

**Disinfection effect of povidone-iodine in aquaculture water of swamp  
eel (*Monopterus albus*)**

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## Abstract

The swamp eel (*Monopterus albus*) is an important commercially farmed fish species in China. However, it is susceptible to *Aeromonas hydrophila* infections, resulting in high mortality and considerable economic loss. Povidone-iodine (PVP-I) is a widely used chemical disinfectant in aquaculture, which can decrease the occurrence of diseases and improve the survival. However, environmental organic matter could affect the bactericidal effectiveness of PVP-I, and the efficacy of PVP-I in aquaculture water is still unknown. In this paper, disinfection assays were conducted to evaluate the effectiveness of PVP-I against the *A. hydrophila* in different types of water. We found that the effective germicidal concentration of PVP-I in outdoor aquaculture water was 25 ppm ~~during~~for 12 hours. In indoor aquaculture water with  $10^5$  CFU/mL bacteria, 10 ppm and 20 ppm of PVP-I could kill 99 % and 100 % of the bacteria, respectively. The minimal germicidal concentration of PVP-I in deionized distilled water and Luria-Bertani broth was 1.25 ppm and 4000 ppm, respectively. Available iodine content assay in LB solutions confirmed that the organic substance had negative impact on the effectiveness of PVP-I, which was consistent with the different efficacy of PVP-I in different water samples. Acute toxicity tests showed that the 24h-LC<sub>50</sub> of PVP-I to swamp eel was 173.82 ppm, which was much lower than the germicidal concentrations in outdoor and indoor aquaculture water, indicating its safety and effectivity to ~~antagonize~~control the *A.*

*hydrophila*. The results ~~presented here would~~ indicated povidone iodine can be helpful  
to for preventing the transmission of *A. hydrophila* in swamp eel aquaculture.

## Introduction

The swamp eel (*Monopterus albus*), belongs to Order Synbranchiformes, Family Synbranchidae, and is widely distributed in southern China, Japan, India and other Southeast Asian countries (FishBase, <http://fishbase.org/>). Due to its great growth performance and rich nutrient content, swamp eel has become a commercially important farmed species in China. However, the intensive and stressful rearing conditions make farmed swamp eels highly susceptible to bacterial pathogens, such as *Aeromonas hydrophila*, which is the main pathogen causing haemorrhagic septicemia and result in high mortality and considerable economic losses (Nielsen et al., 2001; Hossain et al., 2014; ~~Nielsen et al., 2001~~). The clinical signs of infection contains include red and swollen anus, visceral congestion, skin erythematous and gill haemorrhage hemorrhage (Jagoda et al., 2014).

The haemorrhagic hemorrhagic septicemia severely restricts the development prospects of the swamp eel farming industry. To prevent the spread of *A. hydrophila* and outbreak of the diseases in aquaculture, the direct and effective method is to reduce the amount of pathogenic bacteria in the aquaculture water. Povidone-iodine (PVP-I) is an important chemical disinfection widely used to disinfect pathogenic organisms and equipments in aquaculture (Scarfe et al., 2006). It has been reported as a broad-spectrum microbicide with potency to inactivate bacteria, fungi, protozoans, several viruses and some spores (Wutzler et al., 2000). Moreover, PVP-I was

determined to be an animal drug of low regulatory priority by the U.S. Food and Drug Administration, and suggested to act as an egg surface disinfectant due to the lower irritation and toxicity to tissues (USFDA, 2010). However, many environmental factors can affect the efficacy of PVP-I, such as temperature, pH, organic matter, etc. (Amend, 1974). The presence of organic matter can result in a significant decrease in the bactericidal effectiveness of PVP-I (Rodriguez Ferri et al., 2010). By far, most disinfection researches were has been carried out in sterile water (Chang et al., 2015), and only very few reports were mention use of available in aquaculture water for testing (Hershberger et al., 2008).

Swamp eel aquaculture includes outdoor cage culture and indoor tank culture. The former is mainly used for large-scale commercial swamp eel production, and the latter is commonly used in laboratory research. ~~It must be pointed that~~ However, outdoor ~~culture water body is~~ aquaculture ponds are a complex environment containing large amounts of organic matter and suspended solids (Lin, 2002). However, indoor tank culture mainly uses dechlorinated tap water with water changes every day, so the content of organic matter of indoor culture water is very low. Different contents of organic matter may cause differences in disinfection efficiency of PVP-I (Yoneyama et al., 2006). Accordingly, it is necessary to test whether the organic matter in the aquaculture water is able to cause significant negative effect on disinfection.

