

1 [Symbiodinium spp. associated with scleractinian corals from Dongsha Atoll \(Pratas\),](#)
2 [Taiwan, in the South China Sea](#)

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33 **Abstract**

34 Dongsha Atoll (also known as Pratas) in Taiwan is the northernmost atoll in the South
35 China Sea and a designated marine national park since 2007. The marine park's scope of
36 protection covers the bio-resources of its waters in addition to uplands, so it is important to
37 have data logging information and analyses of marine flora and fauna, including their
38 physiology, ecology, and genetics. As part of this effort, we investigated *Symbiodinium*
39 associations in scleractinian corals from Dongsha Atoll through surveys carried out at two
40 depth ranges (shallow, 1-5 m; and deep, 10–15 m) in 2009 and during a bleaching event in
41 2010. *Symbiodinium* composition was assessed using restriction fragment length
42 polymorphism (RFLP) of 28S nuclear large subunit ribosomal DNA (nlrDNA). Our results
43 showed that the 796 coral samples from [seven](#) families and 20 genera collected in 2009 and
44 132 coral samples from [seven](#) families and 12 genera collected in 2010 were associated with
45 *Symbiodinium* C, D and C+D. Occurrence of clade D in shallow water (24.5%) was higher
46 compared to deep (14.9%). Due to a bleaching event in 2010, up to 80% of coral species
47 associated with *Symbiodinium* C underwent moderate to severe bleaching. Using the fine
48 resolution technique of denaturing gradient gel electrophoresis (DGGE) of internal
49 transcribed spacer 2 (ITS2) in 175 randomly selected coral samples, from 2009 and 2010,
50 eight *Symbiodinium* C types and two *Symbiodinium* D types were detected. This study is the
51 first baseline survey on *Symbiodinium* associations in the corals of Dongsha Atoll in the South
52 China Sea, and it shows the dominance of *Symbiodinium* clade C in the population.

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56 **Introduction**

57 Coral reefs provide habitat for numerous marine organisms and are considered among
58 the richest ecosystems on earth. In addition to their ecological values, they are also
59 economically important as they contribute toward food, tourism, coastal protection,
60 aesthetics, and cultural significance to people in coastal areas (Moberg and Folke 1999;
61 Hoegh-Guldberg 2004; Wilkinson 2004) and act as ecosystem engineers (Jones *et al.* 1994;
62 Coleman and Williams 2002). As a result of climate change and anthropogenic disturbances,
63 corals and coral reefs in the recent decades (30-40 years) have suffered an unprecedented
64 decline in terms of species abundance and community degradation (Hoegh-Guldberg 1999;
65 Coles and Brown 2003; Hughes *et al.* 2003; Bellwood *et al.* 2004).

66 Increasing seawater temperature is considered one of the main causes of this decline
67 (Hoegh-Guldberg *et al.* 2007). Generally, corals undergo a phenomenon known as bleaching
68 when confronted with 1.0-2.0 °C above the mean summer average seawater temperatures.

69 Bleaching is a result of the breakdown of symbiosis between the coral host and single celled
70 algae *Symbiodinium*, either due to release of *Symbiodinium* cells by the host or escape of the
71 cells from the host (see Weis 2008). While a majority of corals undergo bleaching, some
72 coral species can resist thermal stress or changes in environmental conditions. This is due
73 either to the ability of the coral host to withstand stress or by associating with a stress-
74 resistant type of *Symbiodinium*, or to a combination of the two (Berkelmans and van Oppen
75 2006; Bhagooli *et al.* 2008; Baird *et al.* 2009).

