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Mitigating NaCl stress in *Vigna radiata* L. cultivars through *Bacillus pseudomycoides*

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Salt stress is one of the significant abiotic stresses that exert harmful effects on plant growth and production. In this study, different concentrations of NaCl were applied to five cultivars of mung bean (*Vigna radiata* L.), along with the inoculation of a salt-tolerant bacterial strain to assess the growth and yield. The bacterial strain was isolated from the saline soil of district Sahiwal, Punjab, Pakistan and identified as *Bacillus pseudomycoides*. Plant growth was monitored at 15-days interval and finally harvested after 120 days at seed set. Both sodium and potassium uptake in above- and below-ground parts was assessed using a flame photometer. Fresh and dry mass, the number of nodules, pods, and seeds per plant, as well as the weight of seeds per plant and the weight of 100 seeds, were significantly reduced with an increase in concentration of NaCl from 3 to 15 dSm⁻¹. Plants exposed to NaCl stress without bacterial inoculum exhibited significant reduction in growth and yield, as compared to the plants with bacterial inoculum as these plants showed a significant increase in studied parameters. An inverse relationship was found between growth parameters and salt concentration. It was found that the cultivar Inqelab mung showed the least reduction in growth and yield traits among studied cultivars, while Ramzan mung showed the maximum reduction. Among all the cultivars, maximum Na⁺ uptake was found in roots while the least in seeds. The study concludes that NaCl stress significantly reduces the growth and yield among mung bean cultivars, but inoculation of *Bacillus pseudomycoides* can alleviate the salt stress. Such findings will be helpful to cultivate the selected cultivars in the soils with varying concentrations of NaCl.

Mitigating NaCl stress in *Vigna radiata* L. cultivars through *Bacillus pseudomycoides*

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11 **Key Words:** Biomass, cultivars, growth, mung bean, salt-tolerant bacteria, stress, yield

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20 **Abstract**

21 Salt stress is one of the major abiotic stresses that harm plant growth and production. In this study,
22 different concentrations of NaCl were applied to five cultivars of *Vigna radiata* L. (mung bean),
23 along with the inoculation of a salt-tolerant bacterial strain to assess the growth and yield. The
24 bacterial isolation was done from the saline soil of district Sahiwal, Punjab, Pakistan and
25 recognized as *Bacillus pseudomycoides*. Plant growth was monitored at 15-days interval and
26 finally harvested after 120 days at seed set. Both sodium and potassium uptake in above- and
27 below-ground parts was assessed using a flame photometer. There were observed significant
28 reduction in fresh and dry mass, the number of root nodules, pods, and seeds per plant as well as
29 total seeds weight and 100 seeds weight, as the concentration of NaCl increased from 3-15 dSm⁻¹.
30 Plants exposed to NaCl stress without bacterial inoculum exhibited significant reduction in growth
31 and yield, as compared to the plants with bacterial inoculum as these plants showed a significant
32 increase in studied parameters. An inverse relationship was found between growth parameters and
33 salt concentration. It was found that the cultivar Inqelab mung showed the least reduction in growth
34 and yield traits among studied cultivars, while Ramzan mung showed the maximum reduction.
35 Among all the cultivars, maximum Na⁺ uptake was found in roots while the least in seeds. The
36 study concludes that NaCl stress significantly reduces the growth and yield among mung bean
37 cultivars, but inoculation of *Bacillus pseudomycoides* can alleviate the salt stress. Such findings
38 will be helpful to cultivate the selected cultivars in the soils with varying concentrations of NaCl.

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42 **Introduction**

43 Salinity imposes enormous pressure on agricultural production due to salt deposition in soil
44 (Shahrasbi *et al.*, 2020). Salt stress has badly affected about 20% of cultivated land and 50% of
45 cropland worldwide (Naveed *et al.*, 2020). The salt-affected area expansion is being accelerated
46 by anthropogenic activities as well as climate change (Arora & Bhatla, 2017). About 50% of
47 agricultural productivity is reduced through salinity in arid and semiarid regions (Kamran *et al.*,
48 2019a). In Pakistan, 6.7 million hectares of land is being degraded annually through salinity,
49 destroying 40,000 hectares of land in Pakistan (Hassan *et al.*, 2021). Salt eventually affects some
50 soils as a result of improper salt ~~removal~~^{management} from the affected site (Saleem *et al.*, 2020). Soluble salts
51 are retained in the soil during agricultural practices due to anthropogenic activities such as
52 improper fertilizer use and disposal, which causes excessive ~~salinification~~^{salinification}, accelerating soil salinity
53 (Zafar *et al.*, 2019). Soil salinization is caused by high evaporation rates, low rainfall, and irrigation
54 with salt-containing water, which results in salinity (Okur & Orcen, 2020).

