

Lessepsian migration and parasitism: richness, prevalence and intensity of parasites in the invasive fish *Sphyraena chrysotaenia* compared to its native congener *Sphyraena sphyraena* off Tunisia

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Background Parasites can play various roles in the invasion of non-native species, but these are still understudied in marine ecosystems. This also applies to invasions from the Red Sea to the Mediterranean Sea via the Suez Canal, the so-called Lessepsian migration. In this study, we investigated the role of parasites in the invasion of the Lessepsian migrant *Sphyraena chrysotaenia* in the Tunisian Mediterranean Sea. **Methods** We compared metazoan parasite richness, prevalence and intensity of *S. chrysotaenia* (Perciformes: Sphyraenidae) with infections in its native congener *S. sphyraena* by sampling these fish species at seven locations along the Tunisian coast. Additionally, we reviewed the literature to identify native and invasive parasite species recorded in these two hosts. **Results** Our results suggest the loss of at least two parasite species of the invasive fish species. At the same time, the Lessepsian migrant has co-introduced three parasite species, during the initial migration to the Mediterranean Sea, that are assumed to originate from the Red Sea of which only one parasite species has been reported during the spread to Tunisian waters. In addition, we found that the invasive fish has acquired six parasite species that are native in the Mediterranean Sea. However, parasite richness, prevalence and intensity were overall much lower in the invasive compared to the native fish host in the Mediterranean Sea. **Discussion** These results suggest that the Lessepsian migrant may affect native fish hosts by potentially altering the dynamics of native and invasive parasite-host interactions via parasite release, parasite co-introduction and parasite acquisition. They further suggest that the lower infection levels in the invasive fish may result in a competitive advantage over native fish hosts (enemy release hypothesis). This study demonstrates that cross-species comparisons of parasite infection levels are a valuable tool to identify the different roles of parasites in the course of Lessepsian migrations.

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23

24 **Abstract**

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 26 still understudied in marine ecosystems. This also applies to invasions from the Red Sea to the
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 28 investigated the role of parasites in the invasion of the Lessepsian migrant *Sphyraena*
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 32 these fish species at seven locations along the Tunisian coast. Additionally, we reviewed the
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34 **Results** Our results suggest the loss of at least two parasite species of the invasive fish species.
 35 At the same time, the Lessepsian migrant has co-introduced three parasite species, during the
 36 initial migration to the Mediterranean Sea, that are assumed to originate from the Red Sea of
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 38 addition, we found that the invasive fish has acquired six parasite species that are native in the
 39 Mediterranean Sea. However, parasite richness, prevalence and intensity were overall much
 40 lower in the invasive compared to the native fish host in the Mediterranean Sea.

41 **Discussion** These results suggest that the Lessepsian migrant may affect native fish hosts by
 42 potentially altering the dynamics of native and invasive parasite-host interactions via parasite
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Introduction

One of the potential explanations for the establishment and subsequent spread of invasive species in marine and other ecosystems is the enemy release hypothesis (Elton, 1958; Keane & Crawley, 2002). This hypothesis states that invasive species may gain a competitive advantage over native species by losing all or part of their natural enemies, such as predators and parasites, during the invasion process. For parasites, such a release or reduction has been documented for a wide range of host taxa including marine species (Torchin, Lafferty & Kuris, 2001; Keane & Crawley, 2002; Torchin et al., 2003; Torchin & Mitchell, 2004; Blakeslee, Fowler & Keogh, 2013). However, invasive hosts do not necessarily lose all their native parasites during an invasion, but can often co-introduce parasites to their non-indigenous range (Lymbery et al., 2014). The likelihood of this co-introduction depends, among others, on the host specificity and life cycle of the respective parasite species. Generalist parasites which infect a larger range of host species and parasites with direct life cycles are more likely to be co-introduced than highly specific parasites or parasites with complex life cycles (i.e., depending on several sequential different host species; Torchin, Lafferty & Kuris, 2002; Poulin & Morand, 2004; Lymbery et al., 2014). In the new range, co-introduced parasites may also infect native hosts (parasite spillover; Prenter et al., 2004, Kelly et al., 2009), with potentially serious impacts on native species and ecosystems (emerging diseases; Daszak, Cunningham & Hyatt, 2000). Finally, invasive species can also acquire native parasites from native host species. This parasite acquisition may have deleterious effects on naïve invasive host species (‘increased susceptibility hypothesis’ *sensu* Colautti et al., 2004), but may also ultimately amplify native parasite population sizes, resulting

in increased parasite loads in native hosts, a phenomenon which is referred to as parasite spillback (Kelly et al., 2009). While many of these mechanisms may result in a competitive advantage for invasive over native species, the magnitude of this advantage will depend on the actual difference in parasite infection levels between invasive and native hosts. Several studies comparing infection levels in invasive and introduced hosts (community studies or cross-species comparisons; *sensu* Colautti et al., 2004; Torchin & Mitchell, 2004) have shown that infection levels are often lower in invasive host species (Georgiev et al., 2007; Dang et al., 2009; Roche et al., 2010; Gendron, Marcogliese & Thomas, 2012). However, such cross-species comparisons of infection levels in invasive and native competitors are surprisingly scarce (Goedknecht et al., 2017). In addition, our knowledge on the role of parasites in biological invasions in general is still limited, especially in the marine realm (Vignon & Sasal, 2010; Goedknecht et al., 2016).

This also applies to species invasions in the Mediterranean Sea, an ecosystem with an extraordinarily high rate of species introductions, with more than 1000 alien species listed (Bilecenoglu et al., 2013). Especially via the Suez Canal, which was opened in 1869, many benthic invertebrates and fish species have migrated from the Indian Ocean over the Red Sea to the Mediterranean Sea, a massive human-initiated invasion referred to as Lessepsian migration (*sensu* Por, 1978). Despite this high migration rate and the resulting high number of species introductions, parasitological investigations of invasive species have been surprisingly rare in this region (Pérez-del-Olmo, Kostadinova & Gibson, 2016) and have only focused on a few host species such as a portunid crab (Galil & Innocenti, 1999), the Lessepsian fishes *Siganus* spp. (Diamant, 2010), *Fistularia commersonii* (Merella et al., 2016), *Etrumeus golanii* (Boussellaa et al., 2016) and *Lagocephalus sceleratus* (Bakopoulos, Karoubali & Diakou, 2017). Initially, 18 species of parasites spread over four taxonomic groups (Monogenea, Crustacea, Protozoa and

Digenea) had been recognized as Lessepsian migrants which have been co-introduced with their hosts to the Mediterranean Sea (Zenetos et al., 2008), and more recently this list has been updated by the addition of many other parasite species (Diamant, 2010, Merella et al., 2016). Given that there are many Lessepsian migrants that have never been under parasitological investigation, the actual list of co-introduced parasite species is likely to be much longer. In addition, invasive hosts may have acquired native parasite species but whether the resulting parasite loads of hosts are actually lower, equal to or higher than the ones in native species in the Mediterranean Sea has scarcely been studied to date.

In the present study, we investigated metazoan parasite infections in the Lessepsian migrant fish *Sphyraena chrysotaenia* Klunzinger, 1884, along the Tunisian coast in the central Mediterranean Sea. This fish species was first recorded in the eastern Mediterranean Sea in 1931 (Spicer, 1931) and has since then spread westwards, after which it was reported for the first time at the Tunisian coast in 2002 (Bradai et al., 2002). It is now a relatively common piscivorous fish of about 20-25 cm length, living in the pelagic and demersal zones to a depth of 50 m in inshore waters where it is captured by local artisanal fisheries (Golani & Ben Tuvia, 1995; Wadie & Riskhallah, 2001; Zouari-Ktari, Bradai & Bouain, 2009). A cross-species comparison of the parasite communities was carried out considering the parasite richness and levels of infection in the invasive fish and in the congeneric native one *Sphyraena sphyraena* Linnaeus, 1758. This native fish species is usually larger than the invasive species (30-60 cm length) but has an overlapping prey spectrum (Kalogirou et al., 2012), lives in the same habitats and is also used by local fisheries (Relini & Orsi Relini, 1997; Allam, Faltas & Ragheb, 2005). By sampling invasive (*S. chrysotaenia*) and native (*S. sphyraena*) fish hosts along the Tunisian coast and by conducting an additional parasitological literature survey, we aimed to answer the following

specific research questions: 1) Is there evidence that *S. chrysotaenia* experienced a release from its native parasites from the Red Sea?, 2) did *S. chrysotaenia* co-introduce parasites from the Red Sea and/or did it acquire native parasites from the Mediterranean Sea? and 3) how do parasite richness, prevalence and intensity in the invasive *S. chrysotaenia* compare with those in the native *S. sphyraena*?

