

Changes in nutrients and decay rate of *Ginkgo biloba* leaf litter exposed to elevated O₃ concentration in urban area

Wei Fu^{1,2}, Xingyuan He¹, Sheng Xu¹, Wei Chen¹, Yan Li¹, Bo Li¹, Lili Su¹ and Qin Ping¹

¹ Key Laboratory of Forest Ecology and Management, Institute of Applied Ecology, CAS, Shenyang, China

² University of Chinese Academy of Sciences, Beijing, China

ABSTRACT

Ground-level ozone (O₃) pollution has been widely concerned in the world, particularly in the cities of Asia, including China. Elevated O₃ concentrations have potentially influenced growth and nutrient cycling of trees in urban forest. The decomposition characteristics of urban tree litters under O₃ exposure are still poorly known. *Ginkgo biloba* is commonly planted in the cities of northern China and is one of the main tree species in the urban forest of Shenyang, where concentrations of ground-level O₃ are very high in summer. Here, we hypothesized that O₃ exposure at high concentrations would alter the decomposition rate of urban tree litter. In open-top chambers (OTCs), 5-year-old *G. biloba* saplings were planted to investigate the impact of elevated O₃ concentration (120 ppb) on changes in nutrient contents and decomposition rate of leaf litters. The results showed that elevated O₃ concentration significantly increased K content (6.31 ± 0.29 vs 17.93 ± 0.40 , $P < 0.01$) in leaves of *G. biloba*, significantly decreased the contents of total phenols (2.82 ± 0.93 vs 1.60 ± 0.44 , $P < 0.05$) and soluble sugars (86.51 ± 19.57 vs 53.76 ± 2.40 , $P < 0.05$), but did not significantly alter the contents of C, N, P, lignin and condensed tannins, compared with that in ambient air. Furthermore, percent mass remaining in litterbags after 150 days under ambient air and elevated O₃ concentration was 56.0% and 52.8%, respectively. No significant difference between treatments was observed in mass remaining at any sampling date during decomposition. The losses of the nutrients in leaf litters of *G. biloba* showed significant seasonal differences regardless of O₃ treatment. However, we found that elevated O₃ concentration slowed down the leaf litter decomposition only at the early decomposition stage, but slightly accelerated the litter decomposition at the late stage (after 120 days). This study provides our understanding of the ecological processes regulating biogeochemical cycles from deciduous tree species in high-O₃ urban area.

Submitted 16 November 2017

Accepted 14 February 2018

Published 5 March 2018

Corresponding author

Sheng Xu, xusheng@iae.ac.cn,
shengxu703@126.com

Academic editor

Paolo Giordani

Additional Information and
Declarations can be found on
page 12

DOI 10.7717/peerj.4453

© Copyright

2018 Fu et al.

Distributed under
Creative Commons CC-BY 4.0

OPEN ACCESS

Subjects Ecology, Plant Science, Climate Change Biology, Ecotoxicology, Environmental Impacts

Keywords *Ginkgo biloba*, Litter decomposition, Elevated O₃ concentration, Litter quality

INTRODUCTION

In recent decades, due to the large increases in the emission of O₃ precursors including NO_x and VOCs around the world, the ground-level O₃ concentrations are constantly increasing, particularly in Asia (Sitch et al., 2007; IPCC, 2013). It is estimated that the O₃ concentration in the troposphere will increase by 40% to 60% by 2100 (Akimoto et al.,

2015). Among air pollutants, O₃ has the most widespread negative impact on terrestrial vegetation. In particular, it may potentially influence biogeochemical cycles of forest ecosystems (Nikolova et al., 2010; Calatayud et al., 2011; Sicard et al., 2016).

Effects of O₃ on forest ecosystems productivity and feedbacks have been widely investigated worldwide (Chappelka & Samuelson, 1998; Paoletti, 2006; De Bauer & Hernández-Tejeda, 2007) and recently were reviewed (Wang et al., 2016). In contrast, O₃ effects on litter decomposition are much less known (Nikula, Vapaavuori & Manninen, 2010). Indirect evidence comes from litter photodegradation studies in semi-arid and arid ecosystems (Austin & Vivanco, 2006). Among the few studies carried out in natural forest ecosystems, Parsons, Bockheim & Lindroth (2008) showed that, over a 23-month observation period on leaf litterbags of aspen and birch reciprocally transplanted to separate the effect of substrate quality from environment effects, increasing O₃ concentration by fumigation slowed down both aspen and birch litter decay rate, exacerbating the effects of elevated CO₂ concentration, but accelerated birch litter decay under ambient CO₂. A negative effect of O₃ fumigation on litter decay rate was also observed for holm oak leaf litter in Mediterranean forest (Baldantoni, Fagnano & Alfani, 2011). Such observations were explained by CO₂- and O₃-mediated changes in litter chemistry, particularly carbohydrates, nitrogen, and tannins. O₃ effects on litter decomposition in urban forests have not yet been explored. Filling such gap is particularly important, as in urban ecosystems, where tropospheric O₃ concentration can be very high due to photochemical air pollution; urban trees play a fundamental role in mitigating air pollution (Manes et al., 2012). Their leaf litter, if decaying faster when exposed to high O₃ concentration, would improve soil chemical properties and promote nutrient cycles, therefore affecting the sustainable development of urban areas (Nikula, Vapaavuori & Manninen, 2010; Xu et al., 2012).

Ginkgo biloba is commonly planted in the cities of northern China and it is one of the main tree species in the urban forest of Shenyang, Liaoning Province, China. According to our recent observations, the highest O₃ concentration at ground level is frequently over 40 ppb or even up to more than 80 ppb during the summer in the urban area of Shenyang city (Xu et al., 2015). For many years, we assessed the effects of elevated O₃ concentration on the eco-physiology of urban trees including *G. biloba* (He et al., 2009; Lu et al., 2009; Li et al., 2011; Xu et al., 2015). Here we aim to complement such previous studies with a manipulative experiment testing O₃ effects on *G. biloba* leaf litter decomposition and chemical features. Based on the results of many previous studies, we hypothesized that O₃ exposure at high concentrations commonly experienced by urban trees, could alter chemical composition and decrease the decomposition rate of this tree leaf litter. In this study, we predicted that elevated O₃ concentration would change the chemical compositions of leaves and decomposition rate of leaf litter. Therefore, the main objectives of this study are (1) to assess the changes of leaf litter quality of *G. biloba* fumigated by elevated O₃ concentration, including the changes in the contents of some nutrients and secondary metabolites, and (2) to evaluate the decomposition rates of leaf litter from this gymnosperm tree species exposed to high O₃ concentration.

MATERIALS & METHODS

Study site and experimental treatments

The study was conducted in the Shenyang Arboretum of the Chinese Academy of Sciences (41°46'N, 123°26'E) located in an urban area (He *et al.*, 2009). The arboretum with a mean elevation of 41 m and an area of 5 ha was founded in 1955, mainly planted with native tree species. There are more than 300 tree species and the forest coverage rate was 53.7% (He *et al.*, 2016). Nowadays, it is a near-natural urban forest (He *et al.*, 2003). Affected by warm temperate-zone semi-humid monsoon climate, this area has an annual average temperature of 6.2 to 9.7 °C. The average temperatures in January and July are −12.6 and 27.5 °C, respectively. The maximum temperature is 38.3 °C and the minimum temperature is −30.5 °C (Xu *et al.*, 2014). The average annual precipitation is 755.4 mm. The frost-free period lasts for 150 d yearly (Xu *et al.*, 2005). The flora of the arboretum located area belongs to the intersection of Changbai Mountain, Northern China, and the Mongolian Floras (Xu *et al.*, 2006).

