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In situ study of environmental factors (temperature and salinity) affecting cohort patterns and growth rates in *Ciona robusta*

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Assessing and understanding the ecological impacts of marine invasive species is important for managing marine ecosystems, especially since their rapid growth, short reproductive cycles, and wide range of water temperature adaptability pose major challenges. In this study conducted in Mokpo, South Korea, we explored the cohort dynamics and environmental dependence of ascidian, *Ciona robusta*, which is considered a widespread invasive species. Through comprehensive observations conducted from June to October 2022, we identified five distinct cohorts, challenging existing assumptions about lifespan and cohort patterns. All separation index values exceeded 2, indicating clear separation of cohorts during the study period. The cohorts had a lifespan of between 4 and 10 weeks, much shorter than the previously reported maximum of 2 years. These differences suggest that local climatic conditions can have a significant impact on lifespan parameters. In addition, growth rates were significantly positively correlated with environmental conditions, particularly temperature. This highlights that *C. robusta* growth rates are particularly sensitive to temperature changes despite their salinity tolerance. Therefore, this study contributes significantly to our understanding of invasive species dynamics in marine ecosystems. In particular, it provides valuable insights for developing management strategies to mitigate the impacts of *C. robusta* due to climate change.

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Abstract

Assessing and understanding the ecological impacts of marine invasive species is important for managing marine ecosystems, especially since their rapid growth, short reproductive cycles, and wide range of water temperature adaptability pose major challenges. In this study conducted in Mokpo, South Korea, we explored the cohort dynamics and environmental dependence of ascidian, *Ciona robusta*, which is considered a widespread invasive species. Through comprehensive observations conducted from June to October 2022, we identified five distinct cohorts, challenging existing assumptions about lifespan and cohort patterns. All separation index values exceeded 2, indicating clear separation of cohorts during the study period. The cohorts had a lifespan of between 4 and 10 weeks, much shorter than the previously reported maximum of 2 years. These differences suggest that local climatic conditions can have a significant impact on lifespan parameters. In addition, growth rates were significantly positively correlated with environmental conditions, particularly temperature. This highlights that *C. robusta* growth rates are particularly sensitive to temperature changes despite their salinity tolerance. Therefore, this study contributes significantly to our understanding of invasive species dynamics in marine ecosystems. In particular, it provides valuable insights for developing management strategies to mitigate the impacts of *C. robusta* due to climate change.

Introduction

To preserve marine ecosystems, understanding and evaluating the impacts of disturbance and pollution caused by marine invasive species is crucial (Whitlatch and Bullard 2007; Locke 2009; Kanamori et al. 2017). In addition, certain invasive species have negative impacts on human economic activities (Schultz et al. 2011; Park et al. 2018). Examples of fouling organisms, particularly those that live in a sessile life, include ascidians, bryozoans, hydrozoans, barnacles, sponges, and mussels (Bosch-Belmar et al. 2019; Shevalkar et al. 2020; Lins and Rocha 2022). Ascidians are a taxonomic group containing many diverse species widely distributed worldwide and are dominant in fouling communities (Miller 1971; Young 1985; Sahade et al. 2004; Shenkar and Swalla 2011). They are one of the major causes of marine invasive species problems due to their rapid growth rate, short reproductive cycle, and lack of crucial predators (Shenkar and Loya 2008; Lynch et al. 2016; Kanamori et al. 2017).

A primary life history characteristic of ascidians is that they are sessile as adults but, in contrast, are free-swimming as larvae (Millar 1971). Larvae typically settle in proximity to their parents (Zhan et al. 2015), though they may also disperse widely, contingent on local hydrological conditions (Fletcher et al. 2013). After development, larvae hatch from the eggs and disperse in search of suitable substrates for metamorphosis. Following larval metamorphosis, they develop into adults through settlement. The ascidian life cycle is influenced by substrate materials, light conditions, pH, temperature, and salinity, which play a crucial role in the survival and growth of marine organisms (Anderson and Underwood 1994; Nandakumar 1995; McKinney and McKinney 2002; Chase et al. 2016; Granot et al. 2017; Jones et al. 2022).

Ciona intestinalis was reclassified into two distinct species based on molecular and morphological studies: *Ciona robusta* (formerly *C. intestinalis* type-A) and C. intestinalis type-B (Brunetti et al. 2015; Gissi et al. 2017). While *C. robusta* corresponds to the previously designated type-A, type-B remains classified under *C. intestinalis*, allowing for clear differentiation between the two types in various geographic regions. *Ciona robusta* is now recognized worldwide as a significant marine invasive species, prompting extensive research on its distribution and spread in various countries (Bouchemousse et al. 2017; Shenkar et al. 2018; Park et al. 2018; Kim et al. 2019a; Bae et al. 2023a, 2023b). In South Korea, *C. robusta* is legally recognized and managed as a "Marine ecosystem disturbing species" (a legal term for marine harmful organisms; Kim et al. 2019b).

