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The braincase anatomy of *Simosaurus gaillardoti* (Diapsida: Sauropterygia) revealed with X-Ray micro-Computed Tomography

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Sauropterygia is a clade of Mesozoic marine reptiles that includes the eosauropterygian Simosaurus gaillardoti Meyer, 1842, classically considered to be a member of Nothosauroidea. The braincase of this species has thus far only been studied in acidprepared specimens. Acid preparation is a destructive technique prone to information loss, e.g. through the dissolution of thin braincase bones. Here, one well-preserved skull (SMNS 16363) that remains partially embedded in matrix has been visualised using X-ray micro-Computed Tomography and the braincase region has been virtually extracted. This braincase provides valuable information on the general shape of the endocast, the existence and shape of epipterygoids, which were previously considered absent in the taxon, the course of cranial nerves, and the bifurcation of the internal carotid arteries along an expanded and broad parabasisphenoid, the latter extending in a tapering cultriform process to the level of the external and internal narial openings. The arrangement of the semicircular canals of the inner ear confirms previously hypothesised adaptations for near-shore aquatic life in the species. The similarities of the braincase anatomy of Simosaurus gaillardoti and Nothosaurus marchicus support the current phylogenetic placement of the former as an early branching member of Nothosauroidea. The cranial flattening observed in nothosaurs relative to the less dorsoventrally flattened skull of *Simosaurus* reflects diverging feeding strategies. Most nothosaurs were fish-trap ambush predators whereas Simosaurus gaillardoti had durophagous, as well as (opportunistic) piscivorous capacities. These results indicate that specialised piscivorous predation using fish-trap dentition could be independently derived in nothosaurs and in pistosauroids (including plesiosaurs).

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Abstract

- 25 Sauropterygia is a clade of Mesozoic marine reptiles that includes the eosauropterygian
- 26 Simosaurus gaillardoti Meyer, 1842, classically considered to be a member of Nothosauroidea.
- 27 The braincase of this species has thus far only been studied in acid-prepared specimens. Acid
- 28 preparation is a destructive technique prone to information loss, e.g. through the dissolution of
- 29 thin braincase bones. Here, one well-preserved skull (SMNS 16363) that remains partially
- 30 embedded in matrix has been visualised using X-ray micro-Computed Tomography and the
- 31 braincase region has been virtually extracted. This braincase provides valuable information on
- 32 the general shape of the endocast, the existence and shape of epipterygoids, which were
- 33 previously considered absent in the taxon, the course of cranial nerves, and the bifurcation of the
- 34 internal carotid arteries along an expanded and broad parabasisphenoid, the latter extending in a
- 35 tapering cultriform process to the level of the external and internal narial openings. The
- 36 arrangement of the semicircular canals of the inner ear confirms previously hypothesised





37	adaptations for near-shore aquatic life in the species. The similarities of the braincase anatomy of
38	Simosaurus gaillardoti and Nothosaurus marchicus support the current phylogenetic placement
39	of the former as an early branching member of Nothosauroidea. The cranial flattening observed
40	in nothosaurs relative to the less dorsoventrally flattened skull of Simosaurus reflects diverging
41	feeding strategies. Most nothosaurs were fish-trap ambush predators whereas Simosaurus
42	gaillardoti had durophagous, as well as (opportunistic) piscivorous capacities. These results
43	indicate that specialised piscivorous predation using fish-trap dentition could be independently
44	derived in nothosaurs and in pistosauroids (including plesiosaurs).
45	
46	Introduction
47	Sauropterygia is a diverse clade of marine reptiles that lived from the Lower Triassic until the K-
48	Pg mass extinction (c. 252 – 66 Ma) (Rieppel, 2000; Scheyer et al., 2014), but recent studies
49	pushed back the possible appearance of marine reptile clades, including the sauropterygians, to
50	the end of the Permian (Wang et al., 2022; Kear et al., 2023, 2024). Sauropterygia is generally
51	considered to include the two clades Placodontiformes and Eosauropterygia (Fig. 1A-C),
52	although there is an ongoing discussion on the internal topology of the tree and the systematics
53	are far from stable. Some studies divide Eosauropterygia into Pachypleurosauria, Pistosauroidea,
54	and Nothosauroidea (e.g., Scheyer et al., 2017; Allemand et al., 2023). Saurosphargidae was
55	recently added to the phylogenetic analysesas well, but its position either within or closely
56	related to Sauropterygia remains contested (e.g., Wang et al., 2022; Wolniewicz et al., 2023).
57	Placodontiformes appeared in the early Anisian (247-242 Ma) in nearshore environments
58	Placodontia in particular featured a highly modified dentition specialised with flat and broad
59	plate-like teeth facilitating their durophagous diet (Rieppel, 2002; Neenan et al., 2013, 2015;
60	Pommery et al., 2021; Laboury et al., 2023). Arguably the best-understood placodontoid genus,
61	including the skull- and braincase anatomy, is <i>Placodus</i> Agassiz, 1833, known from numerous
62	isolated bones and teeth but also associated skeletons (e.g., Agassiz, 1833-1845; Drevermann,
63	1933; Jiang et al., 2010; Nosotti & Rieppel, 2002; Jiang et al., 2008; Neenan & Scheyer, 2012;
64	Klein et al., 2022). Placodus was a bottom feeder, mostly preying on shelled marine
65	invertebrates (Rieppel, 2002; Klein et al., 2015; Xing et al., 2020). Among Eosauropterygia,
66	Pachypleurosauria potentially appeared in the fossil record in the Olenekian (237 – 227 Ma) with
67	taxa such as Majiashanosaurus discocoracoidis (Jiang et al. 2014; Wang et al., 2022), but its



- 68 placement among Sauropterygia has also been contested, as it was also found outside of
- 69 Eosauropterygia (Wolniewicz et al., 2023), or as successive sister taxon (with *Hanosaurus*) to
- 70 Nothosauroidea plus Pachypleurosauria (Li and Liu, 2020). Pachypleurosauria sensu stricto
- 71 mostly fed on small, soft-bodied animals (Rieppel, 2002) and the majority of pachypleurosaurs
- 72 are equipped with teeth consisting of a single dental morphology: small, conical teeth suited for
- 73 processing soft-bodied invertebrates and fish (Rieppel, 2002). Facultatively durophagous forms
- 74 are also known as is evidenced by *Anarosaurus heterodontus* (Klein, 2009). Pistosauroidea,
- 75 which in the Late Triassic gave rise to the globally distributed open-water plesiosaurs including
- 76 the genus *Plesiosaurus*, were piscivorous (Storrs, 1993; Wintrich et al., 2017).
- 77 Nothosauroidea includes Olenekian forms that are among the oldest currently recognised
- 78 sauropterygian fossils (Li & Liu, 2020; Scheyer et al., 2019; Kear et al., 2024). Nothosaurs are
- 79 mostly found in coastal or platform deposits and are considered piscivorous but potentially
- 80 include the more durophagous *Simosaurus* (Storrs, 1993; Rieppel, 1999, 2002; Klein &
- 81 Griebeler, 2018).
- 82 The family Simosauridae contains two species in monospecific genera: Simosaurus gaillardoti
- 83 (Rieppel, 2000; De Miguel Chaves et al., 2018a; Fig. 1D-F) and *Paludidraco multidentatus* (De
- 84 Miguel Chaves et al., 2018b). P. multidentatus is a relatively recently discovered simosaurid that
- 85 was described based on one specimen found in the Keuper facies in central Spain dating from the
- 86 Carnian to Norian (237 208 Ma) (De Miguel Chaves et al., 2018b). S. gaillardoti appeared in
- 87 the Ladinian stage (242 237 Ma) and disappeared in the Carnian stage (237 227 Ma)
- 88 (Rieppel, 2000; Dalla Vecchia, 2008; Klein et al., 2016; De Miguel Chaves et al., 2018a). The
- 89 species can be found in Upper Muschelkalk and Lower Keuper stratigraphic units of mostly the
- 90 western Tethyan region (e.g., Klein et al., 2016) but has also been described in the Middle East
- 91 (Kear et al., 2010; Cabezuelo-Hernández et al., 2024). The phylogenetic position of S. gaillardoti
- 92 remains debated, with most older studies classifying it as a nothosauroid (Fig. 1) despite distinct
- 93 morphological differences from other nothosauroids (Cheng et al., 2016; Klein et al., 2016; De
- 94 Miguel Chaves et al., 2018a; highly nested within Nothosauroidea as sister taxon to
- 95 Germanosaurus: Li & Liu, 2020). Other recent studies found Simosauridae (i.e., Simosaurus and
- 96 Paludidraco) as sister clade to Nothosauridae plus Pistosauroidea (Wang et al., 2022) or
- 97 Simosaurus as the sister taxon to Pistosauridae (Qiao et al., 2022; Wolniewicz et al., 2023; note
- 98 that other nothosaurs were not included in those studies).



99	Simosaurus gaillardoti could reach 3-4 meters in body length. Its skulls are flat and
100	brevirostrine, meaning they feature a broad jaw with a short rostrum (Rieppel, 1994b). S.
101	gaillardoti crania have been shown to exhibit relatively large variation (Rieppel, 1994a; De
102	Miguel Chaves et al., 2018a). This variation, is for example, expressed in the structural openings
103	on the dorsal side of the skull (De Miguel Chaves et al., 2018a). Compared to other nothosaurs,
104	Simosaurus has large, rounded upper temporal fenestra (Fig. 1, 2). Some nothosaurs have a
105	specialised (mostly size-related) piercing dentition with large stout and often strongly recurved
106	fangs followed by small, more gracile and often less-curved teeth (e.g., Rieppel, 2000). This has
107	been interpreted to facilitate ensnaring or trapping fish within the jaws, rather than piercing them
108	(Rieppel, 2002; Laboury et al., 2023). S. gaillardoti, on the other hand, has teeth that are slightly
109	bulbous in shape and more similarly sized (Rieppel 1994a, 2002). This rather suggests a
110	durophagous feeding strategy for S. gaillardoti (Rieppel, 2002). Based on its jaw mechanics, it
111	might have been able to execute a strong and quick snapping bite to process ammonoids and
112	hard-scaled fish (Rieppel, 1994a). The simosaurid P. multidentatus has teeth that, unlike those of
113	S. gaillardoti, appear adapted for filter-feeding (De Miguel Chaves et al., 2018b).
114	In recent years, visualisation with X-ray micro-Computed Tomography (μCT) has become an
115	increasingly popular instrument for palaeontological investigation, especially to study the
116	internal skull- and braincase anatomy, which are usually not easily accessible (Sutton, 2008).
117	Since μCT scanning is a non-destructive technique that reveals internal structures using density
118	differences in the scans (Germain & Ladevèze, 2021; Leyhr, 2023), this provides improved
119	insight into the morphology and internal anatomy of diverse fossils without damaging or
120	sacrificing rare specimens. Utilisation of μCT scanning thus prevents rare specimens from being
121	destroyed by e.g., acid-preparation (e.g. Rieppel, 1994b) or (serial) thin sectioning (e.g., Klein,
122	2010), thus conserving them for improved future methods, while the virtual data can be made
123	easily available and shared without the need to handle the often very delicate and fragile
124	specimens (Sutton et al., 2016). The number of scanned sauropterygian skulls, including
125	segmented braincases in publications, remain scarce however (summarised for example in
126	Allemand et al., 2023). Sauropterygian skulls that were scanned with μCT include, for example,
127	Nothosaurus marchicus (Voeten et al., 2018), Hispaniasaurus cranioelongatus (Marquez-Aliaga
128	et al., 2019), Keichousaurus hui (Liao et al., 2021), Placodus gigas (Neenan & Scheyer, 2012),
129	Henodus chelyops (Pommery et al., 2021), Parahenodus atancensis (De Miguel Chaves et al.,





