

Influencing factors and health risk assessment of microcystins in the Yongjiang river (China) by the Monte Carlo simulation method

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A total of 90 water samples and associated environmental data were collected from three sample points from March–December 2017 from river Yongjiang. The relationship between the environmental factors and the microcystins (MCs) concentration were investigated in this article. The results showed that total phosphate (TP) is the dominant factor affecting the contents of MC-LR and the total content of MCs (TMCs), while pH and total nitrogen/total phosphate (TN/TP) ratio are the main factors affecting the content of MC-RR and MC-YR, respectively. Professional risk assessment software @Risk7.5 was used to evaluate the risk of dietary intake of microcystins (MCs). The result indicated that the health risks of MC-RR from drinking water source were higher than those of MC-LR and MC-YR, and the pollution of MCs would lead to high potential health risks, especially in children. Health risks for children found for MC-RR from drinking water sources were higher than the maximum allowance levels recommended by the US Environmental Protection Agency (USEPA) (1×10^{-4}), and health risks for adults found for MC-RR from drinking water sources were higher than the maximum allowance levels recommended by the International Commission on Radiological Protection (ICRP) (5×10^{-5}). Therefore, it is necessary to strengthen the protection and monitoring of drinking water source for effective control of water pollution and safeguarding of human health.

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21 **ABSTRACT**

22 A total of 90 water samples and associated environmental data were collected from three sample
23 points from March–December 2017 from river Yongjiang. The relationship between the
24 environmental factors and the microcystins(MCs) concentration were investigated in this article.
25 The results showed that total phosphate (TP) is the dominant factor affecting the contents of MC-
26 LR and the total content of MCs (TMCs), while pH and total nitrogen/total phosphate(TN/TP)
27 ratio are the main factors affecting the content of MC-RR and MC-YR, respectively.
28 Professional risk assessment software @Risk7.5 was used to evaluate the risk of dietary intake of
29 microcystins (MCs). The result indicated that the health risks of MC-RR from drinking water
30 source were higher than those of MC-LR and MC-YR, and the pollution of MCs would lead to
31 high potential health risks, especially in children. Health risks for children found for MC-RR
32 from drinking water sources were higher than the maximum allowance levels recommended by
33 the US Environmental Protection Agency (USEPA) (1×10^{-4}), and health risks for adults found
34 for MC-RR from drinking water sources were higher than the maximum allowance levels
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36 Therefore, it is necessary to strengthen the protection and monitoring of drinking water source
37 for effective control of water pollution and safeguarding of human health.

38 **Subjects** Health Ecotoxicology and Environmental Impacts

39 **Keywords** Microcystins; Environmental factors; @Risk7.5; Monte Carlo method; Health risk
40 assessment

41 1. INTRODUCTION

42 Eutrophication of freshwater bodies can result in algal blooms, especially those caused by
43 cyanobacteria. The outbreak of Cyanobacteria blooms secretes a secondary metabolite of the
44 algal toxin that inhibits the growth of other species in water and ensures the growth advantage of
45 algae itself (Ame et al. 2003, Cao et al. 2017). Microcystin is a type of algal toxin consisting of
46 monocyclic peptides with more than 90 isomers (Yang et al. 2017). Of these, the most widely
47 distributed and highly toxic are microcystin-LR (MC-LR), microcystin-RR (MC-RR), and
48 microcystin-YR (MC-YR) (Zegura 2016). These toxins are synthesized in the cells and released
49 after cell rupture, resulting in the appearance of microcystins in the water source.

50 Cyanobacteria blooms appear in eutrophicated waters worldwide, and MCs can be
51 bioaccumulated by aquatic animals and can reach humans, severely endangering human health
52 and leading to illness or death. In 1975, the drinking water source in the small town of
53 Pennsylvania was contaminated with MCs, which resulted in more than half of the local
54 population suffering from acute gastroenteritis (Keleti et al. 1982). In 1996, 116/131 patients,
55 who received routine treatment at the Brazil hemodialysis center, presented abnormal symptoms
56 such as blurred vision, nausea, and vomiting due to MCs contamination, and >50 individuals
57 succumbed to mortality (Pouria et al. 1998). Studies on drinking water showed that in Haimen
58 and Fu Sui, where liver cancer were high incidence in China, the content of MCs in the water

59 were high as well ([Ueno et al. 1996](#)). In 2010, the International Agency for Research on Cancer
60 (IARC) listed microcystins as a “possible human carcinogen” (Group 2B) based on its potential
61 carcinogenicity ([2010](#)).

62 Although MCs have been confirmed to cause acute and chronic damage to the human body,
63 reports on MC-LR, MC-RR, and MC-YR and their risk assessment in Yongjiang river of China
64 are yet lacking. The environmental conditions of water source are crucial in toxin production.
65 However, the factors affecting the production of MCs (nutrient levels and climatic conditions) in
66 Yongjiang have not yet been elucidated. Therefore, the present study investigated the
67 concentration and distribution of MCs in Yongjiang river as influenced by seasonal changes in
68 water quality and the related parameters.

69 Owing to the requirement of prolonged duration and inoperability of toxicological health
70 risk assessments, the present study could not be conducted in a population. Thus, the Monte
71 Carlo simulation mathematical and logical model was used. In recent years, this model has been
72 used in order to understand the behavior of water systems. This would aid in decision-making
73 due to the main advantage of the ability to model any appropriate assumptions of the problem or
74 the system ([Clausen et al. 2017](#)). Moreover, the Monte Carlo simulation of uncertainties was
75 applied in the risk assessment model by collecting limited samples to predict the overall situation.
76 In this regard, the risk uncertainty is intuitively expressed, which is in agreement with the nature
77 of the uncertainty and predictability of the risk; the risk managers and policymakers can provide
78 a favorable intuitive and scientific basis ([Paladino et al. 2017](#), [Sasi et al. 2017](#)). The US

79 Environmental Protection Agency (USEPA) has set Monte Carlo simulation as a basic method in
80 the risk analysis policy (Moolenaar 1996). Currently, American Palisade developed @Risk
81 Monte Carlo software that is loaded into Excel simulation technology for risk analysis based on
82 the professional software. The software is mainly based on the analysis of stochastic simulation
83 method of Monte Carlo that provides various potential results using a variety of probabilistic
84 simulations, the probability of occurrence of events that constitute a risk, forecast the risk of
85 uncertainty quantitatively, and summarize the characterization results. Li et al. (2017) used the
86 Monte Carlo model to assess the quantitative risk of aluminum in Youtiao, which was not found
87 to exceed the provisional tolerable weekly intake (PTWI) set by the joint FAO/WHO Expert
88 Committee on Food Additives (JECFA) for the public. Jia et al.(2018) used the Monte Carlo
89 simulation to evaluate the trace elements in four freshwater fish from a mine-impacted river, and
90 found that consumption did not exert any appreciable adverse impact on human health due to the
91 exposure to trace elements in fish muscle.

