

# Extending the fossil record of late Oligocene non-biting midges (Chironomidae, Diptera) of New Zealand (#105715)

1

First submission

## Guidance from your Editor

Please submit by **20 Oct 2024** for the benefit of the authors (and your token reward) .



### Structure and Criteria

Please read the 'Structure and Criteria' page for guidance.



### Custom checks

Make sure you include the custom checks shown below, in your review.



### Author notes

Have you read the author notes on the [guidance page](#)?



### Raw data check

Review the raw data.



### Image check

Check that figures and images have not been inappropriately manipulated.

If this article is published your review will be made public. You can choose whether to sign your review. If uploading a PDF please remove any identifiable information (if you want to remain anonymous).

## Files

Download and review all files from the [materials page](#).

13 Figure file(s)



## Custom checks

### New species checks



Have you checked our [new species policies](#)?



Do you agree that it is a new species?



Is it correctly described e.g. meets ICZN standard?



# Structure and Criteria

## Structure your review

The review form is divided into 5 sections. Please consider these when composing your review:

1. **BASIC REPORTING**
2. **EXPERIMENTAL DESIGN**
3. **VALIDITY OF THE FINDINGS**
4. General comments
5. Confidential notes to the editor

 You can also annotate this PDF and upload it as part of your review

When ready [submit online](#).

## Editorial Criteria

Use these criteria points to structure your review. The full detailed editorial criteria is on your [guidance page](#).

### BASIC REPORTING

-  Clear, unambiguous, professional English language used throughout.
-  Intro & background to show context. Literature well referenced & relevant.
-  Structure conforms to [Peerj standards](#), discipline norm, or improved for clarity.
-  Figures are relevant, high quality, well labelled & described.
-  Raw data supplied (see [Peerj policy](#)).

### EXPERIMENTAL DESIGN

-  Original primary research within [Scope of the journal](#).
-  Research question well defined, relevant & meaningful. It is stated how the research fills an identified knowledge gap.
-  Rigorous investigation performed to a high technical & ethical standard.
-  Methods described with sufficient detail & information to replicate.

### VALIDITY OF THE FINDINGS

-  **Impact and novelty is not assessed.** Meaningful replication encouraged where rationale & benefit to literature is clearly stated.
-  All underlying data have been provided; they are robust, statistically sound, & controlled.
-  Conclusions are well stated, linked to original research question & limited to supporting results.



The best reviewers use these techniques

## Tip

**Support criticisms with evidence from the text or from other sources**

## Example

*Smith et al (J of Methodology, 2005, V3, pp 123) have shown that the analysis you use in Lines 241-250 is not the most appropriate for this situation. Please explain why you used this method.*

**Give specific suggestions on how to improve the manuscript**

*Your introduction needs more detail. I suggest that you improve the description at lines 57- 86 to provide more justification for your study (specifically, you should expand upon the knowledge gap being filled).*

**Comment on language and grammar issues**

*The English language should be improved to ensure that an international audience can clearly understand your text. Some examples where the language could be improved include lines 23, 77, 121, 128 – the current phrasing makes comprehension difficult. I suggest you have a colleague who is proficient in English and familiar with the subject matter review your manuscript, or contact a professional editing service.*

**Organize by importance of the issues, and number your points**

1. Your most important issue
2. The next most important item
3. ...
4. The least important points

**Please provide constructive criticism, and avoid personal opinions**

*I thank you for providing the raw data, however your supplemental files need more descriptive metadata identifiers to be useful to future readers. Although your results are compelling, the data analysis should be improved in the following ways: AA, BB, CC*

**Comment on strengths (as well as weaknesses) of the manuscript**

*I commend the authors for their extensive data set, compiled over many years of detailed fieldwork. In addition, the manuscript is clearly written in professional, unambiguous language. If there is a weakness, it is in the statistical analysis (as I have noted above) which should be improved upon before Acceptance.*

# Extending the fossil record of late Oligocene non-biting midges (Chironomidae, Diptera) of New Zealand

Viktor O Baranov<sup>Corresp., 1</sup>, Jörg U. Hammel<sup>2</sup>, Daphne E. Lee<sup>3</sup>, Alexander R. Schmidt<sup>4</sup>, Uwe Kaulfuss<sup>5</sup>

<sup>1</sup> Estación Biológica de Doñana, Consejo Superior de Investigaciones Científicas, Sevilla, Spain

<sup>2</sup> Institute of Materials Research, Helmholtz Centre, Geesthacht, Germany

<sup>3</sup> Department of Geology, University of Otago, Dunedin, New Zealand

<sup>4</sup> Department of Geobiology, Georg-August Universität Göttingen, Göttingen, Germany

<sup>5</sup> Department of Animal Evolution & Biodiversity, Georg-August Universität Göttingen, Göttingen, Germany

Corresponding Author: Viktor O Baranov

Email address: viktor.baranov@ebd.csic.es

**Background:** The chironomid subfamily Orthocladiinae is diverse and highly endemic in New Zealand, and thought to reflect a complex biogeographical history. This fauna has been important for developing phylogenetic and biogeographic concepts such as Brundin's writings on Transantarctic relationships but the fossil record that would support these reconstructions has been extremely poor. Here we describe the first fossil species of Chironomidae, subfamily Orthocladiinae, from New Zealand, based on inclusions in amber from the late Oligocene Pomahaka Formation of the South Island. **Methods:** We examined newly excavated fossil tree resin (amber) from the late Oligocene Pomahaka Formation in southern New Zealand for inclusions. Amber pieces containing chironomids were prepared and morphologically investigated using light-microscopy and  $\mu$ CT-scanning. Specimens were taxonomically identified based on identification keys for modern adult chironomid midges. Habitus and key morphological features of each specimen were documented photographically and/or by line drawings. **Results:** Thirteen Chironomidae specimens from Pomahaka amber were identified as members of the subfamily Orthocladiinae Kieffer, which hitherto had no fossil record from New Zealand. *Bryophaenocladus zealandiae* sp. nov. Baranov is the first fossil Southern Hemisphere record of the genus *Bryophaenocladus* Thienemann, which is absent from the extant fauna of the main islands of New Zealand but may be present on the subantarctic Auckland Islands. Two specimens described as Morphotype 1 closely resemble *Bryophaenocladus zealandiae* sp. nov. but are significantly larger, either indicating intraspecific size variation or representing a separate taxon close to *B. zealandiae*. *Gymnometriocnemus extinctus* sp. nov. Baranov is the first fossil record of the widespread extant genus *Gymnometriocnemus* Edwards, today present with a single endemic species in New Zealand. The new fossil taxa belong to

chironomid groups requiring terrestrial or semi-aquatic habitats for larval development, which supports the notion of a humid forest swamp paleoenvironment for the Pomahaka amber source forest.

# Extending the fossil record of late Oligocene non-biting midges (Chironomidae, Diptera) of New Zealand

Viktor Baranov<sup>1</sup>, Jörg U. Hammel<sup>2</sup>, Daphne E. Lee<sup>3</sup>, Alexander R. Schmidt<sup>4</sup>, & Uwe Kaulfuss<sup>5</sup>

<sup>1</sup> Estación Biológica de Doñana, Consejo Superior de Investigaciones Científicas, Sevilla, Andalucía, Spain

<sup>2</sup> Institute of Materials Research, Helmholtz-Zentrum Geesthacht, 22607 Hamburg, Germany

<sup>3</sup> Department of Geology, University of Otago, PO Box 56, Dunedin, New Zealand

<sup>4</sup> Department of Geobiology, University of Göttingen, 37077 Göttingen, Germany

<sup>5</sup> Department of Animal Evolution & Biodiversity, University of Göttingen, 37073 Göttingen, Germany

## Abstract

**Background:** The chironomid subfamily Orthoclaadiinae is diverse and highly endemic in New Zealand, and thought to reflect a complex biogeographical history. This fauna has been important for developing phylogenetic and biogeographic concepts such as Brundin's writings on Transantarctic relationships but the fossil record that would support these reconstructions has been extremely poor. Here we describe the first fossil species of Chironomidae, subfamily Orthoclaadiinae, from New Zealand, based on inclusions in amber from the late Oligocene Pomahaka Formation of the South Island.

**Methods:** We examined newly excavated fossil tree resin (amber) from the late Oligocene Pomahaka Formation in southern New Zealand for inclusions. Amber pieces containing chironomids were prepared and morphologically investigated using light-microscopy and  $\mu$ CT-scanning. Specimens were taxonomically identified based on identification keys for modern adult chironomid midges. Habitus and key morphological features of each specimen were documented photographically and/or by line drawings.

**Results:** Thirteen Chironomidae specimens from Pomahaka amber were identified as members of the subfamily Orthoclaadiinae Kieffer, which hitherto had no fossil record from New Zealand. *Bryophaenocladus zealandiae* sp. nov. Baranov is the first fossil Southern Hemisphere record of the genus *Bryophaenocladus* Thienemann, which is absent from the extant fauna of the main islands of New Zealand but may be present on the subantarctic Auckland Island.

specimens described as Morphotype 1 closely resemble *Bryophaenocladus zealandiae* sp. nov. but are significantly larger, either indicating intraspecific size variation or representing a separate taxon close to *B. zealandiae*. *Gymnometriocnemus extinctus* sp. nov. Baranov is the first fossil record of the widespread extant genus *Gymnometriocnemus* Edwards, today present with a single endemic species in New Zealand. The new fossil taxa belong to chironomid groups requiring terrestrial or semi-aquatic habitats for larval development, which supports the notion of a humid forest swamp paleoenvironment for the Pomahaka amber source forest.

**Keywords:** Chironomidae, fossil insects, palaeoecology, Pomahaka Formation, Zealandia

## Introduction

Non-biting midges (Chironomidae) have historically served as a model group for the development of both modern phylogenetic analysis and historical biogeography (Hennig, 1960; Brundin, 1966). Studies of the Chironomidae fauna of New Zealand have played a major role in understanding transantarctic vicariance patterns (Brundin, 1966; Krosch & Cranston, 2013). In particular, phylogenetic studies of the Podonominae, southern Diamesinae and austral Orthoclaadiinae were seminal for understanding vicariance patterns caused by the break-up of Gondwana (Brundin, 1966; Krosch et al., 2011; Krosch & Cranston, 2013). This renders the fossil record of New Zealand's Chironomidae significant to understand biogeographic patterns in the Southern Hemisphere (Schmidt et al., 2018; Baranov, Haug & Kaulfuss, 2024).

However, our knowledge of the fossil history of Chironomidae in New Zealand has been very limited, so far. Schmidt et al. (2018) reported four specimens of *Bryophaenocladus* (Orthoclaadiinae) from Oligocene amber from the South Island, which are included in our descriptions herein. Baranov, Haug & Kaulfuss (2024) described three morphotypes of immature Chironomidae from Early Miocene lake sediments at Foulden Maar in South Island. Subfossil records of Chironomidae include the larvae of *Corynocera duffi* Deevey, 1955 from Holocene swamp deposits in Canterbury, South Island (Deevey, 1955) and numerous other chironomid taxa identified from various Holocene sites on South Island (Schakau, 1991; Woodward & Schulmeister, 2007; Dieffenbacher-Krall et al., 2008). Phylogenetic reconstructions have suggested that some chironomid lineages such as representatives of Podonominae survived a

postulated “Oligocene bottleneck” or hypothetical complete Oligocene submersion of the New Zealand landmass (Cranston et al., 2010).

Considering this limited fossil record, it is difficult to improve our understanding of the evolutionary history of Chironomidae in New Zealand. Thus, any additional deep time records of Chironomidae from New Zealand are of great value. In this study, we describe two new species of non-biting midges (Chironomidae: Orthoclaadiinae) from Oligocene amber from the South Island of New Zealand. These new discoveries extend our knowledge of the past diversity and historical biogeography of Orthoclaadiinae in New Zealand.

## Geological setting

The Chironomidae specimens studied herein are inclusions in amber from the estuarine late Oligocene Pomahaka Formation in southern New Zealand. Fossiliferous amber was collected from a lignite bed and associated carbonaceous mudstone in a temporary excavation pit on private farmland near Pomahaka River approx. 12 km south of Tapanui (46.04450°S, 169.22292°E) (Fig. 1). The locality is registered as G45/f0107 in the New Zealand Fossil Record File (GNS Science & Geological Society of New Zealand, 2024). Fourier-transform infrared spectroscopy analysis of amber from the site indicates an araucarian, *Agathis*-like parent plant, which is supported by finds of araucarian wood and frequent pollen of *Araucariacites australis* Cookson in Pomahaka Formation sediments (Pocknall, 1982; Lee et al., 2009; Kaulfuss et al., 2024). Within the lignite and underlying carbonaceous mudstones, amber is very common and occurs randomly distributed as mm-sized droplets to dm-sized lumps and blocks, showing no signs of sorting and abrasion by reworking and transport. Combined with reconstructions from sedimentological and palynological data (Pocknall, 1982; Lindqvist, Gard & Lee, 2016), this indicates *in situ* resin deposition and amber formation in a domed forest swamp adjacent to brackish mire or saltmarsh within an estuarine paleoenvironment. A comprehensive facies analysis of Pomahaka Formation was published by Lindqvist, Gard & Lee (2016). The late Oligocene age (Chattian, New Zealand stage Duntroonian, 27.3–25.3 ma) of Pomahaka Formation is based on palynomorph and molluscan biostratigraphy (Wood, 1956; Pocknall, 1982; Beu, Maxwell & Brazier, 1990).

## Materials and Methods



# *Material*

Thirteen Chironomidae inclusions from Pomahaka amber were studied herein, including four specimens reported as *Bryophaenocladus* Thienemann or closely related to it by Schmidt et al. (2018) and nine newly discovered specimens. Three of the *Bryophaenocladus* specimens reported by Schmidt et al. (2018) are fossilized in a single piece of amber and a further specimen in a separate piece. The collection number OU35028.2 collectively assigned to all four specimens in Schmidt et al. (2018) is here replaced by individual numbers for each specimen (OU47579–OU47582). The nine new specimens originate from a single amber piece made up of multiple thin layers formed by successive resin flows but were separated and prepared as individual pieces (Figs. 1B, C). The type material and associated specimens are deposited in the Geology Museum of the Geology Department, University of Otago (OU); collection numbers are provided below in the Systematic Paleontology section.

# *Preparation and imaging*

Layered pieces of amber were microscopically examined for biological inclusions and subsequently separated along surfaces of individual resin flows. In instances where this resulted in the exposure of wings at the surface of the amber piece, wings were photographed with a binocular stereomicroscope (Carl Zeiss Stemi 508 with a Canon EOS 70D digital camera) prior to further preparation. Where possible, the thin and brittle, inclusion-bearing amber shards were ground and polished to obtain dorsal, ventral and/or lateral views of inclusions. Polished amber shards, and those too small and fragile for polishing, were embedded in epoxy resin to stabilise specimens, applying the protocol provided by Sadowski et al. (2021). Epoxy-embedded amber pieces were ground and polished using a grinder/polisher machine (Buehler Eco-Met 250) and CarbiMet silicone carbide abrasive papers (CarbiMet) and/or manually using a set of wet silicone carbide abrasive papers (FEPA P #220–4000).

Specimens were studied with a Carl Zeiss AxioScope A1 compound microscope and photographed with a Canon 5D digital camera. Figures were generated with Helicon Focus (8.2.0) software and enhanced using Adobe® Photoshop CC. Line drawings were prepared with [Inkscape software](#).

