

Ginsenoside Rk1 bioactivity: a systematic review

Abdelrahman Elshafay¹, Ngo Xuan Tinh², Samar Salman³, Yara Saber Shaheen⁴, Eman Bashir Othman⁵, Mohamed Tamer Elhady⁶, Aswin Ratna Kansakar⁷, Linh Tran⁸, Le Van², Kenji Hirayama⁹ and Nguyen Tien Huy^{10,11}

- ¹ Faculty of Medicine, Al-Azhar University, Cairo, Egypt
- ² Faculty of Pharmacy, University of Medicine and Pharmacy, Ho Chi Minh city, Vietnam
- ³ Tanta University Hospital, Tanta, Egypt
- ⁴ Faculty of Medicine, Cairo University, Cairo, Egypt
- ⁵ Department of Medicine, Tripoli Central Hospital, Tripoli, Libya
- ⁶ Department of Pediatrics, Zagazig University Hospitals, Sharkia, Egypt
- ⁷ Dirghayu Guru Hospital and Research Center, Kathmandu, Nepal
- ⁸ Institute of Research and Development, Duy Tan University, Da Nang, Vietnam
- ⁹ Department of Immunogenetics, Institute of Tropical Medicine (NEKKEN), Graduate School of Biomedical Sciences, Nagasaki University, Nagasaki, Japan
- ¹⁰ Evidence Based Medicine Research Group & Faculty of Applied Sciences, Ton Duc Thang University, Ho Chi Minh City, Vietnam
- ¹¹ Department of Clinical Product Development, Institute of Tropical Medicine (NEKKEN), Leading Graduate School Program, and Graduate School of Biomedical Sciences, Nagasaki University, Nagasaki, Japan

ABSTRACT

Ginsenoside Rk1 (G-Rk1) is a unique component created by processing the ginseng plant (mainly Sung Ginseng (SG)) at high temperatures. The aim of our study was to systematically review the pharmacological effects of G-Rk1. We utilized and manually searched eight databases to select in vivo and in vitro original studies that provided information about biological, pharmaceutical effects of G-Rk1 and were published up to July 2017 with no restriction on language or study design. Out of the 156 papers identified, we retrieved 28 eligible papers in the first skimming phase of research. Several articles largely described the G-Rk1 anti-cancer activity investigating "cell viability", "cell proliferation inhibition", "apoptotic activity", and "effects of G-Rk1 on G1 phase and autophagy in tumor cells" either alone or in combination with G-Rg5. Others proved that it has antiplatelet aggregation activities, anti-inflammatory effects, anti-insulin resistance, nephroprotective effect, antimicrobial effect, cognitive function enhancement, lipid accumulation reduction and prevents osteoporosis. In conclusion, G-Rk1 has a significant anti-tumor effect on liver cancer, melanoma, lung cancer, cervical cancer, colon cancer, pancreatic cancer, gastric cancer, and breast adenocarcinoma against in vitro cell lines. In vivo experiments are further warranted to confirm these effects.

Subjects Plant Science, Evidence Based Medicine, Pharmacology **Keywords** Ginsenoside, Systematic review, Rk1, Clinical pharmacology

INTRODUCTION

Ginseng is commonly known as a medicinal herb that is obtained from the roots of genus *Panax* (*Shin*, *Kwon & Park*, *2015*). Ginseng belongs to one of the most ancient herbs

Submitted 5 June 2017 Accepted 16 October 2017 Published 17 November 2017

Corresponding author Nguyen Tien Huy, nguyentienhuy@tdt.edu.vn

Academic editor Doil Choi

Additional Information and Declarations can be found on page 21

DOI 10.7717/peerj.3993

© Copyright 2017 Elshafay et al.

Distributed under Creative Commons CC-BY 4.0

OPEN ACCESS

in traditional medicine and is still widely used today (*Choi et al., 2013*). Ginsenosides are classified based on the steroidal structure and the number of hydroxyl groups/sugar moieties attached to it, such as protopanaxadiol, protopanaxatriol, oleanolic acid (or aglycone oleanolic acid) and ocotillol (*Nag et al., 2015*). The protopanaxadiol group includes Rb1, Rb2, Rb3, Rc, Rd, Rg3, Rh2, Rs1, and Rk1. The protopanaxatriol group includes Re, Rf, Rg1, Rg2, and Rh1 (*Kim, Kim & Shin, 2013*). Ro is classified as an oleanolic acid group (*Tachikawa et al., 1999*). Details of types of ginsenosides are presented in Fig. 1.

The quality and composition of ginsenosides in the ginseng plant are affected by a range of factors such as species, age, part of the plant itself, method of cultivation, harvesting season and preservation methods (*Lim, Mudge & Vermeylen, 2005*; *Schlag & McIntosh, 2006*). Some of the ginsenosides, e.g., Rk1, Rg3, Rg5, F4, are isolated from the heat-processed ginseng, Sun ginseng (SG), but are not detected in raw or air-dried ginseng (*Kim et al., 2000*).

Ginsenosides are widely known to have many pharmacological activities (*Choi*, 2008; *Ernst*, 2010) such as anti-tumor, anti-inflammatory (*Chen et al.*, 2007), anti-fatigue (*Tang et al.*, 2008) and analgesic effects (*Nemmani & Ramarao*, 2003).

Ginseng plant is commonly harvested after four to six years of cultivation and is divided in three types based on the processing methods: (1) fresh ginseng which is less than four years old, (2) white ginseng from four to six years and is oven dried after peeling, (3) red ginseng which is six years and steamed before drying. These processing methods aim to improve the efficacy, safety, and preservation (*Yun*, 2001). SG was recently developed by heat-treatment at high temperature and pressure, which were higher than those applied to the conventional preparation of red ginseng.

SG has shown higher concentrations of less polar ginsenosides, which were either entirely absent or present in trace amounts in conventional red ginseng (*Keum et al.*, 2000; *Kwon et al.*, 2001).

The ginsenoside Rk1 (G-Rk1) is one of the main elements of SG (*Kim et al.*, 2008). Various studies confirmed the anti-cancer effects of G-Rk1 on several neoplastic such as hepatocellular carcinoma and melanoma (*Kim et al.*, 2012; *Kim et al.*, 2008). In recent studies, G-Rk1 was confirmed as a new endothelial barrier enhancer, which is capable of preventing or even blocking the vascular endothelial growth factor (VEGF)-induced vasopermeability in the endothelial cells. This presents the potential of developing pharmaceuticals that may effectively control pathologic vascular leakages (*Maeng et al.*, 2013). Therefore, we aimed to systematically review the bioactivities of G-Rk1 in both human and animals.

METHODS

Protocol and registration

The Preferred Reporting Items for Systematic Reviews and Meta-Analysis (PRISMA) Checklist (*Moher et al.*, 2009) was followed in this systematic review. Our protocol was registered at PROSPERO CRD42016029129 in January 2016.

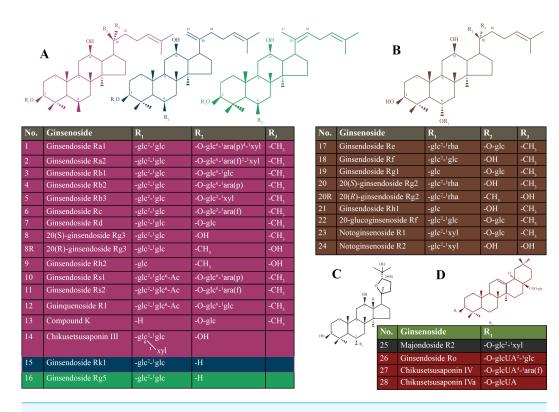


Figure 1 Chemical structure of the ginsenosides types. (A) protopanaxadiol (PPD)-type ginsneosides including Rk1 represented in blue color, Rg5 represented in green color, and the rest of PPD-types are in violet; (B) protopanaxatriol (PPT)-type ginsneosides represented in brown color; (C) Ocotillol- type ginsneoside is represented in gray color; (D) Oleanic acid-type ginsneosides are represented in red color. glc, b-D-glucose; rha, a-L-rhamnose; arap, a-L-arabinose (pyranose); araf, a-L-arabinose (furanose).

Full-size DOI: 10.7717/peerj.3993/fig-1

Eligibility criteria

We selected only original studies published up to July 2017 that provided information about the biological and pharmaceutical effects of G-Rk1. We included articles with G-Rk1 biological effects on human and animals either *in vivo* or *in vitro* with no restriction regarding publication language, publication date, or study design.

We excluded three main types of studies which are: (1) Studies with unreliable extracted data or overlapping data set; (2) studies with only abstract available or no full-text available; (3) books, reviews, meta-analysis studies, conference papers, and thesis. Any disagreement was discussed carefully among three reviewers to get a final decision.

Information sources and search strategies

We conducted electronic searches using eight databases which include: PubMed, Scopus, ISI Web of Science, Google Scholar, SIGLE (System for Information on Grey Literature in Europe), Virtual Health Library (VHL), World Health Organization Global Health Library (GHL), and POPLINE. A Manual search using reference lists of studies was performed to find more relevant studies. The search strategy was performed by (AE, NXT, SS, YSS, EBO, MTE, ARK) and more information on search strategy was provided in Table S1.

Study selection

We selected articles in two phases: (1) Title and abstract screening of all searched articles; (2) full-text screening. The articles which were not in agreement with our inclusion and exclusion criteria were excluded. Three independent reviewers completed these two selecting phases. When disagreement occurred, a consensus decision was made following a discussion with supervisor (NTH).

Data collection process and data items

We prepared our primary extraction form, extracted three papers with it one by one, modified our form after each paper extraction and finally developed the extraction sheet that we used in the remaining articles. Three independent reviewers extracted the data from each paper. A discussion among the three reviewers was held to reach a consensus whenever there was a disagreement in any information retrieved. If three reviewers could not come to an agreement, the supervisor (NTH) was consulted.

The extracted data items included the last name of the first author, year of publication, year of subject recruitment, journal name, study design, country and city of origin of cell lines, the name of the plant, and method of extraction of our targeted material (G-Rk1). If the study included animals, we extracted their species, sex, age, and weight. If it had been done *in vivo*, we extracted the name of the cell line, its origin, the main medium used in terms of either primary (isolated by authors) or commercial cell lines. Also, we extracted the name of the measured parameter, an assay for its measurement, time effect, administration time, active substance name, its concentration, mean, standard deviation, standard error, a *P* value of results and the statistical test. When the data was presented as graphs, we used Web blot digitizer software, and the average of the results from three reviewers was calculated to obtain one result.