Material and methods

Fish sampling

Between October 2012 and July 2015, a total of 107 specimens of *Sphyraena sphyraena* (native fish) and 148 specimens of *Sphyraena chrysotaenia* (invasive fish) were collected at seven fishing localities along the Tunisian coast (off the cities of Sfax, Kerkennah, Skhira, Chebba, Zarat, Zarzis and Sayeda; Fig. 1, Table 1). Fish were mostly bought from local fishermen operating landing trawlers along the Tunisian coast at a depth of about 30 m. In addition, some specimens were recovered from artisanal inshore fishery of Kerkennah and Chebba. Only adult individuals were examined, with total lengths of 25.7-42.7 cm for *S. sphyraena* and 18.5-26.2 cm for *S. chrysotaenia*. Samples were kept fresh or were deep-frozen in individual plastic bags at -10°C, until examination in the laboratory. After defrosting, all fish individuals were identified to species level using Whitehead et al. (1984) and Fisher et al. (1987) and examined for parasites as described below. During our study, Faculty of Sciences of Sfax provided full approval for this purely observational research.

Parasite sampling

We only checked for metazoans parasites, as this was the focus of our study. Fish skin, fins, nasal pits, eyes and buccal cavities were thoroughly examined for the presence of ectoparasites

under a stereomicroscope with incident light. Gill arches were separated by incision, and then placed in petri dishes filled with sea water and examined for the presence of ectoparasites. Internal organs (stomach, pyloric caeca, intestines, heart, liver, spleen, gall bladder and gonads) were separated and individually examined for the presence of endoparasites. Platyhelminthes were fixed between a slide and coverslip with 70% ethanol. Fixed specimens were stained with Semichon's acetic carmine, dehydrated using a graded ethanol series then cleared in clove oil and mounted in Canada balsam. Other parasites (copepods, isopods, nematodes and annelids) were directly fixed in 70% ethanol for later examination. Parasites were identified to the lowest taxonomic level possible using Gibson et al. (2002) for Digenea, Neifar (1995) and Theisen et al. (2017) for Monogenea, Euzet (1994) for Cestoda, Berland (1961) and Petter & Maillard (1988) for Nematoda and the keys of Kabata (2003) for Copepoda.

Literature review

We searched literature databases (Host-Parasite database from the Natural History Museum in London, Web of Science, Google Scholar) for published records of additional parasite species from the Mediterranean Sea and the Red Sea. Search strings included the species names and the different parasite taxa. In addition, we searched the reference sections of publications and our own reference collections for potential further studies reporting parasite infections in the two fish.

Statistical analyses

As sampling effort differed between native ($n = 107$) and invasive fish hosts ($n = 148$), we produced rarefaction curves to identify the level of dependence of species accumulation on

sampling effort. Sample-based rarefaction curves were computed based on sample sizes at the different locations (Gotelli & Colwell 2001) using Estimates S 9.1.0 (Colwell, 2013). Based on recommendations by Walther & Morand (1998) in regard to parasites, we used the nonparametric species estimator Chao2 for our rarefaction analyses. This species estimator algorithm uses the frequency of unique species in samples to estimate the number of missing species in a population (Chao, 2005).

We used general linear models (GLMs) to test for statistical differences between the fish species and among locations in parasite richness of individual fish (Poisson distribution; $n = 255$), infection status of individual fish (infected or uninfected; binomial distribution; $n = 255$) and intensity of infected fish (negative binomial distribution; $n = 127$). In each model, we added host species and location as fixed factors and an interaction term. As the parasite fauna of the two host species was very different, we used the total infection status and intensity of parasite species per host species to compare infection levels between hosts. Although this procedure may obscure the potentially different effects on hosts exerted by different parasite species, this lumping procedure still allows for an approximate comparison of overall infection levels. We did not add fish size as a covariate in the models because preliminary analyses using GLMs did not show an effect of fish size on any of the response variables. All statistical models were run using the statistical software environment R v3.3.0. (R Development Core Team 2016).

Results

After the dissections of 107 individuals of the native *Sphyraena sphyraena* and 148 individuals of the invasive *S. chrysotaenia* caught along the coast of Tunisia, ten different parasite species infecting the two fish were found. Our additional literature survey added another ten parasite

species records from the Mediterranean or Red Sea to the total parasite species list of both fish species (Table 2).

Parasite release

In total, five parasite species were collected from the Lessepsian migrant *Sphyrna chrysotaenia* in Tunisian coastal waters (Table 2). In addition, the literature survey indicated that four other parasite species have been found in this invasive host species elsewhere in the Mediterranean Sea (Table 2). Hence, *S. chrysotaenia* is infected by at least nine parasite species in its invaded range. In its native range, the Red Sea, two parasite species of *S. chrysotaenia* have been reported in the literature (Table 2). In addition, three of the parasite species found in *S. chrysotaenia* in the Mediterranean Sea most likely originate from Red Sea, although published records are not available. This suggests that the Lessepsian migrant harbours at least five parasite species in its native range, the Red Sea (Table 2).

Parasite co-introduction and acquisition

The examination and the literature survey revealed that the Lessepsian migrant *S. chrysotaenia* is infected by three parasite species in the Mediterranean Sea that are assumed to originate from the Red Sea (Table 2). Our results further revealed that the Lessepsian migrant *S. chrysotaenia* has acquired six native parasite species in the Mediterranean Sea (Table 2). Five of those species were found in our survey in Tunisian coastal waters and a fifth species had been noted in *S. chrysotaenia* elsewhere in the Mediterranean Sea (Table 2).

Comparison with the native congeneric species *Sphyrna sphyraena*

The native congeneric species *Sphyraena.sphyraena* was infected with six parasite species in Tunisian coastal waters and four additional species have been described in the literature (Table 2). Of all these parasite species, only one was shared with the introduced fish host *S. chrysotaenia*, the presumably native isopod *Gnathia* sp.. None of the parasites that were co-introduced by the Lessepsian migrant have been found to infect the native congeneric host species (Table 2).

In general, the rarefaction curves did not reach asymptotic levels in the two congeneric host species, indicating that a higher sampling effort may reveal more (albeit rare) parasite species in Tunisian coastal waters (Fig. 2). The species accumulation curves further indicate that the total parasite species richness at a given sampling effort could be higher in the native than in the invasive host species. This was also reflected in the mean parasite richness per individual fish found at the seven locations, which was generally significantly higher in the native *S. sphyraena* than in the invasive *S. chrysotaenia* (GLM: $\beta = 0.705$, $SE = 0.586$, $\Delta_{Deviance} = 110.655$, $df = 253$, $p < 0.001$; Fig. 3). However, at some locations this pattern was reversed, resulting in a significant interaction term ($\Delta_{Deviance} = 3.470$, $df = 241$, $p < 0.01$; Fig. 3).

Likewise, the infection status of fish hosts was generally significantly higher in the native *S. sphyraena* than in the invasive *S. chrysotaenia* (GLM: $\beta = 1.966$, $SE = 1.201$, $\Delta_{Deviance} = 162.599$, $df = 247$, $p < 0.001$; Fig. 4a). This was consistent over all locations as there was no significant difference in overall infection status between locations ($p = 0.294$; Fig.4a). However, due to the absence of infections at two locations, there was a significant interaction term ($\Delta_{Deviance} = 168.19$, $df = 241$, $p < 0.05$; Fig. 4a). Mean intensity of infections also differed between host species, with the native fish being infected with higher numbers of parasites than the invasive fish host (GLM: $\beta = 0.337$, $SE = 0.586$, $\Delta_{Deviance} = 35.420$, $df = 119$, $p < 0.001$; Fig.

4b). There was also a significant effect of location on overall infection intensities ($\Delta_{\text{Deviance}} = 26.295$, $df = 120$, $p < 0.001$), with location 1 showing much lower values than the other locations (Fig. 4b). However, there was no significant interaction between location and host species ($p = 0.249$).

Discussion

Based on samples from the Tunisian coast and additional literature data, our analyses indicate that the invasive Lessepsian migrant *S. chrysotaenia* lost two parasite species during its introduction to the Mediterranean Sea, but that the invasive fish also co-introduced parasites from the Red Sea and acquired one generalist parasite from native fish hosts in the Mediterranean Sea (Table 2). However, parasite richness and infection levels were overall much lower in the invasive compared to the native fish host, suggesting a potential competitive advantage for the Lessepsian migrant.

Parasite release

In its native range (the Red Sea), the invasive *S. chrysotaenia* harbours at least five parasite species (Table 2). Of these five parasite species, only three species have been recorded in the Mediterranean Sea and only one in Tunisian coastal waters (Table 2). Hence, two parasite species, the digenean *Bucephalus sphyraenae* and the monogenean *Pseudolamellodiscus sphyraenae* have been lost in the process of the initial invasion to the Mediterranean Sea. Two other species, the copepod *Nothobomolochus denticulatus* and the isopod *Cymothoa indica*, have been observed elsewhere in the Mediterranean Sea, but not in Tunisian coastal waters (Table 2), suggesting a loss of these parasite species in the course of the spread to the Tunisian coast. This loss of several natural parasite species is consistent with the enemy release hypothesis (Torchin

et al., 2003; Blakeslee, Fowler & Keogh, 2013) and has previously been observed in other Lessepsian migrant fish, the rabbitfish *Siganus rivulatus* (see Diamant, 1989) and partially reported from the bluespotted cornet fish *Fistularia commersonii* (see Merella et al., 2016).

