

9 **ABSTRACT:** Arbuscular mycorrhizal fungi (AMF) played an essential role in complex
10 ecosystems. However, the species diversity and composition of AMF community remain
11 unclear in semi-arid mountain. Further, it is not well understood if the characteristics of AMF
12 community assembly differs for different habitat types, e.g., agricultural arable land, artificial
13 forest land, natural grassland, and bush/wood land. Here, using the high-throughput technology
14 by Illumina sequencing on MiSeq platform, we explored the species diversity and composition
15 of soil AMF communities among different habitat types in a semi-arid mountain (Mid-western
16 region of China). Then we analyzed the effect of nutrient composition and soil texture on AMF
17 community assembly. Our results showed that members of the *Glomus* genera were
18 predominated in all soil types. The distance-based redundancy analysis indicated that the
19 content of water, available phosphorus, and available potassium were the most crucial
20 geochemical factors that significantly affected AMF communities ($p < 0.05$). The analysis of
21 the soil texture confirmed that AMF diversity was negatively correlated with soil clay content.
22 The comparison of AMF diversity among the various habitat types revealed that the artificial
23 forest land had the lowest AMF diversity in comparison with other land types. Our findings
24 suggest that there were differences in species diversity and composition of soil AMF
25 communities among different habitat types. These findings shed new light on the characteristics
26 of community structure and drivers of community assembly in AMF in semi-arid mountains,
27 and point to the potential importance of different habitat types on AMF communities.

28 **Key words:** Illumina sequencing; AMF communities; soil properties; semi-arid field.

29

30 INTRODUCTION

31 Arbuscular mycorrhizal fungi (AMF) play high value for ecosystem restoration and
32 sustainability (Herder *et al.*, 2010; Sanders, 2010; Verbruggen *et al.*, 2010). The majority of
33 land plant species has the potential ability to form symbiotic relationships with AMF, which
34 can significantly enhance plant growth (Lekberg & Koide, 2005), improve soil structure
35 (Piotrowski *et al.*, 2004; Caravaca *et al.*, 2006; Wilson *et al.*, 2009), and contribute to plant
36 resistance to environmental stress (Benjamina, Karl & Johnn, 2009; Balliu, Sallaku & Rewald,
37 2015). And AMF also can maintain ecosystems stability and promote ecosystem development
38 (Larsen, Williams & Kremen, 2005; Fuhrman, 2009; Rosindell, Hubbell & Etienne, 2011).
39 Therefore, to explore the ecological environment in diverse regions, understanding the AMF
40 diversity and biogeography will be of primary importance (Fitter, 2005; Chaudhry *et al.*, 2012).

41 In recent years, lots of studies have reported the AMF community composition in different
42 environmental condition (Öpik *et al.*, 2006; Wubet *et al.*, 2006; Heijden & Scheublin, 2007;
43 Lee, Lee & Young 2008; Krüger *et al.*, 2009). Scholars have argued that the composition of AMF
44 communities will vary along the gradients of land-use intensity under the same climatic
45 conditions and region of agricultural ecosystems (Dumbrell *et al.*, 2010; Fritz *et al.*, 2010;
46 Lekberg *et al.*, 2011; Mirás-Avalos *et al.* 2011, Meadow & Zabinski, 2012). And several papers
47 also confirmed that the AMF distributions are caused by their ability to tolerate high nutrient
48 concentrations in different vegetation soil types (Porrás-Alfaro *et al.*, 2007; Egertonwarburton,
49 Johnson & Allen, 2008; Thomson, Robson & Abbott, 2010). Meanwhile, through the
50 investigation of natural or agricultural habitats, scholars have shown that a high diversity of
51 rhizosphere AMF was found in natural habitat (Öpik *et al.*, 2008; Bonfim *et al.*, 2016), and the
52 AMF communities inhabiting plant roots tended to have a lower diversity in agricultural
53 ecosystems (Daniell *et al.*, 2001; Alguacil *et al.* 2011, Schnoor *et al.* 2011, Bainard *et al.*, 2012).
54 However, most of the previous research works focused on single ecosystems (Helgason *et al.*,
55 1998; Lumini *et al.*, 2010; Verbruggen & Toby, 2012), and there are no comparative analyses
56 on the AMF condition among different soil types under the same climate conditions in semi-
57 arid regions.

58 Hitherto, traditional studies of AMF community composition have been scarce, partly due
59 to the limitations of spore morphological features, which are easily influenced by external
60 disturbances (Oehl *et al.*, 2004), such as integrity of the spores (e.g., ability to identify spores).
61 Due to the above defects, new research technologies are constantly updated. For instance, the
62 development of molecular methods has greatly facilitated the studies of AMF taxonomic and
63 phylogenetic reconstruction and has enhanced the sensitivity of AMF identification and
64 quantification (Lekberg *et al.*, 2007; Helgason & Fitter, 2009; Balestrini *et al.* 2010, Gast *et al.*,

65 2011). Moreover, significant improvements have been made in the analysis of AMF condition
66 by the high-throughput technology (Margulies et al., 2006). The small ribosomal subunit (SSU)
67 has been used to analyze the diversity of AMF. Due to technology advancements, it can provide
68 the most comprehensive reference sequence data set (Öpik et al., 2010), and the sequencing
69 data can provide detailed analyses on AMF communities among complex habitat types (Öpik
70 et al., 2013). In summary, the application of new technologies will greatly improve the study of
71 AMF communities.

