

Association of *HMGCR* rs17671591 and rs3761740 with lipidemia and statin response in Uyghurs and Han Chinese

Ziyang Liu^{1,2,3,*}, Yang Zhou^{1,*}, Menglong Jin³, Shuai Liu¹, Sen Liu¹, Kai Yang¹, Huayin Li¹, Sifu Luo¹, Subinuer Jureti¹, Mengwei Wei¹ and Zhenyan Fu¹

¹ The First Affiliated Hospital, Xinjiang Medical University, Urumqi, Xinjiang, China

² Xinjiang Medical University, Urumqi, Xinjiang, China

³ State Key Laboratory of Pathogenesis, Prevention and Treatment of High Incidence Diseases in Central Asia, Xinjiang Medical University, Urumqi, Xinjiang, China

* These authors contributed equally to this work.

ABSTRACT

Background: Dyslipidemia plays a very important role in the occurrence and development of cardiovascular disease (CVD). Genetic factors, including single nucleotide polymorphisms (SNPs), are one of the main risks of dyslipidemia. 3-hydroxy-3-methylglutaryl-CoA reductase (*HMGCR*) is not only the rate-limiting enzyme step of endogenous cholesterol production, but also the therapeutic target of statins.

Methods: We investigated 405 Han Chinese and 373 Uyghur people who took statins for a period of time, recorded their blood lipid levels and baseline data before and after oral statin administration, and extracted DNA from each subject for SNP typing of *HMGCR* rs17671591 and rs3761740. The effects of *HMGCR* rs17671591 and rs3761740 on lipid levels and the effect of statins on lipid lowering in Han Chinese and Uyghur ethnic groups were studied.

Results: In this study, for rs17671591, the CC vs. TT+CT model was significantly correlated with the level of LDL-C before oral statin in the Uyghur population, but there were no correlations between rs17671591 and the level of blood lipid before oral statin in the Han population. The CC vs. TT+CT and CT vs. CC+TT models were significantly correlated with the level of LDL-C after oral statin in the Uyghur population. There was no significant correlation between rs3761740 with blood lipids before and after oral statin in the Han population. For rs3761740, before oral statin, the CC vs. AA+CA model was significantly correlated with the level of LDL-C, and the CA vs. CC+AA model was significantly correlated with the level of total cholesterol (TC), low density lipoprotein cholesterol (LDL-C), and non-high density lipoprotein cholesterol (HDL-C) in the Uyghur population. After oral statin, the CC vs. AA+CA and CA vs. CC+AA models were significantly correlated with the level of TC, LDL-C, and apolipoprotein (APOB), and the C vs. A model was significantly correlated with the level of TC, triglyceride (TG), LDL-C, and APOB in the Uyghur population. Particularly, the CT vs. CC+TT model of rs17671591 was significantly correlated with the changes of LDL-C after oral statin in the Uyghur population. In this study, we also explored the association of rs17671591 and rs3761740 with the rate of dyslipidemia as a reference.

Submitted 19 January 2024
Accepted 30 August 2024
Published 27 September 2024

Corresponding author
Zhenyan Fu,
fuzhenyan316@126.com

Academic editor
Efe Sezgin

Additional Information and
Declarations can be found on
page 14

DOI 10.7717/peerj.18144

© Copyright
2024 Liu et al.

Distributed under
Creative Commons CC-BY 4.0

OPEN ACCESS

Conclusion: We found that *HMGCR* rs3761740 was correlated with the levels of TC, LDL-C, and non-HDL-C before and after oral statin in Uyghurs, but not with blood lipid levels in the Han population. In the Uyghur population, *HMGCR* rs17671591 was associated with the level of LDL-C before and after oral statin, and also affected the changes of LDL-C after oral statin.

Subjects Genetics, Cardiology, Medical Genetics

Keywords Lipid, *HMGCR*, Statin, Statin resistance, China, SNP, Single nucleotide polymorphisms, Multi-ethnic population, Cardiovascular disease, Statin response

INTRODUCTION

Cardiovascular disease (CVD) is the primary cause of human death, and constitutes significant health and economic burdens worldwide (*Tsao et al., 2023; Ralapanawa & Sivakanesan, 2021; Erbel et al., 2014*). Coronary heart disease kills more than 7.96 million people worldwide in 2006 and 9.48 million in 2016 (*GBD 2016 Causes of Death Collaborators, 2016*). CVD is a complex disease that involves multiple mechanisms and cell types, and is affected by many risk factors such as diabetes, hyperlipidemia, smoking, hypertension, lack of exercise, obesity, and heredity (*Visseren et al., 2021*). Dyslipidemia plays a very important role in the occurrence and development of CVD (*Arvanitis & Lowenstein, 2023; O'Malley et al., 2020*).

Dyslipidemia is a disease characterized by a series of lipid metabolic disorders, such as those involving abnormally elevated plasma levels of lipid and lipoprotein dysfunction (*Wang et al., 2018; Ahmad & Leake, 2019*). Dyslipidemia is mainly characterized by an increase in plasma total cholesterol (TC), low density lipoprotein cholesterol (LDL-C), triglyceride (TG), and a decrease in high density lipoprotein cholesterol (HDL-C) (*Smith, 2019; Xiao et al., 2019*). Dyslipidemia is a major risk factor for CVD according to data from 2009 to 2012, with more than 100 million American adults age 20 and older having total cholesterol levels of 200 mg/dL (5.17 mol /L) or higher, and nearly 31 million having total cholesterol levels of 240 mg/dL (6.20 mol /L) or higher (*Mozaffarian et al., 2016*). Genetic factors, including single nucleotide polymorphisms (SNPs), are a major risk factor for dyslipidemia (*Liu et al., 2017; Lu et al., 2017*).

Mammalian 3-hydroxy-3-methylglutaryl-CoA reductase (*HMGCR*) is an endoplasmic reticulum-localized glycoprotein consisting of a hydrophobic n-terminal domain that spans the cell membrane eight times and a large soluble C-terminal domain that projects into the cytoplasm (*Liscum et al., 1985*). *HMGCR* is a rate-limiting enzyme for cholesterol synthesis and is closely related to plasma cholesterol content (*Luo, Yang & Song, 2020*).

Statins are currently recognized as the primary treatment for lowering blood lipids (*Grundy et al., 2019*). *HMGCR* is both a rate-limiting enzymatic step for endogenous cholesterol production and a therapeutic target for statins, which reduce cholesterol production in the liver (*Istvan & Deisenhofer, 2001*). The reduction of cholesterol in the liver further leads to the upregulation of low-density lipoprotein receptors, which enhances the clearance of TC and LDL-C from the circulation, thereby reducing lipid levels (*Brown & Goldstein, 1997*). However, despite the effectiveness of statins, there is

significant variation in how individual patients respond to them (*Simon et al., 2006*). Pharmacogenetic studies have demonstrated the effect of genetic polymorphism on the wide variability of responses to statins observed in patients (*Reiner, 2014; Schmitz & Drobnik, 2003*). In addition, genome-wide association studies have identified several genetic variants associated with higher or lower responses to statin therapy, primarily in genes associated with cholesterol homeostasis (*Barber et al., 2010*). It has also been reported that *HMGCR* SNPS are associated with lipid levels after statin treatment (*Cuevas et al., 2016*).

HMGCR is the rate-limiting enzyme for cholesterol synthesis and the main target of statin therapy. In this study, *HMGCR* rs17671591 and rs3761740 were selected to evaluate their effects on blood lipid levels and the lipid-lowering effect of statin in Han Chinese and Uyghur populations.

METHOD

Ethical approval of the research protocol

This study was approved by the Ethics Committee of the First Affiliated Hospital of Xinjiang Medical University (number: 220525-06-2305A-Y1) and carried out in accordance with the principles of the Declaration of Helsinki. Each participant received written informed consent, including express permission to conduct DNA analysis and collect relevant clinical data.

