- 1 The role of Central American barriers in shaping the evolutionary history of the northernmost
- 2 glassfrog, *Hyalinobatrachium fleischmanni* (Anura: Centrolenidae)
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20	Abstract	
21	Background. The complex geological history of Central America has been useful for developing	
22	an understanding of the effects of different processes and barriers on the distribution and	
23	diversity of multiple groups of organisms. Anurans are an excellent choice for such studies	
24	because they usually have site fidelity and reduced movement. The objective of this work was to	Deleted: due to their
25	identify the impact of recognized geographic barriers on the genetic structure, phylogeographic	Deleted: limited distribution, Deleted: ,
26	patterns and divergence times of a wide-ranging amphibian species, <i>Hyalinobatrachium</i>	(2000)
27	fleischmanni.	
21	fielsenmann.	
28	Methods. We amplified two coding mitochondrial regions (COI and ND1) and one ribosomal	
29	region (16S) in samples collected from the coasts of Veracruz and Guerrero in Mexico to the	
30	humid forests of Chocó in Ecuador. We examined the biogeographic history of the species	
31	through spatial clustering analysis (BAPS, Geneland and sPCA), Bayesian and maximum	Comment [Office1]: Plural or singular?
32	likelihood reconstructions, and spatiotemporal diffusion analysis.	Comment [Office2]: Plural or singular?
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33	Results. Our data suggest a Central American origin of <u>Hyalinobatrachium</u> fleischmanni and two	Deleted: H.
34	posterior independent dispersals towards North and South American regions. The first clade	
35	comprises individuals from Colombia, Ecuador, Panama and the sister species <u>H. tatayoi</u> ; this	Deleted: Hyalinobatrachium
1 36	clade shows little structure, despite the presence of the Andes mountain range and the long	
37	distances between sampling sites. The second clade consists of individuals from Costa Rica,	
38	Nicaragua, and eastern Honduras, with no apparent structure. The third clade includes individuals	
39	from western Honduras, Guatemala, and Mexico and displays deep population structure.	
40	Discussion. Herein, we synthesize the impact of known geographic areas that act as barriers to	Comment [Office3]: I suggest adding the following:
41	glassfrog dispersal and have demonstrated their effect in differentiating <i>H. fleischmanni</i> into three	- Effect of the Tachira depression The non-monophyly of H. fleischmanni (i.e. H. tatayoi).

- 47 markedly isolated clades. The observed genetic structure is associated with an initial dispersal
- 48 event from nuclear Central America, followed by vicariance, likely occurring during the
- 49 Pliocene. The southern samples are characterized by a very recent population expansion, likely
- 50 related to sea level and climatic oscillations during the Pleistocene, while the structure of the
- 51 northern clade has probably been driven by dispersal through the Isthmus of Tehuantepec and
- 52 isolation by the Motagua–Polochic–Jocotán fault system and the Mexican highlands.

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Comment [Office4]: Unclear what the authors mean with "nuclear"

Introduction

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55	Historical biogeography focuses on the role of the geographic space as a driver of biological
56	processes such as speciation, extinction, and diversification (Cox, Lalde & Moore, 2016). Areas
57	with a complex geological history are characterized by the appearance and disappearance of
58	multiple barriers and corridors in their history. These barriers may significantly affect the gene
59	flow of resident species, leading to allopatric speciation by vicariance, whereas corridors may
60	lead to species dispersal and colonization of new areas (Noss, 1991). The use of molecular data
61	for the reconstruction of species relationships, the development of new methods for
62	biogeographic analyses, and the increase in geological studies in complex regions have
63	revolutionized the understanding of such biological processes (Ronquist & Sanmartin, 2011).
64	Biogeographic studies have integrated information regarding the relationships within or between
65	closely related taxa, providing valuable opportunities to understand how patterns of biodiversity
66	may have been shaped, even at short time scales (Crawford, Bermingham & Polanía-S, 2007;
67	Streicher et al., 2014).
68	Central America is a region with a rather complex biogeographic history and high diversity of
69	habitats and species (Myers et al., 2000; Cavers, Navarro & Lowe, 2003; Iturralde-Vinent, 2006;
70	Daza, Castoe & Parkinson, 2010). The region is delimited to the north by the Isthmus of
71	Tehuantepec (IT) in Mexico and to the south by the Andes in Colombia (Gutiérrez-García &
72	Vázquez-Domínguez, 2012). The geological landscape of Central America has been continuously
73	modified, especially during the last 15 million years (Ma), by major events including the

emergence of the Panama Arc (13-15 Ma, Montes et al., 2015), the posterior closure of the

Panama Isthmian land bridge when it ceased to function as a seaway (~9-10 Ma, Montes et al.,

2012a; Montes et al., 2012b; Ramirez et al., 2016), and the posterior global climatic transitions

77	during the Plio-Pleistocene (Montes et al., 2015). These events triggered the Great American
78	Biotic Interchange, or GABI, involving the replacement of native taxa (extinctions) and the
79	establishment and diversification of colonizing taxa (speciation) on both continents (Marshall et
80	al., 1982; Stehli & Webb, 1985). Ample phylogeographic research in this region has allowed the
81	effects of geomorphology, topographic barriers, volcanic activity, large climate changes,
82	intermittent connections, and corridors on the biota to be described, aiding in our understanding
83	of the influence of past events on the patterns of genetic structure and the geographic distribution
84	of birds (García-Moreno et al., 2004; Cadena, Klicka & Ricklefs, 2007; Arbeláez-Cortés, Nyári
85	& Navarro-Sigüenza, 2010), plants (Cavers, Navarro & Lowe, 2003; Ornelas, Ruiz-Sánchez &
86	Sosa, 2010; Cavender-Bares et al., 2011), reptiles (Hasbun et al., 2005; Venegas-Anaya et al.,
87	2008), mammals (Eizirik et al., 2001; Ordoñez-Garza et al., 2010; Pérez-Consuegra & Vázquez-
88	Domínguez, 2017), and amphibians (Mulcahy, Morrill & Mendelson, 2006; Crawford,
89	Bermingham & Polanía-S, 2007; Wang, Crawford & Bermingham, 2008; Hauswaldt et al.,
90	2011). As a result, diverse geological factors and major barriers have been more frequently
91	correlated with the evolutionary history and dispersal of species (Bagley & Johnson, 2014).
92	Amphibians are excellent systems for studies in which geological and environmental histories are
93	inferred at fine scales, due to their ecology, particularly regarding their terrestrial habits,
94	intolerance to salt water (Beebee, 2005), and marked niche conservatism (Smith, Stephens &
95	Wiens, 2005; Wiens et al., 2006), as well as the restriction of the particular habitats of many
96	species (e.g., Savage, 2002). However, evaluation of the impact of barriers on the
97	phylogeographic patterns of this taxon, extending across the entire Central American region, has

been precluded because most amphibians have small ranges.

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Comment [Office5]: See Castroviejo-Fisher et al. 2014.

99 Glassfrogs (Centrolenidae) comprise a diverse family endemic to the Neotropics, with numerous 100 species and high levels of endemism, mainly distributed among the northern Andes and Central 101 America regions (Castroviejo-Fisher et al., 2014; Mendoza & Arita, 2014). Studies on the 102 dispersal capability of glassfrogs are limited, but these frogs are known to be restricted to 103 streamside habitats (Ruiz-Carranza & Lynch, 1991) and to show site fidelity (Valencia-Aguilar, Castro-Herrera & Ramírez-Pinilla, 2012) and low mobility, with potential genetic subdivision 104 and restricted gene flow (Delia, Bravo-Valencia & McDiarmid, 2017; Robertson, Lips & Heist, 105 2008). The glassfrog Hyalinobatrachium fleischmanni (Boettger, 1893) has one of the widest 106 distributions, ranging from Guerrero and Veracruz states in Mexico, through the lowlands of 107 108 Central America, to the southernmost limit of its distribution in Ecuador (citations needed). 109 Males of the species call from vegetation along the margins of streams, and egg masses are usually laid on the underside of leaves over a stream. This species exhibits site fidelity and 110 111 parental care by males, who attend one or more clutches at the same time (Delia et al., 2010; 112 Savage, 2002; Barrera-Rodríguez, 2000). Tadpoles fall from vegetation into the water, where they develop; they are apparently fossorial, living buried in the leaf litter and bank substrate of 113 streams (Villa & Valerio, 1982). Related species are distributed in different regions of South 114 115 America, including the northern and central Andes, Guiana shield, and Amazon basin, where previous analyses have suggested an Andean origin for H. fleischmanni (Castroviejo-Fisher et al., 116 2014). 117 118 Considering its wide distribution, coupled with its site fidelity, H. fleischmanni is an ideal organism for studying the role that Central American geographic barriers have played in the 119 120 dispersal patterns of lowland glassfrog species. In the present study, we had the following objectives: (1) to reconstruct the historical biogeography that has shaped the evolutionary history

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of *H. fleischmanni*, including dispersal or vicariance events and time of divergence; (2) to evaluate the possible presence of multiple isolated lineages within *H. fleischmanni*; and (3) to identify the impact of recognized geographic barriers on the genetic structure and phylogeographic patterns of *H. fleischmanni* over time. Based on known information about the species, we tested the hypothesis that *H. fleischmanni* had a South American origin and subsequently dispersed into the Central American lowlands after the closure of the Isthmus of Panama. Additionally, we hypothesized that the dispersal of this species in Central America has been limited by various high mountain ranges acting as barriers and that changes in sea level during the Pleistocene had an impact on the genetic structure of the lowland populations (Bagley & Johnson 2014). Hence, our prediction is that the current genetic structure of *H. fleischmanni* reflects patterns of vicariance events driven by dispersal barriers.

