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1	Cranial bone histology of Metoposaurus krasiejowensis (Amphibia, Temnospondyli) from
2	the Late Triassic of Poland
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ABSTRACT

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In this study a detailed description of the 21 skull bones of *Metoposaurus krasiejowensis* from the Late Triassic of Poland is presented. All dermal bones show a diploë structure, with the ornamented external surface. The ridges consist of well vascularized parallel-fibered bone; the valleys are built of an avascular layer of lamellar bone. The dense clumps of thin, well mineralized Sharpey's fibers are preserved. The growth marks are manifested in four ways: a sequence of resting lines; layers of lamellar bone alternated with layers with Interwoven Structural Fibers (ISF); thick, avascular annuli and high vascularized zones; and the alternations of valleys and ridges. The thick middle region consists of cancellous bone, with varying porosity. The thin and less vascularized internal cortex consists of parallel-fibered bone. Calcified cartilage is observed in the quadrate and the exoccipital. The skull bones show strong variability within the microanatomical and histological levels. The histological framework is not bonelimited, but varies even in one single bone; this seems to be related to the specific position of the bone and depends on the local biomechanical loading of the particular part of the skull. The large accumulation of Sharpey's fibers in the occipital condyles indicates the presence of strong muscles and ligaments connecting the skull to the vertebral column. The dynamic processes during the ornamentation deposition are observed indicate that the position of the ridges and grooves change during the growth and could be the specific adaptation to biomechanical conditions and stress distribution during the bone development. In the supratemporal the



46	cementing lines indicated the remodeling process could be involved into the creations of
47	sculpture.
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49	Keywords:
50	dermal bones, skull, microanatomy, growth pattern
51	INTRODUCTION
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53	Metoposaurids were large, up to 3 meters long, late Triassic Temnospondyli with a
54	strongly dorso-ventrally flattened body, and well adapted to aquatic life. The most characteristic
55	and best known part of the Metoposaurus skeleton is the extremely flat parabolic skull with
56	anteriorly located orbits (e.g. Schoch & Milner, 2000).
57	The flat bones of the skull develop via direct transformation of preexisting connective
<mark>58</mark>	tissue (Francillion-Vieillot et al., 1990). The external surface of the dermal bones is
59	characteristically ornamented. A network of raised, reticulate ridges that enclose approximately
60	flat-bottomed, interlocking, polygonal cells is the most common type. The vast majority of these
61	cells are four-, five-, or six-sided, creating a honeycomb- or waffle iron-like texture. In some
62	temnospondyls, this is essentially the only texture present. The second texture type comprises
63	raised, parallel to sub-parallel ridges separated by round-bottomed grooves (Rinehart & Lucas,
64	2013). The function of the ornamentation is still unclear. The best supported hypotheses suggest
65	the increase the surface area for skin supports, increasing the strength of the bone or the
66	protection of blood vessels (summarized in Coldiron, 1974; Witzmann, 2009; Rinehart & Lucas,
66 67	protection of blood vessels (summarized in Coldiron, 1974; Witzmann, 2009; Rinehart & Lucas, 2013).



68	The histology of temnospondyl dermal bones was first described by Gross (1934), who
69	provided a short description of the skull bones of Mastodonsaurus, Metoposaurus and
70	Plagiosternum, and recognized that the dermal bones exhibit a diploë structure. Later,
71	histological studies on the dermal bones in Temnospondyli have focused mainly on morphology
72	vascular network and collagen fibres organization (Bystrow, 1947; Enlow & Brown, 1956;
73	Coldiron, 1974; de Ricqlès, 1981; Castanet et al, 2003; Scheyer, 2007) and were limited only to
74	the few taxa. The systematic studies of dermal bones within numerous tetrapod taxa were
75	provided by Witzmann (2009) and de Buffrénil et al. (2016).
76	Recent the histological studies of metoposaurids have focused mainly on the long bones,
77	ribs and vertebrae (Steyer et al., 2004; Gądek, 2012; Konietzko-Meier & Klein, 2013;
78	Konietzko-Meier & Sander, 2013; Konietzko-Meier et al., 2013; Koniezko-Meier et al., 2014).
79	Up until now the dermal bones of <i>Metoposaurus</i> have not been studied in detail histologically.
80	The only record of the histological description of Metoposaurus diagnosticus dermal bone was
81	given by Gross (1934) and later re-described by Witzmann (2009). However, it is unclear if the
82	illustrated section was derived from the skull or the pectoral girdle, or even if the tested bone-
83	fragment belongs to Metoposaurus at all.
84	The main goal of this study is to present the detailed description of the histology of
85	almost all bones from one skull and determine, if possible, the tendencies and variability of the
86	histological framework from the one <i>Metoposaurus krasiejowensis</i> (Sulej, 2002) skull. The
87	temnospondyl skull functionally represents one skeletal element, however, anatomically, it is a
88	conglomerate of numerous bones various in shape and thickness, having various functions and
89	biomechanical loading (i.e. Fortuny et al., 2011, Fortuny et al., 2012). Another question that
90	arises is if the various functions of the single bones are also reflected in the histology, and if the



- 91 histological data may be useful for interpretations the skull mechanics. The results of this
- 92 question, however, will be published as a separate paper.

MATERIAL AND METHODS

Material – The skull (UOPB 01029; 40 cm in length) of *Metoposaurus krasiejowensis* was studied histologically (Figs. 1 and 2). The roof side of the skull was almost completely preserved, whereas on the palatal side only the fragments of the vomer, parasphenoid, pterygoids, quadrates and exoccipitals were preserved. The species discovered in Poland was originally described as *Metoposaurus diagnosticus krasiejowensis* Sulej, 2002, the subspecies of an older *Metoposaurus diagnosticus* (Meyer, 1842). Brusatte *et al.* (2015) resigned using the subspecies for the German *M. diagnosticus diagnosticus* (von Meyer, 1842) and the Polish *M. diagnosticus krasiejowensis* and suggested to refer both taxa on the separate species level as *M. diagnosticus* and *M. krasiejowensis* instead. This taxonomy is followed in this study.

Locality – The examined material comes from the famous locality in Krasiejów where a large number of disarticulated skeletons have been discovered in the Upper Triassic (Keuper), fine-grained, continental sediments. The bones can be found in two main bone-bearing horizons referred to as the lower and the upper horizon (Dzik & Sulej, 2007). The lower horizon has been deposited on an alluvial plain during a catastrophic mud-flow event (Bodzioch & Kowal-Linka, 2012). The skull presented here, has been excavated from the less than 1 m thick lower bone-bearing layer, which is very rich in *Metoposaurus krasiejowensis* remains accompanied by relatively a high diversified fossil assemblage. Vertebrates are represented by a second temnospondyl, *Cyclotosaurus intermedius* (Sulej & Majer, 2005), a phytosaur (*Palaeorhinus*; see Dzik, 2001), a typical terrestrial tetrapod aetosaur *Stagonolepis olenkae* (Sulej, 2010), pterosaurs



114	(Dzik & Sulej, 2007), until recently undefined till now sphaenodonts and other small tetrapods
115	(Dzik & Sulej, 2007), and fishes (dipnoans described recently by Skrzycki 2014, as well as
116	various actinopterygian and chondrichthyan species). Invertebrates, such as unionid bivalves
117	(Dzik et al., 2000; Dzik & Skawina, 2011; Skawina 2013), cycloids (Dzik, 2008) spinicaudatan
118	crustaceans (Olempska, 2004), fresh-water ostracods (Olempska, 2004, 2011) and some
119	gastropods, are also very common. The upper horizon is restricted to lenses cemented with
120	calcium carbonate, interpreted as a meander deposit (e.g. Gruszka and Zieliński, 2009). It is
121	dominated by strictly terrestrial animals including Stagonolepis and the primitive
122	dinosauromorph Silesaurus opolensis (Dzik, 2003). Aquatic vertebrates such as amphibians and
123	phytosaurs are less common compared to the lower horizon. Apart from that, one fragmentary
124	specimen of the rauisuchian Polonosuchus silesiacus (Brusatte et al., 2010) was excavated
125	between the upper and lower horizons.
126	According to complex stratigraphic studies of the Upper Silesian Keuper, the bone-
127	bearing beds have been deposited in the early Norian times (Racki & Szulc, 2014; Szulc et al.,
128	2015a, b), however, biochronological speculations uphold the Late Carnian age (e.g. Dzik &
129	Sulej 2007; Lucas et al. 2007; Lucas, 2015).
130	Methods – The skull was sectioned in 20 planes (Fig. 2) and the thin sections were
131	prepared according to standard petrographic procedures (Lamm, 2014). The thin sections were
132	ground and polished to a thickness of about 60-80 µm using wet SiC grinding powders (SiC 600,
133	800). Subsequently, the thin sections were studied under a LEICA DMLP light microscope in
134	plane and cross polarized light.
135	The histological nomenclature follows Francillion-Vieillot et al. (1990) and Witzmann
136	(2009). According to Francillion-Vieillot et al. (1990) the annual growth cycle consists of a



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thick, fast growing zone, a thin, slow growing annulus, and a Line of Arrested Growth (LAG). In this study, the term zone is used as in its traditional meaning, for the highly vascularized layer, with lower organization of collagen fibers. The term annulus refers to the layer without vascular canals and higher organization of collagen fibers, but usually similar in thickness as a zone. In the studied material, no clear LAGs can be observed, instead adjacent to the annuli, numerous lines are present. To avoid nomenclature problems, all lines representing the cessation of growth are referred in this study as resting lines, without determination if they occur annually or not (Konietzko-Meier & Sander, 2013) – also see discussion.