92 The present study was conducted to investigate the current status of drinking water sources
93 in the Yongjiang river in China regarding the contamination of MCs. A risk assessment was
94 conducted using the Monte Carlo simulation. These findings provide a basis to undergo an
95 effective control of water pollution and quality in order to protect the human health of the area.

96 **2. MATERIALS AND METHODS**

97 **2.1. Sampling location**

98 Yongjiang river (108°06'-108°49'E, 22°50'-22°52'N) is a major water resource with an
99 average annual flow of 1292 m³/s in the Nanning City area. The nutrient enrichment is the
100 causative for the reproduction of algae. It is the main urban water source in Nanning city, China,
101 and the tributary channel is also a vital transportation route. The surface area of the lake is 2676
102 ha with a maximum depth of 23 m and is used for entertainment as well as a source of water for
103 domestic use, agriculture, fishery, and industry. The nutrients entering the Yongjiang River come
104 from the cultivated land and farms. The nutrient enrichment is causative for the reproduction of
105 algae.

106 **2.2. Sampling**

107 Specifically, water samples were collected from the Yongning river in Nanning City from
108 March to December in 2017. For this, the river section was set in a total of three sampling points:
109 Qingxiu District, Jiangnan District, and Yongning District, respectively. The water samples were
110 collected 3 times/month from each sampling point, i.e., a total of 90 samples were collected in 10
111 months. Sample collection, containers, stabilization, and transportation to the laboratory were in
112 accordance with the methods described in Pesce et al. ([Pesce et al. 2002](#)).

113 **2.3. Water quality analysis**

114 Water parameters [for example, χ^1 =Water Temperature, χ^2 =pH, χ^6 =Dissolved Oxygen
115 (DO)] were measured in situ and [for example, χ^3 =Total Phosphate (TP), χ^4 =PO₄³⁻-P, χ^5 =Total
116 Nitrogen (TN)] were measured in laboratory. The data represented the results obtained from

117 three experiments. The analysis of all the above water samples was in reference to the standard
118 methods ([Wang et al. 2002](#)).

119 **2.4. Determination of MCs**

120 Water samples were collected from the sampling sites and passed through the sand funnel to
121 remove large particles of impurities and were stored at 4°C and protected from light. A total of
122 2000-ml water sample was collected, and 500-mL water sample is passed through the 0.45-µm
123 filter under reduced pressure filtration and extracted two times with 5mL ultra-pure water after
124 five times freezing-thawing at -80 °C/37 °C. After filtering through the 0.45-µm hybrid filter for
125 removing the algal cells, the solution was passed through solid phase extraction (SPE) (500 mg/6
126 mL) (SUPELCO, USA). The SPE was rinsed with 20 mL of 20% methanol and 10 mL deionized
127 distilled water. The toxin was eluted from the stationary phase with 80% methanol (containing
128 0.05% TFA), and each sample was dried in a water bath at 60 °C). The air-dried samples were
129 suspended into 1 mL deionized distilled water for high performance liquid chromatography
130 (HPLC) (Shimadzu LC-10A, Japan) analysis. The solubilized toxin samples were analyzed using
131 HPLC with UV detector at 238 nm and symmetrical C₁₈ column (3.9×150 mm²) (Waters, USA).
132 The mobile phase constituted of 33% acetonitrile and 67% deionized distilled water in 0.1%
133 phosphate buffer (pH=3). The flow rate was set at 1 mL/min. MC-LR and MC-RR (Solarbio,
134 approximate purity, 95%) and MC-YR (Alexis, approximate purity 98%) were used as standards.
135 Furthermore, the concentrations of MCs were determined by calibrating such area under the peak
136 with the corresponding standard curves.

137 2.5. Method of risk assessment

138 2.5.1. Construction of exposure assessment model

139 The three main sources of exposure to pollutants were intake, inhalation, and skin
140 absorption; the proportion of each was different due to the variations in the pollutants. The
141 present study assessed the risk exposure caused by drinking water. The daily exposure to MC-
142 RR, MC-YR, and MC-LR by direct drinking water was assessed using the Monte Carlo
143 simulation by @Risk7.5 software operating platform; the Bootstrap sampling method was used
144 for the quantification of uncertainty. Each Bootstrap sample was simulated with 10,000 Monte
145 Carlo simulations to obtain different percentile values (P5–P95) for determining its uncertainty.
146 The probabilistic assessment method was used to construct the exposure evaluation model
147 ([Zobitz et al. 2011](#)).

148 The mechanism underlying the different exposures and various exposure routes with
149 different exposure dose formula were employed for the exposure assessment model of the health
150 risk of chemical pollutants from the US environmental protection agency. Chronic daily intake
151 (CDI) evaluated the safety of MCs in drinking water and the health risks of different routes in
152 different populations. The chronic daily intake ($\mu\text{g}/\text{kg}/\text{d}$] formula is as follows.

153 Daily exposure of drinking water ([Duy et al. 2000](#), [Funari et al. 2008](#)):

$$\text{CDI} = \frac{C_w \times \text{IR} \times \text{EF} \times \text{E}}{\text{BW} \times \text{AT}}$$

154 Here, C_w is the concentration of pollutants in the water, $\mu\text{g}/\text{L}$; IR is the amount of drinking

155 water, L/d; EF is the frequency of exposure, drinking water for daily necessities, 365d/a; ED is
156 exposure duration, a; BW is the average body weight, kg; AT is the average time, ED×365 d/a.
157 According to the WHO, the standard weight of adults is 70 kg, and the daily drinking amount is
158 2 L/d; while the weight of children is 16 kg and the daily drinking amount is 1 L/d (van et al.
159 2009).

160 2.5.2. Construction of risk description model

161 The characteristics of the pollutants in a water environment are generally divided into genetic,
162 toxic substances (including radioactive pollutants and chemical carcinogens) and somatic toxic
163 substances (for instance, non-carcinogens) as recommended by the USEPA water environmental
164 health risk assessment model. The risk and non-cancer risk MCs from carcinogenic exposure
165 from drinking water is calculated.

166 (1) Health hazard risk model of chemical carcinogens

167 The formula of health hazard risk caused by chemical carcinogens recommended by
168 USEPA (Duy et al. 2000):

$$R_i^c = \frac{1 - \exp(-D_i q_i)}{70}$$

169

170 In the formula, R_i^c is the average personal carcinogenic annual risk of chemical carcinogen
171 i through drinking water, a^{-1} ; 70 indicates the average life expectancy of Chinese population, a;

172 D_i is the daily average exposure to chemical carcinogen i through drinking water, i.e., CDI,
173 $\mu\text{g}/\text{kg}/\text{d}$; q_i is the carcinogenic strength coefficient of the chemical carcinogen i through the
174 drinking water, and currently, there is no recognized carcinogenic intensity coefficient of MCs.
175 Based on the formula of carcinogenic strength coefficient of carcinogens (Hitzfeld et al. 2000),
176 this study deduced the formula as follows:

177

$$\text{CPI} = \frac{(\text{OR}-1) \times \text{LR}}{D}$$

178 Where, CPI is the coefficient of carcinogenic strength estimated from the population data,
179 namely q_i , $\text{kg}/\text{d}/\mu\text{g}/\text{L}$. According to the population of 80,000 inhabitants, the study showed that
180 the person who drank the river water presented a liver cancer OR of 1.246 (Falconer and
181 Buckley 1989, Yeh et al. 1989). MC-LR indicated the risk of cancer in an individual in the whole
182 local population. This parameter was not dimensionless and was calculated according to the risk
183 of cancer during the individual's lifespan in the population of China: 6.2×10^{-3} (Yeh et al. 1989).
184 D indicated the calculation of the average daily life exposure dose, $\mu\text{g}/\text{kg}/\text{d}$. According to the
185 results of the study by Wolf et al., the carcinogenic strength of MC-RR was set to one-tenth of
186 that of MC-LR, while the strength of carcinogenicity of MC-YR and MC -LR was
187 equivalent(Wolf and Frank 2002).

