

Flower diversity and bee reproduction in an arid ecosystem

Jimena Dorado, Diego P. Vázquez

Background. Diverse flower communities are more stable in floral resource production along the flowering season, but the question about how the diversity and stability of resources affect pollinator reproduction remains open. High plant diversity could favor short foraging trips, which in turn would enhance bee fitness. In addition to plant diversity, greater temporal stability of floral resources in diverse communities could favor pollinator fitness because such communities are likely to occupy the phenological space more broadly, increasing floral availability for pollinators throughout the season. In addition, this potential effect of flower diversity on bee reproduction could be stronger for generalist pollinators because they can use a broader floral spectrum. Based on above arguments we predicted that pollinator reproduction would be positively correlated to flower diversity, and to temporal stability in flower production, and that this relationship would be stronger for the most generalized pollinator species. **Materials & Methods.** Using structural equation models, we evaluated the effect of these variables and other ecological factors on three estimates of bee reproduction, and whether such effects were modulated by bee generalization on floral resources. **Results.** Contrary to our expectations, flower diversity had no effect on bee reproduction, stability in flower production had a weakly negative effect on one of the bee reproductive variables, and the strength of the fitness-diversity relationship was unrelated to bee generalization. In contrast, elevation had a weak, non significant negative effect on bee reproduction, despite the narrow elevation range encompassed by our sites. **Discussion.** Flower diversity did not affect the reproduction of the solitary bees studied here. Although high temporal stability in flower production is expected to enhance pollinator reproduction, in our study it had a weakly negative---instead of positive---effect on the average number of brood cells per nest. Other environmental factors that vary with elevation could influence bee reproduction. Our study focused on a small group of closely-related bee species, which cautions against generalization of our findings to other groups of pollinators. More studies are clearly needed to assess the extent to which pollinator demography is influenced by the diversity of floral resources.

1 Flower diversity and bee reproduction in an arid ecosystem.

2 Jimena Dorado^{1,*} and Diego P. Vázquez^{1,2}

3

4 1 Instituto de Argentino de Investigaciones de las Zonas Áridas, CONICET, CC 507, 5500

5 Mendoza, Argentina.

6 2 Instituto de Ciencias Básicas, Universidad Nacional de Cuyo, Centro Universitario,

7 M5502JMA Mendoza, Argentina.

8 * E-mail: jdorado@mendoza-conicet.gov.ar; fax: +54-261-524-4101.

9 Abstract

10 **Background.** Diverse flower communities are more stable in floral resource production along
 11 the flowering season, but the question about how the diversity and stability of resources affect
 12 pollinator reproduction remains open. High plant diversity could favor short foraging trips,
 13 which in turn would enhance bee fitness. In addition to plant diversity, greater temporal stability
 14 of floral resources in diverse communities could favor pollinator fitness because such
 15 communities are likely to occupy the phenological space more broadly, increasing floral
 16 availability for pollinators throughout the season. In addition, this potential effect of flower
 17 diversity on bee reproduction could be stronger for generalist pollinators because they can use a
 18 broader floral spectrum. Based on above arguments we predicted that pollinator reproduction
 19 would be positively correlated to flower diversity, and to temporal stability in flower production,
 20 and that this relationship would be stronger for the most generalized pollinator species.

21 **Materials & Methods.** Using structural equation models, we evaluated the effect of these
 22 variables and other ecological factors on three estimates of bee reproduction, and whether such
 23 effects were modulated by bee generalization on floral resources. **Results.** Contrary to our
 24 expectations, flower diversity had no effect on bee reproduction, stability in flower production
 25 had a weakly negative effect on one of the bee reproductive variables, and the strength of the
 26 fitness-diversity relationship was unrelated to bee generalization. In contrast, elevation had a
 27 weak, non significant negative effect on bee reproduction, despite the narrow elevation range
 28 encompassed by our sites. **Discussion.** Flower diversity did not affect the reproduction of the
 29 solitary bees studied here. Although high temporal stability in flower production is expected to
 30 enhance pollinator reproduction, in our study it had a weakly negative---instead of positive---
 31 effect on the average number of brood cells per nest. Other environmental factors that vary with

32 elevation could influence bee reproduction. Our study focused on a small group of closely-
 33 related bee species, which cautions against generalization of our findings to other groups of
 34 pollinators. More studies are clearly needed to assess the extent to which pollinator demography
 35 is influenced by the diversity of floral resources.