This experiment was carried out in open-top chambers (OTCs). Chambers are 4 m in diameter and 3 m in height, with a 45° sloping frustum and 4-m distance between neighbouring OTCs (Li *et al.*, 2011; Xu *et al.*, 2015). This experiment included control in ambient air (AA, about 40 ppb) and elevated O₃ concentration (EO, 120 ppb). AA and EO had three independent OTC replicates with three OTCs, respectively. Six OTCs were used in total. In 20 May, 2012, nine healthy and uniform five-year-old saplings of *G. biloba* (1.5 m in average height) from a local nursery were selected and planted in each OTC. During the growing season, the saplings were irrigated twice a week and fertilized once at the beginning of the experiment. After 60 d (20 July), the saplings were fumigated with O₃ for 8 h a day from 9:00–17:00, except in bad weather such as thunderstorm conditions. By the end of fumigation on 5 November, 2012, all the yellow (senescent) leaves of *G. biloba* from each chamber were harvested and regarded as leaf litter. These senescent leaves under AA and EO were randomly divided into two parts: one part was dried at 65 °C to constant weight for determining the initial chemical compositions and the other was sufficiently dried, and stored in a big glass container and then kept at a cool place under room temperature for the litter decomposition experiments in OTCs next year (2013).

The leaf litters were decomposed according to the litterbag method (Zhang *et al.*, 2015) by placing them in nylon bags (20 cm × 25 cm) with 1 mm² mesh. Before the experiment, each bag was filled with 8 g samples and numbered. On 19 May, 2013, 15 bags were placed into each OTC to touch the topsoil and separate them at 10-cm intervals each other. A total of 45 bags of AA and EO were prepared, respectively. Before putting the litter bags into OTCs, the topsoil (0–10 cm) of each treatment was gathered after removing weeds and other sundries from the ground. Soil samples were mixed uniformly after the stones, roots and other debris were removed. Homogenized fresh soil was passed through a sieve (pore size was 5 mm) and dried under natural conditions. The basic physical and chemical properties of topsoil (0–10 cm) are shown in Table 1. O₃ fumigation was carried out for 8 h each day (9:00–17:00). The samples were collected once a month (Sampling date is 19 every month from June to October) to be decomposed for 150 days in total. O₃

Table 1 The physical and chemical properties of topsoil (0–10 cm) in OTCs before leaf litter decomposition of *G. biloba*.

Treatments	pH	SWC (%)	ST (°C)	SOC (mg/g)	C (mg/g)	N (mg/g)	P (mg/g)	K (mg/g)	C/N	C/P
AA	6.75 (0.06)	28.25 (1.08)	25.50 (0.82)	9.12 (0.30)	33.23 (0.79)	2.54 (0.03)	4.44 (0.09)	5.70 (0.08)	13.10 (0.40)	7.48 (0.27)
EO	6.67 (0.03)	28.34 (0.97)	25.30 (1.15)	9.20 (0.04)	34.12 (0.19)	2.58 (0.01)	4.35 (0.06)	5.44 (0.43)	13.22 (0.11)	7.85 (0.06)

Notes.

Data are shown mean and standard deviation (SD) in the parenthesis ($n = 3$).

AA, ambient air (control); EO, elevated O₃; SWC, soil water content; ST, soil temperature; SOC, soil organic matter content.

concentrations were monitored every day throughout the growing period (19 May–20 October 2013), at 0.80 m above the ground, in the OTCs of both AA and EO (Fig. 1). The microclimatic conditions in the OTCs during litter decomposition are summarised in Table 2. After washing and drying, the cleaned litters were placed into Kraft envelope bags and oven-dried at a constant temperature of 65° C. Afterwards, leaf litters were weighed and dry mass were pulverised by a high-speed grinding machine (FW-100; Taisite Instrument Co., Ltd., Tianjing, China) and filtered through a sieve (80 meshes, mesh size is 5 mm). The powdered samples were preserved in self-sealing bags for the laboratory measurements.

Chemical analyses and experimental statistics

Carbon (C) and nitrogen (N) contents were determined by an elemental analyser (Vario MACRO Cube, Elementar, Germany). Phosphorus (P) and potassium (K) contents were measured by atomic absorption spectrometry (AA800; Perkin Elmer, San Jose, CA, USA) according to the Mo-Sb colorimetric method and flame photometric method, respectively. Lignin content was determined using the ultraviolet spectrophotometric method (Liyama & Wallis, 1988) and total phenol content was measured according to the method of Julkunen-Tiitto (1985) with a minor modification. The condensed tannins and soluble sugar contents were determined by spectrophotometer (UV-1800, Shimadzu Corp., Kyoto, Japan) according to the butyl alcohol-hydrochloric acid method (Porter, Hrstich & Chan, 1986) and anthrone colorimetric method (Dubois et al., 1956), respectively.

Mass remaining of leaf litter over time, expressed as percentage of the initial value, was calculated according to the assumed simple exponential model formulated by Olson (1963).

$$Y = A_t/A_0 = e^{-kt}$$

Where, Y , A_0 and A_t indicate the remaining rate of litter mass monthly, the initial litter mass (g), and the remaining mass (g) of leaf litter at time t (months), respectively. In addition, e , and k are the base of natural logarithms, and the decomposition coefficient of the litters, respectively. As for t , it is the decomposition time (months) including the half-life of decomposition ($t_{0.5} = \ln(2)/k$).

The remaining rate of nutrient composition was calculated (Pancotto et al., 2003):

$$E = [(M_t \times C_t)/(M_0 \times C_0)] \times 100\%$$

Where, E , M_t , M_0 , C_0 , and C_t represent the remaining (% initial) of nutrient elements, dry mass (g) of leaf litter at the designated time of decomposition, initial dry mass (g), initial

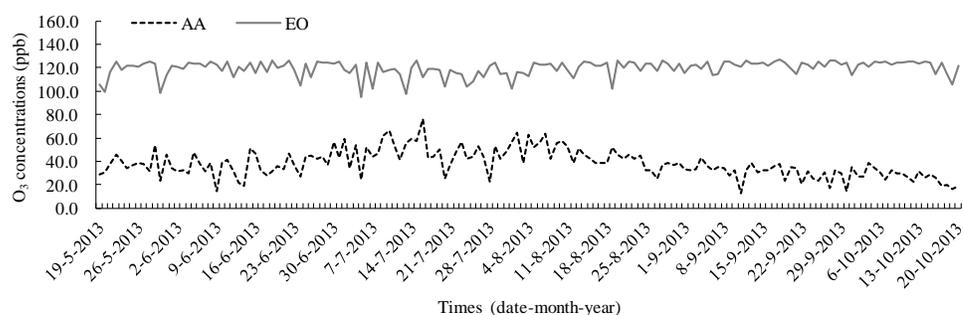


Figure 1 Seasonal variations in O₃ concentrations in OTCs with ambient air (AA) and elevated O₃ (EO) during leaf litter decomposition.