188 (2) Non-carcinogenic health risk assessment model

189 The health risk assessment model recommended by USEPA was used to evaluate the non-

190 carcinogenic health risk of MCs in the Yongjiang river source water. The non-carcinogenic risk
191 was described using a hazard index (HI) by the following formula:

$$192 \quad \text{HI} = \frac{\text{CDI}}{\text{RfD}}$$

193

194 Here, *RfD* is the reference dose for MCs, the internationally accepted *TDI* instead of MC-
195 LR *RfD* was 0.04 µg/kg/d. According to the equivalent toxicity relationship among MC-RR,
196 MC-YR and MC-LR, the *RfD* values of MC-RR and MC-YR were 0.4 and 0.04 µg/kg/d,
197 respectively (Wolf and Frank 2002, Lee et al. 2017).

198 The HI to 1 is usually calculated for the benchmark: HI >1 indicates harm to the human
199 body; the value higher than the reference dose for exposure may be dangerous; HI ≤ 1 indicates
200 that the extent of damage is smaller and the exposure level is lower than the reference dose,
201 which is unlikely to be detrimental (Younes 1999).

202 2.6. Statistical analysis

203 IBM SPSS Statistics 22.0 software was used to perform all descriptive statistical analysis
204 included minimum value, maximum value, mean value, Pearson's correlation analysis and
205 stepwise multiple linear regression. Moreover, the risk assessment of MC-RR, MC-YR, and MC-
206 LR exposure in water source was carried out using @Risk7.5 probabilistic evaluation software
207 based on the Monte Carlo simulation technique.

208 3. RESULTS

209 **3.1. Analysis of MCs by HPLC**

210 Based on the high-performance liquid chromatography (HPLC) analysis method
211 recommended by the national standard (GB/T 20466-2006), the separation conditions were
212 optimized, and the methodological test study conducted according to the experimental apparatus.
213 The order of the peaks and time of each standard substance were as follows: MC-RR (7.599 min),
214 MC-YR (14.125 min), and MC-LR (17.601 min). The test sample was analyzed in 18 min
215 (Figure 1).

216 **3.2. Concentration level, distribution characteristics, and environmental impact factors of** 217 **MCs in source water**

218 **3.2.1. Concentration distribution characteristics of MCs in source water**

219 The concentration of MC-LR, MC-RR, and MC-YR in water samples were detected, the
220 total content of MCs (TMCs) in water is the sum of the content of extracellular MCs (EMCs) and
221 intracellular MCs (IMCs) dissolved in the water, and the results summarized in Table 1.

222 **3.2.2. Seasonal distribution characteristics of MCs in water samples**

223 The seasonal distribution of MCs in Yongjiang river source is shown in Figure 2, which
224 demonstrated that the concentration of MC-YR (refers to the sum of intra and extra and the same
225 below) was significantly lower than the other two MCs. The concentrations of MC-RR and MC-
226 YR peaked in October, respectively.

227 3.2.3. Pearson's correlation analysis of environmental factors and MCs concentration

228 Pearson's correlation analysis of environmental factors and MCs concentration was
229 analyzed and the results shown in Table 2. According to the correlation analysis, the
230 concentration of MC-RR was positively correlated with water temperature and TN ($p < 0.01$),
231 with significant negative correlation with pH ($p < 0.01$); The concentration of MC-YR was
232 negative correlated with TN:TP ratio ($p < 0.05$); The concentration of MC-LR was positively
233 correlated with water temperature, TP and $\text{PO}_4^{3-}\text{-P}$ ($p < 0.01$), with significant negative
234 correlation with DO, TN:TP ratio ($p < 0.01$); The concentration of TMCs was positively
235 correlated with water temperature and TP ($p < 0.01$), with significant negative correlation with pH,
236 DO and TN:TP ratio ($p < 0.01$).

237 3.2.4. Stepwise multiple linear regression analysis of MCs concentration and environmental 238 factors

239 Stepwise multiple linear regression analysis of MCs concentration and environmental
240 factors was shown in Table 3. The results indicate that TP (χ^3) is the dominant factor affecting
241 the contents of MC-LR and TMCs. pH (χ^2) and TN:TP ratio (χ^7) are the primary factors affecting
242 the content of MC-RR and MC-YR, respectively. These findings were in agreement with the
243 results of the correlation analysis.

244 3.3. Assessment of MCs exposure in source waters

245 The risk assessment of MC-RR, MC-YR, and MC-LR exposure in water source was carried

246 out using @Risk7.5 probabilistic evaluation software based on the Monte Carlo simulation
247 technique (Sasaki et al. 2017). The corresponding probability distribution of various parameters
248 used in the assessment was based on @Risk7.5 provided by the standard distribution function.

249 3.3.1. Distribution fitting of the concentration of MCs in source water

250 The present study used @Risk7.5 software to fit the processed samples. The sample data of
251 MCs concentration in source water was characterized as continuous data. The fitting results were
252 Gamma, Invgauss, Lognorm, Expon, and Loglogistic, followed by the optimal fitting distribution
253 model. Three main methods were used to test the goodness of fit: Chi-Sq (Chi-squared) test, K-S
254 (Kolmogorov–Smirnov) test, and A-D (Anderson–Darling) test (Lipton et al. 1995, Cummins et
255 al. 2009). Taken together, the sample fitting results (Table 4) were used to determine the fitting
256 distribution types of the optimal probability of the pollution data: MC-RR had Gamma and
257 Invgauss distribution, MC-YR had Lognorm, Expon, and Loglogistic distribution, and MC-LR
258 had Gamma and Expon distribution. Furthermore, the results of the distribution parameters after
259 fitting and comparison with the sample data parameters are summarized in Table 5.

260 The abscissa in Figures 3, 4, and 5 represent the concentrations of MC-RR, MC-YR, or
261 MC-LR; The concentrations are partitioned, the length of each interval is the group distance, the
262 rectangular area is the frequency of the group, and the ratio of the total sample. The vertical axis
263 is the frequency divided by the group distance obtained, and the curve is the most suitable
264 distribution probability distribution. The fitting result can be visually observed from the
265 coincidence of the rectangle shape and the figure.