36 Key words: pollination, flower diversity, bee fitness

Introduction

There is a consensus that diversity enhances ecosystem functioning (Cardinale et al. 2012). Species diversity provides redundancy in function, so that ecological processes are more stable in more diverse communities (MacArthur 1955, Elton 1958). Stability has multiple definitions, resulting in a variety of diversity–stability relationships (Ives & Carpenter 2007). In plant communities, the diversity–stability relationship has been well studied for biomass production (Caldeira et al. 2005, Tilman, Reich & Knops 2006, Isbell et al. 2009, Hector et al. 2010), and we have recently reported that diverse flower communities are also more temporally stable in terms of floral resource production (Dorado & Vázquez 2014). But the question about how the diversity and stability of resources affect reproduction of pollinators remains open. Wild bees are gaining increasing attention as they are known to enhance fruit set of crops regardless of honey bee abundance (Garibaldi et al. 2013). Thus, generating knowledge that contributes to understand the determinants of the demography and population dynamics of pollinators is becoming a priority, as it can contribute to their conservation.

There is a general positive association between plant and pollinator diversity (e.g., Steffan-Dewenter & Tschamntke 2001, Potts et al. 2003a, Ebeling et al. 2008, Fründ, Linsenmair & Blüthgen 2010), but little is known about the mechanisms behind this relationship. A positive relationship would be expected if the diversity in one group enhanced the populations in the other group, so that greater flower diversity enhanced the demography of pollinator populations, and greater pollinator diversity enhanced the demography of plant populations. For example, blueberries planted next to wild flowers that support rich pollinator assemblages produced higher fruit set, berry weight, and number of mature seeds per berry (Blaauw & Isaacs 2014). Similarly,

a generalist cavity nesting bee species (*Osmia rufa*) reduces its foraging trip duration as the number of plant species increases (Gathmann & Tschardt 2002), which can result in higher energy available for reproduction. Another bee (*Dieunomia triangulifera*) increased its mean reproductive success when collecting high amount of pollen in shorter foraging trips (Minckley et al. 1994), whereas other bees (*Hoplitis adunca* and *Chelostoma rapunculi*) decreased the proportion of brood cells provisioned per time unit with long foraging trips (Zurbuchen et al. 2010). In another species (*Osmia lignaria*), fitness depends on the closeness to natural patches (Williams & Kremen 2007), which usually harbor greater plant diversity; this bee species also feeds its larvae using pollen from different plant species despite higher foraging costs (Williams & Tepedino 2003). Thus, if high plant diversity tends to shorten foraging trips, and short foraging trips enhance bee fitness, plant diversity could indirectly enhance bee fitness. This effect should be stronger for polylectic than oligolectic pollinators, given that the latter are more restricted in their diet.

In addition to plant diversity, greater temporal stability of floral resources in diverse communities (Dorado & Vázquez 2014) could favor pollinator fitness because such communities are likely to occupy the phenological space more broadly than their species-poor counterparts, increasing floral availability for pollinators throughout the season. For example, in multi-species assemblages of herbaceous plants of the genus *Clarkia*, diverse communities provide more resources along the flowering season, sustaining a higher number of pollinator individuals per plant (Moeller 2004). Furthermore, a bumblebee study found that even if floral resources are abundant, high stability of floral resources throughout the flowering season is needed to enhance bumblebee fitness (Wesphal et al. 2009). Thus, both high flower abundance and high temporal

stability of floral resources are likely to enhance pollinator reproduction (Müller et al. 2006, Westphal, Steffan-Dewenter, & Tschardtke 2009).

To evaluate whether there is an effect of flower diversity on pollinator reproduction it is necessary to disentangle the effect of flower abundance, as it could be positively correlated with flower richness, as it happens with biomass in plant communities (Tilman 1999); if so, there could be a spurious positive correlation between flower richness and pollinator fitness. Other local environmental factors, such as elevation or disturbance history, should also be accounted for, as they are known to influence species diversity (Potts et al. 2003b, Grytnes and McCain 2007, Dorado & Vázquez 2014). Structural equation modeling (SEM) represents an excellent tool to assess causal relationships among multiple variables simultaneously (Grace 2006).

Our aim is to study the effect of flower diversity and temporal stability of floral resources on the reproduction of seven species of cavity nesting bees from the Monte desert in Argentina. Based on the above arguments, we expected to find that flower diversity and temporal stability of floral resources correlates positively to three estimates of bee reproduction: average number of brood cells per nest per site, total number of brood cells per site, and total number of nests per site. We also expected to find a positive correlation between the strength of the fitness-diversity correlation and the degree of generalization of each bee species.

