

Taxifolin protects rat against myocardial ischemia/reperfusion injury through modulating mitochondrial apoptosis pathway

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Background: Taxifolin (TAX), a flavonoid has been reported to play an underlying protective role on the cardiovascular disease. Thus, this study is to evaluate the effect and potential mechanisms of TAX on ischemia/reperfusion (I/R) injury. **Methods:** Healthy heart of rat was subjected to I/R using Langendorff apparatus. The hemodynamic parameters were recorded during the perfusion, including heart rate (HR), the left ventricular developed pressure (LVDP), the maximum/minimum rate of left ventricular pressure rise ($+dp/dt_{max}$ and $-dp/dt_{min}$) and Rate pressure product (RPP). Histopathological examination of left ventricular was measured by using Hematoxylin-Eosin (HE) staining. Creatinekinase-MB (CK-MB) and lactate dehydrogenase (LDH) activities in effluent perfusion, and malondialdehyde (MDA), superoxide dismutase (SOD) and glutathione peroxidase (GSH-PX) levels in tissue were assayed. Apoptosis related proteins, such as B-cell lymphoma-2 (Bcl-2), Bcl2-associated X (Bax) and cytochrome c (Cyt-c) also were assayed by elisa. Western blot was employed to determine apoptosis-executive proteins, including caspase 3 and caspase 9. **Results:** The results demonstrated that TAX treatment significantly improve ventricular functional recovery, as evident by the increase in LVDP, $+dp/dt_{max}$, $-dp/dt_{min}$ and RPP, increased the levels of SOD, GSH-PX, whereas suppressed the levels of LDH, CK-MB, MDA. Furthermore, TAX upregulate the level of Bcl-2 protein and downregulate the level of Bax, Cyt-c, caspase 3, caspase 9 protein. **Discussion:** Our results indicated that treatment of TAX remarkably improve cardiac function, regulate oxidative stress and attenuate apoptosis. It is concluded that TAX has a cardioprotective effect against I/R injury through the modulation of mitochondrial apoptosis pathway.

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19

20 **ABSTRACT**

21 **Background:** Taxifolin (TAX), a flavonoid has been reported to play an underlying protective
22 role on the cardiovascular disease. Thus, this study is to evaluate the effect and potential
23 mechanisms of TAX on ischemia/reperfusion (I/R) injury.

24 **Methods:** Healthy heart of rat was subjected to I/R using Langendorff apparatus. The
25 hemodynamic parameters were recorded during the perfusion, including heart rate (HR), the left
26 ventricular developed pressure (LVDP), the maximum/minimum rate of left ventricular pressure
27 rise (+dp/dt_{max} and -dp/dt_{min}) and Rate pressure product (RPP). Histopathological examination of
28 left ventricular was measured by using Hematoxylin-Eosin (HE) staining. Creatinekinase-MB
29 (CK-MB) and lactate dehydrogenase (LDH) activities in effluent perfusion, and

30 malondialdehyde (MDA), superoxide dismutase (SOD) and glutathione peroxidase (GSH-PX)
31 levels in tissue were assayed. Apoptosis related proteins, such as B-cell lymphoma-2 (Bcl-2),
32 Bcl2-associated X (Bax) and cytochrome c (Cyt-c) also were assayed by elisa. Western blot was
33 employed to determine apoptosis-executive proteins, including caspase 3 and caspase 9.

34 **Results:** The results demonstrated that TAX treatment significantly improve ventricular
35 functional recovery, as evident by the increase in LVDP, $+dp/dt_{max}$, $-dp/dt_{min}$ and RPP, increased
36 the levels of SOD, GSH-PX, whereas suppressed the levels of LDH, CK-MB, MDA.
37 Furthermore, TAX upregulate the level of Bcl-2 protein and downregulate the level of Bax, Cyt-
38 c, caspase 3, caspase 9 protein.

39 **Discussion:** Our results indicated that treatment of TAX remarkably improve cardiac function,
40 regulate oxidative stress and attenuate apoptosis. It is concluded that TAX has a cardioprotective
41 effect against I/R injury through the modulation of mitochondrial apoptosis pathway.

42

43 INTRODUCTION

44 Ischemic heart disease is a threat to human health. Extracorporeal circulation and coronary
45 bypass surgery are usually carried out to improve myocardial ischaemia after myocardial
46 infarction occurs. However, when blood supply returns to heart after a period of ischemia will
47 cause additional damage to the heart. The condition was referred to as cardiac
48 ischaemia/reperfusion (I/R) injury (*Braunwald et al., 2012*). Nowadays, I/R injury is a major
49 factor, which often cause to death. During myocardial I/R injury, cardiomyocytes undergo death
50 at a higher frequency, mainly including necrosis and apoptosis (*Gottlieb et al., 1994*). Apoptosis
51 is programmed cell death, which is considered to be the vital pathological processes in acute
52 reperfusion injury (*Konstantinidis et al., 2012*). When the amount of cardiomyocyte decreases,
53 the heart may undergo ventricular remodelling, compensatory cardiac hypertrophy and
54 eventually lead to heart failure (*Pangonyte et al., 2008; Du et al., 2010*). Therefore, exploring the
55 detailed mechanisms that trigger cardiomyocytes death and how to prevent it during I/R injury is
56 still a public issue.

57 In the I/R injury, the morphological changes of cardiomyocytes were observed include
58 microvascular damage, myocardial cell edema, and the body symptoms include myocardial
59 enzyme release, arrhythmias and weak systolic function (*Naito et al., 2000*). It is generally
60 believed that this reperfusion damage is caused by increased free radical activity. When
61 circulating blood decreases, the level of oxygen supply can't keep up with oxygen demand by
62 cardiomyocytes, the aerobic metabolism turns into anaerobic metabolism (*Giordano et al., 2005*).
63 Anaerobic metabolism leads to the production of lactic acid which results in disturbances of
64 ionic homeostasis. A timely reperfusion is crucial for recovery of an ischemic myocardium, but
65 by sudden re-appearance of circulating blood to the dying myocardium, increased reactive
66 oxygen species (ROS) will be produced in reaction to hyperoxia which can make the functional

67 situation of organization worse (*Akhlaghi et al., 2009*).

68 Flavonoid is the most prevalent class of naturally occurring compound and ubiquity in woody
69 and herb plants. It exerts multiple biochemical properties and wide pharmacological effects
70 (*Moon et al., 2006*). Epidemiological studies have shown that flavonoids is associated with a
71 reduced risk of cardiovascular diseases (*Raj Narayana et al., 2001*; *Bjorklund et al., 2017*).
72 Fisetin, a plant-derived bioflavonoid, significantly attenuated the I/R-induced tissue injury,
73 blunted the oxidative stress, and restored the structure and function of mitochondria
74 (*Shanmugam et al., 2018*). Quercetin has been demonstrated improves post ischemic recovery of
75 isolated heart of rats after global ischemia (*Barteková et al., 2010*). Previous research has
76 revealed that TAX exerts anti-inflammatory effects and protects against oxidative stress-induced
77 injury in human endothelial cells (*Guo et al., 2015*) and rat peritoneal macrophages (*Arutyunyan*
78 *et al., 2016*). It also exhibited to possess free radical scavenging, antioxidant and anti-
79 inflammatory effects (*Sun et al., 2014*; *Xie et al., 2017*). Recent studies demonstrated that TAX
80 involved in the amelioration of cardiac disorders associated with against cardiac hypertrophy and
81 fibrosis (*Guo et al., 2015*). TAX is structurally similar to quercetin potentially beneficial effects
82 on the cardiovascular system. But its potential for I/R protection remain unclear. TAX is a
83 potential candidate for the prevention or treatment of I/R injury. However, there was no previous
84 report about whether TAX could influence the injury of I/R in isolated rat hearts. In present
85 study, we aimed to evaluate the cardioprotective effects of TAX and investigated the
86 mechanisms underlying these effects in isolated hearts of rats.