Several processes may be responsible for a loss of parasites in the course of passing the Suez Canal “filter” or “bottleneck” (Por, 1978). First of all, parasites may not be able to cope with the environmental conditions in the canal or in the Mediterranean Sea and do not survive to the migration the new ecosystem. This negative impact of environmental conditions should be particularly relevant for ectoparasites, which are more exposed to the external environment than endoparasites, and may explain the loss of the monogenean *Pseudolamellodiscus sphyraenae*. Second, parasites may survive the passage through the canal, but, in the case of parasites with complex life cycles, they may be unable to find suitable intermediate hosts in the recipient ecosystems. In general, parasite co-introductions seem to be more common in parasites with simple life cycles as the necessity for all hosts being present for parasites with complex life cycles makes invasions less likely (Lymbery et al., 2014; Goedknecht et al., 2016). Such a lack of suitable hosts may explain the loss of the trematode *Bucephalus sphyraenae* as trematodes are generally very host specific in respect to their first intermediate gastropod host (Poulin & Cribb, 2002; Galaktionov & Dobrovolskij, 2003). The respective gastropod host species that serves as first intermediate host for *B. sphyraenae* is not known, but a (co-)introduction of snails is unlikely because of their relatively reduced mobility. Finally, even in the case that all potential hosts are present in the new environment, the number of introduced parasite individuals may be too low to maintain a viable population. In general, propagule pressure is known to be a strong determinant of invasion success in biological invasions in general (Wonham et al., 2000; Forsyth

& Duncan, 2001; Rouget & Richardson, 2003; Colautti et al., 2004). In the case of parasites, propagule pressure may be low if infection levels are low in the native region, reducing both the chances for a co-introduction and for developing viable populations after introduction to the Mediterranean Sea. This could explain the loss of infections with the monogenean *Pseudolamellodiscus sphyraenae* as this species may show only very low infection levels in its native range. These different mechanisms are not only acting during the passage of Lessepsian migrants through the Suez Canal, but also during the spread in the Mediterranean Sea after the initial introduction. This is illustrated by the copepod *Nothobomolochus denticulatus* and the isopod *Cymothoa indica* which have both been co-introduced into the Mediterranean Sea and have been found on *S. chrysotaenia* in the eastern Mediterranean Sea (Trilles & Bariche, 2006; El-Rashidy & Boxshall, 2012), but not in Tunisian waters (our study).

Parasite co-introduction

The present results and literature review revealed that the Lessepsian migrant *S. chrysotaenia* is infected by three parasite species in the Mediterranean Sea that most likely originate from the Red Sea (Table 2). One of these species, the monogenean parasite *Pseudempleurosoma* sp., was recorded in our study in Tunisian waters. There are several reasons why we believe that this parasite species has been co-introduced to the Mediterranean Sea (although this still needs to be confirmed by records from its presumed native range). First of all, the species has not been recorded in any native fish species in the Mediterranean Sea in earlier studies, although fish parasites are relatively well studied in this coastal ecosystem (Oliver, 1987; Neifar & Euzet, 2007; Pérez-del-Olmo, Kostadinova & Gibson, 2016; Chaabane et al. 2016a; Chaabane et al. 2016b). Second, the genus *Pseudempleurosoma*, represented by four

species in the literature, seems to have a strict specificity to its host (Santos, Mourão & Cárdenas, 2001), suggesting that it can only be co-introduced with its host. Finally, with their direct life cycles, monogeneans are likely to be introduced and persist in a new environment. Especially in gregarious fish such as the Sphyraenidae, the transmission of parasites with a direct life cycle can easily take place so that the lifecycle can be maintained in the new environment after an initial introduction.

The two other parasite species, the copepod *Nothobomolochus denticulatus* and the isopod *Cymothoa indica* are less host specific and have also been found on other fish species from the Red Sea (El-Shahawy & Desouky, 2010). These two species have not been recorded in our study in Tunisian waters but have been reported elsewhere in the Mediterranean Sea (Trilles & Bariche, 2006; El-Rashidy & Boxshall, 2012). Both species have not been recorded in any native fish before their invasion but are considered to be co-introduced by the describing authors (Trilles & Bariche, 2006; El-Rashidy & Boxshall, 2012). However, as there are no published records of the two species available from the native range of the host, their invasive status still has to be confirmed. Both species have not (yet) been co-introduced into Tunisian coastal waters in the course of the spread of their hosts after the initial introduction in the Mediterranean Sea, probably due some of the mechanisms discussed above.

Parasite acquisition

Besides being infected with co-introduced parasites, our study also revealed that the Lessepsian migrant *S. chrysotaenia* has acquired six native parasite species in the Mediterranean Sea. Four of those species were found in our survey in Tunisian coastal waters and a fifth species

322 had been noted in *S. chrysotaenia* elsewhere in the Mediterranean Sea (Table 2). We consider
 323 these acquired parasite species to be native species from the Mediterranean Sea due to the
 324 following reasons. The digenean *Lecithochirium* sp. has not been reported from the invasive *S.*
 325 *chrysotaenia*, neither in the Mediterranean Sea nor in the Red Sea (Fischthal, 1982; Nahhas, Sey
 326 & Nakahara, 2006). Given the complex life cycle of the Hemiuridae that need at least one
 327 intermediate host species, and the fact that this genus now contains at least more than 100
 328 species (Surekha & Lakshmi, 2005) which at least 10 are reported from Mediterranean Sea with
 329 scarcely life cycles (Table 3) and that the genus *Lecithochirium* is very specific to its first
 330 intermediate gastropod host (Gibson & Bray 1994), *Lecithochirium* sp. is most likely of
 331 Mediterranean origin and has been acquired after the introduction of *S. chrysotaenia* to the
 332 Mediterranean Sea. Specimens of the copepod *Caligus* sp. reported from *S. chrysotaenia* were in
 333 poor condition and their identification to the species level was impossible. According to the
 334 literature, no caligids were collected from *S. chrysotaenia* in the Red Sea but these parasitic
 335 copepods have been recorded in native Mediterranean fish (Benmansour & Ben Hassine, 1998;
 336 Raibaut, Combes & Benoit, 1998). Although many species are highly host specific there are also
 337 many *Caligus* species with low host specificity (Yuniar, Palm & Walteret, 2007), making a host
 338 switch to the invasive host likely. However, further studies will be needed to ascertain the native
 339 status of *Caligus* individuals found on the invasive host. Annelids of the family Piscicolidae are
 340 considered to be generalists with a broad host specificity and are regularly reported from many
 341 native fish hosts from Tunisian coasts and the Mediterranean Sea in general (Châari & Neifar,
 342 2015). According to the literature, no gnathiids were reported previously from *S. chrysoteania* in
 343 the Red Sea. In contrast, this isopod genus is very common in the Mediterranean Sea and along
 344 the Tunisian coasts, particularly on the skin of Sphyaenidae host species so that it is most likely

native to the area. However, the native status of gnathiids found on the invader. The native Cymothoid *Anilocra physodes* has been reported from many native hosts of the Mediterranean Sea (Innal et al., 2007) and was only observed from the invasive fish *S. chrysotaenia* off Turkish (Innal et al., 2007). Finally, the trematode *Bucephalus labracis*, which was not found in the Lessepsian migrant elsewhere in the Mediterranean Sea (Fischhal, 1982), is considered to be native as it uses a native bivalve (*Ruditapes decussatus*) as first intermediate host, a native fish (*Atherina boyeri*) as second intermediate host and a native fish (*Dicentrarchus labrax*) as definitive host (Paggi & Orecchia (1965); Maillard (1976); Gargouri-Ben Abdallah & Maamouri (2005); Dhrif et al., 2015). The acquisition of these native parasites may generally have potentially adverse effects naïve invasive host species, but also on native species, as the invasive additional hosts can potentially elevate infection levels in native species via parasite spill back. However, whether significant parasite spillback occurs in Tunisian waters remains to be studied.

Cross-species comparison

Our comparison of parasite infections in the Lessepsian migrant with the native congeneric *Sphyraena sphyraena* revealed considerable qualitative and quantitative differences between the parasite communities of the two fish species. The native fish was infected with six parasite species in Tunisian coastal waters and three additional species have been described elsewhere in the Mediterranean Sea (Table 2). Of all these parasite species, only one was shared with the invasive fish host, the presumably native isopod *Gnathia* sp.. As only larval stages of gnathiid parasite can infect fish and all larvae are morphologically very similar, identification to species level was not possible. Native *Gnathia* species have been reported from the study area (Châari & Neifar, 2015) and we assume that the species found in our study is native but without

further genetic work the native-invasive status in the invader cannot be fully clarified. Interestingly, none of the parasites that were co-introduced with the Lessepsian migrant has been found to infect the native congeneric host species. While the total number of parasite species found in the two fish hosts in the Mediterranean Sea was similar (nine in the invasive and ten in the native fish), the mean number of parasite species found in an individual fish was significantly higher in the native compared to the invasive fish. Although the rarefaction analyses suggested that further sampling would likely reveal more rare parasite species for each fish host, this general pattern of a lower richness of the more common parasite species in the invasive host species would remain at higher sampling efforts. In general, the parasite fauna of the invasive fish was mainly composed of generalist native parasites acquired in the new environment and one co-introduced parasite species, while the parasite community in native fish included more specialist parasites. This is consistent with the general idea that invasive hosts often lose specific parasites (especially with complex life cycles) in the course of the introduction and acquire mainly native generalist parasites in the new range (Blakeslee, Fowler & Keogh, 2013; Lymbery et al., 2014).