130	2020), as well as <i>Dolichorhynchops</i> sp. (Sato et al., 2011) and <i>Libonectes morgani</i> (Allemand et
131	al., 2019). Of the mentioned examples, H. cranioelongatus lacks the posterior braincase region,
132	and H. chelyops and K. hui were so far imaged only for dental evaluation and are thus not
133	associated with a segmented braincase (Marquez-Aliaga et al., 2019; Liao et al., 2021; Pommery
134	et al., 2021). In contrast, braincase and/or endocast descriptions based on CT scan data exist for
135	the plesiosaur Dolichorhynchops sp. (Sato et al., 2011) and Libonectes morgani (Allemand et al.,
136	2019).
137	Concerning nothosauroids, Rieppel (1994b) described the anatomy of acid-prepared skulls of
138	Simosaurus gaillardoti and Nothosaurus sp. Although the acid preparation, a destructive
139	technique, led to a greater understanding of the exterior skull morphology, it failed to capture the
140	comprehensive internal anatomy of S. gaillardoti and often thin and fragile bones within the
141	braincase were lost. More recently, the cranial endocast of Nothosaurus marchicus (Voeten et
142	al., 2018) was described, but detailed renderings of the braincase bones were not included at the
143	time. The present study aims to describe the braincase anatomy by providing renderings of the
144	well-ossified posterior region of the skull alongside the associated endocranial voids of a
145	Simosaurus gaillardoti specimen (Fig. 2A-H; Supplementary Fig. 1) that is still largely covered
146	in sediment matrix. Utilising μCT scan data and 3D reconstruction (also of an additional
147	specimen of S. gaillardoti; see Supplementary Fig. 2) are used to explore and interpret the
148	similarities and differences relative to other nothosaurs in their anatomical, morphofunctional,
149	and phylogenetic contexts.
150	
151	Materials & Methods
152	The material studied consists of the cranium of Simosaurus gaillardoti (SMNS 16363), which
153	was found in the Upper Muschelkalk of Murr, Baden-Württemberg, Germany. The cranium is
154	approximately 38 cm in length and is almost complete, but parts have been reconstructed with
155	plaster for display purposes (Fig. 1D-F; Supplementary Fig. 1). In addition, to verify the
156	morphology of some of the braincase of <i>Simosaurus</i> , an additional specimen (GPIT/RE/09313)
157	was partially segmented. This specimen was mostly prepared out of the sediment matrix except
158	the braincase region. The Simosaurus cranial scan data were then also compared to published
159	scan data on Nothosaurus marchicus (TW480000375; Voeten et al., 2018); Placodus gigas

(UMO BT 13; Neenan & Scheyer, 2012), and the plesiosaurs Dolichorhynchops sp. (ROM

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61	29010; Sato et al., 2011) and <i>Libonectes morgani</i> (D1-8213; Allemand et al., 2019). Due to
62	preservational reasons or difficulties related to clear identification of braincase bones, we refrain
63	from making detailed comparisons with the braincase and cranial endocast of Parahenodus
64	atancensis and the available CT scan data of Keichousaurus hui and Hispaniasaurus
65	cranioelongatus.
66	
67	Institutional abbreviations. D1, collections of the Rhinopolis Museum, Gannat, France; GPIT,
68	Paläontologische Sammlung der Universität Tübingen, Germany; ROM, Royal Ontario Museum,
69	Toronto, Ontario, Canada; SMNS, Staatliches Museum für Naturkunde Stuttgart, Germany; TW,
70	collections of Museum TwentseWelle, Enschede, The Netherlands; UMO, Urwelt-Museum
71	Oberfranken, Bayreuth, Germany.
72	
73	Anatomical abbreviations. aCerC, cerebral carotid artery; aIC, internal carotid artery; aip,
74	anterior inferior process of prootic; aPAL, palatine artery; aPT; pterygoid artery; asc, anterior
75	semicircular canal; ba, basilar artery; bo, basioccipital; cav.ep, cavum epiptericum; cba, cavity
76	for basilar artery; cbr, cerebellum; cer, cerebrum; clp, clinoid process of parabasisphenoid
77	complex; CN V, passage for trigeminal cranial nerve (V); CN VII hy; passage for hyomandibular
78	branch of facial cranial nerve (CN VII); CN VIII, passage for vestibulocochlear cranial nerve
79	(VIII); CN IX-XI, passage for glossopharyngeal (IX), vagus (X) and accessory cranial nerves
80	(XI); CN XII, passage for hypoglossal cranial nerve (XII); crc, crus commune; cup, cultriform
81	process; cqp, cranio-quadrate passage; en, external narial opening; end, endocast; ept,
82	epipterygoid; ept-fco, pterygoid facet of the epipterygoid; ex, exoccipital; fm, foramen magnum;
83	fo, fenestra ovalis; ju.f, jugular foramen; la, bony labyrinth; lsc, lateral semicircular canal; m,
84	sediment matrix; mds, median septum; mo, medulla oblongata; o, orbit; op, opisthotic; opt, optic
85	lobe; pa, parietal; para/pin, parapineal/pineal complex; parf, parietal foramen; pa-sq,
86	undifferentiated parietal-squamosal; pbs, parabasisphenoid; pi, paracondylar interstices; po,
87	pons; pr, prootic; psc, posterior semicircular canal; pt, pterygoid; ptf, posttemporal fenestra; sin,
88	sinus system; so, supraoccipital; sq, squamosal; stc, sella turcica; utf, upper temporal fenestra;
89	vCL, lateral head vein (=vena capitis lateralis); ve, vestibule.
90	



191	Data acquisition. Both cranıa were scanned with a Phoenix vltomelx L240 machine at the Accès
192	Scientifique à la Tomographie à Rayons X facility (AST-RX) in the Muséum national d'Histoire
193	naturelle (MNHN), Paris, France. Micro-X-ray tomography was used to scan the SMNS 16363
194	cranium twice, once to image the complete cranium (Fig. 2) and subsequently to obtain a higher-
195	resolution scan of the braincase region. The complete cranium was scanned with a voltage of 160
196	kV and a current of 500 mA. The acquired voxel size is 0.12739793 mm. The higher-resolution
197	scan of the braincase was scanned with a voltage of 165 kV and a current of 500 mA, and the
198	voxel size is $0.06218133~\text{mm}$. The second specimen, GPIT/RE/09313, was scanned using $150kV$
199	at 500 mA, yielding a voxel size of 0.12354325 mm. Part of the scan data were used in a
200	previous study on the evolution of the sauropterygian labyrinth organ (Neenan et al., 2017).
201	
202	Data processing. Using MIMICS Innovation Suite 24.0 (Materialise, Leuven, Belgium), the
203	individual bones have been identified and traced throughout the μCT image stacks, generating a
204	virtual 3D osteology. In SMNS 16363 the bones that could be identified and traced are the
205	parietal, squamosals, pterygoids (plus associated bones dorsal to the pterygoids), basioccipital,
206	supraoccipital, opisthotics, exoccipitals, prootics and parabasisphenoid (Fig. 3-6). Additionally,
207	anatomical voids were virtually extracted and identified as representing nerves, vasculature and
208	other original soft tissue correlates (Fig. 7-9). These include the cranial endocast, inner ear, sinus
209	system, internal carotid arteries, and cranial nerves. The scan also revealed patches of dark,
210	amorphous material, identified as plaster and artificial infilling, which is not easily identified
211	externally in the specimen (Fig. 1; contrast enhanced in the 3D rendering shown in
212	Supplementary Fig. 1). For the purposes of this study, the posterior-most portion of the
213	braincase, from the upper temporal region backwards, has been segmented in detail. An
214	overview of the entire cranium in lesser detail was segmented and visualised in VGStudio MAX
215	2024.1 (Volume Graphics, Heidelberg, Germany). The "Paint and Segment" tool, which utilises
216	machine learning, was used to expedite the segmentation process. With the tool, discrete classes
217	were assigned. Classes denote different types of material present within the scan, as
218	characterised through different radiodensities (expressed in grey levels) and textures. The chosen
219	classes for this scan were air, bone, matrix, and plaster. For every chosen class, a small part of
220	the skull or otherwise representative material within the scan was segmented. The algorithm
221	subsequently explores the rest of the volume, predicts which sections represent assigned classes,



222	and automatically segments these accordingly. Iterative improvements to correct for
223	misclassification were manually implemented until satisfactory results were achieved. The
224	'create ROIs' (region of interest) tool was then used to create a 3D model of the cranium with th
225	assigned classes as different regions of interest. The segmenting tools allowed the matrix
226	between the teeth to be removed, after which the model was checked for any major inaccuracies.
227	In addition, the parabasisphenoid and associated passages of the internal carotid arteries were
228	also segmented in the entire skull scans of SMNS 16363 and GPIT/RE/09313; to reconstruct the
229	complete antero-posterior extend of that bone within the crania (Fig. 2; Supplementary Fig. 2).
230	
231	Results
232	Systematic Palaeontology
233	Sauropterygia Owen, 1860
234	Eosauropterygia Rieppel, 1994 [Rieppel, 1994a]
235	Nothosauroidea Nopcsa, 1928 [p. 172; not Baur, 1889]
236	Simosauridae Huene, 1948
237	Simosaurus gaillardoti Meyer, 1842
238	
239	Osteological aspects
240	As in other sauropterygian reptiles (e.g., Edinger, 1921; Rieppel, 1994b, 2000; Allemand et al.,
241	2023), the braincase of Simosaurus gaillardoti is formed by the basioccipital, supraoccipital, a
242	pair of opisthotics, a pair of prootics, a pair of exoccipitals and the parabasisphenoid. These
243	bones are bordered on the dorsal side by the parietal and squamosal forming the posterior skull
244	roof and on the ventral side by the pterygoids (Fig. 3-6). Rieppel's (1994b) description of the
245	braincase of S. gaillardoti presented the bones, foramina and other structures, which are used as
246	a basis for the current study.
247	In SMNS 16363, the basioccipital, together with the exoccipitals, forms the ventral and
248	ventrolateral borders of the foramen magnum. The basioccipital can be divided into two parts;
249	the occipital condyle that protrudes straight posteriorly from the cranium on the posterior side
250	and the internal bone that rests on the pterygoid at the posterior border of the cranium Fig. 5). It
251	delimits the cranial endocast ventrally and supports the exoccipitals that project dorsolaterally
252	from the basioccipital. Anteriorly, there is a clear gap separating the basioccipital from the



253	parabasisphenoid, although an in vivo cartilaginous connection cannot be ruled out. The
254	exoccipitals extend between the basioccipital and the supraoccipital, form the ventrolateral
255	borders of the foramen magnum, and are laterally flanked by the opisthotics. The exoccipitals
256	participate in the occipital condyle dorsolaterally, but they are mostly separated from each other
257	by the basioccipital. Each exoccipital bone is pierced by one foramen: the jugular foramen (Fig.
258	3), also called the metotic foramen (sensu Rieppel, 1994b). Laterally, the exoccipitals connect
259	with the opisthotics.
260	The sutures between these bones are difficult to trace on the surface and in the scan data of
261	SMNS 16363, so a small lateral connection between the opisthotics and the lateral portion of the
262	basioccipital cannot be excluded.
263	
264	The supraoccipital defines the dorsal border of the foramen magnum and its smooth, concave
265	ventral surface saddles the endocast. It has a medial crest with lateral extensions flanking it on
266	both sides. Anteriorly, this supraoccipital crest bifurcates, creating a Y-shape in dorsal view (Fig.
267	5A). The lateral surface is very irregular and the left and right side are not completely
268	symmetrical in shape (Fig. 5C, D). The opisthotics connect with the exoccipitals and the
269	supraoccipital and rest on the pterygoid laterally. On the dorsal and posterolateral side, the
270	opisthotic meets the squamosal (Fig. 3), and the posttemporal fenestra perforates the suture
271	between these two bones. The opisthotic meets the prootic on a small plane further anterior in the
272	skull, which is visible only in lateral view (Fig. 5E, F). The fenestra ovalis (= fenestra vestibuli)
273	is formed between the opisthotic and the prootic. No structure was found in SMNS 16363 that
274	would indicate a stapes articulating with the fenestra ovalis. The prootic meets the supraoccipital
275	dorsolaterally, but it is not visible in posterior view as it is obscured by the opisthotic,
276	supraoccipital, and exoccipital in SMNS 16363. The prootic is a round-shaped element with a
277	spongy internal bone structure that houses part of the vestibular system. The prootic meets the
278	parietal dorsally, the supraoccipital posterodorsally, the opisthotic posteroventrally and it has an
279	anterior inferior process that meets the parabasisphenoid ventromedially. The anterior border of
280	the prootic frames the oval opening for the trigeminal cranial nerve (CN V) ventrally, posteriorly
281	and dorsally (Fig. 5E; the anterior border of the opening remains free as there is no ossified
282	laterosphenoid in SMNS 16363). The parabasisphenoid consists of two sections: the posterior
283	section includes the sella turcica, posteriorly bounded by a low dorsum sellae, and the anterior



284	section forms wing-like lateral projections and a narrow medial process that projects anteriorly
285	(Fig. 2, 5, 6). The sella turcica (Fig. 5A) is a saddle-shaped structure of the basisphenoid on the
286	ventral side of the cranium, framed by bilateral (antero)dorsal extensions that are identified as
287	the parabasisphenoid clinoid processes. As noted already for other Simosaurus specimens in
288	Rieppel (1994), the sella turcica does not show a median dorsal ridge in the specimens studied
289	herein. Due to damage and subsequent plastering of the pterygoids in SMNS 16363, the anterior
290	extension of the parabasisphenoid and its relationship with the pterygoid is locally difficult to
291	interpret. The wing-like lateral expansions and the medial process, expanding into a median
292	septum anteriorly, form two distinctive anteroposteriorly trending trough-like structures (the
293	cavum epiptericum described for Nothosaurus and Simosaurus by Rieppel, 1994b). Two
294	prominent channels traverse the posterior portion of the parabasisphenoid that house the carotid
295	arteries (see below). In GPIT/RE/09313, the parabasisphenoid and arrangement of the carotid
296	passages was found to have overall the same shape as in the SMNS specimen, although in the
297	former, the median septum and the clinoid processes are slightly more anteriorly expanded and
298	inclined (hook-shaped; Supplementary Fig. 2). Slightly more anteriorly inclined clinoid
299	processes are also visible in SMNS 10360 (Rieppel, 1994: fig. 6). In both our CT scans, the
300	median septum shows a slightly concave posterior margin, a flat and narrow dorsal top, and an
301	excavated anterior margin. A Y-shaped structure of the median septum in cross section as
302	described by Rieppel (1994b: p. 14) could not be observed as both septa were laterally
303	constricted in our studied specimens. The troughs lateral to the median septum are well marked
304	only until the anterior border of the median septum and then taper anteriorly into a cultriform
305	process which extends anteriorly to the level of the external nares. The cultriform process (Fig.
306	2; Supplementary Fig. 2) is laterally crested so the dorsal surface is concave (forming a wide
307	dorsal groove along the process). Grooved and tapering cultriform processes were also described
308	by a wide range of taxa, including for example the neodiapsid Youngina capensis (Gardner et al.,
309	2010), the sauropodomorph dinosaur Massospondylus carinatus (Chapelle & Joiniere, 2018),
310	and the millerettid Milleropsis pricei (Jenkins et al., 2025). In those taxa, the cultriform process
311	does not reach the level of the external nares as is the case in Simosaurus. Towards its tip, the
312	cultriform process seems to bifurcate slightly, which could be due to taphonomic processes. A
313	similar bifurcation was observed in GPIT/RE/09313, but in this specimen the whole anterior
314	palatal region split medially due to taphonomic distortion of the cranium.





