

Microphytoplankton variations during coral spawning at Los Roques, southern-Caribbean

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Phytoplankton drives primary productivity in marine pelagic systems. This is also true for the oligotrophic waters in coral reefs, where natural and anthropogenic sources of nutrients can alter pelagic trophic webs. In this study, microphytoplankton assemblages were characterized for the first time in relation to expected coral spawning dates in the Caribbean. A hierarchical experimental design was used to examine these assemblages in Los Roques archipelago, Venezuela, at various temporal and spatial scales for spawning events in both 2007 and 2008. At four reefs, superficial water samples were taken daily for 9 days after the full moon of August, including days before, during and after the expected days of coral spawning. Microphytoplankton assemblages comprised 100 microalgae taxa at up to 50 cells per mL (mean \pm 8 SD) and showed temporal and spatial variations related to the coral spawning only in 2007. However, chlorophyll *a* concentrations increased during and after the spawning events in both years, and this was better matched with analyses of higher taxonomical groups (diatoms, cyanophytes and dinoflagellates), that also varied in relation to spawning times in 2007 and 2008, but asynchronously among reefs. Heterotrophic and mixotrophic dinoflagellates increased in abundance, correlating with a decrease of the diatom *Cerataulina pelagica* and an increase of the diatom *Rhizosolenia imbricata*. These variations occurred during and after the coral spawning event for some reefs in 2007. For the first time, a fresh-water cyanobacteria species of *Anabaena* was ephemerally found (only 3 days) in the archipelago, at reefs closest to human settlements. Variability among reefs in relation to spawning times indicated that reef-specific processes such as water residence time, re-mineralization rates, and benthic-pelagic coupling can be relevant to the observed patterns. These results suggest an important role of microheterotrophic grazers in re-mineralization of organic matter in coral reef waters and highlight the importance of assessing compositional changes of larger size fractions of the phytoplankton when evaluating primary productivity and nutrient fluxes.

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39 phytoplankton when evaluating primary productivity and nutrient fluxes.

40 **1. Introduction**

41 Phytoplankton drives the energy flow in most marine ecosystems as they are the main
42 primary producers in environments where sufficient light allows for photosynthetic
43 fixation of carbon (C) (Reynolds, 2006). In coral reefs, this productivity appears to be
44 limited by nitrogen (N), as reef waters usually have minimal macronutrient concentrations
45 (Charpy-Roubaud 1990; Furnas et al., 1990; Dizon & Yap, 1999; Koop et al., 2001; Heil et
46 al., 2004). Nutrients and organic compounds enter the typically oligotrophic waters of coral
47 reefs constantly through different pathways, from either natural or anthropogenic sources
48 (Wild et al., 2004; Wolanski et al., 2004; Mumby et al., 2007; Hoegh-Guldberg et al., 2007).
49 For instance, various recirculation processes in reefs drive fluxes of submarine
50 groundwater that might transfer in-land nitrogen into the reefs (Santos et al., 2010). Other
51 natural processes, such as mucus release by corals as well as massive or multi-specific coral
52 spawning events, also represent influxes of labile organic compounds, from which the
53 composition and amount of organic matter released into the water column varies greatly in
54 space and time (Coffroth 1990, 1991; Wild et al., 2004). Anthropogenic sources of nutrients
55 are often derived from land-based pollution, such as coastal development and agriculture
56 on watersheds, among other activities (Burke et al., 2011).

57 Regardless of the source, a surplus of organic or inorganic nutrients in coral reefs can alter
58 the ecosystem biogeochemistry and both the pelagic and benthic food webs (Koop et al.,
59 2001; Eyre et al., 2008). For example, dissolved inorganic nutrients seem to be rapidly
60 incorporated by the phytoplankton (Fabricius et al., 2013), but chronic inputs might lead to
61 higher oxygen demand, through phyto-, bacterio- and zooplankton overgrowth and
62 decomposition (Lapointe & Clark, 1992). Nutrient addition is a well-documented driver of
63 taxonomical and biomass changes in marine phytoplankton assemblages (Piehler et al.,
64 2004; Furnas et al., 2005; Howardth & Marino, 2006; Pirela-Ochoa et al., 2008; Strom,
65 2008; Mutshinda et al., 2013). Seasonal changes in phytoplankton composition in
66 oligotrophic ocean waters, upwelling sites and coastal waters in response to both organic
67 and inorganic nutrient inputs have been reported round the world (Casas et al., 1997; Bode
68 et al., 2001; Oguz et al., 2001; Vuorio et al., 2005; Wu et al., 2005; Harris, 2012).

69 In coral reefs, phytoplankton is dominated by small-size fractions ($<2 \mu\text{m}$) such as the
70 cyanobacteria *Synechococcus* and *Prochlorococcus* (Furnas & Mitchell 1989; Furnas et al.,
71 2005). This small-size fraction has been the focus of studies into the response of pelagic
72 primary producers to coral spawning events, as this response has been mainly evaluated
73 using biomass and pigment concentrations (Eyre et al., 2008; Glud et al., 2008; Wild et al.,
74 2008; Apprill & Rappé, 2011; Patten et al., 2011). However, species of larger sizes (i.e.
75 nano- and microphytoplankton: 2 to 200 μm), especially diatoms, also increase their
76 primary production with nutrient inputs in coral reefs (Furnas, 1991; Crosbie & Furnas,
77 2001). As a result of nutrient input and cell division, larger sizes achieve greater
78 abundances in shallow waters than in oceanic waters (Duyf et al., 2002); suggesting that
79 these microalgae might also play an important role in nutrient uptake and pelagic trophic
80 webs in coral reefs.

81 For example, a nutrient input after a typhoon that occurred in a reef in French Polynesia,
82 was followed by an increase in phytoplankton biomass and primary productivity, and by a
83 change in taxonomic composition of the microphytoplankton (Delesalle et al., 1993). Also,
84 Glud et al. (2008) reported a dinoflagellate bloom in a reef flat after a coral spawning in the
85 Great Barrier Reef; and Horne (2011) detected an increased abundance of the
86 dinoflagellate *Ceratium* spp. 2 days after a spawning event in the Gulf of Mexico. As coral

87 spawning events constitute an input of nutrients in coral reefs (Wild et al., 2004b; Eyre et
88 al., 2008; Glud et al., 2008; Wild et al., 2008; Patten et al., 2011), and growth of various
89 microphytoplankton groups is nitrogen limited (Hausse et al., 2012), it would be expected
90 that this input of nutrients affects the abundance and composition of larger size fraction.

91 Coral spawning events have been used as large natural experiments to understand the
92 effects of an episodic organic matter input on phytoplankton primary productivity and
93 biomass in coral reef systems (Wild et al., 2001, 2004b, 2008; Eyre et al., 2008; Glud et al.,
94 2008; Patten et al., 2011; Apprill & Rappé 2011). After coral spawning, Wild et al. (2004)
95 reported an increase in sediment oxygen demand and Glud et al. (2008) found an increased
96 pelagic and benthic primary production. Similarly, Eyre et al. (2008) showed a post-
97 spawning peak in phytoplankton biomass coinciding with the removal of dissolved
98 inorganic nutrients and changes in nitrogen cycling on the top layer of the sediment.
99 Nutrient additions through mass coral spawning events are thus expected to drive changes
100 in the nitrogen and phosphorous concentrations, and consequently have effects on
101 autotrophic and heterotrophic communities on reefs.

102 Reproductive timing of most common coral species in the Caribbean is well known
103 (Szmant, 1986; Bak & Engle, 1979; Fadlallah, 1983; deGraaf et al., 1999; Budd, 1990;
104 Bassim et al., 2002; Carlon, 2002, Brooke & Young, 2003; Beaver et al., 2004; Bastidas et al.,
105 2005; Severance & Karl, 2006); however, the effects of this event on the reef's pelagic
106 trophic web have not yet been documented in the region. To contribute to this knowledge,
107 herein we describe changes on the composition and abundance of the microphytoplankton
108 assemblages at various spatial and temporal scales at Los Roques, Venezuela during 2007
109 and 2008 coral spawning events. As Los Roques is an oceanic archipelago with a marginal
110 and local source of anthropogenic nutrients, expected coral spawning dates are a good
111 opportunity to assess the effects of a natural source of organic nutrients on the
112 composition and abundance of microphytoplankton separately from those of human
113 sources.

