

A new megaspilid wasp from Eocene Baltic amber (Hymenoptera: Ceraphronoidea), with notes on two non-ceraphronoid families: Radiophronidae and Stigmaphronidae

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Ceraphronoids are some of the most commonly collected hymenopterans, yet they remain rare in the fossil record. *Conostigmus talamasi* Mikó and Trietsch, sp. nov. from Baltic amber represents an intermediate form between the type genus, *Megaspilus*, and one of the most species-rich megaspilid genera, *Conostigmus*. We describe the new species using 3D data collected with synchrotron-based micro-CT equipment. This non-invasive technique allows for quick data collection in unusually high resolution, revealing morphological traits that are otherwise obscured by the amber. In describing this new species, we revise the diagnostic characters for Ceraphronoidea and discuss possible reasons why minute wasps with a pterostigma are often misidentified as ceraphronoids. Based on the lack of ceraphronoid characteristics, we remove *Dendrocerus dubitatus* Brues 1937, Stigmaphronidae, and Radiophronidae from Ceraphronoidea and consider them as *incertae sedis*. We also provide some guidance for their future classification.

1 **A new megaspilid wasp from Eocene Baltic** 2 **amber (Hymenoptera: Ceraphronoidea),** 3 **with notes on two non-ceraphronoid** 4 **families: Radiophronidae and** 5 **Stigmaphronidae**

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18 **ABSTRACT**

19 Ceraphronoids are some of the most commonly collected hymenopterans, yet they remain rare in the
20 fossil record. *Conostigmus talamasi* Mikó and Trietsch, sp. nov. from Baltic amber represents an
21 intermediate form between the type genus, *Megaspilus*, and one of the most species-rich megaspilid
22 genera, *Conostigmus*. We describe the new species using 3D data collected with synchrotron-based
23 micro-CT equipment. This non-invasive technique allows for quick data collection in unusually high
24 resolution, revealing morphological traits that are otherwise obscured by the amber. In describing this
25 new species, we revise the diagnostic characters for Ceraphronoidea and discuss possible reasons
26 why minute wasps with a pterostigma are often misidentified as ceraphronoids. Based on the lack of
27 ceraphronoid characteristics, we remove *Dendrocerus dubitatus* Brues 1937, Stigmaphronidae, and
28 Radiophronidae from Ceraphronoidea and consider them as *incertae sedis*. We also provide some
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30 **INTRODUCTION**

31 Ceraphronoidea is a hymenopteran lineage with an enigmatic phylogenetic position and poorly
32 understood natural history. Their minute body size and parasitoid lifestyle, along with a few antennal and
33 fore wing characters, suggest a close relationship with Proctotrupomorpha (Ronquist et al., 1999; Engel
34 and Grimaldi, 2009). A myriad of other less-obvious morphological traits, however, including parts of
35 the metasomal, genital and mesosomal skeletomuscular systems (Vilhelmsen et al., 2010; Mikó et al.,
36 2013; Ernst et al., 2013), reveal many similarities to non-apocritan Hymenoptera. Even recent molecular
37 phylogenetic studies have failed to place the superfamily with confidence, although they support that the
38 superfamily, indeed, is not closely related to Proctotrupomorpha (Dowton et al., 1997; Heraty et al., 2011;
39 Mao et al., 2015; Klopstein et al., 2013; Peters et al., 2011, 2017; Branstetter et al., 2017). Irrespective of
40 their phylogenetic position, recent ceraphronoids comprise a morphologically well-characterized group
41 that can be readily separated from other hymenopterans based on the following traits.

1. Compact mesosoma In Ceraphronoidea, the pronotum, mesopectus, metapectus, first abdominal tergum, and the metanotum comprise a single, compact sclerite (Fig. 1A). This modification is only found in some wingless hymenopterans, as the presence of the conjunctivae that allows mobility between the above-mentioned mesosomal regions is less important in flightless hymenopterans (Reid, 1941; Keller et al., 2014). The functional consequences of mesosomal compactness in ceraphronoids have never been explored. Most ceraphronoids are able to fly; therefore, mesosomal compactness most likely evolved for a different reason than that of flightless hymenopterans with a similarly compact mesosoma.

2. Orientation of the toruli and antennal bases The ventral position of the antennal insertion sites (toruli) is a well-known characteristic of Ceraphronoidea (Masner, 1993) and it is shared by some other apocritan taxa (e.g. Megalyroidea and Platygastroidea; Sharkey et al. (2012)). The orientations of the torular surface and the antennal base are, however, often overlooked traits of the superfamily. In Ceraphronoidea, the lateral torular margin is elevated relative to the median margin and therefore the antennal foramina, and the antennal scapes are oriented medially in a resting position (Figs 1B, C).

3. Articulation between pronotum and mesoscutum In Ceraphronoidea, the pronotum and the mesoscutum have a unique, ball-and-socket type articulation that corresponds to sharp anterolateral edges on both the mesoscutum and pronotum (Fig. 1A) (Mikó and Deans, 2009). The notauli arise from these articulation (Figs. 2A, B) in Ceraphronoidea. While distinct anterolateral edges on the mesoscutum are present in Megalyridae, they never correspond with ball-and-socket articulations.

4. Wing venation All winged Ceraphronoidea have a stigmal vein that originates from the pterostigma or from the distal portion of the marginal vein posterior to the costal notch (Masner, 1993). Along with this, ceraphronoids have a single vein extending along the anterior margin of the fore wing. This vein is equipped with unique triangular elements (Figs 1D, E), whose functions and origins remain unknown. A single vein on the anterior wing margin is present in numerous other hymenopterans (e.g., aphidiine braconids) but the triangular elements are seemingly specific to Ceraphronoidea (IM, personal observation). In many Chrysidoidea the two wing veins on the anterior wing margin (costal and subcostal veins) are adjacent and may superficially look “fused”, but they are always separated by a faint line (Olmi, 1994; Richards, 1939).