Materials and methods

137 Tissue sampling

Genetic material was obtained across the entire distribution area of the species, from both museum collections and fieldwork (Fig. 1). Fieldwork was performed during the rainy season, in which at least three individuals were collected at each locality. Specimen collection permits were provided by Ministerio de Medio Ambiente, Colombia (Resolution 120 of 24 August 2015) and Secretaría del Medio Ambiente y Recursos Naturales, Mexico (office number 00947/16). Captured specimens were euthanized with a 20% lidocaine hydrochloride (Xylocaine) injection, and all efforts were made to minimize suffering. Liver or muscle tissues were collected in the field and were stored in an RNAlater solution until their use in the laboratory. Specimens were

fixed with 10% formalin, stored in 70% ethanol and deposited in biodiversity collections at 146 147 public research institutions in each country. 148 Molecular techniques 149 DNA was obtained from muscle and liver tissues following the phenol-chloroform extraction protocol of Sambrook & Russell (2006). The quantity and quality of the DNA were verified in 150 151 1% agarose gels and by measuring absorbance using a NanoDrop spectrometer (Thermo Fisher Scientific Inc., Wilmington, DE, USA). Amplification of the mitochondrial COI (658 bp), 16S 152 (895 bp), and ND1 (961 bp) genes was performed following the protocols described by 153 Guayasamin et al. (2008). PCR products were visualized with agarose gels and purified according 154 155 to the EXO-SAP protocol (GE Healthcare, Piscataway, NJ, USA). DNA sequences were obtained with the BigDye Terminator Cycle Sequencing kit (Applied Biosystems). Sequences from H. 156 tatayoi from Venezuela were also included. The sequences were assembled and edited manually 157 158 and were aligned with Geneious 9.1.2. We are aware that analyses based on mitochondrial DNA (mtDNA) provide a limited view of 159 160 species evolution (i.e., matrilineal inheritance). However, due to the extreme difficulty of obtaining genetic material from the nuclear Central America region, we could only perform our 161 analyses with sequences that were directly comparable to those from previous studies. However, 162 mtDNA is a robust indicator of population history and species limits (Avise, 2000; Zink & 163 Barrowclough, 2008), and mtDNA markers have helped to decipher genetic structuring in the 164 form of phylogeographic breaks (phylogenetic splits between mostly distinct geographical 165 lineages) in abundant Central American studies (Bagley & Johnson 2014). In addition, our study 166

Comment [Office7]: Who generated these sequences? Include source of sequences (it should be an additional column in the Supplementary Table 1).

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Comment [Office8]: Explain the difficulties. Are they related to burocratic difficulties or problems associated to gene sequencing?

encompasses the widest geographic distribution of the species, allowing us to evaluate the

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diversification events shaping the evolutionary history of the species.

170 Data analysis

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171 Identification of landscape barriers and genetic diversity

The first step in identifying the main barriers to dispersal was to perform spatial and clustering analyses that are commonly applied to mtDNA sequences. First, we conducted a Bayesian analysis of population structure with BAPS v6.0 (Corander et al., 2008), using the spatial clustering of individuals, considering that the spatial prior may strengthen inferences for sparse molecular data. Second, the spatial location of genetic discontinuities was estimated with Geneland (Guillot et al., 2005), which estimates the number of populations within the geographical area of interest, maps borders between populations, assigns individuals to populations, and detects possible migrants. We ran the model in R under the correlated allele frequency model, without uncertainty regarding spatial locations. We generated 10⁵ iterations to a thinning of 100, with the maximum rate of the Poisson process fixed as the number of individuals. Third, we performed a spatial principal component analysis (sPCA; Jombart, 2008a) in the package adegenet in R (Jombart, 2008b), for which we constructed a neighbor distance net among all coordinates and tested for significant, geographically correlated genetic structure along the main axis, based on a global randomization test. Previous studies have suggested multivariate ordination analyses as an alternative to Bayesian algorithms because they do not make any assumptions about the underlying population genetic model (Jombart, Devillard & Balloux, 2010). Considering that the pool of samples was slightly different for each gene, all analyses were run independently for each gene, with a maximum k value of 10 populations per analysis.

We next defined genetically homogeneous regions obtained through landscape analyses for diversity index estimation. We calculated haplotype (h) and nucleotide diversity (π), globally and by region, with DNAsp (Libardo & Rozas, 2009). Additionally, the distribution of genetic variability at hierarchical levels was estimated using analysis of molecular variance (AMOVA). Additionally, a median-joining haplotype network for each gene was constructed using PopArt (French et al., 2014). Genetic differentiation among regions was estimated based on the F_{ST} statistic with the *poppr* package (Kamvar, Tabima & Grünwald, 2014) and used corrected distances according to the K2P parameter (Kimura, 1980) in MEGA v.7 (Kumar, Strecher & Tamrura 2016).

Phylogenetic analyses

The sequences of all genes were concatenated, and a phylogenetic tree at the intraspecific level was estimated using both likelihood analysis in RAxML (Stamatakis, 2006) and a Bayesian inference approach implemented in MrBayes (Ronquist & Huelsenbeck, 2003). We rooted our phylogeny using the species *Hyalinobatrachium carlesvilai*, *H. mondolfii*, *H. chirripoi* and *H. colymbiphyllum* as outgroups. A list of the specimens and GenBank accession numbers included in this study is presented in Table S1. The best evolutionary model for each non-coding region (16S) and for the coding genes (COI and ND1) was evaluated using PartitionFinder 2 software (Lanfear et al., 2016). Maximum likelihood analysis was conducted using 10,000 rapid bootstrap analyses, the GTR + Γ evolution model and summarized support for the best tree. For Bayesian inference, we ran two independent analyses for 12 million generations, sampling trees and parameter values every 1,000 generations. Burn-in was set to 25%, and the first 3 million generations were therefore discarded.

Comment [Office10]: Add "species", "elevation", "Genbank code" and "source" as columns in this table. The table should include outgroups. "Source" should include the studies that generated the sequences.

212 Divergence times and Bayesian spatiotemporal diffusion analyses To estimate diversification times for the different H. fleischmanni mitochondrial lineages, we 213 214 employed BEAST 1.6.2 (Drummond & Rambaut, 2007). The time to the most recent common ancestor for the main lineages was calculated via Bayesian Markov chain Monte Carlo (MCMC) 215 searches. The ultrametric tree was inferred de novo using the same partition substitution models. 216 217 In the absence of a fossil record for glassfrogs, we based our analysis on previously published divergence times. We used three stem ages for *Hyalinobatrachium* species as calibration 218 constraints, following Castroviejo-Fisher et al. (2014). The most recent calibration point was 219 placed at 2.42 Ma (Confidence Interval (C.I.)=1.63-3.37), representing the divergence between 220 221 H. fleischmanni (USNM 559092) and H. tatayoi (MHNLS 17174). The following calibration node was placed at 7.65 Ma (C.I.=5.93–9.63), representing the divergence between H. 222 fleischmanni-H. tatayoi and H. charlesvilai. The most ancient calibration point corresponded to 223 224 the divergence between H. fleischmanni and H. mondolfii (8.4 Ma, C.I.=6.68–10.52). We implemented an uncorrelated lognormal relaxed molecular clock, and trees were sampled every 225 226 1,000th iteration for 100,000,000 generations, with 20% of the initial samples being discarded as 227 burn-in, after empirical assessment of appropriate chain convergence and mixing with Tracer 1.4 (Rambaut & Drummond, 2007). We constructed the historical demography of the major clades 228 obtained from the phylogenetic results, using Bayesian skyline plots that estimate the posterior 229 distribution of population sizes (Drummond et al., 2005). 230 To reconstruct the ancestral distribution and spatial dispersal of the species, we performed a 231 Bayesian spatiotemporal diffusion analysis in BEAST (v.1.8.4), assuming continuous spatial 232 233 diffusion with a time-heterogeneous random walk model ("Relaxed Random Walk", RRW, 234 Lemey et al. 2010). For this analysis, we used a subset of 34 samples with data for all three genes

