# 1 Small population size and possible extirpation of the

# 2 threatened Malagasy poison frog Mantella cowanii

- 3 Devin Edmonds<sup>1,2</sup>, Raphali R. Andriantsimanarilafy<sup>3</sup>, Angelica Crottini<sup>4,5,6</sup>, Michael J.
- 4 Dreslik<sup>1,2</sup>, Jade Newton-Youens<sup>7</sup>, Andoniana Ramahefason<sup>8</sup>, Christian
- 5 Randrianantoandro<sup>8</sup>, Franco Andreone<sup>9</sup>
- 6 <sup>1</sup>Department of Natural Resources and Environmental Sciences, University of Illinois at
- 7 Urbana-Champaign, Champaign, Illinois, USA
- 8 <sup>2</sup>Illinois Natural History Survey, Prairie Research Institute, University of Illinois at
- 9 Urbana-Champaign, Champaign, IL, USA
- 10 <sup>3</sup>Madagasikara Voakajy, Antananarivo, Madagascar
- 11 <sup>4</sup>CIBIO, Centro de Investigação em Biodiversidade e Recursos Genéticos, Vairão,
- 12 Portugal
- 13 <sup>5</sup>Departamento de Biologia, Faculdade de Ciências, Universidade do Porto, Porto,
- 14 Portugal
- 15 <sup>6</sup>BIOPOLIS Program in Genomics, Biodiversity and Land Planning, CIBIO, Campus de
- 16 Vairão, Vairão, Portugal
- 17 Department of Natural Sciences, Manchester Metropolitan University, Manchester, UK
- 18 <sup>8</sup>Mention Zoologie et Biodiversité Animale, Université d'Antananarivo, Antananarivo,
- 19 Madagascar

21

25

- 20 <sup>9</sup>Museo Regionale di Scienze Naturali, Torino, Italy
- 22 Corresponding Author:
- 23 Devin Edmonds<sup>1,2</sup>
- 24 1816 S Oak Street, Champaign, Illinois, 61820, USA
- 26 Email address: dae2@illinois.edu

#### Abstract

27 28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47 48

49

50

51

52

53

54

55

56

57

58

59

60

61

62

63

Amphibians are experiencing severe population declines, requiring targeted conservation action for the most threatened species and habitats. Unfortunately, we do not know the basic demographic traits of most species, which hinders population recovery efforts. We studied one of Madagascar's most threatened frog species, the harlequin mantella (Mantella cowanii), to confirm it is still present at historic localities and estimate annual survival and population sizes. We surveyed eleven of all thirteen known localities and were able to detect the species at eight. Using a naïve estimate of detection probability from sites with confirmed presence, we estimated 1.54 surveys (95% CI: 1.10-2.37) are needed to infer absence with 95% confidence, suggesting the three populations where we did not detect M. cowanii are now extirpated. However, we also describe two new populations within the scientific literature for the first time. Repeated annual surveys at three sites showed population sizes ranged from 13-137 adults over 3-8 years, with the most intensively surveyed site experiencing a >80% reduction in population size during 2015–2023. Annual adult survival was moderately high (0.529-0.618) and we recaptured five individuals in 2022 and one in 2023 first captured as adults in 2015, revealing the maximum lifespan of the species in nature can reach nine years and beyond. Our results show M. cowanii is characterized by a slower life history pace than other Mantella species, putting it at greater extinction risk. Illegal collection for the international pet trade and continued habitat degradation are the main threats to the species. We recommend conservation efforts continue monitoring populations and reassess the International Union for Conservation of Nature (IUCN) Red List status because M. cowanii, may be Critically Endangered rather than Endangered based on population size and trends.

#### Introduction

Amphibian species are facing extinction rates at least 22 times faster than the average rate during the 10 millennia before industrialization, resulting in their status as the most threatened vertebrate class (Ceballos et al., 2015; Luedtke et al., 2023). Many species are experiencing severe population declines, leading to widespread range contractions through extirpation (e.g., Beyer & Manica 2020; Granados-Martínez et al. 2021; Patla & Peterson 2022). Habitat loss is the largest threat to amphibians, but infectious diseases, invasive species, climate change, overexploitation, and pollution are all responsible for declines and interact in complex ways (Collins, 2010; Grant, Miller & Muths, 2020). Such threats and population trends highlight the immediate need for increased conservation, especially targeted toward the most threatened species and their habitats.

Formatted: Highlight

Deleted: discovered

Formatted: Highlight

Formatted: Highlight

Formatted: Pattern: Clear (Accent 4), Highlight

Deleted: it

The island of Madagascar supports extraordinary amphibian species richness and endemism, with more than 415 described endemic frog species representing five anuran clades of independent origin (Crottini et al., 2012; Antonelli et al., 2022; AmphibiaWeb 2023). Alarmingly, 46.4% of assessed Malagasy frog species are threatened, owing largely to deforestation (Ralimanana et al., 2022; IUCN, 2023). Deforestation has eliminated as much as a quarter of the tree cover on the island over the last 25 years and the rate has only increased since 2005 (Vieilledent et al., 2018; Suzzi-Simmons, 2023). Consequently, many frog species in Madagascar have patchy distributions restricted to isolated pockets of forest in an otherwise inhospitable landscape (Lehtinen & Ramanamanjato, 2006). So far, there have been no documented modern frog species extinctions in Madagascar (Andreone et al., 2008, 2021), but many records of species presence are from biological inventories conducted decades ago in areas with high rates of land use change. Verifying species presence and confirming the extant distribution of threatened species is some of the most vital information for informing conservation (Villero et al., 2017). Relatedly, we know little about frog population trends in Madagascar, even for highly threatened species. The lack of demographic information is not unique to Madagascar; we do not know survival or fertility rates for 87.5% of amphibian species globally (Conde et al., 2019). Baseline estimates of survival, recruitment, and other demographic traits are urgently needed to improve conservation efforts and inform management decisions (Grant, Miller & Muths, 2020).

Some of the most well-known amphibians in Madagascar are the Malagasy poison frogs in the genus *Mantella*. One species (*M. laevigata*) exhibits parental care and all *Mantella* species display aposematic coloration to warn predators of their poisonous skin alkaloids sequestered from prey (Vences et al., 2022). As such, they are familiar examples of convergent evolution with Neotropical dendrobatids (Daly, Highet & Myers, 1984; Chiari et al., 2004; Fischer et al., 2019). While several *Mantella* species are widespread and have been found in degraded habitat and agricultural plantations (e.g., *M. betsileo*, *M. ebenaui*, and *M. viridis*, Vences et al., 1999; Andreone et al., 2006; Crottini et al., 2012), most are restricted to small areas, have highly localized populations, show a recent dramatic demographic decline, and are threatened by ongoing habitat changes (e.g., Crottini et al., 2019). Compounding the threat of habitat loss is overexploitation; thousands of wild poison frogs are exported annually from Madagascar for the international pet trade (Rabemananjara et al., 2007b), though export quotas have been restricted recently to smaller quantities of just six species (CITES, 2022).

The harlequin mantella frog (*M. cowanii*) is one of the most threatened *Mantella* species, with a small and fragmented distribution in the central highlands. This region of Madagascar was formerly a mosaic of grassland, woodland, and subhumid forest,

covering the mountainous area between the island's humid east and dryer west (Yoder et al., 2016). Today the central highlands consist mostly of secondary grasses and land converted for subsistence agriculture and cattle grazing, with little humid forest remaining (Andriambeloson et al., 2021; Ranarilalatiana et al., 2022). Thirteen localities of *M. cowanii* are known from the region: six in a cluster around the village of Antoetra, five 80 km northwest on the Itremo Massif, and two isolated localities located >100 km north of all other known populations, one near Betafo and the other east of Antakasina (Rabibisoa, 2008; Rabibisoa et al., 2009). All populations occur along mountainous streams with large boulders and adjacent wet rockfaces, which are typically covered in wet moss and bryophytes. While some sites have intact gallery forests, others are almost entirely devoid of trees (Fig. 1).

Crucially, some *M. cowanii* populations are known from only one or two scientific expeditions carried out in the early 2000s (e.g., Andreone & Randrianirina, 2003; Andreone et al., 2007; Crottini et al., 2011) and could now be extirpated. Due to its striking black and orange coloration (Fig. 2), *M. cowanii* was heavily exploited for the international pet trade during the 1990s and early 2000s (Andreone, Mercurio & Mattioli, 2006). From 1994–2003, several thousand frogs recorded as *M. cowanii* were exported from Madagascar for commercial purposes, after which legal trade was halted and the export quota was set to zero (Rabemananjara et al., 2007b; CITES, 2022). Despite heavy collection pressure and ongoing habitat loss, the demographic characteristics of *M. cowanii* populations remain largely unknown.

In 2008, a conservation strategy for *M. cowanii* was spearheaded by the IUCN Amphibian Specialist Group of Madagascar and Conservation International, which focused on improving habitat management at two localities near Antoetra (Rabibisoa 2008). A decade later, a workshop was organized to update, build on, and revitalize the initial conservation strategy. The 2018 workshop participants included officials from Malagasy government, academia, biodiversity conservation organizations, and local communities (Edmonds, Andreone & Crottini, 2022). The workshop resulted in the *Mantella cowanii* Action Plan (McAP), which was officially launched in 2021 (Andreone et al., 2020; Rakotoarison, Ndriantsoa & Rabemananjara, 2022). The McAP proposed 38 conservation actions needed for *M. cowanii*, with actions grouped into five themes: habitat protection, scientific research, local development, environmental awareness, and long-term sustainability. We aimed to fill the most critical research needs in the McAP by 1) confirming the presence of *M. cowanii* at localities across its range and 2) if present, estimating the key demographic traits of survival and population size.

