Response diversity of free-floating plants to nutrient stoichiometry and temperature: Growth and turion formation (#8375)

First submission

Please read the **Important notes** below, and the **Review guidance** on the next page. When ready **submit online**. The manuscript starts on page 3.

Important notes

Editor and deadline

Leon Higley / 28 Jan 2016

Files 5 Figure file(s)

2 Raw data file(s)

1 Other file(s)

Please visit the overview page to **download and review** the files

not included in this review pdf.

DeclarationsNo notable declarations are present



Please in full read before you begin

How to review

When ready <u>submit your review online</u>. The review form is divided into 5 sections. Please consider these when composing your review:

- 1. BASIC REPORTING
- 2. EXPERIMENTAL DESIGN
- 3. VALIDITY OF THE FINDINGS
- 4. General comments
- 5. Confidential notes to the editor
- You can also annotate this **pdf** and upload it as part of your review

To finish, enter your editorial recommendation (accept, revise or reject) and submit.

BASIC REPORTING

- Clear, unambiguous, professional English language used throughout.
- Intro & background to show context.
 Literature well referenced & relevant.
- Structure conforms to **PeerJ standard**, discipline norm, or improved for clarity.
- Figures are relevant, high quality, well labelled & described.
- Raw data supplied (See <u>PeerJ policy</u>).

EXPERIMENTAL DESIGN

- Original primary research within Scope of the journal.
- Research question well defined, relevant & meaningful. It is stated how research fills an identified knowledge gap.
- Rigorous investigation performed to a high technical & ethical standard.
- Methods described with sufficient detail & information to replicate.

VALIDITY OF THE FINDINGS

- Impact and novelty not assessed.

 Negative/inconclusive results accepted.

 Meaningful replication encouraged where rationale & benefit to literature is clearly stated.
- Data is robust, statistically sound, & controlled.
- Conclusion well stated, linked to original research question & limited to supporting results.
- Speculation is welcome, but should be identified as such.

The above is the editorial criteria summary. To view in full visit https://peerj.com/about/editorial-criteria/



Response diversity of free-floating plants to nutrient stoichiometry and temperature: Growth and turion formation

Michael J McCann

Free-floating plants, like most groups of aquatic primary producers, can become nuisance vegetation under certain conditions. On the other hand, there is substantial optimism for the applied uses of free-floating plants, including wastewater treatment, biofuel production, and aquaculture. Therefore, understanding the species-specific responses of floating plants to abiotic conditions will inform both management decisions and the beneficial applications of this plant group. I measured the responses of three floating plant species common in the northeast United States (Lemna minor, Spirodela polyrhiza, and Wolffia brasiliensis) to nutrient stoichiometry (nitrogen and phosphorus) and temperature in the laboratory. I also used survey data to determine the pattern of species richness of floating plants in the field and its relationship with the dominance of this group. Floating plant species exhibited unique responses to nutrient stoichiometry and temperature in the laboratory, especially under low temperatures (18°C) and low nutrient conditions (0.5 mg N L⁻¹, 0.083 mg P L⁻¹). Species displayed an apparent tradeoff with different strategies of growth or dormancy. In the field, water bodies with only Lemna minor were the most common; floating plant polycultures were not more dominant. The response diversity observed in the lab may not be associated with the dominance of this group in the field because it is masked by environmental variability, has a weak effect, or is only important during transient circumstances.



- 1 **Title:** Response diversity of free-floating plants to nutrient stoichiometry and temperature:
- 2 Growth and turion formation
- 3 **Author:** Michael J. McCann a, 1
- 4 a Affiliation: Department of Ecology and Evolution, Stony Brook University, 650 Life Sciences
- 5 Building, Stony Brook, New York 11794-5245 USA
- 6 ¹ Present address: Department of Marine and Coastal Sciences, Rutgers University, 71 Dudley
- 7 Road, New Brunswick, New Jersey 08901-8525 USA
- 8 **E-mail:** mccannmikejames@gmail.com



Abstract

10	Free-floating plants, like most groups of aquatic primary producers, can become nuisance
11	vegetation under certain conditions. On the other hand, there is substantial optimism for the
12	applied uses of free-floating plants, including wastewater treatment, biofuel production, and
13	aquaculture. Therefore, understanding the species-specific responses of floating plants to abiotic
14	conditions will inform both management decisions and the beneficial applications of these
15	plants. I measured the responses of three floating plant species common in the northeast United
16	States (Lemna minor, Spirodela polyrhiza, and Wolffia brasiliensis) to nutrient stoichiometry
17	(nitrogen and phosphorus) and temperature in the laboratory. I also used survey data to
18	determine the pattern of species richness of floating plants in the field and its relationship with
19	the dominance of this group. Floating plant species exhibited unique responses to nutrient
20	stoichiometry and temperature in the laboratory, especially under low temperatures (18°C) and
21	low nutrient conditions (0.5 mg N L ⁻¹ , 0.083 mg P L ⁻¹). Species displayed an apparent tradeoff
22	with different strategies of growth or dormancy. In the field, water bodies with only Lemna
23	minor were the most common; floating plant polycultures were not more dominant. The response
24	diversity observed in the lab may not be associated with the dominance of this group in the field
25	because it is masked by environmental variability, has a weak effect, or is only important during
26	transient circumstances.



Introduction

28	Free-floating plants, like most groups of aquatic primary producers, can become nuisance
29	vegetation under certain conditions (Portielje and Roijackers 1995, Janse and Van Puijenbroek
30	1998, Scheffer et al. 2003, Smith 2012). Shallow lakes and ponds, agricultural ditches, and
31	tropical lakes can be dominated by thick mats of floating plants, altering abiotic conditions and
32	reducing biotic diversity (Morris and Barker 1977, Janes et al. 1996, Morris et al. 2003,
33	Verdonschot and Verdonschot 2013). The dominance of this functional group is driven by
34	nutrient enrichment (both nitrogen and phosphorus), and low levels of either of these nutrients
35	can limit floating plant growth (Portielje and Roijackers 1995, Kufel et al. 2010, Smith 2014). In
36	addition to eutrophication, increased temperatures due to climate change may also favor the
37	dominance of this group over other primary producers (Netten et al. 2011, Peeters et al. 2013).
38	On the other hand, there is substantial optimism for the applied uses of free-floating plants, such
39	as wastewater treatment, biofuel production, ecotoxicological assessment, and aquaculture (e.g.,
40	Greenberg et al. 1992, Skillicorn et al. 1993, Ge et al. 2012, Xu et al. 2012, Verma and Suthar
41	2014). Therefore, understanding the species-specific responses of floating plants to nutrients and
42	temperature will have both management implications and beneficial applications. For example, it
43	floating plant species exhibit response diversity (i.e., unique response to abiotic conditions)
44	(Elmqvist et al. 2003), then a more diverse assemblage of floating plants may be more resilient
45	or dominant (Naeem and Wright 2003), and thus, harder to manage. Furthermore, if particular
46	species have unique responses, than those with a desirable suite of traits may be identified for
47	applied uses.

