

Postmortem transport in fossil and modern shelled cephalopods (#29347)

1



First submission

Editor guidance

Please submit by **2 Aug 2018** for the benefit of the authors (and your \$200 publishing discount).



Structure and Criteria

Please read the 'Structure and Criteria' page for general guidance.



Raw data check

Review the raw data. Download from the [materials page](#).



Image check

Check that figures and images have not been inappropriately manipulated.

Privacy reminder: If uploading an annotated PDF, remove identifiable information to remain anonymous.

Files

Download and review all files from the [materials page](#).

7 Figure file(s)

2 Table file(s)

1 Raw data file(s)



Structure your review

The review form is divided into 5 sections. Please consider these when composing your review:

1. BASIC REPORTING
2. EXPERIMENTAL DESIGN
3. VALIDITY OF THE FINDINGS
4. General comments
5. Confidential notes to the editor

 You can also annotate this PDF and upload it as part of your review

When ready [submit online](#).

Editorial Criteria

Use these criteria points to structure your review. The full detailed editorial criteria is on your [guidance page](#).

BASIC REPORTING

-  Clear, unambiguous, professional English language used throughout.
-  Intro & background to show context. Literature well referenced & relevant.
-  Structure conforms to [PeerJ standards](#), discipline norm, or improved for clarity.
-  Figures are relevant, high quality, well labelled & described.
-  Raw data supplied (see [PeerJ policy](#)).

EXPERIMENTAL DESIGN

-  Original primary research within [Scope of the journal](#).
-  Research question well defined, relevant & meaningful. It is stated how the research fills an identified knowledge gap.
-  Rigorous investigation performed to a high technical & ethical standard.
-  Methods described with sufficient detail & information to replicate.

VALIDITY OF THE FINDINGS

-  Impact and novelty not assessed. Negative/inconclusive results accepted. *Meaningful* replication encouraged where rationale & benefit to literature is clearly stated.
-  Data is robust, statistically sound, & controlled.
-  Speculation is welcome, but should be identified as such.
-  Conclusions are well stated, linked to original research question & limited to supporting results.

Standout reviewing tips

3



The best reviewers use these techniques

Tip

Support criticisms with evidence from the text or from other sources

Example

Smith et al (J of Methodology, 2005, V3, pp 123) have shown that the analysis you use in Lines 241-250 is not the most appropriate for this situation. Please explain why you used this method.

Give specific suggestions on how to improve the manuscript

Your introduction needs more detail. I suggest that you improve the description at lines 57- 86 to provide more justification for your study (specifically, you should expand upon the knowledge gap being filled).

Comment on language and grammar issues

The English language should be improved to ensure that an international audience can clearly understand your text. Some examples where the language could be improved include lines 23, 77, 121, 128 – the current phrasing makes comprehension difficult.

Organize by importance of the issues, and number your points

1. Your most important issue
2. The next most important item
3. ...
4. The least important points

Please provide constructive criticism, and avoid personal opinions

I thank you for providing the raw data, however your supplemental files need more descriptive metadata identifiers to be useful to future readers. Although your results are compelling, the data analysis should be improved in the following ways: AA, BB, CC

Comment on strengths (as well as weaknesses) of the manuscript

I commend the authors for their extensive data set, compiled over many years of detailed fieldwork. In addition, the manuscript is clearly written in professional, unambiguous language. If there is a weakness, it is in the statistical analysis (as I have noted above) which should be improved upon before Acceptance.

Postmortem transport in fossil and modern shelled cephalopods

Margaret M. Yacobucci ^{Corresp.} ¹

¹ Department of Geology, Bowling Green State University, Bowling Green, Ohio, United States

Corresponding Author: Margaret M. Yacobucci

Email address: mmyacob@bgsu.edu

The chambered shells of cephalopod mollusks, such as modern *Nautilus* and fossil ammonoids, have the potential to float after death, which could result in significant postmortem transport of shells away from living habitats. Such transport would call into question these clades' documented biogeographic distributions and therefore the many (paleo)biological interpretations based on them. It is therefore imperative to better constrain the likelihood and extent of postmortem transport in modern and fossil cephalopods. Here, I combine the results of classic experiments on postmortem buoyancy with datasets on cephalopod shell form to determine that only those shells with relatively high inflation are likely to float for a significant interval after death and therefore potentially experience postmortem transport. Most ammonoid cephalopods have shell forms making postmortem transport unlikely. Data on shell forms and geographic ranges of early Late Cretaceous cephalopod genera demonstrate that even genera with shell forms conducive to postmortem buoyancy do not, in fact, show artificially inflated biogeographic ranges relative to genera with non-buoyant morphologies. Finally, georeferenced locality data for living nautilid specimens and dead drift shells indicate that most species have relatively small geographic ranges and experience limited drift. *Nautilus pompilius* is the exception, with a broad Indo-Pacific range and drift shells found far from known living populations. Given the similarity of *N. pompilius* to other nautilids in its morphology and ecology, it seems unlikely that this species would have a significantly different postmortem fate than its close relatives. Rather, it is suggested that drift shells along the east African coast may indicate the existence of modern (or recently extirpated) living populations of nautilus in the western Indian Ocean, which has implications for the conservation of these cephalopods.

Postmortem transport in fossil and modern shelled cephalopods

Margaret M. Yacobucci

Department of Geology, Bowling Green State University, Bowling Green, Ohio, 43403, USA

Email address: mmyacob@bgsu.edu

8 Abstract

9 The chambered shells of cephalopod mollusks, such as modern *Nautilus* and fossil
10 ammonoids, have the potential to float after death, which could result in significant postmortem
11 transport of shells away from living habitats. Such transport would call into question these
12 clades' documented biogeographic distributions and therefore the many (paleo)biological
13 interpretations based on them. It is therefore imperative to better constrain the likelihood and
14 extent of postmortem transport in modern and fossil cephalopods. Here, I combine the results of
15 classic experiments on postmortem buoyancy with datasets on cephalopod shell form to
16 determine that only those shells with relatively high inflation are likely to float for a significant
17 interval after death and therefore potentially experience postmortem transport. Most ammonoid
18 cephalopods have shell forms making postmortem transport unlikely. Data on shell forms and
19 geographic ranges of early Late Cretaceous cephalopod genera demonstrate that even genera
20 with shell forms conducive to postmortem buoyancy do not, in fact, show artificially inflated
21 biogeographic ranges relative to genera with non-buoyant morphologies. Finally, georeferenced
22 locality data for living nautilid specimens and dead drift shells indicate that most species have
23 relatively small geographic ranges and experience limited drift. *Nautilus pompilius* is the
24 exception, with a broad Indo-Pacific range and drift shells found far from known living
25 populations. Given the similarity of *N. pompilius* to other nautilids in its morphology and
26 ecology, it seems unlikely that this species would have a significantly different postmortem fate
27 than its close relatives. Rather, it is suggested that drift shells along the east African coast may
28 indicate the existence of modern (or recently extirpated) living populations of nautilus in the
29 western Indian Ocean, which has implications for the conservation of these cephalopods.

30

Introduction

The problem of postmortem transport

Biogeographic distributions strongly influence the ecology, evolution, and extinction of clades. While scientists studying modern organisms can collect data on the observed locations of sampled living specimens, paleontologists must assume that fossil localities provide an accurate estimate of the group's living geographic range. For most marine animal groups, especially benthic invertebrates, that assumption appears to work well, as their hard parts tend to be buried and fossilized at or near their life locations (Kidwell & Flessa, 1996; Tomašových & Kidwell, 2009; Tyler & Kowalewski, 2017). However, cephalopod mollusks with chambered shells present a special challenge—once their soft parts are removed, cephalopod shells can float after death and therefore be picked up and transported by surface currents (Fig. 1). This postmortem transport or "drift" has been observed in living nautilid species, but how commonly it occurs in ancient cephalopod groups has been debated in the scientific literature for over 100 years. If postmortem transport was frequent and extensive, the geographic distributions of fossil cephalopods should be considered unreliable proxies for living ranges. Biogeographic studies of fossil cephalopods require some assurance that locality data are meaningful, especially as paleontologists integrate more quantitative methods to studying paleobiogeographic patterns (Brayard et al., 2015; Ifrim, Lehmann & Ward, 2015; Korn & De Baets, 2015; Lehmann et al., 2015; Wani, 2017; Yacobucci, 2017). Also, as modern nautilids experience greater fishery pressure and habitat disruption due to climate change and other anthropogenic impacts, conservation efforts will need to accurately assess population distributions (Dunstan, Alanis & Marshall, 2010; Dunstan, Bradshaw & Marshall, 2011; Dunstan, Ward & Marshall, 2011a, 2011b; Sinclair et al., 2011; De Angelis, 2012; Barord et al., 2014; Williams et al., 2015; Ward,

Dooley & Barord, 2016; Saunders, Greenfest-Allen & Ward, 2017). It is therefore imperative to determine the likely frequency of postmortem transport in both fossil and modern shelled cephalopods.