# *μCT-scanning*

Four Chironomid specimens preserved in close proximity in nearly opaque amber could not be prepared separately and studied by light microscopy. These specimens (OU47579, OU47580, OU47581 and OU47582) were scanned on the Imaging Beamline P05 (Lytaev et al., 2014) operated by the Helmholtz-Zentrum Hereon at the PETRA III storage ring (Deutsches Elektronen Synchrotron - DESY, Hamburg, Germany), using a photon energy of 18 keV and a sample-to-detector distance of 100 mm. Projections were recorded with a custom 20 MP CMOS imaging system with an effective pixel size of 1.28  $\mu\text{m}$  (Lytaev et al., 2014). For each tomographic scan, 3601 projections were recorded at equal intervals between 0 and  $\pi$ . Reconstruction was carried out by applying a transport of intensity phase retrieval approach and using the filtered back projection algorithm (FBP). This workflow was carried out in a custom reconstruction pipeline using Matlab (Math-Works) and the Astra Toolbox (Moosmann et al., 2014; van Aarle et al., 2015, 2016). Raw projections were binned twice for further processing, resulting in an effective pixel size of the reconstructed volume (voxel) of 2.56  $\mu\text{m}$ . Scanned volumes were reconstructed using Drishti ver. 2.6.6 (Limaye, 2012). To decrease the demands for computer memory, we converted all stacks into 8-bit tiffs, downscaled all tiffs by 50% and subsequently cropped the empty space around the amber piece using Fiji ‘scale’ and ‘crop’ functions (Schindelin et al., 2012). Volumes were rendered in Drishti ver. 2.6.6 (Limaye, 2012).

# *Terminology and taxonomy*

Our morphological terminology is based on Sæther (1980) and Marshall et al. (2017). Specimens were identified using the keys provided by Cranston, Oliver & Sæther (1989), Langton & Pinder (2007), Albu (1974), Andersen & Schnell (2000), Armitage (1987), Du, Wang & Sæther (2011), Epler (2012), Freeman (1959), Hazra & Das (2011), Kaczorowska & Gilka (2002), Lin, Qi & Wang (2012), Makarchenko & Makarchenko (2006), Moubayed & Lods-Crozet (2022), Moyubayed & Langton (2023), Pinder & Armitage (1986), Sæther (1973, 1977, 1983), Sublette & Wirth (1980), Wang, Sæther & Andersen (2001), Wang, Liu & Epler (2004), Wang, Andersen & Sæther (2006) and Willassen (1996).

Leg measurements of specimens are mainly approximated values only, due to the difficulty of measuring the variously oriented legs in the amber.

# **Results**

# Systematic paleontology

Order **Diptera** Linnaeus, 1758

Family **Chironomidae** Newman, 1838

Subfamily **Orthoclaadiinae** Kieffer, 1911

Genus ***Bryophaenocladus*** Thienemann, 1934

***Bryophaenocladus zealandiae* sp. nov.**, Baranov

(Figs. 1C, 2–7; Table 1)

**Holotype.** OU47576, adult male, complete specimen in a piece of translucent, yellowish-orange amber with dimensions of  $8 \times 4 \times 0.5$  mm. Head and thorax covered by cloudy coating (“Verlummung”) ventrally and dorsally, and parts of the thorax and abdomen obscured by numerous bubbles (Figs. 1C, 2, 4, 5).

**Paratypes.** OU47540, OU47572 and OU47575, adult males, generally well preserved but some morphology obscured by air bubbles (Fig. 3)

**Associated specimens.** OU47579 in a semi-translucent piece, OU47580, OU47581 and OU47582 together in one nearly opaque piece, mostly obscured by detritus and air bubbles; all adult males (Figs. 6, 7).

**Derivation of name.** The specific epithet refers to the largely submerged continent Zealandia.

**Type locality and horizon.** Temporary lignite pit, site G45/f0107, near Tapanui, southern New Zealand; Pomahaka Formation, late Oligocene (Chattian, New Zealand stage Duntroonian).

**Diagnosis.** The new species can be easily distinguished from any living and fossil *Bryophaenocladus* species based on the combination of the tapering anal point, hyaline at its distal part, and anvil-shaped inferior volsella, together with gonostylus gently curving distally (Andersen & Schnell, 2000; Baranov, Andersen & Hagenlund, 2015; Wang, Sæther & Andersen, 2001; Wang, Liu & Epler, 2004; Wang, Andersen & Sæther, 2006; ; Seredusz and Wichard, 2007).

## Description

**Habitus:** Total length 1.2–1.7 mm, wing length 0.71–0.96 mm ( $n = 5$ ). Overall light yellowish-brown coloration, with thorax and pedicelli darker than the rest of the body.

**Head:** Eyes bare, kidney-shaped, without dorsomedial extension. Palpomeres’ length in  $\mu\text{m}$  ( $n = 2$ , OU47572, OU47575): 2nd: 23, 3rd: 48–60, 4th: 80, 5th: 92–93 (Fig. 2A). Clypeus square,

with at least 8 setae (paratype OU47575). Palpomere three with a possible small distal protrusion, but condition of specimens not permitting corroboration of that. Antennae with 13 flagellomeres, (flagellomeres measurable on holotype only, length in  $\mu\text{m}$ ): F11: 13.766, F12: 16, F13: 25, F14: 25, F15: 19, F16: 21, F17: 21, F18: 17, F19: 15, F110: 24, F111: 15, F112: 23, F113: 166, AR = 0.7.

**Thorax:** Acrostichals ~~setae~~ strong and decumbent, ~~holotype OU47576: 5 visible, paratype OU47540: 8 visible.~~ Dorsocentrals (holotype OU47576) uniserial, at least 7 on each side; **scutellars uniserial**, at least 8. Postnotum bare. Anepisternum and epimeron without leaf-shaped setae.

**Legs:** Leg elements lengths as listed in Table 1. Terminal tarsomeres without pulvilli, shape of all the flagellomeres cylindrical. Foreleg tibial spurs 10–31  $\mu\text{m}$  (n = 2), midlegs without tibial spur, hindtibia with two spurs, short 6–16  $\mu\text{m}$  (n = 3), long 17–31  $\mu\text{m}$  (n = 3) hindtibia comb made of 7–9 (n = 3) setae. Tarsomeres without pseudospurs. Lateral spines compressed to the shaft of the tibial spurs.

Table 1. Length (in  $\mu\text{m}$ ) of leg segments of *Bryophaenocladius zealandiae* sp. nov. Baranov, males (measured on different numbers of specimens, depending on the preservation of the leg elements of the fossil). Values are given as min–max range and mean.

Leg	Femora	Tibia	Ta1	Ta2	Ta3	Ta4	Ta5
Foreleg	255–547 350 (n=5)	255–570 370 (n=5)	136–260 206 (n=3)	58–100 78 (n=3)	50–84 67 (n=3)	36–45 41 (n=3)	50–65 55 (n=3)
Midleg	255–543 400 (n=6)	248–656 374 (n=4)	136–287 202 (n=3)	66–145 96 (n=3)	67–82 76 (n=3)	57–88 70 (n=3)	44–84 63 (n=3)
Hindleg	305–541 419 (n=6)	290–607 454 (n=6)	202–459 286 (n=6)	81–177 131 (n=5)	85–111 100 (n=5)	45–73 57 (n=5)	43–60 52 (n=4)

**Wings:** 0.71–0.96 mm long (n = 5). Anal lobe strongly reduced. Costal extension ca. 85 µm long (n=1). Cu<sub>1</sub> slightly sinuate. Squama fully fringed, with at least 11 setae (n = 1, holotype) (Figs. 3A, B). Wing membranes without macrotrichia, with coarse punctuation.

**Hypopygium:** Anal point hyaline, bare, 23–40 µm long (n = 3), parallel-sided for the most of the length, widening distally. Gonocoxite 60–140 µm long (n = 3), with large, axe-shaped inferior volsella (Figs. 5A–D, 7C–D). Gonostylus gently curved distally, 43 µm long (n = 1, holotype). Megasetae present, crista dorsalis absent (Figs. 5A–D).

# **Taxonomic notes**

The new species is attributed to the genus *Bryophaenocladus* based on the combination of bare eyes, bare wings, fringed squama, ~~anepisternum and epimeron without leaf-shaped setae~~, lateral spines compressed to the shaft of the tibial spurs, pulvilli absent, acrostichal setae strong and decumbent, comb present on the hindtibia, and anal point hyaline and well developed (Cranston, Oliver & Sæther, 1989). In the absence of a modern, comprehensive revision of the genus *Bryophaenocladus* it is difficult to ascertain relations between the new fossil species and other species of *Bryophaenocladus*. The general shape of the hypopigium, particularly the long, distally expanding anal point, is highly reminiscent of *B. beuki*, Baranov, Andersen & Hagenlund, 2015 from Baltic amber (Baranov, Andersen & Hagenlund, 2015). Among extant taxa, the hypopigium of the new species is quite similar to *B. psilacrus* Makarchenko & Makarchenko, 2006 in the axe-shaped inferior volsella, the long, hyaline anal point, and the gently curving gonostylus, without crista dorsalis, as well as to *B. vernalis* (Goetghebuer, 1921) (Brundin, 1956; Makarchenko & Makarchenko, 2006).

# **Morphotype 1 cf. *Bryophaenocladus zealandiae***

(Figs. 1C, 8, Table 2)

**Material.** OU47573 and OU47574, both complete and fairly well visible within yellowish-orange translucent amber.

# **Description**

**Habitus:** Total length 2 mm, wing length 1 mm. Colour: dark brown head and body, and legs of the lighter-brown colour. **Head:** Eyes bare, kidney-shaped, without dorsomedial extension. **Palpomeres'** length in  $\mu\text{m}$  ( $n = 2$ ): 2nd: 26, 3rd: 45–77, 4th: 46–52, 5th: 83–100 (Figs. 7A–D). Clypeus square. Palpomere three is definitely with a conical protrusion on the distal end. **Thorax:** Acrostichals setae strong and decumbent, 8 present ( $n = 2$ ). Dorsocentrals present but difficult to count, uniserial, at least 4. Postnotum bare. Anepisternum and epimeron without leaf-shaped setae. **Legs:** Leg elements lengths as listed in Table 2. Terminal tarsomeres without pulvilli, shape of all the flagellomeres cylindrical. Foreleg tibial spurs  $16\ \mu\text{m}$  ( $n = 2$ ), midlegs without tibial spur, hindtibia with two spurs, short  $27\ \mu\text{m}$  ( $n = 1$ ), long  $30\text{--}46\ \mu\text{m}$  ( $n = 2$ ) hindtibia comb made of  $6\text{--}8$  ( $n = 2$ ) setae. Tarsomeres without pseudospurs. Lateral spines compressed to the shaft of the tibial spurs. **Wings:** 1 mm long ( $n = 2$ ). Details of venation not observable. **Hypopygium:** Only visible in lateral aspect Anal point hyaline, bare, ca  $50\ \mu\text{m}$  long ( $n = 1$ ). Gonocoxite ca.  $150\ \mu\text{m}$  long ( $n = 1$ ).

Table 2. Length (in  $\mu\text{m}$ ) of leg segments of morphotype 1 cf. *Bryophaenocladus zealandiae* (measured on two specimens). Values are given as min–max range and mean.

Leg	Femora	Tibia	Ta1	Ta2	Ta3	Ta4	Ta5
Foreleg	471–518	461–606	325–415	111–115	101–109	72–136	46–79
Midleg	409–511	471–561	178–250	93–144	65–117	33–49	50–51
Hindleg	489–565	530–570	293–340	153–189	95–142	52–79	62–72

### Taxonomic notes

This morphotype is highly reminiscent of *Bryophaenocladus zealandiae* sp. nov., but is 30–50% larger than this new species. It is unclear if this is an indication of naturally large intraspecific size variation or if these two specimens (OU47573, OU47574) represent a different taxon. Should they indeed be members of *Bryophaenocladus zealandiae* sp. nov., this will corroborate that this species has a distal projection on the end of palpomer 3, supporting affinity with the subgenus *Odontocladus* Albu, 1974.

Genus *Gymnometriocnemus* Edwards, 1932  
*Gymnometriocnemus* (*Gymnometriocnemus* s.str.) *extinctus* sp. nov. Baranov

(Figs. 1C, 9–12, Tables 3, 4)

**Holotype.** OU47546; male, well preserved, except for missing thorax, in a piece of semi-translucent, yellow amber ( $7 \times 5 \times 0.5$  mm) with abundant small air-bubbles (Figs. 1C, 9, 10).

**Paratypes.** OU47577, OU47578; females, complete and well visible in semi-translucent pieces of yellow-brown amber (Figs. 1C, 11, 12).

**Derivation of name.** After latin *exstinguō* or *extinguō*, meaning extinct.

**Type locality and horizon.** Temporary lignite pit, site G45/f0107, near Tapanui, southern New Zealand; Pomahaka Formation, late Oligocene (Chattian, New Zealand stage Duntroonian).

**Diagnosis.** This fossil species can be distinguished from the rest of the known species of *Gymnometriocnemus* on the basis of the combination of the following characters: subapical setae of the antenna weak, gonostylus expanding distally, without obvious crista, but with an sub-oval expansion ventrally, megasetae short and sturdy, inferior volsella subrectangular, narrowing distally, strong sclerotized virga absent (Freeman, 1959; Hergstrom, 1974; Sæther, 1969, 1983; Sasa & Kikuchi, 1995; Stur & Ekrem, 2015; Makarchenko & Makarchenko, 2019). Of all species of *Gymnometriocnemus*, *G. (G. st. str.) extinctus* sp. nov. is most reminiscent of *G. (G. st. str.) marionensis* Sæther, 1969, but differs in having much shorter and sturdier megasetae (Sæther, 1969; Stur & Ekrem, 2015).

## Description

### Adult male (OU47546)

**Habitus:** Total length 2.3 mm, wing length 1.5 mm. Colour: dark brown across the parts of the body.

**Head:** Eyes bare, presence of the dorsomedial extension impossible to ascertain. Palpomeres' length in  $\mu\text{m}$  ( $n = 1$ ): 3rd: 115, 4th: 126–152, 5th: 143 (Fig. 8B). Clypeus square with at least 10 setae. Antennae with 13 flagellomeres, (flagellomeres measurable on holotype only, length in  $\mu\text{m}$ ): F11: 27, F12: 30, F13: 19, F14: 31, F15: 30, F16: 31, F17: 25, F18: 30, F19: 27, F110: 28, F111: 28, F112: 31, F113: 360, AR=1.1 Flagellomere 13 with a crown of gentle sensillae and stronger subapical setae (Fig. 9C).

**Thorax:** Most of the thorax, except for anteropronotum and part of the scutum, missing. Strong anteropronotal setae present (at least two), three dorsocentrals visible.



**Legs:** Leg elements lengths as listed in Table 3. Foreleg tibial spurs 30  $\mu\text{m}$  ( $n = 1$ ), presence and number of other spurs impossible to ascertain. Pulvilli absent.

Table 3. Length (in  $\mu\text{m}$ ) of leg segments of *Gymnometriocnemus extinctus* sp. nov. Baranov, male holotype OU47546.

Leg	Femora	Tibia	Ta1	Ta2	Ta3	Ta4	Ta5
Foreleg	704	741	423	175	222	161	217
Midleg	596	520	468	124	76	-	-
Hindleg	615	783	-	-	-	-	-

**Wings:** 1.5 mm long ( $n = 1$ ). Wing membrane densely covered with macrotrichia. Cu<sub>1</sub> slightly sinuate, costal extension pronounced. Squama apparently bare (Figs. 10A–B).

