

1 The sex lives of ctenophores: the influence of light, body size, and self-fertilization on the reproductive
2 output of the sea walnut, *Mnemiopsis leidyi*

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14 **Abstract**

15 Ctenophores (comb jellies) are emerging as important animals for investigating
16 fundamental questions across numerous branches of biology (e.g., evodevo, neuroscience, and
17 biogeography). Several ctenophore species including, most notably, *Mnemiopsis leidyi*, are
18 known as invasive species, adding to the importance of studying the ecology of these animals.
19 Despite the growing interest, relatively little is known about ctenophore reproduction. Like most
20 ctenophores, *M. leidyi* is a simultaneous hermaphrodite capable of self-fertilization. In this study,
21 we assess the influence of light on spawning, the effect of body size on spawning likelihood and
22 reproductive output, and the cost of self-fertilization on egg viability in *M. leidyi*. Our results
23 suggest that *M. leidyi* spawning is more strongly influenced by circadian rhythms than specific
24 light clues, and that body size significantly impacts spawning and reproductive output. We also
25 find a lower percentage of viable embryos from *M. leidyi* that were spawned alone versus those
26 that were spawned in pairs, suggesting that self-fertilization may be costly in these animals.
27 These results provide critical insight into the reproductive ecology of these ctenophores and
28 provide a fundamental resource for researchers working with *M. leidyi* in the laboratory.

29 **Introduction**

30 Ctenophores (comb jellies) are fascinating planktonic animals most easily recognized by
31 eight rows of cilia that they use as their primary means of locomotion. Recent work suggests
32 ctenophores are the sister group to the rest of all animals and therefore are especially informative
33 as to the state of the most recent common ancestor of animals (Dunn et al. 2008; Hejnol et al.
34 2009; Ryan et al. 2013; Borowiec et al. 2015; Chang et al. 2015; Whelan et al. 2015) but see
35 (Pisani et al. 2015). This phylogenetic position, the availability of nuclear and mitochondrial

Comment [R21]: There are few ctenophore species which are known as invasive species, not several. For many species the distribution is not well known, and thus also the status of invasive species is also not well known. I would be careful with the wording

36 genome sequences (Pett et al. 2011; Ryan et al. 2013), and the ease with which embryos can be
37 collected and observed (Pang & Martindale 2008b) has made the ctenophore *Mnemiopsis leidyi*
38 (Fig. 1) an emergent model system in which to study animal evolution and development (Pang &
39 Martindale 2008a). In addition, since the invasion of *M. leidyi* into European waters from its
40 native range on the Atlantic seaboard (Vinogradov et al. 1989; Reusch et al. 2010) has had
41 profound impacts on European fisheries (Kideys 2002; Oguz, Fach & Salihoglu 2008; Finenko et
42 al. 2013), interest is high in the biogeography and invasion dynamics of *M. leidyi*. Despite the
43 growing importance and utility of *M. leidyi*, the reproductive ecology of these animals is not very
44 well understood.

45 The reproductive biology and life-history of *M. leidyi* has likely played a major role in its
46 ability to invade and establish populations in foreign waters. *M. leidyi*, like most ctenophores, are
47 simultaneous hermaphrodites that have the ability to self-fertilize and have been observed to
48 produce thousands of eggs a day (Baker & Reeve 1974; Costello et al. 2006; Lehtiniemi et al.
49 2012). Offspring may develop from egg to reproductive adult in as few as 13 days (Baker &
50 Reeve 1974; Costello et al. 2012). *M. leidyi* may even produce viable gametes as juveniles
51 (Martindale 1987).

52 A number of studies have described the spawning behavior of *M. leidyi* (Baker & Reeve
53 1974; Pang & Martindale 2008b). Early research suggested that *M. leidyi* spawns as a response
54 to darkness (e.g., sunset) (Freeman & Reynolds 1973), while more recent protocols have stated
55 that *M. leidyi* use light cues to trigger spawning, readily releasing gametes upon exposure to light
56 after spending at least three to four hours in darkness (Pang & Martindale 2008b).

Comment [R22]: Actually, there is quite a few papers on reproductive ecology of *Mnemiopsis leidyi*: reproduction in relation to temperature, salinity, size and prey density, however, not in relation to light cues, or self-fertilization.

Comment [R23]: Maybe something to add: Jaspers et al. 2014: Carbon content of *Mnemiopsis leidyi* eggs and specific egg production rates in northern Europe. J. Plankton Res.

57 Adult *M. leidy* vary dramatically in body size and this variation can affect both the
58 likelihood to spawn and the number of eggs produced (Baker & Reeve 1974; Finenko et al.
59 2006). Animals are more likely to spawn as they grow larger (Baker & Reeve 1974) and larger
60 animals generally produce more eggs per day (Baker & Reeve 1974; Finenko et al. 2006).
61 However, the threshold size before spawning begins has varied from 15mm (Finenko et al. 2006)
62 to 32mm (Baker & Reeve 1974) across studies. It is unclear whether this wide variation in initial
63 spawning size is due to population-specific differences, seasonal timing, or other factors.

64 While self-fertilization may provide the benefit of allowing *M. leidy* to reproduce when
65 conspecifics are not present, it may also come with the cost of inbreeding depression. Inbreeding
66 depression has been shown to affect the viability of offspring in many systems (Charlesworth &
67 Charlesworth 1987; Crnokrak & Roff 1999; Herlihy & Eckert 2002). Rates of self-fertilization
68 and inbreeding depression may be especially high in recently established populations where the
69 population size and genetic diversity are low. Thus, establishing the degree to which self-
70 fertilization is costly in *M. leidy* has particular significance for the management of areas where
71 these ctenophores are invasive. However, to our knowledge, the costs associated with self-
72 fertilization in *M. leidy* have never been thoroughly investigated.