87

88 **MATERIALS & METHODS**

89 **Experimental animals and treatment**

90 Male SD rats (280-300 g each) were obtained from the Laboratory Animal Center of
91 Heilongjiang Medicine University Medical (License Number: SCXK (hei) 2016-0002). The rats
92 were housed under standard conditions with natural light (12 hr) and dark (12 hr) and in a
93 suitable temperature at 22 ± 2 °C. The rats were fed common feedstuff and drank tap water freely
94 during the experimental period. The investigation conformed to Guide for the Care and Use of
95 Laboratory Animals (revised, 1996). All animal experiments were approved by College of
96 Pharmacy of Heilongjiang University of Chinese Medicine, Animal Ethics Committee (Approval
97 number: IACUC-20170903019).

98

99 **Reagents and antibodies**

100 TAX (purity $\geq 98\%$) was purchased from Sigma-Aldrich (St. Louis, MO, USA).
101 Creatinekinase-MB (CK-MB), lactate dehydrogenase (LDH), malondialdehyde (MDA)
102 glutathione peroxidase (GSH-PX) and superoxide dismutase (SOD) assay kit was obtained from
103 Nanjing Jiancheng Bioengineering Institute (Jiangsu, China). Enzyme-linked immunosorbent
104 assay kit about mitochondrial apoptosis (B-cell lymphoma-2 (Bcl-2), Bcl2-associated X (Bax)
105 and cytochrome c (Cyt-c)) was obtained from Cloud-Clone Corp (Houston, TX, USA).

106 Monoclonal primary antibodies anti- β -actin, anti-active caspase 3 and anti-active caspase 9 was
107 purchased from Abcam (MA, USA). Fluorescent-labeled goat anti-rabbit IgG secondary
108 antibodies were obtained from LI-COR Biosciences (Lincoln, Nebraska, USA).

109

110 **Experimental protocol**

111 The rats were randomly divided into four groups (n=8): Normal control group (Control);
112 Myocardial I/R control group (I/R); I/R + TAX treatment group (TAX 5 μ M); I/R + TAX
113 treatment group (TAX 15 μ M). Experimental protocol is shown in Fig. 1. Control group: The
114 hearts were subjected to a continuous perfusion of Krebs–Henseleit (K–H) solution for 120 min.
115 I/R group: The hearts were equilibrated for 30 min, subsequently, global ischemia was
116 performed at 37 °C for 30 min, and next, re-perfused with K–H solution for 60 min. TAX 5 μ M
117 group: The hearts were equilibrated for 30 min, subsequently, global ischemia was performed for
118 30 min at 37 °C, and next, perfused with 5 μ M of TAX-saturated K–H solution for 60 min. TAX
119 15 μ M group: The hearts were equilibrated for 30 min. Subsequently, global ischemia was
120 performed for 30 min, and next, perfused with 15 μ M of TAX-saturated K–H solution for 60 min.

121

122 **Langendorff preparation**

123 After anesthetization via intraperitoneal injection (chloral hydrate solution, 300 mg/kg), rat
124 hearts were quickly removed and subsequently perfused in the Langendorff apparatus. The
125 perfusion was performed for 30 min in a modified K-H buffer gassed with 95 % O₂ and 5 % CO₂
126 at a constant flow velocity and constant temperature (37 °C). The composition of K-H buffer was
127 the following (mM): NaCl 118, MgSO₄ 1.2, KCl 3.2, NaHCO₃ 25, KH₂PO₄ 1.18, CaCl₂ 2.5 and
128 glucose 5.5. After equilibration, 30 min global ischemia was induced followed by 60 min of
129 reperfusion. Control group utilizing the same protocol, but with no ischemia induction was used.
130 Water-filled balloon inserted into the left ventricular cavity was used to monitor hemodynamic
131 parameters. The left ventricular end-diastolic pressure (LVEDP) was maintained at 5-10 mmHg
132 by adjusting the size and position of the balloon. The whole procedure was completed within 2
133 min. The inclusion criteria of experimental samples were a heart rate (HR) of > 250 beats/min
134 and a left ventricular developed pressure (LVDP) of > 75 mmHg in equilibrium phase. The
135 hemodynamic parameters were recorded during the perfusion, including heart rate (HR), LVDP,
136 the maximum/minimum rate of left ventricular pressure rise (+dp/dt_{max} and -dp/dt_{min}). Rate
137 Pressure Product (RPP) = HR × LVDP.

138

139 **Histopathological evaluation of left ventricle sections**

140 For histopathological examination, the cut left ventricle of heart tissues fixed in 10 % neutral
141 formalin at room temperature. After two hours, the flat tissue blocks were embedded in paraffin
142 to make it blocks and then cut into 3 μ m thick tissue sections and stained with H&E. At least
143 three samples from each group were evaluated. The tissue sections were visualized under light
144 microscope (Dewinter technologies, Italy).

145

146 **Estimation of cardiac damage**

147 In present study, heart tissue injury was assessed by determining the concentration of LDH
148 and CK-MB in the perfusate. The LDH and CK-MB content in the perfusate were measured with
149 the assay kit as per the manufacturer's instructions. Samples of the perfusate were collected from
150 the isolated heart at 25 min, 63 min, 90 min and 120 min of perfusion.

151

152 **Measurements of anti-oxidant indices**

153 The hearts tissue was cut into small pieces of tissue and then was grinded with lysate using
154 glass homogenizer. Supernatant of tissue homogenate was frozen for each tissue analysis. MDA,
155 SOD and GSH-PX activity were assessed using commercial elisa kits following the
156 manufacturer's instructions. All enzyme activities were normalized to the total protein
157 concentrations, which were determined using a bicinchoninic acid (BCA) protein assay kit
158 (Beyotime, Shanghai, China).

159

160 **Estimation of Cyt-c, Bcl-2 and Bax levels**

161 Heart tissue samples were weighed, and then minced the tissues to small pieces and
162 homogenized them in specific lysis buffer (w: v = 1:30, 1 mL lysis buffer is added in 30 mg
163 tissue sample) with a glass homogenizer on ice. Then, the homogenates were centrifugated for 5
164 min and collect the supernatant. The levels of Bcl-2 and Bax protein were measured according to
165 manufacturer's instructions of commercial kits (Cloud Clone Corp, Houston, USA).

166

167 **Western blotting analysis**

168 Myocardial tissue samples were lysed with RIPA buffer containing protease inhibitors for 15
169 minutes on ice. The total lysates were clarified by centrifugation, and supernatants were
170 collected. Protein samples (20-25 mg per lane) were loaded on the gels and then separated by 10
171 % sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) under reducing
172 conditions and transferred onto the nitrocellulose membrane (Roche, Mannheim, Germany). The
173 membrane was washed with PBS with 0.1% Tween-20 (PBST) and blocked with 5% skim milk
174 in shaking table for 2 h at room temperature. Then the membrane was washed with PBST and
175 incubated with antigen-specific rabbit IgG antibodies (anti-caspase 3, anti-caspase 9, anti- β -actin;
176 Abcam) diluted 1:1000 in PBST. Next, the membrane was washed with PBST and incubated
177 with fluorescent-labeled goat anti-rabbit secondary antibodies (Lincoln, Nebraska, USA) diluted
178 1:2500 in PBST for 2 h at 4 °C. The target protein bands were scanned using the blot imaging
179 system GelLogic 212 PRO (Carestream, Rochester, NY, USA) after washed in PBST. The
180 obtained images were quantified as final results by image J 1.4.3 (www.imagej.nih.gov/ij). The
181 results were expressed as the fold induction, which compared with the normal control.

182

183 **Statistical Analysis**

184 SPSS16.0 for Windows (SPSS Inc., Chicago, USA) was used for statistical analysis. All data
185 were expressed as mean \pm standard deviation (SD). For comparisons between groups, the one-

186 way ANOVA or student T-test was used where appropriate. The statistical differences was
187 considered significant when $P < 0.05$. # $P < 0.05$ and ## $P < 0.01$ vs. Control group. * $P < 0.01$
188 and ** $P < 0.001$ vs. IR group.

