# Unraveling habitat-driven shifts in alpha, beta, and gamma diversity of hummingbirds and their floral resource (#95322)

First submission

### Guidance from your Editor

Please submit by 31 Mar 2024 for the benefit of the authors (and your token reward) .



#### **Structure and Criteria**

Please read the 'Structure and Criteria' page for general guidance.



#### **Author notes**

Have you read the author notes on the guidance page?



#### Raw data check

Review the raw data.



#### Image check

Check that figures and images have not been inappropriately manipulated.

If this article is published your review will be made public. You can choose whether to sign your review. If uploading a PDF please remove any identifiable information (if you want to remain anonymous).

#### **Files**

Download and review all files from the <u>materials page</u>.

5 Figure file(s)

5 Table file(s)

# Structure and Criteria



### Structure your review

The review form is divided into 5 sections. Please consider these when composing your review:

- 1. BASIC REPORTING
- 2. EXPERIMENTAL DESIGN
- 3. VALIDITY OF THE FINDINGS
- 4. General comments
- 5. Confidential notes to the editor
- You can also annotate this PDF and upload it as part of your review

When ready submit online.

### **Editorial Criteria**

Use these criteria points to structure your review. The full detailed editorial criteria is on your guidance page.

#### **BASIC REPORTING**

- Clear, unambiguous, professional English language used throughout.
- Intro & background to show context.
  Literature well referenced & relevant.
- Structure conforms to <u>PeerJ standards</u>, discipline norm, or improved for clarity.
- Figures are relevant, high quality, well labelled & described.
- Raw data supplied (see <u>PeerJ policy</u>).

#### **EXPERIMENTAL DESIGN**

- Original primary research within Scope of the journal.
- Research question well defined, relevant & meaningful. It is stated how the research fills an identified knowledge gap.
- Rigorous investigation performed to a high technical & ethical standard.
- Methods described with sufficient detail & information to replicate.

#### **VALIDITY OF THE FINDINGS**

- Impact and novelty not assessed.

  Meaningful replication encouraged where rationale & benefit to literature is clearly stated.
- All underlying data have been provided; they are robust, statistically sound, & controlled.



Conclusions are well stated, linked to original research question & limited to supporting results.



# Standout reviewing tips



The best reviewers use these techniques

Т	p

# Support criticisms with evidence from the text or from other sources

# Give specific suggestions on how to improve the manuscript

# Comment on language and grammar issues

# Organize by importance of the issues, and number your points

# Please provide constructive criticism, and avoid personal opinions

Comment on strengths (as well as weaknesses) of the manuscript

### **Example**

Smith et al (J of Methodology, 2005, V3, pp 123) have shown that the analysis you use in Lines 241-250 is not the most appropriate for this situation. Please explain why you used this method.

Your introduction needs more detail. I suggest that you improve the description at lines 57-86 to provide more justification for your study (specifically, you should expand upon the knowledge gap being filled).

The English language should be improved to ensure that an international audience can clearly understand your text. Some examples where the language could be improved include lines 23, 77, 121, 128 – the current phrasing makes comprehension difficult. I suggest you have a colleague who is proficient in English and familiar with the subject matter review your manuscript, or contact a professional editing service.

- 1. Your most important issue
- 2. The next most important item
- 3. ...
- 4. The least important points

I thank you for providing the raw data, however your supplemental files need more descriptive metadata identifiers to be useful to future readers. Although your results are compelling, the data analysis should be improved in the following ways: AA, BB, CC

I commend the authors for their extensive data set, compiled over many years of detailed fieldwork. In addition, the manuscript is clearly written in professional, unambiguous language. If there is a weakness, it is in the statistical analysis (as I have noted above) which should be improved upon before Acceptance.



# Unraveling habitat-driven shifts in alpha, beta, and gamma diversity of hummingbirds and their floral resource

Hellen Martínez-Roldán 1, María José Pérez-Crespo 2, Carlos Lara Corresp. 3

Corresponding Author: Carlos Lara

Email address: carlos.lara.rodriguez@gmail.com

**Background**. Biodiversity, crucial for understanding ecosystems, encompasses species richness, composition, and distribution. Ecological and environmental factors shape species diversity in communities, categorized into alpha (within habitat), beta (between habitats), and gamma (total region) diversity. Hummingbird communities are influenced by habitat, elevation, and seasonality, making them an ideal system for studying these diversities, shedding light on mutualistic community dynamics and conservation strategies. Methods. Over a year-long period, monthly surveys were conducted to record hummingbird species and their visited flowering plants across four habitat types (oak forest, juniper forest, pine forest, and xerophytic shrubland) in Tlaxcala, Mexico. Three locations per habitat type were selected based on conservation status and distance from urban areas. True diversity measures were used to assess alpha, beta, and gamma diversity of hummingbirds and their floral resources. Environmental factors such as altitude and bioclimatic variables were explored for their influence on beta diversity. **Results**. Our data reveal high heterogeneity in species abundance among habitats. For flowering plants, gamma diversity encompassed 34 species, with oak forests exhibiting the highest richness, while xerophytic shrublands had the highest alpha diversity. In contrast, for hummingbirds, 11 species comprised the gamma diversity, with xerophytic shrublands having the highest richness and alpha diversity. Notably, certain floral resources like Loeselia mexicana and Bouvardia ternifolia emerge as key species in multiple habitats, while hummingbirds such as Basilinna leucotis, Selasphorus platycercus, and Calothorax lucifer exhibit varying levels of abundance and habitat preferences. Beta diversity analyses unveil habitat-specific patterns, with species turnover predominantly driving dissimilarity in composition. Moreover, our study delves into the relationships between these diversity components and environmental factors such as altitude and climate variables. Climate variables, in particular, emerge as significant contributors to

<sup>&</sup>lt;sup>1</sup> Doctorado en Ciencias Biológicas, Universidad Autónoma de Tlaxcala, Tlaxcala, Tlaxcala, Mexico

<sup>2</sup> San Pablo 15, Ocototlán 90100, Tlaxcala, Tlaxcala, Mexico

<sup>&</sup>lt;sup>3</sup> Centro de Investigación en Ciencias Biológicas, Universidad Autónoma de Tlaxcala, San Felipe Ixtacuixtla, Tlaxcala, Mexico



dissimilarity in floral resource and hummingbird communities, highlighting the influence of environmental conditions on species distribution. **Conclusions**. Our results shed light on the complex dynamics of hummingbird-flower mutualistic communities within diverse habitats and underscore the importance of understanding how habitat-driven shifts impact alpha, beta, and gamma diversity. Such insights are crucial for conservation strategies aimed at preserving the delicate ecological relationships that underpin biodiversity in these communities.



## Unraveling habitat-driven shifts in alpha, beta, and gamma

## 2 diversity of hummingbirds and their floral resource

3 Hellen Martínez-Roldán<sup>1</sup>, María José Pérez-Crespo<sup>2</sup> and Carlos Lara<sup>3\*</sup> 4 5 6 <sup>1</sup> Doctorado en Ciencias Biológicas, Universidad Autónoma de Tlaxcala, Tlaxcala, México 7 <sup>2</sup> San Pablo 15, Ocotlan 90100, Tlaxcala, México. 8 <sup>3</sup>Centro de Investigación en Ciencias Biológicas, Universidad Autónoma de Tlaxcala, San Felipe 9 Ixtacuixtla, Tlaxcala, México 10 11 \*Corresponding author: 12 Carlos Lara 13 Centro de Investigación en Ciencias Biológicas, Universidad Autónoma de Tlaxcala, San Felipe 14 Ixtacuixtla, Tlaxcala, Mexico 15 Email address: carlos.lara.rodriguez@gmail.com 16 17 **Abstract** 18 19 **Background**. Biodiversity, crucial for une anding ecosystems, encompasses species richness, 20 composition, and distribution. Ecological and environmental factors shape species diversity in 21 communities, categorized into alpha (within habitat), beta (between habitats), and gamma (total 22 region) diversity. Hummingbird communities are influenced by habitat, elevation, and 23 seasonality, making them an ideal system for studying these diversities, shedding light on 24 mutualistic community dynamics and conservation strategies. 25 **Methods**. Over a year-long period, monthly surveys were conducted to record hummingbird 26 species and their visited flowering plants across four habitat types (oak forest, juniper forest, pine 27 forest, and xerophytic shrubland) in Tlaxcala, Mexico. Three locations per habitat type were 28 selected based on conservation status and distance from urban areas. True diversity measures 29 were used to assess alpha, beta, and gamma diversity of hummingbirds and their floral resources.



30	Environmental factors such as altitude and bioclimatic variables were explored for their
31	influence on beta diversity.
32	<b>Results</b> . Our data reveal high heterogeneity in species abundance among habitats. For flowering
33	plants, gamma diversity encompassed 34 species, with oak forests exhibiting the nighest
34	richness, while xerophytic shrublands had the highest alpha diversity. In contrast, for
35	hummingbirds, 11 species comprised the gamma diversity, with xerophytic shrublands having
86	the highest richness and alpha diversity. Notably, certain floral resources like <i>Loeselia mexicana</i>
37	and Bouvardia ternifolia emerge as key species in multiple habitats, while hummingbirds such as
88	Basilinna leucotis, Selasphorus platycercus, and Calothorax lucifer exhibit varying levels of
39	abundance and habitat preferences. Beta diversity analyses unveil habitat-specific patterns, with
10	species turnover predominantly driving dissimilarity in composition. Moreover, our study delves
11	into the relationships between these diversity components and environmental factors such as
12	altitude and climate variables. Climate variables, in particular, emerge as significant contributors
13	to dissimilarity in floral resource and hummingbird communities, highlighting the influence of
14	environmental conditions on species distribution.
15	Conclusions. Our results shed light on the complex dynamics of hummingbird-flower
16	mutualistic communities within diverse habitats and underscore the importance of understanding
17	how habitat-driven shifts impact alpha, beta, and gamma diversity. Such insights are crucial for
18	conservation strategies aimed at preserving the delicate ecological relationships that underpin
19	biodiversity in these communities.
50	
51	
52	
53	
54	
55	
56	
57	
8	
59	
60	