Full-size DOI: 10.7717/peerj.4453/fig-1

Table 2 Microclimatic conditions in OTCs during gas fumigation in 2013.

Treatments	[O ₃] _{mean}	[O ₃] _{max}	AOT40 ⁽¹⁵⁰⁾	RH _{mean}	T _{mean}	[CO ₂]	DPPFD
AA	38.2	76.5	1,167.5	68.3	23.8	372.4	46.9
EO	119.5	126.0	77,246.1	66.9	24.1	368.9	46.5

Notes.

[O₃]_{mean}, average daily (08:–17:00) concentrations of O₃ (ppb); [O₃]_{max}, average maximum daily concentrations of O₃ (ppb); AOT40, cumulative the sum of the differences between the hourly mean ozone concentration in ppb and 40 ppb for each hour of gas exposure; AOT40⁽¹⁵⁰⁾, indicates the accumulated values of AOT40 during the 150-day decomposition experiment (ppb · h); RH_{mean}, average daily air relative humidity (%); T_{mean}, average daily air temperature (°C); AA, ambient air; EO, elevated O₃; [CO₂]_{mean}, average air CO₂ concentration in OTC (μmol mol⁻¹); DPPFD, average daily photosynthetic photo flux density at the canopy level (mol m⁻² day⁻¹).

nutrient content (mg g⁻¹), and nutrient content (mg g⁻¹) of leaf litter at the designated time of decomposition, respectively.

Data analyses were performed using SPSS 18.0 (SPSS Inc., Chicago, IL, USA). General linear model (GLM) was used to evaluate and analyse the dynamics of leaf litter mass remaining, changes in C, N, P, lignin, total phenols, condensed tannin and soluble sugar contents with considering the treatments (two levels), decomposition time (continuous variable), as well as their interaction, as independent factors. All data were presented as mean ± standard deviation. Significant differences between control (ambient air, AA) and elevated O₃ concentration (EO) were tested by *T*-test.

RESULTS

Changes in chemical compositions in leaf litters of *G. biloba* after exposure to EO

In comparison with AA, EO decreased C content (by 26.1%), but increased slightly N content (by 11.3%) in leaves of *G. biloba* by the end of growing season (Fig. 2). EO decreased C/N ratio (by 10.3%). P content decreased under O₃ fumigation. However, changes in C, N, P and C/N were not significant. EO significantly increased the K content by 184.2% (6.31 ± 0.29 vs 17.93 ± 0.40, *P* < 0.01) compared with AA (Fig. 2).

In addition, EO decreased the contents of lignin (by 8.6%) and condensed tannins (by 17.5%). By contrast, the difference between EO and AA was not significant (Fig. 2). EO

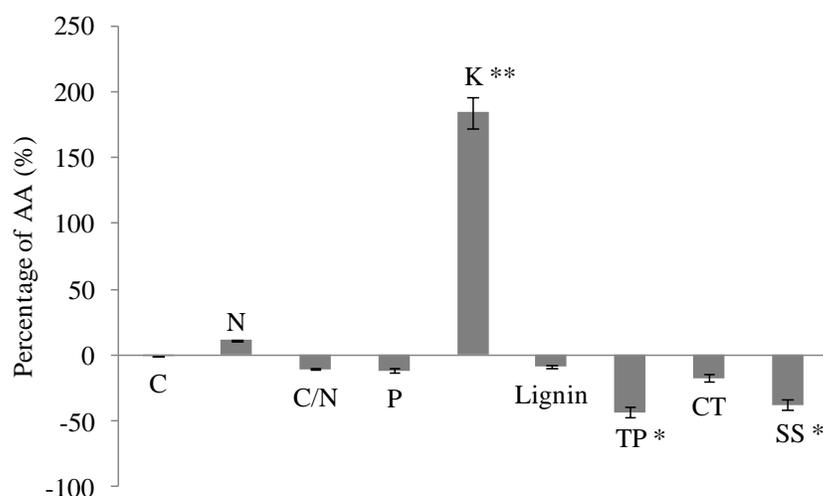


Figure 2 Leaf chemistry changes in *G. biloba* as affected by O_3 concentration.

Full-size [DOI: 10.7717/peerj.4453/fig-2](https://doi.org/10.7717/peerj.4453/fig-2)

significantly decreased the contents of total phenolics (2.82 ± 0.93 vs 1.60 ± 0.44 , $P < 0.05$) and soluble sugar (86.51 ± 19.57 vs 53.76 ± 2.40 , $P < 0.05$) by 43.3% and 37.9% compared with AA, respectively.

Dynamics of leaf litter decomposition of *G. biloba* exposed to EO

Based on the Olson's model, the decay constant and the half-life decomposition time of the leaf litters of *G. biloba* under EO were lower than those AA, showing no significant difference (Table 3). Compared with AA, the mass (dry weight) remaining of leaf litters under EO maintained a higher level at early stage of decomposition (before 60-day sampling point), but decreased with decomposition time after 90 days, and it was 5.8% lower than that of AA at 150-day sampling point (Fig. 3A). No significant difference between AA and EO was observed in mass remaining of *G. biloba* leaf litter over time. O_3 fumigation reduced the remaining of C in leaf litter and a significant difference between AA and EO was found after 120 days (EO decreased by 8.3% compared with that of AA) (Fig. 3B). By contrast, N remaining showed a lower level after 90 days under EO than that of AA. However, no significant difference was found in N remaining between treatments during decomposition (Fig. 3C). P remaining in leaf litter showed higher level under EO than AA, and decreased dramatically after 60 days and reached a lowest value at 90-day sampling point (11.2%) (Fig. 3D).

The remaining of lignin in leaf litters of *G. biloba* showed a significant decreasing trend regardless of O_3 treatment during decomposition (Fig. 3E, Table 4, $P < 0.001$). Compared with the AA, the remaining of lignin under EO was quite lower after 120 days, indicating an increasing of lignin decomposition rate. However, no significant effect was observed in the remaining of lignin under EO (Table 4, $P = 0.249$). After 60 days, the remaining of total phenol content showed a significant decrease under EO at each sampling point (Fig. 3F), compared with AA ($P < 0.05$). A significant interactive effect of O_3 concentration and decomposition time was found for the total phenol remaining (Table 4, $P = 0.002$).

Table 3 Parameters of decomposition rate of *G. biloba* litter leaves exposed to elevated O₃ concentration.

Treatments	Olson exponential decay	R ²	Decay constants (K)	t _{0.5} /a
AA	$y = 0.8512e^{-1.17x}$	0.73	1.17	0.46
EO	$y = 0.8749e^{-1.39x}$	0.84	1.39	0.41

Notes.AA, ambient air; EO, elevated O₃.

No significant difference in each parameter was found between AA and EO.

EO showed no significant effect on the decomposition of condensed tannins during the experiment (Fig. 3G, Table 4, $P = 0.326$). During decomposition, the remaining of soluble sugar showed a significant higher level at each sampling point except for the 90-day sampling point under EO than AA ($P < 0.05$) (Fig. 3H).