**Hypopygium:** With numerous long setae, gonocoxite 114  $\mu\text{m}$  long ( $n = 1$ ). Gonostylus ca. 70  $\mu\text{m}$  long ( $n = 1$ ), expanding distally, without obvious crista, but with sub-oval expansion ventrally, megasetae short and sturdy. Anal point short, cresting top of tergite IX, with several long setae (Figs. 10C–D). Inferior volsella subrectangular, narrowing distally (Figs. 10C–D). Presence of virga impossible to ascertain, but since hypopygium is partially transparent, we can rule out presence of extremely strong and sclerotized virga.

**Adult female** (OU47577, OU47578), tentatively associated with male on the basis of the similarity of the wing structure and setation of the anteropronotum

**Habitus:** Total length 0.9–1.0 mm, wing length 0.63–0.72 mm. Colour: similar to the male holotype.

**Head:** Eyes bare, reniform. Palpomeres length in  $\mu\text{m}$  ( $n = 2$ ): 2nd: 29 ( $n = 1$ , OU47578), 3rd: 40–41, 4th: 41–42, 5th: 75–82 (Figs. 12A–B). Clypeus square with at least 11 setae. Antennae with 5 flagellomeres, ( $n = 2$ , length in  $\mu\text{m}$ ): Fl1: 76 ( $n = 1$ , OU47577), Fl2: 19–29, Fl3: 26–28, Fl4: 22–27, Fl5: 39–40. Flagellomere 5 with a weak but distinct subapical seta (Figs. 12A–B). Pedicellus cup-shaped.

**Thorax:** Acrostichals setae strong and decumbent, 5–8 ( $n = 2$ ). Dorsocentrals biserial, upper row 5, lower row 8. Postnotum bare. Anteropronotum 4. Anepisternum and epimeron without leaf-shaped setae. Prealars 3, humerals 4. Scuterals uniserial, 6.



**Legs:** Leg elements lengths as listed in Table 4. Foreleg tibial spurs 14–15  $\mu\text{m}$  ( $n = 2$ ), midtibial spur 11–15, hindtibial spur 26–37 (length in  $\mu\text{m}$ ). Hindtibial comb made of 8–9 strong setae. Pulvilli absent, empodium feathery.

Table 4. Length (in  $\mu\text{m}$ ) of leg segments of *Gymnometriocnemus extinctus* sp. nov. Baranov, female paratypes (measured on different numbers of specimens, depending on the preservation of leg elements of the fossil). Values are given as min–max range.

Leg	Femora	Tibia	Ta1	Ta2	Ta3	Ta4	Ta5
Foreleg	312	308	109–139	56–70	48–50	28–34	26–46
Midleg	238–282	259–266	92–100	39–49	33–34	23–29	38–47
Hindleg	267–269	211–296	145–171	55–69	70–89	37–40	42–50

**Wing:** 0.63–0.72 mm long ( $n = 2$ ). Wing membrane densely covered with macrotrichia.  $\text{Cu}_1$  slightly sinuate. Squama bare, costal extension pronounced (Fig. 11C). Wing very similar to male holotype (OU47546). Halteres dark-brown in their entirety.

**Female genitalia:** Cerci very small, gonapophysis VIII divided into small mesal lobe and narrow dorsomesal lobe (Figs. 12C–D). Gonocoxite relatively small, with at least 5 strong setae. Tergite IX rounded, undivided (Figs. 12C–D).

# Taxonomic notes

This species is attributed to the genus *Gymnometriocnemus* (subgenus *Gymnometriocnemus*) based on the combination of bare eyes, apical flagellomere with subapical sensillae, wing covered with macrotrichia with pronounced costal extension and bare squama, anteropronotals present, pulvilli absent, presence of crest-like anal point on tergite IX and overall extremely high density of setae on the body (Freeman, 1959; Sæther, 1983; Cranston, Oliver & Sæther, 1989; Stur & Ekrem, 2015). Since the hypopigium and Tergite VIII are partially translucent in the male specimen, and the extremely large and sclerotized virga is not visible, we find its presence unlikely. Since a large and sclerotized virga does not appear to be present, we propose that the new species is a representative of the subgenus *Gymnometriocnemus* s. str. Edwards, 1932 rather than of *Raphidocladus* Sæther, 1983, as the synapomorphy of the latter is the presence of a strong virga (Sæther, 1983; Stur & Ekrem, 2015). This new species can be differentiated from the New Zealand endemic genus *Pterosis* Sublette & Wirth, 1980, which closely resembles

*Gymnometriocnemus*, by the subapical seta, characteristic for *Gymnometriocnemus* but absent in *Pterosis* (Sublette & Wirth, 1980; Sæther, 1983; Stur & Ekrem, 2015). We tentatively associate two females (OU47577, OU47578) with the male holotype OU47546 on the basis of matching wing venation and setation patterns, patterns of anteropronotal setation and general congruence of the female genital morphology with the genus *Gymnometriocnemus* (Sæther, 1977).

## Discussion

### *Faunal affinities and biogeography*

Chironomids as a group are fairly old, with the oldest representatives occurring in the uppermost Triassic of Europe (203 mya) (Ansorge, 1999), although based on dated phylogenies the group is likely significantly older, at least 250 mya (Cranston et al., 2011). The oldest Orthoclaadiinae fossils are known from Lower Cretaceous Lebanese amber (Veltz, Azar & Nel, 2007). A long geological history and rich fossil record has made Chironomids a suitable model group for historical biogeographic analyses. Following Willi Hennig's work on phylogenetic systematic (Hennig, 1959), Lars Brundin became interested in applying principles of cladistic analysis and an emerging understanding of plate tectonics to the analysis of Chironomidae distribution in the Southern Hemisphere (Brundin, 1966). Brundin came to the conclusion that the majority of Chironomidae distribution patterns in Australia, Southern Neotropics and New Zealand can be explained by the break-up of Gondwana (Brundin, 1966). Since then, however, our understanding of the assembly of New Zealand's biota has become more nuanced. In particular the role of dispersal has become more widely appreciated (e.g., Trewick, 2000; Sanmartin et al., 2001).

The composition of the New Zealand Orthoclaadiinae fauna, and Chironomidae in general, reflects a complex history influenced by both trans-Tasman and trans-Antarctic dispersal and vicariance following the break-up of Gondwana (Krosch & Cranston, 2013; Krosch et al., 2011, 2015). The two new species from Pomahaka amber are among the first fossil records of Orthoclaadiinae from New Zealand and allow a valuable but very limited first glimpse of the late Oligocene fauna.

*Bryophaenocladus* has a near worldwide distribution but it appears to be absent from Australia (but larvae have been reported from orchards in western Australia; Cranston, 1996) and New Zealand. When we first recorded *Bryophaenocladus* from New Zealand Oligocene amber

(Schmidt et al., 2018), we noted that there are no formal records of this genus from the extant fauna of New Zealand (Boothroyd & Forsyth, 2011; Ashe & O'Connor, 2012). We also noted that the BOLD V4 system has barcoding records of *Bryophaenocladus* in New Zealand (Schmidt et al., 2018) but, on closer examination, these belong to the two BOLD BINs BOLD:AAM6273 and BOLD:AAG1021. Representatives of these BINs all cluster around the Holarctic species *Bryophaenocladus ictericus* (Meigen, 1830). It is likely that this species has been historically introduced to New Zealand with agricultural produce, as *Bryophaenocladus* larvae are associated with agricultural plants (Cranston, 1987).

While there appear to be no native species of *Bryophaenocladus* on the main islands of New Zealand, it is highly likely that the monotypic *Kuschelius dentifer* Sublette & Wirth, 1980, endemic to the sub-Antarctic Auckland Islands, is in fact a species of *Bryophaenocladus*. Sublette & Wirth (1980) erected the genus *Kuschelius* as intermediate between *Chaetocladus* Kieffer and *Bryophaenocladus*, and distinguished *K. dentifer* from species of *Bryophaenocladus* by the presence of apical setae on the terminal flagellomere of the antenna and slightly diverted spines on the tibial spur of the hind leg (Fig 13 A-E). However, these characters in combination with the structure of the hypopigium and the presence of the distal projection on the distal end of palpomere three fit well within the current definition of *Bryophaenocladus*, subgenus *Odontocladus* Albu, 1974 (Albu, 1974; Armitage, 1987; Moubayed & Langton, 2023). As pointed out by Sæther (1982) and Armitage (1987), *K. dentifer* is almost certainly a *Bryophaenocladus*, very similar to *B. brincki* (Freeman, 1955) originally described from South Africa. Molecular data on *K. dentifer* are not yet available. *Bryophaenocladus zealandiae* sp. nov. Baranov from Pomahaka amber now records the genus *Bryophaenocladus* in Zealandia in the late Oligocene (~26 mya) and documents its post Oligocene extinction in New Zealand, at least on the main islands. The only two other fossils of *Bryophaenocladus* are from Eocene Baltic amber. A modern review of *Bryophaenocladus* and additional fossils are needed to decipher the biogeographic history of this genus.

The genus *Gymnometriocnemus* identified here from Pomahaka amber includes 17 extant species in all major biogeographical regions except Antarctica, with the actual number of global species expected to be much higher (Stur & Ekrem, 2015). Subgenus *Gymnometriocnemus*, to which the new fossil species from Pomahaka amber is assigned, includes 11 species, with *G.*

*wilsoni* Freeman, 1961 occurring in Australia and a single species, the endemic *G. lobifer* (Freeman, 1959) (sometimes misspelled as *Gymnometriocnemis lobifer*) occurring in New Zealand on both the North and South Islands (Ashe & O'Connor, 2012). *Gymnometriocnemus* (*G.s.str.*) *extinctus* sp. nov. Baranov described here from the late Oligocene is the first fossil record of this genus globally. Its discovery in amber from the Pomahaka Formation indicates a long presence of *Gymnometriocnemus* in New Zealand.

#### *Paleoecology of non-biting midges from Pomahaka amber*

It is notable that both newly discovered species of midges from Pomahaka amber belong to Chironomidae groups whose extant representatives have larvae that develop mostly in terrestrial and semi-aquatic habitats (Moller Pillot, 2013), unlike the aquatic habitat seen in larvae of most other chironomids. Larvae of both *Bryophaenocladus* and *Gymnometriocnemus* develop in wet mosses or decaying leaves or similar wet habitats (Strenzke, 1957; Moller Pillot, 2013). Terrestrial and semi-terrestrial Chironomidae are relatively common in various amber deposits worldwide, probably due to their association with mosses and other microhabitats at the bark of the resin-producing trees or on the nearby forest floor (Solórzano-Kraemer et al., 2018). There are two fossil species of *Bryophaenocladus*, *B. beuki* Baranov, Andersen & Hagenlund, 2015 and *B. circumclusus* Seredusz & Wichard, 2007 and a probable larva of this genus (Baranov et al., 2019), all from Eocene Baltic amber. Until now no fossil *Gymnometriocnemus* were known but there are numerous other fossil representatives of Chironomidae whose extant representatives develop in terrestrial habitats, such as *Parametriocnemus*, *Paraphaeocladus*, *Pseudorthocladus*, *Smittia* and *Pseudosmittia* (Zelentsov et al., 2012; Baranov, Andersen & Hagenlund, 2015). High prevalence of the Chironomids with terrestrial larvae in certain amber deposits probably reflects high humidity in amber forest habitats, as relatively high and constant humidity is required by these groups of Chironomids to finish larval development (Strenzke, 1957; Armitage, Cranston & Pinder, 1995; Zelentsov et al., 2012). The finding of terrestrial or semi-aquatic midges in Pomahaka amber is consistent with the paleo-environmental reconstruction. The amber-bearing lignites of Pomahaka Formation formed by in-situ growth and decomposition of wetland forest trees and litter in domed forest swamps (Lindqvist, Gard & Lee, 2016) and the palynomorph assemblage from the lignites includes ferns, shrubs, herbs and reeds associated with moist and

damp habitats and overall indicates high humidity and high rainfall throughout the year (Pocknall, 1982).

## Acknowledgements

We sincerely thank Andrew Morris for allowing access to his property and for generous help with excavating Pomahaka amber. The Geology Department, University of Otago is acknowledged for help with fieldwork logistics and for curating fossil specimens in the collections of the Geology Museum. We are grateful to Dr. Leanne Elder, New Zealand Arthropod Collection, Manaaki Whenua - Landcare Research, for taking photos of *Kuchelius dentifer*, and allowing us to use those photos in this publication. We thank the reviewers xxxxx.

## Funding

Our results build on the earlier project “Captured in amber: ecological complexity in New Zealand’s ancient araucarian forests”, funded by a Marsden Grant (UOO1416) by the Royal Society of New Zealand. This study was funded by the Spanish State Agency for Innovation’s Ramon y Cajal fellowship (RyC2021-032144-I), project title “Climate change in the past and present & Insect decline” (to Viktor Baranov) and by the German Research Foundation (DFG project 429296833), project title “Palaeontology, biogeography and evolution of New Zealand insects”. Open Access Publication Support Initiative from the CSIC Unit of Information Resources for Research (URICI). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

## Competing Interests

The authors declare that they have no competing interests.

## Author Contributions

Viktor Baranov conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.

Joerg Hammel conducted  $\mu$ CT scanning, analysed related data, authored or reviewed drafts of the article, and approved the final draft.

Daphne E. Lee authored or reviewed drafts of the article, and approved the final draft.

Alexander R. Schmidt discovered and prepared some of the amber fossils and commented on the manuscript, conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.

Uwe Kaulfuss conducted fieldwork, prepared some of the amber fossils, conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.

# **Data Availability**

The following information was supplied regarding the data availability: All specimens included in this work are housed at the Geology Museum of the Geology Department, University of Otago (OU), under accession numbers: *Bryophaenocladus zealandiae* sp. nov. Baranov, OU47576 (holotype); OU47540, OU47572, OU47573, OU47574, OU47575 (paratypes); OU47579, OU47580, OU47581, OU47582 (associated material). Morphotype 1, cf. *Bryophaenocladus*, OU47573, OU47574. *Gymnometriocnemus (G.) extinctus* sp. nov. Baranov, OU47546 (holotype); OU47577, OU47578 (paratypes)

# **References**

**Albu P. 1974.** A new subgenus of the genus *Bryophaenocladus* and two new species (Diptera, Chironomidae). In: Brundin L, ed. *Proceedings 5th International Symposium on Chironomidae, Abisko, 7-9 August 1973. Entomologisk Tidskrift Suppl.* **95**:9–12.

**Andersen T, Schnell, ØA. 2000.** New Species of *Bryophaenocladus* Thienemann, 1934 from Tanzania, with Bare Squama (Diptera: Chironomidae), *Aquatic Insects* **22(1)**:48–57.

**Ansorge J. 1999.** *Aenne liasina* gen. et sp. n. – the most primitive non biting midge (Diptera: Chironomidae: Aenneinae subfam. n.) – from the Lower Jurassic of Germany. *Polish Journal of Entomology* **68**:431–443.