48





50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

Although both nitrogen and phosphorus are important drivers of floating-plant dominance, the ratio of both nutrients may have important consequences, especially in multi-species contexts (Smith 2014). Depending on the conditions of the growth medium, species and clones of floating plants differ in their N:P content (reviewed by Landolt and Kandler 1987). For example, Karpati and Pomogyi (1979) reported N:P tissue content ranging from 2.65 for Lemna trisulca to 10.53 for Lemna minor in naturally growing plants. Docauer (1983) reported N:P content of 8.12 for Spirodela polyrhiza, 10.38 for L. minor, and 3.46 and 6.54 for two species of Wolffia (W. borealis and W. columbiana, respectively) when the plants were growing at half of their maximum growth rate. Depending on the nutrient content of the growth medium, tissue N:P in Lemna gibba can range from approximately 3 to nearly 40 (Fulton et al. 2010). These speciesspecific and context-dependent stoichiometric differences are important because nutrient stoichiometry will differ depending on the source of nutrient loading and various other factors, resulting in wide variation in nutrient stoichiometry of different water bodies (Downing and McCauley 1992). If floating plant species are constrained in their nutrient stoichiometry, than this may affect the outcome of competition among floating plants or with other primary producer groups (Sterner and Elser 2002), although nitrogen alone explained most of the outcome of competition among floating plants in the early stages of a field mesocosm experiment (Smith 2014). In addition to vegetative growth on the surface of the water, many species are capable of producing turions - asexually-produced resting bodies that sink to the bottom of the water (Landolt and Kandeler 1987). These structures are typically starch-heavy and allow for

dormancy through winters or adverse conditions (Landolt and Kandeler 1987). Turion

PeerJ

72	production has been reported in S. polyrhiza, some populations of L. minor, and several species
73	of Wolffia, induced by a variety of factors including day length, nutrient and light conditions, and
74	hormones (reviewed by Hillman 1961 and Landolt and Kandeler 1987). Turion production may
75	be important for the persistence of floating plants in both natural and engineered systems.
76	
77	I used laboratory experiments and surveys in lakes and ponds to determine the response diversity
78	(i.e., unique response to abiotic conditions) among floating plant species. To address this
79	question, I performed two laboratory experiments to examine the growth and turion-formation of
80	floating plant species in response to nitrogen, phosphorus, and temperature. The experiments
81	were conducted with three of the most common floating plant species in the northeast United
82	States: Lemna minor L., Spirodela polyrhiza (L.) Schleid., and Wolffia brasiliensis Wedd. To
83	understand whether the response diversity in the laboratory corresponds to increased dominance
84	in the field, I also analyzed a dataset of over 200 freshwater lakes and ponds to determine the
85	association between floating plant species composition and richness and the occurrence of
86	floating plant dominance. If substantial differences exist among floating plant species, then a
87	more diverse assemblage may be more likely to be dominant over a broader range of conditions.
88	In this case, it is expected that water bodies with greater floating plant richness will be more
89	frequently found in a floating plant state.
90	
91	Materials & Methods
92	Laboratory conditions
93	For all experiments, plants were collected from Setauket Mill Pond, East Setauket, New York,
94	USA (40.946061°, -73.115613°) and acclimated under experimental conditions prior to the start



96

97

98

99

100

101

102

103

104

105

106

107

108

109

110

111

112

113

114

of the experiments. Plants were collected in mid-August or mid-June for experiments I and II, respectively. Species were identified according to Crow and Hellquist (2000). Modified Barko-Smart media (Smart and Barko 1985; Szabó et al. 2005) with phosphorus supplied as potassium dihydrogen phosphate and nitrogen supplied as a 1:1 ratio of nitrogen from nitrate and ammonium (potassium nitrate and ammonium chloride) was used as the nutrient media. Micronutrients were supplied to the media by Tropica Aquacare Plant Nutrition Liquid at a concentration of 0.1 mL L⁻¹ (Szabó et al. 2003, 2010). Plants were grown in plastic, multiwell plates with individual well diameters of 22.75 mm containing 4 mL of media. Each well housed a single replicate of an experimental treatment. Multiwell plates were cleaned in 10% hydrochloric acid for at least 1 hour, and then rinsed thoroughly with deionized water prior to using. Living plants (i.e., green) were moved to clean multiwell plates with fresh media every two to four days, depending on the experiment. Any dead (entirely white or brown) where removed. Light was supplied at an intensity of 130-150 µE m⁻² s⁻¹ and a 14:10 hr light:dark photoperiod, which is within the range of many previous studies (Landolt and Kandeler 1987). Temperature-controlled walk-in chambers were used to achieve the target temperatures. Nutrients treatments and species were systematically assigned to wells to ensure that replicates were dispersed across plates and not in adjacent wells. Initial plant area (20 mm² or 5% of the total well area available for growth) was approximately equal for all species within an experiment. Frond number differed because of the size differences among species. In order to prevent crowding, experiments were ended when plants in some treatments filled approximately two-thirds of the well area.

116

117

115

Experiment I: Response to nutrients and temperature



I measured the responses of floating plant species to nutrients and temperature by measuring their growth and turion formation at three nutrient levels (low: 0.5 mg N L⁻¹ and 0.083 mg P L⁻¹; medium: 5 mg N L⁻¹ and 0.83 mg P L⁻¹; or high: 10 mg N L⁻¹ and 1.66 mg P L⁻¹) fully-crossed with three temperatures (18, 24, and 30 °C), for a total of nine treatment combinations. The three nutrient treatments are all at a N:P mass ratio of ~6 and correspond to experimental treatments used in previous studies (e.g., Scheffer et al. 2003; Szabó et al. 2010). At the lowest nutrient level, nutrients were expected to be limiting to growth (Lüönd 1983, Szabó et al. 2010), but for the two highest nutrient levels, nutrients may be saturated (Szabó et al. 2010). Although this is not an exhaustive combination of treatments, these levels sample some of the possible environmental conditions encountered by floating plants in nature, and potentially in engineered applications (e.g., wastewater treatment). Eight replicates of each treatment combination for each species were grown for 12 days. Plants were transferred to new nutrient media 3, 5, 7, and 10 days after the start of the experiment.

