

# cpSNP discovery and genotyping for a *Pinus taeda* breeding population with targeted comparison to related conifers

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## **ABSTRACT**

Pinus taeda (Loblolly pine) is the most important commercial tree species in the southern United States and a significant non-native plantation species in China. Its genetic improvement program has been implemented in South China for 30 years. In this study, the chloroplast (cp) genome of *P. taeda* was sequenced, assembled, and compared with other available chloroplast genomes of Pin aceae using BLAST. Codon usage among 33 species of Pinaceae was analyzed using the relative synonymous codon usage (RSCU) value. The results were then visualized using the pheatmap v1.0.10 in R. The rates of nonsynonymous (Ka) and synonymous (Ks) substitutions in the chloroplast genomes among five species of Pinus were estimated using the seqinr package in R. Additionally, selected single nucleotide polymorphisms (SNPs) were used to genotype 33 individuals from the *P. taeda* breeding population. The *P. taeda* cp genome is 121,530 bp, with certain regions (e.g., ycf1 and ycf2) showing lower sequence conservation compared to other Pinaceae species. Codon usage analysis revealed that codons ending in G or C were not prevalently used, with significant differences in natural selection pressure on chloroplast genes between three species (P. taiwanensis, P. thunbergii, and P. koraiensis) and the other 30 species. Phylogenomic analysis using 36 cp genomes (representing 25 species) resolved Pinus into two subgenera, with P. taeda clustered with P. rigida within the diploxylon pines. Notably, ycf1-based phylogenetic analysis recovered a topology highly congruent (normalized RF = 0.15) with whole-plastome phylogenies. This study validates the single-copy gene ycf1 as a robust and low-cost phylogenetic marker for conifer genus-level reconstruction. The identified SNPs provide valuable molecular markers for genotyping individuals in P. taeda breeding programs, supporting germplasm characterization and management.

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# **INTRODUCTION**

The study of the chloroplast (cp) genome has become a valuable and widely used approach for evolutionary, taxonomic, and phylogenetic analyses of plants. This is due to its simple structure, highly conserved sequence, and single parental (maternal or paternal) inheritance characters (*Wolfe, Li & Sharp, 1987; Daniell et al., 2016; Jakobsson et al., 2007; Nock et al., 2011; Wu & Chaw, 2014; Mao et al., 2023*). Unlike most angiosperms, cpDNA is paternally inherited in all conifers (*Neale, Marshall & Sederoff, 1989; Szmidt, Aldén & Hällgren, 1987; Wagner et al., 1987; Neale, Wheeler & Allard, 1986*). Conifers hold immense ecological and economic value. Understanding the organization of their chloroplast genomes is therefore significant for phylogenetic studies and resolving evolutionary relationships. This knowledge also supports efforts to enhance the commercial use of conifers.

Loblolly pine is the most important commercial tree species in the southern United States due to its fast growth, desirable wood properties, and broad adaptability (*McKeand et al.*, 2003). In China, it is a major non-native plantation species, especially in southern regions (*Gwaze*, *Byram & Lowe*, 2001). Genetic improvement programs for *P. taeda* in China began in the early 1980s. In the 1990s, comprehensive progeny and provenance tests were initiated, leading to advances in selective breeding (*Zhong*, *Chen & Huang*, 1995). Since 2002, China has implemented a breeding strategy (*El-Kassaby & Lstibůrek*, 2009). The paternal inheritance of the conifer cp genome enables paternity verification when the maternal parent is known, thereby facilitating precision breeding programs. Although cp genomes are generally conserved, recent studies have identified variable regions. For example, nine mutational loci were detected in *ycf1* of *P. taeda* (*Li*, 2012). Recent studies have demonstrated that *ycf1* exhibits high variability (*Jiang et al.*, 2017; *Li et al.*, 2020; *Chen et al.*, 2020; *Zhang et al.*, 2023). Such variability, while contrasting with overall cpDNA conservation, offers valuable markers for fine-scale genetic analyses.

While the complete chloroplast genome of P. taeda has been assembled and its broad structure and placement resolved ( $Asaf\ et\ al.$ , 2018), three knowledge gaps remain. First, there is a need for population-level chloroplast single nucleotide polymorphism (cpSNP) discovery within breeding material. Second, a systematic comparison of codon-usage bias across Pinaceae is lacking. Third, the single-copy ycf1 gene has not been empirically assessed as a cost-effective phylogenetic marker. To address these gaps, we generated a high-quality P. taeda cp genome and compared codon-usage patterns across 33 Pinaceae species. We quantified Ka/Ks in five Pinus species, identified 71 novel cpSNPs in 33 breeding individuals, and performed phylogenomic analyses using both whole plastomes and ycf1 across 36 samples (25 species). The resulting cpSNP panel differentiates 72% of breeding individuals, and ycf1 recapitulates the whole-genome tree (normalized RF = 0.15), demonstrating its utility for low-cost conifer phylogenetics and immediate application in germplasm management.

