

The impact of IL-6 and IL-28B gene polymorphisms on treatment outcome of chronic hepatitis C infection among intravenous drug users in Croatia

Zoran Bogdanović,^{1*} Ivana Marinović-Terzić,^{2*} Sendi Kuret,^{3*} Ana Jerončić,^{4*} Nikola Bradarić,⁵ Gea Forempoher,³ Ozren Polašek,⁶ Janoš Terzić,² and Šimun Anđelinović³

*These authors contributed equally

Clinical Hospital Split, Departments of: Internal Medicine, Division of Gastroenterology¹, Pathology³ and Infectious diseases⁵; and University of Split School of Medicine, Departments of: Immunology², Research in Biomedicine and Health⁴, and Public Health⁶, Split 21 000, Croatia.

Corresponding author: Zoran Bogdanović, University of Split, School of Medicine Department of Internal Medicine, Division of Gastroenterology, Spinciceva 1, Split 21000, Croatia; suzbog@gmail.com ; fax: + 38521435820, phone: + 38521453825

Short title: IL genotypes and hepatitis C outcome

26 **ABSTRACT**

27 **Background.** Several genes and their single nucleotide polymorphisms are associated with
28 either spontaneous resolution of hepatitis C infection or better treatment induced viral
29 clearance. We tested a cohort of intravenous drug users (IVDU) diagnosed with chronic
30 hepatitis C (HCV) for treatment response and its association with single nucleotide
31 polymorphisms in Interleukin-6 (rs1800795-IL6) and Interleukin-28B (rs12979860-IL28B)
32 genes.

33 **Methods.** The study included 110 Croatian IVDU positive for anti-HCV antibody.
34 Genotyping was performed by PCR based approach. Patients were treated by standard
35 pegylated-interferon/ribavirin and followed throughout a period of four years, during which
36 sustained virological response (SVR) was determined. All data were analysed with statistical
37 package SPSS 19.0 (IBM Corp, Armonk, NY) and PLINK v1.07 software.

38 **Results.** Patients showed a significantly better response to treatment according to the number
39 of copies of the C allele carried at rs1800795-IL6 ($P=0.034$). All but one of the patients with
40 CC genotype achieved SVR (93%), whereas the response rate of patients with GG genotype
41 was 64%. The association of rs1800795-IL6 with SVR status remained significant after
42 further adjustment for patients' age, fibrosis staging, and viral genotype (OR 2.15, 95%CI
43 1.16-4.68, $P=0.019$). Distributions of allele frequencies at the locus rs12979860-IL28B
44 among the study cohort and the underlying general population were suggestive of a protective
45 effect of CC genotype in acquiring chronic hepatitis C in the Croatian IVDU population.

46 **Discussion.** The rs1800795-IL6 polymorphism is associated with positive response to
47 treatment in IVDU patients positive for HCV infection. A protective role of rs12979860-
48 IL28B CC genotype in acquiring chronic hepatitis C is suggested for Croatian IVDU
49 population.

50

51 INTRODUCTION

52

53 It is estimated that chronic hepatitis C virus (HCV) infection affects nearly 170 million
54 individuals worldwide. It poses as one of the most important and growing threats to public
55 health. Being an intravenous drug user (IVDU) is one of the most important risk factors in
56 acquiring this infection. Both, acute and chronic, hepatitis C infection is a great therapeutic
57 challenge. Only a minority of patients are able to clear the virus, and so do not run the risk of
58 developing HCV induced end liver damage. Among the different and diverse list of factors
59 that influence the therapeutic response, the host's cytokines play a very important role. The
60 cytokine levels are directly influenced by certain gene polymorphisms located within their
61 coding or regulatory regions (Wilson et al. 1997).

62 Among others, IL-6 is reported to be elevated in chronic HCV infection compared to
63 healthy controls (Malaguarnera et al. 1997). The low-producing Interleukin 6 genotype IL-6
64 CC (IL-6 rs1800795 G174C) was associated with spontaneous clearance of hepatitis C virus
65 in patients infected by contaminated blood products (Barrett et al. 2001). On contrary, high
66 producing Interleukin 6 genotypes of the rs1800795 174G/C polymorphism (i.e., GG or GC
67 genotypes) were associated with a greater likelihood of SVR in patients coinfecting with HCV
68 and HIV (Nattermann et al. 2007).

69 Human Interleukin-28B gene encodes interferon lambda-3 (IFN-lambda-3). Interferon
70 lambda has demonstrated antiviral activity against HCV genotype 1 *in vivo* (Pagliaccetti &
71 Robek 2010) and *in vitro* (Muir et al. 2010). It has been shown that a single nucleotide
72 polymorphism (SNP) of IL28B gene (IL-28B rs12979860 C/T) predicts hepatitis C treatment
73 induced viral clearance (Ge et al. 2009) and is associated with spontaneous resolution of
74 hepatitis C infection (Duggal et al. 2013).

[LH1] Comentário: Include hepatitis C therapeutic regimen, since pegylated interferon-ribavirin until the new direct acting antivirals (DAAs)

[LH2] Comentário: Include reference. WHO for example.

[LH3] Comentário: Use abbreviation: HCV

[LH4] Comentário: There are more references to be included

[LH5] Comentário: There are more references to be included

Here, we studied whether interleukin-28B or interleukin-6 (IL-6) promoter ~~single-nucleotide~~ ~~polymorphism~~ SNP affects the response to the standard antiviral treatment in IVDU patients diagnosed with chronic hepatitis C. We determined IL-6 promoter and IL-28B gene polymorphisms in a cohort of 110 patients with chronic hepatitis C. All IVDU positive patients diagnosed with chronic hepatitis C were treated with a standard protocol of peg-interferon alpha-2a and ribavirin. Rates of SVR were compared between IL-6 and IL-28B wild type, heterozygous and homozygous genotypes.