The two type specimens of the present paper are embedded in an approximately 33–55 million-year-old (Gillung and Winterton, 2018; Larsson, 1978; Penney, 2010; Sadowski et al., 2017; Wolfe et al., 2016), somewhat cloudy piece of Baltic amber (Figs 3C, D 4 A–D). These specimens belong in Ceraphronoidea based on the the lyre-shaped notaulus and the orientation of the torulus and antenna (traits that are not obscured). As ceraphronoid experts, we examined the specimens with a light microscope at 230× magnification, and based on their general habitus (*Gestalt*) and the presence of a bifurcated anteromedian projection of the metanoto-propodeo-metapectal complex (Fig. 4A) we first classified them as *Megaspilus*, a genus that has never been recorded from any fossil deposit. However, after looking at the high resolution 3D data (see Supplementary 3D PDF of synchrotron-based micro-CT), we were able to observe otherwise-obscured traits and determine that the new species actually belongs in *Conostigmus*, as part of a species group that shares some key features with *Megaspilus*.

While small body size, reduced wing venation, and the presence of a pterostigma are often used to determine specimens as Ceraphronoidea, these characters occur in almost all hymenopteran superfamilies. Despite the above-listed clear and obvious ceraphronoid synapomorphies, it is common to find smaller Ichneumonoidea and Aculeata, especially Bethyridae and Dryinidae, misidentified as Ceraphronoidea in collections. Perhaps the most commonly misidentified are small specimens of the dryinid genus *Aphelopus* (Fig. 2C). One factor that may contribute to these misidentifications is the use of low-power microscopes for sorting and identification of specimens. Even the most distinct external traits specific to ceraphronoids are obscure with lower magnification and inadequate lighting. In such poor conditions, only those who have trained their eyes by looking at hundreds of ceraphronoid specimens are able to identify these taxa correctly. In some cases, the examination of fossil specimens is similar to studying specimens of recent taxa with a low quality microscope. Morphological traits are usually obscured by artifacts or debris, making them difficult to properly observe (if they can be seen at all).

The discovery of these *Conostigmus* specimens encouraged us to review fossil ceraphronoids and revise the classification of two hymenopteran families that are exclusively represented by fossils and currently classified in Ceraphronoidea. Radiophronidae and Stigmaphronidae are minute, winged wasps

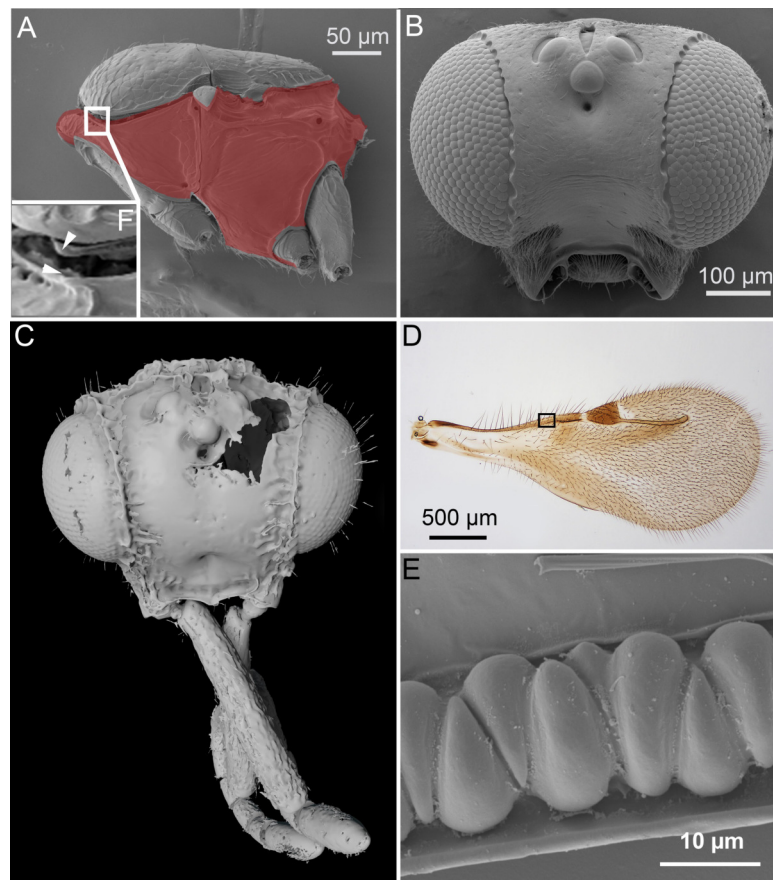


Figure 1. Unique ceraphronoid anatomical traits. (A) SEM micrograph of the mesosoma of *Aphanogmus*. The pronotum, mesopectus, metapectus, metanotum and first abdominal tergum, shown in red, comprise a single sclerite resulting in a compact mesosoma that can be found only in wingless Hymenoptera (e.g. Formicidae workers and Mutillidae females). The mesonotum and the pronotum have a unique ball-and-socket type articulation (smaller box (F) marked with arrowheads), a trait shared by Ceraphronoidea and Megalyroidea. The function of this articulation is unknown. (B) SEM micrograph of the head of *Ceraphron* (subgenus *Eulagynodes*) sp. showing the medially-oriented antennal sockets (toruli). (C) Surface-rendered 3D reconstruction of the head of *Conostigmus talamasi* Mikó and Trietsch nov. sp, bearing medially-oriented toruli and scapes. (D) Brightfield image of the fore wing of *Masner lubomirus* Deans and Mikó 2009 showing the typical ceraphronoid wing venation. There is a single vein along the anterior wing margin equipped with triangular elements and an unbroken stigmal vein that arises from the posterior third of the pterostigma (The region of Figure E is indicated by a small box). (E) SEM micrograph showing the triangular elements on the fore wing of *Conostigmus* sp. The function of these unique elements is unknown.

