

Two new free-living marine species of *Desmodorella* (Nematoda: Desmodoridae) from the continental shelf of Northeastern Brazil, with an emended generic diagnosis and a dichotomous key to the species (#119644)

1

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Two new free-living marine species of *Desmodorella* (Nematoda: Desmodoridae) from the continental shelf of Northeastern Brazil, with an emended generic diagnosis and a dichotomous key to the species

Alex Manoel ^{Corresp., 1, 2}, Patricia F Neres ¹, André M Esteves ¹

¹ Zoologia, Universidade Federal de Pernambuco, Recife, PE, Brazil

² Biologia Animal Graduate Course, Universidade Federal de Pernambuco, Recife, PE, Brazil

Corresponding Author: Alex Manoel
Email address: alexblg08@gmail.com

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Desmodorella cornuta sp. nov. is characterized by a protuberant horn-shaped cuticular projection positioned dorsally in the pharyngeal region, a unique characteristic among other species of the genus. *Desmodorella parabalteata* sp. nov. is distinguished from other species by the presence of a cephalic capsule and cuticle ornamented with vacuoles, multispiral amphidial fovea, longitudinal rows of ridges that are often indistinct under light microscopy, two pairs of lateral rows of spines that are more prominent than the remaining rows, and thin, nearly straight spicules with a slightly swollen proximal end and lacking a capitulum. An emended diagnosis of the genus and a dichotomous key to species are provided.

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Abstract

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Key words: Desmodorinae, Potiguar Basin, Taxonomy, Desmodora, Nematode diversity.

Introduction

The Family Desmodoridae Filipjev, 1922 currently includes six subfamilies, 50 genera, and approximately 430 species (Nemys, 2025). The available literature on the genus *Desmodorella* Cobb, 1933 documents several taxonomic revisions that have altered its status over time (Cobb, 1933; Gerlach, 1950; Wieser, 1954; Gerlach, 1963; Lorenzen, 1976; Verschelde, Gourbault & Vincx, 1998).

After its establishment by Cobb (1933), Gerlach (1950) suggested that *Desmodorella* might represent a subgenus of *Desmodora* de Man, 1889. However, he merely noted this possibility without formally reclassifying the taxon. Subsequently, Wieser (1954), based on Gerlach (1950), reduced *Desmodorella* to a subgenus of *Desmodora*. Similarly, Gerlach (1963) in his review of the Desmodoridae, considered *Desmodorella* as a subgenus of *Desmodora*. He argued that several genera closely related to *Desmodora*, described earlier by Cobb (1920; 1933), should in fact be treated as subgenera of *Desmodora*. In the same study, Gerlach provided a key to the subgenera, using the morphology of the amphidial fovea as the primary diagnostic character.

Lorenzen (1976) disagreed with Gerlach (1963), arguing that amphidial fovea morphology alone was insufficient for differentiating the subgenera *Desmodora* and *Desmodorella*, and thus

synonymized *Desmodorella* with *Desmodora*. In contrast, Verschelde, Gourbault & Vincx (1998) revised the genus *Desmodora* and, while concurring that amphidial fovea morphology was not a reliable distinguishing character, disagreed with the synonymization proposed by Lorenzen. Verschelde, Gourbault & Vincx (1998) reconsidered *Desmodorella* as a valid genus within the Desmodoridae and argued that *Desmodorella* can be easily distinguished from *Desmodora* by the presence of longitudinal rows of ridges or spines along the body—a feature consistently present in *Desmodorella* but absent in *Desmodora*. Additionally, species of *Desmodorella* were noted to have spicules either lacking a capitulum or bearing only a minute one, and without a velum.

Marine representatives of *Desmodorella* have been recorded from the Pacific (Verschelde, Gourbault & Vincx, 1998), Atlantic (Gerlach, 1950; Riera, Núñez & Brito, 2012), Indian (Annapurna *et al.*, 2012), and Antarctic Oceans (Ingels *et al.*, 2006), with habitats ranging from the intertidal zone (Riera, Núñez & Brito, 2012) to deep-sea environments (Verschelde, Gourbault & Vincx, 1998; Fadeeva, Mordukhovich & Zograf, 2016). Occurrences of this genus have also been recorded in freshwater bodies (Gagarin & Nguyen, 2003; Decraemer & Smol, 2006). Along the Brazilian coast, the occurrence of this genus was recorded in dissertations/theses for deep-sea regions in the Campos Basin, Southeast Brazil (Silva, 2012; Moura, 2013) and for the Continental Shelf of the Potiguar Basin, Northeast Brazil (Larrazábal-Filho, 2020).

The present study reports on specimens of *Desmodorella* from the continental shelf of northeastern Brazil, describes two new species, and updates the generic diagnosis by incorporating new morphological characters. Here we also propose a dichotomous key based on male characteristics to facilitate the identification of *Desmodorella* species. Additionally, we highlight the key diagnostic traits that should be jointly considered for accurate species-level differentiation within the genus.

Material and methods

Study area and sampling (Table 1). The sediment used in the study of these animals was obtained from two projects that conducted sampling at different stations along the coast of northeastern Brazil. Table 1 presents details of the collection stations relevant to this study. In both projects the sediment samples were taken in triplicate. For sediment collection, a box-corer or Van Veen grab was used (see Table 1), while meiofauna samples were collected with a 10 cm × 10 cm corer. The collected material was transferred into plastic containers and preserved in a 4% buffered formaldehyde solution.

Laboratory processing. In the laboratory, sediment samples were sieved using a 0.500 mm mesh, followed by a 0.045 mm mesh to retain meiobenthic organisms. The material retained on the 0.045 mm mesh was subsequently extracted using SICOL-40 colloidal silica solution (specific gravity 1.18) (Somerfield, Warwick & Moens, 2005).

Nematodes were counted and extracted under a stereomicroscope using a Dollfus plate. Each specimen was subsequently placed into a small glass container filled with a solution comprising formaldehyde (4%) with glycerin (1%) (Solution 1 – De Grisse, 1969). The procedure for transferring each organism to glycerin was implemented, followed by clearing in accordance with the method outlined by De Grisse (1969). The specimens were then permanently mounted on

glass slides, following the technique described by Cobb (1920). The genus was identified using the identification keys provided by Platt & Warwick (1988) as well as Decraemer & Smol (2006). Species identification was achieved by comparing their characteristics with those detailed in the original descriptions. Illustrations were created with the assistance of an Olympus CX31 optical microscope equipped with a drawing tube. Body measurements were recorded using a mechanical map meter.

For scanning electron microscopy (SEM), specimens were taken from previously mounted glycerin-paraffin slides. These specimens underwent rehydration using distilled water, following the protocol outlined by Abolafia (2015). Subsequently, the specimens were transferred to a meiofauna processing container, as described by Abolafia (2015), and subjected to a gradual dehydration process through a series of graded ethanol concentrations (10% for one day, followed by 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 92%, 95%, and two rounds of 100% on the second day, with transitions between concentrations occurring every two hours). After dehydration, the specimens were dried using a critical point dryer. Finally, the specimens were removed from the container, placed on an aluminum stub that was covered with conductive tape, sputter-coated with gold, and examined using a TM4000 SEM at 10 kV with a backscattered electron (BSE) detector or by combining this with the secondary electron (SE) detector.

The holotype and a female paratype of each species are deposited in the Nematoda Collection at the Museum of Oceanography Prof. Petronio Alves Coelho (MOUFPE), Brazil. Additional paratypes are stored in the Meiofauna Laboratory, Department of Zoology at the Federal University of Pernambuco (NM LMZOO-UFPE).

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Results

SYSTEMATICS

Taxonomic classification, according to Decraemer & Smol (2006)

Class Chromadorea Inglis, 1983

Subclass Chromadoria Pearse, 1942

Order Desmodorida De Coninck, 1965

Suborder Desmodorina De Coninck, 1965

Superfamily Desmodoroidea Filipjev, 1922

Family Desmodoridae Filipjev, 1922

Subfamily Desmodorinae Filipjev, 1922

Genus *Desmodorella* Cobb, 1933

Diagnosis. (Emended from Leduc & Zhao, 2016) Annulated cuticle ornamented with longitudinal rows of ridges or hair-like spines (sometimes indistinct under light microscope). **Cuticular vacuoles present or absent.** Lateral alae absent. **Two pairs of lateral rows with more distinct spines, among the other rows of spines (“false lateral alae”) present or absent. Two to eight longitudinal rows of somatic setae. Horn-shaped cuticular projections present or absent.** Head capsule truncated or rounded, either smooth or ornamented with numerous vacuoles (appearing smooth or wrinkled under SEM). Cephalic setae anterior to or at level of the anterior edge of amphidial fovea. **Rows of subcephalic setae present or absent (additional scattered setae may occur without forming distinct rows).** Large multispiral to loop-shaped amphidial fovea located on head capsule (with the **largest portion situated on the labial region in *D. spineacaudata***). Pharynx with a rounded or oval posterior bulb. Spicules slender or filiform, short to elongated, **lacking a prominent capitulum and velum.** Gubernaculum present, with or without lateral pieces (crurae). **Precloacal supplements present or absent. Tail often conical, sometimes with a cylindrical terminal portion.**

Type species: *Desmodorella tenuispiculum* (Allgén, 1928) Verschelde, Goubault & Vincx, 1998

Valid species of *Desmodorella* Cobb, 1933

The valid species list is based on Verschelde, Goubault & Vincx (1998), Leduc & Zhao (2016) and the Nemys database (2025), with modifications:

Desmodorella abyssorum (Allgén, 1929) Gerlach, 1963

Syn *Desmodora abyssorum* Allgén, 1929

Desmodorella aquaedulcis (Gagarin & Nguyen, 2003) Decraemer & Smol, 2006

Syn *Desmodora aquaedulcis* Gagarin & Nguyen, 2003

Desmodorella balteata Verschelde, Goubault & Vincx, 1998

Desmodorella cornuta **sp. nov.**

Desmodorella curvispiculum (Jensen, 1985) Verschelde, Goubault & Vincx, 1998

Syn *Desmodora curvispiculum* Jensen, 1985

Desmodorella filispiculum (Lorenzen, 1976) Verschelde, Goubault & Vincx, 1998

Syn *Desmodora filispiculum* Lorenzen, 1976

Desmodorella papillostoma (Murphy, 1962) Verschelde, Goubault & Vincx, 1998

- 167 *Syn Desmodora papillostoma* Murphy, 1962
- 168 *Desmodorella parabalteata* **sp. nov.**
- 169 *Desmodorella perforata* (Wieser, 1954) Verschelde, Goubault & Vincx, 1998
- 170 *Syn Desmodora perforata* Wieser, 1954
- 171 *Desmodora wieseri* Inglis, 1968
- 172 *Desmodora wolfgangi* (Inglis, 1968)
- 173 *Xenodesmodora wieseri* Inglis, 1968
- 174 *Desmodorella sanguinea* (Southern, 1914) Verschelde, Goubault & Vincx, 1998
- 175 *Syn Desmodora sanguinea* Southern, 1914
- 176 *Desmodorella schulzi* (Gerlach, 1950) Verschelde, Goubault & Vincx, 1998
- 177 *Syn Desmodora schulzi* Gerlach, 1950
- 178 *Desmodorella sinuata* (Lorenzen, 1976) Verschelde, Goubault & Vincx, 1998
- 179 *Syn Desmodorella sinuata* Lorenzen, 1976
- 180 *Desmodorella spineacaudata* Verschelde, Goubault & Vincx, 1998
- 181 *Desmodorella tenuispiculum* (Allgén, 1928) Verschelde, Goubault & Vincx, 1998
- 182 *Syn Desmodora (Desmodorella) cephalata* Cobb, 1933
- 183 *Desmodora cephalata* (Cobb, 1933)
- 184 *Desmodora cephalia* (Cobb, 1933) Gerlach & Riemann, 1973
- 185 *Desmodora tenuispiculum* Allgén, 1928
- 186 *Desmodorella cephalia* (Cobb, 1933) Gerlach & Riemann, 1973
- 187 *Desmodorella verscheldei* Leduc & Zhao, 2016
- 188 **Invalid species**
- 189 *Desmodorella hirsuta* (Chitwood, 1936) Verschelde, Goubault & Vincx, 1998 (nomen dubium)
- 190 *Desmodorella bullata* (Steiner, 1916) Verschelde, Goubault & Vincx, 1998 (taxon inquirendum)
- 191
- 192 **Description of the new species.**
- 193 *Desmodorella cornuta* **sp. nov.**
- 194 (Table 2; Figures 1–3)

Material studied. Holotype male (MOUFPE 0034), paratype female 1 (MOUFPE 0035), 2 male paratypes (NM-LMZOO/UFPE 511–512) and 2 female paratypes (NM-LMZOO/UFPE 513–514).