Comment [Office11]: Describe the calibration strategy (and limitations) of Castroviejo-Fisher et al. (2014)

235	plus samples lacking some genes but originating from intermediate localities, encompassing the
236	entire distribution of the species. We applied a normally distributed diffusion rate, a coalescent
237	Bayesian Skyride model, and SRD06 substitution models (Shapiro, Rambaut & Drummond
238	2006). We used the jitter option under the TraitLikelihood statistic with a parameter value of 0.1.
239	To summarize the posterior distribution of ancestral ranges using the RRW model, we annotated
240	nodes in a maximum clade credibility tree (MCC) using the program TreeAnnotator v1.7.5. This
241	tree was then used as an input in SpreaD3 (Bielejec et al., 2016) to reconstruct the pattern of
242	spatial diffusion and to visualize lineage diversification across the landscape.
243	Spatial barriers to dispersal
244	We contrasted the previous BEAST analysis with a set of possible geographic elements defined a
245	priori and based on the four geological blocks across Central America (Maya, Chortis,
246	Chorotega, and Choco) (Marshall, 2007; Gutiérrez-García & Vázquez-Domínguez, 2013), as well
l 247	as their northern and southern limits the southern Mexico block (SMB) west of the IT and the
248	South American plate (SAP) east of the Andes, respectively. Three highland barriers were
249	evaluated for their effect on species dispersal: the Motagua-Polochic-Jocotán (MPJ) fault
250	system, the Talamanca Cordillera, and the Andes range. Three previously recognized lowland
251	barriers for lowland amphibians (the Hess Escarpment, HE; western Panama isthmus, WPI; and
252	central Panama isthmus, CPI) were also tested as possible barriers during Pleistocene sea level
253	oscillations (Fig. 1).

Comment [Office12]: Since the study includes H. tatayoi and given the paraphyly of H. fleischmanni, I also would include the Tachira Depression as a barrier.

254 Results

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We generated a final alignment of 2036 base pairs (bp) for 123 samples from 9 countries,

including 13 sequences obtained from the GenBank and BOLD system databases (Table S1). We

with an overall haplotypic diversity h=0.863 and a nucleotide diversity $\pi=1.282$ for the 16S gene. 258 259 By contrast, we found 63 haplotypes for COI, with h=0.979 and $\pi=0.044$, and 45 haplotypes for 260 ND1, with h=0.991 and π =0.042 (Table S2). Landscape analysis, genetic diversity and structure 261 The BAPS results depicted six clusters for 16S, seven for COI and five for ND1. Although the 262 number of clusters varied for the northern and southern regions, three clusters in western Chortis, 263 Chorotega, and eastern Chortis were consistently recovered. The best differentiation was obtained 264 with the 16S sequences, which separated the northern clusters, while the COI sequences could 265 266 differentiate the southern clusters, east-west of the Andes range (Fig. 2a). Geneland showed six, seven and six clusters for 16S, COI and ND1, respectively. Both coding genes showed two 267 clusters on both sides of the Andes range. Separation between the Choco and Chorotega samples 268 269 was found in all cases (Fig. 2b). sPCA performed on individual genotypes revealed a significant, geographically correlated genetic structure for all three genes (nper=999, P=0.001). Eigenvalues 270 271 indicated a higher spatial genetic structure for the main axis, related to the global structure. The 272 first sPCA (regional scale) positive axis for all genes exhibited the greatest variation of genetic 273 distance in relation to the distance network (Fig. 2c). Based on our results, we selected seven genetically homogeneous regions: the North American 274 Pacific, Gulf of Mexico, Maya block, western Chortis, eastern Chortis-Chorotega, Choco block 275 and SAP. The minimum genetic distance between regions was obtained for SAP-Choco 276 (K2P=0.002-0.019), while the maximum was obtained between the North American Pacific and 277 278 Chorotega (K2P=0.041) for 16S, between Maya and SAP for COI (K2P=0.078), and between

did not detect any stop codons in protein-coding genes (COI, ND1). We obtained 25 haplotypes

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Comment [Office13]: Always insert a space before and after =

Comment [Office14]: Since these three genes are linked, shouldn't they be analyzed together?

Comment [Office15]: What does "best differentiation" mean? More genetic structure?

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279	SAP and Chorotega for ND1 (K2P=0.077) (Table 1). The F_{ST} indices between regions was >0.7	
280	for almost all combinations, except between SAP and Chocó for 16S and among the Gulf, Pacific	
281	and Maya regions for ND1 (Table 1).	
282	When comparing the diversity per cluster, the Chorotega population showed the highest	
283	haplotypic diversity for all genes, while the Maya populations exhibited the highest nucleotide	
284	diversity for coding genes. The Gulf population showed the lowest haplotypic diversity, and SAP	Comment [Office16]: Since the shouldn't they be analyzed
285	exhibited the lowest nucleotide diversity (Table S2).	
286	The AMOVA results (Table 2) for all genes indicated that 79-88% of the observed genetic	
287	variability was partitioned between regions, compared with 12-20% within regions (all P <	
288	0.001). Overall, the haplotype networks for the three genes were concordant, with higher	
289	diversity and structure being revealed for COI and ND1 than for 16S. Four mitochondrial	Comment [Office17]: When p
290	haplotype groups were detectable among the entire distribution (Fig. 3), where the concordance	are independent sources of the genes are links, perhap
291	between the haplotype network and the species distribution suggested a deep pattern of	the results of concatenated
292	geographic structuring and differentiation across the complete range. The SAP and Choco regions	
293	shared the same 16S haplotype but showed differences in the COI and ND1 coding genes.	
294	Phylogenetic patterns, times of divergence, and demography	
295	The PartitionFinder output indicated that HKY+I+G, GTR+G, and GTR+G+I were the best	
296	models for 16S, COI, and ND1 respectively. The phylogenetic relationships based on the	
297	Bayesian approach and maximum likelihood showed three main, well-supported clades, although	
298	their relative positions were not fully resolved (Fig. 4). The first clade, designated the "Northern	Comment [Office18]: Are the concatenated genes? Please
299	clade", was divided in two lineages: a large lineage containing all samples from the SMB and	concatenated genes. Treas
300	Maya regions (pp=1) and a smaller one from the western Chortis region (pp=1). The second	

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clade, the "Central clade", was comprised of samples from the eastern Chortis and Chorotega
regions (pp=1). The third clade, the "Southern clade", consisted of samples from the Choco and
SAP regions, including the sister species <i>H. tatayoi</i> from Venezuela (pp=1). The Southern clade
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did not show any structure, displaying a polytomic topology.
The results regarding the estimation of divergence time showed a pattern in which divergence
among the three main clades occurred during the Pliocene (~3.40 Ma, HPD=2.25-4.56 Ma; Fig.
3). With respect to the Northern clade, the split between the lineage from West Chortis and the
remaining samples occurred <u>also during</u> the Pleistocene, <u>in the Gelasian age</u> (~2.19 Ma,
HPD=1.38-3.53), while separation between samples from the Pacific and Gulf+Maya regions
occurred at the beginning of the Calabrian age (~1.76 Ma, HPD=0.86-2.23). Finally, the split
between lineages from the Gulf and Maya regions occurred near the end of the Calabrian age
(~1.51 Ma, HPD=0.62-1.73). The divergence between the Central and Southern clades occurred
at the end of the Pliocene (~2.64 Ma, HPD=1.50-3.68), while splits within each clade began at
the end of the Calabrian age for the Central clade (~0.81 Ma, HPD=0.12-0.71) and during the
Gelasian age (~1.64 Ma, HPD=0.63-1.89) for the Southern clade (Fig. 4).
The 95% credible intervals of the effective population size (BSP results) overlapped along the
entire time period in the Northern clade (Fig. 5a). In the Southern clade (Fig. 5b), the effective
population size did not overlap; this clade exhibited a constant population size and posterior
expansion at approximately 0.1-0.3 Ma. The lack of 95% CI overlap of the more ancient and the
more recent population with a Bayesian posterior probability >0.95 showed that the female
effective sizes did not overlap, providing significant support for a population size change.