315	The parietal is an unpaired and medially placed bone (Fig. 3) of the skull roof and it encloses the
316	cranial endocast dorsally and dorsolaterally. The parietal foramen is well-defined, being medially
317	positioned in the posterior skull table (posterior to the widest mediolateral expansion of the upper
318	temporal fenestrae).
319	The suture between the parietal and the squamosal is partially obliterated externally as well as
320	internally, rendering delimited segmentation of these elements impossible (Fig. 4). The
321	posttemporal fenestra, between the squamosal and the opisthotic, leads to the fenestra ovalis
322	(Fig. 5). The pterygoids form the palate in the braincase region (Fig. 6). Breaks in the pterygoid
323	are filled with plaster mostly on the ventral side (Fig. 6B) thus obscuring large parts of the
324	medial suture between the right and left pterygoid (Fig. 4, 6). Thus, the two pterygoids have been
325	segmented as a single unit. The pterygoid is a relatively flat bone in ventral view, forming a
326	sturdy posteriorly extending quadrate ramus along which the quadrate is sutured laterally (note
327	that the quadrate is not separately segmented herein because only a small portion was visible in
328	the scan data of the higher resolution scan of the braincase region of SMNS 16363). In dorsal
329	view, a thin u-shaped groove extends in posterolateral-anteromedial direction over the pterygoid
330	surface, which anteriorly gets covered by a thin hook-shaped bone with a broad and narrow
331	ventral base. Extending along both sides of the endocast and extending anterodorsally, these
332	bones are interpreted as epipterygoids (Fig. 3, 4, 6), which in contrast to Nothosaurus do not
333	contact the parietal dorsally (Rieppel, 1994b). Within the pterygoid on the posterior side, the
334	foramen for the internal carotid resides (see below). Between the pterygoid and the basioccipital
335	large excavations exist that taper anteriorly (i.e, the paracondylar interstices; Fig. 3F).
336	
337	Cranial endocast
338	Only the posterior portion of the cranial endocast of SMNS 16363, up to the level of the median
339	septum of the parabasisphenoid was segmented out (Fig. 3, 7, 8).
340	Here, the endocranial void is delimited mostly by the dorsal, lateral and and ventral braincase
341	elements. Anteriorly the endocast is not delimited laterally by bone. As such, the lateral edges of
342	the endocast (Fig. 7, 8) are interpolated straight lines between the lowest point on any dorsal
343	bone and the highest point on any ventral bone (Voeten et al. 2018). The endocast is wider
344	dorsally than ventrally, granting it an oblong shape in both posterior and anterior view (Fig. 8).
345	Posterodorsally, the endocast portion is delimited by the supraoccipital, posterolaterally by the





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exoccipitals and opisthotics, posteroventrally by the basioccipital, ventrally by the parabasisphenoid, laterally by the prootics (and by the hook-like epipterygoids), and dorsally/dorsolaterally by the parietal. The endocast smoothly transitions into the sinus system posterodorsally (Fig. 8A, D), which renders delimitation of the cerebral endocast and adjacent sinusal structures locally challenging. The part of the endocast that is delimited by the parietal foramen projects vertically. projects vertically, where it expands slightly in diameter, and is dorsal portion is slightly offset to the left in SMNS 16363. The endocast is vaulted and relatively smooth posterior to the parietal foramen (which might have housed a pineal/parapineal gland; see Smith et al., 2018) and narrows posteriorly. Anterior to the level of the parietal foramen, the cerebral region of the endocast is present but not well defined laterally. On the posteroventral side, the endocast is delimited by the basioccipital, which defines a median groove with two flanking lobes. Anterior to this, the endocast is artificially smoothed during segmentation due to the absence of any ventral constriction. Here, the ventral boundary was interpolated between the basioccipital and the sella turcica. The sella turcica (Fig. 5A) forms an irregular relief on the endocast. Anterior to the sella turcica, the endocast is constricted by plaster and the pterygoid, which produce a bumpy relief until the ventral groove formed by the median septum of the parabasisphenoid is expressed. The internal carotids enter into the cranial endocast ventrally, and only a few of the posterior brain nerves could be traced to exit the endocast laterally (see below).

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Nerves and blood vessels

The internal carotid arteries enter the cranium through the internal carotid foramen on the quadrate ramus of the pterygoid (Fig. 7, 8). The internal carotids then extend along the pterygoid dorsally and ventral to the parabasisphenoid (Fig. 6F). Distinct ridges are visible on the ventral side of the parabasisphenoid tracing the course of the internal carotid arteries. On the level of the clinoid processes of the parabasisphenoid, the carotid arteries bifurcate (Fig. 7A, B). The medial branches are the cerebral carotids, which exist the parabasisphenoid dorsally through two carotid foramina and then merge with the cranial endocast where they cannot be followed anymore. The lateral branches are the palatine arteries. Both palatine arteries can be followed for a short distance more anteriorly, after which they cannot be traced further. The two ventral lobes of the posteroventral endocast (Fig. 8C, D) which are expressed as two anteroposteriorly elongate





377	depressions in the basioccipital (Fig. 5H) likely housed paired basilar arteries that enter here into
378	the endocast. Similar paired basilar arteries were discussed for the mosasaur <i>Platecarpus</i>
379	(Russell, 1967) and have been segmented out also in braincase of Plioplatecarpus peckensis
380	(Cuthbertson et al., 2015). The tubular groove (that partly could be roofed over) extending
381	postolaterally-antero*medially over the pterygoid wings is interpreted to potentially have housed
382	the lateral head vein (=vena capitis lateralis; see Russell, 1967).
383	A complex neurovascular system pervades the pterygoid as could be identified on the left side in
384	SMNS 16363 (Fig. 3, 7, 8). Posteriorly on the ventral surface of the pterygoid, a foramen is
385	present, identified as the likely entry of an artery into the pterygoid, but also potentially housing
386	parts of the palatal branch of the facial cranial nerve (CN VII). This passage within the pterygoid
387	extends as a larger canal that courses anteriorly, parallel to the internal carotid arteries. Smaller
388	branches split off this canal, departing the pterygoid through a number of smaller foramina, three
389	of which open onto the pterygoid dorsally and one foramen on the ventral side. The main canal
390	extends further anteriorly and is visible through the foramen of the CN VII. Slightly posterior to
391	the facial foramen, the canal splits into a complex branching system (leading to the
392	aforementioned smaller foramina), which extends more laterally. At the level of the parietal
393	foramen, the canal exits through the foramen for the presumed palatine branch of CN VII. A
394	similar canal network was likely present also in the right pterygoid as evidenced by few small
395	foramina on the dorsal bone surface, but it is only partially preserved due to the breakage of bone
396	and reconstruction with plaster.
397	The jugular foramen is a structure that opens into the endocast and likely accommodated the
398	cranial nerves IX-XII in SMNS 16363 (Fig. 3F, 7). The posteroventral portion likely
399	accommodated CN XII and the more dorsal part for the passage of CN IX-XI (Fig. 8B, C).
400	Another structure oblique to the semicircular canals of the inner ear would have connected with
401	the cranial endocast, likely housing the vestibulocochlear nerve (CN VIII).
402	
403	Other soft tissue structures
404	Connected to the endocast on the posterior-dorsal part is a large system of cavities (Fig. 8)
405	pertaining to the cranial sinus system (Witmer et al., 2008; Dufeau & Witmer, 2015; Voeten et
406	al., 2018). It can be followed from the connection to the endocast posteriorly between the skull
407	roof bones (i.e., the parietal and squamosals) and the prootic. It then crosses over into the cavity





400	connected to the positemporar renestra between the opisinotic and partetar-squamosar. within the
409	opisthotics, prootics and supraoccipital, the endosseous labyrinth can only be partially
410	reconstructed in SMNS 16363 (Fig. 7-9). The main body of the labyrinth, the vestibule, is oval-
411	shaped. Most of the posterior semicircular canal (SC) could be reconstructed, meeting the
412	anterior SC in the crus commune, whose ventral connection with the vestibule could not be
413	completely reconstructed (Fig. 8E, 9A-D). Of the lateral and anterior SC, only the posterior
414	(opisthotic) portions were traceable, whereas the canals could not be traced much further after
415	entering the prootic. As reconstructed, the right and left labyrinth portions in SMNS 16363
416	generally agree with the morphology seen in the labyrinth extracted from GPIT/RE/09313
417	(Neenan et al., 2017: p. 3853).
418	
419	Discussion
420	Osteological aspects
421	Much of Rieppel's (1994b) original identification of braincase morphology and interpretations of
422	associated soft tissues can be confirmed by the present study of SMNS 16363 and
423	GPIT/RE/09313. The usage of CT scan data of less or unprepared specimens, however, allowed
424	for the identification and re-interpretation of some cranial structures related to the braincase
425	region.
426	One of the surprising finds was the identification of clear epipterygoids in SMNS 16363 (Fig. 3,
427	4, 6, 9E-J). In contrast to Rieppel (1994b) stating the seeming absence of epipterygoids in
428	Simosaurus (based mainly on acid-prepared specimens such as SMNS 10360, 50714 and 50715),
429	two hook-shaped epipterygoids could be identified, whereas their absence in other specimens
430	seems to be thus a preparatory (or preservational) artefact. The left epipterygoid bone has a small
431	perforation in its triangular ventral portion (i.e. the pterygoid facet of the bone; Fig. 3C, 6D, E).
432	However, this hole represents an artifact associated with a mechanical fracture (see Fig. 4B)
433	rather than an original anatomical feature. In GPIT GPIT/RE/09313, the area where the
434	epipterygoids would have been placed, is clearly damaged (Supplementary Fig. 2), as is the
435	region in SMNS 10360 and SMNS 16767. The epipterygoids in SMNS 16363 lack a parietal
436	contact and flank the anterior extend of the posterior ossified part of the parabasisphenoid
437	complex. The latter expands into a median septum as indicated by Rieppel (1994b). The
438	anatomical location of the median septum agrees with the ventral-dorsal extension of the