114 **2. Materials and Methods**

115 **2.1. Study Site**

116 Los Roques National Park is the most important reef complex in Venezuela, as well as in the
117 Southern Caribbean (Casanova et al., 2007). Los Roques is located 160 km north of the
118 coast (66.55-66.95 W 11.65-11.98 N; Fig. 1), and the archipelago encompasses more than
119 50 coralline cays that are protected from wave exposure by two barrier reefs: the eastern
120 barrier and the southern barrier, which are 20 km and 30 km long, respectively (Méndez-
121 Baamonde, 1978). Los Roques was the first marine area in Venezuela to be protected under
122 the category of National Park and is part of the southern corridor, one of the most
123 important areas in the Caribbean region in terms of biodiversity and extension (Rodríguez-
124 Ramírez, 2008). The human population at Los Roques (approximately 2,000) is
125 concentrated in the Northeast, specifically on Gran Roque. In the Southwest there is no
126 permanent settlement and it is relatively less affected by recreational and fishing activities;
127 thus, anthropogenic nutrient sources in the archipelago are marginal when compared with
128 coastal reefs. Studies on biological and physicochemical oceanography are scarce and
129 limited at Los Roques, however there have been studies describing temporal variability in
130 zooplankton and phytoplankton composition and abundance at one or two reefs (González,
131 1989; Spiniello, 1996, Madera & Furderer, 1997), and a seasonal variation has been found
132 in relation to the drought (November-July) and rain (May-October) regimes.

133 **2.2. Sample Collection**

134 To assess variations in microphytoplankton assemblages in relation to coral spawning at
135 Los Roques, a hierarchical design was employed considering the following factors: 1)
136 Locality (fixed, crossed with two levels: Northeast (NE) and Southwest (SW); 2) Reef
137 (random, nested in locality with two levels: Gran Roque (GR) and Madrizquí (MD) on the
138 NE and Dos Mosquises (DMS) and Cayo de Agua (CYA) on the SW); and 3) Period (fixed,
139 crossed with three levels: before, during and after the expected spawning dates). The
140 samples were taken from a motor boat by gently submerging a sterile 1,000 mL plastic
141 bottle in the water down to ~0.5 to 1 m depth until filled. Samples were taken by two
142 different teams at approximately the same hour (9:00-10:00 a.m.) in the NE and SW reefs,
143 while samples were collected less than one hour apart between reefs of the same locality.
144 The samples were then fixed with 10% formalin, stabilized with sodium tetraborate and
145 closed with a sealed cap. Samples were transported to the laboratory and stored in a cold
146 dark place until analysis. Four replicate samples were taken daily for 9 consecutive days

147 after the full moon of August 2007 and 2008 on the basis of the expected dates of
 148 spawning. Expected dates for the first coral species that start spawning in these multi-
 149 specific events (i.e., 2 to 4 days after the full moon of August and September, Szmant, 1986)
 150 marked the end of the “Before” and the start of the “During” sampling period. These dates
 151 are specified for each period and year in Table 1 and were chosen based on extensive
 152 observations of coral spawning dates in the Caribbean (e.g. Van Veghel, 1994, Szmant et al.,
 153 1997; Sánchez et al., 1999; Mendes & Woodly, 2002) and in our study site (Bastidas et al.,
 154 2005).

155 The presence of *Acropora palmata* larvae in the water during the sampling period of this
 156 study was further corroborated through specific antibody signals (Zubillaga, 2010). Briefly,
 157 the procedure consisted of three steps: (1) inoculation of an *Acropora palmata* larvae into
 158 rabbits; these larvae were harvested in the laboratory from bundles collected from *A.*
 159 *palmata* colonies in the field during a spawning event in 2006; (2) extraction of antibodies
 160 from rabbit’s blood samples and (3) immunological assays based on ELISA (Enzyme Linked
 161 Immunoabsorbent Assay) to test for specificity and accuracy of the antibodies (Zubillaga
 162 2010). The presence of larvae in water samples was detected using a spectrophotometer
 163 that measured the colorimetric product of the enzymatic reaction between the antibodies
 164 and the coral larvae (Zubillaga 2010). This method was preferred over direct counts due to
 165 the superior overall accuracy it has for identifying coral larval species (Carlson & Olson,
 166 1993; Coffroth & Mulawka, 1995; Garland et al., 2002).

167 **TABLE 1.** Sampling dates for the Before, During and After periods of coral spawning at Los
 168 Roques, according to the expected dates for the first coral species (*Acropora palmata* and *A.*
 169 *cervicornis*) that start spawning in these multi-specific events

2007		2008	
Period	Date	Period	Date
Before	August 28 th -30 th	Before	August 16 th -18 th
During	August 31 st -September 2 nd	During	August 19 th -21 st
After	September 3 rd -5 th	After	August 22 nd -24 th

170

171 2.3. Environmental variables

172 Surface Sea Water Temperature (SST) and chlorophyll *a* (Chl*a*) concentration (mg/m³)
 173 were obtained for the sampling dates by satellite image analysis (MODIS SCAR; Klein &

174 Castillo, 2010). For this, centroids of the image cells (1 Km x 1 Km spatial resolution) that
175 contained the sampling sites were used to download historical data. No neighbour pixels
176 were used to obtain values of both environmental variables. Due to the proximity between
177 MD and GR, these sampling sites in the northeast fell within the same pixel and thus, SST
178 and Chl_a had the same values for these reefs.

179 **2.4. Sample Processing and Data Analysis**

180 Microalgae from the water column were identified to the lowest possible taxonomic level
181 using a Leika D MIL inverted contrast microscope. Samples were analyzed after 48 h of
182 sedimentation in a 100 mL settling chamber. The settlement periods were used according
183 with the Utermöhl method for oceanic/oligotrophic samples referred to in Hasle (1978).
184 Phase contrast was used on the same sample to enhance cell detection. Magnification
185 power used for microalgae detection (all fields were viewed) was 200x; however, 400x and
186 1000x magnifications were used for identification. The abundance of microalgae species
187 was calculated using the same protocol, based on the volume of the sample taken.
188 Identification to the lowest possible taxonomic level was performed using Peragallo &
189 Peragallo (1897-1908), Cupp (1943), Saunders y Glenn (1969), Ferguson (1968), Marshall
190 (1986), La Barbera (1984, 1993), Sournia (1973, 1986), Sánchez-Suárez (1992, a,b), Tomas
191 (1997), Berárd-Therriault et al. (1999), Díaz-Ramos (2000) and Kravesky et al. (2009).
192 Microalgae species were grouped into diatoms, dinoflagellates, chlorophytes, cyanophytes
193 and coccolithophores and, based on previous work (Jeong, 1994; Jeong & Latz, 1994; Jeong
194 et al., 2004, 2005a, 2005b, 2005c, 2007, 2008, 2010, Du Yoo et al., 2009; Seong et al., 2010),
195 dinoflagellate species were further classified according to trophic functional groups into
196 obligate autotrophic, mixotrophic and heterotrophic species.

197 Shannon diversity index with natural log base were calculated for every sample to facilitate
198 comparisons among the spatial and temporal scales examined (Magurran, 2004).
199 Microphytoplankton community structure was analysed through the attributes richness
200 and abundance (based on the Bray-Curtis index), and composition (based on the Jaccard
201 index). Dissimilarity matrices for both indexes were constructed from the original
202 biological data matrices. Non-metric multidimensional ordination (nMDS) was performed
203 to aid the visualization of temporal patterns and spatial distributions of the samples in
204 terms of the microphytoplankton assemblage. When appropriate, centroids were used to

205 illustrate these patterns. Null hypotheses of no differences in the abundance and
206 composition, as well as the diversity of microphytoplankton, were tested using
207 permutational multivariate analysis of variance (PER-MANOVA, Anderson, 2001). When
208 significant differences ($P(\text{perm}) < 0.05$) were found for certain factors or interaction terms,
209 species contributing to at least 60% of the variability between levels of the terms were
210 identified using the SIMPER routine (Clark & Warwick, 2001). Univariate ANOVAs were
211 performed on the species or taxa selected by the SIMPER routine, and *a posteriori* pair-wise
212 comparisons were performed between levels of the terms with p-values under a 0.05 alpha
213 value. These analyses were performed using PRIMER-E v6 software (Clarke & Gorley,
214 2006)

215 3. Results

216 A total of 100 taxa of microalgae were identified from the water samples; 91 were
217 identified for 2007, 51 for 2008 and 42 were common to both years (Supplementary
218 Material Table 1). Taking both years into account, the most abundant group were the
219 diatoms (Bacillariophyceae), which represented 62% of the microalgae, followed by
220 dinoflagellates (Dinophyceae, 25.4%), and Cyanophytes (Cyanophyceae, 8%).
221 Coccolithophores (Haptophytes) and Chlorophytes represented less than 5% of the total
222 abundance. This two-year trend of group abundances hold true for all reefs in 2008, but in
223 2007, cyanophytes reached highest densities of 43 cells per mL at the NE reefs only (Fig. 2).
224 Most species of diatoms identified (62% of all counted diatoms) were small (6-10 and 16-
225 20 μm), oceanic taxa from the genera *Nitzschia*, *Pseudo-nitzschia*, *Paralia* and *Thalassiosira*.
226 Most taxa of dinoflagellates (48% of the counted cells) were 16-20 μm in size (Fig. 3). The
227 coccolithophores were represented only by the species *Emiliana huxleyi*.
228 Microphytoplankton abundance ranged between 0.26 ± 0.06 and 49.12 ± 8.37 cells per mL
229 (Mean \pm SD) across reefs, years and spawning times. Since the variability between years
230 was high (94% of the variance explained, PERMANOVA table not shown), the
231 microphytoplankton assemblages were analyzed separately for each year to evaluate the
232 effect of the other factors examined. For both years, dissimilarity in microphytoplankton
233 abundance and composition was observed between localities (NE or solid vs. SW or empty
234 symbols in Fig. 4), however, most of the variation in assemblage structure occurred
235 between reefs, particularly in 2008 (Table 2). When considering all taxa, the