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The results regarding Bayesian spatiotemporal diffusion (Fig. 6) highlight the Chorotega and West Chortis region as the most likely ancestral geographic area for *H. fleischmanni*, suggesting two subsequent dispersal events, in which the most recent common ancestor (MRCA) of the Northern clade was distributed in the environs of the Chortis and Maya regions, whereas that of the Southern clade (stem node) was distributed around the Chorotega and Chocó regions. Our results also reflect independent dispersal for samples west of the MPJ fault system and later divergence of the three remaining clusters in the Northern clade around the IT. For the Central clade, an initial range in the Chorotega region south of the Talamanca range was observed, with posterior dispersal towards eastern Chortis. Furthermore, an ancestral range was detected in the Choco block within the Southern clade, with subsequent dispersal towards the south and east, reaching the lowlands east of the Andes range to the south, while two lineages dispersed northwards independently, reaching the southern limit of the Choco region.

Comment [Office20]: Check for consistency. In some other parts of the text you spell it as "Choco".

Discussion

The complex geologic and geographic history of Central America has long intrigued researchers, who have aimed to decipher how different features that act as barriers to or corridors for dispersal have affected the distribution and diversity of multiple taxa (Gutierrez-García & Vázquez-Dominguez, 2013; Bagley & Johnson, 2014). Our results show a deep phylogenetic structure of *H. fleischmanni*, which has differentiated as three well-supported clades, revealing old divergence events dating back to the Pliocene and younger divergence events within clades during the Pleistocene (Fig. 3). Additionally, our results show *H. fleischmanni* to be paraphyletic with *H. tatayoi*, nested within the South American clade.

Comment [Office21]: I would like to see the Discussion focused on analyses on concatenated genes and not gene-by-geny. As mentioned above, all genes used in the study are linked.

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Comment [Office22]: This result has been found by several authors (e.g., Guayasamin et al. 2008, Castroviejo-Fisher et al. 2014)

347 The Southern clade

The Southern clade encompasses samples from the Choco and SAP regions, including the sister 349 species H. tatayoi from Venezuela. Samples ranging from Panama to Venezuela and Ecuador 350 351 grouped together, with no clear phylogenetic separation among them. Nonetheless, haplotype 352 networks and landscape analyses allowed us to identify a partition on both sides of the Andes 353 range for the coding genes (Figs. 2 and 3). The lack of significant structure for the Southern clade is remarkable, considering that the distances between populations reach 1,600 km. In addition, 354 geographic barriers along the Western and Central Panama Isthmus and the Andes range, which 355 are widely recognized as speciation drivers for both highland and lowland species (Bagley & 356 Johnson 2014, Mendoza et al., 2015), seem to not have exerted any effect on the genetic structure 357 358 of this clade. Notably, the genetic distance observed on both sides of the Andes range (COI K2P=1.9%; Table 1) is lower compared to the distances reported for lowland species with a 359 360 higher dispersal capacity from the same region, such as the hummingbird Amazilia amabilis (K2P=2.06%, Mendoza et al., 2016). These results contrast with previous knowledge of the 361 362 ecology of glassfrog species, which have been found to be characterized by site fidelity (Valencia-Aguilar, Castro-Herrera & Ramírez-Pinilla, 2012), low mobility and restricted gene 363 flow even at local scales (Delia, Bravo-Valencia & McDiarmid, 2017; Robertson, Lips & Heist, 364 365 2008). However, most of the previous research on this frog has focused on calling males and reproductive behavior (Delia, Bravo-Valencia, & McDiarmid, 2017), while the dispersal 366 367 capability of females and tadpoles, which can have a significant impact on mtDNA genetic structure, is still unknown. Thus, it is possible that the Choco and SAP regions present adequate 368 369 conditions for tadpole dispersal, allowing range expansion. However, this hypothesis needs to be evaluated based on additional demographic studies and a greater sample size per site. 370

Comment [Office23]: H. tatayoi is not sister to H. fleischmanni.

Comment [Office24]: The study includes only two samples from Venezuela and one from Ecuador. The conclusions would be much stronger if the authors could include more samples from these two countries.

Comment [Office25]: Idem.

Comment [Office26]: Be specific. Which barriers are you referring to?

Comment [Office27]: I don't think the two examples are equivalent. Populations of H. fleischmanni might be connected through the Magdalena Valley.

Our results do not support our initial hypothesis that the species was originally from South America and then dispersed through the Isthmus of Panama. Indeed, the Southern clade is rather young, and the various populations it encompasses differentiated during the last million years (middle Pleistocene). Our results show that this clade has experienced a recent population expansion during the last 100,000-300,000 years, reaching a relatively final stable population size, exhibiting a dispersal route from Central Panama to South America (Figs. 5, 6). Based on the genetic and phylogenetic results for this clade, we suggest that its dispersal towards South America and on both sides of the Andes occurred very recently, likely as a consequence of climatic oscillations during glacial periods (Smith, Amei & Kickla 2012). Under this scenario, there has been insufficient time for effective genetic differentiation to occur, and the phylogenetic reconstruction therefore failed to resolve the divergence detected in the spatial analysis.

The Northern clade

Unlike the Southern clade, the Northern clade shows significant genetic structure in different lineages, ranging from western Chortis (Central America) to lowland forests in Veracruz and Guerrero (Mexico). One remarkable finding was the split between samples from either side of the MPJ fault system, where individuals separated by only 60 km exhibit great genetic distances (K2P=1.6% for 16S, 5.2% for COI and 6.1% for ND1, Table 2), even reaching the limit of the barcode gap for Neotropical amphibians (6% for COI; Lyra et al., 2017). The calibration results showed that samples from these localities have been isolated since the Gelasian age (early Pleistocene; ~2.19 Ma, HPD=1.38-3.53). The MPJ fault system has been recognized as the main barrier to dispersal for multiple species ranging from the Maya to the Chortis blocks (Barrera-Guzman et al., 2012, Rovito et al., 2015, Ornelas, Ruiz-Sánchez & Sosa, 2010), which have effectively acted as a barrier for *H. fleischmanni*.

Comment [Office28]: Given the results, the authors should mention the possibility of cryptic species in H. fleischmanni.

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Within this clade, we found that samples from SMB do not group as a single lineage but instead
display a paraphyletic position in relation to the lineage from the Maya region. These two regions
present the lowest K2P among all comparisons performed in this analysis, except for samples
from either side of the Andes range. The BAPS and sPCA results for COI and ND1 also grouped
samples from these two regions. One possible reason for this finding is that the IT has not acted
as a significant barrier for <i>H. fleischmanni</i> . It is important to highlight that unlike the known
impact of the Western and Central Panama Isthmus, the effect of the IT has mostly been defined
based on montane species (Bryson, García-Vázquez & Riddle, 2011; Jiménez & Ornelas, 2016),
for which it represents a barrier that limits dispersal. Hence, in the case of lowland species such
as <i>H. fleischmanni</i> , the IT might act more as a corridor than a barrier, which is supported here by
the Bayesian spatiotemporal diffusion results (Fig. 6).
On the other hand, the Geneland and phylogenetic results showed a more complex scenario, in
agreement with the presence of three well-defined lineages within the Northern clade, one of
which is located in the north of the SMB, another southward the SMB, and the last in the Maya
block (Fig 2b). This genetic structure is very similar to that observed for the brush-finch <i>Arremon</i>
brunneinucha, distributed in humid montane forests (Navarro-Sigüenza et al., 2008). Samples
brunneinucha, distributed in humid montane forests (Navarro-Sigüenza et al., 2008). Samples
brunneinucha, distributed in humid montane forests (Navarro-Sigüenza et al., 2008). Samples from the Gulf of Mexico and Pacific are clearly isolated by the highlands of the Sierra Madre