The aim of this study was to investigate the germicidal effect of PVP-I on *A. hydrophila* in four different solutions and attempt to determine the effective disinfection concentrations to control the transmission of *A. hydrophila*.

## Metrials and methods

### Bacterial strain

Six isolates of *A. hydrophila* used in this study (Ah 1-6) were isolated from sick *M. albus* by our laboratory and stored at -80 °C until used. Before disinfection assays, all isolates were subcultured at 25 °C overnight on solid-phase Luria-Bertani (LB) agar.

### Water sample preparation

Four different water samples were prepared in this study: outdoor net cage water, dechlorinated tap water, LB broth and deionized distilled water (ddH<sub>2</sub>O). These four types of water samples represented outdoor aquaculture waters, indoor aquaculture waters, eutrophic waters (positive control) and non-nutritive waters (negative control), respectively. Outdoor aquaculture waters were sampled from four different net cages of a swamp eel farm, and some of the swamp eel in one cage appeared-had typical clinical signs of haemorrhagichemorrhagic septicemia. Tap water samples were filtered through 0.22 µm membrane to remove impurities and microbes, then autoclaved at 121 °C for 15 min. LB broth was prepared with deionized distilled water.

### Germicidal test

PVP-I solutions were prepared with different water samples immediately prior to use. For outdoor aquaculture water, 1 mL of each net cage samples were treated with 5 to-100 ppm PVP-I (final concentration) at 25 °C in the dark for different durations (?,?, and ?)time.

For indoor aquaculture water (tap water), ddH<sub>2</sub>O and LB solution, *A. hydrophila* strains were used to evaluate the disinfection effect of PVP-I. Briefly, for indoor aquaculture water, six *A. hydrophila* isolates (Ah 1-6) were incubated in LB liquid media overnight at 25 °C, respectively. Then 5 mL of each culture broth was centrifuged at 12000 rpm for 5 minute at 25 °C. The bacterial pellet was washed with tap water three times and then resuspended and diluted to different concentration (10<sup>3</sup>-10<sup>5</sup> CFU/mL). Disinfection treatments were performed by mixing 900 µL of diluted bacteria solutions and 100 µL PVP-I solution to a 1 mL volume containing 10<sup>3</sup>-10<sup>5</sup> CFU/mL bacteria and 1.25-20 ppm of PVP-I (final concentration). For ddH<sub>2</sub>O

and LB liquid media, only *A. hydrophila* Ah1 isolate (isolated from skin) was used, and the disinfection treatments were performed ~~as same as that~~like those for the in indoor aquaculture water except that the tap water was replaced by ddH<sub>2</sub>O or LB liquid media.

For all above disinfection trials, treatment tubes were incubated at 25 °C in the dark with gentle mixing. At each ~~preconcerted~~nominal exposure time (0, 2, 4, 8 and 12 hours), a 160 µL aliquot was ~~taken out~~transferred to a sterile 1.5 mL microcentrifuge tube, and the equal volume of sodium thiosulfate (0.004 mol/L) was added to neutralize the PVP-I immediately. After neutralization, the mixed solution was tenfold serially diluted, and then 100 µL of each diluted solution was plated onto LB agar plates in triplicate using sterile beads. Plates were incubated at 25 °C for 24 hours and colonies were counted. Every experiment was repeated three times.

#### **Effect of organic matter on available iodine**

LB liquid media was 10-fold serially diluted with ddH<sub>2</sub>O. Then the 2000 ppm of PVP-I solutions were prepared with serially diluted LB solutions. 1 mL of PVP-I solution was mixed with 2 mL of 0.1% soluble starch solution, and the absorption was measured at 585 nm immediately. The undiluted LB broth and ddH<sub>2</sub>O were used as controls. The experiment was repeated three times.