189

190 RESULTS

191 Effects of TAX on Cardiac Parameters of Isolated Hearts

192 We examined whether TAX could protect the hearts of rat against ex vivo I/R injury. The
193 results showed that there was no obvious alteration in average HR of isolated heart in period of
194 reperfusion with TAX or without TAX. In addition, there was no significant HR change between
195 I/R and normal groups during 30 min ischemia and 60 min reperfusion periods (Fig. 2A). After
196 reperfusion, LVDP, $+dp/dt_{max}$ and $-dp/dt_{min}$ from different treatment groups were decreased in
197 varying degrees. For instance, LVDP was significantly increased in TAX 15 μM group compared
198 with I/R group at the end periods of reperfusion (LVDP = 68 mmHg vs. 52 mmHg, $P < 0.05$,
199 Fig. 2B). Compared with the I/R group, TAX 15 μM treatment also significantly improved the
200 heart functional recovery in rat at 60 min of reperfusion (RPP = 15294 mmHg \times beats/min in
201 TAX vs. 10643 mmHg \times beats/min in I/R, $P < 0.01$, Fig. 2E). The results showed that TAX
202 treatment improved the cardiac function recovery of rats during myocardial I/R injury.

203

204 TAX Down-Regulated the Release of LDH and CK-MB

205 At different time points of perfusion, the heart effluents were perfusate. LDH level in total
206 reperfusion process was not conspicuously altered in normal control group. Perfusate LDH
207 activity in the I/R group was improved compared with that of the normal control group after
208 reperfusion and significantly increased at 60 min of reperfusion ($P < 0.01$), and TAX highly
209 reduce enhanced the LDH levels compared with I/R group at 60 min of reperfusion (Fig. 3A).
210 CK-MB release was similar to LDH release. Total expression in the perfusate was not
211 conspicuously change in the control Group, and CK-MB level was not significantly altered in
212 different group in baseline. However, levels in I/R group were markedly higher at 30 min of
213 reperfusion compared with baseline ($P < 0.05$). Interestingly, the CK-MB release in both TAX 5
214 μM and 15 μM Group was significantly decreased at end of reperfusion compared with I/R
215 Group ($P < 0.05$ or $P < 0.01$) (Fig. 3B). These results indicated that TAX could protect cardiac
216 function against I/R injury.

217

218 Effect of Taxifolin on Myocardial Morphology

219 Histopathological examination of myocardial tissue was assessed by hematoxylin-eosin (HE)
220 staining. Typical micrographs of the myocardial structure are shown in Fig. 4. In control group

221 (Fig. 4A), morphology of myocardial tissue is normal. Cardiomyocytes were arranged closely,
222 intercellular space is small, no edema between cells. In contrast, the I/R group (Fig. 4B) showed
223 degenerated muscle fibers and obvious contraction band, severe obvious cells edema, many
224 infiltrated inflammatory cells. As shown in Fig. 4C, TAX at 5 μ M maintained the myocardium
225 with only slight irregularly arranged of fibers and a few contraction bands. As shown in Fig. 4D,
226 TAX 15 μ M group showed orderly cardiomyocytes but a few cells dissolve and degeneration.
227 The results show that treatment with 15 μ M TAX significantly reduced I/R injury in comparison
228 with I/R group.

229

230 **Effect of TAX on I/R-Induced Oxidative Stress in the Myocardium**

231 To explore the cardio-protective mechanism of TAX, the effects of TAX on SOD, GSH-PX
232 and MDA activity were investigated in myocardial tissue in response to I/R injury. As shown in
233 Fig. 5, in TAX 15 μ M group, the SOD and GSH-PX activity were increased significantly in
234 comparison with those of the I/R group ($P < 0.01$), whereas the group pretreated with 5 μ M TAX
235 showed no significant differences. Conversely, these TAX-treated groups showed that MDA
236 production was reduced significantly ($P < 0.01$) in comparison with I/R group.

237

238 **TAX Protects Myocardial Cell from I/R-Induced Mitochondrial Damage**

239 To evaluate if the effect of TAX is mediated through attenuation of the mitochondrial damage,
240 we determined cytochrome c (Cyt-c) in cytosol. Fig. 6A shows that I/R increased cytosolic Cyt-c
241 levels ($P < 0.01$). By comparison, TAX at different doses could reduce I/R-induced increase of
242 Cyt-c levels ($P < 0.01$). The result suggested that TAX attenuated I/R induced Cyt-c release to
243 cytosol. The bcl-2 family protein modulates the release of Cyt-c by regulating mitochondrial
244 transmembrane potential. We also determined Bcl-2 and Bax protein expression levels.
245 Compared with the control group, I/R down-regulated Bcl-2 but up-regulated Bax protein
246 expression (Figs. 6B-6C). TAX increased Bcl-2 levels but decreased Bax levels compared with
247 I/R. These results indicate that TAX may protect mitochondria attenuate apoptosis by regulating
248 the expression of the Bcl-2 family proteins.

249

250 **TAX Attenuates Myocardial I/R-Induced Apoptosis**

251 To explore the potential mechanism of TAX in rats subjected to I/R-induced myocardial
252 injury, we investigated the proteins expressions of active-caspase 3 and active-caspase 9.
253 Compared with control group, the expression of active-caspase 3 was up-regulated in I/R group.
254 While compared with I/R, TAX treatment group significantly reduced the level of active-caspase
255 3 (Fig. 7C). In TAX treated group, it was showed that the expression of active caspase 9 was

256 down-regulated in 15 μM ($P < 0.05$), but did not changed significantly in 5 μM (Fig. 7D). These
257 results indicated that TAX inhibited apoptotic level in heart injury induced by I/R.

258

259 DISCUSSION

260 Growing evidence indicates a therapeutic action of TAX in cardiovascular disease. However,
261 implications of TAX in I/R injury are unclear. This is first study to evaluate the cardioprotective
262 effects of TAX in isolated rat heart subjected to I/R injury. We demonstrated that an important
263 role for TAX improved cardiac function, inhibited oxidative stress and apoptosis in a model of
264 I/R injury in vitro.

265 In the late 19th century, Oscar Langendorff was studying on perfecting isolated heart model.
266 Since then, the procedure has been used to probe pathophysiology of I/R and with the dawn of
267 molecular biology (Bell *et al.*, 2011). Today, the Langendorff heart assay is a predominant in
268 vitro technique used in physiological and pharmacological research. It allows the examination of
269 cardiac contractile strength and heart rate without the complications of an intact animal (Herr *et*
270 *al.*, 2015). Therefore we determine the effect of TAX to cardiac function parameters of isolated
271 heart in myocardial I/R injury using Langendorff equipment. Cardiac functions mainly depend
272 on the contraction and relaxation properties of the ventricular muscle. Changes in cardiac
273 function are strongly linked to the severity of I/R injury (Mehdizadeh *et al.*, 2013). Our analyze
274 data showed I/R can cause marked myocardial dysfunction, including reduction of LVDP,
275 $+\text{dp}/\text{dt}_{\text{max}}$ and $-\text{dp}/\text{dt}_{\text{min}}$. While the TAX treatment groups significantly improved cardiac diastolic
276 dysfunction but did not alter the average HR in isolated heart.

277 LDH is a marker of cellular damage and common disease due to its mass release to plasma
278 during tissue injuries, such as heart failure. CK-MB, expressed extensively in cardiac myocyte,
279 often tested in the serum as an indicator of damage of rhabdomyolysis, myocardial damage and
280 acute kidney injury in clinic (Moghadam-Kia *et al.*, 2016). The increase in the levels of LDH and
281 CK-MB in the organ perfusate after ischemia is direct evidence of cardiac damage (Houshmand
282 *et al.*, 2009). Compared with control group, LDH and CK-MB activity significantly increased in
283 I/R group during myocardial I/R injury. Perfusate LDH and CK-MB activity in the TAX
284 treatment groups, particularly in the 15 μM group, were remarkable reduction than those of the
285 I/R group, which is consistent with the observation of changing cardiac function parameters. In
286 addition, histopathological examination was implemented to assess the effect of TAX on
287 myocardial morphology. The results of pathomorphological research of heart in I/R group show
288 acute myocardial damage and TAX causes favourable morphological changes in heart in I/R
289 injury. The above results support the potential application of TAX as a cardioprotective agent in
290 myocardial I/R injury.