In the 19th and early 20th centuries, some workers noted that the small geographic ranges and concentrations of undamaged fossil cephalopod shells indicated little postmortem transport, while others argued that the lack of soft part preservation (particularly for ammonoids) indicated that the shells must have drifted for some time before sinking to the seafloor (Walther, 1897; Reyment, 1958). The "drift" argument was bolstered by observations in the mid-20th century of transport in modern *Nautilus* in Australia (Iredale, 1944), from the Philippines to Japan (Kobayashi, 1954), to Thailand (Hamada, 1964; Toriyama et al. 1965), and to the Bay of Bengal (Teichert, 1970). Reyment (1958) provided a review of much of this earlier literature. Reyment claimed that postmortem drift was common in both modern and fossil shelled cephalopods (which he described as the minority viewpoint at the time) and investigated the buoyancy of empty shells to determine which shell characteristics were related to greater buoyancy and therefore postmortem transport (see "Cephalopod shell buoyancy" below). Reyment continued throughout his career to argue vigorously that postmortem drift in cephalopods was "the rule rather than the exception" (Reyment, 1970, 1973, 2008) and influenced the position of many other paleontologists (e.g., Stenzel, 1964; House, 1973, 1987; Cecca, 2002) as well as biologists (e.g., Reid, 2016). For example, Chamberlain & Weaver (1978) claimed that modern nautilid shells "often" rise to the surface after death and are "often dispersed by currents--sometimes on an impressive scale...thousands of miles from the habitat of the animals that build them...and similar behavior is inferred for many fossil cephalopods..." (Chamberlain & Weaver, 1978, p. 673-674)

On the other hand, the extent to which postmortem transport has affected the distribution of fossil cephalopods has been questioned. Many fossil localities include very large numbers of well-preserved, intact shells, which is hard to explain by postmortem transport (Kennedy & Cobban, 1976). It is also well-established that many cephalopod taxa or morphotypes are strongly tied to particular depositional environments and sedimentary facies, implying fidelity of dead shells to their living habitat (Lukeneder, 2015). Paleontologists have noted that postmortem transport is likely to remove drift shells from the fossil record entirely, as they degrade over time and end up destroyed when carried by currents into high energy shoreline environments (Hewitt, 1988; Maeda & Wilacher, 1996; Maeda, Mapes & Mapes, 2003; Mapes et al., 2010b; Hembree, Mapes & Goiran, 2014). Fossil cephalopods, therefore, likely represent shells that sank soon after death.

Chamberlain, Ward & Weaver (1981) used buoyancy calculations to argue that most ammonoids, especially those with shells less than 5 cm diameter or found in deeper water settings, would quickly sink to the sea floor after the removal of soft parts, as water flooded the phragmocone (the chambered portion of the shell). A similar argument has been made for Paleozoic and Mesozoic nautiloids (Hewitt & Westermann, 1996; Chirat, 2000), although Chirat (2000) suggested that the Cenozoic nautilid *Aturia* Bronn, 1838 experienced unusually extensive drift because its very long septal neck would have slowed the rate of flooding. Wani et al. (2005) conducted experiments on modern *Nautilus pompilius* Linnaeus, 1758 and showed that the phragmocone floods quickly once the mantle tissue is detached, especially if the shell is small. They argued that only shells over 20 cm in diameter were likely to experience significant postmortem transport, consistent with their claim that observations of long-distance postmortem drift in modern nautilids—while sometimes dramatic—are actually quite rare.

Mapes and colleagues (2010a) pointed out that we typically only encounter modern nautilid shells that have floated into nearshore settings and have little firsthand information about what happens to nautilid shells that sink quickly to the sea floor. Remarkable data from dredged specimens of *Nautilus macromphalus* Sowerby, 1848 show nautilid shells do indeed accumulate on the sea floor in deep water, near their living habitats (Roux, 1990; Roux et al., 1991; Mapes et al. 2010a; Seuss et al., 2015; Tomašových et al., 2016, 2017). Evidence of bioerosion and encrustation has been used as evidence for the duration of transport and exposure at the sea floor in both modern (Seuss et al., 2015; Tomašových et al., 2016, 2017) and fossil (Luci & Cichowolski, 2014) nautilids and in ammonoids (Luci, Cichowolski & Aguirre-Urreta, 2016). Such detailed taphonomic analyses are necessary, as workers have argued that one must not assume postmortem transport but rather search for evidence of it in specific cases (Wani, 2004; Wani 2004; Wani et al. 2005; Wani and Gupta, 2015; Yacobucci, 2015).

Cephalopod shell buoyancy

For a cephalopod shell to experience postmortem drift, it must first become positively buoyant after the death of the animal. Numerous workers have investigated the buoyancy of cephalopod shells, both during life and after death, via physical experiments and mathematical models (Trueman, 1941; Denton & Gilpin-Brown, 1966; Westermann, 1971; Chamberlain & Weaver, 1978; Ward & Martin, 1978; Ward & Greenwald, 1982; Greenwald & Ward, 1987; Jacobs & Chamberlain, 1996; Kröger, 2002; Hammer & Bucher, 2006; Naglik, Rikhtegar & Klug, 2014; Tajika et al., 2014; Hoffman et al., 2015; Naglik et al., 2015), with new imaging and computational techniques (Hoffmann & Zachow, 2011; Hoffmann et al., 2015; Lemanis et al., 2015; Peterman, Barton & Yacobucci, 2018) enabling ever more sophisticated analyses. Workers have established that, in life, modern nautilids are generally neutral to slightly negatively

buoyant (Ward & Martin, 1978; Greenwald & Ward, 1987). Newly formed chambers of the phragmocone are emptied of liquid slowly, at a rate of about 1 mL per chamber per day, via the siphuncle, a tube of tissue extending back from the soft body of the animal through openings in the septal walls that define the chambers (Ward & Martin, 1978). Modern nautilids are capable of partially refilling empty chambers in 10 to 30 hours (for a rate of about 2 mL/day) in response to sudden increases in buoyancy, for instance, if shell is removed by a predator (Ward & Greenwald, 1982). This partial refilling, though, may be insufficient for the animal to regain neutral buoyancy.

After a shelled cephalopod dies and some or all of its soft parts are removed, the shell will become positively buoyant and float until the phragmocone is filled by seawater moving through the siphuncular opening (assuming the phragmocone is intact) (Wani & Gupta, 2015). A critical question is how quickly this flooding of the phragmocone happens. Wani et al. (2005) conducted taphonomic experiments in which freshly killed *N. pompilius* in the Philippines were set on deep (320 m) and shallow (50 m) seafloors. They found that flooding of the phragmocone did not occur immediately postmortem, but that most shells at both depths were flooded within 3 (shallow) to 7 (deep) days. Flooding began when decaying mantle tissue pulled away from the back of the body chamber; presumably removal of soft parts by predation or scavenging would hasten the initiation of chamber filling. Wani et al. (2005) further argued that the rate of flooding would be a function of the radius, thickness, and length of the siphuncular tube. Since fossil ammonoids possessed a siphuncle 1.5 to 3 times as long as that of modern *N. pompilius*, they predicted that ammonoid phragmocones would flood faster than modern nautilids.

Reyment (1973) explicitly connected variations in shell form to the likelihood of positive postmortem buoyancy in ammonoids. He used modern nautilid shells and plastic models of a

146 variety of ammonoid shell morphotypes in laboratory experiments to identify how differing shell
 147 shapes produced different degrees of buoyancy in the empty shells. Reyment (1973) then
 148 performed a principal coordinates analysis on five shell shape parameters (shell diameter,
 149 maximum inflation, body chamber length, ventral inflation, and umbilical width) for 42
 150 Mesozoic ammonoid species. The position of each species on a plot of the first two principal
 151 coordinates indicates its shell shape (Fig. 2). Reyment then mapped out on the plot which shell
 152 shapes had more or less buoyancy postmortem. He concluded that serpenticones (narrow shells
 153 with a wide umbilicus) and oxycones (disc-shapes shells with a high whorl expansion rate) were
 154 less buoyant while the more inflated sphericone shells showed higher postmortem buoyancy.
 155 Hence, it can be predicted that spheroconic ammonoids would be more likely to experience
 156 extensive postmortem transport. 

157 **Linking shell form, postmortem transport potential, and geographic distribution**

158 While Reyment himself insisted that postmortem drift was pervasive among all fossil
 159 cephalopods (2008), his own work (1973) suggested a more complex relationship between shell
 160 form, postmortem buoyancy, and transport potential. Reyment's (1973) morphospace for
 161 Mesozoic ammonoids closely resembles the ternary Westermann morphospace devised by
 162 Ritterbush & Bottjer (2012) to capture morphological variations in regularly coiled ammonoids
 163 (Fig. 3). This resemblance is understandable given the overlap in shell shape parameters that the
 164 two approaches use to construct their morphospaces. Westermann morphospace therefore
 165 becomes a useful framework to assess the link between shell form and postmortem transport
 166 potential.

Given the ongoing debate about the frequency of postmortem transport in both fossil and modern shelled cephalopods and the importance of this question to our understanding of the geographic distributions of these groups, the present research has three goals:

(1) To determine the relationship between cephalopod shell form and postmortem transport potential,

(2) To test the hypothesis that fossil cephalopods with higher postmortem transport potential will show artificially larger geographic ranges than cephalopods with lower transport potential,

(3) To assess the extent of postmortem transport in modern nautilid species.

Ultimately, the intention of this work is to enable fossil cephalopod workers to use shell forms to better predict the likelihood of postmortem transport and to evaluate their confidence in the use of geographic ranges derived from fossil localities as a proxy for the living geographic ranges of the groups under study. In addition, analysis of the geographic distribution of modern living and dead drift nautilid shells will help researchers better evaluate the geospatial extent of these charismatic but threatened species.

Materials & Methods

Datasets

Four datasets were compiled for morphometric and geographic analyses. All datasets are available as Supplementary Material.

(1) Shell shape data for taxa included in Reyment (1973) plus modern nautilids.

Unfortunately, Reyment did not include the raw shell shape data upon which he performed the principal coordinates analysis shown in Fig. 2. Therefore, for each of the genera he included,

190 images of fossil specimens were located in the ammonoid Treatise volumes (Arkell et al., 1957;
 191 Wright, Callomon & Howarth, 1996). In some cases, the images were of the same species
 192 Reyment used, while in other cases a congeneric species had to be used, based on the availability
 193 and quality of preservation of the specimens. These images were used to measure the five shell
 194 shape characters necessary for determining a taxon's position in Westermann Morphospace
 195 (Ritterbush & Bottjer, 2012): shell diameter (D), width of last whorl (b), height of last whorl (a),
 196 height of whorl 180° back from last whorl (a'), and umbilical diameter (UD). In total, 38 species
 197 were included in this dataset. Reyment's assessment of the degree of postmortem buoyancy for
 198 each taxon (Fig. 2) was used to assign each species to one of three categories: buoyant,
 199 intermediate, or non-buoyant.

