

A new species of the odorous frog genus *Odorrana* (Amphibia, Anura, Ranidae) from southwestern China

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The genus *Odorrana* is widely distributed in the mountains of East and Southeastern Asia. An increasing number of new species in the genus have been recognized especially in the last decade. Phylogenetic studies of the *O. schmackeri* species complex with wide distributional range also revealed several cryptic species. Here, we describe a new species in the species complex from Guizhou Province of China. Phylogenetic analyses based on mitochondrial DNA indicated the new species as a monophyly clustered into the *Odorrana* clade and sister to *O. schmackeri*, and nuclear DNA also indicated it as an independent lineage separated from its related species. Morphologically, the new species can be distinguished from its congeners based on a combination of the following characters: (1) having smaller body size in males (snout-vent length < 43.3 mm); (2) head longer than wide; (3) dorsolateral folds absent; (4) tympanum of males large and distinct, tympanum diameter twice as long as width of distal phalanx of finger III; (5) two metacarpal tubercles; (6) relative finger lengths: II < I < IV < III; (7) tibio-tarsal articulation reaching to the level between eye to nostril when leg stretched forward; (8) disks on digits with circum-marginal grooves; (9) toes fully webbed to disks; (10) the first subarticular tubercle on fingers weak; (11) having white pectoral spinules, paired subgular vocal sacs located at corners of throat, light yellow nuptial pad on the first finger in males.

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ABSTRACT

The genus *Odorrana* is widely distributed in the mountains of East and Southeastern Asia. An increasing

number of new species in the genus have been recognized especially in the last decade. Phylogenetic studies of the *O. schmackeri* species complex with wide distributional range also revealed several cryptic species. Here, we describe a new species in the species complex from Guizhou Province of China. Phylogenetic analyses based on mitochondrial DNA indicated the new species as a monophyly clustered into the *Odorrana* clade and sister to *O. schmackeri*, and nuclear DNA also indicated it as an independent lineage separated from its related species. Morphologically, the new species can be distinguished from its congeners based on a combination of the following characters: (1) having smaller body size in males (snout-vent length < 43.3 mm); (2) head longer than wide; (3) dorsolateral folds absent; (4) tympanum of males large and distinct, tympanum diameter twice as long as width of distal phalanx of finger III; (5) two metacarpal tubercles; (6) relative finger lengths: II < I < IV < III; (7) tibio-tarsal articulation reaching to the level between eye to nostril when leg stretched forward; (8) disks on digits with circum-marginal grooves; (9) toes fully webbed to disks; (10) the first subarticular tubercle on fingers weak; (11) having white pectoral spinules, paired subgular vocal sacs located at corners of throat, light yellow nuptial pad on the first finger in males.

INTRODUCTION

Frogs of the genus *Odorrana* Fei, Ye & Huang, 1990 inhabit montane streams in the subtropical and tropical regions of East and Southeast Asia, ranging from the Ryukyu Archipelago of southern Japan, southern

China and Indochina, northeastern India, Myanmar and Malay Peninsula to Sumatra and Borneo (Frost, 2018). Although the systematic relationships of the group had been controversial for decades, it has been recognized as a monophyly in recent years (e.g. Matsui et al., 2005; Ngo et al., 2006; Cai et al., 2007; Che et al., 2007; Stuart, 2008; Wiens et al., 2009; Pyron & Wiens, 2011; Chen et al., 2013). The genus currently contains 58 recognized species (Frost, 2018). Noticeably, in the last decade, 13 species have been described (Li et al., 2008; Tran et al., 2008; Yang, 2008; Bain et al., 2009; Chen et al., 2010a, b; Kuramoto et al., 2011; Mo et al., 2015; Wang et al., 2015; Pham et al., 2016; Saikia et al., 2017). This indicated that the species diversity has been underestimated and more discoveries were obligatory in the genus.

The piebald odorous frog *O. schmackeri* was firstly found in Gaojiayan Town, Changyang County (Co.), Hubei Province (Prov.), China, and named by Boettger (1892) only based on one male specimen and with a negligible description. Thus, Liu & Hu (1961) redescribed *O. schmackeri* based on some specimens from Daba Mountains and Xiushan Co. in Chongqing City of China near to the type locality of the species, and presented the diagnosis characters for the species, such as having great circle brown spots on dorsa and the outer vocal sac below pharynx in males. Afterwards, based on mass of reports and investigations, the species has been suggested to be distributed in Henan, Shaanxi, Gansu, Sichuan, Guizhou, Hubei, Anhui, Jiangsu, Zhejiang, Hunan, Guangdong and Guangxi provinces of China (Fei et al., 2009, 2012), even northern Thailand (Chanard, 2003) and northern Vietnam (Orlov et al., 2002; Nguyen et al., 2005). This group is highly adapted to mountain environments, inhabiting the moist evergreen broad-leaf forests and streams at 200-1400 m altitudes (Fei et al., 2009). Accordingly, it could be assumed that the wide distributional range and strict habitat requirements might promote considerable divergence even speciation due to isolation in the group. Correspondingly, populations ever classified as *O. schmackeri* were suggested to be much variable in

morphology (Fei et al., 2009, 2012), indicating that there might be cryptic species in the group. Indeed, from this group, several new species have been constantly described: *O. nanjiangensis* Fei, Ye, and Jiang, 2007 occurring from Nanjiang County (Co.), Sichuan Province (Prov.); *O. huanggangensis* Chen, Zhou and Zheng, 2010a occurring from Wuyi Mountains in Fujian and Jiangxi provinces; and *O. tianmuis* Chen, Zhou and Zheng, 2010b occurring from Tianmu Mountains in Zhejiang Prov.. However, phylogenetic analyses of Chen et al. (2013) indicated that above four species did not form a monophyly, but were nested with some other *Odorrana* species including two cryptic species, and also strongly rejected the alliance of *O. anlungensis*, *O. yizhangensis* and *O. lungshengensis* with the *O. schmackeri* group that was defined based on morphological data (Fei et al., 2009). So the “*O. schmackeri* species complex” was defined referring to the species resembling *O. schmackeri* on morphology, such as *O. nanjiangensis*, *O. huanggangensis*, *O. hejiangensis*, *O. tianmuis* and several cryptic species (e.g. Li et al., 2015; Zhu, 2016; He, 2017).

Recently, many studies have paid attention to the phylogenetic relationships and diversification of the *O. schmackeri* species complex (e.g. Li et al., 2015; Zhu, 2016). Li et al. (2015) presented a phylogeographic framework for the species complex using 25 populations based on mitochondrial ND2 and two tRNA gene sequences, and proposed seven clades: Clade A was (*O. huanggangensis* + “*O. yizhangensis*” identified by them) from Wuyi Mountains in Fujian Prov., Nanling Mountains in the border of Guangdong and Hunan provinces and mountains in eastern Guizhou Prov.; Clade B was *O. tianmuis* occurring from Huangshan Mountains in Anhui Prov. and Tianmu Mountains in Zhejiang Prov.; Clade C was *O. schmackeri sensu stricto* occupying a more narrow distribution area in Funiu Mountains in Henan and northern Hunan provinces and Daba Mountains in Hubei Prov.; Clade D was proposed as a cryptic species from mountains in Jiangxi Prov.; Clade E was the second cryptic species from mountains of northwestern Guizhou Prov.; Clade F was the third

cryptic species from Funiu Mountains in Henan Prov. and being sympatric with Clade C; and Clade G was the fourth cryptic species from Daba Mountains in Hubei Prov. and also being sympatric with *O. schmackeri sensu stricto* (Fig. 1). As well, Zhu (2016) based on 12S rRNA and 16S rRNA genes showed a broadly similar phylogeographic framework through a more comprehensive sampling with 78 populations, and supplied several different implications: Clade D in Li et al. (2015) should not be recognized as a cryptic species but still be classified as *O. schmackeri*, and thus *O. schmackeri sensu stricto* was in fact distributed in western Henan, southeastern Shanxi, eastern to central Chongqing, northwestern and eastern Hubei, northwestern Hunan, northeastern Guizhou and Jiangxi provinces in China; *O. hejiangensis* occupied a large range around Sichuan Basin even in western Henan and eastern Shanxi provinces; one cryptic species (*Odorrana* sp1 defined by them) had a large range across central Guizhou and Guangxi provinces; and another cryptic species (*Odorrana* sp2 defined by them) occupied a large range around the southern part of Sichuan Basin (Fig. 1). However, because of different samplings and use of different genes in the two studies especially Li et al. (2015) having no morphological data and no releasing of sequences in Zhu (2016), there were still several uncertain points: (1) whether Clade E in Li et al. (2015) containing only two populations (Jinsha Co. and Suiyang Co. in Guizhou Prov.) belonged to *Odorrana* sp1 in Zhu (2016); (2) whether Clade F (the third cryptic species) from Funiu Mountains in Henan Prov. in Li et al. (2015) belonged to *O. hejiangensis*; (3) whether Clade G from Daba Mountains in Li et al. (2015) belonged to *Odorrana* sp2 in Zhu (2016). Anyway, several cryptic species in the species complex have been indicated. To better understand diversification of the species complex, it is necessary and urgent to make more investigations (e.g. distribution range, morphology, molecular phylogenetics, ecology and tadpoles) on the new taxa.

From 2013 to 2017, we carried out a series of biodiversity surveys in Guizhou Prov. of China and

collected many specimens of *Odorrana* superficially resembling *O. schmackeri*. As note, some of them were collected from the population in the Lengshuihe Reserve in Jinsha Co., Guizhou Prov., China (Fig. 1) where the samples of Clade E in Li et al. (2015) indicated as a cryptic species was collected. To distinguish these specimens, we conducted phylogenetic analyses based on mitochondrial DNA and nuclear DNA and morphological comparisons. All analyses consistently supported our specimens from Guizhou Prov. as a new taxon. Hence, we describe it herein as a new species.

METHODS

Sampling

Frogs were collected on September 17 and 21, 2013 and August 3, 4 and 8, 2015, and tadpoles were collected on October 10, 2017. After taking photographs, they were euthanized using isoflurane, and then the specimens were fixed in 10% buffered formalin. Tissue samples were taken and preserved separately in 95% ethanol prior to fixation. Specimens collected in this work were deposited in Chengdu Institute of Biology, Chinese Academy of Sciences (CIB, CAS).

A total of 25 adult specimens of the new taxon including 16 females and nine males were collected from the mountain streams of three localities across Guizhou Province of China (Fig. 1; for voucher information see Table S1). For comparisons, some specimens of the related species were also collected in this study, including 19 *O. schmackeri* specimens from two localities, 23 *O. huanggangensis* specimens from three localities, three *O. yizhangensis* specimens from two localities and two *O. lungshengensis* specimens from one locality (Fig. 1; for voucher information see Table S1).

Ten tadpoles with almost identical morphology were collected from the same place in the stream where

the new taxon was found in the Lengshuihe Nature Reserve of Jinsha Co., Guizhou Prov., China. They were identified as the new taxon because they were almost identical in morphology and one representative of them was genetically close to the adult specimens of the new taxon (see the results). Stages of tadpoles were identified following Gosner (1960).

Collection of molecular data

A total of 18 molecular samples were collected in this study: five containing four adults and one tadpole of the new taxon, five of *O. schmackeri* including two topotypes, three of *O. huanggangensis*, three of *O. yizhangensis* including one topotype, and two of *O. lungshengensis* (for voucher information see Table S2).

Total DNA was extracted using a standard phenol-chloroform extraction protocol (Sambrook et al., 1989). The mitochondrial 12S rRNA, 16S rRNA and ND2 genes and two nuclear protein-coding genes (DOLK and KCNF genes) were amplified and sequenced from our samples. Primer sequences were acquired from literatures for 12S rRNA (Kocher et al., 1989), 16S rRNA (Simon et al., 1994), ND2 (Li et al., 2015), DOLK (Shen et al., 2013) and KCNF (Shen et al., 2013) genes (primer sequences were presented in Table S3). PCR amplification reactions for mitochondrial genes were performed in a 30 µL volume reaction with the following cycling conditions: an initial denaturing step at 95 °C for 4 min; 36 cycles of denaturing at 95 °C for 40 s, annealing at 55 °C (for 12S and 16S)/47 °C-57 °C (for ND2) for 40 s and extending at 72 °C for 70 s, and a final extending step of 72 °C for 10 min. Amplifications of nuclear genes were according to Shen *et al.* (2013). PCR products were purified with spin columns and then were sequenced with both forward and reverse primers same as used in PCR. Sequencing was conducted using an ABI3730 automated DNA sequencer in Shanghai DNA BioTechnologies Co., Ltd. (Shanghai, China). All new sequences were deposited in GenBank (for GenBank Accession numbers see Table S2).