#### **Materials & Methods**

141 Study Sites

We surveyed 11 of the 13 M. cowanii localities known from the literature and identified at the McAP workshop, nine in the Amoron'i Mania Region and two in the Vakinankaratra Region (Fig. 3). Sites ranged in elevation ~1380–2120 m asl. We repeatedly surveyed three sites (Ambatofotsy, Soamasaka, and Fohisokina) during 2020–2024 for 2–7 days/year to estimate demographic traits (Table 1). The sampling effort varied because of logistical constraints and security concerns, limiting our ability to survey for the same number of days annually. Additionally, we combined these data from 2020-2024 with a survey at Fohisokina in 2015, which lasted 20 days to accomplish additional research objectives related to habitat use (Newton-Youens, 2017). The remaining eight sites (Ambinanitelo, Andraholoma, Antakasina, Antsirakambiaty, Bekaraka, Farihimazava, Tsimabeomby, and Vatolampy) were visited up to two times for 1-6 days to confirm species presence. We worked closely with local communities, surveying sites together with people from nearby villages, and asked if there were other places people had seen M. cowanii to help identify additional sites. Only two sites have some form of habitat protection: Fohisokina (also known as Vohisokina) is community-managed with an NGO (Nowakowski & Angulo, 2015), and Antsirakambiaty falls within the boundaries of Itremo Massif Protected Area (Alvarado, Silva & Archibald, 2018). The other sites are unmanaged and without legal protection.

#### Data Collection

Like most other *Mantella* species, *M. cowanii* is highly seasonal in its behavior and is rarely observed before or after the rainy season. Additionally, prior fieldwork suggested *M. cowanii* is mainly active at dawn and dusk, with frogs hidden under refuge during midday (Tsiorisoa Andrianasolo 2016; Newton-Youens 2017). As such, we conducted fieldwork from late November to mid-January, surveying for frogs during 5–8h. In 2015, we also surveyed Fohisokina during 16–18h and combined these data with surveys from the morning of the same day. We did not survey during 16–18h in 2020–2024 or at other sites due to safety concerns about traveling after dark. We searched for frogs visually, walking together in teams of 2–7 people along the stream or adjacent wet rockface. Our search extended opportunistically up to 10–15 m from the stream or wet rock wall. If we heard a frog calling, we used the call to help find its location. Stream segment length varied from 38 m at Bekaraka to 690 m at Ambatofotsy. At Fohisokina, we surveyed along six 50 m-long transects rather than opportunistically throughout the entire site to accomplish additional research objectives (see Newton-Youens 2017).

After capturing a frog, we held it in a plastic bag (or a petri dish in 2015 at Fohisokina), marking the location with a GPS point, flagging tape, and a unique number. We measured snout-to-vent length to the nearest mm with digital calipers or a ruler.

Formatted: Highlight

178 Individuals <22 mm were recorded as subadults under one-year post-metamorphosis 179 based on Guarino et al. (2008). Weight was recorded with a digital scale to the nearest 180 0.01 g, and sex was recorded based on whether an individual had been calling before 181 capture and, if not, body size (Tessa et al. 2009). We also took dorsal and ventral 182 photographs, allowing us to identify recaptured individuals because each frog has a 183 unique ventral pattern (Fig. 4). To a certain extent, such photographic capture-mark-184 recapture techniques can be more accurate and are less invasive than traditional toe-185 clipping or visual implant elastomers (Caorsi, Santos, & Grant 2012; Davis, VanCompernolle, & Dickens 2020). Though software exists to help automate the photo-186 187 matching process (sensu Edmonds et al. 2019), we found photograph angle and quality varied between days, sites, and years, so the automated process was prone to false 188 189 negatives. Instead, one of us (D. Edmonds) visually examined all ventral photographs side-by-side and recorded when there was a match. Following photographs and 190 191 measurements, frogs were released in the location where they were found, typically 192 within 1-3 hours after capture.

We followed all applicable international, national, and institutional guidelines for the care and use of animals. The study methods were approved by the Ministère de l'Environnement et du Développement Durable in permits N°173/20/MEDD/SG/DGGGE/DAPRNE/SCBE.Re, N°439/21/ MEDD/SG/DGGGE/DAPRNE/SCBE.Re, and N°173/22/

198

MEDD/SG/DGGGE/DAPRNE/SCBE.Re and by the University of Illinois Urbana-

Champaign Institutional Animal Care and Use Committee in protocol #21180.

## Analysis

193

194

195

196

197

199

200

201

202

203

204

205

206

207

208

209

210

211

212

213

214

215

We used a robust design capture-mark-recapture model (Pollock 1982) to estimate population size ( $\mathbb{N}$ ) and apparent annual survival ( $\varphi$ ) at Ambatofotsy, Fohisokina, and Soamasaka. These sites were selected because we had sampled them for at least three years, the minimum required for estimating annual survival when detection is imperfect. The robust design uses primary periods when the population is assumed open to births, deaths, immigration, and emigration to estimate  $\varphi$  and secondary periods when the population is assumed closed to estimate N. We used annual surveys from late November to mid-January as open primary periods and days as closed secondary periods (Table 1).

Our analysis compared 17 models, all incorporating site as a group-level effect. We considered models with either constant or site-specific  $\varphi$  and the temporary emigration parameters  $\gamma''$  and  $\gamma'$  either constrained to 0 assuming no movement or set equal assuming random emigration. To account for individual heterogeneity in capture probability (p), we included a random effect of individual on p. Additionally, we compared models with capture probability covariates of site, year, number of surveyors, and survey effort calculated as the number of surveyors multiplied by the survey duration in minutes. We could not include environmental covariates that might be associated with capture probability because environmental variables were not collected consistently across all sites and years. To compare candidate models, we ranked them using Akaike's Information Criterion adjusted for small sample sizes (AICc; Burnham and Anderson 2002). Models with  $\Delta$  AICc < 2 were considered to have support. We analyzed capture-mark-recapture data in program MARK through the *RMark* interface (White & Burnham, 1999; Laake, 2013) and assessed the goodness-of-fit with package *R2ucare* in R version 4.2.0 (Gimenez et al., 2018; R Core Team, 2022).

To infer absence if we did not detect *M. cowanii* at a historic locality, we estimated detection probability with a single-season occupancy model in package *unmarked* (Fiske and Chandler 2011). Data from nine localities with confirmed presence and at least three surveys were used to generate a naïve estimate of detection probability assuming constant detection and occupancy. We then followed Pellet and Schmidt (2005) to estimate the number of surveys needed to detect the species as:

$$N_{min} = \frac{\ln(0.05)}{\ln(1-p)}$$

where  $N_{min}$  is the minimum number of surveys, p is the detection probability, and 0.05 is the confidence level needed to be 95% certain of absence, assuming independent and comparable surveys. Using the number of surveys, we then calculated the confidence level around an observed absence given N number of surveys as:

$$conf = e^{N*\ln(p)}$$

#### Results

#### Verifying Species Presence

We confirmed *M. cowanii* presence at 8 of 11 surveyed localities and identified 2 previously unrecorded populations. However, we failed to detect the species at the historical localities of Andraholoma, Tsimabeomby, and Vatolampy. The naïve detection probability from sites with confirmed presence was 0.86 (95% CI: 0.72–0.93), showing it takes 1.54 surveys (95% CI: 1.10–2.37) to be 95% confident a population is extirpated and 2.37 surveys (95% CI: 1.69–3.65) to be 99% confident. With at least two days of surveys at Andraholoma and Vatolampy during suitable climatic conditions, we can be >97.9% (95% CI: 91.9–99.9%) confident the populations are extirpated and 85.7% (95% CI: 71.7–93.4%) confident there are no *M. cowanii* at Tsimabeomby.

#### Capture Patterns Across Sites

We made 764 captures of 280 individuals across all study sites and years, excluding putative *M. baroni* x *M. cowanii* hybrids. All but 8 frogs were adults. Over half of all captures were at Fohisokina, where we caught 149 individuals 481 times. Six individuals at Fohisokina were recaptured 7 years after initially being caught in 2015 as adults (Fig. 4), one of which was recaptured again in year 8 in 2023. At our second most intensively surveyed site, Soamasaka, we made 137 captures of 40 individuals during annual fieldwork in 2020–2023. Three frogs were recaptured 4 years after their initial encounter in 2020, and 4 were recaptured 3 years apart. At Ambatofotsy, we conducted surveys in 2021, 2022, and 2023 and made 76 captures of 32 individuals, all adults.

#### Annual Survival and Population Sizes

The most parsimonious capture-mark-recapture model had capture probability p and annual adult survival  $\varphi$  varying by site (Table 2). Models with no movement generally performed better than those with random emigration (Table 2). The top model estimated population sizes (N) ranging from 13–137 adult frogs per site across years (Figs. 5–7). The highest N estimate was from Fohisokina in 2015 (137, 95% CI: 120–170) and the lowest from Soamasaka in 2023 (13, 95% CI: 11–22). Fohisokina showed a decreasing population size during 2015–2023 (Fig. 5), whereas Soamasaka and Ambatofotsy were relatively stable over a shorter period (Figs. 6, and 7). There was strong support for site-varying survival (Table 2), with the estimated annual adult survival more precise for Fohisokina than Soamasaka (Fig. 8). For Ambatofotsy, the survival estimate was too imprecise to be informative because the data spanned only three years and only a small number of frogs were captured each year. Overall, the annual survival estimates at Fohisokina and Soamasaka were comparable, although the estimate from Soamasaka was lower than Fohisokina (Fig. 8).

### **Discussion**

Our results demonstrate *M. cowanii* was still present at no less than ten localities in 2022–2023, but the population size is very small for at least three sites (<50 adults per site). Extrapolating across all known localities, in the worst-case scenario, the total adult population size for the species may number <500 individuals. However, frog populations naturally fluctuate in abundance, and a snapshot over several years can easily lead to erroneous conclusions that populations are declining when they are stable (Pechmann et al., 1991; Blaustein, Wake & Sousa, 1994; Meyer, Schmidt & Grossenbacher, 1998). Such fluctuations are typical of species with fast life histories, where fecundity is high and generation time short, thus, demographic rates tend to vary (Sæther et al., 2004, 2013). Additionally, amphibian populations fluctuate more for pond breeding species and less for terrestrial and stream breeding species (Green, 2003).