#### Introduction

- 62 The study of biodiversity, the intricate interplay of life forms within ecosystems, serves as a
- 63 means to depict the structural patterns in communities because it is a key indicator of their
- 64 complexity, interactions, and stability (Tilman, Reich & Knops, 2006; Campbell, Murphy &
- 65 Romanuk, 2011). Its study extends beyond a mere cataloging of species; it involves a
- 66 comprehensive examination of the richness, composition, and distribution of species, spanning
- 67 from local to regional scales (*Jost, 2006*). Ecological factors, both biotic (i.e., species
- 68 interactions) and abiotic (e.g., temperature and precipitation), influence the distribution of
- 69 species and population density within a community (*Pearson & Dawson, 2003*; *Benton, 2009*).
- 70 These environmental and biological factors act as filters that determine which species can
- survive and thrive in a specific area, and their coexistence is contingent upon their specific needs
- and requirements based on competition for resources (Wisz et al., 2013). This way, diversity
- vithin communities is primarily shaped by these ecological processes (*Chesson, 2000*).
- 74 It is widely recognized that species diversity exhibits spatial heterogeneity. For example, at a
- 75 regional scale, significant disparities in species richness have been widely documented among
- habitats (e.g., MacArthur, 1965; Būhning-Gaese, 1997). These spatial trends have given rise to
- 77 the concept of three levels of species diversity: alpha ( $\alpha$ ), beta ( $\beta$ ), and gamma diversity ( $\gamma$ )
- 78 (Whittaker, 1960). The partitioning of biodiversity into three components offers a powerful
- 79 framework to unravel the intricacies of these diversity patterns. Firstly, alpha diversity
- 80 characterizes species richness and abundance within a single habitat, providing insights into the
- 81 structure of local communities. Secondly, beta diversity quantifies the turnover of species
- between habitats, shedding light on the ecological processes driving community assembly and
- 83 turnover. Lastly, gamma diversity, encompassing total species richness across multiple habitats,
- 84 reflects broader regional diversity patterns (Whittaker, 1960). Fundamental topics in ecological
- 85 research have revolved around distribution patterns and mechanisms that maintain species
- 86 diversity across environmental gradients (Lyons & Willig, 2002; McCain, 2009; Wang et al.,
- 87 2017). Understanding these patterns and mechanisms is crucial for devising strategies and
- 88 measures aimed at preserving species diversity in the face of environmental changes.
- 89 Because of their feeding ecology, hummingbirds (Aves: Trochilidae) are closely tied to their
- 90 floral resources (Abrahamczyk & Kessler, 2015). Their extreme specialization in dependence on
- 91 nectar consumption has led these tiny birds to often track the availability of nectar sources by



92	following the blooming of flowers, an ability that enables them to survive and thrive in various
93	habitats across the Americas (Leimberger et al., 2022). The dynamics shaping hummingbird
94	communities have been explored in numerous studies, revealing an intriguing trend.
95	Hummingbird communities in low-lying habitats (≤ 50 m a. s. l.), encompassing both dry and
96	humid forests, experience an upsurge in both species richness and abundance (Buzato, Sazima &
97	Sazima, 2000). In contrast, a different scenario unfolds in habitats surrounded by temperate
98	vegetation at higher and colder elevations (> 2000 m a. s. l. with temperatures around -5°C),
99	such as cloud forests and coniferous forests. In these habitats, there is a tendency for a decrease
100	in the richness and abundance of hummingbird species (Graham et al., 2009; Partida-Lara et al.
101	2018). Interestingly, this general pattern doesn't account for the remarkable species richness in
102	the montane region of the Andes, where elevation has instead generated diverse topographical
103	features that have promoted high speciation rates (Rahbek et al., 2007).
104	In addition to the habitat type's impact on the structure of hummingbird communities,
105	seasonality also exerts an effect due to variations in environmental variables that directly
106	influence the floral resources they utilize, such as precipitation. In this regard, it has been
107	demonstrated that in habitats with scarce precipitation, such as tropical dry forests, the peak
108	flowering of plants visited by hummingbirds primarily occurs during the dry season (Arizmendi
109	& Ornelas, 1990; Bustamante-Castillo, Hernández-Baños & Arizmendi, 2018) Conversely, in
110	temperate environments such as conifeous forests, the flowering peaks of these plants align with
111	the rainy season (Des Granges, 1979; Lara, 2006). In response to this seasonal effect in the
112	environment, there is typically a positive relationship where a greater number of flowers (i.e.,
113	flowering peaks) denotes higher diversity and abundance of hummingbirds at the local level
114	(Cotton, 2007). Therefore, the dynamics of this relationship over time can led hummingbird
115	communities to undergo restructuring (Wolf, Stiles & Hainsworth, 1976; Arizmendi & Ornelas,
116	1990; Lara, 2006).
117	The interaction between hummingbirds and flowers is an ideal context to explore the three
118	diversity components. The diversity of both these groups may be influenced by factors such as
119	resource availability, and habitat specialization. By dissecting the alpha, beta, and gamma
120	diversity patterns within this context, we aim to uncover the mechanisms driving the
121	assemblages and maintenance of these intricate mutualistic communities. Central Mexico is a
122	hotspot of ecological diversity, characterized by its varied topography, altitude gradients, and





climatic variability (Sánchez-Cordero et al., 2005). This ecological heterogeneity provides a
unique backdrop for exploring biodiversity patterns and underlying ecological processes. Among
the states within this region, Tlaxcala, the smallest state in the country (after the capital Mexico
City), holds a unique geographical position that facilitated the collection of comprehensive data
on the diversity of hummingbirds and their flowers across different vegetation types. This
provided insights into the dynamics of these communities within a confined yet ecologically
diverse area. The main goal of our research was to unravel the alpha, beta, and gamma diversity
patterns within hummingbird-flower communities across the most representative habitats of the
region: the oak forest, pine forest, juniper forest, and xerophytic scrubland. These habitats
encompass environmental conditions ranging from typically humid and cold to dry and warm
and are mainly found covering altitudinal ranges from 2400 to 2700 m a.s.l., although pine
forests can be found at elevations as high as 4000 m a.s.l. at the highest point in the region, La
Malinche volcano. Considering the variability in our studied habitats, we expected significant
variations in alpha, beta, and gamma diversity in hummingbird-flower communities across oak
forest, pine forest, juniper forest, and xerophytic scrubland habitats due to their distinct
environmental conditions. Additionally, we hypothesized that abiotic factors such as altitude,
temperature, humidity, and resource availability would influence species composition between
these habitats (beta diversity). Finally, we expected higher alpha diversity in habitats with more
varied conditions, while beta diversity will likely correlate with specific environmental factors
distinguishing each habitat. Our study holds theoretical significance in elucidating the
complexities of alpha, beta, and gamma diversity within mutualistic systems. Moreover, from a
practical standpoint, our findings can inform conservation strategies aimed at preserving the
delicate ecological relationships that underpin the biodiversity of these communities.

#### **Materials & Methods**

148 Study area

From February 2014 to January 2015, samplings were carried out in four types of vegetation (hereafter referred to as "habitats") characteristic of the state of Tlaxcala, Mexico: oak forest (OF), juniper forest (JF), pine forest (PF), and xerophytic shrubland (XS). Based on digital land use and vegetation maps at a 1:250,000 scale, as well as information about the vegetation within

the state of Tlaxcala (INEGI, 2009, 2010; Acosta, Delgado & Cervantes, 1992; Luna, Morrone



154	& Espinoza, 2007), three locations were selected for each habitat (Figure 1). For their selection,
155	these locations met the following requirements: (i) belong to conserved areas according to <i>INEGI</i>
156	(2010), (ii) be distant from areas km), and urban (iii) be separated from each other to ensure
157	sampling independence (average distance between locations greater than 13 km). Subsequently,
158	for each habitat and covering the three selected locations, five 500 m transects were placed with
159	20 m wide bands on each side, and a minimum distance of 100 m between transects. For each
160	transect, its georeference and altitude (m a.s.l.) were obtained using a portable GPS (Garmin
161	Etrex 30). A total of 20 transects were obtained for the four habitat types.
162	In each location, the transects were established in sites that could encompass the dominant tree
163	species for the habitat type. In OF, species of the Quercus genus predominate, such as Q.
164	crassipes, Q. glaucoides, Q. laurina, and Q. mexicana. The dominant tree species in JF is
165	Juniperus deppeana. In PF, characteristic species include Pinus montezumae, P. hartwegii, P.
166	patula, and P. leiophylla. Finally, in XS, dominant species include Yucca filifera, Nolina
167	longifolia, Dasylirion acrotriche, and Opuntia robusta (Figure 1).
168	
169	Sampling of hummingbirds and their flower plants
170	To identify and quantify the abundance of hummingbirds (H) and the flowering plants they
171	visited (FP), monthly surveys were conducted over a 12-month period at five transects
172	established for each habitat type. Sampling was carried out from 8:00 to 13:00 h. During this
173	period, all the hummingbirds detected within the transect were recorded, whether they were
174	observed foraging on the flowers, perched, or in flight. The observed individuals were identified
175	with the assistance of specialized field guides (Williamson, 2001; Arizmendi & Berlanga, 2014).
176	Using this information, we obtained the number of individuals per hummingbird species for each
177	survey.
178	Concurrently, all FP species within a transect (i.e., plants exhibiting tubular flowers, bright
179	colors, and nectar production; Faegri & van Der Pijl, 1979) were recorded. Species that did not
180	fit the proposed ornithophilous syndrome were also included in the records if hummingbirds
181	were observed foraging on them. Floral abundance was measured as the number of open flowers
182	per plant species in each transect. The identification of FP species was conducted using
183	dichotomous keys (Calderón & Rzedowski, 2001). Assessment of sample completeness (sample
184	coverage cov



ver. 2023.03.0+386 (RStudio Team, 2022) with 'iNext' function in the iNEXT package (Hsieh,

