

Large-scale gene flow in the barnacle *Jehlius cirratus* and contrasts with other broadly-distributed taxa along the Chilean coast (#13312)

1

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I commend the authors for their extensive data set, compiled over many years of detailed fieldwork. In addition, the manuscript is clearly written in professional, unambiguous language. If there is a weakness, it is in the statistical analysis (as I have noted above) which should be improved upon before Acceptance.

Large-scale gene flow in the barnacle *Jehlius cirratus* and contrasts with other broadly-distributed taxa along the Chilean coast

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We evaluate the population genetic structure of the intertidal barnacle *Jehlius cirratus* across a broad portion of its geographic distribution using data from the mitochondrial cytochrome oxidase I (COI) gene region. Despite sampling diversity from over 3000km of the linear range of this species, there is only slight regional structure indicated, with overall Φ_{CT} of 0.036 ($p < 0.001$) yet no support for isolation by distance. While these results suggest greater structure than previous studies of *J. cirratus* had indicated, the pattern of diversity is still far more subtle than in other similarly-distributed species with similar larval and life history traits. We compare these data and results with recent findings in four other intertidal species that have planktotrophic larvae. There are no clear patterns among these taxa that can be associated with intertidal depth or other known life history traits.

1 RH: Phylogeography in *Jehlius cirratus*

2 **Large-scale gene flow in the barnacle *Jehlius cirratus* and contrasts with**
3 **other broadly-distributed taxa along the Chilean coast**

4

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13

14 Abstract

15 We evaluate the population genetic structure of the intertidal barnacle *Jehlius cirratus*
16 across a broad portion of its geographic distribution using data from the mitochondrial
17 cytochrome oxidase I (COI) gene region. Despite sampling diversity from over 3000km of
18 the linear range of this species, there is only slight regional structure indicated, with overall
19 Φ_{CT} of 0.036 ($p < 0.001$) yet no support for isolation by distance. While these results suggest
20 greater structure than previous studies of *J. cirratus* had indicated, the pattern of diversity
21 is still far more subtle than in other similarly-distributed species with similar larval and life
22 history traits. We compare these data and results with recent findings in four other
23 intertidal species that have planktotrophic larvae. There are no clear patterns among these
24 taxa that can be associated with intertidal depth or other known life history traits.

25 Introduction

26

27 A persistent question in marine biogeography and population biology involves the
28 interaction of species life history, geographic range, and trait or genealogical diversity
29 within that range. In some cases, genealogical diversity or “structure” (Wares 2016) *within*
30 a species is informative of mechanisms that act to limit other species’ distributional ranges
31 (Dawson 2001; Wares 2002; Wares *et al.* 2001). Of course, these studies often find that
32 organisms with limited larval or juvenile dispersal have greater amounts of structure and
33 less extensive ranges, but there are often exceptions (Marko 2004). It is the variation
34 among species, and the exceptions to the “rules”, that offer continued opportunity to
35 understand marine diversity.

36

37 Early approaches to comparative phylogeography (Dawson 2001; Wares 2002; Wares &
38 Cunningham 2001) focused primarily on regions of co-diversification of intraspecific
39 lineages, e.g. the regions across which species were likely to exhibit structure.
40 Subsequently, Marko (2004) noted that even when species had apparently identical life
41 history and dispersal mechanisms, the distribution of a species across habitats (e.g.
42 intertidal height) could influence their persistence in distinct glacial refugia. However,
43 certainly to understand these associations more taxa should be compared, and Kelly and
44 Palumbi (2010) made explicit comparisons of diversity and population divergence for 50
45 species along the Pacific coast of North America to suggest that species high in the
46 intertidal were perhaps more likely to exhibit spatial genetic structure than those at lower
47 depths. However, within taxa that are more closely related, e.g. the barnacles, this rule does

48 not necessarily hold. The high intertidal *Chthamalus dalli* exhibits no apparent population
49 structure (Wares & Castañeda 2005) relative to the mid-intertidal *Balanus glandula* (Sotka
50 et al. 2004), while other barnacle species in this region also show effectively no structure
51 (Dawson *et al.* 2010).