In the thin sections, the average thickness of the entire bone and of each layer was estimated, expressed as an arithmetical average from three measurements of the thickness of the entire bone/layer. The thickness of the external layer was measured three times on the distance between the border line with middle region and the bottom of valleys and three times as the distance between the border line with middle region and the top of ridges. The mathematical average was calculated from these measurements. The minimum and maximum thicknesses represent the lowest and largest measurement, respectively, for each described layer. As the borders between external cortex/middle region/internal cortex were taken the line where the clearly visible increasing of the remodeling degree of the middle region is visible (the number of the erosion cavities, independent of their size, is visible higher). For the estimation of the ratio between the three components (E – external cortex, M – middle region, I – internal cortex; E.M.I), the average thickness of the external cortex was taken as one and then proportionally the value for middle region and internal cortex were calculated. Note that the internal cortex of dermal bone is oriented to the visceral surface of the body, thus in parasphenoid, pterygoid and vomer the external cortex is then oriented ventrally.



A detailed description of each bone is presented in the Supplementary Material.

162 RESULTS

Microanatomy of dermal bones

Most dermal bones of the skull are flat plates. Only the premaxillas and maxillas possess a more complicated shape (Fig. 3). The premaxilla is built up of three branches: the dental shelf, the alary process (Schoch, 1999) which connects the skull roof to its margin, and the vomeral process, which connects the dental shelf to the vomer (Fig. 3A). The maxilla is built up from two branches, the dorsal branch with an ornamented external cortex, and the ventral branch with the dental shelf (Fig. 3B).

The dermal bones show the clear diploë structure (Fig. 4). The external cortex of the skull-roof bones created variably ornamentations build from a combination of grooves or tubercles and ridges (Table 1, Fig. 1), respectively visible in the cross-section as valleys and ridges (Fig. 4A). The thickness of the flat bones varies from under 0.2 to over 10 millimeters (Table 1), with different proportions between the particular layers. No relation can be observed between the thickness of the external cortex and the thickness of the entire bone, i.e. the external cortex of the tabular and postorbital/jugal about one mm thick, and the average thickness of the skull bones is, respectively, almost ten and six mm (Table 1). The relatively thin squamosum 2 with an average thickness of only about three mm, developed an external cortex which takes up almost half of the bone thickness (Table 1). The largest part of the bone almost always consists of the middle region (which is two to eleven times thicker as the external cortex), with the exception of squamosum 2, where the middle region is the thinnest (Table 1). The internal cortex



is the thinnest of the three layers and composes usually only 20% to 80% of the thickness of external cortex, with the exception of the parasphenoid (Table 1).

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General histology of dermal bones

External cortex – In all sections the external cortex is consists of parallel-fibred bone, whereas in the valleys lamellar bone often occurs (Fig. 4B and C). The elongated osteocyte lacunae with branched canaliculi in the bone matrix are numerous (Fig. 5A). Vascular canals are mostly longitudinally oriented (Fig. 5A, B). The degree of vascularization varies from relatively low in the premaxilla and frontal, moderate in the maxilla and prefrontal, to highly vascularized in the jugal, postorbital, parietal, squamosum, quadratojugal, vomer and parasphenoid. In the nasal, postparietal, postfrontal, tabular and supratemporal, numerous vascular canals within the ridges are visible, which are arranged in rows parallel to the bony surface, whereas the valleys are avascular. The external cortex is dominated by simple vascular canals and primary osteons. In some bones (nasal, lacrimal, prefrontal, tabular, squamosum, vomer and parasphenoid), the secondary osteons and single erosion cavities are visible in the transition region to the middle layer (for details see the Supplementary Material). Typical for the external cortex are distinct collagen fibers (Fig. 5C, E). In some bones (jugal, postorbital, postfronat, postparietal, tabular) they create a thick Intervowen Structural Fibres (ISF) (Fig. 5E). In the premaxilla, maxilla, nasal, lacrimal, jugal, postorbital, postparietal, and quadratojugal, well-mineralized Sharpey's fibers can be observed which are relatively short but numerous, and sometimes packed densely in bundles. In the prefrontal, frontal, postfrontal, parietal, supratemporal, squamosum, tabular and vomer, Sharpey's fibers are rare and occur



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mostly in the deeper parts of the sculptural ridges (Fig. 5C). In the parasphenoid and pterygoid, Sharpey's fibers cannot be observed. Growth marks are expressed in various ways. In the ridges of the lacrimal, frontal, jugal, postfrontal, tabular, quadratojugal and squamosum 2, they are manifested as a sequence of thin resting lines (Fig. 5D). In the postparietal, a sequence of lamellar bone layers and layers consisting of ISF are visible (Fig. 5E). In the quadratojugal, two thick annuli built up of lamellar bone alternate with two highly vascularized zones (Fig. 5F). In the postfronal, squamosum 1, supratemporal, tabular, jugal and quadratojugal, the alternations of valleys and ridges are preserved. The remains of older valleys are filled with the highly vascularized tissue which then constructed the ridges of the next generation (Fig. 5G). In the supratemporal the cementing lines indicated the remodeling process involved into the creations of sculpture is visible (Fig. 5H). **Middle region** – The external cortex transits gradually into the cancellous middle region. The simple vascular canals and primary osteons, of various shapes, are mostly located next to the border between the middle and external regions. A significant part of the middle region is strongly remodeled. The few large erosion cavities (up to 2000 µm in diameter) are present in the premaxilla, nasal, lacrimal, jugal, prefrontal, postparietal, squamosum 1, quadratojugal and all studied bones from the palatal side of the skull. The most highly remodeled middle region occurs in the vomer, where erosion cavities in some areas exceed $\frac{3000 \, \mu m}{2}$ in length (Fig. 6A). In the lacrimal and prefrontal, erosion cavities reach up to the external cortex. The maxilla is dominated by numerous, but medium-sized erosion cavities. In the postfrontal, parietal and supratemporal, erosion cavities are small (less than 500 µm in diameter). In the postfrontal they appear sporadically, whereas occur numerous in parietal and supratemporal (Fig. 6B). The middle region in the tabular and squamosum 2 does not show the typical trabecular structure. The



intensive remodeling is visible; however, the tissue is relatively compact, almost without erosion cavities (Fig. 6C).

Internal cortex – The internal cortex consists of parallel-fibered to lamellar bone. The degree of vascularization varies from very low, almost avascular, in the parietal, postfrontal, supratemporal and squamosum, low in the premaxilla, prefrontal, nasal, and postparietal, moderate in the maxilla, frontal, vomer and parasphenoid, to high in the lacrimal, jugal, postorbital and quadratojugal (Fig. 6D-F). Osteocyte lacunae, showing slightly elongated shapes, are very frequent. Growth marks are visible in form of resting lines. The amount of lines varies from four in the postfrontal and parietal, three in the postparietal, supratemporal and jugal, to two in the nasale (Fig. 6D). In the parasphenoid, well developed zones and annuli can be observed (Fig. 6G). Zones are built of thick, well vascularized layers, while annuli are represented by thinner, avascular layers. The numerous Sharpey's fibers packed in bundles are visible in the tabular and vomer.

Endochondral bones

Quadrate – The partially preserved and well-vascularized cortex consists of parallel-fibered and lamellar bone (Fig. 7A, B). The simple vascular canals occur sporadically, and secondary osteons are more common (Fig. 7A, B). The Sharpey's fibers are very short and occur only in the subsurface parts of the cortex. The elongated osteocyte lacunae are present mainly within the lamellar bone, which outlines the osteons. They do not possess canaliculi. Growth marks cannot be observed.



The central region consists is of spongiosa and is characterized by large pore spaces and irregular trabeculae (Fig. 7C), which contain clumps of calcified cartilage (see also the Supplementary Material).

Exoccipital – The cortex consists of parallel-fibered bone and is relatively well-vascularized (Fig. 7D). The simple vascular are few in number (Fig. 7E, F) and located only in the outermost part of the cortex. The secondary osteons are more frequent (Fig. 7D). Well-mineralized, densely packed bundles of Sharpey's fibers are common and can be seen throughout the entire cortex (Fig. 7D, E). In the exoccipital, the Sharpey's fibers are most abundant and pronounced among all examined bones. Rounded osteocyte lacunae are numerous. Growth marks are absent.

The central region consists of an irregular network of bony trabeculae, with large pore spaces between them (Fig. 7G). In the medial parts of the bone tissue, where trabeculae are poorly developed, accumulations of calcified cartilage are quite common (Fig. 7H).