Type locality. South Atlantic Ocean, continental shelf off the State of Rio Grande do Norte (Potiguar Basin), Brazil; station ME2B2R3 (5°02'30.3"S, 36°23'12.3"W); June 2013; depth: 8.5 m.

Locality of paratypes. Paratype female 1: South Atlantic Ocean, continental shelf off the State of Rio Grande do Norte (Potiguar Basin), Brazil, (5°02'29.6"S, 36°23'11.9"W); June 2013; depth: 8.5 m. **Male paratypes:** (1) South Atlantic Ocean, continental shelf off the State of Rio Grande do Norte (Potiguar Basin), Brazil, (5°01'12.4"S, 36°23'27.6"W); June 2012; depth: 8.1 m; (2) South Atlantic Ocean, continental shelf off the State of Rio Grande do Norte (Potiguar Basin), Brazil, (05°02'29.6"S, 36°23'11.9"W); depth: 8.5 m. **Female paratypes 2 and 3:** South Atlantic Ocean, continental shelf off the State of Rio Grande do Norte (Potiguar Basin), Brazil, (5°01'12.4"S, 36°23'27.6"W); June 2012; depth: 8.1 m.

Etymology. The specific epithet “*cornuta*” refers to the presence of dorsally positioned, horn-shaped cuticular projections.

Holotype male. Body cylindrical (1,254 µm long), narrowest in region between the base of the pharynx and anterior end of testis and widest. Maximum body diameter corresponding to 2.2 times the head diameter. Cuticle coarsely annulated and ornamented with transversal rows of small vacuoles. Cuticle pattern variable along the body. Annules broad and widely spaced in anterior region (first 10 annules below head capsule = 30 µm), narrower and more closely in mid-body (10 annules = 18 µm); broader again from the precloacal region to tail (10 = 20 µm). Twelve longitudinal rows of hair-like spines, sometimes difficult to see under light microscopy, arranged along the body (5–7 µm), often indistinct under light microscope, most visible 85 µm from base of pharynx, extending to precloacal region. At 136 µm from pharynx base, two sublateral rows merge laterally, forming “false lateral alae” (cf. Verschelde, Gourbault & Vincx, 1998) of shorter, more robust spines, extending to testis region. Afterward, rows diverge, and spines morphology returns to regular form. Somatic setae (3–8 µm) arranged in two longitudinal rows (dorsal and ventral) along body, absent in tail region. Dorsal, protuberant horn-shaped cuticular projection (9 µm long) located dorsally at the 14th annule (64.5 µm from anterior end, or 44% of pharynx length). Head capsule long, well-developed, ornamented with numerous small vacuoles below the amphidial fovea. Anterior sensilla arranged in 6+6+4 pattern: six inner labial papillae, six outer labial papillae setiform (about 2 µm long) and four small cephalic setae (3.5 µm long). Cephalic setae corresponding to 16% of the head diameter. Rows of subcephalic setae absent. Two additional setae (about 3 µm long), one dorsal and one ventral, present on posterior head capsule. Amphidial fovea distinctly sclerotized, multispiral, about 3 turns, 43% of corresponding body diameter and located 3.5 µm from anterior end (about 0.2 times the head diameter). Buccal cavity with a strong dorsal tooth and a small ventrosublateral tooth. Pharynx muscular (145 µm long), cylindrical forming slightly oval terminal bulb that occupies 49% of the corresponding body diameter. Nerve ring, secretory-excretory system and cardia not observed. Reproductive system monorchic, with single anterior outstretched testis on left of intestine. Spicules slender (79 µm long), arched ventrally, with slightly swollen proximal end (2.6 times the cloacal body diameter) and without capitulum. Gubernaculum with short lateral crurae. Precloacal supplements absent. Caudal glands indistinct. Tail conical, elongate, about 4 times the cloacal body diameter.

Paratype female 1. Generally similar to male. Body 1,221 μm long, maximum diameter of 57 μm at vulva level (about 2.4 times head diameter). Cuticular annule pattern as in male (first 10 annules below head capsule = 27 μm ; 10 annules in the narrowest body region = 16 μm ; 10 annules in tail = 19 μm). Several incomplete or bifurcated annules present, more visible at pharynx level. Longitudinal rows of hair-like spines and head capsule similar to male. Somatic setae as in male visible along anterior two-thirds of the body. Dorsal horn-shaped cuticular projection (8 μm long) positioned dorsally at 13th annule (56 μm from the anterior end or 39% of pharynx length). Labial region invaginated. Cephalic setae correspond to 13% of head diameter. Amphidial fovea as in male. Basal bulb occupies 38% of corresponding body diameter. Vulva located 858 μm from anterior end, at 70% of body length. Reproductive system didelphic with reflexed ovaries. Tail conical, elongate, 4.3 times cloacal body diameter.

Diagnosis. *Desmodorella cornuta* **sp. nov.** is characterized by the following combination of the features: cuticle coarsely annulated and ornamented with transversal rows of small vacuoles; protuberant horn-shaped cuticular projection located dorsally at 38–44% of the pharynx length; twelve longitudinal rows of hair-like spines arranged along of the body; two pairs of lateral rows with more distinct spines forming “false lateral alae”; head capsule ornamented with numerous small vacuoles; amphidial fovea multispiral (about 3 turns), occupying 42–51% of corresponding body diameter; subcephalic setae absent with additional setae present; tail conical (3.7–4.4 times cloacal/anal body diameter); males with slender, ventrally arched spicules (55–79 μm long; 2–2.6 times cloacal body diameter), with slightly swollen proximal end.

Differential diagnosis (Table 3). *Desmodorella cornuta* **sp. nov.** shares with *D. curvispiculum* (Jensen, 1985) Verschelde, Gourbault & Vincx, 1998, *D. perforata* (Wieser, 1954) Verschelde, Gourbault & Vincx, 1998 and *D. balteata* Verschelde, Gourbault & Vincx, 1998, the following features: head capsule ornamented with numerous small vacuoles, similar spicules length (see Table 3), and absence of subcephalic setae. Additionally, *D. cornuta* **sp. nov.** and *D. balteata* possess “false lateral alae”. However, *D. cornuta* **sp. nov.** is the only known species of *Desmodorella* that exhibits a protuberant, horn-shaped cuticular projection dorsally in the pharynx region (between 38–44% of pharynx length). This unique feature aids in the identification of *D. cornuta* **sp. nov.** and clearly distinguishes it from *D. curvispiculum*, *D. perforata*, *D. balteata* and the other valid species of the genus.

Desmodorella parabalteata **sp. nov.**

(Table 4; Figures 4–8)

Material studied. Holotype male (MOUFPE 0036), paratype female 1 (MOUFPE 0037), 10 male paratypes (NM LMZOO-UFPE 515–524) and 8 female paratypes (NM LMZOO-UFPE 525–532).

Type locality. South Atlantic Ocean, continental shelf of the State of Bahia, Brazil (13°04'10.32"S, 38°25'46.98"W); 11 December 2019; depth: 65 m.

Locality of paratypes. Paratype female 1: South Atlantic Ocean, continental shelf of the State of Sergipe, Brazil, (11°00'00.54"S, 36°49'58.98"W); 10 December, 2019; depth: 54 m. **Male**

paratypes: (1 and 2) Same as holotype locality, Bahia, Brazil, 11 December 2019; depth: 65 m; (3–7) Sergipe, Brazil, (11°00'00.54"S, 36°49'58.98"W), 10 December 2019; depth: 54 m; (8) Sergipe, Brazil, (10°44'59.28"S, 36°25'32.88"W); 09 December 2019; depth: 58 m; (9 and 10) Alagoas, Brazil, (10°07'05.7"S, 35°50'58.0"W); 09 December 2019; depth: 63 m. **Other female paratypes:** (2, 3, 7) Sergipe, Brazil, (11°00'00.54"S, 36°49'58.98"W); 10 December 2019; depth: 54 m; (4, 5, 9) Bahia, Brazil, (13°04'10.32"S, 38°25'46.98"W), 11 December 2019; depth: 65 m; (6, 8) Alagoas, Brazil, (10°07'05.7"S, 35°50'58.0"W); 09 December 2019; depth: 63 m.

Etymology. The specific epithet “*parabalteata*” refers to the morphological similarity of this species to *Desmodorella balteata*.

Holotype male. Body cylindrical, 697.5 µm long; narrowest between base of pharynx and anterior end of testis; widest at level of testis. Maximum body diameter corresponding to 2.1 times head diameter. Cuticle annulated, ornamented with transversal rows of small vacuoles (more evident in pharyngeal region). Cuticle pattern variable along body. Annules broad in anterior pharyngeal region (first 10 annules below head capsule = 16.5 µm), gradually narrowing towards widest body (10 annules = 6 µm) expanding progressively from proximal region of spicule to tail (10 annules = 11 µm). Longitudinal rows of ridges or short spines indistinct in holotype under light microscopy, but visible in paratype male 9 (Fig. 5C). Two sublateral pairs of longitudinal spines converge laterally about 30 µm from base of pharynx, forming “false lateral alae” composed of more robust spines, extending to first third of testis. Somatic setae arranged in six longitudinal rows (four sublateral, one dorsal, and one ventral). Dorsal and ventral rows of somatic setae 2–7 µm long, present along entire body except tail region. Sublateral somatic setae <2–6 µm long, extend from 70 µm behind pharynx to caudal region; smaller and less visible in narrowest region; longer and slightly more robust near precloacal region. Head capsule long, well-developed, ornamented with numerous small vacuoles below amphidial fovea. Anterior sensilla arranged in 6+6+4 pattern: six inner labial papillae, six outer labial setae (about 3 µm long) and four small cephalic setae (4 µm long). Cephalic setae corresponding to 21% of the head diameter. Rows of subcephalic setae absent. Two additional setae, one dorsal and the other ventral (about 2 µm long). Amphidial fovea distinctly sclerotized, multispiral, about 3.5 turns, occupying 52% of body diameter; anterior edge aligned with cephalic setae and located 6.5 µm from anterior end (about 0.3 times head diameter). Buccal cavity with a strong dorsal tooth and a small ventrosublateral tooth. Pharynx muscular 123 µm long, cylindrical with slightly oval terminal bulb (55% of corresponding body diameter). Nerve ring, secretory-excretory system, and cardia not observed. Reproductive system with monorchic with anterior outstretched testis one left of intestine. Spicules slender, 36.5 µm long (1.7 times cloacal body diameter), nearly straight, with slightly swollen proximal end, without capitulum. Gubernaculum funnel-shaped, surrounding distal end of spicules. Precloacal supplements absent. Three caudal glands. Tail conical, elongated, about 5 times cloacal body diameter.