DISCUSSION

Effect of elevated O₃ concentration on chemical composition of *G. biloba* leaf litter

The results of our study showed that elevated O₃ concentration (120 ppb) increased N and K contents in leaf litter of *G. biloba*, but significantly decreased the total phenol content, as well as the C/N and lignin/N ratios. However, the P and lignin contents, as well as the C/P ratio, did not change significantly. Actually, elevated O₃ concentration usually changes the chemical composition of plants (Calatayud et al., 2011; Shang et al., 2018). Booker et al. (2005) found that the changes in N and lignin contents of soybean leaf litter increase significantly, while the soluble sugar content significantly decreased under elevated O₃ concentration (74 ppb), in agreement with our results in this study. Parsons, Lindroth & Bockheim (2004) tested the impact of high O₃ concentration (55 ppb) on leaf litter of *Betula papyrifera* for 12 months. The results demonstrated that the contents of N, lignin, and condensed tannins, as well as the C/N ratio of leaf litter, showed no significant change under elevated O₃ concentration compared to tests in ambient air conditions.

In our study, the increasing of N content in *G. biloba* leaves after high O₃ exposure may be a helpful response to prevent the damage caused by O₃ to some extent (Cao et al., 2016). High N concentration in leaves could make plants adapt to O₃ stress through leaf turnover (Tjoelker & Luxmoore, 1991; Shang et al., 2018). In addition, the C/N ratio was an important indicator for the degree of coordination of C and N metabolism. Reduction of C/N in plants under elevated O₃ concentration showed that the growth of plants was inhibited (Zheng et al., 2011), in agreement with our result that C/N ratio decreased under O₃ fumigation. Zhang et al. (2011) demonstrated that elevated O₃ concentration promoted K absorption during growth of plants. Furthermore, K increased N absorption in order to transfer it into proteins under adverse environment. The increase of K contents in our study was possibly one of the reasons why N content increased under elevated O₃ concentration.

Plant secondary substances play a pivotal role in scavenging the high level of oxygen species caused by ozone at the end of O₃ fumigation (He et al., 2009). Generally, the secondary metabolic enzymes such as phenylalanine ammonia lyase (PAL), peroxidase (POD), and polyphenol oxidase (PPO) are involved in the synthesis of secondary

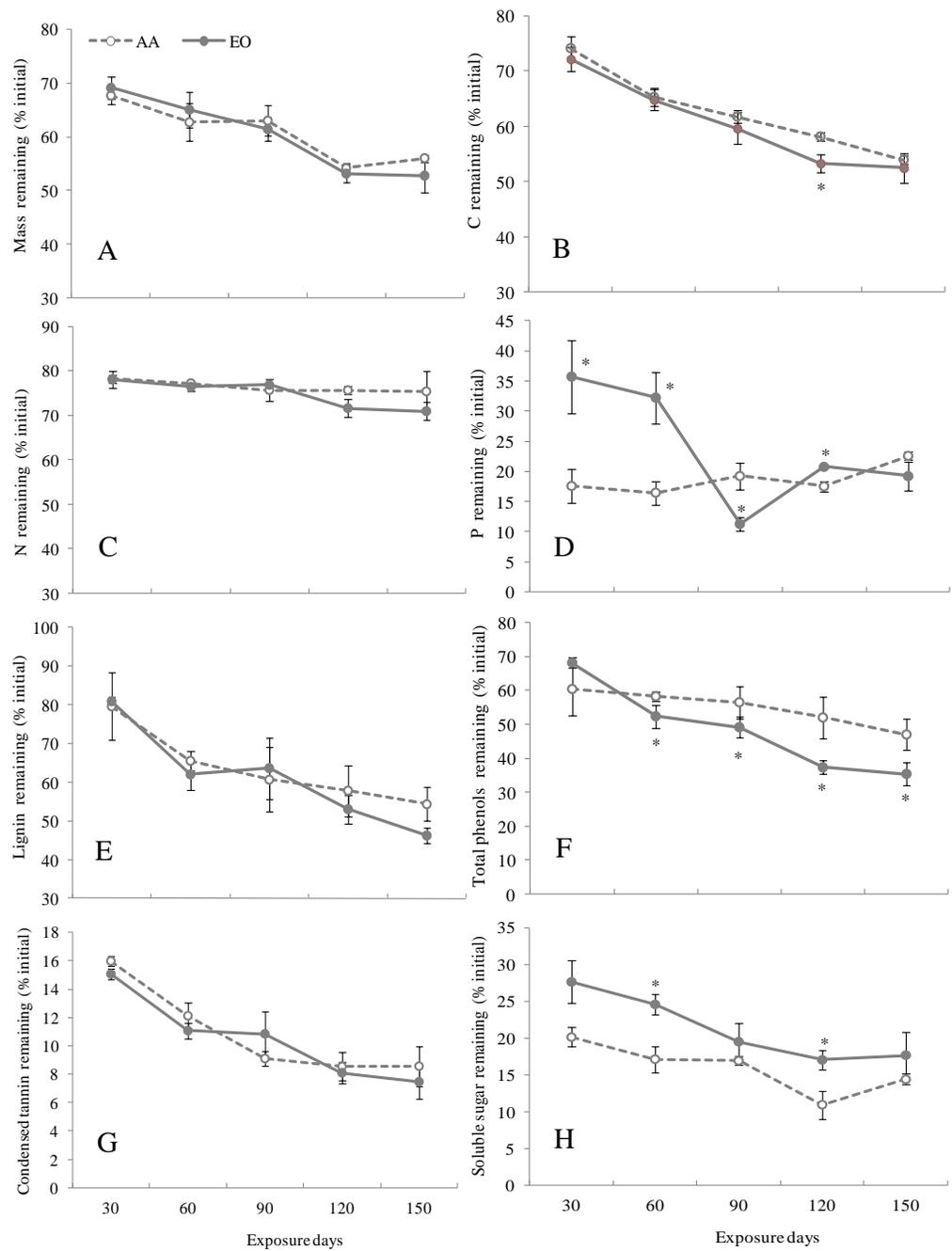


Figure 3 The remaining of mass (A), contents of C (B), N (C) and P (D), lignin (E), total phenolics (F), condensed tannins (G) and soluble sugars (H) in leaf litter of *G. biloba* under elevated O_3 concentration (EO, 120 ppb) and ambient air (AA, 40 ppb) for 150 days.

Full-size DOI: 10.7717/peerj.4453/fig-3

Table 4 Summary of the GLM testing (type III Sum of Squares, in bold values significant at $p < 0.05$) for effects of O₃ treatments and decomposition time (T) on mass and nutrients remaining of leaf litter in *G. biloba*.