263 DISCUSSION

The histological variability – Witzmann (2009) investigated fragments of dermal bones from 20 taxa and concluded that for every taxon, the bone microanatomy and histology were consistent. Intraspecific variability of the histology of dermal bones was only observed in *Mastodonsaurus giganteus* and *Plagiosternum granulosum* and only affects the degree of vascularization and remodeling of the bone (Witzmann, 2009).

Nevertheless, in the *Metoposaurus krasiejowensis* skull, the observed variability is very high and despite bones diploë structure typical for all dermal bones, can be seen in both,



microanatomical and histological levels. The bones show various thicknesses, different
proportions between the layers, variations in the vascularization systems, tissue organizations,
the presence and organization of Sharpey's fibers, degree of remodeling, and growth pattern (see
also the Supplementary Material for the detailed description). The combination of these
characters concludes that each sectioning-plane in the skull represents a unique framework. The
histological properties are not related to the bone, but strictly depend on the particular
sectioning-plane. The transition between the "histological types" is fluent. The jugal and
postorbital, sectioned in the suture region, represent the same microanatomical and histological
framework (Figs. 2 j and po, 3F; Table 1), whereas the squamosum sectioned in the frontal part
of the bone (Fig. 2, sq1) and next to the otic notch (Fig. 2, sq2), showed different architecture on
both microanatomical and histological levels (Figs. 3K, L). This suggests that the histological
framework is not specifically bone-limited, but seems to be related to the specific area of the
skull and i.e. depended on the growth of the entire skull and each bone separately, or to the local
biomechanical loading on the particular part of the skull. Fortuny et al. (2012), based on the
Finite Elements Analysis, showed that the hypothetical biomechanical stress along the skull is
different for each skull-morphotype and depend directly on the shape of the skull and position of
the orbits. If the bone framework is related to the biomechanical loading, the high variability of
temnospondyls skulls results that each taxon might has a unique histological architecture of
homologous bones.
Skeletochronological information – In the external cortex of the <i>Metoposaurus</i> skull,
four types of growth alternation can be observed: numerous resting lines, the sequence of
lamellar layers combined with the high-fibrous tissue (ISF), thick zones and annuli and the
alternation of ridges and valleys (Fig. 5D-G).



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In the lacrimal, frontal, jugal, postfrontal, postparietal, tabular and quadratojugal, the distinct growth marks are manifested as a sequence of numerous thin lines (Fig. 5D). Similar growth marks, present in the external and internal cortices of the dermal bone, have been observed in several temnospondyl taxa (Scheyer, 2007; Witzmann, 2009). In fast-growing amniotes, the presence of several growth lines (Line of Arrested Growth -LAG) next to the surface of bone is known as the External Fundamental System (EFS), and indicates a slowing down of growth, suggesting that the maximum size has been reached (Sander, 2000; Chinsamy-Turan, 2005; Erickson, 2005; Turvey et al., 2005; Sander et al., 2011). In Metoposaurus long bones, similar structures are present in the inner and outer part of the cortex, suggesting that accumulation of external resting lines does not mean the cessation of growth at all, but only the oscillation in growth rate during one season (Konietzko-Meier & Klein, 2013; Konietzko-Meier & Sander, 2013). Then, the complex with accumulation of resting lines is interpreted as the one annulus deposited during one, dry season (Konietzko-Meier & Sander, 2013; Konietzko-Meier & Klein, 2013). As in the long bones, the two highly vascularized parts of the external cortex represent the fast growing zones, and the avascular layers annuli, and the entire external cortex two growth seasons. The sequences of lamellar bone and layers with accumulation of ISF visible in the postprietal (Fig. 5E) can be interpreted as three years of growth. Specific growth marks also represent the alternation of valleys and ridges preserved in the postfronal, squamosum, supratemporal, trabeculare, jugal, and quadratojugal. In these bones, two generations of valleys/ridges are visible (Fig. 5G). However, for all types of growth marks, it is not known how many growth indicators have already been remodeled. In the internal cortex, two to four resting lines are visible; the only exception is the parasphenoid (Fig. 6D). The structure of the internal cortex in the parasphenoid (Fig. 6G) mostly resembles the growth sequence seen in the



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Metoposaurus long bones (Konietzko-Meier & Klein, 2013; Konietzko-Meier & Sander, 2013). The two thick avascular layers represent the annuli, and combined with the two high vascularized zones indicate two growth seasons.

Due to the different number of preserved cycles in one skull the estimation of individual age based on the skeletochronological information is very difficult. Without testing the whole growth series, it is not possible to estimate the amount of remodeled tissue and the growth pattern of each bone and thus, no direct conclusion about the individual age of sectioned skull can be provided.

Cranial sutures were not visible on the skull surface (Gruntmejer, 2012). The disappearance of all traces of sutures on the skull surface during ontogeny is a phenomena often encountered in adult individuals (Moazen et al., 2008). In the completely preserved skeletons of Dutuitosaurus ouazzoui (Dutuit, 1976), a skull of similar length (about 400 mm) as the here described, corresponds with about 142 mm long femur (Dutuit, 1976: pl XXXI; personal observation DKM). Steyer et al. (2004) calculated the individual age of the adult *Dutuitosaurus* femur, comparable in length, for eight to nine years. For the *Metoposaurus* from Poland, large femora are not known; the largest one which was studied histologically is about 90 mm long and is estimated at four years of age (Konietzko-Meier & Klein, 2013). However, comparing skeletochronological data of *Metoposaurus* with that of *Dutuitosaurus* revealed that the femora of overlapping sizes show a similar age in both taxa, whereas the larger femora of *Dutuitosaurus* are older compared to the smaller *Metoposaurus* femora (Konietzko-Meier & Klein, 2013). Thus, on the basis of skeletochronology, a strong developmental plasticity can be excluded for both taxa. The individual age of the Krasiejów skull, based on the comparison with Dutuitosaurus, can be estimated at about eight to ten years. With the preservation of maximum



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three growth layers in the external cortex and two in the internal one, the amount of remodeled growth marks could reach up to six to seven. This indicates a relatively fast remodeling rate of the dermal bones of the skull compared to the long bones.

Remodelling degree – Among the sections from the *Metoposaurus* skull, four main remodeling degrees could be observed in the middle region. The relatively lowest remodeled samples are from the postfrontal, parietal and supratemporal. A few large erosion cavities are present in the premaxilla, nasal, lacrimal, jugal, prefrontal, postparietal, squamosum, quadratojugal and all studied bones from the palatal side of the skull. The maxilla is dominated by numerous, but medium sized erosion cavities. In the middle region of the tabular and squamosum 2, the bone deposition exceeds the bone resorption and it does not represent the typical spongious structure. The increase in remodeling degree is known as a one of the developmental characters. Witzmann (2009) published the detailed histology of dermal bones from a young adult and adult Mastodonsaurus, and observed an increase in remodeling (expressed as an increase of the erosion cavities sizes) in the older specimen. In the *Metoposaurus* skull, different histological stages can be observed among different bones in the skull. The low (postfrontal, parietal and supratemporal) and highly (premaxilla, nasal, lacrimal, jugal, prefrontal, postparietal, squamosum, quadratojugal) remodeled bones seem to represent two stages of the same process, resulting in the increase in porosity of the middle region. This may indicate the sequence of the skull ossification during ontogeny, with the latest ossification of bones occurring on the central part of the skull roof. However, less remodeled samples originate from the groves-ridges regions, whereas the other sections come from the reticulate areas. This confirms the hypothesis presented first by Bystrow (1935) that the polygonal reticulate structures are the center of



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ossification and ridges-grooves areas show the direction and extent of growth from theses ossification centers. In this case, different remodeling degrees, which resemble the ontogenetic change, is the result of longitudinal growth of the bone.

Dynamic of the sculpture pattern – Although dermal sculpture was early recognized as a characteristic for basal tetrapods (e.g. von Meyer, 1858; Fraas, 1889; Fritsch, 1889; Zittel, 1911), the morphogenesis of the sculptures is still questionable (summarized by Witzmann et al., 2010, de Buffrénil et al., 2016). Among extant tetrapods, growth of dermal bony tubercles and ridges has been studied in osteoderms of squamates and in dermal skull bones and osteoderms of crocodiles. In squamates, the presence of pits and ridges on the external surface of osteoderms follows from both local resorption and growth of bone (Zylberberg and Castanet, 1985; Levrat-Calviac & Zylberberg, 1986), whereas in crocodile dermal bones, de Buffrénil (1982) stated that sculpture is mainly the result of local resorption. In contrast, Vickaryous and Hall (2008) found no evidence for morphogenesis of bone sculpture by resorption in *Alligator mississippiensis* and presumed that sculptural ridges develop by preferential bone growth. Concerning basal tetrapods, Bystrow (1935, 1947) showed that the development of bone sculpture in the temnospondyls Benthosuchus, Platyoposaurus and Dvinosaurus took place solely by growth of the bony ridges and tubercles, and resorptive processes were not involved. The thin sections of the dermal bones of skull and pectoral girdle in the basal tetrapods investigated by Witzmann (2009), corroborate Bystrow's findings and show that the dermal sculpture did not develop by local resorption of the bone surface, comparable to the pattern in basal tetrapod osteoderms (Witzmann & Soler-Gijón 2008). According to the last study (de Buffrénil et al., 2016) the involvement of several complex remodeling processes, with the local succession of resorption and reconstruction cycles, is frequent and occurs in all major gnathostome clades. All temnospondyl sections share an



important common feature: the lack of superficial remodeling (resorption and reconstruction cycles). The supratemporal of *Metoposaurus* however show the clearly remodeling process involved into the sculpture creation (Fig. 5H). Because of it is the only one sample it is difficult to generalize the observation, but it proofs that the osteogenic mechanisms involved in the creation and growth are still not clear.

