful prey (Figure 6)

469 The longirostrine taxa also had the lowest heterodonty of modern crocodylians, which is
 470 reminiscent the 'homodont' condition apparent in numerous aquatic predators such as
 471 odontocetes whales (Rommel, 1990). This condition is believed to be ideal for catching and
 472 holding, but not processing, small aquatic prey (MacLeod et al., 2007), as most prey items
 473 consumed are under 10% of their body length (MacLeod et al., 2006). A convergent reduction
 474 in heterodonty within both groups indicates a transition from a multi-functional dental arcade
 475 to one almost exclusively for prey capture. This is clearly the condition in *G. gangeticus*, as it is
 476 almost entirely caniniform along its tooth row and eats primarily fish (Groombridge, 1982). The
 477 other longirostrine taxa, although also primarily caniniform, still displayed the linear shape

478 change typical of other crocodyloids. This may indicate a more versatile diet, these taxa have
 479 also been known to process larger/harder prey items, such as crustaceans, turtles, and
 480 immature primates (Brazaitis, 1973; Groombridge, 1982; Galdikas and Yeager, 1984; Selvaraj,
 481 2012).

 Tooth shape may indicate differences in feeding behavior and processing ability, even
 483 though overlap exists in prey selection. *Alligator mississippiensis* and *C. niloticus* both consume
 484 a wide variety of prey, including both large and small mammals, crustaceans, fish, water fowl,
 485 snakes, turtles, and even each other (McIlenny, 1976; Pooley & Gans, 1976; Groombridge,
 486 1982; Delany  Abercrombie, 1986; Hutton, 1987; Shoop & Ruckdeschel, 1990; Rootes &
 487 Chabreck, 1993; Elsey et al., 2004; Wallace  Leslie, 2008; Gabrey, 2010). A comparison of
 488 controlled feedings of each of these species showed *A. mississippiensis* to fracture and consume
 489 noticeably more bovine skeletal elements than *C. niloticus* (Njau & Blumenschine, 2006;
 490 Drumheller & Brochu, 2014). Our *A. mississippiensis* specimen was generally more molariform
 491 than *C. niloticus*. These teeth would have greater bending strengths to resist breakage when
 492 processing hard material such as bone.

493 Fossil taxa and the appropriateness of analogues

494  Certain fossil taxa were reminiscent of modern counterparts. *Caiman crocodilus* and *C.*
 495 *yacare* are highly similar. This was expected as these species are closely related and were once
 496 considered subspecies, and both eat insects, crustaceans, and fish (Brazaitis, 1973;
 497 Groombridge, 1982; Da Silveira  Magnusson, 1999). Any differences in size and shape
 498 ranges appear to simply be a consequence of the former's incomplete arcades; no distal

maxillary or any dentary crowns were available. Both *Borealosuchus* and *Leidyosuchus* possess dentition somewhat typical of large modern crocodylians (Figure 6d). *Alligator mississippiensis* makes for an excellent dental analogue for one particular *Leidyosuchus* specimen (ROM 01903) as heterodonty and shape regressions overlap consistently. The fact that *Leidyosuchus* lacks any evidence of enlarged distal teeth may be indicative of a difference in the degree these taxa process hard materials, although no taphonomic evidence for this currently exists associated with *Leidyosuchus*. The four specimens of *B. sternbergii* all varied, most likely due to an allometric increase in molariformy (see above), but certain consistencies are apparent. Based on linear shape change, all specimens in this genus were projected to have mid-caniniform and –molariform teeth, with smaller specimens projected to additionally have the high-caniniform mesials and larger specimens having high-molariform distals. The best analogue for this species may be a member of *Crocodylus* where larger individuals have distals approaching high-molariform range such as *C. niloticus*.

The fact that *H. rebouli* surpasses our larger *C. porosus* in size heterodonty, and has a similar upper maximum in relative tooth size, indicates that it may have dealt with similar prey from a mechanical standpoint (Figure 6e). Peirosaurids are believed to be primarily terrestrial crocodyliforms (Tavares et al., 2017), and this dentition argues *H. rebouli* may have focused on mobile, terrestrial vertebrates (Larsson & Sues, 2007). Its large pseudocanines would puncture vertebrates tissue with similar effectiveness to those of *C. porosus*. Noticeable differences are the very large distal procumbent crowns of *H. rebouli*, which suggest potential differences in prey processing. These teeth may have been used for either sheering soft tissue or breaking

bone similar to modern mammalian carnassials, as rolling on land is not an effective means of dismemberment (Fish et al., 2007).

It has been stated that modern taxa do not have, or have secondarily lost, the degree of molariformy commonly found in extinct representatives. Known as “globidonty” (Norell, Clark, & Hutchison, 1994; Brochu, 1999; 2001; Ősi & Barrett, 2011), this term describes enlarged, high-molariform crowns potentially used for durophagy (Figure 6f). *Allognathosuchus* and *Brachychampsa* are ‘textbook’ examples of globidont taxa (Case, 1925; Carpenter and Lindsey, 1980). Although we agree with Brochu (2001, 2004) that *O. tetraspis* is not as extreme, its enlarged distals do overlap with these taxa in both size and shape. *Alligator prenasalis* and “*C. affinis*” distals are similar to *A. sinensis*, also creating a ridge of robust teeth (Mook, 1932). The mechanical capabilities of these particular crowns in ‘modern globidonts’ should be similar to the extinct taxa, which suggests similar processing abilities in the distal regions of the skull. The similarities break down when the rest of the jaw is considered though. The modern globidonts also have caniniform mesials and pseudocanines, suggesting a division of labor along the tooth row. Contrarily, almost all teeth of *A. prenasalis*, *Brachychampsa*, and “*C. affinis*” on the molariform half of the shape-spectrum. These extinct taxa probably did not need to do as much puncturing of compliant substrate, which supports the argument that they may have foraged for mollusks and slow moving turtles (Carpenter and Lindsey, 1980; similar to Salas-Gismondi et al., 2015) rather than being semi-aquatic, ambush predators. *Allognathosuchus*’ pseudocanine suggests a potential division of labor along the tooth row more like modern globidont taxa, but a more complete tooth row is necessary to confirm.

541 The nature of heterodonty in fossil taxa with less than 40% of their tooth row
 542 represented is especially difficult to determine. *Allognathosuchus* was represented here by a
 543 pseudocanine and globidont distals, but previous studies have shown a great degree of tooth
 544 and alveolar size variation that our specimen is missing (Case, 1925; Brochu 2004). The teeth of
 545 *A. krebsi* only occur between D6-D12. This region is so narrow that the slope is a poor indicator
 546 of the shape range. Although  a gavialoid, its teeth are most similar to *M. cataphractus* in
 547 shape, and  may be its most appropriate  analogue. Salas-Gismondi et al., (2016) argued that the
 548 lack of 'telescoped' eyes in *A. krebsi* is indicative of its marine habit (see also Hua & Jouve,
 549 2004). *Mecistops cataphractus* superficially shares this condition, and has been known to
 550 frequent brackish and costal environments (Brazaitis, 1973; Ross and Magnusson, 1989).

551 CONCLUSION

552 Foote's  MD is a reliable method for assessing heterodonty if the tooth row is near
 553 complete, but much of the variability in MD seen here is the result of incompleteness. Foote's
 MD relies on, among other things, the  and mean and the  nple size. Size heterodonty was
 555 greatly underrepresented in one specimen of *O. tetraspis* (AMNH 101417) as its pseudocanines
 556 were missing. These teeth would have deviated greatly from the Grand mean if present, and
 557 their inclusion would most certainly have raised the value. Our *Allognathosuchus* specimen had
 four teeth, with three being globidont and one an average-shaped pseudocanine. ~~The result~~
 559 ~~was a shape Grand mean that the pseudocanine deviated from greatly.~~ That, compounded with
 560 the low sample size, resulted in an inflated shape MD. A similar principle may be applied to
 561 both *A. krebsi* and *B. vorax*; because only a small area of the tooth row was represented all the

individual tooth shape values were close to the Grand mean, so the MD values were deflated. Slope of shape heterodonty is also influenced by the completeness of the tooth row. Several specimens have similar PC1 values for the same positions, but their slopes differed. This is typically due to one or both missing the mesial- or distal-most teeth. This was apparent in *C. crocodilus*; even though it shared almost identical tooth morphology with *C. yacare*, its cranial regression coefficients deviated because the distal 30% of its teeth were missing.

 did not consider all three dimensions here. Living crocodylian teeth are often discussed as conical (Edmund, 1969) or conodont (Hendrickx, Mateus, & Araújo, 2015b). Studies of bending strengths show variation between mesial-distal and labial-lingual axes as well as variation between species (Monfrey, 2017), indicating that functional information may be drawn from the dimension not considered here. This is especially important concerning fossil taxa, as pronounced lateral compression is commonplace. *Boverisuchus vorax* distals plotted similarity to globodont crowns using our method, but they are clearly ziphodont (Brochu, 2001, 2003). Future studies should consider this third dimension at least qualitatively, in order to avoid conflating disparate tooth morphotypes such as these.