Subjects

This is a single-center prospective cohort study of the effects of *HMGCR* rs17671591 and rs3761740 on blood lipid levels before lipid-lowering therapy and statin responsiveness. In this study, patients who were hospitalized in the Heart Center of the First Affiliated Hospital of Xinjiang Medical University from 2008 to 2022 with long-term oral statins were selected, and patients' informed consent and signatures were required. Inclusion criteria: a) Uyghur or Han population aged 30–75; b) first diagnosed with coronary heart disease after admission; c) never took lipid-lowering medication before admission; d) started oral statin (atorvastatin 10 mg/day, rosuvastatin 5 mg/day) lipid-lowering treatment after admission; and e) follow up time ≥ 1 month. Exclusion criteria: a) patients with lost follow-up data; b) patients who adjusted their medication dosage or changed to different types of statins during the follow-up period; c) chronic renal insufficiency; d) fatty liver; e) cirrhosis; f) thyroid disease; or g) patients with genetic sequencing errors. The specific process is shown in [Fig. 1](#). The hospital information platform was used to collect information of patients. Clinical data, blood lipid levels, and biochemical indexes of each patient were collected before oral statin and after oral statin administration for >1 month. The following information was collected: ethnicity, sex, age, blood glucose, alanine aminotransferase (ALT), TC, TG, HDL-C, LDL-C, apolipoprotein A1 (APOA1), apolipoprotein (APOB), lipoprotein a (Lpa), and non-HDL-C, where non-HDL-C = TC-HDL-C. All specimens collected were transported to the Xinjiang CHD VIP Laboratory on dry ice at predetermined intervals. Blood glucose, ALT, TC, TG, HDL-C, LDL-C, APOA1, APOB, and Lpa were all tested by the clinical laboratory of the First Affiliated Hospital of

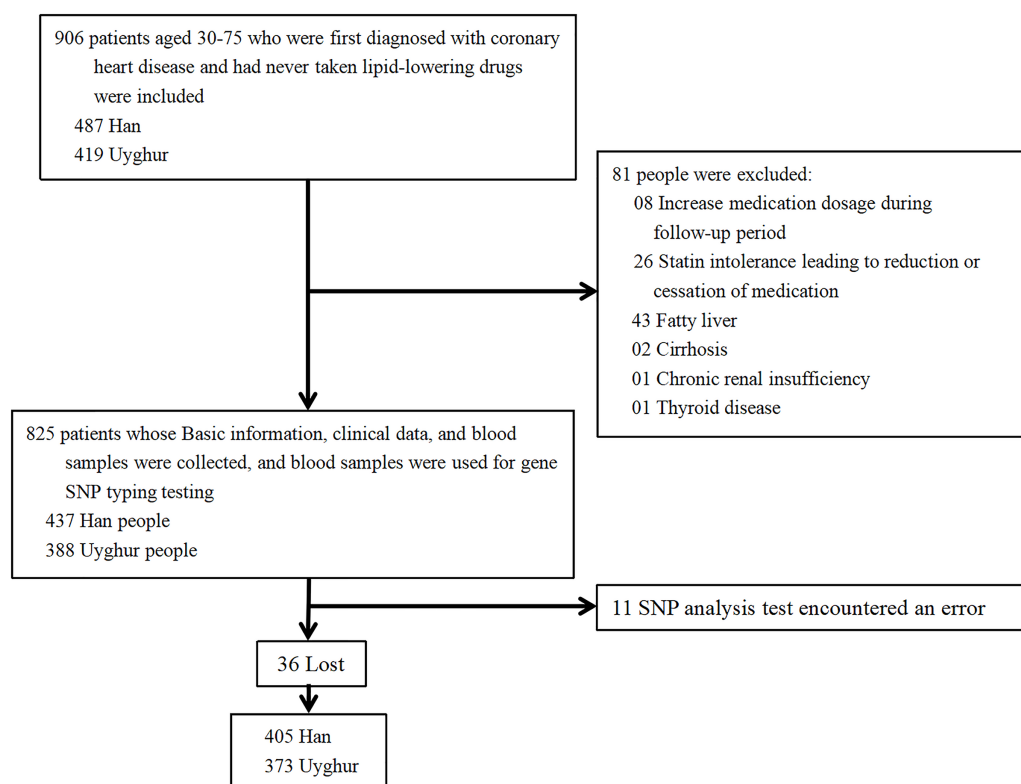


Figure 1 Inclusion and exclusion process of research subjects.

Full-size DOI: 10.7717/peerj.18144/fig-1

Xinjiang Medical University using a biochemical analyzer (Dimension AR/AVL Clinical Chemistry System, Newark, NJ, USA).

Definition of related indicators

According to the 2023 Chinese Lipid Management Guidelines (*Guidelines for the Management of Blood lipids in China, 2023*), high TC was defined as $TC \geq 6.2$ mmol/L, high TG was defined as $TG \geq 2.3$ mmol/L, low HDL-C was defined as $HDL-C < 1$ mmol/L, high LDL-C was defined as $LDL-C \geq 4.1$ mmol/L, low APOA1 was defined as $APOA1 < 1.2$ mmol/L, high APOB was defined as $APOB > 1.1$ mmol/L, high Lpa was defined as $Lpa \geq 300$ mg/L, and high non-HDL-C was defined as $non-HDL-C \geq 4.9$ mmol/L. Changes of lipids ($TC, TG, HDL-C, LDL-C, APOA1, APOB, Lpa, non-HDL-C$) = lipids before oral statin-lipid after oral statin)/(lipids before oral statin).

Genotyping

DNA was extracted using the standard method, and then a custom-by-design 48-Plex SNPscan™ Kit (Cat#:G0104; Genesky Biotechnologies Inc., Shanghai, China) was used for SNP typing. According to the instructions of the kit, DNA underwent denaturation, ligation reaction, and PCR reaction. An ABI3730XL sequencer was used to separate and detect PCR products. The people who carried out genotyping did not know the baseline

data nor other subject indicators. To ensure the quality of genotyping, 4% of the DNA samples were taken again for repeated analysis.

Statistical analysis

SPSS 25.0 was used for statistical analysis. The Hardy-Weinberg equilibrium test was performed by Chi-square test. Continuous variables were expressed as mean \pm standard deviation, and differences between groups were assessed using an independent sample t-test or ANOVA. Comparison of lipid levels before and after statin treatment was analyzed using a paired sample t-test. The rate of categorical variables was expressed and Chi-square test was used to analyze the differences between groups. Multivariate analysis was further adjusted using linear regression models or logistic regression models, because age, sex, and liver function are important factors affecting blood lipids ([Grundy et al., 2019](#)), and adjustment variables included gender, age, and ALT. In addition, a two-tailed *P*-value of less than 0.05 was considered statistically significant.

RESULTS

Individual demographics

A total of 778 subjects were studied, including 405 Han and 373 Uyghur individuals. 75.34% of the Uyghur subjects were male, which was significantly higher than the 61.18% of Han subjects who were male. There were no significant differences in fasting blood glucose, TG, APOB, Lpa, and non-HDL-C between Han and Uyghur subjects before oral statin treatment. The age, TC, LDL-C, HDL-C, and APOA1 of Han subjects before oral statin treatment were significantly higher than those of Uyghur subjects before oral statin treatment. The ALT of Han subjects before oral statin treatment were significantly lower than those of Uyghur subjects before oral statin treatment ([Table 1](#)). In the Han population, oral statin significantly reduced the plasma concentrations of TC, TG, HDL-C, LDL-C, APOA1, APOB, and non-HDL-C, while oral statin did not significantly reduce the plasma concentrations of Lpa. In the Uyghur population, the plasma concentrations of TC, TG, LDL-C, APOA1, APOB, and non-HDL-C were significantly reduced after oral statin, while the plasma concentrations of HDL-C and Lpa were not significantly different before and after oral statin ([Table 2](#)).

Effects of *HMGCR* rs17671591 (SNP1) on blood lipid levels before and after oral statin in Han and Uyghur populations

The distribution of SNP1 genotypes in both Han and Uyghur populations corresponded to the Hardy-Weinberg equilibrium. Before oral statin, there were no correlation between serum lipid levels and SNP1 in the Han population, but there were significant differences in LDL-C levels in the dominant model (CC and CT+TT) in the Uyghur population ([Table S1](#)). Before oral statin, the influence of the SNP1 dominant model (CC and TT +CT) on LDL-C plasma concentration was still statistically significant after multivariate adjustment in the Uyghur population ([Table 3](#)). After oral statin, there was a correlation between plasma APOA1 level and SNP1 in the Han population, and a correlation between plasma LDL-C level and SNP1 in the Uyghur population ([Table S2](#)). After

Table 1 Clinical and metabolic characteristic of subjects.

