Recent advances and challenges on application of tissue engineering for treatment of congenital heart disease

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25   **ABSTRACT**

26   Congenital heart disease (CHD) affects a considerable number of children and adults worldwide.

27   This implicates not only developmental disorders, high mortality, and reduced quality of life but

28   also, high costs for the healthcare systems.   CHD refers to a variety of heart and vascular

29   malformations which could be very challenging to reconstruct the malformed region surgically,

30   especially when the patient is an infant or a child. Advanced technology and research have offered

31   a better mechanistic insight on the impact of CHD in the heart and vascular system of infants,

32   children, and adults and identified potential therapeutic solutions.

33   Many artificial materials and devices have been used for cardiovascular surgery. Surgeons and the

34   medical industry created and evolved the ball valves to the carbon-based leaflet valves and

35   introduced bioprosthesis as an alternative. However, with research further progressing, contracting

36   tissue have been developed in the lab and tissue engineering (TE) could represent a revolutionary

37   answer for CHD surgery. Development of engineered tissue for cardiac and aortic reconstruction

38   for developing bodies of infants and children can be very challenging. Nevertheless, the use of

39   acellular scaffolds, allograft, xenografts, and autografts are already very common. Seeding of cells

40   on the surface and within scaffold is a key challenging factor for use of the above.

41   The use of different types of stem cells has been investigated and proven to be suitable for tissue

42   engineering. They are the most promising source of cells for heart reconstruction in a developing

43   body, even for adults. Some stem cell types are more effective than others, with some

44   disadvantages which may be eliminated in the future.

46 **1.0 INTRODUCTION**

48 Congenital heart diseases (CHD) refer to the abnormal formation of the infant's heart, great
49 thoracic vessels and heart valves during intra-uterine development [1]. CHD is different from the
50 acquired heart diseases which occur because of lifestyle or aging [2]. The abnormalities are
51 structural defects, such as valve defects, intravascular or intracardial stenosis, congenital
52 arrhythmias or cardiomyopathies which greatly affect the early and future life of a CHD patient
53 [1, 3]. People born with CHD need immediate medical care after birth which further continues
54 throughout their lives. In 2010, it was estimated that only in the USA approximately 2.4 million
55 people suffered from CHD and more than half of them were adults [4]. In Europe, for instance, for
56 the period of 2000 to 2005, about 36,000 live births per year were diagnosed with CHD [5]. The
57 number grows bigger when including the unborn that were diagnosed with CHD and died either
58 by the termination of pregnancy or by intrauterine death or even neonate death[5]. In the UK,
59 about 8 in every 1,000 live babies born have a heart or circulatory condition [1]. Some estimate
60 those numbers to be higher and, commonly, percentages of each type of CHD change depend on
61 the geographical area of investigation [6, 7], table 1 further mentions other CHD frequencies for
62 other countries. CHDs not only have an effect on the individual's and their family's life but also,
63 have a huge financial impact on healthcare system. According to NHS England for the financial
64 year 2013/14 the total spent on CHD was £175 million [8] and in the US the total cost for CHD
65 treatment in 2008 was approximately $298 billion [9]. In general, the number of children and adults
66 being diagnosed with CHD increases due to improved technology of diagnostic tools [6].

67 CHD can be diagnosed using transabdominal fetal Doppler echocardiography. Such prognostic
68 protocols are performed in high-risk groups of pregnant women, like those with a family history
69 of CHD [10]. In adults with CHD, the most effective diagnostic practice is transesophageal
70 echocardiography, electrocardiogram, pulse oximetry, X-rays, cardiac catheterization and MRI
71 [11]. The CHDs are managed by surgery, and the efficiency of this approach is largely dependent
72 on the materials that are used during the surgery. These materials are expected to be close to the
73 native cardiac tissue in both structure and function. In structure, CHD could present extremely
74 complicated malformations which cannot be spontaneously or by singular surgical procedure
75 reconstructed, hence the dire need for more research into biomaterials for Tissue Engineering (TE).

76   The recent extensive research focuses on possible ways to fabricate a near ideal tissue. So far, TE

77   appears to be the way forward in creating ideal tissue that can probably mimic the native heart

78   tissue both in structure and function. TE refers to creation of functional three-dimensional tissue

79   using biomaterials and cells for replacement or restoration of damaged organs and/or parts of them.

80   TE is the most promising approach at the present for CHD, as treatment can be "patient-specific"

81   and the engineered tissue could adjust to the developing body of the recipient. Many would think

82   that TE is an idea which conceived and developed in a very recent past. However, it has been

83   proven that tissue regeneration and TE is a concept which was born thousands of years ago, and it

84   has inspired Greek mythology, history, arts, and religion. In arts, religion inspired the well-known

85   painting of "Healing of Justinian" based on the miracle of St. Cosmas and St. Damian, physicians

86   and Christian martyrs who appear to have transplanted the leg of an Ethiopian to the body of a

87   patient [12]. The closest to an artificial replacement of a body part was discovered in Egypt on a

88   mummy which had a wooden replacement of the hallux [13]. However, today, TE involves a

89   combination of creating scaffolds and cell seeding. With regards to the heart, the most commonly

90   used and known artificial parts are the mechanical heart valves and conduits [14, 15]. The first

91   artificial heart valve was placed on live patients only in the last century [14, 16]. In 2003, some

92   evidence was presented to support the regenerative ability of the adult heart [17].

93   This evidence shows the existence of cardiac stem/progenitor cells which can differentiate into

94   new cardiomyocytes and participate in cardiac regeneration [17]. In contrast to the adult heart,

95   infants' and children's hearts regenerate in a larger capacity because the regenerative ability is

96   proportionally correlated to age [18]. Additionally, there is an insufficient number of heart donors

97   which becomes more challenging because of the heterogeneous relation of recipient-donor and the

98   diverse range of CHD. These points result in high mortality rates and further financial costs to the

99   healthcare systems [19]. However, only a small number of preclinical studies have focused on

100   CHD treatments [20, 21]. Stem cells (SC) have been widely investigated mainly for myocardial

101   infarction (MI), as it is currently the leading cause of morbidity and mortality worldwide [22]. Cell

102   seeding is a fundamental component of TE. Several studies have examined the possibility of direct

103   cell delivery in the damaged area, cardiac patch implantation and engineered heart tissue, with the

104   former being the most popular [20, 22, 23]. All possible types of stem cells are under investigation

105   to identify the most appropriate cell types for tissue engineering using in corrective surgery of

106 CHD. This review looks into the congenital heart diseases, biomaterials and scaffolds, and, types

107 of stem cells used in TE.

108

2.0 METHOD

110 This paper was based on review articles and reports in reputable peer-reviewed journals and

111 government websites. The research was conducted using Medline on OvidSP, PubMed, google

112 scholar, website, books, e- books, and reports. The words "congenital heart disease", "tissue

113 engineering", "surgical treatment", "stem cells", "scaffolds", "biomaterials" and a combination of

114 those were used to retrieve literature from the databases.

115

3.0 Congenital Heart Disease: Types, malformations, presentations and interventions

117 CHD includes a diverse range of conditions which shows a variety of symptoms, indications, and

118 malformations detected during pregnancy or after birth [11]. However, these malformations are

119 much influenced by the age of diagnosis [6]. The etiology of CHD is unknown, but it is generally

120 accepted that many factors or a combination of them could contribute to CHD and considered to

121 be caused by multifactorial inheritance. These factors could be genetic, epigenetic or

122 environmental factors such as alcohol and drugs consumption, as well as viral infections like

123 Rubella [11]. The severity of the disease varies, and a number of malformations could be present

124 in each case. Based on the severity of CHD, they are categorized to mild, moderate, and severe

125 CHDs, which the latter is subcategorized to Cyanotic and Acyanotic lesions [19]. The most

126 frequent type of severe CHD is Ventricular Septal Defect (VSD) [24]. VSD could cause

127 myocardial defects which disappear in the first year of the infant's life [24]. Nevertheless, the VSD

128 could also cause some malformations which can be managed only by surgical intervention, that is,

129 infant pulmonary hypertension [6, 24]. The other CHD type is Atrial Septal Defect (ASD) which

130 is usually asymptomatic and in most of the case will only be diagnosed in adulthood [6].

131 Atrioventricular septal defects (AVSD) is mainly observed in trisomy 21 [6]. AVSD is usually

132 characterized by "complete AV-canals with one common AV valve for both ventricles and an

133 interatrial and intraventricular communication" and requires surgical correction. The results of

134 long-term patient follow up after operation have shown very satisfactory survival rate [25].

135   Another type of CHD, tetralogy of Fallot (ToF), is characterized by VSD, pulmonary stenosis,

136   right ventricular hypertrophy and over-riding of the aorta [26]. Infants who suffer from ToF will

137   require immediate surgical intervention for better survival rates and avoid cyanosis, a result of

138   inadequate pulmonary blood flow [26]. Calcific Aortic Valve (CAV), another type of CHD, is a

139   disease which progresses slowly and results to a mild valve thickening and obstructing blood flow,

140   aortic sclerosis or severe calcification with impaired leaflet motion [27]. CAV presents many

141   similarities with arteriosclerosis in adults which is caused by lifestyle or aging [27]. However,

142   CAV is a congenital, progressive disease which could be diagnosed in patients less than one year

143   of age, and those in childhood or even adulthood [28, 29]. Table 1 summarizes the frequencies of

144   congenital heart diseases, while Table 2 summarizes the types of CHDs, presentations and possible

145   management.

146

147   ***Table 1: Frequencies of CHD in some regions***

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149   ***Table 2: Types, presentations and management of CHDs***

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154   Some types of the CHDs are misdiagnosed, undiagnosed, or diagnosed very late in life which could
155   make a successful treatment challenging [64].

156   In contrast to the adult heart, infants' and children's hearts regenerate in a larger capacity because
157   the regenerative ability is proportionally correlated to age [18].   Additionally, there is an
158   insufficient number of heart donors which becomes more challenging because of the
159   heterogeneous relation of recipient-donor and the diverse range of CHD. The mortality rate among
160   all patients who are waiting for any type of organ transplantation is highest in infants who wait for
161   heart transplant [65, 66]. CHDs patients usually require medication, cardiac catheterization or a
162   series of surgical interventions throughout their life [11, 66]. This among other risks increases the
163   chances of HLA-sensitization which eventually makes it more difficult to find a cross-match [66].
164   The main challenge with artificial materials is the alterations in the size and function of the heart
165   from the neonatal period, infanthood, and to adulthood, to which current artificial materials cannot
166   adjust to [11]. The need for early intervention is essential for normal physical and cognitive
167   development [64].

168   The use of bioprosthetics including allografts and xenografts is widely used for treatment of CHD
169   as they present good survival rates in the patient and fewer interventions throughout their lifetime
170   [65]. The observed symptoms, defects, and complications upon bioprosthetic treatment of CHD
171   patient could differ from case to case [24, 64].

172

173   **4.0 Heart, a complex organ for tissue engineering**

174   Unlike other human tissue, cardiac tissue is a more complex tissue considering not only because
175   of its mechanical and structural function but also due to its electrical properties as well. The human
176   heart mainly consists of cardiomyocytes, functions as a blood pump which is regulated by the
177   electrical signal generated by the pacemaker cells in the sinoatrial nodes. This signal is directed
178   and spread through the atrioventricular node to Purkinje fibers, and this is highly important for the
179   direction of blood flow [67]. The diastolic and systolic function of the heart is necessary to be
180   synchronized and adjusted according to the body needs. The isolation of this signal from the rest
181   of cardiac tissue is as important as this electrical signal by himself. This importance is achieved
182   by the extracellular matrix (ECM) of heart which is also responsible for mechanical support and

183 endurance [67]. Aside the functional aspects which briefly mentioned above, the structural aspects
184 (the cardiomyocyte and its ECM) are another factor for strong consideration in the success of TE.
185 To yield an engineered tissue with the best functionality, this engineered tissue must be similar in
186 every sense to the native tissue. The ECM of the heart is mainly a complex mesh of structural
187 elements such as cardio fibroblasts and collagen fibrils, and non-structural elements such as
188 proteoglycans, glycosaminoglycans, and glycoproteins [68, 69] among other components.
189 Repairing the heart using materials which do not have or do not comply with the above
190 characteristics and cannot work in harmony with the host-heart, will result in a non-efficient
191 functioning heart accompanied by a series of complications. The concept and act of repairing the
192 heart using various engineered techniques has evolved over the years from the use of artificial
193 heart valves and grafts to bioprosthesis, and currently forward to the use of biomaterials and
194 scaffolds cells.

195

196 **4.1 Artificial Heart Valve and Grafts**

197 Charles Hufnagel was the first to experiment on animals with an artificial valve which he designed
198 in the 1940's [15]. A few years later, the same type of valve was transplanted into humans [15].
199 Nonetheless, Hufnagel's valve required changes which were succeeded by Harken-Soroff and later
200 by Starr-Edwards ball valve [15]. Following several improvements, the latest version is made of
201 pure carbon as a lighter, smoother material for blood flow and more durable in comparison to other
202 materials [14]. Due to the position of the valve, there is a high transvascular pressure which leads
203 to 'impact wear' and 'friction wear' [14]. Further complications with the use of valve replacements
204 include inflammation around the prosthesis and calcification of the valve itself [70]. The main
205 disadvantage of artificial valves is the thromboembolic risk which leads to lifetime treatment with
206 anticoagulants. This type of treatment involves various complications and brings different risks to
207 patient [14]. Similarly, to valves, artificial vascular grafts have been used for many years as a
208 surgical treatment for CHD [71]. Nevertheless, they provoke an inflammatory response, and they
209 are much less flexible than the body's natural tissue [64]. Originally porous fabric knitted of
210 Dacron and polytetrafluoroethylene (PTFE) was used for stenosis treatment. Later alterations in
211 the porosity of fabric were introduced to prevent the material's corruption [64]. The first attempt
212 to combine biomaterials with patient's cells was made by Wesolowki and colleagues by

213 "preclotting the graft with the patient's blood" [64]. However, future attempts to produce

214 successful results created many doubts about the actual function of cells following citation on clot

215 [64]. Nonetheless, both artificial heart valves and conduits require continuous use of

216 anticoagulants. Remarkably, they do not grow with the patients' heart as patients with CHD are

217 highly likely to require surgical intervention when they are an infant or a child [64, 72].