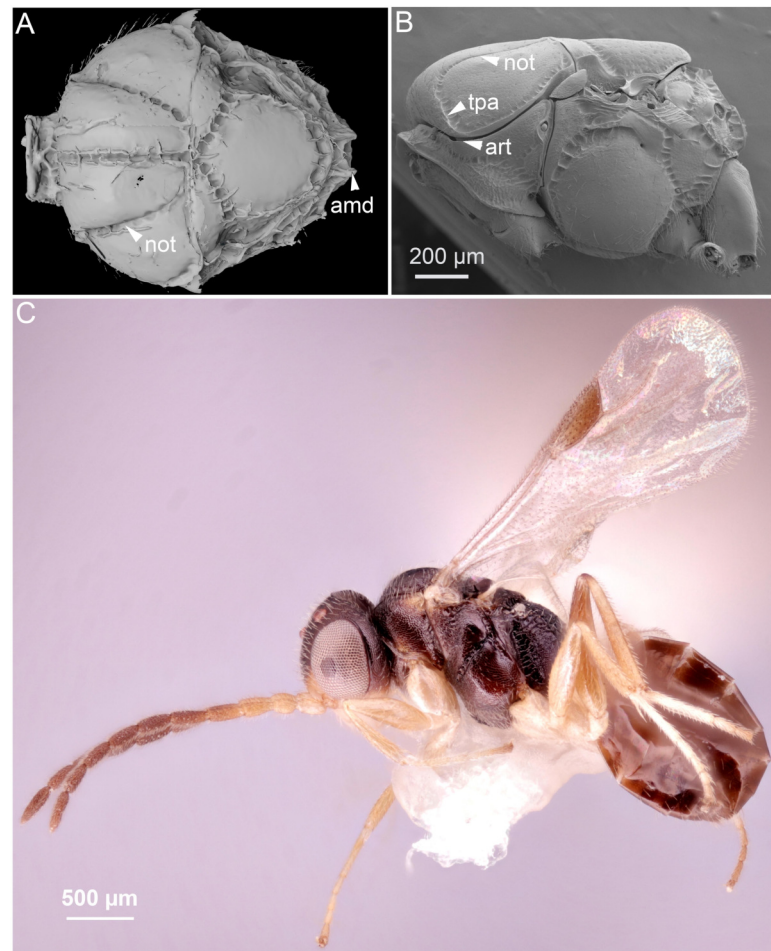


Figure 2. Comparison of ceraphronoid and chrysidoid morphology. (A) Surface rendered 3D reconstruction of the mesosoma of *Conostigmus talamasi* Mikó and Trietsch nov. sp. showing the lyre-shaped notauli (not) and the bifurcated anteromedian projection of the metapectal-propodeal complex (holotype, DEI-GISHym31819). (B) SEM micrograph of the mesosoma of *Trichosteresis glabra* (Boheman 1832). The anterolateral edge of the mesosoma corresponds to a ball-and-socket type articulation (art) between the mesonotum and the pronotum in Megalyroidea and Ceraphronoidea. The notaulus (not) is continuous with the anteromedian transverse sulcus (tpa) that arises from the pronoto-mesonotal articulation in Ceraphronoidea. (C) Brightfield image of the chrysidoid *Aphelopus* sp. These wasps, due to their minute body size, reduced wing venation, and large pterostigma, are often misidentified as Megaspilidae in collections (app=anteromedian projection).

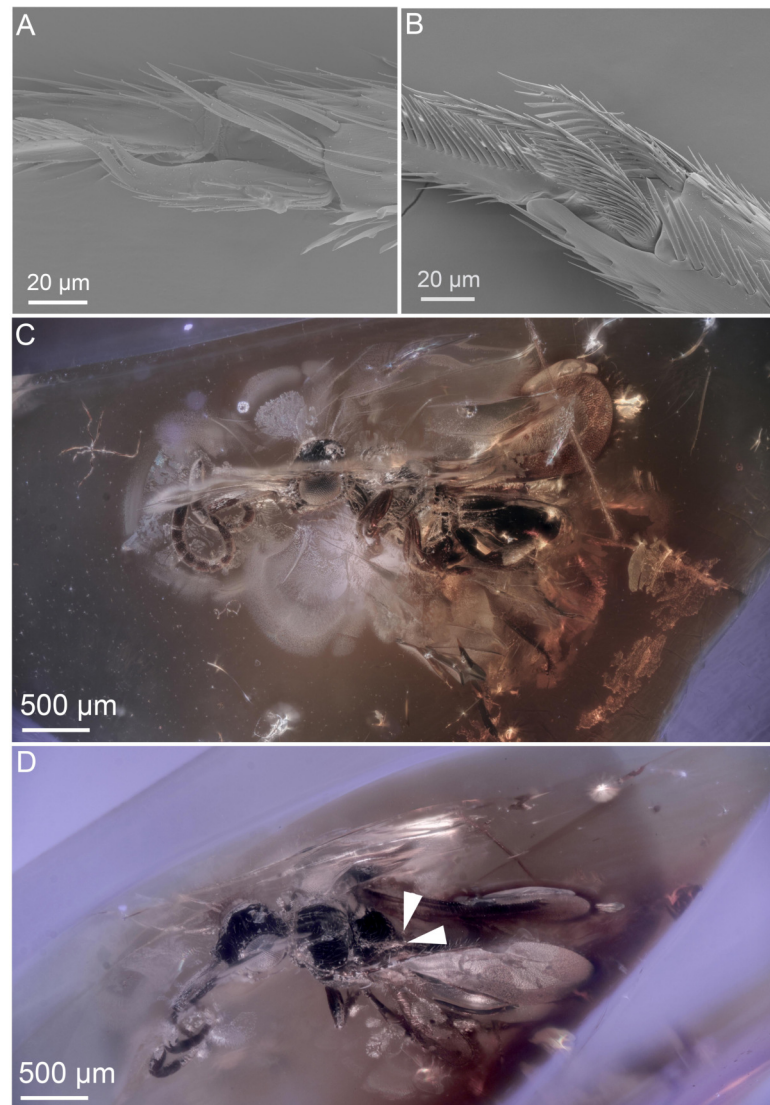


Figure 3. Ceraphronoid morphology in recent and fossilized specimens. (A) SEM micrograph showing the apical protibial spurs in Ceraphronidae. (B) SEM micrograph showing the apical protibial spurs in Megaspilidae. (C) Brightfield image showing the holotype of *Conostigmus talamasi* Mikó and Trietsch, lateral view (DEI-GISHym31819). (D) Brightfield image showing the holotype of *Conostigmus talamasi* Mikó and Trietsch, dorsolateral view, with arrows pointing to the bifurcate anteromedian projection of the metanoto-propodeo-metapectal complex (DEI-GISHym31819).