	Han (<i>n</i> = 405)	Uyghur (<i>n</i> = 373)	<i>P</i>
Sex (male)	249 (61.18%)	281 (75.34%)	<0.001
Age (year old)	60.290 ± 11.020	55.580 ± 8.938	<0.001
Fasting blood glucose (mmol/L)	6.387 ± 2.607	6.426 ± 2.956	0.845
TG (mmol/L)	2.155 ± 1.334	2.295 ± 1.558	0.177
TC (mmol/L)	5.144 ± 0.946	4.911 ± 1.072	0.001
HDL-C (mmol/L)	1.118 ± 0.333	0.962 ± 0.255	<0.001
LDL-C (mmol/L)	3.507 ± 0.752	3.342 ± 0.854	0.004
APOA1 (mmol/L)	1.241 ± 0.276	1.132 ± 0.220	<0.001
APOB (mmol/L)	1.096 ± 0.253	1.061 ± 0.267	0.064
Lpa (mg/L)	237.836 ± 229.412	274.483 ± 289.213	0.051
Non-HDL-C (mmol/L)	4.026 ± 0.952	3.949 ± 1.058	0.284
ALT (U/L)	26.710 ± 20.578	33.310 ± 25.525	<0.001

Note:

A t-test was conducted to generate the *P* values. Abbreviation: TC, total cholesterol; TG, triglycerides; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; APOA1, apolipoprotein A1; APOB, apolipoprotein B; Lpa, lipoprotein a; ALT, alanine aminotransferase.

Table 2 Lipid profile of patients before and after atorvastatin therapy.

Ethnic group	Blood lipid	Before oral statin	After oral statin	<i>P</i>
Han (<i>n</i> = 405)	TG (mmol/L)	2.151 ± 1.342	1.895 ± 1.354	<0.001
	TC (mmol/L)	5.142 ± 0.951	3.873 ± 1.025	<0.001
	HDL-C (mmol/L)	1.122 ± 0.331	1.081 ± 0.292	0.017
	LDL-C (mmol/L)	3.514 ± 0.753	2.352 ± 0.813	<0.001
	APOA1 (mmol/L)	1.241 ± 0.282	1.182 ± 0.253	<0.001
	APOB (mmol/L)	1.100 ± 0.252	0.830 ± 0.254	<0.001
	Lpa (mg/L)	237.703 ± 228.725	245.570 ± 248.344	0.437
	Non-HDL-C (mmol/L)	4.037 ± 0.956	2.798 ± 1.044	<0.001
Uyghur (<i>n</i> = 373)	TG (mmol/L)	2.281 ± 1.522	1.952 ± 1.440	<0.001
	TC (mmol/L)	4.953 ± 1.062	3.992 ± 1.164	<0.001
	HDL-C (mmol/L)	0.992 ± 0.313	0.993 ± 0.282	0.761
	LDL-C (mmol/L)	3.374 ± 0.840	2.524 ± 0.941	<0.001
	APOA1 (mmol/L)	1.151 ± 0.221	1.115 ± 0.276	0.004
	APOB (mmol/L)	1.072 ± 0.267	0.874 ± 0.281	<0.001
	Lpa (mg/L)	265.750 ± 279.740	252.911 ± 270	0.304
	Non-HDL-C (mmol/L)	3.963 ± 1.042	2.992 ± 1.153	<0.001

Note:

A t-test was conducted to generate the *P* values. Abbreviation: TC, total cholesterol; TG, triglycerides; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; APOA1, apolipoprotein A1; APOB, apolipoprotein B; Lpa, lipoprotein a; ALT, alanine aminotransferase.

multivariate adjustment, the influence of the SNP1 allele model (C and T) after oral statin on APOA1 plasma concentration was still statistically significant in the Han population, and the influence of the SNP1 dominant model (CC and CT+TT) and additive model (CT and CC+TT) on LDL-C plasma concentration was still statistically significant in the Uyghur population (Table 3).

Table 3 Association of SNP1 (rs17671591) and SNP2 (rs3761740) with blood lipid level before and after oral statin and after multivariate adjustment.

SNP	Statin	Ethnic	Blood lipids	Gene model		β	P
SNP1	Before oral statin	Uyghur	Blood lipids	CC ($n = 142$)	TT+CT ($n = 231$)	β	P
			LDL-C (mmol/L)	3.220 ± 0.665	3.410 ± 0.945	0.190	0.038
	After oral statin	Han	Blood lipids	CC ($n = 179$)	TT+CT ($n = 226$)	β	P
			APOA1	1.209 ± 0.253	1.156 ± 0.244	-0.048	0.054
			Blood lipids	TT ($n = 36$)	TT+CT ($n = 369$)	β	P
			APOA1	1.220 ± 0.219	1.240 ± 0.281	0.082	0.062
			Blood lipids	C ($n = 548$)	T ($n = 262$)	β	P
			APOA1	1.194 ± 0.247	1.148 ± 0.252	0.041	0.024
		Uyghur	Blood lipids	CC ($n = 142$)	TT+CT ($n = 231$)	β	P
			LDL-C	2.485 ± 0.815	2.704 ± 0.944	0.225	0.02
			Blood lipids	TT ($n = 56$)	TT+CT ($n = 317$)	β	P
			LDL-C	2.485 ± 0.815	2.704 ± 0.944	0.155	0.242
			Blood lipids	CT ($n = 175$)	CC+TT ($n = 198$)	β	P
			LDL-C	2.775 ± 0.923	2.485 ± 0.864	-0.294	0.002
SNP2	Before oral statin	Uyghur	Blood lipids	CC ($n = 326$)	AA+CA ($n = 47$)	β	P
			LDL-C (mmol/L)	3.290 ± 0.726	3.720 ± 1.415	0.431	0.001
			Blood lipids	CA ($n = 44$)	AA+CC ($n = 329$)	β	P
			TC (mmol/L)	5.420 ± 1.712	4.840 ± 0.937	-0.579	0.001
			LDL-C (mmol/L)	3.790 ± 1.440	3.280 ± 0.724	-0.499	<0.001
			Non-HDL-C (mmol/L)	4.439 ± 1.659	3.883 ± 0.933	-0.561	0.001
	After oral statin	Uyghur	Blood lipids	CC ($n = 326$)	AA+CA ($n = 47$)	β	P
			TC (mmol/L)	4.015 ± 1.108	4.532 ± 1.341	0.544	0.002
			LDL-C (mmol/L)	2.564 ± 0.847	3.023 ± 1.159	0.466	0.001
			APOB (mmol/L)	0.881 ± 0.262	1.007 ± 0.311	0.132	0.002
			Blood lipids	CA ($n = 44$)	AA+CC ($n = 329$)	β	P
			TC (mmol/L)	4.576 ± 1.37	4.013 ± 1.104	-0.591	0.001
			LDL-C (mmol/L)	3.105 ± 1.154	2.557 ± 0.846	-0.556	<0.001
			APOB (mmol/L)	1.018 ± 0.312	0.88 ± 0.262	-0.145	0.001
			Blood lipids	C ($n = 696$)	A ($n = 50$)	β	P
			TG (mmol/L)	1.932 ± 1.32	2.373 ± 1.64	-0.486	0.012
			TC (mmol/L)	4.049 ± 1.132	4.493 ± 1.314	-0.468	0.005
			LDL-C (mmol/L)	2.597 ± 0.877	2.952 ± 1.158	-0.358	0.007
			APOB (mmol/L)	0.89 ± 0.267	0.997 ± 0.309	-0.112	0.005

Note: Multivariate analysis was further adjusted using linear regression models and adjustment variables including gender, age, and ALT. The dominant model takes the CC genotype as the reference, the recessive model uses the TT genotype as the reference, and the allele model uses the C allele as the reference. Abbreviation: LDL-C, low-density lipoprotein cholesterol; APOA1, apolipoprotein A1; ALT, alanine aminotransferase.

Effects of *HMGR* rs3761740 (SNP2) on blood lipid levels before and after oral statin in Han and Uyghur populations

The distribution of SNP2 genotypes in both Han and Uyghur populations corresponded to the Hardy-Weinberg equilibrium. No AA genotype was detected in the Han population, but only three individuals with the AA genotype were detected in the Uyghur population.