72 Thus, our study applied the high-throughput sequencing (Illumina platform) to analyze the
73 soil AMF communities in four habitat types, including agricultural arable land, artificial forest
74 land, natural grassland, and bush/wood land, and in contrast to the first two soil habitat types,
75 the last two types were undisturbed (without human interference). All habitat types were located
76 in the Taihang Mountain, which belongs to the semi-arid ecosystem. We aimed to identify the
77 relative importance of soil characteristics on AMF diversity and illustrate the differences in
78 AMF communities among the predominant four soil types. The research would be a valuable
79 contribution toward a clearer understanding on the way human activities changed the
80 composition of the current AMF communities, and the results will facilitate achieving a more
81 precise guidance on local soil reclamation, vegetation restoration, and the maintenance of
82 biodiversity in semi-arid regions.

83 MATERIALS AND METHODS

84 Study Area

85 The research site was located in the south of Taihang Mountain (112°28'–112°30'E,
86 35°01'–35°03'N), a site which belongs to the semi-arid area of China. The climate in the test
87 area is temperate continental monsoon, with an annual average temperature of 14.3 °C and an
88 average annual sunshine rate of 54%; the elevation gradient of our study sites ranged from 231
89 to 432 m above sea level. Soil in the study area is cinnamon (main part is similar to ustalf
90 USDA), and the parent rock was composed mainly of sandstone and shale. The habitat types in
91 this study were bush/wood land, forest land, grassland, and arable land. The bush/wood land
92 included mainly *Vitex negundo* L., *Lespedeza bicolor* Turcz and *Ziziphus jujuba* Mill. var.
93 *spinosa* (Bunge) Hu ex H. F. Chow, Forest land included mainly *Quercus variabilis* Bl.,
94 *Platycladus orientalis* (L.) Franco, and *Robinia pseudoacacia* L. Dominant herbaceous plants
95 in the grassland were *Setaria viridis* (L.) Beauv., *Artemisia princeps* H. Lév. and Vaniot,
96 *Pennisetum alopecuroides* (L.) Spreng., *Arthraxon hispidus* (Thunb.) Makino, and *Rehmannia*
97 *glutinosa* (Gaetn.) Libosch. ex Fisch. et Mey. Finally, the prevalent herbaceous plants in the
98 arable land were *Zea mays* L., *Triticum aestivum* L., *Ipomoea batatas* L., *Brassica campestris*

Deleted: And t

Deleted: analysis

Deleted: ,

Deleted: and d

Deleted: analysis

104 L., and *Lycopersicon esculentum* Mill.

105 **Sample collection**

106 In October 2016, soil samples were collected in triplicate at four sites (W1, BW, WL, and
107 F). The sample collection occurred at the root zone of the plant at a soil depth of 5–10 cm (Table
108 1). Site W1 represented the forest land soil type; site BW had bush/wood soil type; site WL was
109 characterized by grassland soil type; and arable land soil type was represented in site F. These
110 12 soil samples collected were placed in sterile plastic bags and transported in freezing boxes
111 to the laboratory, and they were stored at -70°C until further analysis.

112 **Soil Geochemical Analyses**

113 We analyzed eight different soil factors, including soil pH, water content, available
114 nitrogen (NH_4^+ -N), available potassium (K^+ -K) and phosphate phosphorus (PO_4^{3-} -P). Soil pH
115 was examined by a pH meter (PX-KS06, Guangzhou Puxi Instrument, Guangzhou, China).
116 Water content was measured by drying soil method. And the content of soil clay, silt, and sand
117 was performed by using a Malvern Mastersizer (Mastersizer2000, Malvern Instruments,
118 Malvern, UK). The available nitrogen and available potassium were analyzed by an
119 Autoanalyzer (SEAL-AA3, SEAL Analytical, Milwaukee, WI, USA); phosphate phosphorus
120 analyzed by NaHCO_3 Mo-Sb colorimetric method.

121 **Molecular analyses DNA extraction**

122 50 mg soil was used for metagenomic DNA extraction in each sample, using the Fast DNA
123 Isolation Kit (Q-BIOgene, Heidelberg, Germany). The extracts were stored at -20°C for PCR.
124 1.0% agarose gels for checking DNA concentration and purity.

125 **Miseq sequencing step**

126 Using the 18S rRNA gene and primer sets of AMV4.5N Forward 5'-
127 AAGCTCGTAGTTGAATTTTCG-3' and AMDG R 5'-CCCAACTATCCCTATTAATCAT-3' to
128 amplify the sequences (from soil DNA extracts), the primer had been reported to be acceptable
129 in several previous studies (*Sato et al., 2005*). The initial PCR reactions were similar to the
130 existing studies of Xiao, including :25 μL total volumes, 1-2 μL DNA template, 250 mM dNTPs,
131 0.25 mM of primer, 1X reaction buffer and 0.5U Phusion DNA Polymerase (*Xiao et al., 2005*).