291 Under normal conditions, tissues could maintain the balance between generation and clearance
292 of ROS. However, the balance was disrupted during I/R and cause ROS increase significantly

293 (*Becker et al., 2004*). Excess ROS can oxidize lipids, proteins and DNA, which cause to
294 dysfunction of these molecules and then result in the degeneration of tissue function (*Kleikers et*
295 *al., 2012*). Minimizing ROS production is an important strategy to prevent cardiomyocyte I/R
296 injury (*He et al., 2016*). Therefore, activation of the anti-oxidant enzyme system is a necessary
297 strategy to reduce oxidative stress-induced tissue damage (*Matsushima et al., 2013*). The levels of
298 SOD and GSH-PX rate are used to evaluate tissue per-oxidative injury (*Maciejczyk et al., 2017*).
299 In addition, MDA, an index to evaluate the severity of lipid peroxidation, produced by lipid
300 peroxidation results in destruction of structural proteins and cellular structures (*Pizzimenti et al.,*
301 *2013*). Our results exhibited that SOD and GSH-PX activity was conspicuous increased, whereas
302 MDA level was dramatically decreased by TAX, especially in 15 μ M group. Therefore, we
303 speculated that the TAX exhibits cardioprotective effects by enhancing antioxidant activity and
304 inhibiting free radicals peroxidation.

305 Mitochondrial damage plays an important role in I/R-induced injury. It considered to as the
306 final arbitrators for I/R-induced cell apoptosis (*Powers et al., 2007*). During ischemia and mainly
307 during early phases of reperfusion, excessive of ROS is generated, causing myocardial Ca²⁺
308 overload and opening of the mitochondrial permeability transition pore (mPTP), which can
309 reduce mitochondrial function and finally result in an increase in myocardial cell apoptosis
310 (*Garciaarena et al., 2011; Halestrap et al., 2015*). One of the ways of cell apoptosis is activated
311 by release Cyt-c from the mitochondria to cytosol. In our study, the results showed that TAX is
312 capable of weakening the observed increase in expression of Cyt-c in cytosol. It is very likely
313 that the increased cytosolic content of Cyt-c, which mediates apoptosis, while its expression in
314 mitochondria was not changed (*Lundberg et al., 2004*). Therefore it can make an assumption that
315 down- regulation of Cyt-c as result of TAX whittled apoptotic processes.

316 As an important mitochondrial regulator during myocardial apoptosis, Bcl-2 exerts anti-
317 apoptotic effects by blocking the release of Cyt-c and reducing caspase activity.³⁴ Apoptosis-
318 related proteins, caspase 3 and caspase 9, are also play crucial roles in apoptosis. The caspase
319 apoptotic pathway responds to death signals by releasing apoptosis-inducing factor from the
320 mitochondrial and then translocated to the nucleus (*D'Amelio et al., 2012*). In this study, Bcl-2,
321 an anti-apoptotic protein and Bax, a pro-apoptotic protein were used to assess the effects of TAX
322 on cardiomyocytes apoptosis. The result demonstrated that TAX treatment increases protein
323 expression of Bcl-2, significantly reduces Bax expression when compared with I/R group.
324 Caspase 3 and Caspase 9 were tested to measure the apoptotic level in the isolated heart after I/R
325 injury. We found that increased expression of active form of caspase 3 and caspase 9 under
326 ischemic conditions and decreased expression of them in TAX group. Consistent with those
327 results, treatment with TAX significantly decreased myocardial apoptosis through regulating
328 expression of apoptosis-related proteins, including Bax, Bcl-2, caspase 3 and caspase 9. These
329 findings suggested that the inhibition of apoptosis is closely related to the underlying beneficial
330 effect of TAX in I/R injury.

331

332 CONCLUSIONS

333 In conclusion, TAX exerted cardioprotective effects against I/R injury by inhibiting oxidative
334 stress and cardiac myocyte apoptosis. These phenomena highlight that the mechanism associated
335 with this protection maybe involves modulation of mitochondrial apoptosis pathway. Our find
336 provides a novel of thought for therapeutic development as an adjuvant therapy to I/R injury.

337

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342

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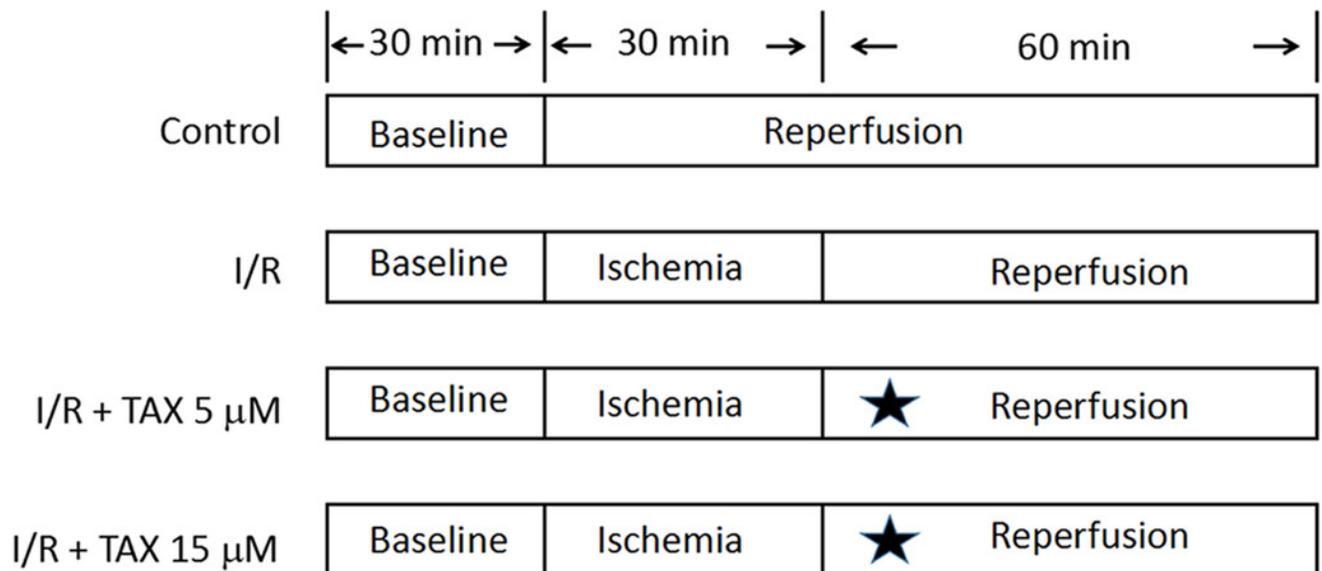
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- 433

Figure 1

Figure 1 Experimental protocol



★ Taxflion and K-H solution for 60 min after 30 min of whole heart ischemia

Figure 2 (on next page)

Figure 2. TAX treatment improved the cardiac function recovery of rats during myocardial I/R injury in vitro model.

(A) Heart rate (HR, beat/per min); (B) left ventricular developed pressure (LVDP, mmHg); (C) maximum rate of left ventricular pressure ($+dp/dt_{max}$, mmHg/s); (D) minimum rate of increase of left ventricular pressure ($-dp/dt_{min}$, mmHg/s); (E) Rate pressure product (RPP, mmHg \times bpm); (F) representative left ventricular pressure records in experimental protocol form different experiment groups.