200 For comparison, shell shape measurements were also made on examples of four living
 201 nautilid taxa: *Nautilus belauensis* Saunders, 1981, *N. macromphalus* Sowerby, 1848, *N.*
 202 *pompilius* Linnaeus, 1758, and *Allonautilus scrobiculatus* Ward & Saunders, 1997. Shell
 203 photographs were accessed via the website Conchology, Inc. (2017). These four taxa are known
 204 to sometimes drift postmortem and were therefore classified as having shell forms that are
 205 buoyant postmortem.

206 (2) Shell shape data from Ritterbush & Bottjer's (2012) ammonoid dataset. In the
 207 supplementary materials for their 2012 paper, Ritterbush and Bottjer supplied the scaled and
 208 normalized Westermann Morphospace parameters U, Th, and w for 177 ammonoid species.
 209 These species represent a broad range of ages, clades, and shell forms.

210 (3) Geographic range and shell shape data for Late Cenomanian and Early Turonian (Late
 211 Cretaceous) cephalopods from Yacobucci (2017). Yacobucci (2017) compiled geographic range
 212 data for 41 Late Cenomanian and 37 Early Turonian ammonoid genera plus the nautilid genus

Eutrephoceras Hyatt, 1894 (which was present in both substages). This dataset therefore offers the opportunity to test whether certain ammonoid shell forms are more likely to show artificially larger geographic ranges due to postmortem drift. Geographic range was estimated as the $\log_{10}$ of the area in square kilometers of a convex hull encompassing all occurrences of each taxon; see Yacobucci (2017) for further details. To determine the position of these genera in Westermann Morphospace, images of specimens of each genus were located in the ammonoid Treatise volumes (Arkell et al., 1957; Wright, Callomon & Howarth, 1996) and the nautiloid Treatise volume (Teichert et al., 1964) for *Eutrephoceras*. These images were used to measure the necessary shell shape characters.

(4) Modern nautilid occurrences from Toriyama et al. (1965) and House (1987). House (1987) compiled descriptive locality information and produced a map for occurrences of both living specimens and dead drift shells of modern nautilids, based partly on occurrences reported in Toriyama et al. (1965). Localities were compiled from both sources and species and place names updated as needed. Each location was then georeferenced to the nearest 0.1 decimal degrees latitude and longitude, using Google Earth. In some cases, a single numbered locality was split into multiple locations, as House (1987) lumped together in his locality descriptions places that are actually relatively far apart (e.g., two separate islands within one island chain). A total of 184 separate nautilid occurrences were included in the final dataset.

Westermann Morphospace and postmortem buoyancy

Five shell shape measurements (D, b, a, a', and UD) were used to calculate the three parameters defining Westermann Morphospace, following the protocol of Ritterbush & Bottjer (2012). First, raw values of the three key shape parameters of involution (umbilical ratio U), shell inflation (thickness ratio Th), and whorl expansion rate (w) were calculated:

$$\text{Raw } U = UD/D$$

$$\text{Raw } Th = b/D$$

$$\text{Raw } w = a/a'$$

Next, these three raw parameters were scaled to fall within the range of values for common ammonoids, using the minimum and maximum values provided in Ritterbush and Bottjer (2012, table 2):

$$\text{Scaled } U = \text{Raw } U / 0.52$$

$$\text{Scaled } Th = (\text{Raw } Th - 0.14) / (0.68 - 0.14)$$

$$\text{Scaled } w = (\text{Raw } w - 1.00) / (1.77 - 1.00)$$

Finally, the scaled values were normalized to range from 0 to 1:

$$U = \text{Scaled } U / (\text{Scaled } U + \text{Scaled } Th + \text{Scaled } w)$$

$$Th = \text{Scaled } Th / (\text{Scaled } U + \text{Scaled } Th + \text{Scaled } w)$$

$$w = \text{Scaled } w / (\text{Scaled } U + \text{Scaled } Th + \text{Scaled } w)$$

These normalized parameters were used to construct ternary plots of Westermann Morphospace. Each corner of the ternary plot represents the maximum end member value for one of the three shell shape parameters. Individual shell forms plot within the triangular morphospace based on the values of these three parameters.

Analysis of relationship between postmortem buoyancy and geographic range

Two approaches were used to assess the prediction that taxa with more buoyant postmortem shell forms would be more likely to drift and therefore have larger geographic ranges. First, the parameter Th was determined to be predictive of whether a taxon was likely to be buoyant or non-buoyant postmortem (see Results below, Fig. 4). Correlations between Th and geographic range were calculated separately for Late Cenomanian and Early Turonian genera

and tested for significance (Fig. 5). Second, the distributions of geographic ranges for buoyant vs. non-buoyant genera were visualized and Mann-Whitney U-tests used to test for significant differences in the medians of these distributions (Fig. 6). All statistical tests were conducted in PAST 3.15 (Hammer, Harper & Ryan, 2001).

Geospatial analysis of modern nautilid occurrences

ArcGIS 10.3.1 (ESRI, 2015) was used to create maps of modern nautilid occurrences and to calculate convex hulls spanning the occurrences for each species.

Results

Westermann Morphospace

The Mesozoic ammonoids whose buoyancy Reymont (1973) investigated plot separately in Westermann morphospace based on their postmortem buoyancy potential (Fig. 4A). The least buoyant taxa (orange circles) and most buoyant taxa (blue circles) mostly fall into separate fields, divided by the line corresponding to a normalized thickness ratio Th ($100 \times \text{whorl width/shell diameter}$) of about 30%. Just two buoyant species fell in the non-buoyant field, and two non-buoyant species fell in the buoyant field. Fourteen of the 17 (82%) ammonoid taxa that Reymont (1973) identified as having an intermediate buoyancy (gray circles) fall in the non-buoyant area. It should be noted that the majority (66%) of Reymont's ammonoid taxa fall within the non-buoyant area. In contrast, the four living nautilid species (*Nautilus belauensis*, *N. macromphalus*, *N. pompilius*, and *Allonautilus scrobiculatus*; dark blue squares), which are known to experience postmortem transport, plot adjacent to the buoyant ammonoids, but with a higher value of w .

The majority (81%) of the 177 ammonoid taxa used in the original Westermann morphospace paper (Ritterbush & Bottjer, 2012), which represent a broad range of groups and time periods, fall within the non-buoyant field (Fig. 4B, open circles). This result suggests that most ammonoid morphotypes would have had a relatively low potential for postmortem flotation. Only ammonoid taxa with relatively wide, inflated shells would have a high potential for experiencing postmortem transport.

Ammonoid genera (plus the nautilid *Eutrephoceras*) from the Cenomanian-Turonian boundary interval plot in both the non-buoyant and buoyant fields (Fig. 4C, green squares). These results lead to the prediction that genera falling in the buoyant field (that is, with a normalized thickness ratio greater than 30%) may show artificially inflated geographic range sizes relative to those contemporaneous taxa with lower postmortem buoyancy, since more buoyant taxa would have been more likely to be transported by surface currents.

Relationship between predicted postmortem buoyancy and geographic range

There is no significant relationship between shell shape and geographic range. Correlations between the normalized thickness ratio and geographic range sizes of Cenomanian-Turonian cephalopods are weak: Early Turonian correlation coefficient $r = 0.161$ ($p = 0.355$) (Fig. 5A); Late Cenomanian correlation coefficient $r = 0.117$ ($p = 0.459$) (Fig. 5B). Hence, the geographic range size of these cephalopod genera is not predictable based on shell form or the postmortem buoyancy inferred from that form, suggested that postmortem transport was not an important factor in controlling observed geographic range.

In addition, no significant difference exists in the distributions of geographic range sizes for cephalopod shells predicted to be postmortem drifters vs. non-drifters. Genera with shell shapes associated with increased postmortem buoyancy (Fig. 6, blue histograms) do not show

larger geographic ranges than less buoyant genera (Fig. 6, orange histograms) in the Late Cenomanian or Early Turonian. Rather, both groups of genera show U-shaped distributions; numerous genera had small geographic ranges regardless of their postmortem transport potential. Mann-Whitney U-tests reveal show no significant differences in the medians of these distributions (Table 1). It seems clear that even if a cephalopod taxon has a shell shape predicted to be more buoyant postmortem, that potential did not in practice result in more postmortem transport and an inflated geographic range.

Modern nautilid geographic distributions

It is interesting to note that Reyment's own buoyancy experiments (Reyment, 1973) showed that fossil cephalopods had a range of postmortem transport potentials, contradicting his persistent claim that most cephalopods experienced extensive transport (Reyment, 2008). The underlying rationale for Reyment's insistence that postmortem drift in shelled cephalopods is the norm was his claim that modern nautilids are known to frequently experience postmortem drift across long distances. This claim is based on occurrences of living nautilid specimens and drift shells reported in Toriyama et al. (1965) and House (1987). As noted in the Introduction, while drift has been observed, actual evidence for long-distance transport of nautilid shells is actually relatively rare (Wani et al., 2005).

Living populations of modern nautilids are known from the western equatorial Pacific, northern Australia, New Guinea, Indonesia, Palau, and the Philippines (Fig. 7A, orange circles). Drift shells, on the other hand, have been found over a much wider area, including further north in the Pacific, around southern Australia, and across the Indian Ocean (Fig. 7A, brown circles). Pooling all nautilid species together makes it appear that nautilids are prone to extensive postmortem transport.

However, investigating the geospatial distributions of individual nautilid species reveals a more complex pattern. Most nautilid species (*N. belauensis*, *N. macromphalus*, *N. stenomphalus* Sowerby, 1848, and *Allonautilus scrobiculatus*) have relatively small living geographic ranges (Fig. 7B, Table 2). Drift shells of these species (darker circles) are rare and remain relatively close to the living population from which they are derived. For example, the one reported location of drifted *N. stenomphalus* falls within the known living range of the species. *N. belauensis* is known only from Palau, while drift shells have been recovered about 1,100 km away in Mindanao, Philippines. *N. macromphalus* lives in the area of New Caledonia, while drift shells have been found 2,100 km away in southeast Australia. While these distances are not trivial, they also do not extend the known range of these species to new ocean basins.