76 Based on various genetic markers (Rowan and Powers 1991a; Rowan and Powers
77 1991b; LaJeunesse 2001; van Oppen *et al.* 2001; Pochon *et al.* 2001; Fabricius *et al.* 2004;
78 Coffroth and Santos 2005; Pochon *et al.* 2006; Stat *et al.* 2009), *Symbiodinium* are
79 genetically diverse and has been classified into clades, types and ectomorphs. Different
80 host-symbiont assemblages can respond differently to diverse conditions, such as
81 temperature, irradiance, and sedimentation disturbance. Most *Symbiodinium* clades/types are

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86 associated with specific host genera or species (LaJeunesse *et al.* 2004a). In many cases,
87 *Symbiodinium* clades/types associate with only one or a few host species. However, studies
88 also have shown that one of the two partners is more flexible (Baker 2003a) and occupies
89 defined ecological niches and roles within and across coral hosts (LaJeunesse *et al.* 2010b;
90 Pochon and Gates 2010; Weber and Medina 2012) based on their physiological responses to
91 various environmental stresses (Iglesias-Prieto *et al.* 2004; Baker 2003a, 2003b; Jones *et al.*
92 2008; Little *et al.* 2004; Rowan 2004; Sampayo *et al.* 2008; Warner *et al.* 2006).

93 *Symbiodinium*-related stress-resistant mechanisms could be a result of strict association
94 with a stress resistant clade/type of *Symbiodinium* or, in case of coral host with multiple
95 *Symbiodinium* association, the result of shuffling between stress sensitive and stress resistant
96 *Symbiodinium* clades/types. The capacity to shuffle *Symbiodinium* clades/types may be key
97 for their acclimatization and/or adaptation (Baker and Romanski 2007; Mieog *et al.* 2007).
98 The general trend is for thermal-tolerant clades/types to supersede thermal-sensitive
99 clades/types under temperature stress (Jones *et al.* 2008; Jones and Berkelmans 2010).

100 As coral and coral reefs face more frequent and intense bleaching events due to climate
101 change and anthropogenic stressors, it is necessary to document the coral-*Symbiodinium*
102 associations from locations which have been ignored or those that are remote. A recent study
103 from western Australia have shown presence of some unique *Symbiodinium* types as well as
104 added to the information on coral-*Symbiodinium* associations and *Symbiodinium* diversity
105 (Silverstein *et al.* 2011). Such studies will help us to understand the current status and predict
106 future response of the corals in new and remote locations.

107 Dongsha Atoll in the South China Sea is one such remote location. Although many
108 studies have been conducted on coral-*Symbiodinium* associations in the South China Sea, no
109 such efforts have been realized in Dongsha Atoll. The Dongsha Atoll Marine National Park
110 was established in 2007 in order to implement legal jurisdiction for conservation efforts.
111 Several studies have been conducted on its reef-building corals despite its remote location.

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119 For example, Ma (1937) studied growth rates in different coral species. Coral biodiversity
120 (17 genera and 45 species) was first described by Yang et al. (1975), followed by Fang et al.
121 (1990), who reported 28 genera and 63 species. Later, Dai et al. (1995) recorded 34 genera
122 and 101 species of scleractinian corals, eight genera and 33 species of octocorals, and one
123 genus and three species of hydrocorallina. Recently, and especially after 1998, interest has
124 arisen on the effects of mass coral bleaching of the coral reef community of Dongsha Atoll.
125 After the exceptionally high 1997-98 summer temperatures during an *El Niño–Southern*
126 *Oscillation* (ENSO) bleaching event, different studies reported a decrease in its coral cover
127 and biodiversity (Fang 1998; Li and Fang 2002; Soong *et al.* 2002). Moreover, Li *et al.*
128 (2000) showed a decline in coral species number due to the extensive use of poisons and
129 explosives from illegal fishing. Before 1998, *Porites* and *Acropora* were the most abundant
130 and widespread genera in Dongsha (Fang *et al.* 1990; Dai *et al.* 1995). Although *Porites* has
131 remained a dominant genus from 1998 to the present, it has shared its dominance with other
132 species belonging to the Merulinidae and Fungiidae (Dai 2013).