Formatted: Pattern: Clear (Yellow)

Deleted: A

Deleted: 5B Deleted: 5C

Deleted: 6

Deleted: 6

Considering *M. cowanii* is a terrestrial stream-breeding frog with comparatively low reproductive output (20–57 eggs per egg mass; Tessa et al. 2009), relatively high annual adult survival, and a maximum lifespan in nature of at least nine years, we believe our results are not an artifact of stochastic fluctuations in population size.

We highlight three possible factors contributing to the >80% population decline at Fohisokina between 2015 and 2023. First, fires burnt much of Fohisokina in November 2020 at the start of their breeding season, just before our survey. Though *M. cowanii* presumably is protected from fire while sheltering in moist rock crevices and caves for much of the year, fire can cause mortality in terrestrial frogs when they are active above ground during the breeding season (e.g., Humphries & Sisson 2012; Potvin et al. 2017). Second, according to a European private breeder, in 2017 more than 100 *M. cowanii* were illicitly offered for sale in Germany, and an unknown number were offered again in 2021. As Fohisokina is the most easily accessed *M. cowanii* locality and was historically a collection site for the pet trade (Rabemananjara et al., 2007b), the frogs were possibly poached from Fohisokina. Lastly, although no records of chytridiomycosis have been confirmed in Madagascar and all frogs we sampled appeared healthy, the amphibian chytrid fungus *Batrachochytrium dendrobatidis* has been reported from the island, and in 2014 was detected on a single *M. cowanii* individual at Soamasaka ~5 km south of Fohisokina (Bletz et al., 2015). Therefore, we cannot rule out disease either.

At Vatolampy, we did not detect M. cowanii after six days of surveys and suspect the population is extirpated. Until our work, the site had not been surveyed since 2003-2004, when Andreone et al. (2007) and Rabemananjara et al. (2007a) collected tissue samples and voucher specimens from the population. Similarly, Andraholoma had not been surveyed since 2009 when the site was visited by one of us (C. Randrianantoandro) for one day and 5 individuals observed. Conversely, we question whether Tsimabeomby, the third site where we did not detect M. cowanii, ever supported a population. We believe the locality was possibly published in error by Rabibisoa (2008) because Tsimabeomby consists of a wet meadow, is without rocks or running water, is isolated from the next nearest population by >3 km, and the local people we worked with had never encountered M. cowanii there whereas they knew of the other populations. Nonetheless, M. cowanii could have been present but undetected during our surveys if the detection probability at Andraholoma, Tsimabeomby, and Vatolampy was lower than elsewhere. Additionally, the naïve estimate of detection probability we used to infer absence did not account for observer, environmental, or temporal factors influencing detection. Still, our team detected M. cowanii at other sites on the days we surveyed Andraholoma, Tsimabeomby, and Vatolampy, illustrating local environmental conditions were favorable for detecting Mantella. We recommend re-surveying the three sites in the coming years to confirm if the populations are extirpated, especially

considering the site-level variation in capture probability we found at sites where *M. cowanii* was present.

328

329

330

331

332

333

334

335

336

337

338

339

340

341

342

343

344

345

346

347

348

349

350

351

352

353

354

355

356

357

358

359

360

361

362

363

364

365

366

Some amphibian species are capable of dispersing long distances to recolonize suitable habitat (Marsh & Trenham 2001; Fonte, Mayer, & Lötters 2019), but we do not expect so for M. cowanii in Madagascar's degraded highland landscape. There are no studies on the movement of Madagascar's poison frogs (but see Andreone et al. 2013), however research on their Neotropical dendrobatid counterparts shows adults rarely move more than a few hundred meters from established territories (e.g., Ringler et al. 2009, Pašukonis et al. 2013; Beck et al. 2017; Pašukonis et al. 2019). Moreover, in a review of the dispersal ability of amphibians, Smith and Green (2005) found nearly half of studied species moved <400m. Historically, adult frogs may have moved between patches when there was more forest in the highlands. However, we suspect Mantella usually passively disperse when tadpoles are flushed between habitat patches during heavy rain. As such, natural recolonization of Vatolampy or Andraholoma is unlikely, especially considering the physiological, movement, and site fidelity constraints amphibians face (Blaustein, Wake & Sousa, 1994). At Vatolampy, the next closest locality is Farihimazava, 1.5 km northeast in a different valley, which supports mostly M. baroni and M. baroni x M. cowanii hybrids (Chiari et al., 2005; Andreone et al. 2007). For Andraholoma, the next closest locality is Andaobatofotsivava >7 km north, though admittedly, the area of the Itremo Massif is poorly explored and there could be additional unrecorded populations between the two.

Previous skeletochronology research estimated the maximum lifespan of M. cowanii at three years (Guarino et al., 2008; Andreone et al., 2011), but we identified individuals at least eight years post-metamorphosis and one individual nine years. Skeletochronology is known to underestimate the ages of older individuals because skeletal growth rings progressively converge with age, and amphibian bone tissue is prone to reabsorption (Eden et al., 2007; Sinsch, 2015). Our results show the advantages of using capture-mark-recapture surveys for estimating demographic traits if resources are available. The long lifespan of M. cowanii is notable when considered together with their reproductive output and body size. The species is one of the largest in the genus, has the largest egg diameter, and has the lowest number of eggs per mass, exemplifying their slow life history compared to other Mantella species (Tessa et al. 2009). All of this aligns with our discovery that M. cowanii has the longest lifespan for the genus. Life history traits often follow altitudinal clines, with slower traits associated with higher altitudes (Hille & Cooper, 2015; Laiolo & Obeso, 2017). Indeed, amphibians tend to live longer and have larger body sizes at higher altitudes (Morrison & Hero, 2003; Andreone et al. 2004); M. cowanii is no exception (Tessa et al. 2009). Such patterns also occur within species across altitudinal gradients (Zhang & Lu, 2012). Considering the M. cowanii from Betafo populations occur ~500m higher than where we

Formatted: Highlight

recorded ~8–9-year-old individuals, frogs from Betafo may live even longer. Such slow life history traits also have conservation implications because they are associated with higher extinction risk (Webb, Brook & Shine, 2002). Given the relatively long lifespan and low reproductive output of *M. cowanii*, the success of recovery efforts for the species may not be as rapid as in other related amphibian species.

Local people brought our attention to two new *M. cowanii* localities during fieldwork, highlighting the value of community engagement when conducting research on threatened species. To our knowledge, the new localities had not been identified before but were noticed after our initial work conducting surveys together with local communities during 2020\_-2021\_, Indeed, people who live in biodiverse rural areas have a unique opportunity to assist in ecological research and monitoring programs (Schmiedel et al., 2016). Such opportunities are especially present in Madagascar (e.g., Dolch et al., 2015; Price, Randriamiharisoa & Klinges, 2023), where most people are subsistence farmers in rural areas and often depend heavily on forest resources. By actively participating and being included in fieldwork, local people recognized the significance of observing *M. cowanii* at new sites, helping inform conservation efforts.

To ensure populations remain extant, we must better identify the causes of declines and the magnitude of threats. Screening populations for Bd and monitoring for illicit M. cowanii in the pet trade are essential actions, while disentangling the complex threat of habitat loss presents additional challenges. The number of trees remaining at sites varies from intact closed-canopy forest to rocky landscapes almost entirely devoid of trees, so the degree to which deforestation is a threat may depend on additional habitat characteristics. Newton-Youens (2017) identified rock caves and refuges as essential habitat features, and speculated they might be used for breeding, though so far, no eggs, tadpoles, or newly metamorphosed individuals have been found in nature. Studies on the microhabitat preferences and activity levels of M. cowanii, like those carried out by Edwards et al. (2019, 2022) for M. aurantiaca and Rasoarimanana, Edmonds & Marquis (2024) for M. baroni, would further help identify the most critical habitat features to protect and the best time to survey sites. Likewise, better information about habitat requirements could be used to locate new sites with unprotected populations we do not know about. All known M. cowanii populations are centered around four isolated sites with likely past connectivity when the highlands were an intact forest-grassland mosaic (Bond, Silander & Ratsirarson, 2023). Fieldwork in the remote areas between the four population centers could uncover additional isolated M. cowanii populations, but time is running out.

#### **Conclusions**

We set out to verify the presence and estimate key demographic traits for one of Madagascar's most threatened frog species and found three historical localities may be

Deleted: 1

Deleted: 2

extirpated while other populations are extremely small. Unfortunately, our results are not unique to Madagascar but represent a global trend in amphibian populations (Stuart et al., 2004; Grant et al., 2016). Amphibians are at the forefront of the extinction crisis, and population monitoring is essential to measure responses to conservation actions and detect declines before recovery is impossible. We used capture-mark-recapture methods to estimate abundance at three localities (Ambatofotsy, Soamasaka, and Fohisokina; Fig. 3), but less costly approaches relying on presence-absence data are likely suitable for monitoring M. cowanii across its entire range (Joseph et al., 2006; Jones, 2011). When enacted with local people as part of a broader program, monitoring can galvanize conservation efforts by adding value to a threatened species and instilling pride in local communities (Andrianandrasana et al., 2005; Danielsen, Burgess & Balmford, 2005). Given that adequate information on a species basic ecology and life history is essential to addressing the causes of decline, we recommend further research run concurrently with conservation efforts and focus on determining the relative impact of disease, illegal trade, and habitat loss. We also recommend reassessing the IUCN Red List status of *M. cowanii*. The species was last assessed in 2014 as Endangered but it may qualify for the Critically Endangered status based on our estimates of

#### Deleted: S

Formatted: Highlight

Formatted: Highlight

#### Acknowledgements

population sizes and trends.

407

408

409

410

411

412

413

414

415

416

417

418

419

420

421

422

423

424

425

426

427

428

429

432

434

We are grateful to Chloe Helsey who assisted with fieldwork in 2015, to L'Homme et l'Environnement and V.O.I. FOMISAME for collaborative efforts at Antoetra, and to

Tiana Randriamboavonjy of the Kew Madagascar Conservation Centre for logistical

support at Itremo. Special thanks go to Association Mitsinjo members Frederic R.

