

# Reassessment of French breeding bird population sizes using citizen science and accounting for species detectability

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## ABSTRACT

Higher efficiency in large-scale and long-term biodiversity monitoring can be obtained through the use of Essential Biodiversity Variables, among which species population sizes provide key data for conservation programs. Relevant estimations and assessment of actual population sizes are critical for species conservation, especially in the current context of global biodiversity erosion. However, knowledge on population size varies greatly, depending on species conservation status and ranges. While the most threatened or restricted-range species generally benefit from exhaustive counts and surveys, monitoring common and widespread species population size tends to be neglected or is simply more challenging to achieve. In such a context, citizen science (CS) is a powerful tool for the long-term monitoring of common species through the engagement of various volunteers, permitting data acquisition on the long term and over large spatial scales. Despite this substantially increased sampling effort, detectability issues imply that even common species may remain unnoticed at suitable sites. The use of structured CS schemes, including repeated visits, enables to model the detection process, permitting reliable inferences of population size estimates. Here, we relied on a large French structured CS scheme (EPOC-ODF) comprising 27,156 complete checklists over 3,873 sites collected during the 2021–2023 breeding seasons to estimate the population size of 63 common bird species using hierarchical distance sampling (HDS). These population size estimates were compared to the previous expert-based French breeding bird atlas estimations, which did not account for detectability issues. We found that population size estimates from the former French breeding bird atlas were lower than those estimated using HDS for 65% of species. Such a prevalence of lower estimations is likely due to more conservative estimates inferred from semi-quantitative expert-based assessments used for the previous atlas. We also found that species with long-range songs such as the Common Cuckoo (*Cuculus canorus*), Eurasian Hoopoe (*Upupa epops*) or the Eurasian Blackbird (*Turdus merula*) had, in contrast, higher estimated population sizes in the previous atlas than in our HDS models. Our study highlights the need to rely on sound statistical methodology to ensure reliable

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ecological inferences with adequate uncertainty estimation and advocates for a higher reliance on structured CS in support of long-term biodiversity monitoring.

**Subjects** Biodiversity, Biogeography, Conservation Biology, Ecology

**Keywords** Bird atlases, Biogeography, Breeding bird surveys, Citizen science, Detectability, Hierarchical distance sampling

## INTRODUCTION

Worldwide bird populations are nowadays caught in the midst of a global, man-driven erosion of biodiversity caused by the synergistic effects of habitat destruction and fragmentation, resource overexploitation, climate change, pollution, pesticide use as well as the global spread of invasive species (Ceballos *et al.*, 2015; Johnson *et al.*, 2017). In Europe, the intensification of agriculture crystallises this phenomenon because the ever-increased use of pesticides and fertilisers has been pointed out as the main driver of current bird population declines (Rigal *et al.*, 2023). Recent researches have pointed out how important species-specific life attributes (*e.g.*, species range and density) and functional traits (*e.g.*, body mass, diet or dispersal) explain long-term bird trends and responses to global changes (Hong *et al.*, 2023; Santini *et al.*, 2023; Germain *et al.*, 2023). Monitoring long-term as well as shorter-term trends have been historically possible through standardised biodiversity—including bird—surveys at a national scale and aggregated at the continental one (Jiguet *et al.*, 2012; Pilotto *et al.*, 2020; Brlík *et al.*, 2021). This monitoring produces comprehensive distribution atlases pointing out current ranges and their historical changes (Keller *et al.*, 2020), as well as long-term population trends (Brlík *et al.*, 2021). Identifying common species declines in the long-term should be accounted for in agricultural and planning policies (Gaston & Fuller, 2008; Rigal *et al.*, 2023) to help reconciling society's needs with the safeguarding of ordinary biodiversity (Couvet & Ducarme, 2014).

Monitoring the success of the implementation of such policies is possible through the measurement of Essential Biodiversity Variables (Jetz *et al.*, 2019; Navarro *et al.*, 2017; Pereira *et al.*, 2013), including the assessment of species distribution range and population abundance or density (Santini *et al.*, 2023). Global monitoring capabilities of species populations have increased over the past few decades as citizen science (CS) has gained prominence across various domains, particularly in ecology (Kullenberg & Kasperowski, 2016). This participatory approach has provided the public with unprecedented opportunities to contribute to biodiversity monitoring through data collection and indirectly through their engagement in policy evaluation (McKinley *et al.*, 2017). The development of CS can be related to the emergence of multiple online databases (Newman *et al.*, 2012) capable of gathering comprehensive datasets across large spatial scales and multiple taxa such as iNaturalist ([www.inaturalist.org](http://www.inaturalist.org)) and BioloVision ([data.bioloVISION.net](http://data.bioloVISION.net)) or taxa-centred, for instance, eBird ([www.ebird.org](http://www.ebird.org)). The emergence of CS databases has thus resulted in an exponential increase in biodiversity monitoring capabilities, ranging from phenological shifts (Hurlbert & Liang, 2012), species distribution (Suzuki-

*Ohno et al., 2017; Matutini et al., 2021*) and abundance estimation (*Callaghan, Nakagawa & Cornwell, 2021*).

Traditionally, species autecology was recenssed in atlases providing temporal snapshots of known distribution and population size estimates using discontinuously gathered data collected during a short period (*Donald & Fuller, 1998*) and allowing long-term changes assessment when repeated over decades (*Keller et al., 2020*). The first attempt of French avifauna compilation dates back to 1936 (*Mayaud, de Balsac & Jouard, 1936*) while the first national atlas was published in 1976 (*Yeatman et al., 1976*) followed by subsequent in 1995 (*Yeatman-Berthelot & Jarry, 1995*) and 2015 (*Issa & Muller, 2015*). Each iteration was marked by a significant increase in participating citizen birders, ranging from 500 to 2,400 to 10,000. However, while the ultimate aim of bird atlases is to acquire even knowledge across multiple species for a given geographical area (typically national-wide inquiries), it is essential to note that rare and threatened species benefit from more in-depth population monitoring than more frequent and abundant ones (*Ingram et al., 2021*). Such exhaustive knowledge of rare species populations is due to interest risen by high extinction risk (*IUCN, 2001*), the need for recovery plans (*Farrier, Whelan & Mooney, 2007*) and narrow distributions allowing exhaustive counts (*Quaintenne et al., 2020*). In contrast, common species (being altogether abundant, widespread; *Rabinowitz, 1981*) are considered least concern (LC). They consequently tend to receive lesser attention than rarer species (*Neeson et al., 2018*) despite being key components of global avian biomass changes (*Gaston & Fuller, 2008; Inger et al., 2015; Whelan, Şekercioğlu & Wenny, 2015; Rigal et al., 2023*).

The principal cause of low quantitative coverage of common birds can be tied to the lack of specific funding stemming from a conservation prioritisation approach (*Brooks et al., 2006; Meine, Soulé & Noss, 2006*) and the trade-off between data quality and data acquisition over large spatial scales (*Devictor, Whittaker & Beltrame, 2010; Kamp et al., 2016*). In France, this incomplete knowledge translates into a significant drop in quality for population size estimates of most common bird species, where 60% of breeding species population estimates are qualified as medium (*i.e.*, state of knowledge of species abundance considered more or less satisfying, but semi-quantitative data are either lacking or outdated, see *Comolet-Tirman et al., 2015*) while for rare and localised species (24% of species), estimates are considered as highly reliable (*Comolet-Tirman et al., 2022*). To account for this discrepancy, a semi-quantitative estimation method was used for the last atlas survey, with population sizes simply inferred from the average number of breeding pairs measured over  $10 \times 10$  km grid cells using abundance classes of 1–9, 10–99, 100–999, 1,000–6,666 (the last upper limit could vary according to knowledge about species densities) multiplied by the number of grid cells known for nesting. Population sizes estimated using this method correspond to lower and upper limits. Lower limits were obtained by calculating the geometric mean of the abundance classes weighted by the number of  $10 \times 10$  grid cells belonging to these abundance classes while upper limits were

obtained by using the arithmetic means (see [Roché, Muller & Sibley, 2013](#) method referred further as *ArGeom*).

However, similarly to other studies ([Kellner & Swihart, 2014](#)), this approach fails to account for species detectability  $p$  defined as the probability of detecting at least one individual of a given species in a particular sampling effort, given that individuals of that species are present in the area of interest during the sampling session ([Boulinier et al., 1998](#)). Numerous studies have previously shown that  $p$  varies with time of day and season ([Skirvin, 1981](#)), observers ([Quinn et al., 2011](#)) and year-specific factors ([Kéry & Schmid, 2004](#)). Omitting species detectability by assuming perfect or constant  $p$  across sampling schemes, observers and habitat types can lead to biased inferences ([Nichols, Thomas & Conn, 2009](#); [Kéry, 2011](#)) and affect the estimation of long-term trends due to its unaddressed variation ([Schmidt, McIntyre & MacCluskie, 2013](#); [Sanz-Pérez et al., 2020](#)).

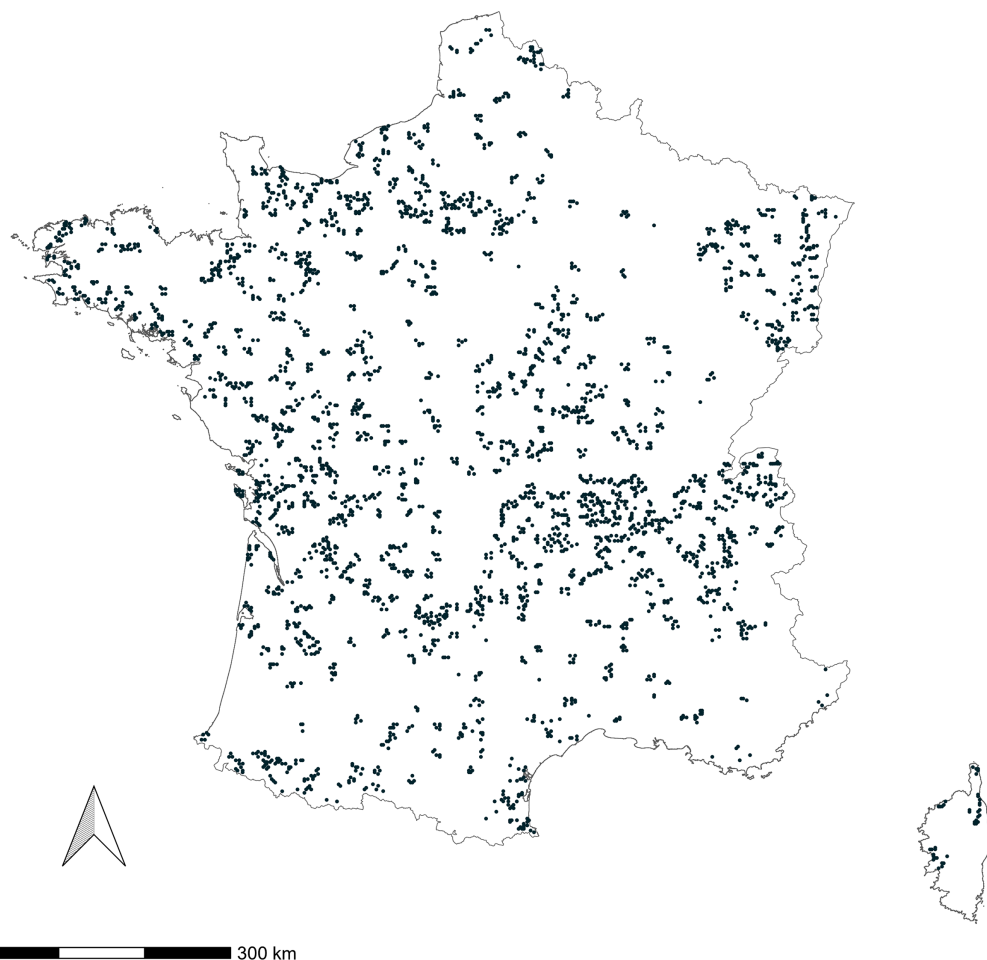
Here, we propose an estimation method enabling a more robust approach of population size estimations. We provide associated uncertainty intervals built upon a revised structured sampling scheme, ensuring data traceability and allowing inferences in the spatial variation of species abundance by formally including the detection process within the modelling framework.

In this study, we aimed at testing whether applying this modelling framework on an unprecedentedly large citizen-based dataset collected over France would (i) provide a new quantitative evaluation of French breeding bird populations and (ii) allow a comparison of population sizes inferred through Hierarchical Distance Sampling (HDS) from those inferred using the previous atlas methodology *ArGeom* across a large part of French avifauna. In particular, we expect that quantifying the influence of species detectability would allow more relevant ecological inferences (*e.g.*, including environmental and sampling effort covariates to the models) to approach closer to a realistic estimation of breeding bird population size at a national level than previously used methodologies. A preprint version of this article has been peer-reviewed and recommended by PCIEcology (<https://doi.org/10.24072/pci.ecology.100683>).