The life cycle of *C. robusta* is of significant consequence with respect to its ecosystem function. It exhibits a dichotomized life history with sessile adults and planktonic larvae (Wilson et al. 2022). This unique life cycle affects ecosystem function in two main ways: (1) The planktonic larvae serve as a major food source for various invertebrates (Bingham and Walters 1989; Boltovskoy and Correa 2015; Rivera-Figueroa et al. 2021). (2) The widespread settlement and growth of adults lead to competition for space with other fouling organisms (Grosberg 1981; Bullard et al. 2004). Moreover, the reproductive and growth patterns of *C. robusta* can have a detrimental impact on local ecosystems through competition with native species. This highlights the necessity of comprehending its reproductive and growth dynamics for the implementation of effective management and conservation strategies (Robinson et al. 2017; Part et al. 2018).

There are at least two generations of *Ciona robusta* and *Ciona intestinalis* (type-B) per year, regardless of geographic region (Caputi et al. 2015; Bouchemousse et al. 2017). However, in warmer climates, reproduction is more intense, resulting in shorter generation periods for populations. For example, in tropical and subtropical regions, more spawning occurs in a year than that in cooler climates (Wilson et al. 2022), and overall growth is relatively faster (Malfant et al. 2017). On the other hand, a study observing oocyte size and gonad development in the cooler climate of Puerto Madryn port, Argentina (from 9 °C in winter to 19 °C in summer), found that reproduction occurs throughout the year, but abundance peaks only once in the fall (Giachetti et al. 2022a; 2022b). The maximum growth length in temperate Kyoto, Japan, has been recorded to be up to 130 mm (Tarallo et al. 2016). Growth also varies not only with temperature but also with feeding. In a laboratory study at a temperature of 15 °C, growth rates varied depending on the type of feed, with a maximum average growth of 11.59 mm in 32 days when fed appropriately (Zupo et al. 2020).

In addition, information on the *in situ* lifespan of *C. robusta* and the number of cohorts present at any one time is scarce. Nevertheless, the lifespan is known to be 2 months to 2 years (Millar 1952; Dybern 1965), and multiple cohorts are reportedly likely present at any one time, depending on temperature (Yamaguchi 1970; 1975). Understanding the cohort and growth patterns of this invasive species can help predict ecosystem changes and develop responses accordingly (Svane 1983; Wong et al. 2011; Lynch et al. 2016). Therefore, the first aim of this study was to closely examine the cohort patterns and growth rates of *C. robusta*, a representative marine invasive species. By correlating these patterns with environmental variables, notably water temperature and salinity, this research aims to unveil the intricate relationships between the lifespan of *C. robusta* and its surrounding ecosystem. Water temperature and salinity have been studied relatively more than other environmental factors (e.g., substrate materials, light conditions and pH; Li et al. 2019; Olivo et al. 2021; Jones et al. 2022) for *C. robusta* and are known to have a decisive effect on *C. robusta* growth (Wilson et al. 2022). Thus, the second aim of this study was to determine if temperature and salinity were the main factors affecting growth rates, even *in situ* environment with multiple variables. The findings of this investigation promise to not only increase the ecological understanding of *C. robusta* but also inform and refine strategies for their management and control. Through a detailed analysis of the cohort dynamics of *C. robusta* under different environmental conditions (especially water temperature), this study seeks to provide valuable insights into the broader challenge of conserving marine biodiversity in situations such as climate change, where the threat of invasive ascidians is increasing.

Materials & Methods

In this study, surveys were conducted at Mokpo Yacht Marina (34°47′2.70″ N, 126°23′21.05″ E) at 2-week intervals from June 2022 to October 2022. The study site, Mokpo Yacht Marina, is located in Mokpo City, Jeollanam-do, South Korea (hereafter referred to as Mokpo; Figure 1). The selected study site was where yachts and ships moored. *C. robusta* were collected from the marina at depths of about 1 m. Sampling continued until October 29, 2022. *C. robusta* was absent in the ninth and tenth surveys, which were conducted on October 15 and October 29, 2022. The objective was to sample at least 50 individuals on each survey. However, due to a lack of live individuals, only 30 were sampled on the last survey (October 1, 2022) where *C. robusta* was last observed. Only undamaged individuals were used to measure body length during the sampling process. We also anesthetized the relaxed body by placing it in a plastic tray (300 × 240 × 70 mm) filled with local seawater and menthol crystals to prevent contraction. The total body length was measured using digital-type Vernier calipers (Digimatic Caliper; Mitutoyo Corporation, Kanagawa, Japan) and imaged using a camera (Tough TG-5; Olympus Corporation, Tokyo, Japan). The measured individuals were placed in 50-mL conical tubes and stored in a portable freezer (–20 °C) for transportation to the laboratory. In the laboratory, each specimen was weighed wet and then vacuum freeze-dried (FDT-8650, Operon, South Korea) for 72 h to measure its dry weight. Wet weight and dry weight were measured (accurate to 0.01 g) using an electric weighing scale (ML4002/01, Mettler Toledo, Switzerland). Water temperature and salinity data were provided by the Korea Hydrographic and Oceanographic Agency of Ocean Data in the Grid Service (https://www.khoa.go.kr, Figure S1). To reduce the error in the length data measured in the field, the length data were calibrated by remeasuring the length in the image using ImageJ (Schneider et al. 2012; National Institutes of Health, Bethesda, MD, USA). In this study, length data served as the primary metric for analysis. To validate the accuracy of length data, dry and wet weights were also measured and subjected to correlation analysis (Figure S2).