Methods

Study area and sampling

This study was conducted in the Monte desert in Villavicencio Nature Reserve, located ca. 40

km north of Mendoza city, Argentina, during the 2008 flowering season (15 October – 8 December 2008; authorized by Dirección de Recursos Naturales Renovables de la Provincia de Mendoza, approval numbers 1130 and 646). We worked in fourteen 100 m × 200 m rectangular study sites (minimum and maximum distance between them were 1.11 km and 14.13 km respectively). These sites lie at 1100-1500 m above sea level, at the ecotone between the Monte desert and the Prepuna biomes (Ambrosetti et al. 1986). The plant community is a 2 m tall shrubland dominated by *Larrea divaricata* (Zygophyllaceae), *Zuccagnia punctata* (Fabaceae), *Prosopis flexuosa* (Fabaceae), *Condalia microphylla* (Rhamnaceae), *Acantholippia seriphoides* (Verbenaceae), and *Opuntia sulphurea* (Cactaceae). We selected sites with contrasting flower abundance, composition and diversity. The region suffers from recurrent fires, which are mostly human-caused and are in fact the most common human disturbance (E. L. Stevani, pers. comm.); the time elapsed since the last fire varied substantially among our study sites (Table S1).

Trap nest sampling

We placed trap nests in 6 points per plot as shown in Fig. S1. Each point had two groups of 24 trap nests consisting of a wood piece with a longitudinal cavity of 5 mm, 8 mm or 11 mm in diameter, and 15 cm of length for the smallest two diameters and 28 cm of length for the largest diameter; wood pieces were arranged as shown in Fig. S2. Trap nests were checked weekly; occupied traps were taken to the laboratory and replaced by empty ones. Each trap nest constitutes one bee nest. Once in the laboratory, nests were opened to record the number of cells; whenever the nest had more than one brood cell, one of them was extracted for pollen identification, and the rest was kept until adult emergence. Pollen identification was done by comparison with a reference collection, prepared including all plant species that flowered at each

study plot (Dorado et. al 2011, Vázquez et. al 2012). The number of emerged adults and their taxonomic identity were recorded in all nests. Although adult number may be a good estimator of female fitness in the absence of larval mortality, because of the high rate of nest parasitism recorded in our study we used instead the number of brood cells per nest as an estimate of female fitness. One species, *Trichothurgus laticeps* Friese, lacks brood cells, as females lay eggs bare amidst a pollen mass; thus, for this species we used the length of the trap cavity occupied by pollen as an estimate of the number of brood cells. For the analysis, we used only the seven bee species that occupied at least 30 trap nests, as we judged smaller sample sizes unreliable for statistical analyses.

Plant sampling

Floral resource availability was studied using flower density. Although he have estimates of the amount of pollen per flower for each species, we decided to use flower density as a measure of resource abundance, as flowers represent the resources packages encountered by pollinators as they forage (see also Vázquez et al. 2009). Flower density was measured weekly at four 8 m × 20 m plots and two 2 m × 50 m transects per site, as described in Fig. S1. We considered weekly sampling intervals adequate, as flowers in our system usually last less than a week. Flower density was estimated multiplying the mean number of flowers per individual by the total number of flowering individuals in the transect or plot when individuals could be distinguished (shrubs and some herbs); we estimated the number of flowers per individual in at least ten individuals of the site. When it was not possible to identify flowering individuals (some herbaceous species), all flowers in a plot or transect were counted. We included in the study all flowering plant species that were assumed to be animal pollinated (we excluded only grass

species).

Statistical analysis

To evaluate the effects of flower diversity and temporal stability of floral resources on bee fitness and to assess the influence of other ecological factors on this relationship, we used structural equation models (hereafter SEM). We built a general initial model to explore the effects of flower richness, flower abundance, time elapsed since the last fire, elevation, and temporal stability in flower production on three bee reproductive parameters (average number of brood cells per nest, total number of brood cells per site and total number of nests per site). To estimate the average number of brood cells per nest we used only data of sites where species were present; for the total number of brood cells per site and the total number of nests per site we used data of all sites, as the absence of a species in a site represented zero abundance. Flower richness was used as a proxy of flower diversity; it was rarefied to remove the effect of flower abundance. Flower abundance was estimated as flower density per site. Time elapsed since the last fire was provided by park rangers (E. L. Stevani, personal communication). Temporal stability in flower production along the season was calculated as the inverse of its coefficient of variation (see Dorado & Vázquez 2014). All variables were standardized subtracting each value by the mean and dividing by the standard deviation. For each bee species, from the initial model we generated more parsimonious nested models by removing variables with small non-significant path coefficients (see models in Fig. S3; see also Maestre et al. 2010).

We evaluated alternative SEM models using a d-separation test (Shipley 2000, 2013) described in S1 section. This analysis allowed us to select the best fitting model based on Akaike's

information criterion (AIC) using a small sampling size. To discriminate among competing models we used the AIC difference, ΔAIC , between a given model and the best-fitting one, i.e., that with the lowest value of AIC. When $\Delta AIC < 3$, models are generally considered to have substantial support; for $3 > \Delta AIC < 7$, models are considered to have considerably less support, while for $\Delta AIC > 10$, models have essentially no support relative to the best model of the set (Richards 2005, Burnham & Anderson 2010). To evaluate whether the studied effects were general for all bee species, we used meta-analytical methods (S2).