## MATERIALS AND METHODS

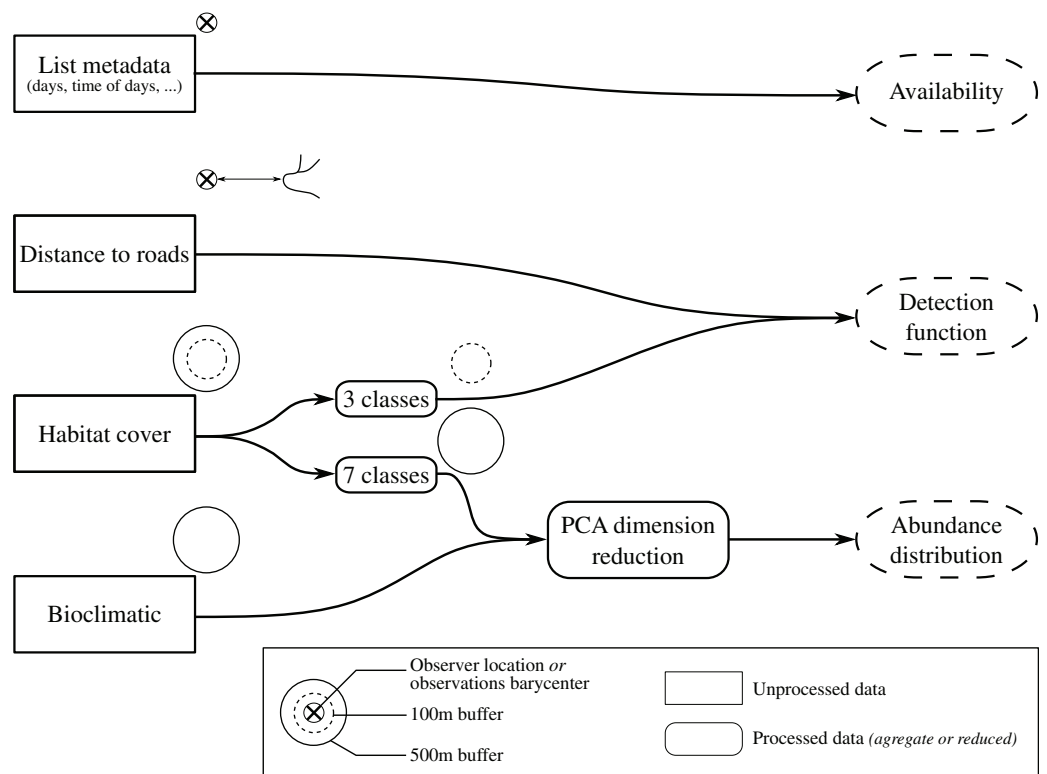
### Sampling protocol

EPOC-ODF (French structured estimation of breeding bird population size) is a French CS monitoring scheme based upon 5-min point counts, where observers are tasked to point locations of recorded individuals, either through visual or auditory detection. Birders can register their field observations directly using the NaturaList smartphone application or transcript later on the data portal Faune-France ([www.faune-france.org](http://www.faune-france.org)). The survey locations corresponded to the centroids of a  $2 \times 2$  km grid, selected from a random sampling. Each location has to be visited three times during the breeding season, from March to June, each consisting of three successive 5-min point counts, to limit chances of duplicated counts while being less demanding in observation effort ([Fuller & Langslow, 1984](#)). After completion, *i.e.*, nine visits during a breeding season, surveyed sites are removed from the sampling pool for the subsequent year, to maximise the number of sites surveyed. See [Materials S1](#) for more details about the sampling design.



**Figure 1** Spatial distribution of surveyed EPOC-ODF locations ( $n = 3,505$ ) over 2021–2023 breeding seasons. [Full-size](#)  DOI: [10.7717/peerj.17889/fig-1](https://doi.org/10.7717/peerj.17889/fig-1)

Over the 2021 and 2023 breeding seasons, 276 distinct species were encountered over 27,156 complete checklists collected over 3,873 pre-selected locations (Fig. 1) by 520 observers. Sampling effort is monitored through local associations tasked to recruit volunteers. The primary focus of the scheme being the monitoring of common breeding bird species, we decided to constrain the number of species considered viable targets of this scheme to 103 out of the 276 species contacted. We narrowed our study to 63 species out of the initial set of 103, comprising only those recorded at a minimum of 150 distinct locations (3.9% of total locations), to have a sample size allowing to reach model convergence. We also applied a temporal filter that considered both observed activity during the breeding season and expert opinion to capture the breeding phenology of each targeted species and exclude possible early or late migrants from population size estimates (see [Materials S2.1](#)).



**Figure 2** Global overview of covariates acquisition, treatments and usage workflow. Data are retrieved over observers' GPS location or approximated using observations barycenter, when unavailable, over two resolutions, 100 m buffer (dotted circles) and 500 m buffer radii (solid circles). Distance to roads is determined by measuring the distance between the nearest road to the observer location or observation barycenter. Habitat cover, in percentage, is aggregate over seven and three classes (see Table S3.1). Seven-class habitat cover and bioclimatic are reduced from PCA keeping the first three dimensions for bioclimatic data and three selected for the seven-class habitat cover.

Full-size DOI: 10.7717/peerj.17889/fig-2

## PCA reduction of environmental covariates

For bioclimatic data, we used 19 variables from WorldClim at 1 km resolution (Fick & Hijmans, 2017), on which we applied a Principal Component Analysis (PCA), keeping the first three axes (82.3% of explained variance), to limit multicollinearity through orthogonal transformation (Cruz-Cárdenas et al., 2014).

We used habitat cover data from Theia OSO at 10 m resolution (Thierion, Vincent & Valero, 2022) and aggregated it according to two different scales: (1) a seven-class corresponding to habitat type (Urban, Annual crops, Perennial crops, Pastures, Grasslands, Forests, Water body/Mineral surfaces) and (2) three-class (Open, Forests, and Artificial) in regards to overall effect on detectability (Fig. 2). Additionally, we conducted PCA dimension reduction on the seven-class aggregation, retaining three of the six PCA (54.71% of explained variance) axes depicting environmental gradients for (i) forest-to-open-field cultures; (ii) open-field cultures-to-pastures and (iii) perennial crop-to-urban Materials S3 for the workflow pipeline and habitat cover aggregation. Distances to roads were measured from ROUTE 500 (Cote et al., 2021). Environmental covariates were



**Table 1** Ensemble of sub-models tested in the secondary candidate set approach (Morin et al., 2020).

States	Sub-models
Detection	~ Distance to roads
	~ Distance to roads + Proportion of artificial lands (100 m)
	~ Distance to roads + Proportion of open lands (100 m)
	~ Distance to roads + Proportion of forests (100 m)
Availability	~ Julian date
	~ Julian date + Julian date <sup>2</sup>
	~ Hour (*)
	~ Hour + Hour <sup>2</sup>
	~ Julian date + Hour
	~ Julian date + Julian date <sup>2</sup> + Hour
	~ Julian date + Hour + Hour <sup>2</sup>
Abundance	~ 3 Bioclimatics PCA axis + 3 Habitat cover PCA axis

**Note:**

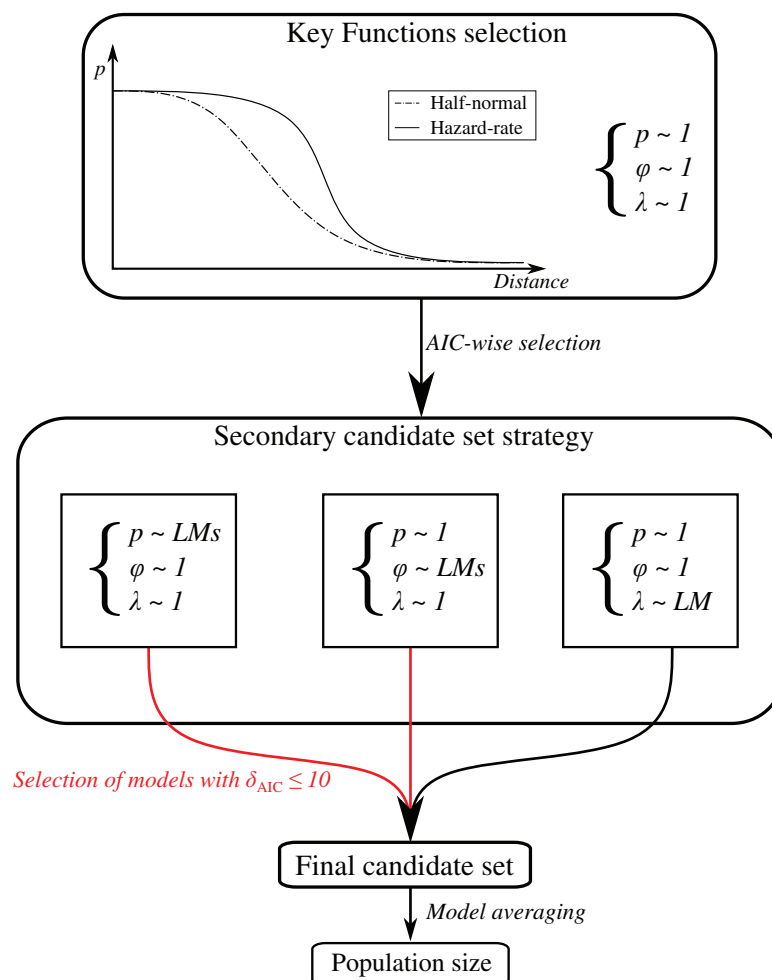
(\*) For the hour effort covariate, we used minutes from sunrise estimated from site longitude, latitude and date of list completion.

extracted over a 500 m buffer radii upon registered observer location (Fig. 2). These distances were chosen according to mean dispersal distances and home range sizes in common European birds (Paradis et al., 1998). The three-class habitat covers were collected upon 100 m circles radii to assess immediate habitat types that could hinder species detection. Whenever the exact location was unavailable, we used the centroid of sightings as a proxy for observer location (Materials S4). We used environmental data collected from a prediction grid covering France at a resolution of  $2 \times 2$  km for PCA dimension reduction. Outcomes from this initial PCA were used to transform environmental data collected from surveyed locations through PCA projections.

### Modelling framework

We used Hierarchical Distance Sampling (HDS) models to estimate the abundance of the target species while accounting for uncertainty arising from the observation process (Chandler, Royle & King, 2011; Kéry & Royle, 2015). We applied a right-side truncation of 5%, removing observation distances above the 95% quantile, for each targeted species to remove extreme distance values for model robustness (Buckland et al., 2001). Then, we divided observation distances into five proportional bin classes based on the maximal observed distance. Models calibration and assessment were done using unmarked 1.2.5 R package (Fiske & Chandler, 2011). Effort covariates were accounted for by incorporating the Julian date and the hours of list realisation (as minutes from sunrise), see Table 1.

Distance sampling key functions, depicting detection probabilities fall-off given distance of observation (Buckland et al., 2001), were chosen between half-normal and hazard-rate based on Akaike Information Criterion (AIC, Akaike, 1974), with other states kept constant.



**Figure 3 Methodological framework for population size estimation.** At first, a key function is determined AIC-wise between half-normal and hazard-rate with other components of the HDS kept constant. The selected key function is then used during the secondary candidate set strategy (Morin et al., 2020), fitting multiple sub-models (Table 1) for each state separately holding others constant. Sub-models with greatest support ( $\delta_{AIC} \leq 10$ ) are then selected in a final candidate set consisting of multiple HDS through combinatorial association (Morin et al., 2020). At this stage, we used sub-models estimated coefficients as starting values to help model convergence. Population size estimates are obtained through model averaging of the final candidate set. LM: Linear Model.

Full-size DOI: 10.7717/peerj.17889/fig-3

We based our modelling framework on a secondary candidate set strategy (Fig. 3), where the detection and availability states of our HDS were fit according to the set of the first candidates while others were kept constant (Morin et al., 2020). For the Poisson process underlying abundance distribution, we used a single model consisting of retained covariate PCAs axes (Table 1). See Table S5.1 for the number of times where each sub-process was included in the final candidate sets.

HDS population size estimates were obtained by averaging retained secondary candidate sets models, based on their relative model performance using AICc (Fig. 3). We excluded the Eurasian Sparrowhawk (*Accipiter nisus*), the Meadow Pipit (*Anthus pratensis*) and the Coal tit (*Periparus ater*), from model averaging and exclusively relied



upon prediction from best final models owing to substantial differences observed among their secondary candidate sets models.