The latest study of de Buffrénil et al. (2016) confirmed, however, that the distinct dynamic processes are involved in the creation and growth of pits and ridges. Authors (de Buffrénil et al., 2016) observed six main patterns of modification of ornamentation image during growth. The simplest one is repetition of the width or position of pits and ridges from one growth stage to the following one. The ridges can drift diverge symmetrically in two opposite directions or the ridges around a given pit may migrate in the same direction. Also the change of size of the ridges is possible providing to the gradual narrowing of pit diameter (convergent ridge drift) or opposite process may occur when the reduction of ridge width is observed. In most drastic case the pits can be entirely filled, and disappear to be replaced in situ by ridges.

The sections from the *Metoposaurus* skull show that the new bone deposition mostly repeats the pattern of sculptures present in the younger stages (Fig. 5D-F); but an ridges drift and valley/ridges alternation my also occur (Fig. 5G). In the last case in such places, where valleys occurred, ridges are being deposited during the next bone generation stage. However, the distance between newly created tops of the ridges is not distinctively different than of the previous generation.

It indicates that the metric pattern of the sculpture is relatively stable, but the position of the ridges and grooves is dynamic during growth as a specific adaptation to different biomechanical loading on the new, larger bone.



Sharpey's fibers – The well-mineralized Sharpey's fibers are present in the cortex of all bones of the *Metoposaurus krasiejowensis* skull. In dermal bones, they are most likely the remains of soft tissue and small muscle attachments (Francillion-Vieillot et al., 1990). In the occipital condyles, Sharpey's fibers are densely packed in bundles and they are much thicker and longer (Fig. 7D) than in the other bones. The Sharpey's fibers occur here in similar amounts like in vertebrae (Konietzko-Meier et al., 2013). Large concentrations of Sharpey's fibers indicate that the exoccipital creates an attachment point for well developed, strong muscles and ligaments which connects the skull to the vertebral column.

Quadrate bone and exoccipital – These bones clearly differ from all other bones of the skull. Their internal structure is very similar to vertebral intercentra. In the case of *Metoposaurus krasiejowensis*, the histology and ossification process of the vertebrae have been detailed described (Konietzko-Meier et al., 2013). The quadrate and exoccipital consists of a trabecular middle region, surrounded by a thin layer of well-vascularized cortex. The presence of cartilage (Fig. 7H) indicates that ossification follows a pattern known for vertebrae with relatively slow ossification of the trabecular part and late development of the periosteal cortex (Konietzko-Meier et al., 2013).

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437	Contributed reagents/materials/analysis tools: DKM KG AB. Wrote the paper: DKM KG AB.
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440	REFERENCES
441	
442	1. Bodzioch A, Kowal-Linka M. 2012. Unraveling the origin of the Late Triassic multitaxic
443	bone accumulation at Krasiejow (S Poland) by diagenetic analysis. Palaeogeography,
444	Palaeoclimatology, Palaeoecology 346/347:25-36.
445	2. Brusatte SL, Nesbitt SJ, Irmis RB, Butler RJ, Benton MJ, Norell MA. 2010. The origin
446	and early radiation of dinosaurs. Earth-Science Reviews 101:68-100.
447	3. Brusatte SL, Butler MJ, Mateus O, Steyer JS. 2015. A new species of Metoposaurus from
448	the Late Triassic of Portugal and comments on the systematics and biogeography of
449	Metoposaurid Temnospondyls. Journal of Vertebrate Paleontology, 35(3): e912988,
450	DOI: 10.1080/02724634.2014.912988.
451	4. Buffrénil de V. 1982. Morphogenesis of bone ornamentation in extant and extinct
452	crocodilians. Zoomorphology 99(2):155-166.



453	5.	Buffrénil de V, Clarac F, Canoville A, Laurin M. 2016. Comparative data on the
454		differentiation and growth of bone ornamentation in Gnathostomes (Chordata:
455		Vertebrata). Journal of Morphology (in press.).
456	6.	Bystrow AP, 1935. Morphologische Untersuchungen der deckknochen Schädels der
457		Stegocephalen. Acta Zoologica 16:65-141.
458	7.	Bystrow AP. 1947. Hydrophilous and zenophilous labyrinthodonts. Acta Zoologica
459		28:137-164.
460	8.	Castanet J, Francillon-Vieillot H, de Ricqlès A. 2003. The skeletal histology of the
461		Amphibia. In H. Heatwole and M. Davies (eds.), Amphibian Biology, Volume 5,
462		Osteology. Surrey Beatty and Sons, Chipping Norton, New South Wales, Australia,
463		1598-1683.
464	9.	Chinsamy-Turan A. 2005. The Microstructure of Dinosaur Bone: Deciphering Biology
465		with Fine-scale Techniques. Johns Hopkins University Press, Baltimore. 1-195.
466	10.	Coldiron RW. 1974. Possible functions of ornament in labyrinthodont amphibians.
467		Occasional Papers of the Museum of Natural History of the University of Kansas,
468		Lawrence, 33:1-19.
469	11.	Dutuit JM. 1976. Introduction à l'etude paleontologique du Trias continental marocain.
470		Description des premiers Stegocephales recueillis dans le couloir d'Argana (Atlas
471		occidental). Memoires du Museum National d'Histoire Naturelle Paris Ser. C 36:1-253.
472	12.	Dzik J, Sulej T, Kaim A, Niedźwiedzki R. 2000. Późnotriasowe cmentarzysko
473		kręgowców lądowych w Krasiejowie na Śląsku Opolskim. Przegląd Geologiczny 48:226
474		35.



- 13. Dzik J. 2001. A new *Paleorhinus* fauna in the early Late Triassic of Poland. *Journal of*
- 476 *Vertebrate Paleontology* 21:625-627.
- 14. Dzik J. 2003. A beaked herbivorous archosaur with dinosaur affinities from the early
- Late Triassic of Poland. *Journal of Vertebrate Paleontology* 23:556-574.
- 15. Dzik J, Sulej T. 2007. A review of the early Late Triassic Krasiejów biota from Silesia,
- 480 Poland. *Palaeontologia Polonica* 64:3-27.
- 481 16. Dzik J. 2008. Gill structure and relationships of the Triassic cycloid crustaceans. *Journal*
- *of Morphology* 269:1501-1519.
- 483 17. Enlow DH, Brown SO. 1956. A comparative histological study of fossil and recent bone
- 484 tissues. I. Texas Journal of Science 8:405-43.
- 18. Erickson GM. 2005. Assessing dinosaur growth patterns: a microscopic revolution.
- 486 Trends in Ecology and Evolution 20(12):677-84.
- 19. Fortuny J, Marcé-Nogué J, de Esteban-Trivigno S, Gil L, Galobart Á. 2011.
- 488 Temnospondyli bite club: ecomorphological patterns of the most diverse group of early
- tetrapods. *Journal of Evolutionary Biology* 24:2040-2054.
- 490 20. Fortuny J, Marce'-Nogue' J, Gil L, Galobart A'. 2012. Skull mechanics and the
- evolutionary patterns of the otic notch closure in Capitosaurs (Amphibia:
- Temnospondyli). *The Anatomical Record* 295:1134-1146.
- 21. Fraas E. 1889. Die Labyrinthodonten der Schwäbischen Trias. *Palaeontographica* 36:1-
- 494 158.
- 495 22. Francillon-Vieillot H, de Buffrénil V, Castanet J, Géraudie J, Meunier F-J, Sire J-Y,
- 296 Zylberberg L, de Ricglès A. 1990. Microstructure and mineralization of vertebrate