In addition to parasite species richness, also total parasite prevalence (parasite infection status in the models) and total infection intensity were generally higher in the native than in the invasive fish. Hence, despite the acquisition of native parasites by the invasive host *S. chrysotaenia*, the native *S. sphyraena* still showed higher total infection levels. This suggests that the invasive host is experiencing lower parasite infection levels compared with the native host species. Several factors may explain the lower infection levels of the invasive compared to the native host species. First, the relatively small size of the invasive fish (20-25 cm length) compared to the native fish (30-60 cm) may cause a space constraint and reduce exposure of the

invasive fish to native parasites. However, the lack of a significant effect of fish size on infection levels in our analyses suggests that body size may only play a minor role. Second, differences in the feeding behaviour of the two fish hosts might explain why their parasite infection levels are different. While the invasive *S. chrysotaenia* narrows its food spectrum to pelagic fish species, the native *S. sphyraena* extended their feeding to supra-benthic species (Kalogirou et al., 2012). The broader food spectrum of the native fish may result in a relative higher exposure to parasites. Third, host suitability of the two fish species may differ and in particular invasive hosts may be less suitable for native parasites due to compatibility issues. Finally, phylogenetic niche conservatism (Wiens & Graham, 2005; Mouillot et al., 2006) and a variety ecological factors such as microhabitat use and life history strategies (Poulin, 2010) may play a role.

The presumably native isopod *Gnathia* sp. was the only parasite genus shared by the two fish hosts in Tunisian coastal waters, with a higher prevalence and intensity in the native *S. sphyraena* than in the invasive *S. chrysotaenia*. However, for both fish species there was only a single record (at location 1 in the invasive host and at location 7 in the native host), compromising a formal statistical comparison, rendering a solid discussion on the subsequent host and parasite populations.

Conclusion

Our study found evidence for the loss of parasite species of the Lessepsian migrant in the course of the introduction as well as for co-introduction of parasites and the acquisition of native parasites. These results suggest that the Lessepsian migrant has the potential to affect native fish

hosts by altering the population dynamics of native parasite species via parasite release, parasite co-introduction and acquisition of native parasites, resulting in increased infection levels in native hosts. They further suggest that the lower infection levels in the invasive host may give them a potential competitive advantage over native hosts. Further studies will be needed to investigate the resulting effects on native parasite dynamics and well as on native fish stocks. This study demonstrated that community studies or cross-species comparisons such as the one presented here, are valuable tools to identify the role of metazoan parasite in Lessepsian migration.

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References

- 433 **Allam SM, Faltas SN, and Ragheb E. 2005.** Stock assessment of Barracudas, genus *Sphyraena*,
434 along the Egyptian Mediterranean Coast. *Egyptian Journal of Aquatic Research* **31**: 281-
435 292.
- 436 **Bakopoulos V, Karoubali I, and Diakou A. 2017.** Parasites of the Lessepsian invasive fish
437 *Lagocephalus sceleratus* (Gmelin 1789) in the eastern Mediterranean Sea. *Journal of*
438 *Natural History* **51**: 421-434. DOI: 10.1080/00222933.2017.1279690
- 439 **Benmansour B, and Ben Hassine OK. 1998.** Preliminary analysis of parasitic copepod species
440 richness among coastal fishes of Tunisia. *Monitore zoologico italiano*. **65**: 341-344 DOI
441 10.1080/11250009809386844
- 442 **Berland B.(1961).** Nematodes from some Norwegian marine fishes. *Sarsia* **2**: 1- 50.
- 443 **Bilecenoglu M, Alfaya JEF, Azzurro E, Baldacconi R, Boyaci YO, Circosta V, Compagno**
444 **LJV, Coppola F, Deidun A, Durgham H, Durucan F, Ergüden D, Fernández-**
445 **Álvarez FÁ, Gianguzza P, Giglio G, Gökoğlu M, Gürlek M, Ikhtiyar S, Kabasakal**
446 **H, Karachle PK, Katsanevakis S, Koutsogiannopoulos D, Lanfranco E, Micarelli P,**
447 **Özvarol Y, Peña-Rivas L, Poursanidis D, Saliba J, Sperone E, Tibullo D, Tiralongo**
448 **F, Tripepi S, Turan C, Vella P, Yokeş MB, Zava B. 2013.** New Mediterranean marine
449 biodiversity records (December, 2013). *Mediterranean Marine Science* **14**: 463-480 DOI
450 10.12681/mms.676
- 451 **Blakeslee AMH, Fowler AE, and Keogh CL. 2013.** Marine invasion and parasite escape:
452 updates and new perspectives. *Advances in Marine Biology*. **66**: 87-169 DOI
453 10.1016/B978-0-12-408096-6.00002-X

- 454 **Boussellaa W, Boudaya L, Derbel H, and Neifar L. 2016.** A new record of the Lessepsian fish
455 *Etrumeus golanii* (Teleostei: Clupeidae) in the Gulf of Gabes, Tunisia, with notes on its
456 parasites. *Cahier de Biologie Marine* **57**: 389-395 DOI 10.21411/CBM.A.843F287A
- 457 **Bradai MN, Saidi B, Ghorbel M, Bouain A, Guelorget O, and Capape C. 2002.** Observations
458 sur les requins du golfe de Gabès (Tunisie méridionale, Méditerranée centrale). *Mesogée*
459 **60**: 61-77.
- 460 **Chaabane A, Justine J-L, Gey D, Bakenhaster MD, and Neifar L. 2016a.**
461 *Pseudorhabdosynochus sulamericanus* (Monogenea, Diplectanidae), a parasite of deep-
462 sea groupers (Serranidae) occurs transatlantically on three congeneric hosts (*Hyporthodus*
463 spp.), one from the Mediterranean Sea and two from the western Atlantic. *PeerJ* **4**:e2233.
464 DOI 10.7717/peerj.2233
- 465 **Chaabane A, Neifar L, Gey D, and Justine J-L. 2016b.** Species of *Pseudorhabdosynochus*
466 (Monogenea, Diplectanidae) from groupers (*Mycteroperca* spp., Epinephelidae) in the
467 Mediterranean and Eastern Atlantic Ocean, with special reference to the
468 "beverleyburtonae group" and description of two new species. *PLoS ONE* **11**: e0159886.
469 DOI 10.1371/journal.pone.0159886
- 470 **Châari M, and Neifar L. 2015.** Parasitic infections in the Mediterranean needle fish *Tylosurus*
471 *acus imperialis* (Teleostei: Belonidae) off Tunisian coast. *Journal of Coastal Life*
472 *Medicine* **3**: 673-676 DOI 10.12980/jclm.3.2015j5-78
- 473 **Chao A. 2005.** Species richness estimation. In: Balakrishnan N, Read CB, Vidakovic B, eds.
474 *Encyclopedia of statistical sciences*. Wiley: New York, 7909-7916.

- Colautti RI, Ricciardi A, Grigorovich IA, and MacIsaac HJ. 2004. Is invasion success explained by the enemy release hypothesis? *Ecology Letters* **7**:721-733 DOI 10.1111/j.1461-0248.2004.00616.x
- Colwell RK. 2013. *EstimateS: Statistical estimation of species richness and shared species from samples*. Version 9. User's Guide and application published at: <http://purl.oclc.org/estimates>.
- Dang C, Montaudouin X, Bald J, Jude F, Raymond N, Lanceleur L, Paul-Pont I, and Caill-Milly N. 2009. Testing the enemy release hypothesis: trematode parasites in the non-indigenous Manila clam *Ruditapes philippinarum*. *Hydrobiologia* **630**: 139-148 DOI 10.1007/s10750-009-9786-9
- Daszak P, Cunningham AA, and Hyatt AD. 2000. Emerging infectious diseases of wildlife: threats to biodiversity and human health. *Science* **287**: 443-449 DOI 10.1126/science.287.5452.443
- Dhrif E, Antar R, Abidli S, and Gargouri L. 2015. Digenean larvae parasitizing *Ruditapes decussatus* (Bivalvia) from Tunisian coasts. *Helminthologia* **52**: 104-112 DOI 10.1515/helmin-2015-0019
- Diamant A. 1989. Lessepsian migrants as hosts: a parasitological assessment of rabbitfish *Siganus luridus* and *S. rivulatus* (Siganidae) in their original and new zoogeographical regions. In: Spanier E, Steinberger Y, Lurin M, eds. *Environmental quality and ecosystem stability*: Environmental Quality ISEEQS Pub, Jerusalem, Israel. **4**: 187-194.
- Diamant A. 2010. Red-Med immigration: a fish parasitology perspective, with special reference to Mxyxosporea. In: Golani D, Appelbaum-Golani B, eds. *Fish Invasions of the Mediterranean Sea: Change and Renewal*. Pensoft Publishers: Sofia-Moscow, 85-97.

