Targeted NGS for species level phylogenomics: “made to measure” or “one size fits all”?

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Targeted high-throughput sequencing using hybrid-enrichment offers a promising source of data for inferring multiple, meaningfully resolved, independent gene trees suitable to address challenging phylogenetic problems in species complexes and rapid radiations. The targets in question can either be adopted directly from more or less universal tools, or custom made for particular clades at considerably greater effort. We applied custom made scripts to select sets of homologous sequence markers from transcriptome and WGS data for use in the flowering plant genus *Erica* (Ericaceae). We compared the resulting targets to those that would be selected both using different available tools (Hyb-Seq; MarkerMiner), and when optimising for broader clades of more distantly related taxa (Ericales; eudicots). Approaches comparing more divergent genomes (including MarkerMiner, irrespective of input data) delivered fewer and shorter potential markers than those targeted for *Erica*. The latter may nevertheless be effective for sequence capture across the wider family Ericaceae. We tested the targets delivered by our scripts by obtaining an empirical dataset. The resulting sequence variation was lower than that of standard nuclear ribosomal markers (that in *Erica* fail to deliver a well resolved gene tree), confirming the importance of maximising the lengths of individual markers. We conclude that rather than searching for “one size fits all” universal markers, we should improve and make more accessible the tools necessary for developing “made to measure” ones.

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Abstract

Targeted high-throughput sequencing using hybrid-enrichment offers a promising source of data for inferring multiple, meaningfully resolved, independent gene trees suitable to address challenging phylogenetic problems in species complexes and rapid radiations. The targets in question can either be adopted directly from more or less universal tools, or custom made for particular clades at considerably greater effort. We applied custom made scripts to select sets of homologous sequence markers from transcriptome and WGS data for use in the flowering plant genus *Erica* (Ericaceae). We compared the resulting targets to those that would be selected both using different available tools (Hyb-Seq; MarkerMiner), and when optimising for broader clades of more distantly related taxa (Ericales; eudicots). Approaches comparing more divergent genomes (including MarkerMiner, irrespective of input data) delivered fewer and shorter potential markers than those targeted for *Erica*. The latter may nevertheless be effective for sequence capture across the wider family Ericaceae. We tested the targets delivered by our scripts by obtaining an empirical dataset. The resulting sequence variation was lower than that of standard nuclear ribosomal markers (that in *Erica* fail to deliver a well resolved gene tree), confirming the importance of maximising the lengths of individual markers. We conclude that rather than searching for "one size fits all" universal markers, we should improve and make more accessible the tools necessary for developing "made to measure" ones.

Keywords: Ericaceae; hybridization enrichment; marker development, next-generation sequencing; phylogeny; targeted sequence capture; target enrichment; transcriptome

Introduction

DNA sequence data is the cornerstone of comparative and evolutionary research, invaluable for inference of population-level processes and species delimitation through to higher level relationships. Sanger sequencing (Sanger, Nicklen & Coulson, 1977) and Polymerase Chain Reaction (PCR) amplification (Saiki et al., 1985) have been standard tools for decades, aided by the development of protocols that can be applied across closely and distantly related organisms. In plants, universal primers such as for plastid (Taberlet et al., 1991), nuclear ribosomal (White et al., 1990) and even single or low copy nuclear (Blattner, 2016) sequences have been widely applied to infer evolutionary histories. Many empirical studies are still limited to these few independent markers, the phylogenetic signal of which may not reflect the true sequence of speciation events (Kingman, 1982; White et al., 1990). Additionally, the resulting gene trees are often poorly resolved, particularly when divergence of lineages was rapid. When it is not possible to generate a robust and unambiguous phylogenetic hypothesis using standard universal markers, protocols for alternative low copy genes are highly desirable (Sang, 2002; Hughes, Eastwood & Bailey, 2006).

With the development of next generation sequencing (NGS) techniques, we now have potential access to numerous nuclear markers allowing us to address evolutionary questions without being constrained by the generation of sequence datasets per se. In principle, the whole genome is at our disposal, but whole genome sequencing (WGS) is currently relatively expensive, time-consuming and computationally difficult, especially for non-model organisms and eukaryote genomes in general (Jones & Good, 2016). These disadvantages will doubtless reduce in the near future, but nevertheless much of the data that might be obtained through WGS is irrelevant for particular purposes. In the case of phylogenetic problems, repetitive elements and multiple copy genes are not useful; neither are sequences that are highly constrained and hence insufficiently variable, nor indeed those that are too variable and impossible to align; nor those subject to strong selection pressure. We need strategies to identify and target sequencing of markers

appropriate for phylogenomic analysis in different clades and at different taxonomic levels, and are currently faced with an array of options.

Different methods, referred to in general as "genome-partitioning approaches", or "reduced-representation genome sequencing", have been developed that are cheaper, faster and computationally less demanding than WGS, and as such are currently more feasible for analyses of numerous samples for particular purposes (Mamanova et al., 2010). These include reduced-site-associated DNA sequencing (RAD-seq; Miller et al., 2007), and similar Genotyping by sequencing (GBS) approaches (Elshire et al., 2011), and whole-transcriptome shotgun sequencing (RNA-seq; Wang, Gerstein & Snyder, 2009). RADSeq and GBS libraries are generated using restriction digestion (followed by size-selection and PCR enrichment) to obtain homologous sequences representing a more or less random subset of the total genomic DNA, whereas RNA-seq uses NGS to retrieve the complete transcriptome of a sample from isolated RNA. These methods can be applied to non-model species (Johnson et al., 2012) but do not necessarily deliver the most informative data for phylogenetic inference. RAD-seq/GBS sequences are short, generally used for obtaining (independent) single nucleotide polymorphisms (SNPs) from across the genome, suitable for population genetic analyses. Transcriptome data cannot be obtained from dried material (such as herbarium specimens), restricting its application. The sequences are functionally conserved and therefore may be more suitable for analysing more ancient divergences, such as the origins of land plants (Wickett et al., 2014). Neither approach is ideal for inferring meaningfully resolved independent gene trees of closely related species as they will inevitably present limited numbers of linked, informative characters.

Alternative approaches can be used to target more variable, longer contiguous sequences involving selective enrichment of specific subsets of the genome before using NGS through PCR based, or sequence capture techniques. PCR based enrichment, or multiplex and microfluidic amplification of PCR products, is the simultaneous amplification of multiple targets (e.g. 48, as used in Uribe-Convers, Settles & Tank, 2016; to potentially hundreds or low thousands per reaction). Although this method dispenses with the need for time-consuming library preparation, it requires prior knowledge of sequences for the design of primers; such primers must be restricted to within regions that are known to be conserved across the study group.

Current targeted sequence capture methods involve hybridization in solution between genomic

DNA fragments and biotinylated RNA "baits" (also referred to as "probes" or the "Capture Library'') between 70 and 120 bp long. Hybridization capture can be used with non-model organisms (as is the case for RAD-seq/GBS and RNA-seq), and shows promising results with fragmented DNA (such as might be retrieved from museum specimens) (Moriarty Lemmon & Lemmon, 2013; Zimmer & Wen, 2015; Hart et al., 2016). Moreover, even without baits specifically designed using organelle genomes, plastid and mitochondrial sequences can also be retrieved during the hybrid-enrichment process (Tsangaras et al., 2014). Use of targeted sequence capture for phylogenetic inference is on the increase but still somewhat in its infancy, with a range of different more or less customised laboratory and bioinformatic protocols being applied to different organismal groups and in different laboratories. The protocols follow two general approaches: One is to design baits for use in specific organismal groups (e.g. Compositae, Mandel et al., 2014; cichlid fish, Ilves & Lopez-Fernandez, 2014; and Apocynaceae, Weitemier et al., 2014□). To this end, conserved orthologous sequences of genes of the species of interest are identified e.g. using a BLASTn or BLASTx search (or equivalent) with transcriptome data, expressed sequences tags (ESTs) and/or WGS. Alternatively, and with considerably less effort, pre-designed sets of more universal baits are used (Faircloth et al., 2012; Lemmon, Emme & Lemmon, 2012). Of the latter, "Ultra Conserved Elements" (UCE) (Faircloth et al., 2012) and "Anchored Hybrid Enrichment" (AHE) (Lemmon, Emme & Lemmon, 2012) approaches have been applied in phylogenetic analyses of animal (e.g. snakes, Pyron et al., 2014; lizards, □Leaché et al., 2014; frogs, Peloso et al., 2016□; and spiders, Hamilton et al., 2016) and plant (*Medicago,* De Sousa et al., 2014□; *Sarracenia,* Stephens et al., 2015; palms, Comer et al., 2016; Heyduk et al., 2016; *Heuchera,* Folk, Mandel & Freudenstein, 2015; *Inga,* Nicholls et al., 2015; and *Protea,* Mitchell et al., 2017) clades.

Universal protocols are an attractive prospect, in terms of reduced cost and effort, and because they might generate broadly comparable data suitable for wider analyses (or even DNA barcoding; Blattner, 2016). However, the resulting sequence markers may not be optimal for all purposes. For phylogenetic inference, low-copy markers are required to avoid paralogy issues, and for successful hybridisation capture similarity of baits to target sequences must fall within c. 75-100% (Moriarty Lemmon & Lemmon, 2013). This places a restriction on more universal markers that will necessarily exclude potentially useful low copy, high variability markers where these are subject to duplications or too variable in particular lineages.

The selection of appropriate sequence markers may therefore be crucial in determining the success of this kind of analysis, especially for non-model species. Transcriptome data for increasing numbers of non-model organisms are available (Matasci et al., 2014) and bioinformatics tools are available that can assist in the selection of markers and design of baits, taking transcriptome and/or whole genome sequences of relevant taxa as input. These include MarkerMiner (Chamala et al., 2015), Hyb-Seq (Weitemier et al., 2014; Schmickl et al., 2016) and BaitsFisher (Mayer et al., 2016). The question for researchers embarking on phylogenomic analyses is whether it is worth the additional cost and effort involved in designing custom baits, and how to select sequence markers in order to get the most information out of a given investment of time and funds.

Our ongoing research addresses the challenge of resolving potentially complex phylogenetic relationships between closely related populations and species of a non-model flowering plant group, the genus *Erica* (Ericaceae; one of 22 families of the asterid order Ericales; (Stevens, 2001)). The c. 700 South African species of *Erica* represent the most species rich 'Cape clade' in the spectacularly diverse Cape Floristic Region (Linder, 2003; Pirie et al., 2016). Analyses of the *Erica* clade as a whole offer a rich source of data in terms of numbers of evolutionary events, and our ability to infer such events accurately is arguably greatest in the most recently diverged species and populations. In such clades, the historical signal for shifts in key characteristics and geographic ranges are in general less likely to have been overwritten by subsequent shifts and (local) extinction. However, phylogenetic inference in rapid species radiations, such as that of Cape *Erica* (Pirie et al., 2016), Andean *Lupinus* (Hughes & Eastwood, 2006) or Lake Malawi cichlid fish (Santos & Salzburger, 2012) presents particular challenges. These include low sequence divergence confounded by the impact of both reticulation and coalescence on population-level processes. To infer a meaningful species tree under such circumstances, we need data suitable to infer multiple, maximally informative, independent gene trees.

