

DOC concentrations across a depth gradient on a Caribbean coral reef (#14691)

1

First submission

Please read the **Important notes** below, the **Review guidance** on page 2 and our **Standout reviewing tips** on page 3. When ready [submit online](#). The manuscript starts on page 4.

Important notes

Editor and deadline

Joseph Pawlik / 13 Dec 2016

Files

1 Table file(s)

Please visit the overview page to [download and review](#) the files not included in this review PDF.

Declarations

Involves a field study on animals or plants.



Please read in full before you begin

How to review






When ready [submit your review online](#). The review form is divided into 5 sections. Please consider these when composing your review:

- 1. BASIC REPORTING**
- 2. EXPERIMENTAL DESIGN**
- 3. VALIDITY OF THE FINDINGS**
4. General comments
5. Confidential notes to the editor





 You can also annotate this PDF and upload it as part of your review

To finish, enter your editorial recommendation (accept, revise or reject) and submit.

BASIC REPORTING

-  Clear, unambiguous, professional English language used throughout.
-  Intro & background to show context. Literature well referenced & relevant.
-  Structure conforms to [PeerJ standards](#), discipline norm, or improved for clarity.
-  Figures are relevant, high quality, well labelled & described.
-  Raw data supplied (see [PeerJ policy](#)).

EXPERIMENTAL DESIGN

-  Original primary research within [Scope of the journal](#).
-  Research question well defined, relevant & meaningful. It is stated how the research fills an identified knowledge gap.
-  Rigorous investigation performed to a high technical & ethical standard.
-  Methods described with sufficient detail & information to replicate.

VALIDITY OF THE FINDINGS

-  Impact and novelty not assessed. Negative/inconclusive results accepted. *Meaningful* replication encouraged where rationale & benefit to literature is clearly stated.
-  Data is robust, statistically sound, & controlled.
-  Conclusions are well stated, linked to original research question & limited to supporting results.
-  Speculation is welcome, but should be identified as such.

The above is the editorial criteria summary. To view in full visit <https://peerj.com/about/editorial-criteria/>

7 Standout reviewing tips

3



The best reviewers use these techniques

Tip

Example

Support criticisms with evidence from the text or from other sources

Smith et al (J of Methodology, 2005, V3, pp 123) have shown that the analysis you use in Lines 241-250 is not the most appropriate for this situation. Please explain why you used this method.

Give specific suggestions on how to improve the manuscript

Your introduction needs more detail. I suggest that you improve the description at lines 57- 86 to provide more justification for your study (specifically, you should expand upon the knowledge gap being filled).

Comment on language and grammar issues

The English language should be improved to ensure that your international audience can clearly understand your text. I suggest that you have a native English speaking colleague review your manuscript. Some examples where the language could be improved include lines 23, 77, 121, 128 - the current phrasing makes comprehension difficult.

Organize by importance of the issues, and number your points

1. Your most important issue
2. The next most important item
3. ...
4. The least important points

Give specific suggestions on how to improve the manuscript

Line 56: Note that experimental data on sprawling animals needs to be updated. Line 66: Please consider exchanging "modern" with "cursorial".

Please provide constructive criticism, and avoid personal opinions

I thank you for providing the raw data, however your supplemental files need more descriptive metadata identifiers to be useful to future readers. Although your results are compelling, the data analysis should be improved in the following ways: AA, BB, CC

Comment on strengths (as well as weaknesses) of the manuscript

I commend the authors for their extensive data set, compiled over many years of detailed fieldwork. In addition, the manuscript is clearly written in professional, unambiguous language. If there is a weakness, it is in the statistical analysis (as I have noted above) which should be improved upon before Acceptance.

DOC concentrations across a depth gradient on a Caribbean coral reef

Benjamin Mueller^{Corresp., 1, 2, 3}, Erik H Meesters⁴, Fleur C van Duyl¹

¹ Department of Marine Microbiology and Biogeochemistry, NIOZ Royal Netherlands Institute for Sea Research and Utrecht University, Den Burg, The Netherlands

² CARMABI Foundation, Willemstad, Curaçao

³ Department of Aquatic Environmental Ecology, University of Amsterdam, Amsterdam, The Netherlands

⁴ Wageningen Marine Research, Den Helder, The Netherlands

Corresponding Author: Benjamin Mueller

Email address: muellerb@gmail.com

The dissolved organic carbon (DOC) pool on tropical coral reefs is mainly fueled by photosynthates released from benthic primary producers (BPP), such as reef algae and scleractinian corals. DOC concentrations near BPP have repeatedly been observed to be elevated compared to those in the surrounding water column. As the DOC release of BPP increases with increasing light availability, elevated DOC concentrations near them will, in part, also depend on light availability. Consequently, DOC concentrations are likely to be higher on the shallow, well-lit reef terrace than in deeper sections on the fore reef slope. We measured *in situ* DOC concentrations and light intensity in close proximity to the reef alga *Dictyota* sp. and the scleractinian coral *Orbicella faveolata* along a depth gradient from 5 to 20 m depth and compared these to background concentrations in the water column. DOC concentrations near *Dictyota* sp. were significantly higher at 10 m than at 5 and 20 m depth. Furthermore, at 10 m DOC concentrations near *Dictyota* sp. were elevated by 15 $\mu\text{mol C L}^{-1}$ compared to background concentrations in the water column, but not at 5 and 20 m. DOC concentrations near *O. faveolata* and in the water column did not differ between depths and concentrations near *O. faveolata* were not elevated compared to background concentrations at any of the tested depths. Our results indicate that DOC concentrations near *Dictyota* sp. can differ along a depth gradient from 5 to 20 m. However, the occurrence of elevated DOC concentrations did not follow a natural light gradient across depth. Instead, a combination of light availability (including a restriction by photoinhibition) and water movement are proposed to interactively determine the DOC concentrations in the close vicinity of BPP across the reef slope.