Figure 4. Brightfield images of the paratype (DEI-GISHym31820) of *Conostigmus talamasi* nov. sp. Mikó and Trietsch. A. Dorsal view, with an arrow pointing to the bifurcated anteromedian projection of the metanoto-propodeo-metapectal complex. B. Left lateral view. C. Ventral view. D. Right lateral view.

that have a distinct pterostigma and, in some cases, two protibial spurs. Besides these traits, however, these two families do not share any other characteristics with Ceraphronoidea. Here, we remove them from the superfamily, leaving them *incertae sedis*, and provide some guidance for their future classification.

MATERIALS AND METHODS

The two specimens for the present study were obtained from the private collection of Baltic amber inclusions of Christel and Hans Werner Hoffeins (Hamburg, Germany) who bought them from commercial source at Amberif in Gdansk. Specimens are embedded in Polyester resin (Voss-Chemie, Uetersen) (Hoffeins, 2001) and are deposited in the Deutsches Entomologisches Institut (Müncheberg, Germany) with the accession numbers: DEI-GISHym31819 (holotype), DEI-GISHym31820 (paratype).

Morphological traits were observed and described using volume and surface rendered 3D models (Figures 1C, 2A, 5, Supplementary 3D PDF). Measurements of anatomical lines (Supplementary Table 1) were performed using the 3D measurement tool in Amira (version 5.6, FEI) using the ASTOR virtual analysis infrastructure at Karlsruhe Institute of Technology (KIT) (Mexner et al., 2017).

Synchrotron X-ray tomography was performed at the UFO imaging station of the KARA Synchrotron Radiation Facility at KIT. In order to achieve high image resolution despite the comparatively large size of the specimen, the latter was scanned in three steps. For each scan, 3,000 equiangularly-spaced radiographic projections were acquired in a range of 180°. The frame rate was set to 70 frames per second, resulting in a scan duration of about 43 seconds. A parallel polychromatic X-ray beam was spectrally filtered by 0.2 mm Al to obtain a peak at about 15 keV. The detector consisted of a thin, plan-parallel lutetium aluminum garnet single crystal scintillator doped with cerium (LuAG:Ce), optically coupled via a Nikon Nikkor 85/1.4 photo-lens to a pco.dimax camera with a pixel matrix of 2008×2008 pixels. The magnification of the optical system was adjusted to 10×, yielding an effective X-ray pixel size of 1.22 μm (dos Santos Rolo et al., 2014). Tomographic reconstruction was performed with the GPU-accelerated filtered back projection algorithm implemented in the software framework UFO (Vogelgesang et al., 2012). The three tomographic volumes were registered and merged with Amira (version 5.6, FEI) using the ASTOR virtual analysis infrastructure at KIT (Mexner et al., 2017).

3D reconstruction followed the protocol described by Ruthensteiner and Heß (2008) and van de Kamp et al. (2014), using Amira for segmentation of every 20th slice of the tomographic volume. Automated interpolation between the labels was performed using the online image segmentation tool Biomedisa (<https://biomedisa.de/>) (Lösel and Heuveline, 2016). CINEMA 4D R18 (Maxon Computer GmbH) was employed for assembly of components, smoothing and polygon reduction. Subsequently, it was imported into Deep Exploration (version 6; Right Hemisphere), saved as Universal 3D file (U3D) and embedded into a PDF document with Adobe® Acrobat® 9 Pro Extended.

Brightfield images of fossil specimens were taken with an Olympus BX43 compound microscope equipped with an Olympus DP73 digital camera. Image stacking was performed with Zerene Stacker (Version 1.04 Build T201404082055; Zerene Systems LLC, Richland, WA, USA). Extended focus images were annotated and modified with Adobe Photoshop 6™ (Adobe Systems, San Jose, CA, USA) using the Adjust/Filter/Unsharp mask and Image/Adjustments/Exposure (Gamma correction) tools.

Taxonomic treatment including natural language (NL) phenotype representations were compiled in mx (<http://purl.org/NET/mx-database>). Terminology of the phenotype statements used in descriptions, are mapped to the Hymenoptera Anatomy Ontology (HAO, available at <http://purl.obolibrary.org/obo/hao.owl>), Phenotypic Quality Ontology (PATO, available at <http://purl.obolibrary.org/obo/pato.owl>), Biospatial Ontology (BSPO, available at <http://purl.obolibrary.org/obo/bspo.owl>) and Common Anatomy Reference Ontology (CARO, available at <http://obofoundry.org/>). Wing venation terminology follows Mikó et al. (2014).

Natural language phenotype representations are in “Entity attribute: value” format. Semantic statements written in OWL Manchester syntax (<http://www.w3.org/TR/owl2-manchester-syntax/>) were generated in Protégé 5.0.0-beta-16 (<http://protege.stanford.edu/>) following Balhoff et al. (2013); Mikó et al. (2014). The OWL (<http://www.w3.org/TR/owl2-overview/>; accessed February 4, 2014) representation of the full data set is stored as a single Resource Description Framework (RDF)-XML file (<http://www.w3.org/TR/REC-rdf-syntax/>; accessed 12 March 2017) in the Github repository (<https://github.com/hymao/hymao-data>).

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new names contained in the electronic version are effectively published under that Code from the electronic edition alone. This published work and the nomenclatural acts it contains have been registered in ZooBank, the online registration system for the ICZN. The ZooBank LSIDs (Life Science Identifiers) can be resolved and the associated information viewed through any standard web browser by appending the LSID to the prefix <http://zoobank.org/>. The LSID for this publication is: [LSID urn:lsid:zoobank.org:pub:0B233959-77FE-46F1-AB82-15C7F816D0BA].

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RESULTS

Conostigmus

The new species belongs in *Conostigmus* based on the presence of a distinct sternaulus (ste: Fig. 5 A), a shorter posterior ocellar line (POL) than ocular-ocellar line (OOL) (Figs 1C, 5B, C) and the presence of a preoccipital carina (Figs 1C, 5A, B, C). These traits are absent from *Dendrocerus*, *Trichosteresis*, and *Platyceraphron*; clavate female antenna and F1 length (proximodistal anatomical line) shorter than the combined length of F2 and F3 are traits that are present in *Conostigmus* and absent from *Megaspilus*.