266 3.3.2. Daily exposure calculation

267 The @Risk7.5 software was utilized for the random extraction of the MCs concentration
268 profiles from the water to calculate the daily exposure of direct drinking water by different
269 populations to MC-RR, MC-YR, and MC-LR. Each simulation cycle was performed for 10,000
270 cycles, and the simulation results are shown in Table 6. In adults and children, a significant
271 difference was observed in daily exposure. P50, P85, P90, and P95 (Table 6) represent the high
272 exposure sites of each population. The MCs exposed to drinking water showed that the children's
273 daily intake was 2-fold higher than that of the adults, suggesting that children are more
274 susceptible to the pollution of MCs than adults.

275 3.4. Risk characterization of MCs in source water

276 3.4.1. Carcinogenic risk of MCs in source water

277 Based on the exposure parameters and carcinogenic risk formula, @Risk7.5 risk analysis
278 software was used to extract the numerical value of MCs concentration in water randomly and
279 calculate the carcinogenic risk of MC-RR, MC-YR, and MC-LR intake by different groups of
280 individuals through direct drinking water. Each simulation cycle of 10,000 displayed the
281 statistical simulation results summarized in Table 7. The carcinogenic annual risk of MC-YR
282 was less than that of MC-LR and MC-RR, and MC-RR was the primary hazard in the source
283 water. The maximum acceptable level and the negligible level of carcinogenic risk for the
284 population recommended by some institutions is listed in Table 8. The annual risk of
285 carcinogenesis of MCs in a water source is 10^{-6} – 10^{-4} . The carcinogenic risk of MC-YR and MC-

286 LR in adults and children is lower than the maximum acceptable risk level designated by USEPA
287 (1×10^{-4}) and ICRP (5×10^{-5}), and the risk of cancer in children is higher than that in adults. The
288 health risks caused by MC-RR from drinking water source for children were significantly higher
289 than the maximum allowance levels recommended by USEPA (1×10^{-4}). Also, the health risks
290 caused by the MC-RR from drinking water source for adults were significantly higher than the
291 maximum allowance levels recommended by ICRP (5×10^{-5}). These statistical details indicated
292 that MC-RR in water bodies exhibited a significant carcinogenic risk to the health of adults and
293 children.

294 **3.4.2. Non-carcinogenic risk of MCs in source water**

295 The exposure parameters and non-carcinogenic hazards index (*HI*) formula was used to
296 calculate the value of different populations through direct drinking water intake of MC-RR, MC-
297 YR, and MC-LR, using @Risk7.5 risk analysis software. Each simulation cycle for a total of
298 10,000 times provided the statistical simulation results (Table 9), which demonstrated that the
299 average non-carcinogenic risk index of MCs in different populations through drinking water
300 intake and the non-carcinogenic risk index of P90 and P95 at high levels of exposure were <1 .
301 This suggested that MCs, which are ingested through drinking water, pose less risk to health. In
302 particular, the non-carcinogenic risk index of MC-RR was less than that of MC-YR, and the non-
303 carcinogenic risk index of MC-YR was less than that of MC-LR, indicating that MC-LR was
304 more hazardous to human health than MC-YR and MC-RR. The MC-RR, MC-YR, and MC-LR
305 display a non-carcinogenic index in children than adults, and thus, MCs are detrimental to

306 children.

307 **4. DISCUSSION**

308 World Health Organization (WHO) suggested that the concentration of MC-LR in drinking water
309 was $<1 \mu\text{g/L}$ (Ibelings, Backer et al. 2015, Menezes, Churro et al. 2017). Australia has developed
310 a long-term safe drinking water guideline with a safety standard of 500 Microcystis cells/mL.
311 The short-term drinking water index was $<0.1 \mu\text{g/L}$ and 5000 Microcystis cells/mL (Nishiwaki-
312 Matsushima, Ohta et al. 1992). Another study suggested that the maximum acceptable level in
313 drinking water was $0.5 \mu\text{g/L}$ of MC-LR, or a lack of equivalent values for other MCs, $1 \mu\text{g/L}$ of
314 total MCs (Dittmann, Neilan et al. 1997).

315 The water quality monitoring in Yongjiang river demonstrated that although no major algal
316 bloom occurred, MC-RR, MC-YR, and MC-LR were present in the water column during the
317 monitoring period. These pollutants indicated that the concentration of MCs in the Yongjiang
318 river was influenced by seasonal changes in water quality-related parameters. Furthermore, the
319 concentration of MC-YR was significantly lower than the other two MCs; This finding was
320 similar to that from others studies, which suggested that the MCs are primarily dominated by
321 MC-RR and MC-LR (Yang, Xie et al. 2006, Bi, Dai et al. 2017). The temperatures in March and
322 April rise gradually; however, the level of MC-LR and MC-RR began to increase in May, which
323 might be attributed to the slow growth of algae. With the significant increase in the growth
324 conditions such as temperature, algal cell growth, accelerated cell aging, the autolysis

325 phenomenon also increased significantly, as well as, the released concentration of MCs in water
326 with respect to MC-RR and MC-YR was elevated. The peak of TMCs content appeared in
327 October, which was related to the higher water temperature in the same period and the suitable
328 climatic conditions. The significant decrease in the TMCs content in November and December
329 might occur due to the decrease in temperature, the variable growth of *Microcystis*, and death of
330 algal cells. These phenomena were similar to those described previously in Tai, Yang-cheng, and
331 Xuanwu lakes in China (Xu, Wang et al. 2010, Li, Gu et al. 2014).

332 Previous studies demonstrated that the concentration of MCs was associated with the
333 nutrient level and climate conditions. Water temperature played a major role in the growth of
334 *Microcystis*. Reportedly, under low temperature, *Microcystis* grew slowly, and its ability to adapt
335 to low temperature was weaker than that of the other algae (Wang et al. 2010). However, the
336 study also found that the influence of water temperature on 3 types of MCs was identical; MC-
337 LR and MC-RR exhibited significant effects depending on the water temperature, while the MC-
338 YR is correlated to the temperature of water had little effect (Table 2). Before October, the water
339 content of MC-RR is low that peaked in October when the water temperature began to decline;
340 Thus, the changes in the process and temperature is not synchronized, indicating the absence of a
341 marked correlation with the water temperature. Intriguingly, water pH was also shown to
342 influence the toxin production in Nanning Yongjiang river. The maximum toxicity was detected
343 at a pH below or above the medium level. As a result, MC-RR and TMCs were negatively
344 correlated with pH (Table 3), which was similar to the results from other studies. Notably, the

345 pH value cannot be used as an appropriate parameter to determine the toxin production. A
346 majority of the blue-green algae can grow well in the water at pH 6.5–7.9 (Wang et al. 2002).
347 The pH of Yongjiang river was within this range. DO showed a negative correlation of MC-LR
348 with TMCs in Yongjiang river. However, some researchers indicated that the increased level of
349 dissolved oxygen is related to the concentration of MCs (Wang et al. 2002). The relationship
350 between toxin and DO could be attributed to the relative stable DO values measured in
351 Yongjiang river. Correlation analysis results indicated that increasing the TP concentration could
352 increase the MCs production, especially that of MC-LR. The current observations were in
353 agreement with those from a study conducted in the large eutrophic Lake Erie in the USA (Harke
354 et al. 2016) that demonstrated positive correlations between TP and the abundance of toxic
355 Microcystis and MCs. Consistent with the trend, Vézic et al. (2002) also found that higher P
356 concentrations were beneficial to the growth of toxic Microcystis. Although TP was a dominant
357 explanatory variable, the effect of TN on the production of MCs could not be ignored. The
358 concentration of MC-YR was negatively correlated with the TN/TP ratio. Previous studies also
359 demonstrated that decreasing the TN/TP ratio concentration could promote the growth and
360 toxicity of Microcystis (Yu et al. 2014, Lei et al. 2015). According to stepwise multiple linear
361 regression, showed TP was found to be the dominant factor affecting the contents of MC-LR and
362 TMCs, and pH and TN/TP ratio as the main factors affecting the content of MC-RR and MC-YR,
363 respectively. These findings were in agreement with the results of correlation analysis.