# **MATERIALS & METHODS**

# DNA extraction and sequencing

Fresh needles of *P. taeda* were collected from the Yingde Research Institute of Forestry in Guangdong Province, P.R. China (24°15'N, 113°25'E). Total cpDNA was extracted from approximately 100 g needles using a high-ionic-strength medium (*Bookjans, Stummann & Henningsen, 1984*). The integrity and purity of DNA were assessed by 1% agarose gel electrophoresis. The purity of the samples was further evaluated by measuring the OD260/280 ratio with a NanoDrop 2000 spectrophotometer (Thermo Fisher Scientific). The DNA concentration was quantified by an Invitrogen Qubit fluorometer. Libraries were prepared in accordance with the manufacturer's instructions with an average insert size of 350 bp. Purified cpDNA was sequenced on an Illumina HiSeq 2500 platform (Science Corporation of Gene, Guangzhou, China) under a paired-end 100 bp mode. Raw read quality was then assessed using fastp (v0.23.2) with the following parameters: a cut window size of 8 nucleotides and a qualified quality Phred score threshold of 20.

### De novo CP genome assembly

The chloroplast assembly software SPAdes (v3.15.5; *Bankevich et al.*, 2012) was used to assemble the chloroplast genome of loblolly pine, with K-mer length set to 79 and 97. The assembly's integrity was verified by mapping reads back to the consensus sequence using Burrows-Wheeler Aligner (BWA; v0.7.17-r1188) and SAMtools (v1.14). The *P. taeda* cp genome was compared with other available cp genomes of Pinaceae by using the CGView Comparison Tool (CCT; *Grant & Stothard*, 2008). Gene annotations were performed using the Clusters of Orthologous Groups (COGs) database. The Basic Local Alignment Search Tool (BLAST) was employed to align the other genomes to *P. taeda*, facilitating the identification of homologous sequences. The complete annotated genome was shown as a circular map using OrganellarGenomeDRAW (OGDRAW; *Greiner, Lehwark & Bock*, 2019). The adenine (A) and thymine (T) distributions were measured based on the results of the AT-skew equation, as follows: AT-skew = (A-T)/(A+T).

#### Codon usage

Codon usage, quantified as relative synonymous codon usage (RSCU) values (*Sharp & Li*, 1987), was determined for all protein-coding genes across the 33 Pinaceae species. Statistical analyses of RSCU distributions were performed, and results were visualized as a heatmap using the pheatmap package (v1.0.10) in R (*R Core Team*, 2022).

## Ka/Ks analysis

The rates of nonsynonymous (Ka) and synonymous (Ks) substitutions within the chloroplast genomes of five *Pinus* species were estimated using seqinR (v3.4-5; *Gouy et al.*, 1984; *R Core Team*, 2022). Histograms of these substitution rates were then created using ggplot2 (v3.0.0) to visualize the distribution of Ka and Ks values.

#### Single nucleotide polymorphisms analysis

There were 32 *P. taeda* cpDNA samples, selected from the Yingde Research Institute of Forestry in Guangdong Province, P.R. China, mixed and sequenced by the Illumina Hiseq

Table 1 Primers for Eighteen SNPs in Pinus taeda.												
No.	Name of Primer	Location	Length (bp)	Sequence (5'-3')	Length of GC% Primer (bp)		PCR Annealing Temperature	SNP				
1	F1	218	540	TTCCCACAACTTTCATACCA	20	40.00%	51	ycf1_98314				
1	R1	757	340	CTTTAGGATAAGCGGGTATT	20	40.00%	51	yeii_20014				
2	F2	277	423	GTCTGATTGGACCATTTGTA			51	ycf2_119153				
2	R2	699	423	TCAATCTTTATGGGTCCTAC	20	40.00%	31	yciz_119133				
3	F3	274	577	TGGTGGAGATGGTGAAGATG 20 50.00%		53	ycf1_101076, ycf1_101085, ycf1_101094, ycf1_101099,					
3	R3	850	5//	AAAGGCCATTAGACTCAGGT	20	45.00%	33	ycf1_101101, ycf1_101103, ycf1_101112, ycf1_101121, ycf1_101130, ycf1_101139, ycf1_101148, ycf1_101157				
	F4	306		GAATATCTAAACCCTGGACT	20	40.00%						
4	D.		576	1777 1 0 0777 0770 0 0 0 1 1 0 1 0 1		4= 000/	51	rpoC1_23167				
	R4	881		ATTAGCTTCTCCCGAACAGA	20	45.00%						
5	F5	251	520	CTTCTCATTTCCAATCCCTG	20	45.00%	53	IGS_23593				
J	R5	770	020	GAAATGGAAGTTTGGGCTCT	20	45.00%		130_25070				
6	F6	243	705	GACCCACATAAGAACAAACG	20	45.00%	53	100 50942 100 50000				
U	R6	947	703	CGGATATGTCCATGATTCACTA	22	40.91%	33	IGS_50842, IGS_50999				

2500 platform with a paired-end 100 bp sequencing strategy. The resulting sequences were aligned against the known reference sequence of *P. taeda* (GenBank accession no. NC\_021440.1). Subsequently, 18 single nucleotide polymorphisms (SNPs) were selected for the detection of 33 individuals in the breeding population of *P. taeda*. Primer-BLAST (https://www.ncbi.nlm.nih.gov/tools/primer-blast/) was used for the primer design (Table 1), and the PCR products were sequenced using the Sanger method.

