

Histological analysis of post-eruption tooth wear adaptations, and ontogenetic changes in tooth implantation in the acrodontan squamate *Pogona vitticeps* (#28010)

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Histological analysis of post-eruption tooth wear adaptations, and ontogenetic changes in tooth implantation in the acrodontan squamate *Pogona vitticeps*

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Teeth have been a focus of research in both extinct and extant taxa alike; a significant portion of dental literature is concerned with dental patterning and replacement. Most non-mammalian vertebrates continuously replace their dentition but an anomalous group of squamates has forgone this process in only having one tooth generation; these squamates all have apically implanted teeth, a condition known as acrodonty. Acrodont dentition and various characteristics attributed to it, including a lack of replacement, have often been defined ambiguously. This study explores this type of implantation through histology in the ontogeny of the acrodont agamid *Pogona vitticeps*. The non-replacing teeth of this squamate provides an opportunity to study wear adaptations, maintenance of occlusion in a non-mammalian system, and most importantly post-eruption changes in the tooth bone interface. In this study the post-eruption changes combined with dental wear likely gives the appearance of acrodont implantation.

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Abstract. Teeth have been a focus of research in both extinct and extant taxa alike; a significant portion of dental literature is concerned with dental patterning and replacement. Most non-mammalian vertebrates continuously replace their dentition but an anomalous group of squamates has forgone this process in only having one tooth generation; these squamates all have apically implanted teeth, a condition known as acrodonty. Acrodont dentition and various characteristics attributed to it, including a lack of replacement, have often been defined ambiguously. This study explores this type of implantation through histology in the ontogeny of the acrodont agamid *Pogona vitticeps*. The non-replacing teeth of this squamate provides an opportunity to study wear adaptations, maintenance of occlusion in a non-mammalian system, and most importantly post-eruption changes in the tooth bone interface. In this study the post-eruption changes combined with dental wear likely gives the appearance of acrodont implantation.

Introduction

Reptilian dentition has been extensively studied in both extinct and extant taxa, and for the vast majority of these taxa, there is constant replacement of teeth, a condition known as polyphyodonty. However, in a subset of reptiles, there has been an evolutionary cessation of replacement, a condition known as monophyodonty. Among lepidosaurian reptiles, this suppression of replacement is limited to Sphenodontidae (Rhynchocephalia), Chamaeleonidae, and Agamidae, with the latter two being grouped within the clade Acrodonta (Squamata) (Pyron et al., 2013). This squamate group is aptly named for the acrodont style implantation of the dentition, and all acrodontians have apically implanted teeth making up the majority of their dentulous surface (Jenkins et al., 2017). This is in contrast to the condition found in most squamates and other reptiles (e.g., Zaher and Rippel, 1999; Delgado et al., 2003a; LeBlanc and Reisz, 2015) in which the tooth is implanted to the lingual surface of the jaw bone, a condition known as pleurodonty that is exemplified in taxa like *Iguana iguana* (Throckmorton, 1976; Montanucci, 2008; Kline and Cullum, 2017). The most studied form of implantation among tetrapods is thecodonty, where the tooth is implanted in a deep socket; this form of implantation is found in all mammals and also occurs within crocodilians and in many extinct archosaurs (e.g., Brown et al., 2015; García and Zurriaguz, 2016). Tooth implantation should not be conflated

with tooth attachment, which refers to the tissue that attaches the tooth to the dentulous bone; this study is primarily concerned with implantation, and how ontogenetic change can influence the appearance of implantation categories.

Acrodonty in Reptilia is largely associated with a lack of tooth replacement (e.g., Zaher and Rippel, 1999; Smirina and Ananjeva, 2007), an association that often alludes to causality but that is never stated outright. At first glance it seems understandable that acrodont implantation

and monophyodonty be associated as most acrodont squamates have both acrodont and pleurodont dentition. In individuals with both pleurodont and acrodont dentition, the pleurodont

teeth are replaced by a new generation of teeth, yet the acrodont teeth are not (Cooper et al. 1970), which seems to have cemented the idea that acrodonty somehow inherently disrupts or inhibits replacement. While extant acrodont squamates are monophyodont, it is important to note that acrodonty is also found in some extinct reptiles (Simões et al., 2015; Haridy et al., 2018) and in non-reptile vertebrates such as piranhas (Shellis and Berkovitz, 1976), all of which replace

their dentition. The supposed function of constant replacement is to avoid excessive wear (Throckmorton, 1979; Benton, 1984; Erickson, 1996); accordingly, higher rates of replacement are often seen in herbivorous taxa, as their fibrous diet and frequency of mastication requires a constant renewal of their dentition. In extreme instances, there are examples of starvation in herbivorous mammals that lack continuous replacement and that have worn their dentition to such a degree such that it is no longer functional (Spencer, 2005). Therefore the lack of

replacement in acrodont squamates raises the question: if acrodontians that lack tooth replacement are relatively long-lived (Zari, 1999; Smirina and Ananjeva, 2017), how are they able to maintain a viable occlusal surface during later stages of life? Monophyodonty also poses two mechanical dilemmas: (1) how do acrodontians adapt to dental wear at a tissue level, and (2) how do the mandibular and maxillary teeth stay in occlusion through ontogeny without replacement or a ligamentous attachment? The former of the two questions has been partially addressed through histology of the acrodontians *Uromastix aegyptia* (Throckmorton, 1979) and *Chamaeleo calypttratus* (Buchtová et al., 2013; Dosedělová et al., 2016a), and members of acrodontan Uromastycinae and Chamaeleoninae (Pyron et al., 2013). These studies found that these acrodontians have a peculiar way to combat wear – they secondarily infill the teeth with bone and/or dentine and remodel the underlying jaw bone into more compact bone. There are six other groups within Acrodonta that presumably also lack replacement and have

undocumented wear adaptations (Pyron et al., 2013). This study tests if the agamid *Pogona vitticeps* (the central bearded dragon); conforms at a tissue level to the wear adaptation patterns of other acrodontians previously documented in literature.

To better understand the relationship between acrodonty, monophyodonty, and wear adaptations, the implantation mode itself should be more thoroughly examined, particularly when it comes to acrodonty as it is the least studied form of implantation. Acrodont implantation has been ambiguously defined at best, with various authors describing the mode of implantation as: (1) apically placed teeth (Edmund, 1966) (2) teeth ankylosed to the margin of the jaw (Motani, 1997) (3) teeth fused to the edge of the jaw bone via undefined tissues (Peyer, 1968). Most definitions used acrodontian squamates or rhynchocephalians as representative groups for acrodont implantation, with acrodonty used as a phylogenetic character (e.g., Zaher and Rieppel, 1999); to ground truth the use of acrodontians as acrodont representatives the natural question becomes: are the acrodont teeth of acrodontians really acrodont? It has been previously proposed that ichthyosaurs and alligators can change implantation types through ontogeny by growth in the jaw ramus and additional ossification on the lingual side (Motani, 1997); the alligators go from pleurodont to thecodont, and the ichthyosaurs become subthecodont from their juvenile state of pleurodonty.

This study explores the ontogenetic change in the teeth of *Pogona vitticeps*, with a focus on comparisons of wear adaptations of this member of Agamidinae to those previously described in Uromastycinae and Chamaeleoninae. This study aims to explain: (1) Do tissue level wear adaptations documented in other acrodontians extend to *P. vitticeps*? (2) How do monophyodont reptiles that lack a ligamentous attachment maintain occlusion through ontogeny? (3) Are the teeth of these reptiles truly acrodont, and does acrodonty limit tooth replacement as indicated by literature? This is the first study to document the change in implantation types through ontogeny due to osteological remodeling of the dentary and the tooth body, with evidence of a change from pleurodont implantation to acrodont implantation in a modern squamate. This has implications on how we view implantation categories as they are likely to be more ontogenetically variable than previously thought.

Materials and Methods

In this study, the genus *Pogona* was represented by the central bearded dragon (*P. vitticeps*). Thirty-seven specimens (S. Figure1) of *P. vitticeps* from the Royal Ontario Museum (ROM) herpetology collection were externally examined, and all measurements were taken by K. Seymour prior to skeletonizing. All the *Pogona vitticeps* specimens were photographed using a Canon EOS40D prior to sectioning. To access normally inaccessible anatomical features of the lower jaw and its dentition, 5 specimens were sectioned in both coronal and longitudinal planes. *Pogona vitticeps* was sectioned at two ontogenetic stages, a juvenile stage (ROM R8234, ROM R8510), and an adult stage (ROM R8507, ROM R8189, ROM R9422). The illustrations and diagrams found in the figures were made using Adobe Photoshop CS6 and Adobe Illustrator CS6.

Histology

All thin sections were made following the ROM histology protocol and were executed in the ROM vertebrate paleontology thin sectioning facility. Specimens were embedded in AP Castolite acrylic resin, vacuumed, and left to cure for a minimum of 24 hours. All specimens were cut using a Buhler Isomet 1000 wafer saw at a low speed of 275rpm. The specimens were mounted on plexiglass slides using Scotch-Weld SF-100 cyanoacrylate. The slides were then mounted on the Hillquist grinding cup and ground down using the grinding cup until optical clarity was achieved; subsequently the specimen was manually ground using progressively finer grit suspensions on glass plates, beginning with a 600-grit silicon carbide powder and working down to a 1-micron aluminum oxide powder. All slides were imaged using a Nikon DS-Fi1 camera mounted to a Nikon AZ 100 microscope fitted with crossed-polarizing and lambda filters and an oblique illumination slider and NIS-Elements software registered to R. R. Reisz of the University of Toronto Mississauga.

Results

External anatomy

The external morphology of both the mandible and dentition is noticeably different between the juvenile and adult specimens of *Pogona vitticeps* (Figs. 1A, B). Several specimens

that represented a relatively complete ontogenetic sequence were examined; for the sake of brevity, two representative specimens, an adult (ROM R8507) and a juvenile (ROM R8234), were chosen for external description. The juvenile specimen (ROM R8234) has fewer tooth positions (16 tooth positions) on the dentary; however, some smaller individuals were found to have as few as 10 teeth (S1. Fig.). There are external mandibular features of ROM R8234 that identify it as juvenile: (1) the symphysis is poorly ossified, (2) the mandible is less robust in bone density, which is externally apparent as a character of bone opacity, and in dorsoventral width, and (3) the mandible is relatively short, with the dentulous region making up more than half the total length. The wear facets on the labial surface of the dentary are not as well developed in the juvenile specimen as those seen in the adults. This specimen of *P. vitticeps* is found to have two teeth implanted in a pleurodont fashion; these occupy the first two tooth positions on the rostral end of the mandible (Fig. 1B). The pleurodont teeth are conical and taper to a point; this represents the most common condition among many, but not all agamids (Cooper et al., 1970). Posterior to the pleurodont dentition, the remaining tooth row has been described in literature to be acrodont and is the focus of this study (Cooper et al., 1970). The acrodont teeth in *P. vitticeps* are triangular and mediolaterally compressed and lack the multi-cuspid morphology that other acrodontians (S. Figure 1), like chameleons and *Uromastyx* display. Compared to the older anterior teeth, the teeth are larger in size posteriorly correlating with age, with the youngest teeth added posteriorly being largest. It appears that the newest tooth is not ankylosed to the jaw bone prior to the development of the next tooth in the series. Instead, the newest tooth appears be attached to the jaw only by soft tissue; a desiccated fibrous tissue separates it from the bone in skeletonized material, (Fig. 2); this is readily recognizable by external observation, as the last two teeth are not oriented at an angle that is congruent with the rest of the dentition (Fig. 1A). This was also observed in several other juvenile specimens in which it is observed only in the posteriormost teeth. Lastly, the newest and last tooth position is growing directly against, and partially resorbing (Fig. 1A), the coronoid process, which does not allow sufficient space for the next more posterior tooth position.