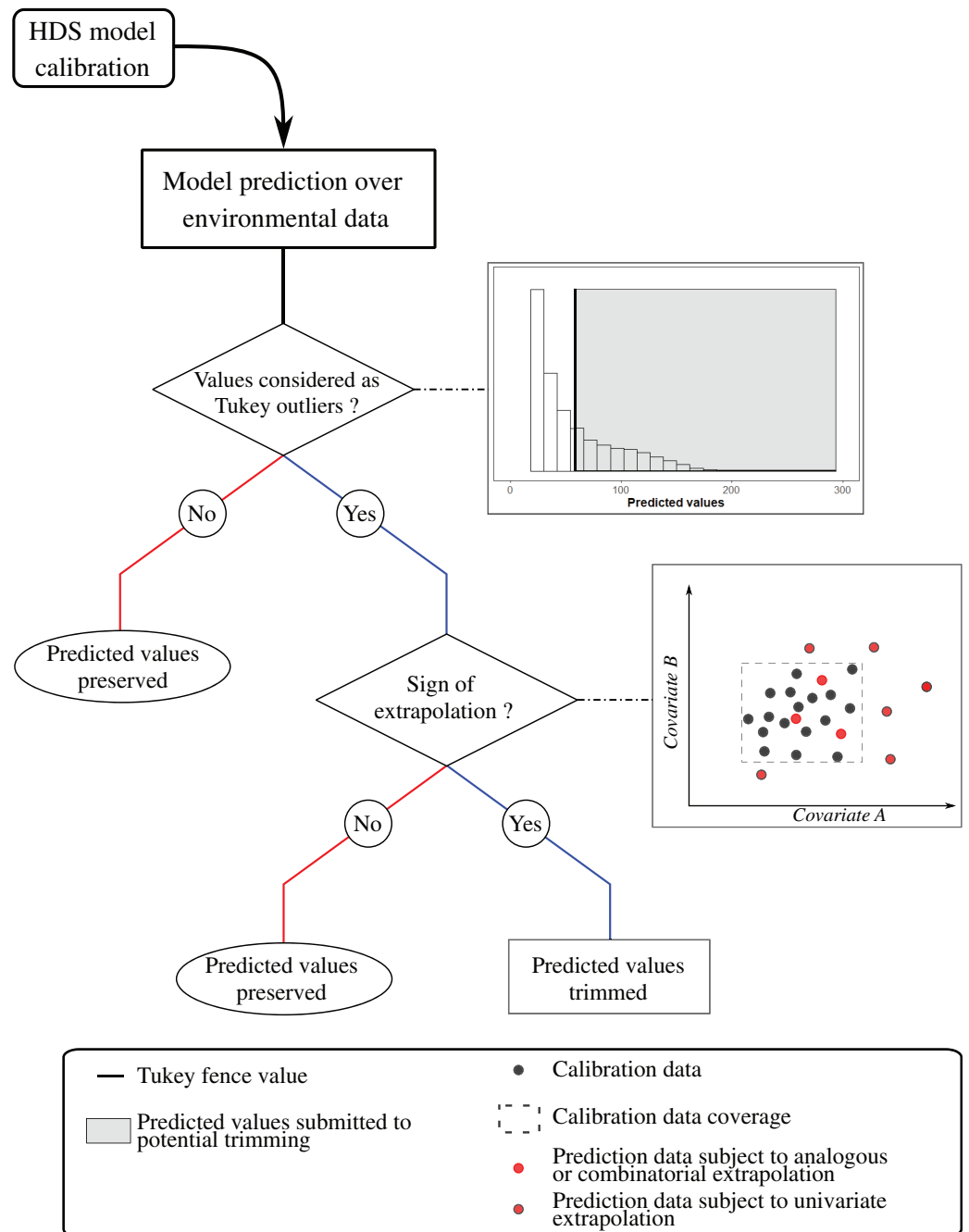
Model goodness-of-fit was assessed using an overdispersion coefficient metric ( $\hat{C}$ ; [Johnson, Laake & Ver Hoef, 2010](#)). We used the chi-square metric as the discrepancy measure between observed and expected counts. Computed  $\hat{C}$  corresponds to the ratio between the chi-square obtained from the fitted model to the mean of bootstrapped chi-squares obtained from simulated datasets based upon estimated parameters ([Kéry & Royle, 2015](#)). All models were fit according to a Poisson (P) distribution after top model assessment and calculation of  $\hat{C}$ , secondary candidate sets with  $\hat{C}$  top models exceeding 1.2 were calibrated using a negative binomial (NB) distribution ([Payne et al., 2018](#)). For a global overview of our modelling approach, see [Materials S5](#). Out of 63 species, we excluded nine species from the analysis; three exhibited signs of underdispersion with  $\hat{C}$  values less than 0.9 while six had  $\hat{C}$  values exceeding 1.5 ([Payne et al., 2018](#)), showing signs of overdispersion, despite being calibrated using a negative binomial distribution, see [Table S2.1](#) for more details.

We assessed the robustness of our estimations to the exclusion of one year of data, corresponding to a third of the global dataset. We compared population size estimates from EPOC-ODF data collected over 2021–2023 to estimates obtained from EPOC-ODF data collected over 2021–2022. Using the 2021–2022 subset, we estimated the population sizes of 30 species, detected in at least 150 distinct sites. From these 30 species, seven mean population sizes estimated using 2021 to 2023 data were outside the confidence intervals estimated from 2021 to 2022 data, with a slightly smaller population size estimated ([Table S2.2](#)) overall highlighting robust estimations.

### Trimming of HDS population size estimate: assessment of model extrapolation

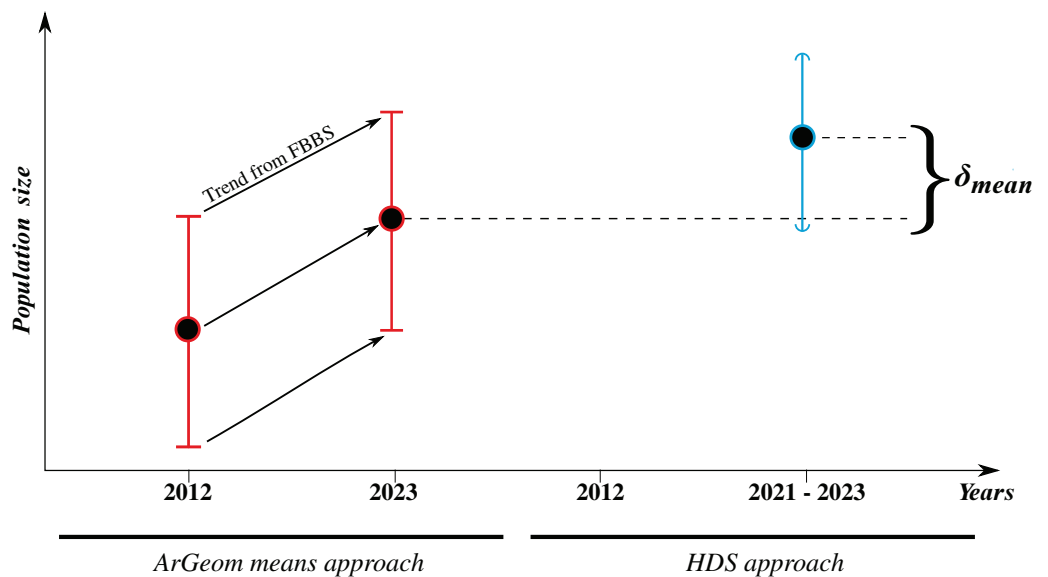
Population sizes were obtained by summing predicted values over the prediction grid. As we intend to predict over a large surface, novel environmental conditions may arise, leading to possible dissimilarities between the environmental gradient collected at survey sites and the environmental gradient over novel conditions ([Elith & Leathwick, 2009](#)). [Mesgaran, Cousens & Webber \(2014\)](#) described two types of extrapolation; (i) novelty type I (NT1) where projected points (*i.e.*, prediction grid) are outside the range of individual covariates collected by the sampling scheme and (ii) novelty type II (NT2) depicting the case when projected points are within univariate range but constitute novel combinations between covariates.

We trimmed predicted values ([Fig. 4](#)) over prediction grid cells showing signs of NT1 extrapolation, to a threshold value determined using a Tukey fence ([Tukey, 1977](#)), estimated from the distribution of predicted values with  $k = 1.5$ . Extrapolation assessments were done using *dsmextra* 1.1.5 R package ([Bouchet et al., 2020](#)). We used this post-prediction treatment to assess population size estimates stability. We measured the coefficient of variation, corresponding to the ratio between the standard deviation and the mean of the “untrimmed” and “outlier-trimmed” estimated range uncertainty. Large coefficients of variation imply great discrepancies in confidence intervals of untrimmed



**Figure 4** Decision tree of the post-prediction treatment. First, we analysed the distribution of predicted abundance values across the prediction grid and detected outliers, using a hinge of  $k = 1.5$  (Tukey, 1977). We compared each environmental condition of the prediction grid cell with the environmental condition collected by the sampling scheme and used for model calibration. When a prediction cell depicted signs of NT1 extrapolation and its estimated abundance was considered as an outlier, we trimmed the predicted grid cell abundance to the Tukey fences value.

Full-size DOI: [10.7717/peerj.17889/fig-4](https://doi.org/10.7717/peerj.17889/fig-4)



**Figure 5** Population size estimates comparison methodology, *ArGeom* in red and HDS in blue. *ArGeom* estimates for 2012 were obtained through calculations using both arithmetic and geometric means (Roché, Muller & Siblet, 2013) and were then updated to 2023 using specific mean trend estimates from FBBS (Table S2.1). Flat intervals signify min and max value estimates, while curved intervals signify confidence intervals. Full-size DOI: 10.7717/peerj.17889/fig-5

and outlier-trimmed estimated uncertainty intervals. This is mainly caused by the spatial filtering from the extrapolation assessment highlighting smaller geographic regions with similar environmental conditions from sampled ones and the trimming of predicted abundance outliers. Species with a coefficient of variation exceeding 30% were removed from the comparison of *ArGeom* and HDS population size estimates (Table S2.3).

### Comparison of *ArGeom* and HDS estimated population sizes

For comparable estimates between *ArGeom* and HDS approaches, we restricted the prediction grid area species-wise for HDS estimation according to the distribution of their known breeding locations, collected over a  $10 \times 10$  km grid during the previous French atlas (Issa & Muller, 2015). To estimate breeding populations of species for which male identification was possible, either male vocalisations or visual distinctions because of sexual dimorphism, an ad-hoc filter was applied (Table S2.1), resulting in HDS estimates reflecting the male counts for those species.

As the *ArGeom* approach estimated species bird population sizes as a number of breeding pairs (Roché, Muller & Siblet, 2013), for species where male identification in the field was impossible (no sexual dimorphism), we used all available data, after applying the phenological filter, and divided HDS estimates by two for comparable estimates with *ArGeom* population sizes. After retrieval of *ArGeom* estimates from the previous atlas (Issa & Muller, 2015), we updated these estimates using recent population trend estimates derived from the French breeding bird survey (FBBS; Jiguet et al., 2012) data spanning 2012–2023 (Table S2.1). Given the absence of a mean estimate in the *ArGeom* approach, we approximated it using the midpoint between the maximum and minimum estimated (Fig. 5).

To study the differences between the two approaches, we measured  $\delta_{mean}$  corresponding to the percentage of the difference between HDS and *ArGeom* estimates.

$$\delta_{mean} = \frac{(Estimates_{ArGeom} - Estimates_{HDS})}{(Estimates_{ArGeom} + Estimates_{HDS})/2}.$$

## Study of variation of estimated population sizes between the two approaches

As species detectability stems from physical traits and vocalisations, phylogenetic related species tend to have the same detectability (*Johnston et al., 2014; Sólymos et al., 2018*). We calibrated a Phylogenetic Generalised Linear Mixed Model (PGLMM) using the *phylolmm* 1.0 (*Li & Bolker, 2019*) R package. We study  $\delta_{mean}$  variations across species while implementing a random effect covariance structured based on phylogenetic relatedness using phylogenetic distances retrieved from *Burleigh, Kimball & Braun (2015)*. The PGLMM model was calibrated using (i) extracted detection probabilities from the availability state estimated through HDS (*Fig. 3* and *Materials S5*) after model averaging of the final candidate sets models in regards to AICc scores, and (ii) *ArGeom* uncertainty as fixed variables. For *ArGeom* uncertainty, corresponding to the difference between maximal and minimal estimated values, we relied on the decimal logarithm to limit variation in  $\delta_{mean}$  solely due to different population size magnitudes.

Response weights consisted of normalised weights from the inverse of uncertainty around FBBS trends between 2012 and 2023 (*Table S2.1*), divided by the mean to limit excessive weight attribution and facilitate model convergence.

## RESULTS

### Species trends over 2012–2023

From 2012 to 2023, out of 63 bird species, 15 showed a significant decrease ( $\bar{x} = -22.79\% \pm 14.84$ ) in total population size, while 16 showed a significant increase ( $\bar{x} = 28.02\% \pm 22.52$ ; see *Materials S2* for species-related FBBS trends).