430 Razafimahefa, Georges Ramarolahy, Samina S. Sam Edmonds, Edupsie

431 Tsimialomanana, and J.E.A. Fanirihasimbolatiana for providing indispensable field

assistance. We thank R. Griffiths, E. Larson, and R. Schooley for offering feedback on

433 an early draft of the manuscript.

437 438 439 440	Alvarado ST, Silva TSF, Archibald S. 2018. Management impacts on fire occurrence: A comparison of fire regimes of African and South American tropical savannas in different protected areas. <i>Journal of Environmental Management</i> 218:79–87. DOI: 10.1016/j.jenvman.2018.04.004.
441 442	AmphibiaWeb. 2023. AmphibiaWeb: Information on amphibian biology and conservation. Available at http://www.amphibiaweb.org (accessed March 9, 2023).
443 444 445 446	Andreone F, Andriantsimanarilafy RR, Crottini A, Edmonds D, Garcia G, Hansen-Hendrikx CM, Rakotoarison A, Razafimanahaka JH. 2020. <i>Mantella cowanii</i> Action Plan 2021–2025 / Plan d'Action <i>Mantella cowanii</i> 2021–2025. Museo Regionale di Scienze Naturali and the Amphibian Survival Alliance, Turin.
447 448	Andreone F, Bergò PE, Mercurio V, Rosa GM. 2013. Spatial ecology of <i>Scaphiophryne gottlebei</i> in the canyons of the Isalo Massif, Madagascar. <i>Herpetologica</i> 69:11–21.
449 450 451 452 453 454 455	Andreone F, Carpenter AI, Cox N, Du Preez L, Freeman K, Furrer S, Garcia G, Glaw F, Glos J, Knox D, Köhler J, Mendelson III JR, Mercurio V, Mittermeier RA, Moore RD, Rabibisoa NHC, Randriamahazo H, Randrianasolo H, Rasoamampionona Raminosoa N, Ravoahangimalala Ramilijaona O, Raxworthy CJ, Vallan D, Vences M, Vieites DR, Weldon C., 2008. The challenge of conserving amphibian megadiversity in Madagascar. <i>PLoS Biology</i> 6:943–946. DOI: 10.1371/journal.pbio.0060118.
456 457 458	Andreone F, Carpenter AI, Crottini A, D'Cruze N, Dubos N, Edmonds D, Garcia G, Luedtke J, Megson S, Rabemananjara FCE, Randrianantoandro JC, Randrianavelona R, Robinson J, Vallan D, Rosa GM, 2021. Amphibian
459 460 461	Conservation in Madagascar: Old and Novel Threats to a Peculiar Fauna. In: Heatwole H, Rödel M-O eds. <i>Status and Threats of Afrotropical Amphibians</i> . Frankfurt am Main: Chimaira, 243.
462 463 464	Andreone F, Giacoma C, Guarino FM, Mercurio V, Tessa G. 2011. Age profile in nine <i>Mantella</i> poison frogs from Madagascar, as revealed by skeletochronological analyses. <i>Alytes</i> 27:73–84.
465 466 467	Andreone F, Mercurio V, Mattioli F. 2006. Between environmental degradation and international pet trade: conservation strategies for the threatened amphibians of Madagascar. <i>Natura - Soc. it. Sci. nat. Museo civ. Stor. nat. Milano</i> 95:81–96.
468 469 470 471	Andreone F, Miaud C, Eusebio Bergò P, Doglio S, Stocco P, Riberon A, Gautier P. 2004. Living at high altitude: testing the effects of life history traits upon the conservation of <i>Salamandra lanzai</i> (Amphibia, Salamandridae). The Italian Journal of Zoology 71: 35–43.

436

References

Deleted: et al.

Deleted: et al

```
474
      Andreone F, Randrianirina JE. 2003. It's not a carnival for the harleguin mantella!
475
           Urgent actions needed to conserve Mantella cowani, an endangered frog from the
476
          high plateau of Madagascar. FrogLog 59: 1-2.
477
       Andreone F, Vences M, Glaw F, Randrianirina JE. 2007. Remarkable records of
478
           amphibians and reptiles on Madagascar's central high plateau. Tropical Zoology
479
           20:19-37.
480
      Andriambeloson JB, Blanco MB, Andriantsalohimisantatra A, Rivoharison T V., Walker
481
          N, Birkinshaw C, Yoder AD. 2021. Living in tiny fragments: a glimpse at the ecology
482
          of Goodman's mouse lemurs (Microcebus lehilahytsara) in the relic forest of
483
           Ankafobe, Central Highlands, Madagascar. Primates 62:887-896. DOI:
484
           10.1007/s10329-021-00947-1.
485
      Andrianandrasana HT, Randriamahefasoa J, Durbin J, Lewis RE, Ratsimbazafy JH.
486
           2005. Participatory ecological monitoring of the Alaotra wetlands in Madagascar.
487
           Biodiversity and Conservation 14:2757-2774. DOI: 10.1007/s10531-005-8413-y.
488
      Antonelli A, Smith RJ, Perrigo AL, Crottini A, Hackel J, Testo W, Faroog H, Torres
489
           Jiménez MF, Andela N, Andermann T, Andriamanohera AM, Andriambololonera S,
490
           Bachman SP, Bacon CD, Baker WJ, Belluardo F, Birkinshaw C, Borrell JS, Cable
191
           S, Canales NA, Carrillo JD, Clegg R, Clubbe C, Cooke RSC, Damasco G, Dhanda
192
           S, Edler D, Faurby S, de Lima Ferreira P, Fisher BL, Forest F, Gardiner LM,
493
           Goodman SM, Grace OM, Guedes TB, Henniges MC, Hill R, Lehmann CER, Lowry
194
           PP, Marline L, Matos-Maraví P, Moat J, Neves B, Nogueira MGC, Onstein RE,
495
           Papadopulos AST, Perez-Escobar OA, Phelps LN, Phillipson PB, Pironon S,
196
           Przelomska NAS, Rabarimanarivo M, Rabehevitra D, Raharimampionona J,
197
           Rajaonah MT, Rajaonary F, Rajaovelona LR, Rakotoarinivo M, Rakotoarisoa AA,
498
           Rakotoarisoa SE, Rakotomalala HN, Rakotonasolo F, Ralaiveloarisoa BA,
199
           Ramirez-Herranz M, Randriamamonjy JEN, Randriamboavonjy T, Randrianasolo
500
           V, Rasolohery A, Ratsifandrihamanana AN, Ravololomanana N, Razafiniary V,
501
           Razanajatovo H, Razanatsoa E, Rivers M, Sayol F, Silvestro D, Vorontsova MS,
502
           Walker K, Walker BE, Wilkin P, Williams J, Ziegler T, Zizka A, Ralimanana H, 2022.
503
           Madagascar's extraordinary biodiversity: Evolution, distribution, and use. Science
504
           378:eaf0869.
505
       Beck KB, Loretto MC, Ringler M, Hödl W, Pašukonis A. 2017. Relying on known or
```

exploring for new? Movement patterns and reproductive resource use in a tadpole-

Beyer RM, Manica A. 2020. Historical and projected future range sizes of the world's

mammals, birds, and amphibians. Nature Communications 11:1-8. DOI:

transporting frog. PeerJ:e3745. DOI: 10.7717/peerj.3745

10.1038/s41467-020-19455-9.

506

507

508

509

510

Deleted: et al.

512 513 514	Blaustein AR, Wake DB, Sousa WP. 1994. Amphibian declines: judging stability, persistence, and susceptibility of populations to local and global extinctions. <i>Conservation Biology</i> 8:60–71. DOI: 10.1046/j.1523-1739.1994.08010060.x.	
515 516 517 518 519	Bletz MC, Rosa GM, Andreone F, Courtois EA, Schmeller DS, Rabibisoa NHC, Rabemananjara FCE, Raharivololoniaina L, Vences M, Weldon C, Edmonds D, Raxworthy CJ, Harris RN, Fisher MC, Crottini A, 2015. Widespread presence of the pathogenic fungus Batrachochytrium dendrobatidis in wild amphibian communities in Madagascar. Scientific Reports 5:1–10. DOI: 10.1038/srep08633.	Deleted: et al
520 521 522	Bond WJ, Silander JA, Ratsirarson J. 2023. Madagascar's grassy biomes are ancient and there is much to learn about their ecology and evolution. <i>Journal of Biogeography</i> 50:614–621. DOI: 10.1111/jbi.14494.	
523 524	Burnham KP, Anderson DR. 2002. <i>Model Selection and Multimodel Inference: A Practical Information-Theoretic Approach</i> . New York: Springer-Verlag.	
525 526 527 528	Caorsi VZ, Santos RR, Grant T. 2012. Clip or snap? An evaluation of toe-clipping and photo-identification methods for identifying individual Southern Red-Bellied Toads, <i>Melanophryniscus cambaraensis</i> . South American Journal of Herpetology 7(2):79–84. DOI: 10.2994/057.007.0210.	
529	Ceballos G, Ehrlich PR, Barnosky AD, Garcia A, Pringle RM, Palmer TM. 2015.	
530 531 532	Accelerated modern human-induced species losses: Entering the sixth mass extinction. <i>Science Advances</i> 1:e1400253–e1400253. DOI: 10.1126/sciadv.1400253.	Formatted
<b>5</b> 31	extinction. Science Advances 1:e1400253-e1400253. DOI:	Formatted
531 532 533 534 535	extinction. Science Advances 1:e1400253—e1400253. DOI: 10.1126/sciadv.1400253.  Chiari Y, Andreone F, Vences M, Meyer A. 2005. Genetic variation of an endangered Malagasy frog, Mantella cowani, and its phylogeographic relationship to the widespread M. baroni. Conservation Genetics 6:1041–1047. DOI: 10.1007/s10592-	
531 532 533 534 535 536 537 538 539	extinction. <i>Science Advances</i> 1:e1400253—e1400253. DOI: 10.1126/sciadv.1400253.  Chiari Y, Andreone F, Vences M, Meyer A. 2005. Genetic variation of an endangered Malagasy frog, <i>Mantella cowani</i> , and its phylogeographic relationship to the widespread <i>M. baroni</i> . <i>Conservation Genetics</i> 6:1041–1047. DOI: 10.1007/s10592-005-9085-6.  Chiari Y, Vences M, Vieites DR, Rabemananjara F, Bora P, Ramilijaona Ravoahangimalala O, Meyer A. 2004. New evidence for parallel evolution of colour patterns in Malagasy poison frogs ( <i>Mantella</i> ). <i>Molecular Ecology</i> 13:3763–3774.	
\$31 532 533 534 \$35 536 537 538 539 540	extinction. <i>Science Advances</i> 1:e1400253—e1400253. DOI: 10.1126/sciadv.1400253.  Chiari Y, Andreone F, Vences M, Meyer A. 2005. Genetic variation of an endangered Malagasy frog, <i>Mantella cowani</i> , and its phylogeographic relationship to the widespread <i>M. baroni</i> . <i>Conservation Genetics</i> 6:1041–1047. DOI: 10.1007/s10592-005-9085-6.  Chiari Y, Vences M, Vieites DR, Rabemananjara F, Bora P, Ramilijaona Ravoahangimalala O, Meyer A. 2004. New evidence for parallel evolution of colour patterns in Malagasy poison frogs ( <i>Mantella</i> ). <i>Molecular Ecology</i> 13:3763–3774. DOI: 10.1111/j.1365-294X.2004.02367.x.	