The aims of this paper are to compare custom versus universal approaches to marker selection, in terms of the predicted sequence lengths and variability and to compare their potential for delivering multiple independent and informative gene trees. In so doing, we generate a tool for low-level phylogenetic inference in *Erica*, we test it experimentally by generating empirical data, and we assess its potential application across a wider group, e.g. the family Ericaceae.

150

151 **Materials & Methods**

152 Our first aim was to identify homologous, single-copy sequence markers for which we could
153 design baits (probes) with similarity of ≥75% (as hybridization between target and probe
154 tolerates a maximum of 25% divergence) that would be predicted to deliver the greatest numbers
155 of informative characters. Baits currently represent a relatively large proportion of the total cost
156 of the protocol (which is expensive on a per sample basis compared to e.g. PCR enrichment). We
157 therefore restricted the total length of hybridisation baits to 692,400 bp (5770 individual 120 bp
158 baits), representing a total "capture footprint" (i.e. sequence length) of 173,100 bp given probe
159 overlap representing 4x coverage. With our lab protocol (see below) this permits dilution of the
160 baits to capture five samples per unit of baits instead of just one. We developed custom-made
161 Python 2.7.6 scripts to identify the wider pool of all potential target sequences from
162 transcriptome and WGS data, as well as applying already available scripts/software for
163 comparison. We subsequently implemented in further scripts different options for prioritising
164 target variability, length and/or intron numbers and lengths to select optimal sequence markers
165 from these pools of potential targets. We then compared the lengths and numbers of the
166 sequences in the different resulting potential and optimal marker sets.

167

168 *Identifying potential target sequences*

169 Our custom-made script (AllMarkers.py; summarised in Fig. 1 available at Github:
170 https://github.com/MaKadlec/Select-Markers/tree/AllMarkers) requires at least two
171 transcriptomes, ideally of taxa closely related to the focal group. Where WGS/genome skimming
172 data of one or more such taxa is available, it can be used too, as in Folk, Mandel & Freudenstein
173 (2015). AllMarkers.py implements the following steps: First, multiple transcriptomes are
174 compared to identify homologues, retaining those found in at least two transcriptomes (and
175 hence likely to also be found in related genomes). We have successfully used up to eight
176 transcriptomes; on eight cores of a fast desktop PC the analyses ran for up to two days.
177 Particularly when larger numbers of larger transcriptomes are compared, an additional filter can
178 be applied prior to this step to remove shorter sequences (e.g. those <1,000 bp) and thereby
179 improve speed. Next, multiple copy sequences (for which homology assessment might be

problematic) are identified, either using BLASTn of transcriptome against WGS, or (when no WGS data is available) by comparison to the classification of proteins as single/mostly single copy across angiosperms by De Smet et al. (2013), using BLASTx following the approach used in MarkerMiner (Chamala et al., 2015). Multiple-copy sequences are then excluded. Finally, a filter for similarity ≥75% is applied. This series of steps is comparable to but differs from those implemented in Hyb-Seq (Weitemier et al., 2014) and in MarkerMiner (Chamala et al., 2015) (Fig. 1), which we also applied here.

The Hyb-Seq pipeline uses transcriptome and WGS sequences of closely related species to select marker sequences. This pipeline employs BLAT (BLAST-like Alignment Tool) to identify single-copy sequences with identity > 99%. After isoform identification, sequences with exons <120 bp and those of total length <960 bp are removed (representing a further filtering of potential targets that is comparable in part to the next steps in our own scripts, as described below), then orthologous sequences are identified using the transcriptome of a closest related species or transcriptomes of four angiosperms (*Arabidopsis thaliana, Oryza sativa, Populus trichocarpa* and *Vitis vinifera*).

For MarkerMiner, WGS data is neither required nor used. This pipeline involves selecting sequences by size in input transcriptomes (we set length parameter to >1000 bp) then using reciprocal BLAST between transcriptomes and a reference proteome to select sequences above 70% similarity. The proteome most closely related to *Erica* implemented in MarkerMiner in August 2016 was that of *Vitis vinifera* (Vitaceae; Vitales; core eudicots; Stevens, 2001). This minimum similarity threshold does not directly reflect that required for successful probe hybridisation, and particularly given comparison to a relatively distantly related proteome (as in this case) can be expected to be conservative. In the final step, MarkerMiner retains putative single copy ortholog pairs following De Smet et al. (2013).

Selection of optimal target sequences from pools of potential targets

The above steps result in potentially large pools of potentially highly suboptimal targets, in particular shorter and/or invariable sequences that, given rapid lineage divergence, may not deliver enough informative characters to discern meaningfully resolved independent gene trees. In order to select optimal markers from these pools given a limited number of baits we designed

a further script (available at Github: https://github.com/MaKadlec/Select-Markers/tree/BestMarkers.py). Depending on the phylogenetic problem to hand (e.g. recent, species level divergence versus older radiations) and available information (e.g. about sequence variability in the focal clade; positions and lengths of potentially more variable introns), various options are possible. In our case, from WGS and transcriptome data we know where introns are likely to be found, but in the absence of sequences from multiple accessions of our ingroup, the only indication of sequence variability comes from comparison of coding regions of relatively distantly related taxa, i.e. single species of *Rhododendron*, *Vaccinium* and *Erica*. We therefore assessed two options: 1) simply selecting the longest sequences. 2) Selecting the longest sequences, but taking into account the (likely) additional length of introns. Using WGS data, we assessed the number and length of introns. For the purpose of ranking potential markers, we decided to use mean intron length in order to avoid favouring the selection of sequences with large introns that a) might not be efficiently captured/sequenced; or b) might not be so large in the focal clade. Finally, the longest sequences were selected that could be captured with our maximum number of baits. Coding regions <120 bp long are shorter than the baits and are likely to be ineffectively captured. For this reason, in the Hyb-Seq approach (Weitemier et al., 2014) all sequences including exons <120 bp are excluded; however, this is at the expense of excluding otherwise optimal markers that may include individual exons of <120 bp. We therefore opted to retain sequences including one or more coding regions ≥120 bp, whilst excluding individual exons <120 bp as potential targets for baits.

In silico *comparison with empirical data*

Our custom scripts (AllMarkers.py and BestMarkers.py), the Hyb-Seq and MarkerMiner pipelines were each applied to the transcriptomes of *Rhododendron scopulorum* (18,307 gene sequences; 1KP project (Matasci et al., 2014) and (diploid) *Vaccinium macrocarpon* (cranberry) (48, 270 sequences, NCBI) (both Ericaceae subfamily Ericoideae; Ericales); and (except for MarkerMiner) WGS of *V. macrocarpon* (NCBI) and *Erica plukenetii* (Le Maitre & Bellstedt, unpublished data). The (potential) length and identity of the resulting targets was compared.

In order to compare these “made to measure” (taxon-specific) targets with those that might be selected using a more “one size fits all” (universal) approach to probe design, we compared

transcriptomes from more distantly related plants 1) of Ericales (*Actinidia chinensis* [Actinidiaceae; 10,000 sequences, NCBI], *Aegiceras corniculatum* [Primulaceae; 49,412 sequences, NCBI], *Camellia reticulata* [Theaceae; 139,145 sequences, NCBI], *Diospyros lotus* [Ebenaceae; 413, 775 sequences, NCBI], *R. scorpulum and V. macrocarpon*); and 2) of eudicots (*Anemone flaccida* [Ranunculales; 46,945 sequences, NCBI], *Dahlia pinnata* [Asterales; 35,638 sequences, NCBI], *Gevuina avellana* [Proteales; 185,089 sequences, NCBI], *Mesembryanthemum crystallinum* [Caryophyllales; 24,204 sequences, NCBI], *Solanum chacoense* [Solanales; 42,873 sequences, NCBI], *Vigna radiata* [Fabales; 78,617 sequences, NCBI], *Vitis vinifera* [Vitales; 52,310 sequences, NCBI] and *R. scorpulum).* Because in this wider context it is no longer appropriate to identify single copy markers on the basis of Ericoideae data alone, we instead used the option to compare to the angiosperm-wide database (De Smet et al., 2013) following an approach similar to MarkerMiner (Chamala et al., 2015). We compared the resulting targets to those of the *Erica*-specific approach, as above.

Generation of a novel empirical dataset

In order to confirm that our scripts can be used to obtain datasets of single-copy markers, we applied them to our empirical study on Cape *Erica*. We used the 132 sequences resulting from our custom scripts, taking into account the potential intron lengths (see results and discussion).

In addition to these targets, we made a small number of ad-hoc modifications of the final dataset, adding further sequences that were not otherwise selected as optimal with the above scripts to the final probe design datasets for the purpose of comparison with other datasets. Two additional targets were rpb2 (as used in phylogenetic reconstruction in *Rhododendron;* Goetsch, Eckert & Hall, 2005) and topoisomerase B (as proposed for use across flowering plants; Blattner, 2016).

Laboratory methods: Plant material was collected in the field under permit (Cape Nature: 0028-AAA008-00134; South Africa National Parks: CRC-2009/007-2014) or obtained from cultivation. DNA was extracted from one sample of *Rhododendron camtschaticum*, supplied by Dirk Albach and Bernhard von Hagen from collections of the Botanic Garden, Carl von Ossietzky Universität, Oldenburg, Germany; and 12 of *Erica* (Table 1) using Qiagen DNAeasy

kits (Qiagen, Hilden, Germany). DNA extraction in *Erica* is generally challenging (Bellstedt et al., 2010) and the quantity and quality of DNA obtained differed considerably between species. To reach the correct amount of DNA required for library preparation, multiple DNA extractions from the same sample were combined.

For library preparation and hybridisation enrichment, we used the Agilent SureSelectXT protocol (G7530-90000), incorporating sample-specific indexes for pooled sequencing. For the library preparation, amount of gDNA used was between 1 and 3 µg, and during the hybridisation and capture step, we used a diluted capture library (1 part Agilent baits solution to 4 of ddH_2O). Sequencing was performed with Illumina NextSeq500 (StarSeq, Mainz, Germany) to generate 25 million paired-end reads of length 150 bp.