439	parabasisphenoid in SMNS 16363 anterior to the level of the parietal foramen, but a dorsal
440	expansion (into a Y-shaped structure) was not observed in both CT-scanned specimens studied
441	herein. Towards anterior, the parabasisphenoid extends as a tapering thin and grooved cultriform
442	process up to the level of the external/internal nares. The process thus likely supported the
443	infraorbital to rostral endocast (i.e., the olfactory tract) ventrally, as was shown also for
444	Nothosaurus marchicus (Voeten et al., 2018).
445	
446	The excavations lateral to the basioccipital have been referred to as eustachian foramen, as they
447	were interpreted as housing the eustachian tube or the internal carotid arteries in Nothosaurus
448	(Koken, 1893). As such, the eustachian tube was interpreted as connecting the middle ear and the
449	pharyngeal region in these animals (Rieppel, 1994b). As shown by the previous study on
450	Nothosaurus marchicus (Voeten et al., 2018), these excavations, termed paracondylar interstices
451	in that study, did neither house the internal carotids, nor did they connect to the pharyngeal
452	region (and thus not serving as a connection with the middle ear). In SMNS 16363, the
453	paracondylar interstices (Fig. 3F) can be followed until the anterior end of the basioccipital, from
454	where they do not connect to the endocast or the middle ear. As in N. marchicus (Voeten et al.,
455	2018), any tiny spaces extending anteriorly to the paracondylar interstices are due to blunt
456	abutting rather than a tight suturing of the braincase bones in this region.
457	In SMNS 16363, the medial pillars of the exoccipitals articulate more posteriorly and
458	posterolaterally with the basioccipital, so that we here consider them to participate in the
459	formation of the occipital condyle, in contrast to what has been reported for some other
460	specimens of Simosaurus gaillardoti (Rieppel, 1994a, b).
461	As discussed by Rieppel (1994b), the location of the fenestra ovalis and structure of adjacent
462	cranial elements are different between Simosaurus (SMNS 10360, 15860, 50714 and 50715) and
463	Nothosaurus (SMNS 59075 and 59076). In Simosaurus, the fenestra ovalis is located between
464	the opisthotic and prootic and is visible in lateral view, whereas the fenestra ovalis is not visible
465	in lateral view in Nothosaurus (Rieppel, 1994b). Similar to Rieppel (1994b), no stapes was
466	encountered in the studied specimens of Simosaurus herein.
467	The posttemporal fenestrae are arranged differently in S. gaillardoti compared to the reduced
468	posttemporal foramen in N. marchicus, which in the latter are medial to the paracondylar
469	Interstices (Voeten et al., 2018) but laterally placed in SMNS 16363. Voeten et al. (2018)





470	rejected the term 'eustachian foramen' (sensu Rieppel, 1994b; De Miguel Chaves et al., 2018a),
471	rendering the placement and geometry of the paracondylar Interstices potentially more flexible,
472	while arguing against the functions otherwise associated with an eustachian foramen.
473	The posttemporal fenestra (Fig. 3F) is located between the opisthotic and squamosal in
474	Simosaurus, whereas the reduced posttemporal foramen in Nothosaurus is bordered by the
475	exoccipital, supraoccipital and in some cases the opisthotic (e.g., Rieppel, 1994b; Voeten et al.,
476	2018; Shang et al., 2022). In SMNS 16363, the posttemporal fenestrae are not as large as
477	observed in <i>Placodus gigas</i> (Sues, 1987), but they are clearly visible and well-defined.
478	Placodonts were obligately durophagous, feeding primarily on hard-shelled prey (Rieppel, 2002).
479	S. gaillardoti was also durophagous but likely supplemented its diet by catching fish using rapid
480	snapping motions (Rieppel, 1994a). N. marchicus, in contrast, captured fish through a quick side-
481	swipe motion with its distinct fish-trap dentition (Rieppel, 2000). This morphofunctional
482	gradient coincides with a shift from large and well-defined posttemporal fenestrae in basal
483	Triassic Sauropterygia to reduced or absent posttemporal foramina in nothosauroids. Progressive
484	reduction of posttemporal fenestrae/foramina thus appears complementary to the development or
485	enlarged upper temporal fenestrae in nothosauroids (Rieppel, 1994b) relative to placodonts,
486	which are the sister taxon to Eosauropterygia. Continued specialisation and the establishment of
487	obligate piscivory with clear fish-trap dentition in nothosaurids may explain the difference
488	between the size of posttemporal fenestrae/foramina in Simosaurus and Nothosaurus,
489	respectively.
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491	Cranial endocast
492	The general shape of the endocast of S. gaillardoti is similar to that of Nothosaurus (Fig. 10),
493	being elongated and tapering anteriorly with an anteroposterior orientation (e.g., Edinger, 1921;
494	Voeten et al., 2018). The endocast of <i>P. gigas</i> is also elongated (Fig. 10C, D), but the shape is
495	anteroposteriorly sigmoidal (Neenan & Scheyer, 2012). The most striking difference between the
496	endocasts of S. gaillardoti (Fig. 10A, B) and N. marchicus (Fig. 10E, F) is the dorsoventral
497	flatness of the latter (Voeten et al., 2018). The endocast of S. gaillardoti is narrower with a
498	vertically oriented oval shape, whereas the endocast of <i>N. marchicus</i> has a more horizontally
499	oriented oval shape. The flatness of the N. marchicus endocast can be attributed to its
500	dorsoventrally flattened skull inferred to be hydrodynamically specialised for lurching ambush
495 496	anteroposteriorly sigmoidal (Neenan & Scheyer, 2012). The most striking difference between the endocasts of <i>S. gaillardoti</i> (Fig. 10A, B) and <i>N. marchicus</i> (Fig. 10E, F) is the dorsoventral
498	vertically oriented oval shape, whereas the endocast of <i>N. marchicus</i> has a more horizontally
	•
300	doisoventially flauched skull inferred to be hydrodynamically specialised for furcilling allibush



501	predation (Voeten et al., 2018). The less flattened endocast of SMNS 16363 is thus likely
502	associated with its less dorsoventrally flattened skull (which also might reflect a requirement for
503	more robusticity/rigidity in a cranium that engages in facultative durophagy), indicating a more
504	broad-spectrum feeding strategy that included both durophagy and piscivory (Rieppel, 1994a).
505	The endocast of S. gaillardoti was also compared with the endocast of Libonectes morgani
506	(Allemand et al., 2019). L. morgani is an elasmosaurid with an anteroposteriorly elongated
507	endocast (Fig. 10d). Unlike S. gaillardoti and N. marchicus, the endocast of L. morgani (Fig.
508	10G, H) is dorsoventrally thick and distinctly thins at the anteroposterior level of the para-/pineal
509	complex. Similarly, the endocast of <i>Dolichorhynchops</i> sp. would be posteriorly thickened based
510	on the reconstructed braincase by Sato et al. (2011), although its anterior endocast portion
511	remains elusive due to the lack of preserved bone in ROM 29010. The endocast of L. morgani
512	also shows two well-defined lobes on the dorsal side posterior to the parapineal/pineal complex
513	(Fig. 10G, H). The posterior lobe was interpreted to represent the cerebellum, and the anterior
514	lobe associated with the optic lobe (Allemand et al., 2019; 2023). The endocast of S. gaillardoti
515	shows a singular lobe posterior to the pineal complex (Fig. 10A), partly segmented as the sinus
516	system. Extrapolating from the condition of <i>L. morgani</i> , this lobe could be either the cerebellum
517	or the optic lobe. The optic lobes are placed more laterally in N. marchicus, which would suggest
518	the referred lobe is the cerebellum in SMNS 16363. However, the cerebellum is often
519	unpronounced in marine reptiles, and here even less developed than in plesiosaurs (Allemand et
520	al., 2019). Thus, whether this lobe is the optic lobe, or the cerebellum remains unclear. The
521	boundary between the pons (posterior-most midbrain) and the medulla oblongata (anterior-most
522	hindbrain) could be recognised in N. marchicus based on a faint flexure (Fig. 10F) (Voeten et al.,
523	2018). In S. gaillardoti, the endocast has a relatively consistent shape at the posterior-most
524	portion, then expands laterally at the anterior end of the basioccipital And this could be the
525	border of the medulla oblongata (Fig. 10A). It must be noted that the ventral border between the
526	basioccipital and the parabasisphenoid is not well delimited due to a gap between both bones
527	(which may have featured a cartilaginous connection during life; Fig. 6, 7). The boundary
528	between pons and medulla oblongata border could thus also be placed more anteriorly in this
529	transitional region.
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Nerves and blood vessels



532	The internal carotid artery enters the skull through the jugular foramen which is more dorsally
533	and medially placed in S . $gaillardoti$ than in N . $marchicus$. The internal carotid arteries also split
534	more anteriorly in S. gaillardoti than in N. marchicus and in plesiosaurs, . In the latter, they split
535	even more anteriorly than in S. gaillardoti, specifically anterior to the basisphenoid (Allemand et
536	al., 2023). The palatine nerve may have been housed by the branch containing the palatine artery
537	in S. gaillardoti, which is the lateral branch after the bifurcation (Rieppel, 1994b).
538	Similar to the condition in Nothosaurus spp. (Rieppel, 1994b; Voeten et al., 2018) only the para-
539	and basisphenoid complex region is ossified and fused in Simosaurus gaillardoti, whereas the
540	more lateral/dorsal sphenoid regions remain un-ossified (i.e., a laterosphenoid is absent). As
541	such, the cranial nerves I-IV can also not be reconstructed using the CT scanned specimens of
542	Simosaurus and only a portion of the CN V is framed by the prootic creating only a partial
543	prootic 'fenestra'. Although Neenan and Scheyer (2012) also indicated the absence of a
544	laterosphenoid in <i>Placodus gigas</i> , a complete prootic fenestra is shown, surrounded by the bone
545	of the prootic. Alternatively, the anterior and anterodorsal part of the fenestra might not be
546	constructed by the prootic but by a thin laterosphenoid instead. A similar condition was, for
547	example, also described in the early branching archosauromorph Tanystropheus hydroides
548	(Spiekman et al., 2020). The laterosphenoid is generally considered a typical structure found in
549	taxa of the archosaur lineage including extant crocodylians and birds (e.g., Kuzim et al. 2021),
550	although Rieppel (1976) argued for referral of the bone in archosaurs as pleurosphenoid instead.
551	In the latter study, the laterosphenoid of snakes was considered to be a non-homologous structure
552	to that of archosaurs, based on differences in cranial development. The term laterosphenoid
553	seemed to have gained traction in archosaur research, however, and a morphologically similar
554	bone to that in the archosaur cranium has been proposed as a homologous element in the cranium
555	of the stem turtle <i>Proganochelys quenstedtii</i> as well (Bhullar & Bever, 2009). Evers et al. (2019),
556	however, argued that the fenestra prootica could not be considered a homologous structure to the
557	trigeminal foramen, a secondarily formed structure in 'anatomically modern turtles' (Scheyer et
558	al., 2022), further raising questions of homology of this part of the reptilian braincase feature, but
559	a review of this feature would go beyond the scope of this article.
560	Rieppel (1994b) hypothesised that the palatine branch of the facial nerve could have emerged
561	from the facial foramen and entered the foramen for the palatine branch of the facial nerve
562	(VIIpl). The course of the facial nerve is unclear in our observed specimen, but this solution





563	seems unlikely considering the presence of the canal within the pterygoid between the facial
564	foramen and VIIpl, as well as the presence of the canal more posteriorly than the facial foramen.
565	According to Rieppel (1994b), each exoccipital in S. gaillardoti would carry two foramina, a
566	larger anterior and a smaller posterior foramen, that house the hypoglossal nerve (CN XII). The
567	specimen SMNS 16363 studied here in detail only shows one opening that corresponds with the
568	jugular foramen, which is also called the metotic foramen in Rieppel (1994b). According to the
569	scan data, we hypothesise that the single jugular foramen in SMNS 16363 corresponds to a
570	combined hypoglossal and metotic foramen.
571	According to Rieppel (1994b), the foramen piercing the prootic ventral to the fenestra ovalis
572	could have housed the hyomandibular branch of the facial nerve (VIIhy). The prootic in SMNS
573	16363 is porous and features a large foramen that correlates with Rieppel's (1994b) observations.
574	Of the cranial nerves exiting the posterior portion of the brain, the glossopharyngeal (CN IX) and
575	vagus (CN X) cranial nerves are intimately related and have been interpreted to enter through the
576	jugular foramen in N. marchicus (Voeten et al. 2018). It seems possible that these nerves,
577	together with the hypoglossal (CN XII) nerve (and the spinal accessory nerve CN XI) passed
578	through the jugular foramen in S. gaillardoti. This condition would differ from P. gigas, in
579	which these nerves do not enter through the same foramen (Neenan & Scheyer, 2012). The
580	hypoglossal (CN XII) nerve in P. gigas exits through a foramen in between the exoccipital and
581	the basioccipital (potentially a derived condition in placodonts), and the glossopharyngeal (CN
582	IX) and vagus (CN X) nerves pass through the jugular foramen. The location where the nerves
583	enter the cranial endocast is different between S. gaillardoti and N. marchicus. The nerves exit
584	more ventrally in N. marchicus than in S. gaillardoti. In S. gaillardoti, the nerves merge into the
585	endocast from the foramen magnum. The course of the nerves can also be followed further
586	anterior in N. marchicus (Voeten et al., 2018), where they merge with the endocast ventral to the
587	foramen magnum.
588	
589	Other soft tissue structures
590	The large dorsal system connected to the endocast is likely the sinus system or houses parts of
591	the middle cerebral vein (Voeten et al., 2018). The structure has a similar placement in N.
592	marchicus but has a smoother transition into the endocast in S. gaillardoti than in N. marchicus,
593	where it emerges relatively perpendicularly to the endocast.