Therefore, the dominant model (CC and CA) and allele model (C and A) were analyzed in the Han population, and the dominant model (CC and AA+CA), additive model (CA and CC+AA), and allele model (C and A) were analyzed in the Uyghur population. Before oral statin, there was no correlation between the blood lipids and SNP2 in the Han population, and there was a correlation between the LDL-C, TC, non-HDL-C, and SNP2 in the Uyghur population (Table S3). After multi-factor adjustment and before oral statin in the Uyghur populations, the influence of the SNP2 dominant model (CC and AA+CA) on LDL-C was still statistically significant, and the SNP2 additive model (CA and CC+AA) still had statistically significant effects on TC, LDL-C, and non-HDL-C (Table 3). After oral statin, there was no correlation between the blood lipids and SNP2 in the Han population, and there was a correlation between the TC, TG, LDL-C, APOB, and SNP2 in the Uyghur population (Table S4). After multi-factor adjustment and oral statin in the Uyghur populations, the influence of the SNP2 dominant model (CC and AA+CA) and additive model (CA and CC+AA) on TC, LDL-C, and APOB was still statistically significant, and the influence of the SNP2 allele model (C and A) on TG, TC, LDL-C, and APOB was still statistically significant (Table 3).

The effect of *HMGCR* rs17671591 (SNP1) on the rate of dyslipidemia before and after oral statin in Han and Uyghur populations

Before oral statin, there was a correlation between the rate of high TC and SNP1 in the Han population, and between the rates of high TC, low HDL-C, high LDL-C, and high non-HDL-C with SNP1 in the Uyghur population (Table S5). After multi-factor adjustment and before oral statin, the additive model (CT and CC+TT) of SNP1 still had statistical significance on the rate of high TC in the Han population. After multi-factor adjustment and before oral statin in the Uyghur population, the dominant model (CC and TT+CT) of SNP1 still had statistical significance in the rates of low HDL-C and high LDL-C in plasma, the recessive model (TT and CC+CT) of SNP1 still had statistical significance in the rates of high TC and high non-HDL-C in plasma, the additive model (CT and CC+TT) of SNP1 still had statistical significance in the rate of low HDL-C in plasma, and the allele model (C and T) of SNP1 still had statistical significance in the rates of high TC, low HDL-C, high LDL-C, high non-HDL-C in plasma (Table 4). After oral statin, there was a correlation between the rates of high APOA1 and SNP1 in the Han population, and a correlation between the rates of high LDL-C with SNP1 in the Uyghur population (Table S6). After multi-factor adjustment and oral statin, the dominant model (CC and TT+CT) and allele model (C and T) of SNP1 still had statistical significance on the rate of high APOA1 in the Han population. The dominant model and allele model of SNP1 still had statistical significance on the rate of high LDL-C in the Uyghur population (Table 4).

The effect of *HMGCR* SNP2 (rs3761740) on the rate of dyslipidemia before and after oral statin in the Han and Uyghur populations

Before oral statin, there was no correlation between the rate of dyslipidemia and SNP2 in the Han population, and there was a correlation between the rate of high TC, high LDL-C, high non-HDL-C, and SNP2 in the Uyghur population (Table S7). After multi-factor

Table 4 The effect of HMGCR SNP1 (rs17671591) and SNP2 (rs3761740) on the rate of dyslipidemia before and after oral statin and after multivariate adjustment.

SNP	Statin	Ethnic	Blood lipids	Gene model		OR	P
SNP1	Before oral statin	Han	Blood lipids	CT (n = 190)	CC+TT (n = 215)	OR	P
			High TC (%)	17.277	8.796	0.415	0.005
		Uyghur	Blood lipids	CC (n = 142)	TT+CT (n = 231)	OR	P
			Low HDL-C (%)	72.340	54.386	0.431	<0.001
			High LDL-C (%)	7.746	14.719	2.070	0.047
			Blood lipids	TT (n = 56)	CC+CT (n = 317)	OR	P
			High TC (%)	19.643	8.833	0.394	0.018
			High Non-HDL-C (%)	25.000	12.618	0.401	0.011
			Blood lipids	CT (n = 175)	CC+TT (n = 198)	OR	P
			Low HDL-C (%)	54.286	67.677	1.878	0.004
			Blood lipids	C (n = 459)	T (n = 287)	OR	P
			High TC (%)	8.279	13.937	1.814	0.014
			Low HDL-C (%)	65.577	54.704	0.624	0.003
			High LDL-C (%)	10.022	15.331	1.637	0.030
			High Non-HDL-C (%)	12.418	17.770	1.559	0.035
	After oral statin	Han	Blood lipids	CC (n = 179)	TT+CT (n = 226)	OR	P
			Low APOA1 (%)	47.191	58.371	1.531	0.043
			Blood lipids	C (n = 548)	T (n = 262)	OR	P
		Uyghur	Low APOA1 (%)	50.368	59.843	0.705	0.028
			Blood lipids	CC (n = 142)	TT+CT (n = 231)	OR	P
			High LDL-C (%)	2.113	8.696	4.514	0.017
			Blood lipids	C (n = 459)	T (n = 287)	OR	P
			High LDL-C (%)	4.585	8.741	0.483	0.018
			Blood lipids	CC (n = 326)	AA+CA (n = 47)	OR	P
			High TC (%)	7.975	27.660	4.403	<0.001
			High LDL-C (%)	10.123	25.532	3.044	0.004
			High Non-HDL-C (%)	12.270	29.787	3.112	0.002
			Blood lipids	CA (n = 44)	AA+CC (n = 329)	OR	P
			High TC (%)	29.545	7.903	0.207	<0.001
SNP2	Before oral statin	Uyghur	High LDL-C (%)	27.273	10.030	0.299	0.002
			High Non-HDL-C (%)	31.818	12.158	0.291	0.001
			Blood lipids	C (n = 696)	A (n = 50)	OR	P
			High TC (%)	9.339	26.000	3.420	<0.001
			High LDL-C (%)	11.207	24.000	2.506	0.009
			High Non-HDL-C (%)	13.506	28.000	2.557	0.005
			Blood lipids	CC (n = 326)	AA+CA (n = 47)	OR	P
			High TC (%)	3.988	13.043	4.190	0.007
	After oral statin	Uyghur	Low HDL-C (%)	61.963	45.652	0.528	0.046
			High LDL-C (%)	4.294	19.565	5.934	<0.001
			High APOB (%)	18.650	39.130	2.965	0.001
			High NonHDL-C (%)	5.828	17.391	4.058	0.003

(Continued)

Table 4 (continued)						
SNP	Statin	Ethnic	Blood lipids	Gene model		OR P
			Blood lipids	C (n = 696)	A (n = 50)	OR P
			High TC (%)	4.604	12.245	0.305 0.013
			Low HDL-C (%)	60.863	46.939	1.732 0.067
			High LDL-C (%)	5.324	18.367	0.234 <0.001
			High APOB (%)	20.000	38.776	0.379 0.002
			High NonHDL-C (%)	6.619	16.327	0.317 0.007
			Blood lipids	CA (n = 44)	CC+AA (n = 329)	OR P
			High TC (%)	13.953	3.951	0.219 0.005
			Low HDL-C (%)	44.186	62.006	1.997 0.037
			High LDL-C (%)	20.930	4.255	0.153 <0.001
			High APOB (%)	39.535	18.790	0.332 0.002
			High NonHDL-C (%)	18.605	5.775	0.221 0.001

Note: Multivariate analysis was further adjusted using logistic regression models and adjustment variables including gender, age, and ALT. The additive model takes the CT genotype as the reference, the dominant model takes the CC genotype as the reference, the recessive model uses the TT genotype as the reference, and the allele model uses the C allele as the reference. Abbreviation: TC, total cholesterol; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; ALT, alanine aminotransferase.

adjustment in the Uyghur populations, the influence of the SNP2 dominant model (CC and AA+CA), additive model (CA and CC+AA), and allele (C and A) models on the rate of high TC, high LDL-C, and high non-HDL-C before oral statin was still statistically significant (Table 4). After oral statin, there was no correlation between the rate of dyslipidemia and SNP2 in the Han population, and there was a correlation between the rate of high TC, high LDL-C, high HDL-C, high APOB, high non-HDL-C, and SNP2 in the Uyghur population (Table S8). After multi-factor adjustment and oral statin in the Uyghur populations, the influence of the SNP2 dominant model (CC and AA+CA) and recessive model (CA and CC+AA) on the rate of high TC, low HDL-C, high LDL-C, high APOB, and high non-HDL-C was still statistically significant, and the influence of SNP2 allele model (C and A) on the rate of high TC, high LDL-C, high APOB, and high non-HDL-C was still statistically significant (Table 4).