Experiment II: Response to nutrient stoichiometry

In a second experiment, I measured the responses of floating plant species to nutrient stoichiometry by measuring their growth and turion formation at all nine combinations of three nitrogen (0.5, 5, and 10 mg N L⁻¹) and three phosphorus levels (0.083, 0.83, and 1.66 mg P L⁻¹), producing a variety of N:P mass ratios, ranging from 0.30 to 120.48 (Table 1). At the lowest treatment level of each nutrient, that nutrient may limit plant growth. Six replicates of each plant species at each of the nine nutrient treatments were grown for 17 days. I transferred plants to new nutrient media 3, 7, 10, and 14 days after the start of the experiment. Plants were grown at approximately 30°C, which had resulted in the maximum growth rates in Experiment I.

PeerJ

1	4	1

142

Growth and turion production

To quantify growth, plants in each replicate of each treatment were photographed (Nikon 143 144 Coolpix 5700 Digital Camera) with backlighting from a light box (Laboratory Supply Company, 145 60 Watts) on days that nutrient media were changed. The two-dimensional plant area on the 146 water surface was measured with ImageJ, version 1.47 (Rasband 2014), using the threshold function on an 8-bit grayscale photo, after dead fronds had been removed by hand (see above). 147 Relative growth rate (RGR) was calculated between each measurement. RGR was calculated as 148 $[\ln(A_2) - \ln(A_1)] / (t_2 - t_1)$, where A is the area of plants in mm², t is time in days, and subscripts 2 149 150 and I indicate two sequential time points in the experiment. Plant thickness or mass was not 151 measured during these experiments, but the overheard surface area is a commonly used measure 152 in many experiments (Landolt and Kandeler 1987). Turions were distinguished as plants that had sunk to the bottom of the experimental vessel, and they typically differed in size, texture, or 153 color from plants on the surface. The number of turions (i.e., asexual resting bodies) produced 154 were counted for each experimental replicate after the live plants had been moved to fresh media. 155 The number of turions was converted to area with equations developed in another study 156 157 (Appendix A). In some replicates, all plants in a replicate died (i.e., bleached white) during the experiment, and were re-started with new plants, assuming that the failed growth was due to 158 159 damage to the plant when handling. These replicates were excluded from analysis if they were 160 not grown for at least 10 days.

161

162

Statistical analysis



163	For all ANOVAs, data were tested for normally distributed residuals with a Shapiro-Wilk test
164	and equal variance among treatment groups with Levene's test. If the data did not meet these
165	assumptions of ANOVA, they were power transformed to ensure that these criteria were met. I
166	performed all statistical analyses in R version 3.0.2 (R Development Core Team 2013).
167	
168	The goal of these experiments was to test whether floating plant species differed in their
169	response to environmental conditions and under what conditions they differed. Therefore,
170	analyses tested for differences between species under particular combinations of environmental
171	conditions (i.e., treatment levels). For both experiments, I performed a one-way ANOVA for
172	each treatment combination to test for an effect of species on the average RGR. I used a Dunn-
173	Šidák correction to adjust p-values for multiple comparisons. When significant treatment effects
174	were found, Tukey's HSD was used to detect differences among species. An alternative
175	approach to analyzing these data is to use factorial ANOVAs to test for the main and interactive
176	effects of species and experimental conditions on growth rates (Appendix B). Since the number
177	of possible pairwise, posthoc comparisons in each experiment is large (351) and species
178	differences under identical conditions (i.e., response diversity) was the focus of this study, this
179	statistical approach is not reported in the main text.
180	
181	In the first experiment (the effect of nutrients and temperature) only W. brasiliensis formed
182	turions. I analyzed the effect of nutrients and temperature on turion area produced (mm² day-1)
183	by W. brasiliensis with a two-way ANOVA. When significant treatment effects were found, I
184	used Tukey's HSD to detect differences among treatment levels. In the second experiment (i.e.,
185	the effect of nitrogen and phosphorus), W. brasiliensis formed turions under all treatment levels





and *S. polyrhiza* under some treatment levels. To detect differences in turion production rate between species at particular nutrient levels, I used one-way ANOVAs at each nutrient level where both *W. brasiliensis* and *S. polyrhiza* formed turions. I used a Dunn-Šidák correction to adjust p-values for multiple comparisons.

Floating plant richness and abundance in natural water bodies

I examined the occurrence and dominance of floating plant species in lakes and ponds with a dataset of 205 freshwater water bodies in Connecticut and Long Island, NY (Appendix C). The data came from two sources: 1) 184 surveys by the Connecticut Agricultural Experiment Station (CAES) in 2005 to 2013, and 2) 21 surveys that I conducted in Long Island, New York and Connecticut, USA in 2011 to 2013. This data set spanned a range of perennial, freshwater lakes and ponds and included the list of floating plant species present in the water body and the maximum floating plant cover (percent of water body covered as quantified through visual observation and mapping) during the late summer (late July to September). See Capers et al. (2007) for a description of the survey methods used for the CAES data. For the Long Island surveys, plant cover was estimated through visual observation similar to methods used in previous studies (Driever et al. 2005, Smith 2012). In this study, I use high floating plant cover as a surrogate for dominance by floating plants, while acknowledging that a consideration of other primary producers (e.g., phytoplankton, submerged vegetation) and covariates is necessary for a rigorous demonstration of complete floating-plant dominance.