The exception to this pattern is *N. pompilius*, which has a much larger living range than the other nautilid species as well as drifted shells observed across a large part of the Indo-Pacific (Table 2, Fig. 7B, green circles). House (1987) and Reymont (2008) claimed that drifted shells in the western Indian Ocean must have derived from living populations in the Philippines, implying travel of dead shells over distances of over 9,000 km. It is not clear, however, why *N. pompilius* would be so much more capable of long-distance postmortem transport compared to the other living nautilid species, which are quite similar in their habitats, shell forms, and postmortem buoyancy (Saunders, 1987; Saunders & Ward, 1987).

Discussion

Postmortem drift in fossil cephalopods

The results presented here support the claim that the majority of ammonoid taxa (including both Paleozoic and Mesozoic groups) would have been considerably less likely to

experience postmortem transport than modern nautilids. Only the subset of ammonoid taxa with relatively inflated shells would experience sufficient postmortem buoyancy to float for any length of time, a prerequisite for extensive transport away from their living habitat. Based on evidence from Late Cretaceous ammonoids, genera with shell forms conducive to postmortem floating did not have larger geographic ranges, indicating that even those fossil cephalopods with a greater chance of postmortem transport did not likely actually experience extensive movement away from their living habitat. These results are consistent with previous workers' arguments for the rarity of extensive postmortem transport of fossil cephalopods, based on taphonomic evidence (Kennedy & Cobban, 1976; Hewitt, 1988; Maeda & Seilacher, 1996; Maeda et al., 2003; Wani, 2004; Wani et al., 2005; Mapes et al., 2010a; Lukeneder, 2015) and buoyancy considerations (Chamberlain, Ward & Weaver, 1981; Hewitt & Westermann, 1996; Chirat, 2000; Wani & Gupta, 2015). They also provide a measure of confidence that fossil locality data are a robust proxy for living biogeographic ranges.

Implications for modern nautilid distributions

The notion that postmortem transport was common in fossil cephalopods ultimately derives from the claim that modern nautilids show frequent and extensive postmortem drift. However, drifted nautilid shells are actually fairly rare (Wani et al., 2005) and only one extant nautilid species, *N. pompilius*, shows widely distributed drift shells. The limited geographic distributions of most nautilid species are consistent with evidence for genetic divergence of geographically separated populations (Wray et al., 1995; Bonnaud, Ozouf-Costaz & Boucher-Rodoni, 2004; Sinclair et al., 2007, 2011; Bonacum et al., 2011; Williams et al., 2015; Vandepas et al., 2016).

The abundance of drifted *N. pompilius* shells in the western Indian Ocean is particularly remarkable, as these sites are quite distant from known living populations (Fig. 7B). Both House (1987) and Reyment (2008) argued that the *N. pompilius* shells that wash up in some numbers on beaches in Kenya and Mozambique must have drifted across the entire Indian Ocean from living populations in the Philippines. As noted above, however, it is difficult to understand why this one species of nautilid would show such different postmortem behavior than its close relatives. As an alternative, it might be possible that these African drift shells are actually being sourced from living populations much closer, in the western Indian Ocean basin (House, 1973; Wani et al., 2005; Matteucci, 2015). Some live *Nautilus* (presumed to be *N. pompilius*) have been reported from Madagascar (Teichert, 1970) and from Mauritius and the Seychelles (by Dr. Anna Bidder, cited in Reyment (1973)), and late Pleistocene fossils of *Nautilus* have been identified on the Bajuni Islands off the Somali coast (Matteucci, 2015). The distances from the isolated islands of the western Indian Ocean to the African mainland are more consistent with the distances traveled by dead shells of the other nautilid species. The possibility of living *N. pompilius* populations within the western Indian Ocean basin is therefore worthy of further investigation, especially in light of new conservation efforts for this iconic marine animal. Such new discoveries are not without precedent. A second species of coelacanth, *L. menadoensis* Pouyaud, Wirjoatmodjo, Rachmatika, Tjakrawidjaja, Hadiaty & Hadie 1999, was identified in Indonesia in 1997, nearly 10,000 km east across the Indian Ocean from the coelacanths' known range in east Africa (Pouyaud et al., 1999). Genetic data suggest these two species have been separated for tens of million years (Holder et al., 1999; Inoue et al., 2005). It seems clear that the marine biota of the Indian Ocean, especially deeper water species like the nautilus and the coelacanth, is still incompletely known.

395

396 **Conclusions**

397 While postmortem transport of cephalopod shells in modern and fossil contexts is
 398 frequently assumed to have been both frequent and extensive, evidence supporting this view is
 399 lacking. Most fossil ammonoids had shell forms that resulted in low postmortem buoyancy; only
 400 highly inflated shells were likely to float for significant periods after death. An analysis of early
 401 Late Cretaceous (Cenomanian-Turonian) cephalopod genera shows that observed geographic
 402 range size was not related to likelihood of postmortem transport, indicating that these observed
 403 fossil ranges are adequate proxies for living geographic ranges. Most living nautilid species have
 404 relatively small geographic ranges with limited dispersal of drift shells. The exception, *Nautilus*
 405 *pompilius*, is widespread throughout the Indo-Pacific, with drift shells found apparently great
 406 distances from known living populations. However, drift shells found along the east African
 407 coast may in reality be derived from cryptic living populations among the isolated islands of the
 408 western Indian Ocean.

409

410 **Acknowledgements**

411 The author would like to thank R. Hoffmann, N. Landman, B. Linzmeier, R. Mapes, and K.
 412 Ritterbush for helpful discussions, and A. Avruch for constructive comments on the manuscript.

413

414 **References**

415 Arkell WJ, Furnish WM, Kummel B, Miller AK, Moore RC, Schindewolf OH, Sylvester-
 416 Bradley PC, Wright CW. 1957. *Treatise on Invertebrate Paleontology, Part L, Mollusca 4*.

Cephalopoda, Ammonoidea. Boulder, Colorado and Lawrence, Kansas: Geological Society of America and University of Kansas.

Barord GJ, Dooley F, Dunstan A, Ilano A, Keister KN, Neumeister H, Preuss T, Schoepfer S, Ward PD. 2014. Comparative population assessments of *Nautilus* sp. in the Philippines, Australia, Fiji, and American Samoa using baited remote underwater video systems. PLoS ONE 9(6): e100799. DOI: 10.1371/journal.pone.0100799.

Bonacum J, Landman NH, Mapes RH, White MM, White A-J, Irlam J. 2011. Evolutionary radiation of present-day *Nautilus* and *Allonautilus*. *American Malacological Bulletin* 29:77-93. DOI: 10.4003/006.029.0221.

Bonnaud L, Ozouf-Costaz C, Boucher-Rodoni R. 2004. A molecular and karyological approach to the taxonomy of *Nautilus*. *Comptes Rendus Biologies* 327:133-138. DOI: 10.1016/j.crv.2003.12.004.

Brayard, A. Escarguel G, Monnet C, Jenks, JF, Bucher, H. 2015. Biogeography of Triassic ammonoids. In: Klug C, Korn D, De Baets K, Kruta I, Mapes RH, eds. *Ammonoid paleobiology: from macroevolution to paleogeography*. Topics in Geobiology 44:163-187. Dordrecht: Springer. DOI: 10.1007/978-94-017-9633-0_7.

Bronn HG. 1838. Lethaea Geognostica oder Abbildungen und Beschreibungen der für die Gebirge-Formationen bezeichnendsten Versteinerungen 2:545-1346.

Cecca F. 2002. Palaeobiogeography of marine fossil invertebrates: concepts and methods. London: Taylor and Francis.

Chamberlain JA Jr, Ward PD, Weaver JS. 1981. Post-mortem ascent of *Nautilus* shells: implications for cephalopod paleobiogeography. *Paleobiology* 7:494-509. DOI: 10.1017/S0094837300025549.

440 Chamberlain JA Jr, Weaver JS. 1978. Equations of motion for post-mortem sinking of
441 cephalopod shells. *Mathematical Geology* 10:673-689. DOI: 10.1007/BF01031898

442 Chirat R. 2000. The so-called 'cosmopolitan palaeobiogeographic distribution' of Tertiary
443 Nautilida of the genus *Aturia* Bronn 1838: the result of post-mortem transport by oceanic
444 palaeocurrents. *Palaeogeography, Palaeoclimatology, Palaeoecology* 157:59-77. DOI:
445 10.1016/S0031-0182(99)00150-9.

446 Conchology, Inc. 2017. Available at <https://www.conchology.be> (accessed 22 June 2017).

447 De Angelis P. 2012. Assessing the impact of international trade on chambered nautilus. *Geobios*
448 45:5-11. DOI: 10.1016/j.geobios.2011.11.005.

449 Denton EJ, Gilpin-Brown JB. 1966. On the buoyancy of the pearly nautilus. *Journal of the*
450 *Marine Biological Association of the U.K.* 46:723-759. DOI: 10.1017/S0025315400033440.

451 Dunstan A, Alanis O, Marshall J. 2010. *Nautilus pompilius* fishing and population decline in the
452 Philippines: a comparison with an unexploited Australian *Nautilus* population. *Fisheries*
453 *Research* 106:239-247. DOI: 10.1016/j.fishres.2010.06.015.

454 Dunstan A, Bradshaw CJA, Marshall J. 2011. *Nautilus* at risk-estimating population size and
455 demography of *Nautilus pompilius*. *PLoS One* 6(2):e16716. DOI:
456 10.1371/journal.pone.0016716.

457 Dunstan AJ, Ward PD, Marshall NJ. 2011a. *Nautilus pompilius* life history and demographics at
458 the Osprey Reef Seamount, Coral Sea, Australia. *PLoS One* 6(2):e16312. DOI:
459 10.1371/journal.pone.0016312.

460 Dunstan AJ, Ward PD, Marshall NJ. 2011b. Vertical distribution and migration patterns of
461 *Nautilus pompilius*. *PLoS One* 6(2):e16311. DOI: 10.1371/journal.pone.0016311.