133 Although the species diversity of scleractinian corals at Dongsha has been investigated
134 extensively (Fang *et al.* 1990; Li *et al.* 2000; Soong *et al.* 2002; Dai *et al.* 1995; Dai 2005,
135 2006, 2008, 2013), no studies have been carried out on the coral- *Symbiodinium* associations.
136 Dongsha Atoll is characterized by well-developed tropical atoll reefs with 281 known
137 scleractinian coral species (Dai 2013). Direct anthropogenic disturbances on the atoll are
138 minimal (about 200 military personnel and park rangers) since Dongsha is a military-
139 exclusive area and a designated marine national park. However, there is occasional human
140 disturbance by fishermen from nearby coastal nations, including China, Philippines, and
141 Vietnam. Hence, the aim of this study was to conduct a baseline survey on *Symbiodinium*
142 associations in the corals of Dongsha Atoll and document the *Symbiodinium* diversity in
143 shallow and deep waters at different sites in the lagoon of the Atoll in 2009 and 2010. In this

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149 paper, we report the results from the preliminary survey carried out in the lagoon of Dongsha

150 Atoll on coral-*Symbiodinium* associations.

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154 **Materials and methods**

155 *Description of the study site and sample collection*

156 Dongsha Atoll (20°35' - 47'N and 116°41' - 55'E), also known as Pratas Island (Fig. 1),
157 is located in the northern South China Sea, 850 km southwest of Taiwan. Dongsha is a
158 circular atoll about 25 km in diameter and its central lagoon covers more than 600 km².
159 Depths in the lagoon range 10-15 m with a maximum depth of 23.7 m (Dai, 2013).
160 Composed of coral debris, Dongsha Island is small (2,860 m long 865 m wide), 2 m above
161 sea level at most, covers a total surface area of 1.74 km², and is located in the western part
162 of the atoll. The circular reef flat around the lagoon spreads over 46 km and its maximum
163 width reaches 2 km.

164 Coral samples were collected in June and September 2009 and September 2010 in the
165 Dongsha Atoll Lagoon (DAL) (Fig. 1). There was a bleaching episode during summer of
166 2010 due to elevated seawater temperatures. Bleached and non-bleached corals were
167 determined by visual survey of color variation between healthy and bleached corals.
168 Sampling was limited to the lagoon due to military-restricted access to the reef crest and
169 outside the atoll. A total of 928 scleractinian coral samples including seven families with 20
170 and 12 genera in 2009 and 2010 respectively. were collected at two depths: 1-5 m (shallow)
171 and 10-15 m (deep). Samples in 2009 and 2010 were collected from nine and four sites,
172 respectively. Samples (~5-10 cm) from coral colonies were randomly collected by hammer
173 and chisel. All the samples were preserved in 70% ethanol (v/v) and kept at 4 °C until
174 processing. Scleractinian corals were identified to species when possible. Scleractinian
175 taxonomy followed Veron and Stafford-Smith (2000), Dai and Horng (2009a, b), and Huang
176 *et al.* (2014). *In situ* seawater temperatures at every site and at both depths were recorded
177 with temperature loggers (HOBO, Pendant™, Onset Computer Corporation, Massachusetts,
178 USA) at 30 min intervals (the periods of record were June 17 to September 15, 2009, and
179 May 28 to September 6, 2010).

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183 *DNA extraction*

184 Total genomic DNA (coral host + *Symbiodinium*) was extracted by the salting-out
185 method (Ferrara *et al.* 2006). Coral tissue was lysed overnight in a 2 mL Eppendorf tube
186 with 200 μ L of lysis buffer [0.25 M Tris, 0.05 M EDTA at pH 8.0, 2% sodium dodecylsulfate
187 (SDS), and 0.1 M NaCl] and 10 μ L of 10 mg mL⁻¹ proteinase E at 55 °C in a water bath.
188 NaCl (210 μ L at 7 M) was added to the lysed tissue in the tube and the sample mixed
189 carefully by inverting the tube. The solution was then transferred to a 2 mL collection tube
190 containing a DNA spin column (Viogene, USA) and centrifuged at 8000 rpm for 1 min. The
191 lysate was washed twice with 500 μ L of ethanol (70%) by centrifuging at 8000 rpm for 1
192 min at each step, with an additional centrifugation step at 8000 rpm for 3 min to dry the spin
193 column. The column was dried further at 37 °C for 15 min and the DNA then eluted by
194 adding 150 μ L of distilled water, with a final centrifugation at 14,000 *g* for 3 min. The quality
195 of genomic DNA was checked using a 1% agarose gel and the concentration of eluted DNA
196 was examined using NanoDrop and then stored at -20 °C for further analysis.