73 In this study, we aim to describe the reproductive cues, effect of body size on spawning,
74 and potential costs of self-fertilization in *M. leidy*. We first investigate spawning cues by placing
75 individuals under different light regimens. We then describe how body size influences spawning
76 likelihood, egg production, and egg viability. Finally, we test whether self-fertilization in *M.*
77 *leidy* is costly by comparing the viability of eggs from ctenophores spawned individually to
78 those spawned with a partner. If self-fertilization is costly, we predict that the offspring of *M.*
79 *leidy* spawning alone will have lower viability than those spawned in groups. Taken together,

Comment [R24]: Smaller sizes has been detected from the European side e.g. Jaspers C, Møller LF, Kiørboe T (2011) Salinity Gradient of the Baltic Sea Limits the Reproduction and Population Expansion of the Newly Invaded Comb Jelly *Mnemiopsis leidy*. PLoS ONE 6(8): e24065. doi:10.1371/journal.pone.0024065

Comment [R25]: Maybe adding a small table showing some results of the previous studies and in which conditions they were recorded? Salinity, temperature, prey density and starvation are known to effect on reproduction of *M. leidy* as well as population specific differences or then a sentence of two demonstrating some of the studies done in relation to other factors.

Comment [R26]: Reference?

80 this study will provide a detailed description of the reproductive ecology of *M. leidy*, supply
81 critical information for studying the invasive impact of these ctenophores, and become a pivotal
82 resource for establishing *M. leidy* as a model system in the laboratory.

83 **Materials & Methods**

84 *Collection*

85 We carefully collected a total of 218 *M. leidy* from the surface waters of Port Orange and
86 St. Augustine, FL using a cteno-dipper (beaker on a stick) and transported them in buckets to the
87 Whitney Laboratory for the Marine Biosciences in St. Augustine, FL between June and October
88 2015. Upon arrival, the ctenophores were transferred first to a large beaker with filtered sea
89 water and then placed in 4" diameter circular glass dishes filled with 250 mL of filtered sea
90 water. We labeled each bowl with a unique identification number and measured the polar length
91 of every ctenophore along the oral/aboral axis to the nearest mm using calipers. Most
92 ctenophores were released after spawning although a few were used for DNA and RNA
93 extraction.

94 *Light effects on spawning and egg production*

95 We tested the protocol described in Pang and Martindale (2008b) using a subset of 64 *M.*
96 *leidy* that we had collected that day (N = 25) or collected and kept overnight in a large kreisel
97 aquarium (N = 39). We did not monitor animals for spawning while they were in the kreisel.
98 Between the hours of 10:00 and 18:00, we placed these animals in dishes in the dark for three to
99 four hours. Upon exposure to light, bowls were monitored over the next two hours for the
100 presence of eggs.

Comment [R27]: I'm missing more detailed information of the sampling e.g. temperature, salinity, light regimes...

Comment [R28]: Mesh size?

Comment [R29]: How long after collection? With or without food? Starvation is known to affect the size of *Mnemiopsis*

Comment [R210]: Exact number? No results shown? Relevance for this study?

Comment [R211]: When were these specimens collected? From what kind of light conditions?

Comment [R212]: Where they fed?

Comment [R213]: Why not?

Comment [R214]: Size of the dishes?

Comment [R215]: Any controls?

101 We conducted a separate set of experiments to test the importance of light cues for
102 spawning on a subset of the *M. leidy* that we had collected from Port Orange (N=66). On the day
103 of collection, we separated each ctenophore into individual 4" diameter bowls filled with 250 mL
104 of filtered seawater and haphazardly assigned individuals to one of four treatments: A) constant
105 light (N = 21), B) 11 hours of light and then four hours of darkness (N = 15), C) seven hours of
106 light and then eight hours of darkness (N = 12), or D) constant darkness (N = 18). All treatments
107 began at 18:00 and ended at 9:00 the next day, at which point we exposed all of the animals to
108 light and immediately recorded whether eggs were present in each bowl.

109 *Size effects on spawning, egg production, and egg viability*

110 In many systems, body size strongly influences reproductive output. We designed an
111 experiment to test the effect of body size on spawning likelihood, egg production, and offspring
112 viability. We tested the effect of size on spawning likelihood using the ctenophores already
113 spawned in the previous light cues experiment (N=66) and an additional 52 *M. leidy* (total N =
114 118) that we collected. We measured the length of every ctenophore along the oral/aboral axis to
115 the nearest mm using calipers and then placed each in their own bowl with 250 mL of filtered
116 seawater. We left the additional 52 animals that had not already been spawned overnight in either
117 constant darkness (N = 26) or in a room with no artificial lights and an uncovered window to
118 experience natural changes in light (N = 26). We immediately recorded whether eggs were
119 present in each bowl on the following morning at 9:00. Since *M. leidy* typically spawn hundreds
120 of eggs, we only considered bowls with at least 15 eggs as having a true spawn. We calculated
121 the effect of size on spawning likelihood using logistic regression and visualized the data with a
122 cubic spline.

Comment [R216]: When? What kind of light conditions?

Comment [R217]: Mesh size?

Comment [R218]: Were these specimens fed?

Comment [R219]: Size distribution?

Comment [R220]: Size distribution

Comment [R221]: How much prior the experiment?

Comment [R222]: Mesh size?