594	The inner ear has been shown to have adaptations to aquatic life, where the semicircular canals
595	have decreased height and/or increased width (Georgi & Sipla, 2008). Neenan et al. (2017)
596	showed that the inner ear altered with changes in locomotion in marine reptiles. The inner ear of
597	Placodus gigas was distinctly larger than the inner ears of all other members of Sauropterygia,
598	following also general shape changes with the transition from near-shore swimmers such as
599	Nothosaurus spp. and S. gaillardoti to pelagic swimmers such as Callawayasaurus colombiensis
600	and Libonectes morgani. The endosseous labyrinth of plesiosaurs was found distinctly more
601	compact, showing anteroposterior shortening and thickening of the semicircular canals (Neenan
602	et al., 2017). These adaptations align with the morphology of the endosseous labyrinths of an
603	extant crocodile (Crocodylus acutus) and an extant sea turtle (Lepidochelys olivacea). The inner
604	ears of the extinct near-shore swimmers resembled that of the crocodylian, whereas the pelagic
605	swimmers look more similar to that of the extant sea turtle. The inner ear of SMNS 16363 (Fig.
606	8E) has a similar morphology to that of GPIT/RE/09313, the <i>Nothosaurus</i> sp. specimen (NME
607	16/4), and the Augustasaurus hagdorni specimen (FMNH PR 1974) used in Neenan et al. (2017)
608	and thus reflects the adaptions for aquatic life in near-shore environments.
609	
610	Palaeoecological implications
611	SMNS 16363 has a braincase morphology that is quite similar to that of <i>Nothosaurus marchicus</i>
612	(specimen TW480000375). The two share similar locations of fenestra, such as the posttemporal
613	fenestra, and overall attachments of bones, such as the placement of the exoccipitals,
614	supraoccipital and opisthotics (Rieppel, 1994b). These conditions are markedly different in
615	placodonts. Additionally, internal structures such as the (para-)pineal complex and the endocast
616	have a broadly similar shape (Fig. 10). The morphology of the inner ear in nothosaurs and
617	simosaurs share the adaptions for near-shore aquatic life as they both feature a smaller
618	endosseous labyrinth than placodonts, but dorsoventrally more elongated semicircular canals
619	compared to the pelagic plesiosaurs (Neenan et al., 2017). Based on these similarities, the present
620	study supports a closer relationship of Simosaurus with other nothosaurs within Nothosauroidea.
621	The near-vertical projection of the parapineal/pineal complex in Simosaurus relative to its subtly
622	forward-slanting orientation in Nothosaurus (Voeten et al., 2018) could reflect a slightly
623	different mode of cranial development to facilitate the more specialised derived cranial
624	architecture in the latter. Nothosauria have been interpreted as piscivorous ambush predators





with fish-trap dentition (Rieppel 2002; Voeten et al., 2018), whereas *Simosaurus gaillardoti* has a more generic (opportunistic) feeding strategy, including facultative durophagy (Rieppel 2002). This is supported here by the dorsoventrally flattened endocast of *Nothosaurus marchicus* (TW480000375; Voeten et al., 2018) relative to the less dorsoventrally flattened endocast (and skull) of SMNS 16363. Durophagy is also seen in the near-shore Placodontiformes which is usually regarded as the most basal sauropterygian clade (Rieppel, 1999, 2000, 2002). Like nothosaurs, pistosaurs have been interpreted as piscivorous (Storrs, 1993). Thus, piscivory with clear fish-trap dentition is interpreted here to have developed multiple times independently in several sauropterygian clades.

Conclusions

The braincase of S. gaillardoti had only been described exteriorly, using acid-prepared skulls and only a limited number of 3D segmented braincases of other Sauropterygia exist. The virtually segmented braincase of SMNS 16363 (and that of the partly segmented GPIT/RE/09313) gives new valuable information not available previously for Simosaurus gaillardoti. This provides a better understanding of the internal morphology of the bones and soft tissue structures, such as cranial nerves and vasculature. It also indicates the existence of epipterygoids in S. gaillardoti not observed earlier in acid-prepared specimens. The inner ear of SMNS 16363 is comparable to that of other near-shore sauropterygians in overall shape, sharing the adaptations for aquatic life but shallower aquatic than the open-water plesiosaurs as was previously indicated (Neenan et al., 2017). The braincase anatomy and the brain endocast support a phylogenetic placement of Simosaurus gaillardoti close to other nothosauroids. Also, piscivory is the currently proposed feeding strategy for most nothosaurs and pistosaurs but not for S. gaillardoti and Paludidraco multidentatus. Thus, clear fish-trap dentition piscivory could be interpreted as independently derived in nothosaurs and pistosaurs. Future research on more related taxa, along with additional scans and segmented endocasts across the phylogeny will provide additional, valuable information about sauropterygian phylogenetic relationships and their evolutionary history.

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663	
664	References
665	Agassiz L. 1833-45. Recherches sur les Poissons Fossiles, Vol. I-V [XLIX+188; XII+646;
666	VIII+422; XVI+318; XII+282]. Neuchâtel: Imprimaire de Petitpierre.
667	Allemand R, Houssaye A, Bardet N, and Vincent P. 2019. Endocranial anatomy of
668	plesiosaurians (Reptilia, Plesiosauria) from the Late Cretaceous (Turonian) of Goulmima
669	(southern Morocco). Journal of Vertebrate Paleontology:e1595636 (1595611 pages) [doi:
670	1595610.1591080/02724634.02722019.01595636].
671	Allemand R, Moon BC, and Voeten DFAE. 2023. The paleoneurology of Ichthyopterygia and
672	Sauropterygia: Diverse endocranial anatomies of secondarily aquatic diapsids. In: Dozo
673	MT, Paulina-Carabajal A, Macrini TE, and Walsh S, eds. Paleoneurology of Amniotes.
674	Cham: Springer, 29-77 [https://doi.org/10.1007/1978-1003-1031-13983-13983_13983].
675	Bhullar B-AS, and Bever GS. 2009. An archosaur-like laterosphenoid in early turtles (Reptilia:
676	Pantestudines). Breviora 518:1-11.
677	Cabezuelo-Hernández A, De Miguel Chaves C, and Pérez-García A. 2024. Sauropterygian
678	remains from the Middle Triassic of Araif El-Naqa as the first identification of
679	Simosauridae (Eosauropterygia) in Egypt. Geobios 87:1-7 [doi:
680	10.1016/j.geobios.2024.1008.1015].
681	Chapelle KEJ, and Choiniere JN. 2018. A revised cranial description of Massospondylus
682	carinatus Owen (Dinosauria: Sauropodomorpha) based on computed tomographic scans
683	and a review of cranial characters for basal Sauropodomorpha. PeerJ 6:e4224 [doi:
684	10.7717/peerj.4224].
685	Cheng Y-N, Wu X-c, Sato T, and Shan H-y. 2016. Dawazisaurus brevis, a new eosauropterygian
686	from the Middle Triassic of Yunnan, China. Acta Geologica Sinica (English Edition)
687	90:401-424.



886	Cuthbertson R, Maddin HC, Holmes R, and Anderson JS. 2015. The braincase and endosseous
689	labyrinth of Plioplatecarpus peckensis (Mosasauridae, Plioplatecarpinae), with functional
690	implications for locomotor behaviour. The Anatomical Record 298:1597-1611 [doi:
691	1510.1002/ar.23180].
692	Dalla Vecchia FM. 2008. First record of Simosaurus (Sauropterygia, Nothosauoidea) from the
693	Carnian (Late Triassic) of Italy. Revista Italiana di Paleontologia e Stratigrafia 114:273-
694	285.
695	de Miguel Chaves C, Ortega F, and Pérez-García A. 2018. Cranial variability of the European
696	Middle Triassic sauropterygian Simosaurus gaillardoti. Acta Palaeontologica Polonica
697	63:315-326 [doi: 310.4202/app.00471.02018].
698	de Miguel Chaves C, Ortega F, and Pérez-García A. 2018. New highly pachyostotic
699	nothosauroid interpreted as a filter-feeding Triassic marine reptile. Biology Letters 14:
700	20180130 [doi: 10.1098/rsbl.2018.0130].
701	de Miguel Chaves C, Serrano A, Ortega F, and Pérez-García A. 2020. Braincase and
702	endocranium of the placodont Parahenodus atancensis de Miguel Chaves, Ortega & Pérez
703	García, 2018, a representative of the highly specialized clade Henodontidae. Comptes
704	Rendus Palevol 19:173-186 [doi: 110.5852/cr-palevol2020v5819a5810].
705	Drevermann F. 1933. Die Placodontier. 3. Das Skelett von Placodus gigas Agassiz im
706	Senckenberg-Museum. Abhandlungen der Senckenbergischen Naturforschenden
707	Gesellschaft 38:321-364.
708	Dufeau, DL, and Witmer, LM. 2015. Ontogeny of the middle-ear air-sinus system in Alligator
709	mississippiensis (Archosauria: Crocodylia). PLoS ONE 10(9):e0137060 [doi:
710	10.1371/journal.pone.0137060].
711	Edinger T. 1921. Über Nothosaurus. I. Ein Steinkern der Schädelhöhle. Senckenbergiana 3:121-
712	129.
713	Gardner NM, Holliday CM, and O'Keefe FR. 2010. The braincase of Youngina capensis
714	(Reptilia, Dipsida): new insights from high-resolution CT scanning of the holotype.
715	Palaeontologia Electronica Vol. 13, Issue 3; 19A:16p [http://palaeo-
716	electronica.org/2010_3/217/index.html].
717	Georgi JA, and Sipla JS. 2008. Comparative and functional anatomy of balance in aquatic
718	reptiles and birds. In: Thewissen JGM, and Nummela S, eds. Sensory Evolution on the



746

747

719 Threshold Adaptations in Secondarily Aquatic Vertebrates [ISBN 978-0-520-25278-3]. 720 Berkeley: University of California Press, 233-256. 721 Germain D, and Ladevèze S. 2021. Methodological Focus A. The New Scalpel: Basic Aspects of CT-Scan Imaging. In: Buffrénil Vd, Ricqlès Ad, Zylberberg L, Padian K, Laurin M, and 722 723 Quilhac A, eds. Vertebrate Skeletal Histology and Paleohistology. Boca Raton: CRC Press, 55-58. 724 725 Huene FRv. 1948. Simosaurus and Corosaurus. American Journal of Science 246:41-43. 726 Jenkins XA, Benson RBJ, Ford DP, Browning C, Fernandez V, Griffiths E, Choiniere J, and 727 Peecook BR. 2025. Cranial osteology and neuroanatomy of the late Permian reptile Milleropsis pricei and implications for early reptile evolution. Royal Society Open Science 728 729 12:241298 [doi: 10.1098/rsos.241298]. Jiang D-Y, Motani R, Hao W-C, Rieppel O, Sun Y-L, Schmitz L, and Sun Z-Y. 2008. First 730 731 record of Placodontoidea (Reptilia, Sauropterygia, Placodontia) from the Eastern Tethys. 732 Journal of Vertebrate Paleontology 28:904-908. Jiang D-Y, Motani R, Tintori A, Rieppel O, Chen G-B, Huang J-D, Zhang R, Sun Z-Y, and Ji C. 733 734 2014. The Early Triassic eosauropterygian Majiashanosaurus discocoracoidis, gen. et sp. nov. (Reptilia, Sauropterygia), from Chaohu, Anhui Province, People's Republic of China. 735 736 Journal of Vertebrate Paleontology 34:1044-1052 [doi: 1010.1080/02724634.02722014.02846264]. 737 Kear B, Rich T, Vickers Rich PV, Ali MA, Al-Mufarrih YA, Matari AH, Al-Masary AM, and 738 739 Halawani MA. 2010. A review of aquatic vertebrate remains from the Middle–Upper 740 Triassic Jilh Formation of Saudi Arabia. Proceedings of the Royal Society of Victoria 122:1-8 [doi: 10.1071/RS10001]. 741 742 Kear BP, Engelschiøn VS, Hammer Ø, Roberts AJ, and Hurum JH. 2023. Earliest Triassic ichthyosaur fossils push back oceanic reptile origins. Current Biology 33:R178-R179 [doi: 743 744 110.1016/j.cub.2022.1012.1053]. Kear BP, Roberts AJ, Young G, Terezow M, Mantle DJ, Santos Barros I, and Hurum JH. 2024. 745

Oldest southern sauropterygian reveals early marine reptile globalization. Current Biology

34:R562-R563 [doi: 510.1016/j.cub.2024.1003.1035].