This step ensured that all analyses were grounded on precise and reliable length measurements, forming the basis of the study's findings.
The body length of the shortest individual collected was 8 mm, and therefore the body length frequency distribution was consisted with 7 mm intervals. Each cohort was determined using the Bhattacharya method (Bhattacharya 1967). FiSAT II software v 1.2.2 (Gayanilo et al. 2005; FAO, Rome, Italy) was used to separate the components of the normal distribution for each survey. The normal distribution was determined iteratively until it could no longer be distinguished using the separation index [(SI) = ΔLk/Δδk, where ΔLk is the difference between the two successive means of the component curves and Δδk is the difference between their estimated standard deviations]. For SI values above 2, cohort separation was found to be statistically reliable. The growth rate of each cohort was calculated using the formula r = (ln Nti+1 - ln Nti)/(ti+1 - ti) (Odum 1971), where 'r' represents the growth rate of the population; 'N' represents the average length of the cohort—Choe and Deibel (2011) used abundance but we used length—'t' represents time, which in this study refers to the order of the survey; 'i' represents a specific point in time, which in this study refers to the number of surveys. We performed t-tests to compare mean lengths between cohorts and multiple linear regression to determine whether the growth rate of cohorts was affected by environmental factors (water temperature and salinity). Growth rate was used as the dependent variable and water temperature and salinity as the independent variables, and the independent variables were tested for normality and equality of variance using the Shapiro-Wilk and Levene test, respectively. The t-tests and multiple linear regression were performed in R (R core team, 2013; R Foundation for Statistical Computing, Vienna, Austria). The t-test was performed by generating normally distributed data using the 'rnorm' function. We also used the 'lm' and 'summary' functions to perform multiple linear regression and model summary statistics, respectively.

Results

A total number of 469 *C. robusta* individuals were collected in this study, with a mean of 58.62 ± 13.77 (mean ± SD) individuals collected in each field survey (Table 1). The shortest and longest individuals (collected on July 9 and August 5) were 8 and 150 mm, respectively. The *C. robusta* body length values measured for cohort analysis and growth rate calculations showed a significant positive correlation with wet ($r^2 = 0.81$) and dry weight ($r^2 = 0.78$). Wet and dry weights were also significantly positively correlated ($r^2 = 0.91$; Figure S1). The maximum observed water temperature during the study period was 26.69 °C (August 26), and the minimum was 22.01 °C (June 27). Salinity was highest at 30.63 PSU (July 13) and lowest at 19.70 PSU (September 9). The ranges for salinity and temperature were 10.92 PSU and 4.68 °C, respectively, indicating a larger range for salinity than for temperature (Figure S2).
Five cohorts were identified over the entire study period (C1–C5). All cohorts had an SI > 2 at each survey time and were significantly separated ($p < 0.05$; Table 1 and S1). C3 was the most observed cohort, with five surveys (June 26 to August 20), and C1 was the least observed cohort (June 26 to July 9), with two surveys. The most cohorts were observed on July 9 (C1–C4), with

four, and the fewest were observed on September 17 and October 1, with one cohort each (C5). Three or more cohorts were consistently observed from the first (June 26) to the fifth survey (August 20), with the last cohort (C5) being the first observed in the fifth survey (Figure 2). The longest cohort was C2 (measured on August 5), with a mean body length of 135.43 ± 15.00 mm, and the shortest cohort was C4 (measured on July 9), with a mean body length of 16.50 ± 9.53 mm. The cohort with the largest range between the maximum and minimum mean body length was C3, which grew from 44.25 ± 12.21 mm (June 26) to 123.50 ± 8.41 mm (August 20), a growth of approximately 79.25 mm. Excluding C1, which had a decrease in mean body length, the cohort with the smallest difference in mean body length was C5, which grew from 28.35 ± 6.09 mm (August 20) to 51.03 ± 6.86 mm (October 1st), a growth of approximately 22.68 mm (Figure 3 and Table 1).

The highest growth rate in this study was 0.6241 for C4, which grew 14.3 mm in mean body length from July 9 (16.50 mm) to July 23 (30.80 mm). In contrast, C1, which lost 3.44 mm in mean body length from June 26 (99.06 mm) to July 9 (95.62 mm), had the lowest growth rate of -0.0353 (Table 1 and S2). To confirm which environmental variables (water temperature and salinity) had a significant effect on growth rate, we first checked the assumptions for multiple linear regression. Normality and homogeneity of variances were verified for the residuals of a preliminary model ($p > 0.05$ for both tests; Table S4). After confirming these assumptions were met, we proceeded with the multiple linear regression analysis. The analysis determined that temperature had a significant effect ($p = 0.000$). The intercept was also significant ($p = 0.024$), indicating a baseline growth rate when other variables are at zero. However, salinity ($p = 0.114$) did not significantly affect growth rate (Table S3). The model summary statistics for the multiple linear regression had an r^2 value of 0.438, an F-value (2, 17) of 8.413, and a p-value of 0.002. Temperature and growth rate were positively correlated, with higher temperatures leading to higher growth rates, and the coefficient of determination was relatively high (r2 = 0.51). On the other hand, salinity and growth rate were negatively correlated, and the coefficient of determination was noticeably lower ($r^2 = 0.18$; Figure 4).