SUBJECTS & METHODS

The study samples

The sample of 112 patients was recruited from the outpatient hepatology unit at the Split Medical Center, Croatia and was followed from September 2007 until November 2013. The sample was drawn from the total of 947 HCV positive patients but only 112 individuals were diagnosed, biopsied, genotyped, treated and followed for a period of four years. Two patients were excluded from further analysis due to an ambiguous SVR status, possible reuse of intravenous drugs and re-infection with a different viral strain, resulting in a cohort of 110 patients. All enrolled patients had an alcohol consumption of less than 14 units per week and other common forms of chronic liver disease were excluded in all cases. Patients were Caucasians with the median (IQR) age at diagnosis of 40 (35-45) years in the SVR group and 41.5 (39-47) in the non-responder (NR) group.

We also used the data from the 10,001 Dalmatians biobank as the source of population-based sample of the underlying, general Croatian population. All controls were apparently healthy subjects with no record of addiction, risky behaviors or detected HCV infection (Polasek 2013; Rudan et al. 2009).

100 Patients' data were collected as a part of standard clinical procedure and the informed consent
101 was obtained prior to participating in the study in all cases. The Ethics Committee of the
102 Clinical Hospital Split approved the study (No: 2181-147-06-01/01-M.J).

103

104 **Diagnosis of HCV infection**

105 A third generation enzyme immunoassay (ELISA; Abbott Diagnostics, Germany) was used to
106 test all subjects for HCV specific antibodies. Reverse-transcriptase polymerase chain reaction
107 (RT-PCR) assay (Amplicor; Roche Diagnostic Systems, New Jersey, USA) was used to test
108 for HCV RNA in all subjects, in order to determine SVR.

109 Include parameters to response definition: SVR and NR. What about relapsers (REL)?

110

111 **Liver Histology**

112 Percutaneous liver biopsy was performed at the time of initial diagnosis and at the beginning
113 of the treatment, using the Trucut biopsy technique (SteryLab, Italy) following informed
114 consent. Inflammation was graded using a histological activity index (HAI) (Knodel et al.
115 1981) and fibrosis (Ishak et al. 1995). Every fifth biopsy was independently validated by two
116 pathologists. A minority of patients was not biopsied either due to secondary coagulopathy or
117 refusal to sign the informed consent and to participate in the procedure.

118

119 **DNA extraction**

120 A salting out technique was used to extract DNA from whole blood or using the QIAmp
121 DNA midi prep kit, (Qiagen Ltd., Crawley, UK). The obtained DNA was used for IL-6
122 promoter and IL-28B genotyping. During this process, all the RNA was removed by
123 incubating the digested preparation with 1.5 ml ribonuclease A (Boehringer Mannheim UK
124 Ltd, East Sussex, UK) per 400 µl of nuclear lysate.

[LH6] Comentário: Check: isn't QIAmp?

125

126 **IL-6 and IL28B genotyping**

127 A 175 base pair (bp) fragment of the human IL-6 gene spanning the promoter IL-6 G-174C
128 region was amplified with gene specific primers (Roche Diagnostics, Alameda, California,
129 USA). The resulting PCR fragments were analyzed with hybridization probes labeled with
130 LightCycler Red 640. The genotypes were identified by running a melting curve with specific
131 melting points (T_m) (Tib Molbiol GmbH, Berlin, Germany).

132 Genetic polymorphism in a SNP located near the IL-28B gene (rs12979860) was
133 determined by enzymatic digestion of the PCR product. DNA fragments were amplified by
134 using specific primers. Primer sequences were 5'-GCCTGTCGTGTACTGAACCA-3', and
135 5'-GCTCAGGGGTCAAATCACAGAAG-3', a PCR product of 143 bp was digested with
136 HhaI enzyme and the resulting fragments of 27, 38, 65 and 78 bp were separated on
137 acrylamide gel.

[LH7] Comentário: Include gel concentration. Include staining if important.

138

139 **Statistical analysis**

140 Data were analysed with the statistical package SPSS 19.0 (IBM Corp, Armonk, NY) and
141 PLINK v1.07 software (Purcell et al. 2007). Absolute numbers and percentages were used to
142 describe categorical data, whereas median and interquartile ranges (IQR) were used to
143 describe quantitative data. One sample binomial test was used to assess distributions of sex,
144 and response to the treatment. The association of response-to-treatment with sex or the viral
145 genotype in infected patients was estimated with the Pearson's chi square test. Furthermore,
146 nonparametric Mann-Whitney test was used to assess age differences, or differences in
147 severity of fibrosis between the response groups.

148 Genetic association tests were mostly performed within PLINK software by using the
149 case and control design. We defined cases as the group of patients that achieved SVR. Full

model association tests were run in PLINK for each SNP using either chi-square or, when appropriate, Fisher exact test; and the best-fit model was identified. Full model included basic allelic, Cochran-Armitage trend, genotypic, dominant gene action and the recessive gene action tests. Additionally, the difference in distribution of IL28B CC carriers vs CT+TT carriers across the response groups was estimated by the Pearson's chi square test using the SPSS v19 software (IBM, Armonk, NY) since the recessive model in PLINK tests only the minor allele and in both our samples (patients and the underlying, healthy general population) the allele C at the IL28B locus was the dominant allele. All significant p-values yielded by genetic tests were further controlled by empirical p-values which were generated by the permutation procedure. Cochran-Mantel-Haenszel statistics was used to test whether the predictive power of the SNP markers was independent of viral genotypes detected in infected patients. Associations of the SVR status with both SNPs were further evaluated by multivariate logistic regression while accounting for covariates/factors: age, viral genotype, stage of fibrosis.