For phylogenetic analyses, we downloaded 12S and 16S gene sequences from GenBank for all those related species especially for their topotypes for which comparable sequences were available (for GenBank Accession numbers see Table S2) based on the previous studies (Chen *et al.*, 2013; Li *et al.*, 2015). As note, ND2 gene sequences for most *Odorrana* species were not sequenced to now, impeding us to examine comprehensive relationships of the genus using this gene. To further understand the divergence between the new taxon and its related species, ND2 sequences for all haplotypes of the *O. schmackeri* species complex in Li *et al.* (2015) were downloaded (for GenBank Accession number see Table S2).

Phylogenetic analyses and genetic distance estimation

Sequences were assembled and aligned using the ClustalW module in BioEdit v. 7.0.9.0 (Hall, 1999) with default settings. The datasets were checked by eye and revised manually if necessary. To avoid bias in alignments, GBLOCKS v. 0.91.b (Castresana, 2000) with default settings was used to extract regions of defined sequence conservation from the length-variable 12S and 16S fragments. No-sequenced fragments were treated as missing data. At last, for phylogenetic analyses of mitochondrial DNA, two datasets were obtained, i.e. three-genes concatenated dataset with 12S (for 61 samples) + 16S (for 61 samples) + ND2 (for 16 samples) and ND2 gene alone dataset (for 112 samples).

Based on the three-genes concatenated dataset, phylogenetic analyses were conducted using maximum likelihood (ML) and Bayesian Inference (BI) methods, implemented in PhyML v. 3.0 (Guindon *et al.*, 2010) and MrBayes v. 3.12 (Ronquist & Huelsenbeck, 2003), respectively. One *Rana chensinensis* was chosen as outgroup according to the previous studies (Pyrone & Wiens, 2011; Chen *et al.*, 2013). To avoid under- or over-parameterization (Lemmon & Moriarty, 2004; McGuire *et al.*, 2007), for the phylogenetic analyses, the best partition scheme and the best evolutionary model for each partition were chosen using PARTITIONFINDER v.

1.1.1 (Lanfear et al., 2012). In this analysis, 12S, 16S genes and each codon position of ND2 gene were defined and Bayesian Inference Criteria (BIC) was used. As a result, the analysis suggested that the best partition scheme is 12S gene/16S gene/each codon position of ND2 gene, and selected TrN+I+G model as the best model for 12S, 16S and the second codon position of ND2 gene and GTR+I+G model as the best model for the other two codon position of ND2 gene. For the ML tree, branch supports were drawn from 10000 non-parametric bootstrap replicates. In BI analyses, the parameters for each partition were unlinked, and branch lengths were allowed to vary proportionately across partitions. Two runs each with four Markov chains were simultaneously run for 50 million generations with sampling every 1000 generations. The first 25% trees were removed as the “burn-in” stage followed by calculations of Bayesian posterior probabilities and the 50% majority-rule consensus of the post burn-in trees sampled at stationarity.

To further visualize the degree of genetic splits among the new taxon and its related species especially members of the *O. schmackeri* species complex recognized by Li et al. (2015), a phylogenetic network using the maximum parsimony method in Spltree v. 4.11.3 (Huson & Bryant, 2006) was constructed based on the ND2 gene sequence dataset. The supports of Spltree lineages were evaluated by 1000 bootstrap replicates.

In addition, to access the genetic divergence between the new taxon and its related species on nuclear DNA, haplotype networks for DOLK and KCNF gene datasets were constructed, respectively, using the maximum parsimony method in TCS v. 1.21 (Clement et al., 2000).

Finally, pairwise uncorrected *p*-distance on the 16S rRNA gene were estimated using MEGA v. 6.06 (Tamura *et al.*, 2011) to evaluate the genetic divergence between *Odorrana* species.

Morphological comparisons

A total of 67 adult specimens including nine males and 16 females of the new taxon, 15 males and four

190 females of *O. schmackeri* and 13 males and ten females of *O. huanggangensis* were measured (for voucher
191 information see Table S1). Ten tadpoles of the new taxon were measured (for voucher information see Table
192 S4). The terminology and methods followed Fei et al. (2009). Measurements were taken with a dial caliper to
193 the nearest 0.1 mm. Twenty one morphometric characters of adult specimens were measured: SVL = snout-
194 vent length (distance from the tip of the snout to the posterior edge of the vent), HDL = head length (distance
195 from the tip of the snout to the articulation of jaw), HDW = maximum head width (greatest width between the
196 left and right articulations of jaw), SL = snout length (distance from the tip of the snout to the anterior corner
197 of the eye), ED = eye diameter (distance from the anterior corner to the posterior corner of the eye), IOD =
198 interorbital distance (minimum distance between the inner edges of the upper eyelids), IND = internasal
199 distance (minimum distance between the inner margins of the external nares), NED = nasal to eye distance
200 (distance between the nasal and the anterior corner of the eye), NSD = nasal to snout distance (distance
201 between the nasal the posterior edge of the vent), IFE = distance between anterior corner of eyes, IAE =
202 distance between posterior corner of eyes, TYD = maximal tympanum diameter, LAL = length of lower arm
203 and hand (distance from the elbow to the distal end of the Finger IV), LW = lower arm width (maximum width
204 of the lower arm), THL = thigh length (distance from vent to knee), TL = tibia length (distance from knee to
205 tarsus), TW = maximal tibia width, TFL = length of foot and tarsus (distance from the tibiotarsal articulation to
206 the distal end of the Toe IV), FL = foot length (distance from tarsus to the tip of fourth toe), FDW = finger disk
207 width (width at the widest part of the disk of finger III), DPW = distal phalanx width (maximal width of distal
208 phalanx of finger III). Ten morphometric characters of larvae were measured: TOL = total length, SVL =
209 snout-vent length, BH = maximum body height, BW = maximum body width, SL = snout length (distance
210 from the anterior corner of the eye to the tip of the snout), SS = snout to spiraculum (distance from spiraculum

to the tip of the snout), MW = mouth width (distance between two corners of mouth), TBW = maximum width of tail base, TL = tail length (distance from base of vent to the tip of tail), TH = tail height (maximum height between upper and lower edges of tail).

In order to reduce the impact of allometry, the correct value from the ratio of each measurement to SVL was calculated and then log-transformed for the following morphometric analyses. One-way analysis of variance (ANOVA) was used to test the significance of differences on morphometric characters between males and females and between different species. The significance level was set at 0.05. To show the spatial distribution of different species on the morphometric characters, principal component analyses (PCA) were performed. These analyses were carried out in the R (R Development Core Team, 2008).

The new species was also compared with all other *Odorrana* species on morphology. Comparative morphological data were obtained from literatures for *O. absita* (Stuart & Chan-ard, 2005), *O. amamiensis* (Matsui, 1994), *O. andersonii* (Boulenger, 1882), *O. anlungensis* (Hu et al., 1973), *O. arunachalensis* (Saikia et al., 2017), *O. aureola* (Stuart et al., 2006), *O. bacboensis* (Bain et al., 2003), *O. banaorum* (Bain et al., 2003), *O. bolavensis* (Stuart & Bain, 2005), *O. cangyuanensis* (Yang, 2008), *O. chapaensis* (Bourret, 1937), *O. chloronota* (Günther, 1876), *O. exiliversabilis* (Li et al., 2001 in Fei et al., 2001b), *O. fengkaiensis* (Wang et al., 2015), *O. geminata* (Bain et al., 2009), *O. gigatympana* (Orlov et al., 2006), *O. grahami* (Boulenger, 1917), *O. graminea* (Boulenger, 1900), *O. hainanensis* (Fei et al., 2001a), *O. hejiangensis* (Deng & Yu, 1992), *O. hosii* (Boulenger, 1891), *O. huanggangensis* (Chen et al., 2010a), *O. heatwolei* (Stuart & Bain, 2005), *O. hmongorum* (Bain et al., 2003), *O. indepressa* (Bain & Stuart, 2005), *O. ishikawae* (Stejneger, 1901), *O. jingdongensis* (Fei et al., 2001a), *O. junlianensis* (Fei & Ye, 2001 in Ye and Fei, 2001), *O. khalam* (Stuart et al., 2005), *O. kuangwuensis* (Liu and Hu, 1966 in Hu et al., 1966), *O. leporipes* (Werner, 1930), *O.*

lipuensis (Mo *et al.*, 2015), *O. livida* (Blyth, 1856), *O. macrotympana* (Yang, 2008), *O. margaretae* (Liu, 1950),
O. mawphlangensis (Pillai & Chanda, 1977), *O. monjerai* (Matsui & Jaafar, 2005), *O. morafkai* (Bain *et al.*,
2003), *O. mutschmanni* (Pham *et al.*, 2016), *O. nanjiangensis* (Fei *et al.*, 2007a), *O. narina* (Stejneger, 1901),
O. nasica (Boulenger, 1903), *O. nasuta* (Li *et al.*, 2001 in Fei *et al.*, 2001b), *O. orba* (Stuart & Bain, 2005), *O.*
rotodora (Yang & Rao, 2008 in Yang, 2008), *O. schmackeri* (Boettger, 1892), *O. sinica* (Ahl, 1927), *O.*
splendida (Kuramoto *et al.*, 2011), *O. supranarina* (Matsui, 1994), *O. swinhoana* (Boulenger, 1903), *O.*
tianmuyii (Chen *et al.*, 2010b), *O. tiannanensis* (Yang & Li, 1980), *O. tormota* (Wu, 1977), *O. trankieni* (Orlov
et al., 2003), *O. utsunomiyaorum* (Matsui, 1994), *O. versabilis* (Liu & Hu, 1962), *O. wuchuanensis* (Xu, 1983
in Wu *et al.*, 1983), *O. yentuensis* (Tran *et al.*, 2008), *O. yizhangensis* (Fei *et al.*, 2007b) and *O. zhaoui* (Li *et al.*,
2008).

Skull scanning

Skulls of two male specimens (voucher number: CIBjs20150803001, CIBjs20150803002) and three
female specimens (voucher number: CIBjs20150804001, CIBGYU20130917003, CIBGYU20130917001) of
the new taxon were scanned in the high-resolution X-ray scanner (Quantum GX micro-CT Imaging System,
PerkinElmer®). The specimens were scanned along the coronal axis at an image resolution of 2000 * 2000.
Segmentation and three-dimensional reconstruction of the CT images were made using VG57 Studio Max 2.2
(Volume Graphics, Heidelberg, Germany).

The Animal Care and Use Committee of Chengdu Institute of Biology, CAS provided full approval for
this purely observational research (Number: CIB2013041102). Field work was approved by the Management
Office of the Lengshuihe Nature Reserve (project number: LSH-201304003).

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RESULTS

Phylogenetic analyses and genetic divergence

Aligned sequence matrix of 12S+16S, ND2, DOLK and KCNF genes contained 1835 bp, 768 bp, 645 bp and 750 bp, respectively. ML and BI analyses based on the 12S+16S+ND2 matrix resulted in essentially identical topologies with high node supporting values (Fig. 2A). All samples of the new taxon occurring from Guizhou Prov. were strongly clustered into a monophyly, which was placed into the genus *Odorrana* and sister to the *O. schmackeri* clade. ND2 splitstree also strongly supported the splits between the new taxon and its related species (Fig. 2B). As note, *Odorrana* sp2 simultaneously revealed by Li et al. (2015) and Zhu (2016) and *Odorrana* sp3 revealed by Li et al. (2015) also occupied an independent lineage, respectively. Only one haplotype was found for all samples of the new taxon, either in KCNF gene or in DOLK gene, and there was no common haplotype between the new species and its related species (Fig. 3).

The genetic distances on 16S gene between the new taxon and its sister species *O. schmackeri* was mean

0.027 (range 0.026–0.028; Table S5), much higher than the intraspecific genetic distance within each of the new taxon, *O. schmackeri*, *O. huanggangensis*, *O. yizhangensis* and *O. lungshengensis* (all intraspecific genetic distance < 0.002). More significantly, it was higher or even much higher than the interspecific genetic distance between many sister species, for example, that between *O. huanggangensis* and *O. tianmuis* (0.014), *O. hainanensis* and *O. fengkaiensis* (0.013), *O. nasuta* and *O. versabilis* (0.013), *O. versabilis* and *O. exiliversabilis* (0.017), *O. morafkai* and *O. banaorum* (0.013), and *O. grahami* and *O. jingdongensis* (0.017; Table S5).