# Phylogenetic analysis

To resolve the phylogenetic position of *P. taeda*, phylogenetic analyses were conducted using both complete cp genomes and the *ycf1* gene sequences from 36 samples. Sequence data were retrieved from NCBI (https://www.ncbi.nlm.nih.gov/), comprising 24 *Pinus* species (*P. rigida*, *P. jaliscana*, *P. oocarpa*, *P. elliottii*, *P. caribaea*, *P. taiwanensis*, *P. thunbergii*, *P. densata*, *P. wallichiana*, *P. strobiformis*, *P. bungeana*, *P. monophylla*, *P. nelsonii*, plus *P. taeda*, *P. greggii*, *P. contorta*, *P. sylvestris*, *P. tabuliformis*, *P. krempfii*, *P. strobus*, *P. massoniana*, *P. lambertiana*, *P. sibirica*, and *P. koraiensis* represented by two samples each) and the outgroup *Cedrus deodara*.

Both datasets were processed identically: multiple sequence alignment used MAFFT v7 (*Katoh, Rozewicki & Yamada, 2019*), and Maximum Likelihood trees were inferred in MEGA X (*Kumar et al., 2018*) with 1,000 bootstrap replicates.

Topological congruence between whole-genome and *ycf1*-only trees was assessed by calculating the normalized Robinson-Foulds (RF) distance using ape (v5.8.1; *Paradis & Schliep*, 2019) in R v4.5.1 (*R Core Team*, 2022).

#### **RESULTS**

## Genome comparison analysis

A total of 33,022,566 base pairs (bp) of raw reads with 38.41% GC content were generated with a paired-end 100 bp read length. The raw sequences of the *P. taeda* cp genome

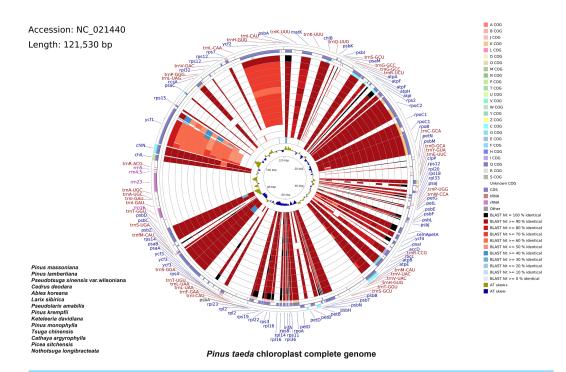


Figure 1 Comparative chloroplast genome analysis of fourteen Pinaceae species against

Pinus taeda. From the outer to the inner color ring: Pinus massoniana, Pinus lambertiana, Pseudotsuga
sinensis var. wilsoniana, Cedrus deodara, Abies koreana, Larix sibirica, Pseudolarix amabilis, Pinus krempfii,
Keteleeria davidiana, Pinus monophylla, Tsuga chinensis, Cathaya argyrophylla, Picea sitchensis, and
Nothotsuga longibracteata. Alignment method: BLASTn alignment of all genomes to Pinus taeda.
Similarity scores: black (100%), red (50–99%), blue (<50%). Four outer narrow rings: Protein-coding
gene positions (Pinus taeda chloroplast genome reference). Color code: Clusters of Orthologous Groups
(COG). Innermost ring: AT skew of P. taeda (positive [+] = A>T; negative [-] = A.

Full-size DOI: 10.7717/peerj.20092/fig-1

were deposited in GenBank with accession number PRJNA1159385, and the assembled sequences with accession number NC\_021440.1. The size of the genome was 121,530 bp, with a GC content of 38.5% (Fig. 1). The positions of all the identified genes in the cp genome and their functional categorization are presented in Fig. 1.

The cp genomes of 14 species of Pinaceae (*P. massoniana*, *P. lambertiana*, *Pseudotsuga sinensis* var. wilsoniana, Cedrus deodara, Abies koreana, Larix sibirica, Pseudolarix amabilis, *P. krempfii*, Keteleeria davidiana, *P. monophylla*, Tsuga chinensis, Cathaya argyrophylla, *Picea sitchensis*, and *Nothotsuga longibracteata*) were selected for comparison with *P. taeda* by using CCT (Fig. 1). The sequence identity between *P. taeda* and other Pinaceae representatives was analyzed, revealing that certain regions were less conserved. For instance, *ycf1* showed sequence identity lower than 70%, while *ycf2* exhibited identities below 80%. Previous studies have confirmed that *ycf1* has an important role in the evolution and classification of *Pinus* (*Daniell et al.*, 2016; *Handy et al.*, 2011; *Georgolopoulos*, *Parducci & Drouzas*, 2016).