236 microphytoplankton assemblage showed little variation in structure among time periods
 237 related to coral spawning (see low variation coefficients for the interaction
 238 Reef(Locality)xPeriod, Table 2). Contrasting with this weak response from the
 239 microphytoplankton structure, Shannon diversity indexes differed significantly among the
 240 spawning times in 2007 (Pseudo-F: 2.37, $p=0.042$) despite, variability between replicate
 241 samples being above 60% (Fig. 5). Taxa diversity differed between the spawning times in
 242 NE reefs. At MD, there were differences Before and During spawning times (*a posteriori*
 243 pairwise comparisons $t=2.16$, $p=0.04$) and at GR, between the During and After periods
 244 ($t=1.97$, $p=0.049$). In 2008, diversity was similar among spawning times but varied
 245 significantly among reefs as in 2007 (Pseudo-F: 10.18, $p=0.001$). In this case, diversity was
 246 lowest at CYA, a SW site, compared with the other reefs ($t=1.56$, $p=0.049$, respectively; Fig.
 247 5).

248 **TABLE 2.** PER-MANOVA based on the Bray–Curtis dissimilarities (no transformation) of
 249 the multivariate abundance of microalgae (102 taxa); on two reefs (random, nested) in the
 250 Northeast and two reefs in the Southwest (“Locality”, fixed) of Los Roques, during, before
 251 and after (“Period”, fixed) coral spawning events in 2007 (A) and 2008 (B). The same
 252 results were obtained for composition (Jaccard index based PERMANOVA) but are not
 253 shown.

254 (A)

Source of Variation	d.f.	MS	Pseudo-F	P(perm)	VC (%)
Locality	1	20104	3.0056	0.1664	4.88
Period	2	2530.7	0.98108	0.4854	0.00
Reef (Lo)	2	6690.3	3.3611	0.0001*	3.42
LoxPE	2	2393.5	0.9279	0.5254	0.00
Re[Lo]xPE	4	2579.7	1.296	0.07	1.28
Residuals	240	1990.5			90.43
Total	251				

255 (B)

Source of Variation	d.f.	MS	Pseudo-F	P(perm)	VC (%)
Locality	1	9933.6	1.3887	0.336	2.78
Period	2	501.9	0.79077	0.5737	0.00

Reef(Lo)	2	7153.3	12.299	0.0001*	13.14
LoxPE	2	501.9	0.79077	0.5724	0.00
Re[Lo]xPE	4	634.7	1.0912	0.3085	0.32
Residuals	276	581.62			83.76
Total	287				

256 Lo: Locality; PE: Period of spawning times; Re: Reefs; Df: Degree of freedom; MS: Mean
 257 Square; VC: Variation Coefficient; P: probability of obtaining a Pseudo-F value similar to the
 258 ones calculated when actually there is no difference between the levels of the factors
 259 evaluated; *: $P(\text{perm}) < 0.05$

260 Variations in microphytoplankton with respect to coral spawning times (Period) were
 261 more evident when analyzed by higher taxonomic groups instead of all taxa. The structure
 262 of diatoms, dinoflagellates and cyanophytes differed between reefs for both years, and
 263 some of these assemblages differed among spawning times at specific reefs (Table 3). In
 264 particular, the structure of dinoflagellates (Pseudo-F=3.93, $p=0.001$) and its trophic groups
 265 (Pseudo-F=8.12, $p=0.001$) differed significantly between spawning times (periods) for
 266 some reefs in 2007 (Fig. 6). Also in 2007, the assemblage of cyanophytes differed After the
 267 coral spawning at NE reefs (Pseudo-F=2.14, $p=0.032$), when the coccoid, filamentous and
 268 the nostocal *Anabaena* sp. peaked in an episodic manner. On the other hand, the
 269 oscillatorial cyanophytes *Lyngbya* and *Spyrogira* were identified only at Dos Mosquises (a
 270 SW reef) on the dates of expected coral spawning. While variations in dinoflagellates and
 271 cyanophytes assemblages occurred in relation to spawning times in 2007, diatoms
 272 assemblage only differed spatially among reefs for both 2007 and 2008 (Pseudo-F=2.048,
 273 $p=0.05$ and Pseudo-F=1.75, $p=0.003$, respectively).

274 Most of the dissimilarity in the assemblage of dinoflagellates among spawning times in
 275 2007 (>70% according to the SIMPER routine) was due to variations in the presence of the
 276 heterotrophic taxa *Protoperdinium* sp., *P. thorianum*, *P. minutum*, *P. excentricum*, *P.*
 277 *conicoides*, *P. pyriforme*, and the mixotrophic species *Neoceratium lineatum* and *Scrippsiella*
 278 *trochoidea* (Supplementary Materials Table 2). For each of these taxa, the univariate
 279 PERMANOVAs showed a significant difference among spawning time periods at MD and at
 280 DMS (a NE reef and a SW reef, respectively). At the SW reefs, the density of mixotrophic
 281 species decreased towards the After period of the spawning times, while the heterotrophic

282 species increased (Fig. 6). At the NE reefs, the abundance of heterotrophic species also
 283 increased After the spawning, whereas the mixotrophic species showed an opposite
 284 pattern to the SW reefs as it increased in abundance. This pattern within locality (NE and
 285 SW) was consistent for all reefs in 2007 (Fig. 6). Correlations between the abundance of
 286 trophic groups were negative and low ($\leq 30\%$), with the exception of mixotrophic and
 287 obligate autotrophs dinoflagellates (64.3%), although this was not statistically significant.
 288 In 2008, dinoflagellates as well as its trophic groups differed only at reefs scales (i.e. GR,
 289 MD, CYA, DMS), since 95% of their abundances occurred at the NE reefs where mixotrophic
 290 species peaked (Fig. 6).

291 **TABLE 3.** Statistically significant ($p < 0.05$) results from the PER-MANOVA analysis, based
 292 on the Bray–Curtis dissimilarities (no transformation) of the univariate abundance of
 293 microalgae taxonomic and trophic groups; on two reefs (random, nested) in the Northeast
 294 and two reef in the Southwest (“Locality”, fixed) of Los Roques, during, before and after
 295 (“Period”, fixed) coral spawning events in 2007 and 2008.

	Source of Variation	Diatoms	Dinoflagellates	DTG	Cyanophytes
2007	Locality (Lo)				
	Period (PE)		*	*	
	Reef[Lo]	*			
	LoxPe				
	Re[Lo]xPe		*	*	*
2008	Locality (Lo)				
	Period (PE)				
	Reef[Lo]	*	*		
	LoxPe				
	Re[Lo]xPe				

296 *: $P(\text{perm}) < 0.05$; DTG: Dinoflagellate trophic groups

297 For diatoms, more than 98% of the dissimilarity between the reefs in both years (SIMPER
 298 analysis) was due to variation in the presence and abundance of *Aptinoptychus* sp.,
 299 *Aulacoseria*, *Cerataulina pelagica*, *Thalassiosira subtilis*, *Rhizosolenia imbricata* and *Melosira*
 300 *varians*. In 2007, significant differences on the abundance of *C. pelagica*, *T. subtilis* and *R.*

301 *imbricata* occurred between reefs and between periods for *R. imbricata* and *C. pelagica* on
 302 SW reefs. *Cerataulina pelagica* was more abundant in the SW reefs, whereas *T. subtilis* was
 303 only identified on NE reefs. Significant correlations between the abundance of these
 304 diatoms and dinoflagellates trophic groups were found for SW reefs. Here, in 2007, *C.*
 305 *pelagica* abundance was negatively and weakly correlated ($r^2=-0.24$; $p=0.055$) with that of
 306 the mixotrophic dinoflagellate *Scropsiella trochoidea*, and positively correlated with that of
 307 heterotrophic dinoflagellates of the genus *Protoperidinium* ($r^2=0.77$; $p=0.0002$; Fig. 7). On
 308 the contrary, *R. imbricata* abundance decreased and was negatively correlated ($r^2=-0.53$;
 309 $p=0.00056$) with the abundance of the heterotrophic *Protoperidinium* species (Fig. 7). In NE
 310 reefs, none of the abundances of *Thalassiosira subtilis*, *Cerataulina pelagica* or *Rhizosolenia*
 311 *imbricata* were correlated to that of dinoflagellates trophic groups.