Morphological comparisons

The results of one-way ANOVA showed that in the new taxon, the males was significantly different from the females on SVL and the ratios of ED, IND, IFE, LAL, TYD, LW and TW to SVL (all *p*-values < 0.05; Table 1). So, morphometric analyses were conducted on males and females, respectively. In PCA for males, the total variation of the first two principal components was 42.23%, and in PCA for females, it is 37.37%. In both males (Fig. 4A) and females (Fig. 4B), on the two-dimensional plots of PC1 versus PC2, the new taxon could be almost separated from *O. schmackeri*. The results of one-way ANOVA indicated that either in males or females, the new taxon was significantly different from *O. schmackeri* and *O. huanggangensis* on many morphometric characters (all *p*-values < 0.05; Tables 1). More detailed descriptions of results from morphological comparisons between the new taxon and its congeners were presented in the following sections for describing the new species.

At all, molecular and morphological results supported that our specimens from Guizhou Prov. of China was a new taxon. It is described as a new species in the following sections:

***Odorrana kweichowensis* sp. nov.**

urn:lsid:zoobank.org:act:95123118-30D2-42B5-B5DB-AB7C32B33F97

Holotype

CIBjs20150803002, adult male (Fig. 5A and B; Fig. 6A and B), collected by S.Z. Li in the Lengshuihe Nature Reserve (27.47361°N, 106.00139°E; elevation 754 m a.s.l.), Jinsha Co., Guizhou Prov., China.

Paratypes

24 specimens (nine adult males and 15 adult females), 14 specimens collected by S.Z. Li from the Lengshuihe Nature Reserve in Jinsha Co., Guizhou Prov., China. Six males: CIBjs20150803001, CIBjs20150803003, CIBjs20150803004, CIBjs20150803005 collected on August 3, 2015 and CIBjs20150804002 on August 4, 2015; eight females: CIBjs20150803006, CIBjs20150803007, CIBjs20150803008 collected on August 3, 2015, CIBjs20150804001, CIBjs20150804003, CIBjs20150804004, CIBjs20150804005 on August 4, 2015 and CIBjs20150808020 on August 8, 2015. Seven specimens collected by S.Z. Li and J. C. Lv from Meitan Co. of Guizhou Prov., China. Three males: CIBGYU20130917004, CIBGYU20130917005 and CIBGYU20130917007 collected by J.C. Lv on September 17, 2013; four females: CIBGYU20130917001, CIBGYU20130917002 and CIBGYU20130917003 by S.Z. Li on September 17, 2013, CIBGYU20130917006 by J.C. Lv on September 17, 2013. Three females: GYU20130921001, CIBGYU20130921002 and CIBGYU20130921003 collected by S.Z. Li on September 21, 2013. Ten tadpoles (CIBjs20171014001-CIBjs20171014010) collected by S.Z. Li on October 10, 2017.

Diagnosis

Odorrana kweichowensis **sp. nov.** is assigned to genus *Odorrana* based upon molecular phylogenetic analyses and the following morphological characters: dorsum is green; tips of digits dilated, tapering, disks with circum-marginal grooves, and vertical diameter longer than horizontal diameter in the disks;

supernumerary tubercle below the base of fingers III and IV; feet fully webbed to disks, without tarsal fold; the first finger thick and nuptial pad distinct.

Odorrana kweichowensis **sp. nov.** could be distinguished from its congeners by a combination of the following characters: (1) having smaller body size in males (snout-vent length < 43.3 mm vs. snout-vent length > 48 mm in many other species); (2) head longer than wider; (3) dorsolateral folds absent; (4) tympanum of males large and distinct, tympanum diameter in males twice as long as width of distal phalanx of finger III; (5) two metacarpal tubercles; (6) relative finger lengths: II < I < IV < III; (7) tibiotarsal articulation reaching to the level between eye to nostril when leg stretched forward; (8) disks on digits with circum-marginal grooves; (9) toes fully webbed to disks; (10) the first subarticular tubercle on fingers weak; (11) having white pectoral spinules, paired subgular vocal sacs located at corners of throat, light yellow nuptial pad on the first finger in males.

Description of holotype

Head longer than wide ($HDL/HDW = 1.08$); top of head flat; snout obtusely rounded in dorsal view ($SL/HDL = 0.41$), rounded in profile, projecting beyond lower jaw; eye large and convex, ED 0.83 times of SL ; $IND = 5.1$ mm, larger than $IOD = 3.2$ mm; $NED = 3.4$ mm larger than $NSD = 2.9$ mm; tympanum circular, large and distinct, twice as long as width of distal phalanx of finger III ($TYD/FDW = 2.84$); vomerine teeth on well-developed ridges; tongue deeply notched posteriorly; paired gular pouches at corners of throat. Forelimbs moderately robust ($LW/SVL = 0.11$); lower arm and hand beyond a half of body length ($LAL/SVL = 0.53$); the first finger slightly longer than the second; finger tips on I–IV dilated to wide, tapering, disks with circum-marginal grooves; nuptial pad on the inner of first finger from base to subarticular tubercles; subarticular tubercles relatively prominent; inner metacarpal tubercle oblong and outer metacarpal tubercle indistinct; no

finger webbing.

Hindlimbs long; tibio-tarsal articulation reaching to the level between eye to nostril when hindlimb adpressed along the side of body; heels overlapping when hindlimbs positioned at right angle to body; singular longer than thigh length; FL 0.85 times SVL; toes slender, relative toe lengths: $I < II < III < V < IV$; toes tapering dilated, disks with circum-marginal grooves; feet fully webbed to disks; web becoming narrower and continuing to the disks as lateral fringe on toes II, III and IV; outer metatarsal tubercle absent; inner metatarsal tubercle present.

Skin shagreened, a number of pustules scattered on dorsum of trunk and flanks; several irregular tubercles scattered on flanks; dorsolateral folds absent; two large glands on the place between below tympanum and above arm; white spinules on throat, chest, fore abdomen, inner side of forearms and upper and lower lips.

Coloration in life

In life, dorsum with bright green network ; large black spots in the center on dorsum, continuing onto dorsal portion of legs to form dark counter-point to bright green irregular bands; flank light yellow with several black spots; tympanum brown-black; upper and lower lip with vertical black bars; arms brown with black transverse bands, thighs with five brown bands and tibiae with six (Fig. 6).

Color in preservative

On dorsum color fades to dark olive with dark brown blotches (Fig. 5A), upper and lower lips marbled brown-black on dirty white; venter variable from white to light yellow; underside of limbs yellowish with white (Fig. 5B); pinkish in the inner side of forearm and thighs; there is a white spot between front corner of eyes; nuptial pad fades to white (Fig. 5).

Variation

Basic statistics for measurements were presented in Table S6. All male specimens were similar in morphology and color pattern, but different from females. SVL in females approximately 1.8 times that in males (SVL mean 73.6 mm, range 62.4–81.1 mm in females, SVL mean 41.0 mm, range 36.2–43.3 mm in males); the ratios of ED, IND, IFE, LAL, TYD and LW to SVL of males are significantly higher than that of females, but the ratios of TW to SVL of males is lower than that of females (Table 1). In some adult females, the black transverse bands on limbs are not obvious (Fig. 6C). In some adult females, chest, chin and ventral surface of limbs scattered with continued light-brown spots (Fig. 7A). Some adult females have black cloud stripe with irregular borders on the ventral side (Fig. 7B), but some don't have (Fig. 6D). In some adult females, dorsum uniform brown with little green impression (Fig. 7C) and some adult females have dorsum uniform brown lacking green impression (Fig. 7D).

Skull description

The skull morphology of the five scanned specimens were almost identical, and thus, only one representative (voucher number: CIBjs20150803001) was presented (Fig. 8) and described as following: skull flat, maxillary teeth well developed, vomerine teeth present; mandible without teeth; nasals large, widely separated from each other, nasals disconnected with the sphenethmoid; sphenethmoid tubular, forms the anterolateral walls of the braincase; frontoparietal roof the braincase and wider posteriorly than anteriorly; prootic large and connect with the exoccipital; a pair of exoccipitals situated the end of the brain; palatines arcuate and long, behind the prevomers; a pair of prevomers obliquely lie anterior to the palatines, vomerine teeth on the prevomers indistinct; parasphenoid in sword shape, supports the braincase ventrally, connects with palatines; a pair of squamosals on the dorsolateral side of the prootic, each squamosal consists of three rami: the zygomatic ramus, the otic ramus, and the ventral ramus; zygomatic ramus pointing to the orbit, otic ramus

shorted, ventral ramus outboard and covered the posterior of pterygoid; a pair of pterygoids, outside of the ventral surface of the squamosal, anterior ramus is the longest, center is inward and leading edge extends to the orbit, medial ramus shorted and attached to the anterior lateral part of the prootic, posterior ramus edge-on and extend to maxillary; a pair of columellae situated ventral to the crista parotica (Fig. 8).

Tadpole description

Body and tail yellowish-brown (Fig. 9A), at Gosner's stage 28-29, TOL 31.6–36.2 mm, SVL 12.1–14.5 mm, other measurements of tadpoles shown in Table S4. Tail 1.5–1.8 times as long as body; TH 25–30% of TL in the 28th stage and 23–28.5% in the 29th stage; BW 45.8% of SVL in the 28th stage and 46.8% in the 29th stage; BH 31.4% of SVL in the 28th stages and 34.9% in the 29th stage; tail fins lightly colored, tail muscles with small black spots; tail depth greater than body depth, dorsal fin arising behind the origin of the tail; maximum depth near mid-length, tip of tail blunt; nostril near snout, eyes positioned dorsally (Fig. 9A); spiracle on the middle left of body (Fig. 9B); keratodont formula I: 3-3/ III: 1-1 (Fig. 9C); both upper and lower lips with labial papillae (Fig. 9C); some additional tubercles at the angles of the mouth, usually with small keratodonts (Fig. 9C).

Sexual dimorphism

SVL in females approximately 1.8 times that in males (SVL mean 73.6 mm, range 62.4–81.1 mm in females; SVL mean 41.0 mm, range 36.2–43.3 mm in males); paired subgular vocal sacs located at corners of throat, pinkish lineae musculinae on dorsal side, nuptial pad light yellow on the inner of first finger from base to subarticular tubercles in males (Figs. 5, 6).

Comparisons

Odorrana kweichowensis **sp. nov.** differs from *O. amamiensis*, *O. andersonii*, *O. bacboensis*, *O.*

400 *cangyuanensis*, *O. chapaensis*, *O. geminata*, *O. graham*, *O. hmongorum*, *O. ishikawae*, *O. jingdongensis*, *O.*
401 *junlianensis*, *O. kuangwuensis*, *O. macrotympana*, *O. margaretae*, *O. mawphlangensis*, *O. mutschmanni*, and
402 *O. wuchuanensis*, by having smaller body size (SVL < 43.30 mm in males in the new species vs. SVL > 48
403 mm in males in other species).

404 *Odorrana kweichowensis* **sp. nov.** differs from *O. absita*, *O. aureola*, *O. banaorum*, *O. bolavensis*, *O.*
405 *exiliversabilis*, *O. gigatympana*, *O. graminea*, *O. hosii*, *O. heatwolei*, *O. indepressa*, *O. khalam*, *O. leporipes*,
406 *O. livida*, *O. monjerai*, *O. narina*, *O. nasica*, *O. nasuta*, *O. orba*, *O. supranarina*, *O. tormota*, *O. trunkieni*, *O.*
407 *utsunomiyaorum*, *O. versabilis*, *O. yentuensis* and *O. zhaoi*, by lacking dorsolateral folds (vs. present in other
408 species).

409 *Odorrana kweichowensis* **sp. nov.** differs from *O. fengkaiensis* by tibiotarsal articulation reaching the
410 level between eye to nostril when leg stretched forward (vs. reaching the level below nostril in the latter); from
411 *O. rotodoar* by tibiotarsal articulation reaching the level between eye to nostril when leg stretched forward (vs.
412 reaching the level beyond eye in the latter); from *O. lipuensis* (vocal sacs absence in males) and *O. hainanensis*
413 (paired internal vocal sacs in males) by paired external vocal sacs located at corners of throat; from *O.*
414 *chloronota* by having head longer than wide (vs. head wider than long in the latter); from *O. morafkai* and *O.*
415 *sinica* by having white pectoral spinules in mature males (vs. absence in the latter); from *O. arunachalensis*, by
416 the relative finger lengths I < II < IV < III (vs. II < I < IV < III in the latter).