548 549 550 551 552 553	OR, Dahlgren JP, Steiner UK, Bland LM, Gomez-Mestre I, Lebreton JD, González Vargas J, Flesness N, Canudas-Romo V, Salguero-Gómez R, Byers O, Bjørneboe Berg T, Scheuerlein A, Devillard S, Schigel DS, Ryder OA, Possingham HP, Baudisch A, Vaupel JW, 2019. Data gaps and opportunities for comparative and conservation biology. <i>Proceedings of the National Academy of Sciences of the United States of America</i> 116:9658–9664. DOI: 10.1073/pnas.1816367116.	Deleted: et al
554 555 556 557	Crottini A., Barbuto M., Casiraghi M., Andreone F. 2011. A rapid amphibian survey at Itremo-Ambatofinandrahana, central Madagascar, with confirmed absence of chytrid fungus and recommendations for future monitoring activities. <i>North-Western Journal of Zoology</i> 7:346–351.	
558 559 560 561	Crottini A., Brown JL, Mercurio V, Glaw F, Vences M, Andreone F. 2012.  Phylogeography of the poison frog <i>Mantella viridis</i> (Amphibia: Mantellidae) reveals chromatic and genetic differentiation across ecotones in northern Madagascar. <i>Journal of Zoological Systematics and Evolutionary Research</i> 50:305–314.	
562 563 564 565	Crottini A, Madsen O, Poux C, Strauß A, Vieites DR, Vences M. 2012. Vertebrate timetree elucidates the biogeographic pattern of a major biotic change around the K-T boundary in Madagascar. <i>Proceedings of the National Academy of Sciences of the United States of America</i> 109:5358–5363. DOI: 10.1073/pnas.1112487109.	Formatted
566 567 568 569	Crottini A, Orozco-terWengel P, Rabemananjara FCE, Hauswaldt JS, Vences M. 2019. Mitochondrial introgression, color pattern variation and severe demographic bottlenecks in three species of Malagasy poison frogs, genus <i>Mantella</i> . <i>Genes</i> 10:317. DOI:10.3390/genes10040317.	
570 571 572	Daly JW, Highet RJ, Myers CW. 1984. Occurrence of skin alkaloids in non-dendrobatid frogs from Brazil (Bufonidae), Australia (Myobatrachidae) and Madagascar (Mantellinae). <i>Toxicon</i> 22:905–919. DOI: 10.1016/0041-0101(84)90182-X.	
573 574 575	Danielsen F, Burgess ND, Balmford A. 2005. Monitoring matters: Examining the potential of locally-based approaches. <i>Biodiversity and Conservation</i> 14:2507–2542. DOI: 10.1007/s10531-005-8375-0.	
576 577 578	Davis HP, VanCompernolle M, Dickens J. 2020. Effectiveness and reliability of photographic identification methods for identifying individuals of a crypitically patterned toad. <i>Herpetological Conservation and Biology</i> 15(1):204–211.	
579 580 581 582	Dolch R, Ndriamiary JN, Randrianasolo M, Ramanantenasoa IA. 2015. Improving livelihoods, training para-ecologists, enthralling children: Earning trust for effective community-based biodiversity conservation in Andasibe, eastern Madagascar. <i>Madagascar Conservation &amp; Development</i> 10:21–28.	
583	Eden CJ, Whiteman HH, Duobinis-Gray L, Wissinger SA. 2007. Accuracy assessment	

585 586	of skeletochronology in the Arizona Tiger Salamander ( <i>Ambystoma tigrinum nebulosum</i> ). Copeia 2007:471–477.	
587 588 589	Edmonds D, Kessler E, Bolte L. 2019. How common is common? Rapidly assessing population size and structure of the frog <i>Mantidactylus betsileanus</i> at a site in east-central Madagascar. <i>Austral Ecology</i> 44:1196–1203. DOI: 10.1111/aec.12797.	
590 591 592	Edmonds D, Andreone F, Crottini A. 2022. Madagascar: An Amphibian Hotspot and the Conservation of the Harlequin Mantella Frog. In: DellaSala DA and Goldstein MI, eds. <i>Imperiled: The Encyclopedia of Conservation</i> . Amsterdam: Elsevier, 260–268.	
593 594 595	Edwards WM, Bungard MJ, Griffiths RA. 2022. Daily activity profile of the golden mantella in the "Froggotron"—A replicated behavioral monitoring system for amphibians. <i>Zoo Biology</i> 41:3–9. DOI: 10.1002/zoo.21650.	
596 597 598 599	Edwards WM, Griffiths RA, Bungard MJ, Rakotondrasoa EF, Razafimanahaka JH, Razafindraibe P, Andriantsimanarilafy RR, Randrianantoandro JC. 2019. Microhabitat preference of the critically endangered golden mantella frog in Madagascar. <i>Herpetological Journal</i> 29:207–213.	Formatted
600 601	Fischer EK, Roland AB, Moskowitz NA, Vidoudez C, Ranaivorazo N, Tapia EE, Trauger SA, Vences M, Coloma LA, O'Connell LA, 2019. Mechanisms of convergent egg	Deleted: et al
602 603	provisioning in poison frogs. <i>Current Biology</i> 29:4145-4151.e3. DOI: 10.1016/j.cub.2019.10.032.	
	, , , , , , , , , , , , , , , , , , , ,	
603 604	10.1016/j.cub.2019.10.032.  Fonte LFM, Mayer M, Lötters S. 2019. Long-distance dispersal in amphibians. <i>Frontiers</i>	
603 604 605 606 607	<ul> <li>10.1016/j.cub.2019.10.032.</li> <li>Fonte LFM, Mayer M, Lötters S. 2019. Long-distance dispersal in amphibians. Frontiers of Biogeography 11(4):e44577. DOI: 10.21425/F5FBG44577.</li> <li>Gimenez O, Lebreton JD, Choquet R, Pradel R. 2018. R2ucare: An R package to perform goodness-of-fit tests for capture—recapture models. Methods in Ecology</li> </ul>	
603 604 605 606 607 608 609 610	<ul> <li>10.1016/j.cub.2019.10.032.</li> <li>Fonte LFM, Mayer M, Lötters S. 2019. Long-distance dispersal in amphibians. Frontiers of Biogeography 11(4):e44577. DOI: 10.21425/F5FBG44577.</li> <li>Gimenez O, Lebreton JD, Choquet R, Pradel R. 2018. R2ucare: An R package to perform goodness-of-fit tests for capture—recapture models. Methods in Ecology and Evolution 9:1749–1754. DOI: 10.1111/2041-210X.13014.</li> <li>Granados-Martínez S, Zumbado-Ulate H, Searle CL, Oliveira BF, García-Rodríguez A. 2021. Niche contraction of an endangered frog driven by the amphibian chytrid</li> </ul>	Deleted: et al

declines. Herpetologica 76:101–107. DOI: 10.1655/0018-0831-76.2.101.