312 Chlorophyll *a* concentrations ranged between 0.35 and 1.03 mg/m³ in 2007 and between
 313 0.039 and 1.3 mg/m³ in 2008. These concentrations increased in days corresponding to the
 314 During and After spawning times, consistently for both years and for all reefs when data
 315 was available (Table 4a). Also, chlorophyll concentrations were more variable in 2008 than
 316 in 2007, and in the SW than in NE reefs for both years (Fig. 8). Sea surface temperatures
 317 ranged between 26.45 and 29.04 °C in 2007 and between 27.4 and 29.1 °C in 2008 (Table
 318 4b). In 2007, SST increased through the sampling period in all reefs but showing a larger
 319 variability in the SW reefs (Fig. 9). In contrast, SST values were very similar among reefs in
 320 2008, reaching higher temperatures During spawning time and decreasing by day 9 within
 321 the After spawning time (Fig. 9).

322 **TABLE 4.** A) Chlorophyll *a* concentration (mg/m³) and B) Seawater Surface Temperature
 323 (°C) for 9 consecutive days, “Before”, “During” and “After” the expected spawning dates for
 324 southern Caribbean acroporids. Data obtained from remote sensing (MODIS SCAR;
 325 1kmx1km resolution).

(A)

Periods	Days	2007			2008			
		NE (MD-GR)	CYA	DMS	Days	NE (MD-GR)	CYA	DMS
Before	28-Aug	NA	NA	NA	16-Aug	0.61	0.039	NA
	29-Aug	0.631	0.387	0.903	17-Aug	0.602	0.042	0.075
	30-Aug	0.67	0.35	NA	18-Aug	NA	0.0545	0.0544

During	31-Aug	0.73	0.42	NA	19-Aug	0.943	0.19	0.23
	01-Sep	NA	NA	NA	20-Aug	0.903	0.280	1.076
	02-Sep	1.03	0.58	NA	21-Aug	0.87	NA	NA
After	03-Sep	0.850	0.643	1.040	22-Aug	0.77	0.60	1.08
	04-Sep	NA	NA	NA	23-Aug	1.23	0.43	1.24
	05-Sep	0.651	0.431	0.826	24-Aug	1.30	NA	NA

(B)

Periods	Days	2007			2008			
		NE (MD-GR)	CYA	DMS	Days	NE (MD-GR)	CYA	DMS
Before	28-Aug	27.57	27.57	27.07	16-Aug	NA	NA	NA
	29-Aug	27.87	27.87	28.23	17-Aug	27.60	27.60	27.98
	30-Aug	27.72	27.72	28.09	18-Aug	27.41	27.41	27.54
During	31-Aug	27.65	27.65	26.45	19-Aug	28.91	28.91	29.15
	01-Sep	27.82	27.82	28.92	20-Aug	28.27	28.27	28.22
	02-Sep	27.94	27.94	29.04	21-Aug	29.01	29.01	28.67
After	03-Sep	28.40	28.40	29.03	22-Aug	28.13	28.13	27.96
	04-Sep	28.65	28.65	28.85	23-Aug	27.95	27.95	28.01
	05-Sep	28.90	28.90	28.93	24-Aug	NA	NA	NA

326

327 4. Discussion

328 Expected variations in microphytoplankton abundance and composition linked to the coral
 329 spawning events were mostly overridden by variations between years and spatial
 330 variations among reefs in Los Roques archipelago. However, when groups of the
 331 microphytoplankton were examined, dinoflagellate assemblages and trophic functional
 332 groups varied in relation to the spawning times. Heterotrophic dinoflagellates increased in
 333 abundance after the spawning event in 2007 for some reefs, while in 2008 this increment
 334 only occurred for mixotrophic dinoflagellates in one reef. Also, diatoms *Cerataulina*
 335 *pelagica* and *Rhizosolenia imbricata* varied simultaneously in abundance, suggesting that
 336 they may be responding to re-mineralization of dissolved nitrogen and grazing activity,
 337 respectively. These results suggest an important role of microheterotrophic grazers in re-
 338 mineralization of organic matter in coral reef waters. Also, our findings support the
 339 relevance of assessing compositional changes of larger size fractions of the phytoplankton
 340 when evaluating primary productivity and nutrient fluxes. A lack of direct measurements
 341 from the nutrient flux-cycling and benthic-pelagic coupling in our study, hampers an in-

342 depth discussion of the patterns observed; however, changes that occurred in a few days
343 on the composition and abundance of some taxa of the microphytoplanktonic groups
344 examined were likely related to macronutrient inputs that resulted from coral spawning, as
345 supported by concurrent increases in chlorophyll *a* concentrations obtained by satellite
346 images.

347 Contrasting results among studies support that phyto- and bacterioplankton and nutrient
348 flux responses to coral spawning are variable across coral reef regions. The subtle changes
349 in microphytoplankton at the lowest taxonomic level observed at Los Roques in relation to
350 coral spawning, contrasted with studies from other regions (Wild et al., 2004; Eyre et al.,
351 2008; Glud et al., 2008; Horne, 2011). In the Great Barrier Reef (GBR), Australia and the
352 Gulf of Mexico, both planktonic and benthic microalgae blooms had been observed within
353 2-5 days following a mass coral spawning event (Wild et al., 2004; Eyre et al., 2008; Glud et
354 al., 2008; Horne, 2011). At Los Roques, chlorophyll *a* concentrations increased During and
355 After the spawning times in all reefs, however, only the abundance and composition of
356 dinoflagellates and cyanophyte assemblages varied with spawning times. Furthermore,
357 changes in cyanophytes consisted of a freshwater taxon that spiked in abundance in 2007,
358 likely due to anthropogenic influence in the NE reefs rather than to a response from spawn
359 material. Also, increased chlorophyll *a* concentrations in relation to spawning in Los
360 Roques was lower than that reported for the GBR (Glud et al., 2008). Differences in the
361 magnitude (e.g. spawned material, number of species and geographic scale) of the
362 spawning events influence the amount of organic matter released to the water column, and
363 thus affect regional differences, which support the uniqueness of the mass coral spawning
364 event at the GBR compared to that occurring in other regions (Harrison & Booth, 2007;
365 Mangubhai & Harrison, 2008). This might explain why in Los Roques and other coral reefs
366 (Aprill & Rappé, 2011), phytoplankton blooms were not detected after massive coral
367 spawning events.

368 In addition to the intensity and magnitude of spawning events across regions, other factors
369 might influence the variability in the phytoplankton response. In this study, the sampling
370 days within spawning periods (Before, During and After) were established based on
371 expected spawning dates for *Acropora palmata*, as this species marks the beginning of the
372 multi-specific coral spawning event in Caribbean shallow waters. However, variations in

373 spawning intensity and time, as reported for this coral species (unpublished data by Miller
374 & Szmant in Key Largo, Florida and La Parguera, Puerto Rico in 2000, Japp between the
375 1970s and 1980s and Miller in 2001), are likely to affect the observed response of
376 microphytoplankton assemblages between years and among spawning periods. In
377 particular, *Acropora palmata* larval abundance suggested a late start in spawning for most
378 reefs in 2008 (i.e. +5 days instead of +2-3 days, supplementary material Table 3). The
379 abundance of *Acropora palmata* larvae in the plankton at the time of this study (Zubillaga,
380 2010) strongly suggested that coral spawning at Los Roques was larger in 2008 than in
381 2007, but in 2008 it lagged with respect to our sampling periods in most of the reefs
382 (supplementary material Table 3). This would have resulted in a During period being
383 similar to a Before period, both with little spawn material; explaining the differences in
384 composition and abundance of microphytoplankton observed between 2007 and 2008 and
385 the observed increase in abundance of diatoms and chlorophyll *a* concentration at one of
386 the SW reefs only in 2008. Days and reefs with higher abundance of coral larvae (Zubillaga,
387 2010) coincided with diatoms being more abundant, a group that requires higher inorganic
388 nutrient concentrations compared to cyanophytes and dinoflagellates (Reynolds, 2006). In
389 contrast, the higher abundance of heterotrophic dinoflagellates in 2007 at various reefs,
390 and of mixotrophic dinoflagellates at the GR reef site in 2008, might indicate a subtle shift
391 in these trophic groups, favouring heterotrophy.

392 A clear pattern of the role of microheterotrophy in the microphytoplankton response to
393 larvae abundance, as it has been reported for phytoplankton biomass and primary
394 productivity (Eyre et al., 2008; Glud et al., 2008), was apparently hidden by the high
395 variability observed among reefs. Previous work in which biomass, primary productivity
396 and/or respiration in sediment and water have peaked shortly after the release of spawned
397 material (Eyre et al., 2008; Glud et al., 2008) were carried out only at one site, on a reef flat.
398 Thus, an examination of these processes (i.e. phytoplankton primary production, biomass
399 and species composition in relation to nutrient re-mineralization) at larger spatial scales in
400 reefs might yield large variability as it was found in this study.