218

219

220 **4.2 Bioprosthetics: Allograft, Xenografts, and Autografts**

221 Allograft heart valves and arterial grafts are collected from deceased humans. In contrast,

222 xenografts are harvested from porcine and bovine animals including heart valves and carotid

223 arteries [14, 16, 70]. Allografts and xenografts came into the picture as an alternative to artificial

224 valves and conduits. Their main advantage is that they do not require lifetime treatment with

225 anticoagulants [14, 73, 74]. Animal tissue is treated with glutaraldehyde [73]. Glutaraldehyde is a

226 five-carbon bifunctional aldehyde used to stabilize tissue to protect from chemical and enzymatic

227 degradation and maintain "its mechanical integrity and natural compliance" [73]. Also, treatment

228 is necessary to reduce immunogenicity of the xenograft by decellularization and sterilization of

229 the tissue [73]. Like xenograft, allografts also need treatment before transplantation and

230 can even be cryopreserved [65]. Despite the great advantage, a number of complications are

231 related to bioprosthetic grafts related to their preparation [73].

232 The risk of cytotoxicity leading to inflammation, as well as the partial loss of mechanical properties

233 of tissue, have been reported [73]. Moreover, calcification is often observed in infants and children

234 with bioprosthetics. Many efforts are being made to find out an alternative treatment for heart

235 bioprosthetics therapy. However, there is a controversy regarding their efficiency [73].

236 In a study carried out by Homann and colleagues, the outcomes of 25 years using allografts and

237 xenografts for reconstruction of the right ventricular outflow tract showed 66% survival at 10-

238 years' follow up. Furthermore, patients with allografts had a mean reoperation-free interval time

239 of 16 years in contrast to the xenograft recipients which this interval time is 10.3 years [65].

240 Allografts may present better outcomes, but they are not in abundance like xenografts. Therefore,

241   many studies are concentrating on the development of tissue valves and vascular grafts created by

242   stem cell-seeded on artificial or natural scaffold [16].

243   The relatively recent "CorMatrix" patch fabricated from the decellularized porcine small intestinal

244   submucosa extracellular matrix (SIS-ECM) mainly composed of collagen, elastin, glycan, and

245   glycoproteins have been introduced into cardiac surgery. SIS-ECM has not only been used in

246   animal models for cardiac surgery [65-77], but also in humans studies, for cardiac and vascular

247   reconstructions such as; augmentation of the tricuspid valve [78], vascular repair of ascending

248   aorta, aortic arch, right ventricular outflow tract, pulmonary artery, valvular reconstruction [79],

249   and closure of septal defects [80]. The study by Witt et al. reported a small risk of stenosis when

250   SIS-ECM is used in the reconstruction of the outflow tracts and great vessels. Interestingly, the

251   SIS-ECM has effectively proved the function in the high-pressure vessels [80]. The pitfall of this

252   study, however, was the short follow up period.

253   A very common surgical practice for CHD is the use of pericardial patches for repairing the septal

254   defect [64]. The autologous pericardium is the best choice for infants as it is free, it does not

255   provoke any immune-response, and it is sterile. Even though it requires some preparation before

256   application, autologous pericardium creates less fibrotic tissue in comparison to Dacron [64].

257   Allograft pericardium is available, but quite a few risk factors are associated with its use [64]

258

259   **4.3 Biomaterials and Scaffolds for tissue engineering**

260   Generally, scaffolds work as a primary base for cells to enhance and produce relevant tissue. The

261   scaffolds should have specific morphological, functional, and mechanical properties to support

262   cells survival and differentiation [23]. Biomaterials used to produce scaffolds should be made of

263   components which will accommodate the above characteristics and create a friendly cell

264   microenvironment [23]. The previously mentioned characteristics apply for all different types of

265   engineered tissue, and the goal is to mimic host tissue in the best possible way. With regards to

266   artificial and bioprosthetic cardiac correction choices, scaffolds and biomaterials should contain

267   various properties such as being biodegradable, biocompatible, flexible and durable and absence

268   of immunogenicity and calcification. Due to the variety of sizes of patients' heart and defects,

269   designed scaffolds should have various size and ability to be able to grow and adapt to the heart

270 [74]. The biomaterial should allow neo-vascularization for adequate oxygenation of the tissue,

271 create minimal scaring tissue and thrombotic risk, as the latter could lead to life treatment with

272 anticoagulants [81, 82]. Furthermore, these biomaterials scaffolds should be bioactive, meaning

273 they should enhance cellularization in vitro and in vivo, and optimize cell efficiency and degrade

274 at a desirable rate [82]. What is more, biomaterials and scaffolds should be in abundance and cost-

275 effective, as high cost could restrict development and use of it in TE as a routine therapeutic choice

276 [83]. The most common biomaterials for cardiac and vascular TE used today are synthetic, and

277 natural polymers [67], and the electrospinning technique has been proven to be the most efficient

278 way to produce scaffolds with these biomaterials [84].

279

280 **4.3.1 Synthetic polymers for cardiac scaffolds**

281 The easiest way to have materials in abundance is to manufacture them. The need for suitable

282 biomaterials for cardiac tissue repair has triggered development of synthetic polymers which are

283 easy to fabricate and manipulate. These polymers can be manipulated with respect to their physical

284 properties, molecular weight, heterogeneity index, and degradation speed [85]. Many synthetic

285 polymers are biocompatible and have excellent mechanical properties which make them a popular

286 choice for sutures and mesh production [67]. Frequently used polymers in cardiac surgery are

287 polyglycolic acid (PGA) and polylactic acid (PLA). These two polymers have been used as a single

288 biomaterial or as 50:50 composite to reconstruct tissue-engineered vascular grafts for treating

289 children with congenital heart disease [86]. Carrier et al. Presented acceptable ultrastructural

290 features and metabolic cell ability when cells were cultured on PGA scaffold in a rotating

291 bioreactor [87, 88]. The rotating bioreactor increases cell adherence and decreases cell damage

292 [87]. The highest concern with synthetic polymers is their toxicity. Therefore, the use of poly-L-

293 lactic acid (PLLA) has increasingly become more of interest. PLLA has demonstrated very good

294 results when combined with bone marrow mesenchymal SC (BM-MSC) for vascular tissue

295 engineering [89]. In vivo studies by Hashi et al showed that nanofibrous scaffolds created with

296 PLLA can remodel in both cellular and ECM content, similar to that of the native artery [89]. Both

297 acellular and were implanted into the common carotid artery of live animal models (rats). The

298 cellular scaffold was cited with MSC and "exhibited very little platelet aggregation on the luminal

299 surfaces" compared to the acellular grafts. This is due to the antithrombogenic property of MSC

300   [89]. PLLA, when degraded in the body, can "be excreted in carbon dioxide and water" [67].

301   Polyurethane, unless copolymerized, is biocompatible but not biodegradable. Polyurethane has

302   been successfully experimented in combination with other materials for cardiac tissue repair, such

303   as siloxane films [90, 91], cellulose [92], urea [93], PLLA [94]. Poly (Ɛ- caprolactone) in

304   combination with other biomaterials have also been proven to be efficient composite for cardiac

305   tissue repair. They have been used in combination with PLLA alone [95, 96] PLLA and collagen

306   [97], polypyrrole and gelatin [98], polyglycolic acid [99], poly (hydroxymethyl glycolide)

307   [100], chitosan and gelatin [101].

308

309   Based on our understanding of the heart as an electroactive tissue, Hitscherich and colleagues have

310   created a piezoelectric scaffold fabricated by electrospinning Polyvinylidene Fluoride-

311   Tetrafluoroethylene (PVDF-TrFE) for cardiac tissue engineering [102]. The combination of

312   synthetic with natural polymers has been suggested to increase cell adherence. However, pure

313   natural polymers have also been examined as an option for polymers [67].

314

315   **4.3.2 Natural polymers for cardiac scaffolds**

316   Natural polymers are biodegradable, biocompatible and easily manipulated matrices composed of

317   complex elements which make up the native tissue [103] The natural polymers used so far for

318   cardiac repair include collagen, gelatin, alginate, silk, fibrin, chitosan and hyaluronic acid [67].

319   Despite their poor mechanical properties, they are good biomaterial for heart TE, as they have high

320   biocompatibility, promote cell-binding and could biodegrade with no "additional treatment or

321   modifications" [67].

322   Collagen is the most widely utilized natural polymer which is the most abundant ECM protein. It

323   functions to guide biological processes, provide structural scaffolding, and tensile integrity [101].

324   Several kinds of literature have reported the use of various collagen types and their modifications

325   in cardiac tissue repair [104-111].

326   Fibrin can be manipulated to create gels, microbeads, and hydrogels [85]. Likewise, biological

327   molecules like the growth factors can be incorporated [112]. Fibrin glue can be used as a stand-

328 alone therapy in cardiac tissue repair as it possesses intrinsic regenerative properties [113]. The
329 success of fibrin patch seeded with human embryonic stem cell-derived cardiac progenitor cells
330 (hESC-CPC) in non-human primate model [114] has resulted in its translation to the first case
331 report of the use of hESC-CPC in severe heart failure with an encouraging patient functional
332 outcome [115]. Other studies on fibrin have demonstrated its efficacy as a sealant after
333 intramyocardial injection [116]; for myocardial tissue repair when seeded with adult stem cells,
334 neonatal cardiac cell, and mesenchymal stem cells [117-119]; to form aortic valves in tissue
335 engineering [120].

336

337 Chitosan has been experimented in several literature as a biomaterial for cardiac regeneration [121,
338 122]. Overall, in cardiomyogenesis, many researchers have agreed on the fact that chitosan seems
339 to be more effective when combined with other factors enhancing integration of stem cells into
340 cardiac tissues [123].

341

342 Alginate, when used alone, has proven to have a remarkable effect on the function of heart models
343 with myocardial infarction. Furthermore, seeding alginate with stem cells has proven to be more
344 efficient in repair of the cardiac tissue [124-127].

345 The use of hyaluronic acid has been shown to be largely dependent on its molecular weight, and
346 several kinds of literature have reported its successful use in cardiac tissue repair [128-131].

347 Evidence of gelatin scaffold placed subcutaneously, and/or on infarcted myocardium in adult rat
348 hearts have shown a good survival of the graft, vessel formation and junctions with recipient rat
349 heart cells [132]. Gelatin was reported to sustain neonatal rat cardiomyocyte tissue in vitro for
350 three weeks [133]; supported the growth of human induced pluripotent stem cell (iPSC)-derived
351 cardiomyocytes [133]; its hydrogel seeded with autologous human cardiac-derived stem cell and
352 basic fibroblast growth factor (bFGF) effectively released bFGF for repair of ischemic
353 cardiomyopathy [134]; and several other studies have shown the efficacy of gelatin as a scaffold
354 for cardiomyogenesis when seeded with cells [135-138]. Fibrinogen/Thrombin-based Collagen
355 Fleece (TachoCombo) have been successfully used to secure hemostasis and enhance complete
356 reconstruction of a large pulmonary artery defect in a canine model [139]. Hence, this biomaterial

357 may be used in reconstruction of the low-pressure pulmonary vessels during a cardiac surgery for
358 a total anomalous pulmonary venous return or transposition of the great vessels.

359 Nevertheless, not all of these polymers can tune well for cardiac TE, and risk of inflammation still
360 exists [140].

361

4.3.3 Native Extracellular Matrix as scaffold

363 Native-specific ECM could be a category itself or part of natural polymers. ECM is collected from
364 animal or donor tissue and processed for culturing cells [141]. Studies have shown that the ratio
365 of native ECM in culture could play a key role in stem cells (SCs) enhancement, differentiation,
366 survival, and phenotype [142]. Other studies have shown that contractible engineered heart patches
367 cultured in ECM mixture and implanted in syngeneic Fischer 344 rats can vascularize, become
368 innervated and survive up to 8 weeks in vivo [141].

369 Furthermore, ECM of decellularized and repopulated hearts and other organs are being used for
370 drug testing [143]. An experiment on decellularized mouse hearts which were repopulated with
371 human cells through coronary vessels exhibited myocardium, vessel-like structures and
372 intracellular Ca2+ transients contracted spontaneously and responded as expected to various drug
373 interventions [144]. It was concluded that heart "ECM could promote proliferation, specific cell
374 differentiation and myofilament formation" [144]. The option of ECM for TE could help overcome
375 the challenges faced using synthetic and other natural biomaterials to replace tissue, valves or
376 organs [145].

377

4.4 Scaffoldless Cell Sheet

379 Another technique, which is independent of scaffolds, has been developed based on the cells'
380 ability to connect via cell-to-cell junction proteins and create ECM [88]. The cells are cultured in
381 normal conditions at 37°C in a temperature-responsive polymer cultures dish. When the culture
382 temperature conditions change, the cells detach from the polymer culture dish as one cell sheet
383 [88]. This technique was developed to avoid inflammatory reactions and fibrotic deposits in the
384 area of graft where scaffold was placed following degradation [88]. A study has shown that

385   contractile chick cardiomyocyte sheets could function effectively around rat thoracic aorta when

386   applied on host myocardium [81]. This cell sheet could synchronize within 1 hour of implantation

387   with the host tissue [81]. Similar results have been shown in 3D structures using several cell sheets

388   aiming to create a thick cardiac patch [146]. The number of sheets is limited, as more than three

389   exhibits poor vascularization. However, the combination of endothelial cells and cardiomyocytes

390   is being examined to promote vascularization before implantation [88]. Table 3 summarizes the

391   pros and cons of the various materials and biomaterials used in tissue engineering.

392

393   *Table 3 Advantages and disadvantages of materials and biomaterials used in TE*

394

5.0 Stem Cells for tissue engineering

396   An equally important point in choosing a suitable biomaterial, scaffold or scaffoldless cell sheet,
397   is the choice of the most appropriate cell types suitable for the TE. Stem cells (SCs) as a known
398   cell source possesses the ability to differentiate toward Cardiac Muscle Cells/cardiomyocytes
399   (CMCs), smooth muscle cells (SMCs) and endothelial cells (ECs), can regenerate cardiac tissue.
400   Based on these properties, they play a key role in TE field. The currently used SCs in TE are
401   summarized in Table 4.