As with the juveniles, several adult individuals were examined, and a representative specimen chosen. Several changes correlated with size and thus presumably with ontogeny can be recognized in the adult specimen of *Pogona vitticeps* (ROM R8507) including an increase in the number of tooth positions. The specimen chosen for sectioning has 17 tooth positions,

although other specimens have been found to have as many as 19 tooth positions; in most specimens a definite tooth count was difficult to ascertain via examination of the external morphology due to extensive wear but was later verified in thin section. Wear on the adult mandible is evident on both the acrodont dentition and the jaw bone. Interestingly, the anterior pleurodont dentition was mostly unworn in the adult specimen or worn to a minimal degree on other adult specimens examined; this is similar to the condition seen in *Agama agama*, which has been documented to replace its anterior pleurodont dentition (Cooper et al. 1970). However, the anterior acrodont teeth are all but worn away, making it difficult to differentiate it from the jaw bone; to circumvent this problem, tooth counts were made under a microscope and later confirmed in thin section when possible. The wear observed on the anterior acrodont dentition was extensive – often there were only traces of the tooth left – which is similar to the dental wear seen in *Uromastix*, a taxon can become functionally edentulous in adulthood (Throckmorton, 1979).

The evidence of extensive wear is not only seen on the teeth but is also found on the mandible itself in the form of wear facets, which are only present on the labial side of the dentary where the maxillary dentition occludes, interdigitating between dentary tooth positions. Wear facets have been characteristically found in acrodont squamates as far back as the Cretaceous (Simões et al., 2015), which are formed due to the maxillary dentition wearing down on the dentary bone during mastication and passive occlusion. These wear facets are present along the posterior two-thirds of the tooth row but are most developed in the posterior region of the dentary of adult specimens of *P. vitticeps*. However, in adult individuals of other acrodontians, like chameleons (Dosedřlová et al., 2016a), the wear facets are found along the entire tooth row. Lastly, it is important to mention that the jaw ontogenetically increases posteriorly in length and dorsoventrally in width, with the articular increasing in robustness, and the coronoid process moving posteriorly relative to the tooth row. This ontogenetic change effectively creates more space for the posterior addition of teeth to the dentary, a feature that is frequently seen in lepidosaurs (Berkovitz and Shellis, 2017).

Histology

Juvenile tissue histology

Longitudinal and coronal sections of the mandible of the juvenile specimen (ROM R8234) were examined (Fig. 2&3). The mandibular ramus in the juvenile specimen is not as well ossified as in the adult (ROM R8507) and has a medial curvature anteriorly; this made obtaining sections of the anterior and posterior dentition within the same sectioning plane difficult. Furthermore, the focus of this study is the non-replacing acrodont dentition, and  therefore, the anterior pleurodont dentition were not included in this study. In longitudinal section, only the dentine is exposed with no other dental tissues (e.g., cementum, enamel) being visible. In longitudinal section, the implantation relationship of the teeth to the jawbone is not easily discernable. However, this plane of section allows for visualization of the incremental maturation of dentition. The most anterior teeth are the oldest, a characterization based on the thickness of dentine, whilst the most posterior teeth are the youngest and have the least amount of dentine infilling. In longitudinal section, the interaction between the individual tooth and its neighboring teeth can be seen in this plane (Fig. 2B-D); there is distinct tissue that attaches the teeth to each other as well as to the jaw bone. This tissue is tentatively identified as alveolar bone (*sensu* LeBlanc and Reisz, 2015) based on the position and presumed function of the tissue rather than on its histological appearance, which does not completely conform to alveolar bone description in literature. The alveolar bone in this specimen has a woven appearance that is less organized than the bone that makes up the dentary. However, it lacks the extensive trabecular or ‘spongy’ appearance that is found in other squamates, such as snakes (Budney, 2004; Budney et al., 2006). This tissue attaches the teeth to each other, as well as to the jaw bone proper, a function that has been attributed to ‘bone of attachment’ (Ananjeva and Smirina, 2007), better known as alveolar bone (Budney et al., 2006; Caldwell, 2007; LeBlanc and Reisz, 2013, 2015; LeBlanc et al., 2016a). The alveolar bone in *P. vitticeps* is also not distinguished from the jaw bone by a reversal line; this is possibly an effect of the plane of section or due to a lack of resorption prior to attachment (Fig. 2). It is important to note that not all the teeth in the juvenile specimen are fully ankylosed to the dentary (Fig. 3); however, each tooth seems to ossify to the adjacent dentition, which occurs prior to full ankylosis and which mirrors what has been documented in *Chamaeleo calypttratus* (Buchtová et al., 2013; Dosedělová et al., 2016a). This observation is based on the youngest tooth (Fig.3), which has yet to ankylose to the dentary, which is attached to the neighboring teeth.

In coronal section, all of the common dental tissues are identifiable, with dentine comprising the bulk of the tooth identified by its characteristic radiating dentinal tubules. The enamel is best visualized in cross-polarized light and is unworn in the youngest teeth (Fig.3B). Because of the mediolateral compression of the tooth, the coronal plane of section is the best plane in which to examine the enamel as it transects the labial and lingual surfaces of the tooth this. The enamel is fairly thick, which has also been reported in *Uromastyx* (Throckmorton, 1979). Due to the lack of wear in the juvenile specimen, the enamel is of equal thickness on the lingual and labial sides of the tooth crown (Fig.3B). On the lingual side of the tooth, the tapering edge of the enamel leads down to a layer of acellular cementum, meeting at the cemento-enamel junction, which defines the boundary between the anatomical crown and root of the tooth. It is likely that the cementum cannot be visualized on the labial side because the alveolar bone has grown to meet the cementum, effectively ankylosing the tooth while obstructing the cementum, or possibly because of resorption of the cementum in this process, although the latter has not been previously documented.

In the coronal section of the juvenile specimen of *P. vitticeps*, it is evident that both the shape of the dentition and the manner in which the tooth is implanted are more congruent with the pleurodont condition, as the lingual side of the tooth is markedly greater in length than that of the labial side (Fig. 3A, C). Although both ends of the tooth are ankylosed to the jaw, it is clear that there is more contact with the jaw bone and more attachment tissues on the labial side, a character of pleurodont implantation. Apart from the obvious labial bias of attachment, alveolar bone can be seen at the base of the tooth, being more woven than the rest of the jaw bone yet apparently lacking the more porous structure that is usually associated with alveolar bone. In coronal section (Fig. 3B), the reversal line defining the boundary between the new alveolar bone and the preexisting jaw bone is visible in cross-polarized light. The labial side of the tooth (Fig. 3C) is shorter and is attached to the jaw bone with more attachment tissue than that present at lingual side; furthermore, the labial side appears to be actively remodeled by osteoblasts and osteoclasts (Fig. 3D). The osteoclast on the labial side of the tooth is identified on the basis of its large size, general shape and the resorption bay that is created in the dentine (Witten and Huyseune, 2009). The high density of bone cell lacunae directly posterior to the osteoclast are identified as osteocytes, which were osteoblasts, indicating that the labial side of the tooth is being resorbed and that bone is being deposited in its place. It is also important to note that the

strut like structure of the cancellous bone likely give the mandible form and function prior to full ossification and maturity.

Lastly, a coronal section through the coronoid process (Fig. 3E), shows the presence of a developing tooth. Its identification as a developing tooth is based on its general shape, as well as the presence of thick enamel relative to the amount of dentine. The presence of more enamel than dentine denotes an early stage of tooth development, as dentine is deposited by odontoblasts later in development (Erickson, 1996; LeBlanc et al., 2016b). The presence of this developing tooth was serendipitously discovered by sectioning the specimen since there were no external indicators that this tooth was buried within the bone of the coronoid process. This new tooth is developing within the coronoid process and is visibly resorbing the bone tissues of both the coronoid and the dentary, effectively making space for itself prior to attachment.

Adult tissue histology

In the longitudinal and coronal sections of the mandible of the adult specimen (ROM R8057), lamellar bone makes up the main body of the dentary (Fig. 3). In coronal section, it can be seen that the large trabeculae seen in the juvenile have been incrementally infilled by lamellar bone, giving the mandible an osteological density that was not present in early ontogeny (Fig 3). As previously reported in agamids (Ananjeva and Smirina, 2007), the bone tissue is not highly vascularized nor actively remodeled, particularly in dentary bone. This is in line with the findings in the adult jaw, where no reversal lines or large areas of remodeling are found. No primary or secondary osteons can be identified in the longitudinal sections. However, in one of the coronal sections, a single osteon could be identified ventral to the tooth implantation site (Fig. 5A). In longitudinal section (Fig. 4C), posterior to the most worn dentition, the wear facets become a marked feature of the dentary's labial surface. The pervasiveness and the depth of the wear facets should not be considered informative in thin section, as the variation of depth is a false impression, and is due to the sectioning plane in combination with the curvature of the dentary. The wear facets form pseudo-pedicels for the tooth remnants; these should not be confused for the 'bony pedicles' that have been identified developmentally in chameleons (Buchtová et al., 2013). These pseudo-pedicels are formed by the wear facets, as the maxillary teeth wear away lamellar bone between the functional dentition on the jaw, leaving the

remaining tooth caps on secondarily formed pedicels of lamellar bone (Fig. 4C-D). This gives the tooth implantation region its distinctly acrodont appearance.

The teeth in the adult specimen, specifically concerning the acrodont teeth, are markedly different than those described in the juvenile (Fig. 5). The unequal shape and implantation of the teeth reported in the juvenile is not recognized in the adult, which has acrodont implantation with no lingual bias (Fig 5B). The anterior pleurodont dentition is shown to maintain its vasculature, which is associated with an open pulp cavity; this is typical of pleurodont dentition among squamates. Perhaps the most interesting feature of the longitudinal sections is seen in the worn acrodont teeth directly posterior to the pleurodont pair (Fig. 4). The teeth are worn to such an extent that often no dentine or enamel is detectable (Fig. 4B), even in thin section. However, the vasculature is maintained and denotes tooth positions, and the pulp cavity remains open and vascularized throughout the functional life of the tooth. Thus, the teeth appear to remain viable into adulthood (Fig. 4E), which is in contrast to the condition reported in *Chameleo* (Dosedřlová et al., 2016a) and in *Uromastyx* (Throckmorton, 1979). The vasculature is best seen in Figures 4 and 5.