415 The Central clade

416 Our results revealed a Central clade without any deep geographic structure, expanding along both 417 the Chortis and Chorotega blocks, which separated from the Southern clade around the end of the 418 Pliocene (~2.64 Ma). We did not find any evidence suggesting differentiation between samples 419 from the Chortis and Chorotega regions, suggesting that the Hess Escarpment has not acted as a 420 significant barrier. However, we must consider the small sample sizes from Honduras and Nicaragua (n=10 samples), which likely limits detailed structural evaluation for this region. Most 421 phylogeographic studies performed in the region known as nuclear Central America face similar 422 problems, with limited or null sampling from northern Honduras (Mulcahy, Morrill, & 423 Mendelson 2006; Castoe et al., 2003; Strecker et al., 2004; Crawford & Smith, 2005) or sampling 424 425 that is biased towards the dry forests of the Pacific coast (Parkinson, Zamudio, & Greene 2000; Hasbún et al., 2005; Vázquez-Miranda, Navarro-Sigüenza & Omland, 2009; Poelchau & 426 Hamrick, 2011), where H. fleischmanni has not been recorded. Our Bayesian spatiotemporal 427 diffusion results showed rapid dispersal from the Chorotega to Chortis blocks, with no apparent 428 429 impact on the genetic structure of these populations (Fig. 6). Nevertheless, additional work is needed to confirm whether the main geographic features present in this region have driven the 430 dispersal of low-mobility species such as H. fleischmanni in humid forests. 431 432 Phylogeographic patterns The three main clades that we identified for *H. fleischmanni* show deep intraspecific divergence, 433 with genetic distance values greater than 2% (16S) and 5% (COI and ND1). Indeed, the 434 landscape analysis, Bayesian spatiotemporal diffusion analyses, and estimated divergence time 435 revealed interesting patterns that allowed us to reconstruct the historical biogeography of these 436 437 frogs and to identify the impact of different geographic barriers on the genetic structure and 438 phylogeographic patterns of H. fleischmanni. Although the main phylogenetic topology and the

three major clades were well supported in Bayesian analyses, the maximum likelihood phylogenetic reconstruction did not resolve these relationships (see Supplementary Material). 440 441 The Bayesian spatiotemporal diffusion analyses suggest that H. fleischmanni originated along the region encompassing the Chorotega and eastern Chortis elements, which contrasts with the South 442 American origin proposed by Castroviejo-Fisher et al. (2014). Interestingly, we found that H. 443 fleischmanni has undergone two dispersal events: one southward to the Chocó region and one 445 northward, reaching the Maya region, followed by vicariance events driven by the effect of the Chortis highlands and the Talamanca range. Considering that divergence times among the three 446 clades are similar (i.e., the isolation of the Northern clade occurred ca. 3.40 Ma (HPD=2.25-4.56 Ma), while that between the two other clades occurred 2.64 Ma (HPD=1.50-3.68)), it is likely 448 that the ancestor of H. fleischmanni arrived in Central America during the Pliocene, soon after the 449 closure of the Isthmus of Panama (Montes et al., 2015). Accordingly, the dispersal-vicariance 450 451 events among the main clades potentially occurred simultaneously or over a very short time, which might explain why the position of the clades and their internal structure were not 452 consistent between the Bayesian and maximum likelihood approaches. 453 454 Regarding the vicariance events for the Central and Southern clades, multiple elements need to be 455 revised. The samples from each cluster that are geographically closest are located on opposite sides of the Talamanca Cordillera in the Chorotega block. The time of divergence of the MRCA 456 for these clades (3.28 Ma, HPD=1.59-3.86) coincides with the estimated age of the intervening 457 mountains (1-2 Ma; Denyer, Alvarado & Aguilar 2000; Marshall et al., 2003), which are 458 recognized as a main driver of speciation (Savage, 2002). The time of divergence also coincides 459 460 with the rise of the sea level during the mid-late Pliocene (~3.5–3 Ma), which generated a 461 seaway, likely reinforcing the western Isthmus of Panama break and therefore acting as a barrier

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Comment [Office29]: Note that all species that are closely related to H. fleischmanni are endemic to South America. Thus, a South American origin of fleischmanni makes a lot of biogeographic sense. The authors should include this in their discussion.

Comment [Office30]: Arrived from where? South America, right?

462	across the Pacific region (Cronin & Dowsett, 1996; Bagley, Hickerson & Johnson 2018). Hence,
463	the central mountain ranges on Costa Rica and Panama and the eustatic sea levels around the
464	western Isthmus of Panama might have increased divergence, as documented for multiple spatial
465	divergence patterns of amphibian species (Crawford Bermingham & Polanía-S; Wang, Crawford
466	& Bermingham., 2008; Bagley, Hickerson & Johnson, 2018).
467	The isolation of the Northern clade from the other two does not entirely correspond to our
468	hypothesis of geographical barriers. Populations from both the Northern and Central clades are
469	distributed throughout the Chortis block, indicating that the MPJ fault system was not the main
470	driver of divergence between clades. On the other hand, our structure (Geneland) results suggest
471	a frontier at the center of the Chortis block, near northeastern Honduras (Fig. 2b). Similar
472	divergence patterns have been observed between two water-dependent subspecies of Caiman
473	crocodilus (Venegas-Anaya et al., 2008), in agreement with the eastern limit of the Chortis
474	highlands (Morrone, 2014; Townsend, 2014). Here, the complex topography resulting from
475	multiple volcanic activities along the Chortis highlands during the last 2 Ma and the presence of
476	dry habitats in the Pacific region (Savage, 2002) may have isolated the H. fleischmanni
477	populations during the late Pliocene. This hypothesis is in agreement with the high species
478	endemism recognized for the region (Anderson et al., 2010; Townsend et al., 2012), in which
479	intensive study is required to evaluate the underestimated regional taxonomic diversity
480	(Townsend & Wilson 2016).

481 Taxonomic implications

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Previous studies have suggested that *H. fleischmanni* is a paraphyletic species in relation to its sister species *H. tatayoi* (Castroviejo-Fisher et al., 2011; Delia, Alvarado & Aguilar, 2017). Here,

Comment [Office31]: The other possibility that should be discussed is that H. tatayoi is a synonym of H. fleischmanni. Are there good morphological and/or acoustic traits that differentiate these species?

we confirm the paraphyly of the species, as the *H. tatayoi* samples are grouped within the Southern clade, lacking significant genetic differences from the western Andes samples.

Furthermore, our overall Bayesian topology coincides with the results obtained by Delia,

Alvarado & Aguilar (2017) for 12S sequences; consequently, we identified three main isolated lineages with large genetic distances that could be considered as candidate species. However, because our analyses are based only on mtDNA differences, we recognize that other sources of evidence (morphologic, acoustic, and/or ecological) are needed to confirm the proposed potential species (Padial et al., 2014).

Conclusions

 We have conducted the most comprehensive analysis of genetic variation and divergence within *H. fleischmanni* to date, producing one of the few phylogenetic and phylogeographic studies for glassfrogs, with the exception of a few studies from Guyana (Castroviejo-Fisher et al., 2011; Jowers et al., 2015), and this is the first such study of a Central American species. Moreover, our results aided in the successful reconstruction of the historical biogeography of these frogs and dispersal and vicariance events during the history *H. fleischmanni* lineages, revealing a higher complexity for the species than expected, especially for the Northern lineage, in which significant population structure was found. Indeed, our results support the Talamanca range, the MPJ fault system, and the Chortis highlands as significant factors exerting effects on the dispersal of lowland amphibians during the late Pliocene and early Pleistocene. Additionally, we suggest that the IT acted as a corridor, rather than a barrier for *H. fleischmanni* during the early Pleistocene, while the Hess Escarpment and the Andes range did not play a significant role as barriers. The

complementary use of phylogenetic and landscape analyses allowed us to perform an adequate

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evaluation of dispersal patterns and potential barriers within this region; hence, our approach can be applied in biogeographic and phylogeographic studies of different taxa.

Acknowledgments

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535	References	Comment [Office32]: Review format. There are several inconsistencies.
536 537 538 539	Anderson DL, House P, Hyman RE, Steiner R, Hawkins HR, Thorn S, Rey MJ, Espinal MR, Marineros LE. 2010. Rediscovery of the Honduran Emerald <i>Amazilia luciae</i> in western Honduras: insights on the distribution, ecology, and conservation of a 'Critically Endangered' hummingbird. <i>Bird Conservation International</i> 20(3):255-262	
540 541 542	Arbeláez-Cortés E, Nyári ÁS, Navarro-Sigüenza AG, 2010. The differential effect of lowlands on the phylogeographic pattern of a Mesoamerican montane species (<i>Lepidocolaptes affinis</i> , Aves: Furnariidae). <i>Molecular Phylogenetics and Evolution</i> 57(2):658-668	
543 544 545	Bacon CD, Silvestro D, Jaramillo C, Smith BT, Chakrabarty P, Antonelli A. 2015. Biological evidence supports an early and complex emergence of the Isthmus of Panama. <i>Proceedings of the National Academy of Sciences</i> 112(19):6110-6115	
546 547	Bacon CD, Molnar P, Antonelli A, Crawford AJ, Montes C, Vallejo-Pareja MC. 2016. Quaternary glaciation and the Great American Biotic Interchange. Geology 44(5):375-378	Comment [Office33]: Italics?
548 549 550	Bagley JC, Johnson JB. 2014. Phylogeography and biogeography of the lower Central American Neotropics: diversification between two continents and between two seas. Biological Reviews 89(4):767-790	
551 552 553	Bagley JC, Hickerson MJ, Johnson JB. 2018 Testing hypotheses of diversification in Panamanian frogs and freshwater fishes using hierarchical approximate Bayesian computation with model averaging. PeerJ Preprints 6, e26623v1 https://doi.org/10.7287/peerj.preprints.26623v1	
554 555 556	Barrera-Guzmán AO, Milá B, Sánchez-González LA, Navarro-Sigüenza AG. 2012. Speciation in an avian complex endemic to the mountains of Middle America (<i>Ergaticus</i> , ves: Parulidae). Molecular phylogenetics and Evolution 62(3):907–920	
557	Beebee TJC 2005. Conservation genetics of amphibians. Heredity 95(6):423-427.	Comment [Office34]: .