DOC concentrations across a depth gradient on a Caribbean coral reef

Mueller B^{1,2,3}, Meesters EH⁴, van Duyl FC¹

¹NIOZ Royal Netherlands Institute for Sea Research, Department of Marine Microbiology and Biogeochemistry, and Utrecht University, Den Burg, Texel, The Netherlands.

²Carmabi Foundation, Willemstad, Curaçao.

³Present address: University of Amsterdam, Department of Aquatic Environmental Ecology, Institute for Biodiversity and Ecosystem Dynamics, Amsterdam, The Netherlands.

⁴Wageningen Marine Research, Den Helder, The Netherlands.

Corresponding author:

Benjamin Mueller

Email address: muellerb@ymail.com

ABSTRACT

The dissolved organic carbon (DOC) pool on tropical coral reefs is mainly fueled by photosynthates released from benthic primary producers (BPP), such as reef algae and scleractinian corals. DOC concentrations near BPP have repeatedly been observed to be elevated compared to those in the surrounding water column. As the DOC release of BPP increases with increasing light availability, elevated DOC concentrations near them will, in part, also depend on light availability. Consequently, DOC concentrations are likely to be higher on the shallow, well-lit reef terrace than in deeper sections on the fore reef slope. We measured *in situ* DOC concentrations and light intensity in close proximity to the reef alga *Dictyota* sp. and the scleractinian coral *Orbicella faveolata* along a depth gradient from 5 to 20 m depth and compared these to background concentrations in the water column. DOC concentrations near *Dictyota* sp. were significantly higher at 10 m than at 5 and 20 m depth. Furthermore, at 10 m DOC concentrations near *Dictyota* sp. were elevated by 15 $\mu\text{mol C L}^{-1}$ compared to background concentrations in the water column, but not at 5 and 20 m. DOC concentrations near *O. faveolata* and in the water column did not differ between depths and concentrations near *O. faveolata* were not elevated compared to background concentrations at any of the tested depths. Our results indicate that DOC concentrations near *Dictyota* sp. can differ along a depth gradient from 5 to 20 m. However, the occurrence of elevated DOC concentrations did not follow a natural light gradient across depth. Instead, a combination of light availability (including a restriction by photoinhibition) and water movement are proposed to interactively determine the DOC concentrations in the close vicinity of BPP across the reef slope.

INTRODUCTION

Dissolved organic carbon (DOC) is the largest pool of reduced carbon on tropical coral reefs (Atkinson & Falter 2003). Typically DOC concentrations are elevated in the reef overlying water compared to the surrounding ocean, suggesting a net production of DOC on coral reefs (Torr  ton et al. 1997; Van Duyl & Gast 2001). Moreover, the lack of a relationship between particulate organic carbon (POC as proxy for planktonic primary producers) and DOC concentrations (Tanaka et al. 2011), and increased DOC concentrations near the bottom compared to the surface water (Van Duyl & Gast 2001) further indicate that benthic primary producers (BPP) are the main source of DOC on tropical coral reefs. Reef algae and scleractinian corals release a

substantial portion of their photosynthetically fixed carbon as DOC into the surrounding water, yet reef algae generally release more DOC than corals (e.g., Haas et al. 2011; Haas et al. 2013b). This algal-derived DOC can promote the growth of opportunistic heterotrophic microbes in the water column as well as in the contact zone between corals and algae (Haas et al. 2013a; Haas et al. 2013b; Nelson et al. 2013). Increased microbial respiration in the coral-algal interface causing anoxia (Gregg et al. 2013; Haas et al. 2013a) in combination with the release of secondary metabolites, can lead to tissue loss or even coral death (Barott & Rohwer 2012; Morrow et al. 2013). Moreover, while most heterotrophic organisms cannot utilize DOC for their nutrition an increasing number of reef sponges is found to predominantly rely on DOC as carbon source (Yahel et al. 2003; De Goeij et al. 2008; Mueller et al. 2014a;). And similar to microbes, sponges also appear to prefer algal- over coral-derived DOC (Rix et al. 2016). In the so-called sponge loop these sponges utilize the energy stored in DOC and make it available to higher trophic levels via subsequent detritus production (Alexander et al. 2014; De Goeij et al. 2013). Both heterotrophic microbes and DOC-feeding sponges are therefore likely to benefit from elevated DOC concentrations with potential consequences for carbon cycling and overall coral reef functioning (e.g., Rohwer & Youle 2010; Barott & Rohwer 2012; De Goeij et al. 2013; Haas et al. 2016).

Elevated DOC concentrations in close proximity to BPP have been repeatedly observed on tropical coral reefs (Van Duyl & Gast 2001; Hauri et al. 2010; Mueller et al. 2014b). However, most studies were conducted in shallow reef areas between 5 and 10 m and little attention was given to deeper reef sections or how DOC concentrations change across depth. Light availability decreases exponentially with depth and is an important environmental parameter that structures benthic communities across the reef slope (e.g. Bak 1974; Veron 2000; Vermeij & Bak 2002). Light availability positively affects the DOC release rates of BPP (Crossland 1987; Haas et al. 2010b; Naumann et al. 2010; Barrón et al. 2014 and references therein). Moreover, also the occurrence of elevated DOC concentrations near them were found to be positively correlated with the availability of light (Mueller et al. 2014b). We therefore hypothesize that DOC concentrations change with depth and that elevated DOC concentrations near BPP are more likely to occur on the shallow, well-lit reef terrace (5 m) than at the drop off (10 m) or in deeper sections of the fore reef slope (20 m). To test this we measured *in situ* DOC concentrations and light intensity in close proximity to the reef alga *Dictyota* sp. and the scleractinian coral

Orbicella faveolata (former *Montastraea annularis*) along a depth gradient from 5 to 20 m depth and compared these to background concentrations in the water column.