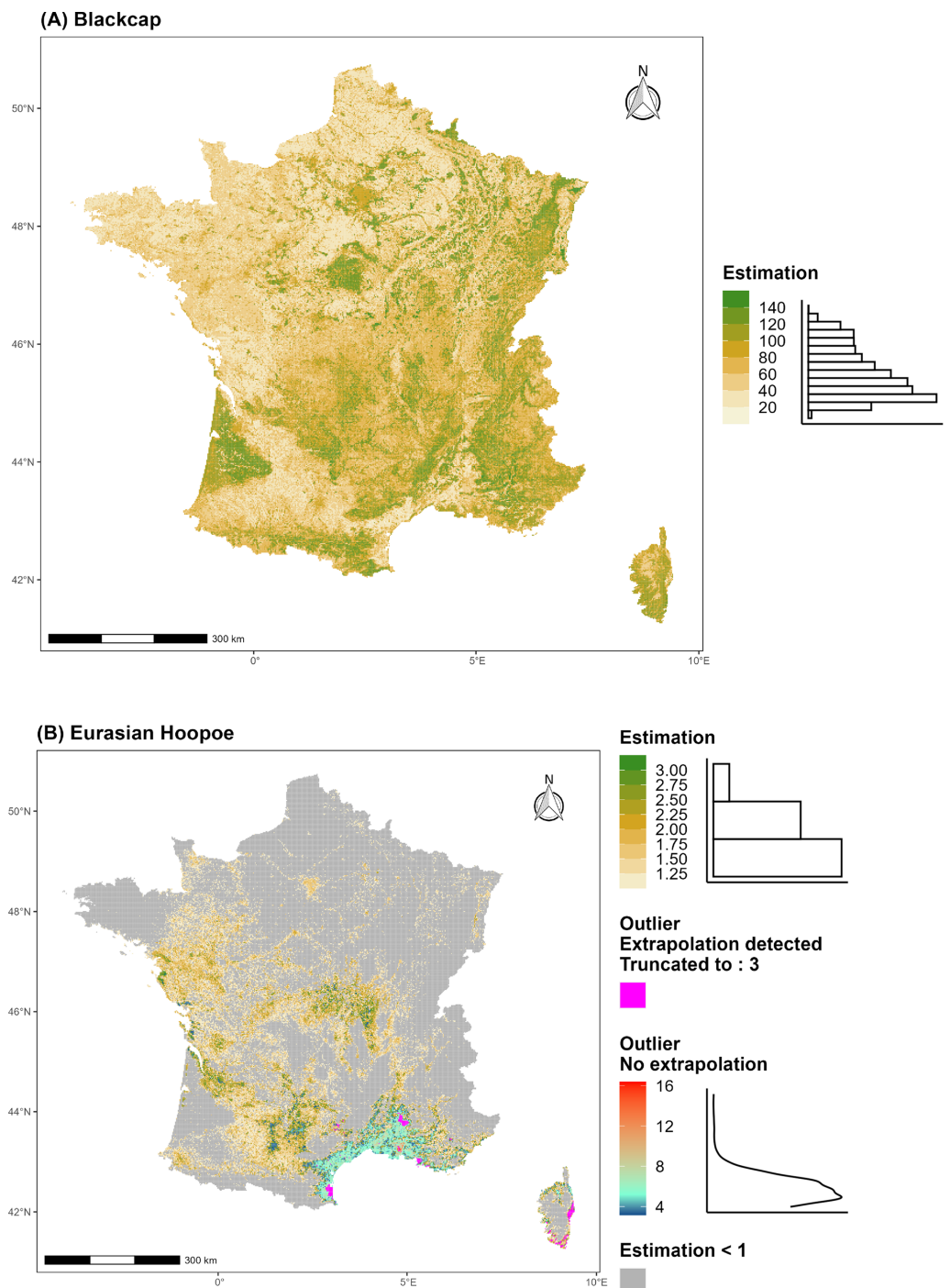
### HDS population size estimations

Out of the 54 species with acceptable values of overdispersion ( $\hat{C}$ ) using the *HDS* approach, we excluded eight species showing large discrepancies in population size estimates (*Materials S2, Table S2.3*) between pre- and post-prediction treatment (*Fig. 2.*).

Out of the remaining 46 species used for comparison between *ArGeom* and *HDS* estimates, *HDS* models showed acceptable values of overdispersion ( $\hat{C}$ ) ranging from 0.94 to 1.2 ( $\bar{x} = 1.07 \pm 0.06$ ) for 38 species calibrated using a Poisson distribution and 1.09 to 1.47 ( $\bar{x} = 1.27 \pm 0.13$ ) for eight species calibrated using a Negative binomial distribution.

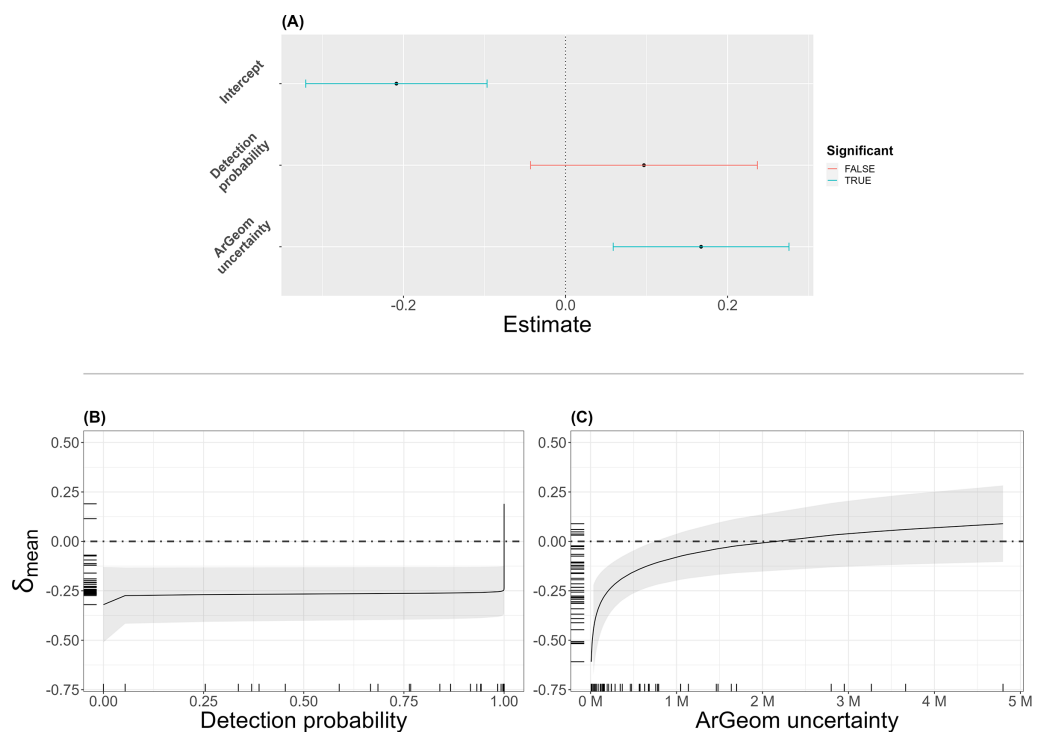
### Population size comparison between *ArGeom* and HDS

Across all species, estimated mean density ranges from 0.09 to 27.51 individuals per square kilometre, while *ArGeom* range uncertainty varies from 3.9 to 6.69 on the decimal logarithm scale corresponding to variations from 7,920 to 4,850,000 in estimated number



**Figure 6** Examples of resulting abundance maps. (A) Blackcap (*Sylvia atricapilla*), and (B) Eurasian Hoopoe (*Upupa epops*). Estimations correspond to the number of male, or potential pairs (see Table S2.3) over a 4 km-squared area. Grid cell predictions are categorised into three groups: (1) those with estimated abundance not considered as outliers depicted with a green colour step gradient and its distribution histogram; (2) those with estimated abundance considered as outliers and not subject to NT1 extrapolation, displayed in a blue-to-red gradient, along with their distribution density; and (3) those with estimated abundance considered as outliers with novel environmental conditions subject to NT1 extrapolation highlighted in pink with the Tukey value used for trimming in the post-prediction treatment.

Full-size DOI: [10.7717/peerj.17889/fig-6](https://doi.org/10.7717/peerj.17889/fig-6)



**Figure 7 Results from the PGLMM.** Confidence intervals of the model coefficient, coefficients significantly different from 0 are represented in blue. Marginal effect plots of population size estimate differences ( $\delta_{mean}$ ) responses.  $\delta_{mean}$  responses are predicted over gradients of focal terms, either species detection probabilities (B) or *ArGeom* reported uncertainties (C), while other covariates are held constant at their mean. Species detection probabilities and *ArGeom* uncertainty are represented on their natural scales, after inverse logit and inverse decimal logarithm transformation, respectively. Dot-dash line corresponds to a  $\delta_{mean}$  of 0, signifying estimated population size convergence by the two approaches, negative and positive values of  $\delta_{mean}$  reflect lower and higher population size estimates of *ArGeom* relative to those obtained using HDS.

Full-size DOI: [10.7717/peerj.17889/fig-7](https://doi.org/10.7717/peerj.17889/fig-7)

of pairs. See [Tables S6.1–S6.3](#) for more details about species estimated population size according to *ArGeom* and HDS approaches.

A comparison between updated *ArGeom* and HDS estimated population sizes showed that HDS estimates were higher than *ArGeom* for 30 of the 46 species tested ([Table S6.1](#)). Our results suggest lower estimates from *ArGeom* ( $\delta_{mean} < -0.2$ ), either for open habitat specialists such as European Stonechat (*Saxicola rubicola*), European Goldfinch (*Carduelis carduelis*) or Eurasian Linnet (*Carduelis cannabina*) than for forest generalists such as Great Spotted Woodpecker (*Dendrocopos major*), Blue Tit (*Cyanistes caeruleus*) or Eurasian Blackcap ([Fig. 6A](#)) with  $\delta_{mean} = -0.629 \pm 0.4$ , over 22 species. Species whose estimations were similar ( $\delta_{mean} \in [-0.2; 0.2]$ , with on average  $\delta_{mean} = -0.006 \pm 0.096$ , over 17 species) between *ArGeom* and HDS included species such as Eurasian Wren (*Troglodytes troglodytes*), Great Tit (*Parus major*), European Robin (*Erithacus rubecula*) and European Nuthatch (*Sitta europea*). Fewer species, mainly characterised by greater maximal observation distances, such as Common Cuckoo (*Cuculus canorus*), Eurasian Hoopoe ([Fig. 6B](#)) and European Blackbird (*Turdus merula*) had higher population sizes estimated by *ArGeom* approach compared to HDS ( $\delta_{mean} > 0.2$ , with on average



$\delta_{mean} = 0.41 \pm 0.148$ , over seven species; see [Materials S6](#) for population size comparison table).

Results from the PGLMM ([Fig. 7A](#)) showed an overall significantly lower *ArGeom* population size estimates ( $-0.209$ , with 95% CI  $[-0.321$  to  $-0.097]$ ,  $P_{val} = 0.001$ ), as well as a significant positive effect of *ArGeom* range intervals ( $0.167$ , with 95% CI  $[0.059$ – $0.276]$ ,  $P_{val} = 0.003$ ) on the differences between the two approaches. Species detection probabilities had no significant effect ( $0.097$ , with 95% CI  $[-0.043$  to  $0.237]$ ,  $P_{val} = 0.176$ ) on  $\delta_{mean}$  variation.

Marginal effect plots from the PGLMM model showed that the mean response of  $\delta_{mean}$  over species detection probability was predominantly negative, ranging from  $-0.45$  to  $-0.1$  ([Fig. 7B](#)), for *ArGeom* uncertainty. This showed that  $\delta_{mean}$  tended towards the convergence of population size estimates ([Fig. 7C](#)) for species with larger estimated interval ranges. There were no signs of multicollinearity ( $VIF < 5$ ; [James et al., 2013](#)) between the two variables.

## DISCUSSION

Our results showed that bird population size estimates from the previous *ArGeom* approach, not accounting for the observation process nor habitat affinity covariates, are predominantly lower than population sizes estimated from the HDS approach, up to 65% of species. While we found that the prior estimated uncertainty ranges from *ArGeom* had a positive effect on the convergence of population sizes estimated by the two methodological approaches (expert estimates based on atlas data *vs* predicted estimates from modelled citizen science data accounting for detection probabilities), we did not find a significant effect of species detection probabilities which could explain the differences between the two approaches. We show that *ArGeom* produces population sizes that are largely lower than those obtained by HDS ( $\delta_{mean} < 0.2$ ) regardless of habitat specialisation or affinity. This is likely due to the methodology used for *ArGeom* that did not account for the detection process nor for species-habitat relationships when extrapolating locally known abundances to unsampled locations.