Conostigmus talamasi Mikó and Trietsch sp. nov.

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Figures 1C, 3B, C, 4, 5

Diagnosis The new species differs from other *Conostigmus* species in the presence of a bifurcated anteromedian projection of the metanoto-propodeo-metaplecto-mesopectal complex (app: Fig. 5C). The anteromedian carina of the syntergite (acs: Fig. 5C) is inserted into the concavity between the projections. The anteromedian projection of the metanoto-propodeo-metaplecto-mesopectal complex in *Conostigmus talamasi* is distinct, but shorter than that of *Megaspilus* species.

Description (Supplementary Table 1)

Body length universal: 2007.29 μ m, 2411.09 μ m.

Head: Head width vs. head height: HW:HH= 1.183, 1.192. Head width vs. interorbital space (HW/IOS) Female: 1.861, 2.011. Dorsal carina of occipital depression count: present. Dorsal carina of occipital depression medial continuity: discontinuous medially. Occipital carina sculpture: crenulate. Median flange of occipital carina count: absent. Submedial flange of occipital carina count: absent. Dorsal margin of occipital carina vs. dorsal margin of lateral ocellus in lateral view: occipital carina is ventral to lateral ocellus in lateral view. Preoccipital lunula count: present. Preoccipital ridge count: present. Preoccipital furrow count: present. Preoccipital furrow anterior extension: adjacent anteriorly to the posterior margin of the median ocellus. Preoccipital furrow anterior region vs posterior region sculpture: posterior region crenulate, anterior region smooth or finely reticulate. Preoccipital furrow anterior region width vs posterior region width: as wide anteriorly as posteriorly. Preoccipital carina count: present. Preoccipital carina shape: interrupted dorsally and represented by irregular, not continuous carinae. Preoccipital carina and occipital carina structure: occipital carina complete, preoccipital carina fused laterally with preorbital carina. Female OOL: POL: LOL: 1.1:1.2:1, 1.2:1.7:1. Postocellar carina count: absent. Preocellar pit count: present. Randomly sized areolae around setal pits on upper face count: absent. Antennal scrobe count: absent. Transverse striation on upper face count: present. Transverse scutes on upper face count: absent. Transverse frontal carina count: absent. Frontal ledge count: absent. Rugose region on upper face count: present. Anterior ocellar fovea shape: fovea not extended ventrally into facial sulcus. Facial pit count: facial pit present. White, thick setae on upper face count: absent. Ventromedian setiferous patch and ventrolateral setiferous patch count: absent. Supraclypeal depression count: absent. Intertorular carina count: present. Median process on intertorular carina count: present, extending from intertorular carina towards dorsal margin of clypeus. Median process of intertorular carina structure: process extends across intertorular area to dorsal margin of clypeus. Intertorular ridge vs. epistomal ridge: fused medially. Intertorular area count: present. Median region of intertorular area shape: convex. Torulus position relative to anterior ocellus and distal margin of clypeus: torulus not reaching epistomal sulcus, closer to distal margin of clypeus than anterior ocellus. Torulo-clypeal carina count: absent. Subtorular carina count: absent. Subantennal groove count: absent. Posterolateral process of gena count: absent. Ocular impression and post ocular orbital carina count: present. Ocular impression

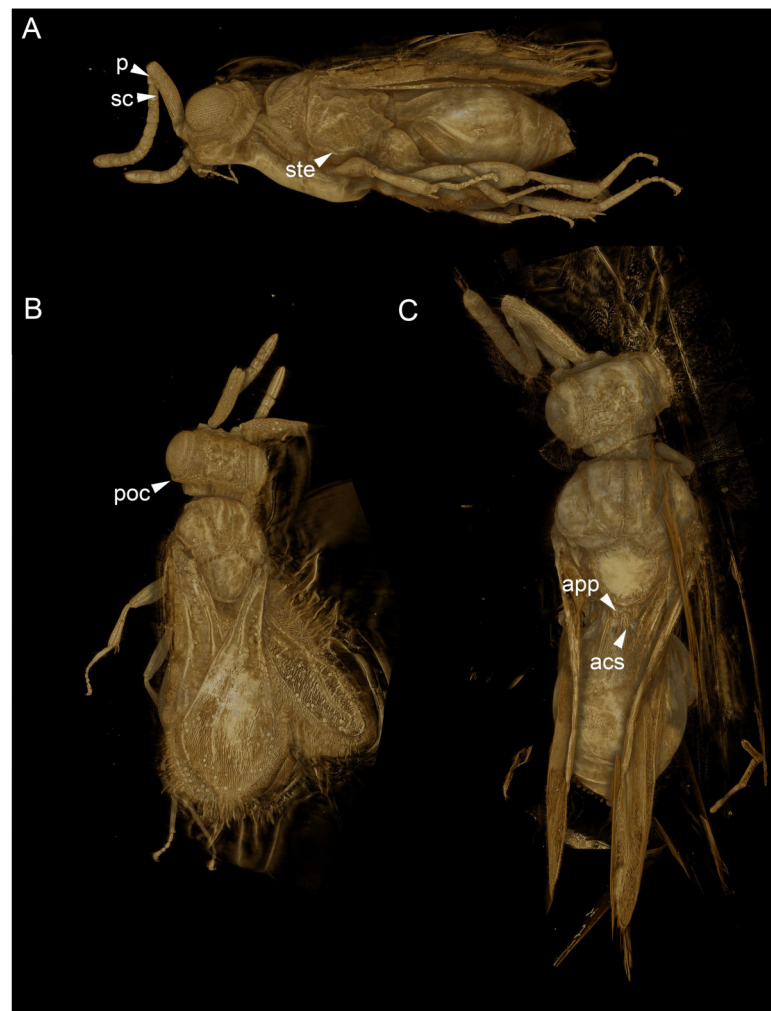


Figure 5. Volume rendered 3D micrographs showing *Conostigmus talamasi* sp. nov. Mikó and Trietsch. A. Paratype, lateral view. B. Paratype, dorsal view. C. Holotype, dorsal view. Abbreviations: acs=anteromedian carina of the syntergite, app=anteromedian projection of the metanoto-propodeo-metapecto-mesopectal complex, p=pedicel, poc=preoccipital carina, sc=scape, ste=ssternaulus

(DEI-GISHym31820).