186 Ma, & Chao, 2016).

187

188

#### True diversity measures

- 189 To assess the structure and differences in H and FP assemblages in the region, we performed an
- analysis of regional diversity (gamma diversity) by considering all habitats as a unit.
- 191 Additionally, we conducted a detailed analysis of local diversity within each habitat (alfa
- diversity), examined how the respective assemblages differ between communities (beta
- diversity), and explored the origins of differences among habitats, including species turnover and
- variations in species richness. Furthermore, we assessed the potential role of environmental
- 195 factors in explaining differences between communities within each habitat. These concepts are
- 196 pivotal for understanding biological processes across diverse habitats, the structure of biological
- 197 communities, and the distribution of species at local and regional level. Their practical
- applications extend to environmental management and conservation of biodiversity.
- Each diversity index, H, can be expressed as its true diversity index or equivalent numbers
- 200 (qD(H)), also referred to as Hill numbers (Jost, 2006; Moreno et al., 2017). Equivalent numbers
- 201 represent the essential components (i.e., species, communities) that a balanced community with
- 202 equally common species would possess, assuming that the diversity index of the balanced
- 203 community matches that of the real community (Jost, 2006, 2010; Pereyra & Moreno, 2013).
- 204 Thus, effective numbers depict the structure of the real community in equivalent units, enabling
- comparisons of the degree of change between communities (*Jost, 2006*; 2007). Effective
- 206 numbers  ${}^{q}D$  derived from the following formula (*Jost, 2007*):

$$207 qD = (\sum_{i=1}^{s} p_i^q)^{1/1-q}$$

208

- where  $p_i$  is the relative frequency of species i, q is the order of true diversity measurement, and S
- 210 is the number of species. The parameter q has an exponential property that determines the
- sensitivity of the index to the relative abundance of species (*Jost, 2006*; 2007). Species richness
- 212 corresponds to the diversity index of order 0 and is insensitive to the relative frequency of
- species. The true diversity measure of order 1 is equivalent to the exponential of Shannon's
- 214 entropy and weights rare and common species proportionally to their abundance. The diversity



215	measures of order 2 are equivalent to Simpson's inverse measures, which favor abundant species
216	while excluding rare ones (Hill, 1973; Jost, 2006; 2007).
217	To measure diversity across the region encompassing the four habitat types, we computed the
218	true gamma diversity index ( ${}^qD\gamma$ ) using the multiplicative partitioning of regional diversity ( ${}^qD\gamma$ )
219	as proposed by Whittaker (1960; 1972), where ${}^{q}D\gamma = {}^{q}D\alpha * {}^{q}D\beta$ , and ${}^{q}D\beta = {}^{\alpha}D\gamma / {}^{q}D\alpha$ . The
220	equivalent numbers, expressed as ${}^{0}\mathrm{D}\alpha$ , denotes the number of species in the communities ( ${}^{0}\mathrm{D}\beta$ )
221	required to match the total species count in the region ( ${}^{0}$ D $\gamma$ ).
222	To evaluate diversity at a local level, we calculated alfa diversity (orders q=0,1,2) for the
223	community composition within each habitat (OF, JF, PF, XS) concerning the H and FP species
224	assemblages.
225	Among communities, changes in species composition are explained by $\beta$ diversity (Whitaker,
226	1960). The $\beta$ diversity can arise from two processes: species turnover (B <sub>3</sub> ) and differences in
227	species richness ( $B_{rich}$ ); both indexes identify the source of disparities between communities
228	(Carvalho, Cardoso & Gomes, 2012). These two components explain $\beta$ diversity additively (B <sub>cc</sub>
229	= $B_{3} + B_{rich}$ ). To derive $\beta$ diversity and its components ( $B_{3}$ , $B_{rich}$ ), three measures were
230	calculated: a) species common to both sites, b) species exclusive to one site, and c) species
231	exclusive to the other site (see formulas in Carvalho, Cardoso & Gomes, 2012). B <sub>cc</sub> represents a
232	proportion of dissimilarity between two communities, where 0 indicates that communities share
233	all species, and 1 corresponds to communities that do not share any species. Additionally,
234	species turnover (B_3) varies from 0 (when species composition is identical) to 1 (when species
235	composition is entirely different). The values of $B_{\text{rich}}$ follow the same scale from 0 to 1 (when
236	species richness is equal or different respectively).
237	Furthermore, following <i>Jost</i> (2007), gamma diversity was calculated for orders $q = 0$ and $q = 1$ ,
238	considering the unequal weighting of H and FP communities. Alpha diversity, essential for
239	understanding each community's composition was assessed for orders 0, 1 and 2. Finally, beta
240	diversity and its components across the four habitats for both communities were computed
241	according to Carvalho, Cardoso & Gomes (2012) and Carvalho et al., (2013). All analysis were
242	performed with RStudio, ver. 2023.03.0+386 (RStudio Team, 2022), using the vegan package
243	······································

The relationship between beta diversity and environmental factors





245	Subsequently, the correlation of $\beta$ diversity (B <sub>cc</sub> , B <sub>3</sub> , B <sub>rich</sub> ) and environmental factors such as
246	altitude, and 22 bioclimatic variables obtained from the WorldClim website
247	(http://www.worldclim.org), was assessed using Mantel tests (Sokal & Rohlf, 1995). For this
248	purpose, the values of each bioclimatic variable were extracted for each transect, and a principal
249	component analysis (PCA) was performed to condense the abiotic variables. Highly correlated
250	variables were removed, as well as those with less contribution to the components explaining
251	>90% of the variance. The selected variables were annual precipitation (Bio12), precipitation of
252	wettest quarter (Bio16), and altitude. Dissimilarity matrices were constructed using the Bray-
253	Curtis method for the selected variables. Simple and partial Mantel tests were conducted with
254	9,999 permutations. The Mantel tests were computed with RStudio, ver. 2023.03.0+386 (RStudio
255	Team, 2022), using the vegan package.
256	
257	Results
258	Abundance of flowering plants and hummingbirds
259	The samplings conducted throughout the study in the four habitat types allowed for the total
260	recording of 34 FP species, which were classified into 22 genera, 17 families, and 11 orders
261	(Supplemental file 1). Of the total quantified flower abundance, 83% was recorded in five FP
262	species: Loeselia mexicana (24%), Bouvardia ternifolia (14%), Castilleja tenuiflora (18%),
263	Penstemon roseus (16%), and Salvia elegans (11%). The last three FP species belong to the
264	order Lamiales (45% of the total abundance). Likewise, <i>L. mexicana</i> , <i>C. tenuiflora</i> , and <i>B.</i>
265	ternifolia were shared species in all four habitat types, thus being characteristic FP species within
266	the region (Figure 2B). Therefore, the description of the results hereafter will be particularly
267	based on these plant sees, as well as in the case of the hummingbird species referred to below.
268	Regarding the H species, considering all the sampled habitats, a total of 11 species were
269	recorded, classified into 9 genera and one family (Trochilidae). In terms of abundance, three H
270	species comprised 86% of the total abundance. Basilinna leucotis was the most abundant
271	hummingbird species in the region (69%), followed in much lower abundance by Selasphorus
272	platycercus (11%), Colibri thalassinus (6.3%), and Calothorax lucifer (5%). The first three H
273	species were recorded in all four habitat types, while C. lucifer was only recorded in XS
274	(Supplemental file 1, Figure 2A).



275	The abundances of the above mentioned FP and H species, exhibited high heterogeneity among
276	the studied habitats. For example, L. mexicana was the most abundant FP species (relative to
277	other plant species present) in the sampled sites of JF (69%) and OF (56%), but very scarce in
278	abundance in PF and XS (6%). Likewise, C. tenuiflora was particularly abundant in XS (47%).
279	In contrast, it was recorded with low abundance in the other habitats (PF = $12\%$ , OF = $0.1\%$ , JF
280	= 0.2%). Regarding B. ternifolia, it was an abundant FP species in JF (27%), XS (21%), and OF
281	(16%), but not in PF (4%). In PF, both P. roseus (41%) and S. elegans (27%) were abundant
282	species in this habitat. In contrast, in OF, the abundance of both species was low (1.5%), while in
283	JF and XS were not recorded (Supplemental file 1, Figure 3F
284	In the case of the H species, their abundances were also highly variable among the sampled
285	habitats. B. leucotis was the most abundant species throughout the study in PF (81%), OF (80%),
286	and JF (78%), while in XS was less abundant (15%). Conversely, S. platycercus was the most
287	abundant in XS (34%), while in other habitats its abundance was less (JF = 12%, OF = 7%, PF =
288	1%). In the case of <i>C. thalassinus</i> , this specie was one of the most abundant in PF (12%) and
289	showed very low abundances in the remaining nabitat types (<4%). Finally, C. lucifer was an
290	abundant species found exclusively in the XS habitat (28%) (Supplemental file 1, Figure 3A).
291	The observed number of FP species and H species in the study seemed to reach an asy ote
292	in relation to our sampling effort across the four sampled habitats (a total of 180 hours of evenly
293	distributed observation efforts for each habitat throughout the study). For FP species, we
294	detected 99.62% for the PF, 99.91% for OF, 99.57% for JF and, 99.95% for XS according to the
295	Chao2 estimator, after conducting 12 samples for each habitat type throughout the study.
296	Likewise, we detected 98.15% of the H species estimated for the PF, 98.40% for OF, 96.25% for
297	JF and 95.16% of those estimated for the XS.
298	
299	True diversity measures
300	Richness at regional level of FP was 34 species ( ${}^{0}D\gamma$ ), with an average local richness ( ${}^{0}D\alpha$ ) of
301	16.5 effective species and 2.06 effective communities ( ${}^{0}D\beta$ ) necessary to account the regional
302	species richness within the region. This implies that on average, $48.5\%$ ( $1/^{0}D\beta$ ) of the total FP
303	species are present in a single habitat. For H assemblages, the average richness ( ${}^{0}D\alpha$ ) was 7.3
304	effective species, representing 66.6% of the total species recorded within the region ( ${}^{0}D\gamma=11$ ).
305	With a ${}^{0}D\beta$ of 1.5 effective communities needed to achieve regional richness, it suggests minimal