Bioinformatic analysis: As the total footprint of selected targets was small, *de novo* assembly was possible. We chose to use MIRA (version 4.0) (Chevreux, Wetter & Suhai, 1999), in part because MIRA can be used to perform both *de novo* assembly and mapping. The two options were used with default parameters for Illumina (overlap value=80 for *de novo* and 160 for mapping assembly; quality level=accurate). Reads were assembled into contiguous sequences (contigs). We then compared using BLASTn against the sequence targets (complete sequences and coding region sequences) as well as against nuclear ribosomal (nrDNA), plastid, and mitochondrial data. Contigs for which overlap with targets was under 100 bp and similarity to target sequences was less than 75% were removed. Using the L-INS-i (iterative refinement method incorporating local pairwise alignment information) method of MAFFT (Katoh et al., 2002), we aligned contigs with each other and with the sequence targets (complete sequences and coding region sequences). Contigs were checked with Gap5 (https://doi.org/10.1093/bioinformatics/btq268) and by comparison to the alignments to identify and confirm remaining separate overlapping contigs without sequence differences. We used custom made scripts to merge and remove redundant contigs, combining only those with identical overlapping sequences (minimum overlap of 30 bp) or which differed by a single base only (in which case this position was coded with IUPAC ambiguity codes). Contigs differing by more than one base or which did not overlap were not combined. This should avoid combining non-continuous contigs representing different copies or alleles at the cost of tending to

overestimate the numbers of such copies where overlap of contigs is incomplete. We then attempted to add to the alignments any <100 bp sequences or sequences under 75% similarity that matched the target according to BLASTn, combining (or not) contigs using the same principles as above.

We excluded alignment positions representing indels or missing data in one or more samples and then calculated the percentage of variable sites per marker, including combined mitochondrial and plastid sequences and individual nrDNA sequences representing Internal and External Transcribed Spacer regions (ITS and ETS) as obtained using Sanger sequencing in previous work (Pirie, Oliver & Bellstedt, 2011; Pirie et al., submitted). Gene trees were inferred using RAxML (Stamatakis, 2014) and used as a rough test for potential paralogy, under the assumption that the ingroup (comprising all samples except *Rhododendron* and the more closely related outgroups *Erica abietina and Erica plukenetii*) is monophyletic.

Results

Similarity, length and overlap of selected markers: "made to measure" versus "one size fits all"

The lengths of sequences selected using the different scripts are presented in Fig. 2. Summary comparisons by method are presented in Table 2 (sequence numbers, lengths and similarity). In general, the additional filter that includes mean intron length resulted in an increased number of shorter targets that might nevertheless deliver greater final sequence lengths, if average lengths of flanking introns are effectively captured (Fig. 2).

Made to measure: We identified 4649 potential markers using our custom script AllMarkers.py. Applying script BestMarkers.py to this pool to optimise for length, two different subsets of optimal markers were obtained: 132 with median length (of coding region) of 2,187 bp when taking intron lengths into account; 79 of median length 2,631 bp when not. Sequence identity was similar (Table 2).

With the Hyb-Seq pipeline, 782 sequences were obtained, which after applying BestMarkers.py, was reduced to 55 of median length 2,157 bp when taking introns into account and 66 of median length 2,184 bp when not. Sequence identity was similar, and similar to that resulting from AllMarkers.py (Table 2).

With MarkerMiner, target sequences are delivered separately for each transcriptome provided. We selected a total pool of 544 potential target sequences, of which 389 are represented in the *R. scopulorum* data and 222 in *V. macrocarpon*. By comparison using our own scripts (available on request) we identified just 67 that were common to both (whereby it should be noted that AllMarkers.py by default retains only those found in at least two transcriptomes). Of the 544 sequences, 519 are indicated by MarkerMiner as mostly single copy and 25 as strictly single copy in angiosperms. After applying BestMarkers.py we retained 254 sequence targets when taking introns into account and 207 sequences when not. Use of MarkerMiner resulted in the selection of greater numbers of shorter and slightly more conserved markers compared to both AllMarkers.py and HybSeq (Table 2, Figs. 2-3).

One size fits all: Applying AllMarkers.py/BestMarkers.py to transcriptomes of Ericales resulted in a pool of 2,354 potential markers and final datasets of 409 sequences when taking introns into account and 171 when not. With the Eudicot transcriptomes, the total pool included 461 potential markers and final datasets 249 (when taking introns into account) and 130 sequences (when not) (Table 2). In the latter, there is a slight increase in similarity (≥85%, similar to MarkerMiner; Fig. 3), and in both, sequences are shorter (Table 2, Fig. 2).

The numbers of markers in common given the different methods for selecting them, before and after applying BestMarkers.py are presented in Fig. 4. Fig. 4a illustrates both the low overlap and large differences in numbers between the complete pools of potential markers identified using the different methods/input data. Expanding in taxonomic scope from *Erica* (identifying single-copy genes on the basis of WGS data) to Ericales and to eudicots (adopting single copy markers from the database of De Smet et al. (2013) resulted in a decrease in numbers of potential markers, and the use of MarkerMiner a further decrease. Fig. 4b illustrates the differences in the optimal markers selected using BestMarkers.py on these pools. There is limited overlap and considerable differences in both target numbers and lengths: overall, AllMarkers.py/BestMarkers.py and HybSeq delivered the longest sequences, whereby the former delivered more markers for the same number of baits. Both the Ericales and eudicot analyses and MarkerMiner delivered greater numbers of shorter sequences.

Empirical data

We performed selective enrichment of 134 markers. Exon sequences used for probe design are presented in supplementary data 1 and sequence alignments in supplementary data 2. With the exception of a single marker, capture was equally effective in the single *Rhododendron* sample and thirteen *Erica* samples. One marker was captured only in *Rhododendron*, and two others was not captured at all. All of the remaining 129 markers plus rpb2 and topoisomerase B were recovered from all thirteen samples analysed. Of these, 6 were single copy without allelic variation; 83 included sequence polymorphisms corresponding to two distinguishable putative alleles in one or more (but not all) individual samples. A further 40 included sequence polymorphisms in all samples which exhibited two or more copies. Of the latter 40, 28 represented paralogs that were easily distinguished on the basis of high sequence divergence in one or more coding region and could thus be segregated into separate matrices of homologous sequences. The remainder (12) included multiple contigs that could not obviously be combined into single homologous sequences or pairs of alleles. Inspection of individual gene trees failed to reject the monophyly of the ingroup in all but five cases.

Comparison of sequence length/variability was limited by uneven sequencing coverage, but we could confirm the capture of complete intron sequences of up to c. 1000 bp and partial introns/flanking non-coding regions of up to c. 500 bp. In addition, large stretches of homologous high copy nuclear ribosomal and mitochondrial sequences were captured for all samples, as well as more fragmented plastid sequences.

Despite incomplete sequencing coverage, the average alignment length of single copy nuclear sequences was 1810 bp, with a range between 823 and 5574 bp. With all gaps and missing data excluded (resulting in alignments of between 327 and 4716 bp), the single copy nuclear sequences presented between 5 and 412 variable positions each, representing a range of 0.5-18% variability. Variability of rpb2 was 3.4%; topoisomerase B: 7.5%; ETS: 22.1%; ITS: 17.9%; mitochondrial: 6.3%; and plastid sequences: 0.54%. A plot of original predicted length of markers (instead of real length since in most cases complete sequences were not obtained) against variability is presented in Fig. 5. There was no obvious relationship between sequence length and variability. A further plot of observed sequence variability against variability of the corresponding transcriptome data (*Rhododendron* compared to *Empetrum*) is presented in Appendix 1; there was also no obvious relationship. Gene trees inferred under ML are documented in Supplementary Material 3 (with further details in Supplementary Material 4),

with six based on selected markers (ITS, mitochondrion, and four single copy nuclear markers that provided the greatest numbers of variable characters) illustrated in Fig. 6.

Discussion

Comparing closely versus distantly related genomes for marker selection

It seems intuitively obvious that optimal markers for a given phylogenetic problem will be those informed by comparison to transcriptomes/WGS of the most closely related representative taxa. With such data, lineage specific gene duplications can be identified and the number of potential targets of appropriate variability maximised. However, the genomic data available for a given focal group (such as transcriptome data from the 1KP project; Matasci set al., 2014) may represent taxa more or less distantly related to it, and particular researchers may or may not wish to go to the trouble of designing and applying custom protocols. Indeed, if an off-the-shelf tool will provide appropriate data, it would be a great deal simpler just to use it. Hence, before embarking on expensive and time-consuming lab procedures, we need to know to what degree targets designed for one group might be applied to more distantly related ones (e.g. in this case the utility of *Erica* baits across Ericaceae, or Ericales); and conversely, how suboptimal baits designed for universal application (e.g. across angiosperms) are likely to be for a given subclade.

Using our own custom scripts, we compared the pools of markers that might be selected on the basis of comparison of relatively closely related genomes with those on the basis of more distantly related ones (i.e. within the subfamily Ericoideae as opposed to within the order Ericales or across eudicots). Our results showed that both the pools and the best marker sets from those pools differed considerably, and that the sequences of the latter were considerably shorter (Table 2, Figs. 2 and 3). On the other hand, sequence variability within Ericales (minimum sequence identity between Ericaceae and Actinidiaceae: 73%) suggests that baits designed for *Erica* are also potentially suited for use at least across Ericaceae, including in *Rhododendron* and *Vaccinium* (both species-rich genera for which such tools might be particularly useful (Kron, Powell & Luteyn, 2002; Goetsch, Eckert & Hall, 2005). In general, our results confirm both the greater potential of custom baits developed for specific clades; and show that once obtained, such tools are nevertheless likely to apply across a fairly broad range of related taxa.

419

420 *The impact of method for marker selection*

421 Having decided to design custom baits, the next question that we might ask is which method to
422 use for probe selection/design. Our results suggest that this is also likely to have a significant
423 impact on the resulting datasets. We compared three approaches to marker selection: our own
424 custom scripts; those presented in the Hyb-Seq approach (Weitemier et al., 2014) and
425 MarkerMiner (Chamala et al., 2015).

426 Of these three, MarkerMiner is arguably the most user-friendly, which is important given that its
427 user base ought ideally to include biologists without extensive bioinformatics skills. However, in
428 our comparisons it fared poorly, delivering the lowest sequence lengths (Table 2). The reasons
429 for this are two-fold: first (and perhaps most importantly), because the transcriptomes used,
430 irrespective of their similarity one to another, are compared to what is likely to be a rather
431 distantly related proteome; second, because the approach for identifying single or low-copy
432 markers involves comparison to a general database (in this case for flowering plants), rather than
433 a case-by-case assessment. Hence, in its current implementation it is to be expected that the most
434 variable sequences will be excluded, as will some that are single copy in the focal group (or with
435 easily discerned paralogs, as was the case here and also at lower taxonomic levels in Budenhagen
436 et al. 2016); and that some that are not single copy in that group will in fact be included. This is
437 reflected in our results by the low number of potential target sequences recovered in total; in the
438 low proportion of those that were recovered also being recovered using our own custom scripts
439 and Hyb-Seq; and in the lower sequence length: the removal of more variable sequences
440 arbitrarily results in the removal of longer ones too (Table 2). This phenomenon is apparently
441 also reflected in the even shorter sequences reported by Budenhagen et al. (2016), using
442 universal angiosperm probes (average 764 bp, derived from targets averaging 343 bp).