	<i>d.f</i>	SS	MS	<i>F</i>	<i>p</i>		<i>d.f</i>	SS	MS	<i>F</i>	<i>p</i>
Mass						Lignin					
O ₃	1	1.31	1.31	0.23	0.637	O ₃	1	43.90	43.90	1.41	0.249
Time	4	963.83	240.95	42.44	<0.001	Time	4	3,102.30	775.57	24.87	<0.001
O ₃ × T	4	31.34	7.83	1.38	0.276	O ₃ × T	4	123.04	30.76	0.99	0.437
Residuals	20	113.55	5.68			Residuals	20	623.67	31.18		
C						TP					
O ₃	1	35.78	35.78	10.04	0.005	O ₃	1	302.88	302.88	16.33	0.001
Time	4	1,504.32	376.08	105.51	<0.001	Time	4	1,995.99	499.00	26.91	<0.001
O ₃ × T	4	15.55	3.89	1.09	0.388	O ₃ × T	4	446.69	111.67	6.02	0.002
Residuals	20	71.29	3.66			Residuals	20	370.91	18.55		
N						CT					
O ₃	1	20.24	20.24	4.82	0.040	O ₃	1	0.94	0.94	1.01	0.326
Time	4	107.98	27.00	6.43	0.020	Time	4	225.17	56.29	60.82	<0.001
O ₃ × T	4	36.94	9.24	2.20	0.106	O ₃ × T	4	8.60	2.15	2.32	0.092
Residuals	20	83.92	4.20			Residuals	20	18.51	0.93		
P						SS					
O ₃	1	200.96	200.96	25.27	<0.001	O ₃	1	217.48	217.48	58.83	<0.001
Time	4	477.90	119.48	15.02	<0.001	Time	4	366.11	917.53	24.77	<0.001
O ₃ × T	4	796.09	199.02	25.03	<0.001	O ₃ × T	4	32.24	8.06	2.18	0.108
Residuals	20	159.04	7.95			Residuals	20	73.90	3.70		

Notes.

TP, total phenols; CT, condensed tannin; SS, soluble sugar.

substances (Tiwari, Sangwan & Sangwan, 2016). Luo & Zhang (2010) found that elevated O₃ concentration significantly decreased the lignin contents of plants by inhibiting the activities of PAL, POD and PPO. In our study, the decrease of lignin content in *G. biloba* leaves could be due to the inhibition of activities of relevant enzyme for synthesising lignin by elevated O₃ concentration.

Phenolic compounds are the main products of secondary metabolism and are important defence substances in plants. Some research demonstrated that elevated O₃ concentrations increased the contents of phenolic compounds in leaves (Yamaji et al., 2003; Peltonen, Vapaavuori & Julkunen-Titto, 2005). In our previous studies, elevated O₃ concentration (80 ppb) significantly increased the total phenolics and condensed tannin contents in leaves of *Quercus mongolica*, leading to the increasing of the antioxidant capacity of plants to O₃ stress (Zhang et al., 2009). In this study, elevated O₃ concentration decreased the contents of condensed tannins and total phenols of *G. biloba* leaves. It could be due to the produce of a large number of free radicals under O₃ stress, leading to many antioxidant substances including phenolic compounds were consumed by the end of the growing season (He et al., 2009). Therefore, the decreases of condensed tannins and total phenolic content in *G. biloba* leaves may increase the sensitivity of the leaf injuries to O₃ (Yamaji et al., 2003). Actually, no effect of elevated O₃ concentrations on the contents of condensed tannins and phenolic compounds was reported by some studies (Lavola, Julkunen-Tiitto & Paakkonen,

1994; Lindroth et al., 2001), not in agreement with our current results due to the differences in tree species and O₃ concentrations to some extent.

Soluble sugar plays an important role in plant metabolism under adverse environments (Liu et al., 2004). Wang et al. (2011) reported that O₃ fumigation (60 ppb, 50% above ambient air) significantly decreased the soluble sugar content of rice in each growth stage. However, Lu et al. (2012) found that the content of soluble sugar in the leaves of *Mangifera indica* increased under O₃ concentration (50 ppb), but decreased significantly under a high-O₃ concentration (200 ppb), in agreement with our result that the soluble sugar contents in leaves of *G. biloba* decreased significantly under elevated O₃ concentration (120 ppb). The reason for this was probably due to the accumulation of glycolytic enzymes in leaves, which accelerated the degradation of sugar components (Tiwari, Sangwan & Sangwan, 2016).

Effect of elevated O₃ concentration on the decomposition rate of *G. biloba* leaf litter

Elevated O₃ concentration not only changes the chemical composition of leaf litter, but also indirectly affects the litter decomposition (Baldantoni, Fagnano & Alfani, 2011). In this study, the remaining mass of dry weight of *G. biloba* leaf litter showed significant positive correlation with the C, N, P, and lignin contents as well as the ratios of C/N and lignin/N, regardless of O₃ treatment (Table 5). This indicated that the higher the contents of C, N, P, and lignin, the higher the mass remaining, and the lower the decomposition rate (Bonanomi et al., 2013). In this study, the mass remaining are larger under elevated O₃ concentration than under ambient air at early decomposition stage, which implied that the decomposition of leaf litter slowed down under O₃ fumigation. This is consistent with most of the previous studies showing that elevated O₃ has adverse effects on litter decomposition (Parsons, Bockheim & Lindroth, 2008; Liu et al., 2009; Baldantoni, Fagnano & Alfani, 2011). At the late stage of decomposition (after 90 days), the mass remaining of leaf litter showed higher level under ambient air than elevated O₃ treatment, which indicated that O₃ slightly increased the decay rate of leaf litter by impacting litter quality in this study. Indeed, Litter quality is the most important factor affecting mass loss and decay rates of nutrients (Bonanomi et al., 2010).

In fact, O₃ exposure has not always led to reduction in litter decomposition rate of tree species (Scherzer, Rebbeck & Boerner, 1998; Kainulainen, Holopainen & Holopainen, 2003). In our recent study, we observed that N mineralization and lignin degradation in leaf litters of *Q. mongolica* under elevated O₃ concentration (120 ppb) were inhibited during early stage of decomposition, but promoted at later stage of decomposition (Su et al., 2016). Indeed, as the major component in leaf litter, lignin has a complex structure and is difficult to decompose (Peng & Liu, 2002). During the decomposition in this experiment, the remaining rates of lignin, condensed tannin and total phenols were lower under elevated O₃ concentration than under ambient air at late decomposition stage, which implied that elevated O₃ concentration promoted the decomposition of leaf litter. The slight promotion of the decomposition rate of leaf litter mainly resulted from the decrease of leaf litter quality induced by O₃ fumigation. Besides, exposure of O₃ at the late decomposition stage of the

Table 5 Spearman correlations coefficients of remaining mass to initial with nutrient contents dynamics during decomposition of *G. biloba* leaf litter.

	Treatments	Remaining mass	C	N	C/N	P	Lignin
Remaining mass	−0.07						
C	−0.07	0.96**					
N	0.12	0.33*	0.28				
C/N	−0.17	0.91**	0.96**	0.17			
P	0.28	0.79**	0.79**	0.48**	0.66**		
Lignin	−0.36*	0.66**	0.66**	0.28	0.75**	0.36*	
Lignin/N	−0.32	0.63**	0.67**	0.19	0.78**	0.34*	0.95**

Notes.

* $P < 0.05$.** $P < 0.01$.

experiment might accelerate the oxidation and decomposition of secondary metabolic substances (Su et al., 2016).