Paratype female 1. Largely similar to male. Body 795 µm long; maximum diameter 51 µm at vulva level. Cuticular annule similar to male (first 10 annules below head capsule = 20 µm; 10 annules in widest body region = 6.5 µm; 10 annules in tail = 13 µm). Longitudinal rows of ridges visible under light microcopy in paratype females 6, 7 and 9 (Fig 6C); indistinct in paratype female 1. Somatic setae (<2–8 µm long) arranged similarly to male, but fewer in number; present along entire body except tail region. Head capsule largely similar to male. Labial region invaginated.

Cephalic setae correspond to 22% of head diameter. Amphidial fovea similar to male. Basal bulb occupies 70% of corresponding body diameter. Vulva 510 µm from anterior end, located at 64% of body length. Reproductive system didelphic with reflexed ovaries. Three caudal glands present. Tail conical, elongated, 4 times cloacal body diameter.

SEM analyses. Male paratypes 8 and 10 and paratype female 3: Head capsule wrinkled (Fig 7B, C and 8B). Cuticular annules in anterior third of body are ornamented with numerous transverse bars (Fig 7B, C, D and 8B). Towards mid-body (narrowest region), bars number decreases and bars elongate forming short spine-like structures arranged in longitudinal rows (Fig 7E). Exact number of longitudinal rows indeterminate. Bifurcated cuticular annules occur along body. Spines of the false lateral alae largest at anterior end of rows, progressively decreasing in size posteriorly (Fig 7E and 8C). After termination of false lateral alae, rows diverge and spines resume morphology seen in other rows (Fig 8D).

Diagnosis. *Desmodorella parabalteata* sp. nov. is characterized by a combination of the following features: cuticle with coarse annulations and ornamented with transversal rows of small vacuoles; cuticle with numerous transverse bars under SEM; longitudinal rows of ridges or short spines (often indistinct under light microscopy) present along body; two pairs of lateral rows with more distinct spines among other rows (false lateral alae); somatic setae arranged in six longitudinal rows (four sublateral, one dorsal, and one ventral); head capsule ornamented with numerous small vacuoles below the amphidial fovea; head capsule wrinkled under SEM; amphidial fovea multispiral, about 3.5 turns, with anterior edge at same level as cephalic setae and occupying 43–59% of corresponding body diameter; subcephalic setae absent; additional setae present; tail elongate-conical (4–6 times cloacal/anal body diameter); males with slender, nearly straight spicules (25–41.5 µm long; 1.4–1.8 times cloacal body diameter), slightly swollen proximally, without capitulum.

Differential diagnosis. *Desmodorella parabalteata* sp. nov. is closely related to *D. balteata*. These two species share several morphological features, including: a cephalic capsule ornamented with numerous small vacuoles below the amphidial fovea; absence of subcephalic setae; multispiral amphidial fovea; longitudinal rows of ridges often indistinct under light microscopy; two pairs of lateral rows with more distinct spines among other rows (false lateral alae) and six longitudinal rows of somatic setae. Together, these traits distinguish both species from the others members of the genus *Desmodorella*.

However, *D. parabalteata* sp. nov. differs from *D. balteata* in the following features: presence of cuticular vacuoles in *D. parabalteata* sp. nov. [versus (vs) absent in *D. balteata*]; number of amphidial turns (3.5 turns in the new species vs 2.6 turns in *D. balteata*); spicule length (25–41.5 µm long in *D. parabalteata* sp. nov. vs 65–85 µm long in *D. balteata*) and spicule morphology (slender, nearly straight spicules with slightly swollen proximal ends and lacking a capitulum in *D. parabalteata* sp. nov. vs slightly curved spicules with a small rounded capitulum in *D. balteata*). Additionally, males of *D. balteata* possess a ventral row of robust precloacal setae, a feature absent in the new species.

373 **Dichotomous identification key for valid species of *Desmodorella* Cobb, 1933**

374	1. Amphidial fovea multispiral (2 or more turns).....	2
375	- Amphidial fovea multispiral (less than 2 turns) or not multispiral.....	3
376	2. Spicules greater than 100 µm long.....	4
377	- Spicule less than 100 µm long.....	5
378	3. Spicules greater than 100 µm long.....	6
379	- Spicule less than 100 µm long.....	9
380	4. Amphidial fovea completely positioned in the main part of the head capsule; vacuolated head capsule present.....	7
382	- Amphidial fovea with anterior edge in lip region and posterior edge in head capsule; smooth head capsule; four ventrosublateral rows of 3–4 thorns on tail.....	<i>D. spineacaudata</i>
385	5. Head capsule not medially bulging; gubernaculum less than $\frac{1}{3}$ of spicule length.....	8
387	- Strongly bulging at the level of amphidial fovea; gubernaculum equivalent to about $\frac{1}{2}$ of spicule length.....	<i>D. abyssorum</i>
389	6. Amphidial fovea cryptospiral; “false lateral alae” absent.....	11
391	- Amphidial fovea loop-shaped; “false lateral alae” present.....	<i>D. verscheldei</i>
392	7. Spicules 220–250 µm, sinuous posteriorly; short subcephalic setae; “false lateral alae” present; 12 longitudinal rows of hair-like spines.....	<i>D. sinuata</i>
395	- Spicules ~about 150 µm, arched; elongated cephalic and subcephalic setae; “false lateral alae” absent.....	<i>D. papillostoma</i>
397	8. “False lateral alae” present (2 pairs of lateral rows of more distinct spines).....	12
399	- “False lateral alae” absent.....	13
400	9. Amphidial fovea loop-shaped (spiral).....	10
401	- Amphidial fovea unispiral.....	<i>D. aquaedulcis</i>
402	10. Elongated subcephalic setae (2 circles with 8 subcephalic setae each); smooth cephalic capsule; 8–12 longitudinal rows of hair-like spines; precloacal supplements as 4 rows of triangular cuticular spines; “false lateral alae” present; shoe-shaped gubernaculum.....	<i>D. schulzi*</i>

- 406 - Short subcephalic setae; vacuolated head capsule posterior to amphids; lamellar
407 gubernaculum.....*D. perforata*
- 408 11. Spicules 240–325 μm ; 16 longitudinal rows of hair-like spines.....*D. filispiculum*
- 409 - Spicules 182–224 μm ; 10 longitudinal rows of hair-like spines
410*D. sanguinea*
- 411 12. Horn-shaped cuticular projection dorsally at pharynx level; 2 longitudinal rows of somatic
412 setae.....*D. cornuta* **sp. nov.**
- 413 - Horn-shaped cuticular projection absent; 6 longitudinal rows of somatic
414 setae.....14
- 415 13. Subcephalic setae absent; 10–14 longitudinal rows of ridges.....*D. curvispiculum*
- 416 - Subcephalic setae present; 12–24 longitudinal rows of ridges.....*D. tenuispiculum***
- 417 14. Cuticular vacuoles absent; spicules 65–85 μm , slightly arched with a tiny rounded capitulum;
418 ventral row of robust precloacal setae
419 present.....*D. balteata*
- 420 - Cuticular vacuoles present; spicules 25–41.5 μm , nearly straight with slightly swollen proximal
421 end, no capitulum.....*D. parabalteata* **sp. nov.**
- 422 (*): Vincx (1983), redescribed *D. schulzi* and mentioned that this species presents “false lateral
423 alae” and 12 longitudinal rows of hair-like spines, following terminology of Verschelde, Goubault
424 & Vincx (1998).
- 425
- 426 (**): the original description of *D. tenuispiculum* lacks data on longitudinal ridge number.
427 Reported counts: 12–20 in *D. cephalata* sensu Chitwood (1936), 16 by Gerlach (1950), 24 by
428 Gerlach (1963), 12 by Boucher (1975), 15 by Platt & Warwick (1988), and 16–18 by Fadeeva,
429 Mordukhovich & Zograf (2016).

430

431 Discussion

432

433 Gerlach (1950) described *Desmodora schulzi* and, years later in his revision (Gerlach,
434 (1963)), synonymized *Desmodora schulzi* with *Heterodesmodora hirsuta* Chitwood, 1936,
435 establishing the new combination *Desmodora hirsuta* (Chitwood, 1936). Vincx (1983) redescribed
436 *Desmodora schulzi*, disagreeing with Gerlach (1963) proposed synonymy and indicating the
437 characteristics that differentiate *Heterodesmodora hirsuta* from *Desmodora schulzi*. Later,
438 Verschelde, Goubault & Vincx (1998) transferred both species to the genus *Desmodorella*,
439 considering them valid and distinct from each other. We agree with Vincx (1983) and Verschelde,
440 Goubault & Vincx (1998) in treating *Desmodorella hirsuta* and *Desmodorella schulzi* as distinct
441 species, and therefore not synonymous. When comparing females of both species, it is possible to
442 note that, with the exception of the total body length and the de Man ratio “c”, other features and
443 body proportions differ (see the comparison between these taxa in the discussion section in Vincx,
444 (1983)). Additionally, although both species share the number of longitudinal rows of spines (*D.*
445 *hirsuta*: 10 rows; *D. schulzi*: 8–10 rows), this feature is not sufficient to synonymize the species.

Similar to the aforementioned species, *D. sanguinea* also has 10 longitudinal rows of spines, and is easily distinguished from *D. schulzi* by comparing the characteristics present in males (spicules length, morphology of the gubernaculum and precloacal supplements). However, we disagree with Vincx (1983) and Verschelde, Goubault & Vincx (1998) regarding the validity of *D. hirsuta*. Since this species was described based on a female (Chitwood, 1936), making it difficult to distinguish it from other *Desmodorella* species, we believe that there is no sustainable evidence to consider it as a valid species. Here, we formally suggest that *Desmodorella hirsuta* (Chitwood, 1936) Verschelde, Goubault & Vincx, 1998 be regarded as a *nomen dubium*.