- 500 **Armitage PD. 1987.** A new species of the genus *Bryophaenocladus* Thienemann, (Diptera:
- 501 Chironomidae) from Tenerife, Canary Islands. *Aquatic Insects* **9(1)**:33–38.
- 502 **Armitage PD, Cranston PS, Pinder LCV (Eds). 1995.** *The Chironomidae*. London: Biology
- 503 and Ecology of Non-biting Midges. Chapman & Hall, 572.
- 504 **Ashe P, O'Connor JP. 2012.** *A world catalogue of Chironomidae (Diptera). Part 2.*
- 505 *Orthoclaadiinae. Two volumes (Sections A, B).* Dublin: Irish Biogeographical Society, 468 + 500.
- 506 **Baranov V, Andersen T, Hagenlund L. 2015.** A new species of *Bryophaenocladus*
- 507 Thienemann, 1934 (Diptera, Chironomidae, Orthoclaadiinae) from Baltic amber. *Norwegian*
- 508 *Journal of Entomology* **62(1)**:53–56.
- 509 **Baranov V, Hoffeins C, Hoffeins HW, Haug JT. 2019.** More than dead males: reconstructing
- 510 the ontogenetic series of terrestrial non-biting midges from the Eocene amber forest. *Bulletin of*
- 511 *Geosciences* **94(2)**:1–13.
- 512 **Baranov VO, Haug JT, Kaulfuss U. 2024.** New records of immature aquatic Diptera from the
- 513 Foulden Maar Fossil-Lagerstätte, New Zealand, and their biogeographic implications. *PeerJ*
- 514 **12**:e17014
- 515 **Beu AG, Maxwell PA, Brazier RC. 1990.** Cenozoic Mollusca of New Zealand. *New Zealand*
- 516 *Geological Survey Paleontological Bulletin* **58**:1–518.
- 517 **Boothroyd I, Forsyth D. 2011.** Checklist of New Zealand Chironomidae (Diptera). National
- 518 Centre for Aquatic Biodiversity and Biosecurity 1–7. Available at
- 519 <https://niwa.co.nz/sites/niwa.co.nz/files/import/attachments/chirolist.pdf> (accessed 3 August
- 520 2023)
- 521
- 522 **Brundin L. 1956.** Zur Systematik der Orthoclaadiinae (Diptera, Chironomidae). Report of the
- 523 Institute of Freshwater Research, Drottingholm, No. 37: 5–185
- 524 **Brundin L. 1966.** Transantarctic relationships and their significance as evidenced by chironomid
- 525 midges. *Kungliga Svenska Vetenskapsakademiens Handlingar* **4(1)**:1–472
- 526 **Cranston PS. 1987.** A non-biting midge (Diptera: Chironomidae) of horticultural significance.
- 527 *Bulletin of Entomological Research* **77(4)**:661–667
- 528 **Cranston PS. 1996.** *Identification Guide to the Chironomidae of New South Wales. AWR*
- 529 *Identification Guide Number 1*. West Ryde: Australian Water Technologies Pty Ltd, 376.
- 530 **Cranston PS, Oliver DR, Sæther OA. 1989.** The adult males of Orthoclaadiinae (Diptera:
- 531 Chironomidae) of the Holarctic region - Keys and diagnoses. In: Wiederholm T, ed.

- 532 *Chironomidae of the Holarctic region. Keys and diagnoses. Part 3. Adult males. Entomologica*
- 533 *Scandinavica Supplement*. Vol. 34: Scandinavian Entomology, 165–352.
- 534 **Cranston PS, Hardy NB, Morse GE, Puslednik L, McCluen SR. 2010.** When molecules and
- 535 morphology concur: the ‘Gondwanan’ midges (Diptera: Chironomidae). *Systematic Entomology*
- 536 **35(4):**636–648
- 537 **Deevey ES. 1955.** Paleolimnology of the upper swamp deposit, Pyramid Valley. *Records of the*
- 538 *Canterbury Museum* **6(4):**291–344
- 539 **Dieffenbacher-Krall AC, Vandergoes MJ, Woodward CA, Boothroyd IK. 2008.** Guide to
- 540 identification and ecology of New Zealand subfossil chironomids found in lake sediment.
- 541 Climate Change Institute, University of Maine, Orono 1–11. Available at
- 542 <http://www.climatechange.umaine.edu/Research/facilities/perl/nzguide.html> (accessed 1 June
- 543 2024)
- 544 **Du J, Wang XH, Saether O. 2011.** Redescriptions of species of *Bryophaenocladus*
- 545 Thienemann, 1934 (Diptera: Chironomidae) described by Brundin (1947). *Zootaxa* **2743(1):**40–
- 546 48
- 547 **Epler JH. 2012.** A brachypterous *Bryophaenocladus* (Diptera: Chironomidae: Orthoclaadiinae)
- 548 with hypopygium inversum from Heggie’s Rock, Georgia, USA. *Zootaxa* **3355(1):**51–61
- 549 **Freeman P. 1955.** Diptera (Nematocera): Chironomidae. In: Hanström B, Brinck P, Rudebeck G
- 550 eds. *South African animal life, volume II*. Stockholm: Almquist & Wiksell, 361–381.
- 551 **Freeman P. 1959.** A study of the New Zealand Chironomidae (Diptera, Nematocera). *Bulletin*
- 552 *British Museum (Natural History), Entomology* **7(9):**395–437
- 553 **Freeman P. 1961.** The Chironomidae (Diptera) of Australia. *Australian Journal of Zoology* **9(4):**
- 554 611–738
- 555 **GNS Science & Geological Society of New Zealand. 2024.** New Zealand Fossil Record File
- 556 [G43/f8500, G45/f0107]. GNS Science. <https://doi.org/10.21420/JQQB-NK89>
- 557 **Hazra N, Das N. 2011.** A new species of *Bryophaenocladus* Thienemann, 1934 (Diptera:
- 558 Chironomidae) from Darjeeling Himalayas, India. *International Journal of Dipterological*
- 559 *Research* **22(3):**139–143
- 560 **Hennig W. 1960.** Die Dipteren-Fauna von Neuseeland als systematisches und
- 561 tiergeographisches Problem. *Beiträge zur Entomologie (= Contributions to Entomology)* **10(3–**
- 562 **4):**221–329



- Hergstrom IA. 1974.** The taxonomy and general biology of some southern Australian Chironomidae (Diptera: Nematocera). PhD thesis. The University of Adelaide, 224.
- Kaczorowska E, Gilka W. 2002.** The first record of *Bryophaenocladus vernalis* [Goetghebuer, 1921][Diptera: Chironomidae] in Poland. *Polskie Pismo Entomologiczne* **71(4)**:355–358
- Kaulfuss U, Szawaryn K, Lee DE, Ruta R. 2024.** The first beetle species described from Oligocene New Zealand amber (Coleoptera: Scirtidae). *Palaeoentomology* **7(4)**:529–538.
- Krosch MN, Baker AM, Mather PB, Cranston PS. 2011.** Systematics and biogeography of the Gondwanan Orthocladiinae (Diptera: Chironomidae). *Molecular Phylogenetics and Evolution* **59(2)**:458–468
- Krosch M, Cranston PS. 2013.** Not drowning,(hand) waving? Molecular phylogenetics, biogeography and evolutionary tempo of the ‘Gondwanan’ midge *Stictocladus* Edwards (Diptera: Chironomidae). *Molecular Phylogenetics and Evolution* **68(3)**:595–603
- Krosch MN, Cranston PS, Baker AM, Vink S. 2015.** Molecular data extend Australian *Cricotopus* midge (Chironomidae) species diversity, and provide a phylogenetic hypothesis for biogeography and freshwater monitoring. *Zoological Journal of the Linnean Society* **175(3)**:496–509
- Langton PH, Pinder LCV. 2007.** *Keys to the adult male Chironomidae of Britain and Ireland, Vol. 1 and 2.* Amleside, Cumbria: Freshwater Biological Association, 239 + 168.
- Lee D, Lindqvist J, Mildenhall D, Bannister J, Kaulfuss U. 2009.** Paleobotany, palynology and sedimentology of Late Cretaceous – Miocene sequences in Otago and Southland. In: Turnbull IM, ed., *Field Trip Guides, Geosciences 09 Conference, Oamaru, New Zealand.* Geological Society of New Zealand Miscellaneous Publication 128B, 39.
- Lin X, Qi X, Wang X. 2012.** Two new species of *Bryophaenocladus* Thienemann, 1934 (Diptera, Chironomidae) from China. *ZooKeys* **208**:51–60
- Limaye A. 2012.** Drishti: a volume exploration and presentation tool. Developments in X-ray Tomography VIII. *Proceedings of the Society of Photo-Optical Instrumentation Engineers (SPIE)* **8506**:85060X. <https://doi.org/10.1117/12.935640>
- Lindqvist JK, Gard HJL, Lee DE. 2016.** Geological setting, sedimentology and biota of the estuarine late Oligocene Pomahaka Formation, New Zealand. *New Zealand Journal of Geology and Geophysics* **59(2)**:352–365

Lytaev P, Hipp A, Lottermoser L., Herzen J, Greving I, Khokhriakov I, Meyer-Loges S, Plewka J, Burmester J, Caselle M, Vogelgesang M, Chilingaryan S, Kopmann A, Balzer M, Schreyer A, Beckmann F. 2014. Characterization of the CCD and CMOS cameras for grating-based phase-contrast tomography. *Developments in X-Ray Tomography IX* (921218): 1–11. <https://doi.org/10.1117/12.2061389>

Makarchenko EA, Makarchenko MA. 2006. Chironomids of the genus *Bryophaenocladus* Thienemann, 1934 (Diptera, Chironomidae, Orthoclaadiinae) from the Russian Far East. *Far Eastern Entomologist* 158:1–24

Makarchenko EA, Makarchenko MA. 2019. Review of the genus *Gymnometriocnemus* Edwards (Diptera, Chironomidae, Orthoclaadiinae) from the Russian Far East. *Euroasian Entomological Journal* 18(5):333–340

Marshall S, Kirk-Spriggs HA, Mullerm BS, Paiero MS, Yau T, Jackson MD. 2017. Key to Diptera families—adults. In: Kirk-Spriggs AH, Sinclair BJ, eds. Manual of Afrotropical Diptera. Volume 1: Introductory Chapters and Keys to Diptera Families. Suricata, Pretoria: South African National Biodiversity Institute. 267-355

Moller Pillot HKM. 2013. *Chironomidae Larvae, Volume 3: Biology and Ecology of the aquatic Orthoclaadiinae*. Zeist: KNNV Publishing, 312.

Moosmann J, Ershov A, Weinhardt V, Baumbach T, Prasad M S, LaBonne C, Xiao X, Kashef J, Hoffmann R. 2014. Time-lapse X-ray phase-contrast microtomography for in vivo imaging and analysis of morphogenesis. *Nature Protocols* 9:294–304. <https://doi.org/10.1038/nprot.2014.033>

Moubayed J, Lods-Crozet B. 2022. On the genus *Bryophaenocladus* Thienemann, 1934. I. Taxonomic notes with description of new species (Diptera: Chironomidae, Orthoclaadiinae). *Euroasian Entomological Journal* 21(4):212–224

Moubayed J, Langton P. 2023. On the genus *Bryophaenocladus* Thienemann, 1934 (Diptera: Chironomidae, Orthoclaadiinae). II. Description of three new species from continental France. *Euroasian Entomological Journal* 22(1):1–10

Pinder LCV, Armitage PD. 1986. The male and female of *Bryophaenocladus muscicola* (Kieffer), based on new material from England (Diptera: Chironomidae). *Insect Systematics & Evolution* 17(2):137–142

- 623 **Pocknall DT. 1982.** Palynology of late Oligocene Pomahaka Estuarine Bed sediments,  
624 Waikoikoi, Southland, New Zealand. *New Zealand Journal of Botany* **20**:263–287
- 625 **Sæther OA. 1969.** Some nearctic Podonominae, Diamesinae, and Orthocladiinae (Diptera:  
626 Chironomidae). *Bulletin of the Fisheries Research Board of Canada* **170**:1–154
- 627 **Sæther OA. 1973.** Four species of *Bryophaenocladus* Thien., with notes on other  
628 Orthocladiinae (Diptera: Chironomidae). *The Canadian Entomologist* **105**(1):51–60.
- 629 **Sæther OA. 1977.** Female genitalia in Chironomidae and other Nematocera: morphology,  
630 phylogenies, keys. *Bulletin of the Fisheries Research Board of Canada* **197**:1–209
- 631 **Sæther OA. 1980.** Glossary of chironomid morphology terminology (Diptera: Chironomidae).  
632 *Entomologica Scandinavica Supplement* **14**:1-51
- 633 **Sæther OA. 1982.** Orthocladiinae (Diptera: Chironomidae) from SE U.S.A., with descriptions of  
634 *Plhudsonia*, *Unniella* and *Platysmittia* n. genera and *Atelopodella* n. subgen. *Entomologica*  
635 *Scandinavica* (= *Insect Systematics & Evolution*) **13**:465–510.
- 636 **Sæther OA. 1983.** A review of Holarctic *Gymnometriocnemus* Goetghebuer, 1932, with the  
637 description of *Raphidocladus* subgen. n. and *Sublettiella* gen. n. (Diptera: Chironomidae).  
638 *Aquatic Insects* **5**(4):209–226.
- 639 **Sadowski E-M, Schmidt AR, Seyfullah LJ, Solórzano-Kraemer MM, Neumann C,**  
640 **Perrichot V, Hamann C, Milke R, Nascimbene PC. 2021.** Conservation, preparation and  
641 imaging of diverse ambers and their inclusions. *Earth-Science Reviews* **220**:103653
- 642 **Sanmartin I, Enghoff H, Ronquist F. 2001.** Patterns of animal dispersal, vicariance and  
643 diversification in the Holarctic. *Biological Journal of the Linnean Society* **73**(4):345–390.
- 644 **Sasa M, Kikuchi M. 1995.** *Chironomidae (Diptera) of Japan*. Tokyo: University of Tokyo  
645 Press, 333.
- 646 **Schakau B. 1991.** Stratigraphy of the fossil Chironomidae (Diptera) from Lake Grasmere, South  
647 Island, New Zealand, during the last 6000 years. *Hydrobiologia* **214**:213–221
- 648 **Schmidt AR, Kaulfuss U, Bannister JM, Baranov V, Beimforde C, Bleile N, Borkent A,**  
649 **Busch A, Conran JG, Engel MS, Harvey M, Kennedy EM, Kerr PH, Kettunen E, Kiecksee**  
650 **AP, Lengeling F, Lindqvist JK, Maraun M, Mildenhall DC, Perrichot V, Rikkinen J,**

651 **Sadowski E-M, Seyfullah LJ, Stebner F, Szwedo J, Ulbrich P, Lee DE. 2018.** Amber  
652 inclusions from New Zealand. *Gondwana Research* **56**:135–146.

653 **Seredszus F, Wichard W. 2007.** Fossil chironomids (Insecta, Diptera) in Baltic amber.  
654 *Palaeontographica Abteilung A* **279(1–3)**:49–91

655 **Solórzano-Kraemer MM, Delclòs X, Clapham ME, Arillo A, Peris D, Jäger P, Stebner F,**  
656 **Peñalver, E. 2018.** Arthropods in modern resins reveal if amber accurately recorded forest  
657 arthropod communities. *Proceedings of the National Academy of Sciences* **115(26)**:6739–6744

658 **Strenzke K. 1957.** Terrestrische Chironomiden. XVI. *Bryophaenocladus nitidicollis*  
659 (Goetgh.)(Diptera: Tendipedidae, Orthoclaadiinae). *Beiträge zur Entomologie (= Contributions to*  
660 *Entomology)* **7(3–4)**:398–411.

661 **Stur E, Ekrem T. 2015.** A review of Norwegian *Gymnometriocnemus* (Diptera, Chironomidae)  
662 including the description of two new species and a new name for *Gymnometriocnemus volitans*  
663 (Goetghebuer) sensu Brundin. *ZooKeys* **508**:127–142.

664 **Sublette JE, Wirth WW. 1980.** The Chironomidae and Ceratopogonidae (Diptera) of New  
665 Zealand's subantarctic islands. *New Zealand Journal of Zoology* **7(3)**:299–378.

666 **Trewick SA. 2000.** Molecular evidence for dispersal rather than vicariance as the origin of  
667 flightless insect species on the Chatham Islands, New Zealand. *Journal of Biogeography*  
668 **27(5)**:1189–1200.