497	skeletal tissues. In: Carter JG [ed.]: Skeletal Biomineralization: Patterns, Processes and
498	Evolutionary Trends, Vol. I. Van Nostrand Reinhold, New York. 471-530.
499	23. Fritsch A. 1889. Fauna der Gaskohle und der Kalksteine der Permformation Böhmens,
500	Vol. 2. Prag: Selbstverlag. 1-132.
501	24. Gądek K. 2012. Palaeohistology of ribs and clavicle of <i>Metoposaurus diagnosticus</i> from
502	Krasiejów (Upper Silesia, Poland). Opole Scientific Society Nature Journal 45:39-42.
503	25. Gross W. 1934. Die Typen des mikroskopischen Knochenbaues bei fossilen
504	Stegocephalen und Reptilien. Zeitschrift für Anatomie und Entwicklungsgeschichte
505	203:731-64.
506	26. Gruntmejer K. 2012. Morphology and function of cranial sutures of the Triassic
507	amphibian Metoposaurus diagnosticus (Temnospondyli) from southwest Poland. In: Jagt-
508	Yazykova E, Jagt J, Bodzioch A, & Konietzko-Meier D. [eds] Krasiejów –
509	palaeontological inspirations. ZPW Plik, Bytom. 34-54.
510	27. Gruszka B, Zieliński T. 2008. Evidence for a very low-energy fluvial system: a case
511	study from the dinosaur-bearing Upper Triassic rocks of Southern Poland. Geological
512	Quarterly 52:239-252.
513	28. Konietzko-Meier D, Sander PM. 2013 Longo bone histology of Metoposaurus
<mark>514</mark>	diagnosticus (Temnospondyli) from the Late Triassic of Krasiejów (Poland) and its
515	paleobiological implications. <i>Journal of Vertebrate Paleontology</i> 35(5):1-16.
516	29. Konietzko-Meier D, Klein N. 2013. Unique growth pattern of Metoposaurus diagnosticus
517	krasiejowensis (Amphibia, Temnospondyli) from the Upper Triassic of Krasiejów,
518	Poland. Palaeogeography Palaeoclimatology Palaeoecology 370:145-157.



519	30. Konietzko-Meier D, Bodzioch A, Sander PM, 2013. Histological characteristics of the
520	vertebral intercentra of Metoposaurus diagnosticus (Temnospondyli) from the Upper
521	Triassic of Krasiejów (Upper Silesia, Poland). Earth and Environmental Science
522	Transactions of the Royal Society of Edinburgh 103:1-14.
523	31. Konietzko-Meier D, Danto M, Gądek K, 2014. The microstructural variability of the
524	intercentra among temnospondyl amphibians. Biological Journal of the Linnean Society
525	112:747-764.
526	32. Levrat-Calviac V, Zylberberg L. 1986. The structure of the osteoderms in the gekko:
527	Tarentola mauritanica. American Journal of Anatomy 176:437-446.
528	33. Lucas SG. 2015. Age and correlation of Late Triassic tetrapods from southern Poland.
529	Annales Societatis Geologorum Poloniae 85(4):627-635.
530	34. Lucas SG, Spielmann JA, Hunt AP. 2007. Bio-chronological significance of Late Triassic
531	tetrapods from Krasiejów, Poland. New Mexico Museum of Natural History and Science
532	Bulletin 41:248-258.
533	35. Meyer E. 1842. Labyrinthodonten—Genera. Neues Jahrbuch für Mineralogie,
534	Geographie, Geologie, Palaeontologie 1842:301-304.
535	36. Meyer H von. 1858. Reptilien aus der Steinkohlenformation in Deutschland.
536	Palaeontographica 6:59-219.
537	37. Moazen M, Curtis N, Evans SE, O'Higgins P, Fagan M.J. 2008. Rigid-body analysis of a
538	lizard skull: Modelling the skull of Uromastyx hardwickii. Journal Biomechanics
539	41:1274-1280.



540	38. Mukherjee D, Ray S, Sengupta DP. 2010. Preliminary observations of the bone
541	microstructure, growth patterns, and life habits of some Triassic Temnospondyls from
542	India. Journal of Vertebrate Paleontology 30(1):78-93.
543	39. Olempska E. 2004. Late Triassic spinicaudatan crustaceans from southwestern Poland.
544	Acta Palaeontologica Polonica 49(3):429-442.
545	40. Olempska E. 2011. Fresh-water ostracods from the Late Triassic of Poland. Joannec
546	Geologie und Paläontologie 11:154-155.
547	41. Lamm ET. 2014. Preparation and sectioning of specimens. In: Padian K, Lamm ET
548	(eds.), Bone histology of fossil tetrapods, advancing methods, analysis, and
549	interpretation. University of California Press. Barkley, Los Angeles, London. 35-54.
550	42. Racki G, Szulc J. 2014. Formacja grabowska – podstawowa jednostka litostratygraficzna
551	kajpru Górnego Śląska. Przegląd Geologiczny 63(2):103-113.
552	43. Ricqlès A. de 1981. Recherches paléohistologiques sur les os longs des tétrapodes. VI
553	Stégocéphales. Annales de Paléontologie 67:141-160.
554	44. Rienhart LF, Lucas SG, 2013. The functional morphology of dermal bone ornamentation
555	in temnospondyl amphibian. In: Tanner LH, Spielmann JA & Lucas SG [eds] The
556	Triassic System. New Mexico Museum of Natural History and Science, Bulletin 61. 524-
557	532.
558	45. Sanchez S, Germain D, de Ricqlès, Abourachid A, Goussard F, Tafforeau P, 2010. Limb-
559	bone histology of temnospondyls: implications for understanding the diversification of
560	palaeoecologies and patterns of locomotion of Permo-Triassic tetrapods. Journal of
561	Evolutionary Biology 1-15.



562	46. Sander PW. 2000. Long bone historogy of the Tendaguru Sauropous. Implications for
563	growth and biology. Paleobiology 26:466-488.
564	47. Sander PM, Klein N, Stein K, Wings O. 2011. Sauropod bone histology and its
565	implications for sauropod biology. In Klein N, K. Remes K, Gee C T & Sander PM [eds]
566	Biology of the Sauropod Dinosaurs: Understanding the Life of Giants. Life of the Past
567	(series ed. J. Farlow). Indiana University Press, Bloomington. 276-302
568	48. Scheyer TM. 2007: Skeletal histology of the dermal armor of Placodontia: the occurrence
569	of 'postcranial fibro-cartilaginous bone' and its developmental implications. Journal of
570	Anatomy 211:737-753.
571	49. Schoch RR. 1999. Comparative osteology of <i>Mastodonsaurus giganteus</i> (Jaeger, 1828)
572	from the Middle Triassic (Lettenkeuper: Longobardian) of Germany (Baden-
573	Württemberg, Bayern, Thüringen). Stuttgarter Beiträge zur Naturkunde Serie B
574	(Geologie und Paläontologie) 278:1-170.
575	50. Schoch RR, Milner AR. 2000. Stereospondyli. Handbuch der Palaeoherpetologie 3B.
576	Munich: Verlag Dr. Friedrich Pfeil 1-230.
577	51. Skawina A, Dzik J. 2011. Umbonal musculature and relationships of the Late Triassic
578	filibranch unionoid bivalves. Zoological Journal of the Linnean Society 163:863-883.
579	52. Skawina A. 2013. Population dynamics and taphonomy of the Late Triassic (Carnian)
580	freshwater bivalves from Krasiejów (Poland). Palaeogeography, Palaeoclimatology,
581	Palaeoecology 379:68-80.
582	53. Skrzycki P. 2015. New species of lungfish (Sarcopterygii, Dipnoi) from the Late Triassic
583	Krasiejów site in Poland, with remarks on the ontogeny of Triassic dipnoan tooth plates.
58/1	Journal of Vertebrate Paleontology 35(5):e964357



54. Steyer JS, Laurin M, Castanet J, de Riegle's A. 2004. First histological and 585 skeletochronological data on temnospondyl growth: palaeoecological and 586 palaeoclimatological implications. Palaeogeography, Palaeoclimatology, Palaeoecology 587 206:193-201. 588 55. Sulei T, 2002. Species discrimination of the Late Triassic temnospondyl amphibian 589 590 Metoposaurus diagnosticus. *Acta Paleontologia Polonica* 47(3):535-546. 56. Sulei T. 2005. A new rauisuchian reptile (Diapsida: Archosauria) from the Late Triassic 591 of Poland. Journal of Vertebrate Paleontology 25(1):78-86. 592 57. Sulej T, Majer D. 2005. The temnospondyl amphibian Cyclotosaurus from the Upper 593 Triassic of Poland. *Palaeontology* 48:157-170. 594 58. Sulei T. 2007: Osteology, variability, and evolution of *Metoposaurus*, a temnospondyl 595 from the Late Triassic of Poland. *Paleontologia Polonica* 64:29-139. 596 59. Sulej T. 2010. The skull of an early Late Triassic aetosaur and the evolution of the 597 stagonolepidid archosaurian reptiles. Zoological Journal of the Linnean Society 158:860-598 881. 599 60. Szulc J, Racki G, Jewuła K. 2015a. Key aspects of the stratigraphy of the Upper Silesian 600 601 middle Keuper, southern Poland. Annales Societatis Geologorum Poloniae 85(4):557-586. 602 61. Szulc J, Racki G, Jewuła K, Środoń J. 2015b. How many Upper Triassic bone-bearing 603 604 levels are there in Upper Silesia (southern Poland)? A critical overview of stratigraphy and facies. Annales Societatis Geologorum Poloniae 85(4):587-626. 605 606 62. Turvey SP, Green OR, Holdaway RN. 2005. Cortical growth marks reveal extended 607 juvenile development in New Zealand moa. *Nature* 435:940-943.