52

53 The particular spatial structure of the species represented in Kelly and Palumbi (2010)
54 varies; however, there is often concordance of population structure among species (Pelc *et*
55 *al.* 2009; Small & Wares 2010) on this coast. Other regions that have been similarly
56 explored – for example, the NW Atlantic coast – have fewer instances of strong population
57 structure aside from regions that are also biogeographic transitions (Altman *et al.* 2013;
58 Díaz-Ferguson *et al.* 2009). Another such example of this concordance of genetic diversity
59 with biogeography was recently published by Haye et al. (2014), looking at species with
60 short-dispersing larval forms around the well-characterized biogeographic transition near
61 30°S latitude along the coast of Chile. Again, the structure of diversity within species was
62 informative to the mechanisms – including shifts in upwelling intensity and nutrient
63 availability (Navarrete *et al.* 2005) – that may limit the distribution of other taxa. As
64 patterns of coastal upwelling are associated with phylogeographic structure in other
65 regions and species (Rocha-Olivares & Vetter 1999; Zakas *et al.* 2009), it merits exploration
66 for how species respond to distinct oceanographic regimes along the Chilean coast.

67

68 Evaluating broad-scale diversity structure on the Chilean coast is of key interest as there
69 are so many oceanographic and biogeographic comparisons to be made between this well-
70 studied coastline and the well-studied Pacific coast of North America (Navarrete *et al.*

71 2008). However, until recently there were few data available for species that spanned most
72 of the length of the Chilean coastline. This scale is of interest because it spans *two* major
73 biogeographic transitions – the region around 30°S noted above, as well as a notable
74 biogeographic transition near 42°S (Thiel *et al.* 2007). While the divergence of taxa near
75 30°S is typically associated with shifts in upwelling and concomitant environmental
76 transitions (Ewers-Saucedo *et al.* 2016; Haye *et al.* 2014), the biogeographic transition at
77 42°S is more likely driven by divergent current flow (Ewers-Saucedo *et al.* 2016).
78 Temperature and salinity both exhibit significant transitions along this coastal region
79 (Acha *et al.* 2004), and thus the dominant biogeographic boundary along the Chilean coast
80 is at about 42°S (Thiel *et al.* 2007).

81

82 Some of the first such work at this spatial scale was done in the direct-developing
83 gastropod *Acanthina monodon* (Sanchez *et al.* 2011) and another gastropod *Concholepas*
84 *concholepas* (Cardenas *et al.* 2009). In *Acanthina*, which has low dispersal potential among
85 locations, strong concordance of intraspecific diversity with the 30°S biogeographic
86 boundary was found, but association with the 42° boundary was less clear. Nevertheless,
87 statistically significant genetic structure and shifts in phenotypic diversity are associated
88 with this region. The gastropod *Concholepas concholepas*, on the other hand, has high
89 potential for pelagic larval dispersal, is similarly distributed along the coast of Chile, but
90 exhibits no significant genetic structure at all (Cardenas *et al.* 2009). These contrasts are
91 wholly in line with predictions based on larval life history.

92

93 Recently, large data sets have become available for other commonly encountered taxa in
94 the Chilean intertidal. Microsatellite data were analyzed in the mussel *Perumytilus*
95 *purpuratus* (Guiñez *et al.* 2016), which both spawns gametes and has a long-lived
96 planktotrophic larva, and this ecosystem engineer exhibited significant structure with two
97 main lineages (separated at approximately 40°S) and isolation by distance within each
98 lineage. Similarly, Ewers-Saucedo *et al.* (2016) explored genetic variation in the high
99 intertidal barnacle *Notochthamalus scabrosus*, with nauplius larvae that have high pelagic
100 larval dispersal potential, and found two primary lineages that mirror the dominant
101 biogeographical pattern of Chile: in the northern Peruvian region only one lineage is found,
102 while both are found in the Intermediate Area that represents the overlap of the Peruvian
103 and Magellanic regions, and only the southern lineage is found south of 42°S. Another
104 barnacle, the edible *picoroco* (*Austromegabalanus psittacus*) exhibits only slight structure
105 along most of the Chilean coast (Pappalardo *et al.* 2016), but nevertheless the observed
106 structure is statistically significant and seems to be associated with the northern (30°S)
107 biogeographic transition.

108

109 To these data we add one more layer: Zakas *et al.* (2009) had explored mitochondrial
110 sequence population structure in the high intertidal barnacle *Jehlius cirratus*, a species that
111 is biologically and ecologically very similar to *Notochthamalus* but found slightly higher in
112 the intertidal (Lamb *et al.* 2014; Shinen & Navarrete 2010, 2014). Zakas *et al.* (2009) found
113 that unlike *Notochthamalus*, there was very little apparent genetic structure in *J. cirratus*.
114 However, that analysis comprised only a small section of the Chilean coast, from ~28-34°S.
115 Here, we expand the sampling of *J. cirratus* to include diversity from ~3500km of coastline,

116 including most of the known distribution (Häussermann & Försterra 2009). As chthamalid
117 barnacles have a propensity to harbor cryptic genetic diversity (Dando & Southward 1981;
118 Meyers *et al.* 2013; Tsang *et al.* 2008; Wares *et al.* 2009; Zardus & Hadfield 2005), we
119 specifically look for any phylogeographic structure that may add to our understanding of
120 coastal biodiversity in Chile. We then more directly compare the whole-coast data
121 described above for the ecological implications of the population structure identified
122 within and among taxa.

