



Carbon capture, photosynthesis, and leaf gas exchange of shade tree species and Arabica coffee varieties in coffee agroforestry systems in Veracruz state, Mexico

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ABSTRACT

Agroforestry systems with native vegetation enhance climate adaptation and mitigation by improving coffee farm resilience, carbon storage, and income diversification. Seven native tree species were pre-selected as shade providers for Veracruz coffee agroforestry systems based on ecological, cultural, and economic criteria. The present study evaluated their physiological performance through above-ground biomass, carbon stocks, and *in-situ* chlorophyll fluorescence and gas exchange measurements under controlled light and temperature conditions. Five *Coffea arabica* varieties were also assessed under these shade canopies using the same leaf-level parameters, and leaf nitrogen and moisture content. *Erythrina americana* and *Persea schiedeana* had the highest carbon sequestration per tree. *E. americana* showed the highest water-use efficiency, whereas *P. schiedeana* showed the lowest transpiration and stomatal conductance, indicating a water-saving strategy via stomatal restriction. These traits reflect their ecological adaptations to shade and microclimate conditions in agroforestry systems. *Inga inicuil* achieved the highest carbon capture per hectare due to high tree density, despite lower individual performance. Species-specific strategies were identified: *Psidium guajava* and *P. schiedeana* exhibited high transpiration but limited carbon gain. *E. americana* and *Inga punctata* formed a drought-resilient group, having a high carbon assimilation and low water loss. Intermediate species (*Heliocarpus appendiculatus*, *Inga vera*, *I. inicuil*) balanced moderate CO₂ assimilation rates with adaptable stomatal response. Photochemical efficiency remained stable across species.

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Shaded *Coffea arabica* var. Oro Azteca had significantly higher leaf nitrogen, moisture, and water-use efficiency than unshaded ones. These differences coincided with lower PAR under shade, aligning with known variations in shaded versus unshaded coffee plants. Principal component analysis showed that PC1 correlated strongly with stomatal conductance and transpiration, driven by *P. guajava* and *P. schiedeana*. PC2 showed a carbon economy trade-off between CO₂ assimilation and internal concentration, dominated by *E. americana*. Collectively, these components highlight stomatal regulation and carbon management as adaptive strategies. Coffee PCA revealed contrasting water-use strategies: PC1 showed inverse stomatal regulation (especially in shaded varieties), and PC2 an energy allocation trade-off between photochemical efficiency and carbon assimilation, with shaded plants maintaining stable CO₂ assimilation regarding unshaded ones. These results demonstrate notable interspecific variation in carbon storage, water-use efficiency, and light conditions among shade trees, offering empirical support for species selection in Veracruz coffee agroforestry.

Subjects Agricultural Science, Plant Science, Climate Change Biology, Natural Resource Management, Environmental Impacts

Keywords Carbon assimilation rates, Stomatal conductance, Water-use efficiency, Photosynthetic photon flux density, Chlorophyll fluorescence, Leaf nitrogen content, Shade tree physiology, *Coffea arabica* varieties, Principal component analysis, Veracruz agroforestry systems

INTRODUCTION

Coffee is one of the most widely traded and consumed agricultural commodities worldwide (FAO, 2022) with 70% produced in Latin America (Baffes, Lewin & Varangis, 2005). Coffee production during 2023 was estimated at 10.1 million tonnes, with an expected growth rate of 5.8% by 2024 (FAO, 2022; ICO, 2023). During 2017–2022, 70% of the world’s coffee production was exported from producing nations to other countries, generating USD 19 billion in revenue and employing 125 million people globally (Panhuysen & Pierrot, 2020; Fairtrade Foundation, 2022).

The coffee industry was valued at US\$132.13 billion in 2024, and globally, there are 12.5 million coffee farms, many located in high-biodiversity zones and managed by smallholders (Donald, 2004; Panhuysen & Pierrot, 2020; Mordor Intelligence, 2024). In Mexico, coffee is produced on approximately 580,000 ha by 481,000 farmers (Ellis et al., 2010; Harvey et al., 2021; USDA FAS, 2023). Approximately 86% of production in the country comes from *Coffea arabica* L. cultivation in shade polycultures that support biodiversity and provide ecosystem services (Beer et al., 1998; Moguel & Toledo, 1999; Dávalos-Sotelo, Morato & Martínez-Pinillos-Cueto, 2008; Jha et al., 2011; Toledo & Moguel, 2012).

Mexican coffee production, which has historically resisted intensification (Rice, 2008), now faces climate-induced land use changes threatening biodiversity (Toledo & Moguel, 2012) and ecosystem services (Beer et al., 1998). Coffee is highly climate-sensitive, with projected range reductions due to shifting temperature, humidity, and rainfall patterns (Bunn et al., 2015; Pham et al., 2019; Bilen et al., 2023). Agroforestry systems mitigate these impacts through shade-regulated microclimates, carbon capture (Verchot et al., 2007;

Noponen et al., 2013; Rahn et al., 2014; Jawo, Kyereh & Lojka, 2022; Terasaki Hart et al., 2023), and soil conservation (*Segura, Kanninen & Suárez, 2006; Lin, 2007; Lin, 2010; Siles et al., 2012; Notaro et al., 2014; Ehrenbergerová, Šenfěldr & Habrova, 2018*), while diversifying income *via* biomass and other attributes of native trees (*Acevedo et al., 1992; Jose, 2009; Häger, 2012; Noponen et al., 2013; Gross et al., 2022*). Optimal shade species should be native to enhance soil fertility and pest control without compromising biodiversity (*Gill & Prasad, 2000; Reigosa et al., 2000; Gliessman, 2015*). The Mexican state of Veracruz exhibits exceptional biodiversity (*Rzedowski, 1978; Estrada-Contreras et al., 2015; Tellez et al., 2020*), making its coffee agroforestry systems particularly valuable for studying carbon sequestration. These systems produce 23% of Mexico's coffee (*Nestel, 1995*) and can store 73.27 Mg C ha⁻¹ in coffee-tall tree combinations (*Ortiz-Ceballos et al., 2020*), retaining 91.2% of forest cover while supporting biodiversity comparable to secondary forests (*Dávalos-Sotelo, Morato & Martínez-Pinillos-Cueto, 2008; Vizcaino-Bravo, Williams-Linera & Asbjornsen, 2020*). This evidence highlights their dual role in climate mitigation and conservation.

Plant photosynthesis is pivotal for CO₂ mitigation through carbon assimilation into biomass (*Fini et al., 2023; Jin et al., 2023*). Accurate carbon sequestration estimates rely on above-ground biomass measurements using allometric equations, yielding realistic estimates of assimilated carbon (C) into forest biomass and, consequently, of carbon cycling in ecosystems (*Liang & Wang, 2020; Araza et al., 2022*). Physiological assessments, particularly leaf gas exchange and chlorophyll fluorescence parameters, offer critical insights into performance and plant acclimation to environmental changes (*Genty, Briantais & Baker, 1989; Sakshaug et al., 1998; Roháček, 2002; Baker & Rosenqvist, 2004; Strasser, Tsimilli-Michael & Srivastava, 2004; Baker, 2008*), enhancing the comprehension of ecosystem carbon dynamics.

Shade trees in agroforestry systems critically regulate microclimatic conditions, affecting the physiological responses of coffee plants and associated tree species. Evidence shows that shade reduces air temperatures by 1–5 °C, alleviating heat stress and maintaining optimal leaf temperatures (20–24 °C) for photosynthesis (*Vaast et al., 2005; Lara-Estrada, Rasche & Schneider, 2023*). These conditions improve chlorophyll fluorescence parameters including Fv/Fm (quantum yield of PSII) by reducing photodamage under excessive irradiance (*Rodríguez-López et al., 2014*). However, shade-adapted leaves typically exhibit lower heat tolerance than sun leaves, as observed in tropical trees like *Inga spectabilis* (Vahl) Willd., with shade leaves exhibiting a reduced threshold for PSII dysfunction (*Slot et al., 2019*). Light availability substantially mediates leaf gas exchange in coffee plants. Shade levels of 30–50% optimise photosynthetic rates through balanced irradiance and photoprotection, whereas excessive shade (>60%) can reduce light-saturated photosynthesis and yield by 10–30% (*DaMatta, 2004; Hagggar et al., 2011; Isaac et al., 2024*). Notably, shade-tree traits such as layered canopies or high leaf nitrogen enhance nutrient cycling and light diffusion, further modifying these physiological responses (*Sauvadet et al., 2019; Isaac et al., 2024*).

Under climate change scenarios, water availability represents a critical challenge for agricultural and natural ecosystems. Water-use efficiency (WUE), calculated in our study as the ratio of CO₂ assimilation to transpiration (CO₂/H₂O) during gas exchange (*Kirkham,*

2005), reflects short-term trade-offs between carbon gain and water loss under controlled conditions and couples plant productivity to water management (Bhattacharya, 2019; Hatfield & Dold, 2019). WUE exhibits interspecific variation according to plant functional traits and environmental conditions (Chaves, Osório & Pereira, 2004; McCarthy, Pataki & Jenerette, 2011). Agroforestry shade enhances WUE through increased ambient humidity (10–20%) and reducing stomatal conductance, although shaded coffee plants frequently exhibit higher mass-specific transpiration due to morphological adaptations like thinner leaves (Lin, 2010; Sarmiento-Soler et al., 2019; de Carvalho et al., 2021; Koutouleas et al., 2022). Collectively, these findings demonstrate shade's role in microclimate stabilisation and photosynthetic optimisation, while highlighting knowledge gaps regarding species-specific physiological trade-offs.

Therefore, the present work examines the ecophysiological characteristics of seven previously selected shade tree species and five *C. arabica* varieties that make up agroforestry systems in central Veracruz, Mexico. For this task, *in-situ* dendrometric measurements of aboveground biomass, chlorophyll fluorescence, gas exchange parameters, and nitrogen and moisture content of leaves located at the understory layer of the selected shade tree species and coffee varieties were conducted under controlled temperature and photosynthetic photon flux density (PPFD) conditions.

We hypothesised that the physiological traits of these seven tree species identified by Flores-Ortiz et al. (2025) and their interactions with coffee varieties growing in shaded environments enhance carbon capture capacity while improving agroforestry system resilience under climate change. These effects are mediated through stress reduction *via* shading and optimised resource-use efficiency. We expect that this information will help decision-making during the selection and management of suitable shade trees based on their ecophysiological characteristics and carbon capture potential in shade coffee agroforestry systems.

MATERIALS AND METHODS

Selection of shade tree species and Arabica coffee varieties in agroforestry systems

Seven native tree species were selected from an initial screening of 50 conducted by Flores-Ortiz et al. (2025) based on conservation status, growth rate, and agroecological utility for coffee production. Species used for firewood or fuel were excluded, along with cultivated *Persea americana* Mill. (Hass avocado) due to its intensive domestication. For the selected shade trees species: *Inga inicuil* Schltdl. and Cham. Ex G. Don (*Ii*); *Inga vera* Willd. (*Iv*); *Inga punctata* Willd. (*Ip*); *Erythrina americana* Mill. (*Ea*); *Psidium guajava* L. (*Pg*); *Persea schiedeana* Turcz. (*Ps*), and *Heliocarpus appendiculatus* Nees (*Ha*), dendrometric, chlorophyll fluorescence, and gas exchange characteristics were measured in trees aged ≤ 30 years.

Due to the heterogeneity of *C. arabica* varieties found in the coffee farms in the region, chlorophyll fluorescence and gas exchange parameters measured were focused on Oro Azteca (Oa), Garnica (G), Costa Rica 95 (Cr), Tipica (T), and Catuai amarillo (Ca)

plants aged 4–6 years. To enable comparison, identical physiological measurements were conducted for Oro Azteca variety plants under unshaded conditions.

This selection aimed to represent a functional diversity of native species with potential for enhancing ecosystem services in coffee farms.

Study area

The shade tree species and coffee varieties studied make up agroforestry systems in a traditional polyculture configuration and in the intermediate secondary succession stage. Coffee plants in unshaded conditions were part of an unshaded monoculture system. The tree density, considering trees between 10–15 m in height and >5 cm in diameter at breast height (DBH, at 1.30 m) for this type of coffee agroforestry system and coffee region, has been reported at $\sim 1,000$ trees ha^{-1} (López-Gómez, Williams-Linera & Manson, 2008; Williams-Linera & Lorea, 2009).

Study sites were located in central Veracruz on shaded and unshaded coffee farms in the municipalities of Teocelo ($19^{\circ}23'36''\text{N}$, $96^{\circ}59'9.4''\text{W}$, at an elevation of 1,117 m a. s. l., average air temperature of 23.43 ± 0.37 °C and $70.4 \pm 5.95\%$ of relative humidity, or RH) and Xico ($19^{\circ}25'23.5''\text{N}$, $96^{\circ}55'42.6''\text{W}$, at an elevation of 1,053 m a. s. l., average air temperature of 27.53 ± 0.86 °C and $60.23 \pm 4.16\%$ of RH for shaded and unshaded conditions, respectively (Fig. 1; Map created using the Free and Open Source QGIS).

To ensure environmental homogeneity, measurements were taken during two consecutive cool-season months (October–November 2022). This period exhibited typical climatic conditions for 2016–2022 trends, with precipitation, temperature, and cloud cover within expected ranges (Weather Spark, 2024). The cool season brought average maxima of 24 °C (Teocelo) and 26 °C (Xico), alongside ≥ 1 mm/h precipitation and 80% cloud cover. Historical records show Teocelo's temperature extremes (21–35 °C, 1945–2020), while Xico ranged from 9–12 °C (minima) to 21–22 °C (maxima, 1966–2023) (Servicio Meteorológico Nacional, <https://smn.conagua.gob.mx/es/climatologia/informacion-climatologica/normales-climatologicas-por-estado?estado=ver> (accessed 17 November 2025)). Both sites share comparable annual precipitation (1,847 and 2,091.8 mm for Xico and Teocelo, respectively; period 1991–2020) and stable conditions due to their proximity.

Sampling strategy

The sampling design was structured to capture intra- and inter-specific ecophysiological variability. For each of the seven shade tree species, three mature and reproductive individuals were randomly selected (individuals = three per species). On each individual, three fully expanded, healthy leaves located on the first lateral branch at the base of the trunk were marked and measured (leaves = three per individual). This yielded in a total of nine measurements per species (measurements = nine). The same protocol was applied to the associated coffee plants under the canopy of each tree species, sampling three individuals per variety and three leaves attached to plagiotropic branches per coffee bush. For statistical analysis, data from the three leaves from the same individual were averaged to obtain a single representative value per individual ($n = 3$ individuals per species/variety). These averages were subsequently used for comparative analyses between species and varieties.