 These limitations aside, the methods proposed here could be very useful in dealing with incomplete fossils. It is common for fossil crocodylian specimens to be lacking many/most of their teeth. The linear nature of tooth shape can predict the shape of these missing teeth. Although slopes may vary between species, a record of the ranges of slopes evident in a certain taxonomic group may be applied to a fossil with few teeth. The preserved tooth/teeth can be plugged into the linear equation, and the shapes of missing teeth may be predicted with a high

degree of certainty. This would result in a more complete representation of the extinct animal's anatomy, useful from the standpoint of both anatomical science and paleontological reconstruction.

Quantifying the teeth of crocodylians will add rigor to future life history studies of the clade. As a quantifiable trait, both tooth shape in a single position and heterodonty as a whole may be incorporated into character matrices of phylogenetic analyses. As we have shown heterodonty  be very real in Crocodylia, descriptors of dentition can describe a numerical range of morphology as opposed to cherry-picking an average tooth or single position.

Monospecific comparisons of multiple specimens can quantify the allometric changes in tooth morphology often alluded to in the literature. The teeth of fossil taxa can be compared statistically to modern taxa to determine the best analogue, and rigorous hypotheses about paleobehavior and paleoecology may be drawn.  Crocodylians, both living and extinct, may be

grouped in dental categories, allowing for species and specimens to be compared to one another. Frequency, size, and hardness of food items may be compared to these categories to determine if a link exists between dental morphotypes and dietary patterns. Crocodylians are used in both performance and actualistic taphonomy studies frequently, and the output of these studies could be correlated with tooth dimensions; tooth shape may be compared to bite-force, death-rolling, and bone-modification.



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608



References:

Adams, T. L., Noto, C. R., & Drumheller, S. (2017). A large neosuchian crocodyliform from the Upper Cretaceous (Cenomanian) woodbine formation of North Texas. *Journal of Vertebrate Paleontology*, 37(4), e1349776.

Aoki, R. (1989). The jaw mechanics in the heterodont crocodilians. *Current Herpetology in East Asia*, 1, 17-21.

Bennett, G. E. (2012). Community structure and paleoecology of crocodyliforms from the upper Hell Creek Formation (Maastrichtian), eastern Montana, based on shed teeth. *Jeffersoniana*, 28, 1-15.

Berkovitz, B. K., & Shellis, R. P. (2017). *The teeth of non-mammalian vertebrates*. Academic Press.

Bookstein, F. L. (1997). *Morphometric tools for landmark data: geometry and biology*. Cambridge University Press.

Borteiro, C., Gutiérrez, F., Tedros, M., & Kolenc, F. (2009). Food habits of the Broad-snouted Caiman (*Caiman latirostris*: Crocodylia, Alligatoridae) in northwestern Uruguay. *Studies on Neotropical Fauna and Environment*, 44(1), 31-36.

Brazaitis, P. (1973). *The identification of living crocodilians*. New York Zoological Society.

- 626 Britt, E. J., Clark, A. J., & Bennett, A. F. (2009). Dental morphologies in gartersnakes
627 (*Thamnophis*) and their connection to dietary preferences.  Journal of Herpetology, 252-
628 259.
- 629 Brochu, C. A. (1999). Phylogenetics, taxonomy, and historical biogeography of Alligatoroidea.
630 Journal of Vertebrate Paleontology, 19(S2), 9-100.
- 631 Brochu, C. A. (2001). Crocodylian snouts in space and time: phylogenetic approaches toward
632 adaptive radiation. American Zoologist, 41(3), 564-585.
- 633 Brochu, C. A. (2003). Phylogenetic approaches toward crocodylian history. Annual Review of
634 Earth and Planetary Sciences, 31(1), 357-397.
- 635 Brochu, C. A. (2004). Alligatorine phylogeny and the status of *Allognathosuchus* Mook, 1921.
636 Journal of Vertebrate Paleontology, 24(4), 857-873.
- 637 Brochu, C. A., & Storrs, G. W. (2012). A giant crocodile from the Plio-Pleistocene of Kenya, the
638 phylogenetic relationships of Neogene African crocodylines, and the antiquity of
639 *Crocodylus* in Africa. Journal of Vertebrate Paleontology, 32(3), 587-602.
- 640 Brown, C. M., VanBuren, C. S., Larson, D. W., Brink, K. S., Campione, N. E., Vavrek, M. J., &
641 Evans, D. C. (2015). Tooth counts through growth in diapsid reptiles: implications for
642 interpreting individual and size-related variation in the fossil record. Journal of
643 Anatomy, 226(4), 322-333.
- 644 Buckley, L. G., & Currie, P. J. (2014). Analysis of intraspecific and ontogenetic variation in
645 dentition of *Coelophysis bauri* (Late Triassic), and implications for the systematics of



isolated theropod teeth: Bulletin 63 (Vol. 63). New Mexico Museum of Natural History

647

and Science.



Busbey, A. B. (1995). The structural consequences of skull flattening in crocodilians. Functional

649

morphology in vertebrate paleontology, 173-192.

650

Busbey, A. B. (1989). Form and function of the feeding apparatus of *Alligator mississippiensis*.

651

Journal of Morphology, 202(1), 99-127.

652

Carpenter, K., & Lindsey, D. (1980). The dentary of *Brachychampsia montana* Gilmore



(Alligatorinae; Crocodylidae), a Late Cretaceous turtle-eating alligator. *Journal of*

654

Paleontology, 1213-1217.



Case, E. C. (1925). Note on a new species of the Eocene crocodilian *Allognathosuchus*, A.

656

wartheni. University of Michigan.



Cheng-kuan, C. (1957). Observations on the life history of Chinese alligator (*Alligator sinensis*

658

Fauvel). *Acta Zoologica Sinica*, 2(004).



Cleuren, J., Aerts, P., & Vree, F. D. (1995). Bite and joint force analysis in *Caiman crocodilus*.

660

Belgian Journal of Zoology (Belgium).

661

Cleuren, J., & de Vree, F. (1992). Kinematics of the jaw and hyolingual apparatus during feeding

662

in *Caiman crocodilus*. *Journal of Morphology*, 212(2), 141-154.



Cleuren, J., & de Vree, F. (2000). Feeding in crocodilians. Feeding: form, function, and evolution

664

in tetrapod vertebrates, 359-394.

- 665 Da Silveira, R., & Magnusson, W. E. (1999). Diets of spectacled and black caiman in the
666 Anavilhanas Archipelago, Central Amazonia, Brazil.  Journal of Herpetology, 181-192.
- 667 D'Amore, D. C. (2009). A functional explanation for denticulation in theropod dinosaur teeth.
668 The Anatomical Record: Advances in Integrative Anatomy and Evolutionary Biology,
669 292(9), 1297-1314.
- 670 D'Amore, D. C. (2015). Illustrating ontogenetic change in the dentition of the Nile monitor
671 lizard, *Varanus niloticus*: a case study in the application of geometric morphometric
672 methods for the quantification of shape–size heterodonty. Journal of  Anatomy, 226(5),
673 403-419.
-  Daniel, W. J. T., & McHenry, C. (2001). Bite force to skull stress correlation-modelling the skull
675 of *Alligator mississippiensis*.
- 676 Davenport, J., Grove, D. J., Cannon, J., Ellis, T. R., & Stables, R. (1990). Food capture, appetite,
677 digestion rate and efficiency in hatchling and juvenile *Crocodylus porosus*. Journal of
678 Zoology, 220(4), 569-592.
- 679 Delany, M. F. (1990). Late summer diet of juvenile American alligators. Journal of Herpetology,
680 24(4), 418-421.
- 681 Delany, M. F., & Abercrombie, C. L. (1986). American alligator food habits in northcentral
682 Florida. The Journal of Wildlife Management,  3-353.
- 683 Dodson, P. (1975). Functional and ecological significance of relative growth in *Alligator*. Journal
684 of Zoology, 175(3), 315-355.

685 Doody, J. S. (2009). Eyes bigger than stomach: prey caching and retrieval in the saltwater
686 crocodile, *Crocodylus porosus*. Herpetological Review, 40(1), 26.

687 Drumheller, S. K., & Brochu, C. A. (2014). A diagnosis of *Alligator mississippiensis* bite marks
688 with comparisons to existing crocodylian datasets. Ichnos, 21(2), 131-146.

689 Drumheller, S. K., & Brochu, C. A. (2016). Phylogenetic taphonomy: a statistical and
690 phylogenetic approach for exploring taphonomic patterns in the fossil record using
691 crocodylians. Palaios, 31(10), 463-478.

 Edmund, A. G. (1962). Sequence and rate of tooth replacement in the Crocodilia. **R. Ont. Mus.,**
693 **Life Sci. Div., Contr.,** 56, 1-42.

694 Edmund, A. G. (1969). Dentition. pp. 117-200 in: C. Gans, A. d'A. Bellairs & TS Parsons (eds.),
695 Biology of the Reptilia, Volume 1: Morphology A.

696 Elsey, R. M., Trosclair III, P. L., & Linscombe, J. T. (2004). The American alligator as a predator of
697 mottled ducks. Southeastern Naturalist, 3(3), 381-390.