364 The Monte Carlo simulation model determined the risk levels and putative human exposure

365 scenarios associated with the blooms in Yongjiang river used for drinking. The whole process of
366 security risk assessment was always accompanied by the uncertainty. The entire process of risk
367 assessment was conducted two steps: exposure assessment and hazard characterization. Although
368 the extrapolation of the experimental results did not lead to certainty, such an extrapolation could
369 be carried out from experimental animals to the general population and from the general
370 population to specific populations (sensitive populations). The variations in human individuals
371 involved parameters such as genetics, age, sex, and environmental (nutritional status) and other
372 factors. On the other hand, missing data or limitations led to uncertainties, including NOVEL,
373 time differences, and lack of exposure data. Recent studies have gradually established superior
374 methods, such as benchmark dose (BMD) and chemical-specific adjustment factor (CSAF), to
375 address and reduce the uncertainty in the risk assessment ([Ibelings et al. 2015](#)). The USEPA and
376 Health Canada have gradually started utilizing the BMD and CSAF methods to develop the
377 health guidance values ([Zeller et al. 2017](#)).

378 Many countries that regulate cyanotoxins in drinking water use a parametric value based on
379 the WHO Guidelines for 1 µg/L MC-LR ([Ibelings et al. 2015](#), [Menezes et al. 2017](#)). With respect
380 to the drinking source waters, most countries use guidance values based on cyanobacterial
381 biomass (cell density, chlorophyll-a, biovolume) indirectly reflecting the potential hazardous
382 MCs concentrations ([Valerio et al. 2009](#)). The risk of health issues due to the presence of other
383 cyanotoxins in drinking waters is deduced; however, the adoption of guidance values for
384 Cyanotoxins in water bodies used for drinking is yet to be investigated ([Lee et al. 2017](#)).

385 **5. CONCLUSION**

386 This study analyzed the influencing factors and the health risk assessment of MCs by Monte
387 Carlo simulation method in Yongjiang river, China. The results showed that TP is the dominant
388 factor affecting the contents of MC-LR and TMCs, while PH and TN/TP ratio are the main
389 factors affecting the content of MC-RR and MC-YR, respectively. The health risk assessment
390 results showed that the risk of MC-RR for human health hazards is higher than that of MC-LR
391 and MC-YR, and children are more vulnerable to MCs contamination than the adults. The risk of
392 carcinogenicity of MC-RR to children health under high exposure was greater than the maximum
393 acceptable risk level recommended by USEPA. The annual risk of carcinogenic exposure in
394 adults with MC-RR was greater than the maximum acceptable risk level recommended by the
395 ICRP. The non-carcinogenic risk index for MCs was <1 . Therefore, MCs in the water bodies
396 should be monitored in regards to the carcinogenic risk to human health.

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Figure 1

HPLC chromatogram of MCs.

(A) HPLC chromatogram of MCs standards, the order of the peaks and time of each standard substance were as follows: MC-RR (7.599 min), MC-YR (14.125 min), and MC-LR (17.601 min).

(B) HPLC chromatogram of MCs water samples, the same time represents the same substance, like A.

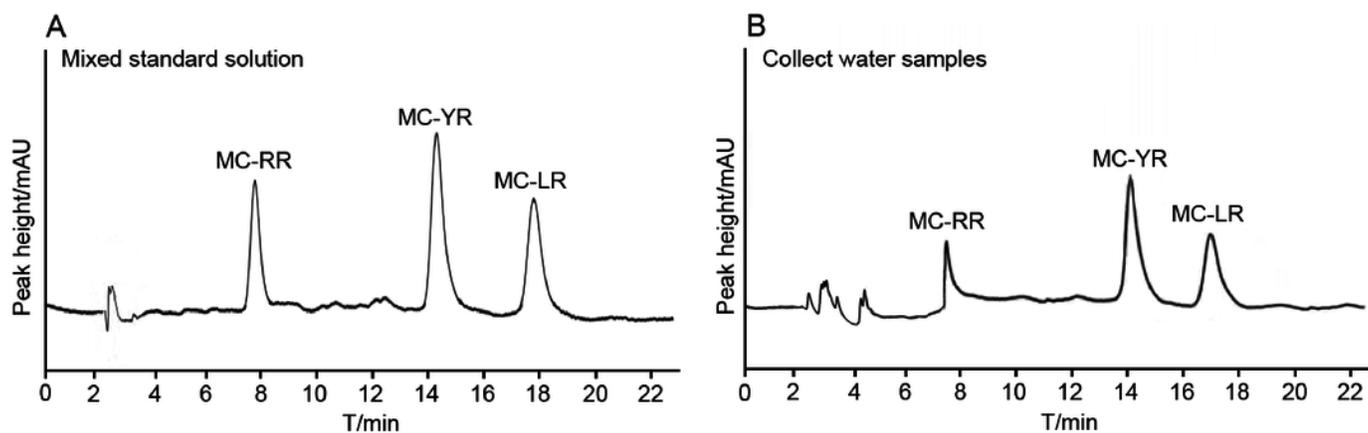


Figure 2

Concentration of MCs in source water in various seasons.

The total content of MCs (TMCs) in water is the sum of the content of extracellular MCs (EMCs) and intracellular MCs (IMCs) dissolved in the water.