202 sculpture: scalloped (foveae composing ocellar impression adjacent, sometimes not separated from each
203 other). Mandibular tooth count: 2. Mandibular lancea count: absent. Maxillary palpomeres count: 5–6.

204 **Antennae:** Flagellar scrobe of the scape count: present. F1 length vs F2+F3 length: F1 shorter than
205 F2+F3. F1 length / F2 length: 1.59, 1.55. F6 length / F7 length+F8 length: 0.59, 0.47. Scape length / F1
206 length+F2 length: 2.1, 2.25. Scape length / F2 length: 5.4, 5.74. Female scape length vs. pedicel length:
207 3.8,4.2. Female F1 length vs. pedicel length: F1 as long as pedicel (1.0-1.1). Female ninth flagellomere
208 length: F9 longer than F7+8.

209 **Mesosoma:** Foveolate sculpture on body count: absent. Mesosoma shape: not compressed laterally,
210 as wide as high or wider than high. Pronope count: present. Transverse pronotal sulcus (anterodorsal
211 branch of pronotal y) count: present. Epomial carina count:absent. Posterodorsal branch of pronotal Y
212 count: present. Ventrolateral invagination of the pronotum count: present. Annular pronotum count:
213 present. Ventromedian region of pronotum and anteroventral region of mesopectus continuity: pronotum
214 and mesopectus continuous ventromedially. Lateroventral invagination of the propleuron count: absent.
215 Mesonotal fossa of the pronotum and pronotal condyle of the mesonotum count: present. Mesonotum
216 anterolateral margin shape: square. Median mesoscutal sulcus count: present. Median mesoscutal sulcus
217 posterior end location: adjacent to transscutal articulation. Scutoscutellar sulcus vs. transscutal articulation
218 location: adjacent. Notaulus count: present. Notaulus posterior end location: adjacent to transscutal
219 articulation. Posterior end of notaulus vs. posterior end of antero-admedian line location: notaulus
220 extends more posteriorly than antero-admedian line. Transscutal articulation completeness: complete.
221 Lateral carina on the mesoscutellum count: absent. Axillular carina count: present. Axillular carina
222 shape: left and right carina continuous posteromedially forming a U-shape carina on the mesoscutellar
223 axillar complex. Axillular setae count: absent. Posterolateral margin of mesoscutellum shape: blunt.
224 Posteromedian process of the mesoscutellum count: absent. Anteromedian projection of the metanoto-
225 propodeo-metapecto-mesopectal complex count: present. Anteromedian projection of the metathorax-
226 propodeum complex shape: bilobed. Anteromedian projection of the metathorax-propodeum complex
227 curvature lateral in view: straight. Sternaulus count: present. Sternaulus length: elongate, exceeding
228 3/4 of mesopleuron length at level of sternaulus. Longitudinal striae extending from crenulae of anterior
229 mesopleural sulcus to mesopleural pit count: absent. Speculum ventral limit: not extending ventrally
230 of pleural pit line. Mesometapleural sulcus count: present. Ventral invagination of mesometapleural
231 sulcus presence: absent. Epicnemial carina count: complete. Epicnemial pit count: absent. Epicnemium
232 posterior margin shape: anterior discriminal pit present; epicnemial carina curved. Mesodiscrimen count:
233 present. Anterior metapleural carina count: absent. Metapleural carina count: present. Metapleural carina
234 vs. propodeal spiracle: metapleural carina extending ventrally of propodeal spiracle. Ventral projection
235 of the metapleural carina count: present. Ventral invagination of the metapleural carina count: absent.
236 Propodeal spiracle dilator muscle apodeme pit location: On metapleural carina. Lateral propodeal carina
237 count: present. Lateral propodeal carina shape: inverted “U” (left and right lateral propodeal carina
238 are adjacent to the antecostal sulcus of the first abdominal tergum submedially). Median propodeal
239 carina count: absent. Posterior propodeal projection count: present. Posterior propodeal projection
240 shape: simple. Propodeal and metacoxal verrucules count: absent. Posterodorsal metapleural area shape:
241 trapezoid. Posterior line of the posterodorsal metapleural area count: present. Transverse line of the
242 metanotum-propodeum vs. antecostal sulcus of the first abdominal tergum: adjacent sublaterally. Carina
243 limiting posteriorly antecosta count: present. Metapecto-propodeal conjunctiva count: present. Posterior
244 margin of nucha in dorsal view shape: straight.

245 **Wings:** Stigmal vein of fore wing count: present. Pterostigma of fore wing count: present. Hind wing
246 reduction: well developed.

247 **Legs:** Calcar shape: bifid. Mesotibial spur count: 2. Mesobasicoxa width vs. metabasicoxa width:
248 metabasicoxa distinctly wider than mesobasicoxa. Posterior mesosomal comb count: absent.

249 **Metasoma:** S1 length vs. shortest width: S1 wider than long. Transverse carina of petiole count:
250 present. Transverse carina on petiole shape: straight. Basal, longitudinal carinae on syntergum count:
251 more than 5. Transverse sulcus of first metasomal sternum count (S1 count): present. Waterston’s
252 evaporatorium count: absent.

253 Locality of type specimens

254 Gulf of Gdańsk (Baltic amber)

Etymology

The new species is named after Elijah Talamas (Florida State Collection of Arthropods), who drew our attention to these unique fossils.

DISCUSSION

Megaspilus vs. *Conostigmus*

Based on the latest phylogenetic analysis (Mikó et al., 2013), and preliminary phylogenomic data from an ongoing molecular study using ultra-conserved elements (UCEs) (Bonnie Blaimer, pers. comm.) *Conostigmus* is polyphyletic and includes *Megaspilus*. Until Dessart's revisions (1972; 1981) of Nearctic and Palaearctic species, *Megaspilus* was a broad taxonomic concept that essentially included all larger megaspiline species with an acute ocellar triangle (*Conostigmus*-type, in contrast to a *Dendrocerus*-type obtuse ocellar triangle), well-defined sternaulus, distinct posterior orbital carina and/or some rugulose sculpture on the frons. Dessart (1972) narrowed the generic concept of *Megaspilus* to include only those species that have an acute ocellar triangle, sternaulus, bifurcated anteromedian projection, and elongate female first flagellomere. The ocellar triangle shape and presence of the sternaulus are shared with numerous *Conostigmus* species, leaving the bifurcated anteromedian projection and the elongate female proximal flagellomeres as diagnostic features for *Megaspilus*.