417 Within the *O. schmackeri* group (*O. anlungensis*, *O. hejiangensis*, *O. lungshengensis*, *O. nanjiangensis*, *O.*
418 *swinhoana*, *O. tianmuyi*, *O. tiannanensis*, *O. yizhangensis*, *O. huanggangensis* and *O. schmackeri*), *Odorrana*
419 *kweichowensis* **sp. nov.** differs from *O. anlungensis* by having white pectoral spinules in mature males (vs.
420 absence in the latter), tibiotarsal articulation reaching the level between eye to nostril when leg stretched

421 forward (vs. reaching the nostril in the latter), web becoming narrower and continuing to the disks of toe IV
 422 (vs. web continuing to the subarticular tubercles of toe IV in the latter); from *O. hejiangensis* by the disks of
 423 fingers with circum-marginal grooves (vs. without circum-marginal grooves on finger I in the latter), two
 424 metacarpal tubercles (vs. three metacarpal tubercles in the latter); from *O. lungshengensis* by the body size of
 425 males with maximum SVL 43.3 mm (vs. SVL 50.0–59.6 mm of males in the latter), tympanum large and
 426 distinct, and beyond twice as long as width of distal phalanx of finger III (vs. tympanum as long as width of
 427 distal phalanx of finger III in the latter), two metacarpal tubercles (vs. without outer metacarpal tubercle in the
 428 latter), tibiotarsal articulation reaching the level between eye to nostril when leg stretched forward (vs.
 429 reaching the tip of snout in the latter); from *O. nanjiangensis* by the body size in males with maximum SVL
 430 43.3 mm (vs. SVL 59.5–66.8 mm in males in the latter), two metacarpal tubercles (vs. without outer
 431 metacarpal tubercle in the latter), tibiotarsal articulation reaching the level between eye to nostril when leg
 432 stretched forward (vs. reaching the tip of snout in the latter); from *O. swinhoana* by the body size in males with
 433 maximum SVL 43.3 mm (vs. SVL 47.7–71.6 mm in males in the latter), tibiotarsal articulation reaching the
 434 level between eye to nostril when leg stretched forward (vs. reaching the tip of snout); from *O. tianmuui* by the
 435 relative finger lengths $II < I < IV < III$ (vs. $I < II < IV < III$ in the latter), two metacarpal tubercles (vs. three
 436 outer metacarpal tubercles in the latter); *O. tiannanensis* by the body size in males with maximum SVL 43.3
 437 mm (vs. SVL 52.5–53.5 mm in males in the latter), tibiotarsal articulation reaching the level between eye to
 438 nostril when leg stretched forward (vs. reaching the level beyond the tip of snout in the latter); *O. yizhangensis*
 439 by the body size in males with maximum SVL 43.3 mm (vs. SVL 47.3–54.0 mm in males in the latter), two
 440 metacarpal tubercles (vs. without outer metacarpal tubercle), tibiotarsal articulation reaching the level between
 441 eye to nostril when leg stretched forward (vs. reaching the tip of snout in the latter); from *O. huanggangensis*

by the relative finger lengths $II < I < IV < III$ (vs. $I < II < IV < III$ in the latter), tibiotarsal articulation reaching the level between eye to nostril when leg stretched forward (vs. reaching the nostril in the latter); from *O. schmackeri*, by the first subarticular tubercle on fingers and supernumerary tubercles being weak (vs. being outstanding in the latter; Fig. 5E, F), and having two outer metacarpal tubercles (vs. outer metacarpal tubercle being indistinct in the latter; Fig. 5H, G).

Odorrana kweichowensis **sp. nov.** differs from *O. schmackeri* by having significantly lower ratios of HDL and SL to SVL in males, having significantly higher ratios of THL, TL and FL to SVL in males, and having significantly higher ratios of IND, THL, TL and FL to SVL in females (all p -values < 0.05 ; Table 1; Table S6); from *O. huanggangensis* by having significantly lower ratios of NSL and IAE to SVL in males, having significantly higher ratios of IND, LAL, THL and TFL to SVL in males, and having significantly higher ratios of LAL, THL, TL and FL to SVL in females (all p -values < 0.05 ; Table 1; Table S6).

Ecology

To present, *Odorrana kweichowensis* sp. nov. has been found in three localities: Lengshuihe Nature Reserve ($27^{\circ}34' - 27^{\circ}26' \text{ N}$, $105^{\circ}57' - 106^{\circ}04' \text{ E}$) in Jinsha Co., Meitan Co. ($27^{\circ}39' - 27^{\circ}42' \text{ N}$, $107^{\circ}33' - 107^{\circ}35' \text{ E}$) and Zheng'an Co. ($28^{\circ}09' - 28^{\circ}20' \text{ N}$, $107^{\circ}30' - 107^{\circ}38' \text{ E}$) in Guizhou Prov. of China. Geographical distances between these localities were from 89 km to 173 km. Population from the Lengshuihe Nature Reserve inhabited broad streams, and near the riparian areas, surrounded by evergreen broadleaved forests (Fig. 10A). Populations from Meitan Co. and Zheng'an Co. inhabited broad slow-flowing rivers surrounded by paddy field (Fig. 10B and C). All of the localities were at elevations 717-766 m. All adult individuals that we found appear on the stones in the streams at night (07:30-12:00 pm) with water pH 6.8-7.1 and water temperature 15-23 °C. Tadpoles could be found at daytime and night. Amplexed individuals could be found in the streams in the type

locality (Fig. 10A). Three sympatric amphibian species *Fejervarya multistriata*, *Rana zhenhaiensis* and *Polypedates megacephalus* were found in Meitan Co. and Zheng'an Co., but only one sympatric amphibian species *Amolops chunganensis* was found in the Lengshuihe Nature Reserve in the type locality.

Etymology

The specific epithet “*kweichowensis*” refers to the distribution of this species, Guizhou Prov., China. The “kweichow” is an old spelling and a transliteration for “Guizhou”. We propose the common English name “Guizhou Odorous Frog” for this species.

DISCUSSION

Morphological similarity of related species in the genus *Odorrana* challenges classifications in the group (Fei et al., 2009; Chen et al., 2013). Integrative taxonomy with multiple evidences, such as genetic divergence, ecological discrepancy, morphometric differences, and so on, turns out to be quite effective, and have already become the main trend. In this study, based on molecular phylogenetic analyses and morphological comparisons, a new species, *Odorrana kweichowensis* sp. nov., was described from mountain streams in Guizhou Prov., China. Phylogenetic analyses based on mitochondrial DNA suggested that the new species belonged to *Odorrana* but was significantly separated from its congeners. Genetic distance based on 16S rRNA gene between it and its sister species *O. schmackeri* was mean 0.027, matching the level about interspecific divergences in amphibians (0.01-0.17; Vences et al. 2005) and being much higher than that between many sister species (of which, most species have been completely recognized as valid species) in *Odorrana*. Moreover, on nuclear protein-coding genes which presented much lower evolutionary rates, the new species was still indicated to occupy an independent lineage separated from other species. These results

confirmed restricted even absent gene flow between the new species and its related species. Finally, the new species was different from its congeners on a lot of morphological characters. At all, multiple evidences supported the validity of the new species.

In this study, *Odorrana kweichowensis* sp. nov. was found in three localities in the northwestern part of Guizhou Prov. of China (Fig. 1). In Li et al. (2015), the new species was only found in two localities in Guizhou Prov.: one was the type locality (Lengshuihe Reserve in Jinsha Co.) of it, and another was the Suiyang Co. closely adjoining Meitan Co. and Zheng'an Co. included in this study. Obviously, it could be speculated that the new species was at least distributed in the northwestern part of Guizhou Prov. of China (Fig. 1). Unfortunately, it was still not sure whether the new species was conspecific with *Odorrana* sp1 recognized by Zhu et al. (2016) in absence of corresponding specimens and sequences. If they were conspecific, the new species should be distributed in a larger range from northern Guizhou Prov. to southern Guangxi Prov., China. In addition, it could not infer whether the populations in northern Thailand and northern Vietnam reported as *O. schmackeri* (Chan-ard, 2003; Orlov et al., 2002; Nguyen et al., 2005; Fig. 1) was conspecific with the new species, because there was also no molecular data and detailed morphological descriptions of them. As note, according to previous studies (Li et al., 2015; Zhu, 2016) and our results, the new species was possibly sympatric with Clade D (*Odorrana* sp2) recognized by Zhu (2016) and *O. hejiangensis* in a narrow area along the boundary between northern Guizhou Prov. and Sichuan Prov. and Chongqing City, China (Fig. 1), though in the localities investigated in this study, we did not find the latter two species in the microhabitats of the new species.

CONCLUSION

We described a new species of the odorous frog genus *Odorrana* (Amphibia, Anura, Ranidae) from Guizhou Prov. of China, and provide evidence for its phylogenetic allocations. *Odorrana kweichowensis* sp. nov. was only known from a narrow range in the northwestern part of Guizhou Prov. of China, and occurred from mountain streams at mid and low elevations similar to most odorous frogs. In our fieldwork, the new species was found to be seriously threatened by local villager and construction of dams and roads. Thus, further more detailed investigations on the species are urgent to ascertain its distributional range and population status. With our description we contributed to a better knowledge of the diversity of the genus *Odorrana* in the southwestern China, and thus suggested that more comprehensive phylogeographic studies would highlight radiation patterns of the group.

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REFERENCES

- Ahl E. 1927 "1925". Über vernachlässigte Merkmale bei Fröschen. *Sitzungsberichte der Gesellschaft Naturforschender Freunde zu Berlin* 1925: 40–47
- Bain RH, Lathrop A, Murphy RW, Orlov NL, Cuc HT. 2003. Cryptic species of a cascade frog from Southeast Asia: taxonomic revisions and descriptions of six new species. *American Museum Novitates* 3417: 1–60
- Bain RH, Stuart BL, Nguyen TQ, Che J, Rao DQ. 2009. A New *Odorrana* (Amphibia: Ranidae) from

- 526 Vietnam and China. *Copeia* **2**: 348–362
- 527 **Bain RH, Stuart BL. 2005.** A new species of cascade frog (Amphibia: Ranidae) from Thailand, with new data
- 528 on *Rana banaorum* and *Rana morafkai*. *Natural History Bulletin of the Siam Society* **53**(1): 3–16
- 529 **Blyth E. 1856 "1855".** Report for October Meeting, 1855. *Journal of the Asiatic Society of Bengal* **24**: 711–
- 530 723
- 531 **Boettger O. 1892.** Katalog der Batrachier-Sammlung im Museum der Senckenbergischen Naturforsch
- 532 Gesellsch, *Frankfurt am Main*, 1–73
- 533 **Boulenger GA. 1882.** Calalogue of the Batrachia Salientias. Ecaudata in the Collection of the British Museum.
- 534 London
- 535 **Boulenger GA. 1891.** On new or little-known Indian and Malayan reptiles and batrachians. *Annals and*
- 536 *Magazine of Natural History*, Series 6, **8**: 288–292
- 537 **Boulenger GA. 1900 "1899".** On the reptiles, batrachians, and fishes collected by the late Mr. John
- 538 Whitehead in the interior of Hainan. *Proceedings of the Zoological Society of London* **1899**: 956–962
- 539 **Boulenger GA. 1903.** Descriptions of new batrachians in the British Museum. *Annals and Magazine of*
- 540 *Natural History*, London, **12**(7): 552–557
- 541 **Boulenger GA. 1917.** Descriptions of new frogs of the genus *Rana*. *Annals and Magazine of Natural History*,
- 542 London, **20**(8): 413–418
- 543 **Bourret R. 1937.** Notes herpétologiques sur l’Indochinese francaise. XIV. Les batrachiens de la collection du
- 544 Laboratoire des Sciences naturelles de l’Université. Descriptions de quinze espèces ou variétés nouvelles.
- 545 *Annexe au Bulletin Général de l’Instruction Publique*, **4**: 5–56
- 546 **Cai HX, Che J, Pang JF, Zhao EM, Zhang YP. 2007.** Paraphyly of Chinese *Amolops* (Anura, Ranidae) and

- phylogenetic position of the rare Chinese frog, *Amolops tormotus*. *Zootaxa* **1531**: 49–55
- Castresana J. 2000.** Selection of conserved blocks from multiple alignments for their use in phylogenetic analysis. *Molecular Biology and Evolution* **17**: 540–552
- Chan-ard T. 2003.** A Photographic Guide to Amphibians in Thailand [In Thai]. Bangkok, Thailand: Darnsutha Press Co., Ltd.
- Che J, Pang JF, Zhao H, Wu GF, Zhao E M, Zhang YP. 2007.** Phylogeny of Raninae (Anura: Ranidae) inferred from mitochondrial and nuclear sequences. *Molecular Phylogenetics and Evolution* **43**: 1–13
- Chen XH, Chen Z, Jiang JP, Qiao L, Lu YQ , Zhou KY, Zheng GG , Zhai XF, Liu JX. 2013.** Molecular phylogeny and diversification of the genus *Odorrana* (Amphibia, Anura, Ranidae) inferred from two mitochondrial genes. *Molecular Phylogenetics and Evolution* **69**: 1196–1202
- Chen XH, Zhou KY, Zheng GM. 2010a.** A new species of odor frog from China (Anura: Ranidae). *Journal of Beijing Normal University (Natural Science)* **46**(5): 606–609
- Chen XH, Zhou KY, Zheng GM. 2010b.** A new species of the genus *Odorrana* from China (Anura, Ranidae). *Acta Zootaxonomica Sinica* **35**(1): 206–211
- Clement M, Posada D, Crandall K. 2000.** TCS: a computer program to estimate gene genealogies. *Molecular Ecology* **9**: 1657–1660
- Deng QX, Yu ZW. 1992.** A new species of the genus *Rana* form China. *Journal of Sichuan Teacher College* **13**(4): 323–326
- Fei L, Hu SQ, Ye CY, Huang YZ. 2009.** Fauna Sinica, Amphibia Vol. 3, Anura Ranidae. Beijing: Science Press
- Fei L, Ye CY, Huang YZ. 1990.** Key to Chinese Amphibians. Chongqing Branch Sci. Technol. Literature