Comment [R223]: Confusing, what was done to the other 66 specimens? Did you use the egg numbers from the previous light cues experiment and the measured size after the experiment?

123 To collect the eggs of the ctenophores that spawned, we poured the water and eggs from
124 each bowl through a 70- μ m filter. The eggs of each ctenophore were then pipetted into separate
125 2" diameter bowls filled with filtered seawater. Eggs were allowed to settle in the bowl before
126 we counted eggs.

Comment [R224]: Mesh size?

127 A number of the ctenophores produced thousands of eggs, making a direct count of all
128 eggs difficult. To address this challenge, we developed a protocol to allow us to estimate the
129 number of eggs in each 2" bowl. We drew a 2" diameter circle and placed a square within the
130 circle so that each point on the square touched the edge of the circle (Fig. 2). Finally, we divided
131 the square into eight equal sized triangles that we labeled 1 – 8. For each ctenophore, we counted
132 the number of eggs in two randomly selected triangles. Two triangles comprise 15.91% of the
133 total area of the circle, and so to estimate the total number of eggs in the dish we multiplied the
134 combined egg count by 6.285. Estimated egg production was log-transformed to increase
135 normality. We then evaluated the correlation between body size and estimated egg production
136 using linear regression for the individuals that spawned (N = 30). The reason the eggs from more
137 *M. leidy* spawns were not counted is because we developed the counting method halfway
138 through the study.

Comment [R225]: Did you test that 2 was enough to detect the variability?

Comment [R226]: What is the total number of spawns?

139 To determine egg viability, we recounted the number of eggs in each dish after 24 hours.
140 *M. leidy* typically develop into juvenile cydippids within 18-24 hours after fertilization
141 (Martindale & Henry 2015). Juveniles can easily be distinguished from undeveloped eggs due to
142 ciliary movement, and since viable embryos can swim away from their original triangle into the
143 water column, we counted the number of undeveloped eggs in the same triangles as in the egg
144 production assay. We then estimated the number of undeveloped eggs in the entire dish using
145 the method described above. Using this estimate we calculated the percent of undeveloped eggs

Comment [R227]: In similar conditions? Temperature?

146 (estimated undeveloped eggs / estimated total eggs) and subtracted that number from one to
147 determine the percentage of viable eggs. We used linear regression to assess the effect of body
148 size on egg viability (N = 30).

149 *Costs of self-fertilization*

150 If self-fertilization is costly, we would expect *M. leidyi* that were spawned alone to have
151 reduced offspring viability compared to those that were spawned in pairs. To test for such a cost,
152 80 *M. leidyi* were randomly placed by themselves or with another individual in a 4” diameter
153 bowl with 250 mL of filtered seawater. Individuals were spawned overnight and the next day we
154 estimated the number of eggs present in each bowl and the percent of viable offspring 24 hours
155 later (see above). We compared estimated egg production and egg viability from ctenophores
156 spawned alone (N = 30 for egg production, N = 29 for egg viability) to ctenophores spawned in
157 pairs (N = 25) using Student’s t-test.

158 All statistical analyses were run in JMP 11.0 (SAS Institute, Cary, NC).

159 **Results**

160 *Spawning cues*

161 Following the recent spawning protocol (Pang & Martindale 2008b), we placed *M. leidyi*
162 in the dark for three to four hours between the hours of 10:00 and 18:00 before exposing them to
163 light. After two hours in light, only five of 39 (12.8%) animals had produced any eggs.
164 Furthermore, the few ctenophores that did spawn often released only a few eggs (median = 18
165 eggs, range 12 – 177 eggs).

166 We next tested the role of light cues in *M. leidyi* spawning. We kept ctenophores in
167 individual bowls overnight in four treatments with varied light cycles and checked each bowl for

Comment [R228]: After how many hours?

Comment [R229]: Why one is missing?

Comment [R230]: Method not results, rephrase

168 eggs the following morning. Almost every ctenophore spawned overnight; we found no
169 difference between ctenophores kept in constant light (20/21 [95%] spawned), four hours of
170 darkness (15/15 [100%] spawned), eight hours of darkness (12/12 [100%] spawned), or constant
171 darkness (17/18 [94%] spawned).

Comment [R231]: Methods not results, rephrase

172 *Size effects on spawning and egg viability*

173 As *M. leidyi* grow larger, the likelihood of spawning significantly increases (Fig. 3,
174 Logistic regression, $N = 118$, $\chi^2 = 62.0$, $p < 0.0001$). All but three ctenophores larger than 30mm
175 spawned overnight, while only one ctenophore smaller than 26mm produced eggs.

Comment [R232]: number of eggs? Any difference there?

176 We saw large variation in the number of estimated eggs spawned (range = 25-3934 eggs,
177 median = 484 eggs). Larger individuals generally produced more eggs (Fig. 4, $N = 30$, $r^2 = .38$,
178 $p < 0.001$). We also found a weak but insignificant positive correlation between body size and
179 egg viability (Fig. 5, $N = 29$, $r^2 = 0.12$, $p = 0.07$).

Comment [R233]: What was the total size distribution of all specimens?

Comment [R234]: Was there difference between the specimens used in the light experiment and the specimens collected only for this experiment? Effect of starvation?

180 *Costs of self-fertilization*

181 We compared the egg production between *M. leidyi* that spawned alone ($N = 30$) with *M.*
182 *leidyi* that spawned in pairs ($N = 25$). We found no difference between treatments in the
183 estimated number of eggs produced (Fig. 6, Student's t-test, t-ratio = 0.005, $p = 1.0$). However,
184 we did find that a higher percentage of offspring from individuals that spawned in pairs ($N = 25$)
185 had developed after 24 hours when compared with individuals that spawned by themselves ($N =$
186 29 , Fig. 7, Student's t-test, t-ratio = 2.3, $df = 52$, $p = 0.025$).