Risk of bias in individual studies

Two independent reviewers assessed all of the selected studies according to the GRADE method (*Guyatt et al.*, 2011) to judge the quality of evidence, and any disagreement was resolved by discussion between them. Items such as limitation, inconsistency, indirectness, imprecision, publication bias, and moderate or large effect size were to be scored as "1" if there is no serious limitation or "0" if there is a serious limitation that has been defined according to GRADE criteria. Then the overall quality was to be scored as "high", "moderate", "low", or "very low" quality, according to their analysis of each study. The supervisor (NTH) was consulted when a disagreement occurs.

Summary measures

Inhibition of cell proliferation, apoptosis, and regulation of protein expression were the main evaluated outcomes.

RESULTS

Study selection

We identified 317 citations using the search strategy. From these, we included 156 articles after removing the duplicates. After that, we examined the title, abstract and further

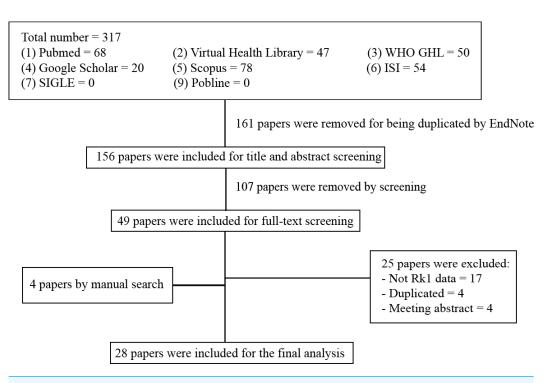


Figure 2 Flowchart of our systematic review - summary of how the systematic search was conducted and eligible studies were identified (PRISMA flow diagram). PRISMA, Preferred Reporting Items for Systematic reviews and Meta-Analyses.

Full-size DOI: 10.7717/peerj.3993/fig-2

excluded 107 articles. We retrieved and evaluated the full-text of the remaining 49 articles, of which 25 articles were excluded, leaving 24 articles that were eligible, in addition to four articles that were retrieved from manually searching the included references. A flowchart described in details the process of identification, inclusion, and exclusion of articles was presented in Fig. 2.

Study characteristics

Out of the 28 studies included, 21 studies were related to the effectiveness of G-Rk1 only and seven studies were reported on the combined effects of G-Rk1 and G-Rg5. The most common study design was *in vitro* study with 22 studies (*Ahn et al.*, 2016; *Ju et al.*, 2012; *Kang et al.*, 2007; *Kang et al.*, 2006; *Kim et al.*, 2012; *Kim et al.*, 2009; *Kim et al.*, 2008; *Kim et al.*, 2013b; *Ko et al.*, 2009; *Kwak & Pyo*, 2016; *Lee et al.*, 2009; *Lee et al.*, 2010; *Lee*, 2014; *Lim et al.*, 2009; *Liu et al.*, 2007; *Park et al.*, 2002; *Ponnuraj et al.*, 2014; *Quan et al.*, 2015; *Ryu et al.*, 2016; *Siddiqi et al.*, 2014; *Toh et al.*, 2011; *Xue et al.*, 2017), while *in vivo* study was less common with only two studies (*Jing et al.*, 2006; *Kim et al.*, 2010). The remaining four articles were both *in vitro* and *in vivo* studies (*Bao et al.*, 2005; *Hu et al.*, 2017; *Maeng et al.*, 2013; *Park et al.*, 2015). A summary of the included studies was presented in Table 1. For G-Rk1, bioactivities and mechanism of actions were summarized in Fig. 3.

Table 1 Study characteristics of included articles.

Author and year	Country	Study design	Cell lines	Parameter assessed
Ko et al. (2009)	South Korea	In vitro	HepG2	Cell viability, cell proliferation, inhibitory activity (IC50)
Lee (2014)	South Korea	In vitro	HepG2	Cell viability
Toh et al. (2011)	Singapore	In vitro	SNU449 (CRL-2234), SNU182 (CRL-2235) and HepG2 (HB-8065)	Cell viability, cell proliferation
Kim et al. (2008)	South Korea	In vitro	HepG2	Cell viability, telomerase activity
Park et al. (2002)	South Korea	In vitro	SK-Hep-1 cells	Cell viability
Lim et al. (2009)	South Korea	In vitro	Junctional proteins (zo-1, occludin and plakoglobin)	ND
Kim et al. (2009)	South Korea	In vitro	3T3-L1 fibroblast cells	Cell viability, lipid accumulation
Kim et al. (2012)	South Korea	In vitro	SK-MEL-2 human melanoma	Cell viability
Ju et al. (2012)	South Korea	In vitro	Platelet	Antiplatelet aggregation activity
Liu et al. (2007)	France	In vitro	Embryonic neural stem cells (neurospheres)	Neurogenic activity
Lee et al. (2009)	South Korea	In vitro	Platelet	Collagen (3–4 μg/L) induced platelet aggregation
Kim et al. (2010)	South Korea	In vivo	ND	TPA-induced mouse ear edema
Maeng et al. (2013)	South Korea	In vitro and In vivo	HREC cells	VEGF-induced retinal endothelial permeability, VEGF-induced destabilization of TJ protein ZO-1, ZO-2 and occludin in membrane and cytosol
Kang et al. (2007)	Japan	In vitro	ND	The OH scavenging inhibition
Kang et al. (2006)	Japan	In vitro	ND	The OH scavenging activities
Lee et al. (2010)	South Korea	In vitro	HUVECs	Cell viability
Kim et al. (2013a)	South Korea	In vitro	Gastric cancer AGS cell	Cell viability, the anticancer activity of ginsenosides after heat processing (IC50)
Bao et al. (2005)	South Korea	<i>In vitro</i> and <i>In vivo</i>	Cortical cell cultures containing neuronal and non-neuronal cells	Cognitive performance, excitotoxicity induced by NMDA and glutamate
Park et al. (2015)	South Korea	<i>In vitro</i> and <i>In vivo</i>	LLC-PK1 cells	Cell viability
Siddiqi et al. (2014)	South Korea	In vitro	The murine cell line, MC3T3-E1	Cell viability, mineralization, ALP activity, collagen and glutathione
Jing et al. (2006)	China	In vivo	ND	Cognitive performance
Ponnuraj et al. (2014)	South Korea	In vitro	3T3-L1 cells	Cell viability, glucose utilization
Ahn et al. (2016)	South Korea	In vitro	HaCaT/RAW 264.7	Anti-inflammation activity
Hu et al. (2017)	China	In vivo	ND	Anti-inflammation activity
Kwak & Pyo (2016)	South Korea	In vitro	A549 cell	Cell viability
Quan et al. (2015)	China	In vitro	A549, HCT-116, HepG2, Hela, MCF-7, and PANC-1 cells	Cell viability
Xue et al. (2017)	China	In vitro	ND	Antimicrobial activity
Ryu et al. (2016)	South Korea	In vitro	ND	Anti-oxidant activity

Notes.

A549 cell, human lung carcinoma; AMA, antimycin A; ALP, alkaline phosphatase; HaCaT, human keratinocyte cell line; HCT-116, human colon carcinoma; Hela, human cervical carcinoma; HepG2, human hepatocellular carcinoma cells; HUVEC, human umbilical vein endothelial cell; HRECs, primary human retina microvascular endothelial cells; LLC-PK1, (pig kidney epithelium, CL-101); MCF-7, human breast adenocarcinoma; NMDA, *N*-methyl-D-aspartate; ND, not defined; PANC-1, human pancreatic cancer; SNU449, SNU182, human liver cancer cell lines; RAW 264.7, the murine macrophage cell line; VEGF, vascular endothelial growth factor; TPA, 12-O-Tetradecanoyl-phorbol-13-acetate; TJ, tight junctions.

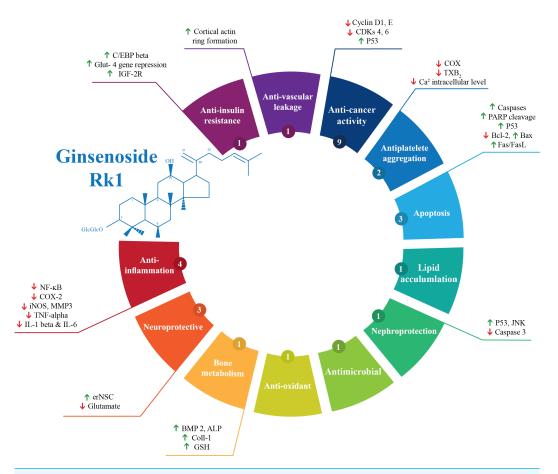


Figure 3 Summary of G-Rk1 bioactivities and its mechanism of actions—ALP, alkaline phosphatase; Bax, BCL2-Associated X Protein; Bcl-2, B-cell lymphoma 2; BMP 2, bone morphogenetic protein-2; COX-2, Cyclooxygenase 2; CDK, cyclin-dependent kinase; erNSC, Epidermal growth factor-responsive neurosphere stem cells; GSH, Glutathione; GLUT-4, Glucose Transporter; IL, interleukin; iNOS, Inducible Nitric Oxide Synthase; IGF, insulin-like growth factor receptor; JNK, Jun N-terminal Kinase; MMP3, Matrix Metalloproteinase 3; NF-kB, Nuclear Factor Kappa B; PARP, Poly ADP (Adenosine Diphosphate)-Ribose Polymerase; TXB-2, Thromboxane B2; TNF-α, tumor necrosis factor. *The number in the small circle indicates the number of studies that report this bioactivity.

Risk of bias across studies

We used the GRADE method (*Guyatt et al.*, 2011) to assess the quality of the included studies. Sixteen studies were categorized as high quality (*Hu et al.*, 2017; *Kang et al.*, 2006; *Kim et al.*, 2009; *Kim et al.*, 2008; *Ko et al.*, 2009; *Kwak & Pyo*, 2016; *Lee et al.*, 2009; *Lee*, 2014; *Lim et al.*, 2009; *Liu et al.*, 2007; *Maeng et al.*, 2013; *Park et al.*, 2002; *Quan et al.*, 2015; *Ryu et al.*, 2016; *Toh et al.*, 2011; *Xue et al.*, 2017). Twelve studies (*Ahn et al.*, 2016; *Bao et al.*, 2005; *Jing et al.*, 2006; *Ju et al.*, 2012; *Kang et al.*, 2007; *Kim et al.*, 2012; *Kim et al.*, 2010; *Kim et al.*, 2013b; *Lee et al.*, 2010; *Park et al.*, 2015; *Ponnuraj et al.*, 2014; *Siddiqi et al.*, 2005; *Jing et al.*, 2006; *Kim et al.*, 2013b; *Park et al.*, 2015; *Ponnuraj et al.*, 2014; *Siddiqi et al.*, 2014) focused on the effectiveness of combined (G-Rk1, G-Rg5), thus, they were

Full-size DOI: 10.7717/peerj.3993/fig-3

downgraded in indirectness item of GRADE factors. Three studies (*Kang et al.*, 2007; *Kim et al.*, 2010; *Lee et al.*, 2010) were not completely pertained to our main outcome since they did not concern mainly with G-Rk1. Four studies (*Kang et al.*, 2006; *Lim et al.*, 2009; *Liu et al.*, 2007; *Park et al.*, 2002) were not downgraded in spite of having insufficient data regarding dose-effect factor as this factor does not belong to the downgraded factors of GRADE method that includes (limitation, inconsistency, indirectness, and imprecision). In contrast, one study (*Lee et al.*, 2010) that was downgraded since it focused on combined (G-Rk1 and G-Rg5) not because of dose effect insufficient data. Another study (*Kim et al.*, 2012) was downgraded because it was used to compare the anti-tumor activity of G-Rk1 versus G-Rk3. However, G-Rk3 has been proven to have a potential antitumor activity. One study (*Ju et al.*, 2012) was downgraded as it has statistical typing mistake of one of its values (Table S2).