Press, Chongqing

Fei L, Ye CY, Jiang JP. 2007b. A new Ranidae frog species China *Odorrana (Odorrana) yizhangensis*

(Ranidae: Anura). *Acat Zoological Sinica* **32**(4): 989–992

Fei L, Ye CY, Jiang JP. 2012. Colored atlas of Chinese Amphibians and their distributions. Chengdu:

Sichuan Publishing House of Science and Technology

Fei L, Ye CY, Li C. 2001a. Descriptions of two new species of the genus *Odorrana* in China (Anura:

Ranidae). *Acta Zootaxonomica Sinica* **26**(1): 108–114

Fei L, Ye CY, Li C. 2001b. Taxonomic studies of *Odorrana versabilis* in China II. Descriptions of two new

species (Amphibian: Ranidae). *Acta Zootaxonomica Sinica* **26**(4): 601–607

Fei L, Ye CY, Xie F. 2007a. A new Ranidae frog species from Sichuan, China *Odorrana (Odorrana)*

nanjiangensis (Ranidae: Anura). *Zoological Research* **28**: 551–555

Fei L, Ye CY. 2001. The color handbook of the amphibians of Sichuan. Beijing: Chinese Forestry Press

Frost DR. 2018. Amphibian Species of the World Version 6.0. American Museum of Natural History, New

York, USA. Available from: <http://research.amnh.org/vz/herpetology/amphibia/> (accessed 11 March 2018)

Gosner K L. 1960. A Simplified Table for Staging Anuran Embryos and Larvae with Notes on Identification.

Herpetologica **16**(3): 183–190

Guindon S, Dufayard JF, Lefort V, Anisimova M, Hordijk W, Gascuel O. 2010. New algorithms and

methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0.

Systematic Biology **59**(3): 307–321

Günther ACLG. 1876 "1875". Third report on collections of Indian reptiles obtained by the British Museum.

Proceedings of the Zoological Society of London **1875**: 567–577

589 **Hall TA. 1999.** BIOEDIT: a user-friendly biological sequence alignment editor and analysis program for
590 Windows 95/98/NT. *In Nucleic acids symposium series* **41**: 95–98

591 **He YX. 2017.** Molecular phylogeny of the genus *Odorrana* inferred from mitochondrial and multiple nuclear
592 genes. Henan Normal University

593 **Hu SX, Zhao EM, Liu CZ. 1966.** A herpetological survey of the Tsinling and Ta-pa shan region. *Acat*
594 *Zoological Sinica* **18**(1): 57–89

595 **Hu SX, Zhao EM, Liu CZ. 1973.** A survey of amphibians and reptiles in Kweichow province, including a
596 herpetofauna analysis. *Acat Zoological Sinica* **19**(2): 149–171

597 **Huson DH, Bryant D. 2006.** Application of phylogenetic networks in evolutionary studies. *Molecular Biology*
598 *and Evolution* **23**: 254–267

599 **Kocher TDWK, Thomas A. 1989.** Dynamics of mitochondrial DNA evolution in mammals: Amplification
600 and sequencing with conserved Primers. *Proceedings of the National Academy of Sciences of the United*
601 *States of America* **86**: 6169–6200

602 **Kuramoto M, Satou N, Oumi S, Kurabayashi A, Sumida M. 2011.** Inter- and intra-island divergence in
603 *Odorrana ishikawae* (Anura, Ranidae) of the Ryukyu Archipelago of Japan, with description of a new
604 species. *Zootaxa* **2767**: 25–40

605 **Lanfear R, Calcott B., Ho SY, Guindon S. 2012.** Partitionfinder: combined selection of partitioning schemes
606 and substitution models for phylogenetic analyses. *Molecular Biology and Evolution* **29**: 1695–1701

607 **Lemmon AR, Moriarty EC. 2004.** The importance of proper model assumption in Bayesian phylogenetics.
608 *Systematic Biology* **53**: 265–277

609 **Li PP, Lu YY, Rao DQ. 2008.** A new species of cascade frog (Amphibia, Ranidae) from Tibet, China. *Acta*

- 610 *Zootaxonomica Sinica* **33**(3): 537–541
- 611 **Li YM, Wu XY, Zhang HB, Yan P, Xue H, Wu XB. 2015.** Vicariance and its impact on the molecular
- 612 ecology of a Chinese Ranid frog species-complex (*Odorrana schmackeri*, Ranidae). *PLOS ONE* **10**(9):
- 613 e0138757
- 614 **Liu CC, Hu SQ.1961.** Chinese Amphibians (Anura). Beijing: Science Press
- 615 **Liu CZ, Hu SQ. 1962.** A survey of amphibians and reptiles in Guangxi province, *Acat Zoological Sinica* **14**:
- 616 73–104
- 617 **Liu CZ. 1950.** Amphibian of western China. *Fieldiana: Zoology Museum, Chicago* **2**: 1–400
- 618 **Matsui M, Jaafar I. 2006.** A new cascade frog of the subgenus *Odorrana* from peninsular Malaysia.
- 619 *Zoological Science* **23**(7): 647–651
- 620 **Matsui M, Shimada T, Ota H, Tanaka-Ueno T. 2005.** Multiple invasions of the Ryukyu Archipelago by
- 621 Oriental frogs of the subgenus *Odorrana* with phylogenetic reassessment of the related subgenera of the
- 622 genus *Rana*. *Molecular Phylogenetics and Evolution* **37**: 733–742
- 623 **Matsui M. 1994.** A taxonomic study of the *Rana narina* complex, with description of three new species
- 624 (Amphibia: Ranidae). *Zoological Journal of the Linnean Society* **111**: 385–415
- 625 **McGuire JA, Witt CC, Altshule DL, Remsen JV. 2007.** Phylogenetic systematics and biogeography of
- 626 hummingbirds: Bayesian and maximum likelihood analyses of partitioned data and selection of an
- 627 appropriate partitioning strategy. *Systematic Biology* **56**: 837–856
- 628 **Mo YM, Chen WC, Wu HY, Zhang W, Zhou SC. 2015.** A new species of *Odorrana* inhabiting complete
- 629 darkness in a karst cave in Guangxi, China. *Asian Herpetological Research* **6**(1): 11–17
- 630 **Ngo A, Murphy RW, Liu WZ, Orlov NL. 2006.** The phylogenetic relationships of the Chinese and

- 631 Vietnamese waterfall frogs of the genus *Amolops*. *Amphibia Reptilia* **27**(1): 81–92
- 632 **Nguyen SV, Ho CT, Nguyen TQ. 2005.** Danh Luc Ech Nhai Va Bo Sat Viet Nam/A Checklist of the
- 633 Amphibians and Reptiles of Vietnam. Hanoi: Nha Xuat Ban Hong Nghiep
- 634 **Orlov NL, Natalia BA, Cuc HT. 2006.** A new cascade frog (Amphibia: Ranidae) from central Vietnam.
- 635 *Russian Journal of Herpetology* **13**(2): 155–163
- 636 **Orlov NL, Ngat LN, Cuc HT. 2003.** A new species of cascade frog from north Vietnam (Ranidae, Anura).
- 637 *Russian Journal of Herpetology* **10**(2): 123–134
- 638 **Orlov, NL, Murphy RW, Ananjeva NB, Ryabov SA, Ho CT. 2002.** Herpetofauna of Vietnam, a checklist.
- 639 Part 1. Amphibia. *Russian Journal of Herpetology* **9**: 81–104
- 640 **Pham CT, Nguyen TQ, Le MD, Bonkowski M, Ziegler T. 2016.** A new species of *Odorrana* (Amphibia:
- 641 Anura: Ranidae) from Vietnam. *Zootaxa* **4084**: 421–435
- 642 **Pillai RS, Chanda SK. 1977.** Two new species of frogs (Ranidae) from Khasi Hills, India. *Journal of the*
- 643 *Bombay Natural History Society* **74**: 136–140
- 644 **Pyron RA, Wiens JJ. 2011.** A large-scale phylogeny of amphibia including over 2800 species, and a revised
- 645 classification of extant frogs, salamanders, and caecilians. *Molecular Phylogenetics and Evolution* **61**:
- 646 543–583
- 647 **R Development Core Team. 2008.** R: a language and environment for statistical computing. R Foundation for
- 648 Statistical Computing. Vienna: Available at <http://www.R-project.org>.
- 649 **Ronquist FR, Huelsenbeck JP. 2003.** MrBayes3: Bayesian phylogenetic inference under mixed models.
- 650 *Bioinformatics* **19**(12): 1572–1574
- 651 **Saikia B, Sinha B, Kharkongor IJ. 2017.** *Odorrana arunachalensis*: A new species of Cascade Frog (Anura:

- Ranidae) from Talle Valley Wildlife Sanctuary, Arunachal Pradesh, India. *Journal of Bioresources*.
- Arunachal Pradesh, India **4**: 30–41
- Sambrook J, Fritsch EF, Maniatis T. 1989.** Molecular Cloning: A Laboratory Manual. New York: Cold
- Spring Harbor Laboratory Press
- Shen XX, Liang D, Feng YJ, Chen MY, Zhang P. 2013.** A versatile and highly efficient toolkit including
- 102 nuclear markers for vertebrate phylogenomics, tested by resolving the higher level relationships of
- the Caudata. *Molecular Biology and Eevolution* **30**(10): 2235–2248
- Simon C, Frari F, Beckenbach A, Crespi B, Liu H, Flook P. 1994.** Evolution, weighting and Phylogenetic
- utility of mitochondrial gene sequences and a compilation of conserved Polymerase chain reaction
- Primers. *Annals of the Entomological Society of America* **87**(6): 651–701
- Stejneger L. 1901.** Diagnoses of eight new batrachians and reptiles from the Riu Kiu Archipelago, Japan.
- Proceedings of the Biological Society of Washington* **14**: 189–191
- Stuart BL, Bain RH. 2005.** Three new species of spinule-bearing frogs allied to *Rana megatypanum* Bain,
- Lathrop, Murphy, Orlov and Ho, 2003 from Laos and Vietnam. *Herpetologica* **61**: 478–492
- Stuart BL, Chan-ard T. 2005.** Two new *Huia* (Amphibia: Ranidae) from Laos and Thailand. *Copeia* **2**: 279–
- 289
- Stuart BL, Chuaynkern Y, Chan-ard T, Inger RF. 2006.** Three new species of frogs and a new tadpole
- from eastern Thailand. *Fieldiana Zoology*, new series **111**: 1–19
- Stuart BL, Orlov NL, Chan-ard T. 2005.** A new cascade frog (Amphibia: Ranidae) from Laos and
- Vietnam. *The Raffles Bull Zool* **53**: 125–131
- Stuart BL. 2008.** The phylogenetic problem of Huia (Amphibia: Ranidae). *Molecular Phylogenetics and*

- 673 *Evolution* **46**: 49–60
- 674 **Tamura K, Stecher G, Peterson D, Fiipiski A, Kumar S. 2011.** MEGA6: molecular evolutionary genetics
- 675 analysis using evolutionary distance. *Molecular Biology and Evolution* **28**: 2725–2729
- 676 **Tran TT, Orlov NL, Nguyen TT. 2008.** A new species of Cascade Frog of *Odorrana* Fei, Yi *et* Huang, 1990
- 677 genus (Amphibia: Anura: Ranidae) from Bac Giang Province (Yen Tu Mountain Range, northeast
- 678 Vietnam). *Russian Journal of Herpetology* **15**: 212–224
- 679 **Vences M, Thomas M, Meijden A, Chiari Y, Vieites DR. 2005.** Comparative performance of the 16S rRNA
- 680 gene in DNA barcoding of amphibians. *Frontiers in Zoology* **2**: 5
- 681 **Wang YY, Lau N, Yang JH, Chen GL, Liu ZY, Pang H, Liu Y. 2015.** A new species of the genus
- 682 *Odorrana* (Amphibia: Ranidae) and the first record of *Odorrana bacboensis* from China. *Zootaxa*
- 683 **3999**(2): 235–254
- 684 **Werner F. 1930.** *Rana leporipes*, a new species of frog from South China, with field notes by R. Mell.
- 685 *Lingnan Science Journal*. Canton **9**: 45–47
- 686 **Wiens JJ, Sukumaran J, Pyron RA, Brown RM. 2009.** Evolutionary and biogeographic origins of high
- 687 tropical diversity in old world frogs (Ranidae). *Evolution* **63**(5): 1217–1231
- 688 **Wu GF. 1977.** A new species of frogs from Huang-Shan, Anhui, *Amolops tormotus* Wu. *Acta Zoologica*
- 689 *Sinica* **23**: 113-115
- 690 **Wu L, Xu RH, Dong Q, Li DJ, Liu JS. 1983.** A new species of *Rana* and records of amphibians from
- 691 Guizhou province. *Acta Zootaxonomica Sinica* **29**(1): 66–70
- 692 **Yang DT, Li SM. 1980.** A New Species of the Genus *Rana* from Yunnan. *Zoological Research* **1** (2): 261–264
- 693 **Yang DT. 2008.** Amphibia and Reptilia of Yunnan. Kunming: Yunnan Publishing Group Corporation,

694 Yunnan Science and Technology Press

695 **Ye CY, Fei L. 2001.** Phylogeny of genus *Odorrana* (Amphibian: Randiae) in China. *Acta Zoologica Sinica*

696 **47(5):** 528–534

697 **Zhu YJ.** 2016. Genetic differentiation of *odorrana schmackeri* species complex. Henan Normal University

Table 1(on next page)

The results of one-way ANOVA with p -values for morphometric comparisons between *Odorrana kweichowensis* sp. nov., *O. schmackeri* and *O. huanggangensis*.