MATERIALS AND METHODS

Fieldwork was performed under the research permit (#2012/48584) issued by the Curaçaoan Ministry of Health, Environment and Nature (GMN) to the CARMABI foundation.

DOC concentrations and light intensity across depth

To quantify DOC concentrations across depth, water samples were taken *in situ* in close proximity (<5 mm) to the abundant reef alga *Dictyota* sp., the scleractinian coral *O. faveolata* and the water column. Both, *Dictyota* sp. and *O. faveolata* are considered holobionts, including epi- and endophytes and associated microbial communities (sensu Barott et al. 2011), jointly affecting the water properties (e.g., DOC concentration) in their close vicinities. Sampling took place on July 24, 2012 at Snake Bay (12° 8' N, 68° 59' W) on the leeward coast of the Island of Curaçao in the Southern Caribbean. The site consists of an approximately 100 m wide sandy reef terrace with patchy coral communities. The reef terrace gradually slopes towards a drop-off that starts around 10 m depth. The reef then slopes down under a steep angle (20-30°; Van Duyl (1985)) and is characterized by a structurally complex reef topography and high coral cover (>30%; De Goeij and Mueller unpubl. data). At midday between 12:00 hrs and 13:00 hrs (when light intensities are the highest) patches of *Dictyota* sp. and colonies of *O. faveolata* were sampled at 5 (reef flat), 10 (drop-off) and 20 m depth (fore reef slope) (each n = 5). In addition, the water column 2 m off the reef bottom was sampled (n = 5) at the same depths and used to indicate background DOC concentrations (i.e., those not directly affected by DOC release of BPP). Sampling started at 20 m depth and 10 and 5 m were sampled consecutively. Per depth approx. 10 min were spent to collect all samples. The sampling procedure described by van Duyl and Gast (2001) and modified by Mueller et al. (2014b) was followed. In short, water samples were collected using 100 ml acid-washed, polypropylene syringes equipped with a flexible silicon tube attached to their tips. The tube was moved slowly above the surfaces of *Dictyota* sp. and *O. faveolata*, respectively, while collecting water. The water column was sampled using a similar syringe. All water samples were collected facing the water current to avoid potential contamination related to the diver's presence. Ambient light intensity (PAR) was recorded simultaneously while sampling (approx. 10 min; sampling intervals 1 min) using a light meter in

a custom-made underwater housing (cosine LI-192SSA underwater quantum sensor connected to LI-1000 data logger; range: PAR 400-700). Water samples were transported (<30 min) to the lab and stored at 4°C until they were processed later that same day.

Processing of DOC samples

Water samples collected were filtered (<20 kPa Hg suction pressure) over a 0.2 µm polycarbonate filter (Whatman, 25 mm). Prior to filtration, filters, glassware and pipette tips were rinsed three times with acid (10 mL 0.4 M HCl) and twice with sample water (10 mL). Afterwards 20 mL of sample water was filtered and the filtrate containing DOC was transferred to pre-combusted (4 h at 450°C) Epa vials (40 mL). Samples were acidified with 6–7 drops of concentrated HCl (38%) to remove inorganic C and stored at 4°C until analysis. DOC concentrations were measured using the high-temperature catalytic oxidation (HTCO) technique in a total organic C analyzer (TOC-VCPN; Shimadzu). The instrument was calibrated with a standard addition curve of Potassium Hydrogen Phthalate (0; 25; 50; 100; 200 µmol C L⁻¹). Consensus Reference Materials (CRM) provided by DA Hansell and W Chen of the University of Miami (Batch 12; 2012; 41-44 µmol C L⁻¹) were used as positive controls for our measurements. Concentrations measured for the batch gave average values (±SD) of 45±3 µmol C L⁻¹. Average analytical variation of the instrument was <3% (5-7 injections per sample).