Effects of *HMGR* rs17671591 (SNP1) and rs3761740 (SNP2) on the change of lipids after oral statin treatment in Han and Uyghur populations

There was no correlation between the change of lipids after oral statin and SNP2 in the Han and Uyghur populations (Table S9). After oral statin, there was a correlation between the change of APOA1 and SNP1 in the Han population, and a correlation between the changes of LDL-C and SNP1 in the Uyghur population (Table S10). After multi-factor adjustment and oral statin, the influence of the SNP1 allele model (C and T) on the change of APOA1 was still statistically significant in the Han population, and the influence of the SNP1 additive model (CT and CC+TT) on the change of LDL-C was still statistically significant in the Uyghur population (Table 5).

Table 5 Association of SNP1 (rs17671591) with changes of the blood lipids after multivariate adjustment.

Han	Change of lipids	C (548)	T (262)	β	P
	changes of APOA1 (%)	1.591 \pm 21.582	4.988 \pm 20.683	3.56	0.028
Uyghur	Change of lipids	CT (175)	CC + TT (198)	β	P
	changes of LDL-C (%)	14.305 \pm 35.39	21.059 \pm 28.397	7.888	0.018

Note:

Multivariate analysis was further adjusted using linear regression models and adjustment variables including gender, age, and ALT. The additive model takes the CT genotype as the reference and the allele model uses the C allele as the reference. Abbreviation: LDL-C, low-density lipoprotein cholesterol; APOA1, apolipoprotein A1; ALT, alanine aminotransferase.

Table 6 Association of SNP1 (rs17671591) with statin resistance rate in the Uyghur population.

		CT (n = 175)	CC+TT (n = 198)	Single factor analysis		Multifactor adjustment	
				OR	P	OR	P
Statin resistance	64 (36.57%)	54 (27.27%)		0.6504	0.049	0.624	0.038
Non-statin resistance	111 (63.43%)	144 (72.73%)					

Note:

Chi-square test was conducted to generate the P values. Multivariate analysis was further adjusted using logistic regression models and adjustment variables including gender, age, and ALT. The additive model takes the CT genotype as the reference.

Effect of *HMGCR* gene polymorphism on statin resistance

Samples with a decreased rate of plasma LDL-C concentration $\leq 10\%$ after oral statin were set as the statin resistance group, and those with a decreased rate of LDL-C $> 10\%$ were set as the non-statin resistance group. It was found that the addition model of SNP1 had a statistically significant effect on the statin resistance rate in the Uyghur population (Table 6). After adjusting for multiple factors, the additive model of SNP1 in the Uyghur population still had a statistically significant effect on the statin resistance rate (Table 6).

DISCUSSION

In this project, we mainly studied the effects of rs17671591 and rs3761740 of *HMGCR* on plasma lipid levels before and after oral statin in Han Chinese and Uyghur populations, as well as the effects on lipid lowering response after oral statin in two populations.

Mammalian *HMGCR* is an ER-localized glycoprotein consisting of a hydrophobic N-terminal domain that spans the membrane eight times, large hydrophilic C-terminal domain located on the cytoplasmic side, transmembrane domains 2–6 that function as a sterol-sensing domain (SSD) that sensitizes *HMGCR* to sterol levels in the endoplasmic reticulum, and a cytoplasmic C-terminal domain that is responsible for converting HMG-CoA to valerate using two NADPH molecules as reducing agents (Liscum et al., 1985). *HMGCR* plays one of the most important roles in cholesterol biosynthesis and is one of the two rate-limiting enzymes in cholesterol biosynthesis. It is also regulated by many factors and is crucial for maintaining lipid homeostasis in the body (Luo, Yang & Song, 2020). *HMGCR* is the main target of statins for lipid reduction (Istvan & Deisenhofer, 2001).

Some studies have proved that effective lipid changes can be seen in the 4th week of atorvastatin dose range of 10–80 mg ([Jones et al., 2005](#)), and this result has also been proved in other studies ([Szapary et al., 2004](#)) where the lipid level remained basically stable after 4 weeks ([Cannon et al., 2004](#); [Niemi et al., 2004](#)), and so this study selected patients who took oral statin for more than 1 month as the study object. In the Clinical Pharmacogenetics Implementation Consortium (CPIC) guidelines, the dosing recommendations were provided according to SLCO1B1, ABCG2, and CYP2C9 genotypes. A dose of atorvastatin at 10 mg once daily and a dose of rosuvastatin at 5 mg once daily is associated with the lowest risk for statin-associated musculoskeletal symptoms ([Cooper-DeHoff et al., 2022](#)), so we chose patients who took 10 mg of atorvastatin daily or 5 mg of rosuvastatin daily as subjects. In this study, we confirmed that oral statin can effectively reduce TC, TG, LDL-C, and APOB, which is consistent with the results of [Jones et al. \(2005\)](#). However, the effect of statins on HDL-C is controversial. [Jones et al.'s \(2005\)](#) study proved that statins can increase HDL-C, while [Szapary et al.'s \(2004\)](#) study proved that statin treatment does not affect the plasma concentration of HDL-C. This difference may be related to the dose of statin or the different ethnic genetic background of the study subjects. In this study, it was proved that the plasma HDL-C concentration of the Han population was significantly reduced after statin treatment, while the plasma HDL-C concentration of the Uyghur population was not significantly changed after statin treatment. Our study also proved that statins can effectively reduce plasma levels of TG, TC, LDL-C, APOA1, APOB, and non-HDL-C in both Han and Uyghur subjects.

The rs17671591 polymorphism is an intergenic variation located near the *HMGCR* gene promoter region (chr5:74,615,021); however, it is not directly located in the gene expression regulatory element or the region affecting the enzyme structure or conformation. The dominant model of rs17671591 polymorphism is associated with HDL-C plasma levels and affects the change of LDL-C and HDL-C plasma concentrations after statin treatment ([Cuevas et al., 2016](#)). The Treating to New Targets (TNT) study also showed an association between *HMGCR* rs17671591 and statin response ([Thompson et al., 2009](#)), but a genome-wide association (GWAS) study also proved that rs17671591 polymorphism was not associated with low density lipoprotein level after taking atorvastatin ([Deshmukh et al., 2012](#)). In this study, we found that the *HMGCR* rs17671591 genotype was associated with the rate of high TC before oral statin, APOA1 and the rate of high APOA1 after oral statin, and the change of APOA1 after oral statin in Han subjects. In the Uyghur population, we found that the *HMGCR* rs17671591 genotype was associated with LDL-C, low HDL-C, high LDL-C, high TC, and high non-HDL-C before oral statin; LDL-C, and high LDL-C after oral statin; and the change of LDL-C after oral statin. The difference of rs17671591 effects between the two ethnic groups may be due to their different genetic backgrounds.

The rs3761740 polymorphism is a variant located in the upstream region of the *HMGCR* gene chr5:75336308 (GRCh38.p14), but it not directly located in gene expression regulatory elements or regions affecting enzyme structure or conformation. Previous studies have not found a correlation between rs3761740 with LDL-C and statins response

to LDL-C effect ([Angelini et al., 2017](#)). However, the results of this study are not completely consistent with this. In the Han population, there was no significant correlation between rs3761740 with blood lipids and lipid-lowering effect of statins. In the Uyghur population, we found that the *HMGCR* rs3761740 genotype was associated with TC, LDL-C, non-HDL-C, the rate of high TC, the rate of high LDL-C, the rate of high non-HDL-C before oral statin; and TC, TG, LDL-C, APOB, highTC, high LDL-C, low HDL-C, high APOB, and high non-HDL-C after oral statin.

The lipid-lowering effect of statins is different for each person. Many genetic mutations are related to the effectiveness of statins. These genes can be divided into two groups. First are those that are directly related to lipoprotein metabolism and affect either LDL production or its catabolism. The second group are drug metabolism-related genes that affect statin pharmacokinetics. Some studies have proved that the genetic variation of *HMGCR* is related to the resistance of statins, but it is not certain if it is due to the ability of statins to inhibit the enzyme or from the greater compensatory upregulation of the *HMGCR* gene ([Sun et al., 2023](#)). High-dose statin therapy is associated with lower blood lipids ([Li et al., 2021](#)), but the risk of intolerance with high-dose statins is also increased ([Cooper-DeHoff et al., 2022](#)). It is very important to predict the lipid-lowering effect of statins and choose low-dose statins that can effectively reduce blood lipids according to the predicted results. In our study, we found that the *HMGCR* rs17671591 additive model (CT and CC+TT) was associated with the change of LDL-C after oral statin in the Uyghur population. This SNP can be used to predict the efficacy of statins in the Chinese Uyghur population and provide strategies for precision therapy.