306	species turnover within the region. Considering the effective communities in H, the species
307	recorded in XS (9 spp.) and OF (2 spp.) contribute to completing the regional richness (Table 1)
308	In terms of the regional diversity ${}^{1}D\alpha$ (equiprobable species) in the FP, an average community
309	calculated 4.4 effective species, while 8.7 effective species were observed in the entire region
310	$(^{1}D\gamma)$ . The communities required to complement $^{1}D\gamma$ are 2 $(^{1}D\beta)$ , indicating that an average
311	community contained 50% of the equiprobable species in the region. For the H assemblages, an
312	average community displayed 2.5 effective species ( ${}^{1}D\alpha$ ), an at the region exhibited 3.3 effective
313	species ( ${}^{1}D\gamma$ ). To complement ${}^{1}D\gamma$ , 1.3 communities were required ( ${}^{1}D\beta$ ), with an average
314	community encompassing 77% of the equiprobable species in the region (Table 1). Regional
315	diversity ( ${}^{1}D\gamma$ ) aligned closely with the abundant species recorded within the region (Figure 2).
316	Regarding alpha diversity ( $\alpha$ ), the habitat with the highest richness ( ${}^{0}D$ ) of FP species was OF
317	(22 species), followed by PF (18 species), and XS (14 species). JF (12 species) had the lowest
318	richness, with the lowest number of effective species of ${}^{1}D$ (2.1) and ${}^{2}D$ (1.8), particularly
319	recording two dominant species (L. mexicana and B. ternifolia) (Figure 3B). In contrast, habitats
320	with the highest number of effective species in orders 1 and 2 are PF ( ${}^{1}D = 5.3$ , ${}^{2}D = 3.8$ ) and XS
321	$(^{1}D = 5.2, ^{2}D = 3.5)$ , respectively (Figure 4B; Teble 2). Consequently, in terms of order 1
322	diversity, on average, PF and XS exhibited 2.5 times more diverse than JF and 1.4 times more
323	diverse than OF. PF presented the five most abundant species within the region (P. roseus, S.
324	elegans, C. ternuiflora, L. mexicana, B. ternifolia) (Figure 2B), while XS shared three species
325	with PF (C. ternuiflora, B. ternifolia, L. mexicana) and had two exclusive abundant species
326	(Salvia chamaedryoides and Salvia melissodora) (Supplemental file 1).
327	The habitat with the highest diversity of H species was XS, recording the highest richness ( $^0D$
328	= 9) and the greatest number of effective species ( ${}^{1}D$ = 5.2, ${}^{2}D$ = 4.2). In this habitat, five
329	abundant hummingbird species were found, two of which ranked among the most abundant
330	species in the region, and one was exclusive to XS (B. leucotis, S. platycercus, S. rufus, A.
331	colubris, and C. lucifer, respectively) (Figure 3A). In contrast, the lowest diversity of H species
332	was observed in OF, PF, and JF, with assemblages having a similar number of effective species.
333	Considering the order 1 diversity measure, XS was, on average, 2.77 times more diverse than
334	OF, JF, and PF (Figure 4A; Table 2).
335	
336	Beta diversity (β)

Beta diversity (β)



337	The B <sub>cc</sub> values obtained among the FP communities indicate dissimilarity ranging from 0.52 to
338	0.77 (where 1 represents maximum dissimilarity). XS is dissimilar compared to the other three
339	habitats (>0.70) (Table 3). The dissimilarity among all communities is primarily attributed to
340	species turnover (B <sub>3</sub> ), except in OF vs. JF, where dissimilarity is attributed to differences in
341	richness (B <sub>rich</sub> ) (Figure 5B ble 3). The H communities have dissimilarity ranging from 0.38 to
342	0.56. Overall, dissimilarity is driven by species turnover (Figure 5A; Table 3). When evaluating
343	the beta diversity between pairs of sites, a very similar trend was found for FP and H. Where, the
344	dissimilarity was mainly due to $B_{\underline{\ \ }3}$ , with low contribution in $B_{\text{rich}}$ . However, the highest values
345	total beta (Bcc) occurred between habitats of FP and low values in assemblages of H, showing
346	more similarity in species composition of H between habitats (Figure 5).
347	
348	The relationship between beta diversity and environmental factors
349	Mantel's simple and partial tests for FP species, between beta components and selected
350	environmental factors in the study, showed a positive correlation in $B_{cc}$ dissimilarities with
351	climate variables and altitude ranging from $r = 0.34$ to $r = 0.45$ . Partial correlations confirm that
352	climate variables contribute more in the relationship. Similar results were obtained for species
353	turnover (B <sub>3</sub> ), with correlation coefficients ranging from $r = 0.27$ to $r = 0.4$ (Table 4). In
354	summary, we found variation in the species turnover rate for both measured variables (altitude
355	and climate variables). However, environmental conditions had a greater effect on the
356	dissimilarity of FP species assemblages. For H species assemblages, differences in $B_{cc} \text{and} B_{\_3}$
357	are explained by climate variables ( $r = 0.45$ ) and not by altitude (Table 4). Correlations for
358	richness differences (B_rich) were not significant in either case (FP and H).
359	
360	Discussion
361	Our study adds a crucial layer of understanding to the intricate ecosystems of our research region
362	by unraveling the complex relationships between flowering plants (FP) and hummingbirds (H).
363	Documenting 34 FP species, spanning 22 genera, 17 families, and 11 orders, underscores the
364	ecological significance of the floral community (Potts et al., 2010; Ollerton, Winfree & Tarrant,
365	2011). The implications of this diversity resonate profoundly, encompassing ecosystem stability,
366	pollination dynamics, and overall biodiversity (Hoehn et al., 2008).



367	The prevalence of five key FP species—Loeselia mexicana, Bouvardia ternifolia, Castilleja
368	tenuiflora, Penstemon roseus, and Salvia elegans—in terms of flower abundance is notable. The
369	flowers of these five FP species are red, which aligns with the fact that 84% of the plants visited
370	by hummingbirds in the Americas are red (Scogin, 1983). These species may hold keystone
371	positions in the ecosystem, influencing community composition and structure (Paine, 1969).
372	This result is consistent with previous suggestions highlighting that North American bird-
373	pollinated flora is dominated by temperate herbaceous lineages, such as Castilleja and
374	Penstemon (Abrahamczyk & Renner, 2015). Therefore, their prominence serves as an indicator
375	of their vital roles in the ecological web. Furthermore, the presence of characteristic FP species
376	shared across all four habitat types underscores their ecological importance and potential role as
377	indicators of habitat health (Lechner, Chan & Campos-Arceiz, 2018).
378	Within the realm of hummingbird diversity, our study identifies 11 recorded species,
379	categorized into 9 genera within the family Trochilidae. Hummingbirds are highly diverse and
380	abundant in the Americas, particularly in tropical and subtropical regions (Howell & Webb,
381	1995). However, their species richness tends to decrease as we move towards higher latitudes
382	and elevations, such as temperate habitats. Despite this, the hummingbird species richness at our
383	study region is relatively higher compared with other temperate forests of North and South
384	America, where up to 13 species may be present (Abrahamczyk & Renner, 2015; López-
385	Segoviano, Bribiesca & Arizmendi, 2018). Typically, hummingbird communities are mainly
386	composed of medium-sized species (Stiles, 1981), of which resident species tend to be the most
387	abundant (Arizmendi & Ornelas, 1990). In our study habitats, seven out of eleven hummingbird
388	species may be considered medium to large-sized (Arizmendi & Berlanga, 2014). Among these,
389	the resident Basilinna leucotis emerges as the dominant hummingbird species, constituting a
390	substantial 69% of the regional hummingbird population. This dominance extends beyond
391	numerical abundance, potentially influencing plant-hummingbird interactions, with cascading
392	effects on plant reproductive success and community structure (Stiles, 1981; Magrach et al.,
393	2020). Interestingly, the second most abundant hummingbird species was the long-distance
394	migrant Selasphorus platycercus. The presence of this species was recorded throughout most of
395	the year in all four habitat types, suggesting that in this region, both resident and winter
396	migratory populations can be found and may even reproduce in these habitats. Based on these
397	findings, it seems that at least some hummingbird species, such as B. leucotis and S. platycercus,



398	demonstrate adaptation to multiple habitat types, suggesting a degree of habitat generalism.
399	These species were found in multiple habitats, indicating they can utilize a range of
400	environments for foraging and nesting.
401	The observed heterogeneity in species abundance across different habitats within our research
402	region offers a captivating glimpse into the tapestry of ecological dynamics. These variations in
403	species abundance likely reflect habitat-specific resource availability, microclimatic conditions,
404	and niche partitioning (Whittaker, 1960; Magurran et al., 2010). This mosaic of habitats beckons
405	researchers to delve deeper into the ecological processes at play. Our findings hold profound
406	implications for conservation and habitat management, underlining the pressing need to consider
407	habitat preferences and ecological niches (Margules & Pressey, 2000). Understanding the
408	intricacies of resource utilization patterns among FP and H species within different habitats
409	guides the strategic prioritization of habitats for protection and conservation, thereby sustaining
410	biodiversity and ecosystem services (Whittaker, Willis & Field, 2001; Krauss et al., 2010).
411	The computation of true gamma diversity ( ${}^qD\gamma$ ) and true beta diversity ( ${}^qD\beta$ ) provides a
412	quantitative foundation for unraveling the regional biodiversity of FP and H species. These
413	metrics, integral to contemporary ecological research (Chao et al., 2014), lay the groundwork for
414	informed regional biodiversity assessments and conservation planning (Jost et al., 2010). The
415	revelation of low species turnover for H assemblage suggests some stability in the species
416	composition across the habitats but higher turnover for FP reflects the presence of habitat
417	specialists alongside widespread species. Species turnover is influenced by the availability and
418	variety of resources within each habitat, which determine the communities composition. As a
419	result, the biota undergoes changes based on the specific requirements for food resources and
420	spatial aspects of the species (Halffter, 1998). This observation highlights the complexities of
421	ecological dynamics within the region, offering insights into the interconnectedness of species
422	and their environments (Vellend et al., 2017; Chase et al., 2011). This nuanced understanding of
423	species turnover has far-reaching implications for ecosystem connectivity and resilience. The
424	presence of habitat specialists signals unique ecological roles and dependencies within their
425	respective ecosystems, urging conservationists to consider the holistic preservation of habitats
426	(Devictor et al., 2007; Cardinale et al., 2012).
427	Our exploration of alpha diversity among different habitats unveils intriguing patterns of
428	species richness and evenness. Habitats such as Pine Forest (PF) and Xeric Scrubland (XS) stand