Considering the high degree of post-ankylosis changes to the tooth-bone interface, it is important to characterize the tissues involved. In the adult specimen (Fig. 5B), the tissues between the tooth and the platform of lamellar bone are particularly interesting when viewed in cross-polarized light, as they are clearly distinct from both the tooth and the lamellar bone that makes up the jaw. The presence of these tissues alone provides evidence of remodeling. In this case, ‘remodeling’ is used in the broad sense rather than to refer strictly to bone remodeling. For example, in this agamid, the dentine that was once present in the lingual side of the tooth of the juvenile has been resorbed and is no longer present, or it is present in a greatly reduced extent in the adult. In both instances, it has been replaced by woven bone. The woven bone can be seen at the base of the teeth in longitudinal section (Fig. 2). However, the true extent of remodeling and change in tooth morphology is best seen in cross-polarized light in coronal section as this plane allows for the best direct comparison to the juvenile teeth in figure 4. Positioned between the dentine and the organized lamellar bone of the jaw is a layer of bone with disorganized fibers previously referred to as woven bone. The lack of organization in this bone indicates the occurrence of relatively fast deposition and is unlikely to be the original alveolar bone that was

identified in the juvenile specimen. The woven appearance under cross-polarized light identifies this as woven bone. In conjunction with its position relative to the teeth, it is tentatively identified here as a remodeling zone (*sensu* Budney, 2004) (Fig. 4C, 5B). In coronal section (Fig. 5B) the remodeling zone is clearly identifiable in cross-polarized light, and the distribution of dental tissues and bone is markedly different than that previously described in the juvenile specimen. In the adult specimen, the anterior acrodont dentition was either completely worn away, or all that remained was a dentine fragment (Fig. 4). The most posterior dentition retained more comparatively more dentine and often with a lingual bias (Fig. 5A), but this was only discernable in thin section. The anterior acrodont teeth in the adult had no evidence of enamel, while the posterior most teeth were the only ones that maintained some enamel, although it is only present on the lingual side. Within the pulp cavity of the adult, there often remains evidence of vasculature (Fig. 4) although it is greatly reduced to the center of the tooth. The overall shape of the tooth is markedly different to that reported in the juvenile, and the external appearance is acrodont like that described in literature (e.g., Budney, 2004).

Discussion

Tooth replacement has been a topic of great interest in recent years in groups ranging from fish to ~~mammals~~ and across broad time scales. There have been several studies documenting the presence of replacement patterns in relation to dental development (e.g., Westergaard and Ferguson, 1990; Richman and Handrigan, 2011; LeBlanc and Reisz, 2015). However, there has been relatively little work on the anomalous squamates that completely cease tooth replacement, the acrodontians. The clade Acrodonta (Cope, 1864) is aptly named for the form of tooth implantation known as acrodonty that this group possesses, namely where the teeth are attached to the apex of the jaw. Grouping based on tooth implantation has divided squamates between acrodontians and pleurodontans (Cope, 1864; Budney, 2004), the latter of which attach their dentition to the lingual margin of the jaw (pleurodont implantation). The classic categories of acrodonty, pleurodonty, and thecodonty have been used as descriptors, as well as phylogenetic characters (Zaher and Rippel, 1999), and continue to be ~~descriptive~~ ~~in~~ ~~even~~ the latest literature (Jenkins et al., 2017) but have been called into question by Estes et al. (1988) who suggested these categories are artificial and are not likely representative of natural groupings.

True teeth in most non-mammalian vertebrates are well known for their extensive replacement patterns through their life (Zaher and Rieppel, 1999; Delgado et al., 2003b; LeBlanc and Reisz, 2015; LeBlanc et al., 2016b); **therefor** any condition that deviates from polyphyodonty is unusual and worthy of study. An important example of non-polyphyodont dentition is seen in mammals, in which there are only two generations of dentition, the deciduous teeth and the permanent teeth. Some mammals such as shrews have even forgone the deciduous phase by resorbing the dentition prior to eruption, functionally giving them one tooth generation (Järvinen et al., 2009). Other than mammals, most fish (but see ratfish, e.g., Huber et al., 2008), amphibians, reptiles and non-mammalian synapsids have continuous tooth replacement through life, making acrodontian squamates an anomaly amongst toothed vertebrates. The lack of replacement comes with a set of challenges, two of which are addressed here. The first challenge is how ~~does an organism grow the jaw~~ whilst having permanently ankylosed dentition and whilst maintaining occlusion with the maxillary dentition? The second challenge is how to maintain a single set of functional teeth through the lifetime of an animal, which essentially is a problem of combating or adapting to wear. These two obstacles are also faced by mammals, which have one primary set of teeth throughout most of their life. In mammals, the issue of maintaining occlusion appears to be solved by maintaining a ligamentous tooth attachment, which allows for the teeth to remain mobile as the mandible grows and remodels (Lumsden and Osborn, 1977; LeBlanc and Reisz, 2013). The issue of wear is at least partially addressed by having much thicker prismatic enamel than that found in most reptiles (Dauphin and Williams, 2008; Kieser et al., 2009). However, the question remains how reptiles that permanently ankylose their dentition to the jaw and those with reptilian enamel adapt to growth and wear.

Wear adaptations

This study found that the unworn teeth on the mandible of the juvenile agamid *P. vitticeps* have a uniform layer of enamel on both the lingual and labial sides, and the enamel is relatively thick in comparison to polyphyodont reptiles such as crocodilians. The thick enamel also seems to occur in two other acrodontians. *Uromastyx* enamel has been documented as ‘thickened’ (Cooper and Poole, 1973; Throckmorton, 1979), and chameleons’ enamel appears thick in recent studies but is not explicitly commented on (Buchtová et al., 2013; Dosedělová et al., 2016b). Furthermore, *Uromastyx*, a herbivorous acrodontian, has been reported to have

thickened prismatic enamel (Throckmorton, 1979), similar to mammalian enamel. This shows that thickening of the enamel is a convergent adaption against wear in both squamates and in mammals.

Enamel thickening is not the only adaptation that acrodontians appear to have evolved in order to combat wear. Previous studies of chameleons and *Uromastyx* have shown that their pulp cavities were infilled with ‘mineralized tissue’ (Dosedělová et al., 2016b) or ‘bone’ (Throckmorton, 1979). The likely purpose of this infilling is to prevent the pulp cavity from being exposed as the external surface of the tooth is worn away; both aforementioned studies also reported the disappearance or significant restriction of vasculature that initially supplied the dentition in early ontogeny. Dosedělová et al. (2016a) also showed increased mineralization in the bone underlying the tooth-bone junction in chameleons, and Throckmorton (1979) found that in *Uromastyx*, the bone below the teeth had become more compact in appearance rather than cancellous. These findings are quite comparable with one another, even though *Uromastyx* and chameleons are on two disparate branches of Acrodonta (Pyron et al., 2013). These results then beg the question: is this (?) pattern more widespread across acrodontians?

This study found comparable results in *Pogona vitticeps*. The pulp cavity is also greatly diminished through ontogeny (Fig. 2B) but is not completely ‘obliterated,’ as seen in *Uromastyx*. The infilling of the pulp cavity in *P. vitticeps* is done through a combination of dentine, and the bone. The progressive infilling of dentine is a normal development for teeth, as odontoblasts continue to sequentially deposit dentine post- eruption. What is abnormal is the bone growing into the pulp cavity and the resorption of the dentine root. In some of the sampled sections of *P. vitticeps*, the teeth fail to show the extent of bone infilling reported in *Uromastyx*, as the matrix that infills the pulp cavity does not seem to have any of the cellular spaces reported in *Uromastyx*. However, in other sections of the same individuals, the teeth experience a lot of remodeling and infilling, often with bone growing over the dentition. This is identified as ‘bone’ rather than ‘mineralized material’ as mentioned by (Dosedělová et al. 2016b) based on the amount of osteocyte lacunae present as well as the presence of osteons. Together the bone and dentine restrict the pulp cavity to such an extent that the tooth can then be worn away without the risk of exposing the vulnerable vasculature and nerves that are within the pulp cavity. Furthermore, the bone making up the dentary is not vascularized and sequentially also becomes

more infilled with bone, making compact bone through ontogeny; this allows for the jaw to be worn in the form of distinct wear facets.

The amount of wear on the adult dentition of *Pogona vitticeps* changes the external morphology significantly. This is common in ungulate mammals (Fortelius and Solounias, 2000) and in herbivorous fossil reptiles (e.g., Reisz, 2006) but has not been reported in other squamates. The dentine and enamel on the labial side of the mandible are worn in the older specimens of *P. vitticeps*, which is compatible with the findings in *Uromastyx* (Throckmorton, 1979), where depending on the tooth position, the enamel and dentine were either completely worn away or were worn to such a degree that the lingual side of the tooth retained a much thicker layer of these two tissues. This pattern is likely caused by the combination of extensive wear from feeding and passive occlusion with the maxillary dentition, as well as the complete lack of tooth replacement. Interestingly, the vasculature that leads to the pulp cavities is still present in the adult specimens of *Pogona vitticeps* indicating that even the most worn teeth probably remained viable through the heavy wear process and likely closed the pulp cavity progressively until they were worn away. This phenomena has also been observed in hadrosaurs, which progressively infill their dentition with dentine as it nears the occlusal surface in their dental batteries and then wear away the entire tooth (LeBlanc et al., 2016b; Bramble et al., 2017).

Maintaining occlusion

Continuous growth of the mandible throughout ontogeny is concurrent with the increased size of the skull. In most polyphyodont taxa, this is not problematic, as constant tooth replacement adjusts for increase in tooth size as well as possible migration (e.g., Haridy et al. 2018); even in mammals, after the animal acquire their permanent set of dentition, a maintained ligamentous attachment of teeth allows for migration and remodeling. However, for acrodont squamates that have ankylosed monophyodont dentition, this becomes problematic – the question becomes how occlusion is maintained as the jaw grows. Through examination of 37 specimens and histological sections, this study of *P. vitticeps* has shown that the growth of the jaw and the initiation of additional tooth development are decoupled processes. This is evident in the juvenile specimen in which the dentition is growing into the coronoid process (Fig. 3E, F),

where the youngest un-erupted tooth is resorbing the ventral portion of the coronoid process in order to continue developing. This indicates that in the early stages of ontogeny, tooth development likely happens at a rate faster than dentary growth. This is reinforced by comparing the juvenile specimen (103mm SV length), which has 16 tooth positions, with the adult, which is more than twice as long (222 mm SV length), only has 17 tooth positions. This supports the hypothesis that attaining the maximum number of teeth occurs relatively early in ontogeny, well before attaining maximum adult size, indicating that tooth development slows and likely stops early in ontogeny, while the various jaw elements continue to grow. The mandible of *P. vitticeps* undergoes many changes through ontogeny, but there is little distortion of the tooth row during growth. This is likely achieved by appositional bone growth of the jaw, with deposition of parallel-fibered bone both internally and externally to achieve the increase in internal ossification, width and length that is seen in the adult and without remodeling or migration of the ankylosed dentition. Through the histological results of this study, which show little bone remodeling, and external observation of 37 specimens (S1), it is hypothesized that occlusion is maintained through the allometric growth of the various portions of the jaw elements. The dentary and its corresponding dentition are the first to reach adult size, while the other jaw elements continue to grow through ontogeny, gradually increasing in robustness but not interfering with occlusion.