443 The Hyb-Seq approach is more similar to our own, but nevertheless results in a different dataset
444 of selected sequences. The main differences lie in the search tool and filters. Our script uses
445 BLAST, whereas Hyb-Seq uses BLAT. BLAT is faster than BLAST, but needs an exact or
446 nearly-exact match to return a hit. Significantly, the exclusion in HybSeq of all sequences
447 including any exons <120 bp is at the loss of markers including variable introns; in our approach
448 the problem of short exon/probe mismatch is avoided simply by ignoring such exons during

449 probe design. The net result is that while both approaches deliver long target sequences, ours can
450 deliver those including more introns (which can therefore be captured using fewer baits).

451

452 *Selecting optimal markers from within a pool of potential candidates*

453 Our approach includes not just a means to select potentially appropriate markers (AllMarkers.py;
454 as is the case with the other approaches compared) but also a second step (BestMarkers.py) that
455 selects putatively optimal markers from amongst that pool. Obviously, it is possible to capture
456 and sequence the entire pool (following Ilves & Lopez-Fernandez, 2014; Mandel et al., 2014;
457 Weitemier et al., 2014). However, by targeting the most appropriate markers, more samples can
458 be analysed less expensively (by dilution of the baits), whilst avoiding expending sequencing
459 effort on a potentially large number of less informative (or perhaps even entirely uninformative)
460 markers.

461 Optimising for intron numbers/length, as implemented in BestMarkers.py would seem
462 appropriate for the purpose of identifying regions that are likely to be both longer and more
463 variable (Folk, Mandel & Freudenstein, 2015): hybrid capture can result in sequencing of
464 potentially long stretches of flanking regions (Tsangaras et al., 2014) without requiring matching
465 baits, and introns should be less constrained, possibly with informative length variation too.
466 Hence, taking into account the additional length of introns in marker selection can result in
467 greater numbers of longer (and likely more variable) obtained sequences. Our empirical results
468 support this approach: sequences showed intron capture of up to 1,000 bp, including regions in
469 which multiple introns are interspersed with short (<120 bp) exons for which no probes were
470 used. Alternatively, if the problem to be addressed represents older divergences (e.g.
471 phylogenetic uncertainty within Ericaceae; Freudenstein, Broe & Feldenkris, 2016) for which
472 length variation in introns would be unhelpful, BestMarkers.py can be used to optimise the
473 length of exons alone.

474 An alternative to optimising for sequence length (with or without taking introns into account)
475 would be to optimise for variability (or combined length and variability). We included this
476 option in BestMarkers.py, but in the absence of data with which to compare within our ingroup,
477 decided *a priori* that we would be more likely to optimise total per sequence variation by
478 selecting on the basis of length alone. This decision was supported by the empirical results: as

might be expected, there was no obvious relationship between sequence length and variability (Fig. 5) and the numbers of informative characters provided by a given target could not be predicted from the similarity of the *Vaccinium* and *Rhododendron* transcriptomes (Appendix 1).

The variability of the data we obtained can be compared to that of nrDNA, plastid and mitochondrial sequences (and which were also obtained here without the need for matching baits due to their high copy number) and to two generally single copy nuclear genes, topoisomerase B and rpb2 (Fig. 5). Consistent with the results presented by Nichols et al. (2015), the variability of the nrDNA spacer regions (ITS and ETS) that are frequently used in empirical studies of plants is at the upper end of that observed in the sequences we obtained (of which topoisomerase B and rpb2 were fairly typical); plastid (and mitochondrial) sequences at the lower end. Given the comparably modest variability of most alternative nuclear markers, this suggests that even in cases where ITS/ETS present sufficient information to infer a well resolved nrDNA gene tree (not the case in Cape *Erica*, Pirie et al., 2011; Fig. 6), considerably longer sequences will be needed to infer comparably resolved independent gene trees. Difficult phylogenetic problems arise when gene trees can be expected to differ, but those inferred from standard markers are not sufficiently resolved to actually reveal it. These are the cases for which targeted capture approaches offer the greatest potential. However, even large numbers of independent markers, if insufficiently variable, may be dominated in phylogenomic analyses by the signal of relatively few (Lanier, Huang & Knowles, 2014). We need to target markers that might deliver a forest of trees, rather than just more bushes, and not all targeted enrichment strategies are optimised to deliver this kind of data.

Conclusions

When sequence variation is appropriate and gene trees are consistent, standard Sanger sequencing of a small number of markers may be all that is required to infer robust and meaningful phylogenetic trees. For species complexes and rapid radiations (either ancient or recent) where this is not the case, the usefulness of sequence datasets will inevitably be limited by the resolution of individual gene trees. Our results suggest that under these circumstances, where the need for NGS and targeted sequence capture, such as hybrid enrichment, is greatest, “made to measure” markers identified using both transcriptome and WGS data of related taxa

509 will deliver results that are superior to those that might be obtained using a more universal "one
510 size fits all" approach. Once available, such markers may nevertheless be useful across a fairly
511 wide range of related taxa: e.g. those presented here, targeted for use in *Erica*, fall within the
512 range of sequence variation that would in principle be applicable across the family Ericaceae.
513 Transcriptome data for many flowering plant groups are now available; these would ideally be
514 complemented with WGS or genome skimming data of one or more focal taxa for use in marker
515 selection. With such data to hand, biologists are still reliant on bioinformatics skills or user-
516 friendly tools (such as MarkerMiner). In either case, the full potential of the techniques will only
517 be harnessed if comparisons to distantly related genomes and generalisations of single/low copy
518 genes across wide taxonomic groups are avoided. We would conclude that rather than searching
519 for "one size fits all" universal markers, we should be improving and making more accessible the
520 tools necessary for developing our own "made to measure" ones.

521

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719

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Figure 1(on next page)

Bioinformatic methods flowcharts

Fig 1 - Flowchart(s) illustrating the methods used for marker selection.

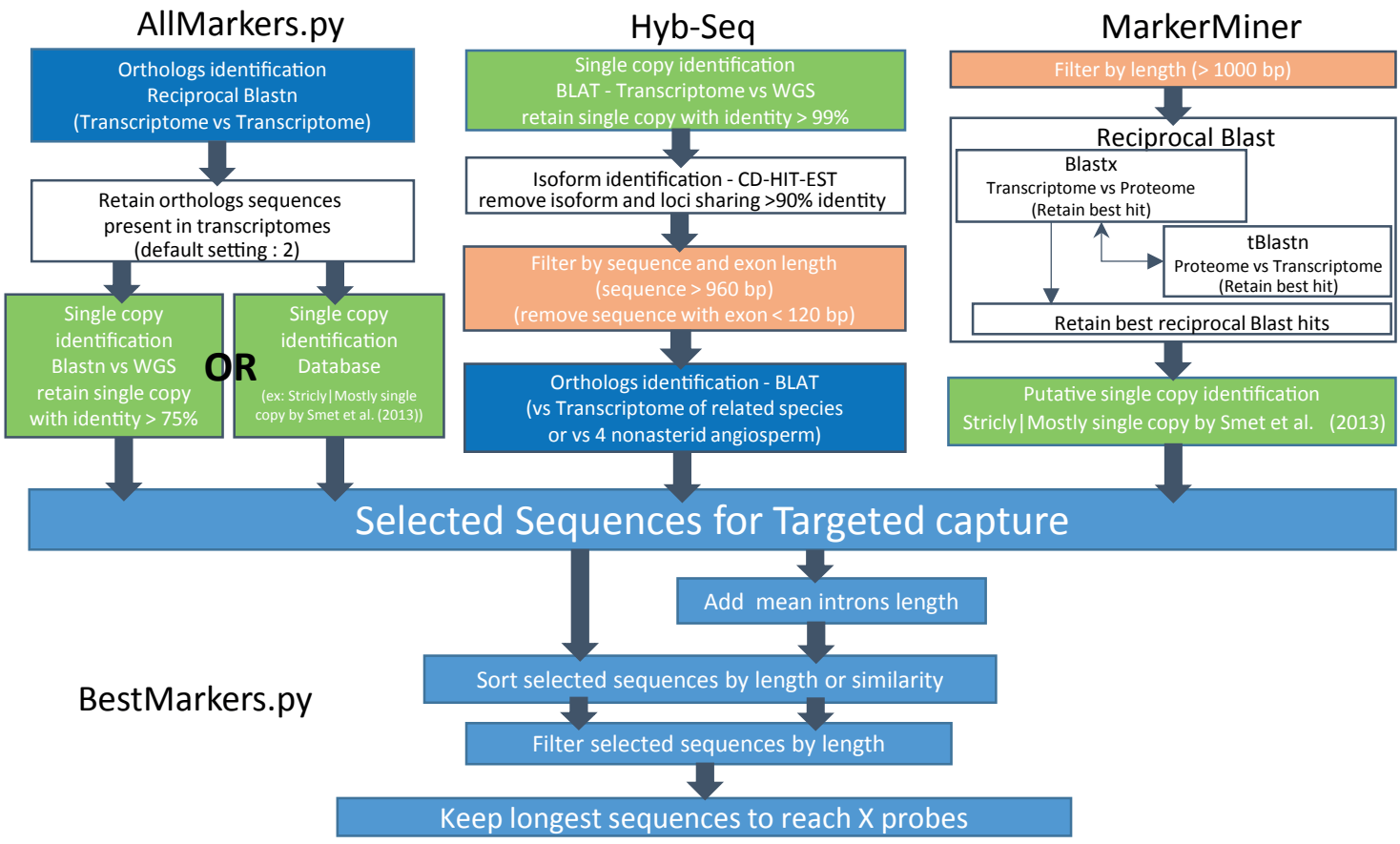

AllMarkers.py
Orthologs identification
Reciprocal Blastn
(Transcriptome vs Transcriptome)
Retain orthologs sequences
present in transcriptomes
(default setting : 2)
Single copy
identification
Blastn vs WGS
retain single copy
with identity > 75%
OR
Single copy
identification
Database
(ex: Stricly|Mostly single
copy by Smet et al. (2013))
Hyb-Seq
Single copy identification
BLAT - Transcriptome vs WGS
retain single copy with identity > 99%
Isoform identification - CD-HIT-EST
remove isoform and loci sharing >90% identity
Filter by sequence and exon length
(sequence > 960 bp)
(remove sequence with exon < 120 bp)
Orthologs identification - BLAT
(vs Transcriptome of related species
or vs 4 nonasterid angiosperm)
MarkerMiner
Filter by length (> 1000 bp)
Reciprocal Blast
Blastx
Transcriptome vs Proteome
(Retain best hit)
tBlastn
Proteome vs Transcriptome
(Retain best hit)
Retain best reciprocal Blast hits
Putative single copy identification
Stricly|Mostly single copy by Smet et al. (2013)
Selected Sequences for Targeted capture
BestMarkers.py
Add mean introns length
Sort selected sequences by length or similarity
Filter selected sequences by length
Keep longest sequences to reach X probes