132 The reactions used a 2720 model Thermal Cycler, and initial PCR amplification was
133 conducted under the steps below: 94°C for 2-min, then 25 cycles of 30-s denaturation at 94°C ,
134 30-s annealing at 56°C , 30-s extension at 72°C , 5-min extension at 72°C . The second step
135 PCR used a template, which come from the first 5 μL product (without dilution). The second

136 step PCR include: one cycle of 3-min at 94 °C, then 8 cycles of 30-s at 94 °C, 56 °C for 30-s
137 and 72 °C for 30-s, and a 5-min extension at 72 °C. The PCR products were separated by
138 electrophoresis (1.5% agarose gel in 0.5 × TBE) and purified using a gel xxtraction kit (Axygen
139 Biosciences, Coming, NY, USA), then the libraries were sequenced by PE300 sequencing on
140 MiSeq v3 Reagent Kit (Illumina) platform (at Tiny Gene Company, Shanghai).

141 **Bioinformatics methods**

142 The sequence reads were analyzed by the combination of software Mothur v. 1.33.3,
143 UPARSE (usearch version v8.1.1756) and R 3.2.2, the original FASTQ files were
144 demultiplexed through the barcode (Schloss *et al.*, 2009). The PE reads for all samples were
145 merged based on mothur. The low quality contigs were removed based on screen.seqs command
146 by the settings filter (maxambig=0, minlength = 200, maxlength =580, the higher threshold can
147 protect some longer sequences, which may be the correct fragment, maxhomop= 8). The
148 decoded data information was aggregated (97% homology) to operational taxonomic units
149 (OTUs) (Edgar, 2013).

Deleted: were

150 BLAST analysis was conducted using the “Nucleotide collection (nr/nt)” database. No
151 threshold was set for E values, alignment length and identity settings. For each OTU
152 representative sequence, a list of top BLAST hits was compiled. Uncultured clones were deleted
153 from the list of top hits. The BLAST getting the highest score was identified as the match’s
154 species.

155 **Statistical Analyses**

156 For the alpha-diversity analysis, Mothur v. 1.33.3 software was used to analyze the OTU
157 richness, Coverage, Chao, and Shannon’s indices as reported earlier by Schloss et al (2009)
158 (Schloss *et al.*, 2009). The values of soil properties and diversity parameters were statistically
159 calculated by SPSS V. 19 software (one-way ANOVA).

Deleted:

Deleted:

160 To identify the AMF relationship in different habitat types, the clustering method based on
161 OTU abundance-based using the R v. 3.1.1. And to identify the AMF associated with different
162 habitat types (agricultural arable land, artificial forest land, natural grassland, and bush/wood
163 land.), we used indicator species analysis approach of Dufrene and Legendre (*Dufrene &*
164 *Legendre P, 1997*).

165 Using the Canoco software (Canoco for Windows 4.5 package) (*Braak & Smilauer, 2002*),
166 we utilized Monte Carlo permutation and distance-based redundancy (db-RDA) tests to explain
167 the correlation between soil AMF and geochemical factors. In addition, the heatmap results of
168 the abundance percentages of AMF genera were obtained by Mothur v. 1.33.3 software. The
169 raw sequence information has been deposited into the NCBI database (Accession No.

Deleted: have

174 SRP116770).

175 RESULTS

176 Soil Properties

177 For the eight geochemical factors measured, the arable land obtained the maximum values
178 of water content, available phosphorus and available potassium (site F). Meanwhile the
179 minimum values of water content and available phosphorus were established in the grassland
180 (site WL). In the bush/wood land (site BW), the maximum values of sand content (average
181 28.9%), but minimum silt content (35.5%) were established (Table 1).

182 AMF Diversity Data and Community Composition

183 In the current study, we have identified a total of 532,841 sequences and 803 OTUs from
184 the total dataset, there were 320,899 sequences belonged to phylum Glomeromycotina
185 (accounting for 60.2%). The number of sequences in each of the samples ranged from 15,095
186 to 35,206, and the number of AMF OTUs ranged from 52 to 83 (genetic distances of 3%). The
187 OTUs' coverage in all soil types reached 99% (Table 2). On the basis of the OTU richness
188 calculated by Chao's index, the grassland observed ~~the~~ greatest AMF value (site WL: 81).
189 Through the analysis of Shannon's index, we discovered that the largest AMF diversity was
190 also present in the grassland (site WL: 3.49–3.52 with an average value of 3.51), followed by
191 the arable land (site F: 3.38–3.46 with an average value of 3.43), bush/wood land (site BW:
192 3.38–3.46 with average 3.42), and the forest land soils (site W1: 2.53–3.15 with an average
193 value of 2.87) (Table 2).

194 Some variations in AMF community composition at the genus level were also detected
195 among all soil samples. The 119 OTUs that could be classified were affiliated with ten AMF
196 genera, whereas those that could not be identified were assigned as unclassified. The *Glomus*
197 were the most abundant genera in all samples: 60%–75% in grassland, 70%–75% in arable land,
198 75%–80% in bush/wood, and 50%–70% in forest land. Meanwhile, their levels varied in the
199 different soil types. *Ambispora* were found in all samples, but a greater abundance was detected
200 in the grassland and arable land samples than in those of the bush/wood and forest land soils
201 (Figure 1).