Discussion

Mokpo, South Korea, is a region where *C. robusta* has been present continuously from spring to fall (Park et al. 2018; Bae et al. 2022; Lee et al. 2022). Because of the temperate climate in the study area, several populations hatch at similar times within a year. Therefore, we conducted this study to distinguish these cohorts and identify their correlations with environmental factors. A total of five cohorts were observed during the study period, and the SI values of all cohorts were > 2, indicating that the cohorts were well distinguished (Hasselblad 1966). At Mokpo, the number of cohorts separated within approximately the same duration of study periods was higher than in other areas. Many studies have documented the seasons when ascidians of the genus *Ciona*, such as *Ciona robusta* (*C. intestinalis* type-A, Bouchemousse et al. 2017) and *Ciona intestinalis* (Mastrototaro et al. 2008; Astudillo et al. 2016; Bouchemousse et al. 2017), dominate the community, but few have distinguished between each cohort. In Nova Scotia (Canada), up to two major recruitment events occur between June and August (Carver et al. 2003). In Naples, Italy, three genetically distinct clusters (cohort) in the population were observed following monthly sampling for 13 months (Caputi et al. 2019). Naples and Nova Scotia are regions with Mediterranean and continental climates, respectively, that are similar to that of Mokpo, although fewer cohorts were recognized than those in Mokpo.

The cohort lifespan of genus *Ciona*, such as *Ciona robusta* (Nakazawa et al., 2019; Beyer et al., 2023) and *Ciona intestinalis* (Millar 1952; Dybern 1965; Beyer et al. 2023), varies from 2 months to 2 years, which contrasts with the results of the present study (approximately 4–10 weeks). The growth, reproduction, and mortality of *C. robusta* are affected by various environmental factors, including natural (temperature, salinity, and microalgae) and anthropogenic (bisphenol A, mercury, copper, and cadmium) factors (Bellas et al. 2001; 2004; Mansueto et al. 2011; Wilson et al. 2022). The yacht mooring where *C. robusta* was sampled has a high diversity of fouling benthic invertebrates and has been the site of a number of related studies. In the Lee et al. (2022) study utilizing artificial substrates at Mokpo, biofouling species such as barnacle (Amphibalanus improvises, etc.), bryozoan (Bugula neritina, etc.), ascidian (*Didemnum vexillum*, etc.), and bivalves (Mytilus galloprovincialis, etc.) were 28 out of 32 total species, and these taxa are the target species of biofouling studies (Yu et al. 2007; Fino et al. 2014; Forrest and Atalah 2017; Bae et al. 2023c). The results of Bae et al. (2022) also showed that of the 23 species of sessile benthic invertebrate observed in Mokpo, 10 were ascidian and 5 were bryozoan, including species such as *Didemnum vexillum*, *Ascidiella aspersa*, *Bugula neritina*, and *Watersipora subtorquata*, which have been the subject of various biofouling studies, including *C. robusta* (Yu et al. 2007; Lynch et al. 2016; Marasinghe et al. 2018; Bae et al. 2023a). Therefore, Mokpo is a site where a variety of biofouling species have been observed, suggesting fewer anthropogenic environmental factors contributing to *C. robusta* mortality. Therefore, the lifespan of *C. robusta* cohorts in temperate climates is presumed to be shorter than the known maximum of 2 years (Millar 1952; Dybern 1965). On the other hand, the results of the present study, where cohort lifespan ranged from 4 to 10 weeks, are more in line with studies that suggest a lifespan of approximately 3 months (Nakazawa et al. 2019; Beyer et al. 2023). For body length, the maximum body length of the *C. robusta* we sampled was 150 mm, with C2 having the longest mean body length of the cohort at 135.43 ± 15.00 mm. These results are similar to those of a study that sampled *C. robusta* in Kyoto, Japan, and found a maximum body length of 130 mm (Tarallo et al. 2016). Therefore, a clear understanding of the lifespan and body length of *C. robusta* in relation to its habitat will help to evaluate effective management and control.

During the study period, five cohorts were observed, suggesting that two or more cohorts existed at different points in time simultaneously. Giachetti et al. (2022b) reported that *C. robusta* have consistently mature gonads throughout the year, supporting our findings. Other reasons for these differences in the number of cohorts present at a point in time and the lifespan of the cohorts are considered to be the influence of environmental factors other than temperature and salinity or methodological differences in analysis. Specifically, live algae and non-live particles ensure the survival of *C. robusta* (Zupo et al. 2020); therefore, these environmental factors may have played a role or previous monitoring studies may not have used a normal distribution to separate cohorts. A wide range of information is available on post-settlement growth rates of juveniles, but linking the information to understand the factors that influence them remains a challenge (Wilson et al. 2022). However, in this study, the growth rates of the separate cohorts allowed us to determine with relative clarity how temperature and salinity affect *C. robusta* growth. Therefore, we analyzed the correlation of growth rates with temperature and salinity, the most representative and influential factors. The multiple linear regression results showed that the growth rate of each *C. robusta* cohort was more relatively correlated with temperature than with salinity, consistent with our findings that *C. robusta* development is most closely associated with increases in temperature (Yamaguchi 1970; 1975).