RESULTS

A total of 110 IVDU patients with elevated liver function tests were diagnosed with a chronic hepatitis C, treated by standard interferon 2-alfa/ribavirin protocol and followed over at least one year for a SVR. The majority of treated patients (78 out of 110, or 71%) achieved SVR (one sample binomial test, $P < 0.001$). Men were significantly more prevalent in our sample than women: 82 (75%) vs. 28 (25%) (one sample binomial test, $P < 0.001$). Demographic and clinical characteristics of the sample are presented in Table 1. As shown in Table 1, non-responders were significantly older ~~that~~ than responders. We found no association of the SVR status with the sex, the initial Ishak score fibrosis stage, or the viral genotype.

175 Distributions of patients' genotypes across response-to-treatment groups are shown for
176 both SNP loci, rs1800795- IL6 and rs12979860- IL28B, in Table 2.

177 With regard to genetic variation at the rs1800795-IL6 locus, we determined Hardy-
178 Weinberg equilibrium in both response groups (exact test, p-values from 0.392 to 0.819).
179 Overall, the genetic association tests indicated that the addition of allele 'C' has protective
180 effect and increases the chance of achieving SVR. Specifically, several genetic association
181 tests confirmed the association between rs1800795-IL6 polymorphism and SVR status, with
182 the Cochran-Armitage trend test providing the best model fit ($\chi^2=4.477$, $df=1$, $p=0.034$,
183 empirical $p=0.039$). Patients with rs1800795-IL6 CC genotype had significantly better
184 ~~sustained virological response~~SVR (14 out of 15, 93 %) compared to those with GC (37 out
185 of 52, 71%) or GG (27 out of 43, 63%) genotypes (Figure 1).

186 After controlling for viral genotypes, the association of rs1800795-IL6 polymorphism
187 with SVR status remained significant (Mantel-Haenszel 2x2 $\chi^2=4.483$, $p=0.034$, with odds
188 ratio of 1.99, 95% CI 1.05-3.78). The association was further evaluated by multivariate
189 logistic regression analysis while accounting for covariates: age, viral genotype, and fibrosis
190 ($F \geq 3$). The result of regression analysis confirmed that the addition of allele 'C' increased the
191 chance of achieving SVR (OR 2.45, 95% CI 1.13-5.30, $P=0.023$). Besides the rs1800795-IL6
192 ~~polymorphysmpolymorphism~~, age was the only covariate that significantly affected SVR
193 status, with each additional year slightly decreasing the chance of SVR response (significance
194 at the level of 0.1, OR 0.95, 95%CI 0.89-1.01, $P=0.096$).

195 The genotype frequencies at the rs12979860-IL28B locus met the Hardy-Weinberg
196 expectations in nonresponders (exact test, $p=0.473$), and deviated from the Hardy-Weinberg
197 equilibrium in SVR cases (i.e. SVR responders, $p=0.004$). When we analysed the genotype
198 frequencies at the rs12979860 locus in the IVDU cohort, and in the underlying, apparently
199 healthy, general-population sample ($n=531$ individuals; HWE, $p=0.999$), we observed that the

frequencies of CC, CT and TT genotypes in our cohort were: 27%, 63%, and 10%; whereas the corresponding frequencies in the population-based sample were 49%, 42%, and 9% (Figure 2). The finding demonstrated a large and significant reduction of CC genotype (z-score test for proportions, $p < 0.001$), and a significant increase in frequency of heterozygous TC genotype ($p < 0.001$) in the IVDU cohort of chronic HCV patients compared to the underlying, apparently healthy population. The frequencies of TT genotype were comparable between the groups ($p = 0.741$).

With regard to the SVR responder and non-responder groups, we did not, however, observe a significant association of SVR response with the allele-carrier groups: CC and CT+TT (chi square=0.118, df=1, $p = 0.732$); although the frequency of CC genotype was in fact somewhat higher in responders (by 3%). The result persisted after we controlled for viral genotypes (Mantel-Haenszel 2x2 $\chi^2 = 0.630$, $p = 0.427$) and after multivariate logistic regression using age, viral genotype and fibrosis stage as additional predictors (data not shown).

DISCUSSION

This study investigated the role of IL-6 and IL-28B gene polymorphisms on SVR in IVDU patients diagnosed with chronic hepatitis C infection. The treatment was conventional and included Peg IFN combined with ribavirin for either 48 weeks (genotype 1) or 24 weeks (genotype 3). We defined sustained virological response as an absence of ~~detectible~~-detectable virus at the end of follow up evaluation and or disease relapses according to the standard definitions (Ghany et al. 2009).

Interleukin-6 was originally discovered as a protein that caused the final differentiation of B cells into immunoglobulin secreting cells (Muraguchi et al. 1988).