#### **Median lethal concentration (LC<sub>50</sub>)**

Healthy swamp eel (11-15 g) were obtained from a commercial farm and acclimated in dechlorinated tap water at 25±1 °C in 10 L aquarium tanks for 2 weeks until use. ~~Daily, eels were~~fed and ~~renewed the tank water~~replaced with fresh water ~~once a day~~. The fishes were starved for 24 hours prior to and during the test to reduce the contaminations by fecal and excess food. Five concentrations of PVP-I were chosen for testing purposes (100, 150, 175, 200 and 250 ppm) and a group without PVP-I was used as the control. ~~The duration of exposure was ??~~. For each concentration and control, three replications were conducted and each replication contained ten swamp eel. Fish were observed every four hours and the dead fish was

immediately removed from the test tanks. The  $LC_{50}$  value was calculated using the Probit analysis (Lin, 2002; Lu et al., 2017). All animal procedures were conducted according to the guidelines for the care and use of experimental animals established by the Ministry of Agriculture of China (No. SCXK YU2005-0001). Animal Care and Use Committee (ACUC) in Jiangxi Agricultural University, who specially approved this study.

## Statistical analysis

SPSS17.0 was used for data analysis. Significance was evaluated by one-way analysis of variance (ANOVA) using LSD test. A value of  $P < 0.05$  was considered to indicate a significant difference.

## Results

### The effect of PVP-I in outdoor aquaculture water

Four outdoor net cage water samples were marked as WA, WB, WC and WD. pH value of these cage water samples were 7.12, 7.25, 7.08 and 7.32, respectively. The water sample WD was taken from a cage in which haemorrhagic septicemia was discovered. According to plate count, the initial cultivable bacterial concentration of these samples were  $(0.70 \pm 0.07) \times 10^3$  CFU/mL,  $(1.59 \pm 0.23) \times 10^3$  CFU/mL,  $(0.38 \pm 0.04) \times 10^3$  CFU/mL and  $(3.96 \pm 0.22) \times 10^3$  CFU/mL, respectively. The germicidal test showed that low concentrations of PVP-I, such as 5 ppm and 10 ppm, could not provide effective disinfection (Fig. 1). When treated with these two concentrations of PVP-I, the average survival decreased in early period, then increased and finally up to about 1.5-2.5 fold and 0.8-1.5 fold of initial bacteria amount, respectively. Increasing the PVP-I concentration to 25 ppm significantly improved the bactericidal effects resulting in  $98.08 \pm 0.17\%$ ,  $98.74 \pm 0.13\%$ ,  $96.49 \pm 0.03\%$  and  $99.66 \pm 0.03\%$  mortality in four outdoor water samples for 12 hours, respectively (Fig. 1). However, higher PVP-I concentrations (50-100 ppm) did not

significantly increase the sterilization rate ( $P>0.05$ ), although 80 ppm and 100ppm could kill all bacteria in WA and WD samples.

### **The effect of PVP-I in indoor aquaculture water**

In small-scale indoor farming, the dechlorinated tap water is often used as aquaculture water. The results showed that the effects of PVP-I on six *A. hydrophila* isolates was similar in tap water, and with the increase of bacterial content, the concentration of PVP-I for complete sterilization increased accordingly. In tap water with  $10^3$  CFU/mL bacteria (Fig. 2A), 1.25 ppm of PVP-I (final concentration) could reduce bacterial counts within 12 hours, but did not completely eliminate the *A. hydrophila*. Increasing the concentration of PVP-I to 2.5 ppm, all bacteria were killed within 4 hours. When the bacterial content was  $10^4$  CFU/mL (Fig. 2B), 10 ppm of PVP-I was required to achieve complete disinfection within 2 hours, and lower concentrations (2.5 ppm and 5 ppm) could not killed all bacteria. When the bacterial content was  $10^5$  CFU/mL (Fig. 2C), although 10 ppm of PVP-I could kill more than 99% of the bacteria, to achieve 100% of sterilization 20 ppm of PVP-I was required. When treated with lower concentrations, the amount of culturable bacteria gradually declined to the lowest in 8 hours, and then increased to different levels.

### **The effect of PVP-I in ddH<sub>2</sub>O and LB liquid media**

LB liquid media and deionized distilled water were used as eutrophic and non-nutritive controls, respectively. The results showed that effective disinfectant concentrations of PVP-I were significantly different between these two types of solutions. In LB liquid media, neither 500 ppm nor 1000 ppm of PVP-I could inhibit the proliferation of *A. hydrophila* at all bacterial concentrations. For example, in LB solution with  $10^3$ ,  $10^4$  and  $10^5$  CFU/mL bacteria (Fig. 3), after 12 hours of incubation with 500 ppm of PVP-I, the number of bacteria increased to 1800, 6050 and 20,000 times to the initial number of bacteria, respectively. Increasing the PVP-I concentration to 2000 ppm, ~~could only worked-controlled bacteria at the lowest bacteria concentration~~ (~~in~~  $10^3$  CFU/mL of LB medium; ~~Fig. 3A~~). To achieve

complete disinfection, 4000 ppm were required and in this concentration all *A. hydrophila* could be killed within 2 hours.