Unlike the remaining of lignin, condensed tannin and total phenols, the remaining of soluble sugar showed higher values under elevated O_3 concentration than ambient air at any sampling point during litter decomposition. The lower decay rate observed for ozone-exposed leaves could be due to changes in both structural and functional molecules that we did not study here or decreases in the activities of enzymes relating to carbohydrate metabolism under O_3 exposure (Peace, Lea & Darrall, 1995). In addition, the higher value of carbohydrate remaining was maintained under O_3 fumigation during decomposition and becoming much smaller difference between AA and EO by the end of the experiment, which might result from inhibiting effect of O_3 on decay rates of leaf litter with high initial values at early decomposition stage. The similar results were observed in some previous studies (Fioretto et al., 2005; Liu et al., 2009).

CONCLUSIONS

In this study, we found that elevated O_3 showed no significant impact on chemical compositions and decay rates of *G. biloba*, although it decreased the contents of C, P, C/N ratio, lignin, total phenols, condensed tannins, and soluble sugars in *G. biloba* leaves by the end of gas fumigation. In fact, O_3 fumigation slightly inhibited the decomposition of *G. biloba* leaf litter at the early stages of decomposition, but increased decomposition rate at late stages of this experiment. During the whole decomposition, the losses of the nutrients in leaf litters of *G. biloba* showed significant seasonal differences regardless of O_3 treatment. Rising atmospheric O_3 concentration will likely elicit species-specific effects on litter production and decomposition in urban forests, retarding decay rates in some species such as birch (Parsons, Bockheim & Lindroth, 2008), while potentially exerting little effect on others. Furthermore, elevated O_3 will not exert its influence on litter decay rates in isolation from other factors including soil microorganism, although the results from our study showed that O_3 in current concentration in this study had little (not significant) effect on decomposition of *G. biloba* leaf litter. Our results indicated that the ground-level O_3 concentrations in some cities of China could significantly alter the chemical compositions

and decomposition rates of at least one deciduous gymnosperm tree species once suffering from long-time exposure of higher O₃ concentration. These changes are likely to have important implications for our understanding of the processes regulating the storage and emission of C from urban forest ecosystems under climate change. Therefore, further research for the long-term O₃ exposure of urban trees (only one growing season in this study) is necessary to determine the nutrient cycling and sustainability of main tree species with different ages including *G. biloba* in urban forest, where O₃ is one of the most widespread of all the gaseous pollutants in urban area.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

This work was jointly supported by the National Natural Science Foundation of China (NSFC, 41675153, 31270518 and 31170573), the Key development program of the Chinese Academy Science (KFZD-SW-302-01) and the Key Project of NSFC (90411019). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors:

National Natural Science Foundation of China: 41675153, 31270518, 31170573.

Chinese Academy Science: KFZD-SW-302-01.

Key Project of NSFC: 90411019.

Competing Interests

The authors declare there are no competing interests.

Author Contributions

- Wei Fu performed the experiments, analyzed the data, contributed reagents/materials/-analysis tools, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Xingyuan He and Wei Chen conceived and designed the experiments, authored or reviewed drafts of the paper, approved the final draft.
- Sheng Xu analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Yan Li analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft, search for the references.
- Bo Li conceived and designed the experiments, performed the experiments, contributed reagents/materials/analysis tools, authored or reviewed drafts of the paper, approved the final draft, revised the paper.
- Lili Su performed the experiments, contributed reagents/materials/analysis tools, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.

- Qin Ping performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.

Data Availability

The following information was supplied regarding data availability:

The raw data is provided in the [Supplemental Files](#).

Supplemental Information

Supplemental information for this article can be found online at <http://dx.doi.org/10.7717/peerj.4453#supplemental-information>.

REFERENCES

- Akimotoa H, Morib Y, Sasaki K, Itano Y. 2015.** Analysis of monitoring data of ground-level ozone in Japan for long term trend during 1990–2010: causes of temporal and spatial variation. *Atmospheric Environment* **102**:302–310 DOI [10.1016/j.atmosenv.2014.12.001](https://doi.org/10.1016/j.atmosenv.2014.12.001).
- Austin AT, Vivanco L. 2006.** Plant litter decomposition in a semi-arid ecosystem controlled by photodegradation. *Nature* **442**:555–558 DOI [10.1038/nature05038](https://doi.org/10.1038/nature05038).
- Baldantoni D, Fagnano M, Alfani A. 2011.** Tropospheric ozone effects on chemical composition and decomposition rate of *Quercus ilex* L. leaves. *Science of the Total Environment* **409**:979–984 DOI [10.1016/j.scitotenv.2010.11.022](https://doi.org/10.1016/j.scitotenv.2010.11.022).
- Bonanomi G, Incerti G, Antignani V, Capodilupo M, Mazzoleni S. 2010.** Decomposition and nutrient dynamics in mixed litter of Mediterranean species. *Plant Soil* **331**:481–496 DOI [10.1007/s11104-009-0269-6](https://doi.org/10.1007/s11104-009-0269-6).
- Bonanomi G, Incerti G, Giannino F, Mingo A, Lanzotti V, Mazzoleni S. 2013.** Litter quality assessed by solid state ^{13}C NMR spectroscopy predicts decay rate better than C/N and Lignin/N ratios. *Soil Biology & Biochemistry* **56**:40–48 DOI [10.1016/j.soilbio.2012.03.003](https://doi.org/10.1016/j.soilbio.2012.03.003).
- Booker FL, Prior SA, Torbert HA, Hu S. 2005.** Decomposition of soybean grown under elevated concentrations of CO_2 and O_3 . *Global Change Biology* **11**:685–698 DOI [10.1111/j.1365-2486.2005.00939.x](https://doi.org/10.1111/j.1365-2486.2005.00939.x).
- Calatayud V, Cerveró J, Calvo E, Garcia FJ, Armiriana JR. 2011.** Responses of evergreen and deciduous *Quercus* species to enhanced ozone levels. *Environment Pollution* **159**:55–63 DOI [10.1016/j.envpol.2010.09.024](https://doi.org/10.1016/j.envpol.2010.09.024).
- Cao JX, Shang H, Chen Z, Tian Y, Yu H. 2016.** Effects of elevated ozone on stoichiometry and nutrient pools of *Phoebe Bournei* (Hemsl.) Yang and *Phoebe Zhennan* S. Lee et FN Wei seedlings in subtropical China. *Forests* **7**:78–88 DOI [10.3390/f7040078](https://doi.org/10.3390/f7040078).
- Chappelka AH, Samuelson LJ. 1998.** Ambient ozone effects on forest trees of the eastern United States: a review. *New Phytologist* **139**:91–108 DOI [10.1046/j.1469-8137.1998.00166.x](https://doi.org/10.1046/j.1469-8137.1998.00166.x).
- De Bauer MDL, Hernández-Tejeda T. 2007.** A review of ozone-induced effects on the forests of central Mexico. *Environmental Pollution* **147**:446–453 DOI [10.1016/j.envpol.2006.12.020](https://doi.org/10.1016/j.envpol.2006.12.020).