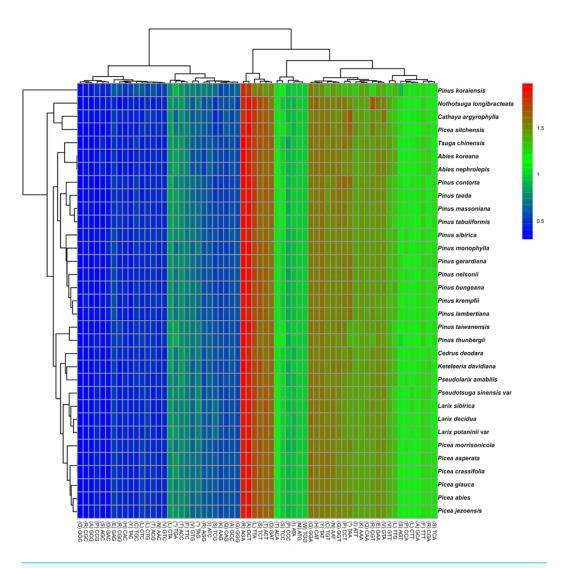


Figure 2 Relative synonymous codon usage (RSCU) heatmap of protein-coding genes across 33 species. Color scale: RSCU values from low (blue, 0.5) to high (red, 2.0); x-x-axis: Codon patterns clustered by UPGMA (*Lance & Williams*, 1967; Euclidean distance).

Full-size DOI: 10.7717/peerj.20092/fig-2

#### Codon usage

The codon usage patterns among the 71 distinct cp protein-coding genes in *P. taeda* were examined (Fig. 2). The 71 protein-coding genes consisted of 20,255 codons, with the majority ending in A or T and the remainder ending in C or G. Codons terminating in A or T exhibited significantly higher RSCU values compared to those ending in C or G, as detailed in Table 1. Statistical analysis of codon usage distributions across 33 Pinaceae species revealed infrequent use of codons ending with G or C, as shown in the heatmaps (Fig. 2). Similar patterns have been reported in other cp genomes (*Delannoy et al.*, 2011; *Raubeson et al.*, 2007; *Ying et al.*, 2016).

Among the 33 analyzed Pinaceae species, codon usage patterns were largely conserved, except for *P. taiwanensis*, *P. thunbergii*, and *P. koraiensis*. Phylogenetic clustering revealed

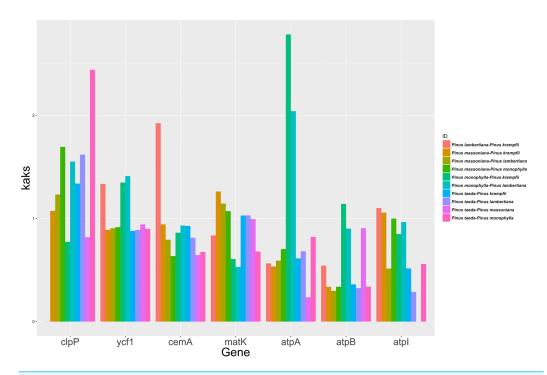


Figure 3 Ka/Ks ratios for the seven Chloroplast protein-coding genes among five *Pinus* species.

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that *P. taiwanensis* and *P. thunbergii* formed a distinct clade, divergent from other *Pinus* species. Additionally, *Picea* and *Abies* occupied phylogenetically distant positions both from *Pinus* and from each other. The clustering results were inconsistent with traditional classification results (*Gernandt et al.*, 2005).

### Ka/Ks analysis

The study analyzed the molecular evolutionary characteristics of seven chloroplast genes (*clpP*, *ycf1*, *cemA*, *matK*, *atpA*, *atpB*, and *atpI*) from five species belonging to *Pinus*. The Ka/Ks ratios of the 10 species pairs of these genes are shown in Fig. 3. Significant differences in the Ka/Ks ratios were found between the seven chloroplast genes. Ka/Ks >1 was detected in seven species pairs for *clpP* and in five species pairs for *matK*. For *ycf1* and *cemA*, Ka/Ks >1 was detected in three species pairs each, while the remaining species pairs had Ka/Ks ratios close to 1. For *atpA*, *atpB*, and *atpI*, Ka/Ks >1 was detected in only one or two species pairs, with the remaining species pairs having Ka/Ks ratios close to 1.

## SNP analysis of the chloroplast genome

Compared with the known reference sequence, 71 SNPs and four indels were identified from the 33 individuals in the breeding population of *P. taeda* (Table 2). Of these, 35 SNPs were located in the gene-coding regions of 11 genes, while the remaining SNPs were found in an intergenic region. The study found that 50 SNPs had mutation frequencies below 15%, which constituted 70.42% of the total number of SNPs. The lower mutation rate indicates that the cp genome of *P. taeda* was conservative. Notably, three SNP loci had

mutation rates between 30% and 50%, with two located in gene-coding regions (*ycf1* and *ycf2*). Specifically, *ycf1* contained 17 mutations (23.94% of the total mutation sites), and *ycf2* had three SNPs, including one with the highest mutation rate of 48%. *R poB* had three SNPs, two of which had the same mutation rate. *R poC2* had two SNPs with mutation rates at 24% and 3%. *R poC1* had two SNPs with a similar mutation rate, as did *psaA*. The intergenic regions between *matK* and *chlB*, *psbJ* and *petA*, and *rps15* and *psaC* contained 14 SNPs, eight of which had mutation rates higher than 10%.