669 **Van Aarle W, Palenstijn WJ, De Beenhouwer J, Altantzis T, Bals S., Batenburg KJ, Sijbers**  
670 **J. 2015.** The ASTRA Toolbox: A platform for advanced algorithm development in electron  
671 tomography. *Ultramicroscopy* **157**:35–47. <https://doi.org/10.1016/j.ultramic.2015.05.002>

672 **Van Aarle W, Palenstijn WJ, Cant J, Janssens E, Bleichrodt F., Dabravolski A., De**  
673 **Beenhouwer J., Batenburg KJ., Sijbers J. 2016.** Fast and flexible X-ray tomography using the  
674 ASTRA toolbox. *Optics Express* **24**:25129–25147. <https://doi.org/10.1364/OE.24.025129>

675 **Veltz I, Azar D, Nel A. 2007.** New chironomid flies in Early Cretaceous Lebanese amber  
676 (Diptera: Chironomidae). *African Invertebrates* **48**:169–191.

677 **Wang XH, Sæther OA, Andersen T. 2001.** Afrotropical *Bryophaenocladus* Thienemann, 1934  
678 (Diptera: Chironomidae). *Studia Dipterologica* **8**:447–462.

679 **Wang XH, Liu Z, Epler JH. 2004.** New species of *Bryophaenocladus* Thienemann from the  
680 Nearctic Region (Diptera: Chironomidae: Orthoclaadiinae). *Zootaxa* **581(1)**:1–10.

- 681 **Wang X, Andersen T, Sæther OA. 2006.** Neotropical *Bryophaenocladus* Thienemann, 1934  
682 (Diptera: Chironomidae). *Studies on Neotropical Fauna and Environment* **41(1)**:19–32.
- 683 **Willassen E. 1996.** A nival *Bryophaenocladus* THIENEMANN, 1934, with reduced wings.  
684 *Annalen des Naturhistorischen Museums in Wien* **98B**:507–512.
- 685 **Wood BL. 1956.** The geology of the Gore subdivision. *New Zealand Geological Survey Bulletin*  
686 **53**:1–128.
- 687 **Woodward CA, Shulmeister J. 2007.** Chironomid-based reconstructions of summer air  
688 temperature from lake deposits in Lyndon Stream, New Zealand spanning the MIS 3/2 transition.  
689 *Quaternary Science Reviews* **26(1-2)**:142–154.
- 690 **Zelentsov NI, Baranov VA, Perkovsky EE, Shobanov NA. 2012.** First records on non-biting  
691 midges (Diptera: Chironomidae) from the Rovno amber. *Russian Entomological Journal*  
692 **21(1)**:79–87.

**Figure 1 Late Oligocene Pomahaka amber.** (A) Map of amber locality G45/f0107 near Tapanui, southern New Zealand. (B) Typical appearance of layered, fossiliferous Pomahaka amber. (C) Epoxy-embedded pieces of Pomahaka amber with newly discovered Chironomidae inclusions.

**Figure 2 *Bryophaenocladus zealandiae* sp. nov. Baranov, holotype OU47576.** (A) Habitus, dorsal view. (B) Habitus, ventral view. (C) Antenna, ventral view. (D) Tibial spurs. Abbreviations: *Ti1*, foreleg tibia; *Ti2*, midleg tibia; *Ti3*, hindleg tibia.

**Figure 3 *Bryophaenocladus zealandiae* sp. nov. Baranov, paratypes.** (A, B) Habitus and hypopygium of paratype OU47540. (C, D) Hypopygium and habitus of paratype OU47575. (E) Habitus of paratype OU47572.

**Figure 4 *Bryophaenocladus zealandiae* sp. nov. Baranov, wing of holotype OU47576.** (A) Photomicrograph. (B) Line drawing. Abbreviations: *An*, anal vein; *B*, brachiolum; *C*, costal vein; *ce*, costal extension; *Cu1*, cubital vein 1; *Cu2*, cubital vein 2; *M1+2*, medial vein 1+2; *M3+4*, medial vein 3+4; *R1*, radial vein 1; *R2+3*, radial vein 2+3; *R4+5*, radial vein 4+5; *RM*, radial medial crossvein; *Sc*, subcostal vein; *Sq*, squama.

**Figure 5 *Bryophaenocladus zealandiae* sp. nov. Baranov, hypopigium of holotype OU47576.** (A) Photomicrograph, dorsal. (B) Line drawing, dorsal. (C) Photomicrograph, ventral. (D) Line drawing, ventral. Abbreviations: *AP*, anal point; *Gc*, gonocoxite; *Gs*, gonostylus; *Ivo*, inferior volsella; *TVIII*, abdominal tergite 8; *TIX* abdominal tergite 9.

**Figure 6 *Bryophaenocladus zealandiae* sp. nov. Baranov, associated specimen OU47579.**

(A) Habitus. (A') Overview of the amber piece containing the specimen. (B) Habitus, opposite side of the body.

**Figure 7 *Bryophaenocladus zealandiae* sp. nov. Baranov, uCT scans of associated**

**specimens.** (A) Habitus of specimens OU47580 and OU47581 in the same amber piece. (B) Habitus of specimen OU47580, dorso-lateral view. (C) Hypopigium of specimen OU47581. (D) Hypopigium (OU47581), marked. Abbreviations: *AP*, anal point; *Gc*, gonocoxite; *Gs*, gonostylus; *TIX*, abdominal tergite 9. (E) Overview of the amber piece containing specimens OU47580, OU47581 and OU47582.

**Figure 8 Morphotype 1 cf. *Bryophaenocladus zealandiae*.** (A) Habitus of specimen OU47574.

(B) Habitus of specimen OU47573. (C) Palpomere 3 (OU47574), arrow marks distal protrusion. (D) Hypopigium, lateral view (OU47574).

**Figure 9 *Gymnometriocnemus extinctus* sp. nov. Baranov, holotype OU47546, male.** (A)

Habitus. (B) Head. (C) Last flagellomere apical setae marked by arrow. (D) Hypopigium, ventral view.

**Figure 10 *Gymnometriocnemus extinctus* sp. nov. Baranov, holotype OU47546.** (A)

Photomicrograph of wing. (B) Line drawing of wing. (C) Photomicrograph of hypopigium, dorsal view. (D) Line drawing of hypopigium, dorsal view. Abbreviations: *AP*, anal point; *Gc*, gonocoxite; *Gs*, gonostylus; *IVo*, inferior volsella; *TIX*, abdominal tergite 9; *C*, costal vein; *Cu1*, cubital vein 1; *M1+2*, medial vein 1+2; *M3+4*, medial vein 3+4; *R1*, radial vein 1; *R2+3*, radial vein 2+3; *R4+5*, radial vein 4+5.

**Figure 11 *Gymnometriocnemus extinctus* sp. nov. Baranov, female paratypes.** (A, B) Habitus

and wing of specimen OU47577. (C) Habitus of specimen OU47578.

**Figure 12** *Gymnometriocnemus extinctus* sp. nov. Baranov, female paratypes. (A) Head of specimen OU47577. (B) Head of specimen OU47578; b' close-up of last flagellomere with apical setae. (C) Female genitalia (OU47578). (D) Female genitalia, marked (OU47578). Abbreviations: *A8*, abdominal segment 8; *Ce*, cerci; *Gca*, gonocoxite apodem; *Gc*, gonocoxite (8); *Gp8*, gonapophysis 8; *TIX*, tergite 9.

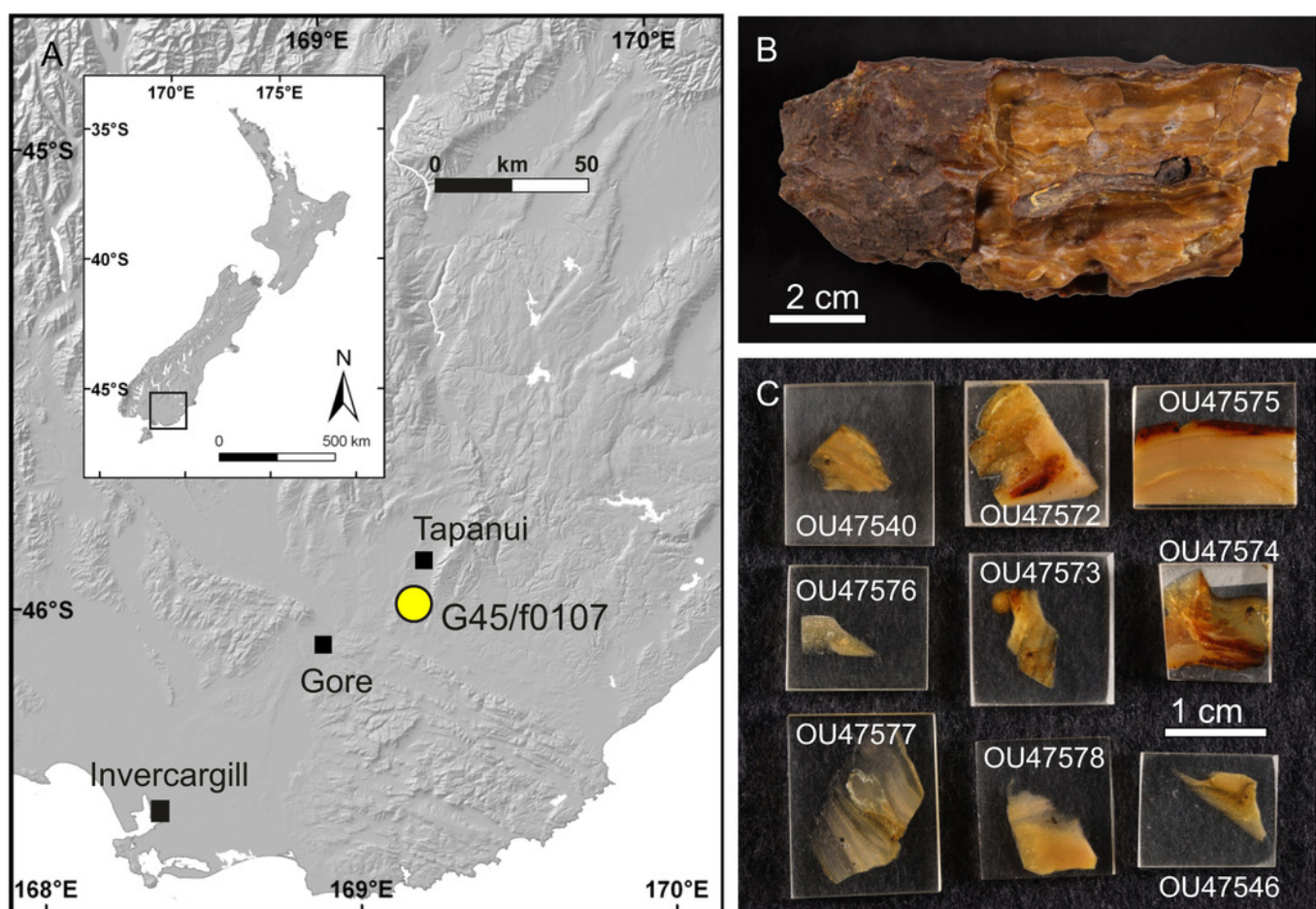
**Figure 13.** Holotype (adult male, number NZAC02044947) of *Kuschelius dentifer* Sublette and Wirth, 1980. (A). Wing. (B) Head, arrow marks a apical protrusion of the 3d palpomere. (C) Hypopigium. (D) Midtibia with spurs and the comb. (E) Hindtibia with the spurs and comb. All photos in this plate are made by Dr. Leanne Elder, licensed under CC BY 4.0 and used with the photographer's explicit permission.



# Figure 1

Figure 1 Late Oligocene Pomahaka amber.

(A) Map of amber locality G45/f0107 near Tapanui, southern New Zealand. (B) Typical appearance of layered, fossiliferous Pomahaka amber. (C) Epoxy-embedded pieces of Pomahaka amber with newly discovered Chironomidae inclusions.



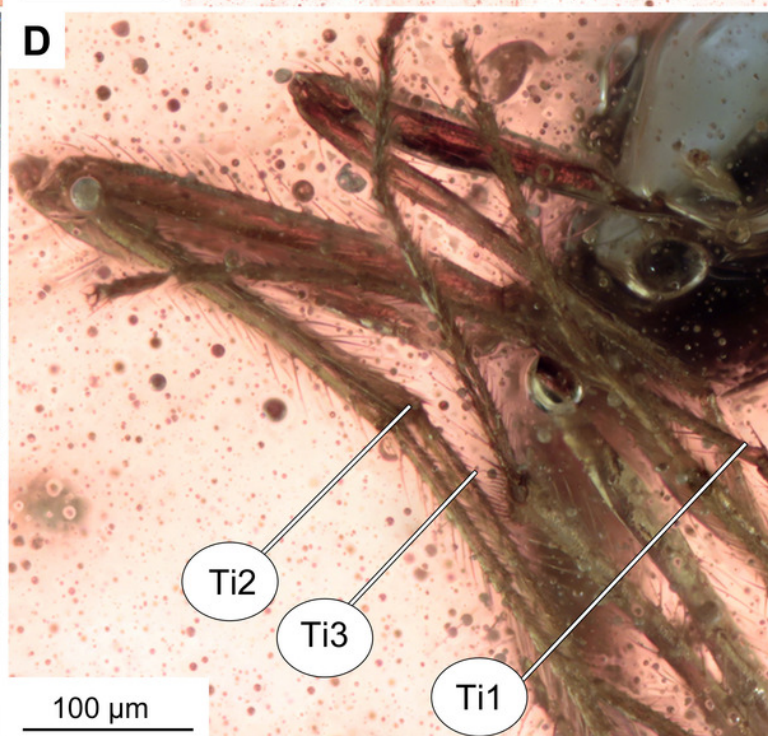
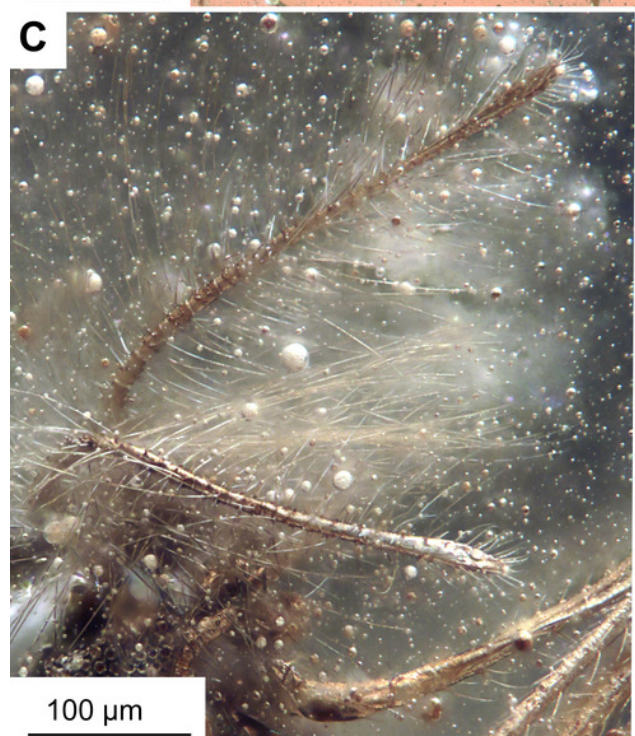
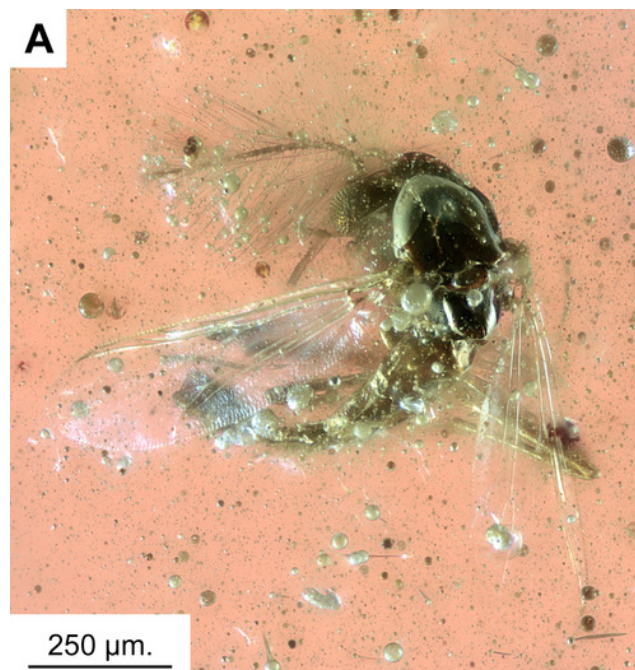
# Figure 2

Figure 2 *Bryophaenocladus zealandiae* sp. nov. Baranov, holotype OU47576.