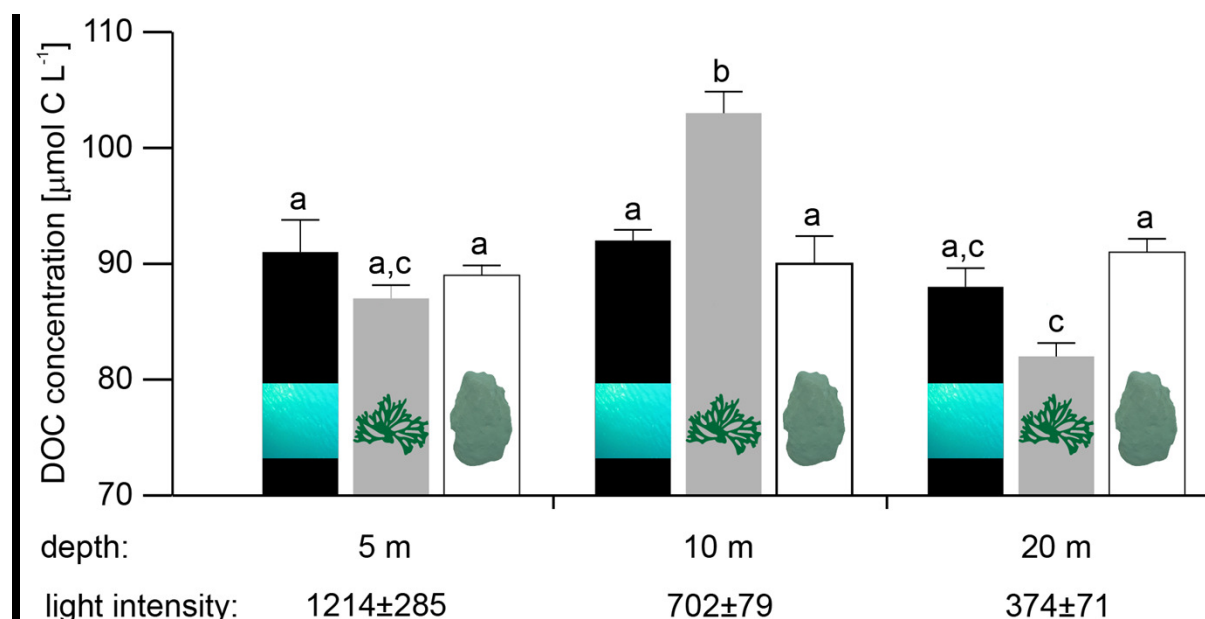
Data analysis

Differences in DOC concentrations at the substrate-water-interface of *Dictyota* sp., *O. faveolata* and the water column from 5, 10 and 20 m were tested using a Kruskal-Wallis test followed by a Mann-Whitney U test in case of significant differences.

RESULTS

In situ DOC concentration in close proximity to *Dictyota* sp. differed significantly across depths (Kruskal-Wallis, p=0.01) (Figure 1 and Supplemental Information for raw data). The distribution of the data from 10 m was different from that at 5 (Mann-Whitney, p=0.02) and 20 m (Mann-Whitney, p=0.01). Estimated mean DOC concentration at 10 m was 107±5 (±SD) µmol L⁻¹ and thus 20 and 25 µmol L⁻¹ higher compared to 5 and 20 m, respectively. No differences in DOC concentrations among depths were observed near *O. faveolata* (Kruskal-Wallis, p=0.93) and in the water column (Kruskal-Wallis, p=0.62). At 10 m depth the distribution of the data of

136 *Dictyota* sp. differed from that of the water column (Mann-Whitney, $p=0.02$), with estimated
 137 mean DOC concentrations near *Dictyota* sp. being elevated by $15 \mu\text{mol L}^{-1}$ compared to
 138 background concentrations. In contrast, the distribution of the data at 5 m (Mann-Whitney,
 139 $p=0.81$) and 20 m depth (Mann-Whitney, $p=0.35$) did not differ between *Dictyota* sp. and in the
 140 water column. Furthermore, estimated mean DOC concentration near *O. faveolata* did not differ
 141 from those in the water column at any of the tested depths. Interestingly, at 20 m estimated mean
 142 DOC concentration near *Dictyota* sp. was significantly lower than near *O. faveolata* (Mann-
 143 Whitney, $p=0.028$). The sampling depths of 5, 10 and 20 m corresponded to a light intensity of
 144 1214 ± 285 , 702 ± 79 and $374 \pm 71 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ (mean \pm SD) during the sampling
 145 (Supplemental Information for raw data).



147 **Figure 1 Mean *in situ* DOC concentrations (n=5, except for water column 10 m and**
 148 ***Dictyota* sp. 5 m with n=4) measured in the water column (2 m off the reef slope; black)**
 149 **and at the substrate-water interfaces of the reef algae *Dictyota* sp. (dark grey) and the**
 150 **scleractinian coral *Orbicella faveolata* (white) at 5, 10 and 20 m depth. Error bars indicate**
 151 **SE. Concentrations with the same letter are not significantly different at $\alpha = 0.05$. Measured *in***
 152 ***situ* light intensity (mean \pm SD) during the sampling is given in $\mu\text{mol photons m}^{-2} \text{s}^{-1}$.**

153

154 DISCUSSION

In this study we investigated DOC concentrations in close proximity to the reef alga *Dictyota* sp., the scleractinian coral *O. faveolata*, and in the water column across a depth gradient from 5 to 20 m. DOC concentrations near *Dictyota* sp. differed between depths, whereas those near *O. faveolata* and in the water column remained similar over the tested depth range. Elevated DOC concentrations compared to the background concentrations in the water column were only observed near *Dictyota* sp. at 10 m, but not at 5 and 20 m depth, or near *O. faveolata* at any of the tested depths.

Elevated DOC concentrations in close proximity to BPP occur when DOC release exceeds removal processes. Consequently, environmental parameters that affect the DOC release of BPP (e.g., light availability (Haas et al. 2010b; Barrón et al. 2012), temperature (Gillooly et al. 2001; Haas et al. 2010b), grazing pressure (Berman & Holm-Hansen 1974), senescence (Khailov & Burlakova 1969), nutrient availability (López-Sandoval et al. 2011; Mueller et al. 2016), hydrodynamic conditions (Wild et al. 2012)), in combination with factors which affect the accumulation of DOC near them (e.g. morphology of the BPP, hydrodynamic conditions (Losee & Wetzel 1993, Escartín & Aubrey 1995), DOC consumption by heterotrophic microbes and sponges (Gast et al. 1999; Yahel et al. 2003; Scheffers et al. 2005; De Goeij et al. 2008)) interactively determine the DOC concentrations in close vicinity to BPP (Figure 2). The lack of elevated DOC concentrations near *Dictyota* sp. at 5 m depth could thus be explained by (1) insufficient DOC release, (2) high DOC removal or (3) a combination of both.