# Individual distinction of the breeding population

A total of 18 SNPs were selected to detect 33 individuals from the breeding population of *P. taeda*. Of these, 13 SNPs were mapped to *ycf1*. Analysis revealed that 32 individuals exhibited SNPs across at 15 different loci, and 23 individuals could be distinguished based on these SNPs. Only one individual's genome matched the reference genome, as shown in Table 3. Among the 23 distinguished individuals, one additional SNP was identified, and seven SNPs were found to be the most prevalent.

Furthermore, three additional SNPs were found in 14 individuals, accounting for 60.87% of the distinguishable SNPs. A total of 13 individuals had two SNPs, which represented 39.40% of the total number of individuals (33). Compared to previous studies, the cpDNA markers had a comparatively high level of variability (*Walter & Epperson*, 2010). The chloroplast simple sequence repeats (cpSSRs) of *P. slvestris* revealed a high level of intra-populational polymorphism across the entire range (*Semerikov et al.*, 2014). Additionally, the variation among populations accounted for 99% of the total variance (*Pazouki et al.*, 2016).

Additionally, the remaining 10 individuals that could not be distinguished, each of which had only one or two SNPs, could be classified into four groups. Introducing more SNPs could potentially help distinguish these individuals. Specifically, individuals P043, P054, and P064 exhibited identical SNP profiles. Given their common origin, they may descend from the same paternal lineage. Similarly, individuals G09, G10, and 243 also had the same SNPs. G09 and G10 had the same origin, while 243 was selected from a different stand. The loblolly pine in China, which were all introduced from the southern United States, might be related to each other.

# Phylogenetic insights from cp Genomes and ycf1

Phylogenetic reconstruction using complete cp genomes resolved the 24 *Pinus* species and the outgroup *C. deodara* into two well-supported clades (Clade A and Clade B; Fig. 4A). The topology contained 33 nodes, with 26 nodes achieving maximal bootstrap support (100%), while the remaining nodes attaining values ranging from 64% to 99%. This bifurcation is consistent with the canonical classification proposed by *Gernandt et al.* (2005). Notably within Clade A, *P. taeda* and *P. rigida* converge into the same branch. *P. taeda* emerged as a focal lineage closely affiliated with *P. jaliscana*, *P. oocarpa*, *P. greggii*, *P. elliottii*, and *P. caribaea*, corroborating clade relationships reported by *Duan et al.* (2016) and *Zeb et al.* (2019); *Zeb et al.* (2022).

Analysis of *ycf1* gene sequences from the same chloroplast genomic resources recovered an overall congruent phylogenetic architecture (Fig. 4B), despite minor topological

Table 2 Mutation sites of the P. taeda chloroplast genome.

Rate

0.11

0.09

0.07

0.1

0.24

0.24

0.24

0

0

 $psbM\!\sim\!trnD\text{-}GCA$ 

 $psbM\!\sim\!trnD\text{-}GCA$ 

 $trnE\text{-}UUC{\sim}clpP$ 

 $psaJ\!\sim\!trnP\text{-}UGG$ 

 $psbJ\!\sim\!petA$ 

 $psbJ\!\sim\!petA$ 

 $psbJ\!\sim\!petA$ 

 $psbJ\!\sim\!petA$ 

 $psbJ\!\sim\!petA$ 

84756

97585

98169

100294

100387

100523

100598

100730

100732

Gene

Coordinate

Ref

SNP/ Indels

Coordinate

29448

29449

30550

34091

37238

37239

37240

37263

38094

G

T

T

T

T

Α

С

-ATCT

-GAAG

4409	A	G	0.1	$matK{\sim}chlB$	39801	A	G	0.18	petA	100910	С	-GATGGTGAA	0	ycf1
4473	С	T	0.03	$matK{\sim}chlB$	41670	С	T	0.1	psaI~accD	101076	T	G	0.01	ycf1
4609	С	T	0.04	$matK{\sim}chlB$	41678	С	G	0.06	psaI~accD	101085	T	G	0.04	ycf1
4910	G	T	0.09	$matK{\sim}chlB$	50842	С	T	0.06	$trnH$ - $GUG\sim trnT$ - $GGU$	101103	T	G	0.11	ycf1
5514	G	A	0.27	$matK{\sim}chlB$	58344	С	A	0.04	petD	101112	T	G	0.07	ycf1
5690	G	A	0.12	$matK{\sim}chlB$	64071	T	С	0.04	rpl22	101121	T	G	0.1	ycf1
9984	A	С	0.23	trnG-GCC	65304	С	A	0.15	rpl2	101130	T	G	0.06	ycf1
10349	A	С	0.06	trnG- $GCC$	70068	T	G	0.04	$trnT$ - $UGU\sim rps4$	101139	G	T	0.4	ycf1
11437	G	A	0.1	atpA	71282	A	С	0.04	trnS-GGA~ycf3	101157	T	G	0.09	ycf1
19229	G	A	0.24	rpoC2	71422	C	G	0.12	trnS-GGA~ycf3	101166	T	G	0.02	ycf1
20388	С	T	0.03	rpoC2	74477	T	G	0.23	ycf 3~psaA	102292	A	С	0.36	ycf1~rps15
22162	A	T	0.15	rpoC1	74535	A	С	0.05	ycf 3~psaA	102658	С	T	0.06	rps15~psaC
23394	T	G	0.1	rpoC1	75726	C	T	0.04	psaA	103514	С	T	0.06	rps15~psaC
24094	С	T	0.04	rpoB	76044	A	G	0.04	psaA	103620	G	T	0.18	rps15~psaC
24666	G	A	0.04	rpoB	81006	T	G	0.04	$trnfM$ - $CAU\sim psbZ$	103785	G	A	0.07	rps15~psaC
26193	С	A	0.12	rpoB	83527	G	T	0.09	$psbD\sim trnT$ - $GGU$	104282	G	A	0.1	rps15~psaC