- Klein N, and Griebeler EM. 2018. Growth patterns, sexual dimorphism, and maturation modeled
 in Pachypleurosauria from Middle Triassic of central Europe (Diapsida: Sauropterygia).
- 750 *Fossil Record* 21:137-157 [doi: 110.5194/fr-5121-5137-2018].
- 751 Klein N. 2009. Skull morphology of *Anarosaurus heterodontus* (Reptilia: Sauropterygia:
- Pachypleurosauria) from the lower Muschelkalk of the Germanic Basin (Winterswijk, the
- 753 Netherlands) *Journal of Vertebrate Paleontology* 29:665-676 [doi:
- 754 610.1671/1039.1029.0327].
- 755 Klein N. 2010. Long bone histology of Sauropterygia from the Lower Muschelkalk of the
- Germanic Basin provides unexpected implications for phylogeny. *PLoS ONE* 5(7): e11613
- 757 [doi: 10.1371/journal.pone.0011613].
- 758 Klein N, Houssaye A, Neenan JM, and Scheyer TM. 2015. Long bone histology and
- microanatomy of Placodontia (Diapsida: Sauropterygia). Contributions to Zoology 84:59-
- 760 84 [doi: 10.1163/18759866-08401005].
- 761 Klein N, Sander PM, Krahl A, Scheyer TM, and Houssaye A. 2016. Diverse aquatic adaptations
- in *Nothosaurus* spp. (Sauropterygia)—inferences from humeral histology and
- 763 microanatomy. *PLoS ONE* 11(7):e0158448 [doi: 10.1371/journal.pone.0158448].
- Klein N, Wintrich T, Hagdorn H, Spiller D, Winkelhorst H, Goris G, and Scheyer TM. 2022.
- 765 Placodus (Placodontia, Sauropterygia) dentaries from Winterswijk, The Netherlands
- 766 (middle Anisian) and Hünfeld, Hesse, Germany (late Anisian) with comments on
- ontogenetic changes. *PalZ* 96:289-302 [doi: 210.1007/s12542-12022-00614-w].
- Koken E. 1893. Beiträge zur Kenntnis der Gattung Nothosaurus. Zeitschrift der Deutschen
- 769 *Geologischen Gesellschaft* 45:337-377.
- 770 Kuzmin IT, Boitsova EA, Gombolevskiy VA, Mazur EV, Morozov SP, Sennikov AG, Skutschas
- PP, and Sues H-D. 2021. Braincase anatomy of extant Crocodylia, with new insights into
- the development and evolution of the neurocranium in crocodylomorphs. *Journal of*
- 773 Anatomy 239:983-1038 [doi: 1010.1111/joa.13490].
- Laboury A, Scheyer TM, Klein N, Stubbs TL, and Fischer V. 2023. High phenotypic plasticity at
- the dawn of the sauropterygian radiation. *PeerJ* 11:e15776 [doi: 10.7717/peerj.15776].
- 776 Leyhr, J. 2023. Musculoskeletal Development in Jawed Vertebrates: Gene function, cis-
- regulation, and 3D phenotypes in zebrafish. (PhD Thesis). *Acta Universitatis Upsaliensis*
- 778 [https://www.diva-portal.org/smash/record.jsf?pid=diva2:1789545].



779	Li Q, and Liu J. 2020. An Early Triassic sauropterygian and associated fauna from South China
780	provide insights into Triassic ecosystem health. Communications Biology 3:63 [doi:
781	10.1038/s42003-020-0778-7].
782	Liao J-l, Lan T, Xu G-h, Li J, Qin Y-j, Zaho M-s, Li X-l, and Wang Y. 2021. Tooth structure and
783	replacement of the Triassic Keichousaurus (Sauropterygia, Reptilia) from South China.
784	Frontiers in Ecology and Evolution 9:741851 [doi: 10.3389/fevo.2021.741851].
785	Marquez-Aliaga A, Klein N, Reolid M, Plasencia P, Villena JA, and Martinez-Perez C. 2017. An
786	enigmatic marine reptile, Hispaniasaurus cranioelongatus (gen. et sp. nov.) with
787	nothosauroid affinities from the Ladinian of the Iberian Range (Spain). Historical Biology
788	31:223-233 [doi: 210.1080/08912963.08912017.01359264].
789	Meyer Hv. 1842. Simosaurus, die Stumpfschnauze, ein Saurier aus dem Muschelkalke von
790	Luneville. Neues Jahrbuch für Mineralogie, Geognosie, Geologie und Petrefaktenkunde
791	1842:184-198.
792	Miedema F, Spiekman SNF, Fernandez V, Reumer JWF, and Scheyer TM. 2020. Cranial
793	morphology of the tanystropheid Macrocnemus bassanii unveiled using synchrotron
794	microtomography. Scientific Reports 10: 12412 [doi: 10.1038/s41598-020-68912-4].
795	Neenan JM, and Scheyer TM. 2012. The braincase and inner ear of Placodus gigas
796	(Sauropterygia, Placodontia)—a new reconstruction based on micro-computed
797	tomographic data. Journal of Vertebrate Paleontology 32:1350-1357 [doi:
798	1310.1080/02724634.02722012.02695241].
799	Neenan JM, Klein N, and Scheyer TM. 2013. European origin of placodont marine reptiles and
800	the evolution of crushing dentition in Placodontia. Nature Communications 4:1621 [doi:
801	10.1038/ncomms2633].
802	Neenan JM, Li C, Rieppel O, and Scheyer TM. 2015. The cranial anatomy of Chinese placodonts
803	and the phylogeny of Placodontia (Diapsida: Sauropterygia). Zoological Journal of the
804	Linnean Society 175:415-428 [doi: 410.1111/zoj.12277].
805	Neenan JM, Reich T, Evers SW, Druckenmiller PS, Voeten DFAE, Choiniere JN, Barrett PM,
806	Pierce SE, and Benson RBJ. 2017. Evolution of the sauropterygian labyrinth with
807	increasingly pelagic lifestyles. Current Biology 27:3852-3858 [doi:
808	3810.1016/j.cub.2017.3810.3069].



309	Nopcsa FB. 1928. The genera of reptiles. Geologica hungarica (Ser Palaeontologica), Budapest,
310	Fasc 1 (1928) 1:163-188.
311	Nosotti S, and Rieppel O. 2002. The braincase of <i>Placodus</i> Agassiz, 1833 (Reptilia,
312	Placodontia). Memorie della Società Italiana di Scienze Naturali e del Museo Civico di
313	Storia Naturale di Milano 31:3-18.
314	Owen R. 1860. Palaeontology, or, a systematic summary of extinct animals and their geological
315	relations. Edinburgh: Adam and Charles Black.
316	Pommery Y, Scheyer TM, Neenan JM, Reich T, Fernandez V, Voeten DFAE, Losko AS, and
317	Werneburg I. 2021. Dentition and Feeding in Placodontia: tooth replacement in Henodus
318	chelyops. BMC Ecology and Evolution 21:136 [doi: 10.1186/s12862-021-01835-4].
319	Qiao Y, Liu J, Wolniewicz AS, Iijima M, Shen Y, Wintrich T, Li Q, and Sander PM. 2022. A
320	globally distributed durophagous marine reptile clade supports the rapid recovery of
321	pelagic ecosystems after the Permo-Triassic mass extinction. Communications Biology 5:
322	1242 [doi: 10.1038/s42003-022-04162-6].
323	Rieppel O. 1976. The homology of the laterosphenoid bone in snakes. Herpetologica 32:426-
324	429.
325	Rieppel O. 1994a. Osteology of Simosaurus gaillardoti and the relationships of stem-group
326	Sauropterygia. Fieldiana (Geology), New Series 28:1-85.
327	Rieppel O. 1994b. The braincases of Simosaurus and Nothosaurus: monophyly of the
328	Nothosauridae (Reptilia: Sauropterygia). Journal of Vertebrate Paleontology 14:9-23.
329	Rieppel O. 1995. The genus <i>Placodus</i> : systematics, morphology, paleobiogeography, and
330	paleobiology. Fieldiana: Geology, New Series 31:1-44.
331	Rieppel O. 1999. Phylogeny and paleobiogeography of Triassic Sauropterygia: problems solved
332	and unresolved. Palaeogeography, Palaeoclimatology, Palaeoecology 153:1-15.
333	Rieppel O. 2000. Sauropterygia I - Placodontia, Pachypleurosauria, Nothosauroidea,
334	Pistosauroidea. Handbuch der Paläoherpetologie [Handbook of Paleoherpetology] Part
335	12A:1-134.
336	Rieppel O. 2002. Feeding mechanics in Triassic stem-group sauropterygians: the anatomy of a
337	successful invasion of Mesozoic seas. Zoological Journal of the Linnean Society 135:33-
338	63.



339	Russell DA. 1967. Systematics and morphology of American mosasaurs. <i>Peabody Museum of</i>
340	Natural History Bulletin No. 23:1-241.
341	Sato T, Wu X-C, Tirabasso A, and Bloskie P. 2011. Braincase of a polycotylid plesiosaur
342	(Reptilia: Sauropterygia) from the Upper Cretaceous of Manitoba, Canada. Journal of
343	Vertebrate Paleontology 31:313-329 [doi: 310.1080/02724634.02722011.02550358].
344	Scheyer TM, Romano C, Jenks J, and Bucher H. 2014. Early Triassic marine biotic recovery: the
345	predators' perspective. PLoS ONE 9(3): e88987 [doi: 10.1371/journal.pone.0088987].
346	Scheyer TM, Neenan JM, Bodogan T, Furrer H, Obrist C, and Plamondon M. 2017. A new,
347	exceptionally preserved juvenile specimen of Eusaurosphargis dalsassoi (Diapsida) and
348	implications for Mesozoic marine diapsid phylogeny. Scientific Reports 7: 4406 [doi:
349	10.1038/s41598-017-04514-x].
350	Scheyer TM, Neuman AG, and Brinkman DB. 2019. A large marine eosauropterygian reptile
351	with affinities to nothosauroid diapsids from the Early Triassic of British Columbia,
352	Canada. Acta Palaeontologica Polonica 64:745-755 [doi: 710.4202/app.00599.02019].
353	Scheyer TM, Klein N, Evers SW, Mautner A-K, and Pabst B. 2022. First evidence of
354	Proganochelys quenstedtii (Testudinata) from the Plateosaurus bonebeds (Norian, Late
355	Triassic) of Frick, Canton Aargau, Switzerland. Swiss Journal of Palaeontology 141:17
356	[doi: 10.1186/s13358-022-00260-4].
357	Shang Q-H, Li C, and Wang W. 2022. Nothosaurus luopingensis n. sp. (Sauropterygia) from the
358	Anisian, Middle Triassic of Luoping, Yunnan Province, China. Vertebrata PalAsiatica
359	60:249-270 [doi: 210.19615/j.cnki.12096-19899.220524].
360	Smith KT, Bhullar B-AS, Köhler G, and Habersetzer J. 2018. The only known jawed vertebrate
361	with four eyes and the bauplan of the pineal complex. Current Biology 28 (7): P1101-
362	1107.e2 [doi: 10.1016/j.cub.2018.02.021].
363	Storrs GW. 1993. Function and phylogeny in sauropterygian (Diapsida) evolution. American
364	Journal of Science 293-A:63-90.
365	Sues H-D. 1987. On the skull of <i>Placodus gigas</i> and the relationships of the Placodontia. <i>Journal</i>
366	of Vertebrate Paleontology 7:138-144 [http://www.jstor.org/stable/4523133].
367	Sutton M, Rahman I., and Garwood R. 2016. Virtual paleontology—an overview. <i>The</i>
368	Paleontological Society Papers 22:1-20.