558 559 560	Bielejec F, Baele G, Vrancken B, Suchard MA, Rambaut A, Lemey P. 2016. SpreaD3: interactive visualisation of spatiotemporal history and trait evolutionary processes. Molecular Biology and Evolution, 33(8):2167-2169	
561 562 563	Bryson RW, García-Vázquez UO, Riddle BR. 2011. Phylogeography of Middle American gophersnakes: mixed responses to biogeographical barriers across the Mexican Transition Zone. Journal of Biogeography 38(8):1570-1584.	
564 565 566	Cadena CD, Klicka J, Ricklefs RE. 2007. Evolutionary differentiation in the Neotropical montane region: molecular phylogenetics and phylogeography of <i>Buarremon</i> brush-finches (Aves-Emberizidae). Molecular Phylogenetics and Evolution 44(3):993–1016	
567 568 569	Castoe TA, Chippindale PT, Campbell JA, Ammerman LK, Parkinson CL. 2003. Molecular systematic of the middle American jumping pitvipers (genus <i>Atropoides</i>) and phylogeography of the <i>Atropoides nummifer</i> complex. Herpetologica 59(3):421–432	
570 571	Castroviejo-Fisher S, Ayarzagüena J, Vila C. 2007. A new species of <i>Hyalinobatrachium</i> (Centrolenidae: Anura) from Serrania de Perijá, Venezuela. Zootaxa 1441(1):51-62	
572 573 574	Castroviejo-Fisher S, Vila C, Ayarzagueena J, Blanc M, Ernst R. 2011. Species diversity of <i>Hyalinobatrachium</i> glassfrogs (Amphibia: Centrolenidae) from the Guiana Shield, with the description of two new species. Zootaxa 3132:1-55	
575 576	Castroviejo-Fisher S, Guayasamin JM, Gonzalez-Voyer A, Vilá C. 2014. Neotropical diversification seen through glassfrogs. Journal of Biogeography 41(1):66-80	Deleted: a
577 578 579	Cavender-Bares J, González-Rodríguez A, Pahlich A, Koehler K, Deacon N. 2011. Phylogeography and climatic niche evolution in live oaks (<i>Quercus</i> series Virentes) from the tropics to the temperate zone. Journal of Biogeography 38(5):962–981	
580 581 582	Cavers S, Navarro C, Lowe AJ ₂ 2003. Chloroplast DNA phylogeography reveals colonization history of a Neotropical tree, <i>Cedrela odorata</i> L., in Mesoamerica. Molecular Ecology 12(6):1451-1460.	Deleted: ,
583 584 585	Crawford AJ, Bermingham E, Polanía-S C. 2007. The role of tropical dry forest as a long-term barrier to dispersal: a comparative phylogeographical analysis of dry forest tolerant and intolerant frogs Molecular Ecology 16(22):4789-4807	
584	barrier to dispersal: a comparative phylogeographical analysis of dry forest tolerant and intolerant	

- 592 Cronin TM, Dowsett HJ. 1996. Biotic and oceanographic response to the Pliocene closing of the
- 593 Central American Isthmus. In: Jackson, JBC, Budd AF, Coates A, eds. Evolution and
- environment in tropical America. The University of Chicago Press, Chicago, 76-104.
- 595 Daza JM, Castoe TA, Parkinson CL, 2010. Using regional comparative phylogeographic data
- from snake lineages to infer historical processes in Middle America. Ecography 33(2):343–354
- 597 Delia J, Cisneros-Heredia DF, Whitney J, Murrieta-Galindo R. 2010. Observations on the
- 598 reproductive behavior of a Neotropical glassfrog, Hyalinobatrachium fleischmanni (Anura:
- 599 Centrolenidae). South American Journal of Herpetology 5(1):1-12
- 600 Delia J, Bravo-Valencia L, McDiarmid RW. 2017. Notes on paternal behavior in
- 601 *Hyalinobatrachium* glassfrogs (Anura: Centrolenidae). Phyllomedusa 16(1):101-107.
- 602 Denyer P, Alvarado GE, Aguilar T. 2000. Historia Geológica. in: P. Denyer and S. Kussmaul
- 603 (Eds.), Geología de Costa Rica. Editorial Tecnológica de Costa Rica, San José, pp. 155-167.
- 604 Drummond AJ, Rambaut A, Shapiro B, Pybus OG. 2005. Bayesian coalescent inference of past
- 605 population dynamics from molecular sequences. Molecular biology and evolution, 22(5):1185-
- 606 1192
- 607 Drummond AJ, Rambaut A. 2007. BEAST: Bayesian evolutionary analysis by sampling trees.
- BMC evolutionary biology, 7(1):214.
- 609 Drummond AJ, Ho SYW, Phillips MJ, Rambaut A. 2006. Relaxed phylogenetics and dating with
- 610 confidence. PLoS Biology 4(5):e88
- 611 Eizirik E, Kim JH, Menotti-Raymond M, Crawshaw Jr. PG, O'Brien SJ, Johnson WE. 2001.
- 612 Phylogeography, population history and conservation genetics of jaguars (*Panthera onca*,
- 613 Mammalia, Felidae). Molecular Ecology 10(1):65–79
- 614 French N, Yu S, Biggs P, Holland B, Fearnhead P, Binney B, Fox A, Grove-White D, Leigh JW,
- 615 Miller W, Muellner P, Carter P, 2014. Evolution of Campylobacter species in New Zealand, In:
- 616 Sheppard SK, Méric G. eds. Campylobacter Ecology and Evolution. Caister Academic Press.,
- 617 Norfolk, pp. 221–240.
- 618 García-Moreno J, Navarro-Sigüenza AG, Peterson AT, Sánchez-González LA. 2004. Genetic
- 619 variation coincides with geographic structure in the common bush-tanager (Chlorospingus
- 620 ophthalmicus) complex from Mexico. Molecular Phylogenetics and Evolution 33(1):186–196.
- 621 Guayasamin JM, Castroviejo-Fisher S, Ayarzagüena J, Trueb L, Vilà C. 2008. Phylogenetic
- 622 relationships of glassfrogs (Centrolenidae) based on mitochondrial and nuclear genes. Molecular
- Phylogenetics and Evolution 48(2):574-595

- 624 Gutiérrez-García TA, Vázquez-Domínguez E. 2013. Consensus between genes and stones in the
- 625 biogeographic and evolutionary history of Central America. Quaternary Research 79(3):311-324
- 626 Hasbún CR, Gómez A, Khöler GG, Lunt DH. 2005. Mitochondrial DNA phylogeography of the
- 627 Mesoamerican spiny-tailed lizards (*Ctenosaura quinquecarinata* complex): historical
- 628 biogeography, species status and conservation. Molecular Ecology 14(10):3095–3107
- Hauswaldt JS, Ludewig AK, Vences M, Pröhl H. 2011. Widespread co-occurrence of divergent
- 630 mitochondrial haplotype lineages in a Central American species of poison frog (Oophaga
- 631 *pumilio*). Journal of Biogeography, 38(4):711–726
- 632 Iturralde-Vinent MA. 2006. Meso-Cenozoic Caribbean paleogeography: implications for the
- 633 historical biogeography of the region. International Geology Review, 48(9):791–827
- 634 Jiménez RA, Ornelas JF. 2016. Historical and current introgression in a Mesoamerican
- 635 hummingbird species complex: a biogeographic perspective. PeerJ 4:e1556
- Jombart T, Devillard S, Dufour A-B, Pontier D. 2008a. Revealing cryptic spatial patterns in
- 637 genetic variability by a new multivariate method. Heredity 101(1):92-103
- Jombart T, 2008b. adegenet: a R package for the multivariate analysis of genetic markers.
- 639 Bioinformatics 24(11):1403-1405
- 640 Jombart T, Devillard S, Balloux F. 2010. Discriminant analysis of principal components: a new
- method for the analysis of genetically structured populations. BMC genetics 11(1):94
- Jowers MJ, Lehtinen RM, Downie RJ, Georgiadis AP, Murphy JC. 2015. Molecular
- 643 phylogenetics of the glass frog *Hyalinobatrachium orientale* (Anura: Centrolenidae): evidence
- 644 for Pliocene connections between mainland Venezuela and the island of Tobago. Mitochondrial
- 645 DNA 26(4):613-618
- 646 Kamvar ZN, Tabima JF, Grünwald NJ. 2014. Poppr: an R package for genetic analysis of
- 647 populations with clonal, partially clonal, and/or sexual reproduction. PeerJ 2:e281
- 648 http://dx.doi.org/10.7717/peerj.281
- 649 Kimura M. 1980. A simple method for estimating evolutionary rates of base substitutions through
- comparative studies of nucleotide sequences. Journal of molecular evolution 16(2):111-120
- Kumar S, Stecher G, Tamura K. 2016. MEGA7: molecular evolutionary genetics analysis version
- 7.0 for bigger datasets. Molecular biology and evolution 33(7):1870-1874
- Lanfear R, Frandsen P., Wright AM, Senfeld T, Calcott B. 2016. PartitionFinder 2: new methods
- 654 for selecting partitioned models of evolution for molecular and morphological phylogenetic
- analyses. Molecular Biology and Evolution, 34(3):772-773