429	out as bastions of high alpha diversity of flowering plants, suggesting the presence of diverse and
430	evenly distributed species assemblages (Magurran, 1988; Grime, 1998). In contrast, Juniper
431	Forest (JF) exhibits lower diversity, beckoning further investigation into the drivers of this
432	pattern, including resource availability and biotic interactions (Connell, 1978; Tilman, 1982).
433	XS, on the other hand, shines as a habitat with high diversity for both FP and H species,
434	especially in terms of order 1 (species accounting almost all of the total abundance and
435	proportionately). Understanding the variations in alpha diversity among habitats has profound
436	implications for crafting effective land management and conservation strategies. Our findings
437	underscore the imperative to prioritize the protection and restoration of diverse habitats to
438	maintain biodiversity and enhance ecosystem resilience (Noss & Cooperrider, 1994).
439	The exploration of beta diversity, especially the dissimilarity among FP and H communities,
440	unveils the uniqueness of species assemblages across habitats. The high dissimilarity observed in
441	Xeric Scrubland (XS) points to the existence of distinctive ecological communities, potentially
442	shaped by factors such as dispersal limitation, environmental gradients, or species interactions
443	(Legendre et al., 2009). The dissimilarity in species composition is primarily due to species
444	turnover, implying unique ecological roles and contributions of different species to each habitat.
445	These findings emphasize the paramount importance of preserving a variety of habitats to
446	safeguard the diverse assemblages they harbor. By prioritizing conservation efforts across
447	heterogeneous landscapes, we promote ecosystem resilience and augment the capacity of these
448	ecosystems to adapt to changing environmental conditions (Pressey et al., 2007; Hobbs et al.,
449	2014).
450	We found a significant relationship between environmental factors (specifically climate
451	variables) and dissimilarities in both FP species and H species assemblages. The positive
452	correlation observed in $B_{cc}$ indicates that as climate variables and altitude vary, the dissimilarity
453	in the composition of FP species increases. Furthermore, the results show that climate variables
454	play a more influential role in this relationship compared to altitude. This suggests that the
455	climatic conditions of a habitat are particularly important in shaping the composition of FP. The
456	variations in species turnover (B <sub>3</sub> ) also align with this pattern, reinforcing the impact of
457	environmental conditions on the diversity and composition of FP species. In the case of
458	hummingbird species, the dissimilarities in $B_{cc}$ and $B_{\underline{\ 3}}$ are mainly influenced by climate



459	variables, not altitude. This emphasizes the significance of climate in determining the
460	composition and diversity of hummingbird species across different habitats.
461	However, our study did not find significant correlations for richness differences (B_rich) for
462	both FP and H species. This implies that differences in species richness between habitats were
463	not strongly related to the measured environmental variables and altitude. Thus, the positive
464	correlations detected between beta diversity and climate variables, offer compelling insights into
465	the potential influence of climate change on species composition within our research region
466	(Bellard et al., 2012). The ramifications of shifting climate conditions extend to alterations in
467	species distributions, impacting ecological dynamics and the provisioning of ecosystem services
468	(Parmesan, 2006).
469	Previous studies have shown that climate change can be particularly threatening to
470	hummingbirds by affecting the phenology of floral resources on which they depend (Inouye et al.
471	2000; McKinney et al. 2012). Even minor changes in blooming dates may be of consequence, as
472	hummingbirds will eventually arrive after flowering begins, which could reduce their nesting
473	success (Aldridge et al. 2011; McKinney et al. 2012). This disruption in the flowering phenology
474	within and among different habitats can affect both latitudinal and altitudinal migration
475	undertaken by hummingbirds following these floral resources. The established interaction
476	networks between hummingbirds and their floral resources should be incorporated into future
477	studies of geographic distribution models and climate change. Thus, our findings accentuate the
478	central role played by environmental conditions in shaping species assemblages (Chase et al.,
479	2011). This knowledge informs the development of effective habitat conservation and restoration
480	strategies that account for the influence of climate and topography on ecosystem structure and
481	function (Sax et al., 2007; Hobbs et al., 2014).
482	
483	Conclusions
484	Our study provides a comprehensive understanding of the abundance, composition, and diversity
485	of flowering plants and hummingbirds across different habitat types. The identified dominant FP
486	and H species play crucial roles in the ecological dynamics of these habitats. Moreover, the
487	analysis of true diversity measures and beta diversity highlights the importance of community
488	species turnover and regional species richness. Habitat variations significantly influence



189	abundance and diversity patterns, emphasizing the need for habitat-specific conservation
190	strategies.
191	The findings of this research not only deepen our knowledge of ecological interactions but also
192	underscore the necessity of considering environmental factors in biodiversity conservation.
193	Understanding how habitats shape the diversity and composition of these critical ecological
194	components is essential for effective conservation and sustainable management of natural
195	ecosystems. These insights are pivotal for future research and conservation efforts, providing a
196	solid foundation for further investigation into the intricate relationships between hummingbirds,
197	flowering plants, and their habitats. By considering the dynamic interplay of environmental
198	variables and biodiversity, we can develop informed strategies to protect and preserve these
199	invaluable ecological partnerships for future generations.
500	
501	Acknowledgements
502	We gratefully acknowledge Sandra Rodríguez, Lucia Salas, Magali Luna, Liliana Xicohténcatl
503	and Juan Manuel González for field assistance. Two anonymous reviewers provided useful
504	comments on previous versions of the manuscript. This work constitutes partial fulfillment of H.
505	M. R.'s doctorate requirements at UATx.
506	
507	References
808	Abrahamczyk S, Kessler M. 2015. Morphological and behavioural adaptations to feed on
509	nectar: how feeding ecology determines the diversity and composition of hummingbird
510	assemblages. <i>Journal of Ornithology</i> <b>156:</b> 333–347. DOI: <u>10.1007/s10336-014-1146-5</u> .
511	Abrahamczyk S, Renner SS. 2015. The temporal build-up of hummingbird/plant mutualisms in
512	North America and temperate South America. BMC Evolutionary Biology 15: 1–12. DOI:
513	10.1186/s12862-015-0388-z.
514	Acosta PR, Delgado MJL, Cervantes SP. 1992. La vegetación del estado de Tlaxcala.
515	Jardín Botánico de Tizatlán. Gobierno del estado de Tlaxcala. Tizatlán, Tlaxcala, Mexico,
516	p.1-31.
517	Aldridge G, Inouye DW, Forrest JR, Barr WA, Miller-Rushing AJ. 2011. Emergence of a
518	mid-season period of low floral resources in a montane meadow ecosystem associated



519	with climate change. <i>Journal of Ecology</i> <b>99:</b> 905–913. DOI: <u>10.1111/j.1365-</u>
520	<u>2745.2011.01826.x</u> .
521	Arizmendi MC, Ornelas JF. 1990. Hummingbirds and their floral resources in a tropical dry
522	forest in Mexico. Biotropica 22: 172-180. DOI: 10.2307/2388410.
523	Arizmendi MC, Berlanga H. 2014. Colibríes de México y Norteamérica, 1st edn. CONABIO,
524	México.
525	Bellard C, Bertelsmeier C, Leadley P, Thuiller W, Courchamp F. 2012. Impacts of climate
526	change on the future of biodiversity. <i>Ecology Letters</i> <b>15(4):</b> 365–377. DOI:
527 528	10.1111/j.1461-0248.2011.01736.x.  Benton MJ. 2009. The Red Queen and the Court Jester: species diversity and the role of biotic
529	and abiotic factors through time. Science 323(5915): 728-732. DOI:
530	10.1126/science.1157719.
531	Būhning-Gaese K. 1997. Determinants of avian species richness at different spatial
532	scales. Journal of Biogeography 24(1): 49–60. DOI: 10.1111/j.1365-
533	<u>2699.1997.tb00049.x</u> .
534	Bustamante-Castillo M, Hernández-Baños BE, Arizmendi MC. 2018. Hummingbird
535	diversity and assemblage composition in a disturbed tropical dry forest of
536	Guatemala. Tropical Conservation Science 11: 1940082918793303. DOI:
537	10.1177/1940082918793303.
538	Buzato S, Sazima M, Sazima I. 2000. Hummingbird-pollinated floras at three Atlantic Forest
539	sites 1. Biotropica 32: 824–841. DOI: 10.1111/j.1744-7429.2000.tb00621.x.
540	Campbell V, Murphy G, Romanuk TN. 2011. Experimental design and the outcome and
541	interpretation of diversity-stability relations. Oikos 120(3): 399-408. DOI:
542	10.1111/j.1600-0706.2010.18768.x.
543	Cardinale BJ, Duffy JE, Gonzalez A, Hooper DU, Perrings C, Venail, P, Narwani A, Mace
544	GM, Tilman D, Wardle DA, Kinzig AP, Daily GC, Loreau M, Grace JB,
545	Larigauderie A, Srivastava DS, Naeem S. 2012. Biodiversity loss and its impact on
546	humanity. Nature 486(7401): 59–67. DOI: 10.1038/nature11148.
547	Carvalho JC, Cardoso P, Gomes P. 2012. Determining the relative roles of species
548	replacement and species richness differences in generating beta-diversity patterns. Global
549	<i>Ecology and Biogeography</i> <b>21(7):</b> 760–771. DOI: <u>10.1111/j.1466-8238.2011.00694.x</u> .