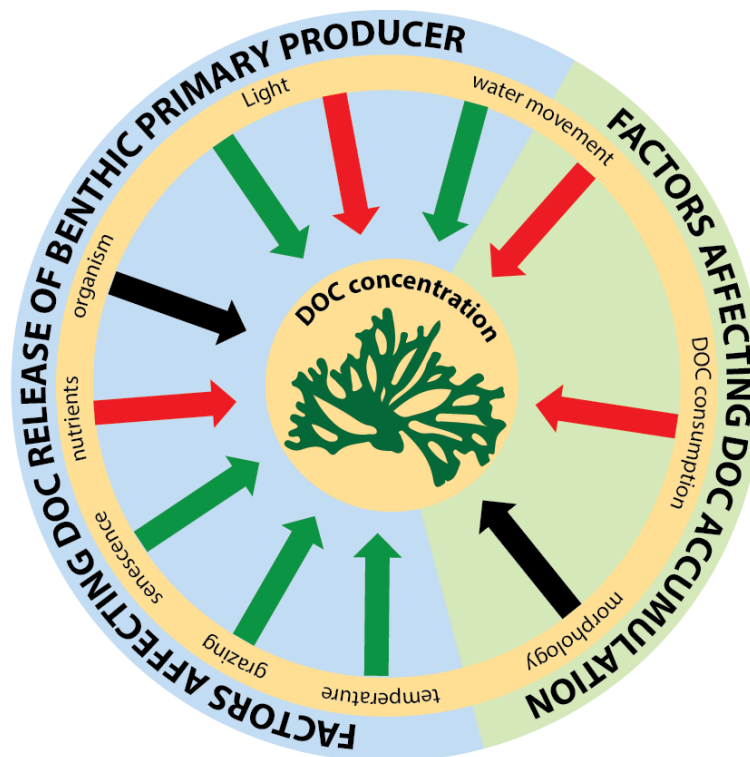


Figure 2 *In situ* DOC concentrations near benthic primary producers are interactively determined by factors that are affecting the DOC release of the benthic primary producers and by those affecting the accumulation of DOC. Green and red arrows indicate positive and negative effects on *in situ* DOC concentrations, respectively. Black arrows indicate the general effect of the organism under consideration and its morphology.

Light availability is generally considered to have a strong positive effect on DOC release of reef algae. However, Haas et al. (2010b) reported that this positive correlation in the reef alga *Caulerpa* sp. only held until a maximum light intensity was reached. At these light intensities DOC release rates steeply decreased to levels comparable to those in the dark. They explained this decrease with the onset of photoinhibition at a species-specific light intensity, which is a common phenomenon in coral reef BPP (Franklin 1994; Hanelt et al. 1994; Brown et al. 1999; Hoegh-Guldberg & Jones 1999; Iglesias-Prieto et al. 2004). Accordingly, photoinhibition likely reduced the DOC release of *Dictyota* sp. at 5 m depth and therefore contributed to the fact that no elevated DOC concentrations in its close proximity were found at this depth.

Similar to light availability, ~~also~~ hydrodynamic conditions can affect *in situ* DOC concentrations near BPP in two ways. Either positively, when water movement increases the metabolism and DOC release rates of BPP by alleviating the limitation of the diffusive boundary layer around them (Carpenter et al. 1991; Lesser et al. 1994; Wild et al. 2012), or negatively, when water movement and water exchange hamper the accumulation of DOC by dilution (Hauri et al. 2010). Water movement generally decreases exponentially as a function of depth (Shashar et al. 1996) and significantly higher water movement rates are reported at 5 compared to 10 or 20 m depth on the reef slope of Curaçao (Vermeij & Bak 2003). Thus, a reduced DOC release rate of *Dictyota* sp. due to photoinhibition in combination with high water movement and water exchange that hamper the accumulation of DOC, could explain the lack of elevated DOC concentrations near *Dictyota* sp. at 5 m depth. It can be further assumed that the negative effect of water movement and water exchange on the accumulation of DOC at 10 m was higher than at 20 m, i.e., a higher DOC release rate was necessary to result in elevated DOC concentrations at 10 m. Yet, despite higher water movement, elevated DOC concentrations near *Dictyota* sp. were only found at 10, but not at 20 m. This suggests that DOC release rates were higher at 10 m than at 20 m, which is in line with the aforementioned positive relation between light availability and DOC release. Interestingly, at 20 m depth DOC concentrations in close proximity to *Dictyota* sp. were depleted compared to concentrations near *O. faveolata* (and lower relative to, but not significantly different from those in the water column). Reduced water movement and thus a prolonged water residence time combined with a low, but steady release of bio-available DOC by *Dictyota* sp., could have stimulated the growth of heterotrophic microbial communities. The bio-available DOC could have further allowed those communities to metabolize otherwise refractory components of the DOC pool and thereby deplete the local DOC stock, as described for the water columns overlying algal dominated reefs (Dinsdale et al. 2008; Haas et al. 2016).