To evaluate whether the effect of flower diversity becomes stronger with increasing pollinator generalization, we performed Spearman's rank correlations between the path coefficient representing the effect of flower richness on each of the three bee reproductive parameters mentioned above, and two measures of the corresponding species degree of generalization. We estimated the degree of diet generalization of each bee species using the species degree and Simpson's diversity index; degree is simply the number of food species consumed from all sites polled, whereas Simpson's index is a function of the number of food items and the proportion in which they were consumed. We used rarefaction to estimate both measures of generalization to make them comparable among bee species, as the number of brood cells was highly variable among nests. A positive correlation between the path coefficient of flower richness on bee reproduction and generalization would support our hypothesis that the reproduction of generalist pollinators is enhanced by flower richness.

All analyses were performed using R statistical software (R Core Team 2013). Rarefaction of flower richness was performed using the rarefy function of the vegan package (Oksanen et al.

2013). Pearson's partial correlations were performed using the `pcor.test` function of the `ppcor` package to obtain independence probabilities and the path coefficients (Kim and Yi 2007, Kim & Yi 2006, Johnson & Dean 2002).

Results

We recorded 598 occupied trap nests by 11 solitary bee species (Table 1).

The complete model assessing the influence of multiple ecological factors on the potential relationship between flower diversity and bee fitness (Model 1, Fig. S3a) showed no effect of flower abundance or time elapsed since the last fire on the bee reproductive variables studied. Although the only significant effect was that of temporal stability on the average number of brood cells per nest, we kept flower richness and elevation in the simplified model because they showed suggestive, albeit non significant, trends. There was a weak positive effect of flower richness on average number of brood cells per nest and a weak negative effect of elevation on the three reproductive variables (Fig S4); however, none of these effects were statistically significant. The simplified model (model 2, Fig. S3b) fitted best according to ΔAIC (8.43) for all species (see S1 and Fig S3); this model fits the data well according to the d-separation test ($p = 0.84$, $df = 2$, $C = 3.69$). In this simplified model, the negative effect of temporal stability in flower production on the average number of cells per nest was weaker than in the complete model (Fig 1 blue error bars; confidence limits of path coefficient for model 1: -0.467, -0.101; confidence limits of path coefficient of model 2: -0.264, -0.004). Also, the simplified model shows a negative trend in the effect of elevation on the total number of cells and nests per site, but this trend is statistically non significant (confidence limits of path coefficients for the total

number of cells and nests per site respectively: -0.465, 0.031, and -0.472, 0.027).

The effect of flower diversity on pollinator reproduction was unrelated to pollinator generalization for any of the bee reproductive variables and generalization indices used (Table 2).

Discussion

Contrary to our expectations, we found no effects of flower diversity and flower abundance on bee reproduction. Thus, flower diversity did not matter for the reproduction of the solitary bees studied here. Although high temporal stability in flower production is expected to enhance pollinator reproduction (Wesphal et al. 2009, Dorado & Vázquez 2014), in our study it had a weakly negative---instead of positive---effect on the average number of brood cells per nest. In contrast, elevation had a non significant negative effect on bee reproduction, despite the narrow elevational range encompassed by our sites (1100–1500 m), which suggests that the intrinsic characteristics of the study sites could have influenced the relationship between floral diversity and bee reproduction.

An explanation of the negative effect of temporal stability on brood cell production concerns a compensatory behavior of females to avoid parasitism. In sites with high temporal stability in flower production, females might lay fewer eggs per nest while building more nests, so as to maximize larval survival per site. This reasoning makes two implicit assumptions. First, that the bee species are parasitized, which we indeed observed for many of the bee species studied here (data not shown). Second, that nesting sites are not limited for the population. In fact, the trap

nest sampling with replacement highly increased the nest availability in our study sites. If this mechanism were responsible for the observed negative effect of temporal stability in flower production on the average number of brood cells per nest, we would expect the number of cells per site to be either unrelated to temporal stability or to be higher in the more temporally stable sites, and the number of nests to be higher in the more temporally stable sites, as females would be laying eggs at their maximum capacity but distributing them in more nests. Matching these expectations, the total number of brood cells per site was unrelated to temporal stability (Fig. 1, flower stability), while the number of nests per site tends to increase with temporal stability for most species, although the effect was statistically non-significant.

An alternative explanation of the negative effect of temporal stability on brood cell production per nest could be that elevation might be weakening the effects of other variables on pollinator reproduction. This is particularly likely considering the positive direct effect of elevation on flower diversity, the positive indirect effect of elevation on stability, and the (nonsignificant) negative effect of elevation on bee reproduction (Fig. 1); these effects could be neutralizing the effect of the flower diversity and temporal stability in flower production on bee reproduction. The upper sites are located in ravines, which are probably wetter and cooler than the lower sites, located in open land. Thus, changes in humidity and temperature associated to elevation could be influencing bee reproduction more strongly than the other ecological factors studied here.