Acrodont implantation as a result of ontogenetic remodeling

Acrodont implantation is defined ambiguously in literature. The mode of implantation has been defined as attachment to the ‘edge’ of a jaw (Peyer, 1968), and teeth ankylosed to the ‘apex’ of the jaw by cement (Edmund, 1969), or as the fusion of dentition to the ‘margin’ of the jaw (Motani, 1997). Eventually, the lack of replacement also became a character of acrodont tooth implantation (Zaher and Rippel, 1999), as the condition is present in acrodont squamates. Implantation categories are pervasive in literature and are often used as phylogenetic characters, but these definitions are problematic and tend to imply an evolutionary progression of tooth implantation from simple to more complex (Budney et al., 2006). In reality, the traditional categories of acrodonty, pleurodonty, and thecodonty are, at best, descriptive terms (Estes and Charles, 1988) (Fig. 6A). *Pogona vitticeps* and likely other agamids change implantation categories through ontogeny, and therefore, they are a good representation as to why implantation

categories should only be used descriptively. The teeth in the juvenile *P. vitticeps* are unequal in shape, with the lingual side being much longer than that of the labial side (Fig. 3), and attachment mostly occurring on the labial side, which are characteristics of pleurodont implantation. Traditionally in literature, the anatomy present in adult individuals of various taxa is described; this likely explains why acrodont squamates got their namesake since in their adult stage, the teeth do appear acrodont. This study reveals how implantation may appear to be acrodont in adulthood through a combination of factors (Fig 7); (1) the gradual infilling of teeth with dentine that allows for progressive wear (2) resorption of the dentine by odontoclasts (Fig3C,D; Fig 6-2), (3) growth on the dentary that infills part of the pulp cavity with woven bone and that reduces the trabecular structure of the dentary bone, (4) the continued remodeling of the dentary which removes traces of buried dentine and the previous pleurodont dentition. Tooth wear then further changes the tooth morphology (5-7) the maxillary dentition occludes with the mandibular dentition wearing away the labial side of the mandibular dentition and wearing away part of the dentary creating wear facets. This progression model shows how teeth that were originally pleurodont in implantation (Fig 6-1) can appear acrodont (Fig 6-7) and have an entirely different morphology due to a combination of dentine infilling, bone remodeling, and tooth wear. It is difficult to ascertain if these secondary remodeling steps would occur in any pleurodont squamate taxa but are suppressed by tooth replacement, or if these are secondarily acquired adaptations in monophyodont taxa to combat wear. Therefore, at least in the agamid *P. vitticeps*, the dentition changes from pleurodont to acrodont in appearance through ontogeny. This indicates that we should not use implantation categories as phylogenetic characters if the ontogenetic stage of the specimen is unknown, as is the case in many fossil taxa.

Conclusion

The dentition of acrodontians has recently been an area of interest in the context of dental development and implantation, as this group is known for its mode of implantation and monophyodonty. The latter is an anomalous occurrence among tetrapods, as intermittent to continuous replacement of teeth is the primitive condition for toothed vertebrates, including most extant fish, amphibians, and amniotes. This lack of replacement has become associated with acrodonty, resulting in monophyodonty becoming a formal characteristic of acrodont implantation in squamates, and previous authors have implied that acrodont implantation is the

cause of monophyodonty. However, this study shows that modern acrodont reptiles do not initially have acrodont implantation early in ontogeny, and there is a distinct ontogenetic change in the morphology and implantation of dentition of the agamid *Pogona vitticeps*. The youngest teeth in juvenile specimens are pleurodont in implantation, with a greater lingual contribution of tooth tissues and an attachment biased towards the labial side. These tissues are secondarily remodeled through a step-wise process of: (1) resorption of dentine, (2) deposition of bone and dentine, (3) wear of the tooth surface, and (4) wear of the jaw bone proper, effectively changing the morphology. These processes give *P. vitticeps* the appearance of acrodont dentition in adulthood. This is an important distinction to make as it signifies that this acrodont squamate, and likely other acrodont reptiles, do not develop acrodont teeth, but rather develop pleurodont teeth like the vast majority of squamates, making their teeth inherently pleurodont, and secondarily acrodont. **Therefore**, the wear adaptations, remodeling, and dental wear all come together to give the appearance of an acrodont mode of implantation. The wear adaptations documented in this study likely evolved due to the lack of replacement. Essentially the monophyodont condition causes the acrodont condition, rather than acrodonty causing monophyodonty, as has been implied in literature.

This raises the question as to how we should code acrodont dentition in squamate phylogenies; is it true acrodonty if this implantation mode is only achieved through secondary remodeling of the teeth and dentary? Finally, another important consideration, and a direction of future studies, is that if all acrodontians share these ontogenetic changes and wear adaptations, then what is the significance of the convergence seen between acrodontian squamates and the ~~other~~ rhynchocephalian *Sphenodon*, which is also reported to have acrodont dentition?

Acknowledgments

Incredible thanks go to Robert R. Reisz and Aaron H. Leblanc who brought this issue to light and encouraged the development of this study. I would like to thank Diane Scott for photographs and SEM images and beaded dragon husbandry advice. Thanks to Bryan Gee for his everlasting patience and editing of this manuscript into presentable form. I would like to thank Kevin Seymour of the Royal Ontario museum (ROM) for assistance in locating appropriate specimens for this study and to thank David Evans for allowing the use of the ROM histology

laboratory and reading early versions of this manuscript. Lastly, I would like to acknowledge
Osiris the bearded dragon whose presence in the lab was essential to the direction of this study.

References

Ananjeva, N., and E. Smirina. 2007. Growth layers in bones and acrodont teeth of the agamid
lizard *Laudakia stoliczka* (Blanford, 1875) (Agamidae, Sauria). *Amphibia-Reptilia*
28:193–204.

Berkovitz BK, Shellis RP. The teeth of non-mammalian vertebrates. Academic Press; 2016 Oct
14.

Bramble, K., A. R. H. LeBlanc, D. O. Lamoureux, M. Wosik, and P. J. Currie. 2017.
Histological evidence for a dynamic dental battery in hadrosaurid dinosaurs. *Scientific*
Reports 7:15787.

Brown, C. M., C. S. Vanburen, D. W. Larson, K. S. Brink, N. E. Campione, M. J. Vavrek, and
D. C. Evans. 2015. Tooth counts through growth in diapsid reptiles: Implications for
interpreting individual and size-related variation in the fossil record. *Journal of Anatomy*
226:322–333.

Buchtová, M., O. Zahradníček, S. Balková, and A. S. Tucker. 2013. Odontogenesis in the Veiled
Chameleon (*Chamaeleo calyptratus*). *Archives of Oral Biology* 58:118–133.

Budney, L. 2004. A survey of tooth attachment histology in squamata: The evaluation of tooth
attachment classification and characters. University of Alberta pp.

Budney, L. A., M. W. Caldwell, and A. Albino. 2006. Tooth socket histology in the Cretaceous
snake *Dinilysia*, with a review of amniote dental attachment tissues. *Journal of Vertebrate*
Paleontology 26:138–145.

Caldwell, M. W. 2007. Ontogeny, anatomy and attachment of the dentition in mosasaurs
(Mosasauridae: Squamata). *Zoological Journal of the Linnean Society* 149:687–700.

Cooper JS, Poole DF. The dentition and dental tissues of the agamid lizard, *Uromastyx*. *Journal*
of Zoology. 1973 Jan;169(1):85–100.

Cope, E. D. 1864. On the characters of the higher groups of Reptilia Squamata: and especially of
the *Diploglossa*. *Academy of Natural Sciences* 16:224–231.

Dauphin, Y., and C. T. Williams. 2008. Chemical composition of enamel and dentine in modern
reptile teeth. *Mineral Mag* 72:247–250.

- 556 Delgado, S., T. Davit-Beal, and J. Y. Sire. 2003a. Dentition and tooth replacement pattern in
557 Chalcides (Squamata; Scincidae). *Journal of Morphology* 256:146–159.
- 558 Dosedělová, H., K. Štěpánková, T. Zikmund, H. Lesot, J. Kaiser, K. Novotný, J. Štembírek, Z.
559 Knotek, O. Zahradníček, and M. Buchtová. 2016a. Age-related changes in the tooth–bone
560 interface area of acrodont dentition in the chameleon. *Journal of Anatomy* 229:356–368.
- 561 Erickson, G. M. 1996. Incremental lines of von Ebner in dinosaurs and the assessment of tooth
562 replacement rates using growth line counts. *Proceedings of the National Academy of*
563 *Sciences of the United States of America* 93:14623–14627.
- 564 Estes, R., and C. Charles. 1988. *Phylogenetic Relationships of the Lizard Families: Essays*
565 *Commemorating Charles L. Camp, illustrate.* (Gregory K. Pregill (ed.)). Stanford University
566 Press, 631 pp.
- 567 Fortelius M, Solounias N. Functional characterization of ungulate molars using the abrasion-
568 attrition wear gradient: a new method for reconstructing paleodiets. *American Museum*
569 *Novitates*. 2000 Oct:1–36.
- 570 García, R. A., and V. Zurriaguz. 2016. Histology of teeth and tooth attachment in titanosaurs
571 (Dinosauria; Sauropoda). *Cretaceous Research* 57:248–256.
- 572 Haridy, Y., A. R. H. LeBlanc, and R. R. Reisz. 2018. The Permian reptile *Opisthodontosaurus*
573 *carrolli*: a model for acrodont tooth replacement and dental ontogeny. *Journal of Anatomy*
574 232:371–382.
- 575 Huber, D. R., M. N. Dean, and A. P. Summers. 2008. Hard prey, soft jaws and the ontogeny of
576 feeding mechanics in the spotted ratfish *Hydrolagus colliei*. *Journal of The Royal Society*
577 *Interface* 5:941–953.
- 578 Hugall, J. F., R. Foster, M. Hutchinson, and M. S. Y. Lee. 2008. Phylogeny of Australasian
579 agamid lizards based on nuclear and mitochondrial genes: implications for morphological
580 evolution and biogeography. 343–358.
- 581 Järvinen, E., M. Tummers, and I. Thesleff. 2009. The role of the dental lamina in mammalian
582 tooth replacement. *Journal of Experimental Zoology Part B: Molecular and Developmental*
583 *Evolution* 312:281–291.
- 584 Jenkins, K. M., M. E. H. Jones, T. Zikmund, A. Boyde, and J. D. Daza. 2017. A review of tooth
585 implantation among rhynchocephalians (Lepidosauria). *Journal of Herpetology* 51:300–306.
- 586 Kieser, J. A., T. Tkatchenko, M. C. Dean, M. E. H. Jones, W. Duncan, and N. J. Nelson. 2009.