202 Correlation among the three factors (AMF Communities, Soil Types and Environmental 203 condition)

204 To determine the differences in soil AMF community, the OTU cluster analysis showed
205 that the 12 soil samples were divided into four Soil Types (Figure 2), and the indicator species

Deleted: a

207 analysis revealed that there were 60 AMF indicators (indicator value > 0.25, p < 0.05) in this 4
208 groups types, it mainly included bush/wood (*Glomus* and *Diversispora* taxa), arable land
209 (*Glomus*, *Septoglomus* and *Rhizophagus* taxa), grassland (*Glomus* and *Septoglomus* taxa),
210 forest land (*Glomus* and *Paraglomus* taxa) (Table S1). The top 50 OTUs of all samples were
211 selected and their abundances compared through the heatmap software, which revealed their
212 relative distributions and abundances of the top 50 OTUs in all samples (Figure 3). There is a
213 listing of all AMF OTUs and their closest matches in Table S2.

214 The distance-based redundancy analysis (db-RDA) showed that there was a significant
215 correlation between the combination of the eight environmental factors and soil AMF
216 community structure, and 81.9% of the soil community variation was attributed to the eight
217 environmental factors (Figure 4 and Table 3). Using the Monte Carlo permutation test, we found
218 that water content ($r^2 = 0.7332$, $p < 0.01$), available phosphorus ($r^2 = 0.7576$, $p < 0.01$), available
219 potassium ($r^2 = 0.7973$, $p < 0.01$), silt ($r^2 = 0.6461$, $p < 0.05$), and sand ($r^2 = 0.6293$, $p < 0.05$)
220 were important properties (Table 3).

221 DISCUSSION

222 As mentioned earlier, the study area was located in the South Taihang Mountains of China,
223 whose climate characterizes the region as a typical semi-arid climate zone. Under natural
224 conditions, the thin soil layer, low forest coverage and much gravel are the characteristics of
225 this area. Its forest types were mainly dominated by human intervention of *Quercus variabilis*
226 Bl and *Platycladus orientalis* (L.), and the vegetation was poor and only limited species could
227 be planted (Zhao, 2007). Thus, improving local soil conditions and promoting plant growth are
228 urgent tasks. The fact that AMF community was a crucial factor in plant growth and soil
229 improvement under environmental stress (Oehl et al., 2004; Smith & Read, 2008). However,
230 some information had remained unexplored for the Taihang Mountain area, such as the
231 distribution of AMF communities, the variation of AMF diversity, and the influence of various
232 soil types on AMF composition. Therefore, in this study, we investigated the AMF communities
233 among the predominant four soil types in the South Taihang Mountain region, and the results
234 will provide valuable reference for improving the local ecological environment.

235 By analyzing the results of the 4 different soil types, the research showed that the diversity
236 of AMF communities in undisturbed grassland soil type was greater than that in artificial forest
237 land (Table 2), it was consistent with Öpik et al. (2008), they discovered that rich biological
238 species composition and low external disturbance may lead to higher diversity of rhizosphere
239 AMF of the natural vegetation soil. And results also showed that the value of Shannon's index
240 in arable land was larger than that in artificial forest land. This outcome might have been caused
241 by the cultivation practices implemented by the farmers, including the application of farmyard

Deleted: on

Deleted: on

Deleted: on

Deleted: on

Deleted: ,

Deleted: and i

Deleted: ie

249 manure (food residues, livestock manure, etc.), which increased the number of microbial
250 communities by raising the level of available nutrients (Helgason & Fitter, 2009). Moreover, it
251 is generally accepted that the organic agriculture farming methods are regarded as a useful
252 measure to increase AMF diversity (Aroca, Porcel & Ruizlozano, 2007), and the farmers in
253 study region usually apply farmyard manure with cultivation methods that were closed to those
254 of organic agriculture farming. On the other hand, probably due to the growth and reproduction
255 of specific AMF communities requiring particular host plant species, which leads to a less
256 abundant community under a single artificial plantation habitat (Long et al., 2010). In general,
257 human disturbance caused changes in the forest land environment, which reduced the
258 transportation and distribution of AMF communities (Yuan et al., 2008), and the artificial forest
259 land had the lowest AMF diversity in comparison with other land types.

260 Meanwhile, the results of the sequence data analysis of AMF community composition
261 showed that members of both genera *Ambispora* and *Glomus* existed in different soil types,
262 including forest land, bush/wood, grassland, and arable land. Nevertheless, the representatives
263 of *Glomus* were identified to be the main genus, and *Glomus* taxon served as indicator species
264 for each habitat. These results are similar to ~~previously published research that~~ confirmed that
265 the species of *Glomus* were the most abundant in the AMF assemblage (Oehl et al., 2005).
266 Some researchers also revealed that although *Rhizophagus*, *Ambispora*, and *Glomus* dominated
267 in soils, only *Glomus* was found in almost all samples from the rhizosphere soil (Giovannetti,
268 Azzolini & Citernesi, 1999; Yang et al., 2010). The influence of certain factors may be the reason
269 why *Glomus* was the dominance members in the AMF assemblage among those of other genera.
270 On the one hand, the species of *Glomus* genus can usually produce large numbers of spores and
271 hypha fragments, which can colonize and extensively spread onto the roots of plants (Öpik et
272 al., 2006). And *Glomus* also has a certain resistance in complex environments (Miransari et al.,
273 2008; Bever et al., 2009; Barto et al., 2011). Therefore, these features facilitate the survival and
274 spread of *Glomus* genus members in a semi-arid mountain, and the emergence of this
275 phenomenon is ~~also the result of adaptation to the local ecological environment.~~