I used a goodness of fit test (G-test, based on a chi-square) to test if all floating plant species richness levels were equally likely to occur (i.e., random), excluding water bodies without





209	floating plants. I used a second G-test to determine if floating plant dominance (≥ 66.67% cover)
210	was equally likely to occur under different levels of floating plant richness. The expected value
211	for each richness level in floating plant dominated water bodies was based on the observed
212	frequency of each floating plant richness level across all water bodies (both dominated and non-
213	dominated). Floating plant richness was categorized as 1, 2, or \geq 3 species to ensure adequate
214	sample sizes in each level.
215	
216	Results
217	Experiment I: Response to nutrients and temperature
218	Average relative growth rate (RGR) was different among species at six of the nine combinations
219	of nutrients and temperature (Table 2, Figure 1). Species growth rates were equal when nutrients
220	and temperatures were high (10 mg N L^{1} and 1.66 mg P L^{1} and 24 or 30 °C) or at 18°C and
221	medium nutrients (5 mg N L ⁻¹ and 0.8 mg P L ⁻¹). Typically, <i>Lemna minor</i> and <i>Spirodela</i>
222	polyrhiza growth rates were equal to each other and both were greater than the growth rate of
223	Wolffia brasiliensis (Figure 1). Only W. brasiliensis formed turions in this experiment. There
224	was a significant effect of nutrients ($F_{2,64}$ = 4.770, p = 0.012), temperature ($F_{2,64}$ = 38.706, p <
225	0.001), and significant interaction ($F_{4,64} = 4.089$, $p = 0.005$) on the turion production rate of W .
226	brasiliensis (Figure 2). At both 18 and 30 °C W. brasiliensis decreased turion production at the
227	highest nutrient level, but at 24 °C turion production increased with nutrient level (Figure 2).
228	
229	Experiment II: Response to nutrient stoichiometry
230	Average RGR differed among species in four of the nine nutrient combinations (Table 3, Figure
231	3). Species differences were found whenever nitrogen was low (0.5 mg N L ⁻¹) or when





232	phosphorus was low and nitrogen was medium (0.08 mg P L ⁻¹ and 5 mg N L ⁻¹). Both <i>S</i> .
233	polyrhiza and W. brasiliensis formed turions in this experiment. W. brasiliensis formed turions at
234	all combinations of nitrogen and phosphorus, whereas S. polyrhiza only formed turions at low
235	nitrogen and medium and high phosphorus or low phosphorus and medium and high nitrogen
236	(Figure 4). At low nitrogen and medium phosphorus (ANOVA, $F_{1,9} = 51.62$, p <0.001), low
237	nitrogen and high phosphorus (ANOVA, $F_{1,10} = 49.91$, p < 0.001), and medium nitrogen and low
238	phosphorus (ANOVA, $F_{1,10} = 49.82$, p <0.001), W. brasiliensis had a greater turion production
239	rate than S. polyrhiza (Figure 4). At only low phosphorus and high nitrogen both species had
240	equal turion production rates (ANOVA, $F_{1,6}$ = 6.64, p = 0.042) (Dunn-Šidák adjusted critical p-
241	value 0.013)
242	
243	Floating plant richness and abundance in natural water bodies
244	Most freshwater lakes and ponds in Connecticut and Long Island, NY did not have any floating
245	plants present (106 of 205, Table 4). Across all water bodies with floating plants present, a total
246	of seven taxa were found. L. minor, S. polyrhiza, and Wolffia spp. were the most common taxa,
247	occurring in 82, 47, and 42 of the 205 water bodies, respectively. The next most common
248	species, L. trisulca, only occurred in 4 lakes and ponds. Among water bodies with floating
249	plants present, the occurrence of different levels of species richness levels was non-random
250	(Table 4, Figure 5a, G-test, $G = 6.909$, $df = 2$, $p < 0.031$). Monocultures were more common than
251	expected and three- and four-species polycultures were less common than expected (Table 4,
252	Figure 5a).



Only twenty water bodies had floating plant cover greater than 66.67%. Among these water bodies, there was no significant association between floating plant richness categories and the frequency of occurrence (Table 4, Figure 5b, G-test, G = 2.430, df = 2, p = 0.7). Although not statistically significant, water bodies dominated by floating plants tended to have three or more species of floating plants; whereas, water bodies not dominated by floating plants tended to have one or two species (Figure 5b). The results of these analyses did not change if a higher threshold for floating plant dominance (e.g., 80% cover) was used or if the analysis was limited to small water bodies (< 5 ha surface area) or water bodies with higher nutrients (total phosphorus > 0.02 mg $P L^{-1}$).

Discussion

In general, the floating plant species in this study exhibited differences (i.e., response diversity) in their average growth rates across nitrogen, phosphorus, and temperature conditions. Differences among species were typically seen under less favorable conditions (i.e., low nutrients and low temperatures), whereas, species typically had similar growth rates under conditions expected to be most favorable for their growth (i.e., high nutrients and high temperature). When differences were detected, *Lemna minor* and *Spirodela polyrhiza* typically grew at rates that were equal to each other and higher than *Wolffia brasiliensis*. On the other hand, *W. brasiliensis* produced more resting bodies across most experimental conditions, whereas *S. polyrhiza* only occasionally produced turions. *L. minor* never produced turions in these experiments. This suggests a tradeoff between producing floating or sinking biomass under these experimental conditions and may explain the lower relative growth rate of *W. brasiliensis*. In the field, a floating plant-dominated state did not occur more frequently in water bodies with





higher floating plant richness, opposite to the expectation if response diversity is important for formation of floating plant dominance.

The apparent tradeoff between growth and resting body production among floating plant species may have important consequences for this functional group. The different strategies among species can allow the floating plant functional group as a whole to have both rapid growth on the water surface and insurance against perturbation via their resting bodies. Therefore, floating plant polycultures may have a combination of strategies that may not be achievable by a single species. For example, the floating plant functional group in a water body with both *L. minor* and *W. brasiliensis* could have both faster growth at low nutrients or temperatures (due to the traits of *L. minor*) and a greater number of resting bodies to re-colonize the water body at the start of a growing season (due to the traits of *W. brasiliensis*). A polyculture of floating plant species and their unique responses to environmental conditions may allow the functional group to attain higher biomass or persist in a water body over a broader range of conditions.