- 587 Microstructure of dental hard tissues and bone in the tuatara dentary, *Sphenodon punctatus*
588 (Diapsida: Lepidosauria: Rhynchocephalia). *Frontiers of Oral Biology* 13:80–85.
- 589 Kline, A. L. W., and D. Cullum. 2017. A long term study of the tooth replacement phenomenon
590 in the young green iguana, *Iguana iguana*. *Society for the Study of Amphibians and*
591 *Reptiles* 18:176–185.
- 592 LeBlanc, A. R. H., and R. R. Reisz. 2013. Periodontal ligament, cementum, and alveolar bone in
593 the oldest herbivorous tetrapods, and their evolutionary significance. *PLoS ONE* 8.
- 594 LeBlanc, A. R. H., and R. R. Reisz. 2015. Patterns of tooth development and replacement in
595 captorhinid reptiles: a comparative approach for understanding the origin of multiple tooth
596 rows. *Journal of Vertebrate Paleontology* 4634:e919928.
- 597 LeBlanc, A. R. H., R. R. Reisz, D. C. Evans, and A. M. Bailleul. 2016a. Ontogeny reveals
598 function and evolution of the hadrosaurid dinosaur dental battery. *BMC Evolutionary*
599 *Biology* 16:152.
- 600 LeBlanc, A. R. H., R. R. Reisz, K. S. Brink, and F. Abdala. 2016b. Mineralized periodontia in
601 extinct relatives of mammals shed light on the evolutionary history of mineral homeostasis
602 in periodontal tissue maintenance. *Journal of Clinical Periodontology* 43:323–332.
- 603 Lumsden, A. G. S., and J. W. Osborn. 1977. The evolution of chewing: A dentist's view of
604 palaeontology. *Journal of Dentistry* 5:269–287.
- 605 Montanucci, R. 2008. Comparative dentition in four iguanid lizards. *Herpetologica* 24:305–315.
- 606 Motani, R. 1997. Temporal and Spatial Distribution of Tooth Implantation in Ichthyosaurs; pp.
607 81–103 in *Ancient marine reptiles*. San Diego: Academic Press.
- 608 Pyron, R., F. T. Burbrink, and J. J. Wiens. 2013. A phylogeny and revised classification of
609 Squamata, including 4161 species of lizards and snakes. *BMC Evolutionary Biology* 13:93.
- 610  Reisz RR. Origin of dental occlusion in tetrapods: signal for terrestrial vertebrate evolution?.
611 *Journal of Experimental Zoology Part B: Molecular and Developmental Evolution*. 2006
612 May 15;306(3):261–77.
- 613 Richman, J. M., and G. R. Handrigan. 2011. Reptilian tooth development. *Genesis* 49:247–260.
- 614 Shellis, R. P., and B. K. B. Berkovitz. 1976. Observations on the dental anatomy of piranhas
615 (Characidae) with special reference to tooth structure. *Journal of Zoology* 180:69–84.

- 616 Simões, T. R., E. Wilner, M. W. Caldwell, L. C. Weinschütz, and A. W. A. Kellner. 2015. A
617 stem acrodontan lizard in the Cretaceous of Brazil revises early lizard evolution in
618 Gondwana. *Nature Communications* 6:8149.
- 619 Smirina, E., and N. Ananjeva. 2017. On the longevity, growth and reproductive characteristics of
620 Lichtenstein's Toadhead Agama, *Phrynocephalus interscapularis* Lichtenstein, 1856
621 (Agamidae, Sauria). *Amphibia-Reptilia* 38:31–39.
- 622 Smirina, E. M., and N. B. Ananjeva. 2007. Growth layers in bones and acrodont teeth of the
623 agamid lizard *Laudakia stoliczkana* (Blanford, 1875)(Agamidae, Sauria). *Amphibia-
624 Reptilia* 28:193–204.
- 625 Spencer, M. A. 2005. Dental functional morphology: How teeth work. *American Journal of
626 Human Biology* 17:384–385.
- 627 Throckmorton, G. S. 1979. The effect of wear on the cheek teeth and associated dental tissues of
628 the lizard *Uromastix aegyptius* (Agamidae). *Journal of Morphology* 160:195–207.
- 629 Throckmorton, G. Y. S. 1976. Oral food processing in two herbivorous lizards, *Iguana iguana*
630 (Iguanidae) and *Uromastix aegyptius* (Agarnidae). *Journal of Morphology* 148:363–390.
- 631 Westergaard, B., and M. W. J. Ferguson. 1990. Development of the dentition in *Alligator
632 mississippiensis*: Upper jaw dental and craniofacial development in embryos, hatchlings,
633 and young juveniles, with a comparison to lower jaw development. *American Journal of
634 Anatomy* 187:393–421.
- 635 Witten, P. E., and A. Huysseune. 2009. A comparative view on mechanisms and functions of
636 skeletal remodelling in teleost fish, with special emphasis on osteoclasts and their function.
637 *Biological Reviews* 84:315–346.
- 638 Zaher, H., and O. Rippel. 1999. Tooth implantation and replacement in Squamates, with special
639 reference to mosasaur lizards and snakes. *American Museum Novitates* 3271:1–19.
- 640 Zari, T. A. 1999. On the reproductive biology of the herbivorous spiny-tailed agamid *Uromastyx
641 philbyi* in Western Saudi Arabia. *Zoology in the Middle East* 19:123–130.
- 642 Zheng, Y., and J. J. Wiens. 2016. Combining phylogenomic and supermatrix approaches, and a
643 time-calibrated phylogeny for squamate reptiles (lizards and snakes) based on 52 genes and
644 4162 species. *Molecular Phylogenetics and Evolution* 94:537–547.

Captions

FIGURE 1. A comparative figure showing the external morphological differences in the dentition and mandibles between juvenile and adult specimens of *Pogona vitticeps*; (A) Right mandibular ramus of juvenile specimen ROM R8234; (B) Right mandibular ramus of adult specimen ROM R8507; (C) superimposed outlines of mandibles of varying ontogenetic stages, showing that most growth occurs in the posterior end of the mandible. **Abbreviations; ad**, acrodont dentition; **cp**, coronoid process; **pd**, pleurodont dentition; **wf**, wear facets. Arrows indicating un-ankylosed teeth. Scale bar= 1 cm.

FIGURE 2. Details of external morphology of the juvenile dentition of *Pogona vitticeps*. (A) ROM R8234 in lingual view showing un-ankylosed dentition, the most posterior growing into the coronoid; (B) ROM R8418 scanning electron microscope image of incomplete juvenile mandible with; (C) closeup showing fibrous alveolar bone between the teeth; (D) closeup showing the lingual contribution of dentine to the teeth; (E) closeup showing the youngest tooth growing into and resorbing the coronoid. **Abbreviations; ab**, alveolar bone; **cr**, coronoid bone; **de**, dentine. Arrows showing un-ankylosed teeth.

FIGURE 3. Longitudinal histological sections of the juvenile mandible of *Pogona vitticeps* with a focus on the dentition. (A) External view of ROM R8234, box outlining the dentition cut; (B) a section of the posteriormost seven tooth positions. (C) a section of the posterior seven tooth positions in cross polarized light; (D) a schematic representation showing the distinct tissues as well as the progressive ankylosis of the teeth. **Abbreviations; ab**, alveolar bone; **de**, dentine; **jb**, jaw bone; **pc**, pulp cavity. Scale bar = 1000um

FIGURE 4. Coronal sections of juvenile mandible of *Pogona vitticeps* with a special focus on tooth histology. (A) Coronal section of the jaw and tooth of a juvenile specimen of *P. vitticeps* ROM R8510 showing minimal ossification of the jaw bone, and pleurodont tooth attachment; (B) close up of a juvenile specimen's (ROM R8510) dentition, showing unworn morphology; (C) coronal section of a juvenile specimen (ROM R8234) showing pleurodont implantation, and remodeling; (D) closeup of the attachment site in ROM R8234, showing the labial side of the dentition being resorbed by the osteoclast; (E) coronal section of the coronoid process in ROM R8234 showing a tooth developing lingually inside the coronoid and dentary; (F) a close up of

675 the tooth in the jaw bone, showing the resorption of the jawbone around newly developing tooth.
 676 **Abbreviations:** **ab**, alveolar bone; **ac**, acellular cementum; **cb**, cancellous bone; **cp**, coronoid
 677 process **de**, dentine; **dt**, developing tooth; **lb**, lamellar bone; **en**, enamel; **ob**, osteoblast **oc**,
 678 osteoclast; **rb**, resorption bay. All un-labeled scale bars = 1 mm.

679 **FIGURE 5.** Longitudinal sections of adult mandible of *Pogona vitticeps* with a focus on tooth
 680 histology. (A) External view of specimen ROM R8507, box outlining the dentulous area
 681 sectioned; (B) broad view showing the variation between the anterior and the posterior dentition;
 682 (C) a closeup of the worn dentition showing wear facets and remodeling zones; (D) broad view
 683 showing the variation between the anterior and the posterior dentition, showing the depth of wear
 684 facets and arrangement of vasculature; (E) a close up of the anterior dentition with open pulp
 685 cavities and associated vasculature. **Abbreviations:** **de**, dentine; **jb**, jaw bone; **pc**, pulp cavity;
 686 **vc**, vascular canal; **wf**, wear facet. The arrows demarcate the extensive vasculature leading to
 687 pulp cavities. Scale bar=1000um

688 **FIGURE 6:** Coronal sections of adult mandible of *Pogona vitticeps* with a focus on tooth
 689 histology. (A) Coronal section of the jaw and tooth of an adult specimen of *P. vitticeps* ROM
 690 R8507 showing the maintained vasculature and worn enamel on the labial side; (B) coronal
 691 section of the jaw and tooth of an adult specimen of *P. vitticeps* ROM R8507 showing the
 692 extensive remodeling of the tooth attachment site as well as the tooth. **Abbreviations:** **de**,
 693 dentine; **lb**, lamellar bone; **en**, enamel; **vc**, vascular canals; **rz**, remodeling zone. Scale
 694 bar=500um

695 **FIGURE 7:** Schematic explanation of implantation changes through ontogeny and tooth wear.
 696 (A) three basic traditional implantation categories, without attachment tissue illustrated; (B)
 697 pleurodont impanated tooth is attached via alveolar bone, yellow arrows show direction of
 698 dentine deposition; (C) purple arrows show direction of dentine resorption by odontoclasts, blue
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 700 (D) blue arrow shows further bone deposition; (E) the tooth appears implanted at the apex of the
 701 dentary; (F) Shows the position of maxillary tooth; (G) the enamel is worn on the labial side; (H)
 702 with advanced wear, the enamel, dentine, and dentary bone are worn on the labial side.

703

704 **Supplementary**

705 **S.FIGURE 1.** Ontogenetic sequence of *P. vitticeps* mandibles that were used to make the growth
 706 schematic in Figure 1C. (A) ROM R8234; (B) ROM R8150; (C) ROM R8227;(D) ROM R9422;
 707 (E) ROM R8507; (F) ROM R8189. Scale bar = 1 cm.

708 **TABLE 1.** A table of all specimens of *P. vitticeps* examined for this study. In orange are specific
 709 specimens either sectioned or figured.