5.1 Embryonic Stem Cells

403   Embryonic stem cells (ESCs) are one of the cell sources which are used in TE approaches. ESCs
404   are derived from the inner cell mass of preimplantation blastocyst [147]. They can differentiate
405   into all different cell types of three germ layers. Human ESC (hESC) could be a good candidate
406   for cardiac tissue engineering. In a study conducted by Landry et al., hESC-derived
407   cardiomyocytes (hESC-CMC) showed very good phenotype including myofibril alignment,
408   density, morphology, contractile performance and gene expression profile which, however, was
409   only confirmed after 80-120 days in vitro culture [148]. Various groups apart from Landry and
410   colleagues conducted studies to confirm the successful differentiation of ESC to cardiomyocytes
411   as presented in a review by Boheler and colleagues in 2002 [149]. Duan et al. investigated how
412   native cardiac ECM could affect hESC differentiation. This group processed porcine hearts to
413   collect digested cardiac ECM which then mixed with collagen to create a hydrogel for cell
414   cultivation purposes. The cultured hESCs on biomaterials comprised of 75% native cardiac porcine
415   ECM and 25% hydrogel with no additional growth factors have shown a great differentiation with
416   cardiac troponin T expression and contractile behavior, compared to the hydrogel with a smaller
417   ratio of native ECM [142]. Based on various studies, ESCs could be a good option for cardiac TE
418   [141]. Additionally, some factors such as ethical concerns, provoked immunogenicity, and risk of
419   tumorigenesis make the ESCs a very controversial choice of cell source for TE [74].

420

421   **5.2 Induced Pluripotent Stem Cells**

422   Another type of SCs which are used in TE is induced pluripotent stem cells (iPSCs). The iPSCs

423   are somatic cells which are reprogrammed to behave like ESC and show the same properties. The

424   iPSCs can differentiate into all three germ layers [150, 151]. Takahashi and his group were the

425   first groups who were able to reprogram the somatic adult cells like fibroblasts to iPSCs using viral

426   vectors to introduce four key factors OCT4, SOX2, c-Myc, and KFL-4 to fibroblasts [150]. This

427   method was used to reprogram fibroblasts to embryonic-like cells and from this state differentiate

428   them into a relevant type of cells [150]. Ludry et al. have also shown that human iPSC-derived

429   cardiomyocytes (hiPSC-CMCs) present the same characteristics as hESC-derived cardiomyocytes

430   in long-term in vitro culture [148]. Lu and colleagues presented the successful repopulation of

431   decellularized cadaveric mouse heart with hiPSC-derived multipotential cardiovascular

432   progenitors [148]. They also demonstrate that the heart ECM promotes proliferation,

433   differentiation and myofilament formation of CMs from the repopulated hiPSC-derived cells.

434   Furthermore, they have checked the electrical coupling of these cells and also examined the

435   constructive ability of repopulated heart using electrocardiogram which presented arrhythmia. Lu

436   et al. have further examined the effects of pharmacological agents on repopulated heart and

437   observed remarkable responses [141]. This model is explored as an option to personalized

438   medicine concerning drug testing/discovery [141, 152]. Specifically, for CHD which presents such

439   a variety of profiles, individual patient-specific human models' development could help to

440   understand how each patient would respond to existing pharmaceutical treatments. Even though it

441   may not be possible to create actual organ heart models with individual clinical features of the

442   disease [153].

443   Nonetheless, like ESC, iPSC has demonstrated tumorigenesis [74]. The group of Leda and

444   colleagues showed a direct transdifferentiation of fibroblasts to functional cardiomyocytes using

445   three key factors, Gata4, Mef2c, and Tbx5, within a very short time and suggested that direct

446   reprogramming could reduce the risk of tumorigenesis [154]. Still, using viral vectors for

447   reprogramming procedure is problematic and involves various risks [74]. Therefore, today more

448   different ways for iPSC production are being used and investigated to find out safer and more

449   effective alternatives for this reprogramming procedure [21, 155]. In the event this problem is

450   solved, iPSC could be the safest type of cell sources for TE as they will not provoke any immune-

451 response and cell harvesting procedure to produce iPSCs is not life-threatening for the patients.

452 Moreover, iPSC raises less ethical concerns in comparison to ESC or fetal SC.

453

454 **5.3 Prenatal, Perinatal, and Postnatal Stem Cells**

455 Prenatal, perinatal and postnatal SCs are other cell sources used in TE, which include chorionic

456 villi derived multipotent SCs, amniotic fluid-derived SCs (AFSCs), umbilical cord blood derived-

457 endothelial progenitor cells (UCB-EPCs) and umbilical cord- or cord blood-derived- multipotent

458 SCs [156]. UCB progenitors, like endothelial progenitor cells (EPCs), have distinctive

459 proliferative properties in comparison to other cells sources [157]. This category of SCs is

460 exceptionally important as the child's own SCs could be used for heart TE, for CHD patients who

461 are diagnosed before birth. Immunogenicity or an additional procedure to harvest autologous SCs

462 from the infant or child could be avoided in this procedure. Furthermore, it has been proven that

463 AFSCs do not form teratomas in contrast to ESCs and iPSCs [158]. All categories of these cells

464 have been investigated with remarkable results on engineered valves and vascular grafts [156, 159-

465 163]. This type of SCs is not applicable to adults who have been diagnosed later in life with CHD.

466

467 **5.4 Adult Stem Cells**

468 **5.4.1 Bone Marrow-derived Stem cells**

469 Apart from UCB, EPCs can be found in the peripheral blood (PB-EPCs) and bone marrow (BM-

470 EPCs) of adults [164,165]. However, bone marrow is a richer source of EPCs in comparison to

471 peripheral blood. In 1997, Asahara et al. identified the CD34+ mononuclear hematopoietic

472 progenitor cells in the peripheral blood which in vitro presented endothelial-like characteristics

473 [164]. The EPCs which are originated from BM considered to play a crucial role in endothelial

474 repair, and they have been suggested for treatment of ischemia patients and vascular TE with very

475 encouraging results [158,160,165,166]. A successful complete endothelium regeneration of

476 decellularized canine carotid arteries has been reported in animal studies using PB-EPCs [167]. In

477 addition to vascular TE, the EPCs have been assessed for tissue-engineered heart valves [168].

478 Bone marrow derived EPCs are greatly involved in de novo vessel formation and
479 neovascularization in pathological conditions like ischemia and cancer [164]. Similar applications
480 to PB-EPCs and prenatal EPCs have been recorded for vascular graft in the congenital heart
481 surgery using bone marrow-derived stem cells [169]. Mirensky and colleagues used sheets of non-
482 woven PGA mesh as a scaffold to create vascular graft in combination with human bone marrow
483 mononuclear cells (BM-MNCs). The results were very encouraging as no aneurysm or thrombotic
484 incidence were reported, despite the absence of anticoagulants. This group suggests this method
485 as a suitable vascular treatment for CHD based on their results from 6 weeks follow up after graft
486 implantation, which has shown signs of degradation, and it was fully accommodated by the host's
487 cells [169]. Nonetheless, the host mice were at full-growth which makes it questionable how
488 successful this application could be in a developing animal model. Interestingly only one week
489 after implantation no human BM-MNCs were detected, which suggests that BM-MNCs play a
490 paracrine role rather than cell replacement [165]. More studies have reported similar results with
491 the same conclusion [74,170]. Furthermore, an investigation on 25 young patients under 30 years
492 old who underwent extracardiac total cardiopulmonary connection with BM-MNCs engineered
493 vascular graft has also presented convincing results. A long-term patient follow-up has shown zero
494 deaths in relation to the implanted grafts, no thromboembolic, hemorrhagic, or infectious
495 complications, however, 6 of them developed grafts stenosis which was treated successfully [74].
496 Despite the encouraging results with BM-MNCs, BM-MSCs still present more advantages. These
497 advantages are, for example, their ability to differentiate into a variety of cell types even progenitor
498 cells; relatively easy procedure for their collection, isolation, storage, and proliferation; presenting
499 a similar phenotype to the valve cells; present anti-thrombogenic properties; and their
500 immunogenicity is manageable [89, 145]. In a comparative study, Vincentelli et al. examined
501 short- and long-term characteristics of the porcine decellularized scaffold which were processed
502 with in-situ injections of BM-MNCs and BM-MSCs, before transplanting in a lamb of animal
503 models [171]. Short-term results did not show any significant differences. However, the 4 months
504 (long-term) follow up has shown a significant decrease of transvalvular and distal gradients, more
505 inflammatory reaction, more structural deterioration as well as calcification, and a thick fibrous
506 pannus around the suture line in the BM- MNCs group. These observations in the BM-MNCs
507 group were significantly different from the BM-MSCs group [171].

508

509 **5.4.2 Cardiac Progenitor Cells**

510 Cardiac progenitor cells (CPCs) or also known as Cardiac Stem Cells (CSCs) are a type of cells
511 which are found in the adult heart and express stem cell factor receptor kinase c-kit+, also shown
512 to be negative for the markers of blood/endothelial cell lineage [17]. CPCs have only been in the
513 spotlight for about 15 years now with the credit given to Beltrami and colleagues as they
514 demonstrated the self-renewing and multipotent characteristics of these cells; differentiating into
515 all three different cardiogenic cell types which are cardiomyocytes, smooth muscle cells and
516 endothelial cells [17]. More recently, a study carried out by Vicinanza and colleagues helped to
517 lay to rest the controversies about c-kit+ cardiac cells by demonstrating that only a small fraction
518 of the c-kit+ adult cardiac stem cells possess the tissue-specific progenitor properties [172]. The
519 study was carried out in adult Wistar rats in which an experimental acute myocardial infarction
520 (MI) was induced and the border of the infarcts were injected with GFP+c-kit+ for the first group,
521 "GFP-expressing CD45-c-kit+ CSCs (CSC^{GFP})" for the second group, and the placebo group were
522 injected with PBS. The study showed that CSC^{GFP} only significantly reduced apoptosis and
523 hypertrophy of the cardiomyocytes, it also significantly reduced scarring and improved ventricular
524 functions [172]. The study further demonstrated that about 10% of the overall c-kit+ cardiac cells
525 are CD45-c-kit+, and only about 10% of these are clonogenic and multipotent. Therefore, it was
526 inferred that only about 1-2% of the total c-kit+ myocardial cells have a demonstrable multipotent
527 CSC phenotype [172]. These c-kit+CD45-(deletion of which also renders the cells CD31-, CD34-)
528 CPCs were also shown to express to some degree Sca-1, Abcg2, CD105, CD166, PDGFR-α, Flk-
529 1, ROR2, CD13, and CD90 [172]. The number of CPCs have however been shown to be
530 significantly higher in neonates but dramatically decreases after the age of 2 [71, 173]. Thus, these
531 CPCs in combination with a suitable scaffold, could be the answer to treat a number of CHD, as
532 CPCs could be collected during palliative surgery or via endomyocardial biopsy [19,64].

533 The potential SCs and biomaterials for TE in CHDs are represented in the figure 1 below. Also,
534 the figure 2 below shows the promising strategies for the treatment of CHDs.

535

536 *Table 4 Scaffolds and SCs used for TE in some study models*

537

539   **Figure 1:** This schema represents the different types of stems that can be used on the biomaterial

540   backbone (depicted as the background characters) for cardiovascular Tissue Engineering (TE). A)

541   Induced pluripotent stem cells (iPSCs) derived from fibroblast. B) Prenatal, Perinatal, and

542   Postnatal Stem cells (PPPSCs) are derived from amniotic fluid, umbilical cord, and chorionic villi.

543   C) Bone Marrow Stem Cells (BMSCs) such endothelial progenitor cells (EPCs) and Mesenchymal

544   stem cells (MSCs) can easily be isolated from the bone marrow. D) Cardiac progenitor cells

545   (CPCs) can be harvested during palliative surgery or endomyocardial biopsy. E) Embryonic stem

546   cells (ESCs) derived from the inner cell mass of the blastocyst

547

548

549

550   **Figure 2** : The schematic diagram represents the potential of Stem Cells (SCs) and Tissue

551   Engineering (TE) for corrective surgical treatment of infants as well as adolescent patients with

552   Congenital Heart Disease (CHD). Various sources for Stem Cells (SCs) are presented here as

553   alternatives to harvesting the appropriate Stem Cells (SCs) which can be used to seed on clinically

554   certified biomaterial scaffolds for reconstructing functional cardiac tissue- engineered grafts.

555   These grafts could be implanted via the corrective surgery into the heart of infants and adolescent

556   patients with Congenital Heart Disease (CHD) for definitive correction of cardiac defects. This

557   optimized cardiac-tissue engineered grafts should have potential to grow in parallel with the child,

558   while are lacking any tumorigenicity, immunogenicity, thrombogenicity, calcification, and other

559   risk factors.

560

561   **7.0 STANDPOINT**

562   Due to the high number of patients as well as newborns who are suffering from CHD and also high

563   costs of their implications, it is necessary and vital to making intensive research for finding an

564   effective treatment for CHD patients. Stem cell research has shown remarkable results in all kinds

565   of tissue engineering including skin [174], cartilage [175], vascular [176], ocular [177] and cardiac

566   tissues [82, 87, 178, 179].

567

568 The importance of engineered cardiac tissue lies in the fact that synthetic non-degradable materials
569 cannot adjust to the patient's developing body. A number of patients who suffer from CHD are
570 adults, and they are more suitable for this type of therapies. However, many patients with severe
571 CHD are infants and children whose body is constantly developing. Although, various synthetic
572 and natural biodegradable biomaterials have been used so far which have shown good results, the
573 one with the best degradation rate is yet to be found. There are various complications related to
574 existing surgical treatments and scaffolds which cannot be ignored. Calcification, inflammatory
575 reaction and life-long anticoagulants treatment are the most important known complications for
576 the conventional methods of CHD treatment [14, 70]. The complexity of CHD makes TE possibly
577 the most suitable solution for treatment of patients with CHD.

578

579

580 **8.0 CONCLUSION**

581 The replacement or correction of a malformation in a complex system like the cardiovascular
582 system could only be successful with tissues which can mimic the native heart and vascular tissues.
583 SCs have opened the door to such treatments. The best SC candidates and biomaterials are yet to
584 be identified, despite the encouraging results. All different types of SCs which have been
585 investigated so far still present some disadvantages. Extensive research would be required to
586 enable deeper understanding, solve drawbacks, and promote SCs use for tissue engineering in the
587 future. All the efforts channeled at obtaining proper legal regulation for using SCs, developing
588 new technologies for scaffold production as well as scaffoldless techniques, developing faster and
589 safer methods for producing patient-specific iPSCs, and research into the effectiveness of SCs in
590 TE for treatment of CHD, predicts a very positive future for patients, researchers and surgeons.