- 656 Libardo P, Rozas J, 2009. DnaSP v5: a software for comprehensive analysis of DNA
- polymorphism data. Bioinformatics 25(11):1451-1452
- 658 Lyra ML, Haddad CF, Azeredo-Espin AML. 2017. Meeting the challenge of DNA barcoding
- Neotropical amphibians: polymerase chain reaction optimization and new COI primers.
- Molecular ecology resources 17(5):966-980
- 661 Marshall LG, Webb SD, Sepkoski JJ, Raup DM. 1982. Mammalian evolution and the great
- 662 American interchange. Science 215(4538):1351-1357
- 663 Marshall JS, Idleman BD, Gardner TW, Fisher DM. 2003. Landscape evolution within a
- retreating volcanic arc, Costa Rica, Central America. Geology 31(5):419-422
- 665 Marshall JS. 2007. The geomorphology and physiographic provinces of Central America. Central
- America: geology, resources and hazards, 1, 75-121.
- 667 Mendoza ÁM, Arita HT. 2014. Priority setting by sites and by species using rarity, richness and
- 668 phylogenetic diversity: the case of neotropical glassfrogs (Anura: Centrolenidae). Biodiversity
- and Conservation 23(4):909-926
- 670 Mendoza ÁM, Ospina OE, Cárdenas-Henao H, García-R JC. 2015. A likelihood inference of
- 671 historical biogeography in the world's most diverse terrestrial vertebrate genus: Diversification of
- direct-developing frogs (Craugastoridae: *Pristimantis*) across the Neotropics. Molecular
- phylogenetics and evolution 85:50-58
- 674 Mendoza ÁM, Torres MF, Paz A, Trujillo-Arias N, López-Alvarez D, Sierra S, Forero F,
- 675 González MA. 2016. Cryptic diversity revealed by DNA barcoding in Colombian illegally traded
- bird species. Molecular ecology resources 16(4):862-873
- 677 Montes C, Cardona A, McFadden R, Morón SE, Silva CA, Restrepo-Moreno S, Ramírez DA,
- 678 Hoyos N, Wilson J, Farris D, Bayona GA, Jaramillo CA, Valencia V, Bryan J, Flores JA, Bayona
- 679 GA. 2012a. Evidence for middle Eocene and younger land emergence in central Panama:
- implications for Isthmus closure. Bulletin, 124(5-6):780-799
- 681 Montes C, Bayona G, Cardona A, Buchs DM, Silva CA, Morón S, Hoyos N, Ramírez DA,
- 682 Jaramillo CA, Valencia V. 2012b. Arc-continent collision and orocline formation: Closing of the
- 683 Central American seaway. Journal of Geophysical Research: Solid Earth, 117(B4)
- Montes C, Cardona A, Jaramillo C, Pardo A, Silva JC, Valencia V, Ayala C, Pérez-Angel LC,
- Rodriguez-Parra LA, Ramirez V, Niño H. 2015. Middle Miocene closure of the Central
- 686 American seaway. Science 348(6231):226-229

Deleted:

- 688 Morrone JJ. 2014. Biogeographical regionalisation of the Neotropical region. Zootaxa 3782(1):1-
- 689 110
- 690 Mulcahy DG, Morrill BH, Mendelson JRI. 2006. Historical biogeography of lowland species of
- 691 toads (Bufo) across the Trans-Mexican Neovolcanic Belt and the Isthmus of Tehuantepec. Journal
- 692 of Biogeography 33(11):1889–1904
- 693 Myers N, Mittermeier RA, Mittermeier CG, Da Fonseca GA, Kent J. 2000. Biodiversity hotspots
- for conservation priorities. Nature 403(6772):853-858
- Navarro-Sigüenza AG, Peterson AT, Nyari A, García-Deras GM, García-Moreno J. 2008.
- 696 Phylogeography of the *Buarremon* brush-finch complex (Aves, Emberizidae) in Mesoamerica.
- 697 Molecular phylogenetics and evolution 47(1):21-35
- Noss RF. 1991. Landscape connectivity: different functions at different scales. Landscape
- 699 linkages and biodiversity. Island Press, Washington DC.
- 700 Ordóñez-Garza N, Matson JO, Strauss RE, Bradley RD, Salazar-Bravo J. 2010. Patterns of
- 701 phenotypic and genetic variation in three species of endemic Mesoamerican *Peromyscus*
- 702 (Rodentia: Cricetidae). Journal of Mammalogy 91(4):848–859
- Ornelas JF, Ruiz-Sánchez E, Sosa V, 2010. Phylogeography of *Podocarpus matudae*
- 704 (Podocarpaceae): pre-Quaternary relicts in northern Mesoamerican cloud forests. Journal of
- 705 Biogeography 37(12):2384–2396.
- 706 Padial JM, Miralles A, De la Riva I, Vences M, 2010. The integrative future of taxonomy.
- 707 Frontiers in zoology, 7(1):16
- 708 Palacios M, Voelker G, Rodriguez LA, Mateos M, Tobler M. 2016. Phylogenetic analyses of the
- 709 subgenus Mollienesia (Poecilia, Poeciliidae, Teleostei) reveal taxonomic inconsistencies, cryptic
- 710 biodiversity, and spatio-temporal aspects of diversification in Middle America. Molecular
- 711 phylogenetics and evolution 103:230-244.
- 712 Parkinson CL, Zamudio KR, Greene HW. 2000. Phylogeography of the pitviper clade
- 713 Agkistrodon: historical ecology, species status, and conservation of cantils. Molecular Ecology
- 714 9(4):411–420
- 715 Pérez-Consuegra SG, Vázquez-Domínguez E. 2017. Intricate evolutionary histories in montane
- 716 species: a phylogenetic window into craniodental discrimination in the *Peromyscus mexicanus*
- 717 species group (Mammalia: Rodentia: Cricetidae). Journal of Zoological Systematics and
- 718 Evolutionary Research 55(1):57-72
- 719 Pinto-Sánchez NR, Ibáñez R, Madriñán S, Sanjur OI, BerminghamE., Crawford AJ. 2012. The
- 720 great American biotic interchange in frogs: multiple and early colonization of Central America by