401 In addition to the amount of material spawned, other local (reef) scale-dependent
402 processes such as re-mineralization of organic matter, might be related to the variations in
403 *chl a* and microphytoplankton observed among reefs in nine days. The response of

404 photosynthetic microalgae to the input of labile organic compounds is mediated by the
405 metabolic activity of the microbial reef community, often called the microbial loop
406 (Anderson & Ducklow, 2001; Pomeroy et al., 2007; Nelson et al., 2013). This process entails
407 great variability as it may operate on time scales of hours to days (Carlson, 2002) and it
408 depends on many factors, from benthic composition to oceanographic characteristics of the
409 reefs (Nelson et al., 2011). Re-mineralization of coral spawn materials is mostly carried out
410 in reef sediments (Westneat & Resing, 1988; Haas et al., 2011, 2013), whereas the role
411 played by bacterioplankton in the process vary from immediate and significant (Wild et al.,
412 2008) to lagged and marginal (Apprill & Rappé, 2011). Thus, the temporal response of the
413 microphytoplankton community to an organic matter input might be controlled by how
414 fast the re-mineralized material is incorporated into the water column through benthic-
415 pelagic coupling (Eyre et al., 2008; Patten et al., 2008; Nelson et al., 2011; Apprill & Rappé,
416 2011). In this sense, during coral spawning events in the Gulf of Mexico, Horne (2011)
417 observed only small changes of nutrient concentrations in water, which suggests a low re-
418 mineralization activity in the water column, but he also reported an increased abundance
419 of dinoflagellates of the genus *Ceratium* spp. Thus, in our study, reef-specific differences in
420 the re-mineralization activity could have contributed towards the variability found at the
421 reef scale in the structure of diatoms assemblages.

422 In this study, functional group and species-specific patterns of microphytoplankton related
423 to the spawning period were observed at some reefs. During and After the coral spawning
424 in 2007, the increased abundance in *Protoperdinium* heterotrophic species, and the
425 concomitant decrease of the mixotrophic *Neoceratium lineatum*, may constitute a response
426 to the coral spawning via abundance of prey. The abundance of heterotrophic
427 dinoflagellates found in this study was very similar to that found by Horne (2011) in the
428 Gulf of Mexico 2 days after the coral spawning. However, at some reefs in Los Roques, the
429 abundance of mixotrophic dinoflagellates was almost double that reported by Horne
430 (2011). Some heterotrophic dinoflagellate species prey preferentially upon heterotrophic
431 bacteria and protozoans and the eggs and larvae of metazoans (Jeong, 1994, 2007, et al.,
432 2010), whereas mixotrophic species more frequently prey upon small-size microalgae like
433 haptophytes, crysophytes, picophytes, raphidophytes (not assessed on this study),
434 chlorophytes, autotrophic dinoflagellates, some diatom species and autotrophic bacteria (Li

435 et al., 2000; Jeong et al., 2005; Berge et al., 2008). The negative correlation between the
436 abundance of heterotrophic dinoflagellates and one of their grazed species, the diatom
437 *Rhizosolenia imbricata* (Willén, 1991), might indicate that preferential feeding occurred in
438 detriment of autotrophic microalgae species. Consistent with this, heterotrophic and
439 mixotrophic dinoflagellates are recognized to have important effects on plankton
440 abundance and composition as microheterotrophic grazers (Lessard & Swift, 1985). Thus,
441 despite increased *Chla* concentrations, grazing activity by heterotrophic dinoflagellates
442 might explain the lack of a significant increase in diatom abundance in relation to spawning
443 periods at some reefs in 2007.

444 In addition to grazing, microheterotrophic phytoplankton (i.e. mixo- and heterotrophic
445 dinoflagellates among other protozoans) maintain nutrient demand through re-
446 mineralization of organic matter in oceanic waters year-round, and their contribution to
447 dissolved inorganic nutrients might be higher than that of zooplankton (Bode et al., 2005).
448 Oligotrophic coral reefs have an efficient recycling of nutrients (Szmant, 2002; Wild et al.,
449 2008) and are able to incorporate a massive coral-spawning organic matter input in less
450 than 6 days through the benthic microbial loop. Because sediment re-mineralized nitrogen
451 is not readily returned to the water column (Eyre et al., 2008), microheterotrophic re-
452 mineralization within the microphytoplankton community might be mediating the
453 response of autotrophic groups to the input of that organic matter. Although the role of
454 these grazers in the re-mineralization of spawned materials in coral reefs has not been
455 evaluated, the positive correlation between the diatom *Cerataulina pelagica* - whose
456 abundance has been related to increase on dissolved nitrate and ammonium availability in
457 the water column (Härnström et al., 2009) - and the heterotrophic dinoflagellates of the
458 genus *Protoperidinium*, might suggest an important role of the latter on the availability of
459 dissolved inorganic nutrients at the SW reefs. A similar increase in dinoflagellate
460 abundance coinciding with a decrease in that of the diatoms was found 2 days after coral
461 spawning in the Gulf of Mexico (Horne, 2011).

462 At Los Roques, high abundance of small rather than medium and large size diatoms,
463 especially in the relatively pristine SW reefs, might indicate an effective nutrient cycling as
464 cell size in microalgae has an influence on nutrient preference and uptake (Koiko et al.,
465 1985; Stolte et al., 1994), with smaller phytoplankters favoured over larger ones in systems

466 driven by regenerated nutrients and rapid cycling of organic matter (Caroppo, 2000).
467 Similarly, the peak abundance in diatoms coinciding with that of mixotrophic
468 dinoflagellates in 2008 at GR might also respond to re-mineralization of spawned materials
469 by the latter. However, the consistency of this pattern was not captured in our 2008
470 sampling probably due to lagged spawn in the other reefs. Also, sampling days for each
471 time period (i.e. Before-During-After) were relatively short (3 days each), and these
472 spawning periods were contiguous. If the response of the larger size phototrophs of the
473 phytoplankton to spawned organic matter is mediated by microheterotrophy, as suggested
474 by our results, more days between sampling periods could have been better for detecting
475 changes in the microphytoplankton assemblages. .

476 Observed microphytoplankton density, although well within previous reports for Dos
477 Mosquises in Los Roques and other oligotrophic waters (e.g. González, 1989; Spiniello,
478 1996, Madera & Furderer, 1997), revealed spatiotemporal differences of the
479 microphytoplankton community in Los Roques at scales previously unexplored.
480 Differences in plankton assemblages known to occur between the NE and SW sectors of the
481 archipelago during the dry season from November to June (González, 1989; Casanova et al.,
482 2007) also hold true for August and September during this study, a period with a weak
483 hydrodynamic influence in the area (Casanova et al., 2007). Our results also indicated a
484 strong reef-based variability within previously thought homogenous sectors (Casanova et
485 al., 2007) and a daily variability within otherwise considered constant seasons (González,
486 1989, Spiniello, 1996; Madera & Furderer, 1997). The presence of the cyanophyte
487 *Anabaena* sp. in August and September on the NE reefs in 2007 constitutes the first report
488 of this species in Los Roques, indicating episodic freshwater inputs, consistent with human
489 settlement at Gran Roque. Algae from this genus can produce toxins of public health
490 concern (Figuereido et al., 2004); however, their disappearance in samples from
491 consecutive days suggests unfavourable water conditions for these populations and/or
492 short water residence time. Toxin-producing dinoflagellate species were also identified, but
493 in low abundance.

494 While our study lacked of direct biomass measures through time, studies elsewhere have
495 not targeted compositional changes on microalgae assemblages. Therefore, a full picture of
496 changes in phytoplankton across regions and reefs in relation to coral spawning is still

497 lacking. Our results highlighted the importance of including taxonomical and functional
498 changes in larger size phytoplankton fractions when evaluating nutrient fluxes and primary
499 and secondary pelagic productivity in coral reefs in relation to coral spawning events, as
500 well as the importance of replication at reef spatial scales for generalizing observed
501 patterns. Manipulative experiments to establish the effect of microheterotrophic grazing
502 and nutrient re-mineralization activity by dinoflagellates on the response of the
503 phytoplankton assemblages to organic matter inputs constitute a logical next step to better
504 comprehend the role of dinoflagellates in reef waters during a nutrient input.
505 Concomitantly, specific processes related to inter-annual variability need to be assessed
506 through the sampling of non-spawning times as well in order to better discriminate
507 phytoplankton responses to coral spawning events. Similarly, large-scale operating
508 oceanographic and/or climatic processes not yet assessed on the study site may also have
509 an important role as structuring factors, as suggested by the amount of inter-annual
510 variability observed in the microphytoplankton structure and composition and the daily
511 patterns of SST and Chla concentrations.