C. robusta is relatively intolerant of low salinity conditions and has a developmental salinity limit of 26 ‰ (Madariaga et al. 2014; Kim et al. 2019b). Outside of these low salinity conditions, *C. robusta* will grow rapidly up to (and beyond) 24.7 °C in the laboratory (Kim et al. 2019b). Notwithstanding, the optimal temperature for development *in situ* was determined to be 14.7–23.7 °C (Caputi et al. 2019). Therefore, the temperature (24.88 °C) and salinity (29.05 PSU) conditions for C4 on July 23, when the growth rate was highest (0.6241), were optimal for *C. robusta* to achieve rapid growth. Correlations with growth rate performed using the Scheirer–Ray–Hare test on *C. robusta* juveniles in the laboratory under four conditions, two each of temperature (12 and 17 °C) and salinity (25 and 35 PSU) in combination, demonstrated statistical significance for all sources ($p < 0.05$; Malfant et al. 2017). However, in the present study, only temperature significantly positively correlated with growth rate. These results are attributed to the salinity measured during the study not persisting below the low salinity limit of 26 PSU. Although conditions below 26 PSU existed in the field, they were relatively short-term, lasting approximately 2 or 7–12 days compared with the 28 days or more in a laboratory study (Malfant et al. 2017). Therefore, the impact of low salinity may have been minimal.

Overall, the current study identified five *C. robusta* cohorts, each with a significant SI, indicating clear distinctions. Contrasting the number of cohorts and growth conditions in Mokpo with those in areas with similar climates but fewer cohorts highlights the importance of environmental factors and methodological approaches in understanding *C. robusta* population dynamics. The lifespan of the *C. robusta* cohort in Mokpo is shorter than previously recorded (Up to two years; Millar 1952; Dybern 1965), estimated to be between 4 and 10 weeks, contrasting with previous studies showing longer lifespans. In addition, growth rates were more strongly correlated with temperature than salinity, highlighting the important role of temperature. Particularly encouraging is the fact that the correlation between cohort growth rate and temperature was demonstrated *in situ* rather than in laboratory settings. Salinity had a less significant correlation with growth rate compared with that of temperature, but this is likely due to the fact that low salinity did not persist long enough at the study site to significantly affect growth rate. However, because this study was limited in geographic and temporal scope and focused primarily on temperature and salinity, it may have overlooked other environmental factors that could affect the growth and spread of *C. robusta*, such as ocean acidification and nutrient levels. Further research should include long-term observations in more diverse geographic locations, incorporating a wider range of environmental variables, and exploring correlations.

Conclusions

Through an *in situ* study, we observed multiple cohorts thriving simultaneously in a temperate climate (Mokpo, South Korea) and concluded that temperature strongly influences the growth rate of *C. robusta*. By understanding the cohort patterns and temperature-dependent growth rates of global invasive species such as *C. robusta*, this research provides foundational knowledge that is instrumental for developing predictive models and management strategies aimed at controlling the spread of invasive species. This understanding is particularly valuable in the context of climate change, as shifts in temperature could alter the invasion dynamics and ecological impacts of such species. The insights gained from this study are poised to significantly benefit fields such as invasive species management, marine conservation, and climate change research by offering a more nuanced approach to predicting and mitigating the effects of invasive species under changing environmental conditions. In future research, a broader understanding of the *C. robusta* cohort would be gained if long-term surveys were conducted in multiple geographic locations (varying climates), including factors such as ocean acidification and nutrient levels that were not addressed in this study.

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Figure 1

Map and photos of the survey site

Map showing the location of the survey site (blue dot) in South Korea (a). Zoomed-in study site (b). Satellite imagery indicating points (red dot) where artificial substrates were installed (c) and foreground photo (b).