Synthesis of results Anti-cancer activity

Cell viability was measured by different assays through the studies including four studies used Cell Counting Kit-8 (CCK8) assay (*Kim et al.*, 2012; *Kim et al.*, 2008; *Kim et al.*, 2013b; *Ko et al.*, 2009), five studies used 3-(4,5-dimethyl-thiazol-2yl) -2,5-diphenyl tetrazolium bromide (MTT) assay (*Kwak & Pyo*, 2016; *Lee et al.*, 2010; *Park et al.*, 2002; *Quan et al.*, 2015; *Siddiqi et al.*, 2014), and one study that used WST-1 assay (*Toh et al.*, 2011) while the final one used EZ-CytoTox cell assay kit (*Park et al.*, 2015) (Table 2).

Liver cancer. Toh et al. (2011) evaluated the inhibitory effects of G-Rk1 (0.25 μg/ml) on cell growth of liver cancer cell lines (human hepatocellular carcinoma cells (HepG2), SNU449, and SNU182). A significant reduction of cell viability caused by G-Rk1 at 0.25 mg/ml was recorded (p < 0.001). The inhibition concentration (IC₅₀) value of G-Rk1 for inhibiting growth in the SNU449 cell line for 48 h was evaluated 0.08 mg/ml (100 μM) by using the WST-1 assay. These results indicated that G-Rk1 is one of the most anti-proliferative ginsenosides of raw and steamed P. notoginseng. Similarly, *Quan et al.* (2015) revealed that the HepG2 cell viability was reduced to 23% and 15% compared to the vehicle control when treated with G-Rk1 at 40 μM and 80 μM for 24 h, respectively (*Quan et al.*, 2015).

Ko et al. (2009) evaluated the effect of G-Rk1 on cell viability of HepG2 cells after 24 h incubation in concentrations of 50, 75, 100 μM in the presence of 0.1 μM taxol which was used as a positive control. Compared with the vehicle control, G-Rk1 (at a dose of 100 μM) inhibited HepG2 cell proliferation by about 40%. When HepG2 cells were exposed to various concentrations of G-RK1 for 24 h (from 50 to 100 μM), the inhibitory effect on growth rate raised significantly, from 8 to 37.5%, in a dose-dependent manner. In addition, the cell viability was also tested when bafilomycin A1 was added to G-Rk1 (100 μM) and then, three independent experiments showed that this co-treatment enhanced HepG2 cell death more than the cells that were treated with 100 μM of G-Rk1 alone. In this experiment, to verify the effects of this combination and exclude cytotoxicity of bafilomycin A1, cytotoxicity was measured after 24 h and no cytotoxicity was detected.

In the study of *Kim et al.* (2008) they assessed the effects of G-Rk1 on cell viability of HepG2 cells. The concentrations of G-Rk1 ranging from 12.5 to 100 μ M with 0.5

Table 2 Summary of anti-cancer activity of G-Rk1.

Author, Year	Cells' origin	Methods/ Cell lines	Methods/ Assays	Methods/ Time effect		Methods/ Treatment					
						Positive control			Active compoun		
					Substance	Concentration	Activity (%)	Substance	Concentration	Activity (%)	
Cell viability	inhibition ^a										
Ko et al. (2009)	South Korea	HepG2	CCK-8 assay	24 h	Taxol	0.1 μΜ	100	G-Rk1	50, 75, 100 μΜ	92, 70, 62.5	Inhibition of cell viability in a dose dependent manner
Toh et al. (2011)	China	SNU449, SNU182, HepG2	WST-1.	48 h (12 h for HepG2)	ND	ND	ND	G-Rk1	250 μg/ml	17.5, 21.1, 18.9 at 24 h for the cell lines respectively	Reduction of cell viability significantly
Kim et al. (2008)	South Korea	HepG2	CCK-8 assay	48 h	Kit- supplied TSR8 and HeLa cell lysate	ND	ND	G-Rk1	25, 50, 75, 100 μM	89.2, 68.6, 45.7, 5.4	The first identification of the biological activity of G-Rk1 against HepG2 cell growth
Park et al. (2002)	South Korea	SK-Hep-1 cells	MTT assay	ND	ND	ND	ND	G-Rk1	5, 10, 20, 50, 100 μM/ml	ND	Inhibition of cell viability
Kim et al. (2012)	South Korea	SK-MEL- 2 human melanoma	Cell viability assay	12 h	ND	ND	ND	G-Rk1	5, 10, 25, 50, 75, 100 μM	100, 96, 93.5, 80, 60.5, 18.2	Inhibition of cell viability of SK-Mel- 2 human melanoma
				48 h	ND	ND	ND	G-Rk1	5, 10, 25, 50, 75, 100 μM	93.3, 90.5, 81, 65.5, 40.5, 8.8	cells when they were incubated with G-Rk1 for 24 h and 48 h, at concentrations (0, 10, 25, 50, 75, 100 µM) in a dose dependent manner
Lee et al.	ND	HUVECs	MTT assay	24 h	ND	ND	ND	G-Rk1	10 μg/ml	86.29	Not related to
(2010)				48 h	ND	ND	ND			87.42	G-Rk1 activity
Kim et al. (2013b)	South Korea	Gastric cancer AGS cell	CCK-8 assay	24 h	ND	ND	ND	G- Rg5/G- Rk1	0, 12.5, 25, 50, 100 μg/ml	100, 98.81, 93.57, 93.57, 37.55, 2.94	Inhibition of cell viability in a dose dependent manner
Kwak & Pyo (2016)	South Korea	A549 cell	MTT assay	24 h	Cisplatin	5, 10, 25, 50, 100 μM	84, 73, 72, 51, 37	G-Rk1	5, 10, 25, 50, 100 μM	87, 83, 83, 73, 18	Inhibition of cell viability in a dose dependent manner

(continued on next page)

Tab	e 2	(continued)	
I av	LC 2	commuea	

Author, Year	Cells' origin	Methods/ Cell lines	Methods/ Assays	Methods/ Time effect	Methods/ Treatment						Conclusions	
	Ü		,			Positive control Active compound				•		
					Substance	Concentration	Activity (%)	Substance	Concentration	Activity (%)	•	
Quan et al. (2015)	China	HepG2 cell	MTT assay	24 h	ND	ND	ND	G-Rk1	5, 10, 20, 40, 80 μM	85, 111, 105, 23, 15	Reduction of cell viability	
		A549 cell								97, 93, 110, 47, 3.6		
		HCT-116 cell								99, 103, 70, 6.5, 5.4		
		Hala cell								113, 116, 119, 36, 11		
		MCF-7 cell								125, 127, 123, 84, 8.6		
		PANC-1 cell								98, 115, 104, 24, 9.9		
Cell prolifer	ation inhibi	tion ^b										
Ko et al. (2009)	South Korea	HepG2	CCK-8 assay	24 h	Taxol	0.1 μΜ	100	G-Rk1	50, 75, 100 μΜ	8, 30, 37.5	G-Rk1 inhibits cell proliferation in the early stage of G-Rk1- induced apoptosis cell line.	
Toh et al. (2011)	China	SNU449	WST-1.	48 h	ND	ND	ND	G-Rk1	100 μΜ	50	G-Rk1 inhibit cell proliferation in the SNU449 cell line	
Apoptotic ac	ctivity											
Kim et al. (2008)	South Korea	HepG2	CCK-8 assay	48 h	ND	ND	ND	G-Rk1	100 μΜ	ND	G-Rk1 induced an increase in the fraction of early apoptotic cells from 0.46 to 16.23%.	
Kim et al. (2012)	South Korea	SK-MEL- 2 human melanoma	FAS/FASL antagonist analysis	24 h	Fas/FasL antagonist Kp 7–6	1 mM	ND	G-Rk1	5, 10, 50, 100 μΜ	96, 93.5, 79.5, 18.7	The apoptotic effect of G-Rk1 might be influenced by other pathways	
Hu et al. (2017)	China	Mice	Expression of Bax	ND	APAP	250 mg/kg	90.7	G-Rk1 + APAP	10, 20 mg/kg	25.5, 39.8	G-Rk1 has apoptotic effect by increasing	
			Expression of Bcl-2				12.6			65.7, 50.5	Bax expression and decreasing Bcl-2	
Votes												

Notes.

APAP, acetaminophen; A549 cell, human lung carcinoma; Bax, BCL2-Associated X Protein; Bcl-2, B-cell lymphoma 2; CCK-8, Cell Counting Kit-8; HaCaT, human keratinocyte cell line; HCT-116, human colon carcinoma; Hela, human cervical carcinoma; HepG2, human hepatocellular carcinoma cells; HUVEC, human umbilical vein endothelial cell; LLC-PK1, (pig kidney epithelium, CL-101); MCF-7, human breast adenocarcinoma; MTT, 3-(4,5-dimethyl-thiazol-2yl) -2,5-diphenyl tetrazolium bromide; MC3T3-E1, (RCB1126, an osteoblast-like cell line derived from C57BL/6 mouse calvarias); PANC-1, human pancreatic cancer; SNU449, SNU182, human liver cancer cell lines.

^ameasured by cell viability (%).

^bmeasured by cell proliferation inhibition (%).

(v/v) dimethyl sulphoxide added as control and incubated the cells for 48 h were used in this study. At 75 and 100 μ M of G-Rk1, the effect of G-Rk1 induced cell death was maximized to 55% and 95% cell death respectively. In addition, the results revealed that the treatment of HepG2 cells with 100 μ M G-Rk1, the fraction of early apoptotic cells increased from 0.46 to 16.23% and the underlying mechanism by which G-Rk1 induces the mitochondria-independent apoptosis can be through the activation of caspase-8, the signaling cascade of the one not associated with Fas-associated death domain expression.