462 ESRI (Environmental Systems Research Institute). 2015. ArcGIS Desktop, Release 10.3.1.
 463 Redlands, California: ESRI.

464 ESRI (Environmental Systems Research Institute). 2016. Ocean Basemap. *Available at*
 465 <https://www.arcgis.com/home/item.html?id=5ae9e138a17842688b0b79283a4353f6>
 466 (accessed 15 September 2017).

467 Greenwald L, Ward PD. 1987. Buoyancy in *Nautilus*. In: Saunders WB, Landman NH, eds.
 468 *Nautilus: the biology and paleobiology of a living fossil*. Topics in Geobiology 6:547-560.
 469 New York: Plenum Press.

470 Hamada T. 1964. Notes on drifted *Nautilus* in Thailand. *Scientific Papers of the College of*
 471 *General Education, University of Tokyo* 14:255-277.

472 Hammer Ø, Bucher H. 2006. Generalized ammonoid hydrostatics modelling, with application to
 473 *Intornites* and intraspecific variation in *Amaltheus*. *Paleontological Research* 10:91-96. DOI:
 474 10.2517/prpsj.10.91.

475 Hammer Ø, Harper DAT, Ryan PD. 2001. PAST: Paleontological statistics software package for
 476 education and data analysis. *Palaeontologia Electronica* 4(1):article 4.

477 Hembree DI, Mapes RH, Goiran C. 2014. The impact of high-energy storms on shallow-water
 478 *Nautilus* (Cephalopoda) taphonomy, Lifou (Loyalty Islands). *Palaios* 29:348-362. DOI:
 479 10.2110/palo.2013.113.

480 Hewitt RA. 1988. Nautiloid shell taphonomy: interpretations based on water pressure.
 481 *Palaeogeography, Palaeoclimatology, Palaeoecology* 63:15-25. DOI: 10.1016/0031-
 482 0182(88)90088-0.

483 Hewitt RA, Westermann GEG. 1996. Post-mortem behaviour of Early Paleozoic nautiloids and
 484 paleobathymetry. *Paläontologische Zeitschrift* 70:405-424. DOI: 10.1007/BF02988081.

Hoffmann R, Lemanis R, Naglik C, Klug C. 2015. Ammonoid buoyancy. In: Klug C, Korn D, De Baets K, Kruta I, Mapes RH, eds. *Ammonoid paleobiology: from anatomy to ecology*. Topics in Geobiology 43:613-648. Dordrecht: Springer. DOI: 10.1007/978-94-017-9630-9_16.

Hoffmann R, Zachow S. 2011. Non-invasive approach to shed new light on the buoyancy business of chambered cephalopods. *IAMG 2011 Publication*. DOI: 10.5242/iamg.2011.0163.

Holder MT, Erdmann MV, Wilcox TP, Caldwell RL, Hillis DM. 1999. Two living species of coelacanths? *Proceedings of the National Academy, U.S.A.* 96:12616-12620.

House MR. 1973. An analysis of Devonian goniatite distributions. In: Hughes NF, eds. *Organisms and continents through time*. Special Papers in Palaeontology 12:305-317.

House MR. 1987. Geographic distribution of *Nautilus* shells. In: Saunders WB, Landman NH, eds. *Nautilus: the biology and paleobiology of a living fossil*. Topics in Geobiology 6:53-64. New York: Plenum Press.

Hyatt A. 1894. Phylogeny of an acquired characteristic. *Proceedings of the American Philosophical Society* 32(143):349-647.

Ifrim C, Lehmann J, Ward P. 2015. Paleobiogeography of Late Cretaceous ammonoids. In: Klug C, Korn D, De Baets K, Kruta I, Mapes RH, eds. *Ammonoid paleobiology: from macroevolution to paleogeography*. Topics in Geobiology 44:259-274. Dordrecht: Springer. DOI: 10.1007/978-94-017-9633-0_10.

Inoue JG, Miya M, Venkatesh B, Nishida M. 2005. The mitochondrial genome of Indonesian coelacanth *Latimeria menadoensis* (Sarcopterygii: Coelacanthiformes) and divergence time

estimation between the two coelacanths. *Gene* 349:227-235. DOI:

10.1016/j.gen.2005.01.008.

Iredale T. 1944. Australian pearly *Nautilus*. *Australian Zoologist* 10:294-298.

Jacobs DK, Chamberlain JA Jr. 1996. Buoyancy and hydrodynamics in ammonoids. In:

Landman NH, Tanabe K, Davis RA, eds. *Ammonoid paleobiology*. Topics in Geobiology 13.

Plenum, New York.

Kennedy WJ, Cobban WA. 1976. *Aspects of ammonite biology, biogeography, and*

biostratigraphy. Special Papers in Palaeontology 17. London: Palaeontological Association.

Kidwell SM, Flessa KW. 1996. The quality of the fossil record: Populations, species, and

communities. *Annual Review of Earth and Planetary Sciences* 24:433-464. DOI:

10.1146/annurev.earth.24.1.433.

Kobayashi T. 1954. A contribution towards Palaeo-Flumenology, science of the oceanic currents

of the past, with a description of a new Miocene *Aturia* from central Japan. *Japanese Journal*

of Geology and Geography 25:35-56.

Korn D, De Baets K. 2015. Biogeography of Paleozoic ammonoids. In: Klug C, Korn D, De

Baets K, Kruta I, Mapes RH, eds. *Ammonoid paleobiology: from macroevolution to*

paleogeography. Topics in Geobiology 44:145-161. Dordrecht: Springer. DOI: 10.1007/978-

94-017-9633-0_6.

Kröger B. 2002. On the efficiency of the buoyancy apparatus in ammonoids: evidences from

sublethal shell injuries. *Lethaia* 35:61-70. DOI: 10.1111/j.1502-3931.2002.tb00068.x.

Lehmann J, Ifrim C, Bulot L, Frau C. 2015. Paleobiogeography of Early Cretaceous ammonoids.

In: Klug C, Korn D, De Baets K, Kruta I, Mapes RH, eds. *Ammonoid paleobiology: from*

macroevolution to paleogeography. Topics in Geobiology 44:229-257. Dordrecht: Springer.
DOI: 10.1007/978-94-017-9633-0_9.

Lemanis R, Zachow S, Füsseis F, Hoffmann R. 2015. A new approach using high-resolution
computed tomography to test the buoyant properties of chambered cephalopod shells.
Paleobiology 41:313-329. DOI: 10.1017/pab.2014.17.

Linnaeus, C. 1758. *Systema Naturae*, 10th edition. Stockholm: Holmiae.

Lukeneder A. 2015. Ammonoid habitats and life history. In: Klug C, Korn D, De Baets K, Kruta
I, Mapes RH, eds. *Ammonoid paleobiology: from anatomy to ecology*. Topics in Geobiology
43:689-791. Dordrecht: Springer. DOI: 10.1007/978-94-017-9630-9_18.

Maeda H, Mapes RH, Mapes G. 2003. Taphonomic features of a Lower Permian beached
cephalopod assemblage from Central Texas. *Palaaios* 18:421-434. DOI: 10.1669/0883-
1351(2003)018<0421:TFOALP>2.0.CO;2.

Mapes RH, Landman NH, Cochran K, Goiran C, Richer de Forges B, Renfro A. 2010a. Early
taphonomy and significance of naturally submerged *Nautilus* shells from the New Caledonia
region. *Palaaios* 25:597-610. DOI: 10.2110/palo.2009.p09-109r.

Mapes RH, Hembree DI, Rasor BA, Stigall A, Goiran C, Richer de Forges B. 2010b. Modern
Nautilus (Cephalopoda) taphonomy in a subtidal to backshore environment, Lifou (Loyalty
Islands). *Palaaios* 25:656-670. DOI: 10.2110/palo.2010.p10-010r.

Matteucci R. 2015. Drifted *Nautilus* shells from the Bajuni Islands (southern Somali coast of
Indian Ocean). *Journal of Mediterranean Earth Sciences* 7:35-50. DOI:
3304/JMES.2015.004.

- Naglik C, Rikhtegar F, Klug C. 2014. Buoyancy of some Palaeozoic ammonoids and their hydrostatic properties based on empirical 3D-models. *Lethaia* 49:3-12. DOI: 10.1111/let.12125.
- Naglik C, Tajika A, Chamberlain J, Klug C. 2015. Ammonoid locomotion. In: Klug C, Korn D, De Baets K, Kruta I, Mapes RH, eds. *Ammonoid paleobiology: from anatomy to ecology*. Topics in Geobiology 43:649-688. Dordrecht: Springer. DOI: 10.1007/978-94-017-9630-9_17.
- Peterman D, Barton CC, Yacobucci MM. In review, 2018. The hydrostatics of Paleozoic ectocochleate cephalopods (Nautiloidea and Endoceratoidea) with implications for modes of life and early colonization of the pelagic zone. Submitted to *Palaeontologia Electronica*.
- Pouyaud L, Wirjoatmodjo S, Rachmatika I, Tjakrawidjaja A, Hadiaty R, Hadie W. 1999. Une nouvelle espèce de coelacanth. Preuves génétiques et morphologiques. *Comptes Rendus de l'Académie des Sciences, Series III, Sciences de la Vie* 322:261-267.
- Reid A. 2016. Post-mortem drift in Australian cuttlefish sepions: its effect on the interpretation of species ranges. *Molluscan Research* 36:9-21. DOI: 10.1080/13235818.2015.1064366.
- Reyment RA. 1958. Some factors in the distribution of fossil cephalopods. *Stockholm Contributions in Geology* 1:97-184 + 7 plates.
- Reyment RA. 1970. Vertically inbedded cephalopod shells. Some factors in the distribution of fossil cephalopods, 2. *Palaeogeography, Palaeoclimatology, Palaeoecology* 7:103-111. DOI: 10.1016/0031-0182(70)90071-4.
- Reyment RA. 1973. Factors in the distribution of fossil cephalopods. Part 3: Experiments with exact models of certain shell types. *Bulletin of the Geological Institutions of the University of Uppsala, N.S.* 4:7-41.