0.05

0.08

0.05

0.26

0.35

0.02

0.13

0.02

0.02

tRNA-Thr~rrn16

ycf1

ycf1

ycf1

ycf1

ycf1

ycfl

ycfl

ycf1

G

G

G

T

G

T

G

G

SNP/ Indels

Rate

Gene

Coordinate

104944

105020

105446

109795

113045

114839

114997

117038

119153

G

T

Α

G

С

C/G

С

G

С

T

+TCTTCC

SNP/ Indels

Rate

0.2

0.13

0.28

0.22

0.09

0.04

0.48

0

0.11/0.01

 $psaC\sim ccsA$ 

 $psaC\sim ccsA$ 

 $psaC\sim ccsA$ 

ycf 2 ycf 2

ycf 2

ycf 2

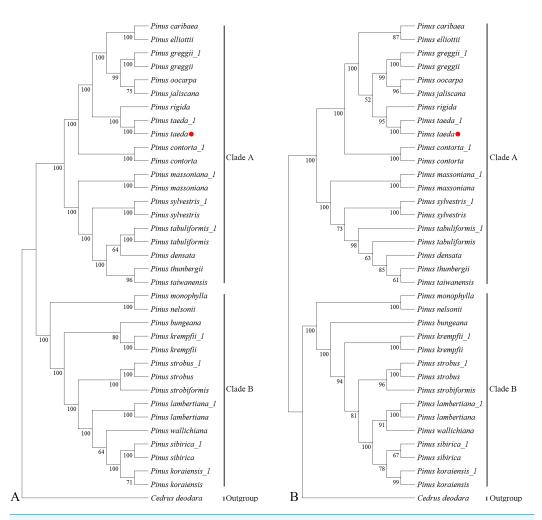
 $rpl~32{\sim}trnV{-}GAC$ 

rps 7~trnL-CAA

Gene

Table 3	Table 3 Fifteen single nucleotide polymorphisms (SNPs) for individual identification in the <i>Pinus taeda</i> breeding population.														
Gene	rpoC1	IGS	IGS	IGS	ycf1	ycf1	ycf1	ycf1	ycf1	ycf1	ycf1	ycf1	ycf1	ycf1	ycf2
Pos	23167	23593	50842	50999	98314	101076	101085	101103	101112	101121	101130	101139	101148	101157	119153
Ref	G	С	C	G	G	T	T	T	T	T	T	G	G	T	G
013	_	-	_	_	T	-	-	-	-	-	G	-	T	-	T
023	-	A	T	A	-	-	-	G	-	-	-	T	-	-	-
024	A	A	T	A	-	-	-	-	-	-	-	T	-	-	-
026	-	A	T	-	-	-	-	G	-	-	-	T	-	-	-
222aA	A	A	T	-	-	-	-	-	-	G	-	-	T	-	-
243	_	-	-	_	-	-	-	G	-	-	-	-	-	-	T
248	-	-	-	-	-	-	-	-	-	-	-	-	T	-	-
252	_	-	-	_	T	-	-	-	-	-	-	-	-	-	T
257	_	-	_	_	N	-	-	-	-	-	-	T	-	-	T
259	-	A	T	A	T	-	-	G	-	-	-	T	-	-	-
262	-	-	-	-	T	-	-	-	-	-	_	T	-	G	T
N3	-	-	-	-	-	-	-	-	-	-	-	T	-	-	-
G01	-	N	T	A	-	-	-	G	-	-	-	T	-	-	-
G10	-	-	-	-	-	-	-	G	-	-	-	-	-	-	T
G9	-	-	-	-	-	-	-	G	-	-	-	-	-	-	T
P012	-	-	-	-	T	G	G	G	G	-	G	-	-	-	T
P034	-	-	-	-	-	-	-	-	-	G	-	T	-	-	-
P043	_	-	-	-	-	-	-	-	-	-	-	-	-	G	T
P052	A	A	Т	A	-	-	-	-	-	G	-	T	-	-	-
P054	-	-	-	-	-	-	-	-	-	-	-	-	-	G	T
P063	-	_	-	-	-	-	-	G	-	-	G	-	_	_	_
P064	_	-	-	-	-	-	-	-	-	-	-	_	-	G	T
Q16	-	_	-	_	-	-	-	-	-	G	-	-	-	-	Т
Q26	_	-	-	-	_	-	-	-	-	-	G	_	-	-	Т
Q6	-	_	_	-	T	-	_	-	_	-	-	_	-	-	Т
S11	A	A	T	A	-	-	-	G	-	G	_	T	-	-	-
S3	_	-	_	_	_	-	_	_	_	-	G	_	-	-	-
S6	-	-	-	-	-	-	-	-	-	-	N	-	-	-	-
W05	-	-	-	-	T	-	-	-	-	-	-	T	T	-	T
W11	-	-	-	-	-	-	-	-	-	-	-	T	-	-	-
W13	-	_	-	-	-	-	-	-	-	-	-	-	-	-	-
W16	A	A	T	A	-	-	-	-	-	-	-	-	-	-	_
W28	-	-	-	-	-	-	-	-	-	-	-	-	-	_	T