To increase their cytotoxicity against Sk-Hep-1 hepatoma cancer cells, *Park et al.* (2002) used steamed ginseng which was separated by HPLC and tested with MTT assay to produce many active ginsenosides including G-Rk1. In this study, they found that the isolated G-Rk1 was associated with an inhibitory effect on cell viability in Sk-Hep1 cells. The growth inhibition concentration of G-Rk1 was 13 μM.

Lung cancer. G-Rk1 was evaluated in human lung cancer A549, and cell viability (% to control) was assessed using MTT assay. At the concentration of 50 μM, there was a statistically significant difference between cisplatin treated cell lines and Rk1 treated cell lines. However, G-Rk1 showed approximately two times higher anticancer activity than Rg5 when treated at 100 μM. After 24 h treatment, the IC50 values of G-Rk1 and cisplatin were 70, and 50 μM, respectively. Several proteins were found to be related to the apoptotic effect of G-Rk1 such as calmodulin-like protein, purine nucleoside phosphorylase, adaptor molecular crk, and transaldolase enzyme were increased while biliverdin reductase, aldehyde dehydrogenase, dihydropteridine reductase, and transactive response DNA binding protein-43 were decreased (*Kwak & Pyo, 2016*). In another study, A549 cell viability was reduced to 47% and 3.6% compared to the vehicle control when treated with G-Rk1 at 40 μM and 80 μM for 24 h, respectively (*Quan et al., 2015*).

Melanoma. To evaluate the inhibitory effect on cell viability of SK-Mel-2 human melanoma cells, *Kim et al.* (2012) incubated these cells with G-Rk1 for 24 and 48 h at different concentrations (0, 10, 25, 50, 75, 100 μM) in a dose-dependent manner. *Erb et al.* (2005) provoked a controversy with the role of FAS and/or FASL in human malignant melanoma. Therefore, the effect of FAS and/or FASL on cell viability was evaluated by *Kim et al.* (2012) by adding Fas/FasL antagonist Kp 7–6 of concentration 1 mM and incubated it for 1 h. Then, the cells were treated with various concentrations of G-Rk1 (1, 5, 10, 50 and 100 μM). The results showed that Kp 7–6 treatment alone did not induce cell death or cell proliferation. Therefore, they concluded that Kp 7–6 has no effect on cell viability when used alone. However, when the cells were treated with Kp 7–6 followed by G-Rk1 (100 μM) treatment, the effect of G-Rk1 was reduced by 32 % compared to the control (no treatment of Kp 7–6). Moreover, they also assessed the induction of apoptosis by G-Rk1 in SK-MEL-2-Human Melanoma and their findings showed that when the concentration of G-Rk1 increased, the number of apoptotic cells also increased. More importantly, the cell lines responded in a dose-dependent manner.

Other types of cancer. Kim et al. (2013b) evaluated the effect of the combination of G-Rg5/G-Rk1 on cell viability of gastric cancer cells. After treatment with this combination at

different concentrations (12.5, 25, 50 and 100 μ M) for 24 h, the results showed an inhibitory effect on cell viability and proliferation of these cells in a dose-dependent manner (99, 93.5, 37.5, 3%) respectively. In another study, cell viability was assessed using different cancer cell lines including human colon carcinoma (HCT-116), human cervical carcinoma (Hela), human breast adenocarcinoma (MCF-7), and human pancreatic cancer (PANC-1). When they were treated with 80 μ M of G-Rk1, cell viability was reduced by 5.4%, 11%, 8.6%, and 9.9%, respectively (*Quan et al.*, 2015).

Antiplatelet aggregation activity

Two studies evaluated the anti-aggregation effects of G-Rk1 both *in vivo* and *in vitro* (*Ju et al.*, 2012; *Lee et al.*, 2009) respectively. *Ju et al.* (2012) compared the antiplatelet aggregation activity of G-Rk1 and acetylsalicylic acid (ASA). The results indicated that G-Rk1 exhibits a stronger antiplatelet aggregation activity than ASA in which the action of G-Rk1 in platelets might be related to arachidonic acid (AA) metabolism. In addition, the alteration of (S) hydroxyl eicosatetraenoic acids and thromboxane B2 levels were determined using an immunoassay kit and UPLC/Q-TOF MS system, respectively. The 12-hydroxyleicosatetraenoic acid level was remarkably decreased in the G-Rk1 group but increased in the ASA-treated group. The thromboxane B2 level in the washed platelets decreased significantly by 66% when treated with 100 μM ASA and 77% when treated with 10 μM G-Rk1 (*Ju et al.*, 2012). They used the colorimetric COX inhibitor screening assay to measure the inhibitory effects of G-Rk1 on COX-1 and COX-2. It was found that G-Rk1 inhibits both COX-1 and COX-2 activities. However, at a concentration of 20 μM, G-Rk1-derived inhibition was higher on COX-2 than on COX-1 (*Ju et al.*, 2012).

Lee et al. (2009) explained in his study that the effect of G-Rk1 on adenosine diphosphate (3–4 μ M) induced platelet aggregation was monitored turbidimetrically by using ASA as a positive control. Both ASA and G-Rk1 showed the dose-dependent inhibitory effect on collagen, AA, and U46619 (9,11-dideoxy-11a,9a-epoxymethanoprostaglandin F2a) (thromboxane A2 mimetic drug)-induced platelet aggregation. However, they showed a negligible effect on adenosine diphosphate-induced aggregation. G-Rk1 exhibited the strongest inhibitory effect on collagen, AA, and U46619-induced platelet aggregation (Lee et al., 2009). G-Rk1 was found to be a potent inhibitor of AA and U46619 -induced platelet aggregation (Table 3).

Anti-inflammatory activity

G-Rk1 was found to have an anti-inflammatory effect by inhibiting NF- κ B levels in the *in vitro* models (*Lee*, 2014). These results were assessed using luciferase assay. HepG2 cells were seeded at 1×10^5 cells/well in a 12-well plate and grown for 24 h. While G-Rk1 was pretreated with dimethyl sulphoxide for 1 h and then it was treated with tumor necrosis factor- α (10 ng/mL), the sulfasalazine was used as positive control. Their data demonstrated the strong inhibitory activity of G-Rk1 on NF- κ B expression with 50% (IC50) value from 0.75 μ M. However, the results revealed that G-Rk1 had cytotoxic effects, which occur in concentrations higher than 10 μ M. Another evaluation of G-Rk1 anti-inflammatory activity (*Kim et al.*, 2010) was its suppressing effect on 12-O-tetradecanoyl- phorbol-13-acetate

Table 3 Summary of the effects of G-Rk1 on antiplatelet aggregation, anti-inflammatory, anti-vascular leakage, nephroprotective effect, neuroprotective effect, bone metabolism, anti-insulin resistance effect, and lipid accumulation.

Author, Year	Cells' origin	Methods/ Cell lines	Methods/ Assays	Methods/ Time effect		Conclusions					
						Positive control		Active compound			•
					Substance	Concentration	Activity	Substance	Concentration	Activity	•
Antiplatelet	t aggregatio	n									
Ju et al. (2012)	South Korea	Platelet	A UPLC/Q-TOF MS system	ND	ASA	50 μM	ND	G-Rk1	50 μM	ND	G-Rk1 strongly inhibited platelet aggregation at 50 μM compared with ASA
Lee et al. (2009)	South Korea	Platelet	Turbidimetrically	ND	ASA	66 μM	50	G-Rk1	3 μΜ	50 ^a	G-Rk1 exhibited 22-fold inhibitory effect of that of ASA on AA-induced aggregation
Anti-inflam	ımatory acti	vity									
Lee (2014)	South Korea	HepG2	NF-κB- luciferase assay	1 h	Sulfasalazine	0.54 μΜ	50	G-Rk1	0.75 μΜ	50 ^a	G-Rk1 exhibited the potentials as anti-inflammatory substance against hepatitis
Kim et al. (2010)	South Korea	Collagen- induced mouse arthritis model	Edema	4 h	ND	ND	ND	G-Rk1	10, 50 mg/kg	9.09, 7.83 ^b	G-Rk1 exhibited anti- inflammatory activity on collagen-induced mouse arthritis model
Ahn et al. (2016)	South Korea	HaCaT/RAW 264.7	TARC/CCL17	1 h	TNF-α	10 ng/mL	157 pg/ml	G-Rg5/G- Rk1 + TNF-α	1, 25, 50 μg/ml	118, 104, 95.4 pg/ml	The results suggesting G-Rg5/G-Rk1 as a promising natural therapy
			MDC/CCL22				243 pg/ml	INI-u		215, 209, 189 pg/ml	in the control of atopic dermatitis
Hu et al. (2017)	China		Expression of TNF- α	1 h	APAP	250 mg/kg	156 ng/L	G-Rk1 + APAP	10, 20 mg/kg	87, 96.7 ng/L	G-Rk1 has a protective effect against APAP
			Expression of IL-1B				1,550 pg/L			1,060, 1,140 pg/L	induced liver injury in mice by decreasing the expression of TNF-α and IL-1B

(continued on next page)