To develop the dichotomous key, the main characteristics that, together, effectively helped distinguish the *Desmodorella* species were: morphology and number of turns of the amphidial fovea; spicule length (short or elongated) and morphology; presence or absence of vacuoles in the head capsule, as well as in the rings along the body; presence or absence and morphology (elongated or short) of the subcephalic setae; presence or absence of two pairs of lateral rows of more distinct spines, among the other rows of spines (referred to by Verschelde, Goubault & Vincx (1998) as “false lateral alae”); number of longitudinal rows of somatic setae; number of longitudinal rows of ridges or spines; and morphology of the precloacal supplements. Although these are relevant characteristics for species identification/differentiation, the presence or absence of subcephalic setae and the number of longitudinal rows of ridges or spines should be analyzed with caution. Subcephalic setae can be lost during specimen preparation, and their presence, in some cases such as in *D. filispiculum*, is inferred from the visualization of the insertion point of the setae (Lorenzen, 1976). The number of longitudinal rows of ridges or short spines can often be difficult to determine, especially through optical microscopy, as mentioned by Verschelde, Goubault & Vincx (1998) when describing *D. balteata*. Despite providing SEM analyses, Verschelde, Goubault & Vincx (1998) did not mention the number of longitudinal rows of spines that occur in *D. balteata* and *D. spineacaudata*. When redescribing *D. tenuispiculum*, Fadeeva, Mordukhovich & Zograf (2016) reported that the visualization of rows was only possible through SEM analyses. Although it was possible to visualize the rows of ridges in some paratypes of *D. parabalteata* **sp. nov.**, the SEM analysis allowed us to demonstrate the configuration of these structures more clearly. However, it was not possible to precisely determine the number of rows that occur in this species, with variation in the number of rows along the body (a greater number of rows in the widest part compared to the median region where the body narrows), along with the occurrence of discontinuous rows. The literature on *D. tenuispiculum* records a large variation in the number of longitudinal rows of ridges present in this species. When describing *D. tenuispiculum*, Allgén (1928) did not indicate the number of rows present in the species. In subsequent redescriptions, the number of rows varied between 12 and 24 (Chitwood, 1936; Gerlach, 1950 and 1963; Boucher, 1975; Platt & Warwick, 1988; Fadeeva, Mordukhovich & Zograf, 2016). Similarly, when redescribing *D. schulzi*, Vincx (1983) reported the presence of 12 longitudinal rows of hair-like spines along the body, while the original description (Gerlach, 1950) reported that there are 8–10 rows. These variations may be due to the difficulty in visualizing and determining the number of rows or may reflect an intraspecific variation regarding this feature. Therefore, it is extremely important that the characteristics found in *Desmodorella* species are analyzed together to determine and identify the species.

Desmodorella cornuta **sp. nov.** possesses a protuberant horn-shaped cuticular projection positioned dorsally in the pharyngeal region. This feature is unique among the *Desmodorella*

species but can be observed in the Desmodoridae genus *Spinonema* Larrazábal-Filho *et al.*, 2019. This genus encompasses species that possess a strongly cuticularized dorsal spine located in the pharyngeal region. However, *Spinonema* species have C-shaped anteriorly oriented lateral alae (without spines) and spicules that may possess a velum. *Desmodorella cornuta* **sp. nov.**, on the other hand, has two pairs of lateral rows with more distinct spines, among the other rows of spines and spicules lacking velum, a combination of characteristics typically found in representatives of the genus *Desmodorella*. The genus *Spinonema* was originally described from specimens found in sediment samples collected in the Potiguar Basin, northeastern coast of Brazil, the same type locality as *Desmodorella cornuta* **sp. nov.** We believe that the occurrence of similar structures in different genera of Desmodoridae may reflect a process of adaptive convergence. The occurrence of a protuberant horn-shaped cuticular projection in the pharyngeal region was included in the diagnosis of the genus.

This contribution expands knowledge of the biodiversity of the genus *Desmodorella*, includes new diagnostic characteristics, updates the list of valid species and highlights the main characteristics that should be analyzed together to distinguish the species of the genus.

Acknowledgements

The authors express their sincere gratitude to the project titled 'Evaluation of benthic and planktonic biota in the offshore portion of the Potiguar and Ceará basins' conducted by the Brazilian oil company Petrobras, particularly to its coordinator, Professor Dr. Paulo Jorge Parreira dos Santos from UFPE, for granting them the opportunity to examine the material associated with this project. We express our gratitude to the Departamento de Zoologia, UFPE for the scanning electron microscopy facilities (FACEPE APQ-0522-2.04/19) at Laboratório Avançado de Microscopia e Imagem (LAMI-UFPE) of the Núcleo de Prospecção e Gestão da Biodiversidade do Nordeste, Universidade Federal de Pernambuco. We would like to thank Dra. Hianna Arely Milca Fagundes Silva, UFPE, for the technical support in obtaining the SEM photographs. The authors express their gratitude to the IAM-FIOCRUZ Core Facilities for providing access to Electron Microscopy Services.

References

- Abolafia J. 2015. A low-cost technique to manufacture a container to process meiofauna for scanning electron microscopy. *Microscopy research and technique*, 78, 771–776. DOI 10.1002/jemt.22538
- Allgén, CA. 1928. Freilebende marine Nematoden von den Campbell- und Staten-inseln. *Nyt Magazin for Naturvidenskaberne, Christiania* LXVI, 261–309.
- Allgén, CA. 1929. Über einige antarktische freilebende marine Nematoden. *Zoologischer Anzeiger*, 84, 126–140.

- 536 Annapurna, C, Bhanu, CV, Srinivasa, RM, Sivalakshmi, MV, Cooper, LMG & Kasivishweshwara,
537 RY. 2012. Free living nematodes along the continental slope off Northeast coast of India. *Journal*
538 *of the Marine Biological Association of India*, 54 (2), 52–60.
539 DOI 10.15373/22778179/NOV2013/173
540
541 Boucher, G. 1975. Nématodes des sables fins infralittoraux de la Pierre Noire (Manche
542 occidentale). I. Desmodorida. *Bulletin du Muséum National d' Histoire Naturelle*, 3^e série, 285,
543 Zoologie 195.
544
545 Chitwood, BG. 1936. Some marine nematodes from North Carolina. *Proceedings of the*
546 *Helminthological Society of Washington*, 3 (1), 1–16.
547
548 Cobb, NA. 1920. One hundred new nemas (type species of 100 new genera). *Contributions to a*
549 *Science of Nematology*, Cobb, 9, 217–343.
550
551 Cobb, NA. 1933. New nemic genera and species, with taxonomic notes. *Journal of Parasitology*,
552 10, 81–94.
553 DOI 10.2307/3272166
554
555 De Coninck LA. 1965. Classe des Nématodes: Systematique des nématodes et sous classe des
556 Adenophorea. In: Grassé PP., ed. *Traité de Zoologie: Anatomie, Systématique, Biologie*, 4, 586–
557 681.
558
559 De Grisse AT. 1969. Redescription ou modification de quelques techniques utilisées dans l'étude
560 des nématodes phytoparasitaires. *Mededelingen van de Rijksfakulteit Landbouwwetenschappen te*
561 *Gent*, 34, 351–369.
562
563 Decraemer W, Smol N. 2006. Orders Chromadorida, Desmodorida and Desmoscolecida. In:
564 Eyualet A., Traunspurger W. & Andrassy I., eds. *Freshwater Nematodes: Ecology and*
565 *Taxonomy*. CABI Publishing, Wallingford, pp. 497–573.
566 DOI 10.1079/9780851990095.0497
567
568 de Man, JG. 1880. Die einheimischen, frei in der reinen Erde und im süßen Wasser lebenden
569 Nematoden monographisch bear-beitet. Vorläufiger Bericht und descriptiv-systematischer Theil.
570 Tijdschrift der Nederlandsche Dierkundige Vereeniging 5:1–104.
571
572 de Man, JG. 1889. Troisième note sur les Nématodes libres de la Mer du Nord et de la Manche.
573 *Mémoires de la Société Zoologique de France*, 2, 182–216.
574
575 Fadeeva, N, Mordukhovich, V & Zograf, J. 2016. Free-living marine nematodes of Desmodorella
576 and Zalonema (Nematoda: Desmodoridae) with description of two new species from the deep sea
577 of the North Western Pacific. *Zootaxa*, 4175 (6), 501–520.
578 DOI 10.11646/zootaxa.4175.6.1
579
580 Filipjev IN. 1922. New data about free-living nematodes of the Black Sea. *Transactions of*
581 *Stavropol Agricultural Institute*, 1, 83–184.

- Gagarin, VG & Nguyen, VT. 2003. Two new species of free-living nematodes from the Cau River (Vietnam). *Zoologicheskii Zhurnal*, 82, 1418–1425. [in Russian]
- Gerlach, SA. 1950. Über einige Nematoden aus der Familie der Desmodoriden. *Zoologischer Anzeiger*, 145, 178–198.
- Gerlach, SA. 1963. Freilebende Meeresnematoden von den Malediven II. *Kieler Meeresforsch*, 19, 67–103.
- Gerlach SA, Riemann F. 1973/1974. The Bremerhaven Checklist of Aquatic Nematodes: A catalog of Nematoda Adenophorea excluding the Dorylaimida. *Veröffentlichungen des Instituts für Meeresforschung in Bremerhaven*, Supplement 4.
- Inglis, WG. 1968. Interstitial nematodes from St. Vincent's Bay, New Caledonia Expédition française sur les recifs coralliens de la Nouvelle Calédonie. *Editions de la Fondation Singer-Polignac, Occasional Publications*, 2, 29–74.
- Ingels, J, Vanhove, S, De Mesel, I & Vanreusel, A. 2006. The biodiversity and biogeography of the free-living nematode genera *Desmodora* and *Desmodorella* (family Desmodoridae) at both sides of the Scotia Arc. *Polar Biology*, 29, 936–949.
DOI 10.1007/s00300-006-0135-4
- Inglis, WG. 1983. An outline classification of the Phylum Nematoda. *Australian Journal of Zoology*, 31, 243–255.
DOI 10.1071/ZO9830243
- Jensen P. 1985. The nematode fauna in the sulphide-rich brine seep and adjacent bottoms of the East Flower Garden, NW Gulf of Mexico. 1. Chromadorida. *Zoological Scripta* 14:247–263
DOI 10.1007/BF00392509.
- Larrazábal-Filho, AL. 2020. Estudo taxonômico da família Desmodoridae (Nematoda: Desmodorida) na Bacia Potiguar, Rio Grande do Norte, Brasil. Doctoral thesis, Universidade Federal de Pernambuco, Recife, 147 pp. *Available at* <https://repositorio.ufpe.br/handle/123456789/38052>
- Larrazábal-Filho, AL, Neres, P, Silva. MC & Esteves, AM. 2019. A new genus with three new species of free-living marine nematodes of the subfamily Desmodorinae (Nematoda: Desmodoridae), from the continental shelf off northeastern Brazil. *Zootaxa*, 4615 (2), 321–342.
DOI 10.11646/zootaxa.4615.2.5
- Leduc, D & Zhao, Z. 2016. Phylogenetic relationships within the superfamily Desmodoroidea (Nematoda: Desmodorida), with descriptions of two new and one known species. *Zoological Journal of the Linnean Society*, 176, 511–536.