622	Green DM. 2003. The ecology of extinction: Population fluctuation and decline in
623	amphibians. Biological Conservation 111:331-343. DOI: 10.1016/S0006-
624	3207(02)00302-6.
005	Overing FM Occas M Terry O Andrews F 0000 Obertlife and of the observe

- Guarino FM, Sacco M, Tessa G, Andreone F. 2008. Short life span of two charismatic
   *Mantella* species: age-structure in the critically endangered *M. cowani* and in the
   syntopic *M. baroni*. In: Androne F ed. *A Conservation Strategy for the Amphibians* of Madagascar. Torino: Museo Regionale di Scienze Naturali, 265–276.
- Hille SM, Cooper CB. 2015. Elevational trends in life histories: revising the pace-of-life framework. *Biological Reviews* 90:204–2013.
- Humphries WJ, Sisson MA. 2012. Long distance migrations, landscape use, and
   vulnerability to prescribed fire of the Gopher Frog (*Lithobates capito*). *Journal of Herpetology* 46:665–670. DOI: 10.1670/11-124.
- 634 IUCN. 2023. The IUCN Red List of Threatened Species. Version 2022-2. *Available at http://www.iucnredlist.org* (accessed March 15, 2023).
- Jones JPG. 2011. Monitoring species abundance and distribution at the landscape scale. *Journal of Applied Ecology* 48:9–13. DOI: 10.1111/j.1365-2664.2010.01917.x.
- Joseph LN, Field SA, Wilcox C, Possingham HP. 2006. Presence-absence versus
   abundance data for monitoring threatened species. *Conservation Biology* 20:1679–
   1687. DOI: 10.1111/j.1523-1739.2006.00529.x.
- Laake JL. 2013. RMark: An R Interface for analysis of capture-recapture data with
   MARK. AFSC Processed Rep. 2013-1. Alaska Fish. Sci. Cent., NOAA, Seattle,
   WA.
- Laiolo P, Obeso JR. 2017. *Life-History Responses to the Altitudinal Gradient*. Cham,
   Switzerland: Springer Nature. DOI: 10.1007/978-3-319-55982-7\_11.
- Lehtinen R, Ramanamanjato J-B. 2006. Effects of rainforest fragmentation and
   correlates of local extinction in a herpetofauna from Madagascar. *Applied Herpetology* 3:95–110. DOI: 10.1163/157075406776984248.
- Luedtke JA, Chanson J, Neam K, Hobin L, Maciel AO, Catenazzi A, Borzée A, Hamidy
   A, Aowphol A, Jean A, Sosa-Bartuano Á, Fong G A, de Silva A, Fouquet A, Angulo
   A, Kidov AA, Muñoz Saravia A, Diesmos AC, Tominaga A, Shrestha B, Gratwicke
   B, Tjaturadi B, Martínez Rivera CC, Vásquez Almazán CR, Señaris C.
- Chandramouli SR, Strüssmann C, Cortez Fernández CF, Azat C, Hoskin CJ,
   Hilton-Taylor C, Whyte DL, Gower DJ, Olson DH, Cisneros-Heredia DF, Santana
- 656 DJ, Nagombi E, Najafi-Majd E, Quah ESH, Bolaños F, Xie F, Brusquetti F, Álvarez

657 658 659 660 661 662 663 664 665 666 667 668 669	FS, Andreone F, Glaw F, Castañeda FE, Kraus F, Parra-Olea G, Chaves G, Medina-Rangel GF, González-Durán G, Ortega-Andrade HM, Machado IF, Das I, Dias IR, Urbina-Cardona JN, Crnobrnja-Isailović J, Yang JH, Jianping J, Wangyal JT, Rowley JJL, Measey J, Vasudevan K, Chan KO, Gururaja KV, Ovaska K, Warr LC, Canseco-Márquez L, Toledo LF, Díaz LM, Khan MMH, Meegaskumbura M, Acevedo ME, Napoli MF, Ponce MA, Vaira M, Lampo M, Yánez-Muñoz MH, Scherz MD, Rödel MO, Matsui M, Fildor M, Kusrini MD, Ahmed MF, Rais M, Kouamé NGG, García N, Gonwouo NL, Burrowes PA, Imbun PY, Wagner P, Kok PJR, Joglar RL, Auguste RJ, Brandão RA, Ibáñez R, von May R, Hedges SB, Biju SD, Ganesh SR, Wren S, Das S, Flechas S V., Ashpole SL, Robleto-Hernández SJ, Loader SP, Incháustegui SJ, Garg S, Phimmachak S, Richards SJ, Slimani T, Osborne-Naikatini T, Abreu-Jardim TPF, Condez TH, De Carvalho TR, Cutajar TP, Pierson TW, Nguyen TQ, Kaya U, Yuan Z, Long B, Langhammer P, Stuart SN,
670 671	2023. Ongoing declines for the world's amphibians in the face of emerging threats. <i>Nature</i> 622:308–314. DOI: 10.1038/s41586-023-06578-4.
672 673	Marsh DM, Trenham PC. 2001. Metapopulation dynamics and amphibian conservation. Conservation Biology 15(1):40–49. DOI: 10.1111/j.1523-1739.2001.00129.x.
674 675 676	Meyer AH, Schmidt BR, Grossenbacher K. 1998. Analysis of three amphibian populations with quarter-century long time-series. <i>Proceedings of the Royal Society B: Biological Sciences</i> 265:523–528. DOI: 10.1098/rspb.1998.0326.
677 678	Morrison C, Hero J-M. 2003. Geographic variation in life-history characteristics of amphibians: a review. <i>Journal of Animal Ecology</i> 72:270–279.
679 680	Newton-Youens J. 2017. The importance of microclimates in the ex situ management of amphibians. MS Thesis. University of Manchester.
681 682	Nowakowski J, Angulo A. 2015. Targeted habitat protection and its effects on the extinction risk of threatened amphibians. <i>FrogLog</i> 23:8–10.
683 684 685 686	Patla DA, Peterson CR. 2022. The slow decline of a Columbia Spotted Frog population in Yellowstone National Park: A cautionary tale from a developed zone within a large protected area. <i>Ecological Indicators</i> 136:108606. DOI: 10.1016/j.ecolind.2022.108606.
687 688 689	Pašukonis A, Ringler M, Brandl HB, Mangione R, Ringler E, Hödl W. 2013. The homing frog: high homing performance in a territorial dendrobatid frog <i>Allobates femoralis</i> (Dendrobatidae). <i>Ethology</i> 119(9):762–768. DOI: 10.1111/eth.12116.
690 691 692	Pašukonis A, Loretto MC, Rojas B. 2019. How far do tadpoles travel in the rainforest? Parent-assisted dispersal in poison frogs. <i>Evolutionary Ecology</i> 33(4):613–623. DOI: 10.1007/s10682-019-09994-z.

Deleted: et al

694 695 696	Pechmann JHK, Scott DE, Semlitsch RD, Caldwell JP, Vitt LJ, Gibbons JW. 1991.  Declining amphibian populations: The problem of separating human impacts from natural fluctuations. <i>Science</i> 253:892–895. DOI: 10.1126/science.253.5022.892.	
697 698 699	Pellet J, Schmidt BR. 2005. Monitoring distributions using call surveys: Estimating site occupancy, detection probabilities and inferring absence. <i>Biological Conservation</i> 123:27–35. DOI: 10.1016/j.biocon.2004.10.005.	
700 701	Pollock KH. 1982. A capture-recapture design robust to unequal probability of capture. Journal of Wildlife Management 46:752–757.	
702 703 704 705	Potvin DA, Parris KM, Smith Date KL, Keely CC, Bray RD, Hale J, Hunjan S, Austin JJ, Melville J, 2017. Genetic erosion and escalating extinction risk in frogs with increasing wildfire frequency. <i>Journal of Applied Ecology</i> 54:945–954. DOI: 10.1111/1365-2664.12809.	Deleted: et al
706 707 708	Price F, Randriamiharisoa L, Klinges DH. 2023. Enhancing demographic diversity of scientist-community collaborations improves wildlife monitoring in Madagascar. Biological Conservation 288:110377. DOI: 10.1016/j.biocon.2023.110377.	
709 710	R Core Team. 2022. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computer, Vienna, Austria. https://www.R-project.org/.	
†11 712 713 714	Rabemananjara FCE, Chiari Y, Ramilijaona OR, Vences M. 2007a. Evidence for recent gene flow between north-eastern and south-eastern Madagascan poison frogs from a phylogeography of the <i>Mantella cowani</i> group. <i>Frontiers in Zoology</i> 4:1. DOI: 10.1186/1742-9994-4-1.	Formatted
715 716 717 718 719	Rabemananjara FCE, Raminosoa NR, Ravoahangimalala OR, Rakotondravony D, Andreone F, Bora P, Carpenter AI, Glaw F, Razafindrabe T, Vallan D, Vieites DR, Vences M, 2007b. Malagasy poison frogs in the pet trade: a survey of levels of exploitation of species in the genus Mantella. Amphibian and Reptile Conservation 5:3–16. DOI: 10.1514/journal.arc.0050020.	Deleted: et al
720 721	Rabibisoa N. 2008. <i>Plan d'Action de Conservation de Mantella cowani (PACM)</i> . Conservation International and the Amphibian Specialist Group of Madagascar.	
722 723 724 725	Rabibisoa N, Randrianasolo H, Anjeriniaina M, Mackinnon J, Andriamamonjisoa A, Randrianantoandro C, Andreone F. 2009. New findings of harlequin mantella improve the conservation status of Madagascar's most threatened frog. <i>Froglog</i> 92:5–8.	
726 727 728	Rakotoarison A, Ndriantsoa S, Rabemananjara F., Rabibisoa NHC, Rakotonanahary TF, Randriamahazo H, Andreone F. 2022. More than 15 years of amphibian conservation in Madagascar under the flag of IUCN SSC Amphibian Specialist	

Group. In: Biaggini M., Corti C., Giacobbe D., Lo Cascio P. eds. Herpetologia

Siciliae - XIII Congresso della Societas Herpetologica Italica (Lipari 22-26)

Siciliae - XIII Congresso della Societas Herpetologica Italica (Lipari 22-26 settembre 2021). Naturalista Siciliano 46:305–312.