187 **Discussion**

188 The ctenophore *Mnemiopsis leidyi* has become an emerging model from which to study
189 evolution and development, especially for understanding early animal evolution (Pang &
190 Martindale 2008a). Additionally, the invasion of *M. leidyi* in European waters has had
191 devastating impacts on fisheries (Shiganova 1998; Kideys 2002; Finenko et al. 2013) and has led
192 to strong interest in these animals. Understanding *M. leidyi* reproductive ecology is a necessary
193 step in establishing it as an important model in the laboratory and may allow for improved
194 management of these animals in afflicted areas.

Comment [R235]: Repetition from the intro

195 Previous work has suggested that *M. leidyi* uses light cues to induce spawning (Freeman
196 & Reynolds 1973; Pang & Martindale 2008b; Martindale & Henry 2015); however, our attempts
197 at replicating this spawning cue failed. Instead, we found that almost every *M. leidyi* over a
198 certain size spawned overnight regardless of the light/dark cycle; even those individuals that
199 were placed under constant light consistently spawned. This result suggests that *M. leidyi* spawns
200 using a circadian rhythm rather than specific light cues, at least when initially brought into the
201 lab. Sequencing data indicate that the *M. leidyi* genome contains a number of orthologs involved
202 in animal circadian rhythm including *Clock* and *ARNTL*. These and other circadian rhythm
203 genes have been associated with reproduction and reproductive timing in a number of systems
204 (Boden & Kennaway 2006; Leder, Danzmann & Ferguson 2006; Liedvogel et al. 2009).
205 Functional genomic analyses into how these circadian-rhythm genes affect spawning could
206 potentially provide solid evidence linking circadian rhythms and *M. leidyi* spawning. Given the
207 phylogenetic position of ctenophores as the sister lineage to the rest of animals (Dunn et al.
208 2008; Ryan et al. 2013; Borowiec et al. 2015; Chang et al. 2015; Whelan et al. 2015), such a
209 study would also address to what extent the genetic circuitry underlying animal circadian rhythm
210 was present in the last common animal ancestor.

Comment [R236]: Difference in number of eggs produced?

Comment [R237]: Reference?

Comment [R238]: Add: but see (Pisani et al. 2015)

211 Previous spawning protocols were described for *M. leidy* populations near Woods Hole,
212 Massachusetts (Pang & Martindale 2008b). To our knowledge, spawning protocols have not
213 previously been described for *M. leidy* in the Atlantic waters of northern Florida. While these
214 two *Mnemiopsis* populations had previously been classified as a separate species (Massachusetts
215 = *Mnemiopsis leidy*, Agassiz 1865, northern Florida = *Mnemiopsis mccradyi* Mayer, 1900), they
216 are now generally considered to be separate populations of the same species (Pang & Martindale
217 2008a; Bayha et al. 2015), although this has yet to be extensively tested genetically. Populations
218 within species may differ in their reproductive timing or cues (e.g. Partecke, Van't Hof &
219 Gwinner 2004; Moore, Bonier & Wingfield 2005) and so it could be that the spawning behavior
220 we observed is unique to the northern Florida population of *M. leidy*. Alternatively, spawning
221 behavior could change across seasons with changes to day length or water temperature.

222 Body size plays an essential role in ctenophore reproduction. Spawning occurs almost
223 exclusively in larger *M. leidy* (>30mm), although a few individuals smaller than 30mm spawned
224 and a few animals larger than 40mm did not spawn (Fig. 3). Interestingly, this result differs from
225 *M. leidy* reproduction in the Caspian Sea where individuals begin spawning at 15 mm and the
226 most common size of spawning individuals is between 20 and 30 mm (Finenko et al. 2006). Why
227 these populations differ in size of reproduction is unclear, but they may be influenced by water
228 temperature, resource abundance, or the low salinity of the Caspian Sea (Finenko et al. 2006).
229 The differences in the non-native *M. leidy* might also be a result of selection for body size or age
230 of reproductive maturity due to selective pressures imposed by ship-ballast transport.

231 Not surprisingly, larger individuals in our study produced more eggs than smaller
232 individuals (Fig. 4). Body size may correspond to nutritional status rather than age (Reeve, Syms
233 & Kremer 1989) and so larger ctenophores may simply be those well fed enough to produce

Comment [R239]: Example of *Mnemiopsis* in the Baltic Jaspers C, Møller LF, Kjørboe T (2011) Salinity Gradient of the Baltic Sea Limits the Reproduction and Population Expansion of the Newly Invaded Comb Jelly *Mnemiopsis leidy*. PLoS ONE 6(8): e24065. doi:10.1371/journal.pone.0024065

Comment [R240]: Examples? References?

234 gametes. The production of gametes is costly (Hayward & Gillooly 2011) and smaller
235 ctenophores preferentially allocate resources to somatic growth rather than gamete production
236 (Reeve, Syms & Kremer 1989). Since larger individuals consume more prey (Bishop 1967;
237 Finenko et al. 2006) they likely have more resources available to produce eggs than smaller
238 individuals.

239 Body size may also affect offspring viability. We found that the percentage of developed
240 eggs after 24 hours increased as individuals grew larger (Fig. 5), although this result was
241 marginally not significant. If body size truly does affect offspring viability it may be due to
242 sperm volume. If sperm are limited, especially in small individuals, larger animals may simply
243 have more sperm available to fertilize eggs. Alternatively, larger animals may provision more
244 resources to their eggs than smaller animals, which may increase egg viability or development
245 speed. This possibility could be tested by comparing the size of eggs across body sizes.