- 627 Lorenzen, S. 1976. Desmodoridae (Nematoden) mit extrem langen Spicula aus Südamerika.
628 *Mitteilungen aus dem Instituto Colombo-Alemán de Investigaciones Científicas*, 8, 63–78.
- 629
- 630 Moura JR. 2013. Nematofauna no talude da Bacia de Campos, Rio de Janeiro, Brasil: uma
631 avaliação batimétrica. Masters dissertation, Universidade Federal de Pernambuco, Recife, 66 pp.
632 Available at <https://repositorio.ufpe.br/handle/123456789/10537> (accessed 19 February 2025).
- 633
- 634 Murphy, DG. 1962. Three undescribed nematodes from the coast of Oregon. *Limnology and*
635 *Oceanography*, 7 (3), 386–389.
- 636
- 637 Nemys eds. 2025. Nemys: world database of nematodes. Available at <https://nemys.ugent.be>
638 (accessed on 19 February 2025).
- 639
- 640 Pearse AS. 1942. *An introduction to parasitology*. Springfield: Charles C. Thomas.
- 641
- 642 Platt, HM & Warwick, RM. 1988. Free-living Marine Nematodes (Part II British Chromadorids)
643 *Synopses of the British fauna (New series)*, 38, 502 pp.
- 644
- 645 Riera, R, Núñez, J & Brito, MC. 2012. Three new records of Desmodorids (Nematoda,
646 Desmodoridae) from sandy seabeds of the Canary islands. *Orsis*, 26, 9–19.
- 647
- 648 Silva MC. 2012. Contribuição de cânions na biodiversidade da nematofauna do Atlântico Sul –
649 Bacia de Campos, Rio de Janeiro, Brasil. Doctoral thesis, Universidade Federal de Pernambuco,
650 Recife, 431 pp. Available at <https://repositorio.ufpe.br/handle/123456789/10252> (accessed 19
651 February 2025).
- 652
- 653 Somerfield PJ, Warwick RM, Moens T. 2005. Meiofauna techniques. In: Eleftheriou A. &
654 McIntyre A., eds. *Methods for the Study of Marine Benthos*. 3rd Edition. Blackwell, Oxford, pp.
655 229–272.
- 656 DOI 10.1002/9780470995129.ch6
- 657
- 658 Southern, R. 1914. Clare Island Survey. Nematelmia, Kinorhyncha, and
659 Chaetognatha. *Proceedings of the the Royal Irish Academy*. 31, part 54, section 3, 1–80.
- 660
- 661 Steiner, G. 1916. Freilebende Nematoden aus der Barentssee. *Zoologische Jahrbücher, Abteilung*
662 *für Systematik, Geographie und Biologie der Tiere*, 39, 511–664.
- 663
- 664 Verschelde, D, Goubault, N & Vincx, M. 1998. Revision of Desmodora with descriptions of new
665 desmodorids (Nematoda) from hydrothermal vents of the Pacific. *Journal of the Marine Biological*
666 *Association of the United Kingdom*, 78, 75–112.
- 667 DOI 10.1017/S0025315400039977
- 668
- 669 Vincx, M. 1983. Redescription and ontogenetic study of *Desmodora schulzi* Gerlach 1950. *Biol.*
670 *Jb. Dodonaea*, 51, 171–179.
- 671
- 672

673 Wieser, W. 1954. Free-living marine nematodes. II. Chromadoroidea. *Lunds Universitets*
 674 *Årsskrift. N. F. Avd.* 50, 1–148.

675

676

Figure 1

Figure 1. *Desmodorella cornuta* sp. nov. Holotype male and paratype female 1.

Holotype male and paratype female 1. Holotype male: (A) whole body overview; (B) cuticle details - 1: at the pharynx level; 2: at the beginning of the false lateral alae; 3: at the end of the of the false lateral alae, (C) anterior region, (D) buccal cavity, (E) spicule and gubernaculum. Paratype female 1: (F) whole body overview, (G) anterior region.

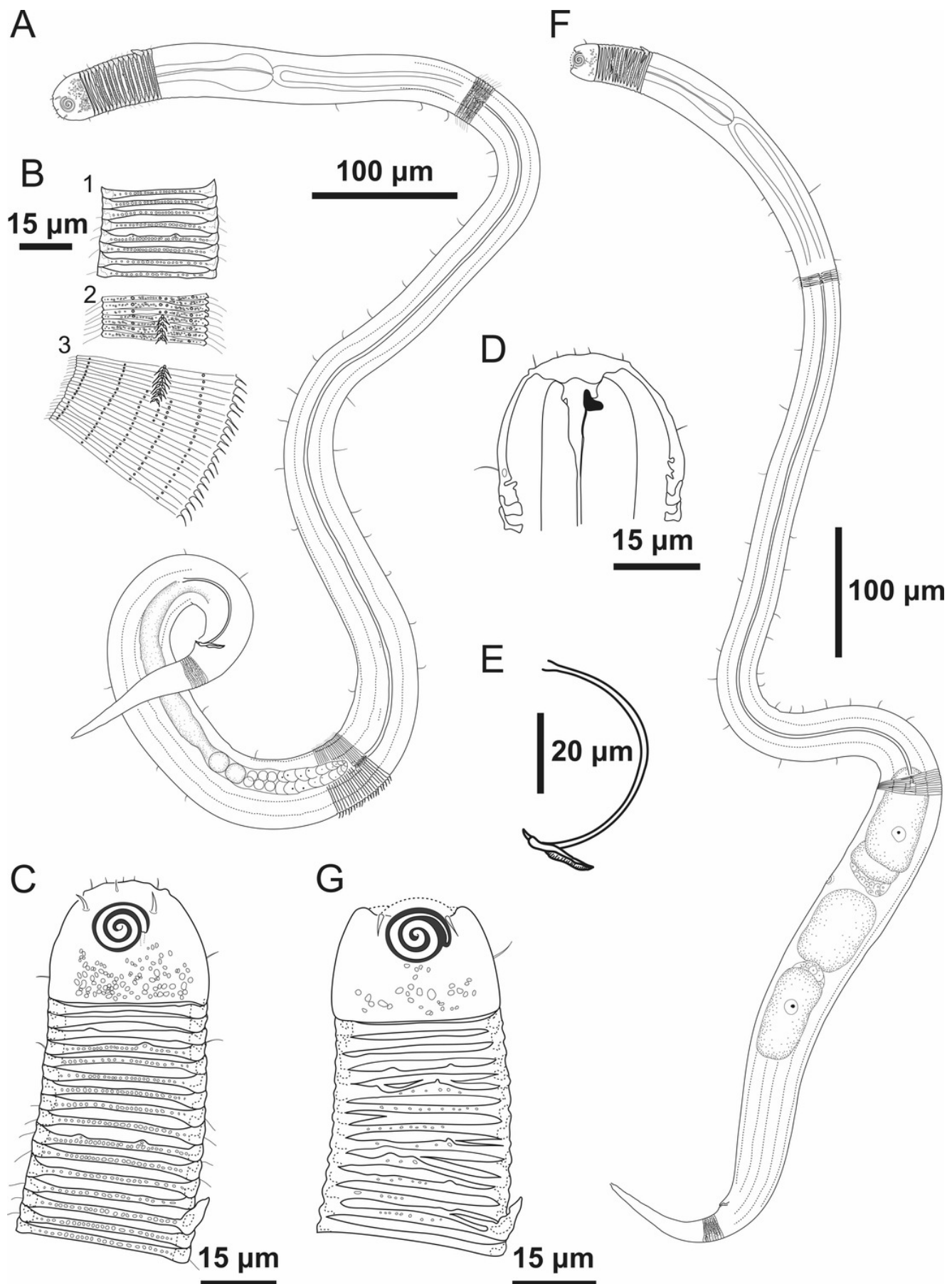


Figure 2

Figure 2. *Desmodorella cornuta* sp. nov. Holotype male.

(A) Anterior end (cs: cephalic setae; amph: amphidial fovea), (B) anterior end (ils: inner labial setae; ols: outer labial setae; dt: dorsal tooth; hscp: horn-shaped cuticula projection), (C) anterior region, (D) beginning of the false lateral alae, (E) end of the of the false lateral alae, (F) cuticular hair-like spine and somatic setae (ss), (G) spicule, (H) gubernaculum (gub), (I) tail.

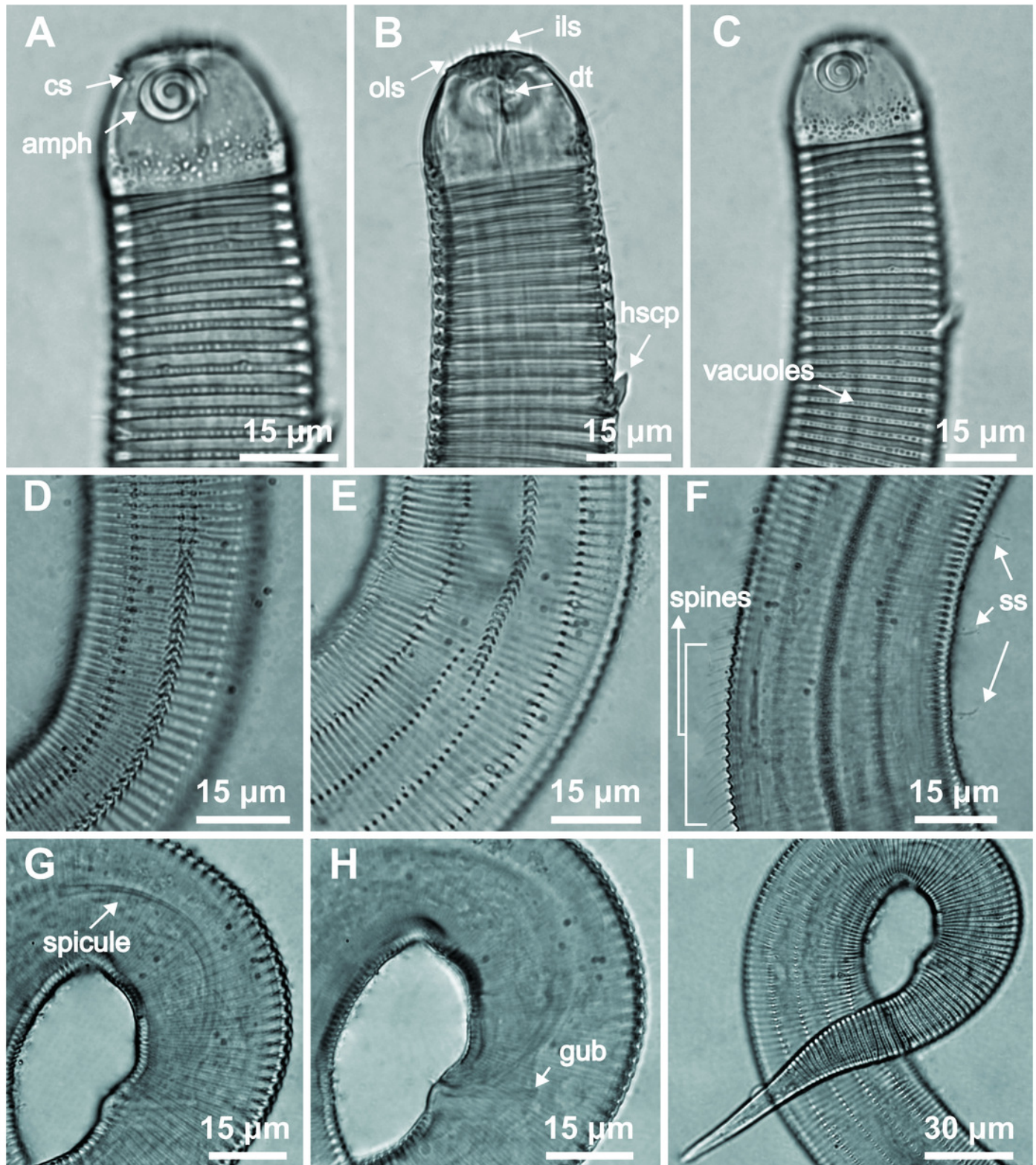


Figure 3

Figure 3. *Desmodorella cornuta* sp. nov. Paratype female 1.