Table 3	(continued)
---------	-------------

origin ar leakage South	Cell lines	Assays	Time effect							-
· ·					Positive control			Active compound		_
· ·				Substance	Concentration	Activity	Substance	Concentration	Activity	-
South										
Korea	HRECs	Sucrose permeability assay	1 h	ND	ND	ND	G-Rk1	10 μg/ml	114.72°	G-Rk1 exhibited an inhibitory effect of VEGF-induced vascular permeability in the mouse retina
Rk1 on lipid	accumulation									
South Korea	3T3-L1 fibroblast cells	Oil red O staining	2 h	ND	ND	ND	G-Rk1	10, 50, 100	0.11, 0.12, 0.08 ^d	G-Rk1 showed inhibitory effect on lipid accumulation in 3T3-L1 adipocytes
ective effect	t									
South Korea	LLC-PK1 cells	EZ-Cytox cell viability assay kit	24 h	EGCG (without cisplatin)	0 μg/ml	99.58%	G-Rg5/G- Rk1		100.0%	G-Rg5 and G-Rk1 showed a protective effect against cisplatin-induced nephrotoxicity in cultured
				EGCG	0, 50, 100, 250 μg/ml	40.76, 46.34, 47.33, 38.65%	G-Rg5/G- Rk1 (cisplatin 25 uM)	0, 50, 100, 250 μg/ml	40.23, 45.23, 57.32, 80.21%	kidney cells and mice
olism										
South Korea	The murine cell line, MC3T3-E1	MTT assay	24 h + 48 h	AMA	60 μg/ml	99.33%	G-Rg5/G- G-Rk1 + AMA	1, 10, 20, 30, 50 μg/ml	109.21, 111.54, 123.43, 131.21, 140.05%	G-Rg5/G-Rk1 enhances cell growth of MC3T3-E1 cells in a dose-dependent manner, also in presence of AMA
ctive effect										
South Korea	Mice	ND	ND	Ethanol	3 g/kg	34.5, 44.22°	G-Rg5/G- G-Rk1	10 mg/kg	45.68, 207.48°	G-Rg5/G-G-Rk1 significantly reversed the memory dysfunction that was induced by ethanol or scopolamine
China	Mice	ND	ND	Ethanol	ND	35, 62°	G-Rg5/G- G-Rk1	2, 10 mg/kg	44, 50/ 145, 184°	The results suggest that those compounds have the ability to improve the acquisition of ethanoltreated mice
	Rk1 on lipid South Korea south Korea south Korea south Korea south Korea	Rk1 on lipid accumulation South 3T3-L1 Korea fibroblast cells ective effect South LLC-PK1 Korea cells olism South The murine Korea cell line, MC3T3-E1 ctive effect South Mice Korea	RkI on lipid accumulation South 3T3-L1 Oil red O staining cells ective effect South LLC-PK1 EZ-Cytox cell viability assay kit south Cells WTT assay cell line, MC3T3-E1 ctive effect South Mice ND Korea ND	RkI on lipid accumulation South 3T3-L1 Oil red O 2 h Korea fibroblast cells ective effect South LLC-PK1 EZ-Cytox cell viability assay kit South The murine Korea cell line, MC3T3-E1 MTT assay 24 h + 48 h ective effect South Mice ND ND	RkI on lipid accumulation South 3T3-L1 Oil red O 2 h ND Korea fibroblast cells ective effect South LLC-PK1 EZ-Cytox cell viability assay kit South Cells Viability assay kit South The murine Korea cell line, MC3T3-E1 MTT assay 24 h + AMA AMA AMA EGCG (without cisplatin) EGCG EGCG EGCG EGCG EGCG ETWE effect South Mice ND ND Ethanol	Rorea permeability assay Rkl on lipid accumulation South 3T3-L1 Oil red O 2 h ND ND South fibroblast cells South LLC-PK1 EZ-Cytox cell viability assay kit South Korea cells South The murine Korea cell line, MC3T3-E1 MTT assay 24 h AMA 60 µg/ml South Korea cell line, MC3T3-E1 MTT assay 24 h AMA 60 µg/ml South Mice ND ND Ethanol 3 g/kg	Rorea permeability assay permeability permeability	Rective effect South So	Kkt on lipid accumulation South Korea 3T3-L1 fibroblast cells Oil red O staining 2h ND ND ND G-Rk1 10, 50, 100 South Korea LLC-PK1 cells EZ-Cytox cell viability assay kit 24 h EGCG (without cisplatin) 0 μg/ml 99.58% G-Rg5/G-Rk1 Korea EGCG (without cisplatin) 0,50,100,250 40.76, μg/ml G-Rg5/G-Rk1 0,50,100, 250 μg/ml 46.34, Rk1 kl1 250 μg/ml 250 μg/ml 250 μg/ml 250 μg/ml 250 μg/ml 250 μg/ml 48 h AMA 60 μg/ml 99.33% G-Rg5/G-G-Rk1 + 50 μg/ml 50 μg/ml 50 μg/ml AMA 50 μg/ml AMA 50 μg/ml AMA 50 μg/ml AMA 44.22* G-Rg5/G-G-Rk1 10 mg/kg China Mice ND ND Ethanol ND 3 g/kg 34.5, G-Rg5/G-G-Rk1 10 mg/kg 2, 10 mg/kg	Rorea Permeability Assay Rorea Rore

(continued on next page)

Table 3 (continued)

Author, Year	Cells' origin	Methods/ Cell lines	Methods/ Assays	Methods/ Time effect		Methods/ Treatment					
					Positive control			Active compound			
					Substance	Concentration	Activity	Substance	Concentration	Activity	
Anti-insulin	n resistance	effect									
Ponnuraj et al. (2014)	South Korea	3T3-L1	MTT assay	24 h	ND	ND	1.36	G-Rg5/G- Rk1	25, 50, 75, 100 μg/ml	1.4, 1.36, 1.33, 1.31 ^f	G-Rk1 increases the IGF- 2R and glucose utilization in adipocytes.
(2017)					Tunicamycin	2 μg/ml	0.73	G-Rg5/G- Rk1 (under Tunicamycin 2 μg/ml)		1.4, 1.47, 1.5, 1.54 ^f	m adipocytes.

Notes.

AA, arachidonic acid; AMA, antimycin A; ASA, acetylsalicylic acid; APAP, acetaminophen; A549 cell, human lung carcinoma; Bax, BCL2-Associated X Protein; Bcl-2, B-cell lymphoma 2; CCK-8, Cell Counting Kit-8; EGCG, Epigallocatechin gallate; HaCaT, human keratinocyte cell line; HCT-116, human colon carcinoma; Hela, human cervical carcinoma; HepG2, human hepatocellular carcinoma cells; HRECs, Primary human retina microvascular endothelial cells; IL-1β, interleukin-1β; LLC-PK1, (pig kidney epithelium, CL-101); MCF-7, human breast adenocarcinoma; MDC/CCL22, macrophage-derived chemokine; MTT, 3-(4,5-dimethyl-thiazol-2yl) -2,5-diphenyl tetrazolium bromide; MC3T3-E1, (RCB1126, an osteoblast-like cell line derived from C57BL/6 mouse calvarias); PANC-1, human pancreatic cancer; SNU449, SNU182, human liver cancer cell lines; TNF- α, tumor necrosis factor-alpha; TARC/CCL17, thymus and activation-regulated chemokine.

^a50% inhibition concentration (IC₅₀) values.

^bwas indicated as the increase in weight of the right ear punch over that of the left (mg).

^c[³H] sucrose permeability (%).

^dmeasured by the optical absorbance at 490 nm.

^eLatency by seconds for learning and testing respectively.

^fCell viability was measured based on absorbance values at 570 and 630 nm.

(TPA) induced mouse ear edema. The right ear of ICR mouse was treated with red ginseng saponin extract, G-Rg3, G-Rg5, and G-Rk1 of 10, and 50 mg/kg and after 30 min, ear edema in both ears was induced by topical application of TPA, which is a potent inflammatory agent. They measured the extent of edema and noticed that the pretreatment with red ginseng saponin extract or G-Rk1 suppresses TPA-induced mouse ear edema, and when administering G-Rk1 orally, the formation of edema was blocked. Hu et al. (2017) showed that in acetaminophen (APAP) induced liver injury in mice, G-Rk1 can be used as a protective agent, as it significantly reduced the levels of tumor necrosis factor (TNF- α) to 87 ng/L and when treated with 10 mg/kg G-Rk1 compared to 156 ng/L when treated with 250 mg/kg APAP. A significant reduction of interleukin-1ß (IL-1ß) was observed with G-Rk1 (Hu et al., 2017). Atopic dermatitis in which keratinocytes and macrophages produce excess chemokines and cytokines, especially thymus and activation-regulated chemokine (TARC/CCL17) and macrophage-derived chemokine (MDC/CCL22), as well as nitric oxide (NO), Ahn's results using G-Rg5/G-Rk1 on TNF-α/ IFN-γ stimulated human keratinocytes cell line (HaCaT cells) showed a significant reduction of TARC/CCL17 expression. Furthermore, using the same combination on the murine macrophage cell line RAW264.7, the secretion of lipopolysaccharide (LPS) mediated NO and reactive oxygen species were significantly reduced, suggesting G-Rg5/G-Rk1 as a promising natural therapy in the control of atopic dermatitis (Ahn et al., 2016) (Table 3).

Effect of G-Rk1 on vascular leakage

A study evaluated the G-Rk1 effect on VEGF (*Maeng et al.*, 2013) by treating primary human retina microvascular endothelial cells with G-Rk1 at a concentration of ($10 \mu g/ml$) for 40 min then stimulating it with 20 $\mu g/ml$ of VEGF to disrupt the cell membrane. Sucrose permeability assay was used to evaluate the endothelial permeability and the results showed that G-Rk1 inhibited VEGF-induced retinal endothelial permeability. They used reverse-transcription polymerase chain reaction (RT-PCR) and densitometric analysis was used to assess translocation of tight junctions (TJ) proteins, and immunostaining was used to evaluate disruption of TJ proteins after the cells were stained with anti-ZO-1, anti-ZO-2, and anti-occludin antibodies. The authors found that G-Rk1 inhibited VEGF effect on TJ protein localization but it did not affect the transcription of TJ proteins (Table 3).

Effect of G-Rk1 on lipid accumulation

Ginseng is known to have effects on obesity ($Kim\ et\ al.$, 2009). In vitro treatment of mouse 3T3-L1 fibroblast cells with G-Rk1 resulted in reducing lipid accumulation, in which these cells differentiated into adipocytes after being treated with various G-Rk1 concentrations (10, 50, 100 μ M) for 2 h at 490 nm optical absorbance ($Kim\ et\ al.$, 2009) (Table 3).

Neuroprotective effect of G-Rk1

The combination of G-Rg5/G-Rk1 had a pronounced effect on the excitotoxic and oxidative stress-induced neuronal cell damage that was tested in primary cultured rat cortical cells (*Bao et al.*, 2005). These cells were cultured *in vitro* for 12–20 days, then exposed to $100 \mu M$ glutamate or N-methyl-D-aspartate for 15 min in the absence or presence of

G-Rg5/G-Rk1. The cell damage was assessed after 20–24 h by measuring LDH activity in the culture media. Data were calculated from cells exposed to the respective excitotoxic insults without ginsenosides. Data presented that approximately 70–80% of the cells were damaged by glutamate or N-methyl-D-aspartate compared to vehicle-treated control cells. The excitotoxic effect was significantly inhibited by G-Rg5/G-Rk1 in a concentration-dependent manner, in which 50% inhibition was achieved at 14.7 μg/mL of G-Rg5/G-Rk1.