- El-Rashidy HH, and Boxshall GA. 2012. Bomolochid copepods (Crustacea: Copepoda: Bomolochidae) parasitizing immigrant and native barracuda (Actinopterygii: Sphyraenidae) caught off the Egyptian Mediterranean coast. *Zoosymposia* **8**: 20-28 DOI 10.11646/zoosymposia.8.1.5
- El-Shahawy IS, and Desouky ARY. 2010. *Myripristis murdjan* (Beryciformes: Holocentridae) a new host record for *Cymothoa indica* (Crustacea, Isopoda, Cymothoidae). *Acta Adriatica* **51**: 103-110.
- Elton CS. 1958. *The ecology of invasions*. Methuen: London, UK.
- Euzet L. 1994. Order Tetracanthidea Carus, 1863. In: Khalil LF, Jones A and Bray RA, ed. *Keys to the cestode parasites of vertebrates*. CAB International, Wallingford, U.K., 149-194.
- Euzet L, and Trilles J-P. 1960. Sur deux Monogènes nouveaux de *Sphyraena sphyraena* (L.) (Teleostei Sphyraenidae). *Bulletin de la Société Zoologique de France*. **85**: 189-198.
- Fischthal JH. 1982. Additional records of digenetic trematodes of marine fishes from Israel's Mediterranean coast. *Proceedings of the Helminthological Society of Washington* **49**: 34-44.
- Fisher W, Schneider M, and Bauchot ML. 1987. Fiches FAO d'identification des espèces pour les besoins de la pêche (Révision 1). *Méditerranée et Mer Noire*. Zone de pêche 37. Vertébrés. Rome FAO **2**: 1-1045.
- Forsyth DM, and Duncan RP. 2001. Propagule size and the relative success of exotic ungulate and bird introductions to New Zealand. *The American Naturalist* **157**: 583-595 DOI 10.1086/320626

- 519 **Galaktionov K, and Dobrovolskij AA. 2003.** *The biology and evolution of trematodes: An*
520 *essay on the biology, morphology, life cycles, transmissions, and evolution of digenetic*
521 *trematodes.* Springer, Dordrecht, Netherlands.
- 522 **Galil BS, and Innocenti G. 1999.** Notes on the population structure of the portunid crab
523 *Charybdis longicollis* Leene, parasitized by the rhizocephalan *Heterosaccus dollfusi*
524 Boschma, off the Mediterranean coast of Israel. *Bulletin of Marine Science* **64**: 451-463.
- 525 **Gargouri M, and Maamouri F. 2005.** The life cycle of *Bucephalus labracis* Paggi and
526 Orecchia, 1965 (Digenea, Bucephalidae), a parasite of *Dicentrarchus labrax* in Tunisia.
527 *Bulletin of the European Association of Fish Pathologists* **25**: 297-302.
- 528 **Gendron AD, Marcogliese DJ, and Thomas M. 2012.** Invasive species are less parasitized than
529 native competitors, but for how long? The case of the round goby in the Great Lakes-St.
530 Lawrence Basin. *Biological Invasions* **14**: 367-384 DOI DOI 10.1007/s10530-011-0083-
531 y
- 532 **Georgiev BB, Sanchez MI, Vasileva GP, Nikolov, PN, and Green AJ. 2007.** Cestode
533 parasitism in invasive and native brine shrimps (*Artemia* spp.) as a possible factor
534 promoting the rapid invasion of *A. franciscana* in the Mediterranean region. *Parasitology*
535 *Research* **101**: 1647-1655 DOI 10.1007/s00436-007-0708-3
- 536 **Gibson DI, and Bray RA. 1994.** The evolutionary expansion and host-parasite relationships of
537 the Digenea. *International Journal for Parasitology* **24**: 1213-1226 DOI 10.1016/0020-
538 7519(94)90192-9
- 539 **Gibson DI, Jones A, and Bray RA. 2002.** *Keys to the Trematoda*, Vol. 1. CABI Publishing:
540 Wallingford, UK DOI 10.1016/S0035-9203(03)90052-6

- 541 **Goedknecht MA, Havermans J, Waser AM, Luttikhuisen PC, Perdomo EV, Camphuysen K**
542 **(CJ), van der Meer J, and Thieltges DW. 2016.** Parasites and marine invasions:
543 Ecological and evolutionary perspectives. *Journal of Sea Research* **113**: 11-27 DOI
544 10.1016/j.seares.2015.12.003
- 545 **Goedknecht MA, Havermans J, Waser AM, Luttikhuisen PC, Vellila E, Camphuysen K**
546 **(CJ), Van der Meer J, and Thieltges DW. 2017.** Cross-species comparison of parasite
547 richness, prevalence, and intensity in a native compared to two invasive brachyuran
548 crabs. *Aquatic Invasions* **12**: 201-212 DOI 10.3391/ai.2017.12.2.08
- 549 **Golani D, and Ben Tuvia A. 1995.** Lessepsian migration and the Mediterranean fisheries of
550 Palestine. In: Armantrout NB, ed. *Conditions of the world's aquatic habits. Proceedings*
551 *of the World Fishery Congress*. New Delhi, India. 279-289.
- 552 **Gotelli NJ, and Colwell RK. 2001.** Quantifying biodiversity: procedures and pitfalls in the
553 measurement and comparison of species richness. *Ecology Letters* **4**: 379-391 DOI
554 10.1046/j.1461-0248.2001.00230.x
- 555 **İnnal D, Kirkim F, and Erk'akan F. 2007.** The parasitic isopods, *Anilocra frontalis* and
556 *Anilocra physodes* (Crustacea; Isopoda) on some marine fish in Antalya Gulf, Turkey.
557 *Bulletin of the European Association of Fish Pathologists* **27**: 239-241.
- 558 **Kabata Z. 2003.** Copepods Parasitic on Fishes. In: Crothers JH, Hayward PJ, eds. *Synopsis of*
559 *British Fauna* (New Series), Vol. 47 (revised). The Linnean Society & The Estuarine &
560 Coastal Sciences Association, London, 274 pp.
- 561 **Kalogirou S, Mittermayer F, Pihl L, and Wennhage H. 2012.** Feeding ecology of indigenous
562 and non-indigenous fish species within the family Sphyraenidae. *Journal of Fish Biology*
563 **80**: 2528-2548 DOI 10.1111/j.1095-8649.2012.03306.x

- 564 **Keane RM, and Crawley MJ. 2002.** Exotic plant invasions and the enemy release hypothesis.
565 *Trends in Ecology & Evolution* **17**: 164-170 DOI 10.1016/S0169-5347(02)02499-0
- 566 **Kelly DW, Paterson RA, Townsend CR, Poulin R, and Tompkins DM. 2009.** Parasite
567 spillback: A neglected concept in invasion ecology? *Ecology* **90**: 2047-2056 DOI
568 10.1890/08-1085.1
- 569 **Kritsky DC, Jiménez-Ruiz FA, and Sey O. 2000.** Diplectanids (Monogeneoidea:
570 Dactylogyridea) from the gills of marine fishes of the Persian Gulf off Kuwait.
571 *Comparative Parasitology* **67**: 145-164.
- 572 **Looss A. 1899.** Weitere Beiträge zur Kenntnis der Trematoden-Fauna Aegyptens, zugleich
573 versuch einer natürlichen Gliederung des Genus *Distomum* Retzius. *Zoologische*
574 *Jahrbucher* **12**: 521-784.
- 575 **Lymbery AJ, Morine M, Kanani HG, Beatty SJ, and Morgan DL. 2014.** Co-invaders: The
576 effects of alien parasites on native hosts. *International Journal for Parasitology:*
577 *Parasites and Wildlife* **3**: 171-177 DOI 10.1016/j.ijppaw.2014.04.002
- 578 **Maillard C. 1976.** Distomatoses de poissons en milieu lagunaire. Dr. Sci. Thesis. Montpellier:
579 Université des Sciences et Techniques du Languedoc.
- 580 **Merella P, Pais A, Follesa MC, Farjallah S, Mele S, Piras MC, and Garippa G. 2016.**
581 Parasites and Lessepsian migration of *Fistularia commersonii* (Osteichthyes,
582 Fistulariidae): shadows and light on the enemy release hypothesis. *Marine Biology* **163**:
583 97-97 DOI 10.1007/s00227-016-2865-3
- 584 **Mouillot D, Krasnov BR, Shenbrot GI, Gaston KJ, and Poulin R. 2006.** Conservatism of
585 host specificity in parasites. *Ecography* **29**: 596-602 DOI 10.1111/j.0906-
586 7590.2006.04507.x

- Nahhas FM, Sey O, and Nakahara G. 2006.** Digenetic trematodes of marine fishes from the Arabian Gulf off the coast of Kuwait. Family Bucephalidae Poche, 1907, and the description of a new species. *Helminthologia* **43**: 147-157 DOI 10.2478/s11687-006-0028-7
- Neifar L. 1995.** Contribution à l'étude de la biodiversité des monogènes parasites de poissons du secteur nord-est de la Tunisie. Mémoire de Diplôme d'Etudes Approfondies. Université de Tunis II.
- Neifar L, and Euzet L 2007.** Five new species of *Pseudorhabdosynochus* (Monogenea: Diplectanidae) from the gills of *Epinephelus costae* (Teleostei: Serranidae). *Folia Parasitologica* **54**: 117-128 DOI 10.14411/fp.2007.017
- Oliver G. 1987.** Les Diplectanidae Bychowsky, 1957 (Monogenea, Monopisthocotylea, Dactylogyridea). Systématique. Biologie. Ontogénie. Écologie. Essai de phylogénèse. Dr. Sci. Thesis. Université de Montpellier. DOI: 10.6084/m9.figshare.1295274
- Paggi L, and Orecchia P. 1965.** Su un nuovo trematode parassita dell'intestino di Morone labrax: *Bucephalus labracis* n. sp. *Parassitologia* **7**: 69-74.
- Pérez-del-Olmo A, Kostadinova A, and Gibson DI. 2016.** The Mediterranean: high discovery rates for a well-studied trematode fauna. *Systematic Parasitology* **93**: 249-256 DOI 10.1007/s11230-016-9626-z
- Petter AJ, and Maillard C. 1988.** Larves d'Ascarides parasites de poissons en Méditerranée occidentale. *Bulletin du Muséum national d'histoire naturelle. Section A, Zoologie, biologie et écologie animales. Muséum national d'Histoire naturelle: 4e série, tome* **10**: 347-369.