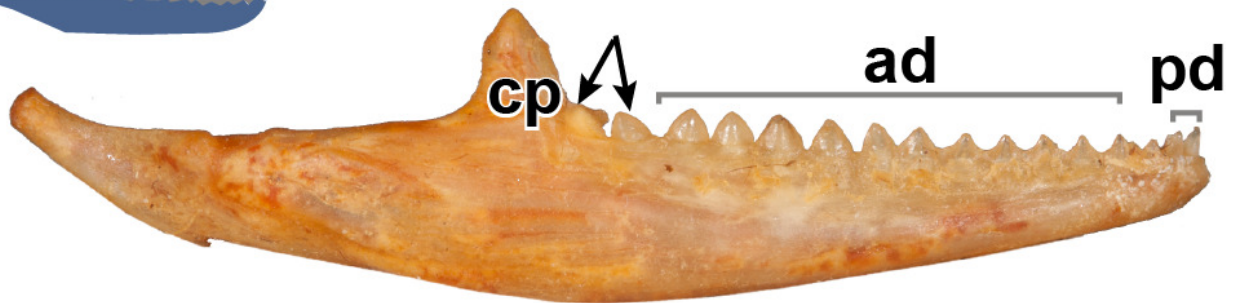
710

Figure 1

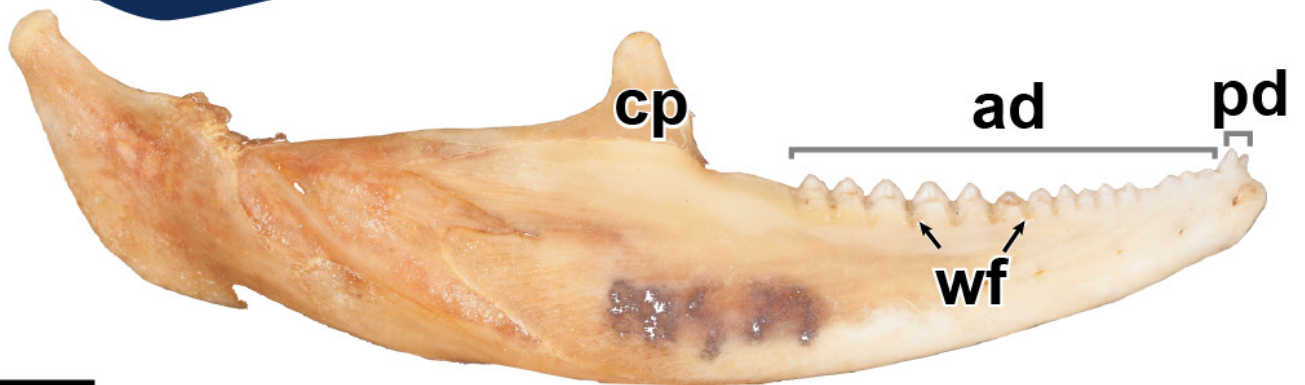
A comparative figure showing the external morphological differences in the dentition and mandibles between juvenile and adult specimens of *Pogona vitticeps*

(A) Right mandibular ramus of juvenile specimen ROM R8234; (B) Right mandibular ramus of adult specimen ROM R8507; (C) superimposed outlines of mandibles of varying ontogenetic stages, showing that most growth occurs in the posterior end of the mandible. **Abbreviations;** **ad**, acrodont dentition; **cp**, coronoid process; **pd**, pleurodont dentition; **wf**, wear facets. Arrows indicating un-ankylosed teeth. Scale bar= 1 cm. Photo credit: Diane Scott

A



B



C

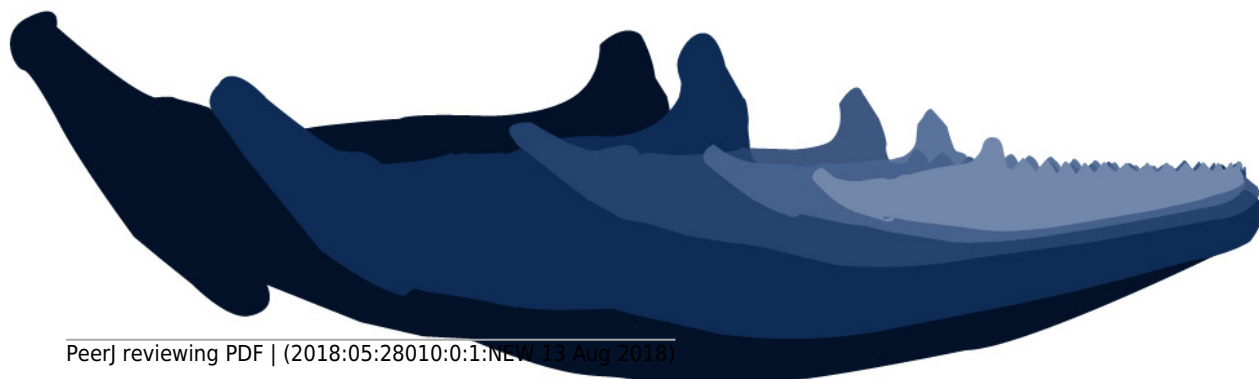


Figure 2

Details of external morphology of the juvenile dentition of *Pogona vitticeps*.

(A) ROM R8234 in lingual view showing un-ankylosed dentition, the most posterior growing into the coronoid; (B) ROM R8418 scanning electron microscope image of incomplete juvenile mandible with; (C) closeup showing fibrous alveolar bone between the teeth; (D) closeup showing the lingual contribution of dentine to the teeth; (E) closeup showing the youngest tooth growing into and resorbing the coronoid. **Abbreviations;** **ab**, alveolar bone; **cr**, coronoid bone; **de**, dentine. Arrows showing un-ankylosed teeth. Image credit: Diane Scott

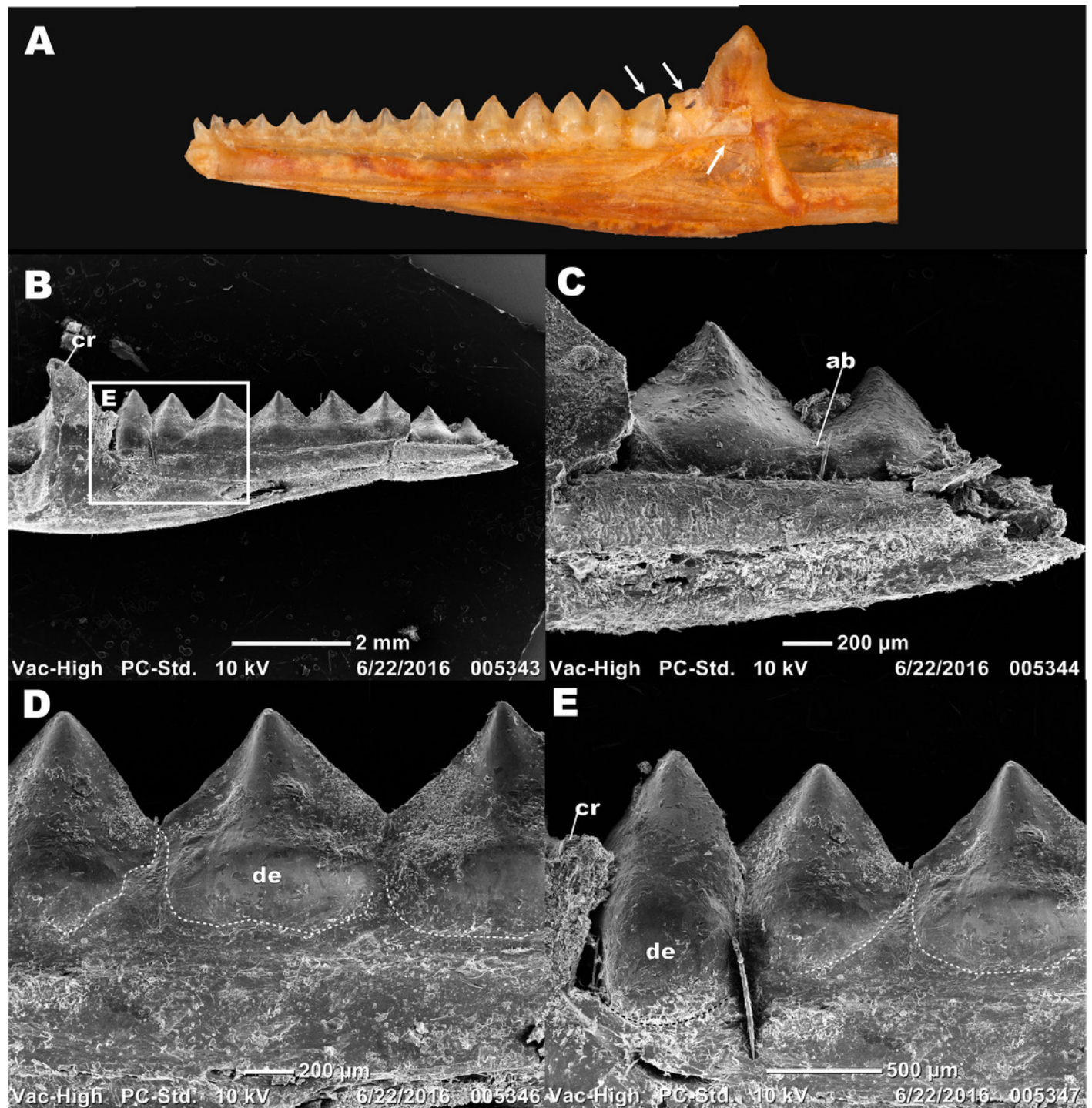


Figure 3

Longitudinal histological sections of the juvenile mandible of *Pogona vitticeps* with a focus on the dentition.

(A) External view of ROM R8234, box outlining the dentition cut; (B) a section of the posteriormost seven tooth positions. (C) a section of the posterior seven tooth positions in cross polarized light; (D) a schematic representation showing the distinct tissues as well as the progressive ankylosis of the teeth. **Abbreviations;** **ab**, alveolar bone; **de**, dentine; **jb**, jaw bone; **pc**, pulp cavity. Scale bar = 1000um