In previous work, Bao et al. (2005) used a passive avoidance test to evaluate the effect of G-Rg5/G-Rk1. The latency in seconds was used to measure the cognitive performance of ethanol-induced amnesia in mice. The mice were orally treated with saline as vehicle and the ratio of G-Rg5/G-Rk1 equal 1:1 with a concentration of 10 mg/kg once a day for 4 days. The latency period of the mice administrated with ethanol was 24.9% less than the one of control mice (without ethanol-treatment), but it was significantly enhanced by the oral administration of G-Rg5/G-Rk1 with 1.2-fold increase than that of the control. The same steps were done, but this time after inducing amnesia with a single injection of scopolamine (3 mg/kg), also G-Rg5/G-Rk1 (10 mg/kg) provided the same enhancing significant result (p < 0.01). In another work, Jing et al. (2006) did the same tests of ethanol-induced amnesia in mice, which were given water as the control and ratio of G-Rg5/G-Rk1 equal 1:1 in the concentration of 10 mg/kg. They found that G-Rg5/G-Rk1 could significantly prolong the latency period by 2.97 folds more than that of the control. These two studies presented that G-Rg5/G-Rk1 would give beneficial results in the memory function of the normal, ethanol or scopolamine-induced amnesia in brains. G-Rk1 was reported to have a significant neurogenic activity in Epidermal growth factor-responsive neurosphere stem cells (erNSCs). However, this activity was less than G-Rg5 (*Liu et al.*, 2007) (Table 3).

Nephroprotective effect of G-Rk1

Park et al. (2015) examined the effect of the G-Rg5/G-Rk1 combination on cisplatin-induced nephrotoxicity in mice at cisplatin concentration 25 μ M and G-Rg5/G-Rk1 concentrations of (0, 50, 100, 250 μ g/ml). Results with EZ-cytotoxic cell viability assay kit showed a significant reduction in cisplatin and induced a reduction in cell viability. This effect was higher than that of Epigallocatechin gallate at the same concentrations as G-Rk1 (Table 3).

Bone metabolism

Siddiqi et al. (2014) evaluated the osteogenic activity of G-Rg5/G-Rk1. MC3T3-E1 cells were treated with differentiation medium (either with or without G-Rg5/G-Rk1) for 12 days at different concentrations in which different substances were added to the culture medium in order to evaluate various effects of G-Rg5/G-Rk1 on differentiated fibroblast. The extent of calcium deposition, which is an indicator of osteoblasts mineralization, was measured by MTT assay. Data were expressed as a percentage of control, which showed that G-Rg5/G-Rk1 protected the extracellular matrix mineralization from antimycin A devastating effects. Besides, it turned out that alkaline phosphatase (ALP) activity evaluated by Smart BCA protein assay kit, increased by two folds after treatment with G-Rg5/G-Rk1 (30–50 μ g/mL).

The effect of G-Rg5/G-Rk1 on cellular collagen was measured using Sirius Red-based colorimetric assay. Results were similar to that of ALP activity in which cellular collagen was markedly increased. When glutathione contents of the cells were measured by glutathione assay kit after exposure to various concentrations of G-Rg5/G-Rk1, data showed that G-Rg5/G-Rk1 increase the level of glutathione in a dose-dependent manner. In order to evaluate gene expression levels, a total RNA was isolated from the cells, which were treated with G-Rg5/G-Rk1 and was amplified by RT-PCR. The results indicated that the maturation and the differentiation of MC3T3-E1 cells were induced by G-Rg5/G-Rk1 mediated BMP-2/Runx2 and the level of expression of Runx2 increased by the action of G-Rg5/G-Rk1 (Table 3).

Anti-insulin resistance effect of G-Rk1

Ponnuraj et al. (2014) assessed the effect G-Rk1 on insulin resistance. 3T3-L1 cells were treated with G-Rg5/G-Rk1 complex at different concentrations where tunicamycin was used to induce stress on the endoplasmic reticulum (ER). As for cell viability, measured with MTT assay, results showed that cells treated with G-Rg5/G-Rk1 complex had overcome the stress which induced by tunicamycin. Cells were made insulin resistant by immersing them into a medium that contains insulin and by treating them with dexamethasone, then with the stress agent and G-Rg5/G-Rk1 complex and were analyzed by glucose oxidase reagent, while tunicamycin was used as a positive control. Results found that the amount of glucose left in the medium is high in the cells treated with tunicamycin and low in the cells treated with G-Rg5/G-Rk1 complex and this was achieved through C/EBP homologous protein-10 (CHOP)-mediated pathway and increase insulin-like growth factor receptor (IGF-2R) (Table 3).

Anti-oxidant effect of G-Rk1

Hydrothermal treatment of primary ginsenosides at 100 C transformed them into either deglycosylated and/or dehydrated ginsenoside. As the hypothermal reaction increase, it yields more 20 (S)-Rg3, Rk1, and Rg5. In addition, when they compared the antioxidant activity between the hydrothermally processed samples at 100 C and others processed by the steaming, they got results that showed that sun ginseng samples were higher in antioxidant activities. However, it results in fewer ginsenosides than these samples which reacted at 120 C (*Ryu et al.*, 2016).

Antimicrobial effect of G-Rk1

Xue et al. (2017) assessed the antimicrobial effect of G-Rk1 measured by the minimum inhibitory concentration (MIC) and minimum bacterial concentration (MBC). Compared to Erythrocin (positive control), G-Rk1 exhibited higher MIC and MBC against different bacterial strains compared to Erythrocin (positive control) with (MIC: 31.3 vs 8 μg/ml; MBC: 125.0 vs 16.0 μg/ml) against *Clostridium perfringens*, (MIC: 16.0 vs 8 μg/ml; MBC: 125.0 vs 31.3 μg/ml) against *Fusobacterium nucleatum, and* (MIC: 62.5 vs 16.0 μg/ml; MBC: 125.0 vs 62.5 μg/ml) against *Porphyromonas gingivalis*. Therefore, G-Rk1 can be a promising cure for halitosis.

DISCUSSIONS

Ginsenosides are active compounds extracted from white or red ginseng (P. Ginseng Meyer). Ginsenosides have shown pharmacological effects on the cardiovascular system (Sun, Liu & Chen, 2016), the immune system (Song, Zang & Hu, 2009), and the central nervous system (Zhou et al., 2014), as well as anti-stress, antioxidant, and anti-cancer activities. Moreover, ginsenosides have shown good results in the treatment of diabetes disease by improving glucose and insulin control in type 2 diabetes in a clinical trial (Vuksan et al., 2008). Antitumor inhibitory effects of ginsenosides have been demonstrated because of their cytotoxicities such as the suppression of tumor angiogenesis and metastasis by G-Rb2 (Sato et al., 1994) and the enhancement of apoptosis by G-Rg3 in various cancer cell lines such as breast cancer (*Kim et al.*, 2013a). Although G-Rk1 has a similar structure to G-Rg3, G-Rk1 could be formed by processing ginseng at high temperature, but its antitumor activities would have been limited. Its pharmacological activity has been assessed on antitumor activity in human hepatocellular carcinoma cells (Kim et al., 2008). Apart from these activities, G-Rk1 has been demonstrated to ameliorate impaired memory function and prevent platelet aggregation (Lee et al., 2009). Furthermore, G-Rg3, G-Rk1, and G-Rg5 exhibited a potential effect in the management of human arthritis (Kim et al., 2010).

In this systematic review, we found that various pharmacological and therapeutic effects of G-Rk1 have been reported in the 28 included studies such as anti-cancer effects (*Kim et al.*, 2008), antiplatelet aggregation activities (*Ju et al.*, 2012; *Lee et al.*, 2009), cognitive function enhancement (*Bao et al.*, 2005), anti-inflammatory effects (*Kim et al.*, 2010; *Lee*, 2014), lipid accumulation reduction (*Kim et al.*, 2009), antioxidant effects (*Ryu et al.*, 2016), anti-insulin resistance (*Ponnuraj et al.*, 2014), and protection against human arthritis and nephrotoxicity (*Kim et al.*, 2010; *Park et al.*, 2015).

Anti-cancer activity is one of the most common bioactivities of G-Rk1. By assessing such studies in "cell viability"/"cell proliferation inhibition" and "apoptotic activity", these studies exhibited the anti-cancer effects of G-Rk1 in *in vitro* studies as well as the combined effect of G-Rg5/G-Rk1 (ratio equal 1:1). In terms of "cell viability", the effects of G-Rk1 on cell viability of HepG2 cells, SNU449, SNU182, SK-Hep-1, SK-Mel-2, Hela, HCT-116, MCF-7, PANC-1, A549, and human malignant melanoma was found significantly in a dose-dependent manner (*Kim et al.*, 2012; *Kim et al.*, 2008; *Ko et al.*, 2009; *Kwak & Pyo*, 2016; *Quan et al.*, 2015). The concentrations of G-Rk1 vary from 0 to 100 μM, and the cytotoxic effect was maximum at 75 and 100 μM (*Kim et al.*, 2008). The effects of G-Rk1 were also evaluated in combination with other chemotherapeutics (Bafilomycin A1)(*Ko et al.*, 2009).

It was found that the enhancement of HepG2 cell death was higher when applying G-Rg5/G-Rk1 combination than that of G Rk1 alone. Furthermore, we also found three relevant studies (*Kim et al., 2013b*; *Park et al., 2015*; *Siddiqi et al., 2014*) that evaluated the effects of G-Rg5/G-Rk1 co-treatment on cell viability of gastric cancer cells, mice, and MC3T3-E1 cells. The authors demonstrated that G-Rg5/G-Rk1 has potential effects on inhibiting cell viability and proliferation in a dose-dependent manner. The combination of G-Rg5/G-Rk1 with others chemotherapies (cisplatin (*Park et al., 2015*), antimycin

A (*Siddiqi et al.*, 2014)) has a greater effect on cell death than using G-Rg5 or G-Rk1 alone. Besides, it was also proved that co-administration of G-Rg5/G-Rk1 with a ratio 1:1 have various effects such as improving the cognitive performance in ethanol-induced amnesia in mice (*Bao et al.*, 2005; *Jing et al.*, 2006), inhibiting the exotoxic and oxidative stress-induced neuronal cell damage (*Bao et al.*, 2005), and stimulating the mineralization of the extracellular matrix of osteoblasts (*Siddiqi et al.*, 2014).

In this systematic review, we found two studies presenting the antiplatelet aggregation activities with the results indicating that G-Rk1 ($10 \mu M$) can be stronger than ASA ($100 \mu M$) regarding the antiplatelet aggregation (Ju~et~al., 2012). Lee et al. (2009) also showed that G-Rk1 inhibited the effects of collagen, AA, and U46619-induced platelet aggregation. G-Rk1 was also indicated as one of the effective anti-inflammatory agents through the inhibition of both COX1 and COX2 activities and NF- κ B levels (Ju~et~al., 2012; Lee, 2014).