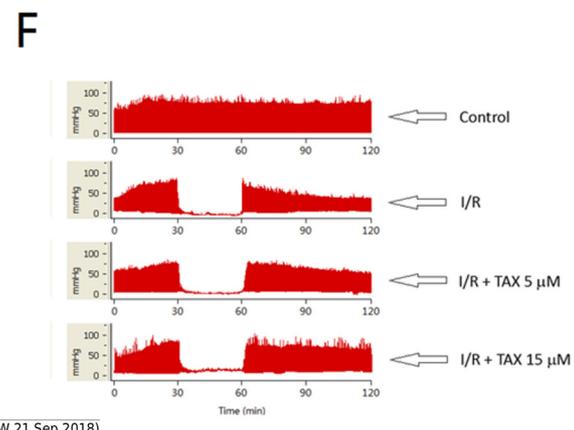
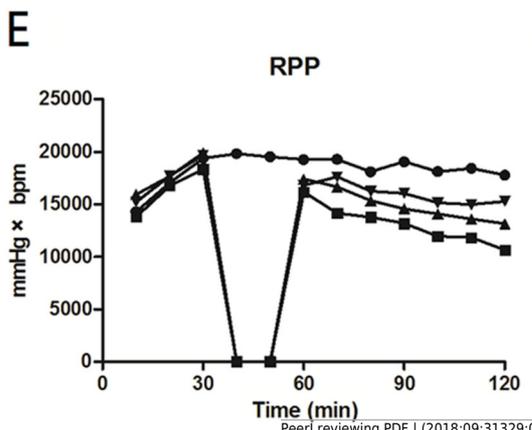
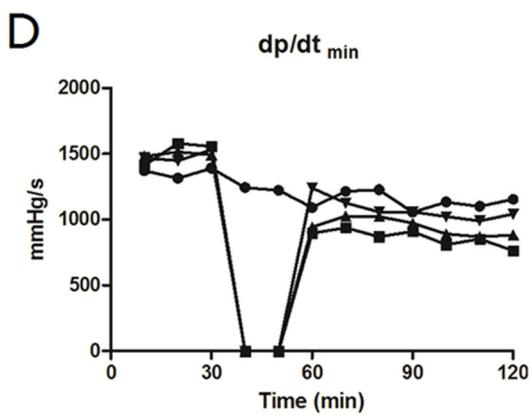
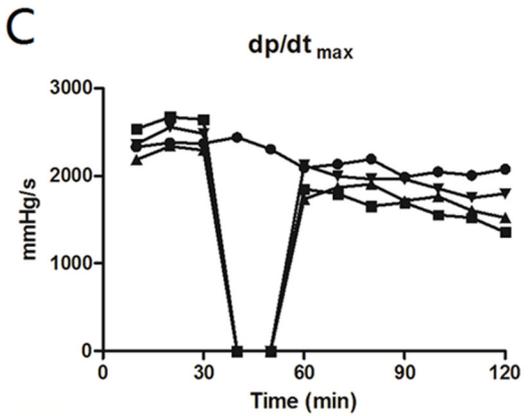
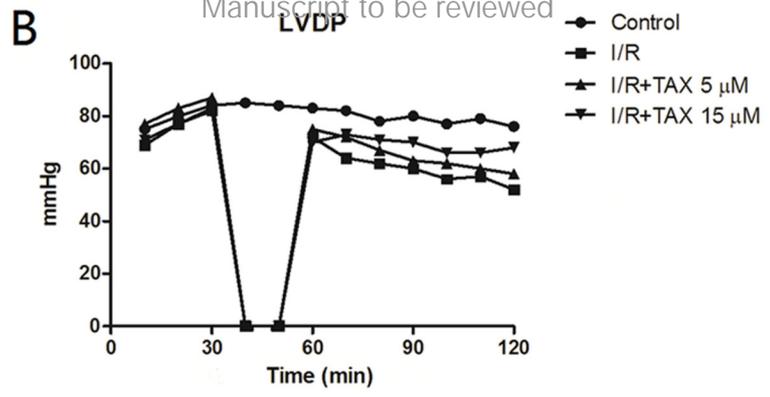
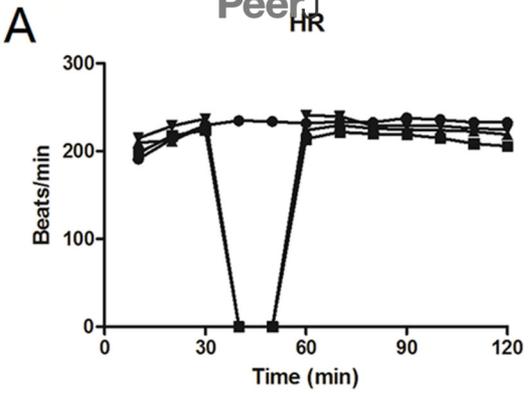


Figure 3(on next page)

Figure 3. Effect of TAX on injury of cardiomyocytes by measurement of LDH and CK-MB.

The levels of perfusate LDH and CK at different time points in the control, I/R and TAX-treat group (5 μ M and 15 μ M) are shown. # $P < 0.05$ and ## $P < 0.01$ compared with the control group; * $P < 0.05$ and ** $P < 0.01$ compared with the I/R group; U/L: international enzyme activity unit per liter.

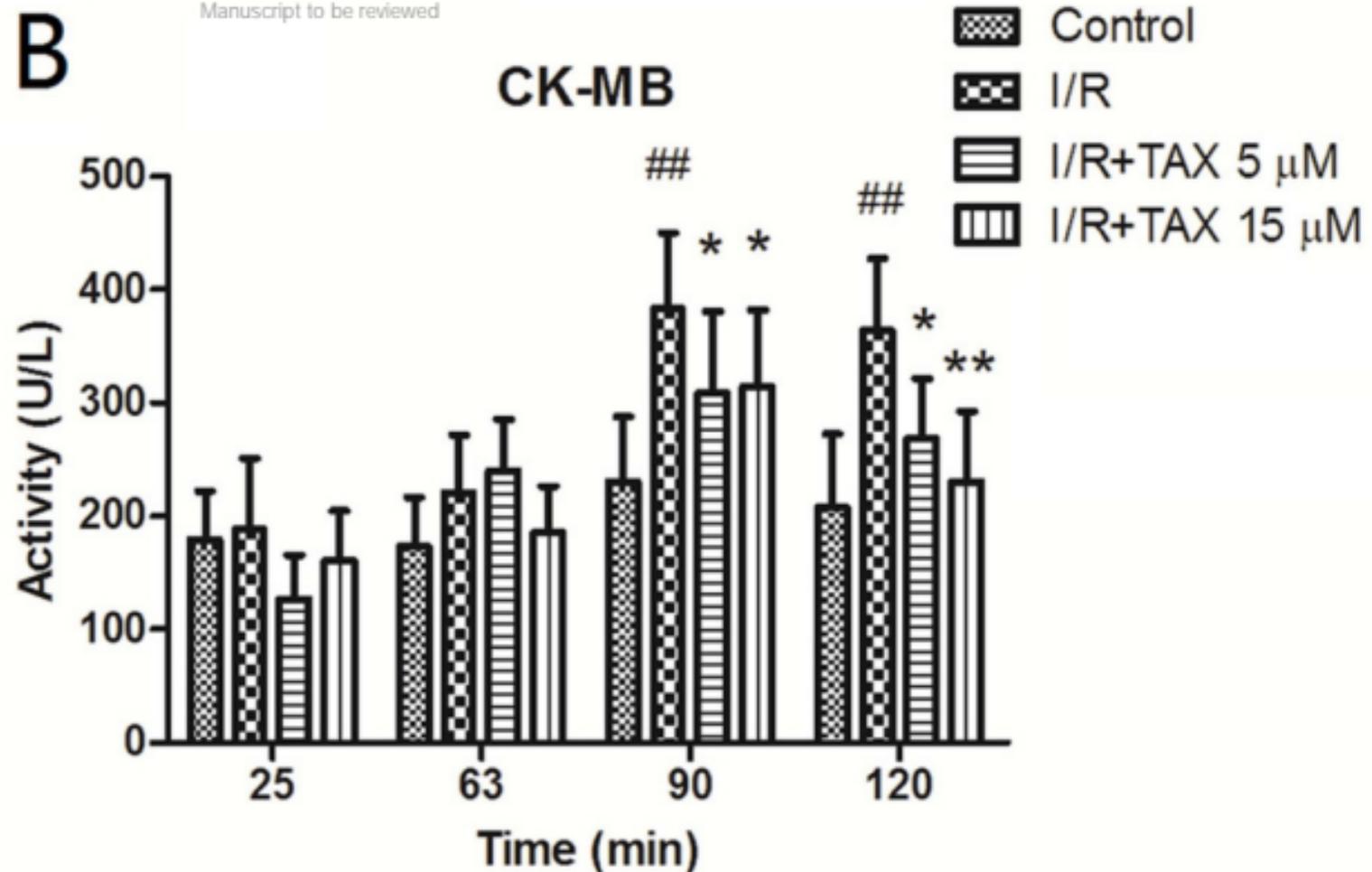
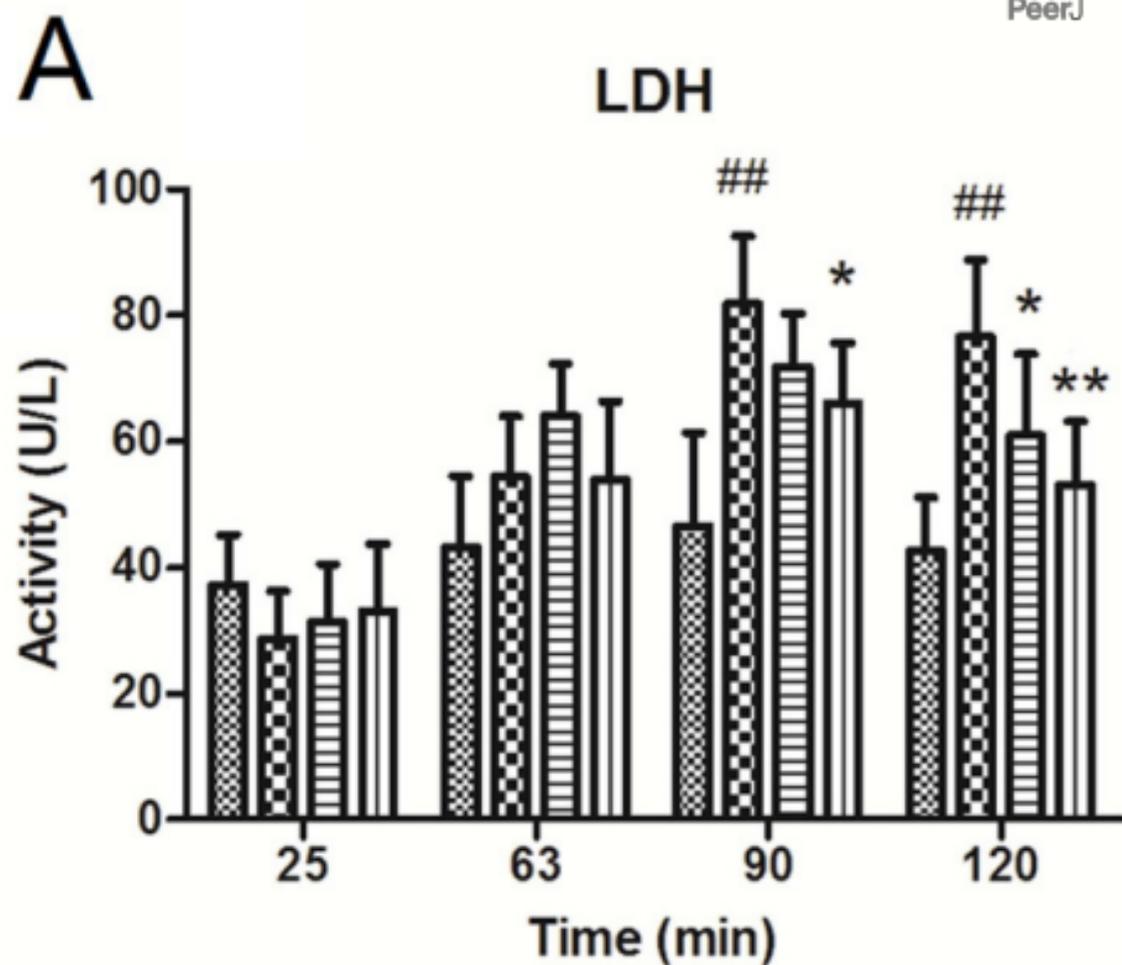
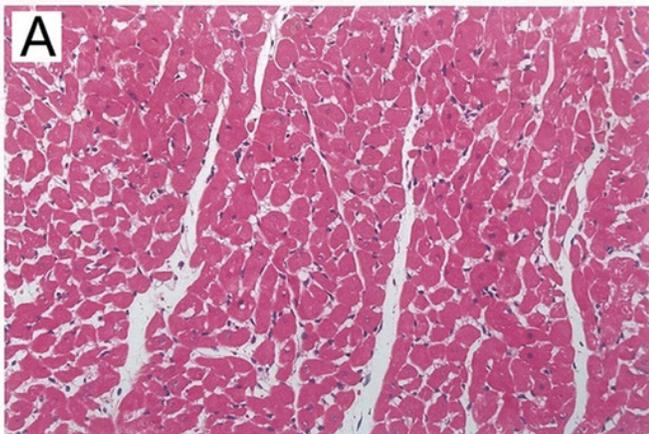