Despite the lack of significant evidence for the effect of species detection probabilities on population size estimations, our results tend to corroborate previous studies where ignored detection processes had likely biased ecological inferences, including species distribution models ([Kéry, Gardner & Monnerat, 2010](#)), population trends ([Norvell, Howe & Parrish, 2003](#); [Schmidt, McIntyre & MacCluskie, 2013](#)) and population sizes ([Kéry, Royle & Schmid, 2005](#)). This lack may be especially true in a context of global change, where avian breeding phenology showed evidence of shifts towards earlier breeding over the years ([Parmesan, 2007](#); [Devictor et al., 2012](#); [Gaüzère & Devictor, 2021](#)) to synchronise with their food sources ([Visser, Holleman & Gienapp, 2006](#); [Michel et al., 2016](#)). Such shifts induce diverse species-related seasonal and inter-annual changes in detectability that need to be accounted for, particularly for schemes spanning over multiple species ([Lehikoinen, 2013](#)).

In France, the *ArGeom* approach was developed in the context of moderate semi-quantitative data collection with an acknowledged uneven participation across the territory ([Roché, Muller & Sibley, 2013](#); [Issa & Muller, 2015](#)). The semi-quantitative data

collection was based on the estimation of the number of breeding pairs over  $10 \times 10$  km grids derived from a mixture of count prospects and expert opinions collected over 1,953 out of 5,879  $10 \times 10$  km grids (Issa & Muller, 2015). As the primary goal of this approach was to give a likely magnitude of population size across the territory (Roché, Muller & Sibley, 2013), ArGeom intervals were produced by the extrapolation from the initially prospected  $10 \times 10$  km grids to all metropolitan grid cells considered suitable for breeding, after the detection of potential and confirmed breeding evidence. Although proved useful and relevant to assess population sizes when large-scale quantitative data on species occurrences are lacking or are unevenly distributed, such a methodology implies greater uncertainty intervals for abundant and broadly distributed species and smaller intervals for scarce and narrowly distributed species when not accounting for measurement uncertainty.

Contrary to generalist species, which have widespread distributions due to broader habitat niche breadths, specialists are generally more localised (Clavel, Julliard & Devictor, 2011) and typically use a smaller range of habitats (Julliard et al., 2006). Despite a growing interest in rare species-focused monitoring (Fontaine et al., 2022), citizen sciences programs are mainly designed for large-scale multi-species surveys (Devictor, Whittaker & Beltrame, 2010). Citizen science schemes balance between a trade-off among data quantity and data quality, corresponding to either the acquisition of a great quantity of unstructured scheme, or the acquisition of standardised data implying replicated visits over randomly sampled locations (Devictor, Whittaker & Beltrame, 2010). As such, in the first case, citizen science schemes could be more prone to false-negative errors, resulting in biased inferences over habitat cover relations due to omission of the detection process (Johnston, Matechou & Dennis, 2022). In the second case, given the small habitat range of specialist species and the scale of the territory sampled (e.g., here the metropolitan French territory), citizen science sampling schemes could be representative of the entire territory sampled but with a higher risk of missing some key habitats and associated specialist species.

### Potential consequences for community-level assessments

A recent study about long-term effects of climate and land use changes on bird communities (Gaiüzère et al., 2020) showed that both generalist cold-dwelling species, such as the Common Chiffchaff (*Phylloscopus collybita*) or the Eurasian Blackcap, and warm-dwelling species, such as the Common Nightingale (*Luscinia megarhynchos*) had the most substantial negative and positive contributions to the trend in Community Thermal Index (CTI), a community-weighted index representing the realised thermal niche of a community based upon species relative abundance and species thermal indices (STI). In the present study, these species tended to have lower population sizes estimated when the detection process was omitted compared to estimates based on our modelling approach. As a result, this could affect the estimations of their contribution to the calculation of community-weighted mean indices, such as CTI, and therefore bias the estimation of the trend in community thermal response and subsequent studies of aggregated indices, which are known to display large regional variation (Rigal & Knape, 2024). We, therefore, suggest that considering the detection process in studies relying on community-weighted indexes

by species' relative abundances could be as important as it is for estimating population sizes.

Community indices such as species diversity (Ricotta, 2005) and functional diversity (Villéger, Mason & Mouillot, 2008; Gaüzère et al., 2019) are commonly use species relative abundance as a basis, without taking into account the detection process (Pillar & Duarte, 2010), despite multiple studies showing it could affect community indices inference (Tingley & Beissinger, 2013; McNew & Handel, 2015; Jarzyna & Jetz, 2016; Richter et al., 2021).

### Conservation implications

Our study also suggested that lower or higher population sizes estimated from *ArGeom* were not randomly distributed among species according to their conservation status. Out of the 46 species estimations used in the comparison analysis, 10 had an unfavourable conservation status in France (*i.e.*, lower than Least Concern, LC; UICN France et al., 2016).

Among these species of conservation concern, two species, European Greenfinch (*Carduelis chloris*) and European Turtle Dove (*Streptopelia turtur*), showed no signs of difference in their population sizes. By contrast, five species, European Stonechat, Barn Swallow (*Hirundo rustica*), Red-backed Shrike (*Lanius collurio*), Eurasian Kestrel (*Falco tinnunculus*) and Willow Warbler (*Phylloscopus trochillus*) considered as NT (Near Threatened) and three species, Eurasian Linnet, European Goldfinch and European Serin (*Serinus serinus*) considered as VU (Vulnerable) had lower population sizes estimated from *ArGeom* than from HDS approach (NT:  $\bar{\delta}_{mean} = -0.608 \pm 0.217$  and VU:  $\bar{\delta}_{mean} = -0.667 \pm 0.146$ ). Our results showed that these species may need a reevaluation of their conservation status and highlight the need to rely on hierarchical models taking account of the detection process in ecological inferences, given that potential misclassification of population conservation status may arise from process noise and observation error (Connors et al., 2014). As conservation policy decisions depend on uncertainty levels (Williams, 2003; Freckleton, 2020), assessing measurement error through the integration of the detection process (Nichols et al., 2011) could provide more reliable ecological inferences (Guillera-Arroita et al., 2014). CS schemes are becoming more and more a reliable source of data to ensure biodiversity monitoring (Chandler et al., 2017) and can, through standardisation (Buckland & Johnston, 2017; Johnston et al., 2019), contribute to the calibration of data-hungry models such as hierarchical models for reliable ecological inferences (Isaac et al., 2020; Kéry & Royle, 2021; Johnston, Matechou & Dennis, 2022).

### Comparison to other European countries

Another way to assess the relevance of the two estimation approaches would be to compare their population size estimates to the ones obtained from other European countries, using a ratio between countries to produce comparable estimates. Such an approach should however be used with caution because it would be limited by comparability in habitat repartitions or biogeographical considerations among different European countries. To go further into inter-country comparisons, we relied on the German population sizes

estimated for the previous European Bird Directive (*BirdLife International, 2021*) obtained from both point count and territory mapping methods (*Gedeon et al., 2015*). For abundant species such as the Blackcap ( $\delta_{mean} = -0.29$ ; German population size expressed in millions of pairs = [7.17–9.49]), both approaches led to similar results than German population estimates, while HDS estimates were closer to German population sizes for the Firecrest (*Regulus ignicapilla*;  $\delta_{mean} = -0.46$ ; [1.92–2.85]) and the Blue Tit ( $\delta_{mean} = -0.42$ ; [5.01–7.41]). For species with higher population sizes estimated by *ArGeom* than HDS ( $\delta_{mean} > 0.2$ ), the Common Cuckoo ( $\delta_{mean} = 0.25$ ; [0.58–0.95]) and the Corn Bunting (*Emberiza calandra*;  $\delta_{mean} = 0.6$ ; [0.25–0.44]) showed estimates of German populations closer to the HDS than the *ArGeom* approach. Finally, for the Common Whitethroat (*Curruca communis*;  $\delta_{mean} = 0.42$ ; [0.93–1.47]), the German population size is closer to *ArGeom* estimates (see [Materials S6.4](#) for additional information).

Regarding magnitudes, both approaches produced similar estimates compared to German ones. However, due to different sampling and modelling methods used, these formal comparisons, although informative, need to be more fully satisfying and highlight the discrepancies in sampling and analytical methods across the European continent (*Keller et al., 2020*). Such differences could be accounted for, either by (i) a global standardisation of schemes, as promoted by the PECBMS (Pan-European Common Bird Monitoring Scheme; *Brlík et al., 2021*) for species trends, but also (ii) through the use of Integrated Models (IM) capable of mobilising data from multiple and somewhat heterogeneous sources (*Isaac et al., 2020*; *Zipkin et al., 2021*).

### Study limitations

Our approach relies on data collected from the EPOC-ODF structured CS schemes, providing data with repeated visits. However, as is, the frequentist framework of *unmarked* R package (*Kellner et al., 2023*) does not permit inferences on social species occurring in large flocks. Taking account of social species during the breeding season (corresponding to 1/10th of the scheme targeted species) would therefore require a Bayesian framework to include the effect of flock size on species detectability (*Clement, Converse & Royle, 2017*).

Given the timeframe and the sampling design, *i.e.*, all sites are not visited every year to maximise the number of total surveyed locations, it is not possible to estimate species demographic parameters, such as survival and recruitment (*Sollmann et al., 2015*; *Schmidt & Rattenbury, 2018*). We also assumed a sex ratio of 1:1 for species without sexual dimorphism, during the breeding season, which could potentially bias estimates for species deviating from this assumption. Taking account of species population structure requires frameworks such as Integrated Population Models (IPM; *Schaub & Ullrich, 2021*) and specific data collection (*King, 2014*), for instance, bagging or nest surveillance.

As obtaining relevant predictions of species abundance over unsampled environmental conditions was one of our main methodological challenges, we used environmental data condensing habitat information (*Tredennick et al., 2021*). To fit our statistical framework, we assumed that most bird species would interact with their habitat following a linear relationship (see [Fig. 3](#)). We therefore used PCA reduction to summarise species linear responses to national-scale habitat gradients including forest-to-open-field cultures,

open-field cultures-to-pastures and perennial crop-to-urban habitats (see [Materials S3](#)). PCA reduction permits model convergence by condensing complex habitat structures to a small number of environmental covariates, though it could bias estimates of species thriving in a specific habitat restricted to the extreme edge of the sampled gradients. Other methods such as Spatially Varying Covariates models (SVC; [Gelfand et al., 2003](#)) could be used to better account for habitat structure complexity across spatial gradients ([Thorson et al., 2023](#)).

Previous studies have shown that unaccounted variations in species availability, considering a constant detection probability or unmodelled variations, could lead to substantial bias in estimated abundance ([Link et al., 2018](#); [Barker et al., 2018](#); [Duarte, Adams & Peterson, 2018](#)). N-mixture biased estimations can be linked to non-assessment of the sampled area, where a smaller or greater sampled effective area could lead to under- or overestimation ([Kéry & Royle, 2015](#)). In our study, as we relied on distance sampling methods, we define an effective sampled area, based upon collected observation distance, but we also assumed that individuals considered exposed to the sampling (*i.e.*, ‘statistically’ available for modelling) could still be undetected due to small species home ranges or plot-specific habitat cover ([Chandler, Royle & King, 2011](#); see [Table 1](#) and [Fig. 2](#), for covariates used to model species detectability and [Materials S5](#) for model formulation). Despite such consideration, for the HDS model, we assumed that detected individuals were homogeneously distributed over the sampled area. Violating this assumption could lead to within-sample variation that needs to be accounted for, otherwise leading to biased estimates ([Mizel, Schmidt & Lindberg, 2018](#)).

Another potential drawback relies on the quantity of data collected through this structured CS scheme. Over the same breeding season, the semistructured scheme EPOC without temporal replicates nor fixed location requirements collected three times the amount of complete checklists as the structured EPOC-ODF scheme, highlighting CS trade-off of scheme standardisation upon data collection over spatial and temporal scales ([Devictor, Whittaker & Beltrame, 2010](#)). One way to address this trade-off would be to apply data integration methods mobilising multiple data sources to be used for ecological inferences ([Zipkin, Inouye & Beissinger, 2019](#); [Zipkin et al., 2021](#)), either by estimating abundance of less recorded species through trait-based associations ([Callaghan, Nakagawa & Cornwell, 2021, 2022](#); [Robinson et al., 2022](#)) or by constructing joint likelihood functions ([Fithian et al., 2015](#); [Fletcher et al., 2019](#)).

## CONCLUSIONS

Our results suggested an overall lower population size estimate of French common breeding birds obtained from the last French Breeding Bird atlas methodology than from the Hierarchical Distance Sampling modelling used in the present work. Using large-scale datasets from citizen science obtained from standard scheme initiatives allowed us to infer the variation in species abundance, while explicitly modelling the detection process separately from the ecological one. Not accounting for the observation process might have resulted in misleading expert-only estimations of population sizes in the previous atlas, at least for some widespread species not benefitting from exhaustive surveys. In conclusion,

our results advocate for more reliance on the use of statistical tools accounting for the detection process, such as hierarchical models, which, in association with large-scale citizen science data, could constitute a standard methodology to estimate reliable abundance from breeding bird atlases or biodiversity surveys deployed at national or geographically broader scales

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### Competing Interests

The authors declare that they have no competing interests.