- 609 **Por FD. 1978.** Lessepsian migration: the influx of Red Sea biota into the Mediterranean by way
610 of the Suez Canal. Ecological studies. Springer-Verlag, Berlin, Germany.
- 611 **Poulin R, and Cribb TH. 2002.** Trematode life cycles: short is sweet? *Trends in Parasitology*
612 **18:** 176-183.
- 613 **Poulin R, and Morand S. 2004.** *Parasite Biodiversity*. Smithsonian Books, Washington DC,
614 USA DOI 10.1017/S003118200521908X
- 615 **Poulin R. 2010.** Network analysis shining light on parasite ecology and diversity. *Trends in*
616 *Parasitology* **26:** 492-498 DOI 10.1016/j.pt.2010.05.008
- 617 **Prenter J, MacNeil C, Dick JTA, and Dunn AM. 2004.** Roles of parasites in animal invasions.
618 *Trends in Ecology & Evolution* **19:** 385-390 DOI 10.1016/j.tree.2004.05.002
- 619 **Raibaut A, Combes C, and Benoit F. 1998.** Analysis of the parasitic copepod species richness
620 among Mediterranean fish. *Journal of Marine Systems* **15:** 185-206 DOI 10.1016/S0924-
621 7963(97)00079-1
- 622 **Ramdane Z, Bensouilah MA, and Trilles J-P 2009.** Étude comparative des crustacés isopodes
623 et copépodes ectoparasites de poissons marins algériens et marocains. *Cybium* **33:** 123-
624 131.
- 625 **R Development Core Team 2016.** R: A language and environment for statistical computing. R
626 Foundation for Statistical Computing, Vienna, Austria. URL [https://www.R-](https://www.R-project.org/)
627 [project.org/](https://www.R-project.org/)
- 628 **Relini M, and Orsi Relini L. 1997.** The two species of barracuda (Sphyraenidae) in the western
629 mediterranean. *Cybium* **21:** 216-222.

- 630 **Roche DG, Leung B, Franco EFM, and Torchin ME 2010.** Higher parasite richness,
631 abundance, and impact in native versus introduced cichlid fishes. *International Journal*
632 *for Parasitology* **40**: 1525-1530 DOI 10.1016/j.ijpara.2010.05.007
- 633 **Rouget M, and Richardson DM. 2003.** Inferring process from pattern in plant invasions: a semi
634 mechanistic model incorporating propagule pressure and environmental variables. *The*
635 *American Naturalist* **162**: 713-724.
- 636 **Santos CP, Mourão ED, and Cárdenas MQ. 2001.** *Pseudempleurosoma gibsoni* n. sp., a new
637 ancyrocephalid monogenean from *Paralanchurus brasiliensis* (Sciaenidae) from off the
638 southeastern coast of Brazil. *Memórias do Instituto Oswaldo Cruz* **96**: 215-219 DOI
639 10.1590/S0074-02762001000200013.
- 640 **Spicer IJ. 1931.** Report of the Department of Agriculture and Forests for the years 1927-30.
641 Jerusalem: Printing Office, Russian Building, 159-160.
- 642 **Surekha P, and Lakshmi C.V. 2005.** *Lecithochirium testelobatus* n. sp. (Digenea: Hemiuridae)
643 from the Lizard fish, *Saurida undosquamis* from Andhrapradesh Coast. *Journal of*
644 *Parasitic Diseases* **29**: 143-146
- 645 **Theisen S, Palm HW, Al-Jufaili SH, and Kleinertz S. 2017.** *Pseudempleurosoma haywardi* sp.
646 nov. (Monogenea: Ancyrocephalidae (sensu lato) Bychowsky & Nagibina, 1968): An
647 endoparasite of croakers (Teleostei: Sciaenidae) from Indonesia. PLoS ONE **12**:
648 e0184376. DOI 10.1371/journal.pone.0184376
- 649 **Torchin ME, Lafferty KD, and Kuris AM. 2001.** Release from parasites as natural enemies:
650 increased performance of a globally introduced marine crab. *Biological Invasions* **3**: 333-
651 345 DOI 10.1023/A:1015855019360

- 652 **Torchin ME, Lafferty KD, and Kuris AM. 2002.** Parasites and marine invasions. *Parasitology*
653 **124:** 137-151 10.1017/S0031182002001506
- 654 **Torchin ME, Lafferty KD, Dobson AP, McKenzie VJ, and Kuris AM. 2003.** Introduced
655 species and their missing parasites. *Nature* **421:** 628-630 DOI 10.1038/nature01346
- 656 **Torchin ME, and Mitchell CE. 2004.** Parasites, pathogens, and invasions by plants and
657 animals. *Frontiers in Ecology and the Environment* **2:** 183-190 DOI 10.1890/1540-
658 9295(2004)002[0183:PPAIBP]2.0.CO;2
- 659 **Townsend S. 2009.** Incorporating sustainable practices for zoos and aquariums: a Triple Bottom
660 Line approach. *International Zoo Yearbook* **43:** 53-63 DOI 10.1111/j.1748-
661 1090.2008.00065.x
- 662 **Trilles JP, and Bariche M 2006.** First record of the Indo-Pacific *Cymothoa indica* (Crustacea,
663 Isopoda, Cymothoidae), a Lessepsian species in the Mediterranean Sea. *Acta*
664 *Parasitologica* **51:** 223-230 DOI 10.2478/s11686-006-0035-3
- 665 **Vignon M, and Sasal, P. 2010.** The use of geometric morphometrics in understanding shape
666 variability of sclerotized haptor structures of monogeneans (Platyhelminthes) with
667 insights into biogeographic variability. *Parasitology International* **59:** 183-191 DOI
668 10.1016/j.parint.2010.01.006
- 669 **Wadie WF, and Riskallah SI 2001.** Fisheries for the genus *Sphyraena* (Perciformes,
670 sphyraenidae) in the southeastern part of the Mediterranean Sea. *Pakistan Journal of*
671 *Science* **10:** 21-34.
- 672 **Walther BA, and Morand S. 1998.** Comparative performance of species richness estimation
673 methods. *Parasitology* **116:** 395-405.

- 674 **Whitehead PJP, Bauchot ML, Hureau JC, Nielsen J, and Tortonese E, eds. 1984.** *Fishes of*
675 *the North-eastern Atlantic and the Mediterranean*. UNESCO, Paris, France.
- 676 **Wiens JJ, and Graham CH. 2005.** Niche conservatism: integrating evolution, ecology, and
677 conservation biology. *Annual Review of Ecology, Evolution, and Systematics* **36**: 519-
678 539. DOI 10.1146/annurev.ecolsys.36.102803.095431
- 679 **Wonham MJ, Carlton JT, Ruiz GM, and Smith LD. 2000.** Fish and ships: relating dispersal
680 frequency to success in biological invasions. *Marine Biology* **136**: 1111-1121.
- 681 **Yamaguti S. 1953.** Parasitic worms mainly from Celebes. Part. 2. Monogenetic Trematodes of
682 fishes. *Acta Medica Okayama* **8**: 204-256.
- 683 **Yuniar AT, Palm HW, and Walter T. 2007.** Crustacean fish parasites from Segara Anakan
684 Lagoon, Java, Indonesia. *Parasitology Research* **100**: 1193-1204
685 DOI 10.1007/s00436-006-0391-9
- 686 **Zenetos A, Meriç E, Verlaque M, Galli P, and Boudouresque CF. 2008.** Additions to the
687 annotated list of marine alien biota in the Mediterranean with special emphasis on
688 Foraminifera and Parasites. *Mediterranean Marine Science* **9**: 119-166 DOI
689 10.12681/mms.146
- 690 **Zouari-Ktari R, Bradai MN, and Bouain A. 2009.** Reproduction and growth of the
691 Yellowstripe Barracuda *Sphyrna chrysotaenia* Klunzinger 1884, in Central
692 Mediterranean. *Reviews in Fisheries Science* **17**: 485-493
693 DOI 10.1080/10641260903082471

Table 1(on next page)

Information on the sampling design of fish hosts collected for this study in Tunisian coastal waters.