123

124

125 **Methods**

126 Specimens of *J. cirratus* were collected from the intertidal in 2004-2013. Field permits were
127 not required from the Subsecretaría de Pesca y Acuicultura for the specimens included in
128 this paper, as they were not "shellfish resources". Sequences of cytochrome oxidase I
129 (n=153) from Zakas et al (2009) were used in this study (Genbank GU126073 –
130 GU126226); additional sequences (n=187) were generated from subsequent samples
131 collected in 2011-2013 using PCR methods as in Zakas et al. (2009). Samples were mostly
132 collected in central Chile (Table 1), but this additional effort also added substantially to
133 information from northern Chile and northern Patagonia.

134 After quality control and alignment of sequence data using CodonCode Aligner v6.0.2
135 (CodonCode Corporation), data were formatted for analysis using Arlequin v3.5.2.2.
136 (Excoffier *et al.* 2005) to identify population structure. Pairwise Φ_{ST} was calculated for all
137 sites and compared to a matrix of pairwise geographic distance for signal of isolation by
138 distance (Wright 1943); this was done both with haplotypic data as well as nucleotide data
139 under a K2P distance model. Additionally, an exact test of differentiation was calculated for
140 all pairs of populations. Analysis of molecular variance (AMOVA) was performed to identify
141 maximal structure along the coast as in Dupanloup *et al.* (2002) and Zakas et al (2009),
142 using an iterative approach for K contiguous spatial groups, increasing K if there were
143 significant patterns of Φ_{SC} within the determined regional groups. Following the results of
144 AMOVA, a haplotype network was generated using PopArt (<http://popart.otago.ac.nz>).
145 Haplotypes were coded by sample location and by regions separated by the iterative
146 AMOVA results that maximize Φ_{CT} to visually identify components of diversity associated

147 with each regional group. Population diversity was also assessed at each sampled location;
148 nucleotide diversity (π) and haplotype diversity (H) are estimated at each location using
149 Arlequin.

150 **Results**

151 New sequences are archived in Genbank under accession numbers KX014910 - KX015034.
152 Site-specific diversity is presented in Table 1; pairwise values of Φ_{ST} are presented in Table
153 2. Only a single sequence was recovered from the northernmost collection site of Arica, so
154 this sequence was included in the Antofagasta sample (results identical when excluded) for
155 statistical purposes. Values of Φ_{ST} are very low and in general not statistically significant
156 (Table 2); the only exceptional locations are Guanaqueros (30°S) and Pichilemu (34°S),
157 each of which tend to exhibit higher differentiation from a broader set of other locations.
158 No population pairs are significantly different under an exact test. Testing these results for
159 a pattern of genetic isolation by distance was not significant (p 0.245).

160 Although only slight structure is exhibited along the Chilean coast in *J. cirratus* (Φ_{ST} of -
161 0.019, $p \sim 1$), there is statistical regional structure detectable with the increased power of
162 sampling at that scale. Our implementation of spatial AMOVA (Zakas et al. 2009) recovered
163 two contrasts for $K=2$ regions in which $\Phi_{CT} > 0.035$ and $p < 0.01$, though similar results are
164 found if the separation among regions is near to either of these locations (Table 3). These
165 local maxima in Φ_{CT} separate Guanaqueros (30°S) and sites to the north from all locations
166 to the south; and Pichilemu (44°S) and all sites to the south from all locations to the north.
167 No significant Φ_{SC} is exhibited in these comparisons. If $K=3$ groups are chosen using these
168 same delineations, Φ_{CT} is comparable (0.03661, $p < 0.001$).

169 From these results, a haplotype network (minimum spanning tree) is presented in Figure
170 1; “northern” diversity (from Guanaqueros northward), “southern” diversity (including
171 Pichilemu and southward sites), and “central” diversity (locations in between), for
172 visualization.