Figure 2(on next page)

Summary of selected exon/predicted marker lengths by method

Fig 2 - Summary of a) exon lengths and b) predicted exon plus intron lengths of markers selected using AllMarkers.py (shades of green), Hyb-Seq (blue) and MarkerMiner (purple) followed by BestMarkers.py. Each pair of plots represents the markers selected when optimising for exon lengths (left) and predicted exon plus intron lengths (right). From left to right, the first three pairs represent markers targeted for Erica/Ericoideae (comparing by method); the final two for Ericales and eudicots respectively (using AllMarkers.py only)

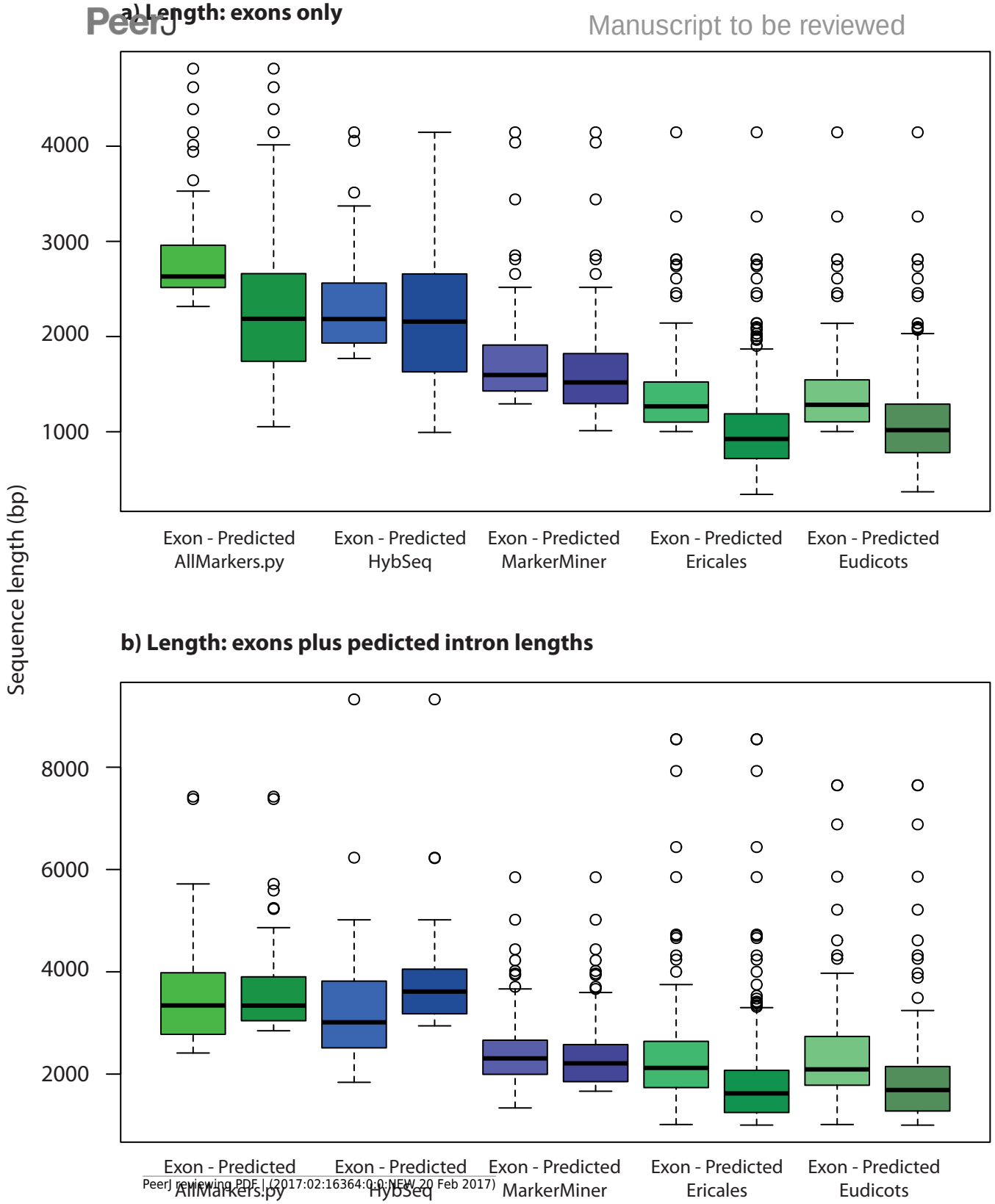

a) Length: exons only
Sequence length (bp)
4000
3000
2000
1000
Exon - Predicted AllMarkers.py
Exon - Predicted HybSeq
Exon - Predicted MarkerMiner
Exon - Predicted Ericales
Exon - Predicted Eudicots
b) Length: exons plus pedicted intron lengths
8000
6000
4000
2000
Exon - Predicted AllMarkers.py
Exon - Predicted HybSeq
Exon - Predicted MarkerMiner
Exon - Predicted Ericales
Exon - Predicted Eudicots

Figure 3(on next page)

Length versus variability of potential and selected sequence markers

Fig 3 - Length versus variability of potential sequence markers (grey dots) and those selected using BestMarkers.py from the pools generated by the different methods (coloured symbols).

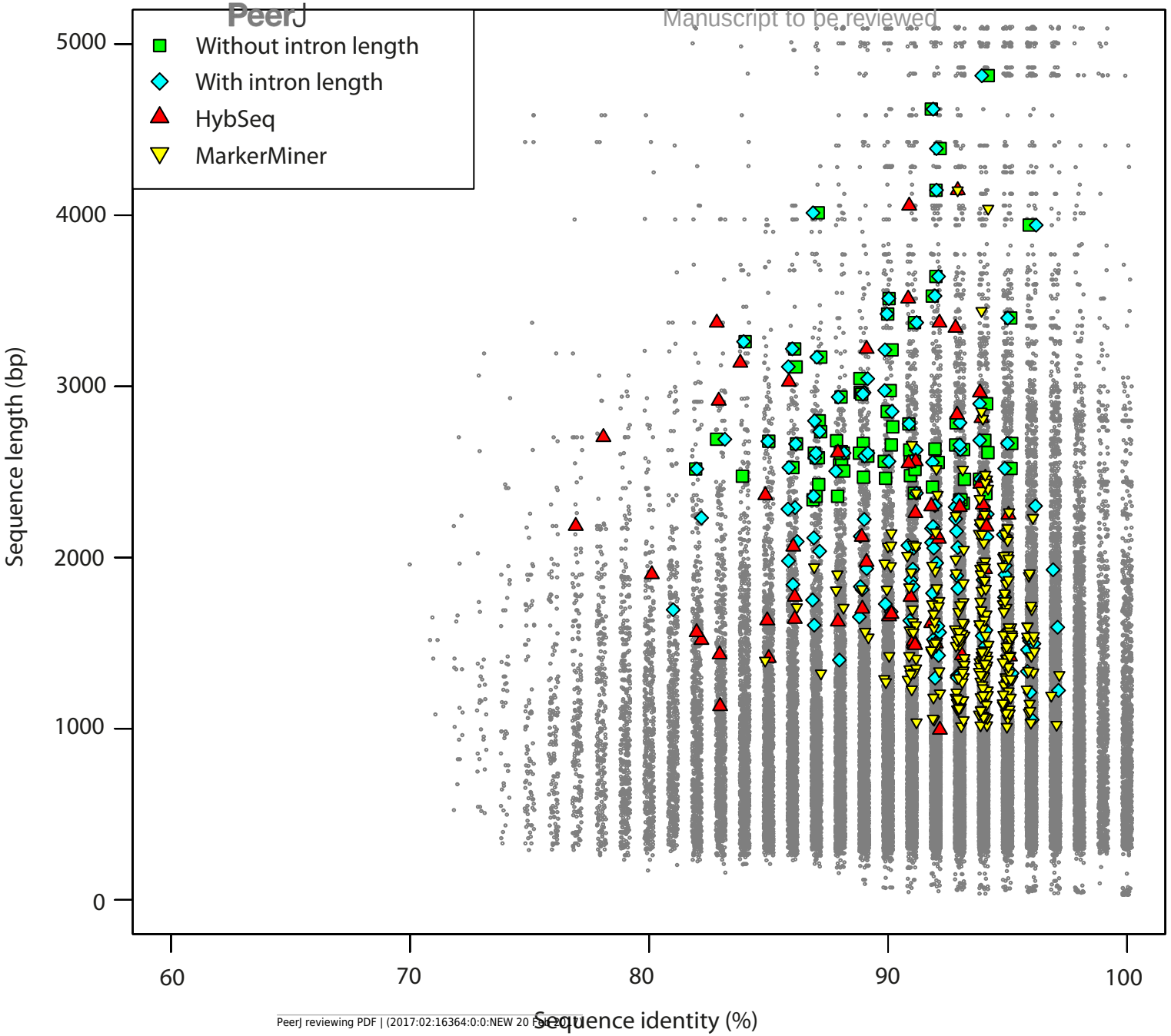
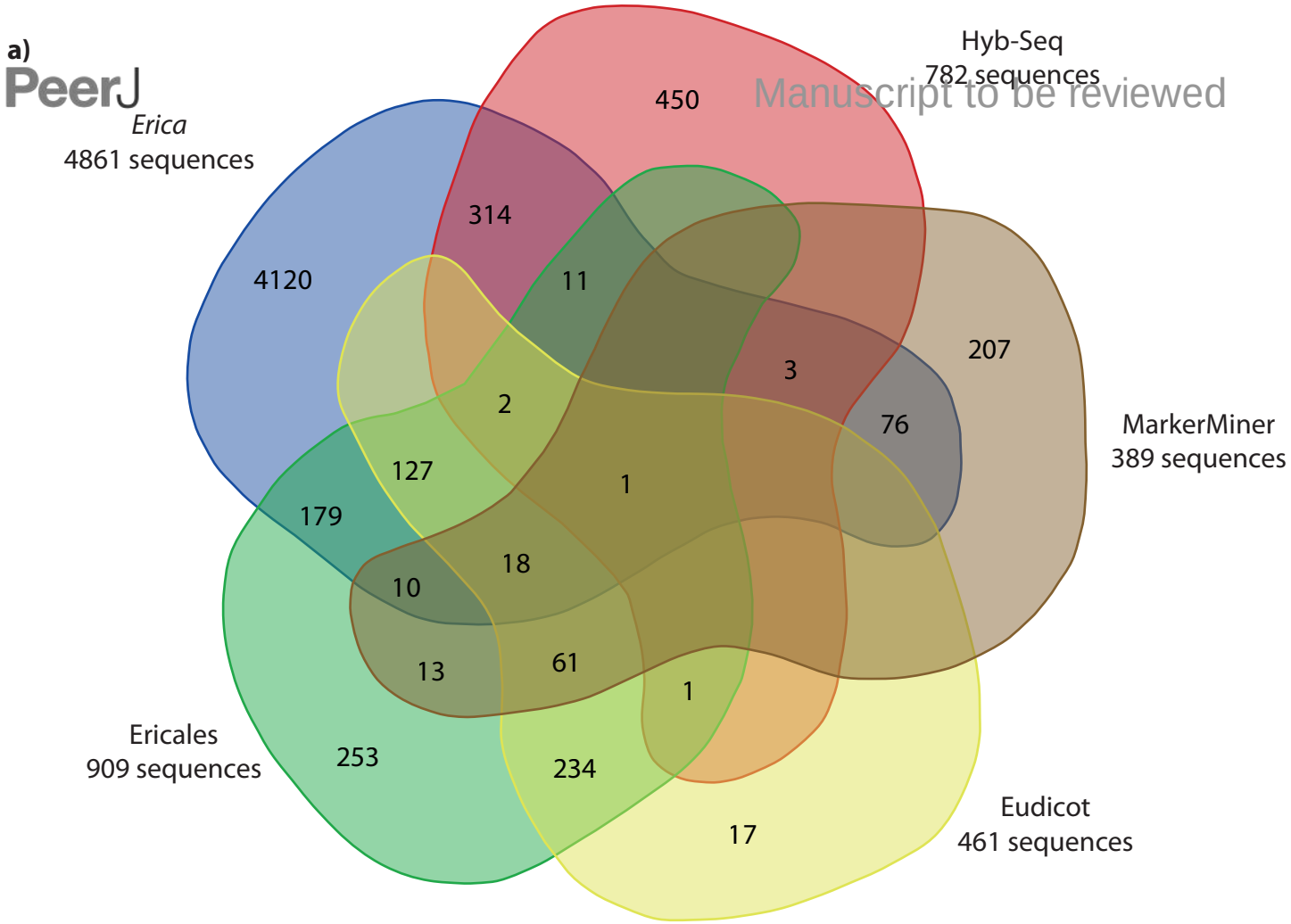