(A) Habitus, dorsal view. (B) Habitus, ventral view. (C) Antenna, ventral view. (D) Tibial spurs.

Abbreviations: *Ti1*, foreleg tibia; *Ti2*, midleg tibia; *Ti3*, hindleg tibia.



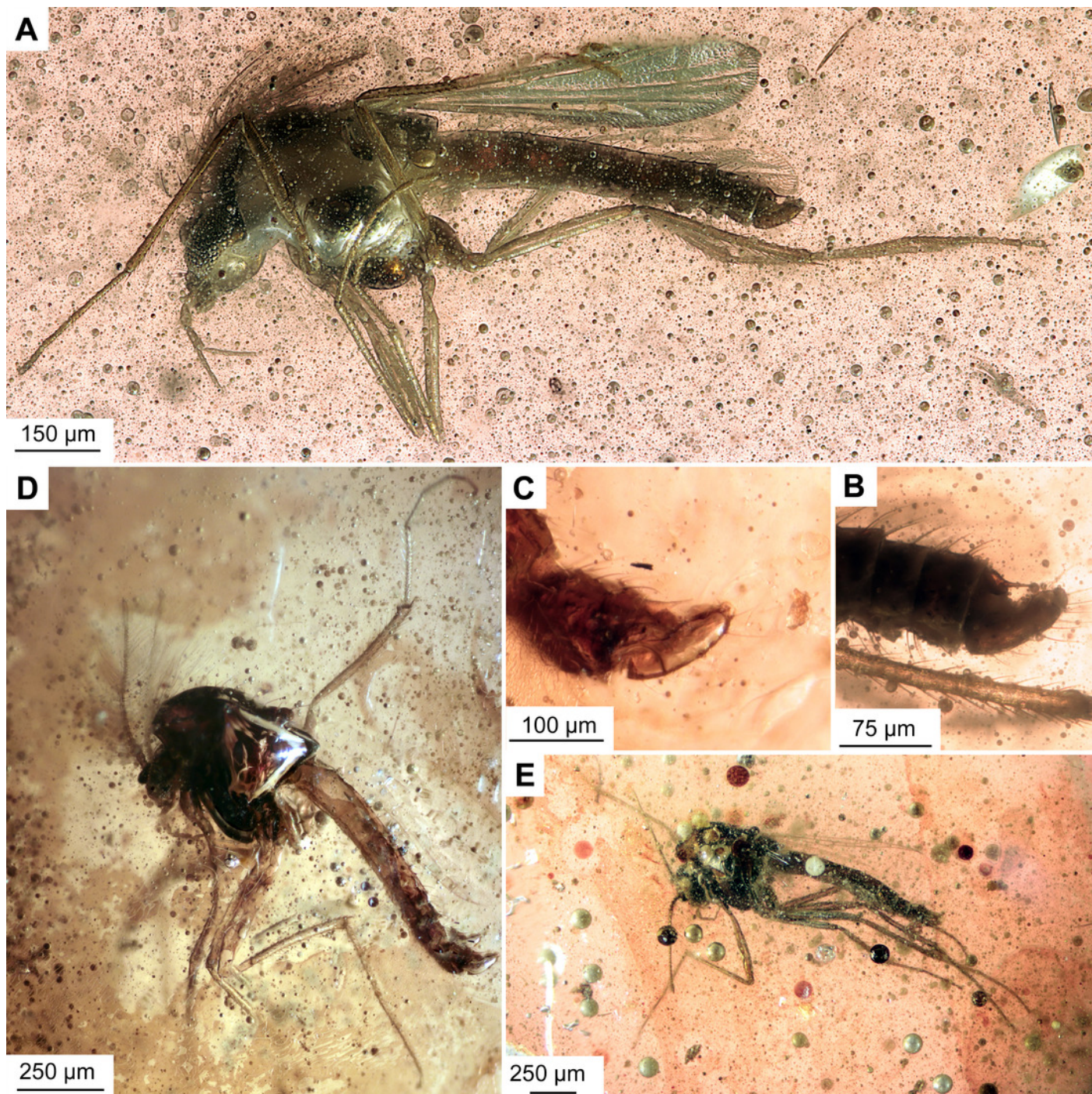


# Figure 3

Figure 3 *Bryophaenocladus zealandiae* sp. nov. Baranov, paratypes.

(A, B) Habitus and hypopygium of paratype OU47540. (C, D) Hypopygium and habitus of paratype OU47575. (E) Habitus of paratype OU47572.



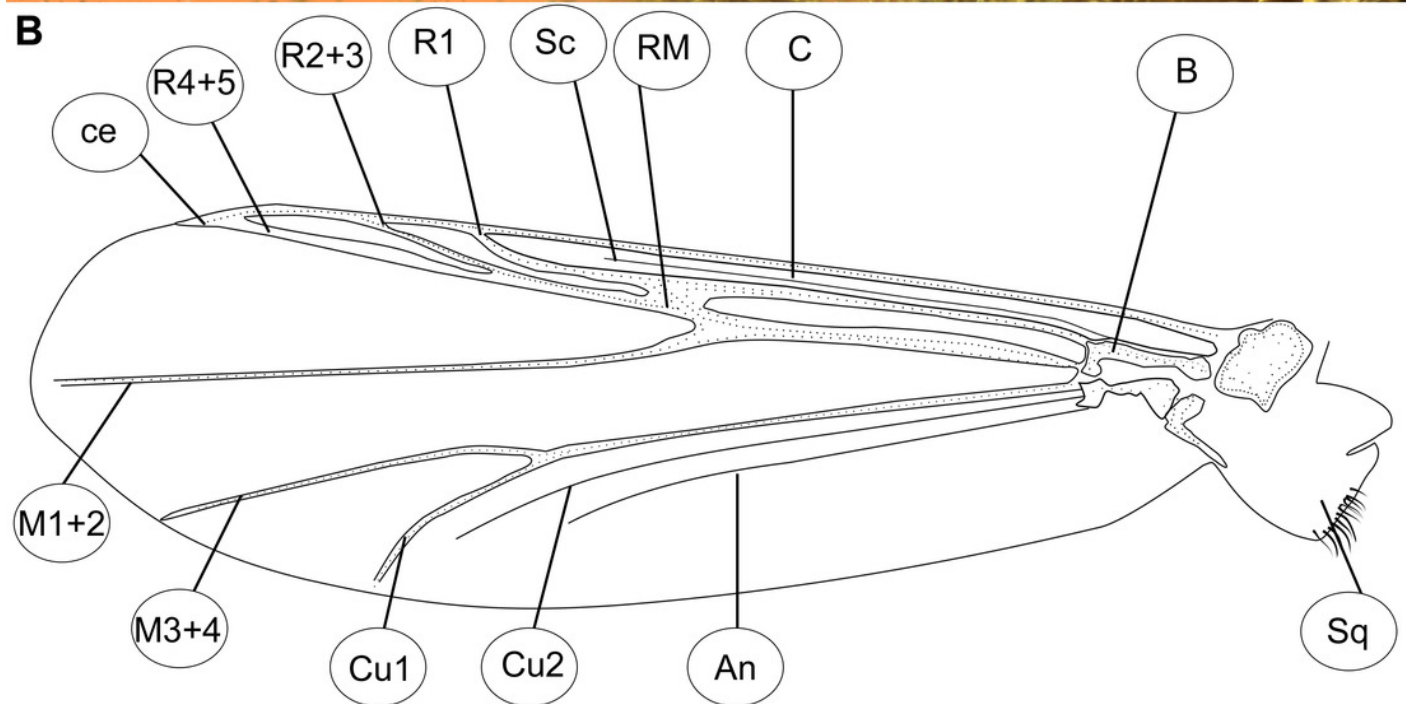
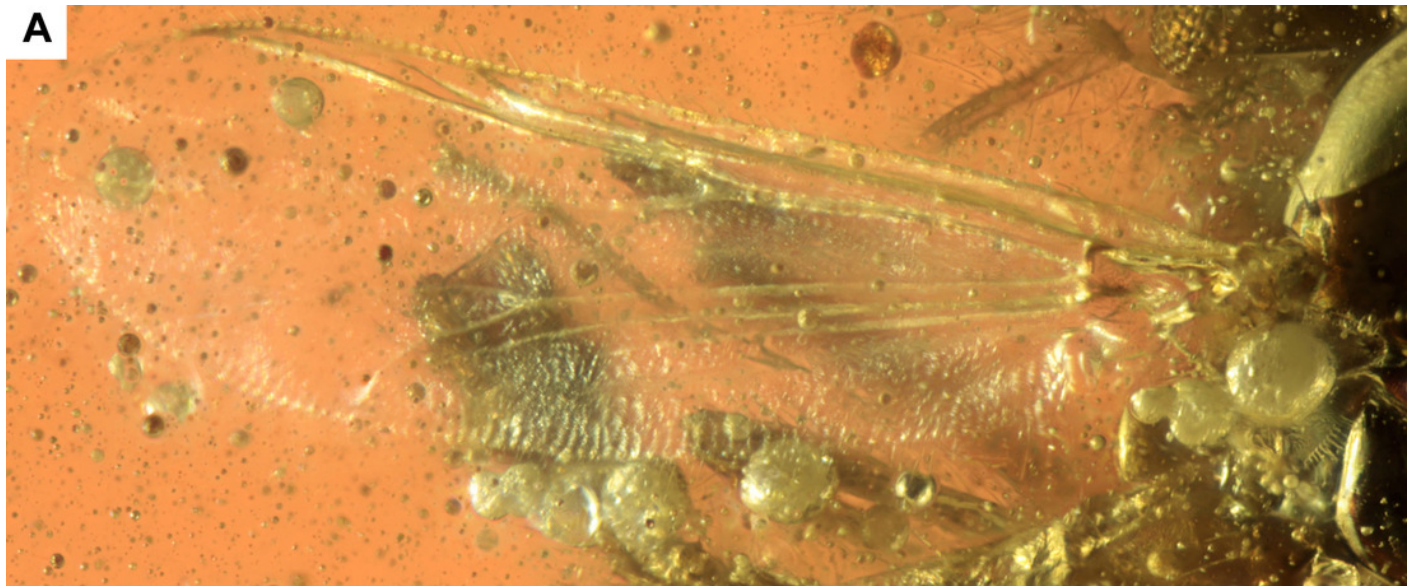


# Figure 4

Figure 4 *Bryophaenocladus zealandiae* sp. nov. Baranov, wing of holotype OU47576.

(A) Photomicrograph. (B) Line drawing. Abbreviations: *An*, anal vein; *B*, brachiolum; *C*, costal vein; *ce*, costal extension; *Cu1*, cubital vein 1; *Cu2*, cubital vein 2; *M1+2*, medial vein 1+2; *M3+4*, medial vein 3+4; *R1*, radial vein 1; *R2+3*, radial vein 2+3; *R4+5*, radial vein 4+5; *RM*, radial medial crossvein; *Sc*, subcostal vein; *Sq*, squama.



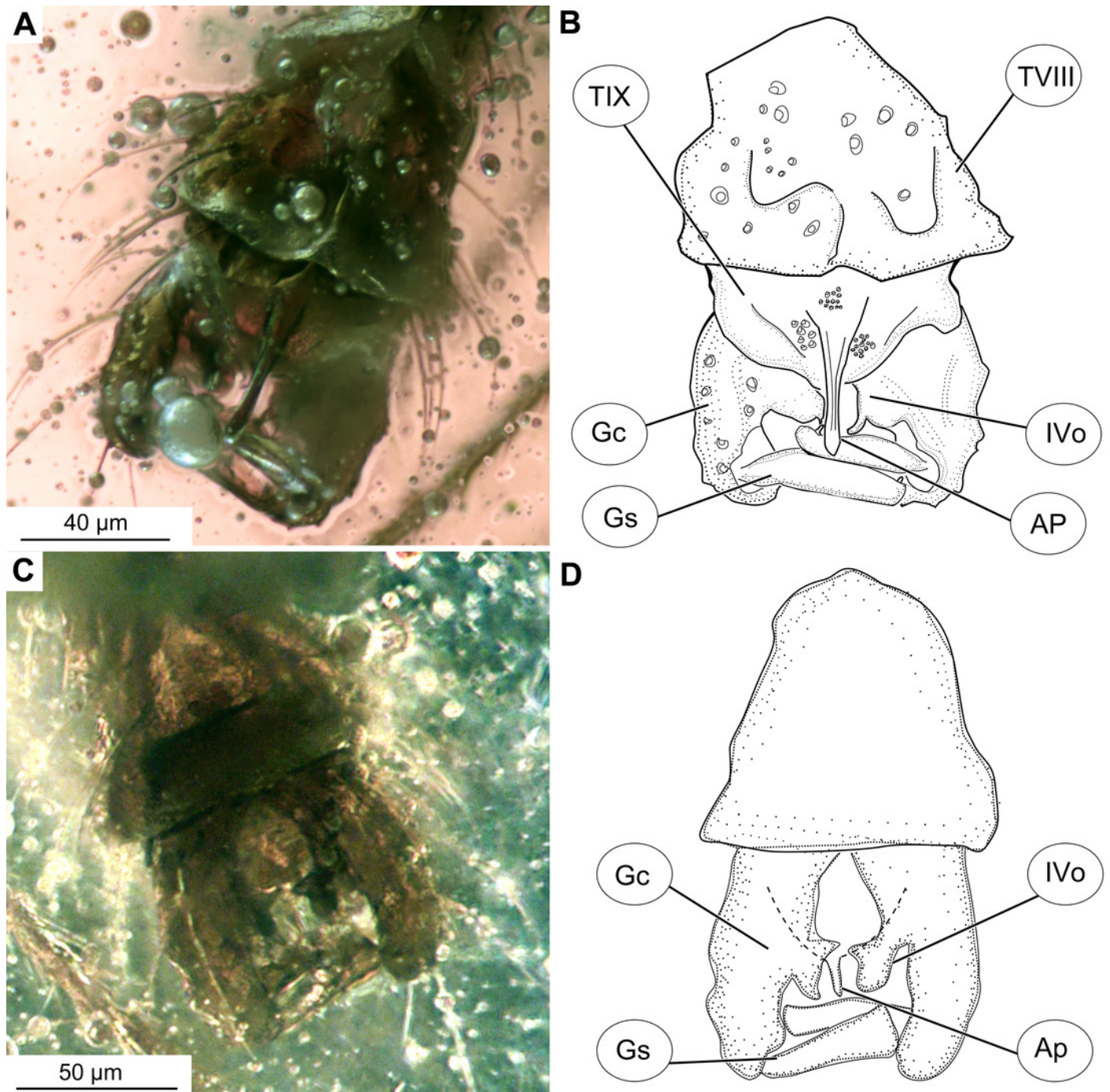


# Figure 5

Figure 5 *Bryophaenocladus zealandiae* sp. nov. Baranov, hypopigium of holotype OU47576.