The bifurcated anteromedian projection of *Megaspilus* is clearly derived from the elevated and medially-projected lateral propodeal carinae. This bifurcated condition can be found in certain *Ceraphron* and *Dendrocerus* (Alekseev, 1978) species, and so far has never been reported from any *Conostigmus* (Mikó et al. 2016; Dessart 1997; CT personal communication). The median portion of the lateral propodeal carina is elevated and forms a bifurcated projection in *Conostigmus talamasi* that is smaller than that of *Megaspilus* and most likely represents an intermediate state.

With the discovery of *Conostigmus talamasi*, the only diagnostic characters for separating *Megaspilus* from *Conostigmus* remain the elongate proximal female flagellomeres. The first female flagellomere is more than 2 times as long as the pedicel in *Megaspilus*, while in other megaspilids, including *Conostigmus*, it is less than 1.5 times as long as the pedicel. The length of the male and female flagellomeres correlate to each other and are important in species-level diagnosis in Megaspilinae (Mikó et al., 2016). The correlation of flagellomere length between different sexes might be related to their courtship behavior as males extensively antennate and repeatedly touch the female antenna during mating (Liebscher, 1972). Consequently, the first male flagellomere is much longer than the scape in *Megaspilus* in contrast with other male megaspilids where the first flagellomere is either shorter or slightly longer than the scape (Mikó et al., 2016; Dessart, 1972, 1981, 1974, 1995, 1997, 1999, 2001).

Ceraphronoid and non-ceraphronoid fossils

To date, there are 18 fossil specimens that share key characteristics with recent Ceraphronoidea (see Supplementary Figure 1), including the compact mesosoma, lyre-shaped notaulus, and the orientation of the antennal bases. Based on their visible morphological characters, these fossils represent (mostly) megaspilid wasps from the Late Cretaceous Santonian to the Early Miocene. There is also a single ceraphronid specimen from the Early Miocene (Supplementary Figure 1).

Wing venation characters are perhaps the most distinctive features shared between these fossils and recent ceraphronoids. In these taxa, a single wing vein is present along the anterior fore wing margin and the stigmal vein is never angled (an angle is present in numerous other taxa in the stigmal vein, marking its intersection with 2RS or r-m veins Mikó et al. (2014)), never tangential to the pterostigma and arising from or anterior to the midpoint of the pterostigma (Fig. 1D, Masner 1993). A single fore wing vein can be found in numerous hymenopteran families, but unlike in these fossils, it is always well separated from the anterior margin.

The remaining fossil hymenopterans currently classified as ceraphronoids (Supplementary Figure 1) lack key ceraphronoid characteristics listed in the introduction. Although they do each possess a pterostigma, the remaining wing venation traits are inconsistent with the superfamily. These taxa might not even be closely related to Ceraphronoidea.

Dendrocerus dubitatus (Brues 1937)

Dendrocerus dubitatus (Brues 1937) was the first described putative fossil ceraphronoid wasp. Brues (1937) explanation for his placement of this species is the following: "This species is undoubtedly very

similar to the large modern genus *Lygocerus*.” Most of the body of the holotype specimen is obscured, however, and it is difficult to understand how Brues was able to prepare a rather detailed description as it was outlined by McKellar and Engel (2011).

However, the wing venation on both fore wings are visible (Supplementary Figure 2A.), and they are cardinally different from that of recent ceraphronoids; the straight stigmal vein arises from the anterior portion of and is tangential with the pterostigma (Supplementary Figure 2A.). The antenna of *D. dubitatus* is composed of only 10 flagellomeres (Supplementary Figure 2B; Brues 1937), a character state that does not occur in any megaspilid taxa. While Ceraphronidae females often have 10 flagellomeres, the vast majority lack the pterostigma (*Trassedia* females have 11 flagellomeres, Cancemi 1996; Mikó et al. 2013; *Masner* is only known from male specimens, Mikó and Deans 2009). From these characters, it is clear that this species is not a ceraphronoid wasp. We consider it as *incertae sedis* and note that it has a wing venation often found in Chrysidoidea, as well as in some Stigmaphronidae.

Stigmaphronidae

Another prospective ceraphronoid fossil, *Allocotidus* Muesebeck 1963, was described and classified with the following explanation (Muesebeck, 1963, pg. 129): “... the specimen ... is incomplete and otherwise in rather poor condition. Enough can be clearly made out, however, to place it in the proctotrupoid family Ceraphronidae.” (Note that at the time of this description Megaspilidae and Ceraphronidae comprised a single family, Ceraphronidae.) This taxon shares only one characteristic with ceraphronoids, the “fused” SC+R vein, although there is a faint line distinctly separating two wing veins at the anterior margin (a characteristic trait of numerous chrysidoids). Kozlov (1975) placed *Allocotidus* into his new family Stigmaphronidae, together with three new genera, *Stigmaphron* Kozlov, *Elasmomorpha* Kozlov and *Hippocoon* Kozlov, and provided a diagnosis largely based on traits shared with the Elasminae (flattened hind coxa, elongate tibial spurs, large mesoscutellar axillae complex, shortened metasoma). For reasons that remain unclear Kozlov (1975), and subsequent authors of stigmaphronid taxa (Engel and Grimaldi, 2009; Ortega-Blanco et al., 2011; McKellar and Engel, 2011) considered these shared traits homoplasious, and classified Stigmaphronidae into Ceraphronoidea.

Stigmaphronidae show polymorphisms both in the number of protibial spurs and in wing venation characters. In some species, the two anterior fore wing veins are not adjacent to each other, the stigmal vein is either broken or arched or straight, and it arises anterior or in the middle of the pterostigma. The wing venation, however, never truly exhibits the characteristics of Ceraphronoidea. None of the stigmaphronid species shares any characteristics with recent ceraphronoids, except that some specimens have two protibial spurs. The presence of the two protibial spurs (Figs 3A, B) has been long considered a plesiomorphic character state in Ceraphronoidea, despite evidence supporting the evolutionary plasticity of the number of tibial spurs in Apocrita (Basibuyuk and Quicke, 1995; Kaartinen and Quicke, 2007; Engel and Grimaldi, 2009). For example, even within Ceraphronoidea the mesotibial spurs are variable between Megaspilidae and Ceraphronidae.