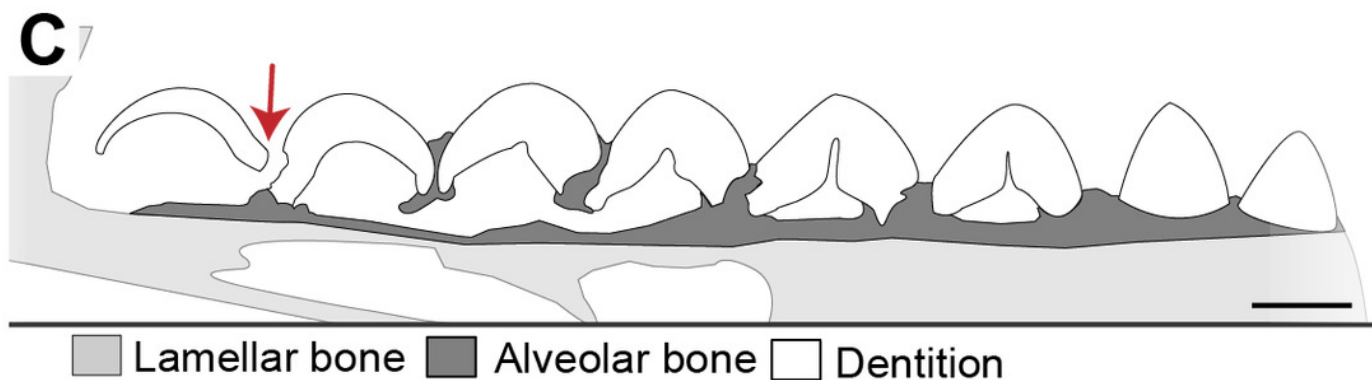
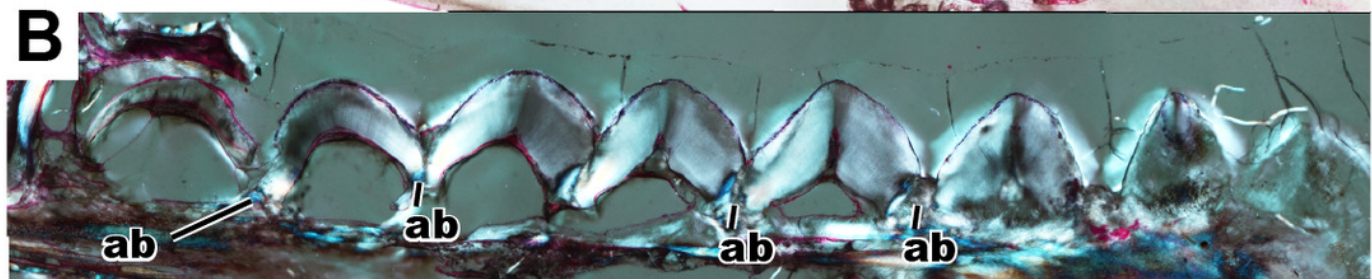
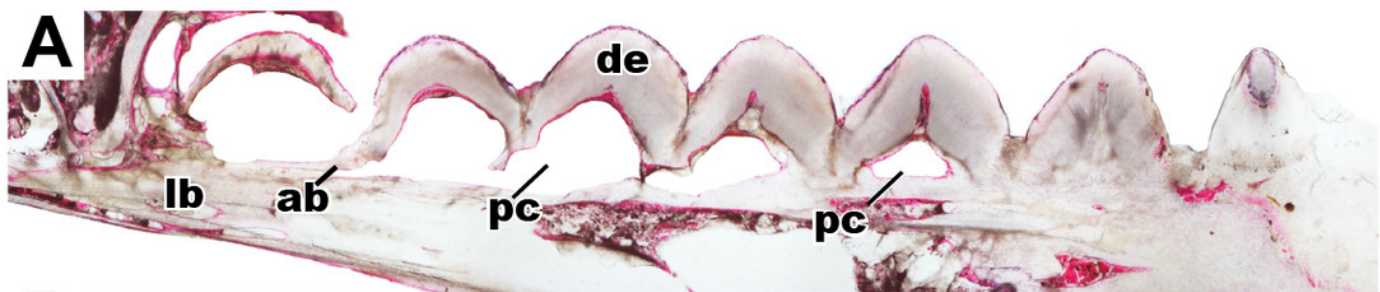
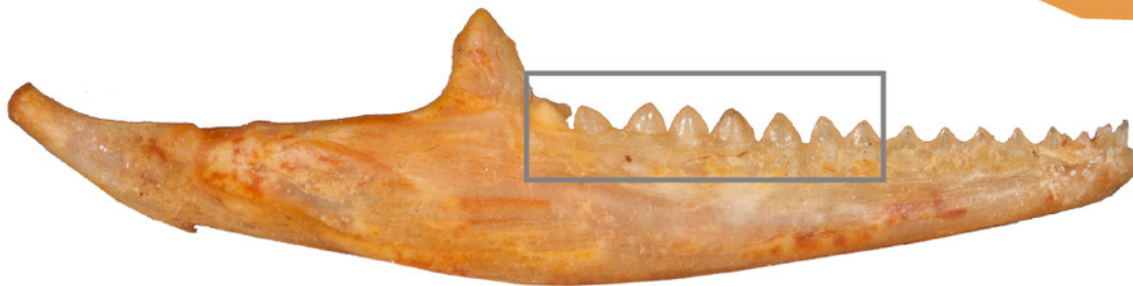
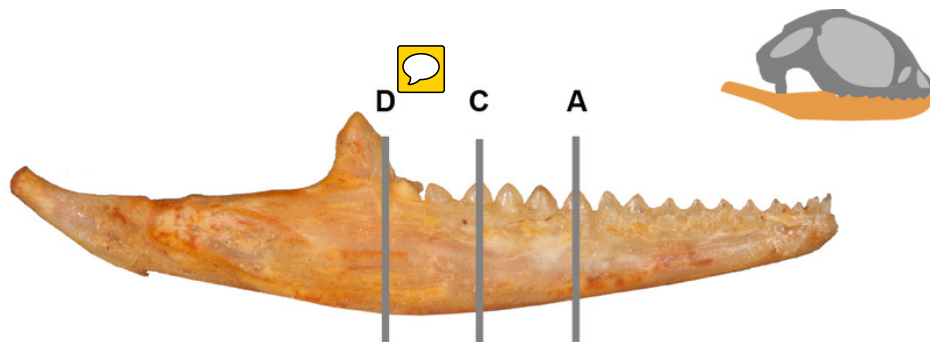


Figure 4

Coronal sections of juvenile mandible of *Pogona vitticeps* with a special focus on tooth histology.

(A) Coronal section of the jaw and tooth of a juvenile specimen of *P. vitticeps* ROM R8510 showing minimal ossification of the jaw bone, and pleurodont tooth attachment; (B) close up of a juvenile specimen's (ROM R8510) dentition, showing unworn morphology; (C) coronal section of a juvenile specimen (ROM R8234) showing pleurodont implantation, and remodeling; (D) closeup of the attachment site in ROM R8234, showing the labial side of the dentition being resorbed by the osteoclast; (E) coronal section of the coronoid process in ROM R8234 showing a tooth developing lingually inside the coronoid and dentary; (F) a close up of the tooth in the jaw bone, showing the resorption of the jawbone around newly developing tooth. **Abbreviations:** **ab**, alveolar bone; **ac**, acellular cementum; **cb**, cancellous bone; **cp**, coronoid process **de**, dentine; **dt**, developing tooth; **lb**, lamellar bone; **en**, enamel; **ob**, osteoblast **oc**, osteoclast; **rb**, resorption bay. All un-labeled scale bars = 1 mm.



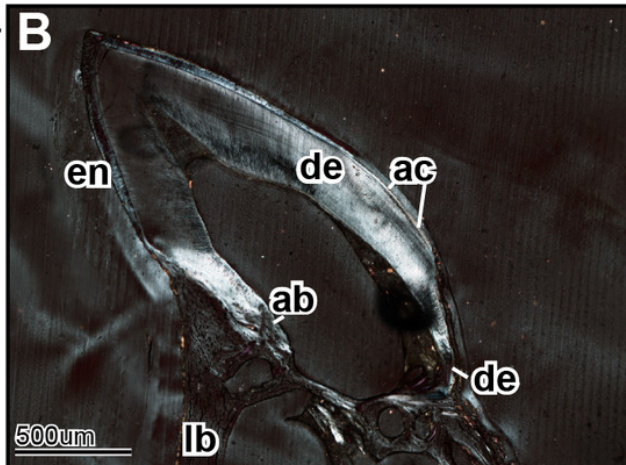


A



← Labial

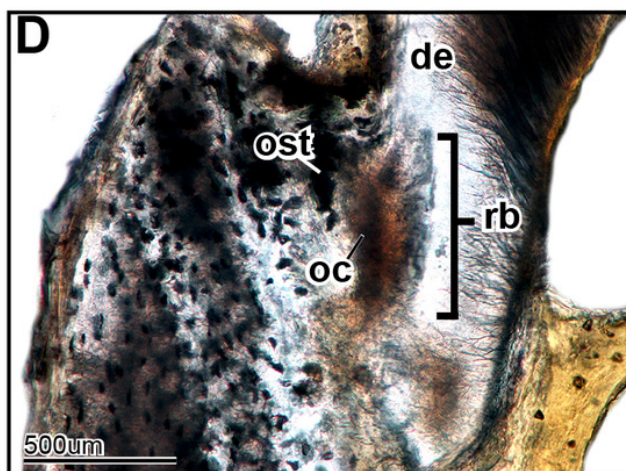
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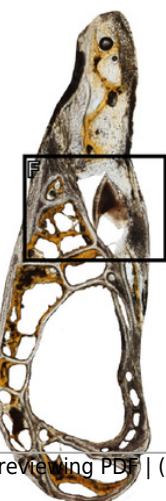
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D



E



F

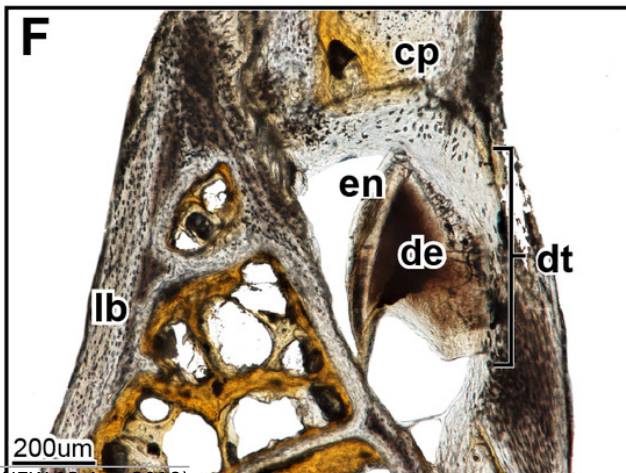


Figure 5

Longitudinal sections of adult mandible of *Pogona vitticeps* with a focus on tooth histology.

(A) External view of specimen ROM R8507, box outlining the dentulous area sectioned; (B) broad view showing the variation between the anterior and the posterior dentition; (C) a closeup of the worn dentition showing wear facets and remodeling zones; (D) broad view showing the variation between the anterior and the posterior dentition, showing the depth of wear facets and arrangement of vasculature; (E) a close up of the anterior dentition with open pulp cavities and associated vasculature. **Abbreviations:** **de**, dentine; **jb**, jaw bone; **pc**, pulp cavity; **vc**, vascular canal; **wf**, wear facet. The arrows demarcate the extensive vasculature leading to pulp cavities. Scale bar=1000um