- 721 the South American genus Pristimantis (Anura: Craugastoridae). Molecular Phylogenetics and
- 722 Evolution 62(3):954-972.
- 723 Poelchau MF, Hamrick JL. 2011. Comparative phylogeography of three common Neotropical
- tree species. Journal of Biogeography 40(4):618-631
- 725 Ramírez DA, Foster DA, Min K, Montes C, Cardona A, Sadove G. 2016. Exhumation of the
- 726 Panama basement complex and basins: Implications for the closure of the Central American
- seaway. Geochemistry, Geophysics, Geosystems 17(5):1758-1777
- 728 Ree RH, Smith SA. 2008. Maximum likelihood inference of geographic range evolution by
- dispersal, local extinction, and cladogenesis. Systematic biology, 57(1):4–14.
- 730 Ree RH, Moore BR, Webb CO, Donoghue MJ. 2005. A likelihood framework for inferring the
- evolution of geographic range on phylogenetic trees. Evolution 59(11):2299–2311
- 732 Rivera-Ortíz FA, Solórzano S, Arizmendi MDC, Dávila-Aranda P, Oyama K. 2016. Genetic
- 733 Diversity and Structure of the Military Macaw (Ara militaris) in Mexico: Implications for
- 734 Conservation. Tropical Conservation Science 10:1-12
- 735 Robertson JM, Lips KR, Heist EJ. 2008. Fine scale gene flow and individual movements among
- 736 subpopulations of *Centrolene prosoblepon* (Anura: Centrolenidae). Revista de biologia tropical,
- 737 56(1):13-26
- 738 Ronquist F, Huelsenbeck JP. 2003. MRBAYES 3: Bayesian phylogenetic inference under mixed
- 739 models. Bioinformatics 19(12):1572–1574.
- Ronquist F, Sanmartín I, 2011. Phylogenetic methods in biogeography. Annual Review of
- 741 Ecology, Evolution, and Systematics 42:441–464.
- 742 Rovito SM, Vásquez-Almazán CR, Papenfuss TJ, Parra-Olea G, Wake DB. 2015. Biogeography
- and evolution of Central American cloud forest salamanders (Caudata: Plethodontidae:
- 744 Cryptotriton), with the description of a new species. Zoological Journal of the Linnean Society
- 745 175(1):150-166
- Ruiz-Carranza PM, Lynch JD. 1991. Ranas Centrolenidae de Colombia I: Propuesta de una
- nueva clasificación genérica. Lozania 57:1–30.
- Sambrook J, Russell DW. 2006. Purification of nucleic acids by extraction with phenol:
- 749 chloroform. Cold Spring Harbor Protocols 2006(1):pdb-prot4455
- 750 Savage JM. 2002. The Amphibians and Reptiles of Costa Rica: A Herpetofauna between Two
- 751 Continents, between Two Seas. University of Chicago press, Chicago.

- 752 Smith SA, Bermingham E. 2005. The biogeography of lower Mesoamerican freshwater fishes.
- 753 Journal of Biogeography 32(10):1835-1854.
- 754 Smith SA, Stephens PR, Wiens JJ. 2005. Replicate patterns of species richness, historical
- biogeography, and phylogeny in Holarctic treefrogs. Evolution 59(11):2433-2450
- 756 Smith BT, Amei A, Klicka J. 2012. Evaluating the role of contracting and expanding rainforest in
- 757 initiating cycles of speciation across the Isthmus of Panama. Proceedings of the Royal Society of
- 758 London B: Biological Sciences, 279(1742):3520-3526.
- 759 Stamatakis A. 2006. RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses with
- thousands of taxa and mixed models. Bioinformatics 22(21):2688-2690
- 761 Stehli FG, Webb SD. 1985. A kaleidoscope of plates, faunal and floral dispersals, and sea level
- changes, in: Stehli FG, Webb SD. eds. The great American biotic interchange. Springer, Boston,
- 763 3-16.
- 764 Streicher JW, García-Vázquez UO, Ponce-Campos P, Flores-Villela O, Campbell JA, Smith EN,
- 765 2014. Evolutionary relationships amongst polymorphic direct-developing frogs in the Craugastor
- 766 rhodopis species group (Anura: Craugastoridae). Systematics and Biodiversity 12(1):1-22
- 767 Townsend JH, Wilson LD, Medina-Flores M, Aguilar-Urbina E, Atkinson BK, Cerrato-Mendoza
- 768 CA, Contreras-Castro A, Gray LN, Herrera-B. LA, Luque-Montes IR, McKewy-Mejía M,
- 769 Alionso Portillo-Avilez A, Stubbs AL, McKewy-Mejía, M. 2012. A premontane hotspot for
- herpetological endemism on the windward side of Refugio de Vida Silvestre Texíguat, Honduras.
- 771 Salamandra 48(2):92-114.
- 772 Townsend JH, 2014. Characterizing the Chortís Block Biogeographic Province: Geological,
- physiographic, and ecological associations and herpetofaunal diversity. Mesoamerican
- 774 Herpetology 1(2):204-252
- 775 Townsend JH, & Wilson LD. 2016. Amphibians of the Cordillera Nombre de Dios, Honduras:
- 776 COI barcoding suggests underestimated taxonomic richness in a threatened endemic fauna.
- 777 Mesoamerican Herpetology, 3(4):910-927.
- 778 Valencia-Aguilar A, Castro-Herrera F, Ramírez-Pinilla MP. 2012. Microhabitats for oviposition
- 779 and male clutch attendance in Hyalinobatrachium aureoguttatum (Anura: Centrolenidae). Copeia
- 780 2012(4):722-731
- 781 Vázquez-Miranda H, Navarro-Sigüenza AG, Omland KE. 2009. Phylogeography of the rufous-
- naped red (Camphylorhynchus rufinucha): Speciation and hybridization in Mesoamerica. The
- 783 Auk 126(4):765–778
- 784 Venegas-Anaya M, Crawford AJ, Escobedo-Galván AH, Sanjur OI, Densmore LD, Bermingham
- 785 E, 2008. Mitochondrial DNA phylogeography of $\it Caiman\ crocodilus$ in Mesoamerica and South

- America. Journal of Experimental Zoology Part A: Ecological and Integrative Physiology
- 787 309(10):614-627
- Villa J, Valerio CE. 1982. Red, white and brown: preliminary observation on the color of the
- centrolenid tadpole (Amphibia: Anura: Centrolenidae). Brenesia 1982(19/20), 1-16.
- 790 Wang IJ, Crawford AJ, Bermingham E. 2008. Phylogeography of the pygmy rain frog
- 791 (Pristimantis ridens) across the lowland wet forest of isthmian Central America. Molecular
- 792 phylogenetics and evolution 47(3):992–1004
- 793 Wiens JJ, Graham CH, Moen DS, Smith SA, Reeder TW. 2006. Evolutionary and ecological
- 794 causes of the latitudinal diversity gradient in hylid frogs: treefrog trees unearth the roots of high
- 795 tropical diversity. The American Naturalist 168(5):579-596
- 796 Yu Y, Harris AJ, Blair C, He X-J. 2015. RASP (Reconstruct Ancestral State in Phylogenies): a
- 797 tool for historical biogeography. RASP (Reconstruct Ancestral State in Phylogenies): a tool for
- historical biogeography. Molecular phylogenetics and evolution 87:46-49.

Figure Captions 799 Fig. 1. Geographic distribution of H. fleischmanni samples. Main geological blocks, delimited by 800 801 geological barriers, are shown. Sample origins are indicated by black dots, while gray polygons show the species distribution according to IUCN. SMB=South Mexican block, IT=Isthmus of 802 Tehuantepec, SAP=South American Plate, MPJ=Motagua-Polochic-Jocotán. 803 804 Fig. 2. Results of the Bayesian (BAPS, Geneland) and multivariate spatial analyses (sPCA) for H. fleischmanni population clustering based on 16S (first column), COI (second column) and 805 ND1 (third column) sequences. For the sPCA analysis, the color of each point is determined in 806 the red-green-blue (RGB) system based on each individual's score on the first (translated to a red 807 808 channel) and second axes (translated to green) of the sPCA. Fig 3. Haplotype networks of mitochondrial DNA haplotypes for 16S (A) COI (B) and ND1 (C) 809 from H. fleischmanni. Hatch marks represent inferred mutational steps. The size of the circle is 810 811 proportional to the number of individuals found for each haplotype. Fig 4. Time-calibrated tree of H. fleischmanni unique haplotypes, inferred from BEAST based on 812 813 the combined ribosomal (16S) and protein-coding (COI, ND1) mitochondrial sequences, with 814 calibration on three nodes indicated by green bars (see Materials and Methods section for details). 815 Blue rectangles over key nodes indicate the 95% highest posterior densities (HPD) of the estimated times of divergence events (in Ma). Clade support is indicated by posterior BI values 816 817 in BEAST and Mr Bayes and by RAxML Bootstrap analysis and is presented in this order separated by a slash. Asterisks at tips represent H. tatayoi samples included in the analysis. The 818 inner map shows the geographic locations of haplotype lineages. Each color in the map coincides 819 820 with the haplogroup obtained in the phylogenetic reconstruction.

821 Fig. 5. Bayesian skyline plots for Northern (A) and Southern (B) clades generated through phylogenetic reconstruction. 822 Fig. 6. Spatial projection of the Bayesian spatiotemporal diffusion analysis of *H. fleischmanni* 823 lineages for three time points, based on the maximum clade credibility (MCC) tree estimated 824 with a "Relaxed Random Walk" model. Lines represent branches of the MCC tree; shaded areas 825 indicate the 95%-HPD uncertainty for the ancestral branches; the shading gradient indicates older 826 (lighter) versus younger (darker) events; and dot color represents the ages of older nodes (darker) 827 and younger tips (lighter). 828

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