- Dubois M, Gilles KA, Hamilton JK, Rebers PA, Smith F. 1956.** Colorimetric method for determination of sugars and related substances. *Analytical Chemistry* **28**:350–356 DOI [10.1021/ac60111a017](https://doi.org/10.1021/ac60111a017).
- Fioretto A, Di Nardo C, Papa S, Fuggi A. 2005.** Lignin and cellulose degradation and nitrogen dynamics during decomposition of three leaf litter species in a Mediterranean ecosystem. *Soil Biology and Biochemistry* **37**:1083–1091 DOI [10.1016/j.soilbio.2004.11.007](https://doi.org/10.1016/j.soilbio.2004.11.007).
- He X, Chen W, Xu W, Liu C, Zhu W, Jin Y, Zhang Y. 2003.** Community ecology in a typical near natural urban forest. *Chinese Journal of Ecology* **22**:162–168.
- He XY, Huang W, Chen W, Dong T, Liu C, Chen Z, Xu S, Ruan Y. 2009.** Changes of main secondary metabolites in leaves of *Ginkgo biloba* in response to ozone fumigation. *Journal of Environment Sciences* **21**:199–203 DOI [10.1016/S1001-0742\(08\)62251-2](https://doi.org/10.1016/S1001-0742(08)62251-2).
- He X, Xu S, Xu W, Chen W, Huang Y, Wen H. 2016.** Effects of climate warming on phenological characteristics of urban forest in Shenyang City, China. *Chinese Geographical Science* **26**:1–9 DOI [10.1007/s11769-015-0782-x](https://doi.org/10.1007/s11769-015-0782-x).
- IPCC. 2013.** *Climate change, 2013: the physical science basis*. Cambridge: Cambridge University Press.
- Julkunen-Tiitto R. 1985.** Phenolic constituents in the leaves of northern willows: methods for the analysis of certain phenolics. *Agricultural and Food Chemistry* **33**:213–217 DOI [10.1021/jf00062a013](https://doi.org/10.1021/jf00062a013).
- Kainulainen P, Holopainen T, Holopainen JK. 2003.** Decomposition of secondary compounds from needle litter of Scots pine grown under elevated CO₂ and O₃. *Global Change Biology* **9**:295–304 DOI [10.1046/j.1365-2486.2003.00555.x](https://doi.org/10.1046/j.1365-2486.2003.00555.x).
- Lavola A, Julkunen-Tiitto R, Paakkonen E. 1994.** Does ozone stress change the primary or secondary metabolites of birch (*Betula pendula* Roth.)? *New Phytologist* **126**:637–642 DOI [10.1111/j.1469-8137.1994.tb02959.x](https://doi.org/10.1111/j.1469-8137.1994.tb02959.x).
- Li XM, Zhang LH, Li YY, Ma LJ, Chen Q, Wang LL, He XY. 2011.** Effects of elevated carbon dioxide and/or ozone on endogenous plant hormones in the leaves of *Ginkgo biloba*. *Acta Physiologiae Plantarum* **33**:129–136 DOI [10.1007/s11738-010-0528-4](https://doi.org/10.1007/s11738-010-0528-4).
- Lindroth R, Kopper BJ, Parsons WFJ, Bockheim JG, Karnosky DFK, Hendrey GR, Pregitzer KS, Isebrands JG, Sober J. 2001.** Consequences of elevated carbon dioxide and ozone for foliar chemical composition and dynamics in trembling aspen (*Populus tremuloides*) and paper birch (*Betula papyrifera*). *Environmental Pollution* **115**:395–404 DOI [10.1016/S0269-7491\(01\)00229-9](https://doi.org/10.1016/S0269-7491(01)00229-9).
- Liu FR, Chen HY, Liu Y, Wei ZM. 2004.** Changes in solute content of different tomato genotypes under salt stress. *Acta Photophysiological Sinica* **30**:99–104.
- Liu L, King JS, Giardina CP, Booker FL. 2009.** The influence of chemistry, production and community composition on leaf litter decomposition under elevated atmospheric CO₂ and tropospheric O₃ in a northern hardwood ecosystem. *Ecosystems* **12**:401–416 DOI [10.1007/s10021-009-9231-y](https://doi.org/10.1007/s10021-009-9231-y).
- Liyama K, Wallis AFA. 1988.** An improved acetyl bromide procedure for determining lignin in woods and wood pulps. *Wood Science and Technology* **22**:271–280.

- Lu T, He XY, Chen W, Yan K, Zhao TH. 2009.** Effects of elevated O₃ and/or elevated CO₂ on lipid peroxidation and antioxidant systems in *Ginkgo biloba* leaves. *Bulletin of Environment Contamination Toxicology* **83**:92–96 DOI [10.1007/s00128-009-9719-3](https://doi.org/10.1007/s00128-009-9719-3).
- Lu G, Huang Y, Chen H, Xu L. 2012.** Effects of ozone on membrane lipid peroxidation and protective enzyme activities of *Thevetia peruviana* and *Mangifera* leaves. *Ecology & Environment Sciences* **21**:1235–1240 DOI [10.1080/01140670709510187](https://doi.org/10.1080/01140670709510187).
- Luo Z, Zhang L. 2010.** Effects of O₃ on lignifications and related enzyme activity in bamboo shoots. *Transaction Chinese Society Agricultural Machinery* **41**:115–117 DOI [10.3969/j.issn.1000-1298.2010.11.022](https://doi.org/10.3969/j.issn.1000-1298.2010.11.022).
- Manes F, Incerti G, Salvatori E, Vitale M, Ricotta C, Costanza R. 2012.** Urban ecosystem services: tree diversity and stability of tropospheric ozone removal. *Ecological Applications* **22**:349–360 DOI [10.1890/11-0561.1](https://doi.org/10.1890/11-0561.1).
- Nikolova PS, Andersen CP, Blaschke H, Matyssek R, Haberle KH. 2010.** Belowground effects of enhanced tropospheric ozone and drought in a beech/spruce forest (*Fagus sylvatica* L./*Picea abies* [L.] Karst). *Environmental Pollution* **158**:1071–1078 DOI [10.1016/j.envpol.2009.07.036](https://doi.org/10.1016/j.envpol.2009.07.036).
- Nikula S, Vapaavuori E, Manninen S. 2010.** Urbanization-related changes in European aspen (*Populus tremula* L): leaf traits and litter decomposition. *Environmental Pollution* **158**:2132–2142 DOI [10.1016/j.envpol.2010.02.025](https://doi.org/10.1016/j.envpol.2010.02.025).
- Olson JS. 1963.** Energy storage and the balance of producers and decomposers in ecological systems. *Ecology* **44**:322–331 DOI [10.2307/1932179](https://doi.org/10.2307/1932179).
- Pancotto VA, Sala OE, Cabello M, Lopez NI, Robson TM, Ballare CL, Caldwell MM, Scopel AL. 2003.** Solar UV-B decreases decomposition in herbaceous plant litter in Tierra del Fuego, Argentina: potential role of an altered decomposer community. *Global Change Biology* **9**:1465–1474 DOI [10.1046/j.1365-2486.2003.00667.x](https://doi.org/10.1046/j.1365-2486.2003.00667.x).
- Paoletti E. 2006.** Impact of ozone on Mediterranean forests: a review. *Environmental Pollution* **144**:463–474 DOI [10.1016/j.envpol.2005.12.051](https://doi.org/10.1016/j.envpol.2005.12.051).
- Parsons WF, Bockheim JG, Lindroth RL. 2008.** Independent, interactive, and species-specific responses of leaf litter decomposition to elevated CO₂ and O₃ in a northern hardwood forest. *Ecosystems* **11**:505–519 DOI [10.1007/s10021-008-9148-x](https://doi.org/10.1007/s10021-008-9148-x).
- Parsons WFJ, Lindroth RL, Bockheim JG. 2004.** Decomposition of *Betula papyrifera* leaf litter under the independent and interactive effects of elevated CO₂ and O₃. *Global Change Biology* **10**:1666–1677 DOI [10.1111/j.1365-2486.2004.00851.x](https://doi.org/10.1111/j.1365-2486.2004.00851.x).
- Peace EA, Lea PJ, Darrall NM. 1995.** The effect of open-air fumigation with SO₂ and O₃ on carbohydrate metabolism in Scots pine (*Pinus sylvestris*) and Norway spruce (*Picea abies*). *Plant, Cell & Environment* **18**:277–283 DOI [10.1111/j.1365-3040.1995.tb00362.x](https://doi.org/10.1111/j.1365-3040.1995.tb00362.x).
- Peltonen PA, Vapaavuori E, Julkunen-Titto R. 2005.** Accumulation of phenolic compounds in birch leaves is changed by elevated carbon dioxide and ozone. *Global Change Biology* **11**:1305–1324 DOI [10.1111/j.1365-2486.2005.00979.x](https://doi.org/10.1111/j.1365-2486.2005.00979.x).
- Peng SL, Liu Q. 2002.** The dynamics of forest litter and its responses to global warming. *Acta Ecologica Sinica* **22**:1534–1544.