173 Discussion

174 As noted in Zakas et al. (2009) there is only slight population structure in *J. cirratus*.
175 Previous efforts had also noted that using alternate statistics such as Hudson’s (Hudson
176 2000) S_{nn} also recovered no signal of structure or pattern of isolation by distance (Wares
177 2014). Here, we **do** identify statistically significant structure that is roughly associated with
178 the 30°S biogeographic transition between the Peruvian and “Intermediate” zones, and
179 there may also be structure further south – but not associated with the boundary at 42°S.
180 Overall, the statistical significance indicated – given that pairwise statistical support was
181 not consistent between permutational tests of Φ_{ST} and pairwise exact tests of population
182 differentiation – suggests little actual spatial variation but sufficient sampling to identify
183 the differential representation of regional samples in the 2 dominant haplotypes found
184 (Figure 1).

185

186 Excluding the direct developer *A. monodon* from further consideration, the studies
187 reviewed earlier and current study include 5 intertidal species with high larval dispersal
188 potential that are distributed and **analyzed** along the length of the Chilean coast.
189 Unfortunately, there is no clear pattern associated with intertidal depth; the species with
190 no or slight population genetic structure (*J. cirratus*, this study; *A. psittacus*, Pappalardo et

191 *al.* 2016; *C. concholepas*, Cardénas *et al.* 2009) are in the highest reaches of the intertidal (*J.*
192 *cirratus*) and the low intertidal (*A. psittacus* and *C. concholepas*). The two species that
193 exhibit significant structure, each with two primary lineages and evidence for isolation by
194 distance within each lineage, are in the high-to-middle intertidal (*N. scabrosus*, Ewers-
195 Saucedo *et al.* 2016; *P. purpuratus*, Guíñez *et al.* 2016).

196

197 Clearly a sample of only 5 taxa is insufficient for statistical consideration. However, what
198 we can indicate is that all 3 barnacles (*A. psittacus*, *J. cirratus*, and *N. scabrosus*) have at least
199 some signal associated with the 30-32° oceanographic transition in upwelling (Lagos *et al.*
200 2005; Navarrete *et al.* 2005); however the two molluscs, the mussel *P. purpuratus* and
201 abalone *C. concholepas* do not. The association of genetic structure with the southern
202 biogeographic boundary near 42°S (Thiel *et al.* 2007) is far more varied; other taxa with
203 shorter distributional ranges that span this biogeographic transition, such as the mussel
204 *Mytilus chilensis*, show little spatial structure at mitochondrial or other putatively neutral
205 markers (L. Besch and Bockrath, unpublished; Areneda *et al.* (2016)) but can be
206 distinguished among different coastal environments by outlier markers (Areneda *et al.*
207 2016) and expression profiling (Núñez-Acuña *et al.* 2012). Ewers-Saucedo *et al.* (2016)
208 note that environmental transitions and current-mediated larval dispersal in this region,
209 where trans-oceanic currents are separated as they reach the continental margin (Acha *et*
210 *al.* 2004), are likely to transport regionally-differentiated diversity along a broad swath of
211 this coastline. Thus, identifying concordant intraspecific diversity patterns among taxa may
212 require a different analytical approach that is model-driven as in Ewers-Saucedo *et al.*
213 (2016).

214

215 There is an expanding interest in exploration of genetic diversity within and among
216 regional populations of intertidal species along the coast of Chile (see Haye et al. 2014 for a
217 recent synthesis). Such data are being used to explore the underlying causes of
218 biogeographic transition (Cardenas *et al.* 2009; Ewers-Saucedo *et al.* 2016; Zakas *et al.*
219 2009), to inform management and aquacultural concerns (Haye & Munoz-Herrera 2013;
220 Núñez-Acuña *et al.* 2012; Pappalardo *et al.* 2016), and better understand how the dynamics
221 of a coastal ocean influence local diversity (Aiken & Navarrete 2014; Broitman *et al.* 2001;
222 Navarrete *et al.* 2005). For example, even with variation among the data and taxa evaluated
223 here, there is a concordance between the genetic transitions exhibited in these taxa and
224 regions of strong upwelling along coastal Chile (Navarrete *et al.* 2005).