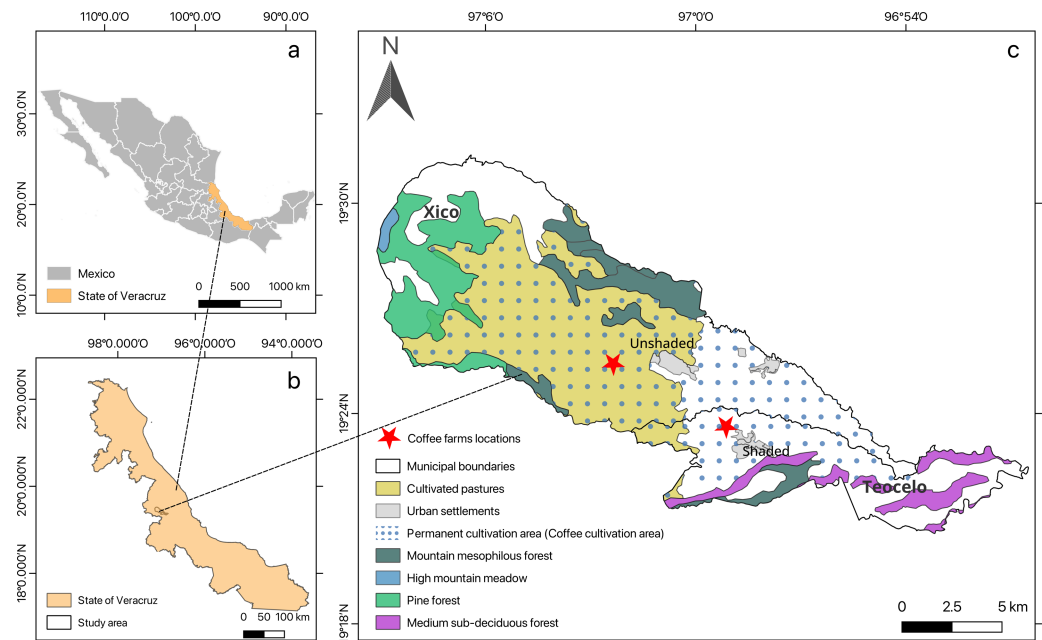


Figure 1 The study area location. (A) Mexico; (B) the state of Veracruz; and (C) the municipalities with the coffee farms where the *in-situ* tests were conducted.

Full-size [DOI: 10.7717/peerj.20255/fig-1](https://doi.org/10.7717/peerj.20255/fig-1)

Dendrometric parameters of shade trees and carbon stocks determination

Measurements were made of the total height and the DBH of three mature and reproductive individuals of each of the seven shade trees. Using these morphometric parameters, the available allometric equations were applied to calculate each tree's above ground biomass or AGB (Table 1). The allometric equations used to calculate the AGBs for *Ha* and *Pg* were at species level, at genus level for *Inga* spp. (*Ii*, *Iv* and *Ip*) and for *Erythrin* a sp. (*Ea*), and at tropical forest level for *Ps* (Rojas-García et al., 2015; Ortiz-Ceballos et al., 2020) (Table 1). AGB was converted to biomass carbon stock (CS) by multiplying by 0.47, representing the standard carbon fraction in tree biomass (IPCC, 2006). CS reflects both tree's ability to grow new cells and its carbon storage potential (IPCC, 2022).

Due to variation in tree age, 10-year normalised CS were obtained by a simple proportionality rule for each tree species. This normalisation enables standardised comparison of carbon storage potential while recognising that species-specific traits, such as shade tolerance and wood density, may influence long-term sequestration rates. Finally, we extrapolated hectare-scale carbon storage using the tree density found in the literature (references in Table 2).

Chlorophyll fluorescence, gas exchange analyses of shade trees and coffee plants

In-situ measurements were performed on leaves from the seven tree species and associated-coffee plants located under the canopy of the seven tree species, which correspond to five varieties of Arabica coffee (Oa, G, Cr, T, and Ca) using a portable infrared gas analyser

Table 1 Dendrometric parameters of shade trees. According with their carbon stock in biomass at 10 years, species are arranged descending. The average of three replicates is presented.

Tree Species	DBH ¹ (m)	Height (m)	Age (years)	Allometric equation	AGB ² (kg tree ⁻¹)	Carbon stock ³ (kg tree ⁻¹)	10-years carbon stock (kg tree ⁻¹)
<i>Persea schiedeana</i> (Ps)	0.35 ± 0.05	14.33 ± 2.08	10.00 ± 4.58	$\text{Exp}((-3.1141) + ((0.9719) * (\text{Ln}(\text{DBH}^2 * H)))$	585.93 ± 124.85	275.38 ± 58.68	275.38 ± 58.68
<i>Erythrina Americana</i> (Ea)	0.58 ± 0.05	11.67 ± 2.02	30.00 ± 0.00	$[0.3700] * [\text{DBH}^{1.9600}]$	1,054.91 ± 175.26	495.81 ± 82.37	165.27 ± 27.46
<i>Inga inicuil</i> (Ii)	0.24 ± 0.00	12.00 ± 3.46	10.00 ± 0.00	$[\text{Exp}[-1.76] * [\text{DBH}^{2.26}]]$	223.29 ± 3.46	104.95 ± 1.63	104.95 ± 1.63
<i>Heliocarpus appendiculatus</i> (Ha)	0.37 ± 0.02	15.67 ± 0.58	20.00 ± 0.00	$[[\text{Exp}[4.9375]] * [[\text{DBH}^2]^{\wedge 1.0583}]] * [1.14]/1000$	331.76 ± 30.59	155.92 ± 14.38	77.96 ± 7.19
<i>Psidium guajava</i> (Pg)	0.08 ± 0.01	5.00 ± 0.00	6.00 ± 1.73	$[0.246689] * [\text{DBH}^{2.24992}]$	28.25 ± 6.71	13.28 ± 3.15	22.13 ± 5.25
<i>Inga vera</i> (Iv)	0.10 ± 0.02	5.00 ± 1.00	11.67 ± 2.89	$[\text{Exp}[-1.76] * [\text{DBH}^{2.26}]]$	30.00 ± 13.21	14.10 ± 6.21	12.09 ± 5.32
<i>Inga punctata</i> (Ip)	0.07 ± 0.00	4.00 ± 0.00	13.33 ± 5.77	$[\text{Exp}[-1.76] * [\text{DBH}^{2.26}]]$	15.33 ± 0.78	7.20 ± 0.37	5.40 ± 0.27

Notes.

¹Diameter at breast height (1.30 m).

²Above-ground biomass (AGB).

³The carbon stock was determined by multiplying the calculate AGBs by the default carbon fraction of 0.47 set by the *IPCC (2006)*.

Table 2 Carbon capture of shade tree species. Estimated carbon stock per hectare using reported densities of the target shade trees species.

Tree Species	≈Trees ha ⁻¹	Carbon stock per hectare (Mg C ha ⁻¹)	Reference (Trees ha ⁻¹)
<i>Persea schiedeana</i> (Ps)	40	11.0152	<i>Soto-Pinto et al. (2001)</i>
<i>Erythrina Americana</i> (Ea)	40	19.8324	<i>Soto-Pinto et al. (2001)</i> and <i>Garza-Lau et al. (2020)</i>
<i>Inga inicuil</i> (Ii)	200	20.99	<i>Barradas & Fanjul (1986)</i>
<i>Heliocarpus appendiculatus</i> (Ha)	40	6.2368	<i>Soto-Pinto et al. (2001)</i> and <i>Romero-Alvarado et al. (2002)</i>
<i>Psidium guajava</i> (Pg)	40	0.5312	<i>Somarriba (1988)</i> and <i>Akter et al. (2022)</i>
<i>Inga vera</i> (Iv)	200	2.82	<i>Garza-Lau et al. (2020)</i>
<i>Inga punctata</i> (Ip)	100	0.72	<i>Valencia et al. (2014)</i> and <i>Soto-Pinto et al. (2001)</i>
Total	660	62.1456	

(IRGA) LI-6400XT (Licor, Lincoln, NE, USA) equipped with a fluorometric cell. For shade trees, we assessed leaves attached to lateral branches closest to the understory layer, between 2–4 m from the ground *via* ladders and climbing equipment. These branches exhibited sympodial growth with a predominantly horizontal or obliquely orientated architecture, characteristic of species such as *Inga* spp. (Troll model), *Ea* (Champagnat model), and *Pg* (Roux model) (*Vester, 2002; de Reffye et al., 2008*). Coffee measurements were taken from leaves attached to plagiotropic branches at 1.30–2 m height.

Chlorophyll fluorescence analysis was performed in dark-adapted leaf tissues (30 min). Minimum (F₀) and maximum fluorescence (F_m) in light-adapted tissues was measured by the saturation pulse method ($\lambda = 630$ nm, $Q > 7,000 \mu\text{mol m}^{-2} \text{s}^{-1}$, 6s). Based on these signals, variable fluorescence in the dark (F_v=F_m-F₀) and quantum efficiency (F_v/F_m) were calculated (*Silva et al., 2010; Rakocevic et al., 2022*). F_v/F_m is frequently used to estimate the photochemical efficiency of PSII (*Niinemets & Kull, 2001; Lepeduš et al., 2005; Zavafer & Mancilla, 2021*). After that, the gas exchange parameters CO₂ assimilation rate, stomatal conductance, transpiration, and intercellular CO₂ (C_i) were measured in the same leaves attached to the branch. All measurements were conducted between 9:00 h and 11:00 h under controlled conditions, with the IRGA operated as an open system with a photon flux density of $1,000 \mu\text{mol m}^{-2} \text{s}^{-1}$, a leaf temperature of 25 °C, and an environmental air CO₂ concentration of ~420 ppm.

Subsequent gas exchange measurements, it proceeded to estimate incident solar radiation in the understory layer between shaded and unshaded systems. PAR was measured under all tree species' canopies and sun-exposed Oro Azteca coffee plants at midday; this period sees peak solar radiation and thermal stress (*Meili et al., 2021; Kohl, Niether & Abdulai, 2024*).

Instantaneous leaf water-use efficiency (WUE = A/E) was calculated as the CO₂ assimilation-transpiration ratio (*Hatfield & Dold, 2019*).

Gravimetric and analytical chemical analysis: moisture and nitrogen content of coffee leaves samples

Following immediately after chlorophyll fluorescence and leaf gas exchange measurements, the same leaves from Oro Azteca coffee plants grown under both shaded and unshaded conditions were excised at the base of the petiole. Collected leaves were stored in sealed plastic containers to prevent moisture loss until further analysis. Subsequently, the samples were processed within the next two days after field sampling. The leaves from each condition were macerated and pooled, then 0.5 g was taken for moisture and nitrogen analysis. Gravimetric methods comparing weight before and after drying determined moisture content based on the following equation:

$$H(\%) = \frac{FW - DW}{FW} \times 100. \quad (1)$$

This parameter refers to the proportion of water present in fresh leaf tissue relative to its total weight, where H (%) is the moisture content expressed as a percentage, FW is the fresh weight, and DW is the dry weight.

We employed the semimicro-Kjeldahl method (Nelson & Sommers, 1980; DOF, 2019) for digested 50 mg of dried leaf material using a digestion/distillation apparatus (Labconco®) to quantify organic nitrogen content.

Statistical analysis

Statistical analyses were performed on the averaged data from nine measurements (described above). The assumption of normality (Shapiro–Wilk, $p > 0.01$) was verified for all datasets. All data met the assumption; thus, no transformations were required.

One-way analysis of variance (ANOVA) was used to identify significant differences ($p < 0.001$) among shade tree species for each parameter. For comparisons involving coffee varieties under different shade conditions, a one-way ANOVA was also employed ($p < 0.001$). Where ANOVA indicated significant differences, the *post-hoc* Tukey's test was applied for pairwise comparisons. Comparisons were conducted for each parameter between all shade tree species. Coffee variety comparisons were performed between the Oro Azteca variety in shaded conditions vs unshaded conditions. Additionally, comparisons were conducted considering only coffee varieties in shaded conditions. WUE statistical differences between shade trees and between coffee varieties were identified following the same methodology ($p < 0.001$). Comparisons of Oro Azteca moisture and nitrogen content between shaded and unshaded conditions were performed using a two-tailed Student's *t*-test ($p < 0.01$) for independent samples.

To analyse the possible clustering of the seven shade tree species and coffee varieties based on their physiological traits, a principal component analysis (PCA) was conducted on both datasets comprising all measured parameters (Fv/Fm, CO₂ assimilation rate, stomatal conductance, transpiration rate, and intercellular CO₂ concentration). PCA analysis proceeded by extracting principal components from the correlation matrix of the variables, with the selection criterion being the retention of components that collectively explained at least 75% of the total variance.

GraphPad Prism[®] version 9.5.1 for macOS (GraphPad Software, San Diego, CA, USA; <http://www.graphpad.com>) was used for all statistical analysis (accessed in January 2023).

RESULTS

Dendrometric parameters of shade trees and carbon stocks determination

Ea and *Ps* showed the highest mean AGB and CS values per tree, while *Ha* and *Ii* displayed intermediate levels, and *Iv*, *Pg*, and *Ip* had the lowest (Table 1). The 10-year normalized CS values showed similar patterns across species, where *Ps* and *Ea* had the highest carbon storage capacities, followed by lower values for the remaining species (Table 1). At the hectare scale, *Ii* achieved the highest annual CS (20.99 Mg C ha⁻¹), followed by *Ea* (19.83 Mg C ha⁻¹), *Ps* (11.01 Mg C ha⁻¹), and *Ha* (6.23 Mg C ha⁻¹), with *Iv*, *Ip* and *Pg* showing the lowest values (0.72 and 0.53 Mg C ha⁻¹, respectively) (Table 2).

Chlorophyll fluorescence, gas exchange and PCA of shade trees and coffee plants

The Fv/Fm values of shade tree species (range 0.74–0.81) and coffee plants (0.73–0.80) showed consistent trends, with no significant differences observed (Table S1). Similarly, no significant differences were found between shaded and unshaded Oa or among other coffee varieties under shaded conditions (Table S2).

CO₂ assimilation rates differed significantly among species ($F_{(6,56)} = 118.3$, $p < 0.001$), with highest values in *Ea* (8.37 ± 0.90 μmol CO₂m²/s), *Pg* (9.18 ± 1.00), and *Ha* (7.99 ± 1.16), and lowest in *Ps* (2.45 ± 0.40), *Iv* (2.21 ± 1.19), and *Ii* (1.67 ± 0.38) (Fig. 2B, Table S1). For Oa coffee, shaded plants showed higher rates than unshaded ($F_{(3,32)} = 54.42$, $p < 0.001$), with *Ip*-Oa (5.68 ± 0.65) and *Ha*-Oa (5.76 ± 0.91) exhibiting the highest values among shaded varieties ($F_{(6,56)} = 16.82$, $p < 0.001$) (Figs. 3A, 4A, Table S2).