55 Salinity causes numerous biochemical and physiological damages in crops, such as imbalanced
56 ionic effect, reactive oxygen species production and lower soil water potential (Rehman *et al.*,
57 2019). Higher concentration of reactive oxygen species disrupts enzymatic processes, cell
58 membranes, as well as the degradation of lipids, proteins, and other biochemicals (Shahid *et al.*,
59 2020). The higher concentration of Na^+ displays a higher Na^+/Ca^+ and Na^+/K^+ ratio, which
60 consequently results in less absorption of K^+ and Ca^+ that disrupts cellular membranes and
61 enzymatic activities, imbalances cellular equilibrium and destroys cellular balance (Kamran *et al.*,
62 2019b). Excess amounts of salt cause oxidative damage by accumulating increased amounts of
63 reactive oxygen species that can cause breakage of nucleic acids, damage plant tissues, and
64 shrinkage of cell and chromatin condensation, which ultimately harms cellular structure (Debnath

65 *et al.*, 2021). An increased amount of salt in the soil profile also results in physiological dryness
66 through less absorption of water, ion toxicity, normal cellular metabolic processes and reduced
67 osmotic potential (Safdar *et al.*, 2019; Garcia-Caparros *et al.*, 2021). Crops show different
68 responses to salt stress at developmental stages (Alsafran *et al.*, 2022). The availability of water
69 and nutrient uptake disturbed ~~sd~~ and ceased ~~es~~ the germination of seeds as well as the growth of plants
70 due to ionic and osmotic influences (Nawaz *et al.*, 2021).

71 Bio-inoculation ~~has been~~ proved highly significant to enhance crop production against different salt stresses
72 (Hyder *et al.*, 2023). Plant growth-promoting bacteria can be used to reduce the harmful effects of
73 abiotic stresses (Mun~~az~~ *et al.*, 2021). Soil is composed of many living and non-living things, such
74 as fungi, bacteria, plants, insects, and minerals, and these create a sense of interaction with each
75 other (Chen *et al.*, 2023). In rhizosphere, soil bacteria help in disease prevention as well as the
76 smooth uptake of nutrients (Kong *et al.*, 2021). The induction of bio-formulation with plant
77 growth-promoting bacteria aids in crop long-term fertility and makes modern approaches more
78 appealing than traditional approaches (Backer *et al.*, 2018).

79 The rhizosphere is the area of soil near the roots where soil microbes are abundant and perform
80 microbial activities (Grover *et al.*, 2021). Rhizo-microbial activity depends on soil properties,
81 climate changes, and host species (Lyu *et al.*, 2020). Bacterial inoculation enhances plant growth
82 through direct and indirect actions (Saleemi *et al.*, 2017). Direct mechanisms involve the easy
83 uptake of nutrients from natural resources or phytohormone secretions that improve plant growth
84 (Basu *et al.*, 2021). Indirect mechanisms include enhancing stress tolerance as well as suppressing
85 plant pathogens (Fatima & Arora, 2021). The diversity of soil and microbes is the backbone of the
86 agroecosystem and increases plant production at a higher rate even under stress (Fatima *et al.*,
87 2020).

88 Beneficial microorganisms dwelling in the soil imposed positive influence on plant metabolism,
89 allowing plants to survive under abiotic stresses like salt, drought, heat, and heavy metal (Luna *et*
90 *al.*, 2007). Various bacterial species such as *Arthobacter*, *Azospirillum*, *Bacillus*, *Enterobacter*,
91 *Klebsiella*, *Pseudomonas*, and *Serratia* secreted different phytohormones mainly indole acetic acid
92 and cytokinin that increased plant biomass, length, and nutrient uptake efficiency (Rosenburg,
93 Gutnik & Rosenburg, 2006). Cytokinins also transferred signal from roots to shoots to manage
94 environmental stresses (Ansari, Ahmad & Pichtel, 2019). Different bacterial strains, such as
95 *Acinetobacter* sp., *B. massilioanrexius*, *B. pseudomycoides* and *B. thuringiensis* have different
96 defense mechanisms against ~~stres~~. *Bacillus pseudomycoides* is a mesophilic, spore forming,
97 facultative anaerobic bacterium that forms rhizoid colonies (Paul *et al.*, 2022). It produced
98 different volatile organic compounds that significantly increased plants biomass, resistance against
99 diseases as well as abiotic stresses, regulated ion uptake and endogenous homeostasis of
100 phytohormones (Farag, Zhang & Ryu, 2013).

101 Increasing salt stress in the soil is the factor that limits the production of crops (Xu *et al.*, 2022).
102 The wide range of plant growth-regulating bacteria interacts with the host and helps in the
103 reduction of harmful ramifications of salinity, boosting soil health (Kumawat, Nagpal & Sharma,
104 2022). The salt-tolerant bacteria indulge in various growth-promoting traits to endure high salt
105 conditions in saline areas (Numan *et al.*, 2018). The salt-tolerant bacteria develop various
106 strategies to overcome the harmful effect of salt such as developing an ion export system to deport
107 extra sodium ions and accumulate chloride and potassium ions to prevent salt stress (Prittesh *et*
108 *al.*, 2020). Some salt-tolerant bacterial strains substitute glucose with lactoprotein, and anionic
109 phospholipase in the membrane to induce a more negative charge for stability and accumulate

110 solutes like amino acid and sugar to sustain the osmotic pressure of cells (Hanelt & Muller, 2013;
111 Zhang *et al.*, 2018; Gupta *et al.*, 2021).