369	Sutton MD. 2008. Tomographic techniques for the study of exceptionally preserved fossils.
370	Proceedings of the Royal Society of London, B 275:1587-1593
371	[doi:1510.1098/rspb.2008.0263].
372	Sutton MD, Rahman IA, and Garwood RJ. (eds.) 2014. Techniques for Virtual Palaeontology.
373	New Analytical Methods in Earth and Environmental Science. Hoboken, New Jersey:
374	Wiley Blackwell. p 200.
375	Voeten DFAE, Reich T, Araújo R, and Scheyer TM. 2018. Synchrotron microtomography of a
376	Nothosaurus marchicus skull informs on nothosaurian physiology and neurosensory
377	adaptations in early Sauropterygia. PLoS ONE 13(1): e0188509 [doi:
378	10.1371/journal.pone.0188509].
379	Voeten DFAE, Albers PCH, and Klein N. 2019. Nothosauroidea from the Vossenveld Formation
380	and their relatives. Grondboor & Hamer Volume 73 nr 5/6 - 2019 - Edition Staringia
381	16:234-244.
382	Wang W, Shang Q, Cheng L, Wu X-C, and Li C. 2022. Ancestral body plan and adaptive
383	radiation of sauropterygian marine reptiles. iScience 25(12):105635 [doi:
384	10.1016/j.isci.2022.105635].
385	Wintrich T, Hayashi S, Houssaye A, Nakajima Y, and Sander PM. 2017. A Triassic
386	plesiosaurian skeleton and bone histology inform on evolution of a unique body plan.
387	Science Advances 3: e1701144 [doi: 10.1126/sciadv.1701144].
388	Witmer LM, Ridgely RC, Dufeau DL, and Semones MC. 2008. Using CT to peer into the past:
389	3D visualization of the brain and ear regions of birds, crocodiles, and nonavian dinosaurs.
390	In: Endo H, and Frey R, eds. Anatomical Imaging Towards a New Morphology [ISBN:
391	978-4-431-76932-3; 105 pp]. Tokyo: Springer, 67-87 [doi: 10.1007/978-4-431-76933-
392	0_6].
393	Wolniewicz A, Shen Y, Li Q, Sun Y, Qiao Y, Chen Y, Hu Y-W, and Liu J. 2023. An armoured
394	marine reptile from the Early Triassic of South China and its phylogenetic and
395	evolutionary implications. eLife 12:e83163 [doi: 10.7554/eLife.83163].
396	Xing L, Klein H, Lockley MG, Wu X-C, Benton MJ, Zeng R, and Romilio A. 2020. Footprints
397	of marine reptiles from the Middle Triassic (Anisian-Ladinian) Guanling Formation of
398	Guizhou Province, southwestern China: The earliest evidence of synchronous style of





899 swimming. Palaeogeography, Palaeoclimatology, Palaeoecology 558: 109943 [doi:

900 10.1016/j.palaeo.2020.109943].



Figure 1

Figure 1: Phylogenetic framework of Sauropterygia and studied specimen.

(A) Hypothesis modified from Allemand et al. (2023 and references therein), reflecting the traditional topology of sauropterygians and *Simosaurus* as sister to other nothosaurs. (B) Hypothesis modified from Li and Liu (2020) in which *Simosaurus* was found being more highly nested within nothosaurs. (C) Hypothesis modified from Wang et al., (2022) showing *Simosaurus* as sister to the clade consisting of nothosaurs and pistosauroids. (D-F) Cranium of *Simosaurus gaillardoti* (SMNS 16363) in dorsal (D), vental (E), and occipital (F) view. Abbreviations: Eo, Eosauropterygia; Eu, Eusauropterygia; No, Nothosauroidea; Pi, Pistosauroidea; Sa, Sauropterygia.

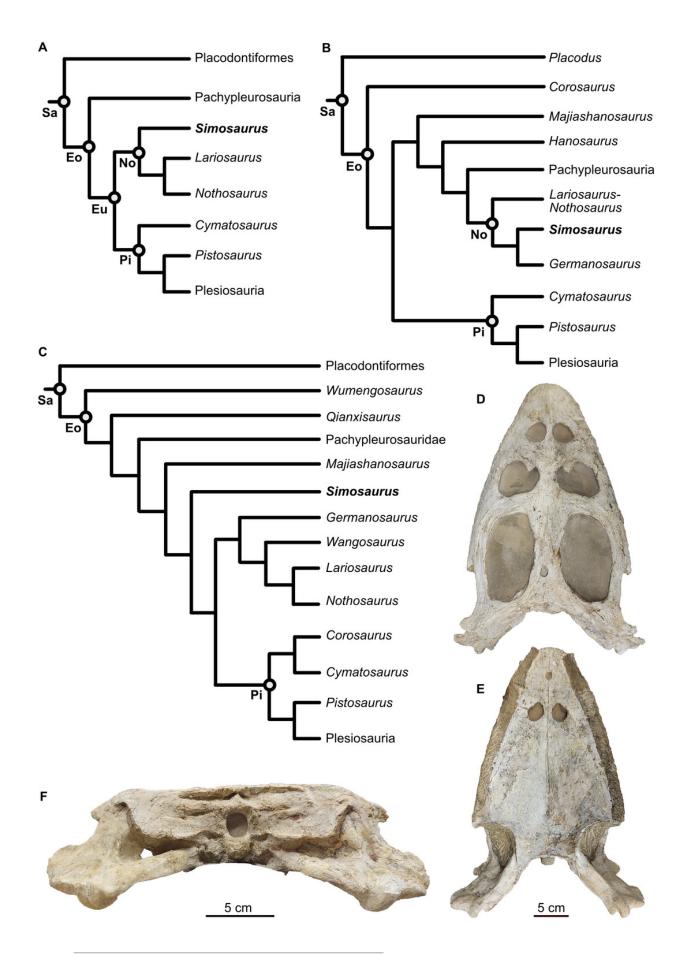
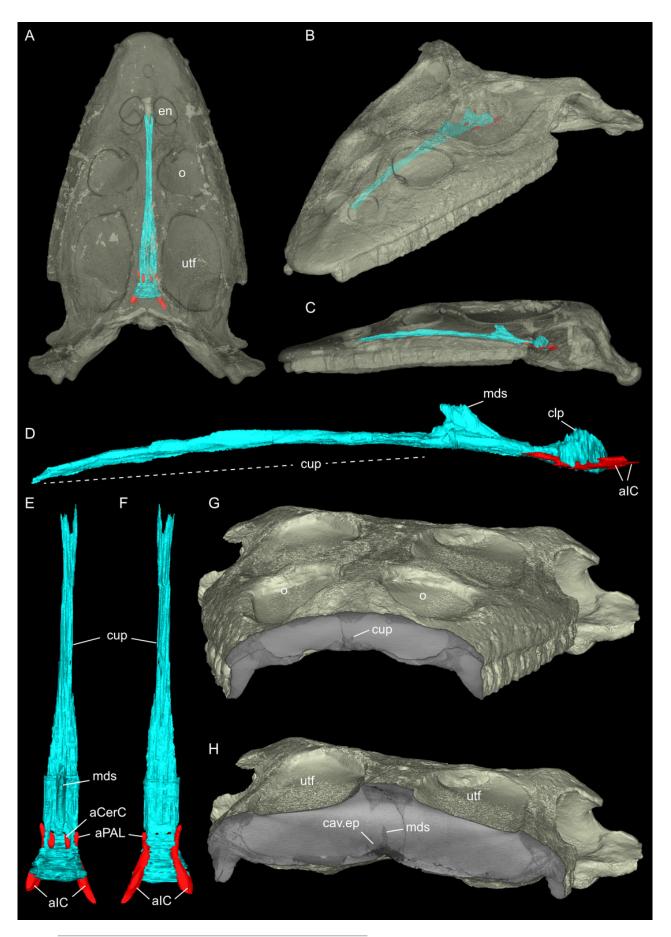




Figure 2: Virtual rendering and partial segmentation of the full skull of *Simosaurus* gaillardoti (SMNS 16363).

(A-C) Partially transparent skull in showing the position and extent of the parabasisphenoid region, in dorsal (A), angled left anterolateral (B), and left lateral (C) view. (D-F) Segmented parabasisphenoid (in turquoise) and associated major blood vessels (in red), in left lateral (D), dorsal (E), and ventral (F) view. (G, H) Angled surface renderings of skull with virtual sections showing the anterior snout (G) and braincase (H) regions in coronal view. Elements not to scale.



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Figure 3: Virtual rendering and segmentation of the full braincase region of *Simosaurus* gaillardoti (SMNS 16363).

Braincase region in dorsal (A), ventral (B), left lateral (C), right lateral with (D) and without cranial endocast and rendered blood vessels (E), and occipital view (F). Elements not to scale.

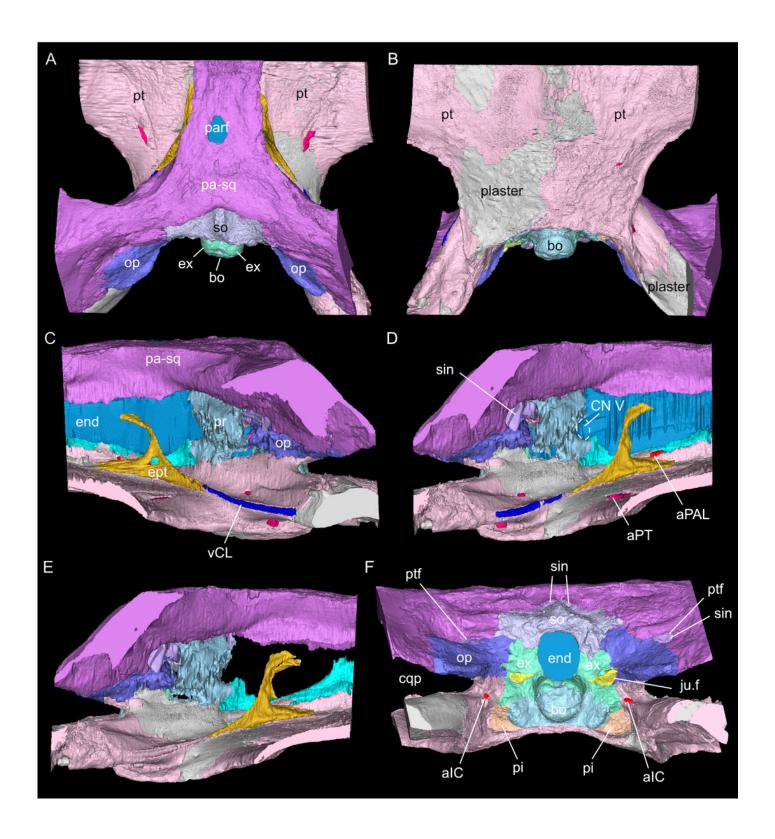
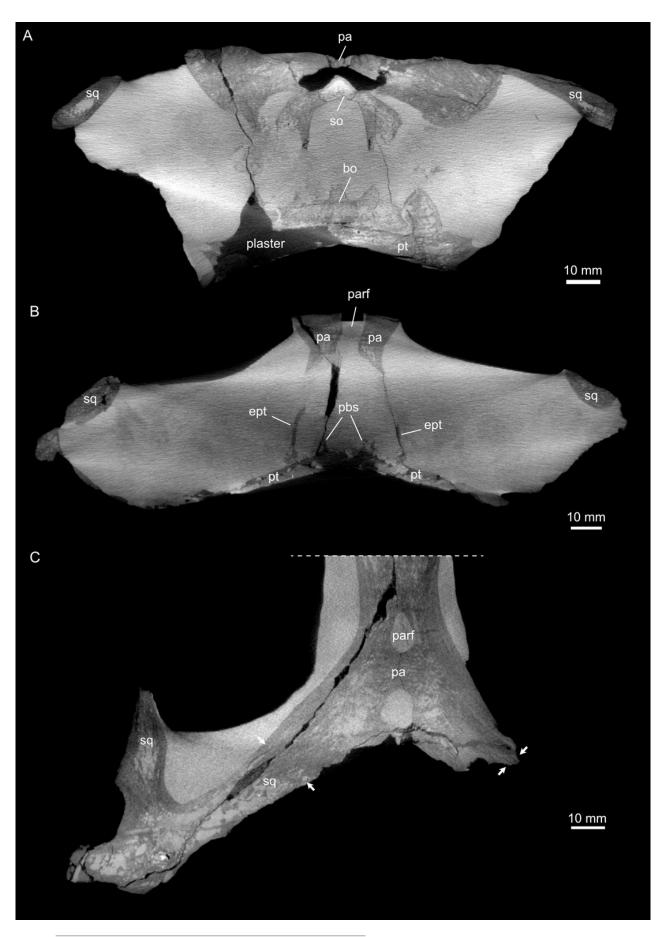




Figure 4: Micro-CT slice data of Simosaurus gaillardoti (SMNS 16363).

(A) Coronal view of braincase region (A) indicating that the bone loss in the specimen occurred mainly due to the breakage of the specimen, as the filled in plaster closely follows the outlines of the preserved bones. (B) Coronal view through braincase at the level of the parietal foramen. The two epipterygoids are clearly visible in articulation with the underlying pterygoids. (C) Axial view of the skull roof bones. Note that the sutures between parietal and the squamosals are only partially visible, potentially indicating incipient fusion of the bones.