(A) anterior region (cs: cephalic setae; amph: amphidial fovea); (B) anterior region (dt: dorsal tooth; hscp: horn-shaped cuticular projection), (C) beginning of the false lateral alae; (D) end of the of the false lateral alae, (E) reproductive system (V: vulva; ant. ov.: anterior ovary), (F) posterior ovary (post. ov.), (G) tail.

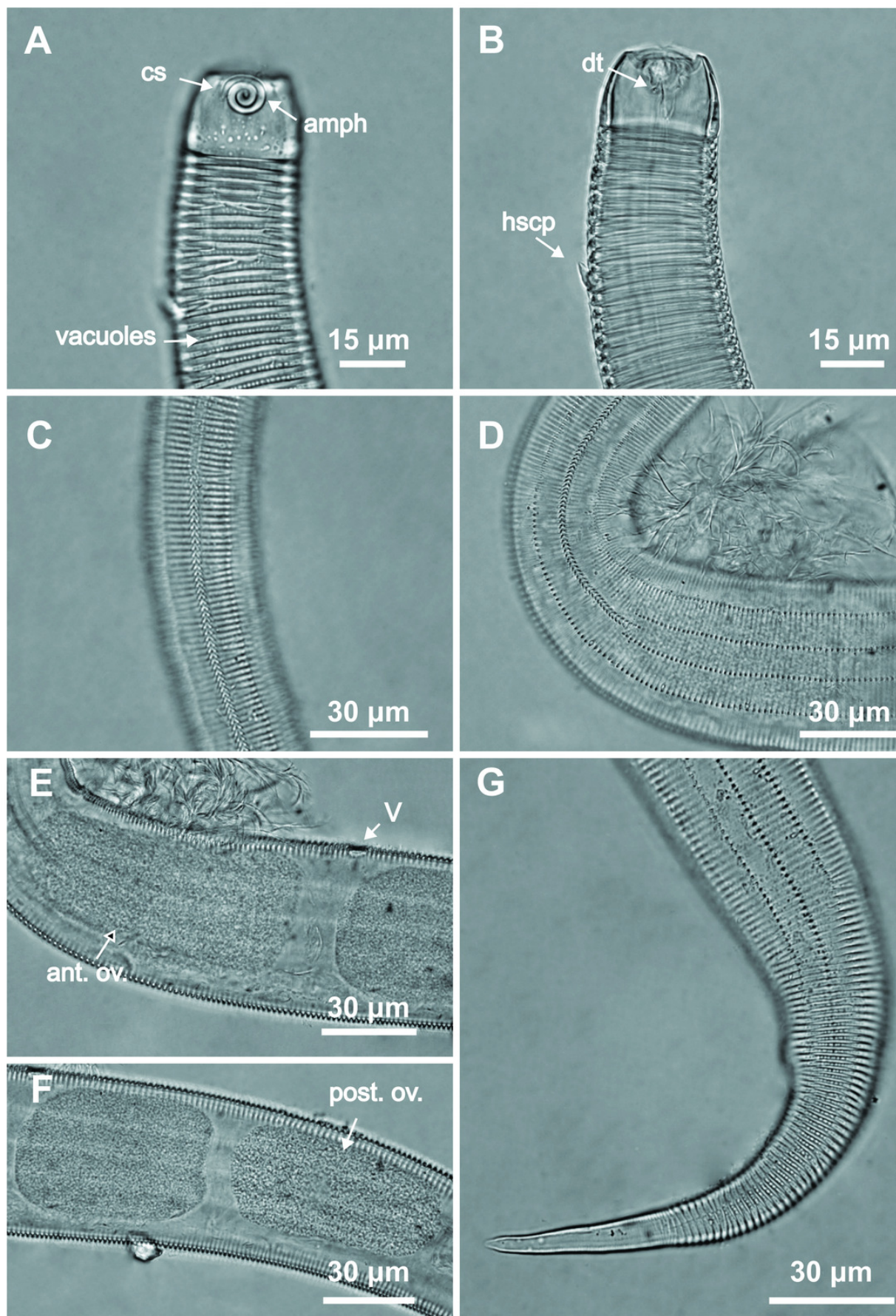


Figure 4

Figure 4. *Desmodorella parabalteata* sp. nov. Holotype male and paratype female 1.

Holotype male and paratype female 1. Holotype male: (A) whole body overview; (B) anterior end (C) buccal cavity, (D) posterior end, Paratype female 1: (E) whole body overview, (F) anterior end.

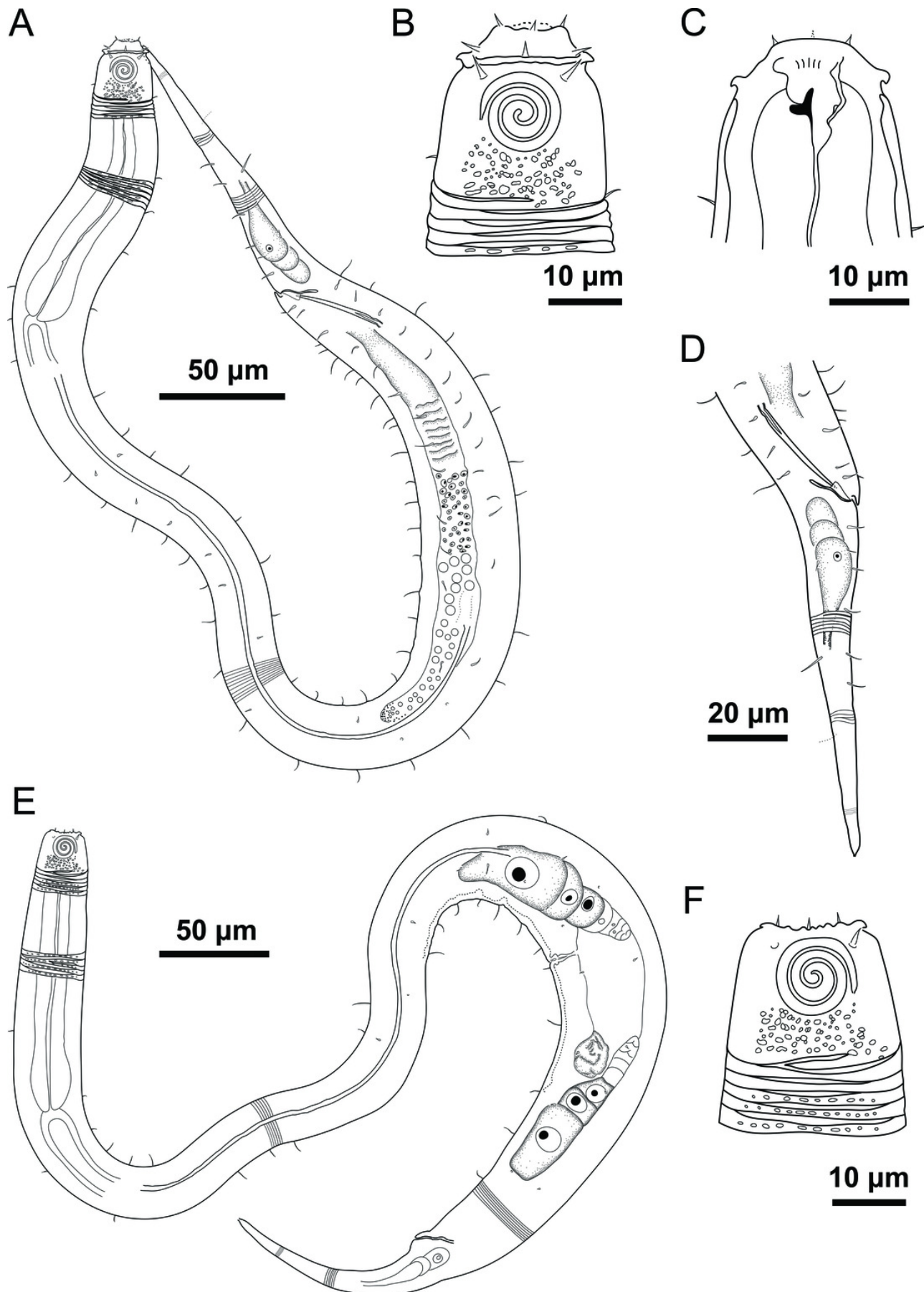


Figure 5

Figure 5. *Desmodorella parabalteata* sp. nov. Holotype male and male paratype 9.

Holotype male: (A) Anterior end (ols: outer labial setae; cs: cephalic setae; amph: amphidial fovea), (B) anterior end (dt: dorsal tooth; hscp: horn-shaped cuticula projection), (D and E) rows of somatic setae, (F) tail, (G) spicule (spic) and gubernaculum (gub). Male paratype 9: (C) longitudinal rows of ridges.