508	63. Vickaryous MK, Hall BK. 2008. Development of the dermal skeleton in <i>Alligator</i>
509	mississippiensis (Archosauria, Crocodylia) with comments on the homology of
510	osteoderms. Journal of Morphology 269:398-422.
511	64. Witzmann F, Soler-Gijón R. 2008. The bone histology of osteoderms in temnospondyl
512	amphibians and in the chroniosuchian Bystrowiella. Acta Zoologica (Stockholm) 91: 96-
513	114. DOI 10.1463-6395.1111/j2008.00385.x
514	65. Witzmann F. 2009. Comparative histology of sculptured dermal bones in basal tetrapods,
515	and the implications for the soft tissue dermis. Palaeodiversity 2:233-270.
516	66. Witzmann F, Scholz H, Müller J, Kardjilov N. 2010. Sculpture and vascularization of
617	dermal bones, and the implications for the physiology of basal tetrapods. Zoological
518	Journal of the Linnean Society 160:302-340.
519	67. Zittel KA. 1911. Grundzüge der Paläontologie (Paläozoologie). II. Abteilung. Vertebrata.
520	Verlag von R. Oldenbourg, München and Berlin. 1-589.
521	68. Zylberberg L, Castanet J. 1985. New data on the structure and the growth of the
522	osteoderms in the reptile Anguis fragilis L. (Anguidae, Squamata). Journal of
523	Morphology 186:327-342.
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633	Fig. 1. The skull of <i>Metoposaurus krasiejowensis</i> (UOPB 01029) from Late Triassic of Poland.
634	(A) Dorsal view of skull; (B) Ventral view of the skull. Scale bar equals 10 cm.
635	
636	Fig. 2. The sectioning planes of the Metoposaurus krasiejowensis skull (UOPB 01029) from
637	Late Triassic of Poland. (A) The skull roof; (B) The palatal side of the skull. The sectioning
638	planes are marked by red lines. Dark grey color indicates preserved parts of the skull; the
639	destroyed or sediment-covered regions are indicated by the light grey color. Scale bar equals 10
640	cm. Abbreviations: $pm = premaxilla$, $m = maxilla$, $n = nasal$, $l = lacrimal$, $prf = prefrontal$, $j = lacrimal$
641	jugal, po = postorbital, pf = postfrontal, f = frontal, p = parietal, st = supratemporal, sq1 =
642	$squamosum\ 1,\ sq2 = squamosum\ 2,\ pp = postparietal,\ t = tabular,\ qj = quadratojugal,\ v = vomer,$
643	ps = parasphenoid, pt = pterygoid, q = quadrate bone, ex = exoccipital.
644	
645	Fig. 3. General microanatomy of the skull bones of <i>Metoposaurus krasiejowensis</i> (UOPB 01029)
646	from Late Triassic of Poland. (A) premaxilla; (B) maxilla; (C) nasal; (D) lacrimal; (E)
647	prefrontal; (F) jugal/postorbital; (G) postfrontal; (H) frontal; (I) parietal; (J) supratemporal; (K)
648	squamosum 1; (L) squamosum 2; (M) postparietal; (N) tabular; (O) quadratojugal; (P)
649	parasphenoid; (Q) vomer; (R) pterygoid; (S) quadrate bone; (T) exoccipital. Scale bar equals 10
650	mm. Abbreviations: ap = alary process, ds = dental shelf, vp = vomeral process.
651	
652	Fig. 4. Detailed microanatomy of the skull bones of <i>Metoposaurus krasiejowensis</i> (UOPB
653	01029) from Late Triassic of Poland, based on the frontal. (A) A valley and two ridges; (B)





654	Enlargement of (A); the external and internal cortex, and trabecular middle region with
655	numerous and large erosion cavities are visible; image in cross-polarized light; (C) The same as
656	(B), but in plane-polarized light. Dashed lines mark the approximately border between the
657	external cortex/middle region/internal cortex. Scale bars equal 10 mm for (A), and 500 μm for
658	(B-C). Abbreviations: g = groove, v = valley, EC = external cortex, MR = middle region, IC =
659	internal cortex, ER = erosion cavities, LB = lamellar bone, PFB = parallel-fibred bone.
660	
661	Fig. 5. Histology of the external cortex of the skull bones of <i>Metoposaurus krasiejowensis</i>
662	(UOPB 01029) from Late Triassic of Poland. (A) Fragment of external cortex of the frontal,
663	image in plane-polarized light; (B) Same as (A), but in cross-polarized light; (C) External cortex
664	of the tabular with distinct Sharpey's fibers in the area of the sculptural ridges, image in cross-
665	polarized light; (D) The resting lines (black arrows) in the cortex of squamosum2, image in
665 666	polarized light; (D) The resting lines (black arrows) in the cortex of squamosum2, image in plane-polarized light; (E) The resting lines expressed as an alternation of lamellar layers and
666	plane-polarized light; (E) The resting lines expressed as an alternation of lamellar layers and
666 667	plane-polarized light; (E) The resting lines expressed as an alternation of lamellar layers and layers with distinct Interwoven Structural Fibers (ISF) in the external cortex of the postparietal,
666 667 668	plane-polarized light; (E) The resting lines expressed as an alternation of lamellar layers and layers with distinct Interwoven Structural Fibers (ISF) in the external cortex of the postparietal, image in cross-polarized light; (F) Zones and annuli present in the external cortex of the
666 667 668	plane-polarized light; (E) The resting lines expressed as an alternation of lamellar layers and layers with distinct Interwoven Structural Fibers (ISF) in the external cortex of the postparietal, image in cross-polarized light; (F) Zones and annuli present in the external cortex of the quadratojugal, image in cross-polarized light; (G) Alternation of valleys and ridges in the
666667668669670	plane-polarized light; (E) The resting lines expressed as an alternation of lamellar layers and layers with distinct Interwoven Structural Fibers (ISF) in the external cortex of the postparietal, image in cross-polarized light; (F) Zones and annuli present in the external cortex of the quadratojugal, image in cross-polarized light; (G) Alternation of valleys and ridges in the postfrontal, image in cross-polarized light; note that remains of lamellar bone in the deep part of
666667668669670671	plane-polarized light; (E) The resting lines expressed as an alternation of lamellar layers and layers with distinct Interwoven Structural Fibers (ISF) in the external cortex of the postparietal, image in cross-polarized light; (F) Zones and annuli present in the external cortex of the quadratojugal, image in cross-polarized light; (G) Alternation of valleys and ridges in the postfrontal, image in cross-polarized light; note that remains of lamellar bone in the deep part of cortex are present, representing the bottom of a valley from an older generation. (H) The
666667668669670671672	plane-polarized light; (E) The resting lines expressed as an alternation of lamellar layers and layers with distinct Interwoven Structural Fibers (ISF) in the external cortex of the postparietal, image in cross-polarized light; (F) Zones and annuli present in the external cortex of the quadratojugal, image in cross-polarized light; (G) Alternation of valleys and ridges in the postfrontal, image in cross-polarized light; note that remains of lamellar bone in the deep part of cortex are present, representing the bottom of a valley from an older generation. (H) The cementing lines (black arrows) visible in the superficial part of the external cortex of the
666 667 668 669 670 671 672	plane-polarized light; (E) The resting lines expressed as an alternation of lamellar layers and layers with distinct Interwoven Structural Fibers (ISF) in the external cortex of the postparietal, image in cross-polarized light; (F) Zones and annuli present in the external cortex of the quadratojugal, image in cross-polarized light; (G) Alternation of valleys and ridges in the postfrontal, image in cross-polarized light; note that remains of lamellar bone in the deep part of cortex are present, representing the bottom of a valley from an older generation. (H) The cementing lines (black arrows) visible in the superficial part of the external cortex of the supratemporal. Dashed lines mark the approximate border between the external cortex/middle