We found no support for the idea that generalist bees are more favored in their reproduction by flower diversity than specialized ones, despite bee species of this study having contrasting degrees of feeding specialization. This result is reasonable given our finding of no general effect

of flower diversity on bee reproduction.

Our study focused on a small group of closely-related bee species, representing less than 5% of the pollinator assemblage in our study area (Chacoff et. al 2012). Our study is in this sense limited, and our finding of no effects of floral diversity on pollinator demography cannot be generalized. More studies are clearly needed to assess the extent to which pollinator demography is influenced by the diversity of floral resources. Furthermore, although we have considered environmental factors, such as elevation or time post-fire, which appeared a priori good candidates to influence bee demography, other environmental factors may also be important. These include humidity and temperature, which should covary with elevation, and other biotic factors such as competition, predation and parasitism.

Acknowledgements

We thank the administration of Villavicencio Natural Reserve for permission to conduct this study, the park rangers for help to find appropriated study sites in the field, Arturo Roig for help with bee identifications, Leticia Escudero, Nydia Vitale and Georgina Amico for laboratory assistance, and members of the Ecological Interactions Lab for helpful comments on the manuscript. JD is a postdoctoral fellow and DPV a career researcher with CONICET. Research was funded through grants from CONICET (PIP 6564), FONCYT (PICT 20805, 1471 and 2010-2779), and BBVA Foundation (BIOCON03-162).

References

Blaauw, B. R., Isaacs, R. 2014. Flower plantings increase wild bee abundance and the

pollination services provided to a pollination-dependent crop. *Journal of Applied Ecology*
51, 809–898.

Burnham, K. P., and Anderson, D. R. 2010. Model selection and multimodel inference. A
practical information – theoretic approach 2nd ed.). New York: Springer.

Caldeira, M. C., A. Hector, M. Loreau, and J. S. pereira. 2005. Species richness, temporal
variability and resistance of biomass production in a Mediterranean grassland. *Oikos* 110, 115–
123.

Cardinale, B. J., J. E. Duffy, A. Gonzalez, D. U. Hooper, C. Perrings, P. Venail, A. Narwani¹, G.
M. Mace, D. Tilman, D. A. Wardle, A. P. Kinzig, G. C. Daily, M. Loreau, J. B. Grace, A.
Larigauderie, D. S. Srivastava & S. Naeem. 2012. Biodiversity loss and its impact on humanity.
Nature, **486**, 59–67.

Chacoff, N. P., D. P. Vázquez, S. B. Lomáscolo, E. L. Stevani, and B. Padrón. 2012. Evaluating
sampling completeness in a desert plant–pollinator network. *Journal of Animal Ecology*
81, 190–200.

Dorado, J. and Vázquez, D. P. 2014. The diversity–stability relationship in floral production.
Oikos 123, 1137–1143.

Dorado, J., D. P. Vázquez, E. L. Stevani, and N. P. Chacoff. 2011. Rareness and specialization
in plant–pollination networks. *Ecology* 92, 19–25.

Ebeling, A., A. M. Klein, J. Sumacher, W. W. Weisser, and T. Tscharntke. 2008. How does
plant richness affect pollinator stability and temporal stability of flower visits? *Oikos* 117,
1808–1815.

Elton, C. S. 1958. Ecology of invasions by animals and plants. With a New Foreward by Daniel
Simberloff 2000. University of Chicago Press.

Fründ J., K. E. Linsenmair, and N. Blüthgen. 2010. Pollinator diversity and specialization in relation to flower diversity. *Oikos* 119, 1581–1590.

Garibaldi L. A., I. Steffan-Dewenter, C. Kremen, J. M. Morales, R. Bommarco, S. A. Cunningham, L. G. Carvalheiro, N. P. Chacoff, J. H. Dudenhöffer, S. S. Greenleaf, A. Holzschuh, R. Isaacs, K. Krewenka, Y. Mandelik, M. M. Mayfield, L. A. Morandin, S. G. Potts, T. H. Ricketts, H. Szentgyörgyi, B. F. Viana, C. Westphal, R. Winfree & A. M. Klein. 2013. Wild pollinators enhance fruit set of crops regardless of honey bee abundance. *Science* 339, 1608–1611.

Gathmann, A., and Tschardtke, T. 2002. Foraging ranges of solitary bees. *Journal of Animal Ecology* 71, 757–764.

Grace, J. B. 2006. Structural equation modeling and natural systems. Cambridge University Press, New York. 365 pp.