Figure 4(on next page)

Overlap of selected markers by method

Fig 4 - Venn diagrams produced using http://bioinformatics.psb.ugent.be/webtools/Venn/ comparing overlap in markers selected given the different methods, superimposed with their numbers. a) The complete pools of potential markers; b) the subsets of markers selected using BestMarkers.py, optimising for total predicted length (exons and introns).



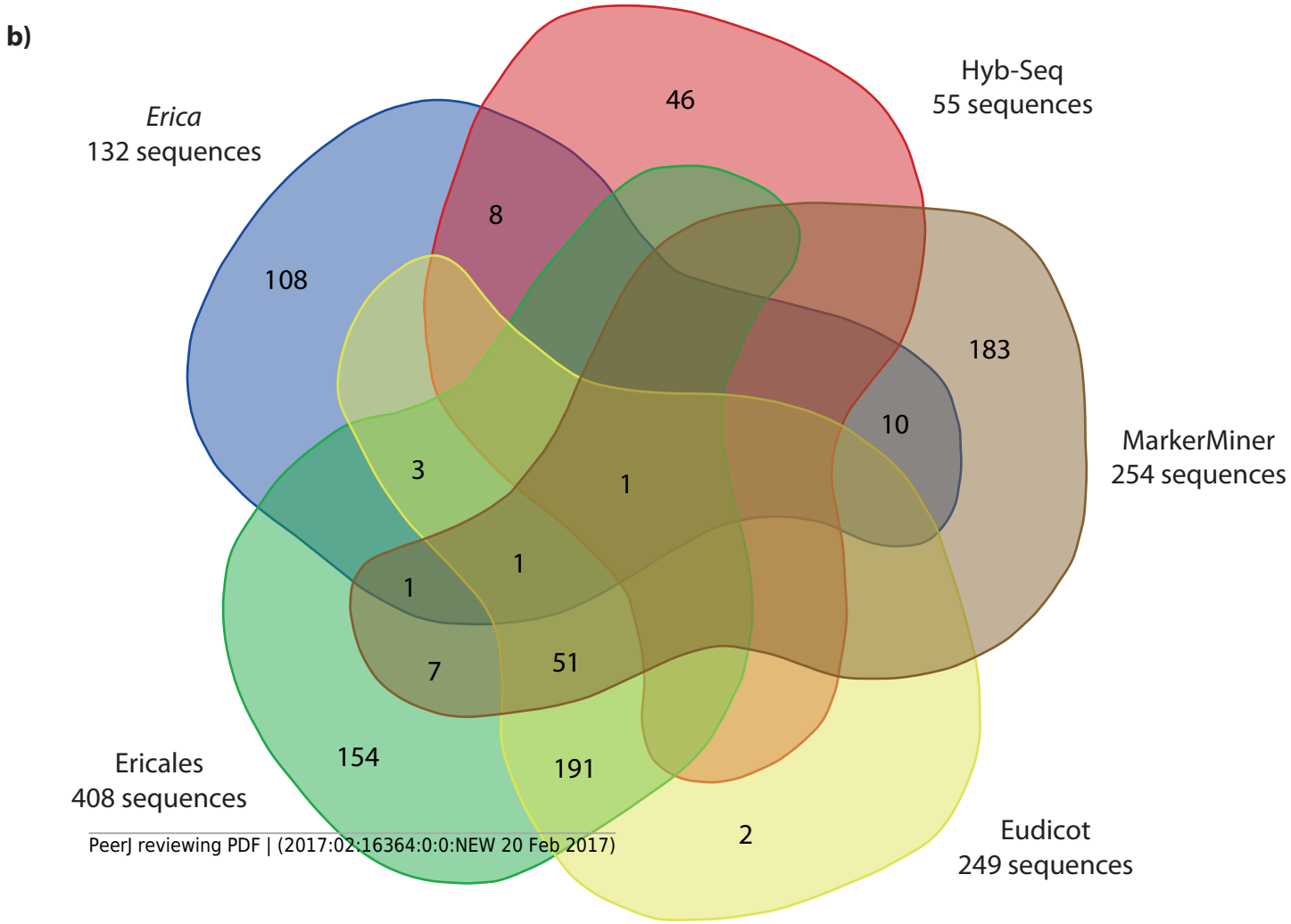


Figure 5(on next page)

Plot of observed variability against predicted sequence length

Fig 5 - Sequence variability observed in the empirical data plotted against predicted sequence length. “Universal” markers rpb2 and topoisomerase B are indicated and plastid, mitochondrial and nrDNA are included with indication of sequence lengths derived from the literature.

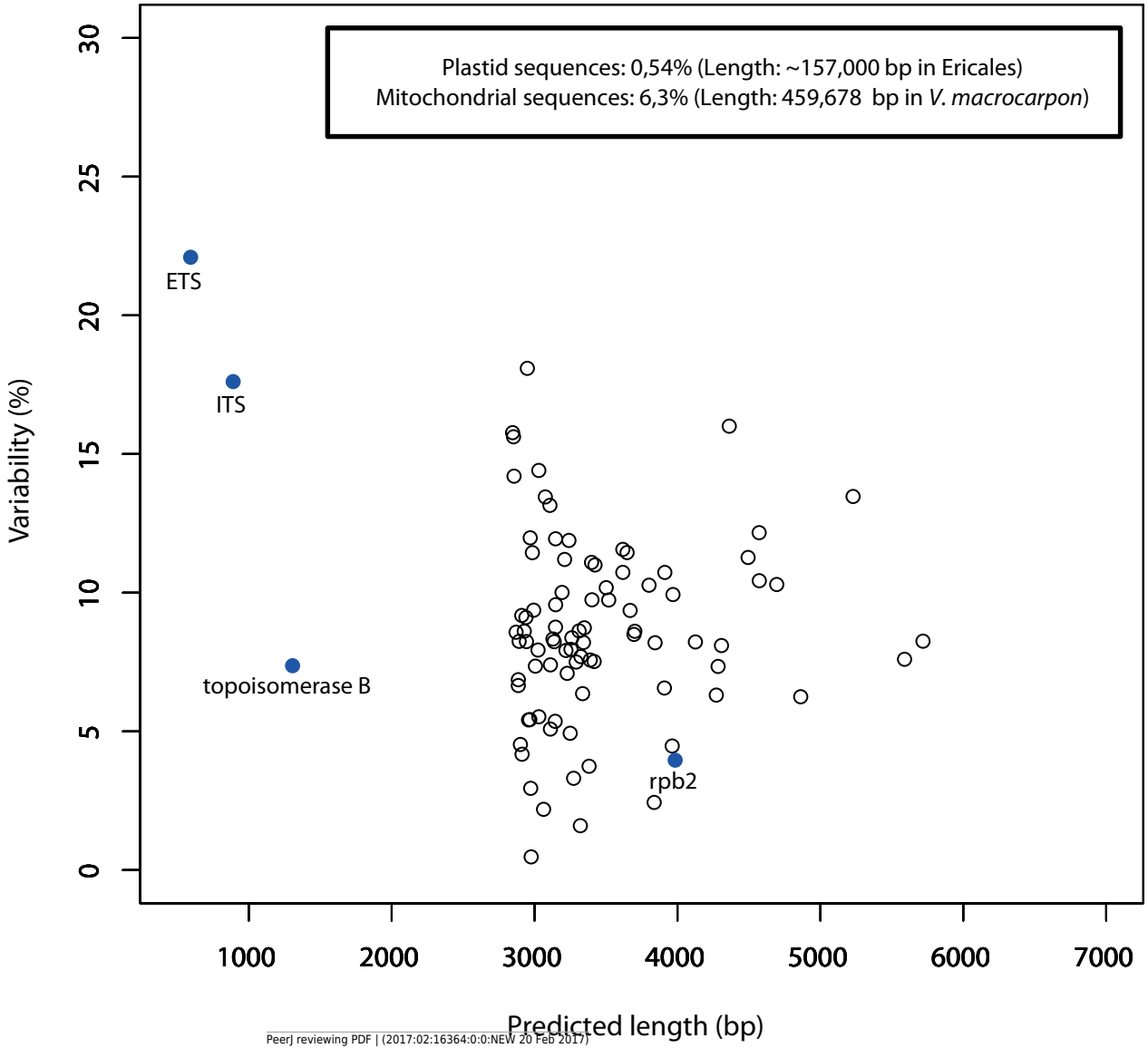
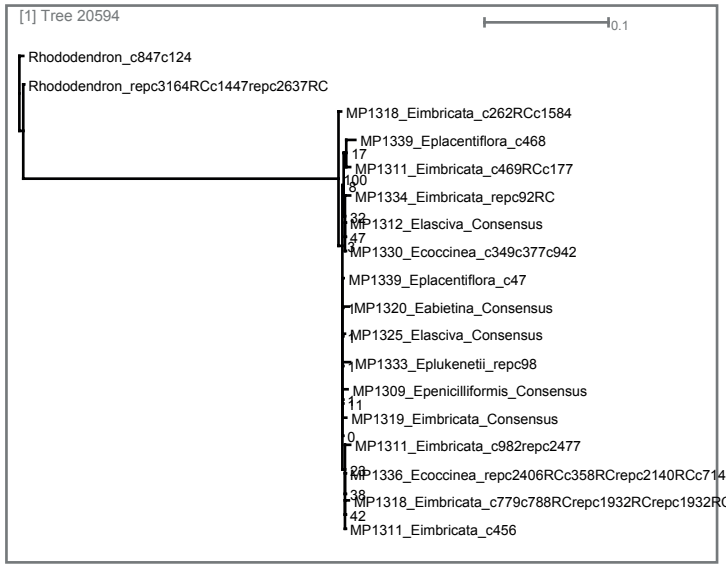
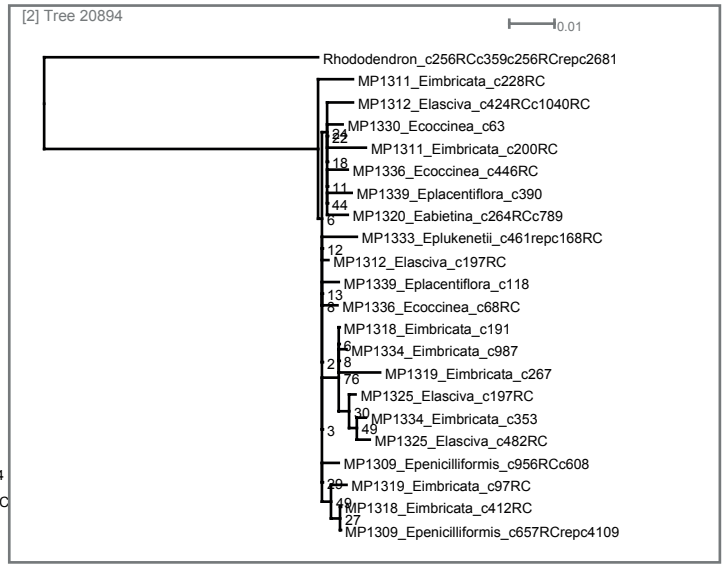