Given are the location numbers used in Figure 1, location name, geographic coordinates, sampling dates and sample sizes (per sex) for the two fish host species (*Sphyraena sphyraena* and *Sphyraena chrysotaenia*).

1

2

Location number	Location name	Geographic coordinates	Sampling dates	<i>Sphyraena sphyraena</i> (native)		<i>Sphyraena chrysotaenia</i> (invasive)	
				Females	Males	Females	Males
1	Sfax	34° 44' 26" N 10° 45' 37" E	22/10/2012 08/03/2013	3	3	9	9
2	Kerkennah	34° 39' 29" N 11° 04' 07" E	10/02/2013 10/03/2013 10/09/2013	20	11	18	15
3	Skhira	34° 17' 57" N 10° 04' 11" E	06/11/2014 04/07/2015	10	8	7	10
4	Chebba	35° 14' 14" N 11° 6' 54" E	05/11/2012	8	1	5	14
5	Zarat	33° 39' 59" N 10° 20' 59" E	07/02/2014	6	1	15	6
6	Zarzis	33° 30' 14" N 11° 06' 43" E	11/05/2015 19/06/2015	11	6	9	7
7	Sayada	35° 40' 7" N 10° 53' 32" E	07/02/2014 03/06/2014	15	4	7	17
Total				73	34	70	78

Table 2 (on next page)

Parasite species of *Sphyraena sphyraena* and *Sphyraena chrysoteania* found in this study and recorded in the literature.

Information on the life cycle and the invasive/native status in the Mediterranean Sea and occurrence in the two host species and regions (Mediterranean Sea and Red Sea). If quantitative data were available, mean prevalence and intensity (\pm SE) in a host species and region are given. + denotes published records of specific parasite species in a host species in a region; (+) denotes an assumed occurrence in a host species and region without published records.

Parasite taxa	Parasite species	Life cycle	Host specificity	Status Med Sea	<i>Sphyraena sphyraena</i> (native range- Med Sea)	<i>Sphyraena chrysotaenia</i> (invasive range - Med Sea)	<i>Sphyraena chrysotaenia</i> (native range - Red Sea)	Reference
Annelida	<i>Piscicolid</i> sp.	complex	generalist	native		0.7% (1 ± 0)		This study
Cestoda	Tetracophyllidea	complex	generalist	native	3.7% (16 ± 0.4)			This study
Copepoda	<i>Caligus</i> sp.	direct	Specialist/generalist?	native		3.8% (1± 0)		This study
	<i>Bomolochus unicirrus</i>	direct	specialist	native	29% (2 ± 0.1)			This study
	<i>Nothobomolochus denticulatus</i>	direct	specialist	invasive		+	(+)	El-Rashidy and Boxshall, 2012
	<i>Pennella filosa</i>	complex	generalist	native	+			Ramdane et al. 2009
	<i>Lecithochirium</i> sp.	complex	generalist	native		12.2% (1.4 ± 0)		This study
Digenea	<i>Didymozoon sphyraenae</i>	complex	specialist	native	85% (5.5 ± 0.4)			This study
	<i>Bucephalus sphyraenae</i>	complex	specialist	not present			+	Nahhas et al. 2006
	<i>Bucephalus labracis</i>	complex	specialist	native		+		Fischthal, 1982
	<i>Plerurus digitatus</i>	complex	generalist	native	+			Looss, 1899
	<i>Gnathia</i> sp.	complex	generalist	native	0.9% (13 ± 0.1)	0.7% (2± 0)		This study
Isopoda	<i>Cymothoa indica</i>	complex	generalist	invasive		+	(+)	Trilles and Bariche, 2006
	<i>Anilocra physodes</i>	complex	generalist	native		+		İnnal et al. 2007
	<i>Pseudempleurosoma</i> sp.	direct	specialist	invasive		4 % (1± 0)	(+)	This study
Monogenea	<i>Chauhanea mediterranea</i>	direct	specialist	native	5.6% (1.4 ± 0.1)			This study
	<i>Pseudolamellodiscus sphyraenae</i>	direct	specialist	not present			+	Yamaguti, 1953; Kritsky et al. 2000
	<i>Cotyloatlantica mediterranea</i>	direct	specialist	native	+			Euzet and Trilles, 1960
	<i>Rhinecotyle crepitacula</i>	direct	specialist	native	+			Euzet and Trilles, 1960
	<i>Anisakis</i> sp.	complex	generalist	native	1.9% (3 ± 0)			This study
Total species richness					10	9	5	
1								
2								
3								
4								

Table 3(on next page)

Parasite species of the genus *Lecithochirium* recorded in the literature, with information on the life cycle (Intermediate host and Definitive host) in the Mediterranean Sea.

Parasites	Intermediate host	Definitive host
<i>Lecithochirium fusiforme</i> Luhe, 1901	<i>Gibbula cineraria</i>	<i>Conger conger</i> <i>Lophius piscatorius</i>
<i>Lecithochirium musculus</i> (Looss, 1907)	<i>Conger myriaster</i> <i>Conger japonicus</i>	<i>Anguilla anguilla</i> <i>Trachurus trachurus</i> <i>Serranus hepatus</i> <i>Crenilabrus cinereus</i> <i>Conger conger</i> <i>Atherina Hepsetus</i> <i>Serranus scriba</i>
<i>Lecithochirium physcon</i> Luhe, 1901	-	<i>Lophius piscatorius</i>
<i>Lecithochirium rufoviride</i> (Rudolphi, 1819)	<i>Gibbula cineraria</i> <i>Blennius pholis</i>	<i>Anguilla anguilla</i>
<i>Lecithochirium texanum</i> (Chandler, 1941)	-	<i>Euthynnus alleteratus</i>
<i>Lecithochirium jaffense</i> Fischthal, 1982	<i>Seriola dumerili</i>	<i>Trachinotus ovatus</i> <i>Epinephelus</i> sp. <i>Echeneis naucrates</i> <i>Pomatomus saltatrix</i> <i>Gobius cobitus</i> <i>Etrumeus golanii</i>
<i>Lecithochirium haifense</i> Fischthal, 1980	-	<i>Atule djeddaba</i>
<i>Lecithochirium magnicaudatus</i> (Fischthal & Kuntz, 1963)	-	<i>Saurida undosquamis</i> <i>Labeo forskalii</i>
<i>Lecithochirium microstomum</i> Chandler, 1935	-	<i>Trichiurus lepturus</i>
<i>Lecithochirium grandiporum</i> (Rudolphi, 1819)	-	<i>Muraena helena</i> <i>Lophius piscatorius</i> <i>Conger conger</i> <i>Saurida tumbil</i>

Figure 1

Sampling locations (1-7) of the native *Sphyraena sphyraena* and the invasive Lessepsian migrant *Sphyraena chrysotaenia* in the central Mediterranean Sea (a) along the Tunisian coast (b) .

For location names, coordinates, sampling dates and sampling effort per species see Table 1.

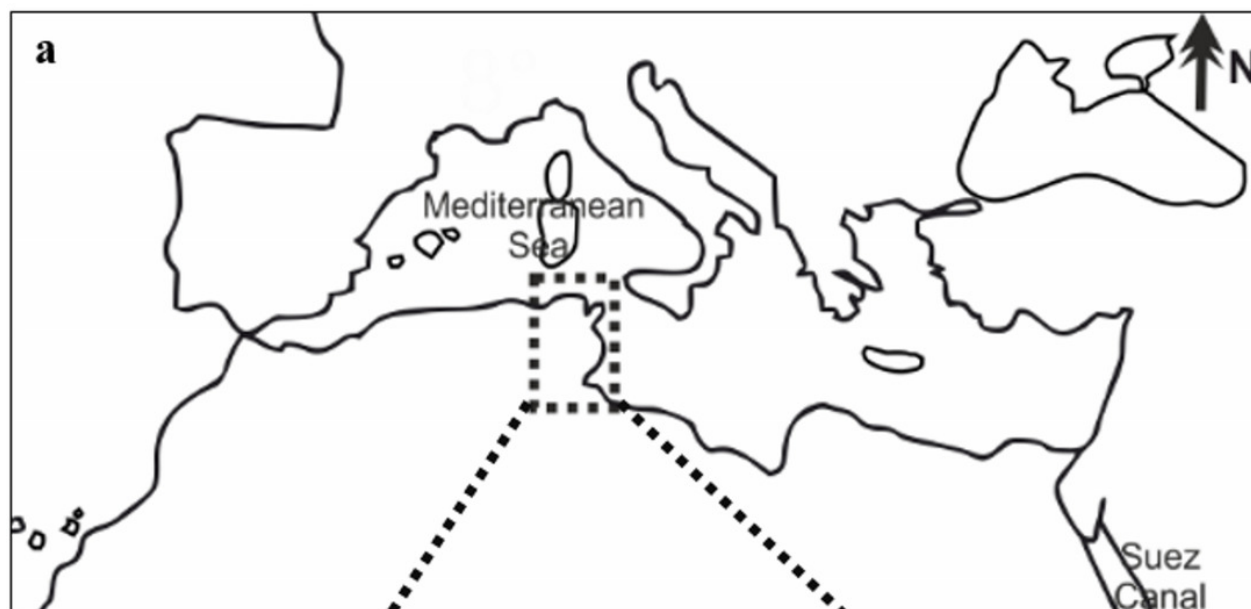


Figure 2

Rarefaction curves for parasite richness for a) the native fish *Sphyraena sphyraena* and b) the invasive Lessepsian migrant *Sphyraena chrysotaenia*.