Transpiration rates varied significantly ($F_{(6,56)} = 65.43$, $p < 0.001$), with highest values in *Pg* (2.59 ± 0.22 mmol H₂O m²/s) and *Ha* (2.34 ± 0.33), and lowest in *Ip* (1.15 ± 0.29), *Ea* (0.71 ± 0.19), and *Ps* (0.48 ± 0.10) (Fig. 2B, Table S1). Among Oa coffee variety, unshaded Oa (0.53 ± 0.04) and *Ip*-Oa (0.53 ± 0.30) showed highest transpiration, while *Ha*-Oa (0.30 ± 0.18) and *Ps*-Oa (0.23 ± 0.02) showed lowest ($F_{(2,32)} = 6.488$, $p < 0.001$) (Fig. 3B, Table S2). Among shaded coffee varieties, *Ii*-G showed the highest transpiration rate (1.02 ± 0.25) while *Pg*-Ca the lowest (0.27 ± 0.06) (Fig. 4B, Table S2).

Stomatal conductance followed similar patterns, with highest values in *Pg* (266.88 ± 29.01 mmol H₂O m²/s), *Iv* (245.44 ± 85.25), and *Ha* (224.88 ± 35.51), and lowest in *Ps* (26.57 ± 12.98) with significant differences between them ($F_{(6,56)} = 19.33$, $p < 0.001$) (Fig. 2C, Table S1). For the Oa cultivar, stomatal conductance varied significantly between unshaded conditions and shaded *Ha*-Oa and *Ps*-Oa ($F_{(3,32)} = 8.697$, $p < 0.001$) (Fig. 3C, Table S2). For shaded coffee, *Iv*-Cr (145.70 ± 67.86) and *Ii*-G (100.17 ± 34.05) showed highest conductance, while *Pg*-Ca (37.76 ± 24.04), *Ip*-Oa (31.65 ± 11.74), and *Ha*-Oa (15.60 ± 4.06) showed lowest ($F_{(6,56)} = 24.02$, $p < 0.001$) (Fig. 4C, Table S2).

C_i values among shade trees exhibited notable variations, with the highest values observed in *Iv* (444 ± 17.34 μmol CO₂/mol), *Ha* (427.44 ± 12.97), and *Ii* (419.78 ± 41.88) and

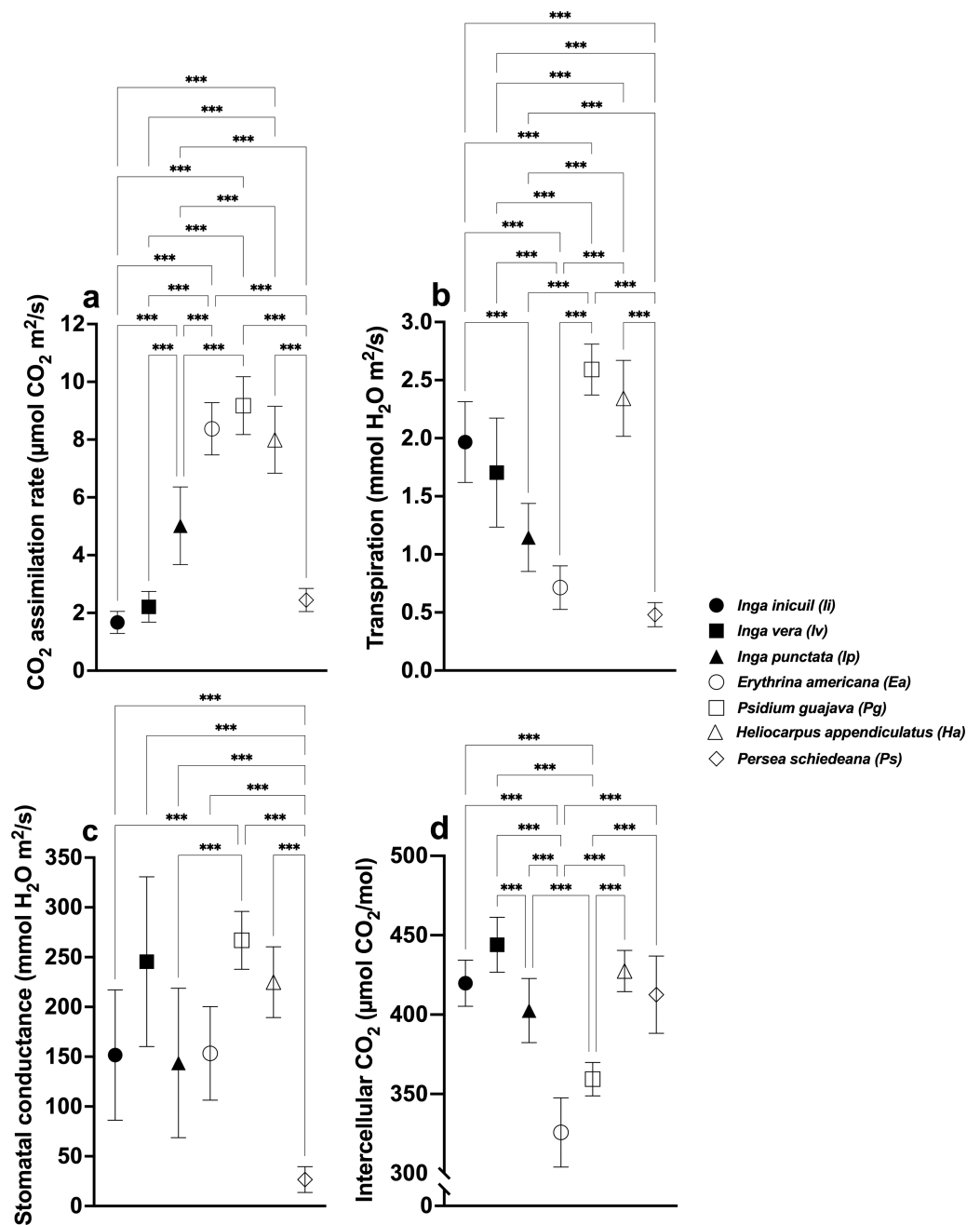


Figure 2 Stomatal parameters of shade trees. (A) CO₂ assimilation rate; (B) Transpiration rate; (C) Stomatal conductance; and (D) Intercellular CO₂. Mean \pm SD ($n = 9$). One-way ANOVA and a *post-hoc* Tukey's test were used to identify significant differences. Comparisons with a p values ≤ 0.001 (***) are depicted.

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lower values in *Pg* (359.33 ± 10.54) and *Ea* (325.78 ± 21.76), with significant differences between them ($F_{(6,56)} = 48.22$, $p < 0.001$) (Fig. 2D, Table S1). For the *Oa* variety, C_i values were influenced by shaded and unshaded conditions, with *Ip*-*Oa* showing the

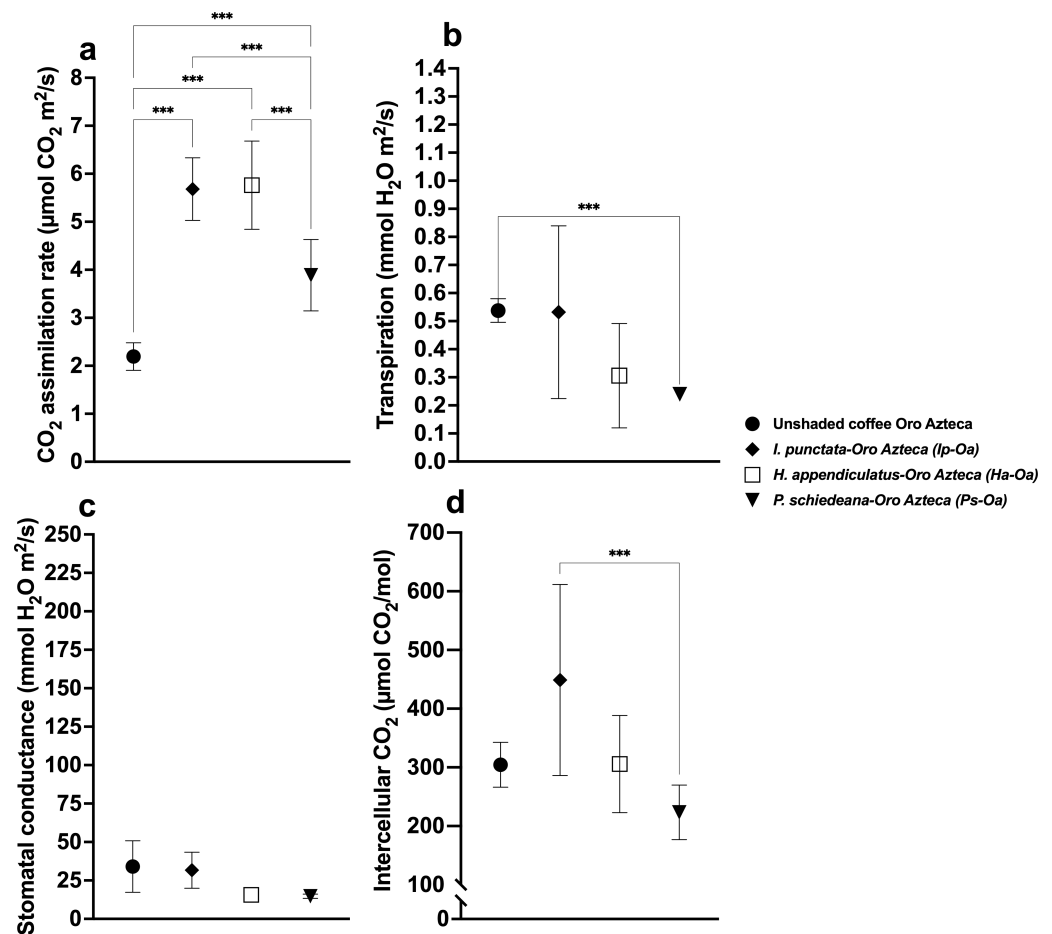


Figure 3 Stomatal parameters of unshaded and shaded coffee plants of the Oro Azteca variety. Shade tree species-coffee associations are indicated in each case. (A) CO₂ assimilation rate; (B) Transpiration rate; (C) Stomatal conductance; and (D) Intercellular CO₂. Mean \pm SD ($n = 9$). One-way ANOVA and a *post-hoc* Tukey's test were used to identify significant differences. Comparisons with a p value ≤ 0.001 (***) are depicted.

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highest values (448.77 ± 82.59) and *Ps*-Oa the lowest (223.44 ± 46.47) ($F_{(6,56)} = 48.22$, $p < 0.001$) (Fig. 3D, Table S2). Among shaded coffee varieties, *Ip*-Oa (448.77 ± 82.59), *Iv*-Cr (443.33 ± 162.69), and *Ii*-G (410.55 ± 26.82) demonstrated higher C_i values, while *Ea*-T (284.88 ± 34.46), *Pg*-Ca (284.88 ± 34.46), *Ha*-Oa, (305.66 ± 82.79) and *Ps*-Oa (223.44 ± 46.47) showed lower values, showing significant differences between them ($F_{(6,56)} = 11.89$, $p < 0.001$) (Fig. 4D, Table S2).

PAR levels differed significantly among shade tree species ($F_{(6,56)} = 64.15$, $p < 0.001$), ranging from $23.3 \pm 4.06 \mu\text{mol m}^{-2} \text{s}^{-1}$ (*Ps*) to 92.2 ± 6.66 (*Ea*) (Table S1). For Oa coffee variety, unshaded coffee received $1,427 \pm 124.0 \mu\text{mol m}^{-2} \text{s}^{-1}$ versus 8.44 (*Ps*-Oa)– 25.11 (*Ip*-Oa) under shade ($F_{(6,56)} = 47.68$, $p < 0.001$) (Table S2). Among shaded coffee varieties, *Iv*-Cr (47.1 ± 9.18) and *Pg*-Ca (32.2 ± 5.38) displayed the highest PAR values, while *Ps*-Oa

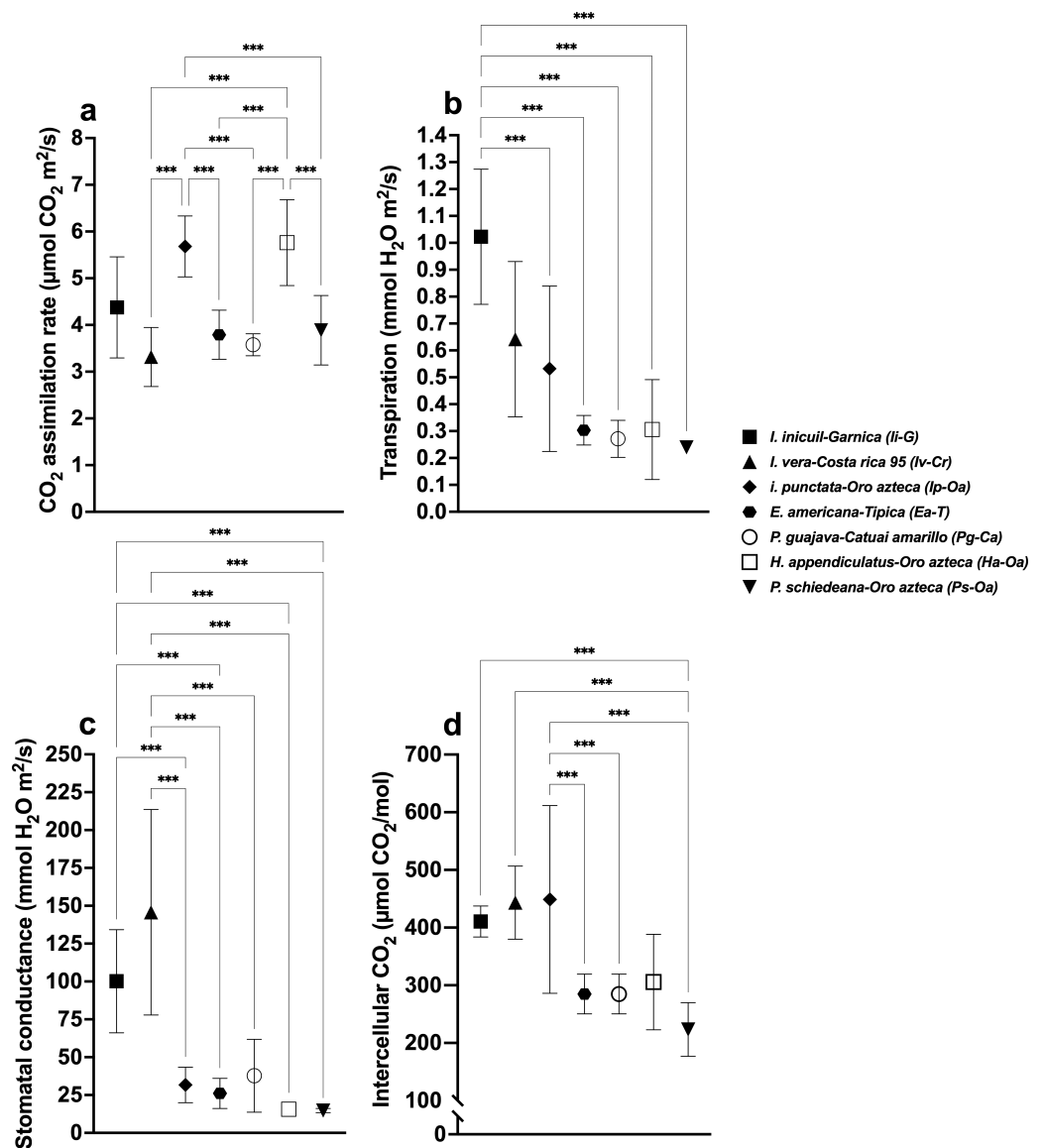


Figure 4 Stomatal parameters of the different *Coffea arabica* varieties in shaded condition. Shade tree species-coffee variety associations are indicated in each case. (A) CO_2 assimilation rate; (B) Transpiration rate; (C) Stomatal conductance; and (D) Intercellular CO_2 . Mean \pm SD ($n = 9$). One-way ANOVA and a *post-hoc* Tukey's test were used to identify significant differences. Comparisons with a p value ≤ 0.001 (***) are depicted.