591

592

593

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601

602

Reference

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1. *National Congenital Heart Disease Audit Report 2012-2015*. (2016) London: National Institute for Cardiovascular Outcomes Research. Retrieved from; https://nicor5.nicor.org.uk/chd/an_paeds.nsf/vwContent/home?Opendocument

2. *British Heart Foundation*. (2016). Retrieved from: www.bhf.org.uk: https://www.bhf.org.uk/research/heart-statistics

3. Okudo M, Benson LD. (2001) Intravascular and intracardiac stents used in congenital heart disease. *Current Opinions in Cardiology*, 16(2): 84-91.

4. Gilboa SM, Devine OJ, Kucik JE, Oster ME, Riehle-Colarusso T, Nembhard WN, Xu P, Correa A, Jenkins K, Marelli AJ. (2016) Congenital Heart Defects in the United States: Estimating the Magnitude of the Affected Population in 2010. *Circulation* 134(2):101-9. doi: 608 10.1161/CIRCULATIONAHA.115.019307.

5. Dolk H, Loane M, Garne E. (2011). Congenital Heart Defects in Europe: Prevalence and Perinatal Mortality 2000-2005. *Circulation* 123(8):841-9. doi:10.1161/CIRCULATIONAHA.110.958405

6. Hoffman JI, Kaplan S. (2002). The incidence of congenital heart disease. *Journal of the American College of Cardiology*, 39 (12): 1890-1900, https://doi.org/10.1016/S0735-1097(02)01886-7

7. van der Linde D, Konings EE, Slager MA, Witsenburg M, Helbing WA, Takkenberg JJ, Roos-Hesselink JW. (2011) Birth Prevalence of Congenital Heart Disease Worldwide: A Systematic Review and Meta-Analysis, *Journal of the American College of Cardiology*, 58(21): 2241-2247. https://doi.org/10.1016/j.jacc.2011.08.025

8. NHS England. (2015). *Proposed National Standards and Service Specifications for Congenital Heart Disease Services: Financial Impact Analysis*. London: NHS England. https://www.england.nhs.uk/wp-content/uploads/2015/06/chd-finance.pdf

9. Lundberg, M. (2013) Cardiovascular Tissue Engineering Research Support at the National Heart, Lung, and Blood Institute. *Circulation Research*, 112(8): 1097-1103. doi:10.1161/CIRCRESAHA.112.300638

10. Nayak K, Chandra G S N, Shetty R, Narayan PK. (2016) Evaluation of fetal echocardiography as a routine antenatal screening tool for detection of congenital heart disease. *Cardiovascular Diagnosis and Therapy*, 6(1): 44-49. doi:10.3978/j.issn.2223-3652.2015.12.01

11. Sun R., Liu M., Lu L., Zheng Y., Zhang P. (2015) Congenital Heart Disease: Causes, Diagnosis, Symptoms, and Treatments. *Cell Biochemistry and Biophysics*, 72(3): 857–860.

635

12. Durant, M. (2018). *The Ethiopian's Leg : Saint Lucy*. [online] Saint-lucy.com. Available at: https://saint-lucy.com/slider/the-ethiopians-leg/ [Accessed 7 Jul. 2018].

13. Finch, J. (2011). The ancient origins of prosthetic medicine. *The Lancet*, 377(9765), pp.548-549.

14. Zilla P, Brink J, Human P, Bezuidenhout D. (2008) Prosthetic Heart Valves: Catering for the few. *Biomaterials*,

640   29(4): 385-406.

641   15. Gott VL, Alejo DE, Cameron DE. (2003) Mechanical heart valves: 50 years of evolution. *The Annals of*
642   *Thoracic Surgery*, 76(6): S2230-S2239.

643   16. Sun R, Liu M, Lu L, Zheng Y, Zhang P. (2003) Thoracic Surgery Directors Association Award.
644   Bone marrow as a cell source for tissue engineering heart valves. *The Annals of Thoracic Surgery*,
645   75(3): 761-767.

646   17. Beltrami AP, Barlucchi L, Torella D, Baker M, Limana F, Chimenti S, Kasahara H, Rota M, Musso E,
647   Urbanek K, Leri A, Kajstura J, Nadal-Ginard B, Anversa P. (2003) Adult Cardiac Stem Cells Are
648   Multipotent and Support Myocardial Regeneration. *Cell*, 114(6): 763–776.
649   https://doi.org/10.1016/S0092-8674(03)00687-1

650   18. Rupp S, Schranz D. (2015) Cardiac Regeneration in Children. *Pediatric Cardiology*, 36(4): 713-718.

651   19. Saxena, A. K. (2010) Congenital Anomalies of Soft Tissues: Birth Defects Depending on Tissue
652   Engineering Solutions and Present Advances in Regenerative Medicine. *Tissue Engineering*,
653   16(5): 455-466. doi: 10.1089/ten.TEB.2009.0700.

654   20. Ebert AD, Diecke S, Chen IY, Wu JC. (2015) Reprogramming and transdifferentiation for
655   cardiovascular development and regenerative medicine: where do we stand? *Molecular Medicine*,
656   7(9), 1090-1103. doi: 10.15252/emmm.201504395.

657   21. Tarui S, Sano S, and Oh H. (2014) Stem cell therapies in patients with single ventricle physiology.

658   *Methodist Debakey Cardiovascular Journal*, 10(2): 77-81.

659   22. Fakoya AOJ. (2017) New Delivery Systems of Stem Cells for Vascular Regeneration in Ischemia.

660   *Frontiers in Cardiovascular Medicine,* 4:7. doi: 10.3389/fcvm.2017.00007.

661   23. Feric NT, Radisic M. (2016) Strategies and Challenges to Myocardial Replacement Therapy.

662   *Stem Cells Transplantation Medicine*, 17(6): 410-416. doi: 10.3390/ijms17060982.

663   24. Penny DJ, Vick GW (2011) Ventricular Septal Defect. *The Lancet*, 377(9771):1103-12. doi:

664   10.1016/S0140-6736(10)61339-6.

665   25. Boening A, Scheewe J, Heine K, Hedderich J, Regensburger D, Kramer HH, Cremer J. (2002)
666   Long-term results after surgical correction of atrioventricular septal defects. *European Journal of*
667   *Cardio-Thoracic Surgery*, 22(2), p167-73.

668   26. Apitz C, Webb GD, Redington A. (2009) Tetralogy of Fallot, .*The Lancet,* 374(9699), 1462–1471,
669   https://doi.org/10.1016/S0140-6736(09)60657-7

670   27. Freeman R, Otto C. (2005) Spectrum of Calcific Aortic Valve Disease Pathogenesis, Disease
671   Progression,   and   Treatment   Strategies.   *Circulation*,   111(24),   3316-3326.   doi:
672   10.1161/CIRCULATIONAHA.104.486738

673   28. Tanya Smith, Cha Rajakaruna, Massimo Caputo, and Costanza Emanueli. (2015) MicroRNAs in
674   congenital heart disease. *Annals of Translational Medicine* 21: 333, doi: 10.3978/j.issn.2305-
675   5839.2015.12.25

676   29. *Medscape*. (2015, August 19). Retrieved from http://emedicine.medscape.com/article/894095-

677   overview.

678 30. Pedra, C., Haddad, J., Pedra, S., Peirone, A., Pilla, C. and Marin-Neto, J. (2009). Paediatric and congenital
679   heart disease in South America: an overview. *Heart*, 95(17), pp.1385-1392.

680 31. CDC (2018). *Atrial Septal Defect Facts | Congenital Heart Defects | NCBDDD | CDC*. [online] Centers for
681   Disease Control and Prevention. Available at:
682   https://www.cdc.gov/ncbddd/heartdefects/atrialseptaldefect.html [Accessed 29 May 2018].

683 32. MayoClinic (2018). *Atrial septal defect (ASD) - Diagnosis and treatment - Mayo Clinic*. [online]
684   Mayoclinic.org. Available at: https://www.mayoclinic.org/diseases-conditions/atrial-septal-defect/diagnosis-
685   treatment/drc-20369720 [Accessed 25 Jan. 2018].

686 33. Krasuski, R. and Bashore, T. (2016). Congenital Heart Disease Epidemiology in the United States. *Circulation*,
687   134(2), pp.110-113.

688 34. van der Linde, D., Konings, E., Slager, M., Witsenburg, M., Helbing, W., Takkenberg, J. and Roos-Hesselink,
689   J. (2011). Birth Prevalence of Congenital Heart Disease Worldwide. *Journal of the American College of
690   Cardiology*, 58(21), pp.2241-2247.

691 35. NHS (2018). *Symptoms*. [online] nhs.uk. Available at: https://www.nhs.uk/conditions/congenital-heart-
692   disease/symptoms/ [Accessed 5 Jun. 2015].

693 36. Calderón-Colmenero, J., Cervantes-Salazar, J., Curi-Curi, P. and Ramírez-Marroquín, S. (2013). Congenital
694   Heart Disease in Mexico. *World Journal for Pediatric and Congenital Heart Surgery*, 4(2), pp.165-171.

695 37. HeartKids (2018). *Congenital Heart Disease - Heartkids site*. [online] Heartkids.org.au. Available at:
696   https://www.heartkids.org.au/congenital-heart-disease [Accessed 5 Jun. 2011].

697 38. Postoev, V., Talykova, L. and Vaktskjold, A. (2014). Epidemiology of cardiovascular malformations among
698   newborns in Monchegorsk (north-west Russia): a register-based study. *Journal of Public Health Research*,
699   3(2):270.

700 39. Patnana, S. and Turner, D. (2018). *Pediatric Hypoplastic Left Heart Syndrome: Background,Pathophysiology,
701   Etiology*. [online] Emedicine.medscape.com. Available at: https://emedicine.medscape.com/article/890196-
702   overview [Accessed 7 Jan. 2017].

703 40. van der Linde, D., Konings, E., Slager, M., Witsenburg, M., Helbing, W., Takkenberg, J. and Roos-Hesselink,
704   J. (2011). Birth Prevalence of Congenital Heart Disease Worldwide. *Journal of the American College of
705   Cardiology*, 58(21), pp.2241-2247.

706 41. Zühlke, L., Mirabel, M. and Marijon, E. (2013). Congenital heart disease and rheumatic heart disease in Africa:
707   recent advances and current priorities. *Heart*, 99(21), pp.1554-1561.

708 42. Canadian Heart Alliance (2018). *Facts and issues*. [online] Canadian Congenital Heart Alliance. Available at:
709   https://www.cchaforlife.org/facts-issues [Accessed 6 Jun. 2018].

710 43. CDC (2018). *Hypoplastic Left Heart Syndrome Facts | Congenital Heart Defects*. [online] Centers for Disease
711   Control and Prevention. Available at: https://www.cdc.gov/ncbddd/heartdefects/hlhs.html [Accessed 8 Nov.
712   2016].

713 44. AHA (2018). *Atrial Septal Defect (ASD)*. [online] Heart.org. Available at:
714   http://www.heart.org/HEARTORG/Conditions/CongenitalHeartDefects/AboutCongenitalHeartDefects/Atrial-
715   Septal-Defect-ASD_UCM_307021_Article.jsp [Accessed 19 Jan. 2018].

716   45. Mai, C., Riehle-Colarusso, T., O'Halloran, A., Cragan, J., Olney, R., Lin, A., Feldkamp, M., Botto, L., Rickard,
717   R., Anderka, M., Ethen, M., Stanton, C., Ehrhardt, J. and Canfield, M. (2012). Selected birth defects data
718   from population-based birth defects surveillance programs in the United States, 2005-2009: Featuring critical
719   congenital heart defects targeted for pulse oximetry screening. *Birth Defects Research Part A: Clinical and*
720   *Molecular Teratology*, 94(12), pp.970-983.

721   46. MayoClinic (2018). *Tricuspid atresia - Symptoms and causes*. [online] Mayo Clinic. Available at:
722   https://www.mayoclinic.org/diseases-conditions/tricuspid-atresia/symptoms-causes/syc-20368392 [Accessed
723   8 Mar. 2018].

724   47. MayoClinic (2018). *Tetralogy of Fallot - Symptoms and causes*. [online] Mayo Clinic. Available at:
725   https://www.mayoclinic.org/diseases-conditions/tetralogy-of-fallot/symptoms-causes/syc-20353477
726   [Accessed 9 Mar. 2018].

727   48. Baffa, J. (2018). *Tetralogy of Fallot - Pediatrics - MSD Manual Professional Edition*. [online] MSD Manual
728   Professional Edition. Available at: https://www.merckmanuals.com/professional/pediatrics/congenital-
729   cardiovascular-anomalies/tetralogy-of-fallot [Accessed 22 Nov. 2016].

730   49. Northwestern Medicine (2018). *Bicuspid Aortic Valve Symptoms*. [online] Northwestern Medicine. Available
731   at: https://www.nm.org/conditions-and-care-areas/cardiovascular-care/center-for-heart-valve-
732   disease/bicuspid-aortic-valve/symptoms [Accessed 7 Jun. 2018].

733   50. MayoClinic (2018). *Bicuspid aortic valve - Overview - Mayo Clinic*. [online] Mayoclinic.org. Available at:
734   https://www.mayoclinic.org/diseases-conditions/bicuspid-aortic-valve/cdc-20385577 [Accessed 7 Jun. 2018].

735   51. MayoClinic (2018). *Patent ductus arteriosus (PDA) - Symptoms and causes*. [online] Mayo Clinic. Available
736   at: https://www.mayoclinic.org/diseases-conditions/patent-ductus-arteriosus/symptoms-causes/syc-20376145
737   [Accessed 25 Jan. 2018].

738   52. Cleveland Clinic (2018). *Patent Ductus Arteriosus (PDA) Treatment Options | Cleveland Clinic*. [online]
739   Cleveland Clinic. Available at: https://my.clevelandclinic.org/health/diseases/17325-patent-ductus-arteriosus-
740   pda/treatment-options [Accessed 7 Jun. 2018].

741   53. MayoClinic (2018). *Pulmonary valve stenosis - Symptoms and causes*. [online] Mayo Clinic. Available at:
742   https://www.mayoclinic.org/diseases-conditions/pulmonary-valve-stenosis/symptoms-causes/syc-20377034
743   [Accessed 8 Jun. 2018].

744   54. Spicer, D., Hsu, H., Co-Vu, J., Anderson, R. and Fricker, F. (2014). Ventricular septal defect. *Orphanet*
745   *Journal of Rare Diseases*, 9(144),pp1-16.

746   55. Carminati, M., Butera, G., Chessa, M., De Giovanni, J., Fisher, G., Gewillig, M., Peuster, M., Piechaud, J.,
747   Santoro, G., Sievert, H., Spadoni, I. and Walsh, K. (2007). Transcatheter closure of congenital ventricular
748   septal defects: results of the European Registry. *European Heart Journal*, 28(19), pp.2361-2368.