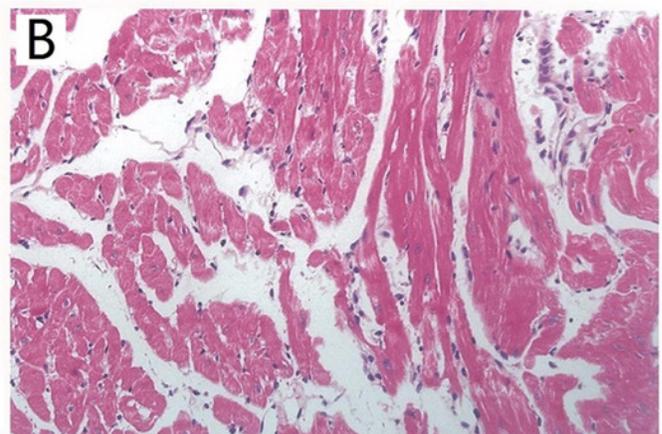
Figure 4

Figure 4. Representative micrographs of HE staining results in various experimental groups.

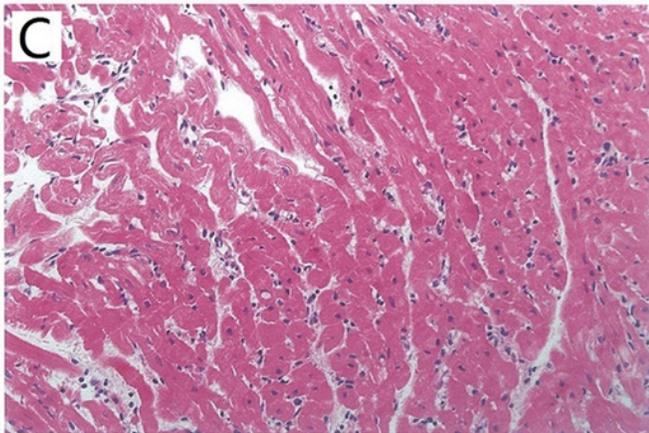
(A) control group; (B) I/R group; (C) treated with TAX 5 μ M group; (D) treated with TAX 15 μ M group, (magnification, $\times 400$).



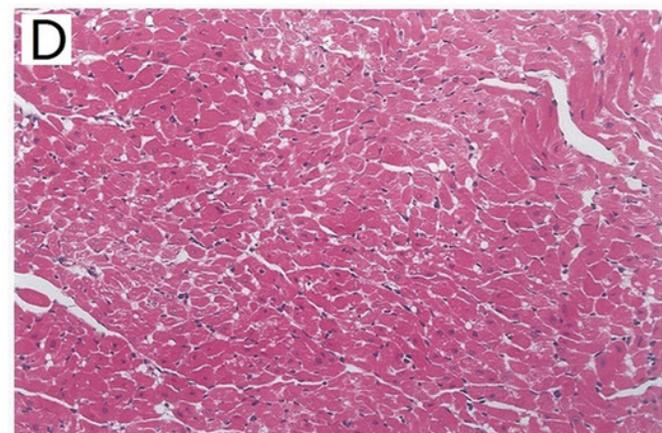
Control



I/R



I/R + TAX 5 μ M



I/R + TAX 15 μ M

Figure 5(on next page)

Figure. 5. Effect of TAX on cardiac the activity of SOD and GSH-PX, contents of MDA.

Values are presented as mean \pm SD. # $P < 0.05$ and ## $P < 0.01$ compared with the control group; * $P < 0.05$ and ** $P < 0.01$ compared with the I/R group. U/mgprot: international enzyme activity unit per milligram tissue protein.