731

732

733

734 735

736

737

738

739 740

741

742

743 744

745

746

747

748

749

750

751

752

753

754

755

756

757

758

759

760

761

762

763 764

765

766

Ralimanana H, Perrigo AL, Smith RJ, Borrell JS, Faurby S, Rajaonah MT, Randriamboavonjy T, Vorontsova MS, Luedtke JA, Chanson J, Neam K, Hobin L, Maciel AO, Catenazzi A, Borzée A, Hamidy A, Aowphol A, Jean A, Sosa-Bartuano <u>Á, Fong G A, de Silva A, Fouquet A, Angulo A, Kidov AA, Muñoz Saravia A, </u> Diesmos AC, Tominaga A, Shrestha B, Gratwicke B, Tjaturadi B, Martínez Rivera CC, Vásquez Almazán CR, Señaris C, Chandramouli SR, Strüssmann C, Cortez Fernández CF, Azat C, Hoskin CJ, Hilton-Taylor C, Whyte DL, Gower DJ, Olson DH, Cisneros-Heredia DF, Santana DJ, Nagombi E, Najafi-Majd E, Quah ESH, Bolaños F, Xie F, Brusquetti F, Álvarez FS, Andreone F, Glaw F, Castañeda FE, Kraus F, Parra-Olea G, Chaves G, Medina-Rangel GF, González-Durán G, Ortega-Andrade HM, Machado IF, Das I, Dias IR, Urbina-Cardona JN, Crnobrnja-Isailović J, Yang JH, Jianping J, Wangyal JT, Rowley JJL, Measey J, Vasudevan K, Chan KO, Gururaja KV, Ovaska K, Warr LC, Canseco-Márquez L, Toledo LF, Díaz LM, Khan MMH, Meegaskumbura M, Acevedo ME, Napoli MF, Ponce MA, Vaira M, Lampo M, Yánez-Muñoz MH, Scherz MD, Rödel MO, Matsui M, Fildor M, Kusrini MD, Ahmed MF, Rais M, Kouamé NGG, García N, Gonwouo NL, Burrowes PA, Imbun PY, Wagner P, Kok PJR, Joglar RL, Auguste RJ, Brandão RA, Ibáñez R, von May R, Hedges SB, Biju SD, Ganesh SR, Wren S, Das S, Flechas S V., Ashpole SL, Robleto-Hernández SJ, Loader SP, Incháustegui SJ, Garg S, Phimmachak S, Richards SJ, Slimani T, Osborne-Naikatini T, Abreu-Jardim TPF, Condez TH, De Carvalho TR, Cutajar TP, Pierson TW, Nguyen TQ, Kaya U, Yuan Z, Long B, Langhammer P, Stuart SN. 2023. Ongoing declines for the world's amphibians in the face of emerging threats. Nature 622:308-314. DOI: 10.1038/s41586-023-06578-4.

Potvin DA, Parris KM, Smith Date KL, Keely CC, Bray RD, Hale J, Hunjan S, Austin JJ, Melville J. 2017. Genetic erosion and escalating extinction risk in frogs with increasing wildfire frequency. *Journal of Applied Ecology* 54:945–954. DOI: 10.1111/1365-2664.12809.

Rabemananjara FCE, Raminosoa NR, Ravoahangimalala OR, Rakotondravony D, Andreone F, Bora P, Carpenter AI, Glaw F, Razafindrabe T, Vallan D, Vieites DR, Vences M. 2007. Malagasy poison frogs in the pet trade: a survey of levels of exploitation of species in the genus *Mantella*. *Amphibian and Reptile Conservation* 5:3–16. DOI: 10.1514/journal.arc.0050020.

Ralimanana H, Perrigo AL, Smith RJ, Borrell JS, Faurby S, Rajaonah MT,
Randriamboavonjy T, Vorontsova MS, Cooke RSC, Phelps LN, Sayol F, Andela N,
Andermann T, Andriamanohera AM, Andriambololonera S, Bachman SP, Bacon
CD, Baker WJ, Belluardo F, Birkinshaw C, Cable S, Canales NA, Carrillo JD, Clegg
R, Clubbe C, Crottini A, Damasco G, Dhanda S, Edler D, Farooq H, de Lima
Ferreira P, Fisher BL, Forest F, Gardiner LM, Goodman SM, Grace OM, Guedes
TB, Hackel J, Henniges MC, Hill R, Lehmann CER, Lowry PP, Marline L, Matos-

Ranarilalatiana T, Razafindraleva HA, Granath G, Bukontaite Malm R, Rakotonirina JC, Razafindranaivo V, Ravaomanarivo LHR, Johansson F, Bergsten J, 2022. Remaining forests on the Central Highlands of Madagascar—Endemic and endangered aquatic beetle fauna uncovered. Ecology and Evolution 12:1–26. DOI: 10.1002/ece3.9580.  Rasoarimanana T, Edmonds D, Marquis O. 2024. Habitat and local climate influence the activity and abundance of Baron's Mantella frog (Mantella baroni). Evolutionary Ecology 38:205–222. DOI: 10.1007/s10682-023-10272-2.  Ravoahangimalala OR, Raminosoa N, Rakotondravony D, Rabemananjara F, Bora P, Razafindrabe TJ. 2004. Donnée sur les Grenouilles du Genre Mantella spp. Endémique de Madagascar. Report number AC22 Doc. 10.3. Département de Biologie Animale, Universite d'Antananarivo.  Ringler M, Ursprung E, Hödl W. 2009. Site fidelity and patterns of short- and long-term movement in the brilliant thighed poison frog Allobates femoralis (Aromobatidae). Behavioral Ecology and Sociobiology 63:1281–1293. DOI: 10.1163/156853891X00329.  Sæther BE, Coulson T, Gretan V, Engen S, Altwegg R, Armitage KB, Barbraud C, Becker PH, Blumstein DT, Dobson FS, Festa-Bianchet M, Gaillard JM, Jenkins A, Jones C, Nicoll MAC, Norris K, Oli MK, Ozqul A, Weimerskirch H, 2013. How life history influences population dynamics in fluctuating environments. American Naturalist 182:743–759. DOI: 10.1086/673497.  Sæther BE, Engen S, Møller AP, Weimerskirch H, Visser ME, Fiedler W, Matthysen E, Lambrechts MM, Badyaev A, Becker PH, Brommer JE, Bukacinski D, Bukacinska M, Christensen H, Dickinson J, Du Feu C, Gehlbach FR, Heg D, Hölker H, Merilä J, Nielsen JT, Rendell W, Robertson RJ, Thomson DL, Török J, Van Hecke P, 2004.  Life-history variation predicts the effects of demographic stochasticity on avian	774 775 776 777 778 779 780 781 782 783	Maraví P, Moat J, Neves B, Nogueira MGC, Onstein RE, Papadopulos AST, Perez-Escobar OA, Phillipson PB, Pironon S, Przelomska NAS, Rabarimanarivo M, Rabehevitra D, Raharimampionona J, Rajaonary F, Rajaovelona LR, Rakotoarinivo M, Rakotoarisoa AA, Rakotoarisoa SE, Rakotomalala HN, Rakotonasolo F, Ralaiveloarisoa BA, Ramirez-Herranz M, Randriamamonjy JEN, Randrianasolo V, Rasolohery A, Ratsifandrihamanana AN, Ravololomanana N, Razafiniary V, Razanajatovo H, Razanatsoa E, Rivers M, Silvestro D, Testo W, Torres Jiménez MF, Walker K, Walker BE, Wilkin P, Williams J, Ziegler T, Zizka A, Antonelli A, (2022): Madagascar's extraordinary biodiversity: Threats and opportunities. <i>Science</i> 378:eadf1466.	(	Deleted: et al
the activity and abundance of Baron's Mantella frog ( <i>Mantella baroni</i> ). Evolutionary Ecology 38:205–222. DOI: 10.1007/s10682-023-10272-2.  Ravoahangimalala OR, Raminosoa N, Rakotondravony D, Rabemananjara F, Bora P, Razafindrabe TJ. 2004. Donnée sur les Grenouilles du Genre <i>Mantella</i> spp. Endémique de Madagascar. Report number AC22 Doc. 10.3. Département de Biologie Animale, Universite d'Antananarivo.  Ringler M, Ursprung E, Hödl W. 2009. Site fidelity and patterns of short- and long-term movement in the brilliant thighed poison frog <i>Allobates femoralis</i> (Aromobatidae).  Behavioral Ecology and Sociobiology 63:1281–1293. DOI: 10.1163/156853891X00329.  Sæther BE, Coulson T, Grøtan V, Engen S, Altwegg R, Armitage KB, Barbraud C, Becker PH, Blumstein DT, Dobson FS, Festa-Bianchet M, Gaillard JM, Jenkins A, Jones C, Nicoll MAC, Norris K, Oli MK, Ozgul A, Weimerskirch H, 2013. How life history influences population dynamics in fluctuating environments. <i>American</i> Naturalist 182:743–759. DOI: 10.1086/673497.  Sæther BE, Engen S, Møller AP, Weimerskirch H, Visser ME, Fiedler W, Matthysen E, Lambrechts MM, Badyaev A, Becker PH, Brommer JE, Bukacinski D, Bukacinska M, Christensen H, Dickinson J, Du Feu C, Gehlbach FR, Heg D, Hötker H, Merilä J, Nielsen JT, Rendell W, Robertson RJ, Thomson DL, Török J, Van Hecke P, 2004.  Life-history variation predicts the effects of demographic stochasticity on avian	†85 786 787	Razafindranaivo V, Ravaomanarivo LHR, Johansson F, Bergsten J, 2022.  Remaining forests on the Central Highlands of Madagascar—Endemic and endangered aquatic beetle fauna uncovered. <i>Ecology and Evolution</i> 12:1–26. DOI:	(	Deleted: et al
Razafindrabe TJ. 2004. Donnée sur les Grenouilles du Genre Mantella spp. Endémique de Madagascar. Report number AC22 Doc. 10.3. Département de Biologie Animale, Universite d'Antananarivo.  Ringler M, Ursprung E, Hödl W. 2009. Site fidelity and patterns of short- and long-term movement in the brilliant thighed poison frog Allobates femoralis (Aromobatidae).  Behavioral Ecology and Sociobiology 63:1281–1293. DOI: 10.1163/156853891X00329.  Sæther BE, Coulson T, Grøtan V, Engen S, Altwegg R, Armitage KB, Barbraud C, Becker PH, Blumstein DT, Dobson FS, Festa-Bianchet M, Gaillard JM, Jenkins A, Jones C, Nicoll MAC, Norris K, Oli MK, Ozgul A, Weimerskirch H, 2013. How life history influences population dynamics in fluctuating environments. American Naturalist 182:743–759. DOI: 10.1086/673497.  Sæther BE, Engen S, Møller AP, Weimerskirch H, Visser ME, Fiedler W, Matthysen E, Lambrechts MM, Badyaev A, Becker PH, Brommer JE, Bukacinska D, Bukacinska M, Christensen H, Dickinson J, Du Feu C, Gehlbach FR, Heg D, Hötker H, Merilä J, Nielsen JT, Rendell W, Robertson RJ, Thomson DL, Török J, Van Hecke P, 2004. Life-history variation predicts the effects of demographic stochasticity on avian	790	the activity and abundance of Baron's Mantella frog (Mantella baroni). Evolutionary		
movement in the brilliant thighed poison frog <i>Allobates femoralis</i> (Aromobatidae).  Behavioral Ecology and Sociobiology 63:1281–1293. DOI:  10.1163/156853891X00329.  Sæther BE, Coulson T, Grøtan V, Engen S, Altwegg R, Armitage KB, Barbraud C,  Becker PH, Blumstein DT, Dobson FS, Festa-Bianchet M, Gaillard JM, Jenkins A,  Jones C, Nicoll MAC, Norris K, Oli MK, Ozgul A, Weimerskirch H, 2013. How life history influences population dynamics in fluctuating environments. <i>American</i> Naturalist 182:743–759. DOI: 10.1086/673497.  Sæther BE, Engen S, Møller AP, Weimerskirch H, Visser ME, Fiedler W, Matthysen E, Lambrechts MM, Badyaev A, Becker PH, Brommer JE, Bukacinski D, Bukacinska  M, Christensen H, Dickinson J, Du Feu C, Gehlbach FR, Heg D, Hötker H, Merilä J, Nielsen JT, Rendell W, Robertson RJ, Thomson DL, Török J, Van Hecke P, 2004.  Life-history variation predicts the effects of demographic stochasticity on avian	793 794	Razafindrabe TJ. 2004. Donnée sur les Grenouilles du Genre <i>Mantella</i> spp.  Endémique de Madagascar. Report number AC22 Doc. 10.3. Département de	(	Formatted
Becker PH, Blumstein DT, Dobson FS, Festa-Bianchet M, Gaillard JM, Jenkins A, Jones C, Nicoll MAC, Norris K, Oli MK, Ozgul A, Weimerskirch H, 2013. How life history influences population dynamics in fluctuating environments. American Naturalist 182:743–759. DOI: 10.1086/673497.  Sæther BE, Engen S, Møller AP, Weimerskirch H, Visser ME, Fiedler W, Matthysen E, Lambrechts MM, Badyaev A, Becker PH, Brommer JE, Bukacinski D, Bukacinska M, Christensen H, Dickinson J, Du Feu C, Gehlbach FR, Heg D, Hötker H, Merilä J, Nielsen JT, Rendell W, Robertson RJ, Thomson DL, Török J, Van Hecke P, 2004. Life-history variation predicts the effects of demographic stochasticity on avian	797 798	movement in the brilliant thighed poison frog <i>Allobates femoralis</i> (Aromobatidae). <i>Behavioral Ecology and Sociobiology</i> 63:1281–1293. DOI:		
Lambrechts MM, Badyaev A, Becker PH, Brommer JE, Bukacinski D, Bukacinska M, Christensen H, Dickinson J, Du Feu C, Gehlbach FR, Heg D, Hötker H, Merilä J, Mielsen JT, Rendell W, Robertson RJ, Thomson DL, Török J, Van Hecke P, 2004. Life-history variation predicts the effects of demographic stochasticity on avian	801 802 803	Becker PH, <u>Blumstein DT</u> , <u>Dobson FS</u> , <u>Festa-Bianchet M</u> , <u>Gaillard JM</u> , <u>Jenkins A</u> , <u>Jones C</u> , <u>Nicoll MAC</u> , <u>Norris K</u> , <u>Oli MK</u> , <u>Ozgul A</u> , <u>Weimerskirch H</u> , 2013. How life history influences population dynamics in fluctuating environments. <i>American</i>	(	Deleted: et al
population dynamics. <i>American Naturalist</i> 164:793–802. DOI: 10.1086/425371.	806 807 808	Lambrechts MM, Badyaev A, Becker PH, Brommer JE, Bukacinski D, Bukacinska M, Christensen H, Dickinson J, Du Feu C, Gehlbach FR, Heg D, Hötker H, Merilä J, Nielsen JT, Rendell W, Robertson RJ, Thomson DL, Török J, Van Hecke P, 2004.	(	Deleted: et al