Grytnes, J. A., and McCain, C. M. 2007. Elevational trends in biodiversity. In: S. Levin (ed.) Encyclopedia of biodiversity. Elsevier, Inc. pp. 1–8.

Hall. Whittaker, J. 1990. Graphical models in applied multivariate statistics. John Wiley & Sons.

Hector, A., Y. Hautier, P. Saner, L. Wacker, R. Bagchi, J. Joshi, M. Scherer-Lorenzen, E. M. Spehn, E. Bazeley-White, M. Weilemann, M. C. Caldeira, P. G. Dimitrakopoulos, J. A. Finn, K. Huss-Danell, A. Jumpponen, C. P. H. Mulder, C. Palmborg, J. S. Pereira, A. S. D. Siamantziouras, A. C. Terry, A. Y. Troumbis, B. Schmid & M. Loreau. 2010. General stabilizing effects of plant diversity on grassland productivity through population asynchrony and overyielding. *Ecology* 91, 2213–2220.

Isbell, F. I. et al.. 2009. Biodiversity, productivity and the temporal stability of productivity:

336 patterns and processes. –*Ecology Letters* 12:443–451.

337 Ives, A. R., and S. R. Carpenter. 2007. Stability and diversity of ecosystems. *Science* 317, 58–

338 62.

339 Johnson, R. A. and W. Dean. 2002. Applied multivariate statistical analysis. –Prentice

340 Kim, S.H. and Yi, S. 2007. Understanding relationship between sequence and functional

341 evolution in yeast proteins. *Genetica* 131, 151–156.

342 Kim, S.H. and Yi, S. 2006. Correlated asymmetry between sequence and functional divergence

343 of duplicate proteins in *Saccharomyces cerevisiae*. *Molecular Biology and Evolution* 23,

344 1068–1075.

345 Lázaro, A. and O. Totland. 2010. Local floral composition and the behaviour of pollinators:

346 attraction to foraging within experimental patches. *Ecological Entomology* 35, 652–661.

347 MacArthur, R. 1955. Fluctuations of Animal Populations and a Measure of Community

348 Stability. *Ecology* 36, 533–536.

349 Maestre F.T., M. A. Bowker, C. Escolar, M. D. Puche, S. Soliveres, S. Maltez-Mouro, P. García-

350 Palacios, A. P. Castillo-Monroy, I. Martáñez & A. Escudero. 2010. Do biotic interactions

351 modulate ecosystem functioning along stress gradients? Insights from semi–arid plant and

352 biological soil crust communities. *Philosophical Transactions of the Royal Society B:*

353 *Biological Sciences* 365, 2057-2070.

354

355 Mincley, R. L., W. T. Wcislo, D. Yanega, and S. L. Buchmann. 1994. Behavior and phenology

356 of a specialist bee (*Dieunomia*) and sunflower (*Helianthus*) pollen availability. *Ecology* 75,

357 1406–1419.

358 Moeller, D. 2004. Facilitative interactions among plants via shared pollinators. *Ecology* 85,

3289–3301.

Müller, A, S. Diener, S. Shnyder, K. Stuts, C. Sedivy & S. Dorn. 2006. Quantitative pollen

requirements of solitary bees: implications for bee conservation and the evolution of bee–

flower relationships. *Biological Conservation* 130, 604–615.

Oksanen, J., F. G. Blanchet, R. Kindt, P. Legendre, P. R. Minchin, R. B. O'Hara, G. L. Simpson,

P. Solymos, M. H. H. Stevens, and H. Wagner. 2013. vegan: Community Ecology

Package.R package version 2.0-8 <http://CRAN.R-project.org/package=vegan>

Potts, S.G., B. Vulliamy, A. Dafni, G. Ne'eman & P. G. Willmer. 2003a. Linking bees and

flowers: how do floral communities structure pollinator communities? *Ecology* 84, 2628–

2642.

Potts, S. G., B. Vulliamy, A. Dafni, G. Ne'eman, C. O'Toole, S. Roberts & P. G. Willmer.

2003b. Response of plant-pollinator communities to fire: changes in diversity, abundance

and floral reward structure. *Oikos* 101, 103–112.

R Core Team 2013. R: A language and environment for statistical computing. R Foundation for

Statistical Computing, Vienna, Austria. URL <http://www.R-project.org/>.

Richards, S. A. 2005. Testing ecological theory using the information–theoretic approach:

examples and cautionary results. *Ecology* 86, 2805–2814.

Roulston, T. H. and Goodell, K. 2011. The role of resources and risks in regulating wild bee

populations. *Annual Review of Entomology* 56: 293–312.

Sargent, R. D. and Ackerly, D. D. 2008. Plant-pollinator interactions and the assembly of plant

communities. *Trends Ecology and Evolution* 23, 123–130.