198 Molecular identification of *Symbiodinium* clades (28S RFLP) and types (ITS2 DGGE)

199 A total of 928 samples (20 genera in 2009 and 12 genera in 2010) were used to analyze
200 *Symbiodinium* clade composition using RFLP method modified from Chen *et al.* (2005a, b).
201 The 5' end of nuclear large subunit ribosomal DNA (nlrDNA) was amplified using a
202 *Symbiodinium*-specific primer set, 28S_{zoox-D1/D2F} (5'-CCT CAG TAA TGG CGA ATG
203 AAC A-3') and 28S_{zoox-D1/D2R} (5'-CCT TGG TCC GTG TTT CAA GA-3') (Loh *et al.*
204 2001). A 25 μ L PCR reaction consisting of 3 μ L DNA (10 ng μ L⁻¹), 2 μ L dNTPs (0.8 mM), 2
205 μ L forward primer (0.16 μ M), 2 μ L reverse primer (0.16 μ M), 3 μ L PCR Buffer (1.2X), 0.5
206 unit *Taq* polymerase (Protech, Taiwan), and 12.5 μ L distilled water was run on a Px2 thermal
207 cycler (Thermo Scientific, USA). The PCR cycling profile consisted of initial denaturation
208 at 95 °C for 1 min followed by 5 cycles of 94 °C for 30 s, 30 cycles of annealing at 55 °C

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211 for 1 min and decreasing 1 °C to a final annealing temperatures of 50 °C and 2 min at 72 °C.
212 The final extension was at 72 °C for 10 min. The PCR product was digested with Rsa I
213 (BioLabs, USA) solution at 37 °C overnight and then run on 3% agarose gel (2% low melting
214 agarose with 1% agarose) at 50 V for 3 h. Bands were stained using ethidium bromide (EtBr)
215 and visualized under ultraviolet radiation.

216 For ITS2 *Symbiodinium* type composition, 168 samples were randomly picked from 17
217 genera in 2009 and 11 genera in 2010. The ribosomal internal transcribed spacer 2 (ITS2)
218 region of *Symbiodinium* was amplified using the primers ITSintfor2, 5'-GAA TTG CAG
219 AAC TCC GTG-3', and ITS2clamp, 5'-CGC CCG CCG CGC CCC GCG CCC GTC CCG
220 CGG GAT CCA TAT GCT TAA GTT CAGCGG GT-3' (LaJeunesse and Trench 2000).
221 Each 50 µl polymerase chain reaction (PCR) comprised 50 ng of genomic DNA, PCR buffer,
222 2.5 mM MgCl₂, 0.4 mM dNTPs, 0.4 µM of each primer, and 2 units of Taq polymerase
223 (Invitrogen, USA). The PCR was run on a Px2 thermal cycler (Thermo Scientific, MA, USA)
224 with a touch-down PCR (LaJeunesse 2002) to ensure specificity. An initial denaturation
225 period at 92 °C for 3 min was followed by 20 cycles of 30 s at 92 °C, annealing from 62 °C
226 to a final annealing temperature of 52 °C with decrements of 0.5 °C, and 30 s at 72 °C. Once
227 the annealing temperature reached 52 °C, a further 20 cycles were performed at that
228 annealing temperature, with a final extension period of 10 min at 72 °C. Each PCR product
229 along with *Symbiodinium* type markers (known ITS2 sequences confirmed by comparing
230 with NCBI GenBank database) was loaded onto an acrylamide denaturing gradient gel (45–
231 80%) and electrophoresed at 115 V for 15 h using a CBS Scientific system (Del Mar, CA,
232 USA). Gels were stained with SYBR Green (Molecular Probes, Eugene, OR, USA) for 15
233 min and photographed for further analysis. Band patterns were confirmed by sequencing the
234 bands cut from the DGGE gel.