112 *Vigna radiata*, commonly known as mung bean, is among the beneficent diet, having ecological
113 and economic values, it's a summer crop, mostly cultivated in arid and semi-arid regions
114 (Shafique *et al.*, 2023). The main dietary components are protein, carbohydrates, fiber, fatty acids,
115 and amino acids (Aasim *et al.*, 2019). Due to nitrogen fixation ability, mung beans can be used for
116 reclamation of soil and enhance soil fertility (Nigar *et al.*, 2023). The annual production of this
117 crop is 30 million tons worldwide, which is 5.1% of the total pulses produced (Sehrawat *et al.*,
118 2018). Mung bean major cultivation lands are Africa, Burma, China, India, Pakistan, and
119 Queensland (Pandey *et al.*, 2021). Mung bean is considered a salt-sensitive crop and, therefore,
120 susceptible to salt stress (Kumar *et al.*, 2023). About 70% mung bean production was reduced at
121 50 mM NaCl concentration which imposed harmful effects on the crop (Dutta & Bera, 2014). The
122 inoculation of salt-tolerant bacteria in mung beans persuades repulsion against sodium uptake,
123 promotes biomass and length of an above and below-ground plant part, and enhances nodulation
124 with nitrogen fixation and rate of photosynthesis with transpiration (Saha, Chattergi & Biswas,
125 2010). So far, the impact of salt-tolerant bacteria on these five varieties has not been studied. The
126 objectives of this current study were 1) screening of five cultivars of *Vigna radiata* for NaCl stress
127 2) utilization of salt tolerant bacteria (*Bacillus pseudomycoides*) to alleviate NaCl stress 3)
128 analyzing the effect of *B. pseudomycoides* inoculation along with salt treatments on growth, yield,
129 and physiological parameters 4) accessing the uptake of Na⁺ and K⁺ in root, stem, leaves and seeds
130 among the five selected cultivars.

131 **Materials and Methods**

132 **Bacterial isolation, screening, and characterization**

133 Three different saline areas in Jahan Khan, district Sahiwal, Punjab, Pakistan, were chosen for soil
134 sampling with geographical location 30° 34' 0" N, 73° 11' 0" E. Salt-tolerant bacteria were isolated
135 from salt-affected soil (Manasi, Rajesh & Rajesh, 2016). Screening of salt-tolerant bacteria was
136 carried out following Zhou *et al.* (2014). Three salt-tolerant bacterial colonies were screened
137 against various concentrations of NaCl ranging from 100 ppm to 30000 ppm salt to determine
138 which concentrations these bacteria can easily survive. Salt-tolerant bacteria samples were
139 prepared and sent to Humanizing Genomics Macrogen in Korea for molecular characterization and
140 identification following Sanjay *et al.* (2018).

141 **Bacterial Inoculum Preparation**

142 Inoculum of bacteria was prepared by adding 8 g of nutrient broth (NB) into one liter of distilled
143 water and bacterial strain loop was inoculated in it. For preparation of slants for inoculum
144 application, it was placed in an incubator at 37 °C for 24 hours.

145 **Seeds collection**

146 Seeds of five different cultivars i.e, NIFA-19, NIFA-17, Ramzan mung, Inqelab mung and Sona
147 mung were collected from the Nuclear Institute for Food and Agriculture Peshawar and the
148 Agriculture Research Institute Dera-Ismael Khan, Pakistan.

149 **Experimental set-up**

150 The pot experiment was conducted to measure plant growth, yield, and physiological parameters
151 for selected cultivars of *V. radiata* as Inqelab mung (V1), NIFA-19 (V2), NIFA-17 (V3), Sona
152 mung (V4) and Ramzan mung (V5). Two set-up were established along the four treatments, one
153 having salt stress without bacterial inoculum ($T_0, T_1, T_2, T_3, T_4, T_5$) and the other having salt stress
154 with bacterial inoculum ($T_0B, T_1B, T_2B, T_3B, T_4B, T_5B$) for all cultivars of mung bean. The seeds
155 of cultivars were germinated in the Botanic Garden and five days old seedlings were treated with

156 NaCl concentration ranging from 3 dSm⁻¹ to 15 dSm⁻¹ and compared with control lacking any
157 application of NaCl. The bacterial inoculum was applied in the pots having germinated seeds; four
158 pots were used for each treatment, with two replicate plants in each pot (Gamalero & Glick, 2022).

159 Hayyat *et al.* (2021) reported the physico-chemical properties of Botanic Garden soil. **Inoculum**
160 **of bacteria was prepared by adding 8 g of nutrient broth (NB) into one liter of distilled water and**
161 **bacterial strain loop was inoculated in it. For preparation of slants for inoculum application, it was**
162 **placed in an incubator at 37 °C for 24 hours.** Prepared broth (10 mL) mixed with distilled water
163 and poured into label pots. The experiment was conducted for 120 days (March to June 2023) in
164 ambient conditions at the Botanic Garden of Government College University Lahore Pakistan.
165 During the experiment the mean day-time temperature was 44°C, relative humidity 26 % and solar
166 radiation 802 watts/m². The plants were monitored on weekly basis for their growth and irrigated
167 appropriately to avoid any water stress; equal amount of water was applied to all pots.