The lack of a relationship between response diversity of this functional group and its ability to become dominant may be due to a variety of factors. In the field, environmental variability may outweigh the relationship between species richness and the formation of the floating plant state. In addition to response diversity, other factors will influence the occurrence of the floating plant state in the field. For example, water bodies in the northeast United States above a size threshold (~ 5 ha) are rarely dominated by floating plants (McCann, *personal observation*). It is also possible that the floating plant response diversity observed in the laboratory only has a small effect on dominance in the field. Although species differences are quantifiable in the laboratory,





300 their magnitude may not be large enough to be detectable in the field. Furthermore, since floating plant dominance is relative uncommon in this region (<10% of water bodies), there may 301 302 be low statistical power to detect a relationship between species richness and floating plant 303 dominance, especially if effect sizes are small. 304 305 It is also possible that response diversity is only important during transient circumstances or conditions rarely encountered in these surveys. Therefore, the response diversity exhibited by 306 307 floating plant species in the lab will not determine whether this functional group is currently in a 308 dominant state in the field. As a result, species-rich water bodies could lose floating plant species due to local extinction and still maintain floating plant dominance. Response diversity may also 309 310 help this functional group form a dominant state in other geographic regions where low 311 temperatures and low nutrients are more common (e.g., the Upper Midwest United States). Rather than allowing floating plants to achieve dominance, the response diversity observed may 312 313 help this functional group persist in a water body, despite unfavorable conditions. Interestingly, L. minor, which did not produce resting bodies in the lab but typically had the fastest growth rate 314 (along with S. polyrhiza), was the most common floating plant species in this region (present in 315 316 82 of 205 water bodies). Despite differences in growth rates and resting body production, S. polyrhiza and W. brasiliensis occurred in a similar number of water bodies (47 and 42, 317 respectively). 318 319 The lower growth rates observed in Experiment II (Figure 3) relative to Experiment I (Figure 1) 320 may be due to the fact that the nutrient media was changed less frequently (every 3-4 days) 321 322 compared to 2-3 days) and nutrient levels likely decreased to a greater extent between media





324

325

326

327

328

329

330

331

332

333

334

335

336

337

338

339

340

341

342

343

344

345

changes in Experiment II. Also, S. polyrhiza only produced turions in Experiment II. The difference in media change frequency may have caused the difference in turion production between experiments, or there may be differences based on the timing when plants were collected from the field (Mid-August and mid-June for Experiments I and II, respectively). Therefore, strict comparison of the growth rates or turion production between experiments should not be done without consideration of the differences in experimental conditions. There are few others studies of the response diversity of the floating plant functional group to temperature, nutrients, or other environmental variables. Lüönd (1983) measured the response of L. minor, S. polyrhiza, and two other species of Lemna to nitrogen and phosphorus at 25°C. All species increased their growth rates in response to increases of both nutrients, as in this study, and all species decreased their growth rate at extremely high nutrient levels (e.g., 1.75 g N L⁻¹, 1.36 g P L⁻¹) (Lüönd 1983). The presence, but not the rate, of turion production was reported for S. polyrhiza. Unfortunately, no statistical comparisons were made to determine if species had unique responses under particular conditions (Lüönd 1983). Lemon et al. (2001) examined the growth of L. minor, S. polyrhiza, and W. borealis, a cogener of W. brasiliensis at 24°C and very high nutrients (33% v/v Hutner's medium, \sim 31 mg N L⁻¹, \sim 23 mg P L⁻¹), and found that W. borealis has the highest growth rate, while S. polyrhiza has the lowest (in terms of frond number, not area growth rate). Results of turion production were not reported (Lemon et al. 2001). Some studies have examined response diversity to variables not included in this study. Floating plants appear to have response diversity to pH (Hicks 1932, McClay 1976). Lemna minor, Spirodela oligorrhiza, and Wolffia arrhiza all have a similar pH range (pH ~3 to 10), but their optimal pH differs, from mildly acidic (W. arrihiza, pH 5.0 or L. minor pH 6.2) to neutral (S. oligorrhiza, pH





7.0) when grown in the lab at 25°C at very high nutrients (\sim 241 mg N L⁻¹ and \sim 32 mg P L⁻¹) (McClay 1976).

While this study was only able to examine a subset of all floating plant species under particular combinations of environmental conditions, it found some conditions where species have response diversity and others where species are redundant. Further studies, including a greater number of species and environmental variables, as well as determining tradeoffs between responses (e.g., growth or resting bodies), are necessary to determine the full breadth of response diversity of this functional group. For example, previous work on floating plant performance under low temperature conditions (~10 °C) shows that species differ in their minimum temperature (Landolt and Kandeler 1987), which may have important consequences for growth of this functional group at the beginning and end of a growing season. Future work should also consider variability in environmental conditions and species composition through space and time. Floating plants are expected to be easily dispersed by waterfowl and other vectors (Barrat-Segretain 1996); therefore, species composition in a waterbody may change through time, with possible consequences for floating plant dominance.

Conclusions

This study has identified differences in three floating plant species common to the northeast United States. Although species differences existed in the laboratory, there was no statistical support that the species richness of floating plants increases their dominance in the field. Although free-floating plants can be viewed as both a nuisance and an opportunity for applied



368	uses, understanding the species-specific responses of these plants to abiotic conditions is
369	essential for both management and applications.
370	
371	Acknowledgements
372	I would like to thank Ishmael Rahim and Eunice Asare for help with the laboratory experiments,
373	Greg Bugbee and the Connecticut Agricultural Experiment Station for data access, and The
374	Frank Melville Memorial Park Foundation, numerous property owners, and the Suffolk County
375	Parks Department for access to water bodies. This manuscript was improved by feedback from
376	Dianna Padilla, Stephen Baines, Heather Lynch, and Gary Mittelbach.
377	
378	Supplemental Information
379	Appendix A. Converting from turion number to turion area
380	Appendix B. Analysis of growth rates using factorial ANOVAs.
381	Appendix C. Properties of 205 surveyed water bodies.
382	Appendix D. Raw data from experiment I (experimentI.xlsx).
383	Appendix E. Raw data from experiment II (experimentII.xlsx).
384	
385	References
386	Barrat-Segretain MH. 1996. Strategies of reproduction, dispersion, and competition in river
387	plants: A review. Vegetatio 123: 13-37.
388	Capers RS, Selsky R, Bugbee GJ, White JC. 2007. Aquatic plant community invisibility and
389	scale-dependent patterns in native and invasive species richness. <i>Ecology</i> 88: 3135-3143.