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(8.44 ± 1.08) and *Ha-Oa* (14.8 ± 6.64) showed the lowest measurements, with significant differences between conditions ($F_{(6,56)} = 47.68$, $p < 0.001$) (Table S2).

WUE values showed significant variation among the seven tree species, ranging from $0.83 \pm 0.15 \mu\text{mol CO}_2/\text{mmol H}_2\text{O}$ (*Ii*) to $16.92 \pm 11.05 \mu\text{mol CO}_2/\text{mmol H}_2\text{O}$ (*Ea*) ($F_{(6,56)} = 5.810$; $p < 0.001$) (Fig. 5A, Table S1). For the *Oa* coffee variety, WUE values ranged from $4.61 \pm 10.06 \mu\text{mol CO}_2/\text{mmol H}_2\text{O}$ in unshaded conditions to $21.62 \pm 11.27 \mu\text{mol CO}_2/\text{mmol H}_2\text{O}$ (*Ha-Oa*) ($F_{(3,32)} = 8.538$; $p < 0.001$) (Fig. 5B, Table S2). Among

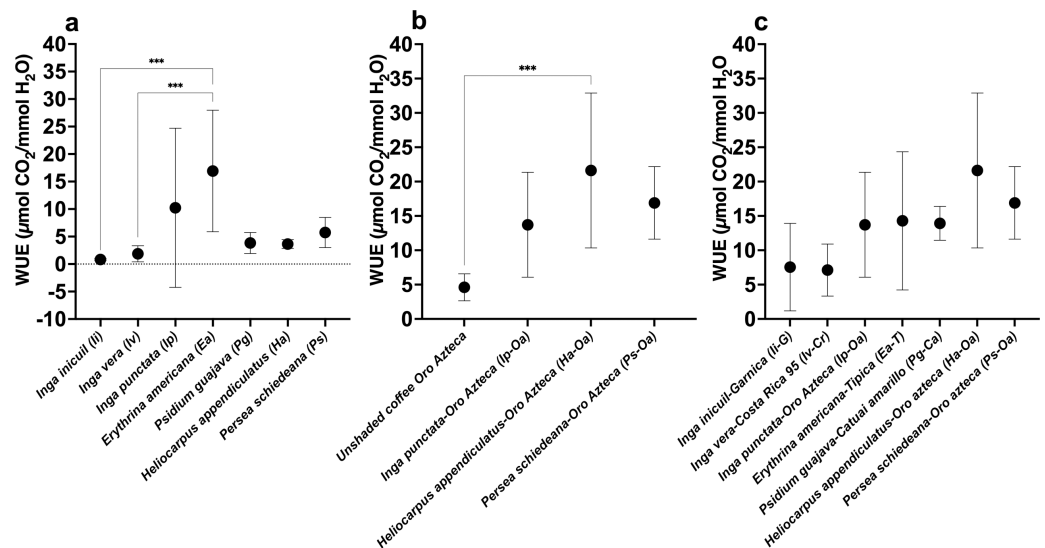


Figure 5 Instantaneous leaf water-use efficiency (WUE) of shade trees and coffee varieties in unshaded and shaded conditions. (A) Shade tree species; (B) Oro Azteca coffee variety; and (C) Shaded Arabica coffee varieties. Mean \pm SD ($n = 9$). One-way ANOVA and a *post-hoc* Tukey's test were used to identify significant differences. Comparisons with a p value ≤ 0.001 (***) are depicted.

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shaded Arabica cultivars, values varied from $7.12 \pm 3.78 \mu\text{mol CO}_2/\text{mmol H}_2\text{O}$ (*lv-Cr*) to $21.62 \pm 11.27 \mu\text{mol CO}_2/\text{mmol H}_2\text{O}$ (*Ha-Oa*) (Fig. 5C, Table S2); no statistically significant differences were observed.

For the PCA for the seven shade tree species, PC1 accounted for 43.57% of the variance (eigenvalue = 2.178), while PC2 contributed 33.43% (eigenvalue = 1.671), cumulatively explaining 77% of the dataset variability (Table S3). In contrast, the PCA for the coffee varieties (including Oro Azteca) under shaded and unshaded conditions revealed a stronger influence of PC1, which explained 49.04% of the variance (eigenvalue = 2.452), with PC2 adding 29.98% (eigenvalue = 1.499), resulting in a slightly higher cumulative variance (79.02%) (Table S4).

For the seven shade tree species, PC1 was strongly associated with stomatal conductance (loading = 0.882) and transpiration (0.866), with *Pg* and *Ps* contributed most to PC1 (42.5% and 37.1%, respectively) (Fig. 6A, Table S3). PC2 showed positive loading for CO_2 assimilation rate (0.594) and negative loading for intercellular CO_2 concentration (-0.937), with *Ea* contributing 73.9% to this axis (Fig. 6A, Table S3).

For coffee varieties, PC1 had negative loadings for stomatal conductance (-0.906), transpiration (-0.897), and intercellular CO_2 (-0.884) (Fig. 6B; Table S4), with *Ps-Oa* (22.0%) and *Ha-Oa* (5.9%) showing highest contributions (Table S4). PC2 showed positive loading for photochemical efficiency ($\text{Fv/Fm} = 0.766$) and negative loading for CO_2 assimilation rate (-0.884) (Fig. 6B, Table S4), with unshaded Oro Azteca contributing 39.3% (Table S4).

Correlation analyses for shade trees, stomatal conductance, and transpiration were strongly positively linked ($r = 0.825$), while CO_2 assimilation and intercellular CO_2 were

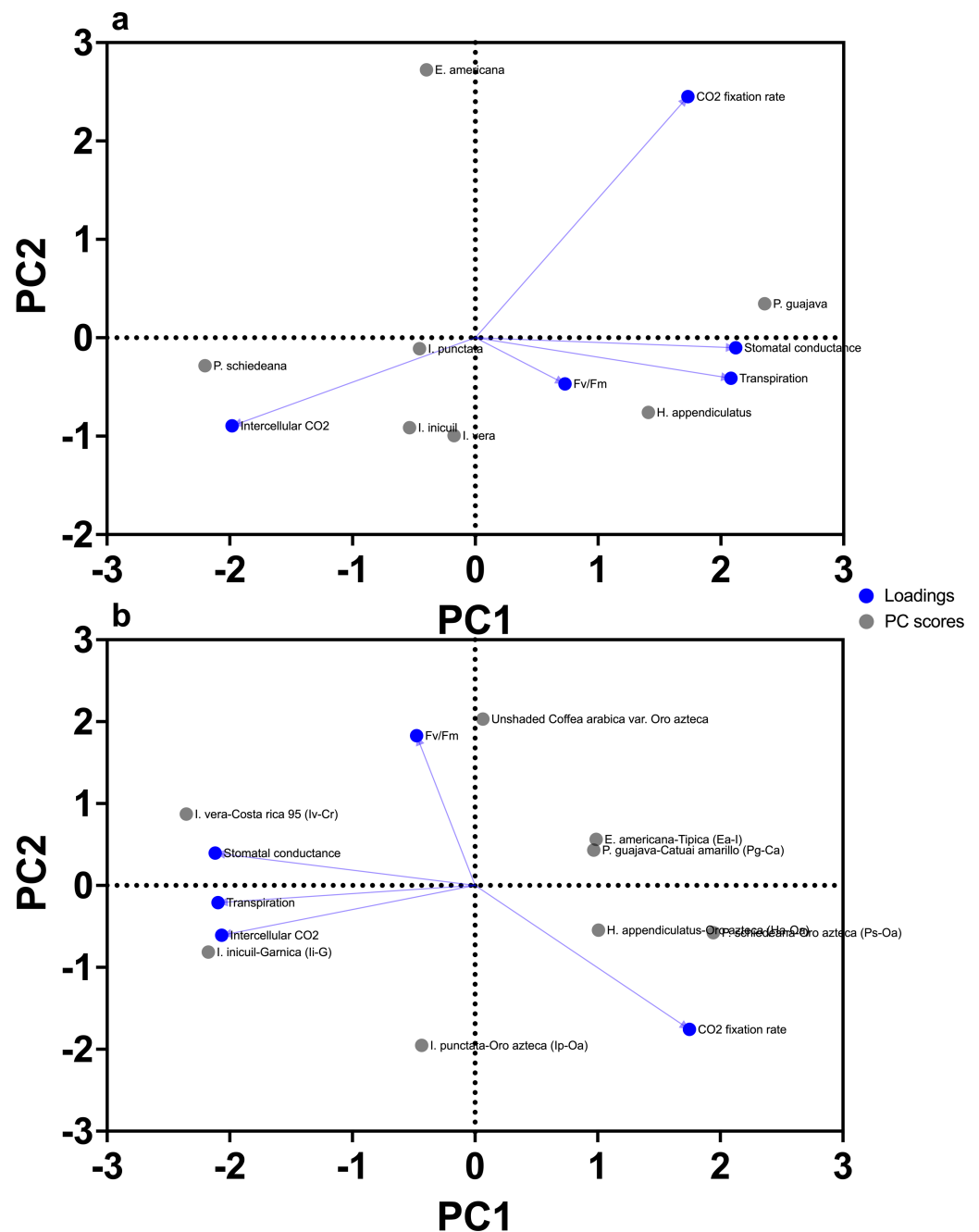


Figure 6 Principal component analysis (PCA) of chlorophyll fluorescence and gas exchange parameters. (A) shade tree species; and (B) Arabica coffee varieties. PC1 represents 43.57% and 49.04% of the total variation for shade trees and Arabica coffee varieties, respectively, while PC2 represents 33.43% and 29.98% of the total variation for shade trees and Arabica coffee varieties, respectively (77% and 79.02% of the total variance for shade trees and Arabica coffee varieties, respectively). Correlation of variables with PCA axes is indicated by blue solid line vectors.

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negatively correlated ($r = -0.689$) (Table S3). For coffee varieties, stomatal conductance, transpiration, and intercellular CO_2 showed strong positive correlations ($r = 0.698$ – 0.731), whereas CO_2 assimilation and F_v/F_m were moderately negatively associated ($r = -0.420$) (Table S4).

Gravimetric and analytical chemical analysis: moisture and nitrogen content of coffee leaves samples

The shaded Oa samples showed a slightly higher moisture content (55.56%) compared to the unshaded samples (53.97%) (Table S5). Also, shaded Oa samples exhibited a significantly higher nitrogen content ($2.83 \pm 0.06\%$) compared to unshaded samples ($2.54 \pm 0.02\%$) ($t(4) = 7.6$; $p < 0.01$) (Table S5).

DISCUSSION

Our study provides a novel, ecophysiology-based framework for selecting shade trees in coffee agroforestry systems by moving beyond traditional ecological or yield-based criteria. We present the first integrated analysis of *in-situ* carbon capture potential, photosynthetic performance, and water-use strategies for seven previously shortlisted native shade tree species and five *Coffea arabica* varieties in Veracruz, Mexico. By combining dendrometric assessments with controlled light and temperature measurements of chlorophyll fluorescence and leaf gas exchange, we unveil distinct functional groups and species-specific trade-offs between carbon assimilation and water conservation. Furthermore, we quantitatively demonstrate how shade mediates key leaf traits in coffee, such as nitrogen and moisture content, leading to improved water-use efficiency. Our findings showed that native shade tree species contributed differentially to carbon capture in coffee agroforestry systems, supporting our hypothesis that their physiological traits enhance both carbon sequestration and system resilience through shading and optimised resource-use efficiency. This work offers empirically supported criteria for designing climate-resilient agroforestry systems that optimise synergies between carbon sequestration, microclimate regulation, and crop physiology.

Dendrometric parameters of shade trees and carbon stocks determination

The seven shade tree species showed distinct photosynthetic traits, growth and biomass allocation patterns, leading to inherent variability in carbon accumulation, making direct interspecific comparisons of AGB and CS unreliable (Poorter *et al.*, 2008; Chave *et al.*, 2014). Works such as that of Garza-Lau *et al.* (2020) showed these types of variations in agroforestry systems in the state of Veracruz. Farmers typically manage tree density through propagation programmes, though optimal balances between competition and productivity need further study. Regional factors like altitude, slope, and shade management affect phenology and influence tree traits and ecosystem services (Bewley & Black, 1994; Cerda *et al.*, 2017; Asanok *et al.*, 2024). These constraints and cultivation practices collectively determine tree densities. Coffee plants carbon sequestration in the study area, although it was not determined in our work, is highly variable (0 – 12 Mg ha^{-1} ; Valdés-Velarde *et al.*,

2022), depending on variety and density, making extrapolation and comparison difficult, even within the same study area.

Three ecological patterns emerge when examining species contributions. First, the *Inga* genus (Fabaceae), particularly *I. inicuil*, achieves remarkable carbon capture through numerical dominance rather than individual tree performance. With 660 trees ha⁻¹, *I. inicuil* accounts for 33.77% of system carbon uptake, achieving 198 Mg C ha⁻¹, nearly double the sequestration reported for other *Inga*, *Erythrina*, and *Musaceae* species (91.64–115.5 Mg C ha⁻¹; [Haber, 2001](#)). Reported densities for *I. inicuil* range from 100 to 800 trees ha⁻¹, averaging 250–350 trees ha⁻¹ ([Barradas & Fanjul, 1986](#); [Soto-Pinto et al., 2001](#)). After 10 years of growth, *I. inicuil* exhibited a CS of 20.9 Mg C ha⁻¹ at 200 trees ha⁻¹, exceeding values for *Inga densiflora* Benth. (24.3 Mg C ha⁻¹ at 400 trees ha⁻¹) with similar age and size parameters ([Salazar-Figueroa, 1985](#); [Kurstén & Burschel, 1993](#)). However, these values were three times lower than those reported for *I. inicuil* in Oaxaca, Mexico (64.3 Mg C ha⁻¹ at 164 trees ha⁻¹; [Hernández-Vásquez et al., 2012](#); [Alessandrini et al., 2011](#); [Tellez et al., 2020](#)), underscoring how regional factors like altitude and microclimate interact with species physiology. When comparing agroforestry systems across different states, it is important to consider that variations in management practices, soil characteristics, and climatic regimes may lead to substantial differences in carbon sequestration. Consequently, direct comparisons between regions should be interpreted with caution, as local variables shape ecosystem functioning. Although *I. punctata* had lower AGB and CS, its high density (representing 20–40% of total trees) contributed significantly to system-level carbon capture ([Soto-Pinto et al., 2001](#)). Incorporating both *I. punctata* and *I. vera* in agroforestry configurations may achieve carbon stocks of 91.64 Mg C ha⁻¹ ([Haber, 2001](#)).