550	Carvalho JC, Cardoso P, Borges PA, Schmera D, Podani J. 2013. Measuring fractions of beta
551	diversity and their relationships to nestedness: a theoretical and empirical comparison of
552	novel approaches. Oikos 122(6): 825–834. DOI: 10.1111/j.1600-0706.2012.20980.x.
553	Chase JM, Kraft NJ, Smith KG, Vellend M, Inouye BD. 2011. Using null models to
554	disentangle variation in community dissimilarity from variation in $\alpha$ -diversity.
555	Ecosphere <b>2(2):</b> 1–11. DOI: <u>10.1890/ES10-00117.1</u> .
556	Chao A, Gotelli NJ, Hsieh TC, Sander EL, Ma KH, Colwell RK, Ellison AM. 2014.
557	Rarefaction and extrapolation with Hill numbers: a framework for sampling and
558	estimation in species diversity studies. Ecological Monographs 84: 45-67. DOI:
559	<u>10.1890/13-0133.1</u> .
560	Chesson P. 2000. Mechanisms of maintenance of species diversity. Annual Review of Ecology
561	and Systematics 31(1): 343–366. DOI: 10.1146/annurev.ecolsys.31.1.343.
562	Connell JH. 1978. Diversity in tropical rain forests and coral reefs: high diversity of trees and
563	corals is maintained only in a nonequilibrium state. <i>Science</i> <b>199(4335):</b> 1302–1310. DOI:
564	10.1126/science.199.4335.1302.
565	Cotton PA. 2007. Seasonal resource tracking by Amazonian hummingbirds. <i>Ibis</i> 149(1): 135–
566	142. DOI: <u>10.1111/j.1474-919X.2006.00619.x</u>
567	Des Granges JL. 1979. Organization of a tropical nectar feeding bird guild in a variable
568	environment. The Living Bird 17: 199–236.
569	Devictor V, Julliard R, Couvet D, Lee A, Jiguet F. 2007. Functional homogenization effect of
570	urbanization on bird communities. <i>Conservation Biology</i> <b>21(3):</b> 741–751. DOI:
571	10.1111/j.1523-1739.2007.00671.x.
572	Faegri K, van Der Pijl L. 1979. The Principles of Pollination Ecology, 3rd edn. Pergamon
573	Press, Oxford, p. 1–256.
574	Graham CH, Parra JL, Rahbek C, McGuire JA. 2009. Phylogenetic structure in tropical
575	hummingbird communities. Proceedings of the National Academy of Sciences 106:
576	19673–19678. DOI: <u>10.1073/pnas.0901649106</u> .
577	Grime JP. 1998. Benefits of plant diversity to ecosystems: immediate, filter and founder
578	effects. <i>Journal of Ecology</i> <b>86(6):</b> 902–910. DOI: <u>10.1046/j.1365-2745.1998.00306.x</u> .
579	Halffter G. 1998. A strategy for measuring landscape biodiversity. <i>Biology International</i> 36: 3–
580	17.



581	Hill MO. 1973. Diversity and evenness: a unifying notation and its consequences. <i>Ecology</i> 54
582	(2): 427–432. DOI: <u>10.2307/1934352</u> .
583	Hoehn P, Tscharntke T, Tylianakis JM, Steffan-Dewenter I. 2008. Functional group diversity
584	of bee pollinators increases crop yield. Proceedings of the Royal Society B: Biological
585	Sciences 275(1648): 2283–2291. DOI: 10.1098/rspb.2008.0405.
586	Hobbs RJ, Higgs E, Hall CM, Bridgewater P, Chapin III FS, Ellis EC, Ewel JJ, Hallett
587	LM, Harris J, Hulvey KB, Jackson ST, Kennedy PL, Kueffer C, Lach L, Lantz TC,
588	Lugo AE, Mascaro J, Murphy SD, Nelson CR, Perring MP, Richardson DM,
589	Seastedt TR, Standish RJ, Starzomski BM, Suding KN, Tognetti PM, Yakob L,
590	Yung, L. 2014. Managing the whole landscape: historical, hybrid, and novel
591	ecosystems. Frontiers in Ecology and the Environment 12(10): 557–564. DOI:
592	<u>10.1890/130300</u> .
593	Howell SN, Webb S. 1995. A guide to the birds of Mexico and northern Central America.
594	Oxford University Press.
595	Hsieh TC, Ma KH, Chao A. 2016. iNEXT: an R package for rarefaction and extrapolation of
596	species diversity (Hill numbers). Methods in Ecology and Evolution 7(12): 1451–1456.
597	DOI: <u>10.1111/2041-210X.12613</u> .
598	Inouye DW, Barr B, Armitage KB, Inouye BD. 2000. Climate change is affecting
599	altitudinal migrants and hibernating species. Proceedings of the National Academy of
600	Sciences 97: 1630–1633. DOI: 10.1073/pnas.97.4.1630.
501	INEGI (Instituto Nacional de Estadística, Geografía e Informática). 2009. Modelo Digital de
502	Elevación, Escala 1:50000. México.
503	INEGI (Instituto Nacional de Estadística, Geografía e Informática). 2010. Carta de uso de
504	suelo y vegetación. Serie IV, escala 1: 250 000. Ciudad de México: Instituto Nacional de
505	Estadística, Geografía e Informática. Website:
606	https://www.inegi.org.mx/temas/usosuelo/default.
507	Jost L 2006. Entropy and diversity. Oikos 113(2): 363–375. DOI: 10.1111/j.2006.0030-
608	<u>1299.14714.x</u> .
509	Jost L. 2007. Partitioning diversity into independent alpha and beta components. Ecology 88
510	(10): 2427–2439. DOI: <u>10.1890/06-1736.1</u> .
611	Jost, L. 2010. The relation between evenness and diversity. <i>Diversity</i> 2(2): 207–232. DOI:



512	10.3390/d2020207.
513	Jost L, DeVries P, Walla T, Greeney H, Chao A, Ricotta C. 2010. Partitioning diversity for
514	conservation analyses. <i>Diversity and Distributions</i> <b>16(1):</b> 65–76. DOI: <u>10.1111/j.1472-</u>
515	<u>4642.2009.00626.x</u> .
616	Krauss J, Bommarco R, Guardiola M, Heikkinen RK, Helm A, Kuussaari M, Lindborg R,
517	Öckinger E, Pärtel M, Pino J, Pöyry J, Raatikainen KM, Sang A, Stefanescu C,
518	Teder T, Zobel M, Steffan-Dewenter I. 2010. Habitat fragmentation causes immediate
519	and time-delayed biodiversity loss at different trophic levels. Ecology Letters 13(5): 597-
520	605. DOI: <u>10.1111/j.1461-0248.2010.01457.x</u> .
521	Lara C. 2006. Temporal dynamics of flower use by hummingbirds in a highland temperate
522	forest in Mexico. Ecoscience 13: 237–29. DOI: 10.2980/1195-
523	6860(2006)13%5B23:TDOFUB%5D2.0.CO;2.
524	Lechner AM, Chan FKS, Campos-Arceiz A. 2018. Biodiversity conservation should be a core
525	value of China's Belt and Road Initiative. Nature Ecology & Evolution 2(3): 4087-409.
626	DOI: <u>10.1038/s41559-017-0452-8</u> .
527	Leimberger KG, Dalsgaard B, Tobias JA, Wolf C, Betts MG. 2022. The evolution, ecology,
528	and conservation of hummingbirds and their interactions with flowering plants.
529	Biological Reviews 97(3): 923–959. DOI: 10.1111/brv.12828.
630	Legendre P, Mi X, Ren H, Ma K, Yu M, Sun IF, He F. 2009. Partitioning beta diversity in a
631	subtropical broad-leaved forest of China. Ecology 90(3): 663-674. DOI: 10.1890/07-
632	<u>1880.1</u> .
633	López-Segoviano G, Bribiesca R, Arizmendi MC. 2018. The role of size and dominance in the
634	feeding behaviour of coexisting hummingbirds. Ibis 160(2): 283–292. DOI:
635	<u>10.1111/ibi.12543</u> .
636	Lyons SK, Willig MR. 2002. Species richness, latitude, and scale-sensitivity. Ecology 83(1):
637	47–58. DOI: <u>10.1890/0012-9658(2002)083%5B0047:SRLASS%5D2.0.CO;2</u> .
638	Luna I, Morrone JJ, Espinosa D. 2007. Biodiversidad de la Faja Volcánica Transmexicana.
639	CONABIO, UNAM, Mexico. p. 255–271.
640	MacArthur RH. 1965. Patterns of species diversity. <i>Biological Reviews</i> 40(4): 510–533. DOI:
541	10.1111/j.1469-185X.1965.tb00815.x.
542	Magrach A, Lara C, Luna UM, Díaz-Infante S, Parker I. 2020. Community-level



543	reorganizations following migratory pollinator dynamics along a latitudinal
544	gradient. Proceedings of the Royal Society B 287(1930): 20200649. DOI:
545	10.1098/rspb.2020.0649.
646	Magurran AE. 1988. Ecological diversity and its measurement. Princeton University Press.
547	Magurran AE, Baillie SR, Buckland ST, Dick JM, Elston DA, Scott EM, Smith RI,
548	Somerfield PJ, Watt AD. 2010. Long-term datasets in biodiversity research and
549	monitoring: assessing change in ecological communities through time. Trends in Ecology
650	& Evolution <b>25(10):</b> 574–582. DOI: <u>10.1016/j.tree.2010.06.016</u> .
651	Margules CR, Pressey RL. 2000. Systematic conservation planning. Nature 405(6783): 243-
652	253. DOI: <u>10.1038/35012251</u> .
653	McCain CM. 2009. Global analysis of bird elevational diversity. Global Ecology and
654	Biogeography <b>18(3)</b> : 346–360. DOI: <u>10.1111/j.1466-8238.2008.00443.x</u> .
655	McKinney AM, CaraDonna PJ, Inouye DW, Barr B, Bertelsen CD, Waser NM. 2012.
656	Asynchronous changes in phenology of migrating Broad-tailed hummingbirds
657	and their early-season nectar resources. <i>Ecology</i> <b>93:</b> 1987–1993. DOI: <u>10.1890/12-</u>
558	<u>0255.1</u> .
559	Moreno CE, Calderón-Patrón JM, Arroyo-Rodríguez V, Barragán F, Escobar F, Gómez-
660	Ortiz Y, Martín-Regalado N, Martínez-Falcón AP, Martínez-Morales MA, Mendoza
661	E, Ortega-Martínez IJ, Pérez-Hernández CX, Pineda E, Pineda-López R, Rios-Díaz
662	CL, Rodríguez P, Rosas F, Schondube JE, Zuria I. 2017. Measuring biodiversity in
663	the Anthropocene: a simple guide to helpful methods. <i>Biodiversity and Conservation</i> 26:
664	2993–2998. DOI: <u>10.1007/s10531-017-1401-1</u> .
665	Noss RF, Cooperrider A. 1994. Saving nature's legacy: protecting and restoring biodiversity.
666	Island Press.
667	Ollerton J, Winfree R, Tarrant S. 2011. How many flowering plants are pollinated by
668	animals? Oikos 120(3): 321–326. DOI: 10.1111/j.1600-0706.2010.18644.x.
669	Paine RT. 1969. A note on trophic complexity and community stability. The American
670	Naturalist 103(929): 91–93. DOI: 10.1086/282586.
571	Parmesan C. 2006. Ecological and evolutionary responses to recent climate change. Annual
672	Review of Ecology, Evolution, and Systematics 37: 637–669. DOI:
573	10.1146/annurev.ecolsys.37.091305.110100.