698 Enax, J., Fabritius, H. O., Rack, A., Prymak, O., Raabe, D., & Epple, M. (2013). Characterization of
699 crocodile teeth: correlation of composition, microstructure, and hardness. Journal of
700  **Structural biology,** 184(2), 155-163.

701 Endo, H., Aoki, R., Taru, H., Kimura, J., Sasaki, M., Yamamoto, M.,  & Hayashi, Y. (2002).
702 Comparative functional morphology of the masticatory apparatus in the long-snouted
703 crocodiles. Anatomia, Histologia, Embryologia, 31(4), 206-213.

Erickson, G. M. (1996). Daily deposition of dentine in juvenile *Alligator* and assessment of tooth replacement rates using incremental line counts. *Journal of Morphology*, 228(2), 189-194.

Erickson, G. M., Gignac, P. M., Lappin, A. K., Vliet, K. A., Brueggen, J. D., & Webb, G. J. W. (2014). A comparative analysis of ontogenetic bite-force scaling among Crocodylia. *Journal of Zoology*, 292(1), 48-55.

Erickson, G. M., Gignac, P. M., Steppan, S. J., Lappin, A. K., Vliet, K. A., Brueggen, J. D., ... & Webb, G. J. (2012). Insights into the ecology and evolutionary success of crocodilians revealed through bite-force and tooth-pressure experimentation. *PLoS One*, 7(3), e31781.



Erickson, G. M., Lappin, A. K., & Vliet, K. A. (2003). The ontogeny of bite-force performance in American alligator (*Alligator mississippiensis*). *Journal of Zoology*, 260(3), 317-327.

Ferguson, M. W. J. (1981). The structure and development of the palate in *Alligator mississippiensis*. *Archives of oral biology*, 26(5), 427-443.

Fish, F. E., Bostic, S. A., Nicastro, A. J., & Beneski, J. T. (2007). Death roll of the alligator: mechanics of twist feeding in water. *Journal of experimental biology*, 210(16), 2811-2818.

Foote, M. (1993). Contributions of individual taxa to overall morphological disparity. *Paleobiology*, 19(4), 403-419.



- 723 Frey, E., & Monninger, S. (2010). Lost in action—the isolated crocodilian teeth from Enspel and
724 their interpretive value. *Palaeobiodiversity and Palaeoenvironments*, 90(1), 65-81.
- 725 Fruchard, C. (2012). The Nile crocodile, a new model for investigating heterodonty and dental
726 continuous renewal in vertebrates. *BioSciences Master Reviews*
- 727 Gerke, O., & Wings, O. (2016). Multivariate and cladistic analyses of isolated teeth reveal
728 sympatry of theropod dinosaurs in the Late Jurassic of northern Germany. *PloS one*,
729 11(7), e0158334.
- 730 Erickson, G. M., Gignac, P. M., Lappin, A. K., Vliet, K. A., Brueggen, J. D., & Webb, G. J. W. (2014).
731 A comparative analysis of ontogenetic bite-force scaling among Crocodylia. *Journal of*
732 *Zoology*, 292(1), 48-55.
- 733 Frazzetta, T. H. (1988). The mechanics of cutting and the form of shark teeth (Chondrichthyes,
734 Elasmobranchii). *Zoomorphology*, 108(2), 93-107.
- 735 Gabrey, S. W. (2010). Demographic and geographic variation in food habits of American
736 alligators (*Alligator mississippiensis*) in Louisiana. *Herpetological Conservation and*
737 *Biology*, 5(2), 241-250.
- 738 Galdikas, B. M., & Yeager, C. P. (1984). Brief report: Crocodile predation on a crab-eating
739 macaque in Borneo. *American Journal of Primatology*, 6(1), 49-51.
- 740 Gignac, P. M., & Erickson, G. M. (2015). Ontogenetic changes in dental form and tooth
741 pressures facilitate developmental niche shifts in American alligators. *Journal of*
742 *Zoology*, 295(2), 132-142.

- 743 Gignac, P., & O'Brien, H. (2016). Suchian feeding success at the interface of ontogeny and
744 macroevolution. *Integrative and comparative biology*, 56(3), 449-458.
- 745 Grigg, G., & Gans, C. (1993). Morphology and physiology of the Crocodylia. *Fauna of Australia*
746 Vol 2A Amphibia and Reptilia. Australian Government Publishing Service, Canberra.326-
747 336.
- 748 Groombridge, B. 1982. The IUCN Amphibia-Reptilia Red Data Book, Part 1: Testudines,
749 Crocodylia, Rhynchocephalia. IUCN, Gland, Switzerland.
- 750 Gunz, P., & Mitteroecker, P. (2013). Semilandmarks: a method for quantifying curves and
751 surfaces. *Hystrix, the Italian Journal of Mammalogy*, 24(1), 103-109.
- 752 Hanson, J. O., Salisbury, S. W., Campbell, H. A., Dwyer, R. G., Jardine, T. D., & Franklin, C. E.
753 (2015). Feeding across the food web: The interaction between diet, movement and
754 body size in estuarine crocodiles (*Crocodylus porosus*). *Austral Ecology*, 40(3), 275-286.
- 755 Hendrickx, C., & Mateus, O. (2014). Abelisauridae (Dinosauria: Theropoda) from the Late
756 Jurassic of Portugal and dentition-based phylogeny as a contribution for the
757 identification of isolated theropod teeth. *Zootaxa*, 3759.
- 758 Hendrickx, C., Mateus, O., & Araújo, R. (2015a). The dentition of megalosaurid theropods. *Acta*
759 *Palaeontologica Polonica*, 60(3), 627-642.
- 760 Hendrickx, C., Mateus, O., & Araújo, R. (2015b). A proposed terminology of theropod teeth
761 (Dinosauria, Saurischia). *Journal of Vertebrate Paleontology*, 35(5), e982797.

762 Hua, S., & Jouve, S. (2004). A primitive marine gavialoid from the Paleocene of Morocco.
763 Journal of Vertebrate Paleontology, 24(2), 341-350.

764 Hutton, J. M. (1987). Growth and feeding ecology of the Nile crocodile *Crocodylus niloticus* at
765 Ngezi, Zimbabwe. The Journal of Animal Ecology, 25-38.

766 Iordansky, N. N. (1964). The jaw muscles of the crocodiles and some relating structures of the
767 crocodilian skull. Anatomischer Anzeiger, 115, 256-280.

768 Kar, S. K., & Bustard, H. R. (1983). Attacks on domestic livestock by juvenile saltwater crocodile,
769 *Crocodylus porosus*, in Bhitarkanika Wildlife Sanctuary, Orissa India. Amphibia-Reptilia,
770 4(1), 81-83.

771 Kieser, J. A., Klapsidis, C., Law, L., & Marion, M. (1993). Heterodonty and patterns of tooth
772 replacement in *Crocodylus niloticus*. Journal of Morphology, 218(2), 195-201.

773 Klingenberg, C. P. (2011). MorphoJ: an integrated software package for geometric
774 morphometrics. Molecular ecology resources, 11(2), 353-357.

775 Langston, W. (1973). The crocodilian skull in historical perspective. Biology of the Reptilia, 4,
776 263-284.

777 Larson, D. W., Brown, C. M., & Evans, D. C. (2016). Dental disparity and ecological stability in
778 bird-like dinosaurs prior to the end-Cretaceous mass extinction. Current Biology, 26(10),
779 1325-1333.

- 780 Larson, D. W., & Currie, P. J. (2013). Multivariate analyses of small theropod dinosaur teeth and
781 implications for paleoecological turnover through time. PLoS One, 8(1), e54329.
- 782 Larsson, H. C. E., & Sidor, C. A. (1999). Unusual crocodyliform teeth from the Late Cretaceous
783 (Cenomanian) of southeastern Morocco. Journal of Vertebrate Paleontology, 19(2), 398-
784 401.
- 785 LeBlanc, A. R., Brink, K. S., Cullen, T. M., & Reisz, R. R. (2017). Evolutionary implications of tooth
786 attachment versus tooth implantation: a case study using dinosaur, crocodilian, and
787 mammal teeth. Journal of Vertebrate Paleontology, 37(5), e1354006.
- 788 Lecuona, A., & Pol, D. (2008). Tooth morphology of *Notosuchus terrestris* (Notosuchia:
789 Mesoeucrocodylia): new evidence and implications. Comptes Rendus Palevol, 7(7), 407-
790 417.
- 791 Luiselli, L., Akani, G. C., & Capizzi, D. (1999). Is there any interspecific competition between
792 dwarf crocodiles (*Osteolaemus tetraspis*) and Nile monitors (*Varanus niloticus ornatus*)
793 in the swamps of central Africa? A study from south-eastern Nigeria. Journal of Zoology,
794 247(1), 127-131.
- 795 MacLeod, C. D., Reidenberg, J. S., Weller, M., Santos, M. B., Herman, J., Goold, J., & Pierce, G. J.
796 (2007). Breaking symmetry: The marine environment, prey size, and the evolution of
797 asymmetry in cetacean skulls. The Anatomical Record: Advances in Integrative Anatomy
798 and Evolutionary Biology, 290(6), 539-545.

799 MacLeod, C. D., Santos, M. B., Lopez, A., & Pierce, G. J. (2006). Relative prey size consumption
800 in toothed whales: implications for prey selection and level of specialisation. Marine
801 Ecology Progress Series, 326, 295-307.