Second, *E. americana* and *P. schiedeana* follow a quality-over-quantity approach. Their substantial trunk diameters and heights enable just 40 trees to capture carbon equivalent to 94.48% and 52.47%, respectively of the carbon captured by 200 *I. inicuil* trees. However, biological constraints, including seed dormancy in *E. americana* ([Bewley & Black, 1994](#); [Bonfil-Sanders, Cajero-Lázaro & Evans, 2008](#)) and extensive crown-canopy development in *P. schiedeana* ([Niembro, 1992](#); [Vázquez-Torres, Campos-Jiménez & Juárez-Fragoso, 2017](#)), naturally restrict their planting densities in managed agroforestry systems.

Third, the complementary roles of remaining species enhance system functionality. *H. appendiculatus*, representing 16–20% of tree strata in Chiapas coffee farms ([Soto-Pinto et al., 2001](#); [Castillo-Capitán et al., 2014](#)), accounted for 10.03% of total carbon assimilation despite representing only 6% of trees in this study. *P. guajava*, which constituted 4–5% of tree density in coffee plantations ([Soto-Pinto et al., 2001](#)), showed the lowest CS values (6.53% of total CS at 10 years), consistent with prior findings ([Nava et al., 2009](#)) and attributable to its average height of 3–8 m ([Heuzé et al., 2017](#)). *P. guajava* can be incorporated into agroforestry systems to enhance carbon storage, particularly in leaves and roots, with a whole calculated CS ranging between 0.27 and 4.19 Mg ha⁻¹ in 2- to 10-year-old orchards ([Naik et al., 2021](#)). Additionally, it provides valuable firewood and fruits ([Somarriba, 1988](#); [Pascarella et al., 2000](#); [Miceli-Méndez, Ferguson & Ramírez-Marcial, 2008](#)).

Our calculated carbon sequestration of shade tree species was highly specific. *I. inicu* achieved system-level dominance through high density, whereas *E. americana* and *P. schiedeana* excelled through high individual tree efficiency. Optimising carbon capture therefore requires strategic species combinations that leverage these complementary ecological patterns and management practices.

Chlorophyll fluorescence, gas exchange and PCA of shade trees and coffee plants

The physiological performance among shade trees and coffee varieties revealed distinct functional strategies shaped by interspecific variation in quantum efficiency, stomatal behaviour, and carbon capture potential. Most species maintained Fv/Fm values above the 0.75 threshold for fully functional PSII ([Genty, Briantais & Baker, 1989](#); [Lepeduš et al., 2005](#)), indicating robust photochemical activity. This divergence underscores how intrinsic physiological traits interact with environmental conditions to determine carbon capture efficiency.

PCA identified three clusters among shade tree species, revealing distinct adaptive strategies:

Stomatal-regulating shade tree species: *P. guajava* and *P. schiedeana*

These exhibited high stomatal conductance and transpiration, dominating PC1 (43.57% variance) with strong loadings (0.882 and 0.866, respectively) and contributions of 42.5% and 37.1%. These high values suggest a prioritisation of carbon assimilation over water loss in these species. [Nava et al. \(2009\)](#) observed a peak CO₂ assimilation at midday followed by an evening decline in *P. guajava* aligning with our observed C_i values. Shaded conditions enhance carbon assimilation ([Idris et al., 2019](#)), though drought vulnerability suggests limited climate adaptability ([Maxwell & Johnson, 2000](#); [Simonin, Limm & Dawson, 2012](#)). Conversely, *P. schiedeana* showed consistently low stomatal conductance and CO₂ assimilation under moderate temperatures (23.43 °C) and high humidity (70.4%). Its limited assimilation coincided with elevated C_i, as has been observed in *P. americana* ([Useche-Carrillo et al., 2022](#)), suggesting saturation kinetics where stomatal closure halts photosynthesis despite available CO₂ ([Sánchez-Díaz & Aguirreolea, 2008](#); [Fricker & Willmer, 2012](#)). While *P. schiedeana*'s low carbon gain limits competitiveness under high VPD, its efficiency in stable, humid microclimates supports its agroforestry roles.

Key divergences emerge in their climate adaptations: *P. guajava*'s diurnal efficiency suits controlled-light systems, whereas *P. schiedeana*'s saturation-prone physiology demands stable humidity. Their contrasting water-use strategies and light-responsive stomata in *P. guajava* ([Idris et al., 2019](#)) versus humidity-dependent conductance in *P. schiedeana* highlight species-specific trade-offs between productivity and resilience.

Carbon-conserving shade tree species: *E. Americana*

This specie exemplified a carbon-conserving strategy, dominating PC2 (33.43% variance) with a 73.9% contribution and highlighting a trade-off between CO₂ fixation (0.594 loading) and C_i (−0.937). It exhibited high CO₂ assimilation rates despite suboptimal Fv/Fm values (<0.75) and lower stomatal conductance. Such traits align with observations

in other *Erythrina* species, where light-saturated photosynthesis couples with high water-use efficiency (Nygren, 1995; Davis & Hidayati, 2019). This behaviour reflects adaptation to low-VPD conditions, where elevated leaf water potential enhances stomatal efficiency (Running, 1976; Dai, 2013; Grossiord et al., 2020). This adaptive mechanism enhanced leaf water potential and assimilation efficiency, contrasting with *Inga* species, which maintained slower stomatal closure (Engineer et al., 2016; Xu et al., 2016). Its pioneer ecology and optimal temperature range ($\sim 28^\circ\text{C}$; García-Mateos, Soto-Hernández & Vibrans, 2001; Palma-García & Gonzales-Rebeles Islas, 2018) support climate resilience, with reduced transpiration under high temperature/radiation enhancing water conservation. WUE analysis revealed reduced transpiration under high temperature and radiation conditions, prioritising water conservation, a trait advantageous for drought-prone agroforestry.

Intermediate species: *I. inicuil*, *I. vera*, *I. punctata*, and *H. appendiculatus*

This group showed balanced traits across both PCs. *I. inicuil*, *I. vera*, and *I. punctata* exhibited lower photosynthetic rates (1.67 ± 0.38 to $5.02 \pm 1.34 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$) than other *Inga* species under controlled conditions (10.60 – $11.65 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$; Dos Santos Pereira et al., 2019), likely due to differences in measurement conditions, such as mature leaves versus younger leaves, light intensity, and temperature. All three species reduced stomatal opening under moderate temperatures (23.43°C) and high humidity (70.4%), limiting water loss more than carbon fixation (Shimshi & Ephrat, 1975). Under elevated temperatures, their slower stomatal closure allowed sustained transpiration (Engineer et al., 2016; Xu et al., 2016), albeit at the cost of reduced WUE. In contrast, *H. appendiculatus* (Malvaceae), a pioneer species, showed higher assimilation (3.7 – $11.6 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$) as has been seen across light regimes (Tinoco-Ojanguren & Percy, 1995), with stomatal conductance and C_i positively correlated with carbon gain (Farquhar & Sharkey, 1982). Its plasticity stems from leaf-level adjustments, increased stomatal density under dim light and thicker leaves in high radiation (Fetcher, Strain & Oberbauer, 1983; Friend, 1984), enabled efficiency in diverse light environments (Stegemann, Timm & Küppers, 1996). Unlike *Inga* species, its photosynthesis was unaffected by light quality (Tinoco-Ojanguren & Percy, 1995), suggesting broader niche tolerance. Under high VPD, *Inga* species conserved water via stomatal closure (Sinclair et al., 2017), while *H. appendiculatus* prioritised carbon fixation, a critical trait for early succession. For agroforestry, this implies *Inga* species stabilise systems under drought, whereas *H. appendiculatus* optimises productivity in variable light.

The observed highest WUE in *I. punctata* and *E. americana*, suggests drought resilience (Beer et al., 1998; Chaves, Osório & Pereira, 2004; Sinclair et al., 2017), though WUE is limited by short-term measurement scales (Medrano et al., 2007).

Notably, coffee varieties exhibited inverse stomatal regulation patterns compared to shade trees, with PC1 loadings of -0.906 for conductance and -0.897 for transpiration, suggesting different water-use strategies under shaded conditions. Arabica coffee clustered into three groups: (1) *P. schiedeana*-Oro Azteca and *H. appendiculatus*-Oro Azteca, with high stomatal regulation and WUE but reduced C_i ; (2) unshaded *C. arabica* var. Oro Azteca, showing light-adapted but carbon-limited photosynthesis; and (3) intermediate varieties (*I.*

iniciul-Garnica, *I. vera*-Costa Rica 95) with balanced traits. Unshaded coffee plants showed low CO₂ assimilation, related to a higher temperature and lower RH regarding shaded plants. Variability in CO₂ assimilation rates between varieties also suggests differential sugar and starch accumulation during photosynthesis (Riaño, 1993; Mosquera-Sanchez et al., 1999).

Previously evidence suggest that unshaded coffee plants showed detrimental effects due to the effect of temperature above 25 °C on stomatal conductance and CO₂ assimilation, since these plants are exposed to higher radiation and VPD, causing the loss guard cell turgor and stomatal resistance (Makino, Nakano & Mae, 1994; Riaño, 1993; Larcher, 1994; Roháček, 2002). On the contrary, shaded plants exhibited more stable CO₂ assimilation rates, possibly linked to moderated microclimatic conditions provided by shade tree canopy.

WUE has been observed to range from 4 to 12.5 μmol CO₂/mmol H₂O in drought-tolerant cultivars (Reis Filho et al., 2022), but our shaded *H. appendiculatus*-Oro Azteca association achieved 21.62 μmol CO₂/mmol H₂O, double these values. Comparative analysis of 21 genotypes revealed generally lower seasonal WUE stability (1.2–3.4 μmol CO₂/mmol H₂O; Tezara et al., 2022), with only unshaded Oro Azteca plants approaching our observed values.

Correlation analyses reinforced these patterns: stomatal conductance and transpiration were strongly positively linked in shade trees ($r = 0.825$) and coffee ($r = 0.698$ – 0.731), while CO₂ assimilation and C_i were negatively correlated in trees ($r = -0.689$), highlighting a conserved carbon-water trade-off. PAR levels beneath canopies differed significantly, with *E. americana* and *Inga* species allowing higher understory PAR (365–379 μmol m⁻² s⁻¹) due to open canopies (Dos Santos Pereira et al., 2019), while *P. schiedeana* and *P. guajava* reduced irradiance by 40–60% (Siles et al., 2012; Idris et al., 2019). *H. appendiculatus* filtered ~60% of full sunlight (Tinoco-Ojanguren & Percy, 1995; Stegemann, Timm & Küppers, 1996), and coffee beneath dense canopies experienced PAR 30–169 times lower than unshaded conditions, mitigating photoinhibition but potentially limiting C₃ photosynthesis (Roháček, 2002; Nava et al., 2009). These PAR disparities highlight trade-offs between photoprotection and light availability for understory crops (Gholipour et al., 2010; Sinclair et al., 2017).

Our results showed how PCA unveils physiological adaptations that can enhance sustainability, productivity, and climate resilience (Hatfield & Dold, 2019), aligning ecological and agricultural goals in water-scarce regions. Three distinct shade tree strategies were observed: *P. guajava* and *P. schiedeana* regulate stomatal gas exchange, *E. americana* conserves carbon with high WUE, and intermediate species like *Inga* species and *H. appendiculatus* balance traits. Coffee plants under shade exhibited more stable carbon assimilation than unshaded ones. A strong carbon-water trade-off was consistent across species, underscoring that optimal agroforestry systems require strategic species pairing based on these complementary physiological adaptations.

Gravimetric and analytical chemical analysis: moisture and nitrogen content of coffee leaves samples

The lower air temperature and light intensity in shaded areas contribute to higher moisture content in coffee leaves by increasing relative humidity, which lowers the VPD ([Schwerbrock & Leuschner, 2017](#)). In contrast, coffee plants in unshaded conditions are subjected to environmental variables more likely to trigger plant stress responses compared to shaded plants and previous evidence on the nitrogen content of leaves has demonstrated that, in stressful situations, increased leaf nitrogen availability promotes the activation and maintenance of photoprotective systems to avert photooxidation ([Fahl et al., 1994](#); [Ramalho et al., 2000](#)), which allows shaded leaves to adapt more efficiently to different irradiation conditions than fully sun-exposed leaves ([Araujo et al., 2008](#)).

Gravimetric and chemical analysis suggests that shaded conditions could have promoted higher leaf moisture content due to increased humidity and lower VPD. Conversely, unshaded coffee plants exhibited traits suggesting environmental stress, with nitrogen likely allocated to support photoprotection mechanisms. These findings highlight a physiological divergence in coffee leaf characteristics mediated by shade management practices.

Some limitations of this study should be considered. The measurements were conducted during the cool season to ensure environmental homogeneity; therefore, physiological responses during warmer or drier periods remain to be investigated. Furthermore, while we infer the effects of shade on microclimate, direct measurements of air temperature, relative humidity, and VPD at the leaf level were not concurrently recorded. Future longitudinal studies across seasons, incorporating continuous microclimate monitoring alongside physiological measurements, would provide a more comprehensive understanding of plant responses to dynamic environmental conditions. Establishing permanent sample plots would also allow for tracking long-term carbon storage and physiological acclimation. Despite these limitations, our study offers a robust snapshot of the physiological mechanisms governing species performance and provides critical insights for the immediate selection and management of shade trees.

CONCLUSIONS

Our findings support the hypothesis that the physiological traits of native shade trees enhance carbon capture and system resilience. We demonstrate that this is mediated through distinct strategies: *E. americana* achieves high individual carbon storage and WUE, supporting resilience in drought-prone scenarios, while *I. inicuil* provides rapid, density-driven carbon capture. Remarkably, shade provision mitigated microclimatic stress for coffee, as evidenced by the higher WUE of shaded Oro Azteca variety plants. Thus, strategic species selection, combining high-carbon species like *E. americana* and *P. schiedeana* with high-tree-density *Inga* species, directly enhances carbon sequestration capacity and improves agroforestry resilience under climate change by optimising resource-use efficiency and reducing physiological stress.

Furthermore, the functional clustering of species reveals a spectrum of adaptive strategies that underline this resilience. For instance, pairing deep-shading species with drought-tolerant ones to mitigate both high radiation and water deficit.