225 Additional work showed that IL-6 and its receptor - sIL-6R α suppress neutrophil recruitment
226 at site of acute inflammation, making way for the influx of monocytes as the inflammatory
227 response is sustained (Kopf et al. 1994). IL-6 is well known pro-inflammatory cytokine with
228 pro-tumorigenic potential (Grivennikov et al. 2009) and is emerging as a key regulatory signal
229 in the development of the newly described pro-inflammatory effectors T-cell subset, so called
230 Th17 cells (Harrington et al. 2005). The IL-6 rs1800795 G allele has been also associated
231 with higher degrees of liver necroinflammation (Falleti et al. 2010) and fibrosis (Cussigh et al.
232 2011).

233 | In our study, allele C at rs1800795-IL6 - a ~~single nucleotide polymorphism~~SNP in the
234 IL-6 gene promoter, was associated with sustained virological response (OR 2.45, 95%CI
235 1.13-5.30, P=0.023). The genotype that confers the highest degree of protection in terms of
236 achieving SVR, rs1800795-IL6 CC, demonstrated an overwhelming lower relapse rate in
237 HCV treated patients (1 out of 15 patients, 7%). Similar results were reported for Italian non-
238 IVDU HCV infected patients thus corroborating the importance of our findings (Cussigh et al.
239 2011). According to prior studies, the CC genotype appears to be associated with significantly
240 lower level of plasma IL-6, whereas the GG and GC genotypes appear to have higher levels
241 of plasma IL-6 (Fishman et al. 1998; Lapinski 2001). This implies a possible connection of
242 IL-6 status with the therapy outcome. The putative low producing IL-6 phenotype may play a
243 protective role against chronic hepatitis C infection by helping to clear the viral particles
244 during standard therapy. Chronic hepatitis C patients with rs1800795-IL6 CC genotype and
245 lower IL-6 serum level may have an attenuated adoptive immune response, directed away
246 from damaging, pro-inflammatory and autoimmune to predominately suppressive and anti-
247 viral inflammatory response.

248 An IL-28B gene single nucleotide polymorphism is located 8 kb upstream of the start
249 codon of IL-28B gene that encodes interferon lambda (IFN- λ) a member of type III IFN

250 family. IFN- λ interacts with a transmembrane receptor to induce a potent antiviral response
251 (Donnelly & Kotenko 2010; Fox et al. 2009; Li & Huang 2007). The antiviral activity is
252 mediated through the activation of the either JAK-STAT (IFN α , λ and γ) or MAPK (IFN α
253 and λ) pathways (Arslani et al. 2013). There is a strong association of genetic variations in IL-
254 28B gene with response to therapy (Ge et al. 2009; Suppiah et al. 2009; Tanaka et al. 2009),
255 and with spontaneous HCV clearance (Duggal et al. 2013). In our cohort of IVDU patients
256 diagnosed with chronic hepatitis C no association between the therapy outcome and the single
257 nucleotide polymorphism in the IL-28B gene, rs12979860-IL28B was identified. However,
258 our data do support involvement of the rs12979860-IL28B CC genotype in both of these
259 patho-physiological/immunological processes: spontaneous HCV clearance and the response
260 to therapy.

261 Firstly, when we compared our cohort of IVDU patients suffering from chronic HCV
262 with the population-based sample taken from the apparently healthy, underlying population;
263 the frequency of the favorable rs12979860-IL28B CC genotype which has been also
264 associated with the spontaneous clearance of HCV, was largely decreased in the patient
265 group. Conversely, the frequencies of genotypes considered to have a neutral effect on
266 acquiring HCV infection/response to therapy were either increased in patients (TC) or showed
267 no difference between the samples (TT). In other words, the protective genotype rs12979860-
268 IL28B CC was found at much lower frequency in infected IVDU individuals with chronic
269 HCV (27%) than in the underlying, healthy population (49%), thus pointing towards the
270 selective loss of CC homozygotes in the patient population.

271 In line with our finding, Gelinas et al. have observed significantly higher prevalence of
272 the responder genotype rs12979860 CC in a group of IVDU who were spontaneous resolvers
273 from a HCV infection than in a baseline population of IVDU users (Gelinas et al. 2013);
274 suggesting a dilution of CC genotype in chronic IVDU HCV patients. Additionally, also

275 | supporting our findings, Ezzikouri et al. found that patients who had cleared HCV
276 | spontaneously were from 2.7 to 4.7 times more likely to carry CC genotype than the TC, or
277 | the TT genotype, respectively (Ezzikouri et al. 2013) while Montes-Cano et al. observed the
278 | CC genotype in 73% of individuals with spontaneous resolution of HCV infection versus only
279 | 46% in individuals with the persistent infection (Montes-Cano et al. 2010).

280 | Secondly, our results still suggest the positive effect of rs12979860-IL28B CC
281 | genotype in acquiring SVR. In particular, the distribution of rs12979860-IL28B genotypes in
282 | the cohort of IVDU chronic patients significantly deviated from HWE-P only in the group of
283 | responders, whereas in non-responders, and in healthy controls the rs12979860-IL28B
284 | genotypes followed the HWE. Since the case (SVR responder) genotypes will only be in
285 | HWE under the multiplicative genetic model (Lewis & Knight 2012), the departure from this
286 | equilibrium, if found exclusively in cases, can be expected in relatively small samples of
287 | patients over a range of genetic models and is indicative of the actual association to the trait
288 | under study (Wittke-Thompson et al. 2005). In addition, we have observed somewhat higher
289 | percentage of CC genotype in SVR cases, although this percentage did not reach a statistical
290 | significance.