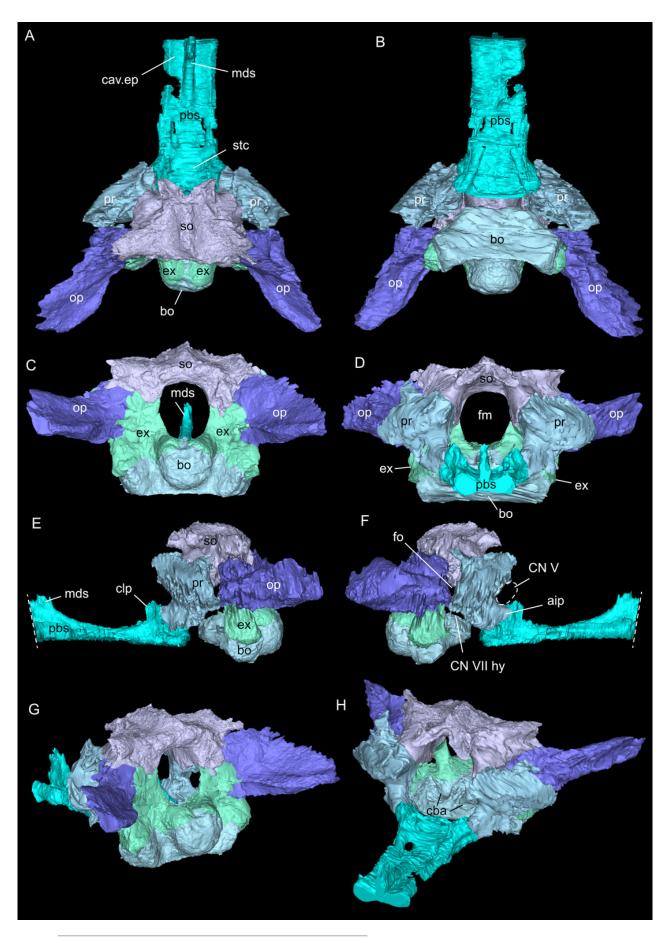


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Figure 5: Virtual rendering and segmentation of bony elements forming the braincase of *Simosaurus gaillardoti* (SMNS 16363).

Braincase in dorsal (A), ventral (B), occipital (C), rostral (D), left lateral (E), right lateral (F), angled right posterolateral (G), and angled left anterolateral view (H). Dashed lines in (E) and (F) indicate anterior delimitation of the parabasisphenoid in this higher resolution-, but spatially limited micro-CT scan. Elements not to scale.



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Figure 6: Virtual rendering and segmentation of the floor of the braincase of *Simosaurus* gaillardoti (SMNS 16363).

Braincase floor elements in dorsal (A), ventral (B), left lateral (C), right lateral (D), angled left anterolateral (E), and dorsal view (F), the latter with segmented blood vessels shown. Elements not to scale.

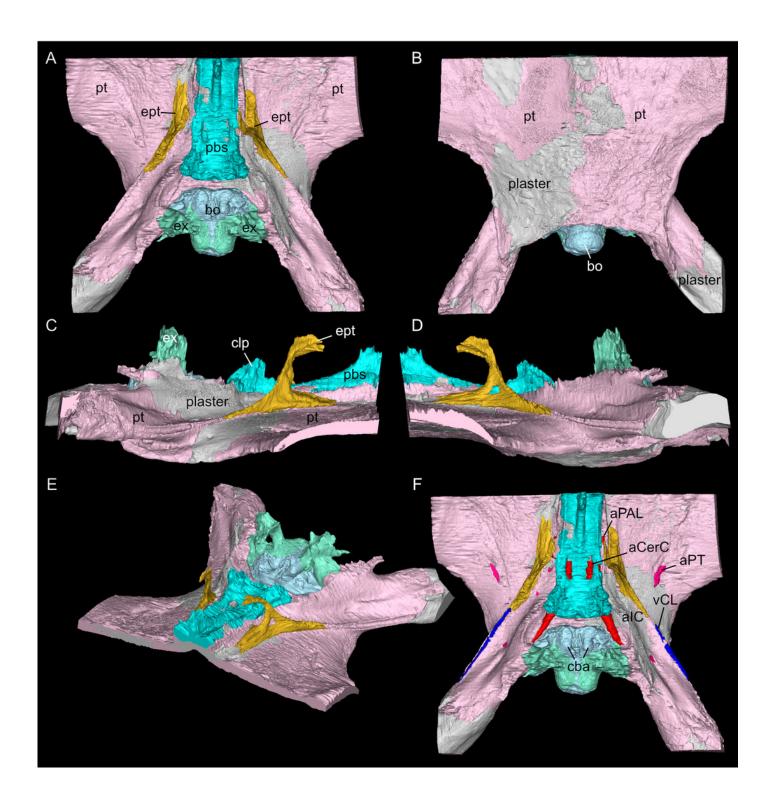




Figure 7: Segmented posterior cranial endocast and casts of blood vessels, nerves and bony labyrinth of *Simosaurus gaillardoti* (SMNS 16363) surrounded by selected transparent braincase bones.

Endocast and selected endocranial voids in dorsal (A), ventral (B), rostral (C), occipital (D), left lateral (E), and angled left posterolateral view (F). Elements not to scale.

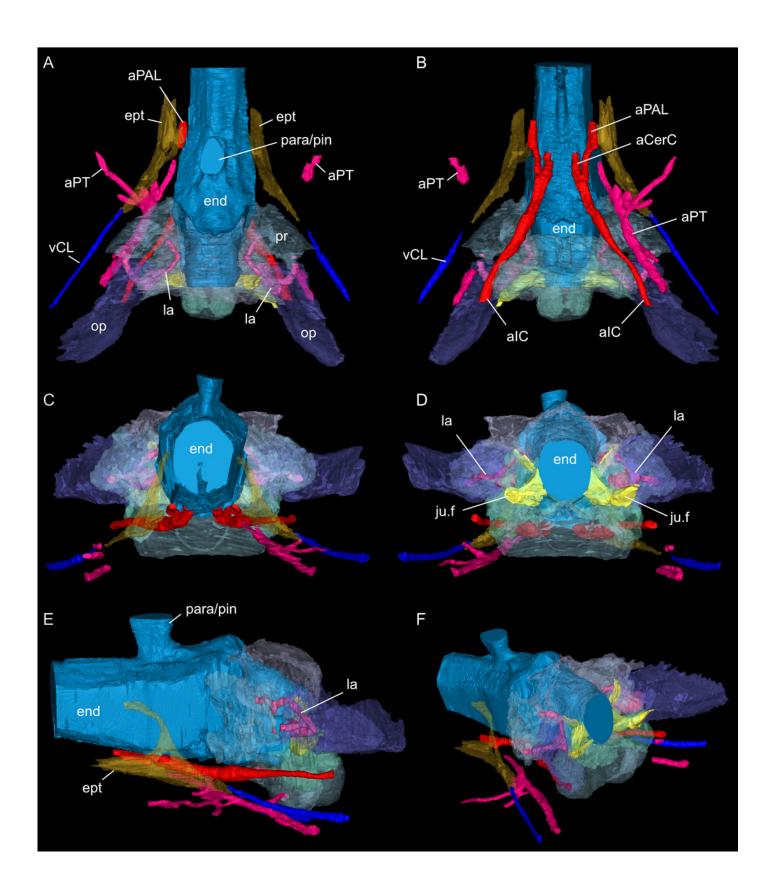




Figure 8: Segmented posterior cranial endocast and selected cranial voids (blood vessels, nerves, bony labyrinth, and part of sinus system) of *Simosaurus gaillardoti* (SMNS 16363).

Endocast and selected endocranial voids in dorsal (A), ventral (B), rostral (C), angled left posterolateral (D) views. (E) Right bony labyrinth in medial view. (F) Isolated cranial endocast in dorsal view. Elements not to scale.

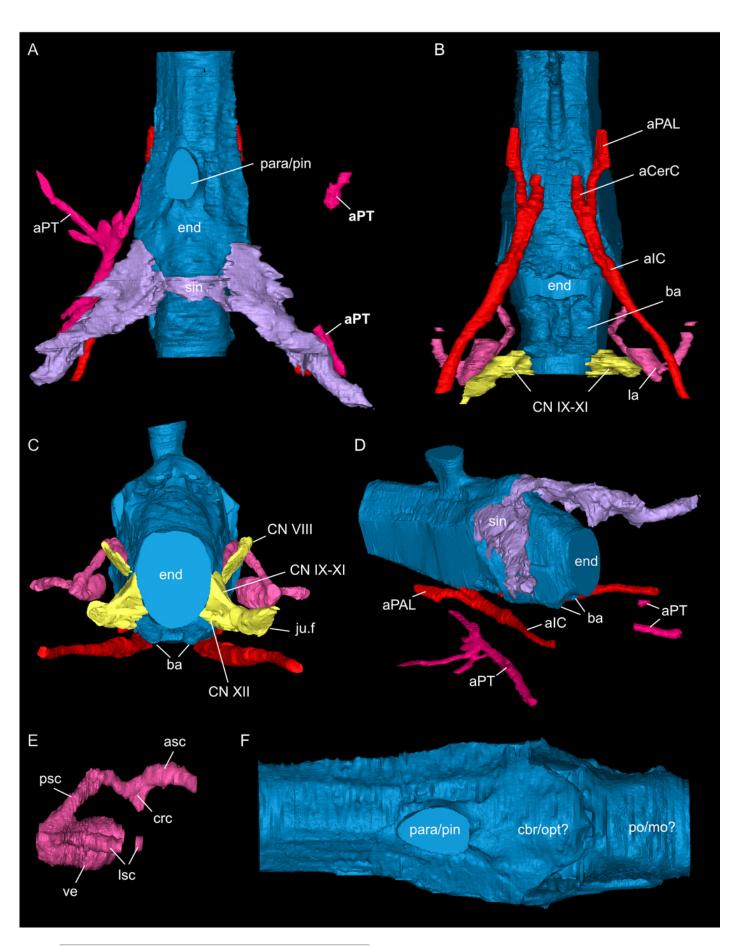




Figure 9: Segmented associated prootic, opisthotic and bony labyrinth (A-D), as well as right epipterygoid (E-J) of *Simosaurus gaillardoti* (SMNS 16363).

Inner ear region in occipital (A), rostral (B), ventral (C), and dorsal view. Right epipterygoid in lateral (E), medial (F), rostral (G), occipital (H), dorsal (I), and ventral (J) view. Elements not to scale.

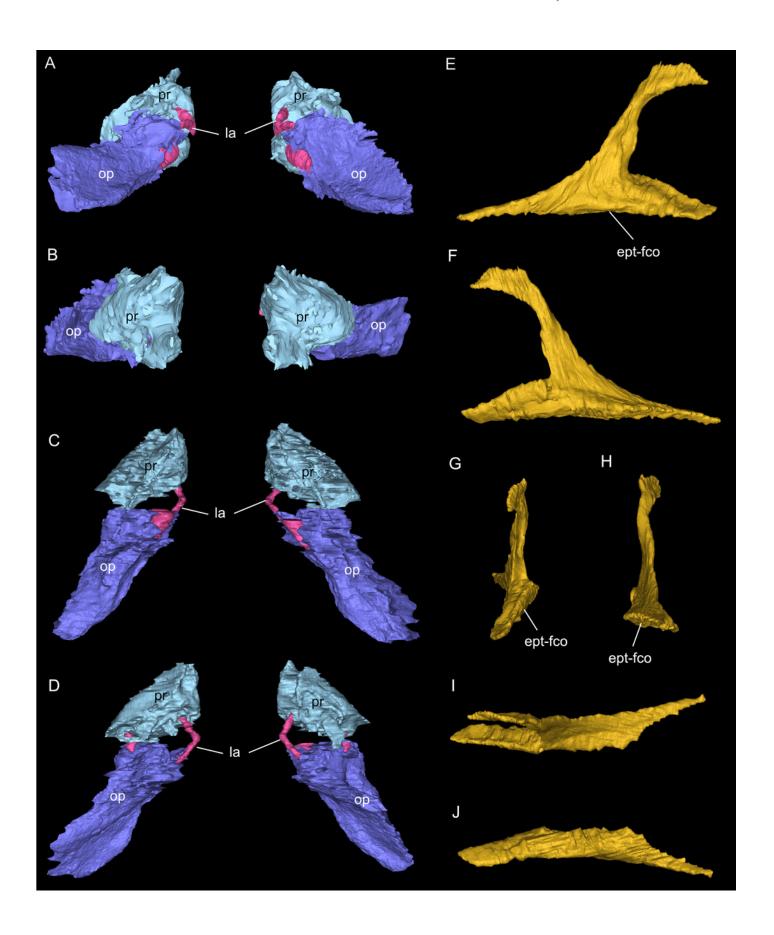




Figure 10: Comparison of segmented sauropterygian endocasts.

(A, B) Simosaurus gaillardoti SMNS 16363; this study; (C, D) Placodus gigas (UMO BT 13; modified after Neenan & Scheyer, 2012); (E, F) Nothosaurus marchicus (TW480000375; modified after Voeten et al., 2018); (G, H) Libonectes morgani (D1-8213, modified after Allemand et al., 2019). Note that the cranial endocast of *P. gigas* was mirrored to reflect the direction of the other endocasts (anterior is towards the left). Images in (A), (C), (E), and (G) show the endocasts in dorsal, images in (B), (D), (F), and (H) in lateral view. Passages for cranial nerves are indicated in yellow, blood vessels in dark red, and the bony labyrinth in lighter red/pink. Elements not to scale.