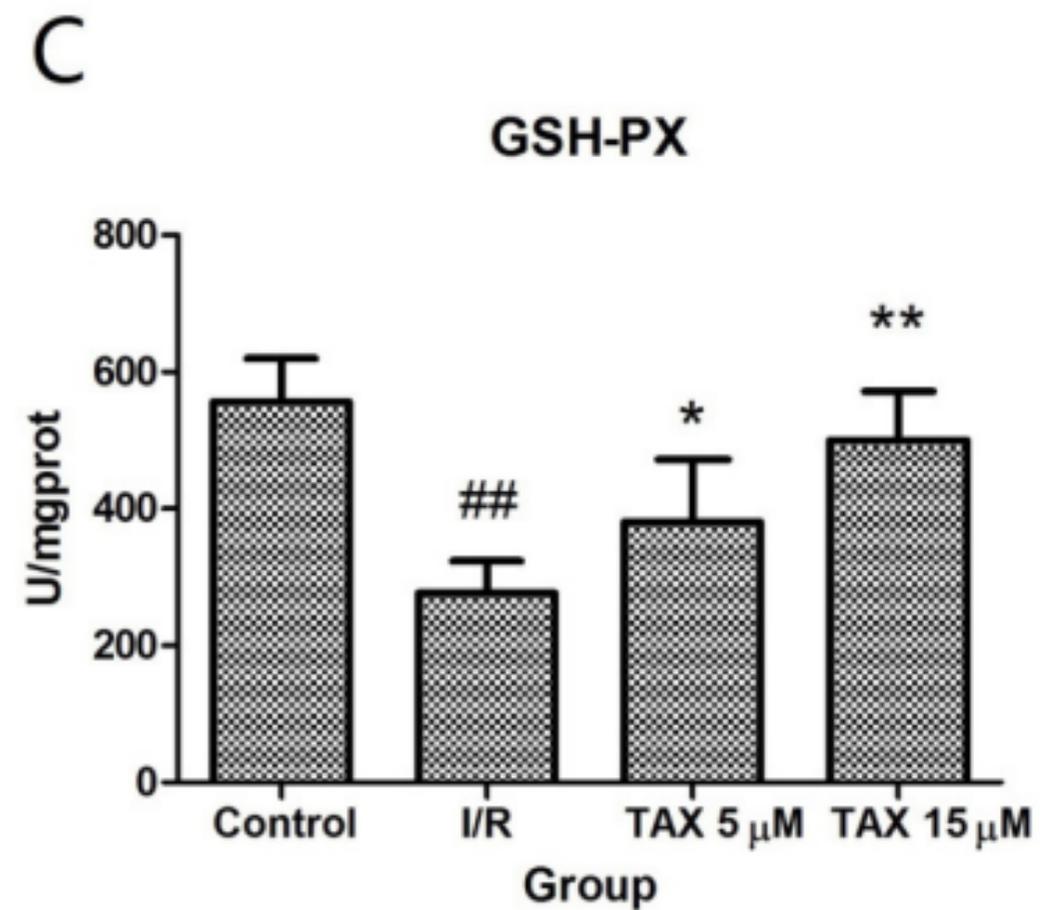
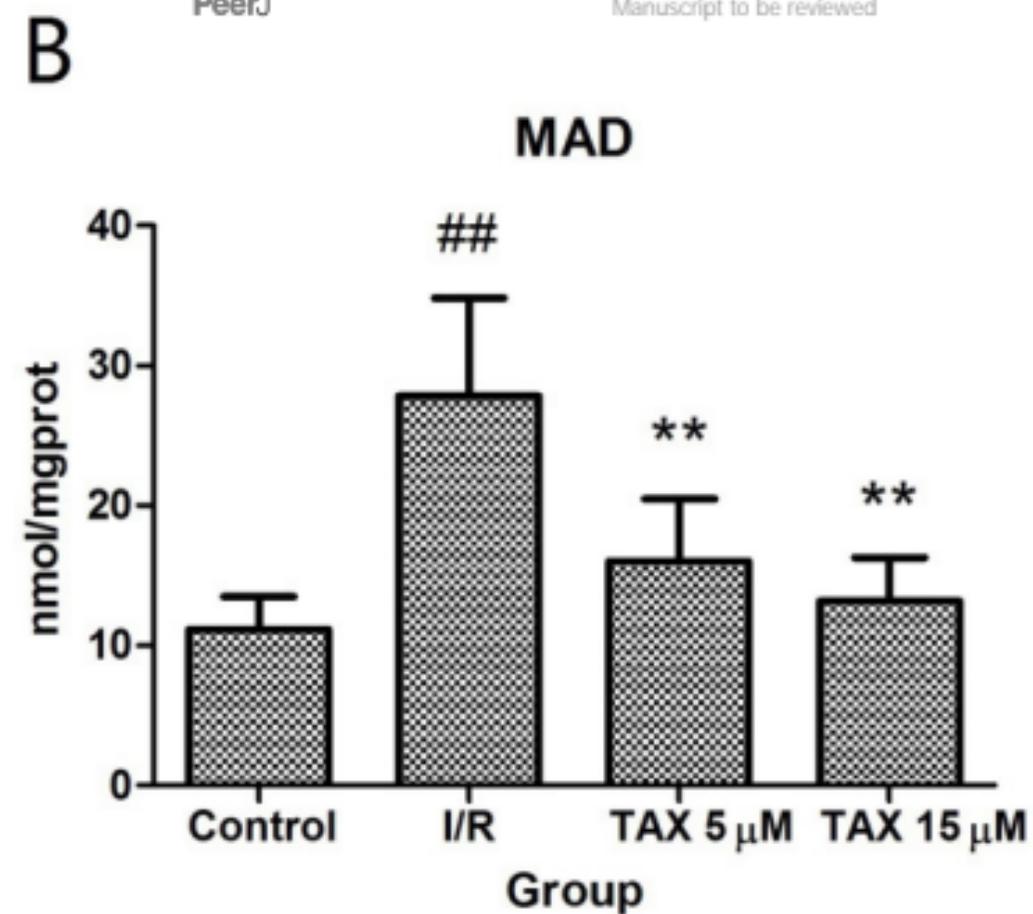
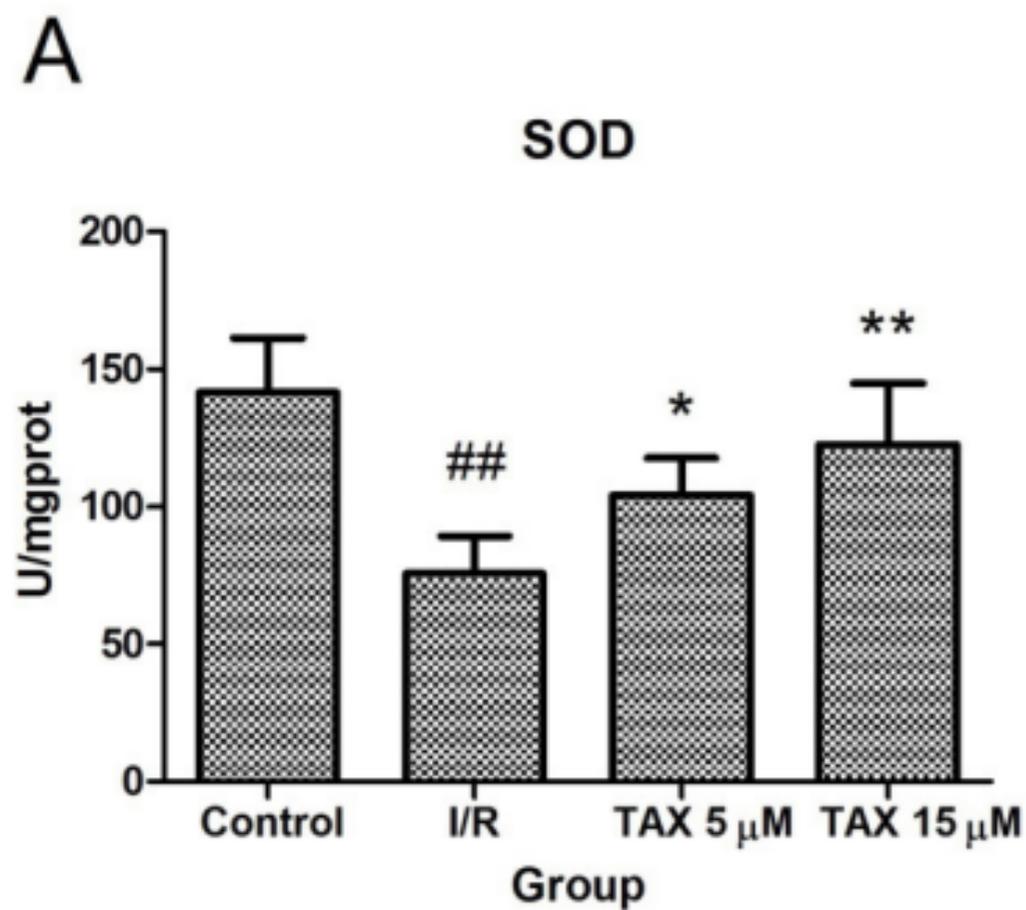
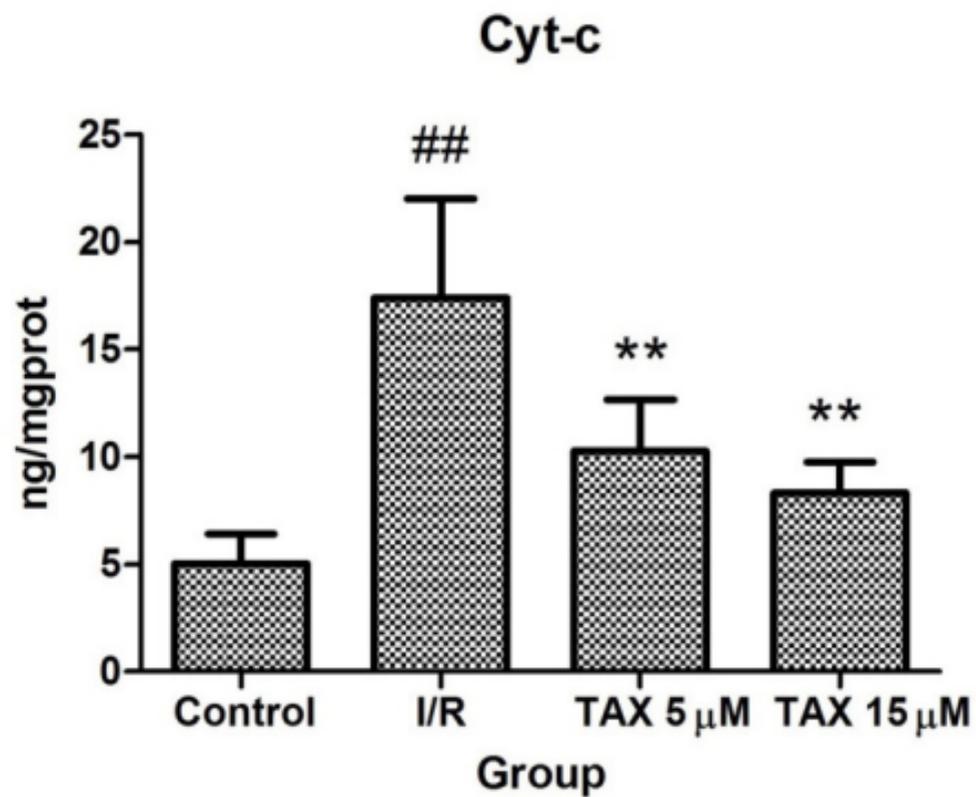