815 \$16 817 818	Schmiedel U, Araya Y, Bortolotto MI, Boeckenhoff L, Hallwachs W, Janzen D, Kolipaka SS, Novotny V, Palm M, Parfondry M, Smanis A, Toko P, 2016. Contributions of paraecologists and parataxonomists to research, conservation, and social development. <i>Conservation Biology</i> 30:506–519.	Deleted: et al
819 820	Sinsch U. 2015. Review: skeletochronological assessment of demographic life-history traits in amphibians. <i>Herpetological Journal</i> 25:5–13.	
821 822 823	Stuart SN, Chanson JS, Cox NA, Young BE, Rodrigues ASLL, Fischman DL, Waller RW. 2004. Status and trends of amphibian declines and extinctions worldwide. Science 306:1783–1786. DOI: 10.1126/science.1103538.	
824 825	Suzzi-Simmons A. 2023. Status of deforestation of Madagascar. <i>Global Ecology and Conservation</i> 42:e02389. DOI: 10.1016/j.gecco.2023.e02389.	
826 827 828 829	Tessa G, Mattioli F, Vincenzo M, Andreone F. 2009. Egg numbers and fecundity traits in nine species of <i>Mantella</i> poison frogs from arid grasslands and rainforests of Madagascar (Anura: Mantellidae). <i>Madagascar Conservation &amp; Development</i> 4:113–119.	
830 831 832	Tsiorisoa Andrianasola G. 2016. Etudes ecologique et biologique de <i>Mantella cowani</i> (Boulenger, 1882) dans la savane de Fohisokina, haute terre centrale Malgache. MS Thesis. Universite d'Antananarivo.	
833	Vences M, Glaw F, Bohme W. 1999. A review of the genus <i>Mantella</i> (Anura, Ranidae,	
834 835	Mantellinae): taxonomy, distribution and conservation of Malagasy poison frogs. <i>Alytes</i> 17:3–72.	Formatted
		Formatted
835 836 837	Alytes 17:3–72.  Vences M, Crottini A, Rabemananjara FCE, Randrianantoandro C, Tsiorisoa GA. 2022.  Mantellidae, Mantellinae: Mantella and Wakea. In: Goodman SM, ed. The New	Formatted
835 836 837 838 839 840 841	Alytes 17:3–72.  Vences M, Crottini A, Rabemananjara FCE, Randrianantoandro C, Tsiorisoa GA. 2022.  Mantellidae, Mantellinae: Mantella and Wakea. In: Goodman SM, ed. The New Natural History of Madagascar. Princeton: Pinceton University Press, 1360–1365.  Vieilledent G, Grinand C, Rakotomalala FA, Ranaivosoa R, Rakotoarijaona J-R, Allnutt TF, Achard F. 2018. Combining global tree cover loss data with historical national forest-cover maps to look at six decades of deforestation and forest fragmentation	Formatted
835 836 837 838 839 840 841 842 843	<ul> <li>Alytes 17:3–72.</li> <li>Vences M, Crottini A, Rabemananjara FCE, Randrianantoandro C, Tsiorisoa GA. 2022.         Mantellidae, Mantellinae: Mantella and Wakea. In: Goodman SM, ed. The New Natural History of Madagascar. Princeton: Pinceton University Press, 1360–1365.</li> <li>Vieilledent G, Grinand C, Rakotomalala FA, Ranaivosoa R, Rakotoarijaona J-R, Allnutt TF, Achard F. 2018. Combining global tree cover loss data with historical national forest-cover maps to look at six decades of deforestation and forest fragmentation in Madagascar. Biological Conservation 222:189–197. DOI: 10.1101/147827.</li> <li>Villero D, Pla M, Camps D, Ruiz-Olmo J, Brotons L. 2017. Integrating species distribution modelling into decision-making to inform conservation actions.</li> </ul>	Formatted
835 836 837 838 839 840 841 842 843 844 845	<ul> <li>Vences M, Crottini A, Rabemananjara FCE, Randrianantoandro C, Tsiorisoa GA. 2022. Mantellidae, Mantellinae: Mantella and Wakea. In: Goodman SM, ed. The New Natural History of Madagascar. Princeton: Pinceton University Press, 1360–1365.</li> <li>Vieilledent G, Grinand C, Rakotomalala FA, Ranaivosoa R, Rakotoarijaona J-R, Allnutt TF, Achard F. 2018. Combining global tree cover loss data with historical national forest-cover maps to look at six decades of deforestation and forest fragmentation in Madagascar. Biological Conservation 222:189–197. DOI: 10.1101/147827.</li> <li>Villero D, Pla M, Camps D, Ruiz-Olmo J, Brotons L. 2017. Integrating species distribution modelling into decision-making to inform conservation actions. Biodiversity and Conservation 26:251–271. DOI: 10.1007/s10531-016-1243-2.</li> <li>Webb JK, Brook BW, Shine R. 2002. What makes a species vulnerable to extinction? Comparative life-history traits of two sympatric snakes. Ecological Research 17:59–</li> </ul>	Formatted

851	marked animals. Bird Study 46:S120-S139. DOI: 10.1080/00063659909477239.
852 853 854 855 856 857	Yoder AD, Campbell CR, Blanco MB, Dos Reis M, Ganzhorn JU, Goodman SM, Hunnicutt KE, Larsen PA, Kappeler PM, Rasoloarison RM, Ralison JM, Swofford DL, Weisrock DW. 2016. Geogenetic patterns in mouse lemurs (genus <i>Microcebus</i> ) reveal the ghosts of Madagascar's forests past. <i>Proceedings of the National Academy of Sciences of the United States of America</i> 113:8049–8056. DOI: 10.1073/pnas.1601081113.
858 859 860 861	Zhang L, Lu X. 2012. Amphibians live longer at higher altitudes but not at higher latitudes. <i>Biological Journal of the Linnean Society</i> 106:623–632. DOI: 10.1111/j.1095-8312.2012.01876.x.