246 Most ctenophores are simultaneous hermaphrodites with the ability to self-fertilize
247 (Martindale & Henry 2015), but it is unknown whether self-fertilization is costly in these
248 animals. Self-fertilization may lead to inbreeding depression which has been shown to have a
249 suite of negative effects, such as reduced fecundity or viability, in many systems (Charlesworth
250 & Charlesworth 1987; Crnokrak & Roff 1999; Herlihy & Eckert 2002). We have shown that *M.*
251 *leidyi* individuals spawning alone had a lower percentage of developed offspring after 24 hours
252 than ctenophores that spawned in pairs (Fig. 7). What contributes to this apparent cost to self-
253 fertilization is unclear. It could be that spawning pairs simply fertilize more eggs than individuals
254 spawning alone, which could occur if sperm are limited. Another possibility could be that the
255 percentage of eggs fertilized did not differ between treatments but that fewer fertilized eggs
256 developed for individuals spawning alone. Although we did not differentiate between

Comment [R241]: Repetition from the introduction

257 unfertilized eggs and non-developing embryos in this study, we did commonly observe embryos
258 that appeared to have arrested development after only a few stages of cell division. This
259 possibility is consistent with a reduction in offspring viability due to inbreeding depression.

Comment [R242]: From single ones or from both?

260 Interestingly, ctenophores in pairs did not produce more eggs than those spawning alone
261 (Fig. 6). The average size of the ctenophores did not differ between treatments, suggesting that,
262 when paired, ctenophores either reduce the number of eggs spawned or only one of the two
263 ctenophores spawned eggs. This latter option, referred to as egg-trading, may indicate the
264 intriguing possibility that ctenophores alternate between releasing sperm and eggs when in pairs
265 or groups. Egg-trading has been reported in other simultaneously hermaphroditic systems
266 including sea slugs, tobacco fish, and polychaetes (Leonard & Lukowiak 1984; Sella 1985;
267 Petersen 1995). This behavior could be used to reduce the chance of self-fertilization in *M.*
268 *leidyi*. However, the underlying assumption of egg-trading is that individuals spawn with the
269 same partners multiple times; we would not expect this to be the case in *M. leidyi* under natural
270 circumstances since movement is largely governed by water flow.

271 Our results also suggest that individuals may be more efficient when spawning alone than
272 with others. Despite the reduced percentage of developing eggs, more viable offspring were
273 produced per individual when spawned alone than when paired. However, we only spawned each
274 ctenophore once. Individuals spawning alone may require a longer refractory period for
275 gametogenesis before spawning again than paired individuals that alternate between releasing
276 eggs and sperm. Comparing the reproductive output and viability between paired and single
277 individuals over multiple days could provide more resolution on the costs associated with self-
278 fertilization.

Comment [R243]: Really? Even you: "find a lower percentage of viable embryos from *M. leidyi* that were spawned alone versus those that were spawned in pairs, suggesting that self-fertilization may be costly in these animals"

279 The ability to self-fertilize almost certainly enhances the capability of ctenophores to
280 spread when invading new areas. However, the costs to self-fertilization that we've demonstrated
281 may at least slow down their invasive capabilities. These costs may be especially high at the
282 initial stages of an invasion when population numbers and genetic diversity are low. Our self-
283 fertilization experiment only examined one stage of development (i.e., 24 hours after spawning)
284 in one generation and yet we still found evidence that self-fertilization is costly. Additional costs
285 likely do not appear until later in life or after multiple generations of self-fertilized offspring. An
286 experiment investigating the multi-generational effects of self-fertilization may provide a clearer
287 picture of the hurdles, or lack of hurdles, *Mnemiopsis* faces when initially invading a new area.

288 **Conclusions**

289 Due to their evolutionary position as sister taxa to all other animals (Ryan et al. 2013),
290 ctenophores in general, and *M. leidy* in particular, are quickly emerging as new model systems
291 from which to understand evolution, development, regeneration, and even human disease (Pang
292 & Martindale 2008a; Maxwell et al. 2014). Understanding the reproductive ecology of
293 ctenophores is a necessary step in establishing these animals as tractable models for these areas
294 of research. This study has reinforced the importance of body size in *M. leidy* reproduction and
295 has provided the first suggestions that self-fertilization may be costly in ctenophores. However,
296 ctenophore reproduction in natural systems is still very much a mystery. For example, little is
297 known about how common it is for *M. leidy* to self-fertilize in the wild. We have shown that
298 spawning likely follows a circadian rhythm, which may be a mechanism to increase the odds of
299 out-crossing if all animals spawn simultaneously. If self-fertilization is indeed costly, additional
300 adaptations to increase the chance of out-crossing are likely. This work provides a fundamental

301 resource for researchers working with *M. leidy* in their laboratory, as well as, a jumping-off
302 point from which future studies of *M. leidy* reproductive biology can be launched.

303 **Acknowledgements**

304 We acknowledge Marta Chiodin, Kira Carreira, Leslie Babonis, Bailey Steinworth, and
305 Allison Zwarycz for help with collecting *Mnemiopsis leidy*. We thank Mark Martindale and
306 David Simmons for advice on ctenophore husbandry and spawning.