390	Crow GE, Hellquist CB. 2000. Aquatic and wetland plants of Northeastern North America:
391	Volume 2. Angiosperms: Monocotyledons. Madison: University of Wisconsin Press.
392	Docauer DM. 1983. A nutrient basis for the distribution of the Lemnaceae. Ph.D. Dissertation,
393	University of Michigan.
394	Downing JA, McCauley E. 1992. The nitrogen: phosphorus relationship in lakes. Limnology and
395	Oceanography 37: 936-945.
396	Driever SM, Van Nes EH, Roijackers RMM. 2005. Growth limitation of Lemna minor due to
397	high plant density. Aquatic Botany 81: 245-251.
398	Elmqvist T, Folke C, Nyström M, Peterson G, Bengtsson J, Walker B, Norberg J. 2003.
399	Response diversity, ecosystem change, and resilience. Frontiers in Ecology and the
400	Environment 1: 488-494.
401	Fulton BA, Brain RA, Usenko S, Back JA, Brooks BW. 2010. Exploring Lemna gibba thresholds
402	to nutrient and chemical stressors: Differential effects of triclosan on internal
403	stoichiometry and nitrate uptake across a nitrogen:phosphorus gradient. Environmental
404	Toxicology and Chemistry 29: 2363-2370.
405	Ge X, Zhang N, Phillips GC, Xu J. 2012. Growing Lemna minor in agricultural wastewater and
406	converting the duckweed biomass to ethanol. Bioresource Technology 124: 485-488.
407	Greenberg BM, Huang XD, Dixon DG. Applications of the aquatic higher plant Lemna gibba for
408	ecotoxicological risk assessment. Journal of Aquatic Ecosystem Health 1: 147-155.
409	Hicks LE. 1932. Ranges of pH tolerance of Lemnaceae. <i>Ohio Journal of Science</i> 32: 237-244.
410	Hillman WS. 1961. The Lemnaceae, or duckweeds: A review of the descriptive and experimental
411	literature. Botanical Review 27: 221-287.



12	Janes RA, Eaton JW, Hardwick K. 1996. The effects of floating mats of <i>Azolla filiculoides</i> Lam
113	and Lemna minuta Kunth on the growth of submerged macrophytes. Hydrobiologia 340
114	23-26.
15	Janse JH, Van Puijenbroek PJTM. 1998. Effects of eutrophication in drainage ditches.
116	Environmental Pollution 102: 547-552.
17	Karpati V, Pomogyi P. 1979. Accumulation and release of nutrients by aquatic macrophytes.
18	Symposium Biologica Hungarica 19: 33-42.
119	Kufel L, Strzałek M, Konieczna A, Izdebska K. 2010. The effect of Stratiotes aloides L. and
20	nutrients on the growth rate of Lemna minor L. Aquatic Botany 92: 168-172.
121	Landolt E, Kandeler R. 1987. The family of Lemnaceae – a monographic study. Volume 2.
122	Zurich: Veröffentlichungen des Geobotanischen Institutes der ETH
123	Lemon GD, Posluszny U, Husband BC. 2001. Potential and realized rates of vegetative
124	reproductive in Spirodela polyrhiza, Lemna minor, and Wolffia borealis. Aquatic Botan
125	70: 79-87.
126	Lüönd AH. 1983. The family of Lemnaceae – a monographic study. Volume 3. Zurich:
127	Veröffentlichungen des Geobotanischen Institutes der ETH.
128	McClay CL. 1976. The effect of pH on the population growth of three species of duckweed:
129	Spirodela oligorrhiza, Lemna minor, and Wolffia arrhiza. Freshwater Biology 6: 125-
130	136.
131	Morris K, Bailey PC, Boon PI, Hughes L. 2003. Alternative stable states in the aquatic
132	vegetation of shallow urban lakes. II. Catastrophic loss of aquatic plants consequent to
133	nutrient enrichment. Marine & Freshwater Research 54: 201-215.



434	Morris PF, Barker WG. 1977. Oxygen transport rates through mats of Lemna minor and Wolffia
435	sp. and oxygen tension within and below the mat. Canadian Journal of Botany 55: 1926-
436	1932.
437	Naeem S, Wright JP. 2003. Disentangling biodiversity effects on ecosystem functioning:
438	Deriving solutions to a seemingly insurmountable problem. <i>Ecology Letters</i> 6: 567-579.
439	Netten JJC, Van Zuidam J, Kosten S, Peeters ETHM. 2011. Differential response to climatic
440	variation of free-floating and submerged macrophytes in ditches. Freshwater Biology 56:
441	1761-1768.
442	Peeters ETH, Van Zuidam JP, Van Zuidam BG, Van Nes EH, Kosten S, Heuts PGM, Roijackers
443	RMM, Netten JCC, Scheffer M. 2013. Changing weather conditions and floating plants
444	in temperate drainage ditches. Journal of Applied Ecology 50: 585-593.
445	Portielje R, Roijackers RMM. 1995. Primary succession of aquatic macrophytes in experimental
446	ditches in relation to nutrient input. Aquatic Botany 50: 127-140.
447	R Development Core Team, 2013. R: A Language and Environment for Statistical Computing. R
448	Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0. Available
449	at: http://www.R-project.org.
450	Rasband WS. 1997-2014. ImageJ. U. S. National Institutes of Health, Bethesda, Maryland, USA,
451	Available at: http://imagej.nih.gov/ij/.
452	Scheffer M, Szabó S, Gragnani A, Van Nes EH, Rinaldi S, Kautsky N, Norberg J, Roijackers
453	RMM, Franken RJM. 2003. Floating plant dominance as a stable state. Proceedings of
454	the National Academy of Sciences of the United States of America 100: 4040-4045.
455	Skillicorn P, Spira W, Journey W. 1993. Duckweed Aquaculture: A new aquatic farming system
456	for developing countries. The World Bank, Washington, D.C.



457	Smart RM, Barko JW. 1985. Laboratory culture of submerged freshwater macrophytes on
458	natural sediments. Aquatic Botany 21: 251-263.
459	Smith SDP. 2012. Identifying and evaluating causes of alternative community states in wetland
460	plant communities. Oikos 121: 675-686.
461	Smith SDP. 2014. The roles of nitrogen and phosphorus in regulating the dominance of floating
462	and submerged aquatic plants in a field mesocosm experiment. Aquatic Botany 112: 1-9.
463	Sterner RW, Elser JJ. 2002. Ecological Stoichiometry: The Biology of Elements from Molecules
464	to the Biosphere. Princeton: Princeton University Press.
465	Szabó S, Roijackers R, Scheffer M. 2003. A simple method for analyzing the effects of algae on
466	the growth of Lemna and preventing algal growth in duckweed bioassays. Archiv für
467	Hydrobiologie 157: 567-575.
468	Szabó S, Roijackers R, Scheffer M, Borics G. 2005. The strength of limiting factors for
469	duckweed during algal competition. Archiv für Hydrobiologie 164: 127-140.
470	Szabó S, Scheffer M, Roijackers R, Waluto B, Braun M, Nagy PT, Borics G, Zambran L. 2010.
471	Strong growth limitation of a floating plant (Lemna gibba) by the submerged macrophyte
472	(Elodea nuttallii) under laboratory conditions. Freshwater Biology 55: 681-690.
473	Verdonschot RCM, Verdonschot PFM. 2013. Shading effects of free-floating plants on drainage-
474	ditch invertebrates. Limnology 15: 225-235.
475	Verma R, Suthar S. 2014. Synchronized urban wastewater treatment and biomass production
476	using duckweed Lemna gibba L. Ecological Engineering 64: 337-343.
477	Xu J, Zhao H, Stomp A, Cheng JJ. 2012. The production of duckweed as a source of biofuels.
478	Biofuels 3: 589-601.