225 What remains unsatisfying is our ability to predict – based on what we know of life history,
226 ecology, and other parameters of a given taxon – which species are likely to exhibit
227 structure across a certain region. Haydon et al. (1994) first noted the problem of both
228 stochastic and deterministic contributions to biogeography and overall population
229 structure. Certainly some ‘significant’ phylogeographic structure may simply represent the
230 interaction of genealogical processes and modest limitations on gene flow (Irwin 2002).
231 However, the most direct contrast of the taxa included here involves the barnacles *N.*
232 *scabrosus* and *J. cirratus*, which are ecologically nearly indistinguishable (Lamb *et al.* 2014;
233 Shinen & Navarrete 2010, 2014) with little known distinction in larval life history. In fact,
234 though *N. scabrosus* exhibits significant phylogeographic structure (Ewers-Saucedo *et al.*
235 2016), the larvae of *N. scabrosus* appear to require longer times in the plankton and longer
236 times for cyprid metamorphosis than *J. cirratus* (Venegas *et al.* 2000). Whether the cause

237 for this contrast in population structure is ecological, physiological, or simply **fortune**
238 remains unclear.

239

240

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247

248

249 **Literature Cited**

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Table 1 Collection sites, number of individuals per sampling site (n) and summary statistics of genetic variability for *Jehlius cirratus*.

Site (South Latitude)	sampled	haplotypes	haplotype diversity	nucleotide diversity (π)
Antofagasta/Arica (18.49°)	31	27	0.978±0.020	0.012±0.009
Huasco (28.46°)	41	25	0.945±0.022	0.009±0.003
Temblador (29.40°)	21	16	0.948±0.040	0.009±0.006
Guaqueros (30.20°)	24	18	0.942±0.040	0.011±0.006
Punta Talca (30.95°)	23	14	0.893±0.052	0.008±0.004
Los Molles (32.25°)	28	23	0.971±0.024	0.011±0.007
Monte Mar (32.95°)	28	24	0.987±0.014	0.011±0.006
El Quisco (33.45°)	29	25	0.988±0.013	0.010±0.006
Las Cruces (33.49°)	17	16	0.993±0.023	0.012±0.006
Matanzas (33.95°)	24	20	0.975±0.024	0.011±0.006
Pichilemu (34.42°)	32	24	0.958±0.025	0.010±0.008
Niebla (39.85°)	25	17	0.957±0.024	0.014±0.008
Añihue (43.85°)	8	7	0.964±0.077	0.016±0.009
Isla Madre de Dios (50.42°)	7	3	0.667±0.160	0.009±0.004

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391 **Table 2** Pairwise Φ_{ST} values among sites (indicated as header) for mitochondrial COI sequence data in *J. cirratus*. Statistically
 392 significant ($p < 0.01$) comparisons are bolded and in blue. The sample from Antofagasta includes the single available sequence
 393 from Arica.