573 Reyment RA. 2008. A review of the post-mortem dispersal of cephalopod shells. *Palaeontologia*
574 *Electronica* 11:12A. DOI:

575 Ritterbush KA, Bottjer DJ. 2012. Westermann Morphospace displays ammonoid shell shape and
576 hypothetical paleoecology. *Paleobiology* 38:424-446. DOI: 10.1666/10027.1.

577 Roux M. 1990. Underwater observations of *Nautilus macromphalus* off New Caledonia.
578 *Chambered Nautilus Newsletter* 60:1.

579 Roux M, Bouchet P, Bourseau JP, Gaillard C, Grandperrin R, Guille A, Laurin B, Monniot C,
580 Richer de Forges B, Rio M, Segonzac M, Vacelet J, Zibrowius H. 1991. L'environnement
581 bathyal au large de la Nouvelle-Calédonie: Résultats préliminaires de la campagne CALSUB
582 et conséquences paléoécologiques. *Bulletin, Société Géologique de France* 162:675-685.

583 Saunders WB. 1981. A new species of *Nautilus* from Palau. *Veliger* 24:1-7.

584 Saunders WB. 1987. The species of *Nautilus*. In: Saunders WB, Landman NH, eds. *Nautilus: the*
585 *biology and paleobiology of a living fossil*. Topics in Geobiology 6:35-52. New York:
586 Plenum Press.

587 Saunders WB, Ward PD. 1987. Ecology, distribution, and population characteristics of *Nautilus*.
588 In: Saunders WB, Landman NH, eds. *Nautilus: the biology and paleobiology of a living*
589 *fossil*. Topics in Geobiology 6:137-162. New York: Plenum Press.

590 Saunders WB, Greenfest-Allen E, Ward PD. 2017. Demographic disequilibrium in living
591 nautiloids (*Nautilus* and *Allonautilus*): Canary in the coal mines? PLoS ONE
592 12(7):e0179811. DOI: 10.1371/journal.pone.0179811.

593 Seuss B, Hembree DI, Wisshak M, Mapes RH, Landman NH. 2015. Taphonomy of backshore
594 versus deep-marine collected *Nautilus macromphalus* conchs (New Caledonia). *Palaios*
595 30:503-513. DOI: 10.2110/palo.2014.057.

596 Sinclair W, Briskey L, Aspden W, Pegg GG. 2007. Genetic diversity of isolated populations of
597 *Nautilus pompilius* in the Coral Sea. *Reviews in Fish Biology and Fisheries* 17:223-235.
598 DOI: 10.1007/s11160-006-9030-x.

599 Sinclair W, Newman SJ, Vianna GS, Williams S, Aspden WJ. 2011. Spatial subdivision and
600 generic diversity in populations on the east and west coasts of Australia: the multi-faceted
601 case of *Nautilus pompilius* (Mollusca, Cephalopoda). *Reviews in Fisheries Science* 19:52-61.
602 DOI: 10.1080/10641262.2010.533794.

603 Sowerby GB. 1848. Monograph of the genus *Nautilus*. *Thesaurus Conchyliorum* 2:463-465.

604 Stenzel HB. 1964. Living *Nautilus*. Pp. K59-K93 in: Moore RC, ed. *Treatise on Invertebrate*
605 *Paleontology, Part K, Mollusca 3. Cephalopoda-General Features-Endoceratoidea-*
606 *Actinoceratoidea-Nautiloidea-Bactritoidea*. Boulder, Colorado and Lawrence, Kansas:
607 Geological Society of America and University of Kansas.

608 Tajika A, Naglik C, Morimoto N, Pascual-Cebrian E, Hennhöfer DK, Klug C. 2014. Empirical
609 3D-model of the conch of the Middle Jurassic ammonite microconch *Normannites*, its
610 buoyancy, the physical effects of its mature modifications and speculations on their function.
611 *Historical Biology* 27:181-191. DOI: 10.1080/08912963.2013.872097.

612 Teichert C. 1970. Drifted *Nautilus* shells in the Bay of Bengal. *Journal of Paleontology* 44:1129-
613 1145.

614 Teichert C, Kummel B, Sweet WC, Stenzel HB, Furnish WM, Glenister BF, Erben HK, Moore
615 RC, Nodine Zeller DE. 1964. *Treatise on Invertebrate Paleontology, Part K, Mollusca 3.*
616 *Cephalopoda-General Features-Endoceratoidea-Actinoceratoidea-Nautiloidea-Bactritoidea.*
617 Boulder, Colorado and Lawrence, Kansas: Geological Society of America and University of
618 Kansas.

Tomašových A, Kidwell SM. 2009. Fidelity of variation in species composition and diversity partitioning by death assemblages: time-averaging transfers diversity from beta to alpha levels. *Paleobiology* 35:94-118. DOI: 10.1666/08024.1.

Tomašových A, Schlögl J, Kaufman DS, Hudáčková N. 2016. Temporal and bathymetric resolution of nautiloid death assemblages in stratigraphically condensed oozes (New Caledonia). *Terra Nova* 28:271-278. DOI: 10.1111/ter.12218.

Tomašových A, Schlögl J, Biroň A, Hudáčková N, Mikuš T. 2017. Taphonomic clock and bathymetric dependence of cephalopod preservation in bathyal, sediment-starved environments. *Palaios* 32:135-152. DOI: 10.2110/palo.2016.039.

Toriyama R, Sato T, Hamada T, Komalarjun P. 1965. *Nautilus pompilius* drifts on the west coast of Thailand. *Japanese Journal of Geology and Geography* 36:149-161.

Trueman AE. 1941. The ammonite body chamber with special reference to the buoyancy and mode of life of the living ammonite. *Quarterly Journal of the Geological Society of London* 96:339-383.

Tyler CL, Kowalewski M. 2017. Surrogate taxa and fossils as reliable proxies of spatial biodiversity patterns in marine benthic communities. *Proceedings of the Royal Society B* 284:20162839. DOI: 10.1098/rspb.2016.2839.

Vandepas LE, Dooley FD, Barord GJ, Swalla BJ, Ward PD. 2016. A revisited phylogeography of *Nautilus pompilius*. *Ecology and Evolution* 6:4924-4935. DOI: 10.1002/ece3.2248.

Walther J. 1897. Über die Lebensweise fossiler Meeresthiere. *Zeitschrift der deutschen geologischen Gesellschaft* 49(2):209-273.

- Wani R. 2004. Experimental fragmentation patterns of modern *Nautilus* shells and the implications for fossil cephalopod taphonomy. *Lethaia* 37:113-123. DOI: 10.1080/00241160410006420.
- Wani R. 2017. Geological duration of ammonoids controlled their geographical range of fossil distribution. *PeerJ* 5:e4108. DOI: 10.7717/peerj.4108.
- Wani R, Gupta NS. 2015. Ammonoid taphonomy. In: Klug C, Korn D, De Baets K, Kruta I, Mapes RH, eds. *Ammonoid paleobiology: from macroevolution to paleogeography*. Topics in Geobiology 44:555-598. Dordrecht: Springer. DOI: 10.1007/978-94-017-9633-0_20.
- Wani R, Kase T, Shigeta Y, De Ocampo R. 2005. New look at ammonoid taphonomy, based on field experiments with modern chambered nautilus. *Geology* 33:849-852. DOI: 10.1131/G21712.1.
- Ward PD, Greenwald L. 1982. Chamber refilling in *Nautilus*. *Journal of the Marine Biological Association of the U.K.* 62:469-475. DOI: 10.1017/S0025315400057404.
- Ward PD, Martin AW. 1978. On the buoyancy of the pearly Nautilus. *Journal of Experimental Zoology* 205:5-12. DOI: 10.1002/jez.1402050103.
- Ward PD, Saunders WB. 1997. *Allonautilus*: a new genus of living nautiloid cephalopod and its bearing on phylogeny of the Nautilida. *Journal of Paleontology* 71:1054-1064. DOI: 10.1017/S0022336000036039.
- Ward P, Dooley F, Barord GJ. 2016. *Nautilus*: biology, systematics, and paleobiology as viewed from 2015. *Swiss Journal of Palaeontology* 135:169-185. DOI: s13358-016-0112-7.
- Westermann GEG. 1971. Form, structure and function of shell and siphuncle in coiled Mesozoic ammonoids. *Life Science Contributions, Royal Ontario Museum* 78:1-39

Williams RC, Jackson BC, Duvaux L, Dawson DA, Burke T, Sinclair W. 2015. The genetic structure of *Nautilus pompilius* populations surrounding Australia and the Philippines. *Molecular Ecology* 24:3316-3328. DOI: 10.1111/mec.13255.

Wray CG, Landman NH, Saunders WB, Bonacum J. 1995. Genetic divergence and geographic diversification in *Nautilus*. *Paleobiology* 21:220-228. DOI: 10.1017/S009483730001321X.

Wright CW, with Callomon JH, Howarth MK. 1996. *Treatise on Invertebrate Paleontology, Part L, Mollusca 4, Revised. Volume 4: Cretaceous Ammonoidea*. Boulder, Colorado and Lawrence, Kansas: Geological Society of America and University of Kansas.

Yacobucci MM. 2015. Macroevolution and paleobiogeography of Jurassic–Cretaceous ammonoids. In: Klug C, Korn D, De Baets K, Kruta I, Mapes RH, eds. *Ammonoid paleobiology: from macroevolution to paleogeography*. Topics in Geobiology 44:189-228. Dordrecht: Springer. DOI: 10.1007/978-94-017-9633-0_8.

Yacobucci MM. 2017. Marine life in a greenhouse world: cephalopod biodiversity and biogeography during the early Late Cretaceous. *Paleobiology* 43:587-619. DOI: 10.1017/pab.2017.3.