Different genetic models are supported by their assumed inheritance pattern, although no optimal models have been established ([Horita & Kaneko, 2015](#)). The levels of significance obtained from different models may be similar, but not exactly the same ([Horita & Kaneko, 2015](#); [Clarke et al., 2011](#)). Although the analysis of multiple models may lead to concerns about multiple comparisons, the unified direction and association obtained may further support the association between SNP and disease status ([Horita & Kaneko, 2015](#); [Wu et al., 2015](#)). In this study, different gene models were established to verify the relationship between each gene model, blood lipids, and statin response, and the consistent influence on the direction and association of each model can better support the conclusions.

The differences between this study and previous studies and between Han and Uyghur populations in this study are not surprising, as lipid concentrations are a complex feature of environmental and genetic factors and their interactions. Because genetic studies rely primarily on genetic diversity, other unexplained interactions are often excluded, including traits such as age, sex, ethnic origin, sample size, heterogeneity among patients, study design, use of different endpoints, evaluation time, drug dosage, strict inclusion/exclusion criteria, and underlying disease. As a result, it becomes very difficult to draw the same conclusions after comparing different studies, so the results tend to apply only to specific populations.

This study also has some limitations. First of all, the multivariate adjustment factors included in this study only included age, sex, and liver function as possible risk factors, but

did not include other potential risk factors affecting blood lipids including diet, exercise, smoking, drinking, and BMI. Second, the follow-up time of this study was ≥ 1 month and was not fixed, but we ensured that each subject did not change the lipid-lowering drugs and dosage during the study period. Some studies have proved that blood lipids tend to be stable after oral statin for 4 weeks (Cannon *et al.*, 2004; Niemi *et al.*, 2004), so the unfixed time had little effect on blood lipids after taking drugs.

CONCLUSION

In this article, we mainly studied the effects of *HMGCR* rs17671591 and rs3761740 on plasma lipid levels before and after oral statin in Han Chinese and Uyghur populations, as well as the effects on lipid lowering response after oral statin in two populations. We found that the SNP of *HMGCR* was associated with the lipid-lowering effect of statins and provides the possibility to predict the reactivity of statins in different individuals.

ACKNOWLEDGEMENTS

We thank all of the patients for participating in the study.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

This work was supported by National key Research and Development program of China [grant number: 2021YFC2500605]; The National Natural Science Foundation of China [grant numbers: 81970380]; State Key Laboratory of Pathogenesis, Prevention and Treatment of High Incident Disease in Central Asia [grant numbers: xyd2021C002]; State Key Laboratory of Pathogenesis, Prevention and Treatment of High Incidence Diseases in Central Asia Fund [SKL-HIDCA-2022-XXG3]. State Key Laboratory of Pathogenesis, Prevention and Treatment of High Incidence Diseases in Central Asia Fund [SKL-HIDCA-2023-36]. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors:

National Key Research and Development Program of China: 2021YFC2500605.

National Natural Science Foundation of China: 81970380.

State Key Laboratory of Pathogenesis.

Prevention and Treatment of High Incident Disease in Central Asia: xyd2021C002, SKL-HIDCA-2022-XXG3, SKL-HIDCA-2023-36.

Competing Interests

The authors declare that there are no competing interests associated with the manuscript.

Author Contributions

- Ziyang Liu conceived and designed the experiments, analyzed the data, authored or reviewed drafts of the article, and approved the final draft.

- Yang Zhou conceived and designed the experiments, analyzed the data, authored or reviewed drafts of the article, and approved the final draft.
- Menglong Jin performed the experiments, prepared figures and/or tables, and approved the final draft.
- Shuai Liu analyzed the data, prepared figures and/or tables, and approved the final draft.
- Sen Liu analyzed the data, prepared figures and/or tables, and approved the final draft.
- Kai Yang performed the experiments, prepared figures and/or tables, and approved the final draft.
- Huayin Li performed the experiments, prepared figures and/or tables, and approved the final draft.
- Sifu Luo performed the experiments, prepared figures and/or tables, and approved the final draft.
- Subinuer Jureti performed the experiments, prepared figures and/or tables, and approved the final draft.
- Mengwei Wei performed the experiments, prepared figures and/or tables, and approved the final draft.
- Zhenyan Fu conceived and designed the experiments, authored or reviewed drafts of the article, and approved the final draft.

Human Ethics

The following information was supplied relating to ethical approvals (*i.e.*, approving body and any reference numbers):

The Ethics Committee of Xinjiang Medical University (number: 220525-06-2305A-Y1).

Data Availability

The following information was supplied regarding data availability:

The raw measurements are available in the [Supplemental Files](#).

Supplemental Information

Supplemental information for this article can be found online at <http://dx.doi.org/10.7717/peerj.18144#supplemental-information>.

REFERENCES

- Ahmad F, Leake DS. 2019. Lysosomal oxidation of LDL alters lysosomal pH, induces senescence, and increases secretion of pro-inflammatory cytokines in human macrophages. *Journal of Lipid Research* 60(1):98–110 DOI 10.1194/jlr.M088245.
- Angelini S, Rosticci M, Massimo G, Musti M, Ravegnini G, Consolini N, Sammarini G, D’Addato S, Rizzoli E, Botbayev D, Borghi C, Cantelli-Forti G, Cicero AF. 2017. Relationship between lipid phenotypes, overweight, lipid lowering drug response and KIF6 and HMG-CoA Genotypes in a subset of the Brisighella heart study population. *International Journal of Molecular Sciences* 19(1):49 DOI 10.3390/ijms19010049.
- Arvanitis M, Lowenstein CJ. 2023. Dyslipidemia. *Annals of Internal Medicine* 176(6):ITC81 DOI 10.7326/AITC202306200.
- Barber MJ, Mangravite LM, Hyde CL, Chasman DI, Smith JD, McCarty CA, Li X, Wilke RA, Rieder MJ, Williams PT, Ridker PM, Chatterjee A, Rotter JI, Nickerson DA, Stephens M,