802 Martin, J. E. (2007). New material of the Late Cretaceous globidontan *Acynodon iberoccitanus*
803 (Crocodylia) from southern France. Journal of Vertebrate Paleontology, 27(2), 362-372.

804 McHenry, C. R., Clausen, P. D., Daniel, W. J., Meers, M. B., & Pendharkar, A. (2006).
805 Biomechanics of the rostrum in crocodilians: a comparative analysis using finite-element
806 modeling. The Anatomical Record Part A: Discoveries in Molecular, Cellular, and
807 Evolutionary Biology, 288(8), 827-849.

808 McIlhenny, E. A. (1976). The alligator's life history. Ten Speed Pr.

809 Mitteroecker, P., Gunz, P., Windhager, S., & Schaefer, K. (2013). A brief review of shape, form,
810 and allometry in geometric morphometrics, with applications to human facial
811 morphology. Hystrix, the Italian Journal of Mammalogy, 24(1), 59-66.

812 Monfroy, Q. T. (2017). Correlation between the size, shape and position of the teeth on the
813 jaws and the bite force in Theropoda. Historical Biology, 29(8), 1089-1105.

814 Mook, C. C. (1932). A study of the osteology of *Alligator prenasalis* (Loomis). Museum.

815 Narváez, I., Brochu, C. A., Escaso, F., Pérez-García, A., & Ortega, F. (2015). New crocodyliforms
816 from southwestern Europe and definition of a diverse clade of European Late
817 Cretaceous basal eusuchians. PloS one, 10(11), e0140679.

- 818 Nifong, J. C., & Silliman, B. R. (2013). Impacts of a large-bodied, apex predator (*Alligator*
819 *mississippiensis* Daudin 1801) on salt marsh food webs. *Journal of experimental marine*
820 *biology and ecology*, 440, 185-191.
- 821 Njau, J. K., & Blumenschine, R. J. (2006). A diagnosis of crocodile feeding traces on larger
822 mammal bone, with fossil examples from the Plio-Pleistocene Olduvai Basin, Tanzania.
823 *Journal of Human Evolution*, 50(2), 142-162.
- 824 Njau, J. K., & Blumenschine, R. J. (2012). Crocodylian and mammalian carnivore feeding traces
825 on hominid fossils from FLK 22 and FLK NN 3, Plio-Pleistocene, Olduvai Gorge, Tanzania.
826 *Journal of human evolution*, 63(2), 408-417.
- 827 Norell, M., Clark, J. M., & Hutchison, J. H. (1994). The Late Cretaceous alligatoroid
828 *Brachychampsia montana* (Crocodylia): new material and putative relationships.
829 *American Museum novitates*; no. 3116.
- 830 Novas, F. E., Pais, D. F., Pol, D., Carvalho, I. D. S., Scanferla, A., Mones, A., & Riglos, M. S. (2009).
831 Bizarre notosuchian crocodyliform with associated eggs from the Upper Cretaceous of
832 Bolivia. *Journal of Vertebrate Paleontology*, 29(4), 1316-1320.
- 833 Osborn, J. W. (1998). Relationship between growth and the pattern of tooth initiation in
834 alligator embryos. *Journal of dental research*, 77(9), 1730-1738.
- 835 Ósi, A., & Barrett, P. M. (2011). Dental wear and oral food processing in *Caiman latirostris*:
836 analogue for fossil crocodylians with crushing teeth. *Neues Jahrbuch für Geologie und*
837 *Paläontologie-Abhandlungen*, 261(2), 201-207.

- 838 Ósi, A., Clark, J. M., & Weishampel, D. B. (2007). First report on a new basal eusuchian
839 crocodyliform with multicusped teeth from the Upper Cretaceous (Santonian) of
840 Hungary. Neues Jahrbuch für Geologie und Paläontologie-Abhandlungen, 243(2), 169-
841 177.
- 842 Pauwels, O. S., Barr, B., Sanchez, M. L., & Burger, M. (2007). Diet records for the dwarf crocodile
843 (*Osteolaemus tetraspis tetraspis*) in Rabi Oil Fields and Loango National Park,
844 Southwestern Gabon. Hamadryad, 31(2), 258-264.
- 845 Perez, S. I., Bernal, V., & Gonzalez, P. N. (2006). Differences between sliding semi-landmark
846 methods in geometric morphometrics, with an application to human craniofacial and
847 dental variation. Journal of anatomy, 208(6), 769-784.
- 848 Peyer, B. (1968). Comparative odontology. University of Chicago Press.
- 849 Pierce, S. E., Angielczyk, K. D., & Rayfield, E. J. (2008). Patterns of morphospace occupation and
850 mechanical performance in extant crocodilian skulls: a combined geometric
851 morphometric and finite element modeling approach. Journal of morphology, 269(7),
852 840-864.
- 853 Pooley, A. C. (1989). Food and feeding habits. Crocodiles and alligators, 76-91.
- 854 Pooley, A. C., & Gans, C. (1976). The Nile crocodile. Scientific American, 234(4), 114-125.
- 855 Rohlf, F. J. (2010). Tps Series. Department of Ecology and Evolution, State University of New
856 York, Stony Brook.

- 857 Rohlf, F. J. (2013). TpsRelw 1.53. Dept. of Ecology and Evolution, State Univ. of New York at
858 Stony Brook, Stony Brook (NY).
- 859 Rootes, W. L., & Chabreck, R. H. (1993). Cannibalism in the American alligator. *Herpetologica*,
860 99-107.
- 861 Rommel, S. (1990). Osteology of the bottlenose dolphin. *The bottlenose dolphin*, 29-49.
- 862 Ross, C. A., & Magnusson, W. E. (1989). Living crocodilians. *Crocodiles and alligators*, 58-73.
- 863 Sadleir, R. W., & Makovicky, P. J. (2008). Cranial shape and correlated characters in crocodilian
864 evolution. *Journal of evolutionary biology*, 21(6), 1578-1596.
- 865 Salas-Gismondi, R., Flynn, J. J., Baby, P., Tejada-Lara, J. V., Wesselingh, F. P., & Antoine, P. O.
866 (2015). A Miocene hyperdiverse crocodylian community reveals peculiar trophic
867 dynamics in proto-Amazonian mega-wetlands. *Proceedings of the Royal Society of*
868 *London B: Biological Sciences*, 282(1804), 20142490.
- 869 Salas-Gismondi, R., Flynn, J. J., Baby, P., Tejada-Lara, J. V., Claude, J., & Antoine, P. O. (2016). A
870 new 13 million year old gavialoid crocodylian from proto-Amazonian mega-wetlands
871 reveals parallel evolutionary trends in skull shape linked to longirostry. *PloS one*, 11(4),
872 e0152453.
- 873 Sampaio, P. R. M., da Silva, M. N., de Matos, S. A., de Matos, L. R. A., & Acosta, M. (2013). First
874 report of predation by a caiman (*Paleosuchus trigonatus*, Crocodylia: Alligatoridae) on a
875 caecilian (*Caecilia marcusii*, Gymnophiona: Caecilidae). *Salamandra*, 49(4), 227-228.

- 876 Santos, S. A., Nogueira, M. S., Pinheiro, M. S., Campos, Z., Magnusson, W. E., Mourao, G. M.
877 (1996). Diets of *Caiman crocodilus yacare* from different habitats in the Brazilian
878 pantanal. *Herpetological Journal*, 6, 111-117.
- 879 Selvaraj, G. (2012). Herpetological notes: *Tomistoma schlegelii* (False Gharial). *Diet. Herpetol*
880 *Rev*, 43, 608-609.
- 881 Schwarz-Wings, D., Rees, J., & Lindgren, J. (2009). Lower cretaceous mesoeucrocodylians from
882 Scandinavia (Denmark and Sweden). *Cretaceous Research*, 30(5), 1345-1355.
- 883 Sheets, H. D. (2012). IMP software series. Computer software and manual]. Buffalo, New York:
884 Canisius College.
- 885 Sheets, H. D., Kim, K., & Mitchell, C. E. (2004). A combined landmark and outline-based
886 approach to ontogenetic shape change in the Ordovician trilobite *Triarthrus becki*. In
887 *Morphometrics* (pp. 67-82). Springer, Berlin, Heidelberg.
- 888 Sheets, H. D., & Zelditch, M. L. (2013). Studying ontogenetic trajectories using resampling
889 methods and landmark data. *Hystrix, the Italian Journal of Mammalogy*, 24(1), 67-73.
- 890 Shimada, K. (2001). On the concept of heterodonty. *Journal of Fossil Research*, 34(2), 52-54.
- 891 Shimada, K. (2002). Dental homologies in lamniform sharks (Chondrichthyes: Elasmobranchii).
892 *Journal of Morphology*, 251(1), 38-72.