Shown are the observed species richness accumulation curves (S_{obs}) which is the mean number of species among runs and the predicted number of species (\pm SE) based on the Chao2 estimator algorithm.

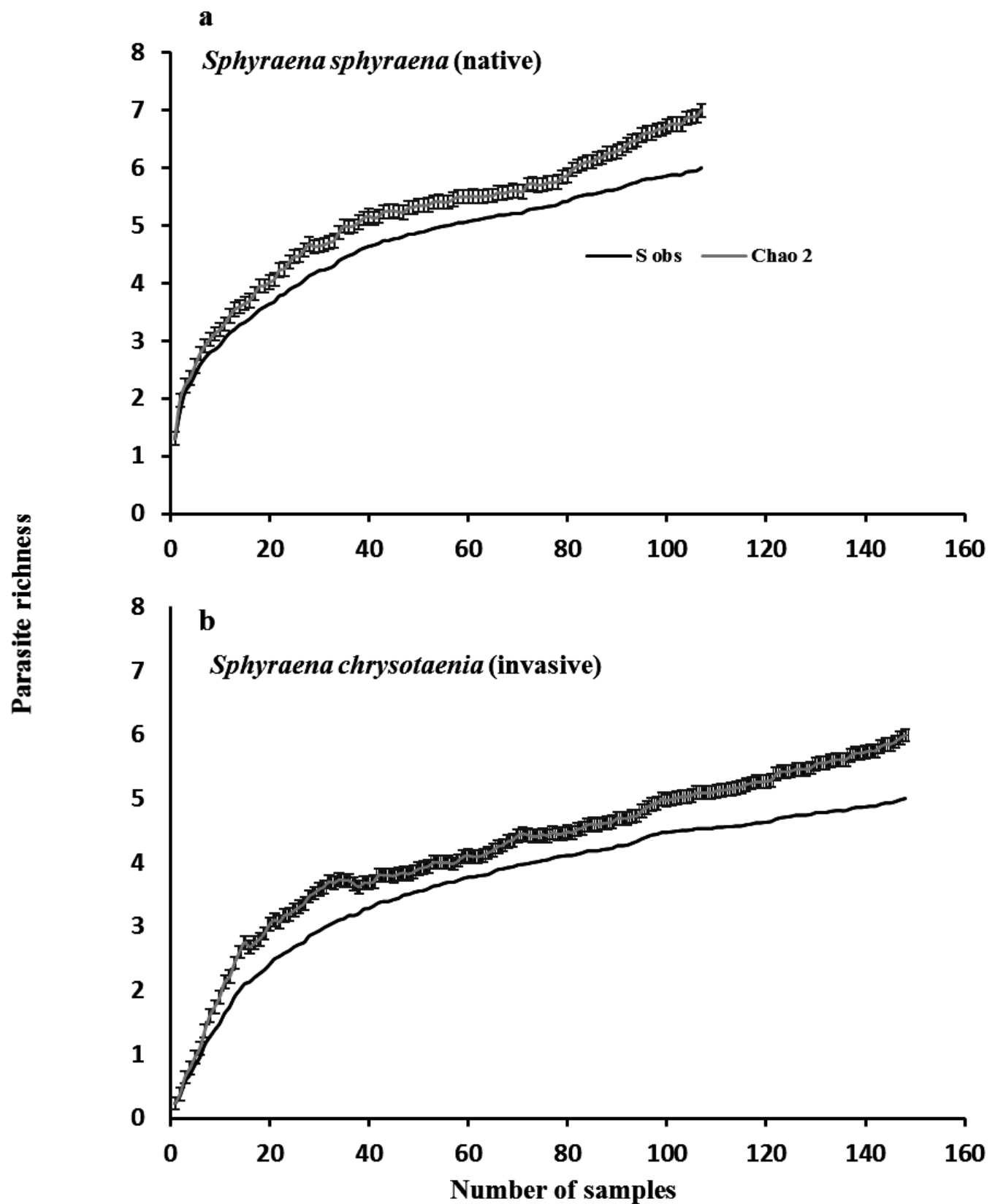


Figure 3

Mean parasite species richness per individual fish.

(including non-infected hosts; \pm SE) at the seven locations in the native fish *Sphyraena sphyraena* (n = 107) and the invasive congeneric *Sphyraena chrysotaenia* (n = 148).

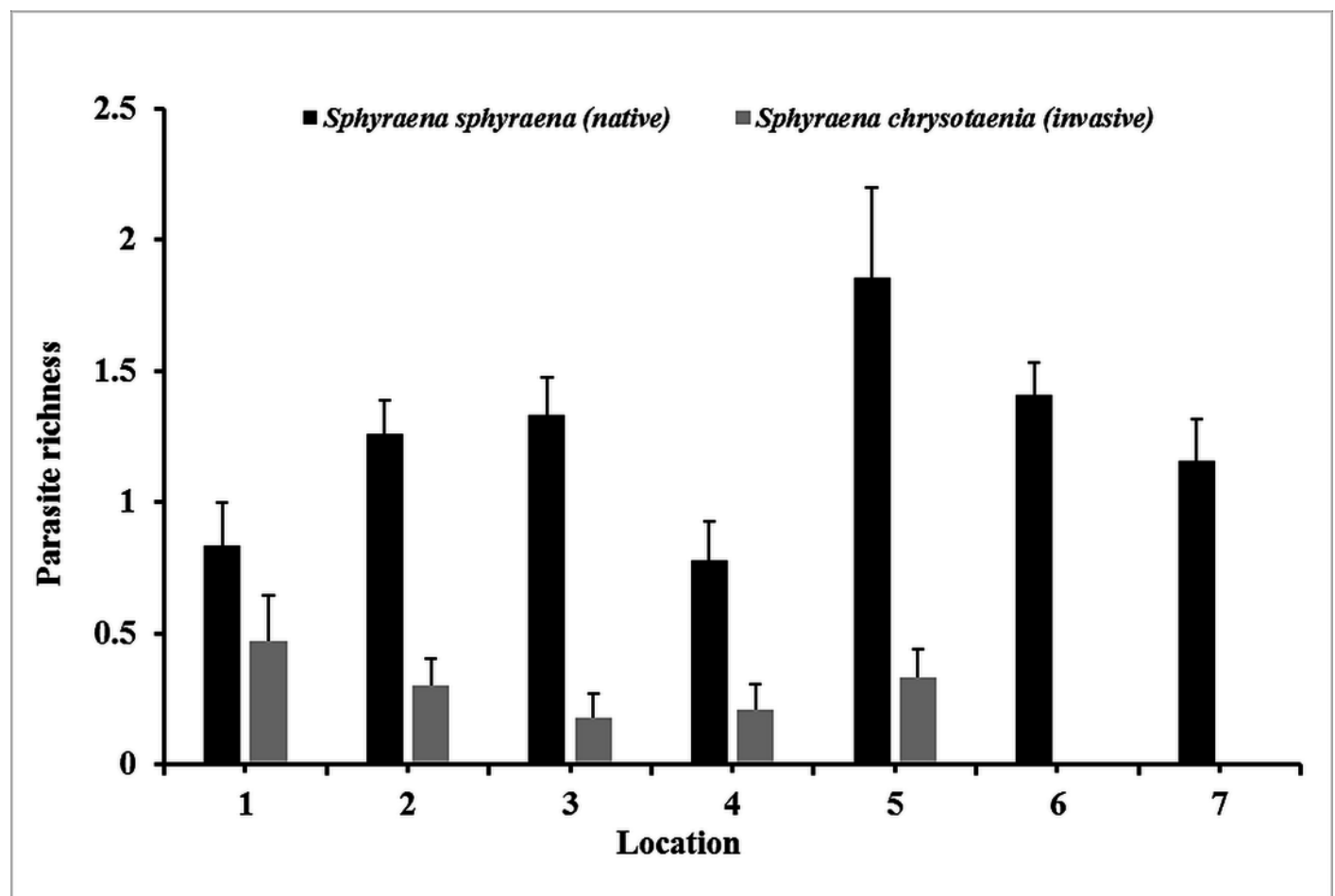


Figure 4

a) Mean total prevalence of all parasites in hosts and b) mean intensity of total parasite loads in native *Sphyraena sphyraena* and invasive *Sphyraena chrysotaenia* at the seven sampling locations.

Mean total prevalence (i.e. hosts infected by 1 or more parasite species); and mean intensity loads (i.e. sum of all parasite individuals found in a host) For sample size (n) per host species and location, see Table 1.