Sharpey's fibers, LB = lamellar bone, ISF = Interwoven Structural Fibers, A = annulus, Z = 676 zone, r = ridge, v = vallev. 677 678 Fig. 6. The details of the histology of the middle region and internal cortex of the skull (UOPB 679 01029) bones of *Metoposaurus krasiejowensis* from Late Triassic of Poland. (A) Large erosion 680 cavities present in the middle region of the vomer; image in plane-polarized light; (B) Poorly 681 remodeled, well vascularized middle region of the parietal; image in plane-polarized light; (C) 682 Poorly vascularized fragment of the middle region of the squamosum2; image in plane-polarized 683 light; (D) Avascular internal cortex with resting lines (black arrows) visible in the parietal; image 684 in plane-polarized light; (E) Moderately vascularized internal cortex of the premaxilla; image in 685 plane-polarized light; (F) Highly vascularized internal cortex of the jugal; image in plane-686 polarized light; (G) Alternation of thick annuli and zones visible in the internal cortex of the 687 parasphenoid; image in cross-polarized light. Dashed lines mark the approximately border 688 between the external cortex/middle region/internal cortex. Scale bars equal 100 µm for (D) and 689 (G) and 500 μm for other photographs. Abbreviations: IC = internal cortex, MR = middle region, 690 EC = external cortex, VC = vascular canals, A = annulus, Z = zone. 691 692 Fig. 7. Histological details of the quadratum (A-C) and exooccipitale (D-H) of *Metoposaurus* 693 694 krasiejowensis skull (UOPB 01029) from Late Triassic of Poland. (A) Fragment of cortex of the 695 quadratum; image in plane-polarized light; (B) The same as (A), but in cross-polarized light; (C) Trabecular bone of the quadrate bone; image in plane-polarized; (D) Fragment of cortex of the 696 exooccipitale with distinct Sharpey's fibers; image in cross-polarized light; (E) Close-up of (D), 697 698 note that the Sharpey's fibers are also visible in plane-polarized light; (F) The same as (E) but in





699	cross-polarized light; (G) Trabeculae visible in the central part of the exooccipitale, image in
700	plane-polarized light; (H) Remains of calcified cartilage preserved in the trabeculae part of
701	exoccipital, image in cross-polarized light. Scale bars equal 500 μm for (C), (D) and (G), and
702	100 μm for other photographs. Abbreviations: C = cortex, TR= trabecular region, OL = osteocyte
703	lacunae, PO = primary osteons, PFL = parallel-fibred bone, VC = vascular canals, ER = erosion
704	cavities, SO = secondary osteons, SF = Sharpey's fibers, CC = calcified cartilage.
705	



Table 1(on next page)

Microanatomy of the sampled bones of Metoposaurus krasiejowensis[i] skull (UOPB 01029).



1 Microanatomy of the sampled bones of *Metoposaurus krasiejowensis* skull (UOPB 01029).

bone	ornamentation	min-max thickness (μm)	average thickness (μm) ¹	E:M:I ²	thickness of the external cortex (µm)
premaxilla (pm)	not preserved	3000 - 5000	4081.2	1:5.4:0.6	583.0
maxilla (m)	not preserved	4000-12000	10720.0	1:11.8:0.6	800.0
nasal (n)	relatively high ridges (about 1000 μm)	1000-8000	1404.0	1:3.6:0.6	270.0
lacrimal (l)	medium high (500 μm), steep ridges and wide grooves	2250-8000	2790.0	1:7.5:0.8	300.0
prefrontal (prf)	medium high (500 μm) and steep ridges	4000-6000	5022.0	1:3.7:0.7	930.0
jugal (j)	high ridges (about 1000 μm) and wide grooves	4000-6500	5900	1:4.2:0.7	1000.0
postorbital (po)	high ridges (about 1000 μm) and wide grooves	4000-6500	5900	1:4.2:0.7	1000.0
postfrontal (pf)	low ridges (about 300 µm) and shallow grooves	3000-5000	3212.0	1:5.7:0.6	440.0
frontal (f)	high ridges (about 1000 μm) and wide grooves	4000-5000	4686.0	1:5.5:0.6	660.0
parietal (p)	high ridges (about 1000 μm) and narrow pits	1000-4000	1960.0	1:3.5:0.4	400.0
supratemporal (st)	high ridges (about 1500 μm) and wide grooves	3000-5000	5130.0	1:4.4:0.3	900.0
squamosum 1 (sq1)	very high ridges (up to 2000 μm) and wide grooves	3000-7000	4700.0	1:2.2:0.2	1100.0
squamosum 2 (sq2)	high ridges (about 1000 μm) and wide grooves	1200-3900	3050.0	1:0.3:0.5	1750.0
postparietal (pp)	steep, high ridges (about 1500 µm) and polygonal pits	7500-10500	6020.0	1:3.1:0.2	1400.0
tabular (t)	high ridges (about	10000-	9760.0	1:6.5:0.8	1000.0



	1000 μm) and wide pits	12000			
quadratojugal (qj)	high ridges (about 1000 μm) and wide	5000-7000	5100.0	1:2.9:0.6	900.0
	grooves	2000 (000	2270.0	1 2 0 0 7	(50.0
vomer (v)	no clear sculpture	3000-6000	3379.0	1:2.8:0.7	650.0
parasphenoid (ps)	no clear sculpture	2000-5000	3750.0	1:5:1.5	900.0
pterygoid (pt)	no clear sculpture	5500-8000	5750.0	1:5.4:1	550.0
quadrate bone	-	diameter 20 mm			
(q)*					
exoccipital (ex)*	-	diameter 20	mm		

- ¹The average thickness of entire bone was estimated in thin sections, expressed as an arithmetical
- 3 average from three measurements of the thickness of a bone taken on the bottom of valleys and
- 4 the top of ridges; ² For the estimation of ratio between external cortex (E), medial region (M) and
- 5 internal cortex (I): E:M:I, the thickness of external cortex was taken as one and then
- 6 proportionally the value for medial region and internal cortex were calculated;
- 7 *Non-dermal bone



Figure 1(on next page)

The skull of Metoposaurus krasiejowensis [i](UOPB 01029) from Late Triassic of Poland.

(A) **Dorsal view of skull; (B)** Ventral view of the skull. Scale bar equals 10 cm.

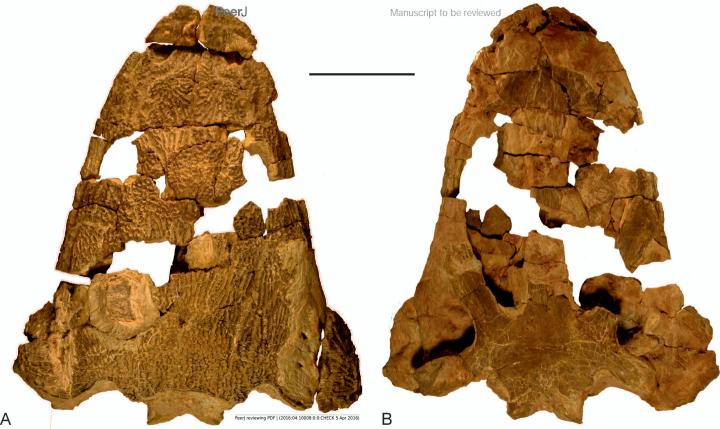




Figure 2(on next page)

The sectioning planes of the Metoposaurus krasiejowensis[i] skull (UOPB 01029) from Late Triassic of Poland.

(A) **The skull roof; (B)** The palatal side of the skull. [b]The sectioning planes are marked by red lines. Dark grey color indicates preserved parts of the skull; the destroyed or sediment-covered regions are indicated by the light grey color. Scale bar equals 10 cm. Abbreviations: pm = premaxilla, m = maxilla, n = nasal, l = lacrimal, prf = prefrontal, j = jugal, po = postorbital, pf = postfrontal, f = frontal, p = parietal, st = supratemporal, sq1 = squamosum 1, sq2 = squamosum 2, pp = postparietal, t = tabular, qj = quadratojugal, v = vomer, ps = parasphenoid, pt = pterygoid, q = quadrate bone, ex = exoccipital.

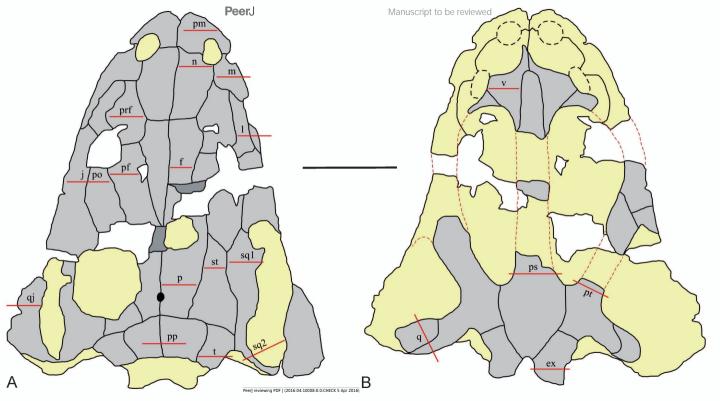




Figure 3(on next page)

General microanatomy of the skull bones of Metoposaurus krasiejowensis[i] (UOPB 01029) from Late Triassic of Poland.

- (A) premaxilla; (B) maxilla; (C) nasal; (D) lacrimal; (E) prefrontal; (F) jugal/ postorbital;
- (G) postfrontal; (H) frontal; (I) parietal; (J) supratemporal; (K) squamosum 1; (L)

squamosum 2; (M) postparietal; (N) tabular; (O) quadratojugal; (P) parasphenoid; (Q)

vomer; (R) pterygoid; (S) quadrate bone; (T) exoccipital. Scale bar equals 10 mm.

Abbreviations: ap = alary process, ds = dental shelf, vp = vomeral process.





Figure 4(on next page)

Detailed microanatomy of the skull bones of Metoposaurus krasiejowensis ([i]UOPB 01029) from Late Triassic of Poland, based on the frontal.