235 *Statistical analysis*

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237 Shallow and deep-water temperature comparisons were analyzed by paired *t*-tests and average
238 temperature values presented as means ± standard deviation (SD). The frequency of daily maximum
239 temperature fluctuation at each location and depth was calculated followed by difference in
240 temperature between shallow and deep waters. Finally, the percentage of over 30°C temperature
241 during the monitoring period was calculated. The *Symbiodinium* clade values in 2009 and 2010 were
242 converted to percentage within and between genera and within and between two depths. The
243 proportion of *Symbiodinium* clades in coral genera in 2009 and 2010 and between shallow and deep
244 waters was analyzed using a chi-square test. All graphs were drawn using Aabel (Ver. 3.0, Gigawiz
245 Ltd. Co., USA) or Datagraph (Visual Data Tools, USA) software for the Macintosh platform.

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Mean daily temps, mean hourly temps? Please elaborate here
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250 **Results**

251 *Seawater temperature*

252 Seawater temperature was recorded from June 17 to September 15, 2009, and May 28 to September
253 6, 2010 (Fig. 2A and B, Supplementary data files S1-S3). During the 2009 period, temperatures at
254 shallow waters (29.55 ± 0.23 °C) were higher than at deep waters (28.82 ± 0.19 °C; Fig. 2A; Table
255 1; paired *t*-test, $p < 0.001$). Similarly, in 2010, temperatures at shallow waters (29.94 ± 0.29 °C)
256 were higher than at deep waters (29.72 ± 0.32 °C; Fig. 2A; Table 1; paired *t*-test, $p < 0.001$).
257 Overall, ΔT was 0.62 ± 0.3 °C in 2009 and 0.22 ± 0.14 °C in 2010. Daily temperature differences
258 between shallow and deep waters in 2009 were 0.5-1.0 °C and 0-0.5 °C in 2010. In 2009, during the
259 period monitored, 36% of the temperatures recorded at the shallow waters exceeded 30.0 °C,
260 whereas only 13% of the temperatures recorded at the deep waters reached this value. In 2010, for
261 shallow and deep waters, 61% and 55% of the temperatures recorded exceeded 30.0 °C. This was
262 also confirmed by the heat map of seawater temperatures measured at reef base and reef top during
263 June- September and May-September in 2009 and 2010 (Fig. 2 B)

264 In this study, samples were collected from nine locations in the DAL. Since the sampling was
265 random and uneven within and between locations and between two sampling times (2009 and
266 2010), we present results from 2009 and 2010 by combining the data for locations and depths,
267 rather than discussing the results from individual locations and depths. Hence, we present the
268 overall picture of coral-*Symbiodinium* associations in DAL. Data of *Symbiodinium* clades in coral
269 genera at different locations is provided as supplementary data file S4.

270

271 *Symbiodinium* clade composition in 2009

272 A total of 796 samples from 20 genera and seven families were collected in 2009 (Fig.
273 3 and 4). RFLP band patterns revealed the presence of *Symbiodinium* clades C, D and C +
274 D.