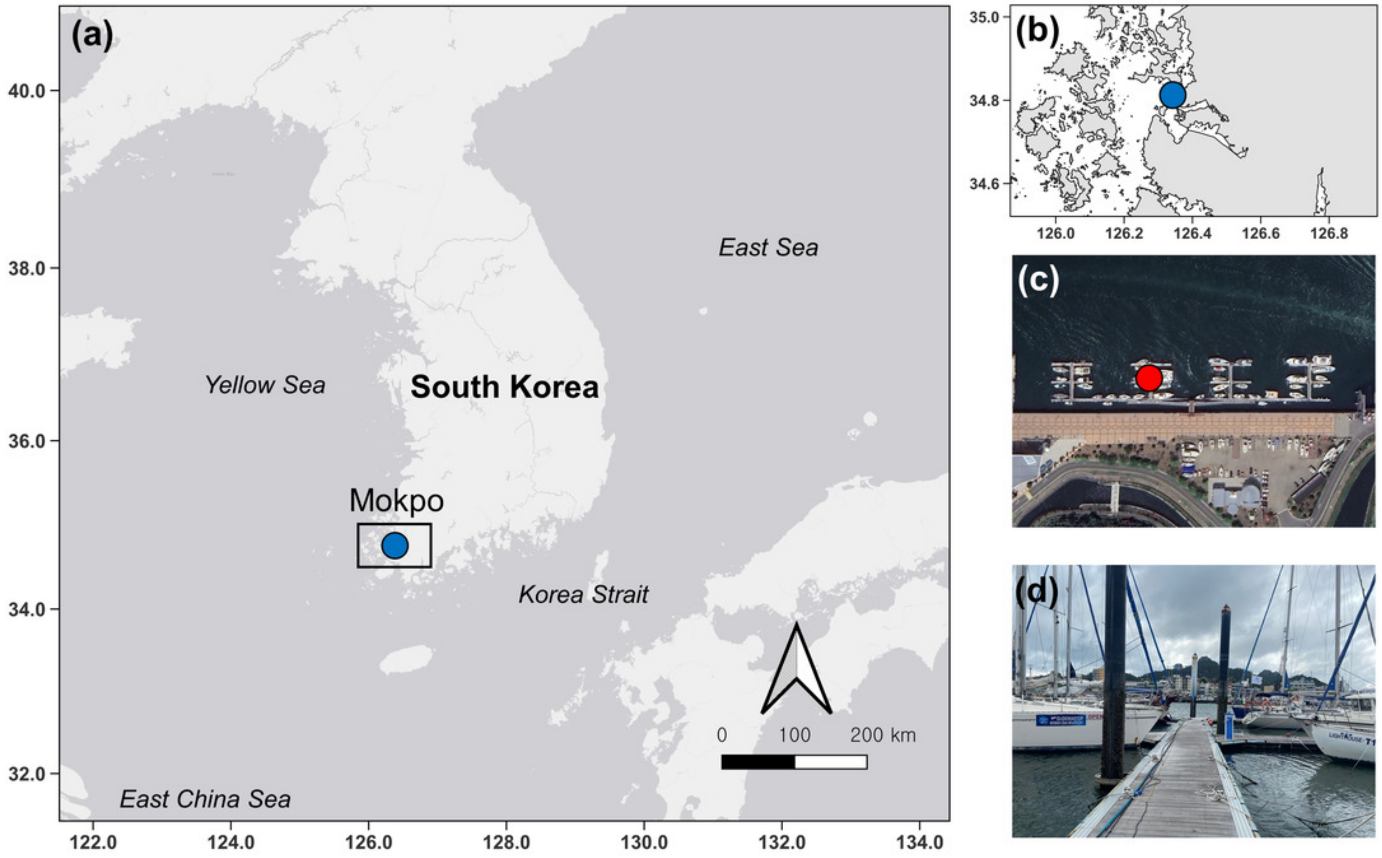

Figure 2

Frequency distribution of body length in *Ciona robusta*

Frequency distribution of body length in *Ciona robusta* samples collected between June 26, 2022, and October 1, 2022, numbered by 2-week interval (1–8). Individual cohorts were defined as the normally distributed components of the sample distribution.