291 | Similar to our results, Searberg et al. also did not found the association between
292 | rs12979860-IL28B genotypes and the spontaneous clearance of HCV in men who have sex
293 | with men (Seaberg et al. 2015). There ~~is~~are obviously a large number of factors:
294 | demographic, viral, and human genetic factors; which influence HCV viremia and the results
295 | on distribution of particular SNPs should be interpreted in a larger context. The fact that the
296 | frequency of rs12979860 CC genotype varies in different ethnic groups or geographical areas
297 | adds to this complexity. In particular, several studies have estimated that East Asians had a
298 | high percentage of CC genotype, whereas the frequency of this genotype was intermediate in
299 | Europeans and minor frequency in African cohorts (Ge et al. 2009; Thomas et al. 2009).

Concerning the geographic variability of rs12979860 CC genotypes, previous studies on Caucasian patients who were infected with the HCV viral genotype 1 estimated the prevalence of CC genotypes to be between 35% and 39% (Cavalcante et al. 2012; Montes-Cano et al. 2010; Nattermann et al. 2011). The percentage of CC genotypes observed in our patients infected with the same viral genotype was 27% (95% CI 19-36%) which is somewhat lower, suggesting that the impact of IL-28B polymorphism on acquiring of infection or spontaneous clearance of HCV might be more prominent in the Croatian population. Having in mind that the population differences in rates of spontaneous clearance have also been proposed for IVDU patients (Fischer et al. 2004; Miedzinski & Taylor 2008), the impact of IL-28B polymorphism on the spontaneous clearance of HCV in Croatian population should be investigated in more details in order to increase our knowledge on the therapeutic effectiveness of pegylated-interferon/ribavirin on rs12979860 genotypes in different populations, particularly in high-HCV-risk IVDU population.

313

314 **CONCLUSIONS**

In conclusion, we have identified that the rs1800795-IL6 CC genotype is associated with significantly better sustained virological response to the standard Peg IFN and ribavirin treatment in IVDU/HCV patients. Also, findings point towards strong protective role of rs12979860-IL28B CC genotype in acquiring chronic hepatitis C infection in Croatian IVDU population. Finally, among all covariates, age is the most important, where every additional year slightly decreases the chance of SVR response. Further prospective and large scale clinical studies are necessary to confirm our results before we can prospectively identify IVDU and HCV patients for whom therapy is likely to be successful.

323

324 **ACKNOWLEDGEMENT:**

325 The project was funded by Croatian Ministry of Science, grant number 258-2160800-0333 to
 326 ŠA and by Croatian Ministry of Science, Sport and Education grant number 216-000000-3348
 327 to JT.

329 REFERENCES:

- 330
- 331 Arslani N, Gajzer B, Papes D, Rajkovic Z, Altarac S, Zore Z, and Filipovic-Zore I. 2013. A
 332 new approach for transversalis fascia reinforcement in Lichtenstein's inguinal hernia
 333 repair. *Surg Today* 43:211-214. 10.1007/s00595-012-0232-7
- 334 Barrett S, Goh J, Coughlan B, Ryan E, Stewart S, Cockram A, O'Keane JC, and Crowe J.
 335 2001. The natural course of hepatitis C virus infection after 22 years in a unique
 336 homogenous cohort: spontaneous viral clearance and chronic HCV infection. *Gut*
 337 49:423-430.
- 338 Cavalcante LN, Abe-Sandes K, Angelo AL, Machado TM, Lemaire DC, Mendes CM, Pinho
 339 JR, Malta F, Lyra LG, and Lyra AC. 2012. IL28B polymorphisms are markers of
 340 therapy response and are influenced by genetic ancestry in chronic hepatitis C patients
 341 from an admixed population. *Liver Int* 32:476-486. 10.1111/j.1478-
 342 3231.2011.02653.x
- 343 Cussigh A, Falletti E, Fabris C, Bitetto D, Cmet S, Fontanini E, Bignulin S, Fornasiere E,
 344 Fumolo E, Minisini R, Pirisi M, and Toniutto P. 2011. Interleukin 6 promoter
 345 polymorphisms influence the outcome of chronic hepatitis C. *Immunogenetics* 63:33-
 346 41. 10.1007/s00251-010-0491-7
- 347 Donnelly RP, and Kotenko SV. 2010. Interferon-lambda: a new addition to an old family. *J*
 348 *Interferon Cytokine Res* 30:555-564. 10.1089/jir.2010.0078
- 349 Duggal P, Thio CL, Wojcik GL, Goedert JJ, Mangia A, Latanich R, Kim AY, Lauer GM,
 350 Chung RT, Peters MG, Kirk GD, Mehta SH, Cox AL, Khakoo SI, Alric L, Cramp
 351 ME, Donfield SM, Edlin BR, Tobler LH, Busch MP, Alexander G, Rosen HR, Gao X,
 352 Abdel-Hamid M, Apps R, Carrington M, and Thomas DL. 2013. Genome-wide
 353 association study of spontaneous resolution of hepatitis C virus infection: data from
 354 multiple cohorts. *Ann Intern Med* 158:235-245. 10.7326/0003-4819-158-4-
 355 201302190-00003
- 356 Ezzikouri S, Alaoui R, Rebbani K, Brahim I, Fakhir FZ, Nadir S, Diepolder H, Khakoo SI,
 357 Thursz M, and Benjelloun S. 2013. Genetic variation in the interleukin-28B gene is
 358 associated with spontaneous clearance and progression of hepatitis C virus in
 359 Moroccan patients. *PLoS One* 8:e54793. 10.1371/journal.pone.0054793
- 360 Falletti E, Fabris C, Vandelli C, Colletta C, Cussigh A, Smirne C, Fontanini E, Cmet S,
 361 Minisini R, Bitetto D, Toniutto P, and Pirisi M. 2010. Genetic polymorphisms of
 362 interleukin-6 modulate fibrosis progression in mild chronic hepatitis C. *Hum Immunol*
 363 71:999-1004. 10.1016/j.humimm.2010.06.006
- 364 Fischer B, Haydon E, Rehm J, Krajden M, and Reimer J. 2004. Injection drug use and the
 365 hepatitis C virus: considerations for a targeted treatment approach--the case study of
 366 Canada. *J Urban Health* 81:428-447. 10.1093/jurban/jth128
- 367 Fishman D, Faulds G, Jeffery R, Mohamed-Ali V, Yudkin JS, Humphries S, and Woo P.
 368 1998. The effect of novel polymorphisms in the interleukin-6 (IL-6) gene on IL-6