variations. The resultant tree likewise recovered Clades A and B across 33 nodes, with 16 nodes exhibiting 100% bootstrap support and the remaining nodes having bootstrap values ranging from 52% to 99%. While *P. jaliscana*, *P. oocarpa*, and *P. greggii* retained their close clustering with *P. taeda* within Clade A, *P. elliottii* and *P. caribaea* failed to group with this lineage. Instead, they occupied distinct positions with bootstrap supports of 92% at their respective nodes.



**Figure 4 Phylogenetic trees of Pinaceae from 36 plastomes and** *ycf1***.** Phylogenetic relationships inferred by maximum likelihood (ML) from 36 complete chloroplast genome sequences representing 25 species of Pinaceae. (A) Whole-chloroplast genome phylogeny. (B) *ycf1* gene phylogeny. Major clades are indicated: Clade A, subgenus *Pinus* (two-needle pines); Clade B, subgenus *Strobus* (single-needle pines). The red dot represents the position of *P. taeda* (NC\_021440.1).

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Quantitative assessment of topological concordance revealed minimal divergence between the whole-plastome and *ycf1*-based phylogenies, evidenced by a normalized RF distance of 0.15. This strong congruence highlights the efficacy of *ycf1* as a single genomic marker for resolving genus-level evolutionary relationships in *Pinus*.

## **DISCUSSION**

The complete cp sequence of *P. taeda*, a dominant non-native plantation species in China, has been assembled, annotated, and analyzed. This species is a valuable resource for investigating the intra- and interspecific evolutionary history of plants (*Birky Jr*, 1978; *Birky Jr*, 2001; *Chase et al.*, 1993; *McCauley*, 1995; *Newton et al.*, 1999; *Provan, Powell &* 

*Hollingsworth*, 2001; *Petit et al.*, 2003). The cp genome consisted of 121,530 bp and lacked inverted repeats, which are commonly found in most angiosperms.

Comparative genomic analysis between *P. taeda* and 14 species of Pinaceae showed that certain regions, such as *ycf1*, were less conserved. This finding was confirmed in a previous study (*Jiang et al.*, 2017; *Li et al.*, 2020; *Chen et al.*, 2020; *Zhang et al.*, 2023). The highest sequence diversity was identified in two regions of *ycf1*. These regions can be targeted in all pine subsections using three primer combinations (*Parks, Liston & Cronn*, 2011; *Handy et al.*, 2011). Recently, *ycf1* has been identified as an essential component of the protein translocon at the chloroplast's inner envelope membrane (*Kikuchi et al.*, 2013). Seven variable sites and nine haplotypes were found in an 840 bp fragment of the DNA-coding region of *ycf1*. These variable sites and haplotypes have been evaluated in the *Pinus* subsection *Australes* and have unquestionable value for studying evolution in the group (*Ortiz-Martínez & Gernandt*, 2016). The present study corroborates that *ycf1*, with 17 mutations identified, plays a significant role in individual distinction in *P. taeda*.

It is probable that the chloroplast genes of *P. taeda* exhibit large differences in the natural selection pressure during the evolutionary process. This is in comparison to the chloroplast genes of other species belonging to the same genus (*Kober & Pogson, 2013*; *Nasrullah et al., 2015*; *Kwon et al., 2016*). However, most species of the same genus have similar codon biases and are basically clustered together. These findings suggest that the codon usage bias of chloroplast genes is closely related to the genetic similarity among species, reflects the evolutionary relationships between species, and could provide additional insights when used in conjunction with phylogenetic analysis to investigate the evolutionary relationships and molecular evolutionary mechanisms of species.