The protibial spurs are particularly important in Hymenoptera systematics, as the anterior spur has evolved into an antenna cleaning device. This trait is an important synapomorphy for Hymenoptera (Sharkey et al., 2012; Basibuyuk and Quicke, 1995; Vilhelmsen et al., 2010). The posterior spur has been reported as well developed, reduced, or absent in non-apocritans and is usually absent from apocritan taxa. There are known exceptions for two putatively unrelated braconid genera (*Rhamnura* and *Bathyaulax* Basibuyuk and Quicke 1995; Kaartinen and Quicke 2007) and recent Ceraphronoidea. This spur is difficult to differentiate from other apical, often unicellular protibial spines and trichoid sensilla (results of the evagination of the membrane of a single epidermal cell) in smaller specimens, even in recent taxa.

It is difficult for us to provide any guidance on how to reclassify stigmaphronids as, given the great polymorphism in tibial spurs, wing venation, and metasomal morphology (Ortega-Blanco et al., 2011), this taxon is likely polyphyletic. Based on the wing venation of Cretaceous chrysidoids, it is possible that stigmaphronids belong in Aculeata. A cenchrus-like area on the metanotum of one species (Engel and Grimaldi, 2009) suggests that at least this stigmaphronid might be closely related to some non-apocritan lineages. We consider Stigmaphronidae *incertae sedis*.

Radiophronidae

As with Stigmaphronidae, the authors of Radiophronidae failed to provide a robust explanation for why they classified this family into Ceraphronoidea. The only character state this taxon might share with Ceraphronoidea, besides the small body size, is the presence of two protibial spurs (Ortega-Blanco

et al., 2010). These spurs, however, are difficult to observe in known specimens (Figs 1C1, 4C in Ortega-Blanco et al., 2010). Radiophronidae also lacks the most important ceraphronoid wing characteristics; they have two wing veins along the proximo-anterior margin of the fore wing instead of one, and the shape of the pterostigma is more elongate than in Ceraphronoidea. If the authors interpreted the fossil correctly, the pronotum of *Microstaphron* is visible in dorsal view and extends posteriorly, while the mesonotum is reduced. Similar modifications can be found in some *Ecnomothorax* (Megaspilinae), *Ecitonetes* (Ceraphronidae), and *Lagynodes* (Lagynodinae) species, but the enlarged pronotum and reduced mesonotum always corresponds to the reduction or absence of wings (Brues, 1902; Dessart and Masner, 1965; Dessart, 1966). Radiophronidae have well-developed wings. On the other hand, the pronotum is usually visible dorsally in Chrysidoidea, e.g. Bethyidae, which have similar wing venation to that of Radiophronidae (Richards, 1939).

Ceraphronoid male genitalia is unique among Apocrita in that they have an independent, moveable apical sclerite, the harpe Mikó et al. (2013). The harpe is absent from the gonostyle-volsella complex of Radiophronidae (Ortega-Blanco et al., 2010). This condition—the absence of a harpe—can only be found in three distantly related recent ceraphronoid species: *Trichosteresis glabra*, *Aetholagynodes stupendus* and *Dendrocercus wollastoni* (Mikó et al., 2013).

Based on the wing venation, mesosomal, and male genitalia morphology, Radiophronidae most likely represents another unique lineage of Cretaceous chrysoid wasps and should be considered *incertae sedis*.

Is Ceraphronoidea the most structurally diverse hymenopteran superfamily?

Superfamilies, among the highest taxonomic ranks (family-level) that are regulated by the ICZN (1999), serve as important taxa for communicating about Hymenoptera evolution (see Sharkey et al., 2012). They represent the highest functional and pragmatic taxa, defined, in part, by their natural history and a set of distinct morphological characteristics. It is critical, therefore, that they remain monophyletic and free from extraneous, unrelated taxa.

Recent and fossil Ceraphronidae and Megaspilidae, including *Conostigmus talamasi*, comprise a monophyletic group that can be clearly defined using numerous apomorphic traits (Masner, 1993). The inclusion of Stigmaphronidae, Radiophronidae and, until recently, the stephanoid Aptenoperissidae and the trigonaloid Maimetshidae (Zhang et al., 2018; Perrichot et al., 2011) within Ceraphronoidea, however, results in a polyphyletic morass that cannot be confidently diagnosed. The resulting taxon would have a variable number of protibial spurs and flagellomeres, the presence and absence of a wasp waist, tarsal plantulae, and cenchri, and either a compact or unabridged mesosoma—i.e., traits that are characteristic of other, well defined superfamilies. Ceraphronoidea has clearly been treated as a waste bin for minute fossil taxa with a pterostigma. Based on the characteristics of those fossils we remove these taxa from Ceraphronoidea.

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SUPPLEMENTARY INFORMATION

Supplementary Figure 1. Timeline of fossils that have been classified as Ceraphronoidea. Names above the timeline are now considered as Ceraphronoidea, while names below are taxa removed from Ceraphronoidea and considered as *incertae sedis* (R=Radiophronidae, S=Stigmaphronidae, M=Megaspilidae, Ce=Ceraphronidae; references cited on the Figure: Rasnitsyn (1991); Engel and Grimaldi (2009); Ortega-Blanco et al. (2011, 2010); Muesebeck (1963); Kozlov (1975); McKellar and Engel (2011); Brues (1937); Alekseev and Rasnitsyn (1981); Alekseev (1995); Brues (1940); Szabó and Oehlke (1986); Dessart (1977); Engel (2013); Peñalver and Engel (2006) **Supplementary Figure 2.** Brightfield images of the holotype

of *Dendrocerus dubitatus* Brues 1937. This species is considered now as *incertae sedis*. **Supplementary 3D PDF.** 3D PDF of *Conostigmus talamasi* sp. nov. holotype (DEI-GISHym31819). **Supplementary Table 1.** Measurements of anatomical lines.

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