574	Partida-Lara R, Enríquez PL, Pérez JRV, de Bonilla EPD. 2018. Estructura espacio-temporal
675	de la diversidad taxonómica y funcional de colibríes en la reserva de la biosfera el
676	Triunfo, Chiapas, Mexico. Ornitología Neotropical 29: 37-50. DOI:
677	10.58843/ornneo.v29i1.229.
678	Pearson RG, Dawson TP. 2003. Predicting the impacts of climate change on the distribution of
679	species: are bioclimate envelope models useful?. Global Ecology and Biogeography
680	<b>12(5):</b> 361–371. DOI: <u>10.1046/j.1466-822X.2003.00042.x</u> .
681	Pereyra LC, Moreno CE. 2013. Divide y vencerás: revisión de métodos para la partición de la
682	diversidad regional de especies en sus componentes alfa y beta. Revista Chilena de
683	Historia Natural <b>86(3)</b> : 231–240. DOI: <u>10.4067/S0716-078X2013000300001</u> .
684	Potts SG, Biesmeijer JC, Kremen C, Neumann P, Schweiger O, Kunin WE. 2010. Global
685	pollinator declines: trends, impacts and drivers. Trends in Ecology & Evolution 25(6):
686	345–353. DOI: <u>10.1016/j.tree.2010.01.007</u> .
687	Pressey RL, Cabeza M, Watts ME, Cowling RM, Wilson KA. 2007. Conservation planning in
688	a changing world. Trends in Ecology & Evolution 22(11): 583-592. DOI:
589	10.1016/j.tree.2007.10.001.
590	Rahbek C, Gotelli NJ, Colwell RK, Entsminger GL, Rangel TFL, Graves GR. 2007.
591	Predicting continental-scale patterns of bird species richness with spatially explicit
592	models. Proceedings of the Royal Society B: Biological Sciences 274(1607): 165–174.
593	DOI: <u>10.1098/rspb.2006.3700</u> .
594	RStudio Team. 2022. RStudio: integrated development environment for R. RStudio, PBC,
595	Boston, MA, USA. Website: http://rstudio.com.
696	Sánchez-Cordero V, Cirelli V, Munguial M, Sarkar S. 2005. Place prioritization for
597	biodiversity content using species ecological niche modeling. Biodiversity Informatics 2:
598	11–23. DOI: <u>10.17161/bi.v2i0.9</u> .
599	Sax DF, Stachowicz JJ, Brown JH, Bruno JF, Dawson MN, Gaines SD, Grosberg RK,
700	Hastings A, Holt RD, Mayfield MM, O'Connor MI, Rice WR. 2007. Ecological and
701	evolutionary insights from species invasions. Trends in Ecology & Evolution 22(9): 465-
702	471. DOI: <u>10.1016/j.tree.2007.06.009</u> .
703	Scogin R. 1983. Visible floral pigments and pollinators. In: Jones CE, Little RJ eds. Handbook
704	of experimental pollination biology, New York: Scientific and Academic Editions, p.



705	160–172.
706	Stiles FG. 1981. Geographical aspects of birdflower coevolution, with particular reference to
707	Central America. Annals of the Missouri Botanical Garden 68(2): 323-351. DOI:
708	<u>10.2307/2398801</u> .
709	Sokal RR, Rohlf FJ. 1995. Biometry. 2nd edn. San Francisco: W. H. Freeman & Co, p. 859.
710	Tilman D. 1982. Resource competition and community structure. Princeton University Press.
711	Tilman D, Reich PB, Knops JM. 2006. Biodiversity and ecosystem stability in a decade-long
712	grassland experiment. Nature 441(7093): 629-632. DOI: 10.1038/nature04742.
713	Vellend M, Baeten L, Becker-Scarpitta A, Boucher-Lalonde V, McCune JL, Messier J,
714	Myers-Smith IH, Sax D F. 2017. Plant biodiversity change across scales during the
715	Anthropocene. Annual Review of Plant Biology 68(1): 563-586. DOI: 10.1146/annurev-
716	arpla nt-042916-040949.
717	Wang CT, Long RJ, Wang QJ, Ding LM, Wang MP. 2007. Effects of altitude on plant-
718	species diversity and productivity in an alpine meadow, Qinghai-Tibetan
719	plateau. Australian Journal of Botany 55(2): 110–117. DOI: 10.1071/BT04070.
720	Wisz MS, Pottier J, Kissling WD, Pellissier L, Lenoir J, Damgaard CF, Dormann CF,
721	Forchhammer MC, Grytnes JA, Guisan A, Heikkinen RK, Høye TT, Kühn I, Luoto
722	M, Maiorano L, Nilsson MC, Normand S, Öckinger E, Schmidt NM, Termansen M,
723	Timmermann A, Wardle DA, Aastrup P, Svenning JC. 2013. The role of biotic
724	interactions in shaping distributions and realised assemblages of species: implications for
725	species distribution modelling. <i>Biological Reviews</i> <b>88(1):</b> 15–30. DOI: <u>10.1111/j.1469-</u>
726	<u>185X.2012.00235.x</u> .
727	Whittaker RH. 1960. Vegetation of the Siskiyou mountains, Oregon and California. Ecological
728	Monographs <b>30(3):</b> 279–338. DOI: <u>10.2307/1943563</u> .
729	Whittaker RH. 1972. Evolution and measurement of species diversity. <i>Taxon</i> 21: 213–251.
730	DOI: <u>10.2307/1218190</u> .
731	Whittaker RJ, Willis KJ, Field R. 2001. Scale and species richness: towards a general,
732	hierarchical theory of species diversity. <i>Journal of Biogeography</i> <b>28(4):</b> 453–470. DOI:
733	10.1046/j.1365-2699.2001.00563.x.
734	Williamson S. 2001. A field guide to hummingbirds of North America. Houghton Mifflin
735	Harcourt, Boston.



736	Wolf LL, Stiles FG, Hainsworth FR. 1976. Ecological organization of a tropical, highland
737	hummingbird community. The journal of Animal Ecology 45(2): 349–379. DOI:
738	10.2307/3879.
739	
740	



### Table 1(on next page)

Multiplicative partition of the gamma diversity (true diversity, modified from Jost, 2007) into its components: D $\gamma$  (regional diversity), D $\beta$  (effective communities), and D $\alpha$  (average alpha).

 $D\alpha$  and  $D\gamma$  are expressed in the same units of species, while  $D\beta$  is expressed in communities. Superscripts correspond to diversity values of orders 0 and 1, based on Hill numbers representing the effective number of species or communities.





5

2 Table 1. Multiplicative partition of the gamma diversity (true diversity, modified from Jost,

3 2007) into its components: Dy (regional diversity), D $\beta$  (effective communities), and D $\alpha$  (average

4 alpha). D $\alpha$  and D $\gamma$  are expressed in the same units of species, while D $\beta$  is expressed in

communities. Superscripts correspond to diversity values of orders 0 and 1, based on Hill

numbe 6 7 Gamma diversity rs Dγ diversity Hummingbirds **Flowering Plants** represe 8 and 0D $^{1}D$ O $^{1}D$ 9 nting its components 34 Dγ 11 3.3 8.7 the 10 1.5 1.3 Dβ 2.06 2 effecti 11 7.3 2.5 16.5 4.4  $D\alpha$ 12 ve

13 number of species or communities.

14

15 16

17

18

19



## Table 2(on next page)

Alpha diversity (true diversity, modified from *Jost, 2006*) of hummingbirds and their flowering plants in oak forest (OF), juniper forest (JF), pine forest (PF), and xerophytic shrubland (XS).

Superscripts correspond to diversity values of orders 0, 1, and 2, represented by Hill numbers, reflecting the effective number of species.



- 1 Table 2. Alpha diversity (true diversity, modified from *Jost, 2006*) of hummingbirds and their
- 2 flowering plants in oak forest (OF), juniper forest (JF), pine forest (PF), and xerophytic
- 3 shrubland (XS). Superscripts correspond to diversity values of orders 0, 1, and 2, represented by
- 4 Hill numbers, reflecting the effective number of species.

			Alpha	diversity		
	Н	Hummingbirds		Flowering Plan		ıts
Habitat Type	-0D	$^{1}D$	$^{2}\mathrm{D}$	$^0\mathrm{D}$	$^{1}D$	$^{2}D$
OF	7	2.2	1.5	22	3.7	2.6
PF	7	2	1.4	18	5.3	3.8
JF	6	2.2	1.5	12	2.1	1.8
XS	9	5.2	4.2	14	5.2	3.5

6

7



## Table 3(on next page)

Beta diversity based on the partition of total beta diversity ( $B_{cc}$ ), species replacement [ $B_{\underline{\ 3}}$ ] and species richness differences [ $\beta_{rich}$ ]) for hummingbirds and their flowering plants.