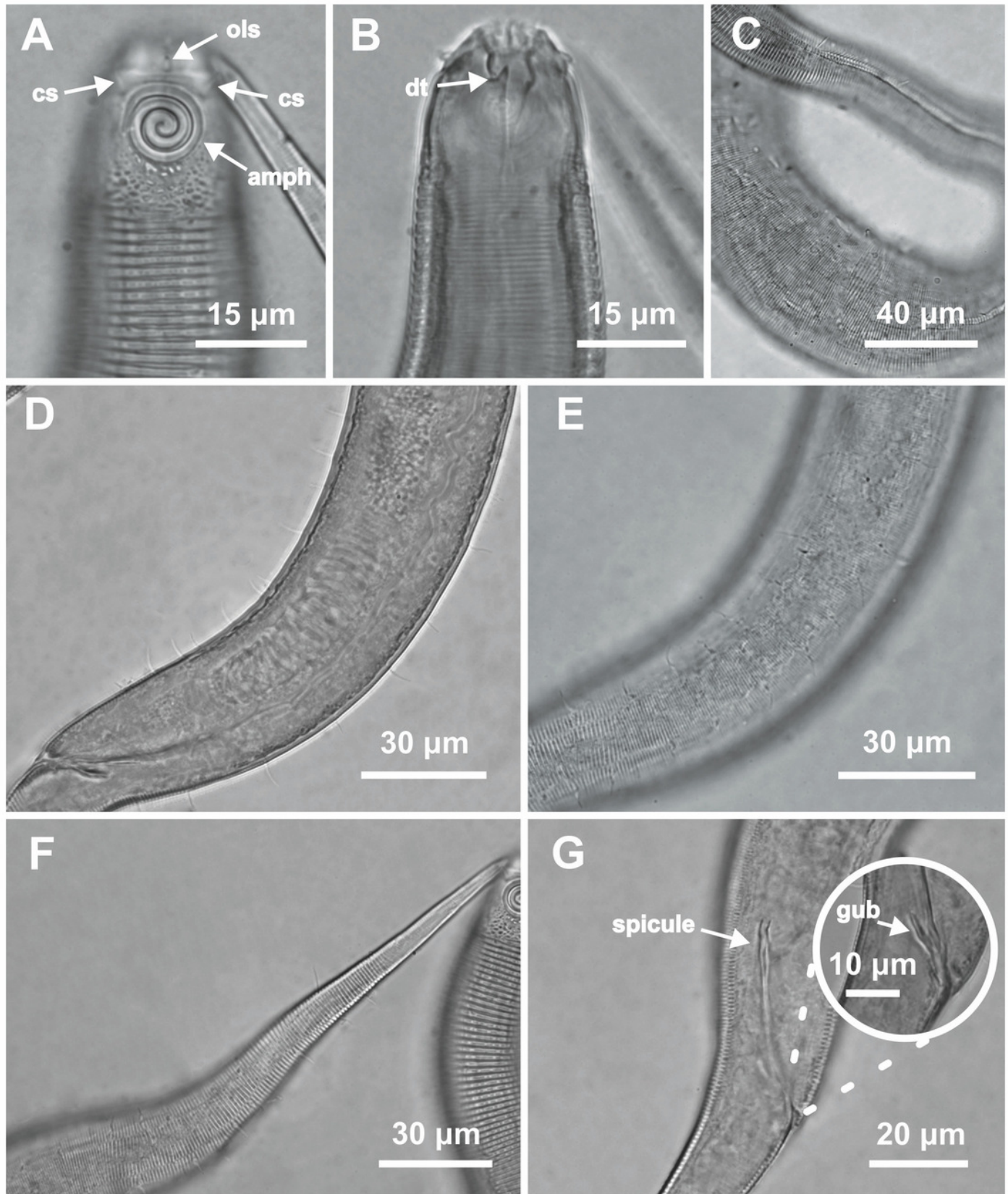


Figure 6

Figure 6. *Desmodorella parabalteata* sp. nov. Female paratype 1 and female paratype 9.

Female paratype 1: (A) anterior end (amph: amphidial fovea), (B) buccal cavity (dt: dorsal tooth), (D) vulva region (V: vulva), (E) anterior ovary (ant. ov.), (F) posterior ovary (post. ov.).

Female paratype 9: (C) longitudinal rows of ridges.

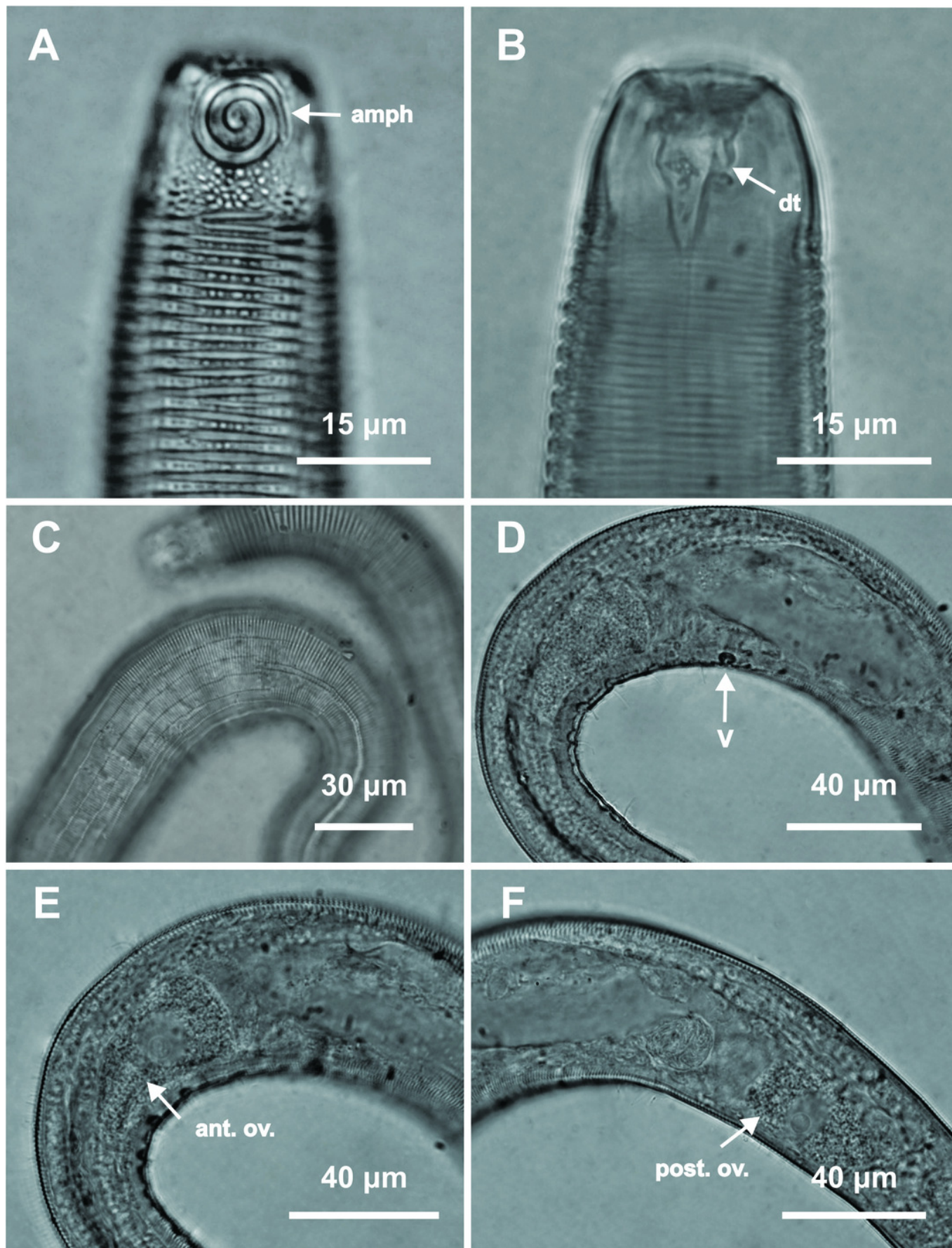


Figure 7

Figure 7. *Desmodorella parabalteata* sp. nov. Male paratypes 8 and 10, SEM photographs.

Paratype male 10: (A) whole body overview; (B) anterior end (ils: inner labial setae; ols: outer labial setae; amph: amphidial fovea); (E) false lateral alae and longitudinal rows of ridges; (F) beginning of the false lateral alae. Paratype male 8: (C) anterior end (cs: cephalic seta; amph: amphidial fovea; ads: additional setae); (D) cuticular ornamentation at the pharynx level (ss: somatic setae).

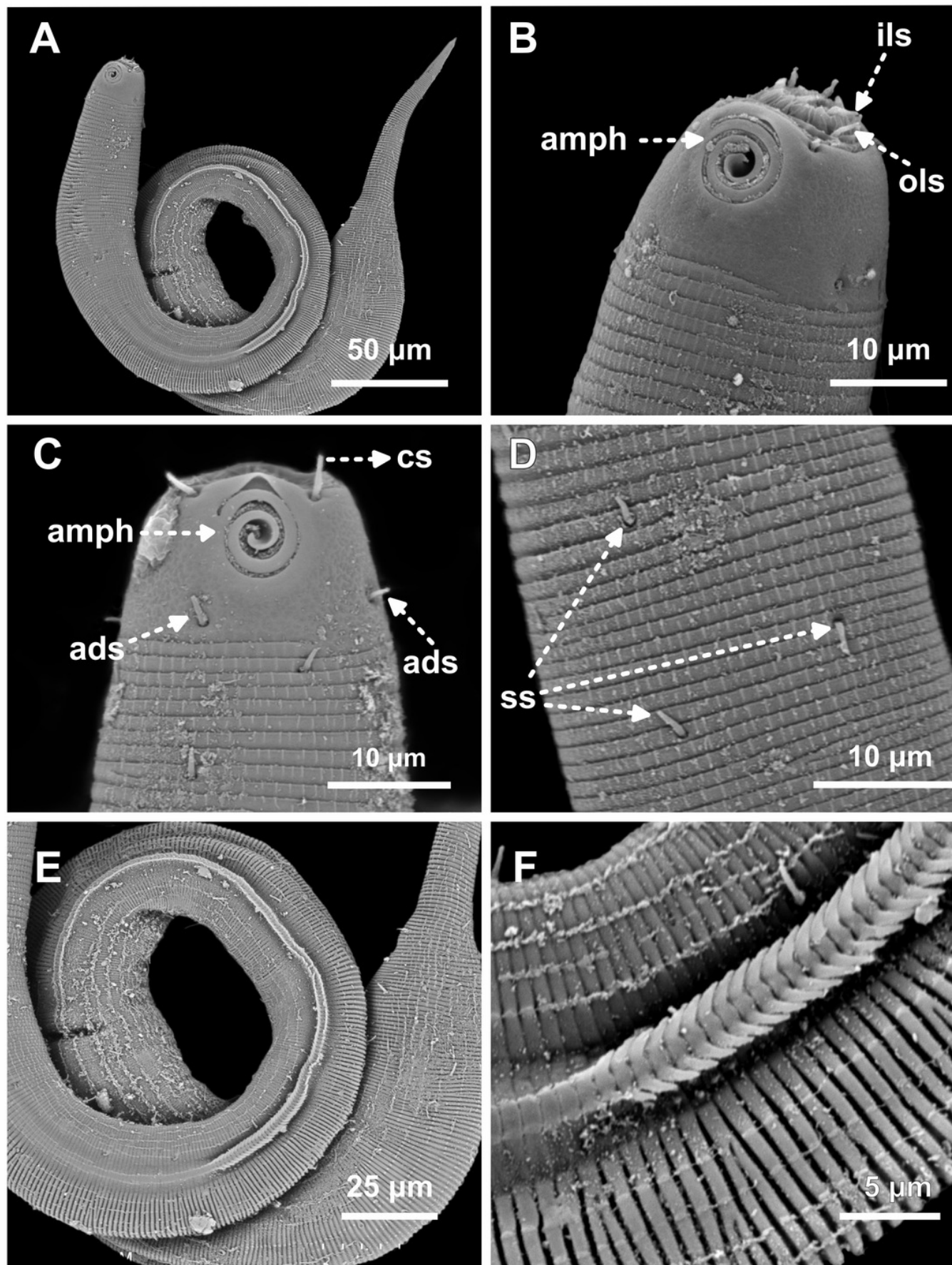


Figure 8

Figure 8. *Desmodorella parabalteata* sp. nov. Paratype female 3, SEM photographs.

(A) whole body overview; (B) anterior end (amph: amphidial fovea; ss: somatic setae); (C) false lateral alae and longitudinal rows of ridges; (D) posterior end of the false lateral alae.