749   56. Minette, M. and Sahn, D. (2006). Ventricular Septal Defects. *Circulation*, 114(20), pp.2190-2197.

750   57. NIH (2018). *Hirschsprung Disease | NIDDK*. [online] National Institute of Diabetes and Digestive and Kidney
751   Diseases. Available at: https://www.niddk.nih.gov/health-information/digestive-diseases/hirschsprung-disease
752   [Accessed 7 Jun. 2018].

753   58. Martins, P. and Castela, E. (2018). *Transposition of the great arteries*. [online] BMC. Available at:
754   https://ojrd.biomedcentral.com/articles/10.1186/1750-1172-3-27 [Accessed 8 Jun. 2018].

755 59. Cincinnati Children Health (2018). *Hirschsprung Disease in Infants & Children | Diagnosis & Treatment*.
756   [online] Cincinnatichildrens.org. Available at: https://www.cincinnatichildrens.org/health/h/hirschsprung
757   [Accessed 5 Jun. 2018].

758 60. Khan, A., Ramsey, K., Ballard, C., Armstrong, E., Burchill, L., Menashe, V., Pantely, G. and Broberg, C.
759   (2018). Limited Accuracy of Administrative Data for the Identification and Classification of Adult Congenital
760   Heart Disease. *Journal of the American Heart Association*, 7(2), p.e007378.

761 61. Safi, L., Liberthson, R. and Bhatt, A. (2016). Current Management of Ebstein's Anomaly in the Adult. *Current
762   Treatment Options in Cardiovascular Medicine*, 18(9).

763 62. Skybchyk, V. and Melen, Y. (2017). Calcific Aortic Valve Stenosis: Current Approaches to the Diagnosis,
764   Medical and Surgical Treatment. *HYPERTENSION*, 0(6.50), pp.74-80.

765 63. Johnson MC, Swanson PE, Johnson DC, Schuessler RB, Cox JL. (1998), An experimental model of small
766 intestinal submucosa as a growing vascular graft. *Journal of Thoracic and Cardiovascular Surgery*, 116(5), 805–
767 811.

768 64. Torok, R. (2015). Coarctation of the aorta: Management from infancy to adulthood. *World Journal of
769   Cardiology*, 7(11), p.765.

770 65. Dodson, R. (2014). *Comprehensive surgical management of congenital heart disease*. 2nd ed. Florida: CRC
771   press.

772 66. Homann M, Haehnel JC, Mendler N, Paek SU, Holper K, Meisner H, Lange R. (2000)
773   Reconstruction of the RVOT with valved biological conduits: 25 years' experience with allografts
774   and xenografts. *European Journal of Cardio-Thoracic Surgery*, 17(6), 624-630.
775   https://doi.org/10.1016/S1010-7940(00)00414-0

776 67. Files MD, Boucek RJ. (2012). 'Shovel-Ready' applications of stem cell advances for pediatric heart
777   disease. *Current Opinion In Pediatric*, 24(5), 577–583, doi:
778   10.1097/MOP.0b013e328357a4cf.

779 68. Kaiser N, Coulombe K. (2016) Physiological inspired cardiac scaffolds for tailored in vivo function
780   and heart regeneration. *Biomedical Materials*, 10(3): 034003. doi:10.1088/1748-
781   6041/10/3/034003

782 69. Rienks M, Papageorgiou A, Frangogiannis N, Heymans S. (2014) Myocardial Extracellular Matrix: An
783   Ever-Changing and Diverse Entity. *Circulation Research*. 114(5):872-888.
784   https://doi.org/10.1161/CIRCRESAHA.114.302533

785 70. Brown L. (2005) Cardiac extracellular matrix: a dynamic entity. *AJP: Heart and Circulatory Physiology*.
786   289(3), H973-H974. https://doi.org/10.1152/ajpheart.00443.2005

787 71. Chaikof EL. (2007), The Development of Prosthetic Heart Valve: Lessons in Form and Function. *The New
788   England Journal of Medicine*, 357:1368-1371. doi: 10.1056/NEJMp078175

789 72. Ratner, B., Hoffman, A., Schoen, F. and Lemons, J. (2004). *Biomaterials science*. 2nd ed. Amsterdam: Elsevier
790   Academic Press.

791 73. Hibino N, McGillicuddy E, Matsumura G, Ichihara Y, Naito Y, Breuer C, Shinoka T. (2010) Late-   term
792   results of tissue-engineered vascular grafts in humans. *The Journal of Thoracic and   Cardiovascular
793   Surgery*, 139(2), 431-436.

794 74. Schmidt CE, Baier JM. (2010) Acellular vascular tissues: natural biomaterials for tissue repair and tissue
795   engineering. *Biomaterials*, 21 (22), 2215-2231.

796 75. Kurobe H, Maxfield MW, Breuer CK, Shinoka T. (2012) Concise review: tissue-engineered
797   vascular grafts for cardiac surgery: past, present, and future. *Stem Cells Translational Medicine*,
798   1(7):566-71. doi: 10.5966/sctm.2012-0044.

799 76. Johnson MC, Swanson PE, Johnson DC, Schuessler RB, Cox JL. (1998), An experimental model of
800   small intestinal submucosa as a growing vascular graft. *Journal of Thoracic and Cardiovascular*
801   *Surgery*, 116(5), 805–811.

802 77. Ruiz CE, Iemura M, Medie S, Varga P, Van Alstine WG, Mack S, Deligio A, Fearnot N, Beier UH,
803   Pavcnik D, Hijazi ZM, Kiupel M. (2005) Transcatheter placement of a low-profile
804   4biodegradable pulmonary valve made of small intestinal submucosa: a long-term study in a swine
805   model. *Journal of Thoracic and Cardiovascular Surgery.* 130(2), 477–484.
806   706https://doi.org/10.1016/j.jtcvs.2005.04.008

807 78. Wainwright JM, Hashizume R, Fujimoto KL, Remlinger NT, Pesyna C, Wagner WR, Tobita K,
808   Gilbert TW, Badylak SF. (2012) Right ventricular outflow tract repair with a cardiac biologic
809   scaffold. *Cells Tissues Organs 195(1-2)*. 159–70. https://doi.org/10.1159/000331400

810 79. Scholl FB, Boucek MM, Chan K-C, Valdes-Cruz L, Perryman R. (2010) Preliminary experience with
811   cardiac reconstruction using decellularized porcine extracellular matrix scaffold: human
812   applications in congenital heart disease. *World Journal of Pediatric and Congenital Heart Surgery*,
813   1(1) 713   132-136.

814 80. Quarti A, Nardone S, Colaneri M, Santoro G, Pozzi M. (2011) Preliminary experience in the use of an
815   extracellular matrix to repair congenital heart diseases. *Interactive CardioVascular and Thoracic*
816   *Surgery* 13(6), 569–572, https://doi.org/10.1510/icvts.2011.280016.

817 81. Witt RG, Raff G, Van Gundy J, Rodgers-Ohlau M, Si MS. (2013) Short-term experience of porcine small
818   intestinal submucosa patches in paediatric cardiovascular surgery. European Journal of
819   Cardiothoracic Surgery 44(1), 72–76, https://doi.org/10.1093/ejcts/ezs638

820 82. Miyagawa S, Roth M, Saito A, Sawa Y, Kostin S. (2011) Tissue-Engineered Cardiac Constructs for
821   Cardiac Repair. *The Annals of Thoracic Surgery*, 91(1), 320-329.
822   https://doi.org/10.1016/j.athoracsur.2010.09.080

823 83. Avolio, E., Caputo, M., Madeddu, P. (2015) Stem cell therapy and tissue engineering for correction of
824   congenital heart disease. *Frontiers in Cell and Developmental Biology 3(39)*, 1-17.
825   https://doi.org/10.3389/fcell.2015.00039

826 84. Williams, D. (2004) Benefit and risk in tissue engineering. *Materials Today*, 7(5), 24–29. doi:
827   10.1016/S1369-7021(04)00232-9

828 85. Zhao G, Zhang X, Lu TJ, Xu F. (2015) Recent Advances in Electrospun Nanofibrous Scaffolds for
829   Cardiac Tissue Engineering. *Advanced Functional Materials.* 9(8): 5726–5738. doi:
830   10.3390/ma9080614

831 86. Maeda K, Tiwari-Pandey R, Ruel M, Suuronen E. (2015) Matrix Therapies for Cell Support and
832   Cardiac Repair. In: Suuronen E, Ruel M, ed. by. *Biomaterials for Cardiac Regeneration*. London:
833   Springer; p. 117-146.

834

87. Sugiura T, Matsumura G, Miyamoto S, Miyachi H, Breuer CK, Shinoka T. (2018) Tissue-engineered vascular in children with congenital heart disease: Intermediate term followup. *Seminars in Thoracic and Cardiovascular surgery*. doi: 10.1053/j.semtcvs.2018.02.002.

88. Carrier RL, Papadaki M, Rupnick M, Schoen FJ, Bursac N, Langer R. (1999) Cardiac tissue engineering: cell seeding, cultivation parameters, and tissue construct characterization. *Biotechnology and Bioengineering*, 5;64(5), 580-9.

89. Masuda S, Shimizu T, Yamato M, Okano T. (2008) Cell sheet engineering for heart tissue repair. *Advanced Drug Delivery Reviews*, 60(2), 277-285.

90. Hashi CK, Zhu Y,   Yang G, Young WL, Hsiao BS, Wang K, Chu B, Li S. (2007) Antithrombogenic property of bone marrow mesenchymal stem cells in nanofibrous vascular grafts. *Proceedings of the National Academy of Sciences of the United States of America*, 104 (29) p11915-11920. https://doi.org/10.1073/pnas.0704581104.

91. Baheiraei N, Gharibi R, Yeganeh H, Miragoli M, Salvarani N, Di Pasquale E, Condorelli G. (2015) Electroactive polyurethane/siloxane derived from castor oil as a versatile cardiac patch, part I: Synthesis, characterization, and myoblast proliferation and differentiation. *Journal of Biomedical Materials Research Part A.*, 104(3):775-787.

92. Baheiraei N, Gharibi R, Yeganeh H, Miragoli M, Salvarani N, Di Pasquale E, Condorelli G. (2016) Electroactive polyurethane/siloxane derived from castor oil as a versatile cardiac patch, part II: HL-1 cytocompatibility and electrical characterizations. *Journal of Biomedical Materials Research Part A*. 104(6):1398-1407.

93. Su W, Ho C, Shih T, Wang C, Yeh C. (2016) Exceptional biocompatibility of 3D fibrous scaffold for cardiac tissue engineering fabricated from biodegradable polyurethane blended with cellulose. International Journal of Polymeric Materials and Polymeric *Biomaterials* , 65(14):703-711.

94. Hernández-Córdova R, Mathew D, Balint R, Carrillo-Escalante H, Cervantes-Uc J, Hidalgo-Bastida L, Hernández-Sánchez F. (2016) Indirect three-dimensional printing: A method for fabricating polyurethane-urea based cardiac scaffolds. *Journal of Biomedical Materials Research Part A.*, 104(8):1912-1921.

95. Tomecka E, Wojasinski M, Jastrzebska E, Chudy M, Ciach T, Brzozka Z. (2017) Poly(l-lactic acid) and polyurethane nanofibers fabricated by solution blow spinning as potential substrates for cardiac cell culture. *Materials Science and Engineering: C.*, 75, 305-316

96. Sharifpanah F, Reinhardt M, Schönleben J, Meyer C, Richter M, Schnabelrauch M, Rode C, Wartenberg A, Bekhite M, Sauer H, Wartenberg M. (2017) Embryonic stem cells for tissue

836

837   biocompatibility, angiogenesis, and inflammation testing. *Cells Tissues Organs*, 204: 1-12.

838   https://doi.org/10.1159/000471794

839

840   97. Centola M, Rainer A, Spadaccio C, De Porcellinis S, Genovese JA, Trombetta M. (2010)
841   Combining electrospinning and fused deposition modelling for the fabrication of a hybrid vascular
842   graft. *Biofabrication*. 2(1):014102.; doi: 0.1088/1758-5082/2/1/014102.

843   98. Mukherjee S, Reddy Venugopal J, Ravichandran R, Ramakrishna S, Raghunath M. (2011)
844   Evaluation of the Biocompatibility of PLACL/Collagen Nanostructured Matrices with
845   Cardiomyocytes as a Model for the Regeneration of Infarcted Myocardium. *Advanced Functional*
846   *Materials*., 21(12):2291-2300.

847   99. Kai D, Prabhakaran M, Jin G, Ramakrishna S. (2011) Polypyrrole-contained electrospun conductive
848   nanofibrous membranes for cardiac tissue engineering. *Journal of Biomedical Materials Research*
849   Part A., 99A(3):376-385.

850   100. Sugiura T, Hibino N, Breuer C, Shinoka T. (2016) Tissue-engineered cardiac patch seeded with
851   human induced pluripotent stem cell derived cardiomyocytes promoted the regeneration of host
852   cardiomyocytes in a rat model. *Journal of Cardiothoracic Surgery*., 11(1). doi: 10.1186/s13019-
853   016-0559-z

854   101. Castilho M, Feyen D, Flandes-Iparraguirre M, Hochleitner G, Groll J, Doevendans P, Vermonden T,
855   Ito K, Sluijter JPG, Malda J. (2017)   Melt Electrospinning Writing of Poly-Hydroxymethylglycolide-
856   co-ε-Caprolactone-Based Scaffolds   for Cardiac Tissue Engineering. *Advanced Healthcare*
857   *Materials*., 6(18):1700311

858   102. Pok S, Myers JD, Madihally SV, Jacot JG. (2013) A multi-layered scaffold of a chitosan and gelatin

859   hydrogel supported by a PCL core for cardiac tissue engineering. *Acta Biomater*., 9: 5630-5642.

860   103. Hitscherich P, Wu S, Gordan R, Xie L-H, Arinzeh T, Lee EJ. (2016) The effect of PVDF-TrFE
861   scaffolds on stem cell derived cardiovascular cells. *Biotechnol Bioeng*., 113:1577–85.
862   doi:10.1002/bit.25918

863   104. Nelson D, Ma Z, Fujimoto K, Hashizume R, Wagner W. (2011) Intra-myocardial biomaterial
864   injection therapy in the treatment of heart failure: Materials, outcomes and challenges. *Acta*
865   *Biomaterialia*. 7(1):1-15.