No elevated DOC concentrations were observed near the scleractinian coral *O. faveolata* at any of the sampling depths. In general, the DOC release of scleractinian corals is more variable than that of reef algae and an increasing number of studies suggest that scleractinian corals only contribute marginally to the local DOC on tropical coral reefs (e.g., Haas et al. 2010a; Naumann et al. 2010; Haas et al. 2011). Furthermore, the massive morphology of *O. faveolata* is less likely to restrict water exchange than the bushy thalli of *Dictyota* sp. and is thereby less favorable for the accumulation of DOC in its vicinity (Stocking et al. 2016). Given the positive effect of light

availability on the DOC release by BPP, we expected to find significantly higher DOC concentrations on the shallow and well-lit reef terrace compared to deeper reef sections, following the natural light gradient across depth. Surprisingly, significant differences in the mean DOC concentrations between the sampled depths were only observed in *Dictyota* sp., but not in *O. faveolata* or the water column. The absence of significant differences in DOC concentrations across the water column was also observed in other studies (Torréton et al. 1997; Nelson et al. 2011). To date only Slattery & Lesser (2015) reported a significant decline in DOC concentration with depth from coral reefs on the Bahamas, albeit this decrease occurred at mesophotic depths below 30 m. This may indicate that at least above mesophotic depths, DOC released by BPP is either quickly taken up by DOC feeding organisms (i.e. heterotrophic bacteria and reef sponges) and/or mixed and diluted throughout the reef overlying water column.

CONCLUSION

While light availability has a strong positive effect on the DOC release of BPP, the occurrence of elevated DOC concentrations near them did not follow a natural light gradient across the reef slope in our study system. Instead, a combination of light availability, which affects the release of DOC (including the restriction by photoinhibition) and water movement, which affects the accumulation of DOC, are proposed to interactively determine the DOC concentrations in the close vicinity of BPP.

ACKNOWLEDGEMENTS

We thank the staff of Carmabi for their hospitality and logistic support during the field work. We further thank V. Chamberland, T. Holtrop, Y. Mulders, E. van der Ent, R. van der Zande and K. Vane for their help during the field work. We are grateful to S. Gonzalez for his contribution to the DOC analysis. The manuscript benefitted greatly from comments by M. Vermeij on earlier versions of the manuscript.

REFERENCES

Alexander BE, Liebrand K, Osinga R, van der Geest HG, Admiraal W, Cleutjens JP, Schutte B, Verheyen F, Ribes M, van Loon E, and de Goeij JM. 2014. Cell turnover and detritus production in marine sponges from tropical and temperate benthic ecosystems. *PLoS One* 9:e109486. 10.1371/journal.pone.0109486

- Atkinson MJ, and Falter JL. 2003. In: Black KD, and Shimmield GB, eds. *Biogeochemistry of marine systems*. Oxford: Blackwell Publishing.
- Bak RPM. 1974. Available light and other factors influencing growth of stony corals through the year in Curacao: *Proc. 2nd Int. Coral Reef Symp., 1974* 2: p. 229-233.
- Barott KL, Rodriguez-Brito B, Janouskovec J, Marhaver KL, Smith JE, Keeling P, and Rohwer FL. 2011. Microbial diversity associated with four functional groups of benthic reef algae and the reef-building coral *Montastraea annularis*. *Environmental Microbiology* 13:1192-1204. 10.1111/j.1462-2920.2010.02419.x
- Barott KL, and Rohwer FL. 2012. Unseen players shape benthic competition on coral reefs. *Trends in Microbiology* 20:621-628. 10.1016/j.tim.2012.08.004
- Barrón C, Apostolaki E, and Duarte C. 2012. Dissolved organic carbon release by marine macrophytes. *Biogeosciences Discuss* 9:1529-1555. doi:10.5194/bgd-9-1529-2012
- Barrón C, Apostolaki ET, and Duarte CM. 2014. Dissolved organic carbon fluxes by seagrass meadows and macroalgal beds. *Frontiers in Marine Science* 1. 10.3389/fmars.2014.00042
- Berman T, and Holm-Hansen O. 1974. Release of photoassimilated carbon as dissolved organic matter by marine phytoplankton. *Marine Biology* 28:305-310. 10.1007/BF00388498
- Brown BE, Ambarsari I, Warner ME, Fitt WK, Dunne RP, Gibb SW, and Cummings DG. 1999. Diurnal changes in photochemical efficiency and xanthophyll concentrations in shallow water reef corals : evidence for photoinhibition and photoprotection. *Coral Reefs* 18:99-105. 10.1007/s003380050163
- Carpenter RC, Hackney JM, and Adey WH. 1991. Measurements of primary productivity and nitrogenase activity of coral reef algae in a chamber incorporating oscillatory flow. *Limnology and Oceanography* 36:40-49. 10.4319/lo.1991.36.1.0040
- Crossland CJ. 1987. In situ release of mucus and DOC-lipid from the corals *Acropora variabilis* and *Stylophora pistillata* in different light regimes. *Coral Reefs* 6:35-42. 10.1007/BF00302210
- De Goeij JM, van den Berg H, van Oostveen MM, Epping EHG, and van Duyl FC. 2008. Major bulk dissolved organic carbon (DOC) removal by encrusting coral reef cavity sponges. *Marine Ecology Progress Series* 357:139-151. 10.3354/meps07403