transcription and plasma IL-6 levels, and an association with systemic-onset juvenile chronic arthritis. *J Clin Invest* 102:1369-1376. 10.1172/JCI2629

Fox BA, Sheppard PO, and O'Hara PJ. 2009. The role of genomic data in the discovery, annotation and evolutionary interpretation of the interferon-lambda family. *PLoS One* 4:e4933. 10.1371/journal.pone.0004933

Ge D, Fellay J, Thompson AJ, Simon JS, Shianna KV, Urban TJ, Heinzen EL, Qiu P, Bertelsen AH, Muir AJ, Sulkowski M, McHutchison JG, and Goldstein DB. 2009. Genetic variation in IL28B predicts hepatitis C treatment-induced viral clearance. *Nature* 461:399-401. 10.1038/nature08309

Gelinas JF, Fabre T, Willems P, Leung RC, George J, Willems B, Bruneau J, and Shoukry NH. 2013. IL28B SNP screening and distribution in the French Canadian population using a rapid PCR-based test. *Immunogenetics* 65:397-403. 10.1007/s00251-013-0688-7

Ghany MG, Strader DB, Thomas DL, and Seeff LB. 2009. Diagnosis, management, and treatment of hepatitis C: an update. *Hepatology* 49:1335-1374. 10.1002/hep.22759

Grivennikov S, Karin E, Terzic J, Mucida D, Yu GY, Vallabhapurapu S, Scheller J, Rose-John S, Cheroutre H, Eckmann L, and Karin M. 2009. IL-6 and Stat3 are required for survival of intestinal epithelial cells and development of colitis-associated cancer. *Cancer Cell* 15:103-113. 10.1016/j.ccr.2009.01.001

Harrington LE, Hatton RD, Mangan PR, Turner H, Murphy TL, Murphy KM, and Weaver CT. 2005. Interleukin 17-producing CD4⁺ effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages. *Nat Immunol* 6:1123-1132. 10.1038/ni1254

Ishak K, Baptista A, Bianchi L, Callea F, De Groote J, Gudat F, Denk H, Desmet V, Korb G, MacSween RN, and et al. 1995. Histological grading and staging of chronic hepatitis. *J Hepatol* 22:696-699.

Knodell RG, Ishak KG, Black WC, Chen TS, Craig R, Kaplowitz N, Kiernan TW, and Wollman J. 1981. Formulation and application of a numerical scoring system for assessing histological activity in asymptomatic chronic active hepatitis. *Hepatology* 1:431-435.

Kopf M, Baumann H, Freer G, Freudenberg M, Lamers M, Kishimoto T, Zinkernagel R, Bluethmann H, and Kohler G. 1994. Impaired immune and acute-phase responses in interleukin-6-deficient mice. *Nature* 368:339-342. 10.1038/368339a0

Lapinski TW. 2001. The levels of IL-1beta, IL-4 and IL-6 in the serum and the liver tissue of chronic HCV-infected patients. *Arch Immunol Ther Exp (Warsz)* 49:311-316.

Lewis CM, and Knight J. 2012. Introduction to genetic association studies. *Cold Spring Harb Protoc* 2012:297-306. 10.1101/pdb.top068163

Li M, and Huang D. 2007. On-column refolding purification and characterization of recombinant human interferon-lambda1 produced in Escherichia coli. *Protein Expr Purif* 53:119-123. 10.1016/j.pep.2006.11.011

Malaguarnera M, Di Fazio I, Romeo MA, Restuccia S, Laurino A, and Trovato BA. 1997. Elevation of interleukin 6 levels in patients with chronic hepatitis due to hepatitis C virus. *J Gastroenterol* 32:211-215.

Miedzinski L, and Taylor G. 2008. Ethnicity and spontaneous clearance of hepatitis C in HIV-HCV coinfecting patients. *Can J Infect Dis Med Microbiol* 19:316.

Montes-Cano MA, Garcia-Lozano JR, Abad-Molina C, Romero-Gomez M, Barroso N, Aguilar-Reina J, Nunez-Roldan A, and Gonzalez-Escribano MF. 2010. Interleukin-28B genetic variants and hepatitis virus infection by different viral genotypes. *Hepatology* 52:33-37. 10.1002/hep.23624

Muir AJ, Shiffman ML, Zaman A, Yoffe B, de la Torre A, Flamm S, Gordon SC, Marotta P, Vierling JM, Lopez-Talavera JC, Byrnes-Blake K, Fontana D, Freeman J, Gray T, Hausman D, Hunder NN, and Lawitz E. 2010. Phase 1b study of pegylated interferon lambda 1 with or without ribavirin in patients with chronic genotype 1 hepatitis C virus infection. *Hepatology* 52:822-832. 10.1002/hep.23743

Muraguchi A, Hirano T, Tang B, Matsuda T, Horii Y, Nakajima K, and Kishimoto T. 1988. The essential role of B cell stimulatory factor 2 (BSF-2/IL-6) for the terminal differentiation of B cells. *J Exp Med* 167:332-344.