Shipley, B. 2000. Cause and correlation in biology. A user's guide to path analysis, structural

equations, and causal inference. Cambridge University Press.

Shipley, B. 2013. The AIC model selection method applied to path analytic models compared using a d-separation test. *Ecology* 94, 560–564.

Steffan-Dewenter, I., and Tschardtke, T. 2001. Succession of bee communities on falds. *Ecography* 24, 83–93.

Tilman D., P. B. Reich & J. M. H. Knops. 2006. Biodiversity and ecosystem stability in a decade-long grassland experiment. *Nature* 441, 629–632.

Tilman, D. 1999. Diversity by default. *Science* 283, 495–496.

Vázquez, D. P., S. B. Lomáscolo, M. B. Maldonado, N. P. Chacoff, J. Dorado, E. L. Stevani, and N. L. Vitale. 2012. The strength in plant–pollination interactions. *Ecology* 93, 719–725.

Westphal, C., I. Steffan-Dewenter, and T. Tschardtke. 2009. Mass flowering oilseed rape improves early colony growth but not sexual reproduction of bumblebees. *Journal of Applied Ecology* 46, 187–193.

Williams, N. M. and C. Kremen. 2007. Resource distribution among habitats determine solitary bee offspring production in a mosaic landscape. *Ecological Applications* 17, 910–921.

Williams, N. M. and V. J. Tepedino. 2003. Consistent mixing of near and distant resources in foraging bouts by the solitary mason bee *Osmia lignaria*. *Behavioral Ecology* 14, 141–149.

Zurbuchen, A., S. Cheesman, J. Klaiber, A. Muller, S. Hein, and S. Dorn. 2010. Long foraging distances impose high costs on offspring production in solitary bees. *Journal of Animal Ecology* 79, 674–681.

402 Figure 1. Box-plot summarizing the path coefficients of model 2 (see Fig. S3) for the seven bee
 403 species studied here. Model 2 describes the effect of flower diversity (estimated using flower
 404 richness), temporal stability of flower production along the flowering season (estimated as the
 405 inverse of the coefficient of variation of the weekly flower abundance mean), and elevation (m
 406 above sea level). The ordinates represents the path coefficients; the abscissa represent the effect
 407 of the above ecological variables on three bee reproductive variables: A, average brood cell
 408 number per nest per site; B, total number of brood cells per site; C, total number of nests per site.
 409 Black error bars are the maximum and minimum values, and blue error bars are the ninety-five
 410 percent confidence limits of path coefficients obtained from bootstrap sampling of the
 411 distribution of path coefficients, and black.

Table 1(on next page)

Table 1

Number of nests per species. We used in this study the species that had more than 30 nests.

Table 1. Number of nests per species. We used in this study the species that had more than 30 nests.

Bee species	Occupied trap nests
<i>Anthidium andinum</i> Jörgensen	6
<i>Anthidium decaspilum</i> Moure	54
<i>Anthidium rubripes</i> Friese	31
<i>Anthidium vigintipunctatum</i> Friese	39
<i>Megachile</i> sp. A	222
<i>Megachile</i> sp. C	17
<i>Megachile ctenophora</i> Holmberg	74
<i>Mourecozelles triciliatus</i> Toro y Cabezas	3
<i>Trichothurgus laticeps</i> Friese	59
<i>Xylocopa ordinaria</i> Smith	88
<i>Xylocopa splendidula</i> Lepertier	5

Table 2(on next page)

Table 2

Correlation coefficients between effect of flower diversity on pollinator reproduction and bee generalization.

1 Table 2. Correlation coefficients between effect of flower diversity on pollinator reproduction
2 and bee generalization.

Generalization index	Reproductive variable	Correlation coefficient (<i>r</i>)	<i>p</i> -value	<i>N</i>
degree	Average number of cells per nest	- 0.21	0.66	7
degree	Total number of cells per site	0.42	0.35	7
degree	Total number of nests per site	0.39	0.39	7
Simpson's diversity index	Average number of cells per nest	0.14	0.78	7
Simpson's diversity index	Total number of cells per site	0.46	0.30	7
Simpson's diversity index	Total number of nests per site	0.28	0.55	7

3

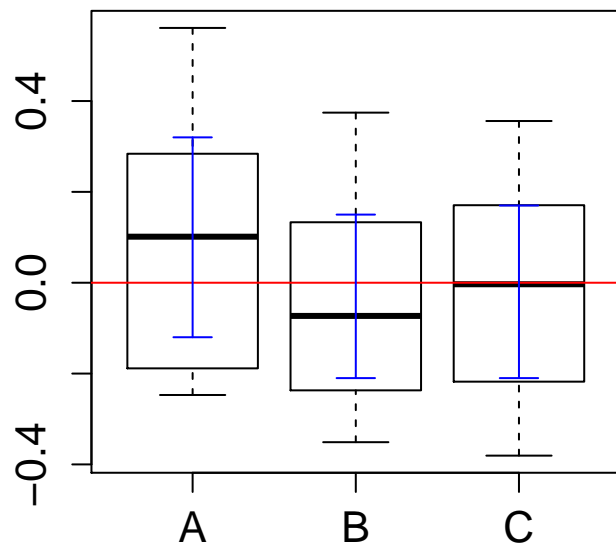
4

Figure 1(on next page)