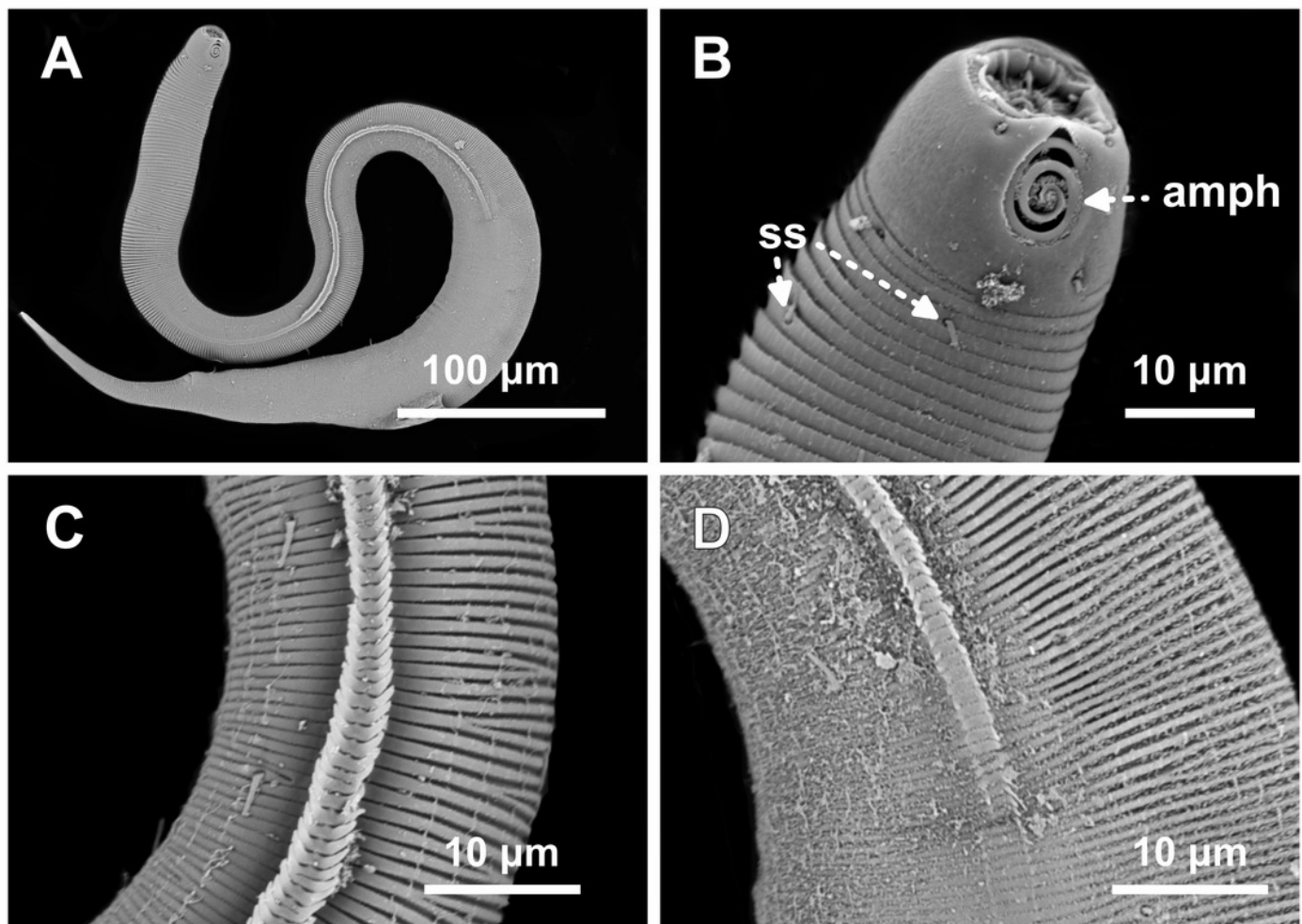


Table 1 (on next page)

Table 1. Collection stations, respective coordinates, depth, and collection gear. The samples were collected from the continental shelf in northeast Brazil, South Atlantic.

Project	Station	Latitude (S)	Longitude (W)	Depth (m)	Gear
Evaluation of benthic and planktonic biota in the offshore portion of the Potiguar and Ceará basins	ME2B2 R1	05°02'29.6"	36°23'11.9"	8.5	van Veen grab
	ME2B2 R3	05°02'30.3"	36°23'12.3"	8.5	
	ME2B3 R2	05°01'12.4"	36°23'27.6"	8.1	
UFPE S.O.S. SEA	14	10°07'05.7"	35°50'57.96"	63	Box-corer
	16	10°44'59.28"	36°25'32.88"	58	
	17	11°00'00.54"	36°49'58.98"	54	
	23	13°04'10.32"	38°25'46.98"	65	

Table 2 (on next page)

Table 2. Morphometric data of *Desmodorella cornuta* sp. nov. The measurements are expressed in micrometers, or if noted, as percentages or ratios. Not applicable (*); not available for measurement (-); a, b, c, c' = de Man's ratios (1880).

<i>Desmodorella cornuta</i> sp. nov.	Holotype (Male)	Male paratypes (n=2)	Paratype (Female 1)	Other female Paratypes (n=2)
Body length	1,254	1,107–1,092	1,221	1,014–1,074
Outer labial setae length	2	2–2.5	-	-
Cephalic setae length	3.5	3–4	3	3–4
Head diameter at level of the cephalic setae	21.5	20.5–22	23.5	18.5–23
Cephalic setae in relation to head diameter at the cephalic setae level (%)	16%	14–19.5%	13%	16%–17%
Distance from anterior end to amphidial fovea	3.5	6–6.5	-	2.5–4
Amphidial fovea diameter (maximum width)	11.5	11–11.5	11	11
Body diameter at level of the amphidial fovea	26.5	26–26.5	25.5	21.5–25.5
% of the amphidial fovea diameter in relation to corresponding body diameter	43%	42–50%	43%	43–51%
Pharynx length	145	142–143	142	137–138
Distance between the horn-shaped cuticular projection to anterior end	64,5	54.5–61.5	56	53.5–56
Length of horn-shaped cuticular projection	9	8.5–9	8	9
Position of the horn-shaped cuticular projection in relation to the pharynx length (%)	44%	38–43%	39%	39%–41%
Pharyngeal bulb diameter	17	19	13	19
Body diameter at level of the pharyngeal bulb	34.5	31	34.5	30–32
% of basal bulb diameter in relation to corresponding body diameter	49%	61%	38%	59–63%
Body diameter at the level of the pharynx end	35	29–30.5	35	30–31
Maximum body diameter	48	34–55	57	42–44.5
Anal or cloacal body diameter	30	26–27	25.5	24–25
Tail length	118	109.5–114	108.5	93–106
Length of spicule along arc	79	55–71.5	*	*
Length of spicule along cord	41	50–64	*	*
Length of gubernaculum	17	17	*	*
Length of gubernaculum in relation to length of spicule along arc (%)	21.5%	24%	*	*
Length of spicule along arc in relation to cloacal body diameter	2.6	2–2.6	*	*
Distance from anterior end to vulva	*	*	858	750–756
Position of vulva from anterior end (%)	*	*	70%	70–75%
Body diameter in vulva region	*	*	57	42–44.5
Anterior ovary length	*	*	81	155.5–159
Posterior ovary length	*	*	77.5	106–108
Reproductive system length	385.5	273	98.5	130–141
% of reproductive system in relation to body length	31%	25%	8%	12–14%
a	26	20–32	21	24
b	8.7	7.7	8.6	7.4–7.8
c	10.6	9.6–10	11.3	9.6–11.6
c'	4	4–4.4	4.3	4.4–3.7

Table 3 (on next page)

Table 3 . Comparison of species *Desmodorella cornuta* sp. nov. with morphologically similar species.

a, b, c = de Man's ratios (1880); parameter absent (-); parameter present (+). (*) = Two pairs of lateral rows of more distinct spines, among the other rows of spines (referred to by Verschelde, Gourbault & Vincx (1998) as "false lateral alae"). (**) = Protuberant horn-shaped cuticular projection positioned dorsally in the pharyngeal region.

	<i>Desmodorella curvispiculum</i>	<i>D. perforata</i>	<i>D. balteata</i>	<i>D. cornuta</i> sp. nov.
Body length (µm)	1004–1042	1850–1410	867–1078	1014–1254
a	17–26	28.6–32.4	15.7–26.3	32–20
b	7.3–7.6	7.4–8.8	5.9–7.6	7.4–8.7
c	10.9–11.5	11.6–13.5	8.7–12.3	9.6–11.6
Spicule length (µm)	76	52	85–65	55–79
False lateral alae*	-	-	+	+
Subcephalic setae	-	-	-	-
Horn-shaped cp. **	-	-	-	+

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Table 4(on next page)

Table 4. Morphometric data of *Desmodorella parabalteata* sp. nov. The measurements are expressed in micrometers, or if noted, as percentages or ratios.

Not applicable (*); not available for measurement (-); a, b, c, c' = de Man's ratios (1880).

<i>Desmodorella parabalteata</i> sp. nov.	Holotype (Male)	Male paratypes (n= 10)	Paratype (Female 1)	Other female paratypes (n= 9)
Body length	697.5	685.5–817.5	795	583–814.5
Outer labial setae length	3	2	2.5	2
Cephalic setae length	4	4	4	4
Head diameter at level of the cephalic setae	19.5	-	-	16–17
Cephalic setae in relation to head diameter at the cephalic setae level (%)	21%	-	-	24–25%
Distance from anterior end to amphidial fovea	6.5	5–7	-	5–8
Amphidial fovea diameter (maximum width)	11.5	10–11.5	11	9–12
Body diameter at level of the amphidial fovea	22	20–23	21.5	19–23
% of the amphidial fovea diameter in relation to corresponding body diameter	52%	43–55%	51%	43–59%
Pharynx length	123	113.5–133.5	131.5	109–129
Pharyngeal bulb diameter	18	19–25.5	21.5	17–23
Body diameter at level of the pharyngeal bulb	32.5	30.5–36	31	28–32.5
% of basal bulb diameter in relation to corresponding body diameter	55%	62–73%	70%	56–75%
Body diameter at the level of the pharynx end	29	24.5–34	30	25–32
Maximum body diameter	40.5	37–51	51	39–51
Anal or cloacal body diameter	21	17.5–24.5	23	19–23
Tail length	113	88–113.5	99	72–107
Length of the non-annulated tail end	13	13–17	15.5	9–23.5
Length of spicules	36.5	25–41.5	*	*
Length of gubernaculum	14.5	12–18	*	*
Length of gubernaculum in relation to length of spicules (%)	40%	37–56%	*	*
Length of spicule along arc in relation to cloacal body diameter	1.7	1.4–1.8	*	*
Distance from anterior end to vulva	*	*	510	389.5–525
Position of vulva from anterior end (%)	*	*	64%	64–68%
Body diameter in vulva region	*	*	51	39–51
Anterior ovary length	*	*	148	108–270
Posterior ovary length	*	*	130	97–271.5
Reproductive system length	232.5	217–317	210	142.5–229.5
% of reproductive system in relation to body length	33%	27–39%	26%	23–28%
a	17	14–21	16	13–17
b	6	6–6.5	6	5–6
c	6	7–8	8	7–9
c'	5	4–6	4	4–5