Importantly, we also demonstrate that these shade tree traits mediate improved resilience in coffee, as hypothesised. The significantly higher WUE and stable carbon assimilation observed in shaded *C. arabica* var. Oro Azteca, particularly under *H. appendiculatus* and *P. schiedeana*, provide direct evidence of stress reduction *via* moderated microclimates. This contrasts with the limitations seen in unshaded plants, confirming that shading is a key mechanism for optimising resource-use efficiency in associated crops.

Consequently, our findings suggest a shift in agroforestry planning from a focus solely on tree density to a trait-based selection framework. The key criteria for species selection must integrate physiological performance data, specifically carbon assimilation rates, WUE, and shade density, to match species to local environmental constraints and production objectives. This relationship is essential for maximising the documented benefits of tree-crop relationships, ultimately supporting more sustainable and climate-resilient coffee production systems.

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Competing Interests

The authors declare there are no competing interests.

Author Contributions

- Daniel Cabrera-Santos conceived and designed the sampling strategy, performed the field measurements, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.
- Patricia Dávila conceived and designed the sampling strategy, authored or reviewed drafts of the article, and approved the final draft.
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- Tiziana Ulian conceived and designed the experiments, analyzed the data, authored or reviewed drafts of the article, and approved the final draft.

Data Availability

The following information was supplied regarding data availability:

The AGB parameters of the tree species, the photosynthetic parameters of the tree and coffee species studied, and raw data obtained by PCA analysis are available in the [Supplementary Files](#).

Supplemental Information

Supplemental information for this article can be found online at <http://dx.doi.org/10.7717/peerj.20255#supplemental-information>.

REFERENCES

- Acevedo G, Beinroth FH, Dubee BC, Esnard AM, Hernandez PJ, Liegel LH, Lugo-Lopez MA. 1992. *Organic carbon sequestration in the soils of Puerto Rico: a case study of a tropical environment*. Mayagüez, Puerto Rico: Dayton Publishing Services.
- Akter R, Hasan MK, Kabir KH, Darr D, Roshni NA. 2022. Agroforestry systems and their impact on livelihood improvement of tribal farmers in a tropical moist deciduous forest in Bangladesh. *Trees, Forests and People* 10:100315 DOI 10.1016/j.tfp.2022.100315.
- Alessandrini A, Biondi F, Di Filippo A, Ziaco E, Piovesan G. 2011. Tree size distribution at increasing spatial scales converges to the rotated sigmoid curve in two old-growth beech stands of the Italian Apennines. *Forest Ecology and Management* 262(11): 1950–1962 DOI 10.1016/j.foreco.2011.08.025.
- Araujo WL, Dias PC, Moraes GA, Celin EF, Cunha RL, Barros RS, DaMatta FM. 2008. Limitations to photosynthesis in coffee leaves from different canopy positions. *Plant Physiology and Biochemistry* 46(10):884–890 DOI 10.1016/j.plaphy.2008.05.005.
- Araza A, De Bruin S, Herold M, Quegan S, Labriere N, Rodriguez-Vega P, Avitabile V, Santoro M, Mitchard ETA, Ryan C, Phillips OL, Willcock S, Verbeek H, Carreiras J, Hein L, Schelhaas MJ, Pacheco-Pascagaza AM, Da Conceição Bispo P, Vaglio-Laurin G, Vieilledent G, Slik F, Wijaya A, Lewis SL, Morel A, Liang J, Sukhdeo H, Schepaschenko D, Cavlovic J, Gilani H, Lucas R. 2022. A comprehensive framework for assessing the accuracy and uncertainty of global above-ground biomass maps. *Remote Sensing of Environment* 272:112917 DOI 10.1016/j.rse.2022.112917.
- Asanok L, Krueama K, Pakketanang J, Chiangrang P. 2024. Variation of shade tree composition and carbon stock of smallholder coffee agroforestry systems along an elevation gradient in Khun Mae Kuang Forest area, northern Thailand. *Agroforest Systems* 98:3045–3060 DOI 10.1007/s10457-024-01073-9.
- Baffes J, Lewin B, Varangis P. 2005. Coffee: market setting and policies. In: *Global agricultural trade and developing countries*. Washington, D.C.: World Bank.
- Baker NR. 2008. Chlorophyll fluorescence: a probe of photosynthesis *In Vivo*. *Annual Review of Plant Biology* 59(1):89–113 DOI 10.1146/annurev.arplant.59.032607.092759.
- Baker NR, Rosenqvist E. 2004. Applications of chlorophyll fluorescence can improve crop production strategies: an examination of future possibilities. *Journal of Experimental Botany* 55(403):1607–1621 DOI 10.1093/jxb/erh196.
- Barradas VL, Fanjul L. 1986. Microclimatic characterization of shaded and open-grown coffee (*Coffea arabica* L.) plantations in Mexico. *Agricultural and Forest Meteorology* 38(1–3):101–112 DOI 10.1016/0168-1923(86)90052-3.
- Beer J, Muschler R, Kass D, Somarriba E. 1998. Shade management in coffee and cacao plantations. *Forestry Sciences* 38:139–164 DOI 10.1007/978-94-015-9008-2_6.

- Bewley J, Black M. 1994. *Seeds: physiology of development and germination*. 2nd edition. New York: Plenum Press.
- Bhattacharya A. 2019. Water-use efficiency under changing climatic conditions. In: Bhattacharya A, ed. *Changing climate and resource use efficiency in plants*. London: Academic Press, 111–180 DOI 10.1016/b978-0-12-816209-5.00003-9.
- Bilen C, El Chami D, Mereu V, Trabucco A, Marras S, Spano D. 2023. A systematic review on the impacts of climate change on coffee agrosystems. *Plants* 12(1):102 DOI 10.3390/plants12010102.
- Bonfil-Sanders C, Cajero-Lázaro I, Evans R. 2008. Germinación de semillas de seis especies de *Bursera* del centro de México. *Agrociencia* 42(7):827–834.
- Bunn C, Läderach P, Ovalle Rivera O, Kirschke D. 2015. A bitter cup: climate change profile of global production of Arabica and Robusta coffee. *Climatic Change* 129:89–101 DOI 10.1007/s10584-014-1306-x.
- Castillo-Capitán G, Ávila-Bello CH, López-Mata L, De León González F. 2014. Structure and tree diversity in traditional popoluca coffee agroecosystems in the Los Tuxtlas Biosphere Reserve, Mexico. *Interciencia* 39(9):608–619.
- Cerda R, Allinne C, Gary C, Tixier P, Harvey CA, Krolczyk L, Mathiot C, Clement E, Aubertot JN, Avelino J. 2017. Effects of shade, altitude and management on multiple ecosystem services in coffee agroecosystems. *European Journal of Agronomy* 82:308–319 DOI 10.1016/j.eja.2016.09.019.
- Chave J, Réjou-Méchain M, Búrquez A, Chidumayo E, Colgan MS, Delitti WBC, Duque A, Eid T, Fearnside PM, Goodman RC, Henry M, Martínez-Yrizar A, Mugasha WA, Muller-Landau HC, Mencuccini M, Nelson BW, Ngomanda A, Nogueira EM, Ortiz-Malavassi E, Péliissier R, Ploton P, Ryan CM, Saldarriaga JG, Vieilledent G. 2014. Improved allometric models to estimate the aboveground biomass of tropical trees. *Global Change Biology* 20(10):3177–3190 DOI 10.1111/gcb.12629.
- Chaves MM, Osório J, Pereira JS. 2004. Water use efficiency and photosynthesis. In: Bacon MA, ed. *Water use efficiency in plant biology*. Oxford: Blackwell Publishing, 42–74.
- Dai AG. 2013. Increasing drought under global warming in observations and models. *Nature Climate Change* 3:52–58 DOI 10.1038/nclimate1633.
- DaMatta FM. 2004. Ecophysiological constraints on the production of shaded and unshaded coffee: a review. *Field Crops Research* 86:99–114 DOI 10.1016/j.fcr.2003.09.001.
- Dávalos-Sotelo R, Morato IM, Martínez-Pinillos-Cueto E. 2008. Almacenamiento de carbono. In: Manson RH, Hernández-Ortíz V, Gallina S, Mehltreter K, eds. *Agroecosistemas cafetaleros de Veracruz biodiversidad, manejo y conservación*. México: Instituto de Ecología, Instituto Nacional de Ecología, 223–233.
- Davis L, Hidayati N. 2019. Carbon dioxide absorption and physiological characteristics of selected tropical lowland tree species for revegetation. *Conference Series: Earth and Environmental Science, Tangerang, Indonesia* 591:012039 DOI 10.1088/1755-1315/591/1/012039.

- De Carvalho AF, Fernandes-Filho EI, Daher M, Gomes L de C, Cardoso IM, Fernandes RBA, Schaefer CEGR. 2021.** Microclimate and soil and water loss in shaded and unshaded agroforestry coffee systems. *Agroforestry Systems* **95**:119–134 DOI [10.1007/s10457-020-00567-6](https://doi.org/10.1007/s10457-020-00567-6).
- De Reffye P, Heuvelink E, Barthelemy D, Cournede PH. 2008.** Plant growth models. In: Jorgensen SE, Fath B, eds. *Encyclopedia of ecology*. London: Academic Press, 2824–2837 DOI [10.1016/B978-008045405-4.00217-2](https://doi.org/10.1016/B978-008045405-4.00217-2).
- Diario Oficial de la Federación (DOF). 2019.** NMX-Y-359-SCFI-2019: alimentos para animales-determinación de proteína cruda método de Kjeldahl en alimentos balanceados e ingredientes mayores official method. Mexico city: Secretaría de Economía, 984.13, A-D. Available at https://www.dof.gob.mx/nota_detalle.php?codigo=5567749&fecha=13/08/2019#gsc.tab=0.
- Donald PF. 2004.** Biodiversity impacts of some agricultural commodity production systems. *Conservation Biology* **18**:17–38 DOI [10.1111/j.1523-1739.2004.01803.x](https://doi.org/10.1111/j.1523-1739.2004.01803.x).
- Dos Santos Pereira HA, Da Costa GS, Schilling AC, Mielke MS, Sanches MC, Dalmolin ÂC. 2019.** Photosynthesis, growth, and biomass allocation responses of two Inga species to contrasting light. *Acta Physiologiae Plantarum* **41**:174 DOI [10.1007/s11738-019-2966-y](https://doi.org/10.1007/s11738-019-2966-y).
- Ehrenbergerová L, Šenfeldr M, Habrova H. 2018.** Impact of tree shading on the microclimate of a coffee plantation: a case study from the Peruvian Amazon. *Bois Et Forest Des Tropiques* **334**(4):13 DOI [10.19182/bft2017.334.a31488](https://doi.org/10.19182/bft2017.334.a31488).
- Ellis EA, Baerenklau KA, Marcos-Martínez R, Chávez E. 2010.** Land use/land cover change dynamics and drivers in a low-grade marginal coffee growing region of Veracruz, Mexico. *Agroforestry Systems* **80**:61–84 DOI [10.1007/s10457-010-9339-2](https://doi.org/10.1007/s10457-010-9339-2).
- Engineer CB, Hashimoto-Sugimoto M, Negi J, Israelsson-Nordström M, Azoulay-Shemer T, Rappel WJ, Iba K, Schroeder JI. 2016.** CO₂ sensing and CO₂ regulation of stomatal conductance: advances and open questions. *Trends in Plant Science* **21**(1):16–30 DOI [10.1016/j.tplants.2015.08.014](https://doi.org/10.1016/j.tplants.2015.08.014).
- Estrada-Contreras I, Equihua M, Castillo-Campos G, Rojas-Soto O. 2015.** Climate change and effects on vegetation in Veracruz, Mexico: an approach using ecological niche modelling. *Acta Botánica Mexicana* **112**:73–93.
- Fahl JI, Carelli MC, Vega J, Magalhaes AC. 1994.** Nitrogen and irradiance levels affecting net photosynthesis and growth of young coffee plants. (*Coffea arabica* L.). *Journal of Horticultural Science* **69**:161–169 DOI [10.1080/14620316.1994.11515262](https://doi.org/10.1080/14620316.1994.11515262).
- Fairtrade Foundation. 2022.** Coffee farmers. Fairtrade Foundation. Available at <https://www.fairtrade.org.uk/farmers-and-workers/coffee/>.
- Farquhar GD, Sharkey TD. 1982.** Stomatal conductance and photosynthesis. *Annual Review of Plant Physiology* **33**(1):317–345 DOI [10.1146/annurev.pp.33.060182.001533](https://doi.org/10.1146/annurev.pp.33.060182.001533).
- Fetcher N, Strain BR, Oberbauer SF. 1983.** Effects of light regime on the growth, leaf morphology, and water relations of seedlings of two species of tropical trees. *Oecologia* **58**(3):314–319 DOI [10.1007/bf00385229](https://doi.org/10.1007/bf00385229).

- Fini A, Vigevani I, Corsini D, Węzyk P, Bajorek-Zydroń K, Failla O, Cagnolati E, Mielczarek L, Comin S, Gibin M, Pasquinelli A, Ferrini F, Viskanac P. 2023.** CO₂-assimilation, sequestration, and storage by urban woody species growing in parks and along streets in two climatic zones. *Science of the Total Environment* **903**:166198 DOI [10.1016/j.scitotenv.2023.166198](https://doi.org/10.1016/j.scitotenv.2023.166198).
- Flores-Ortiz CM, Davila P, Rodríguez-Arevalo I, Manson RH, Toledo-Garibaldi M, Cabrera-Santos D, Salguero MA, Vázquez FG, Cobos J, Gianella M, Bell E, Way M, Mattana E, Ulian T. 2025.** Prioritisation of native trees for enhancing carbon sequestration in shade-grown coffee plantations in the State of Veracruz (México): linking conservation and ecological traits to community needs. *Agroforestry Systems* **99**:55 DOI [10.1007/s10457-025-01155-2](https://doi.org/10.1007/s10457-025-01155-2).
- Food and Agriculture Organization of the United Nations (FAO). 2022.** Markets and trade: coffee. FAO. Available at <https://www.fao.org/markets-and-trade/commodities/coffee/en/>.
- Fricker M, Willmer C. 2012.** *Stomata*. Netherlands: Springer Science and Business Media.
- Friend DJC. 1984.** Shade adaptation of photosynthesis in *Coffea arabica*. *Photosynthesis Research* **5**(4):325–334 DOI [10.1007/BF00034977](https://doi.org/10.1007/BF00034977).
- García-Mateos R, Soto-Hernández M, Vibrans H. 2001.** Erythrina Americana Miller, (Colorín; Fabaceae), a versatile resource from Mexico: a review. *Economic Botany* **55**:391–400 DOI [10.1007/BF02866562](https://doi.org/10.1007/BF02866562).
- Garza-Lau R, Maldonado-Torres R, Álvarez-Sánchez ME, Torres-Rivera JA. 2020.** Characterization of tree species associated with coffee cultivation. *Revista Mexicana de Ciencias Agrícolas* **11**(1):25–32 DOI [10.29312/remexca.v11i1.2210](https://doi.org/10.29312/remexca.v11i1.2210).
- Genty B, Briantais JM, Baker NR. 1989.** The relationship between the quantum yield of photosynthetic electron transport and quenching of chlorophyll fluorescence. *Biochimica Et Biophysica Acta (BBA)—General Subjects* **990**(1):87–92 DOI [10.1016/s0304-4165\(89\)80016-9](https://doi.org/10.1016/s0304-4165(89)80016-9).
- Gholipoor M, Prasad PVV, Mutava RN, Sinclair TR. 2010.** Genetic variability of transpiration response to vapor pressure deficit among sorghum genotypes. *Field Crops Research* **119**(1):85–90 DOI [10.1016/j.fcr.2010.06.018](https://doi.org/10.1016/j.fcr.2010.06.018).
- Gill AS, Prasad JVNS. 2000.** Allelopathic interactions in agroforestry systems. In: *Allelopathy in ecological agriculture and forestry*. Dordrecht: Springer DOI [10.1007/978-94-011-4173-4_13](https://doi.org/10.1007/978-94-011-4173-4_13).
- Gliessman S. 2015.** *Agroecology: the ecology of sustainable food systems*. 3rd Edition. Boca Raton, Florida: CRC Press/Taylor and Francis, 406 DOI [10.1201/b17881](https://doi.org/10.1201/b17881).
- Gross CD, Bork EW, Carlyle CN, Chang SX. 2022.** Agroforestry perennials reduce nitrous oxide emissions and their live and dead trees increase ecosystem carbon storage. *Global Change Biology* **28**:5956–5972 DOI [10.1111/GCB.16322](https://doi.org/10.1111/GCB.16322).
- Grossiord C, Buckley TN, Cernusak LA, Novick KA, Poulter B, Siegwolf RTW, Sperry JS, McDowell NG. 2020.** Plant responses to rising vapor pressure deficit. *New Phytologist* **226**:1550–1566 DOI [10.1111/nph.16485](https://doi.org/10.1111/nph.16485).