307

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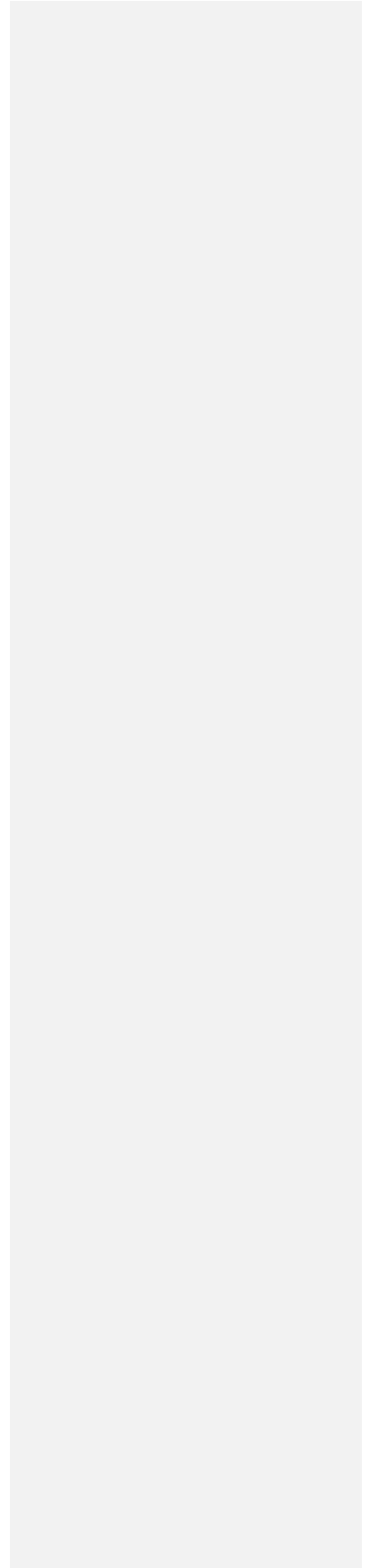


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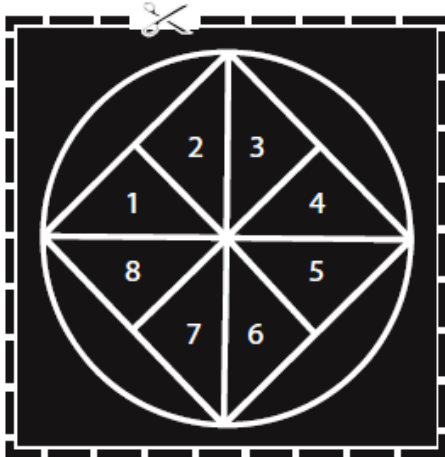
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Fig. 1 *Mnemiopsis leidyi*

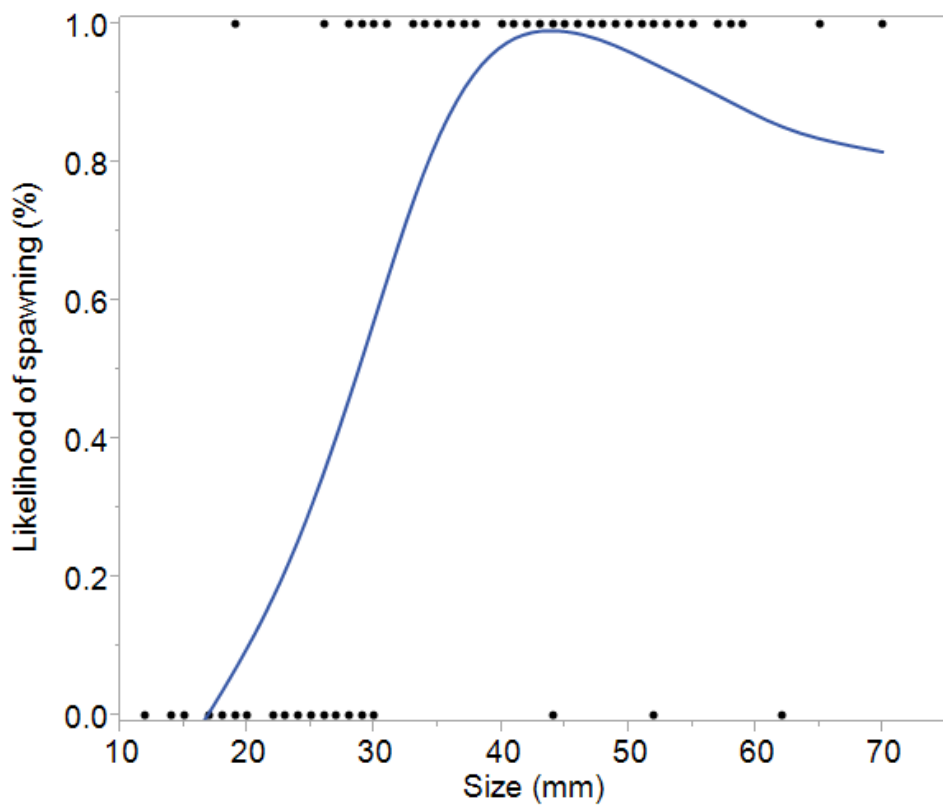


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Fig. 2 The diagram used to estimate egg numbers. Each triangle (labeled 1 – 8) represents 7.96% of the total area of the circle. We counted the eggs in two triangles and then multiplied the total by 6.285 to estimate the total number of eggs in the dish. Scaled to actual size used for 2" bowls.