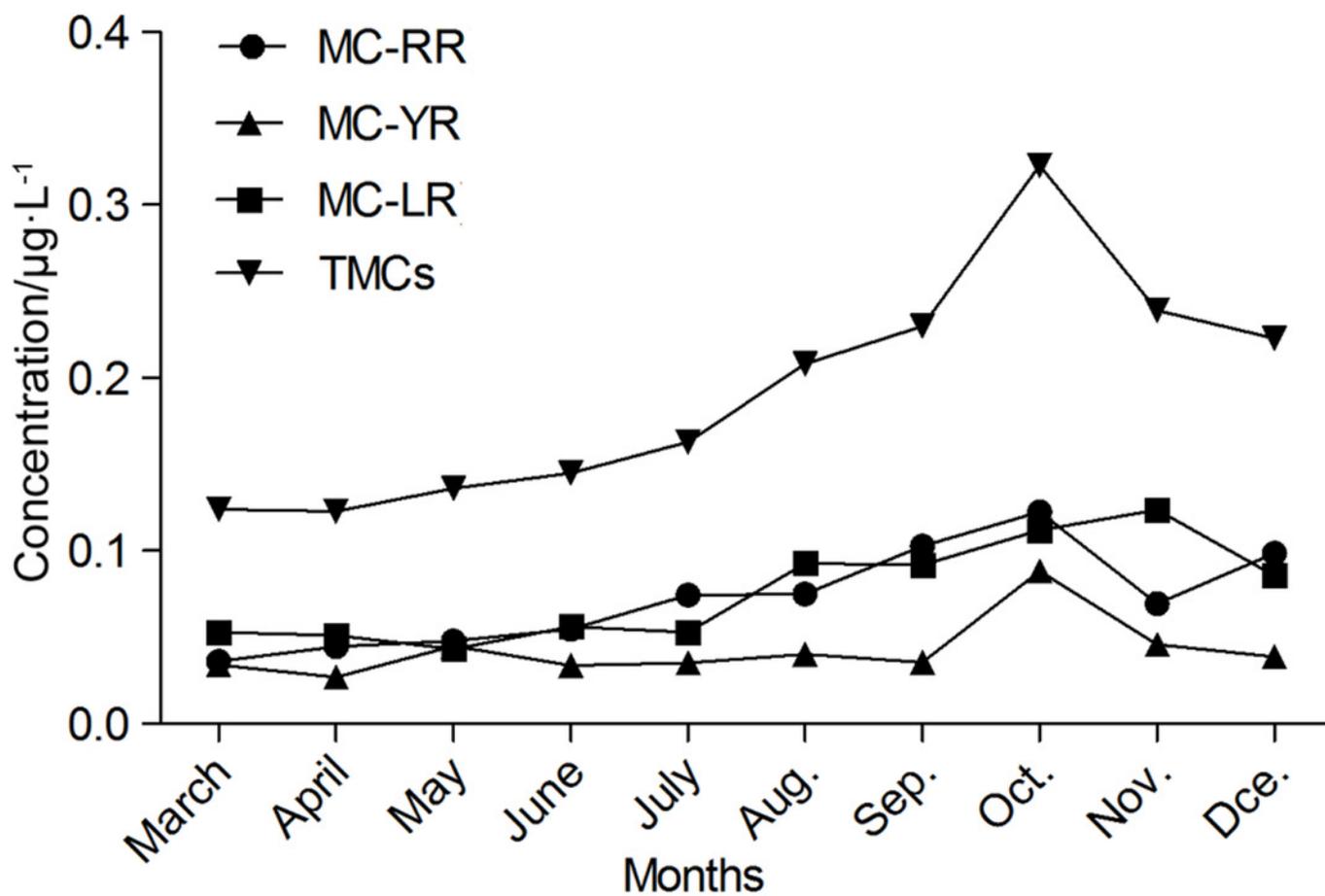


Figure 3

Probability distribution graph after fitting of MC-RR in source water.

The data comparison revealed that the optimal fitting distribution of the most suitable concentration of MC-RR in source water was Gamma (μ 0.073, s 0.061) (first number μ as the position parameter and the second number s as the scale parameter).

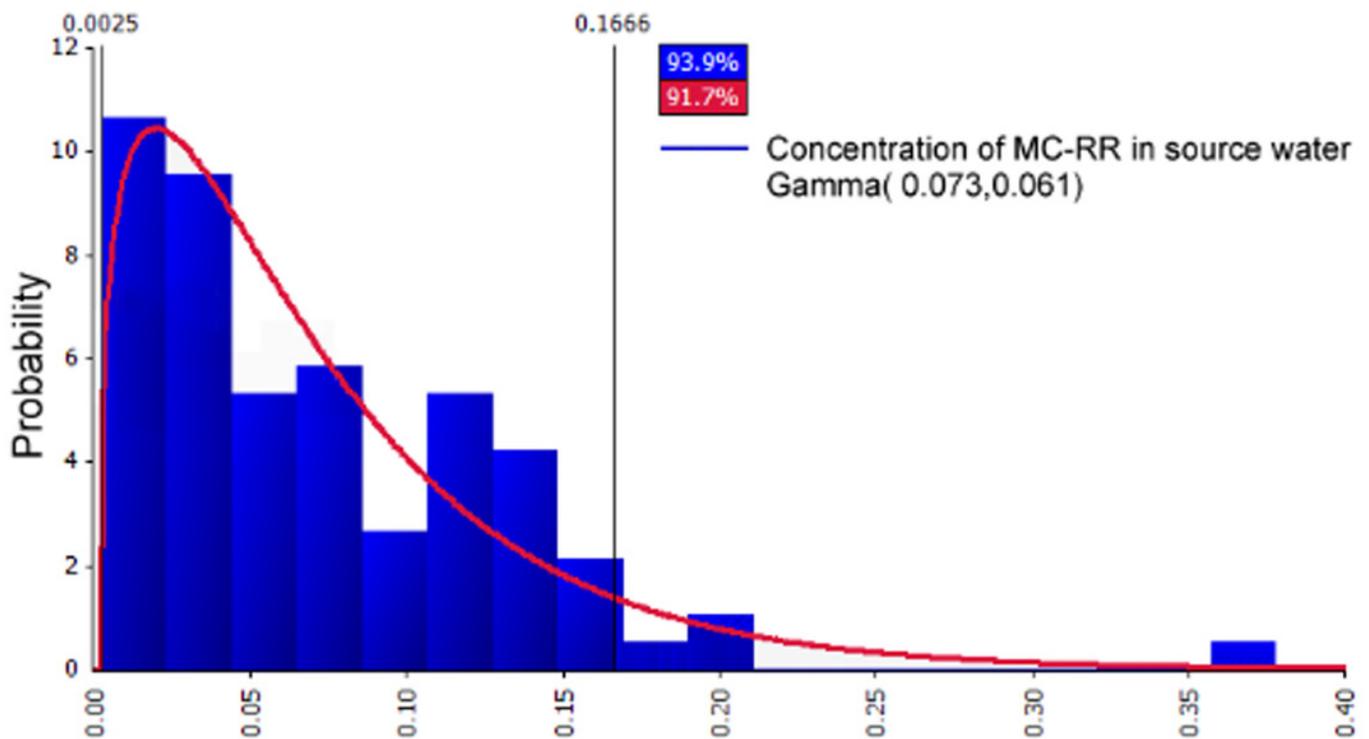


Figure 4

Probability distribution graph after fitting of MC-YR in source water.

The data comparison showed that the best-fitted distribution of MC-YR concentration was Lognorm (μ 0.047, s 0.071).