- Krauss RM. 2010. Genome-wide association of lipid-lowering response to statins in combined study populations. *PLOS ONE* 5(3):e9763 DOI 10.1371/journal.pone.0009763.
- Brown MS, Goldstein JL. 1997. The SREBP pathway: regulation of cholesterol metabolism by proteolysis of a membrane-bound transcription factor. *Cell* 89(3):331–340 DOI 10.1016/S0092-8674(00)80213-5.
- Cannon CP, Braunwald E, McCabe CH, Rader DJ, Rouleau JL, Belder R, Joyal SV, Hill KA, Pfeffer MA, Skene AM. 2004. Pravastatin or atorvastatin evaluation and infection therapy-thrombolysis in myocardial infarction 22 investigators. Intensive versus moderate lipid lowering with statins after acute coronary syndromes. *New England Journal of Medicine* 350(15):1495–1504 DOI 10.1056/NEJMoa040583.
- Clarke GM, Anderson CA, Pettersson FH, Cardon LR, Morris AP, Zondervan KT. 2011. Basic statistical analysis in genetic case-control studies. *Nature Protocols* 6(2):121–133 DOI 10.1038/nprot.2010.182.
- Cooper-DeHoff RM, Niemi M, Ramsey LB, Luzum JA, Tarkiainen EK, Straka RJ, Gong L, Tuteja S, Wilke RA, Wadelius M, Larson EA, Roden DM, Klein TE, Yee SW, Krauss RM, Turner RM, Palaniappan L, Gaedigk A, Giacomini KM, Caudle KE, Voora D. 2022. The clinical pharmacogenetics implementation consortium guideline for SLCO1B1, ABCG2, and CYP2C9 genotypes and statin-associated musculoskeletal symptoms. *Clinical Pharmacology & Therapeutics* 111(5):1007–1021 DOI 10.1002/cpt.2557.
- Cuevas A, Fernández C, Ferrada L, Zambrano T, Rosales A, Saavedra N, Salazar LA. 2016. HMGCR rs17671591 SNP determines lower plasma LDL-C after atorvastatin therapy in Chilean individuals. *Basic & Clinical Pharmacology & Toxicology* 118(4):292–297 DOI 10.1111/bcpt.12493.
- Deshmukh HA, Colhoun HM, Johnson T, McKeigue PM, Betteridge DJ, Durrington PN, Fuller JH, Livingstone S, Charlton-Menys V, Neil A, Poulter N, Sever P, Shields DC, Stanton AV, Chatterjee A, Hyde C, Calle RA, DeMicco DA, Trompet S, Postmus I, Ford I, Jukema JW, Caulfield M, Hitman GA. 2012. Genome-wide association study of genetic determinants of LDL-c response to atorvastatin therapy: importance of Lp(a). *Journal of Lipid Research* 53(5):1000–1011 DOI 10.1194/jlr.P021113.
- Erbil R, Aboyans V, Boileau C, Bossone E, Bartolomeo RD, Eggebrecht H, Evangelista A, Falk V, Frank H, Gaemperli O, Grabenwöger M, Haverich A, Iung B, Manolis AJ, Meijboom F, Nienaber CA, Roffi M, Rousseau H, Sechtem U, Sirnes PA, Allmen RS, Vrints CJ, ESC Committee for Practice Guidelines. 2014. 2014 ESC guidelines on the diagnosis and treatment of aortic diseases: document covering acute and chronic aortic diseases of the thoracic and abdominal aorta of the adult. The task force for the diagnosis and treatment of aortic diseases of the European society of cardiology (ESC). *European Heart Journal* 35(41):2873–2926 DOI 10.1093/eurheartj/ehu281.
- GBD 2016 Causes of Death Collaborators. 2016. Global, regional, and national age-sex specific mortality for 264 causes of death, 1980–2016: a systematic analysis for the global burden of disease study 2016. *Lancet* 390(10100):1151–1210 DOI 10.1016/S0140-6736(17)32152-9.
- Grundy SM, Stone NJ, Bailey AL, Beam C, Birtcher KK, Blumenthal RS, Braun LT, de Ferranti S, Faiella-Tommasino J, Forman DE, Goldberg R, Heidenreich PA, Hlatky MA, Jones DW, Lloyd-Jones D, Lopez-Pajares N, Ndumele CE, Orringer CE, Peralta CA, Saseen JJ, Smith SC Jr, Sperling L, Virani SS, Yeboah J. 2019. 2018 AHA/ACC/AACVPR/AAPA/ABC/ACPM/ADA/AGS/APhA/ASPC/NLA/PCNA guideline on the management of blood cholesterol: a report of the American college of cardiology/American Heart association task force on clinical practice guidelines. *Circulation* 139(25):e1082–e1143 DOI 10.1161/CIR.0000000000000625. Erratum in: *Circulation* 148(7):e5.

- Guidelines for the Management of Blood lipids in China. 2023.** Joint expert committee for the revision of Chinese blood lipid management guidelines. *Chinese Journal of Cardiovascular Diseases* 51(3):221–255 DOI 10.3760/cma.j.cn112148-20230119-00038.
- Horita N, Kaneko T. 2015.** Genetic model selection for a case-control study and a meta-analysis. *Meta Gene* 5(2):1–8 DOI 10.1016/j.mgene.2015.04.003.
- Istvan ES, Deisenhofer J. 2001.** Structural mechanism for statin inhibition of HMG-CoA reductase. *Science* 292(5519):1160–1164 DOI 10.1126/science.1059344.
- Jones PH, McKenney JM, Karalis DG, Downey J. 2005.** Comparison of the efficacy and safety of atorvastatin initiated at different starting doses in patients with dyslipidemia. *American Heart Journal* 149(1):e1 DOI 10.1016/j.ahj.2004.07.025.
- Li S, Liu HH, Guo YL, Zhu CG, Wu NQ, Xu RX, Dong Q, Li JJ. 2021.** Improvement of evaluation in Chinese patients with atherosclerotic cardiovascular disease using the very-high-risk refinement: a population-based study. *The Lancet Regional Health-Western Pacific* 17:100286 DOI 10.1016/j.lanwpc.2021.100286.
- Liscum L, Finer-Moore J, Stroud RM, Luskey KL, Brown MS, Goldstein JL. 1985.** Domain structure of 3-hydroxy-3-methylglutaryl coenzyme A reductase, a glycoprotein of the endoplasmic reticulum. *Journal of Biological Chemistry* 260(1):522–530 DOI 10.1016/S0021-9258(18)89764-2.
- Liu DJ, Peloso GM, Yu H, Butterworth AS, Wang X, Mahajan A, Saleheen D, Emdin C, Alam D, Alves AC, Amouyel P, Di Angelantonio E, Arveiler D, Assimes TL, Auer PL, Baber U, Ballantyne CM, Bang LE, Benn M, Bis JC, Boehnke M, Boerwinkle E, Bork-Jensen J, Bottinger EP, Brandslund I, Brown M, Busonero F, Caulfield MJ, Chambers JC, Chasman DI, Chen YE, Chen YI, Chowdhury R, Christensen C, Chu AY, Connell JM, Cucca F, Cupples LA, Damrauer SM, Davies G, Deary IJ, Dedoussis G, Denny JC, Dominiczak A, Dubé MP, Ebeling T, Eiriksdottir G, Esko T, Farmaki AE, Feitosa MF, Ferrario M, Ferrieres J, Ford I, Fornage M, Franks PW, Frayling TM, Frikke-Schmidt R, Fritsche LG, Frossard P, Fuster V, Ganesh SK, Gao W, Garcia ME, Gieger C, Giulianini F, Goodarzi MO, Grallert H, Grarup N, Groop L, Grove ML, Gudnason V, Hansen T, Harris TB, Hayward C, Hirschhorn JN, Holmen OL, Huffman J, Huo Y, Hveem K, Jabeen S, Jackson AU, Jakobsdottir J, Jarvelin MR, Jensen GB, Jørgensen ME, Jukema JW, Justesen JM, Kamstrup PR, Kanoni S, Karpe F, Kee F, Khera AV, Klarin D, Koistinen HA, Kooner JS, Kooperberg C, Kuulasmaa K, Kuusisto J, Laakso M, Lakka T, Langenberg C, Langsted A, Launer LJ, Lauritzen T, Liewald DCM, Lin LA, Linneberg A, Loos RJE, Lu Y, Lu X, Mägi R, Malarstig A, Manichaikul A, Manning AK, Mäntyselkä P, Marouli E, Masca NGD, Maschio A, Meigs JB, Melander O, Metspalu A, Morris AP, Morrison AC, Mulas A, Müller-Nurasyid M, Munroe PB, Neville MJ, Nielsen JB, Nielsen SF, Nordestgaard BG, Ordovas JM, Mehran R, O'Donnell CJ, Orho-Melander M, Molony CM, Muntendam P, Padmanabhan S, Palmer CNA, Pasko D, Patel AP, Pedersen O, Perola M, Peters A, Pisinger C, Pistis G, Polasek O, Poulter N, Psaty BM, Rader DJ, Rasheed A, Rauramaa R, Reilly DF, Reiner AP, Renström F, Rich SS, Ridker PM, Rioux JD, Robertson NR, Roden DM, Rotter JI, Rudan I, Salomaa V, Samani NJ, Sanna S, Sattar N, Schmidt EM, Scott RA, Sever P, Sevilla RS, Shaffer CM, Sim X, Sivapalaratnam S, Small KS, Smith AV, Smith BH, Somayajula S, Southam L, Spector TD, Speliotes EK, Starr JM, Stirrups KE, Stitzel N, Strauch K, Stringham HM, Surendran P, Tada H, Tall AR, Tang H, Tardif JC, Taylor KD, Trompet S, Tsao PS, Tuomilehto J, Tybjaerg-Hansen A, van Zuydam NR, Varbo A, Varga TV, Virtamo J, Waldenberger M, Wang N, et al. 2017.** Exome-wide association study of plasma lipids in >300,000 individuals. *Nature Genetics* 49(12):1758–1766 DOI 10.1038/ng.3977.