- 893 Shimada, K., Sato, I., & Moriyama, H. (1992). Morphology of the tooth of the American Alligator
894 (*Alligator mississippiensis*): the fine structure and elemental analysis of the cementum.
895 Journal of morphology, 211(3), 319-329.
- 896 Shimada K (2004) The relationship between the tooth size and total body length in the
897 sandtiger shark, *Carcharias taurus* (Lamniformes: Odontaspidae). J Fossil Res 37, 76–
898 81.
- 899 Shimada K, Seigel JA (2005) The relationship between the tooth size and total body length in
900 the goblin shark, *Mitsukurina owstoni* (Lamniformes: Mitsukurinidae). J Fossil Res 38,
901 49–56.
- 902 Shoop, C. R., & Ruckdeschel, C. A. (1990). Alligators as predators on terrestrial mammals.
903 American Midland Naturalist, 407-412.
- 904 Smith, J. B., & Dodson, P. (2003). A proposal for a standard terminology of anatomical notation
905 and orientation in fossil vertebrate dentitions. Journal of Vertebrate paleontology,
906 23(1), 1-12.
- 907 Subalusky, A. L., Fitzgerald, L. A., & Smith, L. L. (2009). Ontogenetic niche shifts in the American
908 Alligator establish functional connectivity between aquatic systems. Biological
909 Conservation, 142(7), 1507-1514.
- 910 Tavares, S. A. S., Branco, F. R., de Souza Carvalho, I., & Maldanis, L. (2017). The
911 morphofunctional design of *Montealtosuchus arrudacamposi* (Crocodyliformes, Upper
912 Cretaceous) of the Bauru Basin, Brazil. Cretaceous Research, 79, 64-76.

- 913 Taylor, J. A. (1979). The foods and feeding habits of subadult *Crocodylus porosus* Schneider in
914 northern Australia. *Wildlife Research*, 6(3), 347-359.
- 915 Thorbjarnarson, J. B. (1990). Notes on the feeding behavior of the gharial (*Gavialis gangeticus*)
916 under semi-natural conditions. *Journal of Herpetology*, 24(1), 99-100.
- 917 Torices, A., Reichel, M., & Currie, P. J. (2014). Multivariate analysis of isolated tyrannosaurid
918 teeth from the Danek Bonebed, Horseshoe Canyon Formation, Alberta, Canada.
919 *Canadian Journal of Earth Sciences*, 51(11), 1045-1051.
- 920 Van Drongelen, W. & Dullemeijer, P. (1982). The feeding apparatus of *Caiman crocodilus*; a
921 functional-morphological study. *Anatomischer Anzeiger*, 151(4), 337-366.
- 922 Valkenburgh, B. V., & Ruff, C. B. (1987). Canine tooth strength and killing behaviour in large
923 carnivores. *Journal of Zoology*, 212(3), 379-397.
- 924 Verdade, L. M. (2000). Regression equations between body and head measurements in the
925 broad-snouted caiman (*Caiman latirostris*). *Revista Brasileira de Biologia*, 60(3), 469-482.
- 926 Wallace, K. M., & Leslie, A. J. (2008). Diet of the Nile crocodile (*Crocodylus niloticus*) in the
927 Okavango Delta, Botswana. *Journal of Herpetology*, 42(2), 361-368.
- 928 Walmsley, C. W., Smits, P. D., Quayle, M. R., McCurry, M. R., Richards, H. S., Oldfield, C. C., ... &
929 McHenry, C. R. (2013). Why the long face? The mechanics of mandibular symphysis
930 proportions in crocodiles. *PLoS One*, 8(1), e53873.

- 931 Webb, G. J. W., Manolis, S. C., & Buckworth, R. (1982). *Crocodylus johnstoni* in the McKinlay
- 932 river area, NTI Variation in the diet, and a new method of assessing the relative
- 933 importance of prey. Australian Journal of Zoology, 30(6), 877-899.
- 934 Webb, G. J. W., & Messel, H. (1978). Movement and dispersal patterns of *Crocodylus porosus* in
- 935 some rivers of Arnhem Land, Northern Australia. Wildlife Research, 5(2), 263-283.
- 936 Westergaard, B., & Ferguson, M. W. J. (1986). Development of the dentition in *Alligator*
- 937 *mississippiensis*. Early embryonic development in the lower jaw. Journal of Zoology,
- 938 210(4), 575-597.
- 939 Westergaard, B., & Ferguson, M. W. J. (1987). Development of the dentition in *Alligator*
- 940 *mississippiensis*. Later development in the lower jaws of embryos, hatchlings and young
- 941 juveniles. Journal of Zoology, 212(2), 191-222.
- 942 Westergaard, B., & Ferguson, M. W. (1990). Development of the dentition in *Alligator*
- 943 *mississippiensis*: upper jaw dental and craniofacial development in embryos, hatchlings,
- 944 and young juveniles, with a comparison to lower jaw development. American Journal of
- 945 Anatomy, 187(4), 393-421.
- 946 Wilberg, E. W. (2017). Investigating patterns of crocodyliform cranial disparity through the
- 947 Mesozoic and Cenozoic. Zoological Journal of the Linnean Society, 181(1), 189-208.
- 948 Wu, X. B., Xue, H., Wu, L. S., Zhu, J. L., & Wang, R. P. (2006). Regression analysis between body
- 949 and head measurements of Chinese alligators (*Alligator sinensis*) in the captive
- 950 population. Animal biodiversity and conservation, 29(1), 65-71.

951 Young, M. T., Brusatte, S. L., Beatty, B. L., De Andrade, M. B., & Desojo, J. B. (2012).
 952 Tooth-on-tooth interlocking occlusion suggests macrophagy in the Mesozoic marine
 953 crocodylomorph *Dakosaurus*. *The Anatomical Record: Advances in Integrative Anatomy*
 954 and *Evolutionary Biology*, 295(7), 1147-1158.

955 Zahradnicek, O., Buchtova, M., Dosedelova, H., & Tucker, A. S. (2014). The development of
 956 complex tooth shape in reptiles. *Frontiers in physiology*, 5, 74.

957 Zelditch, M. L., Sheets, H. D., & Fink, W. L. (2003). The ontogenetic dynamics of shape disparity.
 958 *Paleobiology*, 29(1), 139-156.

959 Zelditch, M., Swiderski, D., Sheets, D. H., & Fink, W. (2004). *Geometric morphometrics for*
 960 *biologists: A primer*: Elsevier Academic Press. Waltham, MA.

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TABLE LEGENDS

Table 1: Size, disparity, and regression information for modern crocodylian specimens. Head lengths are in millimeters. Morphological disparity (MD) is for all teeth for each specimen. “N” represents number for teeth available versus number of positions total. Regression information is for Principal Component 1 plotted against tooth position.

Table 2: Size, disparity, and regression information for fossil crocodylian specimens. Head lengths are in millimeters. Morphological disparity (MD) is for all teeth for each specimen. “N” represents number for teeth available versus number of positions total. Regression information is for Principal Component 1 plotted against tooth position.

FIGURE LEGENDS

Figure 1: Methodology for data collection. A) skulls were selected based on number and condition of teeth present (data was not collected from this photo). B) Each tooth was photographed individually, and the margins were traced (Rohlf, 2016). C) Each outline was converted to 30 equidistant semilandmarks which were then slid. D) Tooth shape variance was represented by vector diagrams, with the mean depicted as points and deviation by vector lines.

Figure 2: Principal component one. Vector diagrams indicate the maximum range of variance (vectors) from the mean (points) for both cranial and dentary teeth.

Figure 3: Average tooth size (above) and shape (below) plotted against skull shape. Gray error bars indicate standard deviation as a consequence of heterodonty. Solid markers depict extant specimens, and hollow markers depict extinct. The solid regression line indicates only extant specimens, while the dashed line indicates all specimens. Maximum shape range is represented through vector diagrams.

Figure 4: Foote's morphological disparity (A, B) and ranges (C, D) for both size and shape. Size is represented by centroid size and shape is represented by Principal Component 1. For each species, all teeth in the sample are represented. For MD, multiple individuals from a single species are listed in a single column, and those with lower values are superimposed in front of those with higher values. Ranges are a composite of all members of a species.

Figure 5: Heterodonty by tooth position. Centroid Size (CS) and Principal Component one (PC1) for extant Alligatoroidea (top), Crocodyloidea (middle), and Gavialoidae (bottom) plotted

against position along the arcade. Welch's ANOVA output comparing positions is listed for each graph with multiple specimens.

Figure 6: Direct comparisons between selected extant and extinct taxa. The "size" axis represents normalized centroid size, and the "shape" axis represents principal component one. A) modern taxa with high molariformy, B) Caimaninae with high size-heterodonty, C) longirostrine taxa, D) mid-heterodont fossil taxa, E) crocodyloids and peirosaurids with high-heterodonty, F) fossil globidont taxa. Note: *Allognathosuchus*, *Argochampsia krebsi*, and *B. sternbergii* were not size normalized by their own skull length, as indicated by empty symbols.

Table 1(on next page)

Size, disparity, and regression information for modern crocodylian specimens.

Head lengths are in millimeters. Morphological disparity (MD) is for all teeth for each specimen. “N” represents number for teeth available versus number of positions total. Regression information is for Principal Component 1 plotted against tooth position.