168 **Mid and Final Harvesting**

169 During mid-harvest, fresh and dry weight of plant parts (above and below ground) was measured.
170 At final harvest, weight (fresh and dry) of plant parts along with other yield attributes viz. number
171 of pods, total seeds number, total seeds weight per plant and weight of 100 seeds were assessed
172 (Desai *et al.*, 2022).

173 **Physiological measurements**

174 The physiological parameters i.e. maximum photosynthetic rate, stomatal conductance, and
175 transpiration rate were measured in the morning from 10:30 am to 11:30 am under ambient
176 conditions before mid-harvest by using IRGA LCA4. The chlorophyll contents were determined
177 using a spectrophotometer (Spectroscan 80D) (Yoshida, Forno & Cock, 1971). The intrinsic water
178 use efficiency of different mung bean cultivars was calculated using the following (Ferreira *et al.*,
179 2018).

180 **Na⁺ and K⁺ uptake**

181 In order to assess the Na⁺ and K⁺ uptake, the ash of leaves, stem, seeds, and roots of studied
182 cultivars was prepared by heating the samples at 550 °C for three hours in muffle furnace
183 (Gallenhammp Size 2). 5 mL of 2N HCl was used to dissolve prepared ash and placed for fifteen
184 minutes followed by the addition of 50 mL distilled water. The solution was filtered by using
185 Whatman No. 42 filter paper to have an extract of particular plant part. The sodium and potassium
186 uptake was assessed by using flame photometer (S20 Spectro lab).

187 **Data Analysis**

188 Statistical analyzes of data was done with Statistix v. 8.1 using a one-way ANOVA (Duncan's
189 multiple range test (DMRT)) involving a comparison of the means across treatments. The
190 descriptive statistics, correlations between parameters and graphics were carried out using Sigma
191 Plot 14.0 version.

192 **Results**

193 **Bacterial screening, identification, and characterization**

194 Three isolated bacterial samples were selected for further screening. After few screenings, only
195 one bacterial strain was chosen for inoculation due to higher tolerance against NaCl. The pure
196 isolate was gram-positive, non-motile, long rod-shaped, and spore-forming under light
197 microscope. Identified as *Bacillus pseudomycoides*, Figure 1 shows the homology of isolated
198 bacterial strain.

199 **Effect of NaCl on *V. radiata* cultivars Biomass**

200 Table 1 represents that salt treatment showed a significant difference in growth and yield among
201 all cultivars while the *Bacillus pseudomycoides* inoculum coped with drastic effect of NaCl.
202 During mid-harvest, plant fresh weight showed a significant increase (63.8% - 93.8%) with

203 *Bacillus pseudomycoides* inoculum while a significant decrease (57.5% - 13.6%) was recorded
204 among all the cultivars having salt treatment only. Similarly, plant dry weight with *Bacillus*
205 *pseudomycoides* inoculum showed a significant increase ranging from (72.3% -84.8%) while
206 decrease (38.6%-16.4%) due to NaCl stress (Figure S1, Supplementary Material).

207 At final harvest, a significant increase (71.8%-94.3%) in fresh weight of plant with *Bacillus*
208 *pseudomycoides* inoculum was observed across all the five cultivars while a significant decrease
209 (57.3% - 14.5%) with salt stress (Figure S2, Supplementary Material). The plant dry weight also
210 showed significant increase (67.3%-92.2%) in plants with *Bacillus pseudomycoides* inoculum
211 while decrease from (45.9% - 14.7%) due to salt stress among five selected cultivars (Figure 2).
212 All five studied cultivars of *V. radiata*, Inqelab mung, NIFA-19 mung, NIFA-17 mung, and Sona
213 mung and Ramzan mung showed significant differences in dry weight (above and below ground)
214 (Figure 2 a,b,c,d,e). The measured differences in the fresh and dry weight of cultivars were;
215 V5<V4<V3<V2<V1.

216 **Effect of NaCl on Yield**

217 The yield attributes such as number of nodules, number of pods per plant, number of seeds per pod
218 per plant, weight of seeds per pod per plant, and weight of 100 seeds also showed reduction in
219 plants without inoculum of bacteria from T₀-T₅ in all cultivars. The inoculum of *Bacillus*
220 *pseudomycoides* provided more yield than plants with salt stress only. There were significant
221 differences observed in yield parameters among the *Vigna radiata* cultivars, Inqelab mung (Figure
222 3 a,f,k,p,u) NIFA-19 mung (Figure 3 b,g,l,q,v), NIFA-17 mung (Figure 3 c,h,m,r,w), Sona mung
223 (Figure 3 d,i,n,s,x), and Ramzan mung (Figure 3 e,j,o,t,y). The percentage increase was observed
224 among all cultivars in number of root nodules (74.3% - 87.5%), pods (65.9% - 92.8%), seeds
225 (72.8% - 94.5%), weight of seeds (64.7% - 85.9%) and weight of 100 seeds (74.9% - 94.8%) in

226 plants with *Bacillus pseudomycoides* inoculum while percentage decrease in number of root
227 nodules (32.8% - 12.7%), pods (27.9% - 9.6%), seeds (33.7% - 15.7%), weight of seeds (45.8% -
228 15.6%) and weight of 100 seeds (25.7% - 7.9%) due to NaCl stress.