(A) Photomicrograph, dorsal. (B) Line drawing, dorsal. (C) Photomicrograph, ventral. (D) Line drawing, ventral. Abbreviations: *AP*, anal point; *Gc*, gonocoxite; *Gs*, gonostylus; *IVo*, inferior volsella; *TVIII*, abdominal tergite 8; *TIX* abdominal tergite 9.



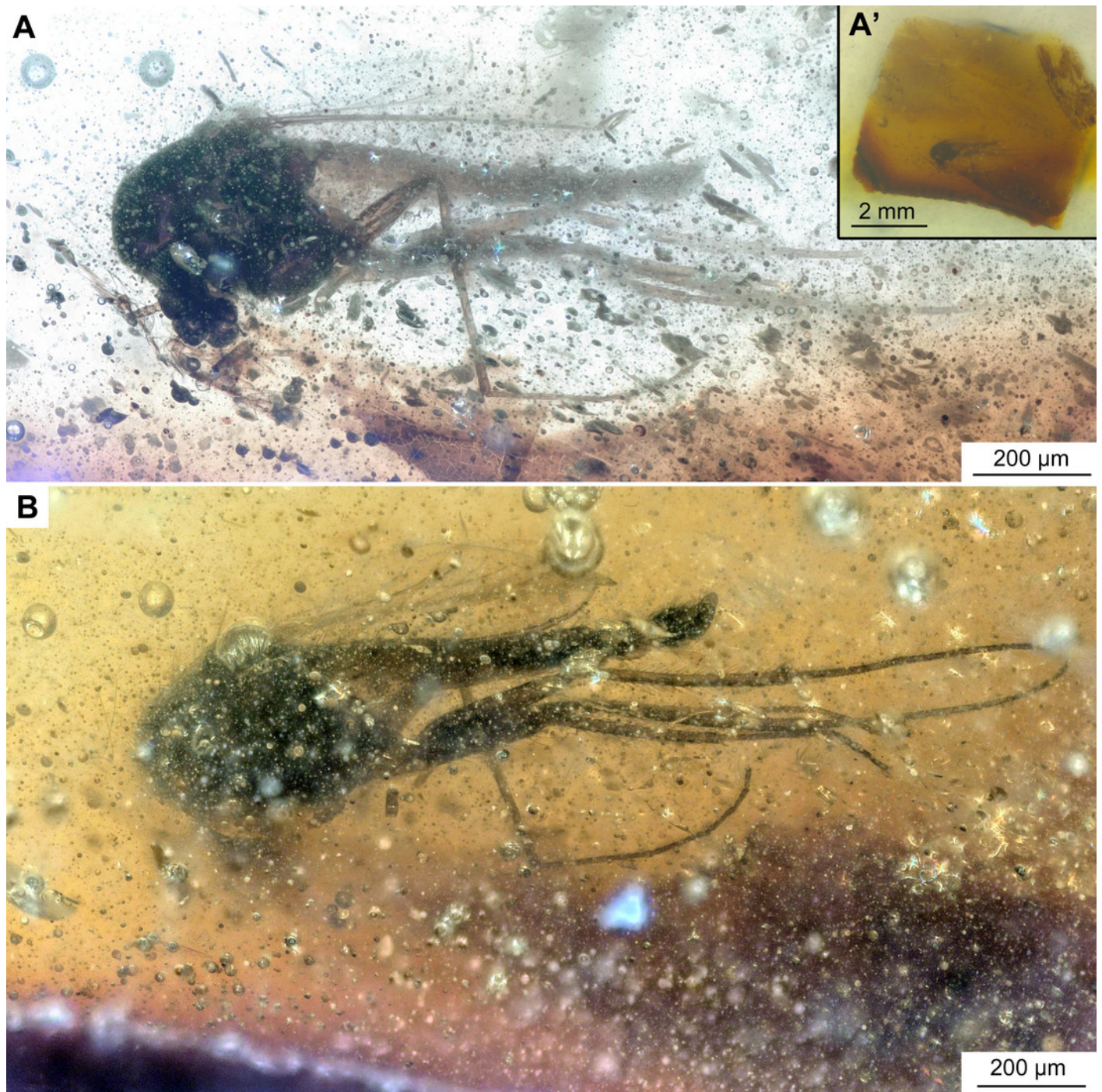


# Figure 6

Figure 6 *Bryophaenocladus zealandiae* sp. nov. Baranov, associated specimen OU47579.

(A) Habitus. (A') Overview of the amber piece containing the specimen. (B) Habitus, opposite side of the body.



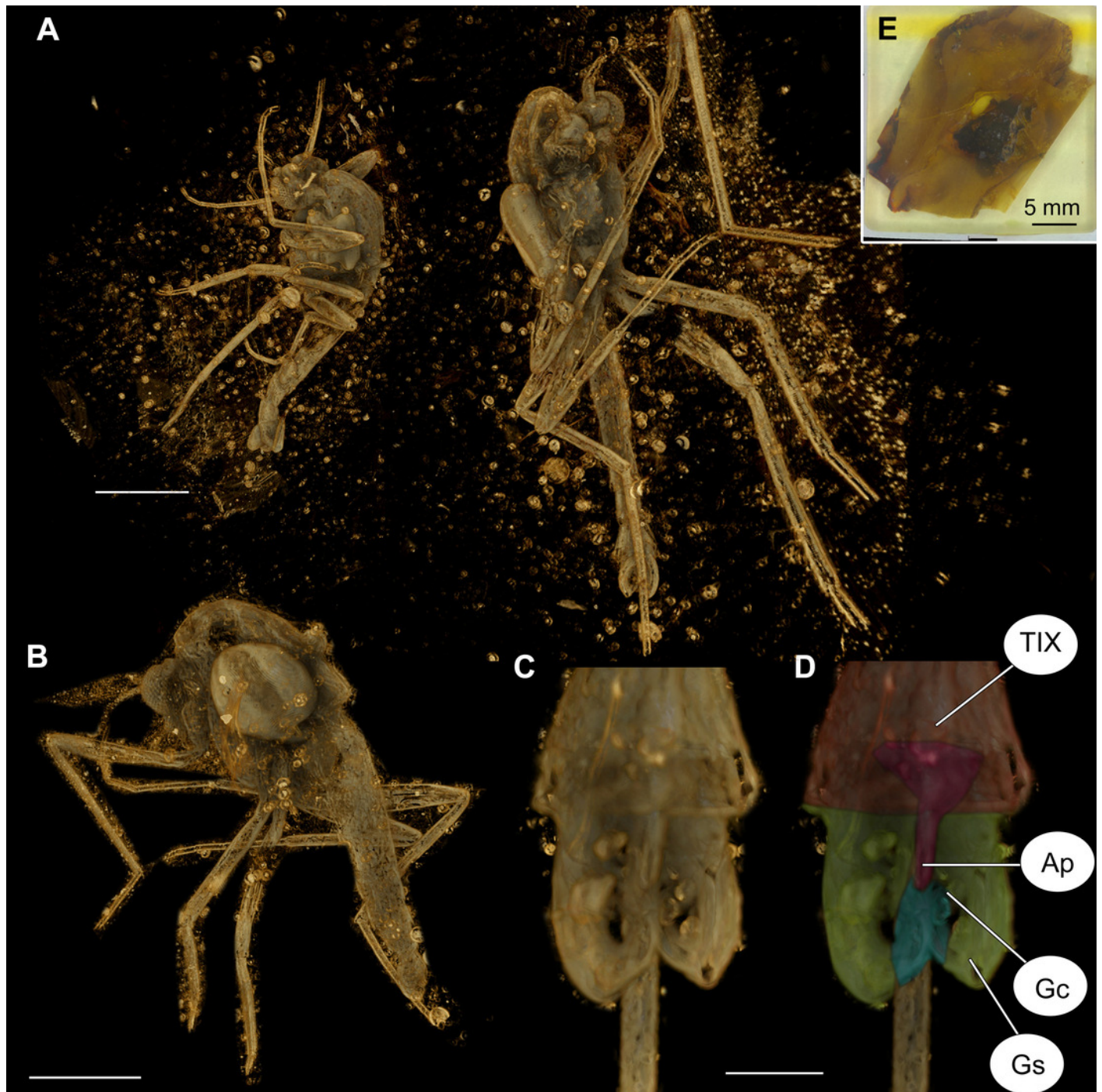


# Figure 7

Figure 7 *Bryophaenocladus zealandiae* sp. nov. Baranov, uCT scans of associated specimens.

(A) Habitus of specimens OU47580 and OU47581 in the same amber piece. (B) Habitus of specimen OU47580, dorso-lateral view. (C) Hypopigium of specimen OU47581. (D) Hypopigium (OU47581), marked. Abbreviations: *AP*, anal point; *Gc*, gonocoxite; *Gs*, gonostylus; *TIX*, abdominal tergite 9. (E) Overview of the amber piece containing specimens OU47580, OU47581 and OU47582.



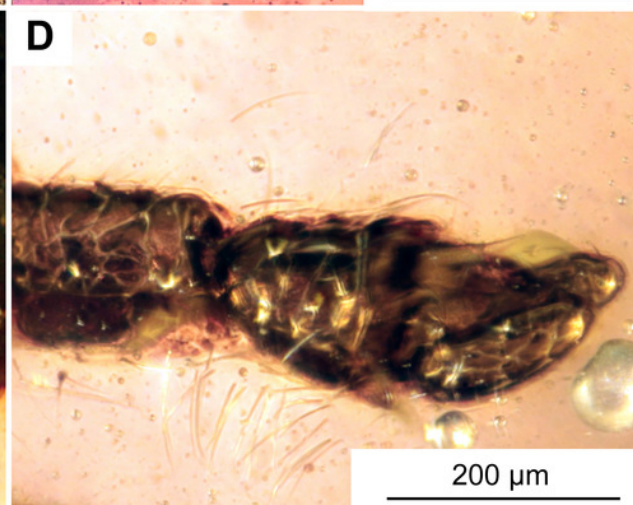
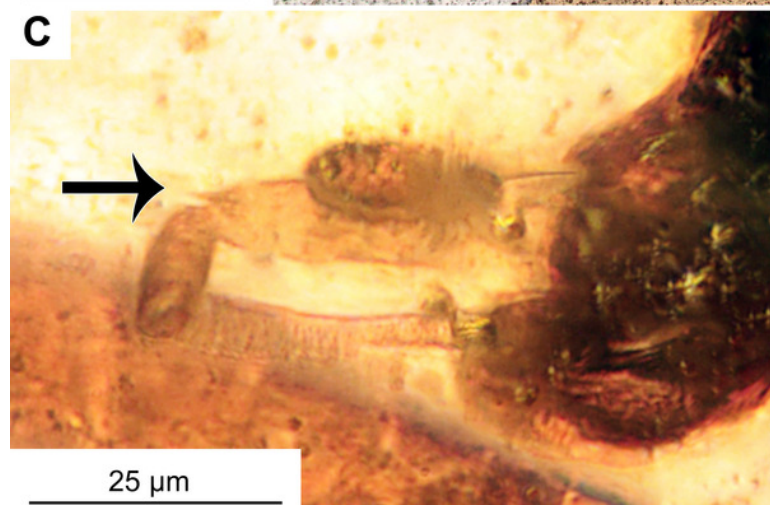
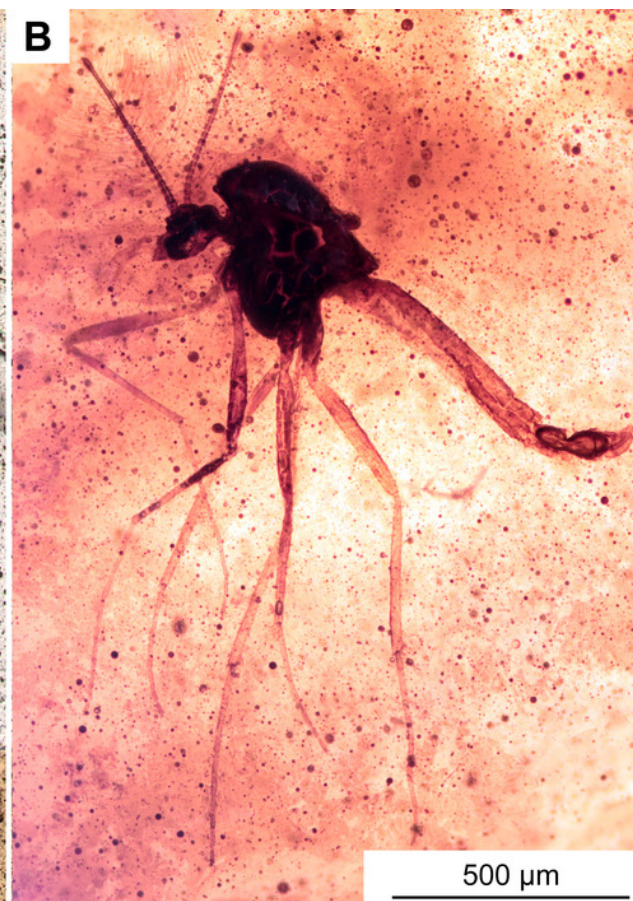


# Figure 8

Figure 8 Morphotype 1 cf. *Bryophaenocladus zealandiae*.

(A) Habitus of specimen OU47574. (B) Habitus of specimen OU47573. (C) Palpomere 3 (OU47574), arrow marks distal protrusion. (D) Hypopigium, lateral view (OU47574).