866   105. Herpel E, Pritsch M, Koch A, Dengler T, Schirmacher P, Schnabel P. (2006) Interstitial fibrosis in the
867   heart: differences in extracellular matrix proteins and matrix metalloproteinases in end-stage
868   dilated, ischaemic and valvular cardiomyopathy. *Histopathology*. 48 (6), 736-747.

869   106. Serpooshan V, Zhao M, Metzler S, Wei K, Shah P, Wang A, Mahmoudi M, Malkovskiy AV, Rajadas
870   J, Butte MJ, Bernstein D, Ruiz-Lozano P. (2016) The effect of bioengineered acellular collagen patch
871   on cardiac remodeling and ventricular function post myocardial infarction. *Biomaterials*. 34(36),
872   9048-9055.

873   107. Dawson J, Schussler O, Al-Madhoun A, Menard C, Ruel M, Skerjanc I. (2011) Collagen scaffolds
874   with or without the addition of RGD peptides support cardiomyogenesis after aggregation of mouse
875   embryonic stem cells. *In Vitro Cellular & Developmental Biology – Animal*, 47(9):653-664.

876   108. Yu H, Zhao H, Huang C, Du Y. (2017) Mechanically and Electrically Enhanced CNT–Collagen
877   Hydrogels As Potential Scaffolds for Engineered Cardiac Constructs. *ACS Biomaterials Science &*
878   *Engineering*, 3(11), 3017–3021 doi: 10.1021/acsbiomaterials.6b0062

879   109. Sun H, Zhou J, Huang Z, Qu L, Lin N, Liang C, Dai R, Tang L, Tian F. (2017) Carbon nanotube-
880   incorporated collagen hydrogels improve cell alignment and the performance of cardiac constructs.
881   *International Journal of Nanomedicine,* 12:3109-3120.

882   110. Frederick J, Fitzpatrick J, McCormick R, Harris D, Kim A, Muenzer J, Marotta N, Smith MJ, Cohen JE,
883   Hiesinger W, Atluri P, Woo YJ. (2010) Stromal Cell-Derived Factor-1 Activation of Tissue-
884   Engineered Endothelial Progenitor Cell Matrix Enhances Ventricular Function After Myocardial
885   Infarction by Inducing Neovasculogenesis. *Circulation.* 122(11 suppl: 1):S107-S117.

886   111. Hsieh SC, Chen HJ, Hsu SH, Yang YC, Tang CM, Chu MY, Lin PY, Fu RH, Kung ML, Chen
887   YW, Yeh BW, Hung HS. (2016) Prominent Vascularization Capacity of Mesenchymal Stem Cells in
888   Collagen–Gold Nanocomposites. *ACS Applied Materials & Interfaces*. 8(42):28982-29000.

889   112. Nie SP, Wang X, Qiao SB, Zeng QT, Jiang JQ, Liu XQ, Zhu XM, Cao GX, Ma CS. (2010)
890   Improved myocardial perfusion and cardiac function by controlled-release basic fibroblast growth
891   factor using fibrin glue in a canine infarct model. *Journal of Zhejiang University Science* B.,
892   11(12):895-904.

893   113. Menasché P, Vanneaux V, Fabreguettes JR, Bel A, Tosca L, Garcia S, Bellamy V, Farouz Y, Pouly J,
894   Damour O, Périer MC, Desnos M, Hagège A, Agbulut O, Bruneval P, Tachdjian G, Trouvin JH,
895   Larghero J. (2014) Towards a clinical use of human embryonic stem cell-derived cardiac progenitors: a
896   translational experience. *European Heart Journal,* 36(12):743-750.

897   114. Gil-Cayuela C, Roselló-LLetí E, Ortega A, Tarazón E, Triviño J, Martínez-Dolz L, González-
898   Juanatey JR, Lago F, Portolés M, Rivera M. (2016) New Altered Non-Fibrillar Collagens in Human
899   Dilated Cardiomyopathy: Role in the Remodeling Process. *PLOS ONE.*, 11(12):e0168130.

900   115. Menasché P, Vanneaux V, Hagège A, Bel A, Cholley B, Cacciapuoti I, Parouchev A,
901   Benhamouda N, Tachdjian G, Tosca L, Trouvin JH, Fabreguettes JR, Bellamy V, Guillemain
902   R, SuberbielleBoissel C, Tartour E, Desnos M, Larghero J. (2015) Human embryonic stem cell-
903   derived cardiac progenitors for severe heart failure treatment: first clinical case report. *European*
904   *Heart Journal*, 36(30):2011-7.
905

906   116. Terrovitis J, Lautamäki R, Bonios M, Fox J, Engles JM, Yu J, Leppo MK, Pomper MG, Wahl
907   RL, Seidel J, Tsui BM, Bengel FM, Abraham MR, Marbán E. (2009). Noninvasive Quantification and
908   Optimization of Acute Cell Retention by In Vivo Positron Emission Tomography After
909   Intramyocardial Cardiac-Derived Stem Cell Delivery. *Journal of the American College of*
910   *Cardiology*, 54(17):1619-1626.

911   117. Llucià-Valldeperas A, Soler-Botija C, Gálvez-Montón C, Roura S, Prat-Vidal C, Perea-Gil I,
912   Sanchez B, Bragos R, Vunjak-Novakovic G, Bayes-Genis A. (2016) Electromechanical
913   Conditioning of Adult Progenitor Cells Improves Recovery of Cardiac Function After
914   Myocardial Infarction. *STEM CELLS Translational Medicine*. 6(3):970-981.

915   118. Tao Z, Mohamed M, Hogan M, Gutierrez L, Birla R. (2014) Optimizing a spontaneously contracting
916   heart tissue patch with rat neonatal cardiac cells on fibrin gel. *Journal of Tissue Engineering and*
917   *Regenerative Medicine*. 11(1):153-163.

918   119. Ichihara Y, Tano N, Fields L, Murugesu N, Shintani Y, Ishida M, Sato N, Suzuki K. Abstract
919   14735: (2017) Epicardial Placement of Instantly Produced Fibrin Glue- Incorporating
920   Mesenchymal Stromal Cells for the Treatment of Heart Failure [Internet]. *Circulation*. from:
921   http://circ.ahajournals.org/content/134/Suppl_1/A14735
922

923   120. Moreira R, Neusser C, Kruse M, Mulderrig S, Wolf F, Spillner J, Schmitz-Rode T,
924   Jockenhoevel S, Mela P. (2016) Tissue-Engineered Fibrin-Based Heart Valve with Bio-Inspired
925   Textile Reinforcement. *Advanced Healthcare Materials*. 5(16):2113-2121.

926   121. Camci-Unal G, Annabi N, Dokmeci M, Liao R, Khademhosseini A. (2014) Hydrogels for cardiac tissue
927   engineering. *NPG Asia Materials*., 6(5):e99.

928
929   122. Wang H, Shi J, Wang Y, Yin Y, Wang L, Liu J, Liu Z, Duan C, Zhu P, Wang C. (2014)
930   Promotion of cardiac differentiation of brown adipose derived stem cells by chitosan hydrogel
931   for repair after myocardial infarction. *Biomaterials*. 35(13):3986-3998.

932   123. Chopra S, Mahdi S, Kaur J, Iqbal Z, Talegaonkar S, Ahmad F. (2006) Advances and potential
933   applications of chitosan derivatives as mucoadhesive biomaterials in modern drug delivery. *Journal*
934   *of Pharmacy and Pharmacology*. 58(8):1021-1032

935   124. Wang L, Shansky J, Borselli C, Mooney D, Vandenburgh H. (2012) Design and Fabrication of a
936   Biodegradable, Covalently Crosslinked Shape-Memory Alginate Scaffold for Cell and Growth Factor
937   Delivery. *Tissue Engineering Part A,* 18(19-20):2000-2007.

938

939   125. Landa N, Miller L, Feinberg M, Holbova R, Shachar M, Freeman I, Ci. (2008) Effect of Injectable
940   Alginate Implant on Cardiac Remodeling and Function After Recent and Old Infarcts in Rat.
941   *Circulation*. 117(11):1388-1396.

942   126. Leor J, Tuvia S, Guetta V, Manczur F, Castel D, Willenz U, Petneházy O, Landa N, Feinberg
943   MS, Konen E, Goitein O, Tsur-Gang O, Shaul M, Klapper L, Cohen S. (2009) Intracoronary
944   Injection of In Situ Forming Alginate Hydrogel Reverses Left Ventricular Remodeling After
945   Myocardial Infarction in Swine. *Journal of the American College of Cardiology*. 54(11):1014-
946   1023.

947   127. Sabbah H, Wang M, Gupta R, Rastogi S, Ilsar I, Sabbah M, Kohli S, Helgerson S, Lee RJ. (2013)
948   Augmentation of Left Ventricular Wall Thickness With Alginate Hydrogel Implants Improves Left
949   Ventricular Function and Prevents Progressive Remodeling in Dogs With Chronic Heart Failure.
950   *JACC: Heart Failure*. 1(3):252-258.

951   128. Bonafè F, Govoni M, Giordano E, Caldarera C, Guarnieri C, Muscari C. (2014) Hyaluronan and
952   cardiac regeneration. *Journal of Biomedical Science*. 21: 100. doi: 10.1186/s12929-014-0100-4

953   129. Yoon SJ, Fang YH, Lim CH, Kim BS, Son HS, Park Y, Sun K. (2009) Regeneration of ischemic heart
954   using hyaluronic acid-based injectable hydrogel. Journal of Biomedical Materials Research Part B*:*
955   *Applied Biomaterials*. 91B (1):163-171.

956   130. Yang MC, Chi NH, Chou NK, Huang YY, Chung TW, Chang YL, Liu HC, Shieh MJ, Wang SS.
957   (2010) The influence of rat mesenchymal stem cell CD44 surface markers on cell growth, fibronectin
958   expression, and cardiomyogenic differentiation on silk fibroin – Hyaluronic acid cardiac patches.
959   *Biomaterials*. 31(5):854-862.

960   131. Göv E, Kenar H, Halbutoğullari Z, Arğa K, Karaöz E. (2016) Cardiomyogenic
961   differentiation potential of human lipoaspirate-derivedstem cells on hyaluronic acid/gelatin plasma gels.
962   *Turkish Journal of Biology*. 40:369-379.

963   132. Li RK, Jia ZQ, Weisel RD, Mickle DA, Choi A, Yau TM. (1999) Survival and function of bioengineered
964   cardiac grafts. *Circulation 100(19)*, 1163-1169.

965   133. McCain M, Agarwal A, Nesmith H, Nesmith A, Parker K. (2014) Micromolded gelatin hydrogels
966   for extended culture of engineered cardiac tissues. *Biomaterials*. 35(21):5462-5471.

967   134. Yacoub M, Terrovitis J. (2013) CADUCEUS, SCIPIO, ALCADIA: Cell therapy trials using
968   cardiac-derived cells for patients with post myocardial infarction LV dysfunction, still evolving. *Global*
969   *Cardiology Science and Practice*. (1):3: 6-8

970   135. Takehara N, Tsutsumi Y, Tateishi K, Ogata T, Tanaka H, Ueyama T, Takahashi T, Takamatsu T,
971   Fukushima M, Komeda M, Yamagishi M, Yaku H, Tabata Y, Matsubara H, Oh H. (2008)
972   Controlled Delivery of Basic Fibroblast Growth Factor Promotes Human Cardiosphere-Derived
973   Cell Engraftment to Enhance Cardiac Repair for Chronic Myocardial Infarction. *Journal of the*
974   *American College of Cardiology*. 52(23):1858-1865.

975   136. Navaei A, Saini H, Christenson W, Sullivan R, Ros R, Nikkhah M. (2016) Gold nanorod-incorporated
976   gelatin-based conductive hydrogels for engineering cardiac tissue constructs. *ActaBiomaterialia*.
977   41:133-146.

978   137. Kudová J, Procházková J, Vašiček O, Perečko T, Sedláčková M, Pešl M, Pacherník J, Kubala L.
979   (2016) HIF-1alpha Deficiency Attenuates the Cardiomyogenesis of Mouse Embryonic Stem Cells.
980   *PLOS ONE*. 11(6):e0158358.

981   138. Cristallini C, Cibrario Rocchietti E, Gagliardi M, Mortati L, Saviozzi S, Bellotti E, Turinetto V,
982   Sassi MP, Barbani N, Giachino C. (2016). Micro- and Macrostructured PLGA/Gelatin Scaffolds
983   Promote Early Cardiogenic Commitment of Human Mesenchymal Stem Cells In Vitro. *Stem Cells*
984   *International*. 2016:1-16.

985   139. Okada M, Yamashita C, Okada M, Okada K. (1995). Role of Endothelin-1 in Beagles With
986   Dehydromonocrotaline-Induced Pulmonary Hypertension. *Circulation*. 92(1):114-119.
987

988   140. Sakai T, Li RK, Weisel RD, Mickle DA, Kim ET, Jia ZQ, Yau TM. (2001). The fate of a tissue-
989   engineered cardiac graft in the right ventricular outflow tract of the rat. *Journal of Thoracic and*
990   *Cardiovascular Surgery 121(5)*, 932-942.

991   141. Zimmermann, W., Melnychenko, I., Eschenhagen, T. (2004). Engineered heart tissue for
992   regeneration of diseased hearts. *Biomaterials 25(9)*, 1639-1647.

993   142. Duan Y, Liu Z, O'Neill J, Wan LQ, Freytes DO, Vunjak-Novakovic G. (2011). Hybrid gel
994   composed of native heart matrix and collagen induces cardiac differentiation of human embryonic stem

995 cells without supplemental growth factors. *Journal of Cardiovascular Translational Research 4(5)*, 605-
996 615.

997 143. Lewis, T. (2016). *The Scientist*. Retrieved from the-scientist.com: http://mobile.the-
998   scientist.com/article/46588/will-organs-in-a-dish-ever-replace-animal-models.

999 144. Lu TY, Lin B, Kim J, Sullivan M, Tobita K, Salama G, Yang L. (2013). Repopulation of
1000   decellularized mouse heart with human induced pluripotent stem cell-derived
1001   cardiovascular progenitor cells. *Nature Communications*. 4:2307. doi: 10.1038/ncomms3307

1002 145. Iop L, Renier V, Naso F, Piccoli M, Bonetti A, Gandaglia A, Pozzobon M, Paolin A, Ortolani F,
1003   Marchini M, Spina M, De Coppi P, Sartore S, Gerosa G. (2009). The influence of heart valve leaflet
1004   matrix characteristics on the interaction between human mesenchymal stem cells and decellularized
1005   scaffolds. *Biomaterials 30(25)*, 4104–4116.