Genus	Species	Number	Head Length	MD (size)	MD (shape)	Host bone
<i>Alligator</i>	<i>mississippiensis</i>	AMNH 71621	407.81	0.00052	0.0205	cranium mandible
<i>Alligator</i>	<i>sinensis</i>	AMNH 23900	161.87	0.00040	0.0241	cranium mandible
		AMNH 23907	139.89	0.00034	0.0170	cranium mandible
<i>Caiman</i>	<i>yacare</i>	AMNH 97297	206.96	0.00084	0.0300	cranium mandible
		AMNH 97300	347.30	0.00102	0.0217	cranium mandible
<i>Crocodylus</i>	<i>acutus</i>	AMNH 7856	486.60	0.00051	0.0208	cranium mandible
		AMNH 7857	436.66	0.00073	0.0251	cranium mandible
<i>Crocodylus</i>	<i>niloticus</i>	AMNH 23471	639.27	0.00064	0.0143	cranium mandible
		AMNH 142494	N/A	N/A	0.0277	cranium mandible
<i>Crocodylus</i>	<i>palustris</i>	AMNH 75707	400.85	0.00063	0.0203	cranium mandible
		AMNH 96134	236.82	0.00031	0.0250	cranium mandible
<i>Crocodylus</i>	<i>porosus</i>	AMNH 66639	426.22	0.00071	0.0182	cranium mandible
		AMNH 94957	575.23	0.00115	0.0179	cranium mandible
<i>Crocodylus</i>	<i>siamensis</i>	AMNH 49231	381.95	0.00049	0.0339	cranium mandible
		AMNH 72640	251.86	0.00042	0.0289	cranium

						mandible
<i>Gavialis</i>	<i>gangeticus</i>	AMNH 131377	428.63	0.00023	0.0110	cranium mandible
<i>Mecistops</i>	<i>cataphractus</i>	AMNH 107634	330.37	0.00035	0.0137	cranium mandible
<i>Osteolaemus</i>	<i>tetraspis</i>	AMNH 101417	247.98	0.00026	0.0236	cranium mandible
		AMNH 117801	203.14	0.00121	0.0239	cranium mandible
		AMNH 117802	149.97	0.00049	0.0264	cranium mandible
<i>Paleosuchus</i>	<i>palpebrosus</i>	AMNH 93812	164.41	0.00071	0.0210	cranium mandible
		AMNH 97328	212.42	0.00112	0.0241	cranium mandible
<i>Paleosuchus</i>	<i>trigonatus</i>	AMNH 58136	228.20	0.00091	0.0291	cranium mandible
		AMNH 137174	188.83	0.00055	0.0161	cranium mandible
<i>Tomistoma</i>	<i>schlegelii</i>	AMNH 113078	449.56	0.00033	0.0180	cranium mandible

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N	m	b	r²	p	0.95 confidence	
17:20	0.3718	0.0342	0.8488	<0.0001	0.2855	0.4582
18:20	0.4203	0.0390	0.8820	<0.0001	0.3388	0.5018
15:19	0.4228	0.0608	0.8398	<0.0001	0.3122	0.5334
14:19	0.3898	0.0604	0.8965	<0.0001	0.3065	0.4731
15:19	0.4037	0.0415	0.8718	<0.0001	0.3109	0.4964
15:19	0.3147	0.0779	0.8095	<0.0001	0.2232	0.4062
13:20	0.5163	-0.0207	0.8703	<0.0001	0.3841	0.6486
15:20	0.4553	-0.0080	0.9522	<0.0001	0.3942	0.5165
14:20	0.4812	-0.0605	0.8360	<0.0001	0.3472	0.6153
16:20	0.4257	-0.0180	0.8304	<0.0001	0.3154	0.5359
15:18	0.4179	-0.0520	0.8805	<0.0001	0.3257	0.5102
11:15	0.4049	-0.0424	0.9060	<0.0001	0.3066	0.5033
10:18	0.4042	-0.0580	0.7943	0.0005	0.2365	0.5720
13:15	0.4218	-0.0301	0.8908	<0.0001	0.3238	0.5199
16:19	0.3287	-0.0064	0.8035	<0.0001	0.2355	0.4219
15:15	0.3485	-0.0419	0.8417	<0.0001	0.2579	0.4391
11:19	0.3530	0.0639	0.8241	0.0001	0.2300	0.4760
11:15	0.4699	-0.0221	0.8724	<0.0001	0.3344	0.6054
9:19	0.3473	-0.0252	0.9072	0.0001	0.2480	0.4465
12:15	0.4753	0.0113	0.9176	<0.0001	0.3750	0.5756
14:19	0.4691	-0.0254	0.8365	<0.0001	0.3386	0.5995
9:15	0.3642	-0.0143	0.8966	0.0001	0.2537	0.4748
11:19	0.4053	-0.0970	0.8253	0.0001	0.2646	0.5459
10:15	0.3515	-0.1030	0.8275	0.0003	0.2207	0.4824
15:19	0.4037	-0.0142	0.8783	<0.0001	0.3137	0.4938
9:15	0.2896	-0.0392	0.7868	0.0014	0.1549	0.4244
8:19	0.6219	-0.0505	0.8926	0.0004	0.4064	0.8373
12:15	0.5154	-0.0615	0.8591	<0.0001	0.3683	0.6625
16:19	0.6298	-0.0499	0.8213	<0.0001	0.4614	0.7981

12:15	0.3921	-0.0374	0.8814	<0.0001	0.2907	0.4934
22:28	0.0963	-0.2404	0.4500	0.0006	0.0466	0.1459
24:26	0.1942	-0.2153	0.5749	<0.0001	0.1204	0.2681
15:18	0.3457	-0.1549	0.8265	<0.0001	0.2508	0.4406
15:15	0.2757	-0.1393	0.9136	<0.0001	0.2249	0.3265
12:17	0.4274	-0.0094	0.7938	0.0001	0.2739	0.5809
9:14	0.5613	-0.0506	0.8028	0.0011	0.3127	0.8100
12:17	0.4923	-0.0301	0.8541	<0.0001	0.3489	0.6356
11:14	0.4432	0.0027	0.8192	0.0001	0.2862	0.6002
12:17	0.4296	0.0134	0.6648	0.0001	0.2547	0.6045
14:14	0.4656	0.0163	0.8694	<0.0001	0.3520	0.5791
19:20	0.4627	-0.0214	0.8683	<0.0001	0.3705	0.5549
21:22	0.3804	0.0034	0.8727	<0.0001	0.3106	0.4501
18:20	0.4648	-0.0341	0.8598	<0.0001	0.3654	0.5643
13:22	0.4898	0.0015	0.8896	<0.0001	0.3753	0.6043
16:20	0.5404	-0.0660	0.9131	<0.0001	0.4448	0.6359
21:22	0.4078	0.0248	0.7765	<0.0001	0.3028	0.5129
14:20	0.5228	-0.0732	0.8433	<0.0001	0.3811	0.6646
15:22	0.3673	-0.0074	0.7522	<0.0001	0.2410	0.4937
13:21	0.3346	-0.1641	0.7179	0.0003	0.1954	0.4738
18:19	0.2887	-0.1628	0.8109	<0.0001	0.2148	0.3626

Table 2(on next page)

Size, disparity, and regression information for fossil crocodylian specimens.

Head lengths are in millimeters. Morphological disparity (MD) is for all teeth for each specimen. “N” represents number for teeth available versus number of positions total. Regression information is for Principal Component 1 plotted against tooth position.

Genus	Species	Number	Head Length	MD (size)	MD (shape)	Host bone
<i>Alligator</i>	<i>prenasalis</i>	ROM 01375	317.20	0.00030	0.0143	cranium
<i>Allognathosuchus</i>	<i>sp.</i>	UCMP 150180	N/A	N/A	0.0511	mandible
<i>Argochampsa</i>	<i>krebsi</i>	ROM 73872	N/A	N/A	0.0136	mandible
<i>Borealosuchus</i>	<i>sternbergii</i>	NMNH 6533	336.21	0.00003	0.0123	cranium
		UCMP 126099	209.05	0.00070	0.0201	cranium
		UCMP 130435	N/A	N/A	0.0210	mandible
		UCMP 131769	N/A	N/A	0.0158	mandible
<i>Boveriosuchus</i>	<i>vorax</i>	UCMP 170767	N/A	N/A	0.0114	mandible
<i>Brachychampsa</i>	<i>sp.</i>	ROM 68491	624.15	0.00032	0.0108	cranium mandible
<i>Caiman</i>	<i>crocodilus</i>	UCMP 42844	301.93	0.00084	0.0186	cranium mandible
"Crocodylus"	<i>affinis</i>	UCMP 131090	534.10	0.00037	0.0161	cranium
		NMNH 4048	N/A	N/A	0.0118	mandible
<i>Hamadasuchus</i>	<i>rebouli</i>	ROM 52620	416.92	0.00108	0.0272	cranium
<i>Leidyosuchus</i>	<i>canadensis</i>	ROM 01903	463.35	0.00069	0.0202	cranium
<i>Leidyosuchus</i>	<i>sp.</i>	ROM 1425	N/A	N/A	0.0136	mandible

1

2

N	m	b	r²	p	0.95 confidence	
7:16	0.2430	0.1597	0.9638	0.0001	0.1888	0.2971
4:19	0.4690	0.0785	0.9938	0.0031	0.3561	0.5818
5:17	0.5499	-0.1334	0.9562	0.0039	0.3336	0.7661
8:23	0.4539	0.0546	0.9179	0.0002	0.3183	0.5894
7:23	0.5223	-0.0850	0.9026	0.0010	0.3250	0.7196
5:20	0.5318	-0.0320	0.9798	0.0012	0.3914	0.6721
11:20	0.3622	-0.0103	0.7930	0.0002	0.2227	0.5017
6:18	0.5937	0.0753	0.9062	0.0034	0.3286	0.8589
12:18	0.1981	0.1923	0.9162	0.0000	0.1559	0.2403
6:20	0.1559	0.2097	0.8106	0.0144	0.0513	0.2606
7:20	0.3036	-0.0521	0.7237	0.0152	0.0879	0.5193
14:20	0.4063	-0.0140	0.8369	0.0000	0.2935	0.5191
11:15	0.2000	0.1133	0.9363	0.0000	0.1607	0.2394
6:19	0.2025	0.1932	0.5669	0.0840	-0.0432	0.4483
14:20	0.5420	-0.0274	0.9550	0.0000	0.4680	0.6160
20:23	0.3967	0.0303	0.9522	0.0000	0.3527	0.4408
8:19	0.3365	0.0486	0.9570	0.0000	0.2653	0.4078

Figure 1(on next page)

Methodology for data collection.