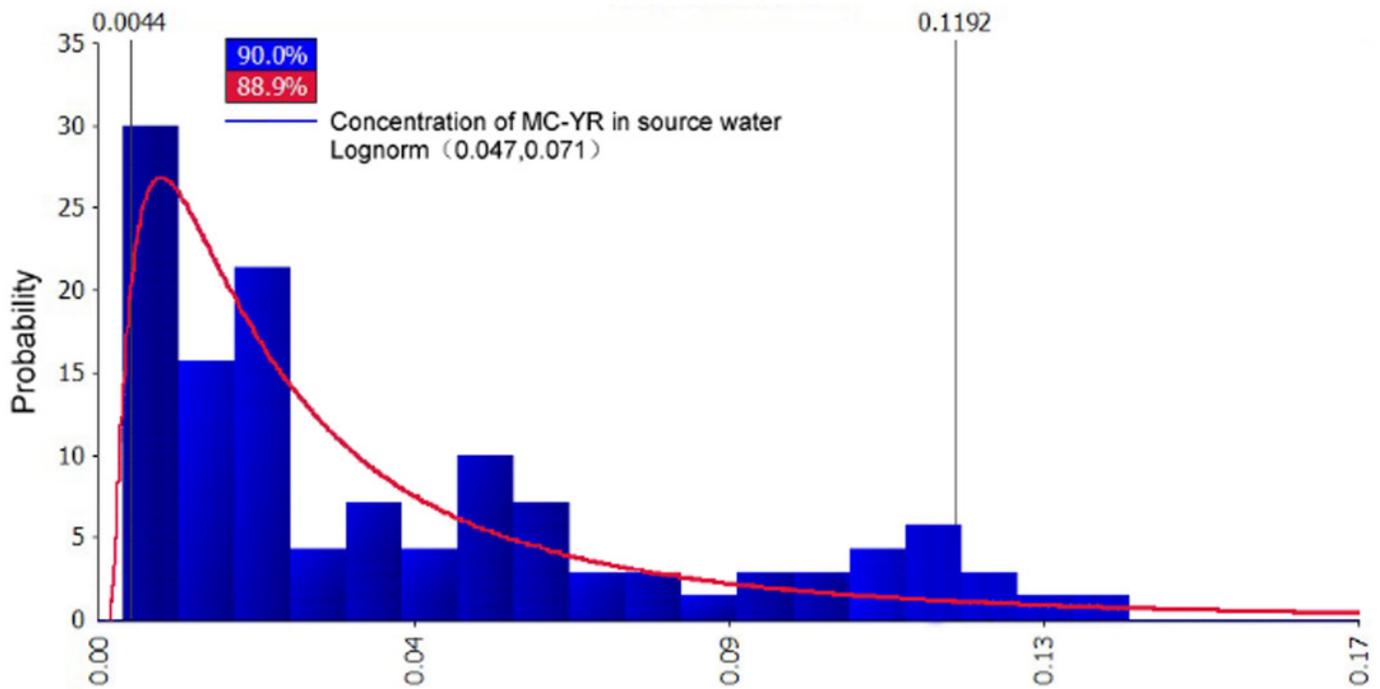


Figure 5

Probability distribution graph after fitting of MC-LR in source water.

The data comparison showed that the best-fitted distribution of MC-LR concentration was Gamma (μ 0.076, s 0.070).

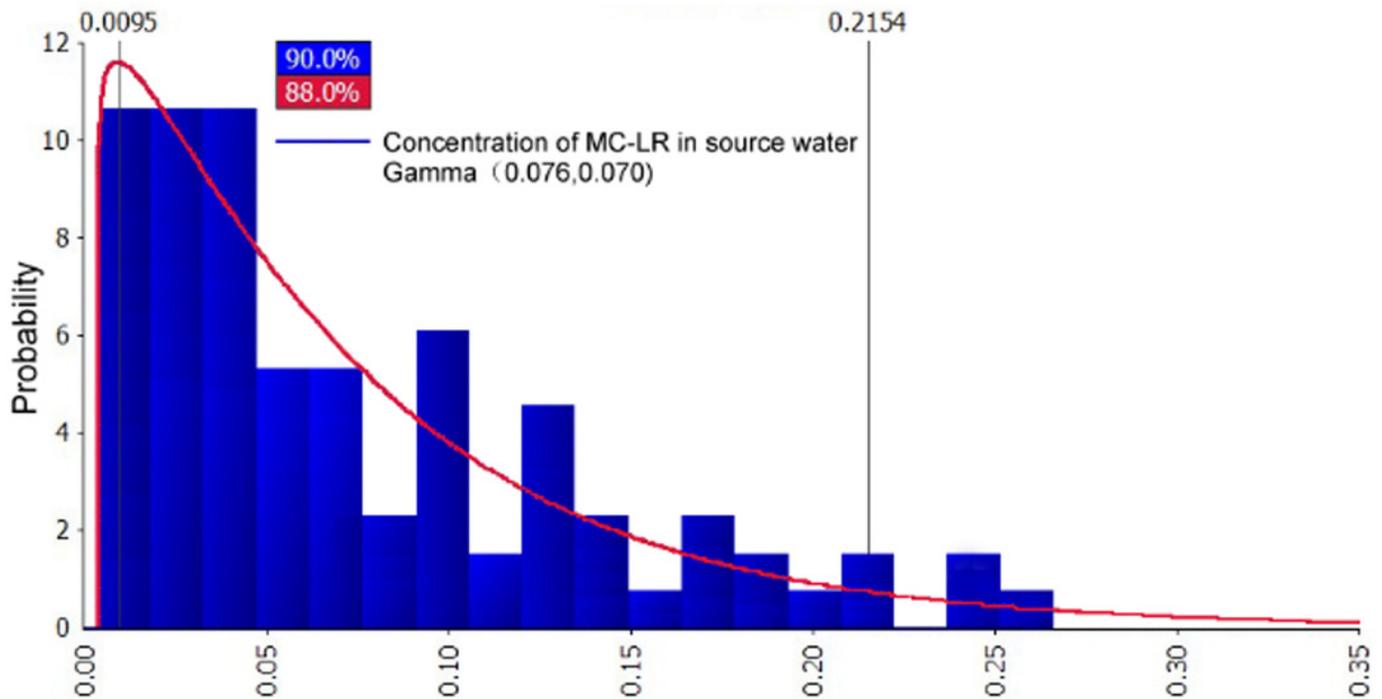


Table 1 (on next page)

Results of analysis of MCs in water samples

Toxin types	Number of samples	EMCs	IMCs	TMCs (EMCs+IMCs) concentration ($\mu\text{g/L}$)	
		Detection Rate/%	Detection Rate/%	Range	Average
		MC-LR	90	77.78	76.67
MC-RR	90	74.44	77.78	0.0224–0.3783	0.0727
MC-YR	90	64.44	56.67	0.0329–0.1433	0.0424

Table 2 (on next page)