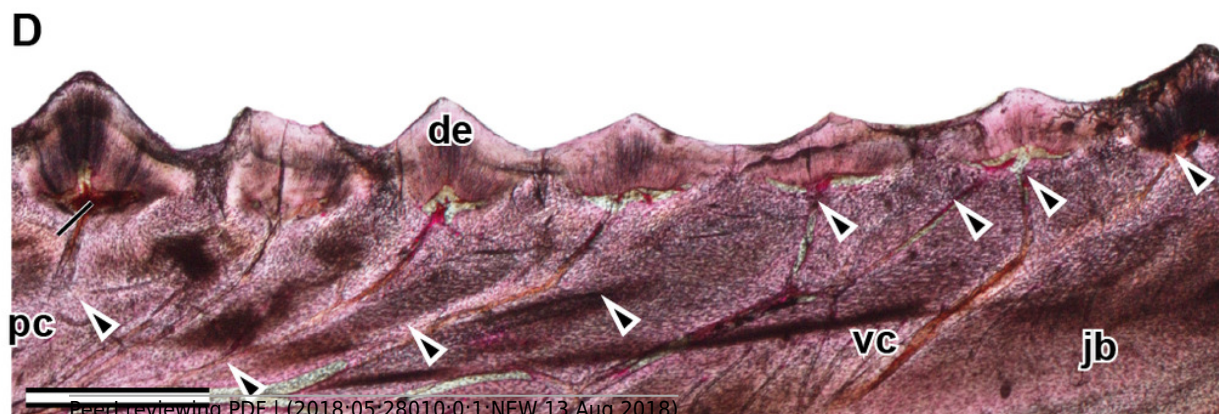
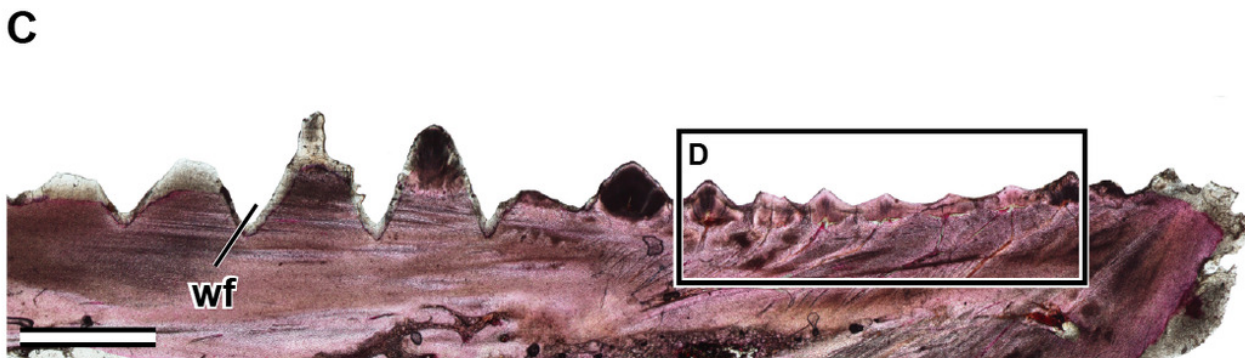
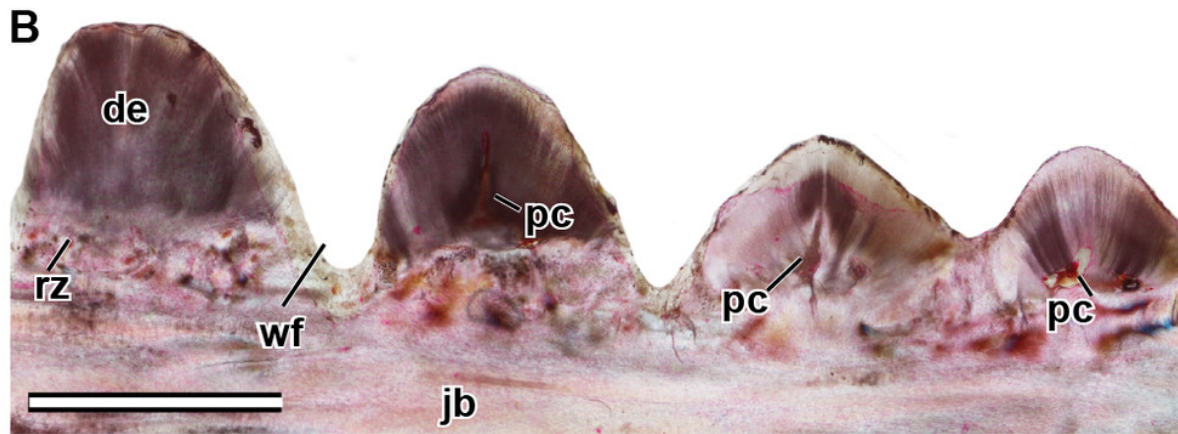


Figure 6

Coronal sections of adult mandible of *Pogona vitticeps* with a focus on tooth histology.

(A) Coronal section of the jaw and tooth of an adult specimen of *P. vitticeps* ROM R8507 showing the maintained vasculature and worn enamel on the labial side; (B) coronal section of the jaw and tooth of an adult specimen of *P. vitticeps* ROM R8507 showing the extensive remodeling of the tooth attachment site as well as the tooth.  **Abbreviations:** **de**, dentine; **lb**, lamellar bone; **en**, enamel; **vc**, vascular canals; **rz**, remodeling zone. Scale bar=500um

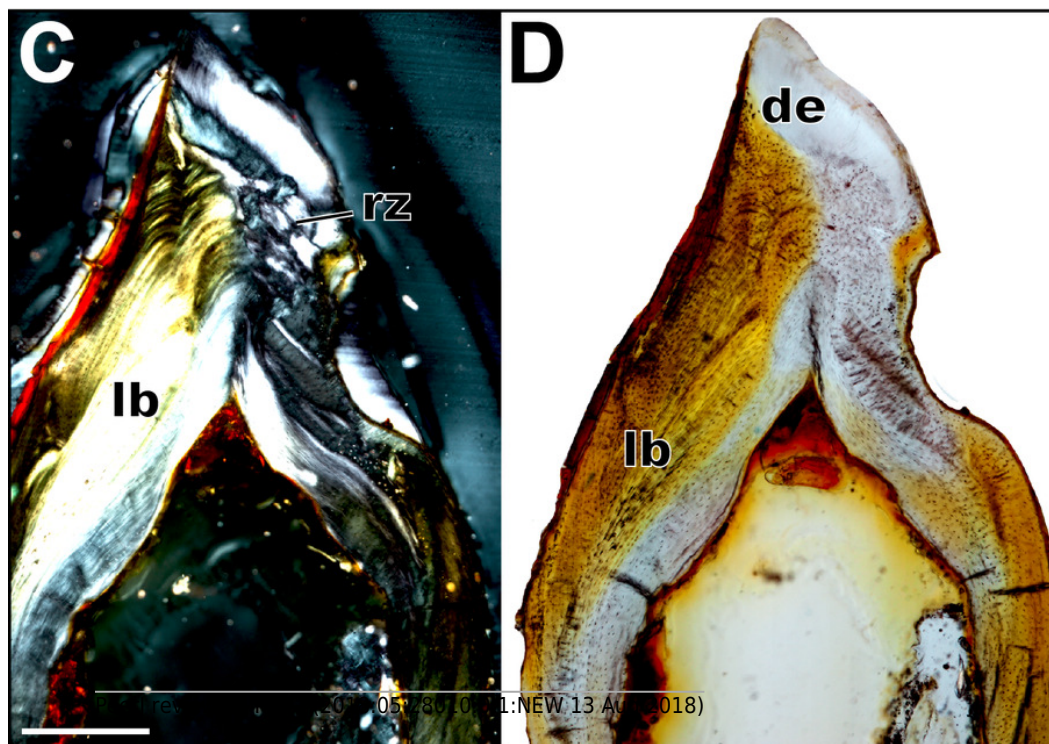
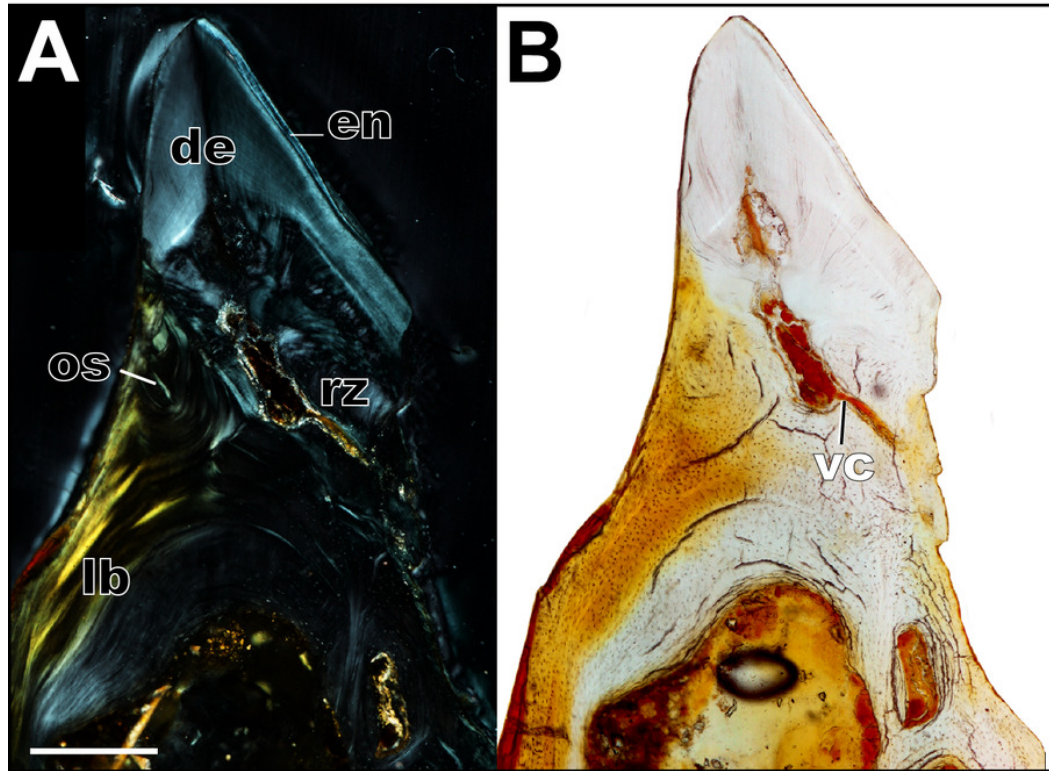
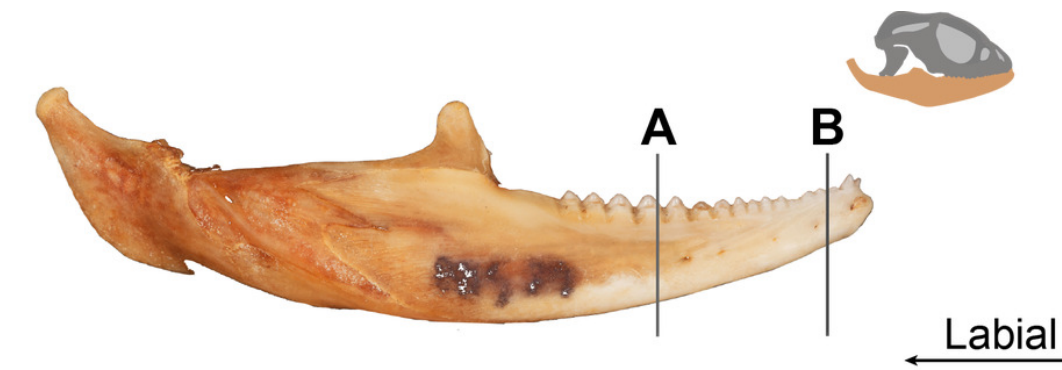


Figure 7

Schematic explanation of implantation changes through ontogeny and tooth wear.

(A) three basic traditional implantation categories, without attachment tissue illustrated; (B) pleurodont impanated tooth is attached via alveolar bone, yellow arrows show direction of dentine deposition; (C) purple arrows show direction of dentine resorption by odontoclasts, blue arrow shows direction of bone deposition by odontoblasts, trabecula in the dentary is reduced; (D) blue arrow shows further bone deposition; (E) the tooth appears implanted at the apex of the dentary; (F) Shows the position of maxillary tooth; (G) the enamel is worn on the labial side; (H) with advanced wear, the enamel, dentine, and dentary bone are worn on the labial side.