276 Moreover, our investigation established that water content is a significant factor which has
277 an obvious effect on the AMF communities. This finding is similar to the results of existing
278 studies (Sieverding, Toro & Mosquera, 1989), which confirmed that the variations in the water
279 content can contribute to changes in the physiological status of local AMF and its ecological
280 niche directly, and water content can also indirectly exert an impact on the utilization of soil
281 nutrients by AMF community. In addition, our research also confirmed that there are significant
282 relationships between the available phosphorus, available potassium, and soil AMF community
283 structure. These interactions may be due to the influence that soil nutrients can have on the

Deleted: some scholars

Deleted: , who

Deleted: aslo

287 growth of local AMF communities as the lack of nutrients inhibits the production and separation
288 of spores (Zaller, Frank & Drapela, 2011). Thus, this work confirmed that environmental
289 factors can drive the composition and distribution of AMF communities.

290 Furthermore, the composition of AMF communities seems to have been strongly
291 influenced by the soil texture distribution, our results showed that the content of silt and sand
292 were significant related to the soil AMF community communities (Table 1), and the AMF
293 diversity was higher in the samples from low-clay but high-sand content soil types. The
294 appearance of the result was probably due to the fact that AMF is an aerobic organism, and the
295 lower clay content provided better aeration, which was advantageous for plant root growth and
296 soil humus decomposition, leading also to accelerated fungal propagation (Torrecillas et al.,
297 2014). The research confirmed that AMF communities was negatively correlated with soil clay
298 content.

299 CONCLUSIONS

300 In conclusion, this study first delineated the species diversity and composition of AMF
301 community in Taihang Mountain, China. The members of the *Glomus* genus were predominant
302 in all soil types. The findings also suggested that nutrient composition and soil texture were the
303 most important factors affecting AMF community. Moreover, there were differences in species
304 diversity and composition of soil AMF communities among different habitat types. These
305 findings shed new light on the characteristics of community structure and drivers of community
306 assembly in AMF in semi-arid mountains, and point to the potential importance of different
307 habitat types on AMF communities.

308 ACKNOWLEDGEMENTS

309 The authors are grateful to the staff of the Xiaolangdi Ecological Station in Henan Province,
310 China, for the provision of the soil materials and testing ground. We also would like to thank
311 the Tiny Gene Bio-Tech (Shanghai) Co., Ltd. for their high-throughput sequence technology.

312 REFERENCES

- 313 **Alguacil MM, Torrecillas E, Caravaca F, Fernández DA, Azcón R, and Roldán A. 2011.** The
314 application of an organic amendment modifies the arbuscular mycorrhizal fungal
315 communities colonizing native seedlings grown in a heavy-metal-polluted soil. *Soil*
316 *Biology & Biochemistry* **43**:1498-1508.
- 317 **Aroca R, Porcel R, and Ruizlozano JM. 2007.** How does arbuscular mycorrhizal symbiosis
318 regulate root hydraulic properties and plasma membrane aquaporins in *Phaseolus vulgaris*
319 under drought, cold or salinity stresses? *New Phytologist* **173**:808-816.
- 320 **Bainard LD, Dai M, Gomez EF, Torres-Arias Y, Bainard JD, Sheng M, Eilers W, and Hamel**
321 **C. 2015.** Arbuscular mycorrhizal fungal communities are influenced by agricultural land
322 use and not soil type among the Chernozem great groups of the Canadian Prairies. *Plant*

323 and *Soil* **387**:351-362.

324 **Balestrini R, Magurno F, Walker C, Lumini E, and Bianciotto V. 2010.** Cohorts of arbuscular
325 mycorrhizal fungi (AMF) in *Vitis vinifera*, a typical Mediterranean fruit crop.
326 *Environmental Microbiology Reports* **2**:594-604.

327 **Balliu A, Sallaku G, and Rewald B. 2015.** AMF Inoculation Enhances Growth and Improves the
328 Nutrient Uptake Rates of Transplanted, Salt-Stressed Tomato Seedlings. *Sustainability*
329 **7**:15967-15981.

330 **Barto EK, Hilker M, Müller F, Mohny BK, Weidenhamer JD, and Rillig MC. 2011.** The Fungal
331 Fast Lane: Common Mycorrhizal Networks Extend Bioactive Zones of Allelochemicals in
332 Soils. *Plos One* **6**:e27195.

333 **Benjamina S, Karl C, and Johnn K. 2009.** Plant and fungal identity determines pathogen
334 protection of plant roots by arbuscular mycorrhizas. *Journal of Ecology* **97**:1274–1280.

335 **Bever JD, Richardson SC, Lawrence BM, Holmes J, and Watson M. 2009.** Preferential
336 allocation to beneficial symbiont with spatial structure maintains mycorrhizal mutualism.
337 *Ecology Letters* **12**:13–21.