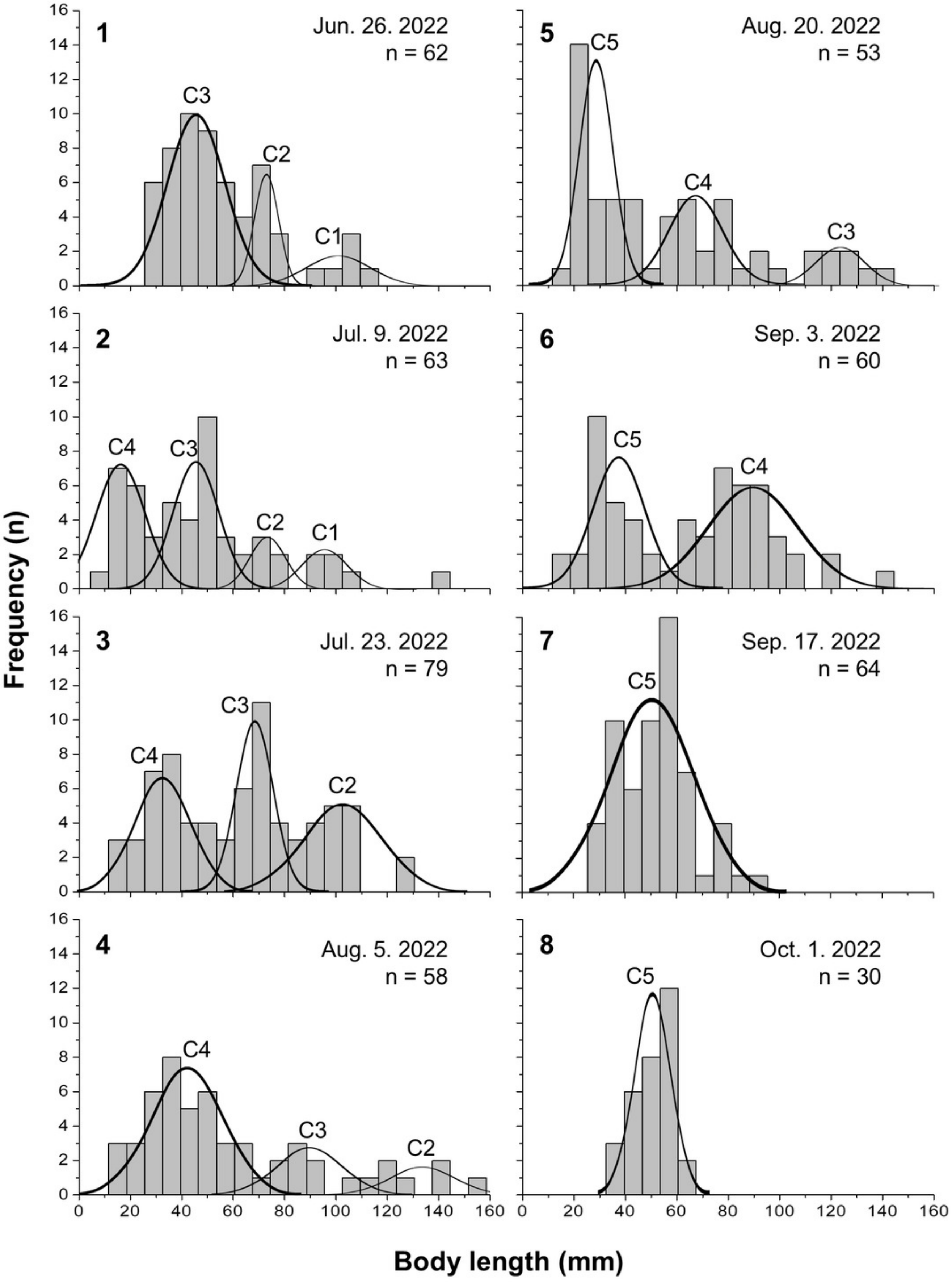
1
Jun. 26. 2022
n = 62
C3
C2
C1
2
Jul. 9. 2022
n = 63
C4
C3
C2
C1
3
Jul. 23. 2022
n = 79
C4
C3
C2
4
Aug. 5. 2022
n = 58
C4
C3
C2
5
Aug. 20. 2022
n = 53
C5
C4
C3
6
Sep. 3. 2022
n = 60
C5
C4
7
Sep. 17. 2022
n = 64
C5
8
Oct. 1. 2022
n = 30
C5
Frequency (n)
Body length (mm)
0
2
4
6
8
10
12
14
16
0
20
40
60
80
100
120
140
160