### Author Contributions

- Jean Nabias conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.



- Luc Barbaro conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.
- Benoît Fontaine conceived and designed the experiments, authored or reviewed drafts of the article, and approved the final draft.
- Jérémy Dupuy conceived and designed the experiments, authored or reviewed drafts of the article, and approved the final draft.
- Laurent Couzi conceived and designed the experiments, authored or reviewed drafts of the article, and approved the final draft.
- Clément Vallé analyzed the data, authored or reviewed drafts of the article, and approved the final draft.
- Romain Lorrilliere conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.

### Data Availability

The following information was supplied regarding data availability:

The data, scripts and additional figures and tables are available at Zenodo:

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### Supplemental Information

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## REFERENCES

- Akaike H.** 1974. A new look at the statistical model identification. *IEEE Transactions on Automatic Control* **19**(6):716–723 DOI [10.1109/TAC.1974.1100705](https://doi.org/10.1109/TAC.1974.1100705).
- Barker RJ, Schofield MR, Link WA, Sauer JR.** 2018. On the reliability of n-mixture models for count data. *Biometrics* **74**(1):369–377 DOI [10.1111/biom.12734](https://doi.org/10.1111/biom.12734).
- BirdLife International.** 2021. *European red list of birds*. Brussels: Office of the European Union.
- Bouchet PJ, Miller DL, Roberts JJ, Mannocci L, Harris CM, Thomas L.** 2020. dsmextra: extrapolation assessment tools for density surface models. *Methods in Ecology and Evolution* **11**(11):1464–1469 DOI [10.1111/2041-210X.13469](https://doi.org/10.1111/2041-210X.13469).
- Boulinier T, Nichols JD, Sauer JR, Hines JE, Pollock KH.** 1998. Estimating species richness: the importance of heterogeneity in species detectability. *Ecology* **79**(3):1018–1028 DOI [10.1890/0012-9658\(1998\)079\[1018:ESRTIO\]2.0.CO;2](https://doi.org/10.1890/0012-9658(1998)079[1018:ESRTIO]2.0.CO;2).
- Brlík V, Šilarová E, Škorpilová J, Alonso H, Anton M, Aunins A, Benkő Z, Biver G, Busch M, Chodkiewicz T, Chylarecki P, Coombes D, de Carli E, del Moral JC, Derouaux A, Escandell V, Eskildsen DP, Fontaine B, Foppen RPB, Gamero A, Gregory RD, Harris S, Herrando S, Hristov I, Husby M, Ieronymidou C, Jiquet F, Kålås JA, Kamp J, Kmecl P, Kurlavičius P, Lehikoinen A, Lewis L, Lindström Å, Manolopoulos A, Martí D, Massimino D, Moshøj C, Nellis R, Noble D, Paquet A, Paquet J-Y, Portolou D, Ramírez I,**

- Redel C, Reif J, Ridzoň J, Schmid H, Seaman B, Silva L, Soldaat L, Spasov S, Staneva A, Szép T, Florenzano GT, Teufelbauer N, Trautmann S, van der Meij T, van Strien A, van Turnhout C, Vermeersch G, Vermouzek Z, Vikstrøm T, Voříšek P, Weiserbs A, Klvaňová A. 2021. Long-term and large-scale multispecies dataset tracking population changes of common European breeding birds. *Scientific Data* 8(1):21 DOI 10.1038/s41597-021-00804-2.
- Brooks TM, Mittermeier RA, da Fonseca GAB, Gerlach J, Hoffmann M, Lamoreux JF, Mittermeier CG, Pilgrim JD, Rodrigues ASL. 2006. Global biodiversity conservation priorities. *Science* 313:58–61 DOI 10.1126/science.1127609.
- Buckland ST, Anderson DR, Burnham KP, Laake J, Borchers D, Thomas L. 2001. *Introduction to distance sampling: estimating abundance of biological populations*. Oxford, New York: Oxford University Press.
- Buckland ST, Johnston A. 2017. Monitoring the biodiversity of regions: key principles and possible pitfalls. *Biological Conservation* 214(6):23–34 DOI 10.1016/j.biocon.2017.07.034.
- Burleigh JG, Kimball RT, Braun EL. 2015. Building the avian tree of life using a large-scale, sparse supermatrix. *Molecular Phylogenetics and Evolution* 84:53–63 DOI 10.1016/j.ympev.2014.12.003.
- Callaghan CT, Nakagawa S, Cornwell WK. 2021. Global abundance estimates for 9,700 bird species. *Proceedings of the National Academy of Sciences of the United States of America* 118(21):e2023170118 DOI 10.1073/pnas.2023170118.
- Callaghan CT, Nakagawa S, Cornwell WK. 2022. Reply to Robinson et al.: data integration will form the basis of future abundance estimates. *Proceedings of the National Academy of Sciences of the United States of America* 119(10):e2117920119 DOI 10.1073/pnas.2117920119.
- Ceballos G, Ehrlich PR, Barnosky AD, García A, Pringle RM, Palmer TM. 2015. Accelerated modern human-induced species losses: entering the sixth mass extinction. *Science Advances* 1(5):e1400253 DOI 10.1126/sciadv.1400253.
- Chandler RB, Royle JA, King DI. 2011. Inference about density and temporary emigration in unmarked populations. *Ecology* 92(7):1429–1435 DOI 10.1890/10-2433.1.
- Chandler M, See L, Copas K, Bonde AMZ, López BC, Danielsen F, Legind JK, Masinde S, Miller-Rushing AJ, Newman G, Rosemartin A, Turak E. 2017. Contribution of citizen science towards international biodiversity monitoring. *Biological Conservation* 213(Part 1):280–294 DOI 10.1016/j.biocon.2016.09.004.
- Clavel J, Julliard R, Devictor V. 2011. Worldwide decline of specialist species: toward a global functional homogenization? *Frontiers in Ecology and the Environment* 9(4):222–228 DOI 10.1890/080216.
- Clement MJ, Converse SJ, Royle JA. 2017. Accounting for imperfect detection of groups and individuals when estimating abundance. *Ecology and Evolution* 7(18):7304–7310 DOI 10.1002/ece3.3284.
- Comolet-Tirman J, Quaintenne G, Sibley J-P, Wroza S, Bal G, Champagnon J, Couzi L, Czajkowski M-A, Denis P, Frochot B, Muller Y. 2022. Estimated bird populations breeders in mainland France. *Alauda* 90:133–150 (in French).
- Comolet-Tirman J, Sibley J-P, Witté I, Cadiou B, Czajkowski M-A, Deceuninck B, Jiguet F, Landry P, Quaintenne G, Roché J-E, Sarasa M, Touroult J. 2015. Statuts et tendances des populations d'oiseaux nicheurs de France. Bilan simplifié du premier rapportage national au titre de la Directive Oiseaux. *Alauda* 83(1):35–76.
- Connors BM, Cooper AB, Peterman RM, Dulvy NK. 2014. The false classification of extinction risk in noisy environments. *Proceedings of the Royal Society B: Biological Sciences* 281(1787):20132935 DOI 10.1098/rspb.2013.2935.

- Cote C, Troncon C, Troncon C, Troncon C. 2021. ROUTE 500® Version 3.0—Descriptif de contenu., 27. Available at <https://geoservices.ign.fr/route500>.
- Couvet D, Ducarme F. 2014. Reconciliation ecology, from biological to social challenges. *Revue d'ethnoécologie* 6:16 DOI 10.4000/ethnoecologie.1979.
- Cruz-Cárdenas G, López-Mata L, Villaseñor JL, Ortiz E. 2014. Potential species distribution modeling and the use of principal component analysis as predictor variables. *Revista Mexicana de Biodiversidad* 85(1):189–199 DOI 10.7550/rmb.36723.
- Devictor V, van Swaay C, Brereton T, Brotons L, Chamberlain D, Heliölä J, Herrando S, Julliard R, Kuussaari M, Lindström Å, Reif J, Roy DB, Schweiger O, Settele J, Stefanescu C, Van Strien A, Van Turnhout C, Vermouzek Z, WallisDeVries M, Wynhoff I, Jiguet F. 2012. Differences in the climatic debts of birds and butterflies at a continental scale. *Nature Climate Change* 2(2):121–124 DOI 10.1038/nclimate1347.
- Devictor V, Whittaker RJ, Beltrame C. 2010. Beyond scarcity: citizen science programmes as useful tools for conservation biogeography. *Diversity and Distributions* 16(3):354–362 DOI 10.1111/j.1472-4642.2009.00615.x.
- Donald PF, Fuller RJ. 1998. Ornithological atlas data: a review of uses and limitations. *Bird Study* 45(2):129–145 DOI 10.1080/00063659809461086.
- Duarte A, Adams MJ, Peterson JT. 2018. Fitting N-mixture models to count data with unmodeled heterogeneity: bias, diagnostics, and alternative approaches. *Ecological Modelling* 374:51–59 DOI 10.1016/j.ecolmodel.2018.02.007.
- Elith J, Leathwick JR. 2009. Species distribution models: ecological explanation and prediction across space and time. *Annual Review of Ecology, Evolution, and Systematics* 40(1):677–697 DOI 10.1146/annurev.ecolsys.110308.120159.
- Farrier D, Whelan R, Mooney C. 2007. Threatened species listing as a trigger for conservation action. *Environmental Science & Policy* 10(3):219–229 DOI 10.1016/j.envsci.2006.12.001.
- Fick SE, Hijmans RJ. 2017. WorldClim 2: new 1-km spatial resolution climate surfaces for global land areas. *International Journal of Climatology* 37(12):4302–4315 DOI 10.1002/joc.5086.
- Fiske I, Chandler R. 2011. unmarked: an R package for fitting hierarchical models of wildlife occurrence and abundance. *Journal of Statistical Software* 43(10):1–23 DOI 10.18637/jss.v043.i10.
- Fithian W, Elith J, Hastie T, Keith DA. 2015. Bias correction in species distribution models: pooling survey and collection data for multiple species. *Methods in Ecology and Evolution* 6(4):424–438 DOI 10.1111/2041-210X.12242.
- Fletcher RJ, Hefley TJ, Robertson EP, Zuckerberg B, McCleery RA, Dorazio RM. 2019. A practical guide for combining data to model species distributions. *Ecology* 100(6):e02710 DOI 10.1002/ecy.2710.
- Fontaine A, Simard A, Brunet N, Elliott KH. 2022. Scientific contributions of citizen science applied to rare or threatened animals. *Conservation Biology* 36(6):e13976 DOI 10.1111/cobi.13976.
- Freckleton RP. 2020. Conservation decisions in the face of uncertainty. In: Vickery JA, Ockendon N, Pettorelli N, Brotherton PNM, Sutherland WJ, Davies ZG, eds. *Conservation Research, Policy and Practice Ecological Reviews*. Cambridge: Cambridge University Press, 183–195 DOI 10.1017/9781108638210.011.
- Fuller RJ, Langslow DR. 1984. Estimating numbers of birds by point counts: how long should counts last? *Bird Study* 31(3):195–202 DOI 10.1080/00063658409476841.
- Gaston KJ, Fuller RA. 2008. Commonness, population depletion and conservation biology. *Trends in Ecology & Evolution* 23(1):14–19 DOI 10.1016/j.tree.2007.11.001.