Plastid sequences: 0,54% (Length: ~157,000 bp in Ericales)
Mitochondrial sequences: 6,3% (Length: 459,678 bp in V. macrocarpon)
ETS
ITS
topoisomerase B
rpb2
Variability (%)
0
5
10
15
20
25
30
1000
2000
3000
4000
5000
6000
7000
Predicted length (bp)

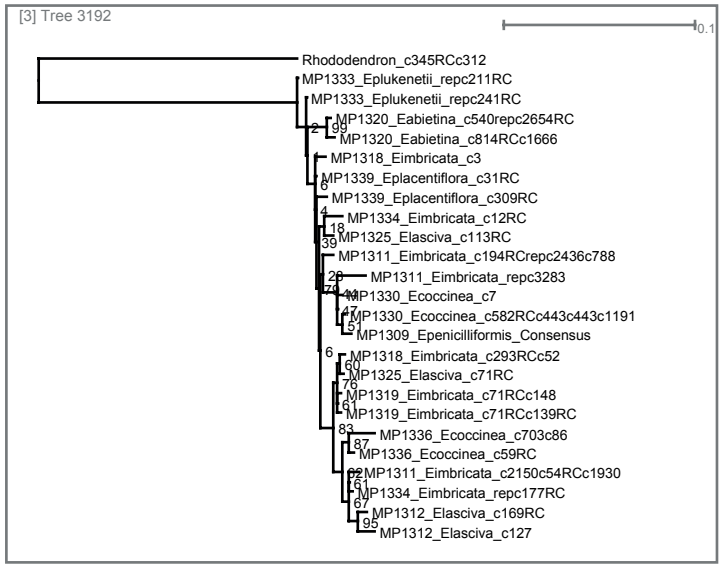
Figure 6(on next page)

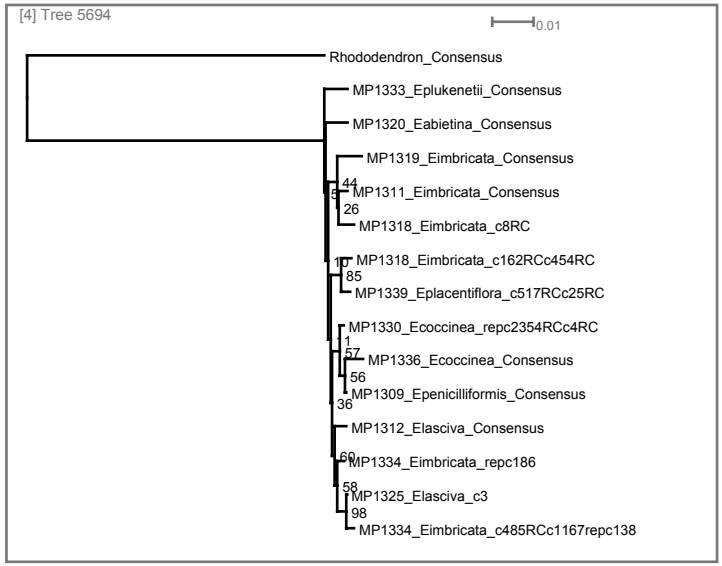
Selected gene trees

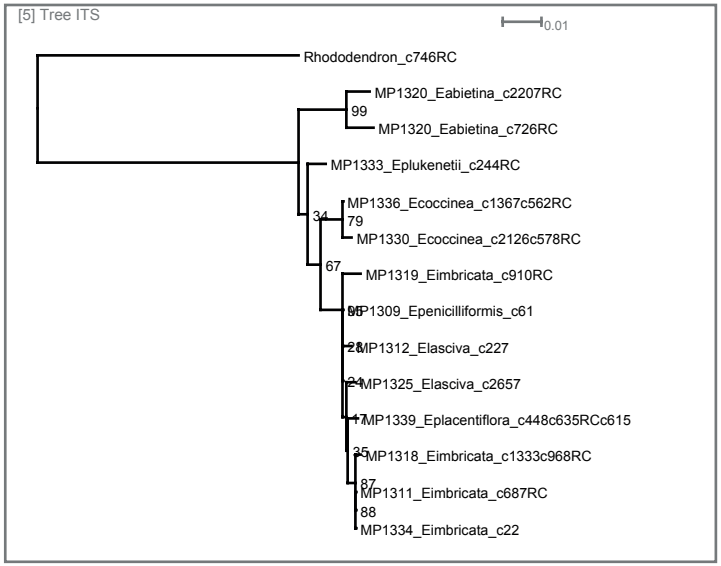
Fig 6 - Selected gene trees inferred under maximum likelihood with RAxML, presented using Dendroscope 3.5.7 (http://dendroscope.org/). The four nuclear markers that showed the highest numbers of variable characters are presented along with those based on ITS and mitochondrial sequences. Terminals correspond to species names and collection codes (Table 1) appended by codes corresponding to one or more contigs that were merged using custom scripts. Some taxa are represented twice in some trees due to the presence of alleles, including two distinct copies of ITS in *E. abietina* ssp. *aurantiaca* (confirming previous work using cloning; Pirie et al., submitted). Scale bars represent substitutions per site, branch labels represent bootstrap support.


[1] Tree 20594
0.1
Rhododendron_c847c124
Rhododendron_repc3164RCc1447repc2637RC
MP1318_Eimbricata_c262RCc1584
MP1339_Eplacentiflora_c468
MP1311_Eimbricata_c469RCc177
MP1334_Eimbricata_repc92RC
MP1312_Elasciva_Consensus
MP1330_Ecoccinea_c349c377c942
MP1339_Eplacentiflora_c47
MP1320_Eabietina_Consensus
MP1325_Elasciva_Consensus
MP1333_Eplukenetii_repc98
MP1309_Epenicilliformis_Consensus
MP1319_Eimbricata_Consensus
MP1311_Eimbricata_c982repc2477
MP1336_Ecoccinea_repc2406RCc358RCrepc2140RCc714
MP1318_Eimbricata_c779c788RCrepc1932RCrepc1932RC
MP1311_Eimbricata_c456


[2] Tree 20894
0.01
Rhododendron_c256RCc359c256RCrepc2681
MP1311_Eimbricata_c228RC
MP1312_Elasciva_c424RCc1040RC
MP1330_Ecoccinea_c63
MP1311_Eimbricata_c200RC
MP1336_Ecoccinea_c446RC
MP1339_Eplacentiflora_c390
MP1320_Eabietina_c264RCc789
MP1333_Eplukenetii_c461repc168RC
MP1312_Elasciva_c197RC
MP1339_Eplacentiflora_c118
MP1336_Ecoccinea_c68RC
MP1318_Eimbricata_c191
MP1334_Eimbricata_c987
MP1319_Eimbricata_c267
MP1325_Elasciva_c197RC
MP1334_Eimbricata_c353
MP1325_Elasciva_c482RC
MP1309_Epenicilliformis_c956RCc608
MP1319_Eimbricata_c97RC
MP1318_Eimbricata_c412RC
MP1309_Epenicilliformis_c657RCrepc4109


[3] Tree 3192
0.1
Rhododendron_c345RCc312
MP1333_Eplukenetii_repc211RC
MP1333_Eplukenetii_repc241RC
MP1320_Eabietina_c540repc2654RC
MP1320_Eabietina_c814RCc1666
MP1318_Eimbricata_c3
MP1339_Eplacentiflora_c31RC
MP1339_Eplacentiflora_c309RC
MP1334_Eimbricata_c12RC
MP1325_Elasciva_c113RC
MP1311_Eimbricata_c194RCrepc2436c788
MP1311_Eimbricata_repc3283
MP1330_Ecoccinea_c7
MP1330_Ecoccinea_c582RCc443c443c1191
MP1309_Epenicilliformis_Consensus
MP1318_Eimbricata_c293RCc52
MP1325_Elasciva_c71RC
MP1319_Eimbricata_c71RCc148
MP1319_Eimbricata_c71RCc139RC
MP1336_Ecoccinea_c703c86
MP1336_Ecoccinea_c59RC
MP1311_Eimbricata_c2150c54RCc1930
MP1334_Eimbricata_repc177RC
MP1312_Elasciva_c169RC
MP1312_Elasciva_c127