512 This study evaluated the structure of larger size fractions of the phytoplankton during coral
513 spawning for the first time in the Caribbean, showing variations in dinoflagellate
514 assemblages and its trophic groups, as well as in some diatom species. These variations
515 highlighted the potential impact of heterotrophic dinoflagellates on coral reefs primary
516 productivity and organic matter cycling. Thus, by examining the structure of
517 microphytoplankton at multiple spatiotemporal scales in Los Roques archipelago, this
518 study provides a first approximation of the effect of coral spawning on these assemblages.

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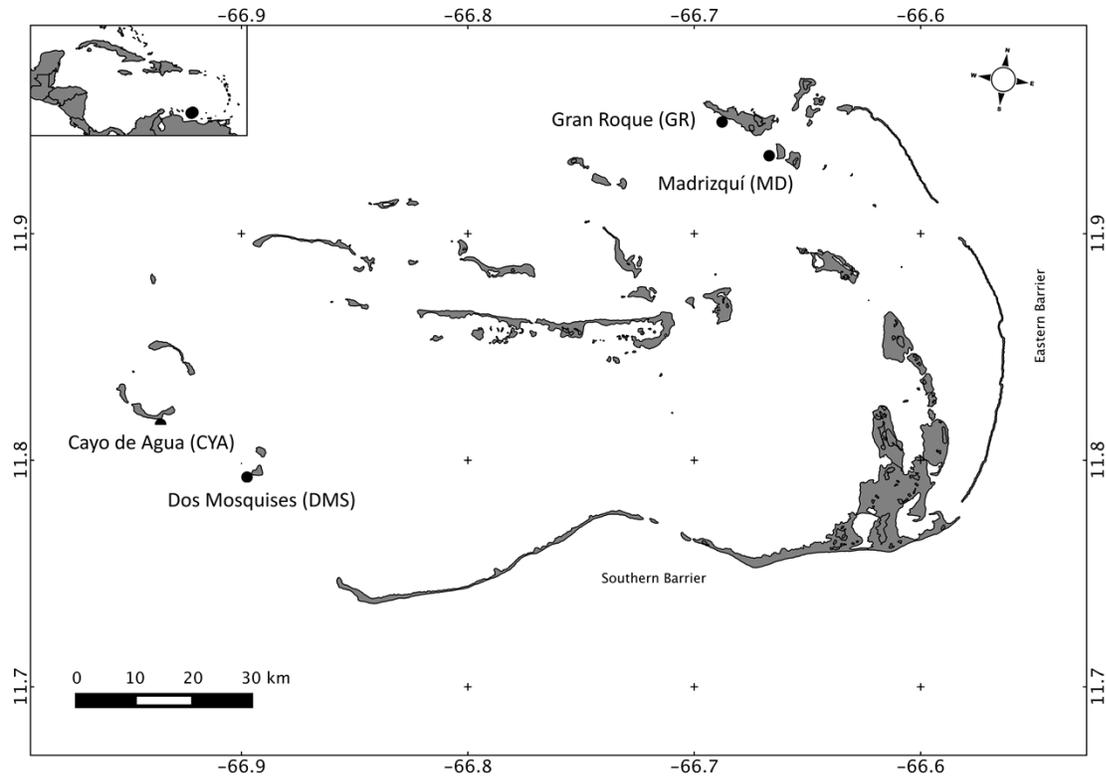
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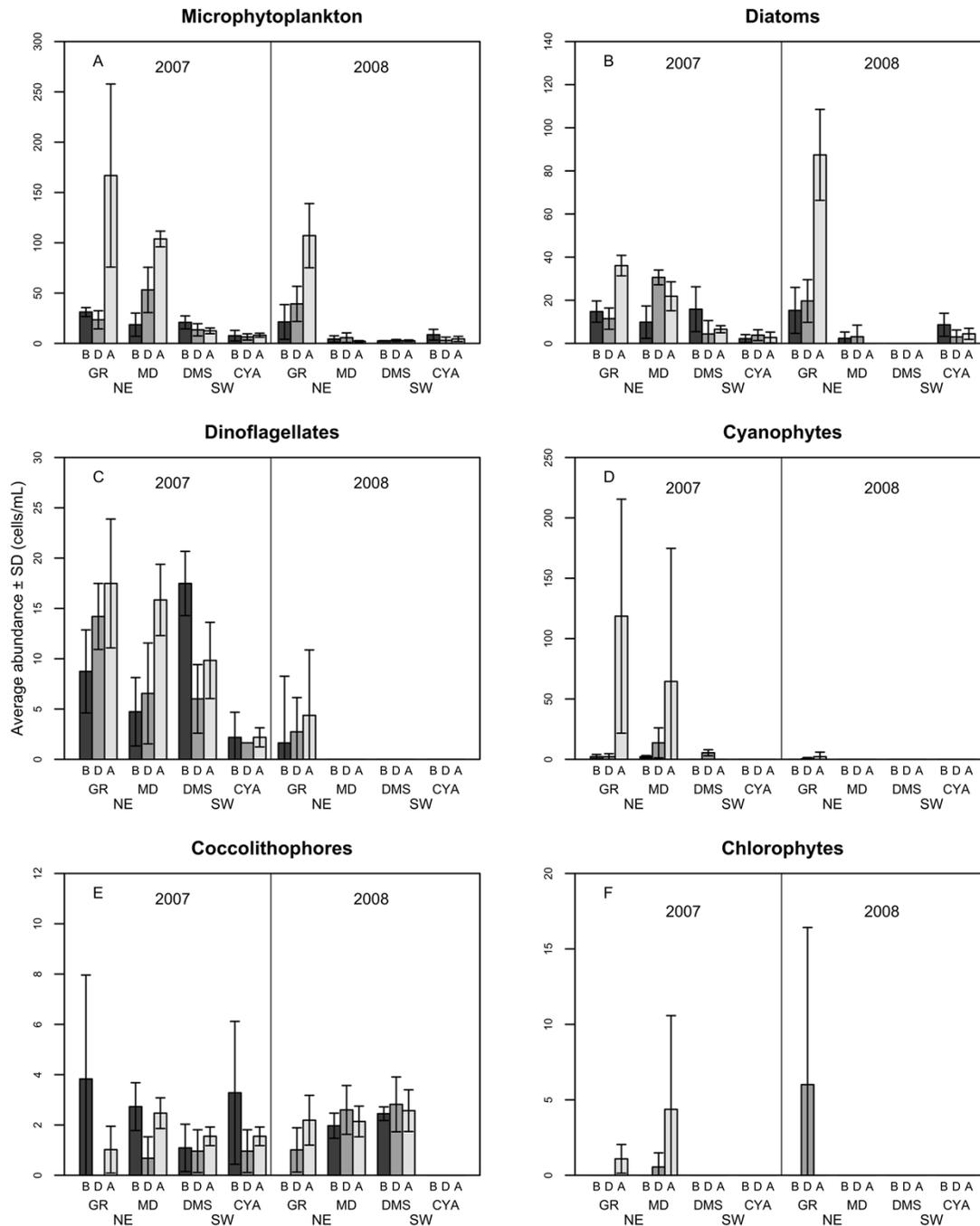
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883 1. Figures



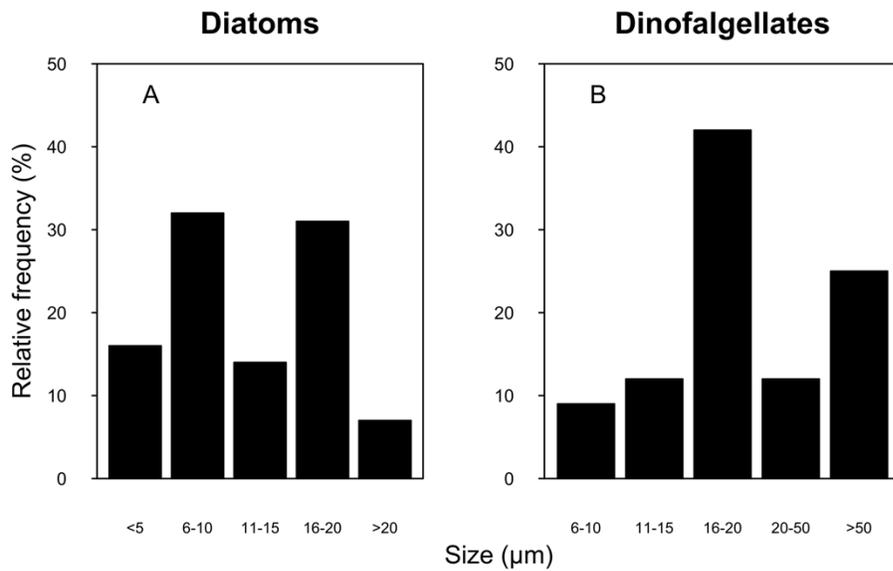
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885 **Figure 1.** Map of Los Roques National Park, Venezuela, Southern Caribbean, showing the four
886 reefs sampled at the Northeast (NE) and Southwest (SW) sectors.