Figure 6(on next page)

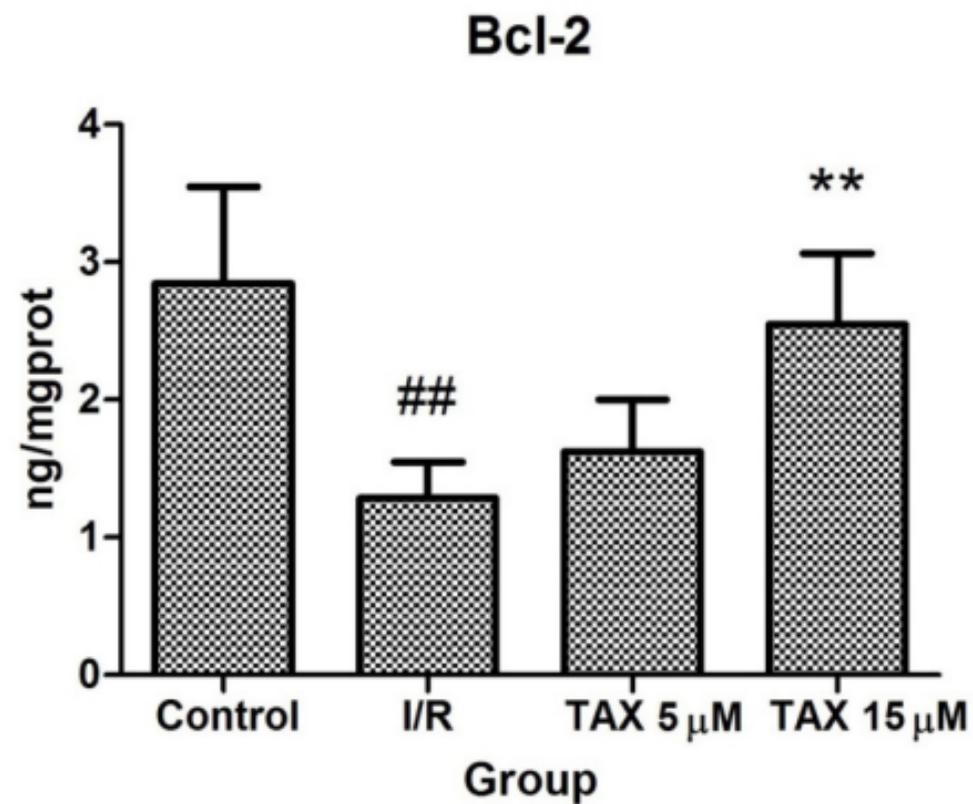
Figure 6. Effect of TAX on the expression of Cyt-c, Bax and Bcl-2 protein.

P < 0.05, ## P < 0.01 vs. Control group; * P < 0.05, ** P < 0.01 vs. IR group. ng/mgprot indicate the nanogram level of the target protein per milligram total protein.

A



B



C

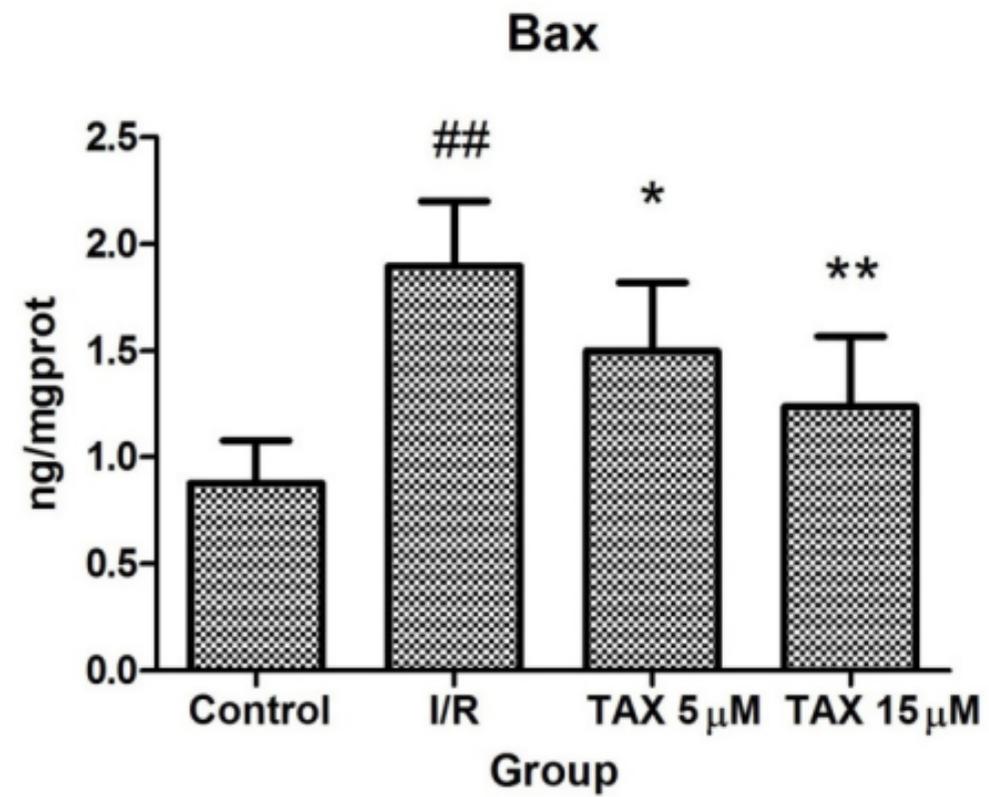


Figure 7 (on next page)

Fig. 7. The changes in the levels of caspase 3, and caspase 9 at the end of reperfusion.