In comparison, the bactericidal efficiency of PVP-I in deionized distilled water was much higher (Fig. 4). In  $10^5$  CFU/mL of deionized distilled water, as low as 1.25 ppm of PVP-I was enough to kill all bacteria in 8 hours. Lower bacterial concentrations resulted in less sterilization times (4 hours for  $10^4$  CFU/mL and 2 hours for  $10^3$  CFU/mL). Increasing the PVP-I concentration to 5 ppm, the sterilization time could be shortened to 2 hours at all bacterial concentrations.

### Available iodine measurement

LB medium contained a large amount of organic matter, so it was used to evaluate the effect of organic matter on the available iodine content in this study. The results showed that the concentration of organic matter could significantly affect the available iodine content, and the higher LB concentrations lead to the lower available iodine contents (Fig. 5). In 2 g/L PVP-I solutions, when the concentration of LB was less than 1%, the content of available iodine was equivalent with that in ddH<sub>2</sub>O. With the increase of LB concentration, the available iodine content decreased rapidly, and when the LB concentration was 20%, the effective iodine content almost reduced to zero.

### LC<sub>50</sub> test

Median lethal dose of PVP-I was calculated using the Probit analysis (Table S1). According to the equation:  $\text{Probit}(p) = -5.214 + 0.03 \times \lg(\text{dose})$  ( $\chi^2 = 6.343$ ), the 24h-LC<sub>50</sub> of PVP-I to swamp eel was 173.82 ppm. The individuals that survived the LC<sub>50</sub> test did not die anymore within a month following exposure.

### Discussion

Disease is a main threat in aquaculture production and disinfection of water bodies has been used as an important measure to prevent waterborne pathogen transmission

in aquaculture (Scarfe et al., 2006). The present disinfection tests were mainly carried out in sterile water (Hershberger et al., 2008; Mainous et al., 2010), while very few in aquaculture water. However, there is a great differences between sterile water and aquaculture water, and the results obtained in sterile water are not suitable for aquaculture water. The aim of this study was to evaluate the effective concentration of PVP-I against *A. hydrophila* in outdoor aquaculture water and indoor aquaculture water.

Our study showed that PVP-I was effective to prevent *A. hydrophila* proliferation in outdoor and indoor aquaculture water and there were water-specific differences in susceptibility. The effective germicidal concentration of PVP-I in outdoor aquaculture water, indoor aquaculture water (tap water), eutrophic water (LB broth), and non-nutritive water (ddH<sub>2</sub>O) were 25 ppm, 10 ppm, 4000 ppm and 1.25 ppm, respectively. These water-specific differences in bactericidal effect might be mainly caused by organic matter. It had been proved by serially diluted LB solution, in which the higher ratio of organic matter (LB medium) lead to the lower concentration of available iodine, and 20% LB was sufficient to neutralize all the free iodine in 2000 ppm PVP-I solutions. This result was also supported by many reports which indicated that the efficacy of PVP-I declined when organic matter (e.g. blood, fish mucus, amino acids, or simple aromatic compounds) was in present (Truesdale and Luther, 1995; Yoneyama et al., 2006). Similar results also had been observed in seawater that the content of molecular iodine reduced within hours when added to seawater (Truesdale et al., 1995). ~~Although the content of organic matter is not very rich in normal aquaculture, but enough to weaken the germicidal effect. Meanwhile,~~ Considering the differences in different aquaculture water, it was necessary to determine the effective disinfectant concentration before use. It should be also noted that, in addition to the effects of a small amount of organic matter in tap water, the differences of disinfection between tap water and deionized distilled water might be caused by inorganic matter, which had been proved that the hardness of the water could affect the available iodine content by influencing pH (Amend, 1974).