1006 146. Yamato, M., & Okano, T. (2004). Cell sheet engineering. *Materials Today*. Volume 7, Issue 5, Pages 42-47.
1007   https://doi.org/10.1016/S1369-7021(04)00234-2

1008 147. Boroviak T., Loos R., Bertone P., Smit A., Nichols J. (2014). The ability of inner cell mass cells to self-renew
1009   as embryonic stem cells is acquired upon epiblast specification. *Nature Cell Biology 16*, 513–525.

1010 148. Lundy SD, Zhu WZ, Regnier M, Laflamme MA. (2013). Structural and Functional Maturation of
1011   Cardiomyocytes Derived from Human Pluripotent Stem Cells. *Stem Cells and Development*.
1012   Jul 15;22(14):1991-2002. doi: 10.1089/scd.2012.0490.

1013 149. Boheler KR, Czyz J, Tweedie D, Yang HT, Anisimov SV, Wobus AM. (2002). Differentiation of
1014   Pluripotent Embryonic Stem Cells Into Cardiomyocytes. *Circulation Research*. Aug 9;91(3):189-
1015   201.

1016 150. Takahashi K, Tanabe K, Ohnuki M, Narita M, Ichisaka T, Tomoda K, Yamanaka S. (2007).
1017   Induction of pluripotent stem cells from adult human fibroblasts by defined factors. *Cell*. Nov
1018   30;131(5):861-72.

1019 151. Yu J, Vodyanik MA, Smuga-Otto K, Antosiewicz-Bourget J, Frane JL, Tian S, Nie J, Jonsdottir GA,
1020   Ruotti V, Stewart R, Slukvin II, Thomson JA. (2007). Induced pluripotent stem cell lines derived
1021   from human somatic cells. *Science*. Dec 21;318(5858):1917-20

1022 152. Bosman A, Edel MJ, Blue G, Dilley RJ, Harvey RP, Winlaw DS. (2015). Bioengineering and Stem
1023   Cell Technology in the Treatment of Congenital Heart Disease. *Journal of Clinical Medicine*. Apr
1024   22;4(4):768-81. doi: 10.3390/jcm4040768.

1025 153. Caputo M, Saif J, Rajakaruna C, Brooks M, Angelini GD, Emanueli C. (2015). MicroRNAs in
1026   vascular tissue engineering and post-ischemic neovascularization. *Advanced Drug Delivery Reviews*.
1027   Jul 1;88:78-91. doi: 10.1016/j.addr.2015.05.003.

1028 154. Ieda M, Fu JD, Delgado-Olguin P, Vedantham V, Hayashi Y, Bruneau BG, Srivastava D. (2010). Direct
1029   Reprogramming of Fibroblasts into Functional Cardiomyocytes by Defined Factors. *Cell*. Aug
1030   6;142(3):375-86. doi: 10.1016/j.cell.2010.07.002.

1031 155. Lüningschrör P, Hauser S, Kaltschmidt B, Kaltschmidt C. (2013). MicroRNAs in pluripotency,
1032   reprogramming, and cell fate induction. *BiochimicaetBiophysicaActa (BBA) - Molecular Cell
1033   Research*. Aug;1833(8):1894-903. doi: 10.1016/j.bbamcr.2013.03.025.

156. Webera, B., Zeisbergera, S., Hoerstrup, S. (2011). Prenatally harvested cells for cardiovascular tissue engineering: Fabrication of autologous implants prior to birth. *Placenta*. Oct;32 Suppl 4:S316-9. doi: 10.1016/j.placenta.2011.04.001.

157. Murohara T, Ikeda H, Duan J, Shintani S, Sasaki Ki, Eguchi H, Onitsuka I, Matsui K, Imaizumi T. (2000). Transplanted cord blood-derived endothelial precursor cells augment postnatal neovascularization. *The Journal of Clinical Investigation*. Jun;105(11):1527-36.

158. De Coppi P, Bartsch G Jr, Siddiqui MM, Xu T, Santos CC, Perin L, Mostoslavsky G, Serre AC, Snyder EY, Yoo JJ, Furth ME, Soker S, Atala A. (2007). Isolation of amniotic stem cell lines with potential for therapy. *Nature Biotechnology*. Jan;25(1):100-6.

159. Schmidt D, Mol A, Breymann C, Achermann J, Odermatt B, Gössi M, Neuenschwander S, Prêtre R, Genoni M, Zund G, Hoerstrup SP. (2006). Living autologous heart valves engineered from human prenatally harvested progenitors. *Circulation*. Jul 4;114(1 Suppl):I125-31.

160. Schmidt D, Breymann C, Weber A, Guenter CI, Neuenschwander S, Zund G, Turina M, Hoerstrup SP. (2004). Umbilical Cord Blood Derived Endothelial Progenitor Cells for Tissue Engineering of Vascular Grafts. *The Annal of Thoracic Surgery*. Dec;78(6):2094-8

161. Schmidt D, Achermann J, Odermatt B, Breymann C, Mol A, Genoni M, Zund G, Hoerstrup SP. (2007). Prenatally fabricated autologous human living heart valves based on amniotic fluid derived progenitor cells as single cell source. *Circulation*. 11;116(11 Suppl):I64-70.

162. Yao, C. (2016). Combination of Endothelial Progenitor Cells, Extracellular Matrix and Aligned Eletrospun Polyhydroxyalkanoate Scaffold for Vascular Tissue Engineering. *Cytotherapy*. 18(6) Supplement, S75. https://doi.org/10.1016/j.jcyt.2016.03.149

163. Petsche Connell J, Camci-Unal G, Khademhosseini A, Jacot JG. (2013). Amniotic Fluid-Derived Stem Cells for Cardiovascular Tissue Engineering Applications. *Tissue Engineering*. Aug;19(4):368-79. doi: 10.1089/ten.TEB.2012.0561

164. Asahara T, Murohara T, Sullivan A, Silver M, van der Zee R, Li T, Witzenbichler B, Schatteman G, Isner JM.(1997). Isolation of putative progenitor endothelial cells for angiogenesis. *Science*. Feb 14;275(5302):964-7.

165. Zisch, A. (2004). Tissue engineering of angiogenesis with autologous endothelial progenitor cells. *Current Opinion in Biotechnology*. 15(5):424-9.

166. Olausson M, Kuna VK, Travnikova G, Bäckdahl H, Patil PB, Saalman R, Borg H, Jeppsson A, Sumitran-Holgersson S. (2014). In Vivo Application of Tissue-Engineered Veins Using Autologous Peripheral Whole Blood: A Proof of Concept Study. *Ebiomedicine*. Sep22;1(1):72-9. doi: 10.1016/j.ebiom.2014.09.001.

167. Zhou M, Liu Z, Liu C, Jiang X, Wei Z, Qiao W, Ran F, Wang W, Qiao T, Liu C. (2012). Tissue engineering of small-diameter vascular grafts by endothelial progenitor cells seeding heparin-coated decellularized scaffolds. *Journal of Biomedical Materials Research*. 100(1):111-20. doi: 10.1002/jbm.b.31928.

168. Sales VL, Mettler BA, Engelmayr GC Jr, Aikawa E, Bischoff J, Martin DP, Exarhopoulos A, Moses MA, Schoen FJ, Sacks MS, Mayer JE Jr. (2011). Endothelial Progenitor Cells as a Sole Source for Ex Vivo Seeding of Tissue-Engineered Heart Valves. *Tissue Engineering*. 16(1):257-67. doi: 10.1089/ten.TEA.2009.0424.

169. Mirensky TL, Hibino N, Sawh-Martinez RF, Yi T, Villalona G, Shinoka T, and Breuer CK. (2010). Tissue-engineered vascular grafts: does cell seeding matter? *Journal of Pediatric Surgery*. 45(6): 1299–1305. doi: 10.1016/j.jpedsurg.2010.02.102.

170. Fernandes S, Chong JJH, Paige SL, Iwata M, Torok-Storb B, Keller G, Reinecke H, Murry CE. (2015). Comparison of Human Embryonic Stem Cell-Derived Cardiomyocytes, Cardiovascular Progenitors, and Bone Marrow Mononuclear Cells for Cardiac Repair. *Stem Cell Reports* 5(5): 753–762. doi: 10.1016/j.stemcr.2015.09.011.

171. Vincentelli A, Wautot F, Juthier F, Fouquet O, Corseaux D, Marechaux S, Le Tourneau T, Fabre O, Susen S, Van Belle E, Mouquet F, Decoene C, Prat A, Jude B. (2007). In vivo autologous recellularization of a tissue-engineered. *Surgery*. Aug;134(2):424-32.

172. Vicinanza C, Aquila I, Scalise M, Cristiano F, Marino F, Cianflone E, Mancuso T, Marotta P, Sacco W, Lewis FC, Couch L, Shone V, Gritti G, Torella A, Smith AJ, Terracciano CMN, Britti D, Veltri P, Indolfi C, Nadal-Ginard B, Ellison-Hughes GM, and Torella D (2017). Adult cardiac stem cells are multipotent and robustly myogenic: c-kit expression is necessary but not sufficient for their identification. *Cell Death and Differentiation*. 24, 2101-2116.

173. Bosman A, Edel MJ, Blue G, Dilley RJ, Harvey RP, Winlaw DS. (2011). Bioengineering and Stem Cell Technology in the Treatment of Congenital Heart Disease. *Circulation*. Apr 22;4(4):768-81. doi: 10.3390/jcm4040768.

174. Alessandri, G., Emanueli, C., & Madedu, P. (2004). Genetically Engineered Stem Cell Therapy for Tissue Regeneration. *Annals of the New York Academy of Science*. May; 1015:271-84. DOI:10.1196/annals.1302.023.

175. Metcalfe AD and Ferguson MWJ. (2007). Tissue engineering of replacement skin: the crossroads of biomaterials, wound healing, embryonic development, stem cells, and regeneration. *The Royal Society Interface*. Jun 22; 4(14): 413–437. doi: 10.1098/rsif.2006.0179

176. Makris EA, Gomoll AH, Malizos KN, Hu JC, Athanasiou KA (2015). Repair and tissue engineering techniques for articular cartilage. *Nature Reviews Reumatology*. 11(1): 21–34. doi: 10.1038/nrrheum.2014.157

177. Chlupác, J., Filová, E., Bacáková, E. (2009) Blood vessel replacement: 50 years of development and tissue engineering paradigms in vascular surgery. *Physiological Research* ;58 Suppl 2:S119-39.

178. Karamichos, D. (2015) Ocular Tissue Engineering: Current and Future Directions. *Journal of Functioning Biomaterials*. Feb 17;6(1):77-80. doi: 10.3390/jfb6010077.

179. Kane NM, Thrasher AJ, Angelini GD, Emanueli C. (2014). Concise Review: MicroRNAs as Modulators of Stem Cells and Angiogenesis. *Stem Cell*. May;32(5):1059-66. doi: 10.1002/stem.1629.

Frequencies of CHDs in some regions

Frequencies of CHD in some regions	
United State	Affects 1% of live births [33]
South America	Colombia: 1.2 per 1000 live births Brazil (Minas Gerais): 9.58 in 1000 live births Brazil (Londrina): 5.49 in 1000 live births [30]
Mexico	Affects 6-8 per 1,000 newborns. Drawing to the conclusion that there about 12,000 or 16,000 babies living with CHD [36]
Asia	Affects 9.3 per 1,000 live births [34]
Europe	Affects 8.2 per 100 live births [40]
United kingdom	Affects about 9 in every 1,000 babies [35]
Russia	Affects 2.7-3.8 per 1,000 newborns estimating as 86 newborns per year being affected with CHD [38]
Australia	Affects 8-10 cases per 1,000 live births. Resulting in 2,400-3,000 newborns with CHD each year. About 65,000 adults are living with CHD [37]
Africa	Mozambique: 2.3 in 1000 live births Northern Nigeria: 9.3% (122 of 1312 patients) [41]
Canada	Affects 1 in 80-100 live births [42]

Types, presentations and management of CHDs

Types of CHD	Presentations of CHD	Medical & Surgical Therapeutic Approach to CHD
Atrial Septal Defects	➤ This defect manifests as a hole in the wall (septum) that separates the chambers above (atria) from those below (ventricles). ➤ The volume of blood that flows through the lungs is increased over time due to the hole caused by the atrial defect resulting in damage to the blood vessels in the lungs. ➤ Frequent respiratory or lung infections ➤ Difficulty breathing ➤ Tiring when feeding (infants) ➤ Shortness of breath when being active or exercising ➤ Skipped heartbeats or a sense of feeling the heartbeat ➤ A heart murmur, or a whooshing sound that can be heard with a stethoscope ➤ Stroke ➤ Swelling of legs, feet, or stomach area [31].	➤ **Medical monitoring:** the patient is monitored to see if the atrial septal defect would close on its own. ➤ **Medications:** beta blockers (to maintain a regular heartbeat) or anticoagulants (to help reduce blood clots). ➤ **Surgery:** can be done through Cardiac catheterization or Open-heart surgery. ➤ **Follow-up care** [31, 32]
Hypoplastic Left Heart	➤ This defect affects the normal blood flow through the heart. The left side of the heart does not form correctly and as such it is considered a critical congenital heart defect. ➤ The following structures on the left side of the heart is affected: a. The left ventricle is underdeveloped. b. The mitral valves are not formed. c. The aortic valve is not formed. d. The ascending aorta is	➤ **Medication:** inpatient medications include prostaglandin E1, Dopamine and Potassium Chloride and outpatient medications are Furosemide, Digoxin and Captopril [39]. ➤ **Nutrition:** feeding tube or special high-calorie formula. ➤ **Surgery:** a. Norwood Procedure: performed on the infant within 2 weeks of a

Syndrome	underdeveloped.	baby's life.
	➤ The left side of the heart cannot pump oxygen-rich blood. ➤ Rapid, difficult breathing ➤ Pounding heart ➤ Weak pulse ➤ Poor feeding ➤ Being unusually drowsy or inactive ➤ Ashen or bluish color ➤ Dilated pupils ➤ Lackluster eyes that seem to stare [43, 44].	b. Bi-directional Glenn Shunt Procedure: done on an infant around 4 to 6 months of age. c. Fontan Procedure: performed on an infant around 18 months to 3 years of age [43]
Hypoplastic Left Heart Syndrome *(continued)*		
Tricuspid Atresia	➤ Occurs in which the tricuspid valve is not formed leading to the underdevelopment of the right ventricle. ➤ The right side of the heart can't pump sufficient blood to the lungs. ➤ Problems breathing ➤ Ashen or bluish skin color (cyanosis) ➤ Poor feeding ➤ Extreme sleepiness ➤ Slow growth and poor weight gain ➤ Edema of the abdomen, legs,	➤ **Medications:** prostaglandins like Alprostadil IV to keep open the ductus arteriosus. ➤ **Nutrition:** feeding tube ➤ **Surgery:** a. Atrial Septostomy: performed in the first few days or weeks of a baby's life b. Banding c. Shunt Procedure: done within the first 2 weeks of a baby's life.