- De Goeij JM, van Oevelen D, Vermeij MJA, Osinga R, Middelburg JJ, de Goeij AFPM, and
Admiraal W. 2013. Surviving in a marine desert: The sponge loop retains resources
within coral reefs. *Science* 342:108-110. 10.1126/science.1241981
- Dinsdale EA, Pantos O, Smriga S, Edwards RA, Angly F, Wegley L, Hatay M, Hall D, Brown E,
Haynes M, Krause L, Sala E, Sandin SA, Thurber RV, Willis BL, Azam F, Knowlton N,
and Rohwer F. 2008. Microbial ecology of four coral atolls in the northern Line Islands.
PLoS One 3:e1584. 10.1371/journal.pone.0001584
- Escartín J, and Aubrey DG. 1995. Flow Structure and Dispersion within Algal Mats. *Estuarine,
Coastal and Shelf Science* 40:451-472. <http://dx.doi.org/10.1006/ecss.1995.0031>
- Franklin LA. 1994. The effects of temperature acclimation on the photoinhibitory responses of
Ulva rotundata Blid. *Planta* 192:324-331.
- Gast GJ, Jonkers PJ, van Duyl FC, and Bak RPM. 1999. Bacteria, flagellates and nutrients in
island fringing coral reef waters: Influence of the ocean, the reef and eutrophication.
Bulletin of Marine Science 65:523-538.
- Gillooly JF, Brown JH, West GB, Savage VM, and Charnov EL. 2001. Effects of size and
temperature on metabolic rate. *Science* 293:2248-2251. 10.1126/science.1061967
- Gregg AK, Hatay M, Haas AF, Robinett NL, Barott K, Vermeij MJA, Marhaver KL, Meirelles
P, Thompson F, and Rohwer F. 2013. Biological oxygen demand optode analysis of coral
reef-associated microbial communities exposed to algal exudates. *PeerJ* 1:e107.
10.7717/peerj.107
- Haas A, Jantzen C, Naumann M, Iglesias-Prieto R, and Wild C. 2010a. Organic matter release by
the dominant primary producers in a Caribbean reef lagoon: implication for in situ O₂
availability. *Marine Ecology Progress Series* 409:27-39. 10.3354/meps08631
- Haas AF, Naumann MS, Struck U, Mayr C, el-Zibdah M, and Wild C. 2010b. Organic matter
release by coral reef associated benthic algae in the Northern Red Sea. *Journal of
Experimental Marine Biology and Ecology* 389:53-60.
<http://dx.doi.org/10.1016/j.jembe.2010.03.018>
- Haas AF, Nelson CE, Wegley Kelly L, Carlson CA, Rohwer F, Leichter JJ, Wyatt A, and Smith
JE. 2011. Effects of coral reef benthic primary producers on dissolved organic carbon and
microbial activity. *PLoS One* 6:e27973. 10.1371/journal.pone.0027973

311 Haas AF, Gregg AK, Smith JE, Abieri ML, Hatay M, and Rohwer F. 2013a. Visualization of
312 oxygen distribution patterns caused by coral and algae. *PeerJ* 1:e106. 10.7717/peerj.106

313 Haas AF, Nelson CE, Rohwer F, Wegley-Kelly L, Quistad SD, Carlson CA, Leichter JJ, Hatay
314 M, and Smith JE. 2013b. Influence of coral and algal exudates on microbially mediated
315 reef metabolism. *PeerJ* 1:e108. 10.7717/peerj.108

316 Haas AF, Fairouz MF, Kelly LW, Nelson CE, Dinsdale EA, Edwards RA, Giles S, Hatay M,
317 Hisakawa N, and Knowles B. 2016. Global microbialization of coral reefs. *Nature*
318 *Microbiology*:16042.

319 Hanelt D, Li J, and Nultsch W. 1994. Tidal dependence of photoinhibition of photosynthesis in
320 marine macrophytes of the South China Sea. *Botanica Acta* 107:66-72.

321 Hauri C, Fabricius KE, Schaffelke B, and Humphrey C. 2010. Chemical and physical
322 environmental conditions underneath mat- and canopy-forming macroalgae, and their
323 effects on understory corals. *PLoS One* 5:e12685. 10.1371/journal.pone.0012685

324 Hoegh-Guldberg O, and Jones RJ. 1999. Photoinhibition and photoprotection in symbiotic
325 dinoflagellates from reef-building corals. *Marine Ecology Progress Series* 183:73-86.

326 Iglesias-Prieto R, Beltran V, LaJeunesse T, Reyes-Bonilla H, and Thome P. 2004. Different algal
327 symbionts explain the vertical distribution of dominant reef corals in the eastern Pacific.
328 *Proceedings of the Royal Society of London, Series B: Biological Sciences* 271:1757-
329 1763.

330 Khailov KM, and Burlakova ZP. 1969. Release of dissolved organic matter by marine seaweeds
331 and distribution of their total organic production to inshore communities. *Limnology and*
332 *Oceanography* 14:521-527. 10.4319/lo.1969.14.4.0521

333 Lesser MP, Weis VM, Patterson MR, and Jokiel PL. 1994. Effects of morphology and water
334 motion on carbon delivery and productivity in the reef coral, *Pocillopora damicornis*
335 (Linnaeus): Diffusion barriers, inorganic carbon limitation, and biochemical plasticity.
336 *Journal of Experimental Marine Biology and Ecology* 178:153-179.
337 [http://dx.doi.org/10.1016/0022-0981\(94\)90034-5](http://dx.doi.org/10.1016/0022-0981(94)90034-5)

338 López-Sandoval D, Fernández A, and Marañón E. 2011. Dissolved and particulate primary
339 production along a longitudinal gradient in the Mediterranean Sea. *Biogeosciences*
340 *Discuss* 8:815-825.