Figure 1

Modern *Nautilus*

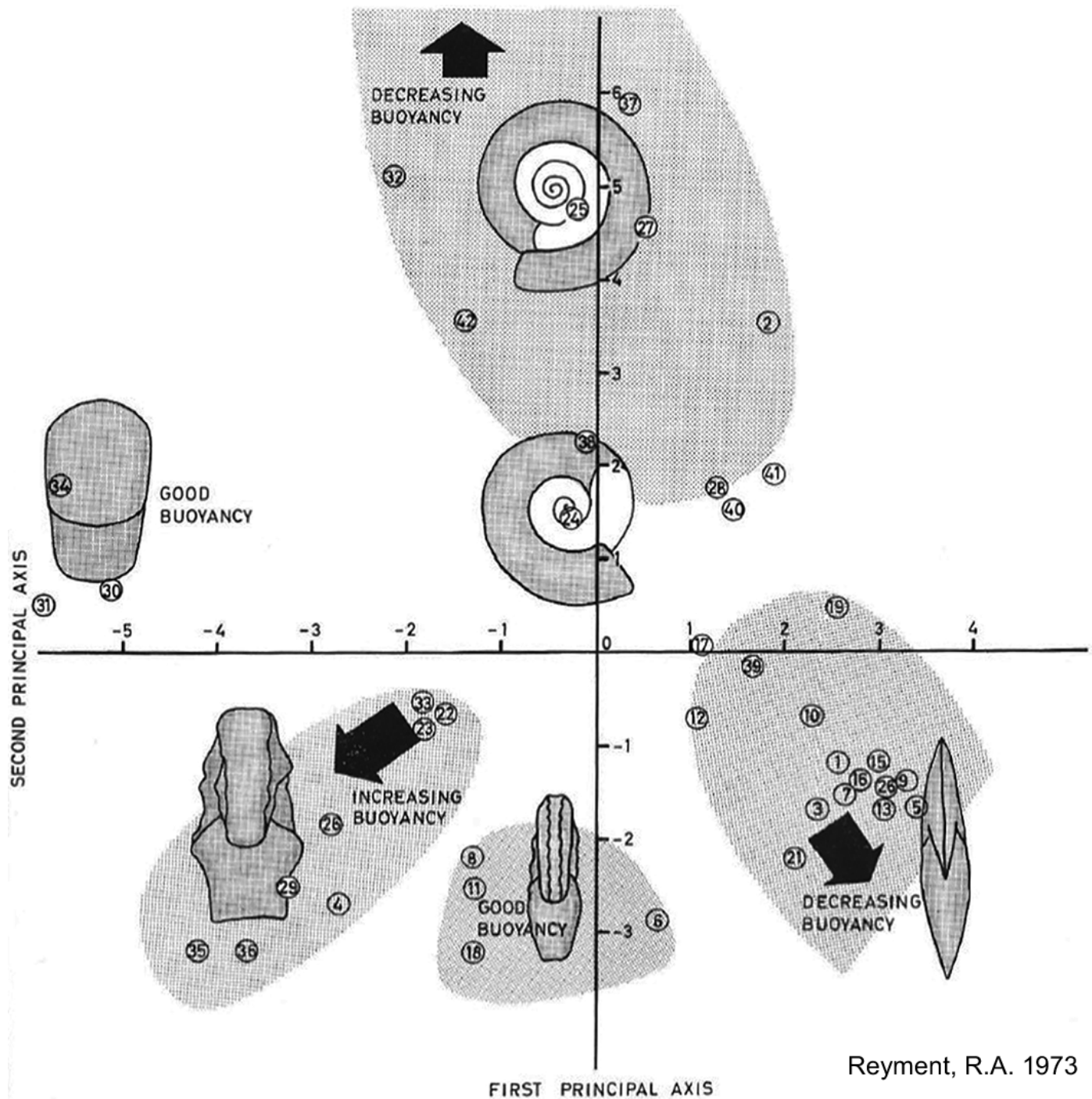
Living *Nautilus* in Palau. Photo source: Public domain image accessed from <https://pxhere.com/en/photo/660195>.



Figure 2

Relationship between shell form and postmortem buoyancy

Plot of first two principal coordinates derived from Reyment's 1973 analysis of five shell parameters on 42 Mesozoic ammonoid species (numbered circles). Each position on the plot reflects a particular shell shape. Reyment indicated predicted postmortem buoyancy of different shell forms on the plot, based on his experimental work. Image source: Reyment (1973), fig. 32, p. 34.



Reyment, R.A. 1973

Figure 3(on next page)

Westermann Morphospace

Schematic representation of Westermann morphospace (Ritterbush & Bottjer, 2012) as a ternary diagram. Each corner of the diagram represents the maximum value for one of the three shell shape parameters used to construct the morphospace: umbilical ratio U (serpenticones), whorl thickness ratio Th (spherocoines), and whorl expansion rate w (oxycones). Any planispiral shell form will plot within the ternary diagram based on its values for these three parameters. Note the similarity of Westermann morphospace to Reyment's (1973) principal coordinates plot (Fig. 2).

U

50

50

W

U

50

Th

serpenticone

spherocone

Th

For more information, please contact the publisher at info@wiley.com



Figure 4(on next page)

Buoyant vs. non-buoyant cephalopods in Westermann Morphospace

(A) Reyment's (1973) taxa (circles) and modern nautilids (dark blue squares) in Westermann Morphospace. Modern nautilids include *Nautilus belauensis* Saunders, 1981, *N. macromphalus* Sowerby, 1848, *N. pompilius* Linnaeus, 1758, and *Allonautilus scrobiculatus* Ward & Saunders, 1997. Color coding indicates predicted postmortem buoyancy: buoyant-blue, intermediate-gray, not buoyant-orange. Light blue line represents a thickness ratio of 30%, generally separating buoyant from non-buoyant shell forms. (B) Addition of ammonoid taxa reported in Ritterbush & Bottjer (2012; open circles). Most ammonoids fall within the non-buoyant postmortem field of morphospace. (C) Addition of Late Cenomanian and Early Turonian cephalopod genera (green squares). These taxa show a mix of shell forms predicted to be buoyant and non-buoyant postmortem.

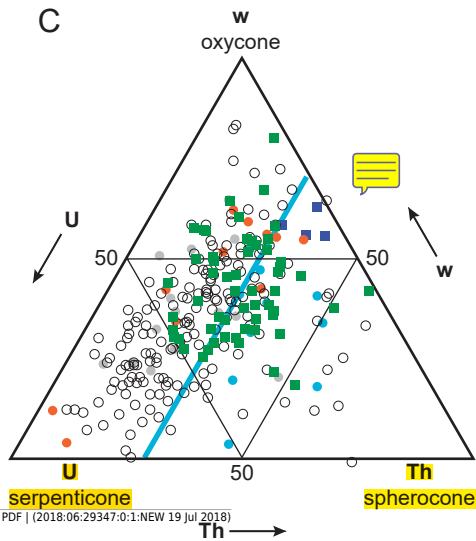
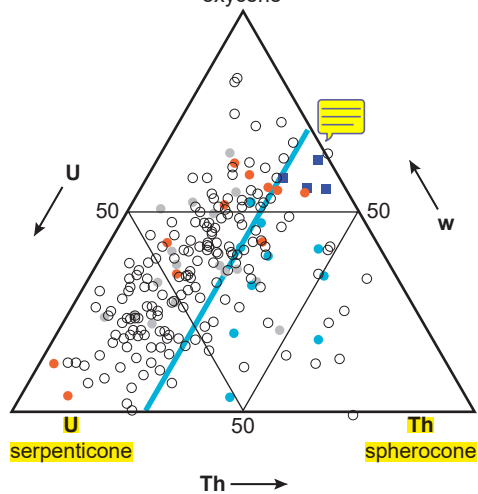
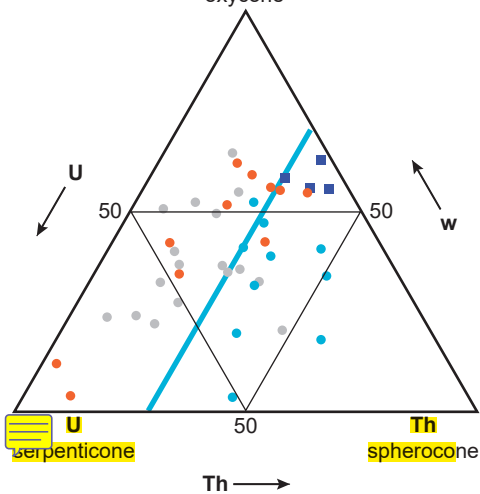
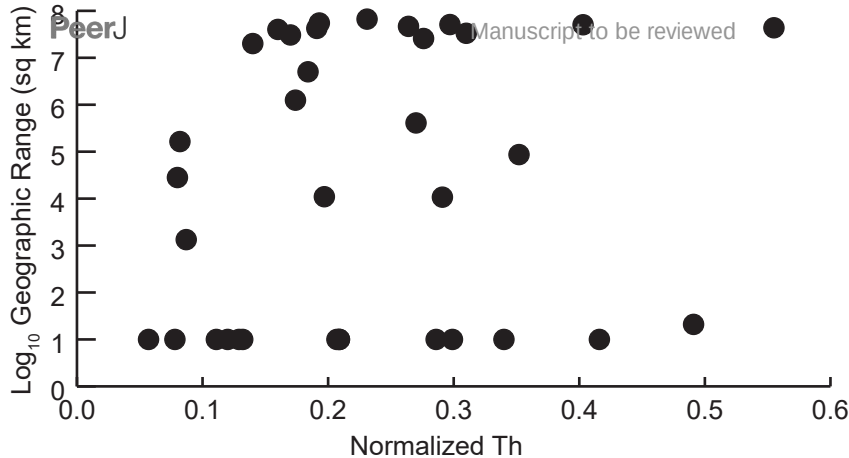


Figure 5(on next page)

Geographic range vs. shell shape

Scatterplots of geographic range (expressed as the $\log_{10}$ of the area in square kilometers spanned by each genus' occurrences) versus normalized thickness ratio (Th) (reflecting degree of shell compression for that genus). (A) Early Turonian genera. Correlation coefficient $r = 0.161$ ($p = 0.355$). (B) Late Cenomanian genera. Correlation coefficient $r = 0.117$ ($p = 0.459$).