338 **Bonfim JA, Vasconcellos RL, Gumiere T, De LCMD, Oehl F, and Nogueira Cardoso EJ. 2016.**
339 Diversity of Arbuscular Mycorrhizal Fungi in a Brazilian Atlantic Forest Toposequence.
340 *Microbial Ecology* **71**:164.

341 **Braak CJFT, and Smilauer P. 2002.** CANOCO Reference Manual and CanoDraw for Windows
342 User's Guide: Software for Canonical Community Ordination (version 4.5). *Ithaca Ny Usa*
343 *Www*.

344 **Caravaca F, Alguacil MM, Azcon R, and Roldan A. 2006.** Formation of stable aggregates in
345 rhizosphere soil of *Juniperus oxycedrus*: Effect of AM fungi and organic amendments.
346 *Applied Soil Ecology* **33**:30-38.

347 **Chaudhry V, Rehman A, Mishra A, Chauhan PS, and Nautiyal CS. 2012.** Changes in Bacterial
348 Community Structure of Agricultural Land Due to Long-Term Organic and Chemical
349 Amendments. *Microbial Ecology* **64**:450-460.

350 **Daniell TJ, Husband R, Fitter AH, and Young JPW. 2001.** Molecular diversity of arbuscular
351 mycorrhizal fungi colonising arable crops. *Fems Microbiology Ecology* **36**:203-209.

352 **Dufrene M, and Legendre P. 1997.** Species Assemblages and Indicator Species: The Need for a
353 Flexible Asymmetrical Approach. *Ecological Monographs* **67**:345-366.

354 **Dumbrell AJ, Nelson M, Helgason T, Dytham C, and Fitter AH. 2010.** Idiosyncrasy and
355 overdominance in the structure of natural communities of arbuscular mycorrhizal fungi: is
356 there a role for stochastic processes? *Journal of Ecology* **98**:419–428.

357 **Edgar RC. 2013.** UPARSE: highly accurate OTU sequences from microbial amplicon reads. *Nature*
358 *Methods* **10**:996.

359 **Egertonwarburton LM, Johnson NC, and Allen EB. 2008.** Mycorrhizal community dynamics
360 following nitrogen fertilization: A cross-site test in five grasslands. *Ecological Monographs*
361 **77**:527-544.

362 **Fitter AH. 2005.** Presidential Address: Darkness Visible: Reflections on Underground Ecology.
363 *Journal of Ecology* **93**:231-243.

- 364 **Fritz O, Endre L, Arno B, Karl S, Robert B, Marcelvander H, and Ewald S. 2010.** Soil type and
 365 land use intensity determine the composition of arbuscular mycorrhizal fungal communities.
 366 *Soil Biology & Biochemistry* **42**:724-738.
- 367 **Fuhrman JA. 2009.** Fuhrman JA.. Microbial community structure and its functional implications.
 368 *Nature* **459**: 193-199. *Nature* **459**:193.
- 369 **Gast CJVD, Gosling P, Tiwari B, and Bending GD. 2011.** Spatial scaling of arbuscular
 370 mycorrhizal fungal diversity is affected by farming practice. *Environmental Microbiology*
 371 **13**:241–249.
- 372 **Giovannetti M, Azzolini D, and Citernesi AS. 1999.** Anastomosis formation and nuclear and
 373 protoplasmic exchange in arbuscular mycorrhizal fungi. *Applied & Environmental*
 374 *Microbiology* **65**:5571-5575.
- 375 **Heijden MGAVD, and Scheublin TR. 2007.** Functional traits in mycorrhizal ecology: their use for
 376 predicting the impact of arbuscular mycorrhizal fungal communities on plant growth and
 377 ecosystem functioning. *New Phytologist* **174**:244-250.
- 378 **Helgason T, Daniell TJ, Husband R, Fitter AH, and Young JPW. 1998.** Ploughing up the wood-
 379 wide web? *Nature* **394**:431. *Nature* **394**:431.
- 380 **Helgason T, and Fitter AH. 2009.** Natural selection and the evolutionary ecology of the arbuscular
 381 mycorrhizal fungi (Phylum Glomeromycota). *Journal of Experimental Botany* **60**:2465.
- 382 **Herder GD, Isterdael GV, Beeckman T, and Smet ID. 2010.** The roots of a new green revolution.
 383 *Trends Plant Sci. Trends in Plant Science* **15**:600-607.
- 384 **Krüger M, Stockinger H, Krüger C, and Schübler A. 2009.** DNA- based species level detection
 385 of Glomeromycota: one PCR primer set for all arbuscular mycorrhizal fungi. *New*
 386 *Phytologist* **183**:212-223.
- 387 **Larsen TH, Williams NM, and Kremen C. 2005.** Extinction order and altered community structure
 388 rapidly disrupt ecosystem functioning. *Ecology Letters* **8**:538-547.
- 389 **Lee J, Lee S, and Young JPW. 2008.** Improved PCR primers for the detection and identification of
 390 arbuscular mycorrhizal fungi. *Fems Microbiology Ecology* **65**:339-349.
- 391 **Lekberg Y, and Koide RT. 2005.** Is plant performance limited by abundance of arbuscular
 392 mycorrhizal fungi? A meta-analysis of studies published between 1988 and 2003.
 393 **168**:189-204.
- 394 **Lekberg Y, Koide RT, Rohr JR, Aldrich-Wolfe L, and Morton JB. 2007.** Role of Niche
 395 Restrictions and Dispersal in the Composition of Arbuscular Mycorrhizal Fungal
 396 Communities. *Journal of Ecology* **95**:95-105.
- 397 **Lekberg Y, Meadow J, Rohr JR, Redecker D, and Zabinski CA. 2011.** Importance of dispersal
 398 and thermal environment for mycorrhizal communities: lessons from Yellowstone National
 399 Park. *Ecology* **92**:1292-1302.
- 400 **Long LK, Yao Q, Guo J, Yang RH, Huang YH, and Zhu HH. 2010.** Molecular community
 401 analysis of arbuscular mycorrhizal fungi associated with five selected plant species from
 402 heavy metal polluted soils. *European Journal of Soil Biology* **46**:288-294.
- 403 **Lumini E, Orgiazzi A, Borriello R, Bonfante P, Bianciotto V, Bonfante P, Visick K, and**
 404 **Ohkuma M. 2010.** Disclosing arbuscular mycorrhizal fungal biodiversity in soil through