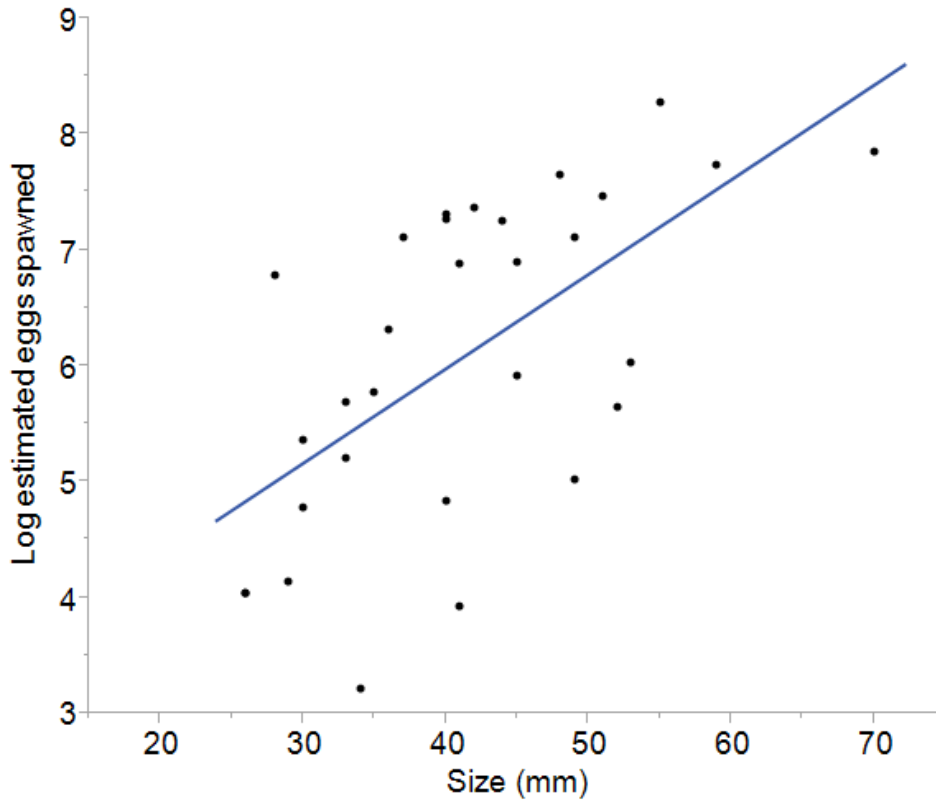


Comment [R244]: Here, I would also like to see the specimens which didn't span (marked with another symbol)

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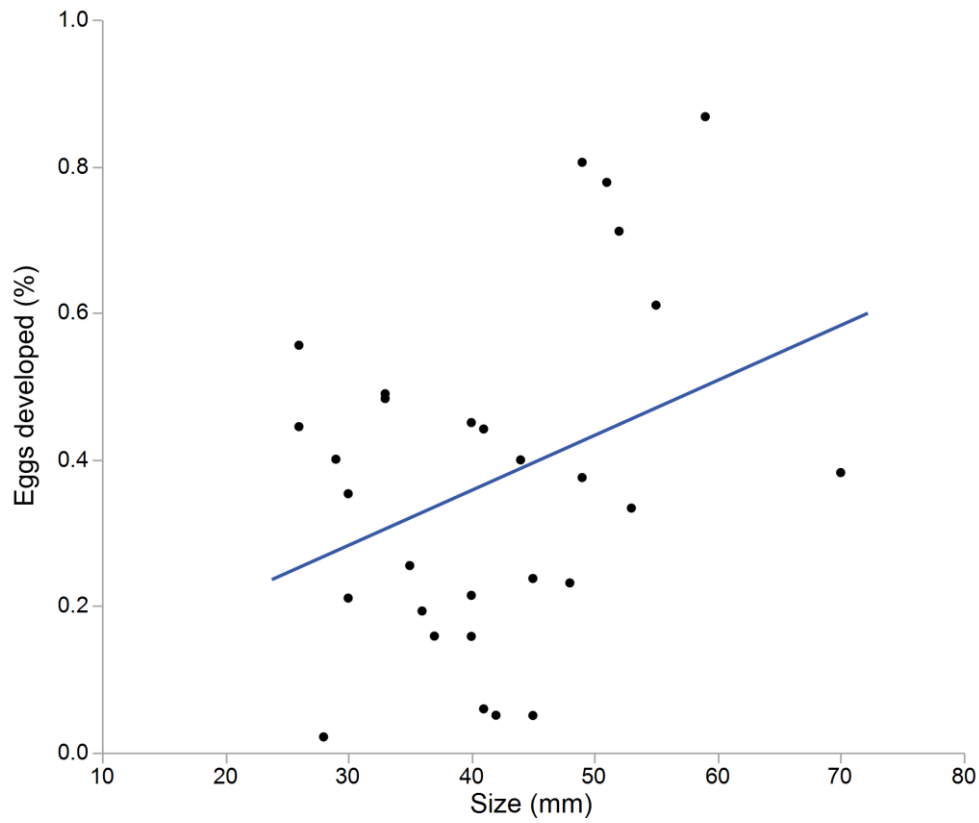
Fig. 3 Cubic spline showing the effect of body size on the likelihood to spawn. Individuals smaller than 26mm rarely spawned while those larger than 30mm almost always spawned. Lambda value of cubic spline set to 1.