Although, more than ten of our included studies reported that G-Rk1 has an anti-cancer effect against different cancer cell lines, all of them were *in vitro* studies with no *in vivo* or clinical studies. Unlikely, it was reported that G-Rg3 has an anti-cancer effect in both *in vitro* and *in vivo* (*Shan et al.*, *2014*). A recent meta-analysis of randomized clinical trials revealed that G-Rg3 combined with chemotherapy for non-small-cell lung cancer could enhance the overall survival rate and alleviate the chemotherapy-induced side effects (*Xu et al.*, *2016*). The shortage of *in vivo* or clinical studies to assess the G-Rk1 anti-cancer effect may raise many questions regarding the effect of G-Rk1 in patients and whether it differs from its *in vitro* action. In addition, what alterations that may occur in the patients. Therefore, there is a need for *in vivo* experiments to confirm the G-Rk1 anti-cancer activity and its mechanism.

Regarding the methodological approaches, several limitations were encountered. One of them is that we could not find any clinical study that used G-Rk1 in patients or healthy people. Out of 317 studies, we included 28 studies using our criteria, they were *in vitro* studies and *in vivo* animals. Based on the GRADE method, seven studies remained because of indirectness of evidence (*Ahn et al.*, 2016; *Bao et al.*, 2005; *Jing et al.*, 2006; *Kim et al.*, 2013b; *Park et al.*, 2015; *Ponnuraj et al.*, 2014; *Siddiqi et al.*, 2014) and inability to explain heterogeneity in results (*Bao et al.*, 2005). To date, there is a shortage of literature regarding clinical studies and the clinical use of G-Rk1 to treat some diseases in patients, and it consequently prohibits the clinical analysis.

CONCLUSIONS

In general, G-Rk1 has a significant anti-tumor effect on liver cancer, melanoma, lung cancer, cervical cancer, colon cancer, pancreatic cancer, gastric cancer, and breast adenocarcinoma against *in vitro* cell lines. Furthermore, *In vivo* experiments are necessary to confirm these effects. Additionally, G-Rk1 has demonstrated several pharmacological effects such as antiplatelet aggregation, anti-inflammatory, anti-oxidant, antimicrobial, anti-insulin resistance, neuroprotective, nephroprotective, and anti-lipid accumulation effects. All of these results support the clinical effects of G-Rk1 and demonstrate the promising possibility to develop the G-Rk1-based treatments, either alone or in combination with G-Rg5, for the previously mentioned conditions.

ACKNOWLEDGEMENTS

The authors wish to thank Eman Al Sagheir, (Faculty of Medicine, Zawia University, Libya) for her efforts to revise and improve the English of our manuscript.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

This study was supported in part by a "Grant-in-Aid for Scientific Research (B)" (16H05844, 2016–2019 for Nguyen Tien Huy) from Ministry of Education, Culture, Sports, Science, and Technology (MEXT) of Japan and by the Japan Initiative for Global Research Network on Infectious Diseases (J-GRID) for Kenji Hirayama. The funders had no role in study design, data collection, and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors:

Ministry of Education, Culture, Sports, Science and Technology (MEXT) of Japan: 16H05844, 2016–2019.

Japan Initiative for Global Research Network on Infectious Diseases (J-GRID).

Competing Interests

Authors Abdelrahman Elshafay, Ngo Xuan Tinh, Samar Salman, Yara Saber Shaheen, Eman Bashir Othman, Mohamed Tamer Elhady, Aswin Ratna Kansakar, Linh Tran, and Le Van are members of Online Research Club (ORC). Nguyen Tien Huy is the founder of ORC.

Author Contributions

- Abdelrahman Elshafay, Ngo Xuan Tinh, Samar Salman, Yara Saber Shaheen and Eman Bashir Othman conceived and designed the experiments, performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, wrote the paper, prepared figures and/or tables, reviewed drafts of the paper.
- Mohamed Tamer Elhady, Aswin Ratna Kansakar and Linh Tran conceived and designed the experiments, performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, wrote the paper, reviewed drafts of the paper.
- Le Van, Kenji Hirayama and Nguyen Tien Huy conceived and designed the experiments, performed the experiments, contributed reagents/materials/analysis tools, wrote the paper, reviewed drafts of the paper.

Data Availability

The following information was supplied regarding data availability:

The raw data is included in Tables 1–3.

Supplemental Information

Supplemental information for this article can be found online at http://dx.doi.org/10.7717/peerj.3993#supplemental-information.

REFERENCES

- Ahn S, Siddiqi MH, Aceituno VC, Simu SY, Zhang J, Perez ZE, Kim YJ, Yang DC. 2016. Ginsenoside Rg5:Rk1 attenuates TNF-α/IFN-γ-induced production of thymus- and activation-regulated chemokine (TARC/CCL17) and LPS-induced NO production via downregulation of NF-κB/p38 MAPK/STAT1 signaling in human keratinocytes and macrophages. *In Vitro Cellular & Developmental Biology-Animal* 52:287–295 DOI 10.1007/s11626-015-9983-y.
- Bao HY, Zhang J, Yeo SJ, Myung CS, Kim HM, Kim JM, Park JH, Cho J, Kang JS. 2005. Memory enhancing and neuroprotective effects of selected ginsenosides. *Archives of Pharmacal Research* 28:335–342 DOI 10.1007/BF02977802.
- Chen LW, Wang YQ, Wei LC, Shi M, Chan YS. 2007. Chinese herbs and herbal extracts for neuroprotection of dopaminergic neurons and potential therapeutic treatment of Parkinson's disease. *CNS & Neurological Disorders-Drug Targets* **6**:273–281 DOI 10.2174/187152707781387288.
- **Choi J, Kim TH, Choi TY, Lee MS. 2013.** Ginseng for health care: a systematic review of randomized controlled trials in Korean literature. *PLOS ONE* **8**:e59978 DOI 10.1371/journal.pone.0059978.
- **Choi KT. 2008.** Botanical characteristics, pharmacological effects and medicinal components of Korean Panax ginseng C A Meyer. *Acta Pharmacologica Sinica* **29**:1109–1118 DOI 10.1111/j.1745-7254.2008.00869.x.
- Erb P, Ji J, Wernli M, Kump E, Glaser A, Buchner SA. 2005. Role of apoptosis in basal cell and squamous cell carcinoma formation. *Immunology Letters* **100**:68–72 DOI 10.1016/j.imlet.2005.06.008.
- Ernst E. 2010. Panax ginseng: an overview of the clinical evidence. *Journal of Ginseng Research* 34:259–263 DOI 10.5142/jgr.2010.34.4.259.
- Guyatt G, Oxman AD, Akl EA, Kunz R, Vist G, Brozek J, Norris S, Falck-Ytter Y, Glasziou P, DeBeer H, Jaeschke R, Rind D, Meerpohl J, Dahm P, Schunemann HJ. 2011. GRADE guidelines: 1. Introduction-GRADE evidence profiles and summary of findings tables. *Journal of Clinical Epidemiology* 64:383–394 DOI 10.1016/j.jclinepi.2010.04.026.
- Hu JN, Xu XY, Li W, Wang YM, Liu Y, Wang Z, Wang YP. 2017. Ginsenoside Rk1 ameliorates paracetamol-induced hepatotoxicity in mice through inhibition of inflammation, oxidative stress, nitrative stress and apoptosis. *Journal of Ginseng Research* DOI 10.1016/j.jgr.2017.07.003.
- Jing Z, Shi-rong W, Quan-cheng C, Long PH, Kang JS. 2006. Effects of ginsenosides Rg3(R), Rg3(S) and Rg5/Rk1 on memory improvement of ethanol treated mice. *Journal of Jilin Agricultural University* 28:1–3 DOI 10.13327/j.jjlau.2006.03.012.

- Ju HK, Lee JG, Park MK, Park SJ, Lee CH, Park JH, Kwon SW. 2012. Metabolomic investigation of the anti-platelet aggregation activity of ginsenoside Rk(1) reveals attenuated 12-HETE production. *Journal of Proteome Research* 11:4939–4946 DOI 10.1021/pr300454f.
- Kang KS, Kim HY, Baek SH, Yoo HH, Park JH, Yokozawa T. 2007. Study on the hydroxyl radical scavenging activity changes of ginseng and ginsenoside-Rb2 by heat processing. *Biological and Pharmaceutical Bulletin* 30:724–728

 DOI 10.1248/bpb.30.724.
- Kang KS, Kim HY, Yamabe N, Yokozawa T. 2006. Stereospecificity in hydroxyl radical scavenging activities of four ginsenosides produced by heat processing. *Bioorganic & Medicinal Chemistry Letters* 16:5028–5031 DOI 10.1016/j.bmcl.2006.07.071.
- Keum YS, Park KK, Lee JM, Chun KS, Park JH, Lee SK, Kown H, Surh YJ. 2000. Antioxidant and anti-tumor promoting activities of the methanol extract of heat-processed ginseng. *Cancer Letters* **150**:41–48 DOI 10.1016/S0304-3835(99)00369-9.
- Kim BM, Kim DH, Park JH, Na HK, Surh YJ. 2013a. Ginsenoside Rg3 induces apoptosis of human breast cancer (MDA-MB-231) cells. *Journal of Cancer Prevention* 18:177–185 DOI 10.15430/JCP.2013.18.2.177.
- **Kim HJ, Kim P, Shin CY. 2013.** A comprehensive review of the therapeutic and pharmacological effects of ginseng and ginsenosides in central nervous system. *Journal of Ginseng Research* **37**:8–29 DOI 10.5142/jgr.2013.37.8.
- Kim JS, Joo EJ, Chun J, Ha YW, Lee JH, Han Y, Kim YS. 2012. Induction of apoptosis by ginsenoside Rk1 in SK-MEL-2-human melanoma. *Archives of Pharmacal Research* 35:717–722 DOI 10.1007/s12272-012-0416-0.
- **Kim KR, Chung TY, Shin H, Son SH, Park KK, Choi JH, Chung WY. 2010.** Red ginseng saponin extract attenuates murine collagen-induced arthritis by reducing proinflammatory responses and matrix metalloproteinase-3 expression. *Biological and Pharmaceutical Bulletin* **33**:604–610 DOI 10.1248/bpb.33.604.
- Kim SN, Lee JH, Shin H, Son SH, Kim YS. 2009. Effects of *in vitro*-digested ginsenosides on lipid accumulation in 3T3-L1 adipocytes. *Planta Medica* **75**:596–601 DOI 10.1055/s-0029-1185358.
- Kim WY, Kim JM, Han SB, Lee SK, Kim ND, Park MK, Kim CK, Park JH. 2000. Steaming of ginseng at high temperature enhances biological activity. *Journal of Natural Products* 63:1702–1704 DOI 10.1021/np990152b.
- **Kim YJ, Kwon HC, Ko H, Park JH, Kim HY, Yoo JH, Yang HO. 2008.** Anti-tumor activity of the ginsenoside Rk1 in human hepatocellular carcinoma cells through inhibition of telomerase activity and induction of apoptosis. *Biological and Pharmaceutical Bulletin* **31**:826–830 DOI 10.1248/bpb.31.826.
- **Kim YJ, Yamabe N, Choi P, Lee JW, Ham J, Kang KS. 2013b.** Efficient thermal deglycosylation of ginsenoside Rd and its contribution to the improved anticancer activity of ginseng. *Journal of Agricultural and Food Chemistry* **61**:9185–9191 DOI 10.1021/jf402774d.