Table 1. N:P mass ratios produced by nine combinations of nitrogen and phosphorus in Experiment II.

		Nitrogen (mg L ⁻¹)		
		0.5	5	10
Phosphorus (mg L ⁻¹)	0.083	6.02	60.24	120.48
	0.83	0.60	6.02	12.05
	1.66	0.30	3.01	6.02

PeerJ

Table 2. One-way ANOVAs for the effect of species on the average relative growth rate (RGR)

486 of floating plants at nine combinations of nutrients and temperature.

Treatment		Averag	Average RGR		
Nutrients	Temperature (°C)	F-statistic	p-value		
low	18	11.403	< 0.001		
	24	39.83	< 0.001		
	30	30.14	< 0.001		
medium	18	5.703	0.011		
	24	7.172	0.004		
	30	8.136	0.002		
high	18	12.44	< 0.001		
	24	4.106	0.031		
	30	4.325	0.027		

487

488

489

Note: Degrees of freedom for all ANOVAs were 2 and 21, except for at low nutrients and 30°C,

where df = 2, 20. Dunn-Šidák adjusted critical p-value is 0.0057. Nutrient levels are low = 0.5

490 mg N L^{-1} and 0.083 mg P L^{-1} , medium = 5 mg N L^{-1} and 0.83 mg P L^{-1} , or high = 10 mg N L^{-1}

491 and 1.66 mg P L⁻¹.

PeerJ

- **Table 3.** One-way ANOVAs for the effect of species on the average relative growth rate (RGR)
- 493 of floating plants at nine combinations of nitrogen and phosphorus.

Treatment		Averag	Average RGR		
Nitrogen	Phosphorus	F-statistic	p-value		
low	low	21.24	< 0.001		
	medium	60.61	< 0.001		
	high	14.1	< 0.001		
medium	low	14.08	< 0.001		
	medium	0.985	0.396		
	high	1.666	0.222		
high	low	0.506	0.613		
	medium	4.727	0.026		
	high	1.283	0.306		

494

- Note: Degrees of freedom for all ANOVAs were 2 and 15. Dunn-Šidák adjusted critical p-value
- 496 is 0.005. Nitrogen levels are low = 0.5 mg N L^{-1} , medium = 5 mg N L^{-1} , and high = 10 mg N L^{-1} .
- 497 Phosphorus levels are low = $0.083 \text{ mg P L}^{-1}$, medium = 0.83 mg P L^{-1} , and high = 1.66 mg P L^{-1} .



Table 4. The frequency of floating plant species compositions and the frequency of floating plant
 cover exceeding two-thirds of the surface area of freshwater lakes and ponds in Connecticut and
 Long Island, NY.

Floating plant		Frequency of	Frequency floating
species richness	Species composition	occurrence	plant cover >66.67%
4	A, LM, SP, W	1	1
	LM, LV, SP, W	1	0
	LM, R, SP, W	1	0
3	LM, LT, SP	2	0
	LM, SP, W	18	7
	LT, SP, W	1	0
	All ≥3 species polycultures	24	8
2	A, W	1	1
	LM, SP	14	2
	LM, W	13	2
	SP, W	2	0
	All 2 species polycultures	30	5
1	A	1	1
	LM	32	4
	LT	1	0
	SP	7	0
	\mathbf{W}	4	2
	All monocultures	45	7
0	None	106	0
	TOTAL	205	20

502

Note: A = Azolla sp., LM = Lemna minor, LT = L. trisulca, LV = L. valdiviana, R = Riccia sp.,

503 $\mathbf{SP} = Spirodela\ polyrhiza$, $\mathbf{W} = Wolffia\ \mathrm{sp}$. Taxa used in the laboratory experiments are

indicated by bold letters.





506	Figure captions
507	Fig. 1 Effect of nutrients and temperature on relative growth rate (RGR) of three species of
508	floating plants. Error bars are standard errors. Post-hoc comparisons among species are for each
509	response variable at each level of nutrients and temperature. Arrows indicate a species that is
510	statistically different (Tukey's HSD $p > 0.05$) at a given nutrient and temperature level. LM =
511	$\label{eq:lemnaminor} \textit{Lemna minor, SP} = \textit{Spirodela polyrhiza}, \ \text{and WB} = \textit{Wolffia brasiliensis}$
512	
513	Fig. 2 Effect of nutrients and temperature on turion formation. Error bars are standard errors.
514	Shared letters indicate no difference between treatment levels (Tukey's HSD $p > 0.05$) for W .
515	brasiliensis turion production. LM = Lemna minor, SP = Spirodela polyrhiza, and WB = Wolffia
516	brasiliensis
517	
518	Fig. 3 Effect of nitrogen and phosphorus on relative growth rate (RGR) of three species of
519	floating plants. Error bars are standard errors. Post-hoc comparisons among species are for each
520	response variable at each level of nitrogen and phosphorus. Arrows indicate a species that is
521	statistically different (Tukey's HSD $p > 0.05$) at a given nutrient and temperature level. N:P
522	ratios are indicated in parentheses above the horizontal axis. $LM = Lemna \ minor$, $SP = Spirodela$
523	polyrhiza, and WB = $Wolffia$ $brasiliensis$
524	
525	Fig. 4 Effect of nitrogen and phosphorus on turion formation of three species of floating plants.
526	When more both L. minor and S. polyrhiza produced turions at a particular treatment level,
527	significant differences between those species are indicated by unique letters (Tukey's HSD,