A) skulls were selected based on number and condition of teeth present (data was not collected from this photo). B) Each tooth was photographed individually, and the margins were traced (Rohlf, 2016). C) Each outline was converted to 30 equidistant semilandmarks which were then slid. D) Tooth shape variance was represented by vector diagrams, with the mean depicted as points and deviation by vector lines.

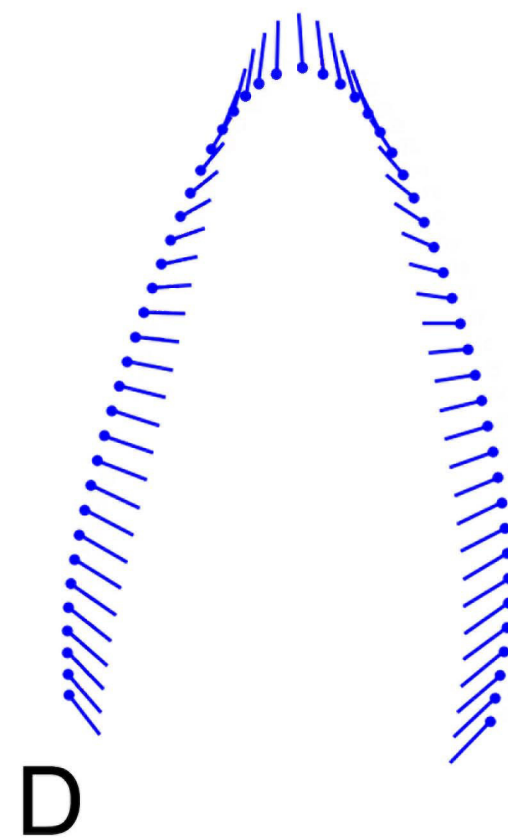
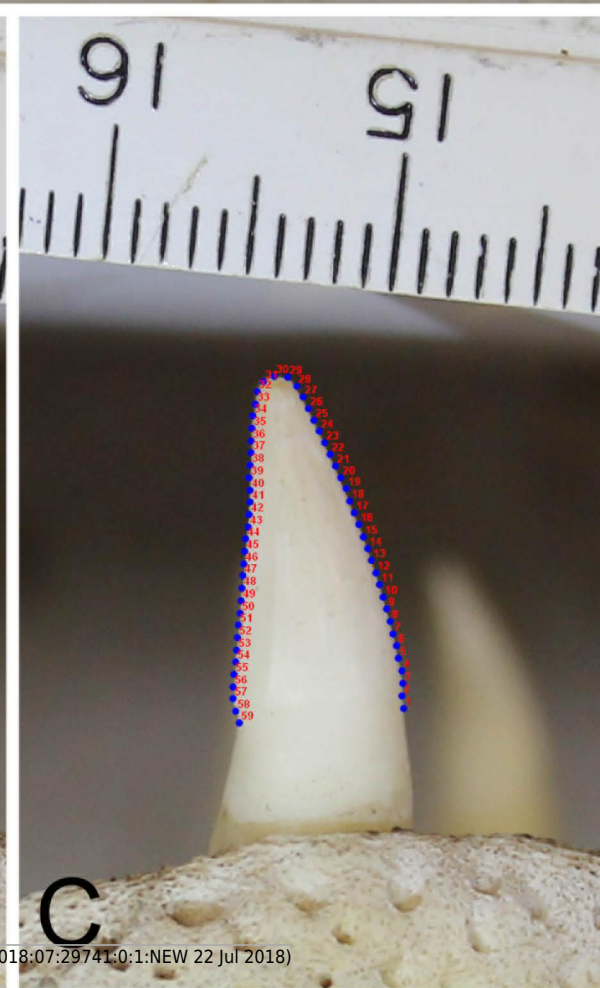
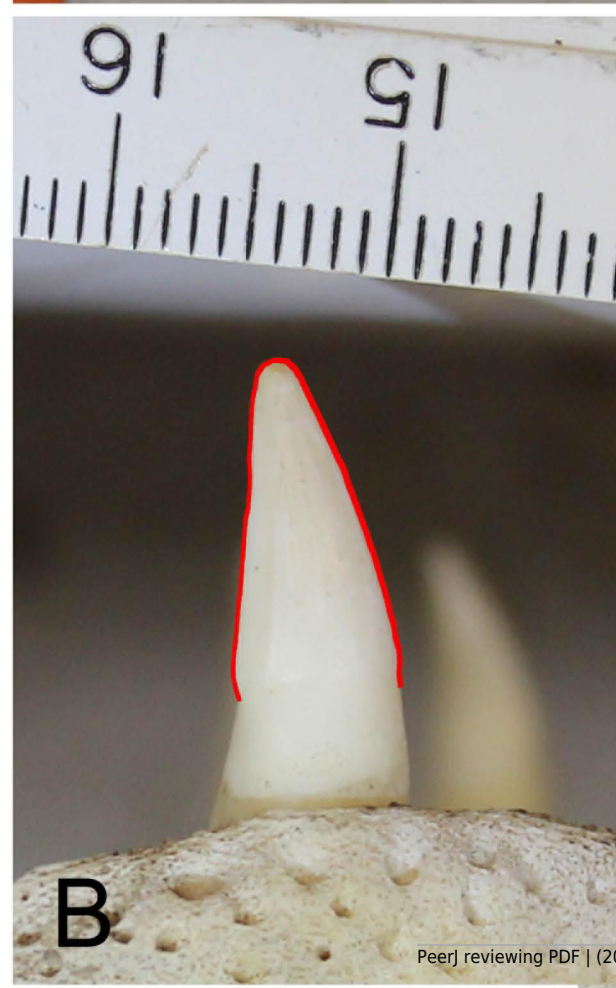


Figure 2 (on next page)

Principal component one.

Vector diagrams indicate the maximum range of variance (vectors) from the mean (points) for both cranial and dentary teeth.

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Figure 3 (on next page)

Average tooth size (above) and shape (below) plotted against skull shape.

Gray error bars indicate standard deviation as a consequence of heterodonty. Solid markers depict extant specimens, and hollow markers depict extinct. The solid regression line indicates only extant specimens, while the dashed line indicates all specimens. Maximum shape range is represented through vector diagrams.