Disinfection can be used not only as a preventive measure in ponds or for equipments, but also as a remedial measure after the outbreak of the disease. The majority of commercially available disinfectants are very effective when ~~worked-used~~ at high concentrations within short contact time. However, for fish, prolonged exposure to high concentrations of disinfectants might be dangerous (Bergmann et al., 2017). Especially for extensive outdoor aquaculture, it is impossible to change the water after disinfection. Therefore, it was necessary to determine the toxicity of disinfectants to fish (LeValley, 1982). In this study, the 24h-LC<sub>50</sub> for *M. albus* was 173.82 ppm, which was much higher than the effective germicidal concentration in aquaculture water, suggesting that PVP-I could be used ~~as a safety~~ as a disinfectant for the culture of *M. albus*. On the other hand, the commercial standard procedure for PVP-I is to pour directly into the water with the final concentration of 0.075-0.093 ppm for prevention and 0.093-0.125 ppm for treatment (Wang et al., 2015). Our data showed that these recommended concentrations were far less than the effective concentrations and would not have any germicidal effect on *A. hydrophila*.

Compared with higher PVP-I concentration, the lower PVP-I concentrations reduced the toxicity to *M. albus*, but ~~leading to worse~~ reduced the bactericidal effect. For example, in outdoor aquaculture water, indoor aquaculture water (tap water) and LB medium, when treated with non-lethal concentration, the number of ~~culturable~~ cultivable bacteria decreased first, then increased during 12 hours. Similar phenomenon was also reported in seawater when treated with some disinfectants (sodium hypochlorite, bleaching powder, formalin) for 12 hours (Wang et al., 2015). The increase of bacteria in later period might be due to the decrease of available iodine content and the proliferation of surviving bacteria. On the one hand, the content of available iodine decreased significantly with the passage of time (Chang et al., 2015); on the other hand, the organic matter in the solution would neutralize free iodine (Takeda et al., 2016). Meanwhile, the organic matter in water samples could supply nutrients for the proliferation of the surviving culturable bacteria. Our results ~~were tally with~~ support this ~~speculate~~ hypothesis; the earliest rise and largest increase

of the number of *A. hydrophila* happened in LB broth which contains the most abundant organic matter, followed by outdoor aquaculture water and tap water. The lowest effective bactericidal concentration and no bacterial increase were found in deionized distilled water, which did not contain any organic matter. Moreover, Kersters et al. (1996) reported ~~there had been proved~~ that *A. hydrophila* was able to grow and proliferation in nutrient-poor filtered and autoclaved tap water ( ~~Kersters et al., 1996~~), which made it hard to control the proliferation of *A. hydrophila* effectively.

It should be noted that we did not consider the viable but non-culturable (VBNC) state of *A. hydrophila*. Although there was no work about the relationship between the VBNC of *A. hydrophila* and disinfection, it has been demonstrated that stressed and starved *A. hydrophila* could enter a VBNC state (Pianetti et al., 2008; Rahman et al., 2001). ~~And~~ Disinfectants, such as hypochlorous acid, ~~had been proved to similarly~~ have induced *Escherichia coli* and *Salmonella typhimurium* into the VBNC state (Oliver et al., 2005). In this study, when PVP-I concentrations were lower than the effective sterilization concentration, the bacteria might not be completely killed and some of them went into the VBNC state. So the early decline in the number of bacteria might be caused by death cells and VBNC state cells together. As for the subsequent increase in the number of bacteria in outdoor aquaculture water, tap water and LB medium, whether it was associated with the resuscitation and growth of some mildly injured VBNC cells, further study was needed. But, there ~~were~~ ~~was~~ some evidences ~~showed~~ that VBNC bacteria could resuscitate and proliferate under certain conditions (Dukan et al., 1997).

This study confirmed the effect of organic matter on PVP-I sterilization and suggested that aquatic organic matter should be considered when PVP-I was used in aquaculture. The results showed the great bactericidal activities of the PVP-I in different aquaculture waters, and we recommended 25 ppm for outdoor aquaculture and 20 ppm for indoor treatment, which were ~~the~~ safe and effective ~~in the~~ ~~for~~ normal swamp eel farming.

## Conclusions

In conclusion, the organic matter in aquaculture water has negative influence on the bactericidal effectiveness of PVP-I. The minimum effective bactericidal concentration of PVP-I in indoor aquaculture water and outdoor aquaculture water were 10 ppm and 20 ppm, respectively. The results presented here would be helpful to suggest povidone iodine could help prevent the transmission of *A. hydrophila* in swamp eel aquaculture.

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