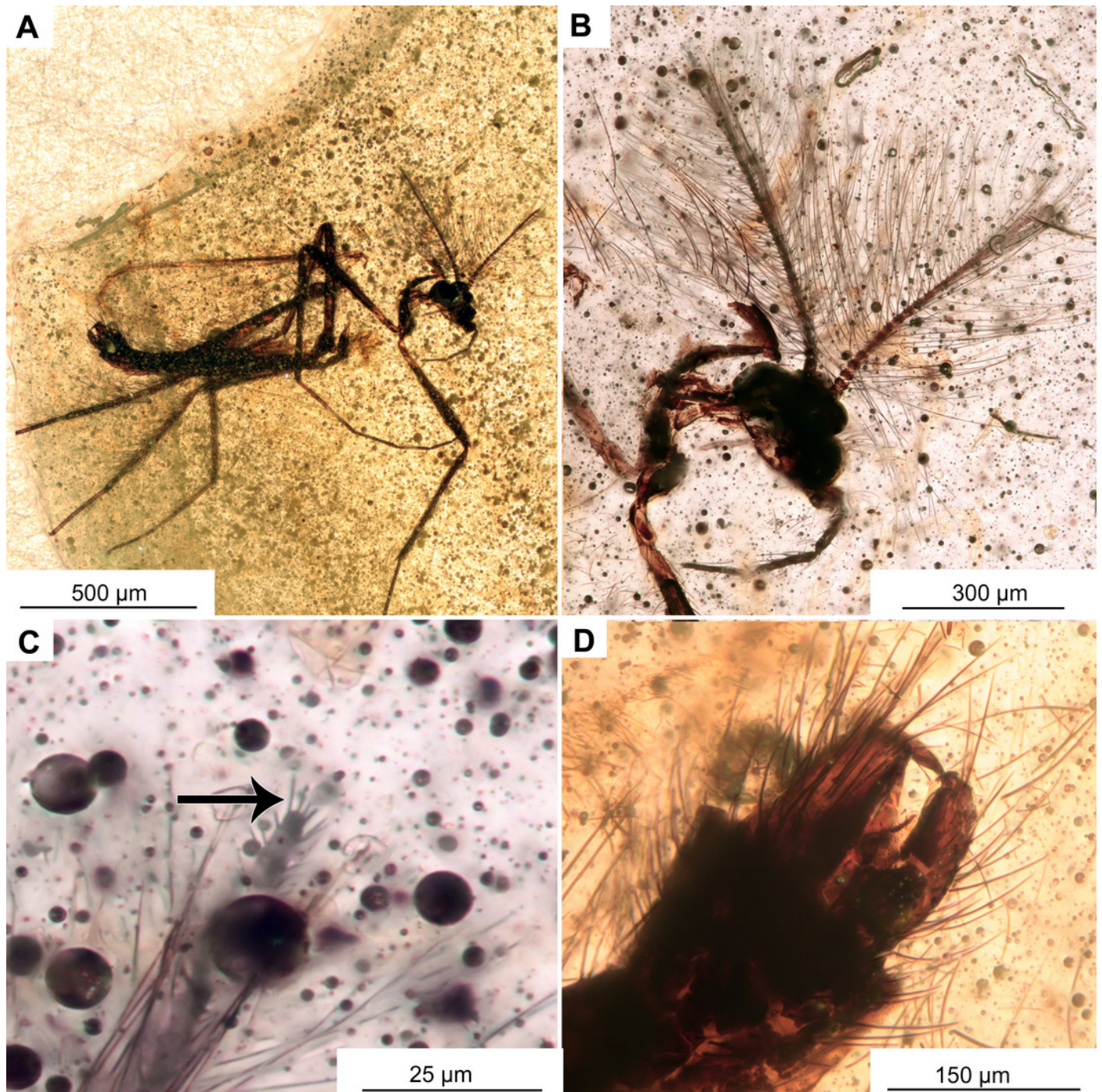


# Figure 9

Figure 9 *Gymnometriocnemus extinctus* sp. nov. Baranov, holotype OU47546, male.

(A) Habitus. (B) Head. (C) Last flagellomere apical setae marked by arrow. (D) Hypopigium, ventral view.



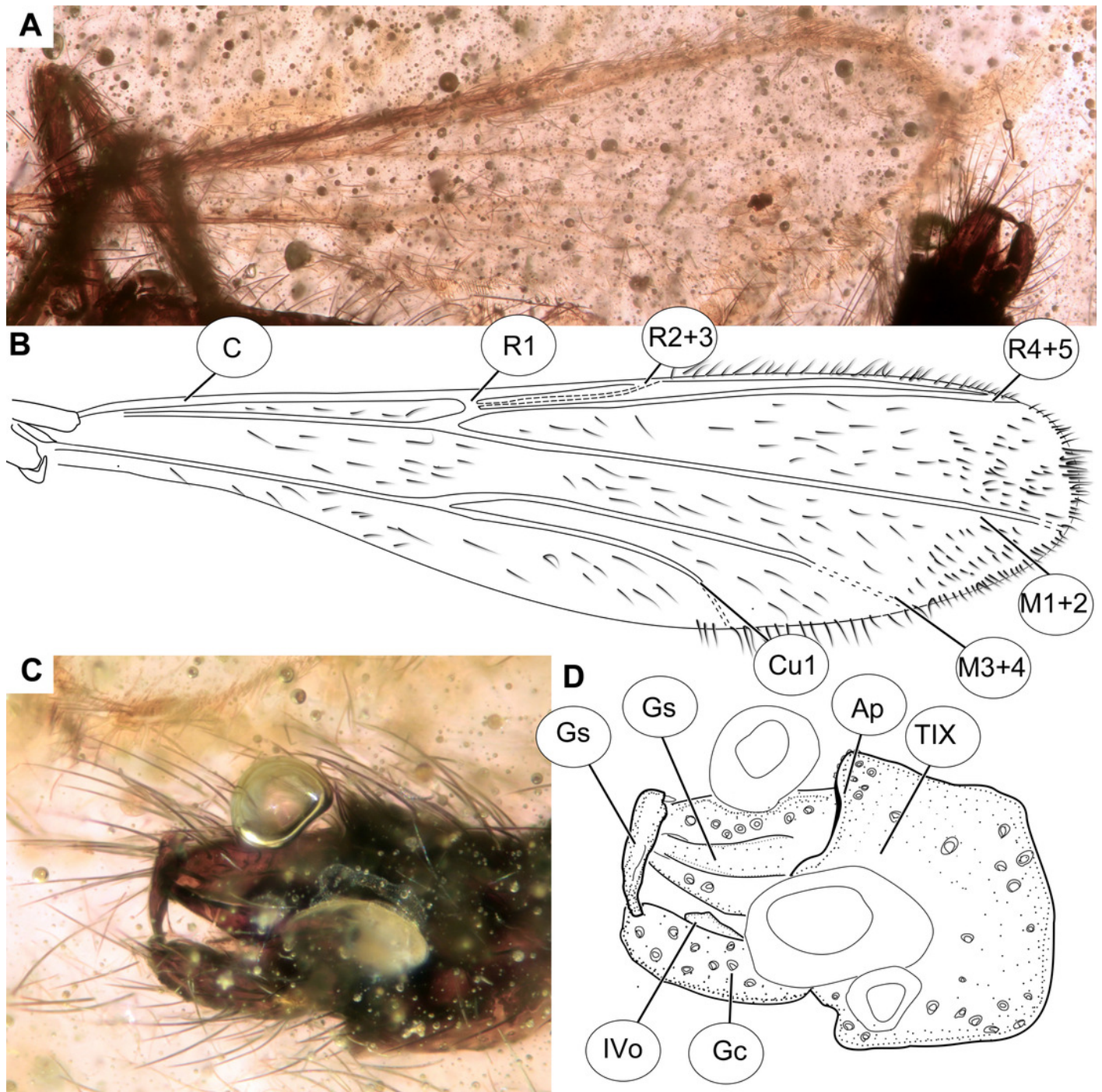


# Figure 10

Figure 10 *Gymnometriocnemus extinctus* sp. nov. Baranov, holotype OU47546.

(A) Photomicrograph of wing. (B) Line drawing of wing. (C) Photomicrograph of hypopigium, dorsal view. (D) Line drawing of hypopigium, dorsal view. Abbreviations: *AP*, anal point; *Gc*, gonocoxite; *Gs*, gonostylus; *IVo*, inferior volsella; *TIX*, abdominal tergite 9; *C*, costal vein; *Cu1*, cubital vein 1; *M1+2*, medial vein 1+2; *M3+4*, medial vein 3+4; *R1*, radial vein 1; *R2+3*, radial vein 2+3; *R4+5*, radial vein 4+5.



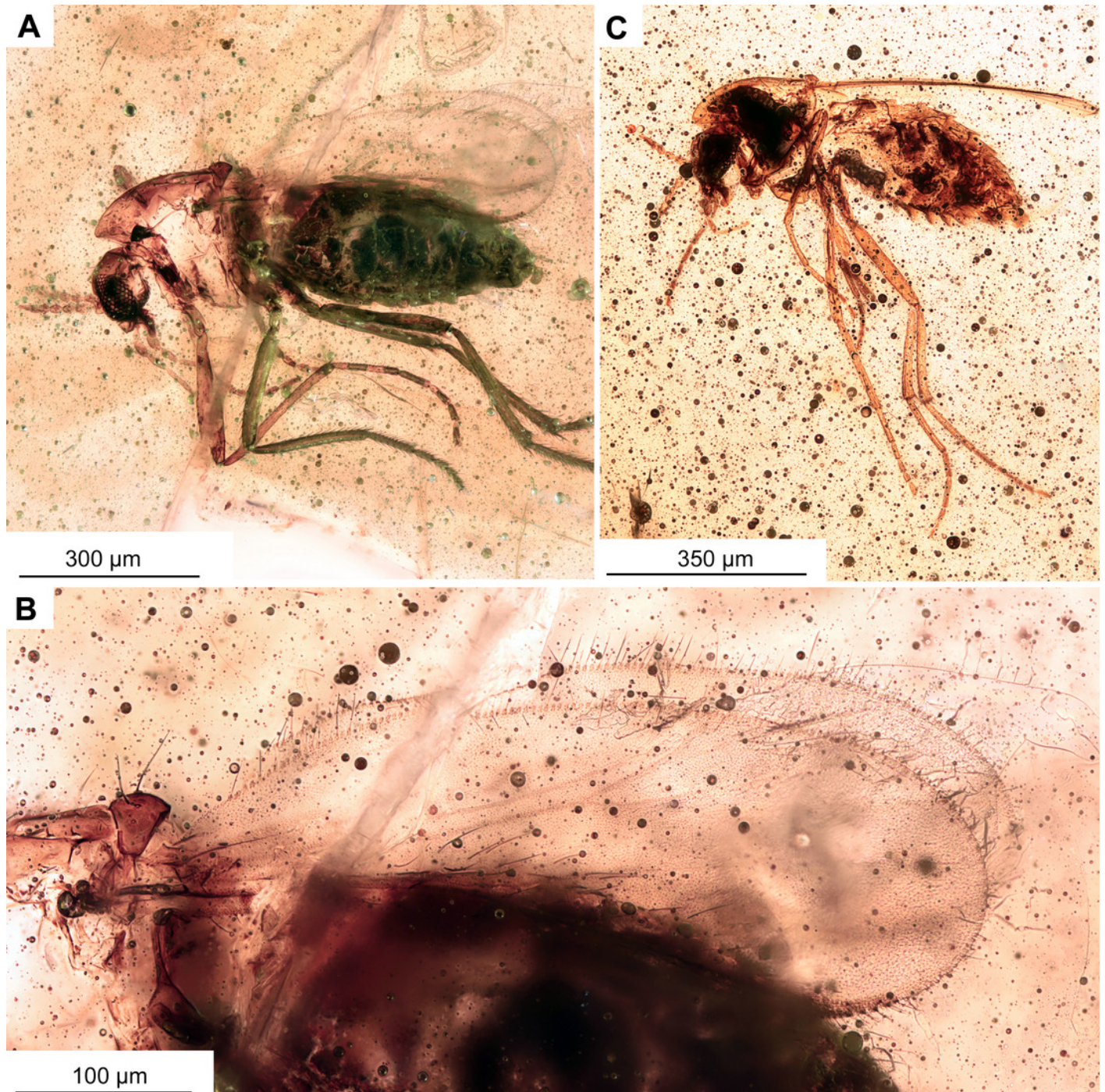




# Figure 11

Figure 11 *Gymnometriocnemus extinctus* sp. nov. Baranov, female paratypes.

(A, B) Habitus and wing of specimen OU47577. (C) Habitus of specimen OU47578.

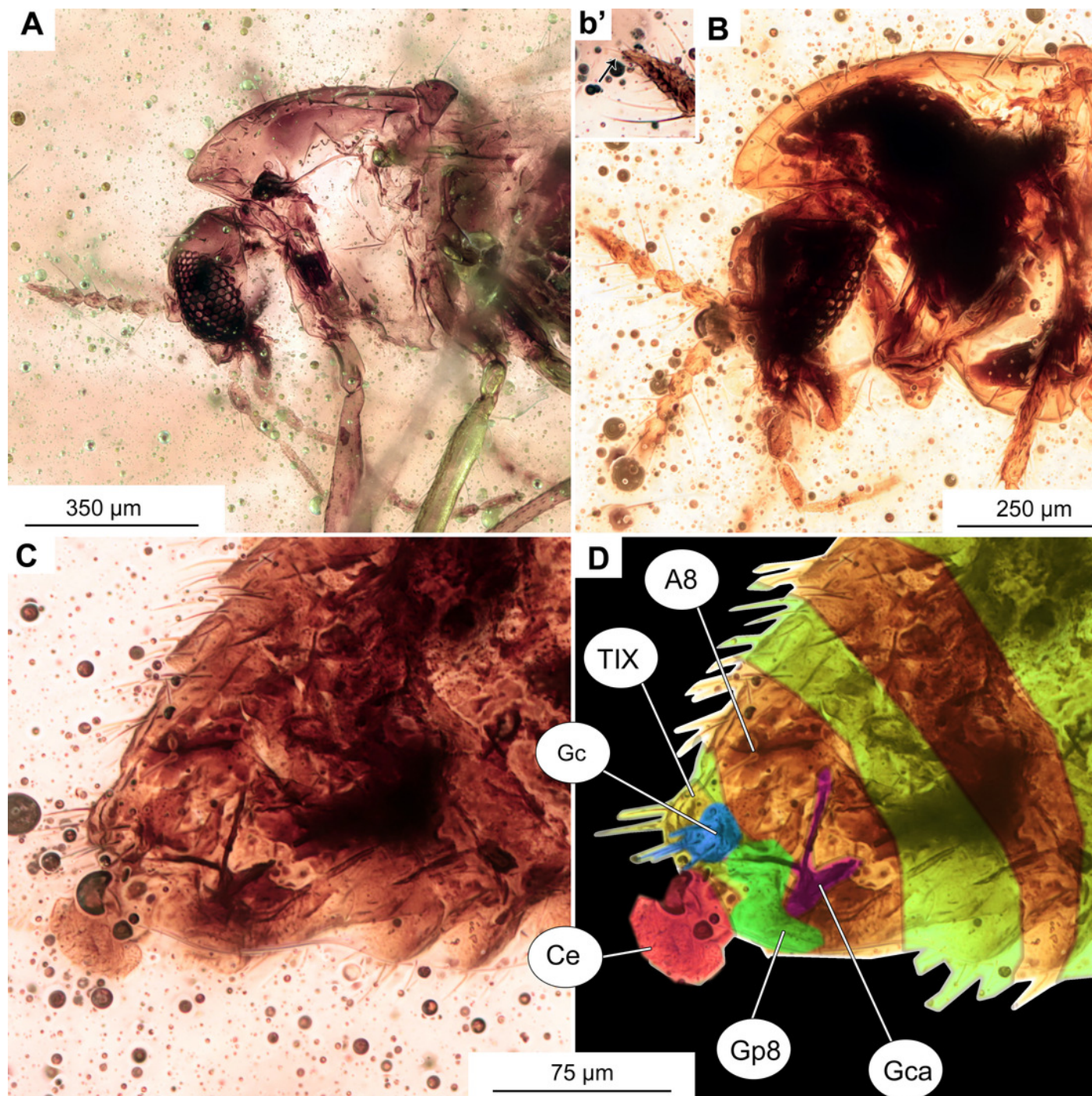


# Figure 12

Figure 12 *Gymnometriocnemus extinctus* sp. nov. Baranov, female paratypes.

(A) Head of specimen OU47577. (B) Head of specimen OU47578; b' close-up of last flagellomere with apical setae. (C) Female genitalia (OU47578). (D) Female genitalia, marked (OU47578). Abbreviations: A8, abdominal segment 8; Ce, cerci; Gca, gonocoxite apodem; Gc, gonocoxite (8); Gp8, gonapophysis 8; TIX, tergite 9.







# Figure 13

Figure 13. Holotype (adult male, number NZAC02044947) of *Kuschelius dentifer* Sublette and Wirth, 1980.

(A). Wing. (B) Head, arrow marks a apical protrusion of the 3d palpomere. (C) Hypopigium. (D) Midtibia with spurs and the comb. (E) Hindtibia with the spurs and comb. All photos in this plate are made by Dr. Leanne Elder, licensed under CC BY 4.0 and used with the photographer's explicit permission.