229 **Effect of NaCl on Gaseous exchange and Na⁺, K⁺ Uptake**

230 The significant differences were found among the studied cultivars regarding their maximum rate
231 of photosynthesis, transpiration, stomatal conductance and water use efficiency (Figure 4 a-t).
232 Plant with inoculum of *Bacillus pseudomycoides* showed percentage increase of photosynthetic
233 rate (75.3%-83.8%), transpiration rate (53.85%-87.1%) and stomatal conductance (67.7%-81.8%)
234 while percentage decrease of photosynthetic rate (25.8%-9.4%), transpiration rate (36.2%-17.3%)
235 and stomatal conductance (47.4%-13.5%) due to salt stress among all cultivars. Water use
236 efficiency also showed increase from 68.3% - 92.1% only with salt stress, the decrease was
237 observed from 28.3% - 13.8% with inoculum of *Bacillus pseudomycoides* in all cultivars. The up-
238 take of Na⁺ and K⁺ varied in both the above and below ground parts of all varieties. The Na⁺
239 concentration was higher in roots than stems and leaves, while the minimum amount of Na⁺ was
240 detected in seeds. The amount of Na⁺ was ranging from 0-2.77 g kg⁻¹ in roots (Figure 5 a-e),
241 0.40-2.83 g kg⁻¹ in stems (Figure 5 f-j), 0.13-2.26 g kg⁻¹ in leaves (Figure 5 k-o) and 0.09-1.90 g
242 kg⁻¹ in seeds (Figure 5 p-t). Similarly, the concentration of K⁺ ranged from 6.73-11.56 g kg⁻¹ in
243 roots (Figure 6 a-e), 7.25-10.83 g kg⁻¹ in stems (Figure 6 f-j), 7.20-10.40 g kg⁻¹ in the leaves (Figure
244 6 k-o) and 7.03-8.76 g kg⁻¹ in the seeds (Figure 6 p-t), across the five cultivars and all treatments
245 with and without *Bacillus pseudomycoides* inoculum.

246 Significant difference in the Na⁺/K⁺ ratio was also found in the studied cultivars (Figure S3,
247 Supplementary Material). The increase in ratio was found in order 63.2% - 91.2% in roots, 72.7%
248 - 88.3% in stem, 73.8% - 87.6% in leaves, 69.3% - 91.6% in seeds with *Bacillus pseudomycoides*

249 while decrease as 38.4% - 16.7% in roots, 43.8% - 17.7% in stem, 34.8% - 12.7% in leaves, 28.4%
250 - 11.7% in seeds with NaCl stress among all the cultivars.

251 **Effect of NaCl on Chlorophyll contents**

252 Figure 7 (k-o) shows the total chlorophyll contents across the studied cultivars, chlorophyll a
253 ranged from $0.84 - 2.53 \text{ mg g}^{-1}$ (Figure 7 a-e), chlorophyll b $0.39 - 2.72 \text{ mg g}^{-1}$ (Figure 7 f-j) and
254 total chlorophyll contents $1.23 - 3.95 \text{ mg g}^{-1}$ (Figure 7 k-o). There were observed significant
255 increase in chlorophyll a (68.4% - 87.5%) and chlorophyll b (65.8% - 84.7%) contents with
256 *Bacillus pseudomycoides* inoculum while decrease in chlorophyll a (31.8% - 13.7%) and
257 chlorophyll b (27.6% - 9.8%) contents due to salt stress. There were also found significant increase
258 in total chlorophyll contents 73.8% - 92.8% with *Bacillus pseudomycoides* inoculum while
259 decrease 28.4% - 11.8% with NaCl stress among all cultivars. Chlorophyll a contents were higher
260 than chlorophyll b contents under all treatments in all cultivars.

261 **Relationship among the variables**

262 A significantly inverse relationship between fresh ($R^2=0.97-0.98$) and dry ($R^2 = 0.99-0.80$) and
263 plant biomass against salt concentration was observed across all the five cultivars. The inverse
264 relationship was also observed among the number of root nodules ($R^2= 0.98-0.99$), pods per plant
265 ($R^2= 0.97-0.96$), seeds per plant ($R^2= 0.98-0.86$) and weight of seeds ($R^2= 0.98-0.96$) against salt
266 concentration (Figures 8).