- Porter LJ, Hrstich LN, Chan BG. 1986. The conversion of procyanidins and prodelphinidins to cyanidin and delphinidin. *Phytochemistry* 25:223–230
DOI 10.1016/S0031-9422(00)94533-3.
- Scherzer AJ, Rebbeck J, Boerner REJ. 1998. Foliar nitrogen dynamics and decomposition of yellow-poplar and eastern white pine during four seasons of exposure to elevated ozone and carbon dioxide. *Forest Ecology and Management* 109:355–366
DOI 10.1016/S0378-1127(98)00290-4.
- Shang B, Feng Z, Li P, Calatayud V. 2018. Elevated ozone affects C, N and P ecological stoichiometry and nutrient resorption of two poplar clones. *Environmental Pollution* 234:136–144 DOI 10.1016/j.envpol.2017.11.056.
- Sicard P, Augustaitis A, Belyazid S, Calfapietra C, Marco AD, Fenn M, Bytnerowicz A, Grulke N, He S, Matyssek R, Serengil Y, Wieser G, Paoletti E. 2016. Global topics and novel approaches in the study of air pollution, climate change and forest ecosystems. *Environmental Pollution* 213:977–987 DOI 10.1016/j.envpol.2016.01.075.
- Sitch S, Cox PM, Collins WJ, Huntingford C. 2007. Indirect radiative forcing of climate change through ozone effects on the land-carbon sink. *Nature* 448:791–794
DOI 10.1038/nature06059.
- Su L, Xu S, Fu W, He X, Chen W, Zhao Y, Ping Q. 2016. Effects of elevated O₃ on the leaf litter decomposition and nutrient release of *Quercus mongolica* in city. *Chinese Journal of Applied Ecology* 27:373–379.
- Tiwari P, Sangwan RS, Sangwan NS. 2016. Plant secondary metabolism linked glycosyltransferases: an update on expanding knowledge and scopes. *Biotechnology Advances* 34:714–739 DOI 10.1016/j.biotechadv.2016.03.006.
- Tjoelker MG, Luxmoore RJ. 1991. Soil nitrogen and chronic ozone stress influence physiology, growth, and nutrient status of *Pinus taeda* L. and *Liriodendron tulipifera* L. seedlings. *New Phytologist* 119:69–81 DOI 10.1111/j.1469-8137.1991.tb01009.x.
- Wang B, Shugart HH, Shuman JK, Lerdau MT. 2016. Forests and ozone: productivity, carbon storage, and feedbacks. *Scientific Reports* 6:22133 DOI 10.1038/srep22133.
- Wang Y, Wang X, Zhou X, Yang L, Zhu J, Kobayashi K, Wang Y. 2011. Impacts of tropospheric ozone concentration enrichment on lodging resistance of rice Wuyuanjing 21. *Jiangsu Journal of Agricultural Science* 27:1167–1173.
- Xu W, Chen W, He X, Xu S, Zhang Y, Wen H. 2012. Litterfall amount and dynamics in urban forest of Shenyang. *Chinese Journal of Applied Ecology* 23:2931–2939.
- Xu W, He X, Chen W, Hu J, Wen H. 2006. Responses of Shenyang urban tree phenology to climate warming. *Chinese Journal of Applied Ecology* 17:1777–1781.
- Xu S, He X, Chen W, Huang Y, Zhao Y, Li B. 2015. Differential sensitivity of four urban tree species to elevated O₃. *Urban Forest & Urban Greening* 14:1166–1173
DOI 10.1016/j.ufug.2015.10.015.
- Xu W, He X, Chen W, Liu C, Sun Y. 2005. Microclimate characters of urban forest in Shenyang City. *Chinese Journal of Applied Ecology* 16:1650–1654.
- Xu S, Xu W, Chen W, He X, Huang Y, Wen H. 2014. Leaf phenological characters of main tree species in urban forest of Shenyang. *PLOS ONE* 9:e99277
DOI 10.1371/journal.pone.0099277.

- Yamaji K, Julkunen-Tiitto R, Rousi M, Freiwald V, Oksanen E. 2003.** Ozone exposure over two growing seasons alters root-to-shoot ratio and chemical composition of birch (*Betula pendula* Roth). *Global Change Biology* **9**:1363–1377
[DOI 10.1046/j.1365-2486.2003.00669.x](https://doi.org/10.1046/j.1365-2486.2003.00669.x).
- Zhang G, He X, Tang L, Yan K, Chen W, Xu S, Li X. 2009.** Effects of elevated ozone on phenolic substances content and total antioxidative capacity of *Quercus mongolica* leaves. *Chinese Journal of Applied Ecology* **20**:725–728.
- Zhang X, Liu Z, Yu Q, Luc NT, Bing Y, Zhu B, Wang W. 2015.** Effect of petroleum on decomposition of shrub-grass litters in soil in Northern Shaanxi of China. *Journal of Environmental Sciences* **33**:245–253 [DOI 10.1016/j.jes.2014.12.013](https://doi.org/10.1016/j.jes.2014.12.013).
- Zhang X, Zhang H, Yin W, Wang X, Dong B, Sheng H, Feng K, Zhu J. 2011.** Effects of elevated atmospheric O₃ on N, P, and K concentrations in soil and wheat plant during wheat growth season. *Chinese Journal of Ecology* **30**:1637–1641.
- Zheng F, Wang X, Hou P, Zhang W, Lu F, Ouyang Z. 2011.** Influences of elevated ozone on growth and C, N, S allocations of rice. *Acta Ecologica Sinica* **31**:1479–1486.