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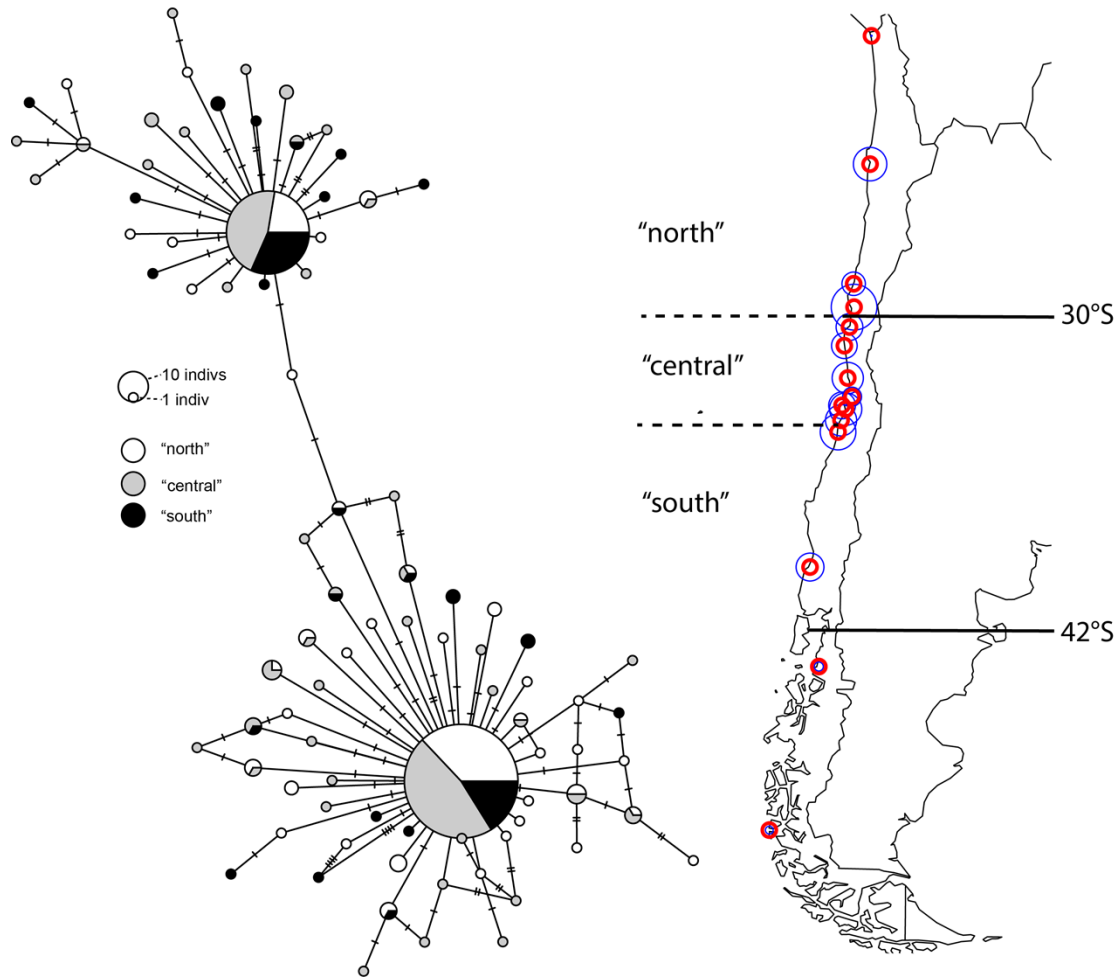
Antofagasta	Huasco	Temblador	Guanaqueros	Punta Talca	Los Molles	Monte Mar	El Quisco	Las Cruces	Matanzas	Pichilemu	Niebla	Añihue
-0.10721												
-0.02397	-0.10075											
-0.06007	0.00344	-0.09836										
-0.00797	-0.07271	0.01272	-0.01539									
-0.01641	-0.09486	-0.01873	-0.07157	0.00493								
-0.07084	0.01909	-0.06296	0.05349	-0.0808	-0.03693							
-0.17547	-0.01582	-0.18666	0.02576	-0.1819	-0.15953	-0.03391						
-0.00509	-0.06798	0.00201	-0.02185	-0.02005	0.01097	-0.08597	-0.16477					
-0.07137	0.01015	-0.05613	0.04841	-0.0811	-0.04482	-0.0131	-0.02592	-0.07314				
0.06509	0.01927	0.10959	0.10642	0.01976	0.085	-0.01377	-0.10077	0.04336	-0.02223			
-0.03313	-0.0885	0.01678	-0.04187	-0.04029	-0.02781	-0.09641	-0.21442	-0.03887	-0.10159	-0.01699		
-0.01175	0.02556	-0.00176	0.07232	-0.03869	0.00933	-0.03799	-0.02988	-0.04939	0.00464	0.02127	-0.05271	
-0.0777	0.01877	-0.04544	0.08615	-0.11043	-0.08512	0.04286	-0.00793	-0.07119	0.03113	-0.09806	-0.13056	0.04426

398 **Table 3** Iterative AMOVA for K=2 regions of sequence diversity. Site is listed as dividing
 399 *that location and all sites to the north* from all locations to the south. The northernmost 2
 400 sites (Arica, Antofagasta) were pooled for analysis as were the southernmost 2 sites
 401 (Añihue, Madre de Dios). Strongest values of Φ_{CT} (by magnitude and p-value) indicated in
 402 bold. Similar value of Φ_{CT} (0.0366, $p < 0.001$) is obtained with K=3 and the regions
 403 separated as in Figure 1.
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Site	Φ_{CT}	p-value
Huasco	0.01406	0.16
Temblador	0.01977	0.11
Guaqueros	0.03679	<0.001
Punta Talca	0.02623	0.03
Los Molles	0.03215	<0.01
Monte Mar	0.02998	0.01
El Quisco	0.02896	<0.01
Las Cruces	0.03463	<0.01
Matanzas	0.03615	<0.005
Pichilemu	0.00076	0.55
Niebla	0.00635	0.64

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413 **Figure 1.** Minimum-spanning tree of mitochondrial COI diversity in *J. cirratus*. Regional designations are generated from
 414 maximal Φ_{CT} values along the coast. Map to right of figure indicates the hypothesized transitions of species and genetic
 415 diversity noted from previous work (30°S, 42°S) and the regional separation of diversity supported by analyses of molecular

416 variance in this study (“north”, “central”, and “south”). Red circles indicate sample locations along the coast; blue circles
417 represent log-transformed sample size (see Table 1).