267 **Discussion**

268 In this study, five cultivars of *V.radiata* were screened for their NaCl stress tolerance and the salt
269 tolerant bacterial strain *Bacillus pseudomycoides* was used to test whether it can mitigate the
270 applied stress. The selected five cultivars showed significant differences regarding their tolerance
271 to NaCl stress. The application of isolated bacterial strain i.e, *Bacillus pseudomycoides* along the

272 salt stress resulted in enhances ^d growth and yield. This salt stress screening of cultivars and the
273 alleviation through *Bacillus pseudomycoides* provided the novel information to cultivate *V. radiata*
274 cultivars in salt affected areas. Anjana, Rawat & Goswami (2023) identified *Bacillus*
275 *pseudomycoides* as halophilic bacteria by 16S rRNA sequencing. Studies also showed that gram-
276 positive bacteria belonging to genus *Bacillus* were largely found in saline area (Javed *et al.*, 2023).
277 Among the five studied cultivars, Inqelab mung showed more resistance to salt application than
278 other cultivars. This could be due to the more nitrogen fixation ability and more uptakes of
279 minerals, as it had larger number of root nodules and root hairs, balanced phytohormones, and
280 production of reactive oxygen species as well as osmotic adjustment of this cultivar (Mansoor *et*
281 *al.*, 2017). Rehman *et al.* (2019) also reported the interaction of rhizobium strains with mung bean
282 cultivars “Inqelab mung” that showed bacterial strains elevated yield and physiological parameters
283 by increasing nutrient absorption, systematic resistance against pathogens, phosphorus
284 solubilization as well as production of phytohormones. Khattak *et al.* (2020) compared yield of
285 NIFA-17 mung and Ramzan mung in breeding experiment. NIFA-17 was declared as commercial
286 crop from 2017 in Khyber Pakhtunkhawa because it provided more yield in terms of seed
287 production, seed size and number of pods than Ramzan mung. Khattak, Saeed & Ibrar, (2019) also
288 compared NIFA-19 and Ramzan mung. NIFA-19 was recommended as new commercial variety
289 in 2018 because it gave more yields in term of seed size, number and production than Ramzan
290 mung. Ramzan mung was also evaluated as hybridized cultivars with short height, stiff stem,
291 earlier maturity, and moderate grain while no resistance against diseases detected (Khattak, Ashraf
292 & Alam, 2006).
293 Our findings showed that salt concentrations, *Bacillus pseudomycoides* inoculum and selected
294 cultivars had a significant impact to increase biomass and yield of different mung bean cultivars,

295 indicating that all these three factors could significantly ^{??} the yield among the mung means. In our
296 study, salt stress decreased biomass and yield of mung bean cultivars while inoculum of *Bacillus*
297 *pseudomycoides* improved it which was in line with previous study by Malook *et al.* (2020). A
298 substantial imbalance in osmotic and water relation is caused by salinity stress which declined
299 growth. Salinity stress can be alleviated by using plant growth promoting bacteria was also found
300 by Negacz *et al.* (2022). Inoculation of plant growth-promoting bacteria *Acinetobacter bereziniae*,
301 *Enterobacter ludwigii* and *Alcaligenes faecalis* ameliorated the harmful effect of saline stress in
302 *Pisum sativum*. It also enhanced biomass and nutrient uptake efficiency. Our findings support
303 Sare, Gontia & Tiwari, (2022) showing that biomass and yield of mung bean cultivars decreased
304 with salt stress because Na^+ ion level increased which induced oxidative stress, harmed ionic
305 balance, membrane integrity and photosynthetic efficiency. The inoculation of *B. pseudomycoides*
306 improved biomass and yield significantly reported by Katsenios *et al.* (2022) who treated sweet
307 corn with different types of plant growth promoting bacteria that increased growth and yield in
308 order: *B. mojavensis*> *B. subtilis*> *B. pumilus*> *B. pseudomycoides* compared to control. Li *et al.*
309 (2022) also characterized ability of *Bacillus pseudomycoides* as heavy metal (Cu) resistant
310 bacteria. The salt stress negatively affected root structure; Yousef *et al.* (2023) also found that
311 NaCl stress imposed bad effect on French beans morphological attributes, causing enzymatic
312 degradation, metabolism, physiological rate alteration, and an imbalance in osmotic potential
313 which destroyed root structure. *Bacillus* sp. also worked symbiotically to improve *V.radiata*
314 growth and yield that stabilized metabolism and osmotic balance (Ahmed *et al.*, 2020).
315 In our findings, root nodules, play significant role in nitrogen fixation, also suffered from salt stress
316 as found by Dogra *et al.* (2013) that number of root nodules decreased in chickpeas under salt
317 stress and its affect was ameliorated by *Mesorhizobium cicero* which improved root nodules and

318 nitrogen fixation ability. Hnini, Taha & Aurag (2022) also found rhizobacteria in *Vachellia tortilis*
319 and co-inoculation of *E. hormaechei*, *P. moraviensis* bacteria in *Sulla flexuosa* L. (Hamane *et al.*,
320 2023) developed more root nodules enhancing root colonizing potential and counteracting nutrient
321 imbalance due to salt stress.