Figure 3

Time series of mean body length

Time series of mean body length (with standard deviation) for each cohort (1–5) observed over the study period.

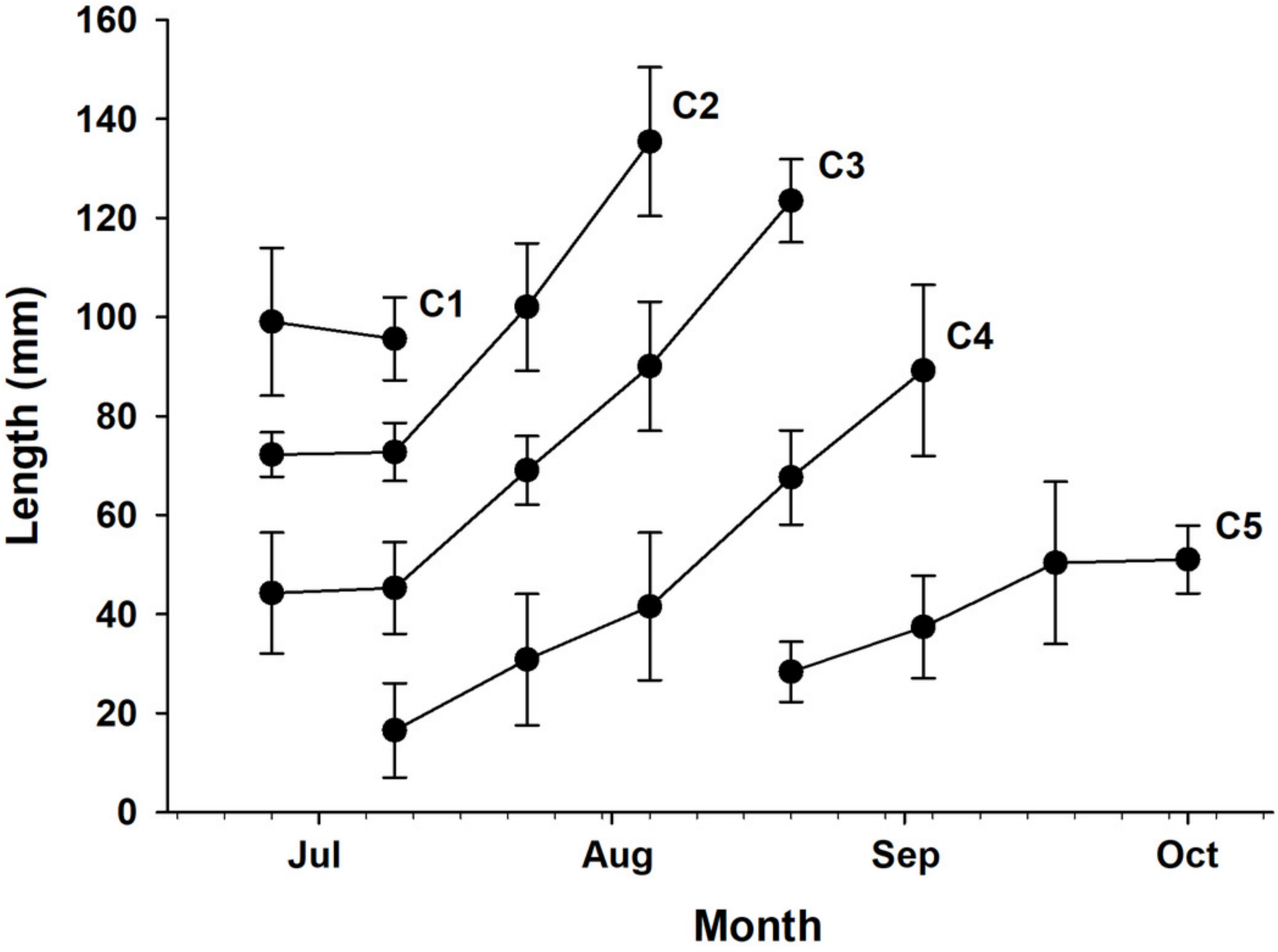

Figure 4

Pearson correlation of growth rate with environmental factors

Pearson correlation of growth rate with water temperature (a) and salinity (b), where the black line represents the regression line.

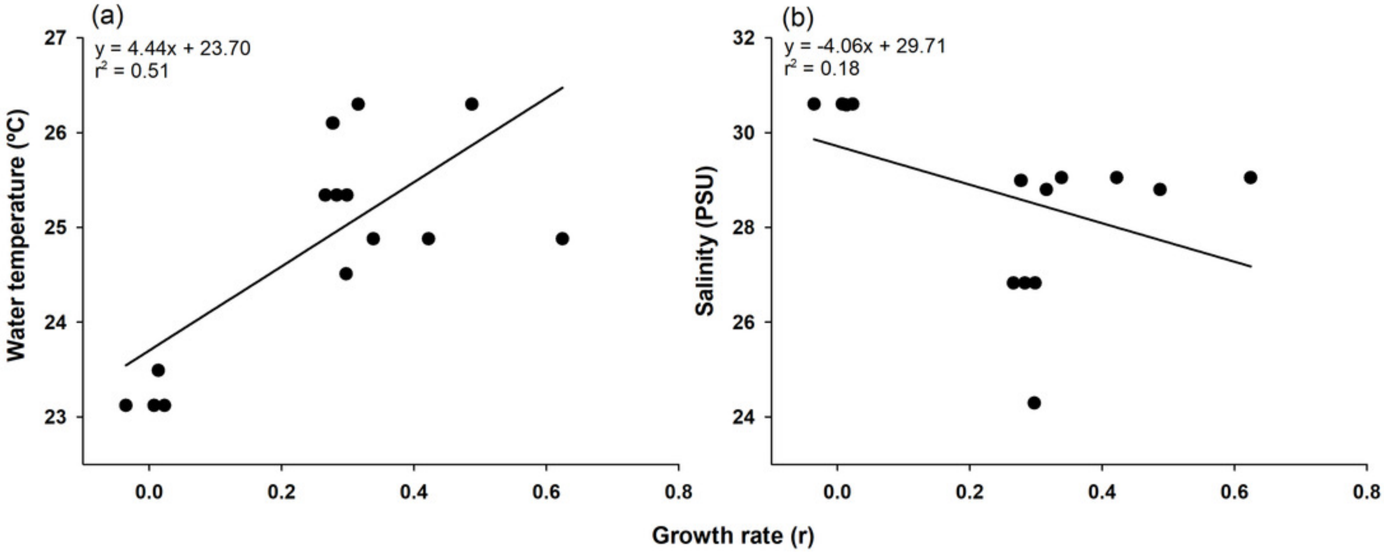

Table 1(on next page)

Characteristics of *Ciona robusta* cohorts

Population measurements and characteristics of cohorts (C1–C5) across different survey dates in 2022.

Date	Survey number	Cohort	Mean body length (mm)	SD	Population	Separation index	n
Jun. 26. 2022	1	C3	44.25	12.21	44		62
		C2	72.22	4.52	10	3.34	
		C1	99.06	14.91	8	2.76	
Jul. 9. 2022	2	C4	16.50	9.53	25		63
		C3	45.28	9.25	24	3.06	
		C2	72.75	5.89	6	3.63	
		C1	95.62	8.34	7	3.21	
Jul. 23. 2022	3	C4	30.80	13.24	31		79
		C3	69.04	6.94	24	3.79	
		C2	102.05	12.88	24	3.33	
Aug. 5. 2022	4	C4	41.52	14.91	38		58
		C3	90.07	13.06	12	3.47	
		C2	135.43	15	8	3.23	
Aug. 20. 2022	5	C5	28.35	6.09	27		53
		C4	67.60	9.55	19	5.02	
		C3	123.50	8.41	7	6.22	
Sep. 3. 2022	6	C5	37.39	10.32	25		60
		C4	89.22	17.27	35	3.76	
Sep. 17. 2022	7	C5	50.34	16.40	64		64
Oct. 1. 2022	8	C5	51.03	6.86	30		30