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277 *Symbiodinium* Clade C occurred in all 20 genera at shallow and deep waters in 2009.
278 *Leptastrea* (100%), *Fungia* (100%), and *Porites* (100%) were found to harbor only
279 *Symbiodinium* clade C at shallow and deep waters. *Cyphastrea* (100%), *Hydnophora*
280 (100%), *Montipora* (100%), and *Psammocora* (100%) were associated only with
281 *Symbiodinium* clade C at deep waters. At shallow waters, the proportion of *Symbiodinium*
282 clades was 64.18% (clade C), 24.5% (clade D), and 11.34% (clades C+D). And, at deep
283 waters, the proportion of *Symbiodinium* clades was 78.6% (clade C), 14.9% (clade D), and
284 6.53% (clades C+D). The clade D proportion in comparison to clade C was significantly
285 different at shallow and deep waters (chi-square test, $p = 0.007$). The clade D proportion was
286 high at shallow waters (24.5%) compared to deep waters (14.9%). At both depths, most
287 corals were associated with > 50% of *Symbiodinium* clade C, the exceptions being *Oxypora*
288 (33 % in shallow and 44% in deep), *Hydnophora* (22% in shallow), *Echinopora* (9% in
289 shallow and 24% in deep), *Favites* (33% in shallow), *Coelastrea* (37% in shallow), *Astrea*
290 (25% in shallow), *Pectinia* (38% in shallow) and *Goniopora* (43% in shallow), which were
291 associated > 50% of the time with clades D or C+D. However, care should be taken when
292 interpreting the results from this data since the sample number was less than ten for most of
293 these genera.

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294 Compared to other genera, Clade D proportions in *Turbinaria* and *Coelastrea* were
295 significantly different when comparing shallow to deep (chi-square test, $p < 0.05$). The clade
296 D proportion of *Turbinaria* at shallow waters (39.5%) was higher than at deep waters
297 (10.8%), and in *Coelastrea* was 46.7% at shallow waters and 15% at deep waters.

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299 *Symbiodinium* clade composition in corals during the 2010 bleaching episode

300 A total of 132 samples from 12 genera and seven families were sampled in 2010 from
301 shallow and deep waters (Fig. 3 and 4). *Symbiodinium* clade D showed significant
302 differences (chi-square test, $p < 0.001$) between bleached and non-bleached corals. Clade D

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306 proportions were extremely high in non-bleached corals (62.8%) compared to bleached
307 corals (10.9%). Bleached corals were dominated by *Symbiodinium* clade C at each location.
308 *Astreopora*, *Psammocora*, *Fungia*, *Cyphastrea*, *Porites*, and *Oxypora*, all associated with
309 clade C, experienced bleaching in Dongsha Lagoon in the summer of 2010, and *Pavona*,
310 *Coelastrea* and *Echinopora* still experienced bleaching even though they were associated
311 with clades D or C+D.

312

313 *Symbiodinium* type composition in corals in 2009 and 2010

314 There was no particular trend in the *Symbiodinium* type composition between [the](#) two
315 sampling years due to random sampling of the coral colonies (Table 2). However,
316 *Symbiodinium* composition was dominated by different types of *Symbiodinium* C. ITS2
317 DGGE analysis of DNA samples revealed the presence of eight *Symbiodinium* types (C1,
318 C1b, C3, C15, C21a, C27, C30 and C40 Table 2). C15 was mainly associated with
319 *Montipora* and *Porites*. The composition of *Symbiodinium* type D consisted of only two
320 types; *Symbiodinium glynii* (D1) and *Symbiodinium trenchii* (D1a), either separately or in
321 combination, depending on the [coral host](#) genus. The two species of *Symbiodinium* D are
322 separated based on a one base-pair difference on a DGGE gel. Because of this fine
323 difference, we ran the gels with appropriate markers for *S. glynii* and *S. trenchii* and also
324 sequenced the cut bands from the ITS2 DGGE gels to reveal the presence of *S. glynii* in
325 several genera of corals sampled from DAL (Fig S1).

326 Out of 17 genera analyzed in 2009, the genera *Montipora*, *Acropora*, *Pavona*,
327 *Oxypora*, *Echinopora* *Favites*, *Astrea*, *Coelastrea*, *Gonipora*, *Hydnopora* and *Pachyseries*
328 were associated with *S. glynii* and/or *S. trenchii* (Table 2). Out of 12 genera analyzed in
329 2010, the genera *Pavona*, *Turbinaria*, *Oxypora*, *Echinopora*, *Astrea* and *Gonipora* were
330 associated with *S. glynii* and/or *S. trenchii* (Table 2).