OK, *O. kweichowensis* sp. nov.; OS, *O. schmackeri*; OH, *O. huanggangensis*. Significance level: * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$. Abbreviations for the morphometric characters refer to Methods section.

Males of <i>OK</i> vs. Females of <i>OK</i>		In males		In females	
		<i>OK</i> vs. <i>OS</i>	<i>OK</i> vs. <i>OH</i>	<i>OK</i> vs. <i>OS</i>	<i>OK</i> vs. <i>OH</i>
SVL	0.0001***	0.3160	0.2370	0.0810	0.4520
HDL	0.5560	0.0030**	0.5740	0.0900	0.0540
HDW	0.6240	0.2100	0.7650	0.0560	0.9000
SL	0.9840	0.0130*	0.1130	0.0740	0.9860
ED	0.0001***	0.4730	0.2510	0.1560	0.9130
IOD	0.1470	0.0910	0.2370	0.4520	0.2880
IND	0.0210*	0.5770	0.0330*	0.0290*	0.8340
NED	0.1470	0.2790	0.8480	0.4940	0.8860
NSD	0.8830	0.0980	0.0430*	0.3060	0.0310*
IFE	0.0001***	0.2140	0.3870	0.0740	0.6930
IAE	0.0001***	0.0490*	0.0310*	0.2320	0.2060
TYD	0.0001***	0.1960	0.2980	0.9260	0.1040
LAL	0.3340	0.0950	0.0310*	0.0320*	0.0000***
LW	0.0040**	0.1090	0.1390	0.3510	0.6420
THL	0.7560	0.0110*	0.0240*	0.0270*	0.0000*
TL	0.0920	0.0170*	0.1970	0.0420*	0.0040**
TW	0.0220*	0.0840	0.1430	0.1500	0.0160*
TFL	0.9930	0.0540	0.0290*	0.0500	0.0390
FL	0.2690	0.0170*	0.2410	0.0270	0.0050**
FDW	0.3400	0.0840	0.5660	0.9480	0.5520
DPW	0.5490	0.1040	0.6290	0.4520	0.6510

Figure 1

Sampling localities in this study.

Localities 1-8 were all in China. 1, Jinsha County (Co.), Guizhou Province (Prov.); 2, Zheng'an Co., Guizhou Prov.; 3, Meitan Co., Guizhou Prov.; 4, Leishan Co., Guizhou Prov.; 5, Jiangkou Co., Guizhou Prov.; 6, Sangzhi Co., Hunan Prov.; 7, Changyang Co., Hubei Prov.; 8, Ruyuan Co., Guangdong Prov.. According to literatures and results in this study, the distributional range of each related species was surrounded by dotted lines on the map. *Odorrana* sp1 and *Odorrana* sp2 were proposed in Zhu (2016); *Odorrana* sp3 was corresponding to the third cryptic species (Clade F) proposed in Li et al. (2015). Different species were denoted as different colors and shapes.

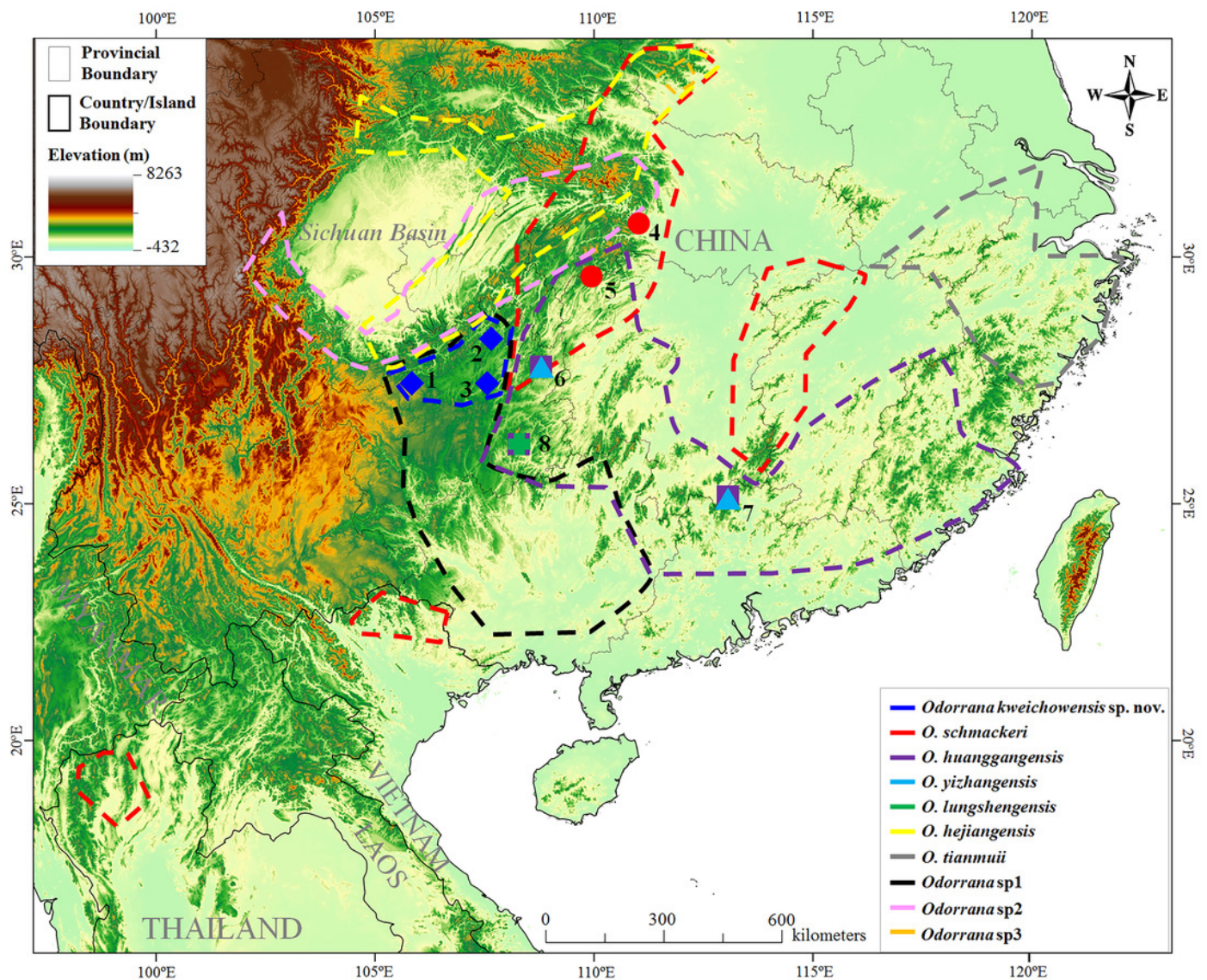


Figure 2

Phylogenetic relationships of *Odorrana kweichowensis* sp. nov. and its congeners.

(A) Maximum likelihood (ML) tree reconstructed based on the three genes (12S + 16S + ND2) concatenated dataset. (B) Phylogenetic network of *O. kweichowensis* sp. nov. and its related species reconstructed by the software Splittree based on ND2 gene sequences. In ML tree, bootstrap supports (BS) from ML analyses/Bayesian posterior probabilities (BPP) from BI analyses were noted beside nodes. In Splittree network, numbers on branches are BS. *Odorrana* sp2 was proposed in Zhu (2016); *Odorrana* sp3 was corresponding to the third cryptic species (Clade F) in Li et al. (2015). Different related species of *Odorrana kweichowensis* sp. nov. were denoted as different colors.

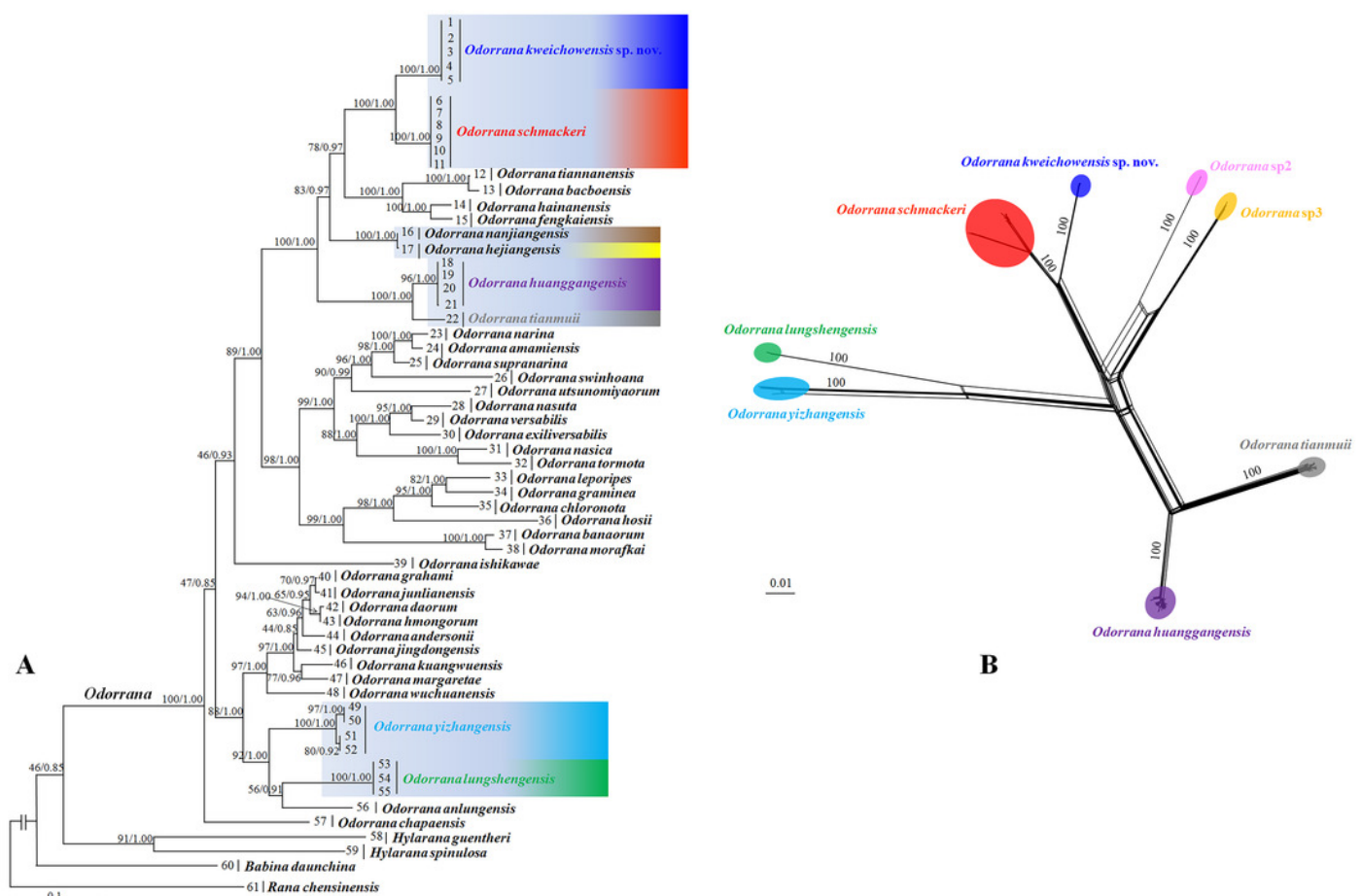


Figure 3

Haplotype networks of *O. kweichowensis* sp. nov. and its related species constructed based on the nuclear gene sequences.