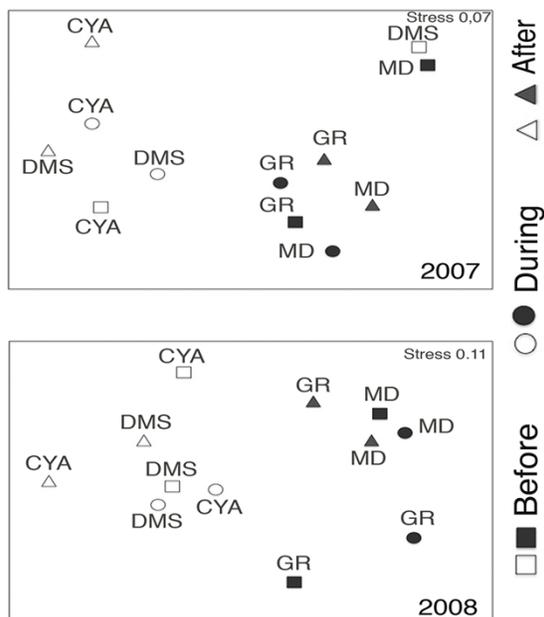
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888 **Figure 2.** Average density of microphytoplankton (cells per mL) of Microphytoplankton (A),
 889 Diatoms (B), Dinoflagellates (C) Cyanophytes (D), Coccolithophores (E) and Chlorophytes (F)
 890 in four reefs (GR, MD, DMS, CYA) located at the Northeast (NE) and Southwest (SW) localities
 891 of Los Roques. B, D, A refers to Before, During and After the coral spawning events of 2007 and
 892 2008. Density values are displayed with different scales for each of the taxonomic groups.



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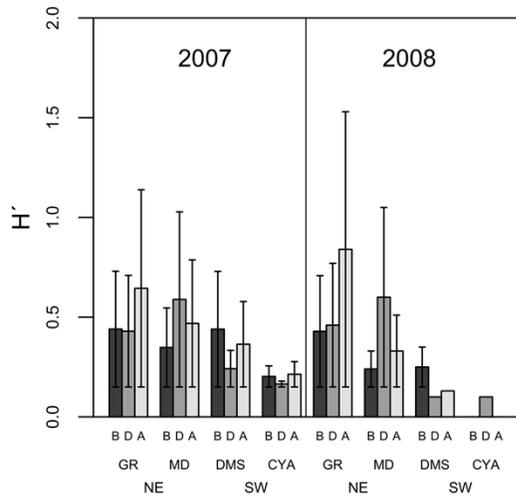
894 **Figure 3.** Relative frequency of the Diatoms (A) and Dinoflagellate (B) groups size classes
 895 sampled in four reefs (GR, MD, DMS, CYA) located at the Northeast (NE) and Southwest (SW)
 896 localities of Los Roques during 9 consecutive days in August and September 2007 and 2008.



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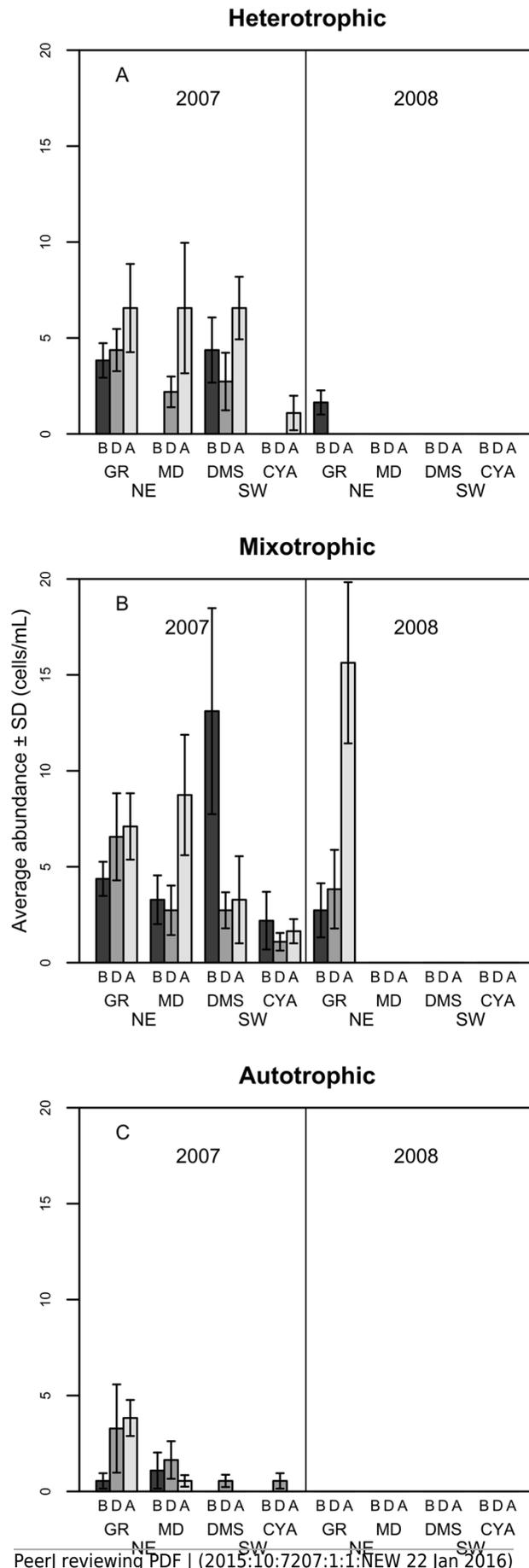
898 **Figure 4.** Nonmetric Multidimensional Ordination (nMDS) based on centroids for the
 899 microphytoplankton assemblages in four reefs (GR, MD, DMS, CYA), located in the Northeast
 900 (empty symbols) and Southwest (filled symbols) of Los Roques.

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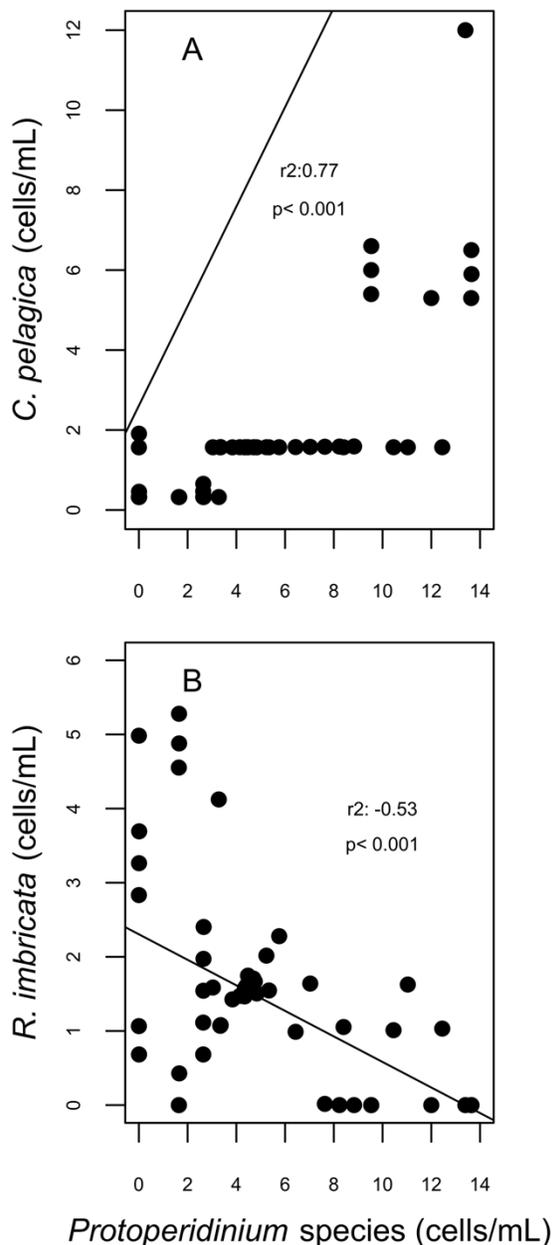


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903 **Figure 5.** Average Shannon Diversity index values ($H' \pm SD$) of microphytoplankton in four
 904 reefs (GR, MD, DMS, CYA) located at the Northeast (NE) and Southwest (SW) localities of Los
 905 Roques. B, D, A refer to Before, During and After the coral spawning events of 2007 and 2008.