- Haber J. 2001.** International MBA. Informe de la Finca Santa Elena, productora de café orgánico. Tapachula, Chiapas, México. Available at <https://www.cec.org/files/documents/publications/1790-finca-santa-elena-mexican-shade-coffee-farm-en.pdf>.
- Häger A. 2012.** The effects of management and plant diversity on carbon storage in coffee agroforestry systems in Costa Rica. *Agroforestry Systems* **86**:159–174 DOI [10.1007/s10457-012-9545-1](https://doi.org/10.1007/s10457-012-9545-1).
- Haggar J, Barrios M, Bolaños M, Merlo M, Moraga P, Munguia R, Ponce A, Romero S, Soto G, Staver C, Virginio E de MF. 2011.** Coffee agroecosystem performance under full sun, shade, conventional and organic management regimes in Central America. *Agroforestry Systems* **82**:285–301 DOI [10.1007/s10457-011-9392-5](https://doi.org/10.1007/s10457-011-9392-5).
- Harvey CA, Pritts AA, Zwetsloot MJ, Jansen K, Pulleman MM, Armbrecht I, Avelino J, Barrera JF, Bunn C, García JH, Isaza C, Munoz-Ucros J, Pérez-Alemán CJ, Rahn E, Robiglio V, Somarriba E, Valencia V. 2021.** Transformation of coffee-growing landscapes across Latin America. A review. *Agronomy for Sustainable Development* **41**:62 DOI [10.1007/s13593-021-00712-0](https://doi.org/10.1007/s13593-021-00712-0).
- Hatfield JL, Dold C. 2019.** Water-use efficiency: advances and challenges in a changing climate. *Frontiers in Plant Science* **10**:103 DOI [10.3389/fpls.2019.00103](https://doi.org/10.3389/fpls.2019.00103).
- Hernández-Vásquez E, Campos-Ángeles GV, Enríquez del Valle JR, Rodríguez-Ortiz G, Velasco-Velasco VA. 2012.** Captura de carbono por *Inga jinicuil* Schltdl: En un sistema agroforestal de café bajo sombra. *Revista Mexicana de Ciencias Forestales* **3**(9):11–21.
- Heuzé V, Tran G, Bastianelli D, Lebas F. 2017.** Guava (*Psidium guajava*) Feedipedia. Available at https://agritrop.cirad.fr/582480/7/ID582480_ENG.pdf (accessed on 03 August 2023).
- Idris A, Linatoc AC, Bin Abu Bakar MF, Takai ZI. 2019.** Effect of light intensity on the gas exchange characteristics and total pigment content of *Psidium guajava*. *IOP Conference Series: Earth and Environmental Science* **269**:012020 DOI [10.1088/1755-1315/269/1/012020](https://doi.org/10.1088/1755-1315/269/1/012020).
- International Coffee Organization (ICO). 2023.** Coffee report and outlook 2023. Available at https://icocoffee.org/documents/cy2023-24/Coffee_Report_and_Outlook_December_2023_ICO.pdf.
- Intergovernmental Panel on Climate Change (IPCC). 2006.** IPCC guidelines for national greenhouse gas inventories—volume 4 agriculture, forestry and other land use. In: *Prepared by the National Greenhouse Gas Inventories Programme*. Japan: IGES.
- Intergovernmental Panel on Climate Change (IPCC). 2022.** Summary for policymakers. Climate change 2022: mitigation of climatechange. contribution of working group III to the sixth assessment report of the intergovernmental panel on climate change. Cambridge University Press.
- Isaac ME, Gagliardi S, Ordoñez JC, Sauvadet M. 2024.** Shade tree trait diversity and functions in agroforestry systems: a review of which traits matter. *Journal of Applied Ecology* **61**:1159–1173 DOI [10.1111/1365-2664.14652](https://doi.org/10.1111/1365-2664.14652).

- Jawo TO, Kyereh D, Lojka B. 2022. The impact of climate change on coffee production of small farmers and their adaptation strategies: a review. *Climate and Development* 15(2):93–109 DOI 10.1080/17565529.2022.2057906.
- Jha S, Bacon CM, Philpott SM, Rice RA, Méndez VE, Läderach P. 2011. A review of ecosystem services, farmer livelihoods, and value chains in shade coffee agroecosystems. In: *Integrating agriculture, conservation and ecotourism: examples from the field. Issues in agroecology—present status and future prospectus*, vol. 1, Dordrecht: Springer DOI 10.1007/978-94-007-1309-3_4.
- Jin S, Zhang E, Guo H, Hu C, Zhang Y, Yan D. 2023. Comprehensive evaluation of carbon sequestration potential of landscape tree species and its influencing factors analysis: implications for urban green space management. *Carbon Balance Manage* 18:17 DOI 10.1186/s13021-023-00238-w.
- Jose S. 2009. Agroforestry for ecosystem services and environmental benefits: an overview. *Agroforestry Systems* 76:1–10 DOI 10.1007/s10457-009-9229-7.
- Kirkham MB. 2005. Water-use efficiency. In: *Encyclopedia of soils in the environment*. 315–322 DOI 10.1016/b0-12-348530-4/00441-0.
- Kohl T, Niether W, Abdulai I. 2024. Impact of common shade tree species on microclimate and cocoa growth in agroforestry systems in Ghana. *Agroforest Systems* 98:1579–1590 DOI 10.1007/s10457-024-01029-z.
- Koutouleas A, Sarzynski T, Bordeaux M, Bosselmann AS, Campa C, Etienne H, Turreira-García N, Rigal C, Vaast P, Ramalho JC, Marraccini P, Raebild A. 2022. Shaded-coffee: a nature-based strategy for coffee production under climate change? A review. *Frontiers in Sustainable Food Systems* 6:877476 DOI 10.3389/fsufs.2022.877476.
- Kursten E, Burschel P. 1993. CO₂ mitigation by agroforestry. *Water, Air and Soil Pollution* 70(1–4):533–544 DOI 10.1007/BF01105020.
- Lara-Estrada L, Rasche L, Schneider U. 2023. Exploring the cooling effect of shading for climate change adaptation in coffee areas. *Climate Risk Management* 42:100562 DOI 10.1016/j.crm.2023.100562.
- Larcher W. 1994. Photosynthesis as a tool for indicating temperature stress events. In: Schulze ED, Caldwell MM, eds. *Ecophysiology of photosynthesis*. Berlin: Springer Study Edition, 261–277 DOI 10.1007/978-3-642-79354-7_13.
- Lepeduš H, Viljevac M, Cesar V, Ljubešić N. 2005. Functioning of the photosynthetic apparatus under low and high light conditions in chlorotic spruce needles as evaluated by *in vivo* chlorophyll fluorescence. *Russian Journal of Plant Physiology* 52(2):165–170 DOI 10.1007/s11183-005-0024-7.
- Liang S, Wang J. 2020. Aboveground biomass. In: Liang S, Wang J, eds. *Advanced remote sensing*. 2nd edition. London: Academic Press, 543–580.
- Lin BB. 2007. Agroforestry management as an adaptive strategy against potential microclimate extremes in coffee agriculture. *Agricultural and Forest Meteorology* 144(1–2):85–94 DOI 10.1016/j.agrformet.2006.12.009.

- Lin BB. 2010.** The role of agroforestry in reducing water loss through soil evaporation and crop transpiration in coffee agroecosystems. *Agricultural and Forest Meteorology* 150:510–518 DOI 10.1016/j.agrformet.2009.11.010.
- López-Gómez AM, Williams-Linera G, Manson RH. 2008.** Tree species diversity and vegetation structure in shade coffee farms in Veracruz, Mexico. *Agriculture, Ecosystems and Environment* 124(3–4):160–172 DOI 10.1016/j.agee.2007.09.008.
- Makino A, Nakano H, Mae T. 1994.** Effects of growth temperature on the response of ribulose- 15 biphosphate carboxylase, electron transport components and sucrose synthesis enzymes to leaf nitrogen in rice and their relationships to photosynthesis. *Plant Physiology* 105:1231–1238 DOI 10.1104/pp.105.4.1231.
- Maxwell K, Johnson GN. 2000.** Chlorophyll fluorescence—a practical guide. *Journal of Experimental Botany* 51:659–668 DOI 10.1093/jexbot/51.345.659.
- McCarthy HR, Pataki DE, Jenerette GD. 2011.** Plant water-use efficiency as a metric of urban ecosystem services. *Ecological Applications* 21:3115–3127 DOI 10.1890/11-0048.1.
- Medrano H, Bota J, Cifre J, Flexas J, Ribas-Carbó M, Gulías J. 2007.** Eficiencia en el uso del agua por las plantas. *Investigaciones Geográficas* 43:63–84.
- Meili N, Manoli G, Burlando P, Carmeliet J, Chow WTL, Coutts AM, Roth M, Velasco E, Vivoni ER, Fatichi S. 2021.** Tree effects on urban microclimate: diurnal, seasonal, and climatic temperature differences explained by separating radiation, evapotranspiration, and roughness effects. *Urban Forestry and Urban Greening* 58:126970 DOI 10.1016/j.ufug.2020.126970.
- Miceli-Méndez CL, Ferguson GB, Ramírez-Marcial N. 2008.** Seed dispersal by cattle: natural history and applications to neotropical forest restoration and agroforestry. In: Myster R, ed. *Post-agricultural succession in the neotropics*. New York: Springer, 165–191 DOI 10.1007/978-0-387-33642-8_7.
- Moguel P, Toledo VM. 1999.** Biodiversity conservation in traditional coffee systems of Mexico. *Conservation Biology* 13(1):11–21 DOI 10.1046/j.1523-1739.1999.97153.x.
- Mordor Intelligence. 2024.** Global coffee market. 2024–2029. Available at [https://samples.mordorintelligence.com/46502/Sample%20-%20Global%20Coffee%20Market%20\(2024%20-%202029\)%20-%20Mordor%20Intelligence1706520790961.pdf](https://samples.mordorintelligence.com/46502/Sample%20-%20Global%20Coffee%20Market%20(2024%20-%202029)%20-%20Mordor%20Intelligence1706520790961.pdf).
- Mosquera-Sanchez LP, Riaño Herrera NM, Arcila-Pulgarin J, Ponce-Davila CA. 1999.** Fotosíntesis, respiración y fotorrespiración en hojas de café *Coffea* sp. *Cenicafé* 50(3):215–221.
- Naik SK, Sarkar PK, Das B, Singh AK, Bhatt BP. 2021.** Biomass production and carbon stock in *Psidium guajava* orchards under hot and sub-humid climate. *Current Science* 120(10):1627–1635 DOI 10.18520/cs/v120/i10/1627-1635.
- Nava D, Ramírez-Ramírez I, Peña Valdivia CB, Díaz-Villaseñor G, González-Hernández VA. 2009.** Características del intercambio de gases en hojas de guayabo. (*Psidium guajava* L.). *Revista Chapingo Serie Horticultura* 15(2):119–126.
- Nelson DW, Sommers LE. 1980.** Total nitrogen analysis of soil and plant tissues. *Journal of Association of Official Analytical Chemists* 63(4):770–778 DOI 10.1093/jaoac/63.4.770.

- Nestel D. 1995. Coffee in Mexico: international market, agricultural landscape and ecology. *Ecological Economics* 15(2):165–178 DOI 10.1016/0921-8009(95)00041-0.
- Niembro RA. 1992. *Árboles útiles de México: naturales e introducidos*. Mexico city: Limusa, 206 p.
- Niinemets Ü, Kull O. 2001. Sensitivity of photosynthetic electron transport to photoinhibition in a temperate deciduous forest canopy: photosystem II center openness, non-radiative energy dissipation and excess irradiance under field conditions. *Tree Physiology* 21:899–914 DOI 10.1093/treephys/21.12-13.899.
- Noponen MRA, Healey JR, Soto G, Hagggar JP. 2013. Sink or source-The potential of coffee agroforestry systems to sequester atmospheric CO2 into soil organic carbon. *Agriculture, Ecosystems & Environment* 175:60–68 DOI 10.1016/j.agee.2013.04.012.
- Notaro KA, Medeiros EV, Duda GP, Silva AO, Moura PM. 2014. Agroforestry systems, nutrients in litter and microbial activity in soils cultivated with coffee at high altitude. *Scientia Agricola* 71:87–95 DOI 10.1590/S0103-90162014000200001.
- Nygren P. 1995. Leaf CO2 exchange of *Erythrina poeppigiana* (Leguminosae: Phaseolae) in humid tropical field conditions. *Tree Physiology* 15(2):71–83 DOI 10.1093/treephys/15.2.71.
- Ortiz-Ceballos GC, Vargas-Mendoza M, Ortiz-Ceballos AI, Mendoza Briseño M, Ortiz-Hernández G. 2020. Aboveground carbon storage in coffee agroecosystems: the case of the central region of the State of Veracruz in Mexico. *Agronomy* 10:382 DOI 10.3390/agronomy10030382.
- Palma-Garcia JM, Gonzales-Rebeles Islas C. 2018. *Recursos arbóreos y arbustivos tropicales para una ganadería bovina sustentable*. México: Dirección General de Publicaciones, Universidad de Colima.
- Panhuyzen S, Pierrot J. 2020. Coffee barometer 2020. Belgium: Coffee Collective 2020. Available at <https://coffeebarometer.org/home2020/>.
- Pascarella J, Aide T, Serrano M, Zimmerman JK. 2000. Land-use history and forest regeneration in the Cayey Mountains, Puerto Rico. *Ecosystems* 3:217–228 DOI 10.1007/s100210000021.
- Pham Y, Reardon-Smith K, Mushtaq S, Cockfield G. 2019. The impact of climate change and variability on coffee production: a systematic review. *Climatic Change* 156:609–630 DOI 10.1007/s10584-019-02538-y.
- Poorter L, Wright SJ, Paz H, Wright IJ. 2008. Are functional traits good predictors of demographic rates? Evidence from five Neotropical forests. *Ecology* 89(7):1908–1920 DOI 10.1890/07-0207.1.
- Rahn E, Läderach P, Baca M, Cressy C, Schroth G, Malin D, Van Rikxoort H, Shriver J. 2014. Climate change adaptation, mitigation and livelihood benefits in coffee production: where are the synergies? *Mitigation and Adaptation Strategies for Global Change* 19:1119–1137 DOI 10.1007/s11027-013-9467-x.
- Rakocevic M, Marchiori PER, Zambrosi FCB, Machado EC, Maia A de HN, Ribeiro RV. 2022. High phosphorus supply enhances leaf gas exchange and growth of young Arabica coffee plants under water deficit. *Experimental Agriculture* 58:e30 DOI 10.1017/S0014479722000266.