- Gaüzère P, Barbaro L, Calatayud F, Princé K, Devictor V, Raison L, Sirami C, Balent G. 2020. Long-term effects of combined land-use and climate changes on local bird communities in mosaic agricultural landscapes. *Agriculture, Ecosystems & Environment* **289**(4):106722 DOI [10.1016/j.agee.2019.106722](https://doi.org/10.1016/j.agee.2019.106722).
- Gaüzère P, Devictor V. 2021. Mismatches between birds' spatial and temporal dynamics reflect their delayed response to global changes. *Oikos* **130**(8):1284–1296 DOI [10.1111/oik.08289](https://doi.org/10.1111/oik.08289).
- Gaüzère P, Doucier G, Devictor V, Kéfi S. 2019. A framework for estimating species-specific contributions to community indicators. *Ecological Indicators* **99**:74–82 DOI [10.1016/j.ecolind.2018.11.069](https://doi.org/10.1016/j.ecolind.2018.11.069).
- Gedeon K, Grüneberg C, Mitschke A, Sudfeldt C, Eickhorst W, Fischer S, Flade M, Frick S, Geiersberger I, Koop B, Kramer M, Krüger T, Roth N, Ryslavy T, Stübing S. 2015. *Atlas deutscher Brutvogelarten: atlas of german breeding birds*. Münster, Westf: Dachverband Deutscher Avifaunisten.
- Gelfand AE, Kim HJ, Sirmans CF, Banerjee S. 2003. Spatial modeling with spatially varying coefficient processes. *Journal of the American Statistical Association* **98**(462):387–396 DOI [10.1198/016214503000170](https://doi.org/10.1198/016214503000170).
- Germain RR, Feng S, Chen G, Graves GR, Tobias JA, Rahbek C, Lei F, Fjeldså J, Hosner PA, Gilbert MTP, Zhang G, Nogués-Bravo D. 2023. Species-specific traits mediate avian demographic responses under past climate change. *Nature Ecology & Evolution* **7**(6):862–872 DOI [10.1038/s41559-023-02055-3](https://doi.org/10.1038/s41559-023-02055-3).
- Guillera-Arroita G, Lahoz-Monfort JJ, MacKenzie DI, Wintle BA, McCarthy MA. 2014. Ignoring imperfect detection in biological surveys is dangerous: a response to 'fitting and interpreting occupancy models'. *PLOS ONE* **9**(7):e99571 DOI [10.1371/journal.pone.0099571](https://doi.org/10.1371/journal.pone.0099571).
- Hong P, Li Z, Yang Q, Deng W, Xu Y, Tobias JA, Wang S. 2023. Functional traits and environment jointly determine the spatial scaling of population stability in North American birds. *Ecology* **104**(4):e3973 DOI [10.1002/ecy.3973](https://doi.org/10.1002/ecy.3973).
- Hurlbert AH, Liang Z. 2012. Spatiotemporal variation in avian migration phenology: citizen science reveals effects of climate change. *PLOS ONE* **7**(2):e31662 DOI [10.1371/journal.pone.0031662](https://doi.org/10.1371/journal.pone.0031662).
- Inger R, Gregory R, Duffy JP, Stott I, Voříšek P, Gaston KJ. 2015. Common European birds are declining rapidly while less abundant species' numbers are rising. *Ecology Letters* **18**(1):28–36 DOI [10.1111/ele.12387](https://doi.org/10.1111/ele.12387).
- Ingram DJ, Ferreira GB, Jones KE, Mace GM. 2021. Targeting conservation actions at species threat response thresholds. *Trends in Ecology & Evolution* **36**(3):216–226 DOI [10.1016/j.tree.2020.11.004](https://doi.org/10.1016/j.tree.2020.11.004).
- Isaac NJB, Jarzyna MA, Keil P, Dambly LI, Boersch-Supan PH, Browning E, Freeman SN, Golding N, Guillera-Arroita G, Henrys PA, Jarvis S, Lahoz-Monfort J, Pagel J, Pescott OL, Schmucki R, Simmonds EG, O'Hara RB. 2020. Data integration for large-scale models of species distributions. *Trends in Ecology & Evolution* **35**(1):56–67 DOI [10.1016/j.tree.2019.08.006](https://doi.org/10.1016/j.tree.2019.08.006).
- Issa N, Muller Y. 2015. *Atlas of birds of metropolitan France: nesting and presence winter*. Paris: Delachaux (in French).
- IUCN. 2001. IUCN Red List categories and criteria, version 3.1. IUCN. Available at <https://portals.iucn.org/library/node/7977>.
- James G, Witten D, Hastie T, Tibshirani R. 2013. *An introduction to statistical learning: with applications in R*. Cham: Springer.

- Jarzyna MA, Jetz W. 2016. Detecting the multiple facets of biodiversity. *Trends in Ecology & Evolution* 31(7):527–538 DOI 10.1016/j.tree.2016.04.002.
- Jetz W, McGeoch MA, Guralnick RP, Ferrier S, Beck J, Costello MJ, Fernandez M, Geller GN, Keil P, Merow C, Meyer C, Muller-Karger FE, Pereira HM, Regan EC, Regan E, Schmeller DS, Turak E. 2019. Essential biodiversity variables for mapping and monitoring species populations. *Nature Ecology and Evolution* 3(4):539–551 DOI 10.1038/s41559-019-0826-1.
- Jiguet F, Devictor V, Julliard R, Couvet D. 2012. French citizens monitoring ordinary birds provide tools for conservation and ecological sciences. *Acta Oecologica* 44(3):58–66 DOI 10.1016/j.actao.2011.05.003.
- Johnson CN, Balmford A, Brook BW, Buettel JC, Galetti M, Guangchun L, Wilmshurst JM. 2017. Biodiversity losses and conservation responses in the Anthropocene. *Science* 356(6335):270–275 DOI 10.1126/science.aam9317.
- Johnson DS, Laake JL, Ver Hoef JM. 2010. A model-based approach for making ecological inference from distance sampling data. *Biometrics* 66(1):310–318 DOI 10.1111/j.1541-0420.2009.01265.x.
- Johnston A, Hochachka WM, Strimas-Mackey ME, Gutierrez VR, Robinson OJ, Miller ET, Auer T, Kelling ST, Fink D. 2019. Best practices for making reliable inferences from citizen science data: case study using eBird to estimate species distributions. *bioRxiv* DOI 10.1101/574392.
- Johnston A, Matechou E, Dennis EB. 2022. Outstanding challenges and future directions for biodiversity monitoring using citizen science data. *Methods in Ecology and Evolution* 14(1):103–116 DOI 10.1111/2041-210X.13834.
- Johnston A, Newson SE, Risely K, Musgrove AJ, Massimino D, Baillie SR, Pearce-Higgins JW. 2014. Species traits explain variation in detectability of UK birds. *Bird Study* 61(3):340–350 DOI 10.1080/00063657.2014.941787.
- Julliard R, Clavel J, Devictor V, Jiguet F, Couvet D. 2006. Spatial segregation of specialists and generalists in bird communities. *Ecology Letters* 9(11):1237–1244 DOI 10.1111/j.1461-0248.2006.00977.x.
- Kamp J, Oppel S, Heldbjerg H, Nyegaard T, Donald PF. 2016. Unstructured citizen science data fail to detect long-term population declines of common birds in Denmark. *Diversity and Distributions* 22(10):1024–1035 DOI 10.1111/ddi.12463.
- Keller V, Herrando S, Voříšek P, Franch M, Kipson M, Milanese P, Martí D, Anton M, Klvaňová A, Kalyakin MV, Bauer H-G, Foppen RPB. 2020. *European breeding bird Atlas 2: distribution, abundance and change*. Barcelona: European Bird Census Council & Lynx Edicions.
- Kellner KF, Smith AD, Royle JA, Kéry M, Belant JL, Chandler RB. 2023. The unmarked R package: twelve years of advances in occurrence and abundance modelling in ecology. *Methods in Ecology and Evolution* 14(6):1408–1415 DOI 10.1111/2041-210X.14123.
- Kellner KF, Swihart RK. 2014. Accounting for imperfect detection in ecology: a quantitative review. *PLOS ONE* 9(10):e111436 DOI 10.1371/journal.pone.0111436.
- Kéry M. 2011. Towards the modelling of true species distributions. *Journal of Biogeography* 38(4):617–618 DOI 10.1111/j.1365-2699.2011.02487.x.
- Kéry M, Gardner B, Monnerat C. 2010. Predicting species distributions from checklist data using site-occupancy models. *Journal of Biogeography* 37(10):1851–1862 DOI 10.1111/j.1365-2699.2010.02345.x.



- Kéry M, Royle JA. 2015. *Applied hierarchical modeling in ecology: analysis of distribution, abundance and species richness in R and BUGS: Volume 1: prelude and static models*. Amsterdam, Boston: Academic Press.
- Kéry M, Royle JA. 2021. Chapter 12—conclusions. In: Kéry M, Royle JA, eds. *Applied Hierarchical Modeling in Ecology: Analysis of Distribution, Abundance and Species Richness in R and BUGS*. Cambridge: Academic Press, 717–723 DOI 10.1016/B978-0-12-809585-0.00012-0.
- Kéry M, Royle JA, Schmid H. 2005. Modeling avian abundance from replicated counts using binomial mixture models. *Ecological Applications* 15(4):1450–1461 DOI 10.1890/04-1120.
- Kéry M, Schmid H. 2004. Monitoring programs need to take into account imperfect species detectability. *Basic and Applied Ecology* 5(1):65–73 DOI 10.1078/1439-1791-00194.
- King R. 2014. Statistical ecology. *Annual Review of Statistics and Its Application* 1(1):401–426 DOI 10.1146/annurev-statistics-022513-115633.
- Kullenberg C, Kasperowski D. 2016. What is citizen science?—a scientometric meta-analysis. *PLOS ONE* 11(1):e0147152 DOI 10.1371/journal.pone.0147152.
- Lehikoinen A. 2013. Climate change, phenology and species detectability in a monitoring scheme. *Population Ecology* 55(2):315–323 DOI 10.1007/s10144-012-0359-9.
- Li M, Bolker B. 2019. First release of phylogenetic comparative analysis in lme4-verse. DOI 10.5281/zenodo.2639887.
- Link WA, Schofield MR, Barker RJ, Sauer JR. 2018. On the robustness of N-mixture models. *Ecology* 99(7):1547–1551 DOI 10.1002/ecy.2362.
- Matutini F, Baudry J, Pain G, Sineau M, Pithon J. 2021. How citizen science could improve species distribution models and their independent assessment. *Ecology and Evolution* 11(7):3028–3039 DOI 10.1002/ece3.7210.
- Mayaud N, de Balsac HH, Jouard H. 1936. *Inventory of birds of France*. Paris: Study Society Ornithological (in French).
- McKinley DC, Miller-Rushing AJ, Ballard HL, Bonney R, Brown H, Cook-Patton SC, Evans DM, French RA, Parrish JK, Phillips TB, Ryan SF, Shanley LA, Shirk JL, Stepenuck KF, Weltzin JF, Wiggins A, Boyle OD, Briggs RD, Chapin SF, Hewitt DA, Preuss PW, Soukup MA. 2017. Citizen science can improve conservation science, natural resource management, and environmental protection. *Biological Conservation* 208:15–28 DOI 10.1016/j.biocon.2016.05.015.
- McNew LB, Handel CM. 2015. Evaluating species richness: biased ecological inference results from spatial heterogeneity in detection probabilities. *Ecological Applications* 25(6):1669–1680 DOI 10.1890/14-1248.1.
- Meine C, Soulé M, Noss RF. 2006. “A Mission-Driven Discipline”: the growth of conservation biology. *Conservation Biology* 20:631–651 DOI 10.1111/j.1523-1739.2006.00449.x.
- Mesgaran MB, Cousens RD, Webber BL. 2014. Here be dragons: a tool for quantifying novelty due to covariate range and correlation change when projecting species distribution models. *Diversity and Distributions* 20(10):1147–1159 DOI 10.1111/ddi.12209.
- Michel NL, Smith AC, Clark RG, Morrissey CA, Hobson KA. 2016. Differences in spatial synchrony and interspecific concordance inform guild-level population trends for aerial insectivorous birds. *Ecography* 39(8):774–786 DOI 10.1111/ecog.01798.
- Mizel JD, Schmidt JH, Lindberg MS. 2018. Accommodating temporary emigration in spatial distance sampling models. *Journal of Applied Ecology* 55(3):1456–1464 DOI 10.1111/1365-2664.13053.