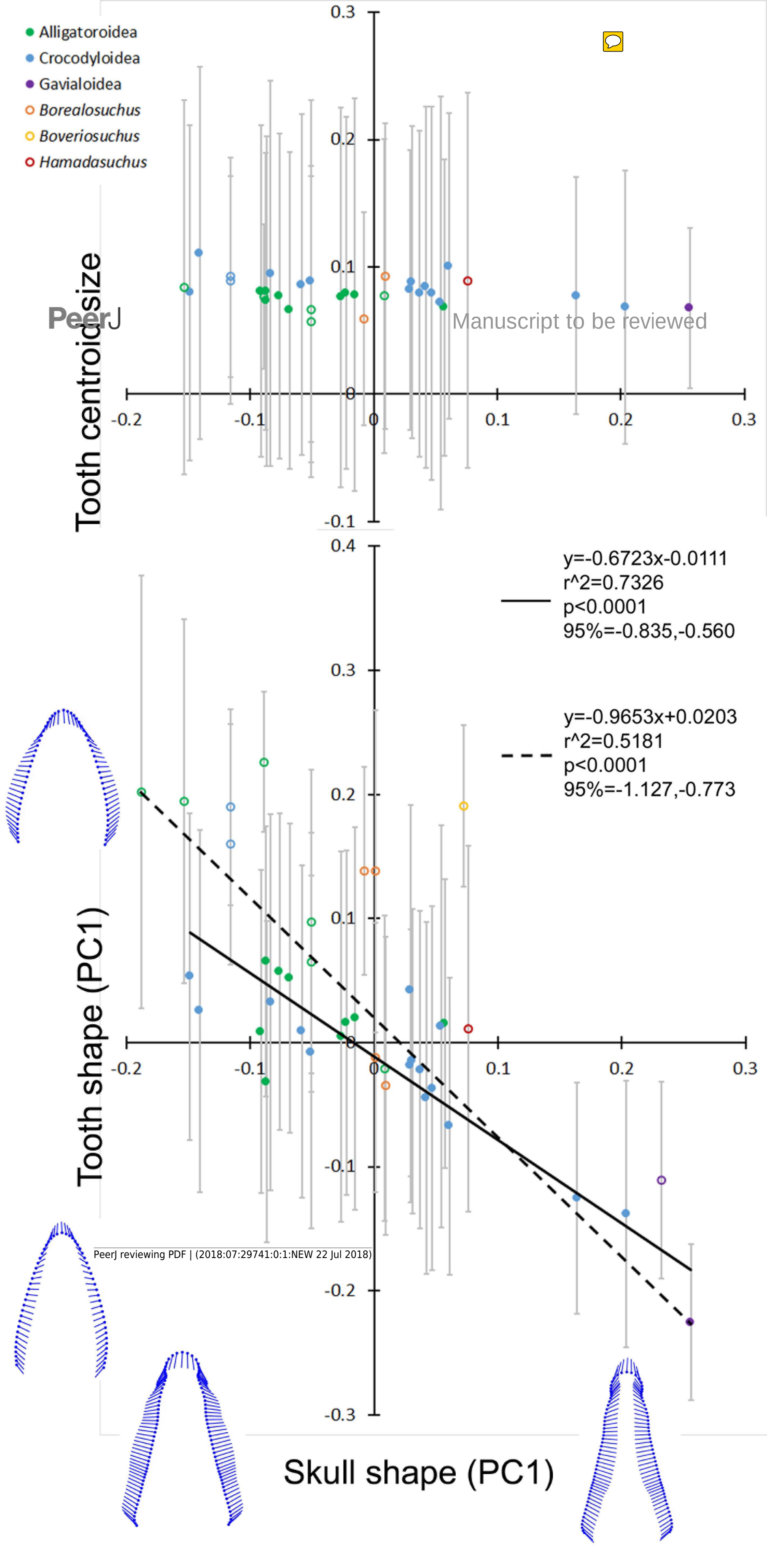


Figure 4(on next page)

Foot's morphological disparity (A, B) and ranges (C, D) for both size and shape.

Size is represented by centroid size and shape is represented by Principal Component 1. For each species, all teeth in the sample are represented. For MD, multiple individuals from a single species are listed in a single column, and those with lower values are superimposed in front of those with higher values. Ranges are a composite of all members of a species.

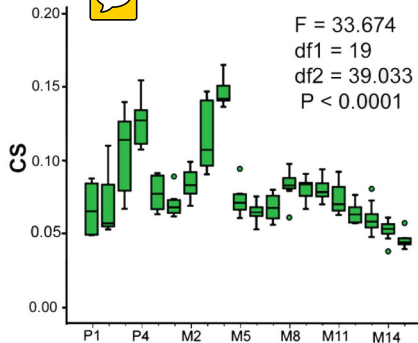
Figure 5(on next page)

Heterodonty by tooth position.

Centroid Size (CS) and **Principal Component one** (PC1) for extant Alligatoroidea (**top**), Crocodyloidea (middle), and Gavialoidae (bottom) plotted against position along the arcade. Welch's ANOVA output comparing positions is listed for each graph with multiple specimens.

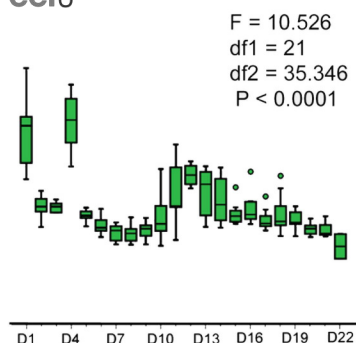


Cranium



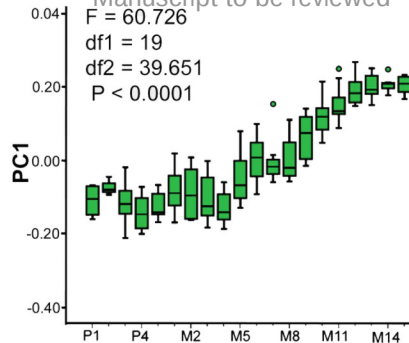
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Dentary

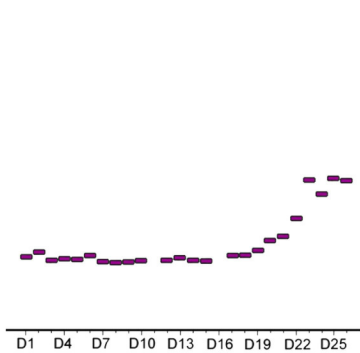
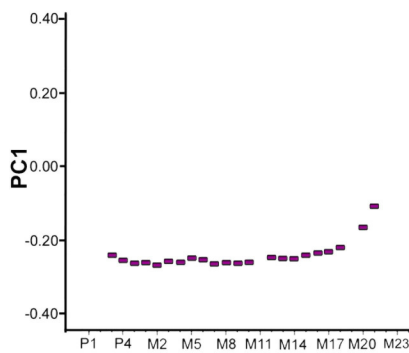
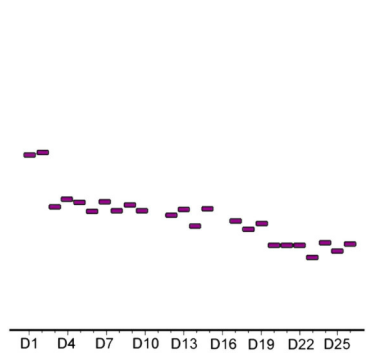
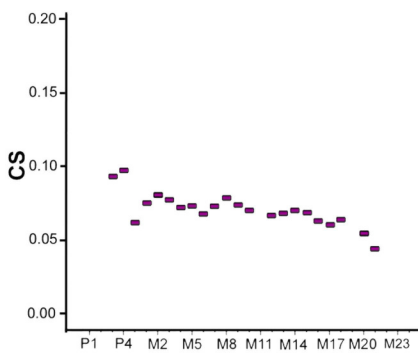
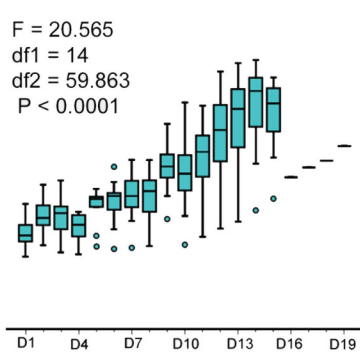
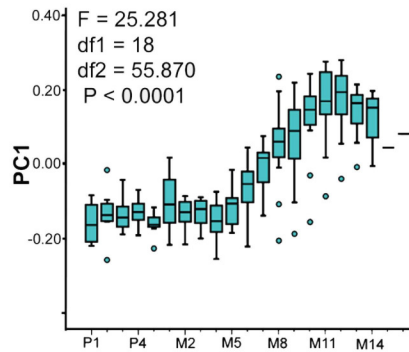
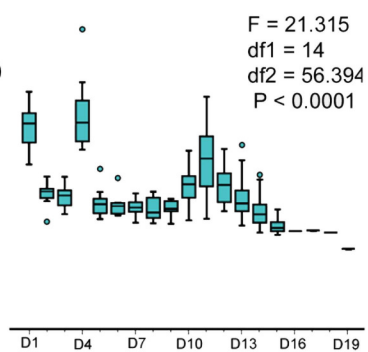
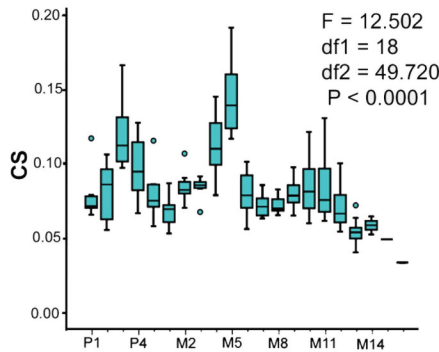
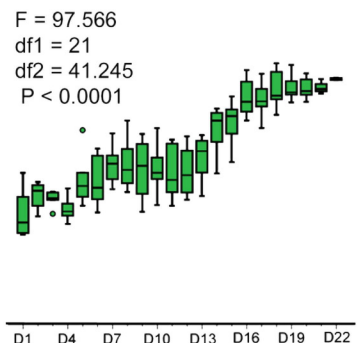


Manuscript to be reviewed

Cranium



Dentary



Position

Position

Position

Position

Figure 6(on next page)

Direct comparisons between selected extant and extinct taxa.

The “size” axis represents normalized centroid size, and the “shape” axis represents principal component one. A) modern taxa with high molariformy, B) Caimaninae with high size-heterodonty, C) longirostrine taxa, D) mid-heterodont fossil taxa, E) crocodyloids and peirosaurids with high-heterodonty, F) fossil globidont taxa. Note: *Allognathosuchus*, *Argochampsa krebsi*, and *B. sternbergii* were not size normalized by their own skull length, as indicated by empty symbols.