Correlation coefficients between MCs and the influencing factors

***Correlation is significant at the 0.05 level (2-tailed).**

****Correlation is significant at the 0.01 level (2-tailed).**

Environmental Factors	Correlation Coefficient			
	MC-RR	MC-YR	MC-LR	TMCs
Water Temperature	0.436**	0.085	0.480**	0.614**
PH	-0.729**	-0.029	-0.164	-0.566**
DO	-0.063	-0.076	-0.768**	-0.570**
TN	0.286**	-0.179	-0.079	0.055
TP	0.043	0.073	0.851**	0.610**
PO ₄ ³⁻ -P	-0.047	-0.071	0.399**	0.204
TN:TP ratio	0.040	-0.229*	-0.434**	-0.347**

Table 3 (on next page)

Results of stepwise multiple linear regression

MCs	Fitting equation	Correlation coefficient R	Adjusted R ²	Tolerance	VIF
	$y=116.848+46.449\chi^3$ (1)	0.581	0.337	1	1
TMCS	$y=805.526+42.979\chi^3-92.886\chi^2$ (2)	0.716	0.513	0.989	1.001
	$y=538.069+35.557\chi^3-75.968\chi^2+6.452\chi$ (3)	0.741	0.549	0.693	1.443
MC-RR	$y=799.874-99.263\chi^2$ (4)	0.718	0.515	1	1
	$y=676.095-91.169\chi^2+3.42\chi$ (5)	0.759	0.576	0.846	1.181
MC-YR	$y=49.805-1.024\chi^7$ (6)	0.246	0.060	1	1
MC-LR	$y=11.216+41.829\chi^3$ (7)	0.806	0.650	1	1
	$y=63.349+32.481\chi^3-6.653\chi^6$ (8)	0.848	0.718	0.678	1.476

Table 4(on next page)

Fitting distribution and related parameters of MCs in source water ($\mu\text{g/L}$)

MCs	Fitting of Distribution	Distributed parameters		Fit test sort			50%	90%	95%
		Mean	Std Dev	K-S	A-D	Chi- Sq	Confiden	Confidence	Confidenc
							ce value	value	e value
MC-RR	Gamma	0.073	0.061	1	1	1	0.056	0.154	0.194
	Invgauss	0.073	0.066	5	2	2	0.053	0.154	0.202
MC-YR	Lognorm	0.047	0.071	4	2	1	0.026	0.103	0.154
	Expon	0.042	0.039	2	1	4	0.030	0.093	0.120
	Loglogistic	0.081	-	1	4	6	0.026	0.124	0.216
MC-LR	Gamma	0.076	0.070	1	1	2	0.055	0.168	0.216
	Expon	0.075	0.073	2	2	1	0.053	0.170	0.221

Table 5 (on next page)

Estimated value of the quantile for overall sample in different theoretical distributions

Projects	Pollutant	Real value($\mu\text{g/L}$)	Types	Predicted value ($\mu\text{g/L}$)	Relative difference of real value (%)
P50	MC-RR	0.058	Gamma	0.056	3.01
			Invgauss	0.053	7.81
			Lognorm	0.026	8.82
	MC-YR	0.024	Expon	0.030	25.17
			Loglogistic	0.026	10.03
	MC-LR	0.057	Gamma	0.055	2.02
Expon			0.053	6.10	
P75	MC-RR	0.011	Gamma	0.099	13.00
			Invgauss	0.095	16.91
			Lognorm	0.053	13.88
	MC-YR	0.062	Expon	0.057	8.17
			Loglogistic	0.056	9.41
	MC-LR	0.112	Gamma	0.104	6.79
Expon			0.104	7.46	
P90	MC-RR	0.136	Gamma	0.154	12.93
			Invgauss	0.154	13.09
			Lognorm	0.103	4.31
	MC-YR	0.108	Expon	0.093	14.10

			Loglogistic	0.124	15.12
	MC-LR	0.166	Gamma	0.168	1.09
			Expon	0.170	2.46
	MC-RR	0.167	Gamma	0.194	16.26
			Invgauss	0.202	21.06
			Lognorm	0.154	29.08
P95	MC-YR	0.119	Expon	0.120	0.58
			Loglogistic	0.216	80.78
	MC-LR	0.215	Gamma	0.216	0.26
			Expon	0.221	2.56
	MC-RR	0.378	Gamma	0.285	24.71
			Invgauss	0.321	15.02
			Lognorm	0.325	127.11
P99	MC-YR	0.143	Expon	0.183	27.56
			Loglogistic	0.735	412.77
	MC-LR	0.266	Gamma	0.326	22.62
			Expon	0.338	27.03

Table 6 (on next page)

Daily exposure to MC intake through drinking water ($\mu\text{g}/\text{kg}/\text{d}$)

Projects	MC-RR		MC-YR		MC-LR	
	Adult	Child	Adult	Child	Adult	Child
Mean	0.002	0.005	0.001	0.003	0.002	0.005
P50	0.002	0.004	0.001	0.001	0.002	0.004
P85	0.004	0.008	0.003	0.006	0.004	0.009
P90	0.004	0.009	0.003	0.007	0.005	0.010
P95	0.005	0.010	0.003	0.007	0.006	0.013

Table 7 (on next page)

Carcinogenic exposure of MCs from source water

Projects	MC-RR		MC-YR		MC-LR	
	Adult	Child	Adult	Child	Adult	Child
Mean	1.27×10^{-5}	1.37×10^{-5}	3.34×10^{-6}	5.76×10^{-6}	5.20×10^{-6}	8.10×10^{-6}
P50	1.40×10^{-5}	1.43×10^{-5}	2.21×10^{-6}	4.40×10^{-6}	4.69×10^{-6}	8.30×10^{-6}
P85	5.43×10^{-5}	1.13×10^{-4}	6.93×10^{-6}	1.09×10^{-5}	9.18×10^{-6}	1.28×10^{-5}
P90	5.47×10^{-5}	1.45×10^{-4}	7.60×10^{-6}	1.16×10^{-5}	9.85×10^{-6}	1.32×10^{-5}
P95	8.63×10^{-5}	1.83×10^{-4}	8.11×10^{-6}	1.20×10^{-5}	1.11×10^{-5}	1.38×10^{-5}

Table 8 (on next page)

Maximal acceptable level and negligible level recommended by different institutions

Institutions	Maximum risk level	Ignore the level of risk	Remarks
USEPA	1×10^{-4}	--	Radiation
ICRP	5×10^{-5}	--	Radiation
Royal Association of England	1×10^{-6}	1×10^{-7}	--
Holland Environmental Protection Agency	1×10^{-6}	1×10^{-8}	Chemical contaminants
Swedish Environmental Protection Agency	1×10^{-6}	--	Chemical contaminants

Table 9 (on next page)

Non-carcinogenic exposure risk of MCs using source water

Projects	MC-RR		MC-YR		MC-LR	
	Adult	Child	Adult	Child	Adult	Child
Mean	0.005190	0.011352	0.030260	0.066190	0.054490	0.119200
P50	0.004123	0.009019	0.017090	0.037380	0.040400	0.088380
P85	0.009157	0.020031	0.067480	0.147610	0.104530	0.228660
P90	0.009722	0.021267	0.077190	0.168840	0.118790	0.259860
P95	0.011903	0.026038	0.085150	0.186270	0.153860	0.336580

1