- Morin DJ, Yackulic CB, Diffendorfer JE, Lesmeister DB, Nielsen CK, Reid J, Schaub EM. 2020. Is your ad hoc model selection strategy affecting your multimodel inference? *Ecosphere* 11(1):e02997 DOI 10.1002/ecs2.2997.
- Navarro LM, Fernández N, Guerra CA, Guralnick R, Guralnick RP, Kissling WD, Londoño MC, Muller-Karger FE, Turak E, Balvanera P, Costello MJ, Delavaud A, Serafy GYE, Ferrier S, Geijzendorffer IR, Geller GN, Jetz W, Kim ES, Kim H, Martin CS, McGeoch MA, Mwampamba TH, Nel JL, Nicholson E, Pettorelli N, Schaeppman ME, Skidmore AK, Sousa-Pinto I, Pinto IS, Vergara SG, Vihervaara P, Xu H, Yahara T, Gill MJ, Pereira HM. 2017. Monitoring biodiversity change through effective global coordination. *Current Opinion in Environmental Sustainability* 29:158–169 DOI 10.1016/j.cosust.2018.02.005.
- Neeson TM, Doran PJ, Ferris MC, Fitzpatrick KB, Herbert M, Khoury M, Moody AT, Ross J, Yacobson E, McIntyre PB. 2018. Conserving rare species can have high opportunity costs for common species. *Global Change Biology* 24(8):3862–3872 DOI 10.1111/gcb.14162.
- Newman G, Wiggins A, Crall A, Graham E, Newman S, Crowston K. 2012. The future of citizen science: emerging technologies and shifting paradigms. *Frontiers in Ecology and the Environment* 10(6):298–304 DOI 10.1890/110294.
- Nichols JD, Koneff MD, Heglund PJ, Knutson MG, Seamans ME, Lyons JE, Morton JM, Jones MT, Boomer GS, Williams BK. 2011. Climate change, uncertainty, and natural resource management. *The Journal of Wildlife Management* 75(1):6–18 DOI 10.1002/jwmg.33.
- Nichols JD, Thomas L, Conn PB. 2009. Inferences about landbird abundance from count data: recent advances and future directions. In: Thomson DL, Cooch EG, Conroy MJ, eds. *Modeling Demographic Processes In Marked Populations Environmental and Ecological Statistics*. Boston, MA: Springer Us, 201–235 DOI 10.1007/978-0-387-78151-8\_9.
- Norvell RE, Howe FP, Parrish JR. 2003. A seven-year comparison of relative-abundance and distance-sampling methods. *The Auk* 120(4):1013–1028 DOI 10.1093/auk/120.4.1013.
- Paradis E, Baillie SR, Sutherland WJ, Gregory RD. 1998. Patterns of natal and breeding dispersal in birds. *Journal of Animal Ecology* 67(4):518–536 DOI 10.1046/j.1365-2656.1998.00215.x.
- Parmesan C. 2007. Influences of species, latitudes and methodologies on estimates of phenological response to global warming. *Global Change Biology* 13(9):1860–1872 DOI 10.1111/j.1365-2486.2007.01404.x.
- Payne EH, Gebregziabher M, Hardin JW, Ramakrishnan V, Egede LE. 2018. An empirical approach to determine a threshold for assessing overdispersion in Poisson and negative binomial models for count data. *Communications in Statistics: Simulation and Computation* 47(6):1722–1738 DOI 10.1080/03610918.2017.1323223.
- Pereira HM, Ferrier S, Walters M, Geller GN, Jongman RHG, Scholes RJ, Bruford MW, Brummitt N, Butchart SHM, Cardoso AC, Cardoso AC, Coops NC, Dulloo E, Faith DP, Freyhof J, Gregory RD, Heip CHR, Heip CHR, Heip CHR, Heip CHR, Hoft R, Hoft R, Höft R, Hurtt GC, Jetz W, Karp DS, McGeoch MA, Obura D, Onoda Y, Pettorelli N, Reyers B, Sayre R, Scharlemann JPW, Stuart SN, Turak E, Walpole M, Wegmann M. 2013. Essential biodiversity variables. *Science* 339(6117):277–278 DOI 10.1126/science.1229931.
- Pillar VD, Duarte LdS. 2010. A framework for metacommunity analysis of phylogenetic structure. *Ecology Letters* 13(5):587–596 DOI 10.1111/j.1461-0248.2010.01456.x.
- Pilotto F, Kühn I, Adrian R, Alber R, Alignier A, Andrews C, Bäck J, Barbaro L, Beaumont D, Beenaerts N, Benham S, Boukal DS, Bretagnolle V, Camatti E, Canullo R, Cardoso PG, Ens BJ, Everaert G, Evtimova V, Feuchtmayr H, García-González R, Gómez García D, Grandin U, Gutowski JM, Hadar L, Halada L, Halassy M, Hummel H, Huttunen K-L, Jaroszewicz B, Jensen TC, Kalivoda H, Schmidt IK, Kröncke I, Leinonen R, Martinho F,

- Meeseburg H, Meyer J, Minerbi S, Monteith D, Nikolov BP, Oro D, Ozoliņš D, Padedda BM, Pallett D, Pansera M, Pardal MÁ, Petriccione B, Pipan T, Pöyry J, Schäfer SM, Schaub M, Schneider SC, Skuja A, Soetaert K, Sprinĝe G, Stanchev R, Stockan JA, Stoll S, Sundqvist L, Thimonier A, Van Hoey G, Van Ryckegem G, Visser ME, Vorhauser S, Haase P. 2020. Meta-analysis of multidecadal biodiversity trends in Europe. *Nature Communications* 11(1):3486 DOI 10.1038/s41467-020-17171-y.
- Quaintenne G, Gaudard C, Béchet A, Benmergui M, Boutteaux J-J, Cadiou B, Camberlein P, Chapalain F, Croset F, Culioli J-M, Dalloyau S, Debout G, Dubois P, Dulac P, Flitti A, Gallien F, Gendre N, Girard O, Havet S, Vincent-Martin N. 2020. Rare and endangered breeding bird survey in France in 2016–2017. *Ornithos* 27-2:73–111.
- Quinn JE, Brandle JR, Johnson RJ, Tyre AJ. 2011. Application of detectability in the use of indicator species: a case study with birds. *Ecological Indicators* 11(5):1413–1418 DOI 10.1016/j.ecolind.2011.03.003.
- Rabinowitz D. 1981. 17. Seven forms of rarity. In: Synge H, Linnean Society of London, King's College, eds. *The Biological Aspects of Rare Plant Conservation: Proceedings of an International Conference Held at King's College, Cambridge, England, 14–19 July 1980. A Wiley-Interscience Publication*. Chichester: Wiley, 205–217.
- Richter A, Nakamura G, Agra Iserhard C, da Silva Duarte L. 2021. The hidden side of diversity: effects of imperfect detection on multiple dimensions of biodiversity. *Ecology and Evolution* 11(18):12508–12519 DOI 10.1002/ece3.7995.
- Ricotta C. 2005. On parametric diversity indices in ecology: a historical note. *Community Ecology* 6(2):241–242 DOI 10.1556/ComEc.6.2005.2.12.
- Rigal S, Dakos V, Alonso H, Auniņš A, Benkó Z, Brotons L, Chodkiewicz T, Chylarecki P, de Carli E, del Moral JC, Domşa C, Escandell V, Fontaine B, Foppen R, Gregory R, Harris S, Herrando S, Husby M, Ieronymidou C, Jiguet F, Kennedy J, Klvaňová A, Kmecl P, Kuczyński L, Kurlavičius P, Kálás JA, Lehikoinen A, Lindström Å, Lorrillière R, Moshøj C, Nellis R, Noble D, Eskildsen DP, Paquet J-Y, Pélassié M, Pladevall C, Portolou D, Reif J, Schmid H, Seaman B, Szabo ZD, Szép T, Florenzano GT, Teufelbauer N, Trautmann S, van Turnhout C, Vermouzek Z, Vikstrøm T, Voříšek P, Weiserbs A, Devictor V. 2023. Farmland practices are driving bird population decline across Europe. *Proceedings of the National Academy of Sciences of the United States of America* 120(21):e2216573120 DOI 10.1073/pnas.2216573120.
- Rigal S, Knappe J. 2024. Investigating the heterogeneity within Wild bird indices in Europe. *Biological Conservation* 290:110452 DOI 10.1016/j.biocon.2024.110452.
- Robinson OJ, Socolar JB, Stuber EF, Auer T, Berryman AJ, Boersch-Supan PH, Brightsmith DJ, Burbidge AH, Butchart SHM, Davis CL, Dokter AM, Di Giacomo AS, Farnsworth A, Fink D, Hochachka WM, Howell PE, La Sorte FA, Lees AC, Marsden S, Martin R, Martin RO, Masello JF, Miller ET, Moodley Y, Musgrove A, Noble DG, Ojeda V, Quillfeldt P, Royle JA, Ruiz-Gutierrez V, Tella JL, Yorio P, Youngflesh C, Johnston A. 2022. Extreme uncertainty and unquantifiable bias do not inform population sizes. *Proceedings of the National Academy of Sciences* 119(10):e2113862119 DOI 10.1073/pnas.2113862119.
- Roché J-E, Muller Y, Siblet J-P. 2013. A simple method for estimating bird populations common breeding species in France. *Alauda* 81:241–268 (in French).
- Santini L, Tobias JA, Callaghan C, Gallego-Zamorano J, Benítez-López A. 2023. Global patterns and predictors of avian population density. *Global Ecology and Biogeography* 32(7):1189–1204 DOI 10.1111/geb.13688.

- Sanz-Pérez A, Sollmann R, Sardà-Palomera F, Bota G, Giralt D. 2020. The role of detectability on bird population trend estimates in an open farmland landscape. *Biodiversity and Conservation* 29(6):1747–1765 DOI 10.1007/s10531-020-01948-0.
- Schaub M, Ullrich B. 2021. A drop in immigration results in the extinction of a local woodchat shrike population. *Animal Conservation* 24(3):335–345 DOI 10.1111/acv.12639.
- Schmidt JH, McIntyre CL, MacCluskie MC. 2013. Accounting for incomplete detection: What are we estimating and how might it affect long-term passerine monitoring programs? *Biological Conservation* 160(2):130–139 DOI 10.1016/j.biocon.2013.01.007.
- Schmidt JH, Rattenbury KL. 2018. An open-population distance sampling framework for assessing population dynamics in group-dwelling species. *Methods in Ecology and Evolution* 9(4):936–945 DOI 10.1111/2041-210X.12932.
- Skirvin AA. 1981. Effect of time of day and time of season on the number of observations and density estimates of breeding birds. In: Ralph CJ, Scott JM, eds. *Estimating Numbers of Terrestrial Birds Studies in Avian Biology*. Chicago, Illinois: Cooper Ornithological Society, 271–274.
- Sólymos P, Matsuoka SM, Stralberg D, Barker NKS, Bayne EM. 2018. Phylogeny and species traits predict bird detectability. *Ecography* 41(10):1595–1603 DOI 10.1111/ecog.03415.
- Sollmann R, Gardner B, Chandler RB, Royle JA, Sillett TS. 2015. An open-population hierarchical distance sampling model. *Ecology* 96(2):325–331 DOI 10.1890/14-1625.1.
- Suzuki-Ohno Y, Yokoyama J, Nakashizuka T, Kawata M. 2017. Utilization of photographs taken by citizens for estimating bumblebee distributions. *Scientific Reports* 7(1):11215 DOI 10.1038/s41598-017-10581-x.
- Thierion V, Vincent A, Valero S. 2022. Theia OSO land cover map 2020. Zenodo DOI 10.5281/zenodo.6538861.
- Thorson JT, Barnes CL, Friedman ST, Morano JL, Siple MC. 2023. Spatially varying coefficients can improve parsimony and descriptive power for species distribution models. *Ecography* 2023(5):e06510 DOI 10.1111/ecog.06510.
- Tingley MW, Beissinger SR. 2013. Cryptic loss of montane avian richness and high community turnover over 100 years. *Ecology* 94(3):598–609 DOI 10.1890/12-0928.1.
- Tredennick AT, Hooker G, Ellner SP, Adler PB. 2021. A practical guide to selecting models for exploration, inference, and prediction in ecology. *Ecology* 102(6):e03336 DOI 10.1002/ecy.3336.
- Tukey J. 1977. *Exploratory data analysis*. Reading, Massachusetts, USA: Addison-Wesley.
- UICN France, MNHN, LPO, SEOF, ONCFS. 2016. *The Red List of threatened species in France—Chapter Birds of metropolitan France*. Paris: IUCN (in French).
- Villéger S, Mason NWH, Mouillot D. 2008. New multidimensional functional diversity indices for a multifaceted framework in functional ecology. *Ecology* 89(8):2290–2301 DOI 10.1890/07-1206.1.
- Visser ME, Holleman LJM, Gienapp P. 2006. Shifts in caterpillar biomass phenology due to climate change and its impact on the breeding biology of an insectivorous bird. *Oecologia* 147(1):164–172 DOI 10.1007/s00442-005-0299-6.
- Whelan CJ, Şekercioğlu Ç.H, Wenny DG. 2015. Why birds matter: from economic ornithology to ecosystem services. *Journal of Ornithology* 156(S1):227–238 DOI 10.1007/s10336-015-1229-y.
- Williams BK. 2003. Policy, research, and adaptive management in avian conservation. *The Auk* 120(1):212–217 DOI 10.1642/0004-8038(2003)120[0212:PRAAMI]2.0.CO;2.

- Yeatman L, Ridel Y, d'ornithologie S française, France D de la protection de la nature. 1976.** *Atlas of breeding birds of France from 1970 to 1975*. Paris: Ornithological Society of France (in French).
- Yeatman-Berthelot D, Jarry G. 1995.** *New Atlas of Breeding Birds of France 1985–1989*. Paris: Ornithological Society of France (in French).
- Zipkin EF, Inouye BD, Beissinger SR. 2019.** Innovations in data integration for modeling populations. *Ecology* **100(6)**:1–3 DOI [10.1002/ecy.2713](https://doi.org/10.1002/ecy.2713).
- Zipkin EF, Zylstra ER, Wright AD, Saunders SP, Finley AO, Dietze MC, Itter MS, Tingley MW. 2021.** Addressing data integration challenges to link ecological processes across scales. *Frontiers in Ecology and the Environment* **19(1)**:30–38 DOI [10.1002/fee.2290](https://doi.org/10.1002/fee.2290).