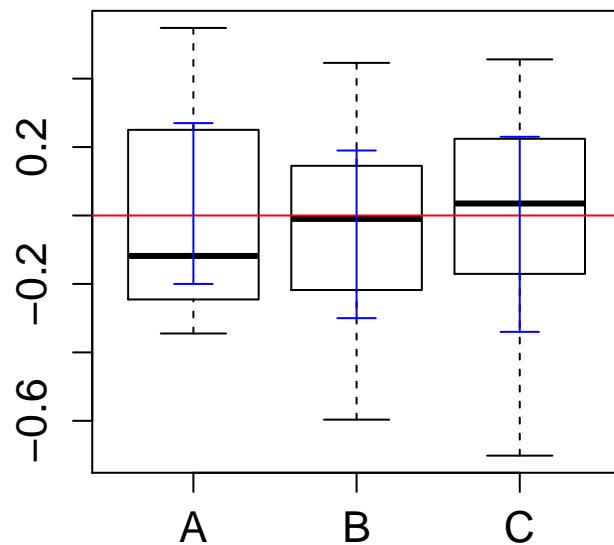
Figure 1

Figure 1. Box-plot summarizing the path coefficients of model 2 (see Fig. S3) for the seven bee species studied here. Model 2 describes the effect of flower diversity (estimated using flower richness), temporal stability of flower production along the flowering season (estimated as the inverse of the coefficient of variation of the weekly flower abundance mean), and elevation (m above sea level). The ordinates represents the path coefficients; the abscissa represent the effect of the above ecological variables on three bee reproductive variables: A, average brood cell number per nest per site; B, total number of brood cells per site; C, total number of nests per site. Black error bars are the maximum and minimum values, and blue error bars are the ninety-five percent confidence limits of path coefficients obtained from bootstrap sampling of the distribution of path coefficients, and black.

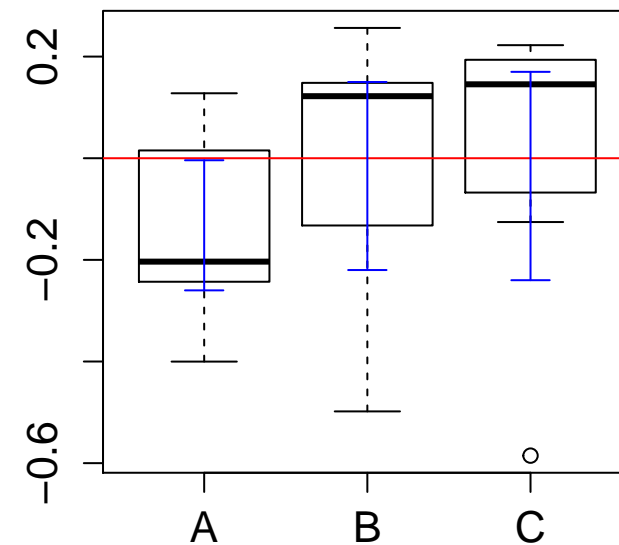
Flower diversity



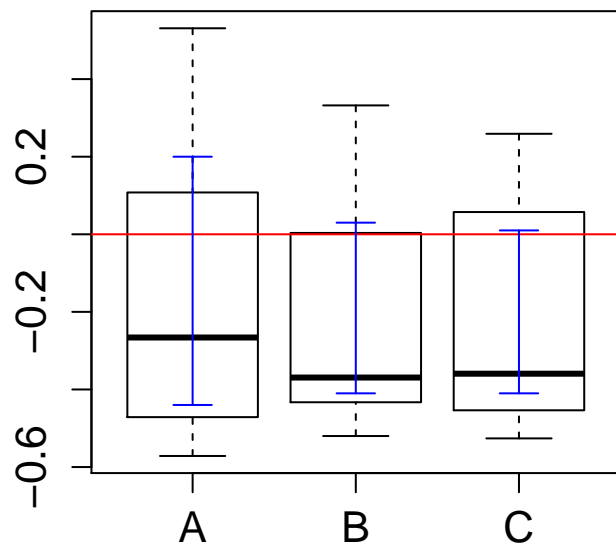
Flower diversity total effect



Flower stability



Elevation



Elevation total effect