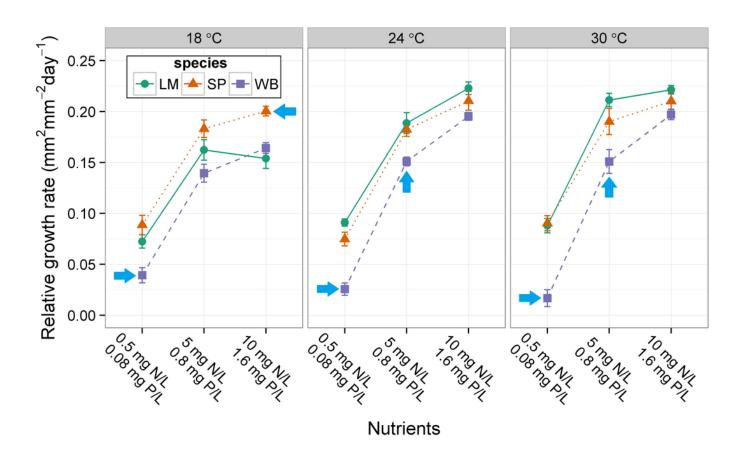
PeerJ

528	p>0.05). Error bars are standard errors. LM = <i>Lemna minor</i> , SP = <i>Spirodela polyrhiza</i> , and WB =
529	Wolffia brasiliensis
530	
531	Fig. 5 Floating plant species richness in a) all water bodies with floating plants present, and b)
532	water bodies with floating plant cover >66.67% of the water surface. Dashed lines indicate
533	expected value if random



Effect of nutrients and temperature on relative growth rate (RGR) of three species of floating plants.

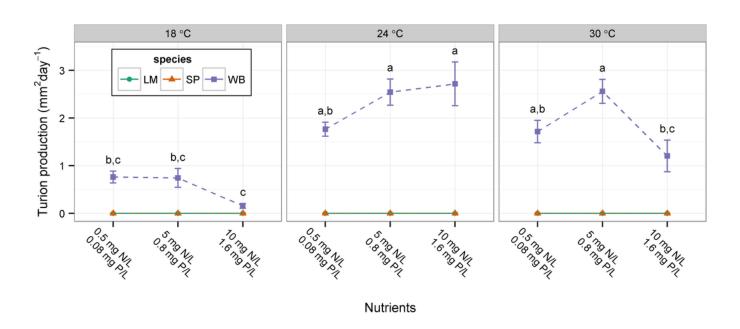
Effect of nutrients and temperature on relative growth rate (RGR) of three species of floating plants. Error bars are standard errors. Post-hoc comparisons among species are for each response variable at each level of nutrients and temperature. Arrows indicate a species that is statistically different (Tukey's HSD p > 0.05) at a given nutrient and temperature level. LM = $Lemna\ minor$, SP = $Spirodela\ polyrhiza$, and WB = $Wolffia\ brasiliensis\ I$





Effect of nutrients and temperature on turion formation.

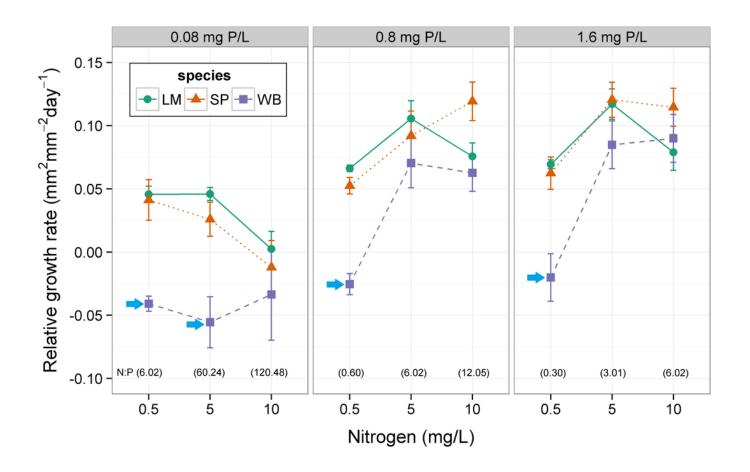
Effect of nutrients and temperature on turion formation. Error bars are standard errors. Shared letters indicate no difference between treatment levels (Tukey's HSD p > 0.05) for W. brasiliensis turion production. LM = Lemna minor, SP = Spirodela polyrhiza, and WB = Wolffia brasiliensis





Effect of nitrogen and phosphorus on relative growth rate (RGR) of three species of floating plants.

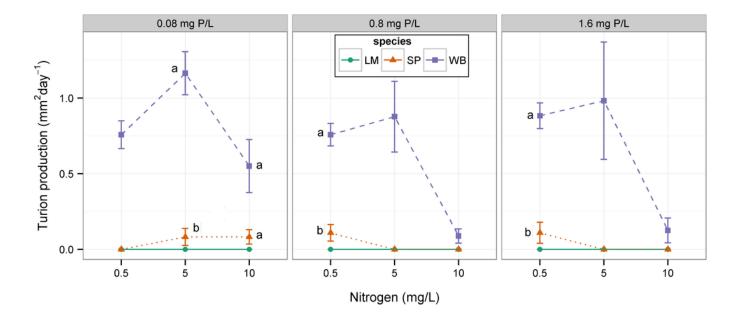
Effect of nitrogen and phosphorus on relative growth rate (RGR) of three species of floating plants. Error bars are standard errors. Post-hoc comparisons among species are for each response variable at each level of nitrogen and phosphorus. Arrows indicate a species that is statistically different (Tukey's HSD p > 0.05) at a given nutrient and temperature level. N:P ratios are indicated in parentheses above the horizontal axis. $LM = Lemna\ minor$, $SP = Spirodela\ polyrhiza$, and $SP = Spirodela\ polyrhiza$





Effect of nitrogen and phosphorus on turion formation of three species of floating plants.

Effect of nitrogen and phosphorus on turion formation of three species of floating plants. When more both L. minor and S. polyrhiza produced turions at a particular treatment level, significant differences between those species are indicated by unique letters (Tukey's HSD, p>0.05). Error bars are standard errors. $LM = Lemna\ minor$, $SP = Spirodela\ polyrhiza$, and $SP = Spirodela\ polyrhiza$, and $SP = Spirodela\ polyrhiza$





Floating plant species richness.

Floating plant species richness in $\bf a$) all water bodies with floating plants present, and $\bf b$) water bodies with floating plant cover >66.67% of the water surface . Dashed lines indicate expected value if random