- **Ko H, Kim Y, Park J, Park JY, Yang HO. 2009.** Autophagy inhibition enhances apoptosis induced by ginsenoside Rk1 in hepatocellular carcinoma cells. *Bioscience, Biotechnology, and Biochemistry* **73**:2183–2189 DOI 10.1271/bbb.90250.
- **Kwak JH, Pyo JS. 2016.** Characterization of apoptosis induced by ginsenosides in human lung cancer cells. *Analytical Letters* **49**:843–854 DOI 10.1080/00032719.2015.1079208.
- Kwon SW, Han SB, Park IH, Kim JM, Park MK, Park JH. 2001. Liquid chromatographic determination of less polar ginsenosides in processed ginseng. *Journal of Chromatography A* 921:335–339 DOI 10.1016/S0021-9673(01)00869-X.
- **Lee JG, Lee YY, Kim SY, Pyo JS, Yun-choi HS, Park JH. 2009.** Platelet antiaggregating activity of ginsenosides isolated from processed ginseng. *Pharmazie* **64**:602–604 DOI 10.1691/ph.2009.9577.
- **Lee SM. 2014.** Anti-inflammatory effects of ginsenosides Rg5, Rz1, and Rk1: inhibition of TNF-alpha-induced NF-kappaB, COX-2, and iNOS transcriptional expression. *Phytotherapy Research* **28**:1893–1896 DOI 10.1002/ptr.5203.
- Lee S, Maharjan S, Kim K, Kim NJ, Choi HJ, Kwon YG, Suh YG. 2010. Cholesterolderived novel anti-apoptotic agents on the structural basis of ginsenoside Rk1. *Bioorganic & Medicinal Chemistry Letters* 20:7102–7105 DOI 10.1016/j.bmcl.2010.09.071.
- Lim SC, Maeng YS, Kwon JY, Kang MH, Hyang JH, Kim YH, Kwon YK, Park YW. 2009. The effect of ginsenoside Rk1 in junctional protein of severe preeclamptic placenta. *Korean Journal of Obstetrics and Gynecology* **52**:303–308.
- **Lim W, Mudge KW, Vermeylen F. 2005.** Effects of population, age, and cultivation methods on ginsenoside content of wild American ginseng (Panax quinquefolium). *Journal of Agricultural and Food Chemistry* **53**:8498–8505 DOI 10.1021/jf051070y.
- **Liu JW, Tian SJ, Barry JD, Luu B. 2007.** Panaxadiol glycosides that induce neuronal differentiation in neurosphere stem cells. *Journal of Natural Products* **70**:1329–1334 DOI 10.1021/np070135j.
- Maeng YS, Maharjan S, Kim JH, Park JH, Suk Yu Y, Kim YM, Kwon YG. 2013. Rk1, a ginsenoside, is a new blocker of vascular leakage acting through actin structure remodeling. *PLOS ONE* **8**:e68659 DOI 10.1371/journal.pone.0068659.
- Moher D, Liberati A, Tetzlaff J, Altman DG, The PRISMA Group. 2009. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *BMJ* 339:Article b2535 DOI 10.1136/bmj.b2535.
- Nag SA, Qin JJ, Wang W, Wang MH, Wang H, Zhang R. 2015. Ginsenosides as anticancer agents: *in vitro* and *in vivo* activities, structure-activity relationships, and molecular mechanisms of action. *Frontiers in Pharmacology* **3**:Article 25 DOI 10.3389/fphar.2012.00025.
- **Nemmani KV, Ramarao P. 2003.** Ginsenoside Rf potentiates U-50,488H-induced analgesia and inhibits tolerance to its analgesia in mice. *Life Sciences* **72**:759–768 DOI 10.1016/S0024-3205(02)02333-0.
- Park IH, Piao LZ, Kown SW, Lee YJ, Cho SY, Park MK, Park JH. 2002. Cytotoxic dammarane glycosides from processed ginseng. *Chemical and Pharmaceutical Bulletin* **50**:538–540 DOI 10.1248/cpb.50.538.

- Park JY, Choi P, Kim T, Ko H, Kim HK, Kang KS, Ham J. 2015. Protective effects of processed ginseng and its active ginsenosides on cisplatin-induced nephrotoxicity: *in vitro* and *in vivo* studies. *Journal of Agricultural and Food Chemistry* **63**:5964–5969 DOI 10.1021/acs.jafc.5b00782.
- **Ponnuraj SP, Siraj F, Kang S, Noh HY, Min JW, Kim YJ, Yang DC. 2014.** Amelioration of insulin resistance by Rk1 + Rg5 complex under endoplasmic reticulum stress conditions. *Pharmacognosy Research* **6**:292–296 DOI 10.4103/0974-8490.138257.
- Quan K, Liu Q, Wan JY, Zhao YJ, Guo RZ, Alolga RN, Li P, Qi LW. 2015. Rapid preparation of rare ginsenosides by acid transformation and their structure-activity relationships against cancer cells. *Scientific Reports* 5:8598 DOI 10.1038/srep08598.
- Ryu J, Lee HW, Yoon J, Seo B, Kwon DE, Shin UM, Choi KJ, Lee YW. 2016. Effect of hydrothermal processing on ginseng extract. *Journal of Ginseng Research* DOI 10.1016/j.jgr.2016.12.002.
- Sato K, Mochizuki M, Saiki I, Yoo YC, Samukawa K, Azuma I. 1994. Inhibition of tumor angiogenesis and metastasis by a saponin of Panax ginseng, ginsenoside-Rb2. *Biological and Pharmaceutical Bulletin* 17:635–639 DOI 10.1248/bpb.17.635.
- Schlag EM, McIntosh MS. 2006. Ginsenoside content and variation among and within American ginseng (Panax quinquefolius L.) populations. *Phytochemistry* 67:1510–1519 DOI 10.1016/j.phytochem.2006.05.028.
- Shan X, Fu Y-S, Aziz F, Wang X-Q, Yan Q, Liu J-W. 2014. Ginsenoside Rg3 inhibits melanoma cell proliferation through down-regulation of histone deacetylase 3 (HDAC3) and increase of p53 acetylation. *PLOS ONE* 9:e115401 DOI 10.1371/journal.pone.0115401.
- **Shin BK, Kwon SW, Park JH. 2015.** Chemical diversity of ginseng saponins from Panax ginseng. *Journal of Ginseng Research* **39**:287–298 DOI 10.1016/j.jgr.2014.12.005.
- Siddiqi MH, Siddiqi MZ, Ahn S, Kang S, Kim YJ, Veerappan K, Yang DU, Yang DC. **2014.** Stimulative effect of ginsenosides Rg5:Rk1 on murine osteoblastic MC3T3-E1 cells. *Phytotherapy Research* **28**:1447–1455 DOI 10.1002/ptr.5146.
- **Song X, Zang L, Hu S. 2009.** Amplified immune response by ginsenoside-based nanoparticles (ginsomes). *Vaccine* **27**:2306–2311 DOI 10.1016/j.vaccine.2009.02.040.
- **Sun Y, Liu Y, Chen K. 2016.** Roles and mechanisms of ginsenoside in cardiovascular diseases: progress and perspectives. *Science China Life Sciences* **59**:292–298 DOI 10.1007/s11427-016-5007-8.
- Tachikawa E, Kudo K, Harada K, Kashimoto T, Miyate Y, Kakizaki A, Takahashi E. 1999. Effects of ginseng saponins on responses induced by various receptor stimuli. *European Journal of Pharmacology* 369:23–32 DOI 10.1016/S0014-2999(99)00043-6.
- **Tang W, Zhang Y, Gao J, Ding X, Gao S. 2008.** The anti-fatigue effect of 20(R)-ginsenoside Rg3 in mice by intranasally administration. *Biological and Pharmaceutical Bulletin* **31**:2024–2027 DOI 10.1248/bpb.31.2024.
- **Toh D, Pate DN, Chan EC, Teo A, Neo SY, Koh HL. 2011.** Anti-proliferative effects of raw and steamed extracts of Panax notoginseng and its ginsenoside constituents on human liver cancer cells. *Chinese Medicine* **6**:Article 4 DOI 10.1186/1749-8546-6-4.

- Vuksan V, Sung MK, Sievenpiper JL, Stavro PM, Jenkins AL, Di Buono M, Lee KS, Leiter LA, Nam KY, Arnason JT, Choi M, Naeem A. 2008. Korean red ginseng (Panax ginseng) improves glucose and insulin regulation in well-controlled, type 2 diabetes: results of a randomized, double-blind, placebo-controlled study of efficacy and safety. *Nutrition, Metabolism & Cardiovascular Diseases* 18:46–56 DOI 10.1016/j.numecd.2006.04.003.
- Xu T, Jin Z, Yuan Y, Wei H, Xu X, He S, Chen S, Hou W, Guo Q, Hua B. 2016. Ginsenoside Rg3 serves as an adjuvant chemotherapeutic agent and VEGF inhibitor in the treatment of non-small cell lung cancer: a meta-analysis and systematic review. *Evidence-Based Complementary and Alternative Medicine* 2016:Article 7826753 DOI 10.1155/2016/7826753.
- Xue P, Yao Y, Yang XS, Feng J, Ren GX. 2017. Improved antimicrobial effect of ginseng extract by heat transformation. *Journal of Ginseng Research* 41:180–187 DOI 10.1016/j.jgr.2016.03.002.
- **Yun TK. 2001.** Panax ginseng—a non-organ-specific cancer preventive? *The Lancet Oncology* **2**:49–55 DOI 10.1016/s1470-2045(00)00196-0.
- Zhou J, Zhang HA, Lin Y, Liu HM, Cui YM, Xu Y, Zhao N, Ma JM, Fan K, Jiang CL. 2014. Protective effect of ginsenoside against acute renal failure via reduction of renal oxidative stress and enhanced expression of ChAT in the proximal convoluted tubule and ERK1/2 in the paraventricular nuclei. *Physiological Research* 63:597–604.