(A) DOLK gene. (B) KCNF gene. Different species were denoted as different colors.

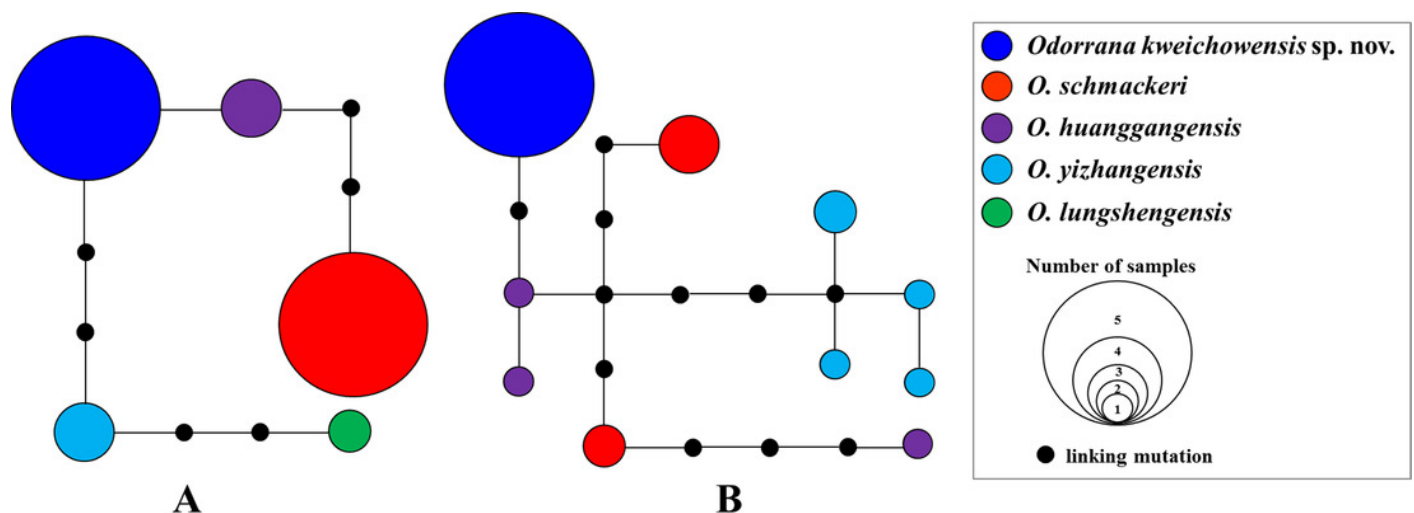


Figure 4

Plots of principal component analyses of *Odorrana kweichowensis* sp. nov., *O. schmackeri* and *O. huanggangnesis*.

(A) Males. (B) Females. PC1, the first principal component; PC2, the second principal component. Different species were denoted as different colors and shapes.

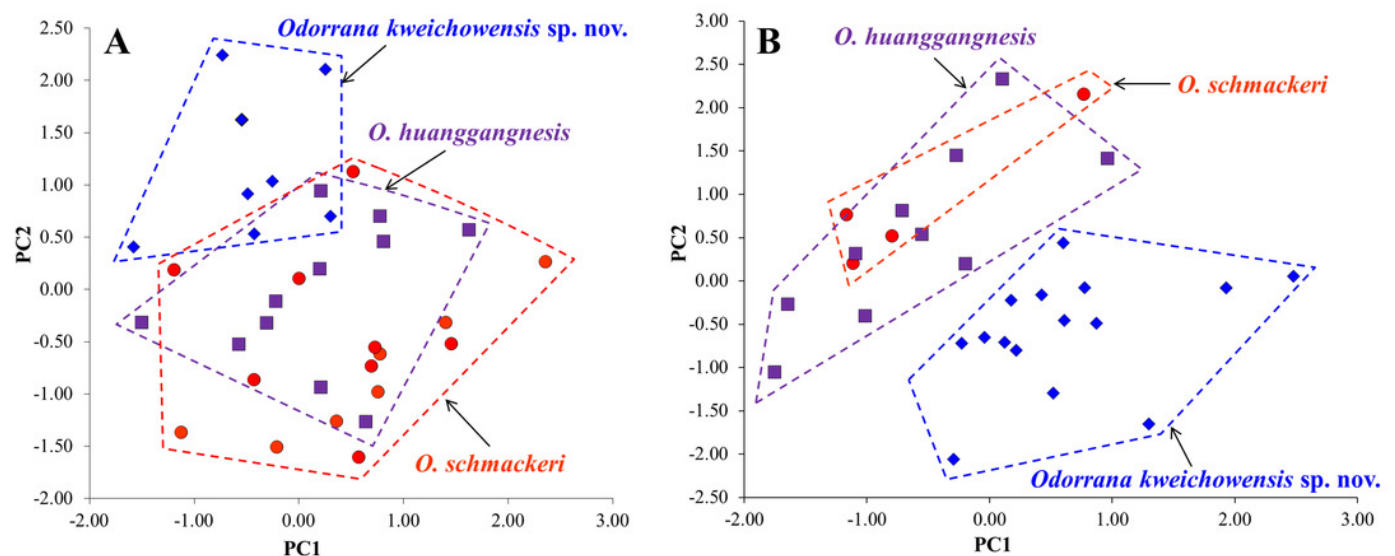


Figure 5

Comparisons of the holotype (voucher number: CIBjs20150803002) of *Odorrana kweichowensis* sp. nov. and one male specimen (voucher number: CIBsz2012062003) of *O. schmackeri*.

(A) and (B), dorsal view and ventral view of specimen CIBjs20150803002, respectively. (C) and (D), dorsal view and ventral view of specimen CIBsz2012062003, respectively. (E) Ventral view of hand of specimens CIBjs20150803002. (F) Ventral view of hand of specimen CIBsz2012062003. (G) Ventral view of foot of specimen CIBjs20150803002. (H) Ventral view of foot of specimen CIBsz2012062003. Photographs by S.Z. Li.

**Note: Auto Gamma Correction was used for the image. This only affects the reviewing manuscript. See original source image if needed for review.*

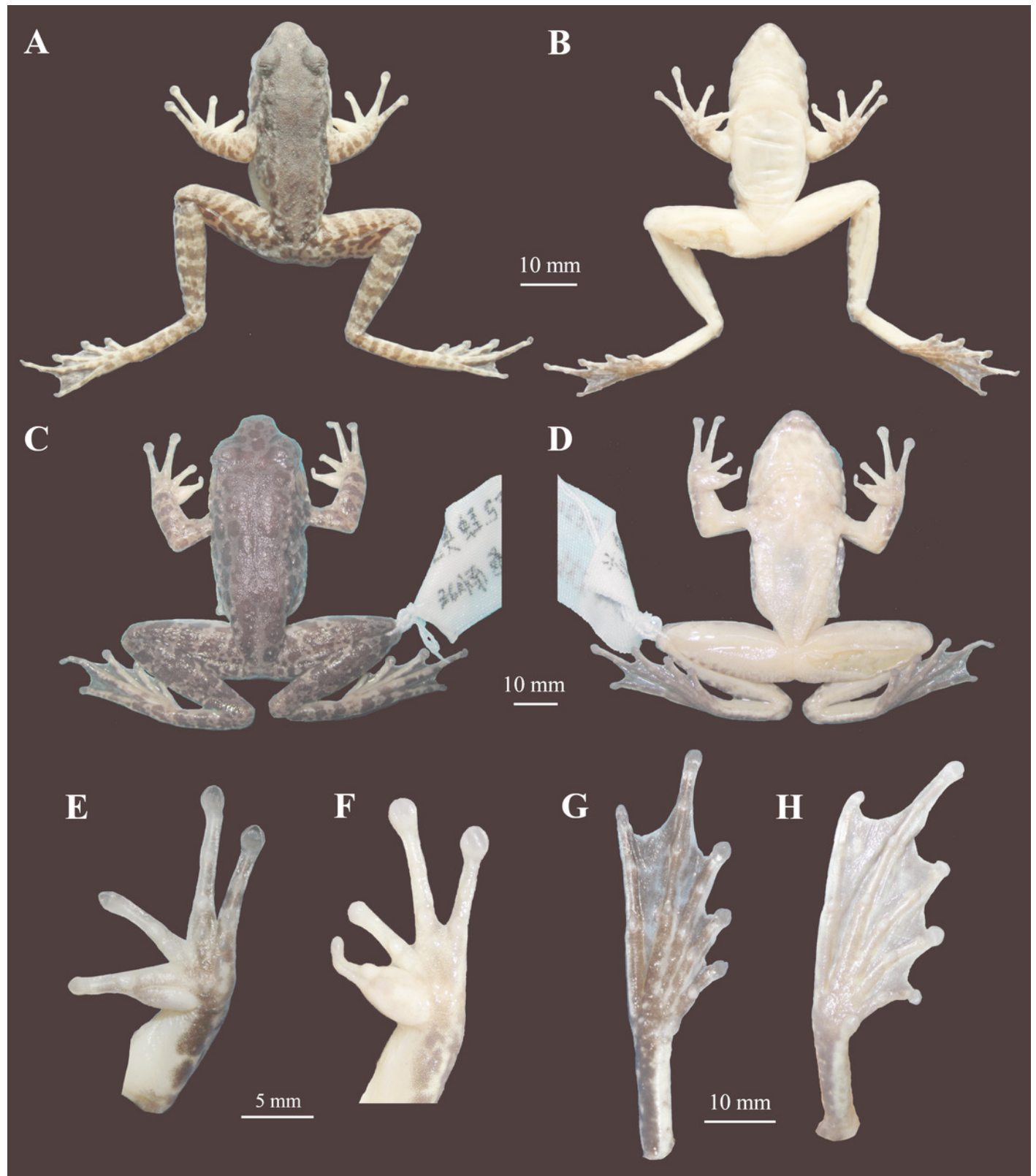


Figure 6

Living *Odorrana kweichowensis* sp. nov. from its type locality, Lengshihe Nature Reserve in Jinsha County, Guizhou Province, China.

(A) and (B), dorsolateral view and ventral view of an adult male (voucher number: CIBjs20150803002), respectively. (C) and (D), dorsolateral view and ventral view of an adult female (voucher number: CIBjs20150803006), respectively. Photographs by S.Z. Li.

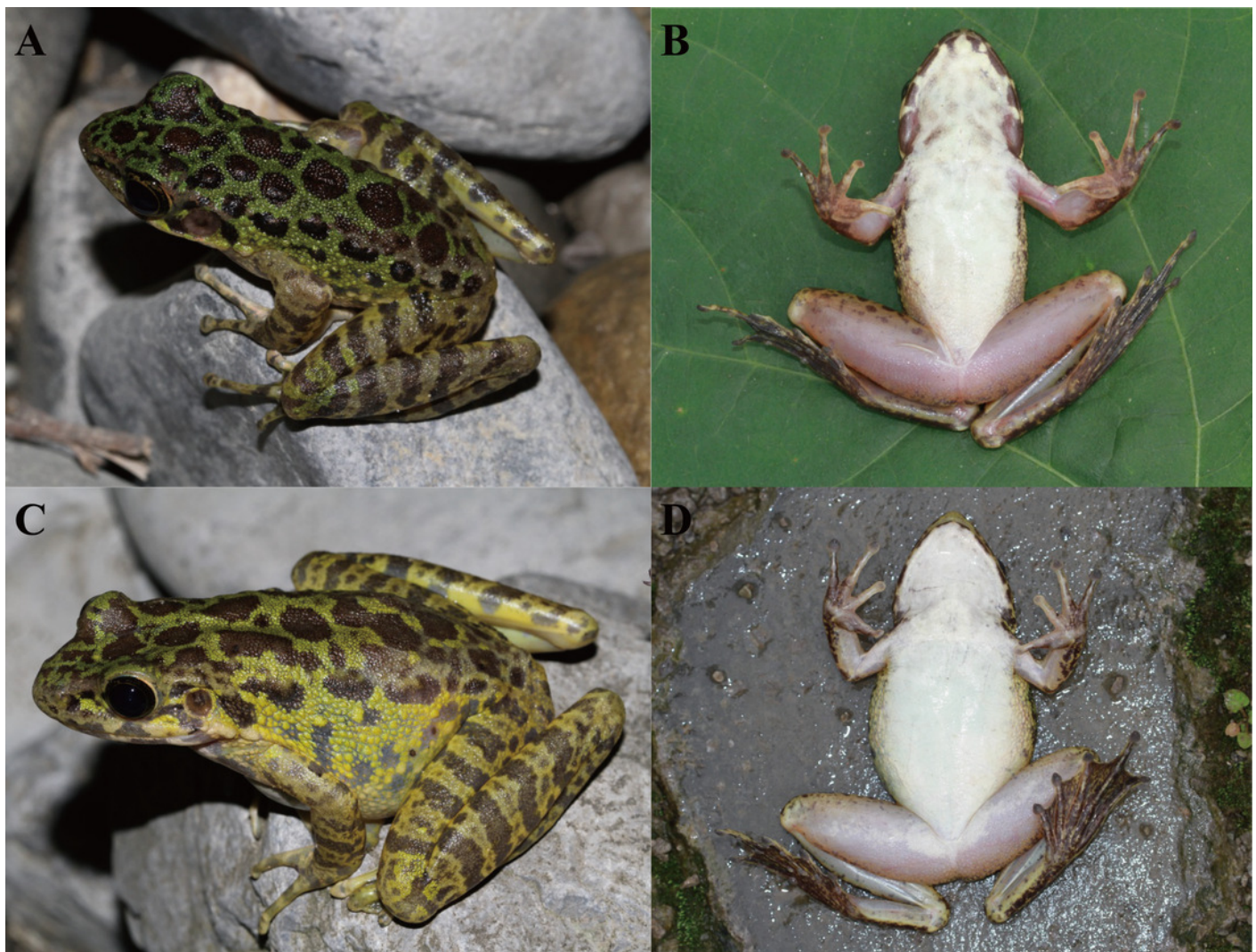


Figure 7

Color variations in *Odorrana kweichowensis* sp. nov..