This analysis was carried out across four sampled habitat types: oak forest (OF), juniper forest (JF), pine forest (PF), and xerophytic shrubland (XS).

- 1 Table 3. Beta diversity based on the partition of total beta diversity (B<sub>cc</sub>), species replacement
- 2  $[B_3]$  and species richness differences  $[\beta_{rich}]$ ) for hummingbirds and their flowering plants. This
- 3 analysis was carried out across four sampled habitat types: oak forest (OF), juniper forest (JF),
- 4 pine forest (PF), and xerophytic shrubland (XS).

		Beta diversity						
	H	ummingbir	ds	Flowering Plants				
Habitat type	$B_{3}$	$\mathrm{B}_{\mathrm{rich}}$	$\mathrm{B}_{\mathrm{cc}}$	$\mathrm{B}_{\_3}$	$\mathrm{B}_{\mathrm{rich}}$	$\mathrm{B}_{\mathrm{cc}}$		
OF vs. JF	0.25	0.13	0.38	0.09	0.43	0.52		
OF vs. PF	0.44	0	0.44	0.43	0.14	0.57		
OF vs. XS	0.36	0.18	0.55	0.43	0.29	0.71		
JF vs. PF	0.44	0.11	0.56	0.50	0.25	0.75		
JF vs. XS	0.20	0.30	0.50	0.60	0.10	0.70		
PF vs. XS	0.20	0.20	0.40	0.62	0.15	0.77		

6



### **Table 4**(on next page)

Correlation results (Mantel tests) between beta diversity of hummingbirds and their flowering plants, altitude and climatic variables were analyzed for each locality.

Additionally, we conducted Partial Mantel tests to examine the results after eliminate the effects of altitude (Climate Variables-Altitude) and climatic variables (Altitude-Climate Variables).



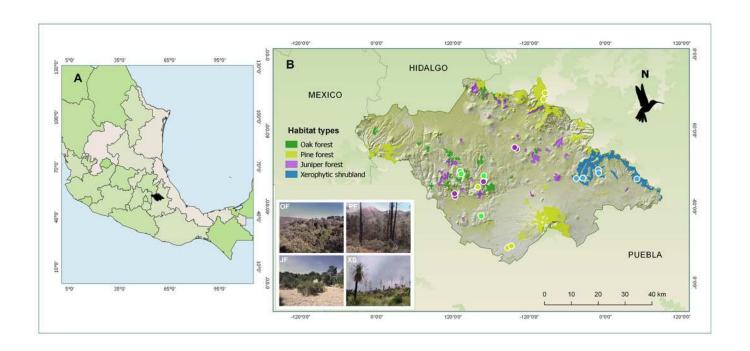
- 1 Table 4. Correlation results (Mantel tests) between beta diversity of hummingbirds and their
- 2 flowering plants, altitude and climatic variables were analyzed for each locality. Additionally,
- 3 we conducted Partial Mantel tests to examine the results after eliminate the effects of altitude
- 4 (Climate Variables-Altitude) and climatic variables (Altitude-Climate Variables).

	Hummingbirds							
	Altitude		Climate Variables		Climate Variables- Altitude		Altitude-Climate Variables	
	r	p	r	p	r	p	r	p
Всс	0.01	0.50	0.45	<0.0 1	0.45	<0.01	0.03	0.41
B_3	0.20	0.91	0.31	<0.0 1	0.30	<0.01	-0.19	0.90
Bric h	0.22	0.13	0.01	0.41	0.03	0.34	0.22	0.13

					Flowering Pla	nts		
	Altitude		Climate Variables		Climate Variables- Altitude		Altitude-Climate Variables	
	$\overline{r}$	<i>p</i>	$\overline{r}$		r	$\overline{P}$	r	$\overline{P}$
Всс	0.34	<0,0 1	0.45	<0.0 1	0.51	<0.01	0.42	<0.01
B_3	0.27	0.01	0.40	<0.0 1	0.44	<0.01	0.33	<0.01
Bric h	0.05	0.54	-0.12	0.93	-0.12	0.94	-0.06	0.57

Maps showing the monitored habitats and locations in the state of Tlaxcala, Mexico, where the diversity of hummingbirds and their flowering plants was studied.

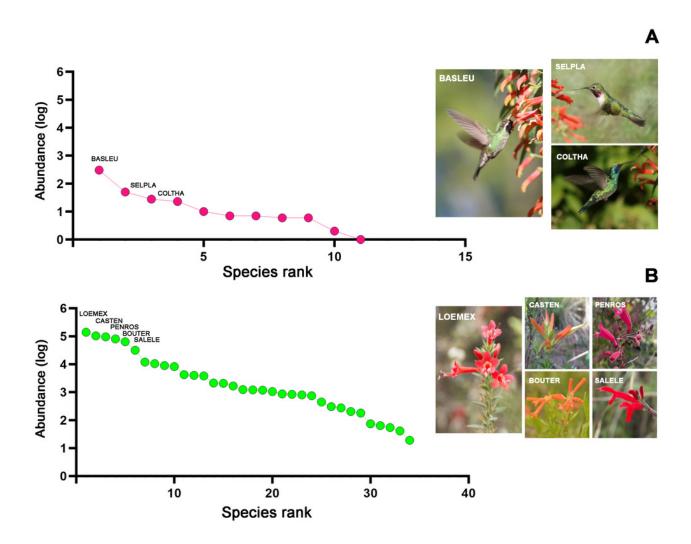
(A) Geographical location. (B) Monitored locations within each habitat. The colored circles represent the transects established for each habitat type: Oak forest (OF) in green, jurier forest (JF) in purple, pine forest (PF) in yellow, and xerophytic shrubland (XS) in blue. Sources: ESRI, Garmin, INEGI, (2009). Uso del suelo y vegetación, escala 1:250000, serie IV. 2009, and Qgis version 2.18, 2016. QGIS Geographic Information System. QGIS Association. <a href="http://www.qgis.org">http://www.qgis.org</a>. Photo credit: Hellen Martínez-Roldán.



Rank/abundance plots for hummingbirds and their flowering plants species at the regional level in Tlaxcala, Mexico.

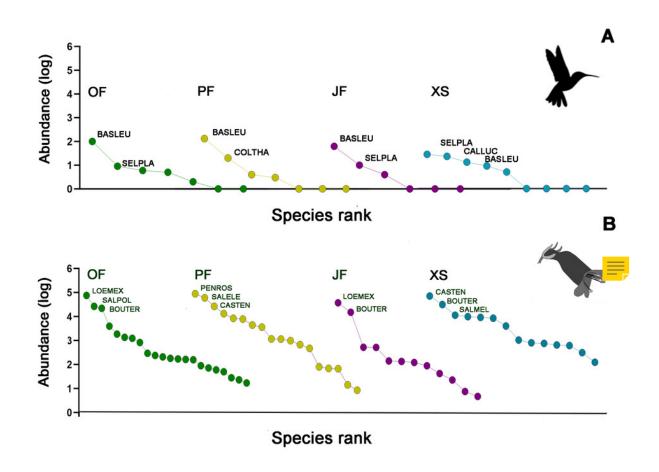
Rank/abundance curves show the distribution of hummingbird and plant species from most to least abundant. (A) *Basilinna leucotis* (BASLEU), *Selasphorus platycercus* (SELPLA), and *Colibri thalassinus* (COLTHA) highly dominate in all sampled habitat types, while (B) *Loeselia mexicana* (LOEMEX), *Castilleja tenuiflora* (CASTEN), *Penstemon roseus* (PENROS), *Bouvardia ternifolia* (BOUTER), and *Salvia elegans* (SALELE) were the most abundant plant species within the region. Photo credit: Ubaldo Marquez-Luna, Hellen Martínez-Roldán, Juan Manuel González, María José Pérez-Crespo and Carlos Lara.





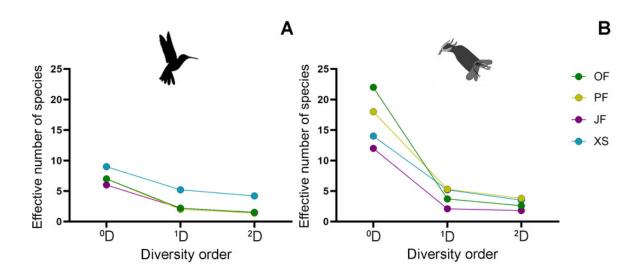
Rank/abundance plots for the hummingbirds and their flowering plant species by each sampled habitat type.

(A) Basilinna leucotis (BASLEU), Selasphorus platycercus (SELPLA), Colibri thalassinus (COLTHA) and Calothorax lucifer (CALLUC) highly dominate in all sampled habitat types, while (B) Loeselia mexicana (LOEMEX), Castilleja tenuiflora (CASTEN), Penstemon roseus (PENROS), Bouvardia ternifolia (BOUTER), Salvia elegans (SALELE), S. polystachya (SALPOL) and S. melissodora (SALMEL) were the most abundant plant species in each sampled habitat type: Oak forest (OF), juniper forest (JF), pine forest (PF), and xerophytic shrubland (XS).



Alpha diversity profiles of hummingbirds and their flowering plant species in the four sampled habitat types.

By following the true diversity concept (*Jost*, *2006*), we obtained the diversity profiles for (A) hummingbirds and (B) flowering plants, showing variation in the number of effective species for each sampled habitat type: Oak forest (OF), juniper forest, pine forest (PF), and xerophytic shrubland (XS). Superscripts correspond to diversity values of orders 0, 1, and 2; values for orders 1 and 2 are shown as Hill numbers, representing the effective number of species.





Contribution of species turnover and differences in species richness to beta diversity of hummingbirds and flowering plants.

Plots show beta diversity of (A) hummingbirds and (B) their flowering plant species, where each segment shows the proportion of each component for each habitat pair: Oak forest (OF), juniper forest (JF), pine forest (PF), and xerophytic shrubland (XS).