322 Our results are also supported by Medeiros & Bettoli (2021) who investigated *Bacillus* sp. on
323 tomatoes which gave more yield than plants under only stress. It might happened due to increased
324 phytohormones, more ROS scavenging ability as well uptake of minerals and reduced osmotic
325 stress. *Bacillus pseudomycoides* can activate different enzymes such as lipase, pectinase and
326 cellulase that play a major role to minimize the stress and uplifted yield Knezevic *et al.* (2021).
327 Under salt stress, chlorophyllase break down different photosynthetic pigments, chlorophyll and
328 other enzymes which produced more reactive oxygen species. The chlorophyll contents, rate of
329 photosynthesis, transpiration and stomatal conductance negatively affected by salt stress. Our
330 findings were similar to AbdelMotlb *et al.* (2023) that stress reduced chlorophyll contents and
331 physiological attributes in green gram. The bacterial inoculum enhanced these parameters by
332 increasing electron transport and photosynthetic activity while lowering xylem balancing pressure
333 and stomatal conductance. More positive results were observed in Inqelab mung, and decreasing
334 influence was shown by Ramzan mung. *B. pseudomycoides* also affected water use efficiency of
335 mung bean cultivars against salt stress as reported Obadi *et al.* (2023) in tomato, it might happened
336 due to osmotic stress, imbalanced ion uptake as well as cellular damages, affect the efficiency of
337 cellular process, including water use. *Rhizobium leguminosarum*, a plant growth promoting
338 bacteria showed resistance against drought stress and improved chlorophyll contents and
339 physiological attributes and yield of plants.

340 The ion uptake results showed that there was comparatively more sodium and less potassium
341 uptake by roots than stems, leaves, and seeds in plant with only salt stress while inoculum of
342 *Bacillus pseudomycoides* improved potassium uptake. Inqelab mung showed less uptake of sodium
343 among all varieties, whereas Ramzan mung presented more uptakes and less resistance against salt
344 stress. Salt stress inflexed more sodium ion in plant cell that initiated osmotic stress, damaged
345 plant tissue, disrupts cellular processes and membrane integrity, Leontidou *et al.* (2020) also found
346 similar result in tomato under salt stress. Higher levels of salt, compartment more Na^+ ions in plant
347 cells that drained plant energy for combating stress rather it could be used for growth. In the
348 presence of excess NaCl salt, the root plasma membrane depolarizes as Na^+ influx which activate
349 guard cell outward rectifying potassium channels. As a result, Na^+ ion diffuses into cell that
350 decrease cytosolic K^+ and increase Na^+ content. Soil-inhabitant bacteria mitigated the uptake of
351 sodium ions by forming an ion export system and prevent cell damage. Inoculation of *Rhizobium*
352 sp. and *Pseudomonas* sp. activated ion exclusion mechanisms that removed extra Na^+ from roots
353 part that prevented seeds and leaf from uptake of salt demonstrated by Ahmed *et al.* (2011). In
354 current study, *Bacillus pseudomycoides* played major role to overcome salt stress and enhanced
355 mung growth and yield also found (ElSharawy, Eid & Ebrahem, 2023) *B. pseudomycoides* a
356 promising bio-control agent against *Alternaria* early blight in tomato.

357 Our results showed *Bacillus psedomycooides* inoculum lowered Na^+/K^+ ratio that boosted plant
358 nodulation, yield and physiological attributes against applied stress. Salt stress elevated Na^+/K^+
359 ratio which disrupts ion homeostasis due to accumulation of Na^+ . Similar report presented by
360 Zhang *et al.* (2018) & Jiang *et al.* (2019) that salt stress in rice, cucumber and *Cicer arietinum*
361 increased Na^+/K^+ ratio by inducing oxidative stress, reducing antioxidant system and disrupt
362 cellular functions while inoculation of *Rhizobium* sp. proved significant against salt stress. These

363 results are in agreement with previous findings of Muahtaq *et al.* (2021). It was supposed that
364 bacterial inoculum supported different mechanisms as intra-cellular Na^+ sequestration, cytosolic
365 potassium retention, osmotic adjustment through synthesis of compatible solutes that enhanced
366 crop growth and yield even under stress.

367 Conclusion

368 The results of the study showed that the combined effect of NaCl stress and inoculum of *Bacillus*
369 *pseudomycoides* significantly improve the growth, yield, and physiological attributes of five
370 different cultivars of *Vigna radiata* L. The Na^+ from NaCl applications accumulated in root
371 epidermal cells, impeding plant water uptake and causing oxidative stress. Plants cope with
372 stressed environments through inoculation of *Bacillus pseudomycoides*, which produces
373 phytohormones, ion equilibrium, and easy uptake of nutrients. All parameters decreased as salt
374 stress increased from 3-15 dSm⁻¹. Among all cultivars, Inqelab mung showed the highest resistance
375 to stress, while Ramzan mung demonstrated the lowest resistance.