Nattermann J, Timm J, Nischalke HD, Olbrich A, Michalk M, Tillmann HL, Berg T, Wedemeyer H, Tenckhoff H, Wiese M, Kullig U, Gobel U, Capka E, Schiefke I, Guthof W, Grungriff K, Konig I, Roggendorf M, Sauerbruch T, and Spengler U. 2011. The predictive value of IL28B gene polymorphism for spontaneous clearance in a single source outbreak cohort is limited in patients carrying the CCR5Delta32 mutation. *J Hepatol* 55:1201-1206. 10.1016/j.jhep.2011.03.011

Nattermann J, Vogel M, Berg T, Danta M, Axel B, Mayr C, Bruno R, Tural C, Klausen G, Clotet B, Lutz T, Grunhage F, Rausch M, Nischalke HD, Schewe K, Bienek B, Haerter G, Sauerbruch T, Rockstroh JK, and Spengler U. 2007. Effect of the interleukin-6 C174G gene polymorphism on treatment of acute and chronic hepatitis C in human immunodeficiency virus coinfecting patients. *Hepatology* 46:1016-1025. 10.1002/hep.21778

Pagliaccetti NE, and Robek MD. 2010. Interferon-lambda in HCV Infection and Therapy. *Viruses* 2:1589-1602. 10.3390/v2081589

Polasek O. 2013. Future of biobanks - bigger, longer, and more dimensional. *Croat Med J* 54:496-500.

Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MA, Bender D, Maller J, Sklar P, de Bakker PI, Daly MJ, and Sham PC. 2007. PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet* 81:559-575. 10.1086/519795

Rudan I, Marusic A, Jankovic S, Rotim K, Boban M, Lauc G, Grkovic I, Dogas Z, Zemunik T, Vatauvuk Z, Bencic G, Rudan D, Mulic R, Krzelj V, Terzic J, Stojanovic D, Puntaric D, Bilic E, Ropac D, Vorko-Jovic A, Znaor A, Stevanovic R, Biloglav Z, and Polasek O. 2009. "10001 Dalmatians:" Croatia launches its national biobank. *Croat Med J* 50:4-6.

Seaberg EC, Witt MD, Jacobson LP, Detels R, Rinaldo CR, Margolick JB, Young S, Phair JP, and Thio CL. 2015. Spontaneous Clearance of the Hepatitis C Virus Among Men Who Have Sex With Men. *Clin Infect Dis* 61:1381-1388. 10.1093/cid/civ562

Suppiah V, Moldovan M, Ahlenstiel G, Berg T, Weltman M, Abate ML, Bassendine M, Spengler U, Dore GJ, Powell E, Riordan S, Sheridan D, Smedile A, Fragomeli V, Muller T, Bahlo M, Stewart GJ, Booth DR, and George J. 2009. IL28B is associated with response to chronic hepatitis C interferon-alpha and ribavirin therapy. *Nat Genet* 41:1100-1104. 10.1038/ng.447

Tanaka Y, Nishida N, Sugiyama M, Kurosaki M, Matsuura K, Sakamoto N, Nakagawa M, Korenaga M, Hino K, Hige S, Ito Y, Mita E, Tanaka E, Mochida S, Murawaki Y, Honda M, Sakai A, Hiasa Y, Nishiguchi S, Koike A, Sakaida I, Imamura M, Ito K, Yano K, Masaki N, Sugauchi F, Izumi N, Tokunaga K, and Mizokami M. 2009. Genome-wide association of IL28B with response to pegylated interferon-alpha and ribavirin therapy for chronic hepatitis C. *Nat Genet* 41:1105-1109. 10.1038/ng.449

Thomas DL, Thio CL, Martin MP, Qi Y, Ge D, O'Huigin C, Kidd J, Kidd K, Khakoo SI, Alexander G, Goedert JJ, Kirk GD, Donfield SM, Rosen HR, Tobler LH, Busch MP, McHutchison JG, Goldstein DB, and Carrington M. 2009. Genetic variation in IL28B

468 and spontaneous clearance of hepatitis C virus. *Nature* 461:798-801.
469 10.1038/nature08463
470 Wilson AG, Symons JA, McDowell TL, McDevitt HO, and Duff GW. 1997. Effects of a
471 polymorphism in the human tumor necrosis factor alpha promoter on transcriptional
472 activation. *Proc Natl Acad Sci U S A* 94:3195-3199.
473 Wittke-Thompson JK, Pluzhnikov A, and Cox NJ. 2005. Rational inferences about departures
474 from Hardy-Weinberg equilibrium. *Am J Hum Genet* 76:967-986. 10.1086/430507
475
476

477

478 **Table 1. Demographic and clinical characteristics of IVDU patients by the response-to-**
479 **treatment status**