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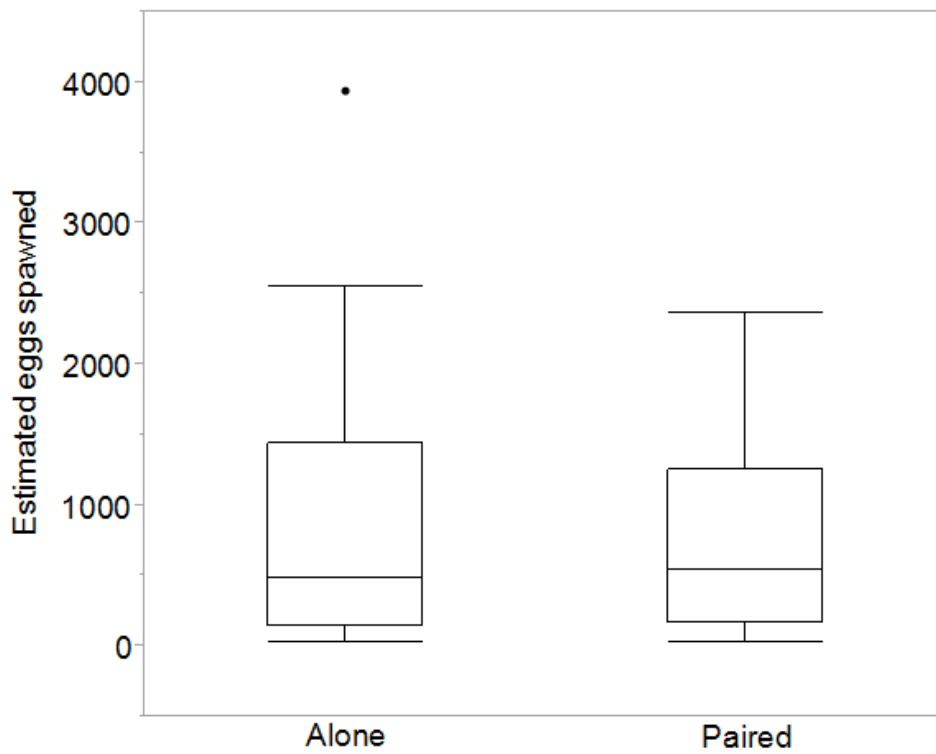
Fig. 4 The effect of body size on egg production. Larger individuals generally produced more eggs than smaller individuals ($N = 30$, $r^2 = .38$, $p < 0.001$). Only those animals that spawned 15 or more eggs are included in the analysis and figure.



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Fig. 5 Correlation between body size and egg viability. Body size positively correlated the percentage of eggs that developed after 24 hours, although the result was marginally not significant ($N = 29$, $r^2 = 0.12$, $p = 0.07$).

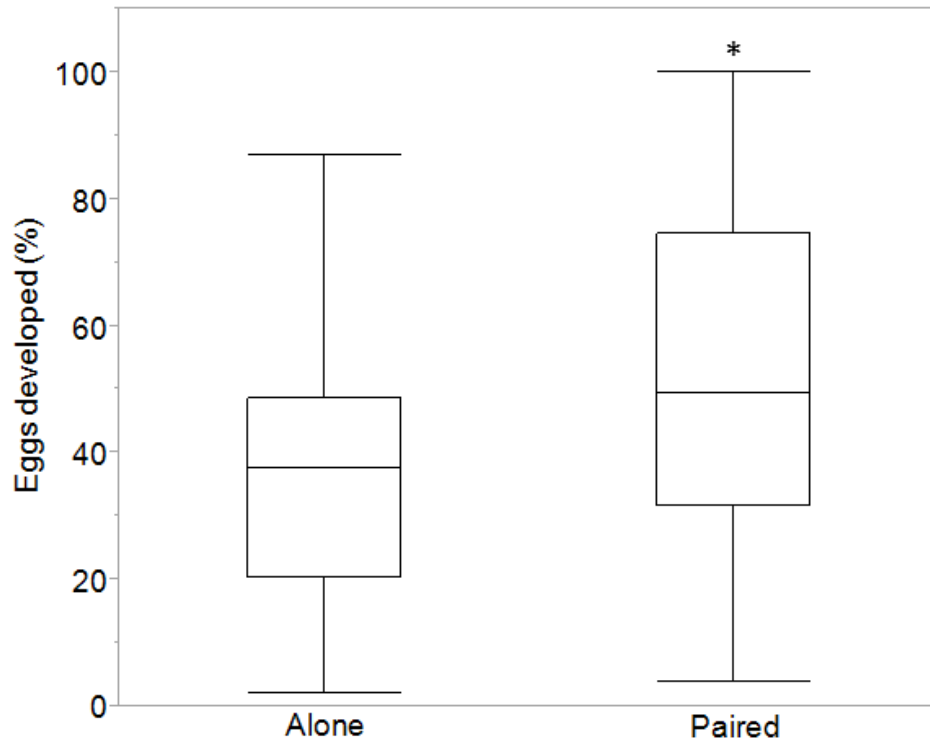


Comment [R245]: Is the number of eggs per individual or per two *Mnemiopsis* when spawning together?

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Fig. 6 The estimated number of eggs spawned for individuals spawning alone (N = 29) and in pairs (N = 25). Surprisingly, two *M. leidy* spawning together did not produce more eggs than individuals spawning alone (Student's t-test, t-ratio = 0.005, p = 1.0). The data point above the Alone box plot indicates an individual that spawned an estimated 3,934 eggs. Removing that data point does not change the overall findings of the analysis.



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Fig. 7 The percentage of eggs developed after 24 hours for individuals spawning alone (N = 29) and in pairs (N = 25). A higher percentage of eggs developed for *M. leidyi* in pairs, possibility suggesting a cost to self-fertilization (Student's t-test, t-ratio = 2.3, df = 52, p = 0.025). Asterisk indicates significant difference across treatments.