- 341 Losee RF, and Wetzel RC. 1993. Littoral flow rates within and around submersed macrophyte
342 communities. *Freshwater Biology* 29:7-17. 10.1111/j.1365-2427.1993.tb00739.x
- 343 Morrow KM, Liles MR, Paul VJ, Moss A, and Chadwick NE. 2013. Bacterial shifts associated
344 with coral–macroalgal competition in the Caribbean Sea. *Marine Ecology Progress
345 Series* 488:103-117. 10.3354/meps10394
- 346 Mueller B, de Goeij JM, Vermeij MJ, Mulders Y, van der Ent E, Ribes M, and van Duyl FC.
347 2014a. Natural diet of coral-excavating sponges consists mainly of dissolved organic
348 carbon (DOC). *PLoS One* 9:e90152. 10.1371/journal.pone.0090152
- 349 Mueller B, van der Zande RM, van Leent PJM, Meesters EH, Vermeij MJA, and van Duyl FC.
350 2014b. Effect of light availability on dissolved organic carbon release by Caribbean reef
351 algae and corals. *Bulletin of Marine Science* 90:875-893. 10.5343/bms.2013.1062
- 352 Mueller B, den Haan J, Visser PM, Vermeij MJA, and van Duyl FC. 2016. Effect of light and
353 nutrient availability on the release of dissolved organic carbon (DOC) by Caribbean turf
354 algae. *Sci Rep* 6:23248. 10.1038/srep23248
- 355 Naumann MS, Haas A, Struck U, Mayr C, el-Zibdah M, and Wild C. 2010. Organic matter
356 release by dominant hermatypic corals of the Northern Red Sea. *Coral Reefs* 29:649-659.
357 10.1007/s00338-010-0612-7
- 358 Nelson CE, Alldredge AL, McCliment EA, Amaral-Zettler LA, and Carlson CA. 2011. Depleted
359 dissolved organic carbon and distinct bacterial communities in the water column of a
360 rapid-flushing coral reef ecosystem. *Isme j* 5:1374-1387. 10.1038/ismej.2011.12
- 361 Nelson CE, Goldberg SJ, Wegley Kelly L, Haas AF, Smith JE, Rohwer F, and Carlson CA.
362 2013. Coral and macroalgal exudates vary in neutral sugar composition and differentially
363 enrich reef bacterioplankton lineages. *Isme j* 7:962-979. 10.1038/ismej.2012.161
- 364 Rix L, de Goeij JM, van Oevelen D, Struck U, Al-Horani FA, Wild C, and Naumann MS. 2016.
365 Differential recycling of coral and algal dissolved organic matter via the sponge loop.
366 *Functional Ecology*:n/a-n/a. 10.1111/1365-2435.12758
- 367 Rohwer F, and Youle M. 2010. *Coral reefs in the microbial seas*: Plaid Press.
- 368 Scheffers SR, Bak RPM, and Duyl FCv. 2005. Why is bacterioplankton growth in coral reef
369 framework cavities enhanced? *Marine Ecology Progress Series* 299:89-99.
370 10.3354/meps299089

- Shashar N, Kinane S, Jokiel PL, and Patterson MR. 1996. Hydromechanical boundary layers over a coral reef. *Journal of Experimental Marine Biology and Ecology* 199:17-28. [http://dx.doi.org/10.1016/0022-0981\(95\)00156-5](http://dx.doi.org/10.1016/0022-0981(95)00156-5)
- Slattery M, and Lesser MP. 2015. Trophic ecology of sponges from shallow to mesophotic depths (3 to 150 m): Comment on Pawlik et al.(2015). *Marine Ecology Progress Series* 527:275-279.
- Stocking JB, Rippe JP, and Reidenbach MA. 2016. Structure and dynamics of turbulent boundary layer flow over healthy and algae-covered corals. *Coral Reefs*:1-13. [10.1007/s00338-016-1446-8](https://doi.org/10.1007/s00338-016-1446-8)
- Tanaka Y, Miyajima T, Watanabe A, Nadaoka K, Yamamoto T, and Ogawa H. 2011. Distribution of dissolved organic carbon and nitrogen in a coral reef. *Coral Reefs* 30:533-541. [10.1007/s00338-011-0735-5](https://doi.org/10.1007/s00338-011-0735-5)
- Torréton J, Pagès J, Dufour P, and Cauwet G. 1997. Bacterioplankton carbon growth yield and DOC turnover in some coral reef lagoons. *Proc 8th Int Coral Reef Symp.* p 947-952.
- Van Duyl FC. 1985. *Atlas of the living reefs of Curaçao and Bonaire (Netherlands Antilles)*. Utrecht.
- Van Duyl FC, and Gast GJ. 2001. Linkage of small-scale spatial variations in DOC, inorganic nutrients and bacterioplankton growth with different coral reef water types. *Aquatic Microbial Ecology* 24:17-26. [10.3354/ame024017](https://doi.org/10.3354/ame024017)
- Vermeij MJA, and Bak RPM. 2002. How are coral populations structured by light? Marine light regimes and the distribution of *Madracis*. *Marine Ecology Progress Series* 233:105-116. [10.3354/meps233105](https://doi.org/10.3354/meps233105)
- Vermeij MJA, and Bak RPM. 2003. Species-Specific Population Structure of Closely Related Coral Morphospecies Along a Depth Gradient (5 - 60 M) Over a Caribbean Reef Slope. *Bulletin of Marine Science* 73:725-744.
- Veron JEN. 2000. Corals of the World, vol. 1–3. *Australian Institute of Marine Science, Townsville*.
- Wild C, Laforsch C, Mayr C, Fuß R, and Niggel W. 2012. Effect of water currents on organic matter release by two scleractinian corals. *Aquatic Ecology* 46:335-341. [10.1007/s10452-012-9404-1](https://doi.org/10.1007/s10452-012-9404-1)

401 Yahel G, Sharp JH, Marie D, Häse C, and Genin A. 2003. In situ feeding and element removal in
 402 the symbiont-bearing sponge *Theonella swinhoei*: Bulk DOC is the major source for
 403 carbon. *Limnology and Oceanography* 48:141-149. 10.4319/lo.2003.48.1.0141