405 a land-use gradient using a pyrosequencing approach. *Environmental Microbiology*
406 **12**:2165-2179.

407 **Margulies M, Egholm M, Altman WE, Attiya S, Bader JS, Bemben LA, Berka J, Braverman**
408 **MS, Chen YJ, and Chen Z. 2006.** Genome sequencing in microfabricated high-density
409 picolitre reactors. *Nature* **437**:376-380.

410 **Meadow JF, and Zabinski CA. 2012.** Linking symbiont community structures in a model
411 arbuscular mycorrhizal system. *New Phytologist* **194**:800-809.

412 **Mirás-Avalos JM, Antunes PM, Koch A, Khosla K, Klironomos JN, and Dunfield KE. 2011.**
413 The influence of tillage on the structure of rhizosphere and root-associated arbuscular
414 mycorrhizal fungal communities. *Pedobiologia* **54**:235-241.

415 **Miransari M, Bahrami HA, Rejali F, and Malakouti MJ. 2008.** Using arbuscular mycorrhiza to
416 alleviate the stress of soil compaction on wheat (*Triticum aestivum* L.) growth. *Soil*
417 *Biology & Biochemistry* **40**:1197-1206.

418 **Oehl F, Sieverding E, Ineichen K, Ris EA, Boller T, and Wiemken A. 2005.** Community structure
419 of arbuscular mycorrhizal fungi at different soil depths in extensively and intensively
420 managed agroecosystems. *New Phytologist* **165**:273–283.

421 **Oehl F, Sieverding E, Mäder P, Dubois D, Ineichen K, Boller T, and Wiemken A. 2004.** Impact
422 of long-term conventional and organic farming on the diversity of arbuscular mycorrhizal
423 fungi. *Oecologia* **138**:574-583.

424 **ÖPIK M, Moora M, Liira J, and Zobel M. 2006.** Composition of root- colonizing arbuscular
425 mycorrhizal fungal communities in different ecosystems around the globe. *Journal of*
426 *Ecology* **94**:778-790.

427 **Opik M, Moora M, Zobel M, Saks U, Wheatley R, Wright F, and Daniell T. 2008.** High diversity
428 of arbuscular mycorrhizal fungi in a boreal herb-rich coniferous forest. *New Phytologist*
429 **179**:867–876.

430 **Opik M, Vanatoa A, Vanatoa E, Moora M, Davison J, Kalwij JM, Reier U, and Zobel M. 2010.**
431 The online database MaarjAM reveals global and ecosystemic distribution patterns in
432 arbuscular mycorrhizal fungi (Glomeromycota). *New Phytologist* **188**:223.

433 **Öpik M, Zobel M, Cantero JJ, Davison J, Facelli JM, Hiiesalu I, Jairus T, Kalwij JM, Koorem**
434 **K, and Leal ME. 2013.** Global sampling of plant roots expands the described molecular
435 diversity of arbuscular mycorrhizal fungi. *Mycorrhiza* **23**:411-430.

436 **Piotrowski JS, Denich T, Klironomos JN, Graham JM, and Rillig MC. 2004.** The effects of
437 arbuscular mycorrhizas on soil aggregation depend on the interaction between plant and
438 fungal species. *New Phytologist* **164**:365–373.

439 **Porrás-Alfaro A, Herrera J, Natvig DO, and Sinsabaugh RL. 2007.** Effect of long-term nitrogen
440 fertilization on mycorrhizal fungi associated with a dominant grass in a semiarid grassland.
441 *Plant and Soil* **296**:65-75.

442 **Rosindell J, Hubbell SP, and Etienne RS. 2011.** The unified neutral theory of biodiversity and
443 biogeography at age ten. *Ecology* **26**:340-348.

444 **Sanders IR. 2010.** [Designer] mycorrhizas: Using natural genetic variation
445 in AM fungi to increase plant growth. *Journal of Cultural Studies* **3**:1081-1083.