- Lu X, Peloso GM, Liu DJ, Wu Y, Zhang H, Zhou W, Li J, Tang CS, Dorajoo R, Li H, Long J, Guo X, Xu M, Spracklen CN, Chen Y, Liu X, Zhang Y, Khor CC, Liu J, Sun L, Wang L, Gao YT, Hu Y, Yu K, Wang Y, Cheung CY, Wang F, Huang J, Fan Q, Cai Q, Chen S, Shi J, Yang X, Zhao W, Sheu WH, Cherny SS, He M, Feranil AB, Adair LS, Gordon-Larsen P, Du S, Varma R, Chen YI, Shu XO, Lam KSL, Wong TY, Ganesh SK, Mo Z, Hveem K, Fritsche LG, Nielsen JB, Tse HF, Huo Y, Cheng CY, Chen YE, Zheng W, Tai ES, Gao W, Lin X, Huang W, Abecasis G, GLGC Consortium, Kathiresan S, Mohlke KL, Wu T, Sham PC, Gu D, Willer CJ. 2017. Exome chip meta-analysis identifies novel loci and East Asian-specific coding variants that contribute to lipid levels and coronary artery disease. *Nature Genetics* 49(12):1722–1730 DOI 10.1038/ng.3978.
- Luo J, Yang H, Song BL. 2020. Mechanisms and regulation of cholesterol homeostasis. *Nature Reviews Molecular Cell Biology* 21(4):225–245 DOI 10.1038/s41580-019-0190-7.
- Mozaffarian D, Benjamin EJ, Go AS, Arnett DK, Blaha MJ, Cushman M, Das SR, de Ferranti S, Després JP, Fullerton HJ, Howard VJ, Huffman MD, Isasi CR, Jiménez MC, Judd SE, Kissela BM, Lichtman JH, Lisabeth LD, Liu S, Mackey RH, Magid DJ, McGuire DK, Mohler 3rd ER, Moy CS, Muntner P, Mussolino ME, Nasir K, Neumar RW, Nichol G, Palaniappan L, Pandey DK, Reeves MJ, Rodriguez CJ, Rosamond W, Sorlie PD, Stein J, Towfighi A, Turan TN, Virani SS, Woo D, Yeh RW, Turner MW, American Heart Association Statistics Committee, Stroke Statistics Subcommittee, Writing Group Members. 2016. Executive summary: heart disease and stroke statistics–2016 update: a report from the American heart association. *Circulation* 133(4):447–454 DOI 10.1161/CIR.0000000000000366.
- Niemi M, Schaeffeler E, Lang T, Fromm MF, Neuvonen M, Kyrklund C, Backman JT, Kerb R, Schwab M, Neuvonen PJ, Eichelbaum M, Kivistö KT. 2004. High plasma pravastatin concentrations are associated with single nucleotide polymorphisms and haplotypes of organic anion transporting polypeptide-C (OATP-C, SLCO1B1). *Pharmacogenetics* 14(7):429–440 DOI 10.1097/01.fpc.0000114750.08559.32.
- O’Malley PG, Arnold MJ, Kelley C, Spacek L, Buelt A, Natarajan S, Donahue MP, Vagichev E, Ballard-Hernandez J, Logan A, Thomas L, Ritter J, Neubauer BE, Downs JR. 2020. Management of dyslipidemia for cardiovascular disease risk reduction: synopsis of the 2020 updated U.S. department of veterans affairs and U.S. department of defense clinical practice guideline. *Annals of Internal Medicine* 173(10):822–829 DOI 10.7326/M20-4648.
- Ralapanawa U, Sivakanesan R. 2021. Epidemiology and the magnitude of coronary artery disease and acute coronary syndrome: a narrative review. *Journal of Epidemiology and Global Health* 11(2):169–177 DOI 10.2991/jegh.k.201217.001.
- Reiner Z. 2014. Resistance and intolerance to statins. *Nutrition, Metabolism, and Cardiovascular Diseases* 24(10):1057–1066 DOI 10.1016/j.numecd.2014.05.009.
- Schmitz G, Drobnik W. 2003. Pharmacogenomics and pharmacogenetics of cholesterol-lowering therapy. *Clinical Chemistry and Laboratory Medicine* 41(4):581–589 DOI 10.1515/CCLM.2003.088.
- Simon JA, Lin F, Hulley SB, Blanche PJ, Waters D, Shiboski S, Rotter JI, Nickerson DA, Yang H, Saad M, Krauss RM. 2006. Phenotypic predictors of response to simvastatin therapy among African-Americans and Caucasians: the cholesterol and pharmacogenetics (CAP) study. *The American Journal of Cardiology* 97(6):843–850 DOI 10.1016/j.amjcard.2005.09.134.
- Smith DA. 2019. Review: in dyslipidemia or atherosclerotic CVD, alirocumab and evolocumab vs control each reduce MI and stroke. *Annals of Internal Medicine* 171(10):JC56 DOI 10.7326/ACPJ201911190-056.

- Sun L, Wolska A, Amar M, Zubirán R, Remaley AT. 2023. Approach to the patient with a suboptimal statin response: causes and algorithm for clinical management. *The Journal of Clinical Endocrinology & Metabolism* 108(9):2424–2434 DOI 10.1210/clinem/dgad153.
- Szapary L, Horvath B, Marton Z, Alexy T, Kesmarky G, Habon T, Szots M, Koltai K, Juricskay I, Czopf J, Toth K. 2004. Short-term effect of low-dose atorvastatin on haemorrheological parameters, platelet aggregation and endothelial function in patients with cerebrovascular disease and hyperlipidaemia. *CNS Drugs* 18(3):165–172 DOI 10.2165/00023210-200418030-00003.
- Thompson JF, Hyde CL, Wood LS, Paciga SA, Hinds DA, Cox DR, Hovingh GK, Kastelein JJ. 2009. Comprehensive whole-genome and candidate gene analysis for response to statin therapy in the treating to new targets (TNT) cohort. *Circulation: Cardiovascular Genetics* 2(2):173–181 DOI 10.1161/CIRCGENETICS.108.818062.
- Tsao CW, Aday AW, Almarzooq ZI, Anderson CAM, Arora P, Avery CL, Baker-Smith CM, Beaton AZ, Boehme AK, Buxton AE, Commodore-Mensah Y, Elkind MSV, Evenson KR, Eze-Nliam C, Fugar S, Generoso G, Heard DG, Hiremath S, Ho JE, Kalani R, Kazi DS, Ko D, Levine DA, Liu J, Ma J, Magnani JW, Michos ED, Mussolino ME, Navaneethan SD, Parikh NI, Poudel R, Rezk-Hanna M, Roth GA, Shah NS, St-Onge MP, Thacker EL, Virani SS, Voeks JH, Wang NY, Wong ND, Wong SS, Yaffe K, Martin SS, American Heart Association Council on Epidemiology and Prevention Statistics Committee and Stroke Statistics Subcommittee. 2023. Heart disease and stroke statistics-2023 update: a report from the American heart association. *Circulation* 147(8):e93–e621 DOI 10.1161/CIR.0000000000001123.
- Visseren FLJ, Mach F, Smulders YM, Carballo D, Koskinas KC, Bäck M, Benetos A, Biffi A, Boavida JM, Capodanno D, Cosyns B, Crawford C, Davos CH, Desormais I, Di Angelantonio E, Franco OH, Halvorsen S, Hobbs FDR, Hollander M, Jankowska EA, Michal M, Sacco S, Sattar N, Tokgozoglu L, Tonstad S, Tsioufis KP, van Dis I, van Gelder IC, Wanner C, Williams B, ESC National Cardiac Societies, ESC Scientific Document Group. 2021. 2021 ESC guidelines on cardiovascular disease prevention in clinical practice. *European Heart Journal* 42:3227–3337 DOI 10.1093/eurheartj/ehab484.
- Wang J, Bai Y, Zhao X, Ru J, Kang N, Tian T, Tang L, An Y, Li P. 2018. oxLDL-mediated cellular senescence is associated with increased NADPH oxidase p47phox recruitment to caveolae. *Bioscience Reports* 38(3):BSR20180283 DOI 10.1042/BSR20180283.
- Wu L, Hu Y, Li D, Jiang W, Xu B. 2015. Screening toll-like receptor markers to predict latent tuberculosis infection and subsequent tuberculosis disease in a Chinese population. *BMC Medical Genetics* 16:19 DOI 10.1186/s12881-015-0166-1.
- Xiao J, Luo J, Hu A, Xiao T, Li M, Kong Z, Jiang L, Zhou Z, Liao Y, Xie C, Chu B, Miao H, Li B, Shi X, Song BL. 2019. Cholesterol transport through the peroxisome-ER membrane contacts tethered by PI(4,5)P2 and extended synaptotagmins. *Science China Life Sciences* 62(9):1117–1135 DOI 10.1007/s11427-019-9569-9.