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334 **Discussion**

335 This is the first study to investigate *Symbiodinium* association in the corals of Dongsha
336 Atoll Lagoon (DAL). The main results of this study are that (1) *Symbiodinium* composition
337 in DAL consists of two clades, C and D; (2) eight *Symbiodinium* C types were detected
338 compared to only two *Symbiodinium* D types; and (3) up to 80% of coral species associated
339 with *Symbiodinium* C underwent moderate to severe bleaching in the year 2010. Since we
340 have utilized both RFLP and DGGE for analysis, to avoid confusion, throughout the
341 discussion, clades and types will not be used and instead we will just use either
342 *Symbiodinium* C or *Symbiodinium* D. We also use species name for those *Symbiodinium* types
343 that have been designated formally.

344 Out of 928 samples collected from 20 (in 2009) and 12 (in 2010) genera of scleractinian
345 corals, 598 hosted *Symbiodinium* C, suggesting its dominance in the South China Sea (see
346 Table 3). The association of corals with *Symbiodinium* C, in addition to physiological
347 factors, could also be due to seawater temperatures being generally below the bleaching
348 threshold in Dongsha (Fig. 2, Table 1).

349

350 *Symbiodinium* association in corals in 2009

351 In 2009, despite *Symbiodinium* C being dominant, it is interesting to highlight the
352 differences seen in *Symbiodinium* D. There was higher proportion (24.5%) in shallow waters
353 compared to deep waters (14.9%). Moreover, the occurrence of *S. glynii* and *S. trenchii* was
354 higher in corals sampled from the shallow waters. The frequency of temperature fluctuations
355 > 1°C at shallow waters was higher than at deep waters, which might be one of the reasons
356 for the higher proportions of *Symbiodinium* D at shallow than at deep waters.

357 In the present study, *Astreopora*, *Pavona*, *Montipora*, *Psammocora*, *Porites*, *Fungia*
358 and *Leptastrea* showed stable associations with *Symbiodinium* C at shallow and deep waters
359 in DAL, suggesting that either host might be physiologically adjusted to temperature stress

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370 or their associated *Symbiodinium* C might have greater photosynthetic stability under
 371 thermal stress (Berkelmans *et al.* 2004; Thornhill *et al.* 2005; van Oppen *et al.* 2005).
 372 *Turbinaria*, *Oxypora*, *Hydnophora*, *Coelastrea*, and *Favites*, which were dominated by
 373 *Symbiodinium* D at shallow compared to deep waters, suggest plasticity in these corals to
 374 warmer waters via *Symbiodinium* D. The coral *Echinopora* had a stable and dominant
 375 association with *Symbiodinium* D at shallow and deep waters. In the present study, the
 376 proportion of *Symbiodinium* D showed a significant difference only in some locations, which
 377 might be due to difference distribution of coral host associated with *Symbiodinium* D.
 378
 379 *Symbiodinium* association in corals in 2010 during a bleaching event
 380 In 2010, scleractinian corals at Dongsha suffered from high seawater temperature stress
 381 caused by a La Niña event. *Astreopora*, *Cyphastrea*, *Fungia*, *Psammacora* associated with
 382 clade C including the genus *Porites*, which is generally considered as stress resistant
 383 (associated with *Symbiodinium* C15) experienced bleaching. Moreover, coral genera
 384 *Pavona*, *Coelastrea*, *Echinopora*, which were associated with clade C or C+D, also
 385 experienced bleaching. Bleaching was seen in 84 % of corals associated with clade C
 386 compared to only 10.87% of corals associated with clade D. Bleaching in those corals
 387 associated with clade D could be due to seawater temperature above threshold limits during
 388 the summer of 2010. In a previous study, the critical coral bleaching temperature threshold
 389 at Dongsha was determined to be 29.6 °C (Dai, 2008). During May 28 to September 6, 2010,
 390 average temperatures at shallow waters (29.9 °C) and deep waters (29.7 °C) within DAL
 391 exceeded the critical value and may have induced coral bleaching (Fig. 2, Table 1).
 392 Consistent above 30 °C seawater temperature for more than a week, both in shallow and
 393 deep waters, could have resulted in extensive bleaching in corals in 2010 (Fig. 2, Table 1).
 394 As a result of random and uneven sampling, it is not possible to analyze the correlation
 395 between *Symbiodinium* clades and seawater temperature results. We suggest that future

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Deleted: diversity (habitat diversity) and abundance (habitat availability), and these may be the factors that influence *Symbiodinium* composition.