(A) **A valley and two ridges; (B)** Enlargement of (A); the external and internal cortex, and trabecular middle region with numerous and large erosion cavities are visible; image in cross-polarized light; (C)[b] The same as (B), but in plane-polarized light. Dashed lines mark the approximately border between the external cortex/middle region/internal cortex. Scale bars equal 10 mm for (A), and 500 μ m for (B-C). Abbreviations: g = groove, v = valley, EC = external cortex, MR = middle region, IC = internal cortex, ER = erosion cavities, LB = lamellar bone, PFB = parallel-fibred bone.

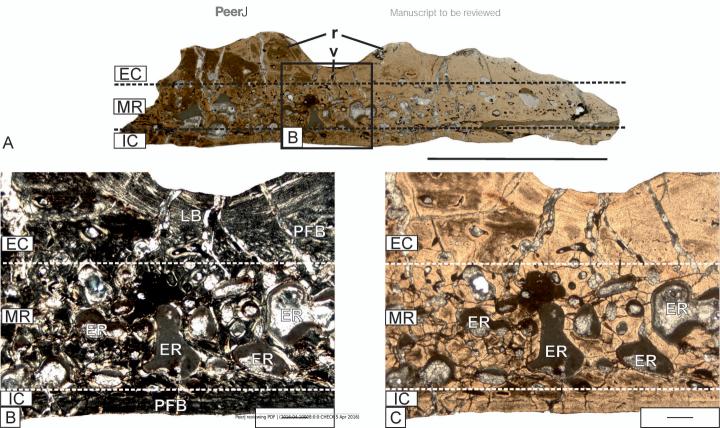




Figure 5(on next page)

Histology of the external cortex of the skull bones of Metoposaurus krasiejowensis [i](UOPB 01029) from Late Triassic of Poland.

(A) Fragment of external cortex of the frontal, image in plane-polarized light; (B) Same as (A), but in cross-polarized light; (C) External cortex of the tabular with distinct Sharpey's fibers in the area of the sculptural ridges, image in cross-polarized **light; (D)** The resting lines (black arrows) in the cortex of squamosum2, image in planepolarized light; (E) The resting lines expressed as an alternation of lamellar layers and layers with distinct Interwoven Structural Fibers (ISF) in the external cortex of the postparietal, image in cross-polarized light; (F) Zones and annuli present in the external cortex of the quadratojugal, image in cross-polarized light; (G) Alternation of valleys and ridges in the postfrontal, image in cross-polarized light; note that remains of lamellar bone in the deep part of cortex are present, representing the bottom of a valley from an older generation. (H) The cementing lines (black arrows) visible in the superficial part of the external cortex of the supratemporal. Dashed lines mark the approximate border between the external cortex/middle region/internal cortex. Scale bars equal for (A-E) and (H) 100 μ m and for (F)-(G) 500 μ m. Abbreviations: OL = osteocyte lacunae, FLB = fibro-lamellar bone, PO = primary osteons, SF = Sharpey's fibers, LB = lamellar bone, ISF = Interwoven Structural Fibers, A = annulus, Z = zone, r = ridge, v = annulusvalley.

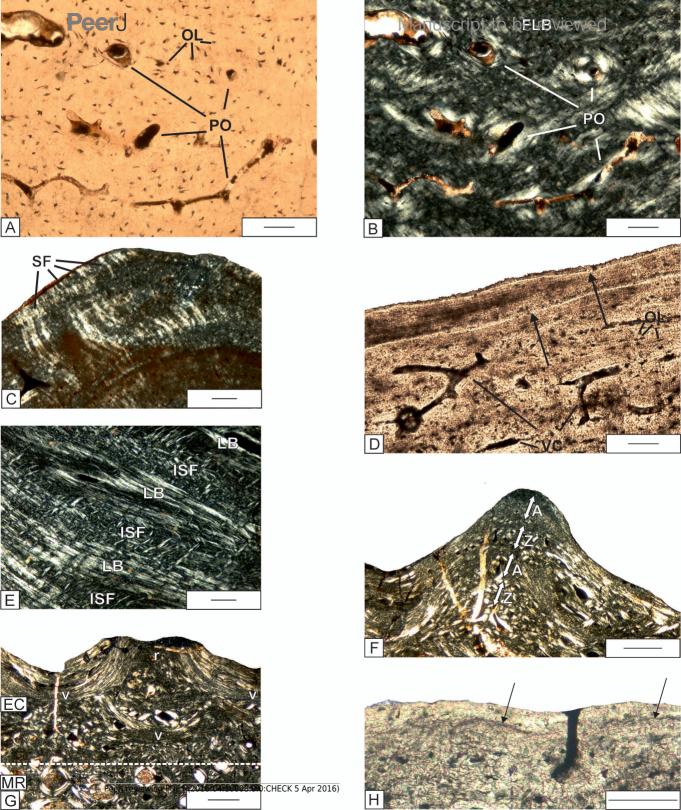




Figure 6(on next page)

The details of the histology of the middle region and internal cortex of the skull (UOPB 01029) bones of Metoposaurus krasiejowensis[i] from Late Triassic of Poland.

(A) Large erosion cavities present in the middle region of the vomer; image in plane-polarized light; (B) Poorly remodeled, well vascularized middle region of the parietal; image in plane-polarized light; (C) Poorly vascularized fragment of the middle region of the squamosum2; image in plane-polarized light; (D) Avascular internal cortex with resting lines (black arrows) visible in the parietal; image in plane-polarized light; (E) Moderately vascularized internal cortex of the premaxilla; image in plane-polarized light; (F) Highly vascularized internal cortex of the jugal; image in plane-polarized light; (G)[b] Alternation of thick annuli and zones visible in the internal cortex of the parasphenoid; image in cross-polarized light. Dashed lines mark the approximately border between the external cortex/middle region/internal cortex. Scale bars equal 100 μ m for (D) and (G) and 500 μ m for other photographs. Abbreviations: IC = internal cortex, MR = middle region, EC = external cortex, VC = vascular canals, A = annulus, Z = zone.

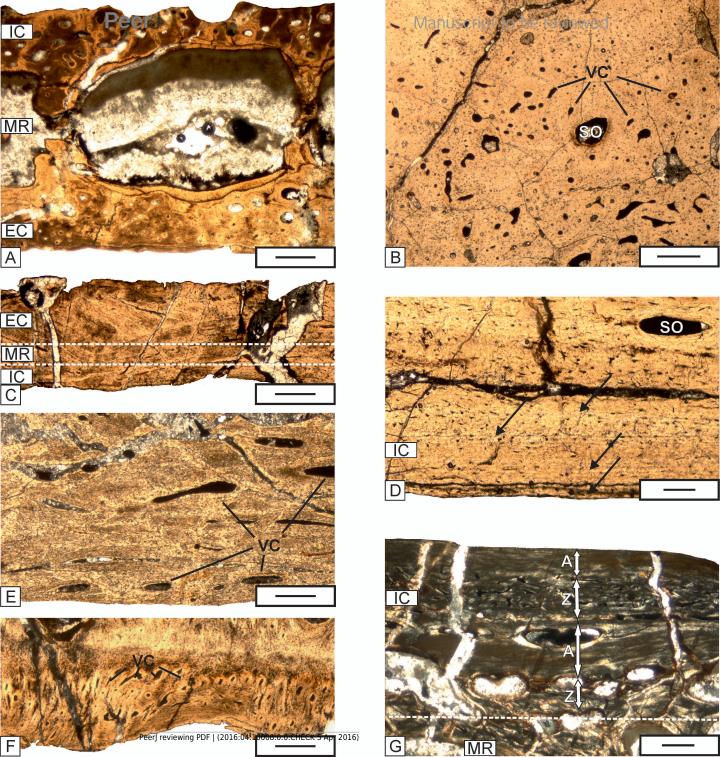




Figure 7(on next page)

Histological details of the quadratum (A-C) and exooccipitale (D-H) of Metoposaurus krasiejowensis [i]skull (UOPB 01029) from Late Triassic of Poland.

(A) Fragment of cortex of the quadratum; image in plane-polarized light; (B) The same as (A), but in cross-polarized light; (C) Trabecular bone of the quadrate bone; image in plane-polarized; (D) Fragment of cortex of the exooccipitale with distinct Sharpey's fibers; image in cross-polarized light; (E) Close-up of (D), note that the Sharpey's fibers are also visible in plane-polarized light; (F) The same as (E) but in cross-polarized light; (G) Trabeculae visible in the central part of the exooccipitale, image in plane-polarized light; (H) Remains of calcified cartilage preserved in the trabeculae part of exoccipital, image in cross-polarized light. Scale bars equal 500 μm for (C), (D) and (G), and 100 μm for other photographs. Abbreviations: C = cortex, TR= trabecular region, OL = osteocyte lacunae, PO = primary osteons, PFL = parallel-fibred bone, VC = vascular canals, ER = erosion cavities, SO = secondary osteons, SF = Sharpey's fibers, CC = calcified cartilage.