- 446 **Sato K, Suyama Y, Saito M, and Sugawara K. 2005.** A new primer for discrimination of
 447 arbuscular mycorrhizal fungi with polymerase chain reaction-denature gradient gel
 448 electrophoresis. *Grassland Science* **51**:179–181.
- 449 **Schloss PD, Westcott SL, Ryabin T, Hall JR, Hartmann M, Hollister EB, Lesniewski RA,**
 450 **Oakley BB, Parks DH, and Robinson CJ. 2009.** Introducing mothur: open-source,
 451 platform-independent, community-supported software for describing and comparing
 452 microbial communities. *Appl Environ Microbiol* **75**:7537-7541.
- 453 **Schnoor TK, Lekberg Y, Rosendahl S, and Olsson PA. 2011.** Mechanical soil disturbance as a
 454 determinant of arbuscular mycorrhizal fungal communities in semi-natural grassland.
 455 *Mycorrhiza* **21**:211-220.
- 456 **Sieverding E, Toro S, and Mosquera O. 1989.** Biomass production and nutrient concentrations in
 457 spores of va mycorrhizal fungi. *Soil Biology & Biochemistry* **21**:69-72.
- 458 **Smith SE, and Read D. 2008.** 5–Mineral nutrition, toxic element accumulation and water relations
 459 of arbuscular mycorrhizal plants. *Mycorrhizal Symbiosis*:145-187.
- 460 **Thomson BD, Robson AD, and Abbott LK. 2010.** Effects of phosphorus on the formation of
 461 mycorrhizas by *gigaspora calospora* and *glomus fasciculatum* in relation to root
 462 carbohydrates. *New Phytologist* **103**:751-765.
- 463 **Torrecillas E, Alguacil MDM, Roldán A, Díaz G, Montesinosnavarro A, and Torres MP. 2014.**
 464 Modularity Reveals the Tendency of Arbuscular Mycorrhizal Fungi To Interact Differently
 465 with Generalist and Specialist Plant Species in Gypsum Soils. *Applied & Environmental*
 466 *Microbiology* **80**:5455-5466.
- 467 **Verbruggen E, and Toby KE. 2010.** Evolutionary ecology of mycorrhizal functional diversity in
 468 agricultural systems. *Evolutionary Applications* **3**:547-560.
- 469 **Verbruggen E, Van DH, MARCEL G. A, Weedon JT, Kowalchuk GA, and Rölting WFM. 2012.**
 470 Community assembly, species richness and nestedness of arbuscular mycorrhizal fungi in
 471 agricultural soils. *Molecular Ecology* **21**:2341.
- 472 **Wilson GW, Rice CW, Rillig MC, Springer A, and Hartnett DC. 2009.** Soil aggregation and
 473 carbon sequestration are tightly correlated with the abundance of arbuscular mycorrhizal
 474 fungi: results from long-term field experiments. *Ecology Letters* **12**:452-461.
- 475 **Wubet T, Weiß M, Kottke I, Teketay D, and Oberwinkler F. 2006.** Phylogenetic analysis of
 476 nuclear small subunit rDNA sequences suggests that the endangered African Pencil Cedar,
 477 *Juniperus procera* , is associated with distinct members of Glomeraceae. *Mycological*
 478 *Research* **110**:1059-1069.
- 479 **Xiao E, Krumins V, Song T, Xiao T, Ning Z, Lan X, and Sun W. 2016.** Correlating microbial
 480 community profiles with geochemical conditions in a watershed heavily contaminated by
 481 an antimony tailing pond ☆. *Environmental Pollution* **215**:141-153.
- 482 **Yang R, Zan ST, Tang JJ, Xin C, and Qian Z. 2010.** Variation in community structure of
 483 arbuscular mycorrhizal fungi associated with a Cu tolerant plant - *Elsholtzia splendens*.
 484 *Applied Soil Ecology* **44**:191-197.
- 485 **Yuan YW, Vestberg M, Walker C, Hurme T, Zhang X, and Lindström K. 2008.** Diversity and
 486 infectivity of arbuscular mycorrhizal fungi in agricultural soils of the Sichuan Province of

487 mainland China. *Mycorrhiza* **18**:59-68.
488 **Zaller JG, Frank T, and Drapela T. 2011.** Soil sand content can alter effects of different taxa of
489 mycorrhizal fungi on plant biomass production of grassland species. *European Journal of*
490 *Soil Biology* **47**:175.
491 **Zhao Y. 2007.** Analysis and evaluation on degraded ecosystem ecological characterizes of
492 vegetation restoration process in the hill areas of the Taihang Moutains. (Doctoral
493 dissertation) *Henan Agricultural University*.(in Chinese)
494

495 **Figures:**

496 Figure 1. Abundance percentages of AMF genera for all soil samples.

497 Figure 2. Clustering analysis of AMF communities based on OTU abundance for each soil.

498 Figure 3. Heat map of top 50 OTUs in all samples. The color intensity (log scale) in each panel

499 shows the percentage of a genus in a sample, referring to color key at the bottom.

500 Figure 4. Distance-based redundancy (db-RDA) tests used to interpret the correlations between

501 the AMF communities and environmental properties.

502

Tables:

Table 1. Geochemical characteristics of the soil samples and other information of the site of the present study.

Table 2. The results of data in the present study.

Table 3. Monte Carlo permutation tests were used to detect the relationship between community composition and soil variables.