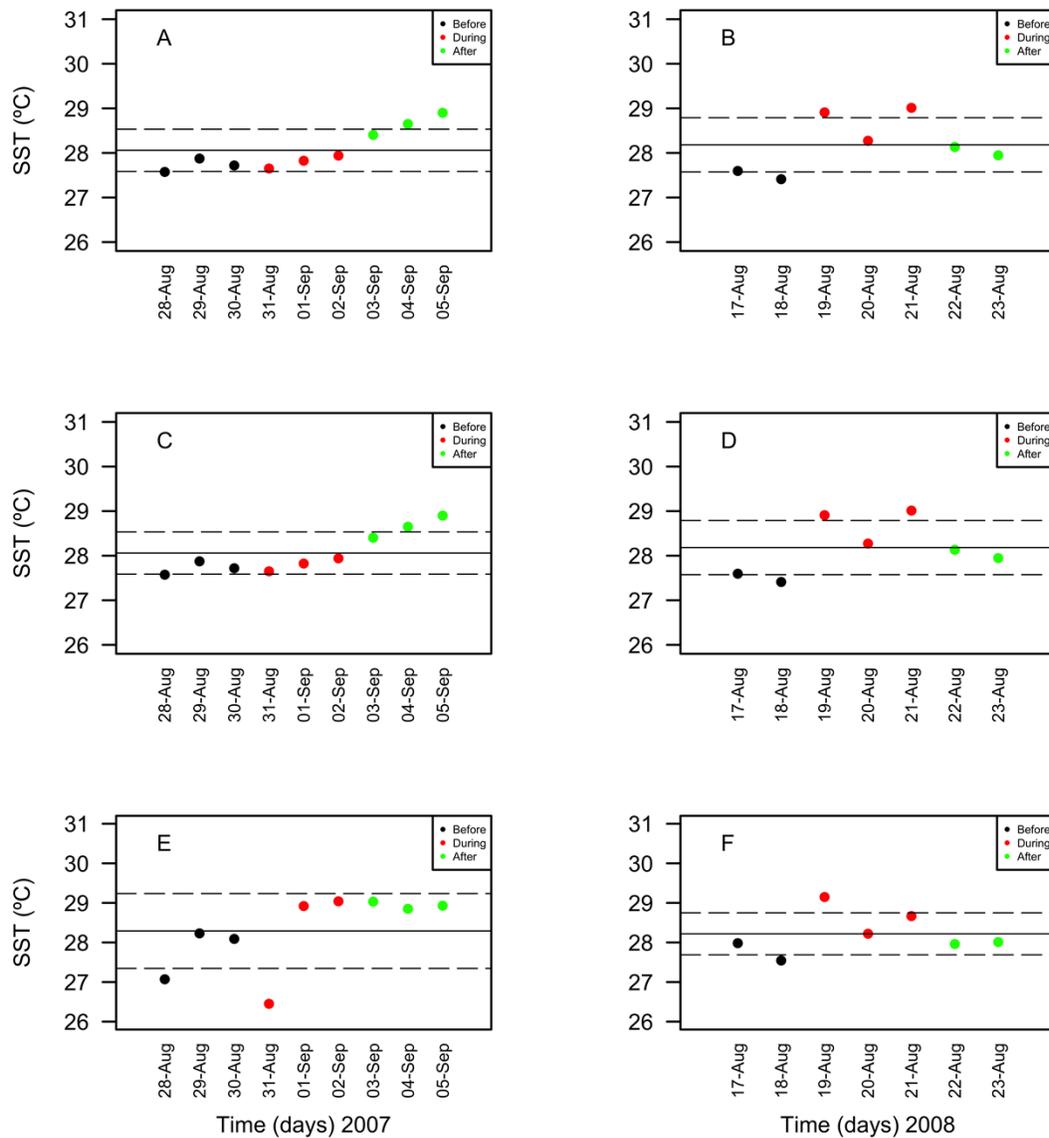
907 **Figure 6.** Density of Heterotrophic (A), Mixotrophic (B) and Autotrophic (C) dinoflagellates
908 (cells per millilitre, Mean \pm SD) in four reefs (GR, MD, DMS, CYA) located at the Northeast
909 (NE) and Southwest (SW) localities of Los Roques. B, D, A refers to Before, During and After
910 the coral spawning event of 2007.



911

912 **Figure 7.** Correlation plots on abundances of *Cerataulina pelagica* (A) and *Rhizosolenia*
913 *imbricata* (B) and the abundance of *Protoperidinium* species; corresponding to the spawning

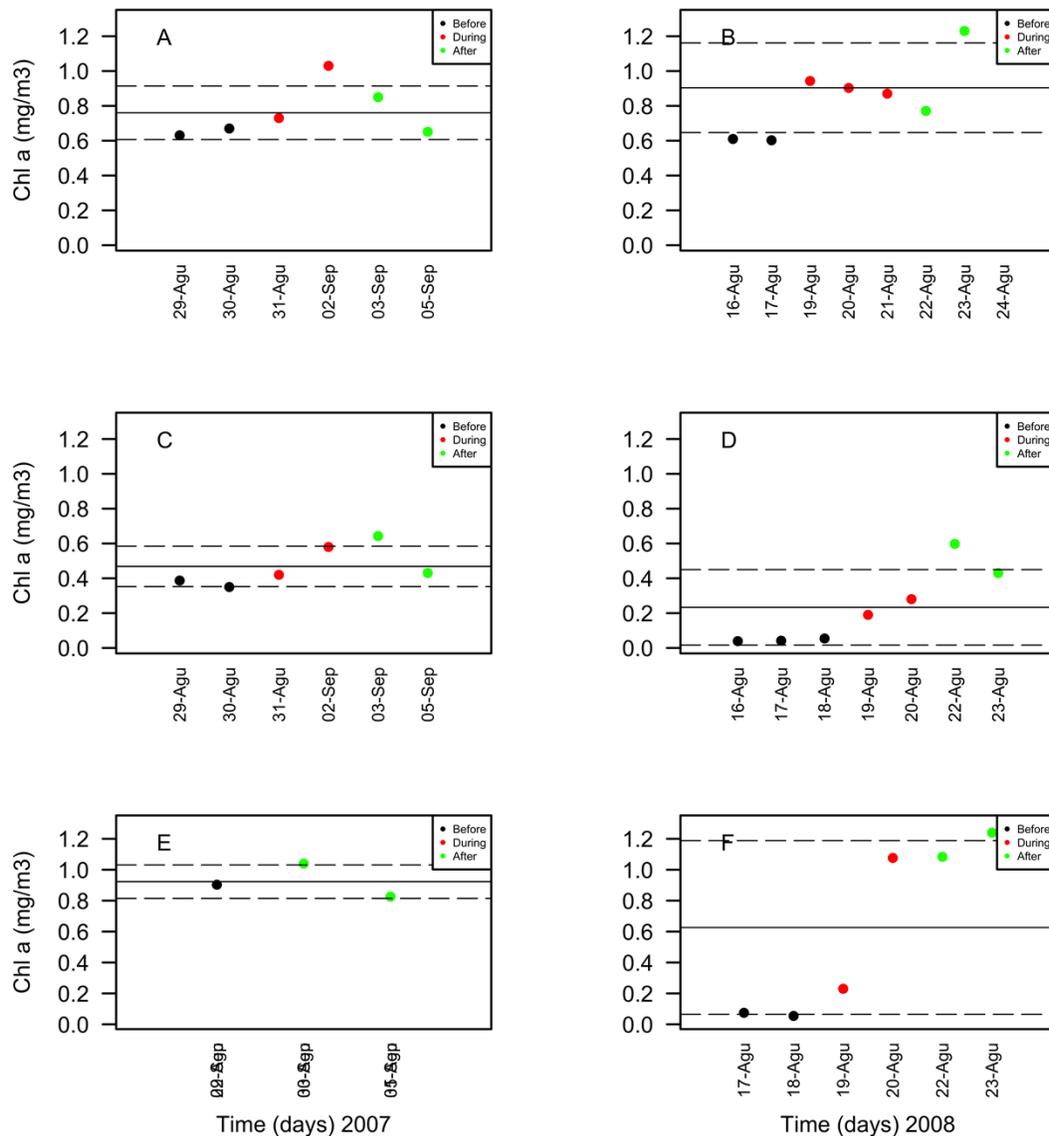
914 event of 2007 in two reefs (DMS, CYA) located at the Southwest (SW) sector of Los Roques.
 915 Only statistically significant ($p < 0.05$) correlations, with Spearman r^2 values are shown.
 916 Abundance values in cell per millilitres are displayed with different scales for each of the
 917 correlation plots.



918

919 **Figure 8.** Surface seawater temperature (SST in °C) values obtained through remote sensing
 920 (MODIS SCAR, 1km x 1Km resolution) for the nine-days sampling period corresponding to
 921 Before (black), During (red) and After (green) expected coral spawning times for the study site,

922 at the Northeast (A-B) and Southwest (SW) Cayo de Agua (C-D) and Dos Mosquises (E-F) reefs
 923 of Los Roques in 2007 and 2008.



924

925 **Figure 9.** Chlorophyll *a* (Chl *a* in mg/m³) values obtained through remote sensing (MODIS
 926 SCAR, 1km x 1Km resolution) for the nine-days sampling period corresponding to Before
 927 (black), During (red) and After (green) expected coral spawning times for the study site, at the
 928 Northeast (A-B) and Southwest (SW) Cayo de Agua (C-D) and Dos Mosquises (E-F) reefs of
 929 Los Roques in 2007 and 2008.