The current study surveyed the evolutionary characteristics of seven chloroplast genes across five species of Pinus. The findings indicated that for clpP and matK, most species pairs exhibited Ka/Ks ratios >1. In contrast, ycf1 and cemA showed ratios approximately half of that value, while the remaining species pairs for these genes had ratios near 1. For atpA, atpB, and atpI, Ka/Ks >1 was observed in only one or two species pairs, while the ratios for the majority of pairs were well below 1. These results suggest that clpP and matK experienced predominantly positive selection, ycf1 and cemA experienced either positive or neutral selection, and atpA, atpB, and atpI were under purifying selection during Pinus evolution (Yang, 1998; Zhang, 2005). This variation may be attributed to the encoded proteins playing different roles in growth and metabolism processes (*Hahn*, 2005; *Ransay*, Rieseberg & Ritland, 2009; Cork & Purugganan, 2004). cpSNP markers helped distinguish individuals among populations of loblolly pine. Distinguishing individuals is crucial for identifying the male parent of open- or mixed-pollination offspring. Given the paternal inheritance of cpDNA in *Pinus*, the male parent can be identified by using the molecular marker developed from SNPs in the cp genome. Fifteen SNPs successfully distinguished 23 individuals within the P. taeda breeding population. The inability to distinguish the remaining individuals suggests they may be closely related. P. taeda was introduced to China nearly 120 years ago, and the individuals in the breeding population of the current study were selected from plantations or had just been introduced from the southern United States. The genetic relationships among individuals in the P. taeda breeding population

were previously undefined. The study of the SNPs in the cp genome may provide a new method for paternal identification in the breeding program of *P. taeda*.

Chloroplast genomes are valuable tools for plant phylogenetic analysis, with important insights often coming from the examination of protein-coding genes (*Eckert & Hall*, 2006; *Gernandt, Liston & Piñero*, 2003; *Moore et al.*, 2010; *Parks, Cronn & Liston*, 2012; *Zhu et al.*, 2016a; *Zhu et al.*, 2016b). These earlier studies provided the foundation for the current investigation into the loblolly pine chloroplast genome. This study performed a phylogenetic analysis using the maximum likelihood (ML) method. The dataset comprised 36 chloroplast genomes and the *ycf1* gene from 24 *Pinus* species, with *C. deodara* as the outgroup. Phylogenetic trees based on both the complete chloroplast genomes and the single *ycf1* gene consistently resolved the 25 species (which were represented by 36 samples) into two major clades: the subgenus *Strobus* (single needle section, clade B) and subgenus *Pinus* (double-needle section, clade A). Particularly significant is the placement of *P. taeda* within Clade A, where it forms a sister relationship with *P. rigida* while maintaining close affinity to *P. jaliscana*, *P. oocarpa*, and *P. greggii*. This configuration aligns with and extends earlier findings by *Zeb et al.* (2019); *Zeb et al.* (2022), resolving previous ambiguities regarding loblolly pine's phylogenetic position.

Notably, analysis of the *ycf1* gene region alone recovered a highly congruent phylogenetic architecture (Fig. 4B), despite minor topological variations affecting *P. elliottii*, *P. caribaea* and *C. deodara* placements. The high degree of concordance between whole-plastome and single-gene topologies-quantified by a normalized RF distance of 0.15-validates *ycf1* as a phylogenetically informative marker for genus-level reconstructions in conifers. This finding substantiates earlier suggestions (*Zhu et al.*, 2016a; *Zhu et al.*, 2016b) regarding the utility of protein-coding genes, while demonstrating that targeted sequencing of *ycf1* has comparable resolution to resource-intensive complete plastome analyses in resolving major clades (*e.g.*, subgenera) in *Pinus*.

#### CONCLUSIONS

We sequenced and deposited the 121,530-bp chloroplast genome of *Pinus taeda* (GenBank: NC\_021440.1). Regions such as *ycf1* and *ycf2* showed markedly lower identity (<70% and <80%, respectively) than 14 other *Pinus* species. Phylogenomic analyses placed *P. taeda* within the diploxylon clade, sister to *P. rigida*. Comparative codon-usage patterns across 33 Pinaceae species revealed signatures of natural selection. In the breeding population, 71 cpSNPs distinguished 72% of individuals; the single-copy gene *ycf1* recapitulated whole-plastome relationships (normalized RF = 0.15), validating it as an efficient barcode. These cpSNPs and the mutation map provide readily applicable markers for paternal identification in *P. taeda* breeding programs, while the hypervariable *ycf1* gene serves as a highly informative marker for future genus-wide phylogenetic and germplasm studies.

# **ADDITIONAL INFORMATION AND DECLARATIONS**

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# **Competing Interests**

The authors declare there are no competing interests.

#### **Author Contributions**

- Ling Wang conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.
- Jipeng Mao analyzed the data, prepared figures and/or tables, and approved the final draft.
- Kaibin Jiang analyzed the data, prepared figures and/or tables, and approved the final draft.
- Zhengyu Wu analyzed the data, prepared figures and/or tables, and approved the final draft.
- Chunxin Liu analyzed the data, prepared figures and/or tables, and approved the final draft.
- Huagong Ning performed the experiments, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.
- Haibiao Zhou analyzed the data, authored or reviewed drafts of the article, and approved the final draft.
- Jiehu Chen analyzed the data, authored or reviewed drafts of the article, and approved the final draft.
- Shaowei Huang analyzed the data, prepared figures and/or tables, and approved the final draft
- Tianyi Liu conceived and designed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.

### **Data Availability**

The following information was supplied regarding data availability:

The raw data is available at CNGBdb: CNP0006272.

The raw data is available at GenBank: PRJNA1159385 and NC 021440.1.

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