(A) and (B), dorsolateral view and ventral view of an adult female from Jinsha County, Guizhou Province, China, respectively. (C) Dorsolateral view of a female from Meitan County, Guizhou Province, China. (D) Dorsolateral view of a female from Zheng'an County, Guizhou Province, China. Photographs by S.Z. Li.

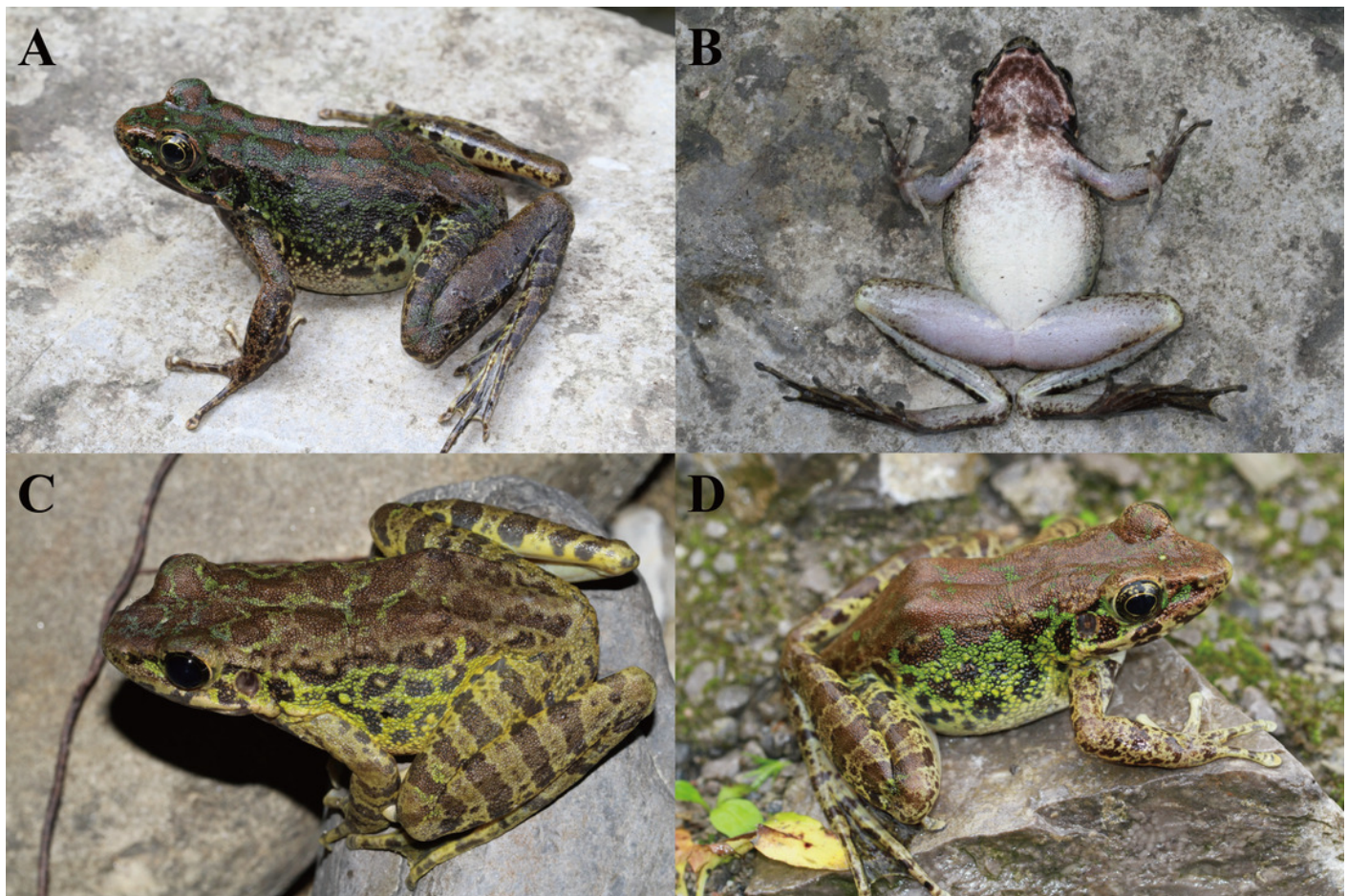


Figure 8

Skull of *Odorrana kweichowensis* sp. nov..

(A) Dorsal view. (B) Ventral view. 1, maxillary; 2, nasal; 3, frontoparietal; 4, pterygoid; 5, squamosal; 6, exoccipital; 7, prootic; 8, maxillary teeth; 9, prevomer; 10, palatine; 11, mandible; 12, sphenethmoid; 13, parasphenoid; 14, columella. Drawings by S.Z. Li.

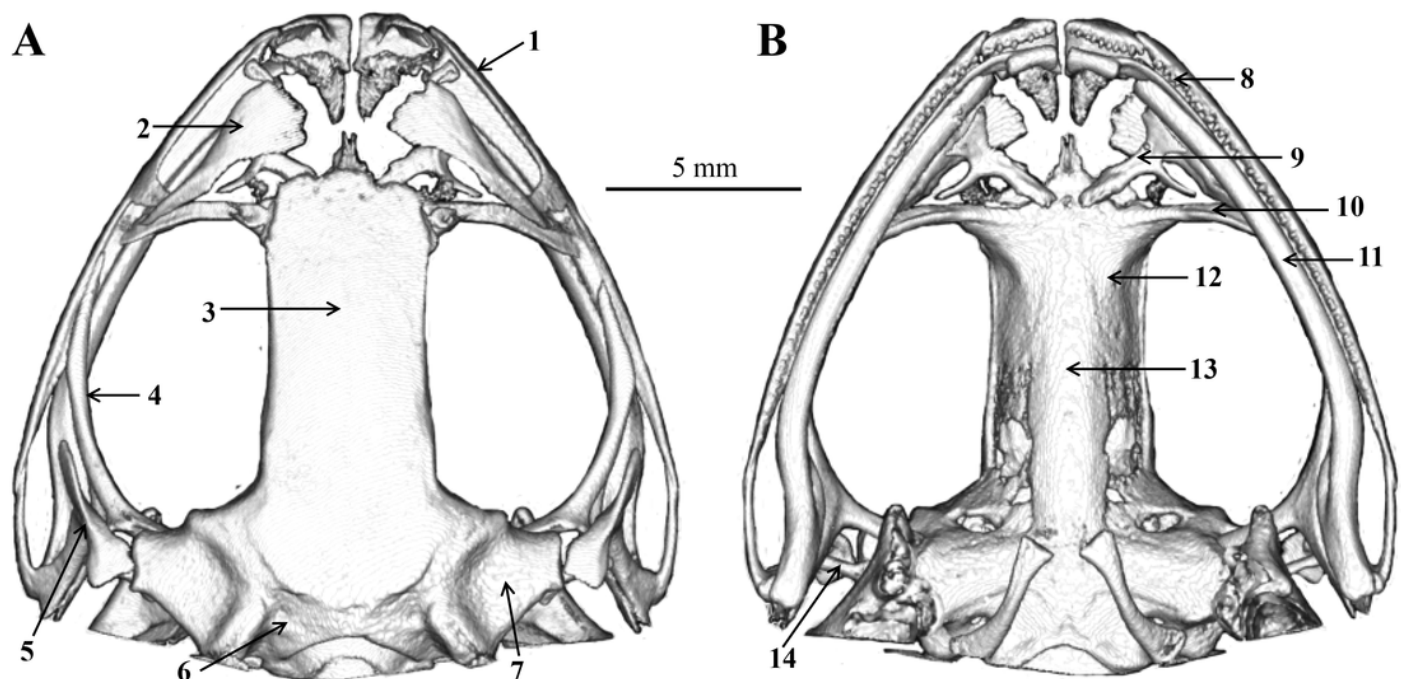


Figure 9

Tadpole of *Odorrana kweichowensis* sp. nov.

(A) Dorsolateral view of specimen CIBJS20171014001 in life. (B) Dorsal view of specimen CIBJS20171014001 in preservative. (C) Structure of the mouth of specimen CIBJS20171014001. 1, spiracle; 2, upper keratodonts; 3, lower keratodonts; 4, labial papillae on upper lips; 5, labial papillae on lower lips; 6, additional tubercles at the angles of mouth. Photographs by S.Z. Li.

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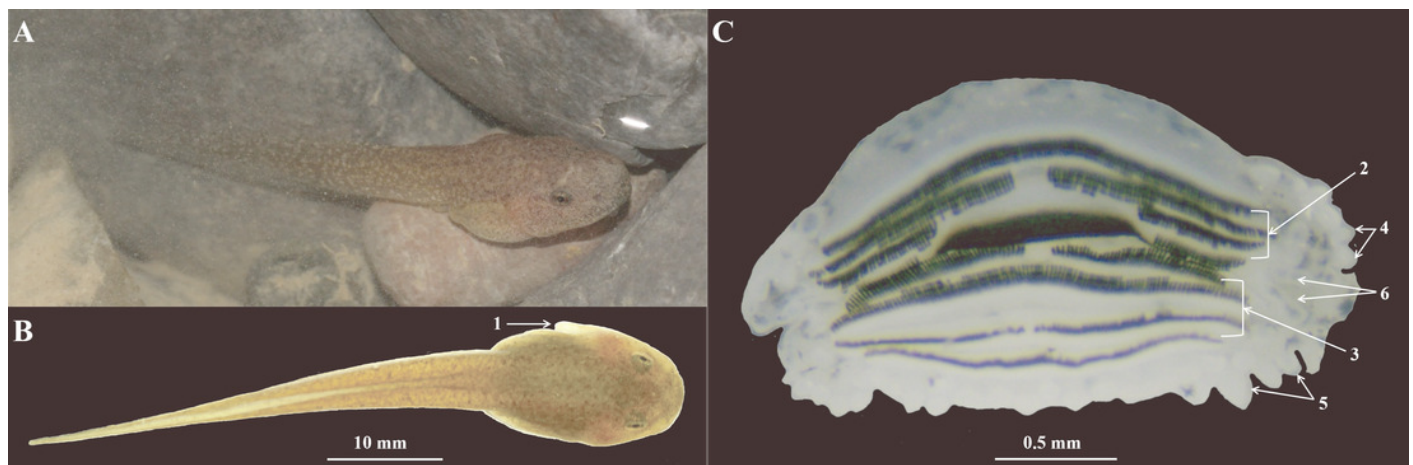


Figure 10

Habitats of *Odorrana kweichowensis* sp. nov..

(A) Habitats in the type locality, Lengshuihe Nature Reserve, Jinsha County, Guizhou Province, China; *insert* is the photo for one pair of amplexed male (smaller) and female (larger) found on the stone in the stream. (B) Habitats in Xieba Town, Zheng'an County, Guizhou Province, China. (C) Habitats in Shilian Town, Meitan County, Guizhou Province, China. Photographs by S.Z. Li.