	ankles and feet [45, 46].	d. Bi-directional Glenn Procedure: performed around 4 to 6 months of the baby's life. **e.** Fontan Procedure: done around 2 years of age [45].
Tetraogy of Fallot	➢ Has a combination of four heart defects. This defect is a combination of pulmonary stenosis, ventricular septal defect, overriding aorta and right ventricular hypertrophy. ➢ Cyanosis ➢ Shortness of breath ➢ Rapid breathing especially during feeding or exercise ➢ Fainting ➢ Clubbing of fingers and toes ➢ Poor weight gain ➢ Fatigue during play or exercise ➢ Prolonged crying ➢ Irritability ➢ Heart murmur due to pulmonary stenosis [47, 48].	➢ **Medication:** Prostaglandin E_1 infusion. ➢ **Surgery:** a. Temporary surgery (palliative surgery): improve blood flow to the lungs. b. Intra-cardiac repair: done during the first year after birth [47].
Tetraogy of Fallot *(continued)*		
	➢ Has only two (bicuspid) cusps instead of three. ➢ A bicuspid aortic valve may result in the heart's aortic valve to narrow (aortic valve stenosis) which prevents the	➢ **Surgery:** a. Aortic valve replacement b. Balloon valvuloplasty c. Aortic valve repair d. Aortic root and

Bicuspid Aortic Valve	valve from opening completely, which reduces or blocks blood flow from the heart to the body. ➤ Trouble breathing ➤ Chest pain or pressure ➤ Fatigue ➤ Heart racing ➤ Light-headedness ➤ Fainting [49].	ascending aorta surgery [50]
Patent Ductus Arteriosus	➤ A persistent opening between the two major blood vessels leading from the heart. ➤ Large patent arteriosus can cause poorly oxygenated blood to flow in the wrong direction. ➤ Poor eating leads to poor growth. ➤ Sweating with crying or eating ➤ Persistent fast breathing or breathlessness ➤ Easy tiring ➤ Rapid heart taste [51].	➤ **Medications:** NSAIDS (Advil, Infant's Motrin), or indomethacin (Indocin) [51] ➤ **Surgery:** Video-assisted thoracic surgical (VATS) repair ➤ **Catheter procedure:** Trans-catheter occlusion [52] ➤ **Watchful waiting**

Pulmonic Valve Stenosis	➤ This defect affects the pulmonic valve in which a deformity on or near the valve causes it to be smaller and as such slows the blood flow.
	➤ The narrowing is due to the underdevelopment of the valve during fetal growth. The cusps maybe defective or too thick or may not separate from each other well.
	➤ Heart murmur
	➤ Fatigue
	➤ Shortness of breath, especially during exertion
	➤ Chest pain
	➤ Fainting [53]
		➤ **Surgery:**
		  a. Balloon valvulplasty
		  b. Open-heart surgery [53]

Pulmonic Valve Stenosis *(continued)*		

	➢ A fissure connecting the two ventricles of the heart. Size varies with each patient [54]
	➢ It can occur isolated or in association with other CHDs.
	➢ There are three kinds: muscular, periventricular and supra-crystal. These are based on location within the septum [55]
	➢ **Blood shunting:** Depending on the size of the hole, blood flows from the left ventricle to the right.
	➢ **Pulmonary hypertension**: The shunting of blood flow leads to increased ventricular output to the pulmonary artery. With time, this can lead to pulmonary hypertension.
	➢ **Eisenmenger's Syndrome:** Rise in pulmonary vascular resistance leads to increase in right ventricular pressure. This can lead to reverse shunting of blood from the right to left. This leads to cyanosis [54]
	➢ Patients could also display clubbing [55]
	➢ Growth retardation: Increased blood flow to the lungs results in an increase in lung compliance. This increases the energy demand for respiration. Thus, an energy deficit is created where the infant does not consume as much calories
		➢ Smaller holes resolve themselves with time.
		➢ **Surgery:** Usually done on larger fissures. It is indicated where patients express symptoms of heart failure, left heart overload and history of endocarditis.
		➢ **Percutaneous techniques:**
		  a. These do not require opening the patient up.
		  b. Trans catheter approach: A catheter is threaded from an artery in the legs, or groin into the heart. A device is then placed to obstruct the hole in the ventricle [54, 55].

Ventricular Septal Defect	as is burned. This impedes growth.
	➢ **Airway Obstruction:**
	  a. Increased pulmonary blood flow increases the size of the pulmonary arteries. This can cause the physical obstruction of large and small airways.
	  b. There is also the possibility of the incidence of pulmonary edema due to increased blood flow. The combination of these events can lead to respiratory distress. Thus, symptoms such as wheezing, and tachypnea can be observed [54].
	  c. Holosystolic/pansystolic murmur on auscultation [56]	

Ventricular Septal Defect *(continued)*		
Total Anomalous Pulmonary Venous Condition	➢ The pulmonary veins are attached to the right atrium instead of the left. ➢ Usually associated with atrial septal defect. ➢ **Cyanosis:** Oxygenated blood from the lungs is pumped into the right atrium. It mixes with deoxygenated blood and passes through the atrial septal defect into the left atrium. This decreases oxygen supply to the body leading to cyanosis. ➢ **Pulmonary hypertension:** Some patients have constricted pulmonary veins that lead to pulmonary hypertension. This leads to pulmonary effusion. ➢ **Hypovolemia**: Some patients manifest with a narrow or restrictive atrial septal defect. This significantly reduces the blood flow to the body leading to hypovolemia [57].	➢ **Surgery:** The pulmonary veins are surgically reattached to the left atrium. ➢ **Cardiac catheterization:** For the patients with a restricted atrial septal defect, a balloon pump is used to widen the fissure until corrective surgery can be carried out [57]
	➢ A condition whereby the aorta and the pulmonary arteries are transposed. The aorta arises from the right ventricle and	➢ **Medications:** Prostaglandin E1 is administered to keep the ductus arteriosus open.

Trans-Position of the Great Arteries **Trans-Position of the Great Arteries** *(continued)*	leads to the lungs. The pulmonary artery arises from the left ventricle and leads to the body. ➢ It is comorbid with ventricular septal defect and patent ductus arteriosus. ➢ **Cyanosis:** Mixing of blood leads to supply of poorly oxygenated blood to the body [58]	➢ **Surgery:** a. Balloon atrial septostomy: A catheter is threaded through the foramen ovale. A balloon is inflated to rip a fissure in the atrium [58]
Truncus Arterious	➢ A condition where the truncus arteriosus of a fetus does not differentiate into an aorta and pulmonary vein. Thus, the patient only has one vessel exiting the heart ➢ **Cyanosis:** This leads to mixing of oxygenated and deoxygenated blood. Thus, the oxygen supply to the body is decreased ➢ **Congestive heart failure:** the excess of volume of blood flow to the heart increases pressure in the lungs. This would eventually lead to cardiac failure. ➢ Usually comorbid with ventricular septal defect [59]	➢ **Surgery:** The truncus arteriosus must be separated into two vessels. This would allow separate blood flow channels to the heart and body [59]

	➢ A congenital malformation of the tricuspid valve
	➢ The posterior and septal leaflets are displaced downwards. This leads to a downward enlargement of the right atrium.
	➢ The walls of the right atrium
		➢ **Surgery:**
		  a. Only required where patient manifests severe symptoms.
		  b. Cone procedure: Where the anterior septal leaflet is maneuvered and

Ebstein's Anomaly	become thin. ➢ It can be comorbid with patent foramen ovale or atrial septal defect. ➢ Patients can be asymptomatic but could also present with symptoms. ➢ **Cyanosis:** Shunting of the blood between patent foramen ovale and atrial septal defect leads to blood mixing between the left and right sides of the heart. This can lead to cyanosis. ➢ **Conduction irregularities:** Some patients present with arrhythmias [60]	sewn to the true annulus. This attachment causes it to be conical in shape. c. Valve replacement d. Where the defective valves can be surgically replaced [60]
Ebstein's Anomaly *(continued)*		
	➢ Characterized by a restriction to blood flow from the right ventricle to the pulmonary artery. It could be due to malformation of the pulmonary valve or of the pulmonary artery itself. ➢ It can manifest with a ventricular septal defect	➢ **Surgery:** a. A shunt must be created between the pulmonary artery and the aorta. This can be done by administration of prostaglandin E to keep the ductus

Pulmonary Atresia	➢ where there are collateral arteries supplying the lungs. ➢ It could also manifest without a ventricular septal defect. Here the right ventricle is usually hypoplastic. It would usually be comorbid with a patent ductus arteriosus [61].	arteriosus open. It could also be doe surgically. b. Fontan's procedure: Done for patients with a hypoplastic right ventricle. The right atrium is surgically connected to the pulmonary artery [61].
Aortic Stenosis	➢ Defect of the aortic valve that restricts its opening. ➢ It can lead to ventricular hypertrophy which can eventually lead to heart failure. ➢ There is also a possibility of development of atrial fibrillation. ➢ Presence of systolic murmur [62]	➢ **Valve replacement :** There are two approaches: a. Surgically b. Trans-catheter approach [62].
Coarctation of the Aorta	➢ It is the constriction of the proximal end of the aorta leading to restriction to blood flow. ➢ Patients can present with acidosis, cardiac failure, as well as shock after ductus arteriosus closes [63]	➢ **Surgery:** a. Coarcted potion can be resected. And the ends of the artery can be re-anastomosed. b. A patch can also be used to surgically dilate the artery. ➢ **Balloon Angioplasty:** A catheter is threaded in to the aorta. A balloon is inflated to enlarge the aorta.

Advantages and disadvantages of materials and biomaterials used in TE

	Artificial Prosthesis	Biological Prosthesis	Biomaterial Scaffolds	Scaffoldless Tissue
Advantages	• Available in abundance • Many different sizes • Long term results available	• Available in abundance • No requirement for life-long treatment with anticoagulants	• Good mechanical properties • Ultrastructural features • Cell adherence • Biocompatibility • Biodegradable	• No need for scaffold • Spontaneous and synchronous pulsation • Could create tubular construct • Can grow with
Disadvantages	• Impact & friction wear • Inflammation • Calcification of valve • Less flexible than natural tissue • Life-long treatment with anticoagulants	• Risk of cytotoxicity • Inflammation • Loss of mechanical properties • Calcification in infants and children • Immunological reactions	• Some present toxicity • Risk of inflammation • Not all tune well with heart	• Limited number of cell sheets (max 3) • Poor vascularization in more than 3 cell-sheets

Scaffolds and SCs used for TE in some study models

Engineered tissue	Scaffold	Type of SCs	Study models	Reference
Heart valve	Synthetic biodegradable non-woven PGA mesh	Human Chorionic villi-derived cells & hCB- EPCs	Culture in bioreactor	164
	Synthetic biodegradable	hAFSCs	Culture in bioreactor	166
	porcine decellularized scaffold	BM-MSCs & BM-MSCs	Lambs	176
Vascular graft	Various synthetic biodegradable	Human Umbilical CB-EPCs	Static conditions & biomimetic flow system	165
	biodegradable non-woven PGA	BM-MNCs	Mice	174
	Biodegradable PLA & PGA	BM-MNCs	Human	79

Figure 1

Schematic of the different types of stems that can be used on the biomaterial backbone for cardiovascular Tissue Engineering (TE)

This schema represents the different types of stems that can be used on the biomaterial backbone (depicted as the background characters) for cardiovascular Tissue Engineering (TE). A) Induced pluripotent stem cells (iPSCs) derived from fibroblast. B) Prenatal, Perinatal, and Postnatal Stem cells (PPPSCs) are derived from amniotic fluid, umbilical cord, and chorionic villi. C) Bone Marrow Stem Cells (BMSCs) such endothelial progenitor cells (EPCs) and Mesenchymal stem cells (MSCs) can easily be isolated from the bone marrow. D) Cardiac progenitor cells (CPCs) can be harvested during palliative surgery or endomyocardial biopsy. E) Embryonic stem cells (ESCs) derived from the inner cell mass of the blastocyst

Figure 2

Promising strategies for CHDs treatment

The schematic diagram represents the potential of Stem Cells (SCs) and Tissue Engineering (TE) for corrective surgical treatment of infants as well as adolescent patients with Congenital Heart Disease (CHD). Various sources for Stem Cells (SCs) are presented here as alternatives to harvesting the appropriate Stem Cells (SCs) which can be used to seed on clinically certified biomaterial scaffolds for reconstructing functional cardiac tissue-engineered grafts. These grafts could be implanted via the corrective surgery into the heart of infants and adolescent patients with Congenital Heart Disease (CHD) for definitive correction of cardiac defects. These optimized cardiac-tissue engineered grafts should have the potential to grow in parallel with the child, while are lacking any tumorigenicity, immunogenicity, thrombogenicity, calcification, and other risk factors.

Umbilical cord blood
Heart biopsy of newborn

Peripheral blood
sample

Bone marrow
sample

Heart biopsy
sample

Somatic cell biopsy
from patient

Cell isolation

Cell isolation

Cell isolation

Cell isolation

Direct transdifferentiation/reprogramming

Reprogramming

AFSCs
UCB-EPCs

PB-EPCs
PB-MNCs

BM-EPCs
BM-MNCs

CPCs

Patient-specific
iPSCs

Differentiation

Differentiation

Differentiation

Differentiation

Differentiation

CMCs + ECs + SMCs

CMCs + ECs + SMCs

CMCs + ECs + SMCs

CMCs + ECs + SMCs

CMCs + ECs + SMCs

CMCs + ECs + SMCs

Engineered
prosthetic grafts (EPGs)

Purified appropriate
cells
+

Engineered
prosthetic grafts (EPGs)

Clinically certified
Biomaterial scaffolds

Implantation of EPGs
via corrective surgery

Implantation of EPGs
via corrective surgery

EPGs implantation to
Infant CHD patient

Adaptability of EPGs with growing
body & accepable heart correction

EPGs implantation to
adult CHD patient

Adaptability of EPGs with body
& accepable heart correction