A



B

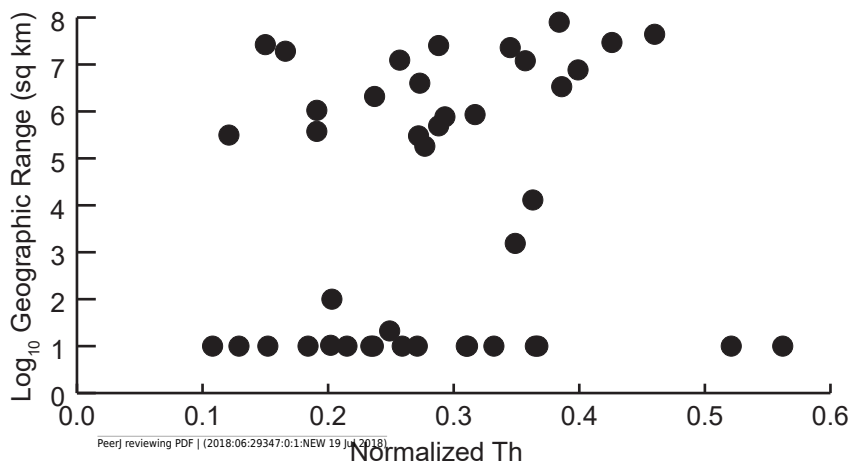
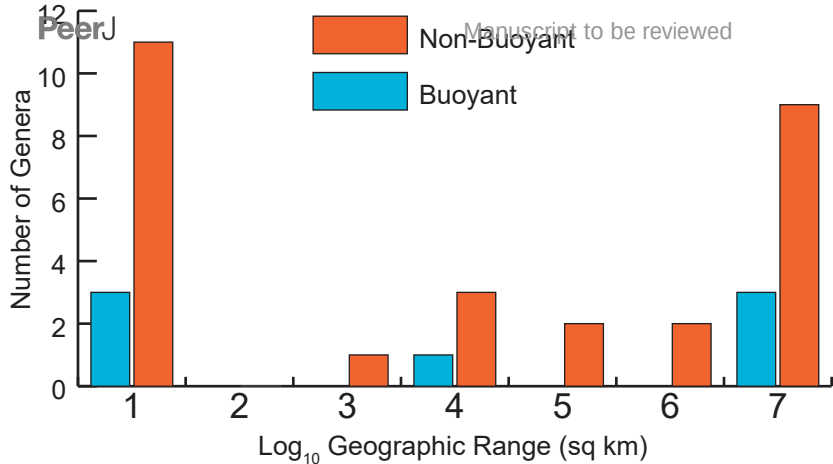


Figure 6(on next page)

Distributions of geographic ranges for taxa with buoyant vs. non-buoyant postmortem shell forms

(A) Early Turonian genera. (B) Late Cenomanian genera. Distributions are not significantly different, implying that taxa with buoyant shell forms more likely to drift postmortem do not, in fact, show larger geographic ranges.

A



B

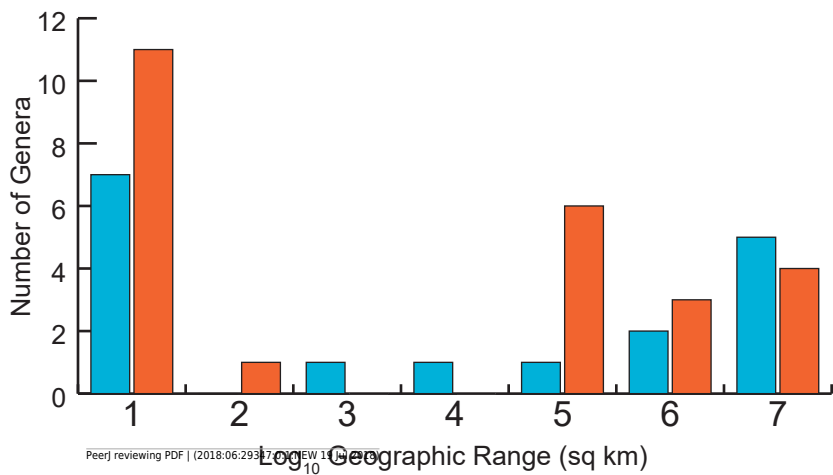


Figure 7 (on next page)

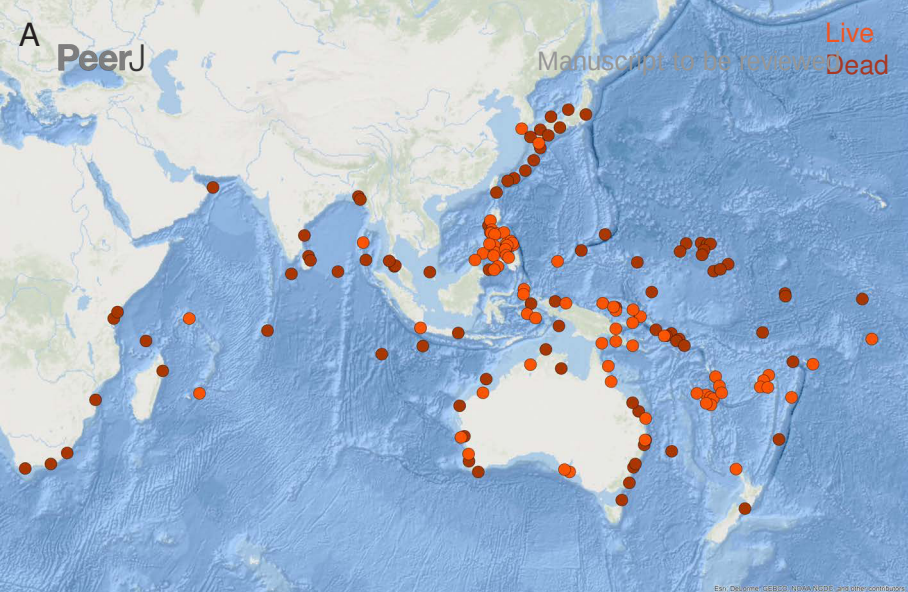
Geographic distributions of modern nautilid species

(A) All modern nautilid occurrences, including live specimens (orange) and dead shells (brown). Drifted shells span a larger area than known living occurrences. (B) Occurrences color-coded by species (live in lighter color, dead in darker color), with convex hulls enclosing all occurrences for each species. Most nautilid species do not have large geographic areas, even including shells that have drifted postmortem. The exception is *N. pompilius*, which has the largest range of drifted shells. The abundance of drifted shells in eastern Africa suggests the possibility of cryptic living populations of *N. pompilius* in the western Indian Ocean. Base map of Indo-Pacific region from ESRI (2016).

A

PeerJ

Manuscript to be reviewed

Live
Dead

B

N. belauensis
N. macromphalus
N. pompilius
N. stenomphalus
A. scrobiculatus

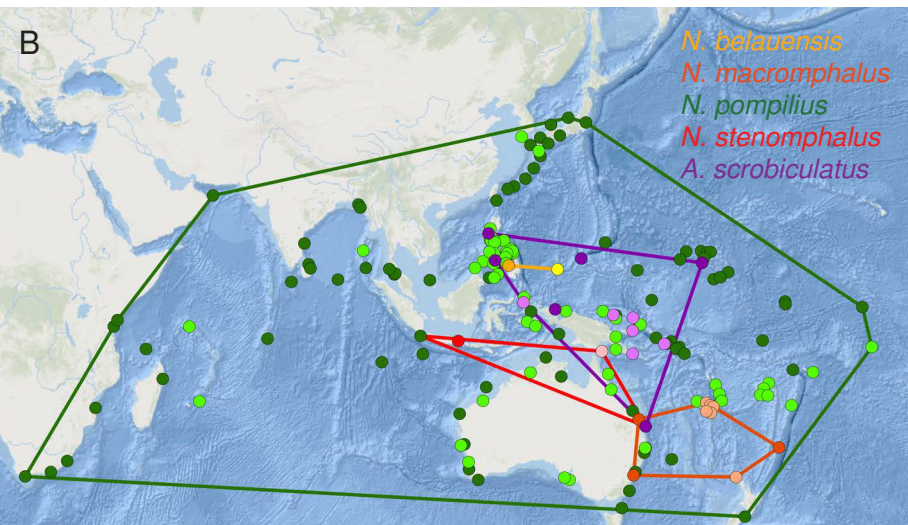


Table 1(on next page)

Comparisons of median geographic range sizes for Late Cenomanian and Early Turonian cephalopod genera

Shells that are buoyant versus not buoyant postmortem are defined as having a normalized thickness ratio of more than versus less than 30% (see Fig. 4).

Table 1: Comparisons of median geographic range sizes for Late Cenomanian and Early Turonian cephalopod genera. Shells that are buoyant versus not buoyant postmortem are defined as having a normalized thickness ratio of more than versus less than 30% (see Fig. 4).

	Median log ₁₀ geographic range (sq km)	Count	p(same median) from Mann- Whitney U-test
Late Cenomanian - Buoyant	4.11	17	0.589
Late Cenomanian - Not Buoyant	5.26	25	
Early Turonian - Buoyant	4.94	7	0.688
Early Turonian - Not Buoyant	4.24	28	

Table 2(on next page)

Geographic range sizes of modern nautilids

Range sizes are represented as the area in square kilometers of convex hulls around nautilid occurrences (Fig. 7B)

Table 2: Geographic range sizes of modern nautilids. Range sizes are represented as the area in square kilometers of convex hulls enclosing nautilid occurrences (Fig. 7B).

Species	Living geographic range (sq km)	Living geographic range plus drift shells (sq km)
<i>Allonautilus scrobiculatus</i>	1,458,480	11,808,955
<i>Nautilus belauensis</i>	500	2,200
<i>N. macromphalus</i>	185,977	4,369,601
<i>N. stenomphalus</i>	3,713,991	3,713,991
<i>N. pompilius</i>	76,428,763	131,322,000