	SVR, N=78	NR, N=32	Statistics
Sex, N (%)			
men	56 (72%)	26 (81%)	Chi square test, p=0.301
women	22 (28%)	6 (19%)	
Age, median (IQR)	40 (35-45)	41.5 (39-47)	Mann-Whitney, p=0.041
Fibrosis stage, N (%)			
Mean Ishak score †	3 (2.0-3.0)	2 (1.0-3.0)	Mann-Whitney, p=0.077
Mean Knodell score ‡	7 (5.0-9.0)	6 (5.0-9.0)	Mann-Whitney, p=0.803
Viral genotype, N (%)			
1	51 (73%)	19 (27%)	Chi square test P=0.706
3	27 (68%)	13 (33%)	

481 SVR-sustained viral response; NR-non responders

482 † Liver fibrosis was scored by Ishak fibrosis score (0-6) as previously described. (Ishak et al.
483 1995)¹⁰

484 ‡ Histology activity index by Knodell score (0-18) in chronic active hepatitis. (Knodell et al. 1981)⁹

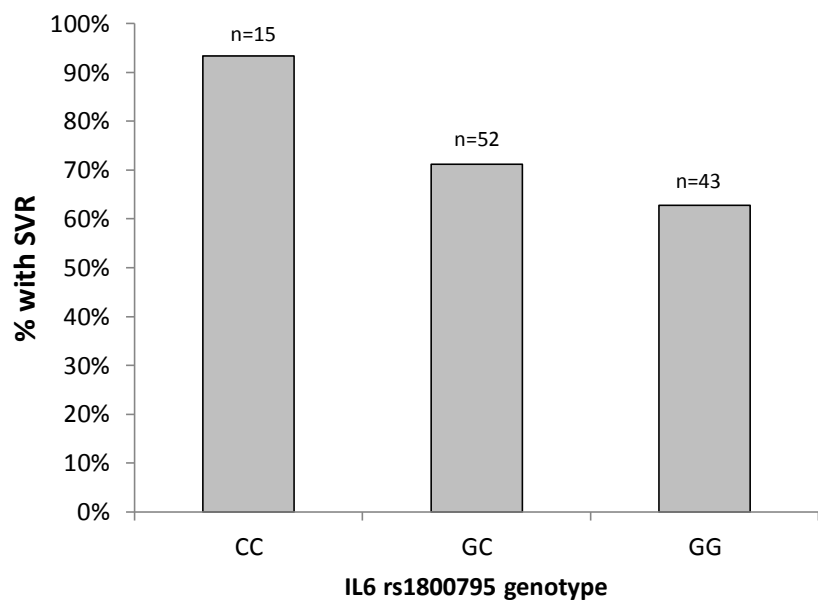
485
486
487

Table 2. Distributions of rs1800795- IL6 and rs12979860- IL28B genotypes, by the response-to-treatment group

SNP	Genotype	SVR, N=78	NR, N=32	Total
IL-28B, N (%)	CC	22 (28%)	8 (25%)	30 (27%)
	CT	50 (64%)	19 (59%)	69 (63%)
	TT	6 (8%)	5 (16%)	11 (10%)
IL-6, N (%)	GG	27 (35%)	16 (50%)	43 (39%)
	GC	37 (47%)	15 (47%)	52 (47%)
	CC	14 (18%)	1 (3%)	15 (14%)

488 SVR-sustained viral response; NR-non responders

489



490

491 **Figure 1.** The percentage of patients with SVR by the rs1800795-IL6 genotypes

492 | n = total number of patients with each genotype.

493

494

495

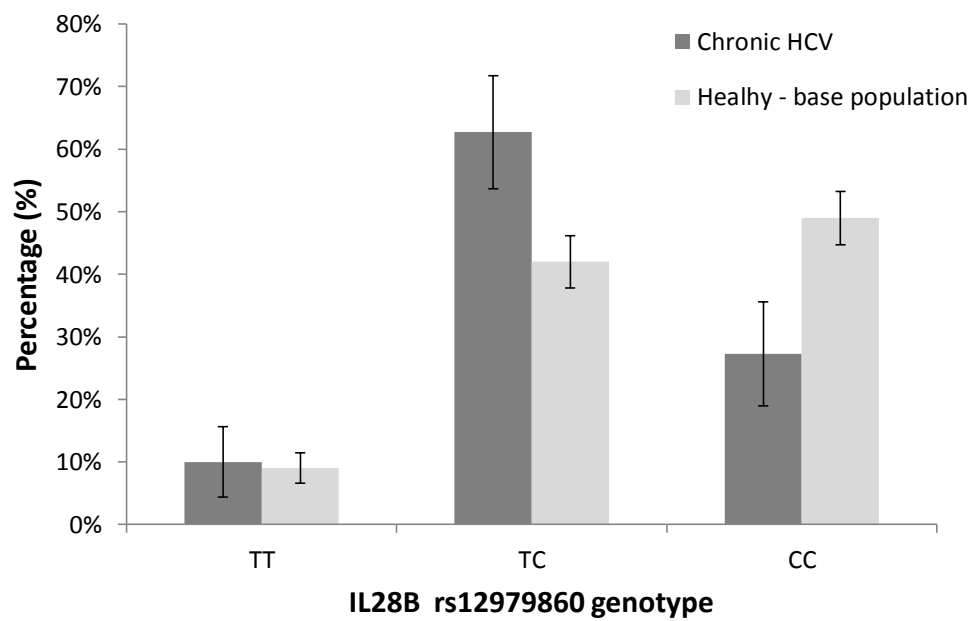


Figure 1. The distribution of rs12979860-IL28B genotypes among: IVDU with chronic HCV (in black bars), and the underlying (base), apparently healthy population (in gray bars). Shown are percentages with accompanying 95% CI. Show statistically significances using * above the bars.