- Ramvalho J, Pons TL, Groeneveld HW, Azinheira HG, Nunes MA. 2000. Photosynthetic acclimation of high light conditions in mature leaves of *Coffea arabica* L.: role of xanthophylls, quenching mechanisms and nitrogen nutrition. *Australian Journal of Plant Physiology* 27(1):43–51 DOI 10.1071/PP99013.
- Reigosa MS, Gonzalez L, Souto XC, Pastoriza JE. 2000. Allelopathy in forest ecosystems. In: *Allelopathy in ecological agriculture and forestry*. Dordrecht: Springer DOI 10.1007/978-94-011-4173-4_12.
- Reis Filho RJC, Carneiro MS de S, Pereira ES, Furtado RN, Morais Neto de LB, Magalhães JA, Alves FG da S, Lopes MN. 2022. Biomass components and water use efficiency in cactus pear under different irrigation systems and harvest frequencies. *Revista Brasileira de Zootecnia* 51:e20210093 DOI 10.37496/rbz5120210093.
- Riaño NM. 1993. Fotosíntesis en hojas de café. Centro Nacional de Investigaciones de Café, Cenicafe. Informe anual de labores de la Disciplina de Fisiología vegetal 1992–1993. 26–46.
- Rice R. 2008. Agricultural intensification within agroforestry: the case of coffee and wood products. *Agriculture, Ecosystems and Environment* 128:212–218 DOI 10.1016/j.agee.2008.06.007.
- Rodríguez-López NF, Martins SC, Cavatte PC, Silva PEM, Morais LE, Pereira LF, Reis JV, Ávila RT, Godoy AG, Lavinski AO, DaMatta FM. 2014. Morphological and physiological acclimations of coffee seedlings to growth over a range of fixed or changing light supplies. *Environmental and Experimental Botany* 102:1–10 DOI 10.1016/j.envexpbot.2014.01.008.
- Roháček K. 2002. Chlorophyll fluorescence parameters: the definitions, photosynthetic meaning, and mutual relationships. *Photosynthetica* 40:13–29 DOI 10.1023/A:1020125719386.
- Rojas-García F, De Jong BHJ, Martínez-Zurimendi P, Paz-Pellat F. 2015. Database of 478 allometric equations to estimate biomass for Mexican trees and forests. *Annals of Forest Science* 72(6):835–864 DOI 10.1007/s13595-015-0456-y.
- Romero-Alvarado Y, Soto-Pinto L, García-Barrios L, Barrera-Gaytán JF. 2002. Coffee yields and soil nutrients under the shades of *Inga* sp. vs. multiple species in Chiapas, Mexico. *Agroforestry Systems* 54:215–224 DOI 10.1023/A:1016013730154.
- Running SW. 1976. Environmental control of leaf water conductance in conifers. *Canadian Journal of Forest Research* 6:104–112 DOI 10.1139/x76-013.
- Rzedowski J. 1978. *Vegetación de México*. México, D.F. Mexico: Limusa.
- Sakshaug E, Bricaud A, Dandonneau Y, Falkowski PG, Kiefer DA, Legendre L, Morel A, Parslow J, Takahashi M. 1998. Parameters of photosynthesis: definitions, theory and interpretation of results. *Journal of Plankton Research* 20(3):603–603 DOI 10.1093/plankt/19.11.1637.
- Salazar-Figueroa R. 1985. Producción de leña y biomasa de *Inga densiflora* Benth en San Ramón, Costa Rica. Silvoenergía (CATIE) 3. Available at <https://repositorio.catie.ac.cr/handle/11554/3510>.

- Sánchez-Díaz M, Aguirreolea J. 2008. Transpiración y control estomático. In: Azcón-Bieto J, Talón M, eds. *Fundamentos de Fisiología Vegetal*. España: McGraw-Hill Interamericana, 41–56.
- Sarmiento-Soler A, Vaast P, Hoffmann MP, Rötter RP, Jassogne L, van Asten PJ, Graefe S. 2019. Water use of *Coffea arabica* in open versus shaded systems under smallholder’s farm conditions in Eastern Uganda. *Agricultural and Forest Meteorology* 266:231–242 DOI 10.1016/j.agrformet.2018.12.006.
- Sauvadet M, den Meersche KV, Allinne C, Gay F, De Melo Virginio Filho E, Chauvat M, Becquer T, Tixier P, Harmand J-M. 2019. Shade trees have higher impact on soil nutrient availability and food web in organic than conventional coffee agroforestry. *Science of the Total Environment* 649:1065–1074 DOI 10.1016/j.scitotenv.2018.08.291.
- Schwerbrock R, Leuschner C. 2017. Foliar water uptake, a widespread phenomenon in temperate woodland ferns? *Plant Ecology* 218:555–563 DOI 10.1007/s11258-017-0711-4.
- Segura M, Kanninen M, Suárez D. 2006. Allometric models for estimating aboveground biomass of shade trees and coffee bushes grown together. *Agroforestry Systems* 68:143–150 DOI 10.1007/s10457-006-9005-x.
- Shimshi D, Ephrat J. 1975. Stomatal behavior of wheat cultivars in relation to their transpiration, photosynthesis, and yield. *Agronomy Journal* 67:326–331 DOI 10.2134/agronj1975.00021962006700030011x.
- Siles P, Centeno H, Cabezas W, Staver C, Jürgen B, Bustamante O. 2012. Preliminary characterisation of common legume tree species as coffee shade for climate change readiness. In: *Proceedings of resilience of agricultural systems against crises*. Göttingen—Kassel/Witzenhausen.
- Silva L, Marchiori PER, Maciel CP, Machado EC, Ribeiro RV. 2010. Fotossíntese, relações hídricas e crescimento de cafeeiros jovens em relação à disponibilidade de fósforo. *Pesquisa Agropecuária Brasileira* 45:965–972 DOI 10.1590/S0100-204X2010000900005.
- Simonin KA, Limm EB, Dawson TE. 2012. Hydraulic conductance of leaves correlates with leaf lifespan: implications for lifetime carbon gain. *New Phytologist* 193(4):939–947 DOI 10.1111/j.1469-8137.2011.04014.x.
- Sinclair TR, Devi J, Shekoofa A, Choudhary S, Sadok W, Vadez V, Riar M, Ruffy T. 2017. Limited-transpiration response to high vapor pressure deficit in crop species. *Plant Science* 260:109–118 DOI 10.1016/j.plantsci.2017.04.007.
- Slot M, Krause GH, Krause B, Hernández GG, Winter K. 2019. Photosynthetic heat tolerance of shade and sun leaves of three tropical tree species. *Photosynthesis Research* 141:119–130 DOI 10.1007/s11120-018-0563-3.
- Somarriba E. 1988. Pasture growth and floristic composition under the shade of guava. *Psidium guajava* L. trees in Costa Rica. *Agroforestry Systems* 6:15–162 DOI 10.1007/BF02344752.

- Soto-Pinto L, Romero-Alvarado Y, Caballero-Nieto J, Segura WG. 2001.** Woody plant diversity and structure of shade-grown-coffee plantations in Northern Chiapas, Mexico. *Revista de Biología Tropical* **49**(3):977–987.
- Stegemann J, Timm HC, Küppers M. 1996.** Light environment and photosynthesis of an understorey and a pioneer species from a premontane rainforest of Costa Rica. *Revista Pensamiento Actual* **2**:61–68.
- Strasser RJ, Tsimilli-Michael M, Srivastava A. 2004.** Analysis of the chlorophyll a fluorescence transient. In: *Chlorophyll a Fluorescence. Advances in photosynthesis and respiration*, vol. 19. Dordrecht: Springer DOI 10.1007/978-1-4020-3218-9_12.
- Tellez O, Mattana E, Diazgranados M, Kühn N, Castillo-Lorenzo E, Lira R, Montes-Leyva L, Rodriguez I, Flores Ortiz CM, Way M, Dávila P, Ulian T. 2020.** Native trees of Mexico: diversity, distribution, uses and conservation. *PeerJ* **8**:e9898 DOI 10.7717/peerj.9898.
- Terasaki Hart DE, Yeo S, Almaraz M, Beillouin D, Cardinael R, Garcia E, Kay S, Lovell ST, Rosenstock TS, Sprengle-Hyppolite S, Stolle F, Suber M, Thapa B, Wood S, Cook-Patton SC. 2023.** Priority science can accelerate agroforestry as a natural climate solution. *Nature Climate Change* **13**:1179–1190 DOI 10.1038/s41558-023-01810-5.
- Tezara W, Duicela-Guambi LA, Reynel-Chila VG, Nazareno-Ortiz R, Bolaños Ortega MJ. 2022.** Seasonal changes in gas exchange and yield of 21 genotypes of *Coffea arabica*. *Botanical Sciences* **100**(4):1000–1101 DOI 10.17129/botsci.3023.
- Tinoco-Ojanguren C, Pearcy RW. 1995.** A comparison of light quality and quantity effects on the growth and steady-state and dynamic photosynthetic characteristics of three tropical tree species. *Functional Ecology* **9**(2):222 DOI 10.2307/2390568.
- Toledo VM, Moguel P. 2012.** Coffee and sustainability: the multiple values of traditional shaded coffee. *Journal of Sustainable Agriculture* **36**(3):353–377 DOI 10.1080/10440046.2011.583719.
- United States Department of Agriculture Foreign Agricultural Service (USDA FAS). 2023.** Coffee annual Mexico. Mexico City. Available at https://apps.fas.usda.gov/newgainapi/api/Report/DownloadReportByFileName?fileName=Coffee%20Annual_Mexico%20City_Mexico_MX2023-0024.pdf.
- Useche-Carrillo NV, Ayala-Arreola J, Campos-Rojas E, Barrientos-Priego A. 2022.** Relationships between stomatal and gas exchange characteristics of the leaf blade in ‘Colín V-33’ avocado seedlings. *Revista Chapingo Serie Horticultura* **28**(3):133–144 DOI 10.5154/r.rchsh.2021.05.008.
- Vaast P, Angrand J, Franck N, Dauzat J, Génard M. 2005.** Fruit load and branch ring-barking affect carbon allocation and photosynthesis of leaf and fruit of *Coffea arabica* in the field. *Tree Physiology* **25**:753–760 DOI 10.1093/treephys/25.6.753.
- Valdés-Velarde E, Vázquez-Domínguez LP, Tinoco-Rueda JÁ, Sánchez-Hernández R, Salcedo-Pérez E, Lagunes-Fortiz E. 2022.** Ecosystem service of carbon stored in coffee plantations under shade in agroforestry systems. *Revista Mexicana de Ciencias Agrícolas* **13**(spe28):287–297 DOI 10.29312/remexca.v13i28.3283.

- Valencia V, García-Barrios L, West P, Sterling EJ, Naeem S. 2014. The role of coffee agroforestry in the conservation of tree diversity and community composition of native forests in a Biosphere Reserve. *Agriculture, Ecosystems and Environment* 189:154–163 DOI 10.1016/j.agee.2014.03.024.
- Vázquez-Torres M, Campos-Jiménez J, Juárez-Fragoso M. 2017. Árboles tropicales de Veracruz. Veracruz: Universidad Veracruzana, Dirección Editorial. Available at <https://libros.uv.mx/index.php/UV/catalog/book/QC016>.
- Verchot LV, Kandji ST, Tomich TP, Ong CK, Albrecht A, Mackensen J, Bantilan CC, Anupama KV, Palm CA, Van Noordwijk M. 2007. Climate change: linking adaptation and mitigation through agroforestry. *Mitigation and Adaptation Strategies for Global Change* 12:901–918 DOI 10.1007/s11027-007-9105-6.
- Vester HFM. 2002. Architectural models in the tree flora of the Yucatan Peninsula. *Botanical Sciences* 71:45–57 DOI 10.17129/botsci.1662.
- Vizcaíno-Bravo Q, Williams-Linera G, Asbjornsen H. 2020. Biodiversity and carbon storage are correlated along a land use intensity gradient in a tropical montane forest watershed, Mexico. *Basic and Applied Ecology* 44:24–34 DOI 10.1016/j.baae.2019.12.004.
- Weather Spark. 2024. Weather Spark: the average weather in Teocelo and Xico, Veracruz, Mexico. Databases. Retrieved from <https://es.weatherspark.com/y/8665/Clima-promedio-en-Teocelo-M%C3%A9xico-durante-todo-el-a%C3%B1o>; <https://es.weatherspark.com/y/7587/Clima-promedio-en-Xico-M%C3%A9xico-durante-todo-el-a%C3%B1o>.
- Williams-Linera G, Lorea F. 2009. Tree species diversity driven by environmental and anthropogenic factors in tropical dry forest fragments of central Veracruz, Mexico. *Biodiversity and Conservation* 18:3269–3293 DOI 10.1007/s10531-009-9641-3.
- Xu Z, Jiang Y, Jia B, Zhou G. 2016. Elevated-CO₂ response of stomata and its dependence on environmental factors. *Frontiers in Plant Science* 7:657 DOI 10.3389/fpls.2016.00657.
- Zavafer A, Mancilla C. 2021. Concepts of photochemical damage of photosystem II and the role of excessive excitation. *Journal of Photochemistry and Photobiology C: Photochemistry Reviews* 47:100421 DOI 10.1016/j.jphotochemrev.2021.100421.