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408 studies in DAL should focus on obtaining samples from tagged coral colonies in different
409 seasons and depths to understand the dynamics of coral-*Symbiodinium* associations.

410

411 *Symbiodinium* type composition in corals in 2009 and 2010

412 From ITS2 DGGE analysis, DAL corals were found to be associated with [eight](#)
413 *Symbiodinium* C (*Symbiodinium goreau* (C1), C1b, C3, C15, C21a, C27, C30 and C40) and
414 this number is greater than in the corals at Kenting, southern Taiwan (Keshavmurthy *et al.*
415 2014). Conversely, only two *Symbiodinium* D types were observed: *Symbiodinium glynii*
416 (D1) and *Symbiodinium trenchii* (D1a). The presence of *Symbiodinium* D in corals sampled
417 in 2010 was more obvious, and we believe this might have been due to the influence of
418 seawater temperature (Fig. 2, Table 1). Our results also showed that *S. glynii* was dominant
419 in many coral genera, both in 2009 (12 genera out of 20) and 2010 (7 genera out of 12),
420 indicating that *S. glynii* may not be specific to any particular host species (LaJeunesse 2002;
421 Baker 2003). This is the first work to show the presence of *S. glynii* in such a large number
422 of coral genera and hence, we believe that *S. glynii* may not be as specific or rare, as
423 [previously thought](#) (see LaJeunesse *et al.* 2010a).

424

425 *Conclusions*

426 The present study has shown that *Symbiodinium* C is dominant at Dongsha Atoll in the
427 South China Sea corals and that natural stressors, such as [elevated](#) seawater temperatures, can
428 influence *Symbiodinium* associations in Dongsha Atoll corals. The baseline information provided on
429 the composition of *Symbiodinium* in corals from Dongsha Atoll through the present work will help,
430 carrying out comprehensive and detailed studies on the diversity of *Symbiodinium* in corals over
431 time. By analyzing data from normal and coral-bleaching years, we have shown that the
432 composition of *Symbiodinium* in some corals is different between years in remote and undisturbed
433 locations such as Dongsha Atoll. Such studies are important for understanding the future responses

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439 of corals to climate change in remote atolls. With increasing threats to coral health, mainly from
440 temperature-induced bleaching events as seen in the Great Barrier Reef in 2016, studies
441 documenting the *Symbiodinium* associations in corals of Dongsha Atoll will help to educate and
442 inform park managers of Dongsha Atoll Marine National Park managers about appropriate
443 management and conservation measures. The focus on the status of coral reefs in the South China
444 Sea, including Dongsha Atolls, has gained attention in the past couple of years, particularly due to
445 the recent destruction via land-filling of several reefs several atolls in the South China Sea.
446 Although offshore atoll reefs in the South China Sea are found to be in better condition than near-
447 shore reefs (Hughes *et al.* 2013), they can still undergo bleaching and mortality as a result of
448 climate change (Graham *et al.* 2006) and other stressors (Hughes *et al.* 2013). Trans boundary
449 cooperation in the South China Sea region has become important for the maintenance and
450 conservation of the various atolls and islands in the South China Sea (McManus *et al.* 2010). As
451 part of the conservation program in Dongsha Atoll, it is necessary to combine the influence of
452 natural disturbances and the effects of anthropogenic stressors, such as fishing activities, in order to
453 understand the alterations of the present and their influences on future coral communities.

454

455 **Acknowledgements**

456 We thank the staff of the Dongsha Atoll Marine National Park, Dongsha Atoll Research Station,
457 Ministry of Science and Technology (MOST), Taiwan and support from members of the Coral Reef
458 Ecology Lab, Biodiversity Research Center, Academia Sinica (BRCAS).

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