[4] Tree 5694
0.01
Rhododendron_Consensus
MP1333_Eplukenetii_Consensus
MP1320_Eabietina_Consensus
MP1319_Eimbricata_Consensus
MP1311_Eimbricata_Consensus
MP1318_Eimbricata_c8RC
MP1318_Eimbricata_c162RCc454RC
MP1339_Eplacentiflora_c517RCc25RC
MP1330_Ecoccinea_repc2354RCc4RC
MP1336_Ecoccinea_Consensus
MP1309_Epenicilliformis_Consensus
MP1312_Elasciva_Consensus
MP1334_Eimbricata_repc186
MP1325_Elasciva_c3
MP1334_Eimbricata_c485RCc1167repc138


[5] Tree ITS
0.01
Rhododendron_c746RC
MP1320_Eabietina_c2207RC
MP1320_Eabietina_c726RC
MP1333_Eplukenetii_c244RC
MP1336_Ecoccinea_c1367c562RC
MP1330_Ecoccinea_c2126c578RC
MP1319_Eimbricata_c910RC
MP1309_Epenicilliformis_c61
MP1312_Elasciva_c227
MP1325_Elasciva_c2657
MP1339_Eplacentiflora_c448c635RCc615
MP1318_Eimbricata_c1333c968RC
MP1311_Eimbricata_c687RC
MP1334_Eimbricata_c22

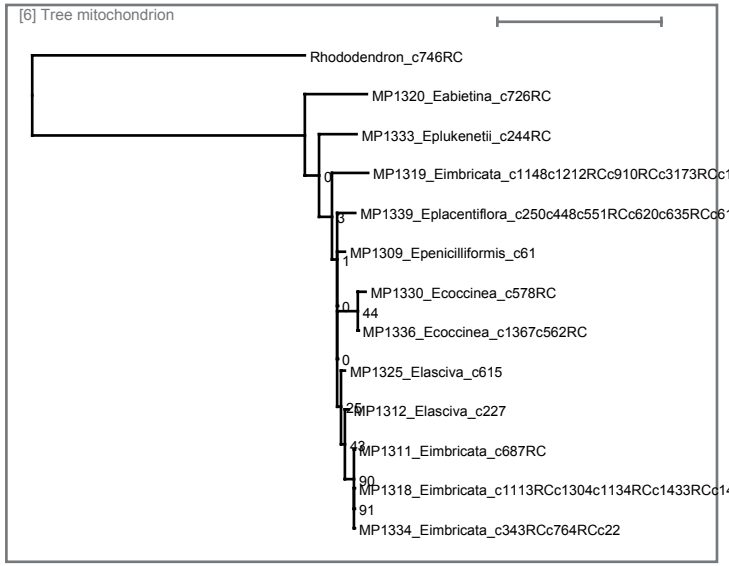

[6] Tree mitochondrion
Rhododendron_c746RC
MP1320_Eabietina_c726RC
MP1333_Eplukenetii_c244RC
MP1319_Eimbricata_c1148c1212RCc910RCc3173RCc1162c
MP1339_Eplacentiflora_c250c448c551RCc620c635RCc615c88
MP1309_Epenicilliformis_c61
MP1330_Ecoccinea_c578RC
MP1336_Ecoccinea_c1367c562RC
MP1325_Elasciva_c615
MP1312_Elasciva_c227
MP1311_Eimbricata_c687RC
MP1318_Eimbricata_c1113RCc1304c1134RCc1433RCc1462R
MP1334_Eimbricata_c343RCc764RCc22

Table 1(on next page)

Voucher details

Table 1 - Samples used for DNA extraction and their collection localities. Vouchers were lodged at herbarium NBG (MP: Pirie).

1 Table 1: Samples used for DNA extraction and their collection localities. Vouchers were lodged
2 at herbarium NBG (MP: Pirie).

Voucher	Sample #	Species	Locality (unless specified, within the Western Cape, South Africa)
MP1320	*78*	*E. abietina* L. ssp. *aurantiaca*	Du Toit's Pass
MP1330	*74*	*E. coccinea* L.	RZE, Greyton
MP1336	*81*	*E. coccinea* L.	Groot Hagelkraal
MP1318	*72*	*E. imbricata* L.	Flouhoogte
MP1319	*73*	*E. imbricata* L.	Stellenbosch
MP1334	*74*	*E. imbricata* L.	Groot Hagelkraal
MP1311	*69*	*E. imbricata* L.	Boskloof
MP1312	*80*	*E. lasciva* Salisb.	Boskloof
MP1325	*83*	*E. lasciva* Salisb.	Albertinia
MP1309	*71*	*E. penicilliformis* Salisb.	Boskloof
MP1339	*75*	*E. placentiflora* Salisb.	Cape Hangklip
MP1333	*82*	*E. plukenetii* L.	Groot Hagelkraal
	68	*R. camtschaticum* Pall.	Oldenburg Botanical Garden, Germany (cultivated)

3

4

Table 2(on next page)

Attributes of selected markers

Table 2 - Range, median and average length of selected markers in *Rhododendron*, with and without taking introns into account, and similarities to homologues in *Vaccinium.*

Table 2: Range, median and average length of selected markers in *Rhododendron*, with and without taking introns into account, and similarities to homologues in *Vaccinium*.

		Length of CR (bp)		Similarity (%)		Predicted length (bp)	
		Range	Mean	Range	Mean	Range	Mean
			Median		Median		Median
			sd		sd		sd
Erica	AllMarkers.py (without intron length) - 79 seq	2316-4815	2834	82-96	90	2412-7425	3541
			2631		90		3342
			535		3,1		998
	AllMarkers.py (with intron length) - 132 seq	1053-4815	2287	81-97	91	2847-7425	3579
			2187		92		3339
			736		3,5		773
	HybSeq (without intron length) - 66 seq	1170-4146	2350	77-95	89	1839-9326	3285
			2184		91		3013
			549		5		1181
	HybSeq (with intron length) - 55 seq	993-4146	2226	77-95	89	2943-9326	3835
			2157		91		3614
			719		5		1032
	MarkerMiner (without intron length) - 207 seq	1293-4146	1726	85-97	93	1338-5849	2411
			1596		94		2307
			419		2		649
	MarkerMiner (with intron length) - 254 seq	1011-4146	1600	85-97	93	1665-5849	2329
			1518		94		2210
			454		2		611
Ericales	AllMarkers.py (without intron length) - 171 seq	1002-4146	1400	82-97	93	1014-8546	2389
			1266		93		2121
			460		2,6		1153
	AllMarkers.py (with intron length) - 408 seq	342-4146	1014	82-97	93	1003-8546	1830
			924		93		1623
			458		2,3		928
Eudicots	AllMarkers.py (without introns length) - 130 seq	1002-4146	1427	85-97	93	1014-7657	2379
			1283		93		2093
			487		2,4		1089
	AllMarkers.py (with introns	369-4146	1112	85-97	93	1002-7647	1895
			1017		94		1689

	length) - 249 seq		494		2,2		960

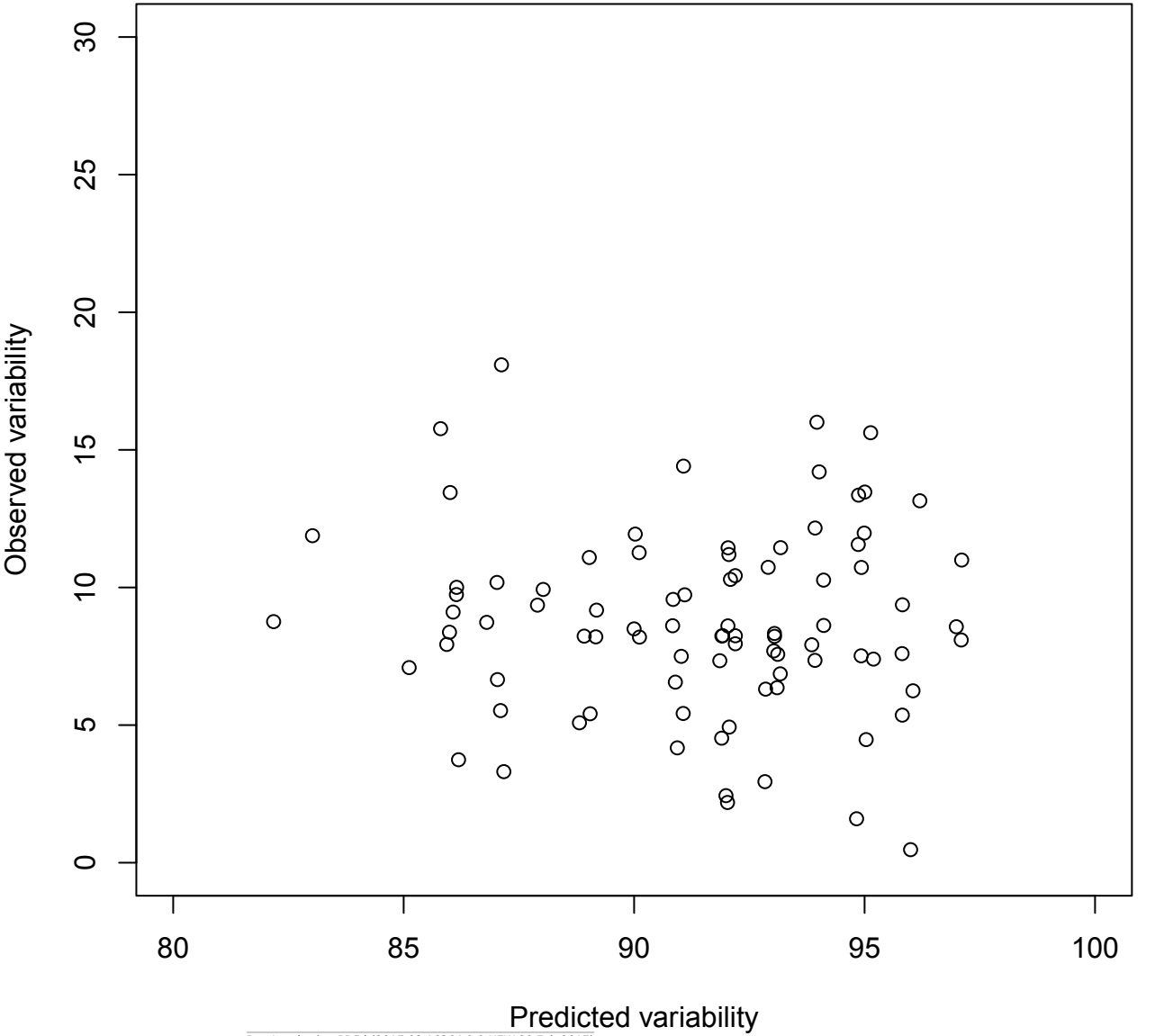
3

Figure 7 (on next page)

Observed against predicted variability of markers

Appendix 1 - Observed variability of selected markers (empirical data) against predicted variability based on *Rhododendron* and *Vaccinium* transcriptome data.

Appendix 1


Observed variability
0
5
10
15
20
25
30
80
85
90
95
100
Predicted variability