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668 **Table S1: Physio-chemical analysis of Botanic Garden Soil**

Attributes	Garden soil
Texture	Loamy Sand
pH	8.23 ± 0.00
EC (dSm ⁻¹)	0.01 ± 0.00
N (%)	8.2 ± 0.05
P (%)	0.01 ± 0.00
K (ppm)	3.4 ± 0.1
Na (ppm)	6.95 ± 0.15
Ca ⁺⁺ (%)	0.11 ± 0.00

Mg ⁺⁺ (%)	0.07 ± 0.00
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Figure 1(on next page)

Figures

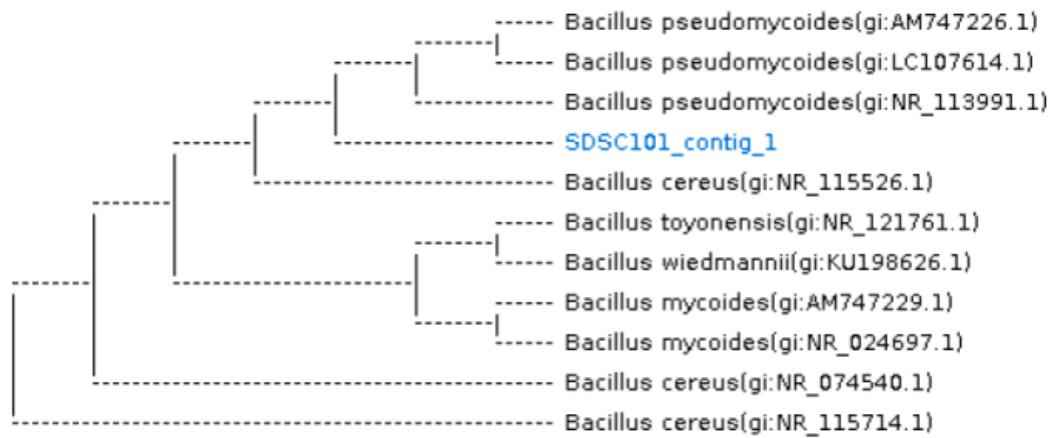
Figures

Figure: 1 Dendrogram showing homology of collected bacteria with *Bacillus pseudomycoides* (16S rRNA report). SDSC101_contig_1 showing the present study bacterial sample.

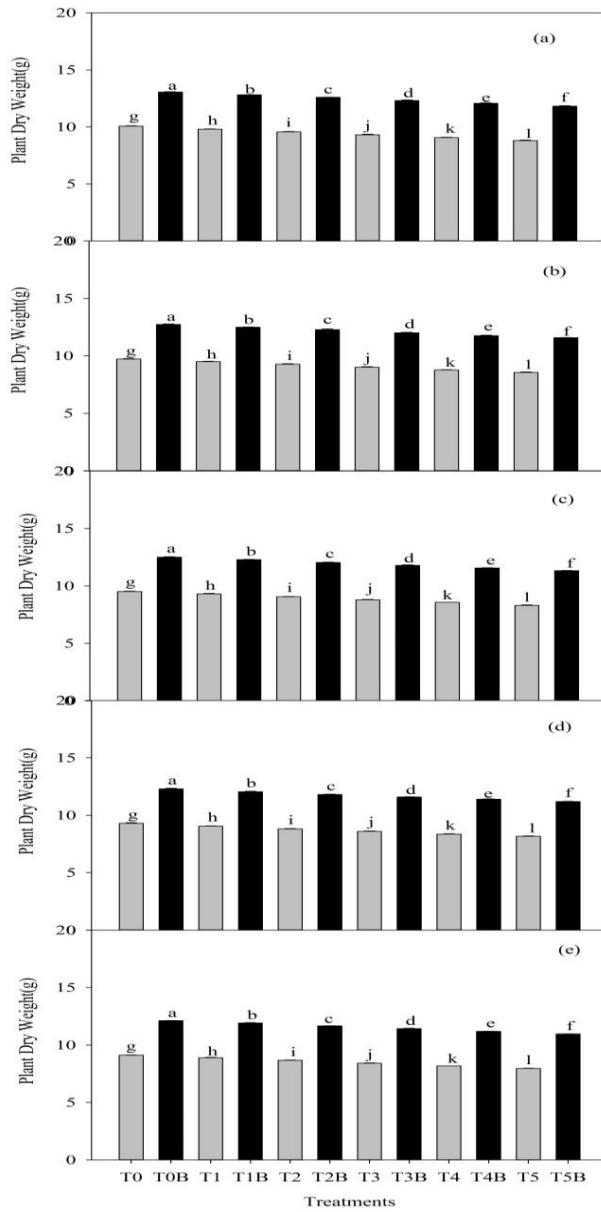


Figure 2: Effect of NaCl on different cultivars of Mung bean (*Vigna radiata* L.), Mean dry weight of plant parts (above and below ground) V1(a), V2(b), V3(c), V4(d) and V5(e), Bars with different letter (s) exhibit significant differences from each other. Grey bars indicate NaCl treatment and black bars indicate NaCl treatment along with *B. pseudomycoides* inoculum.

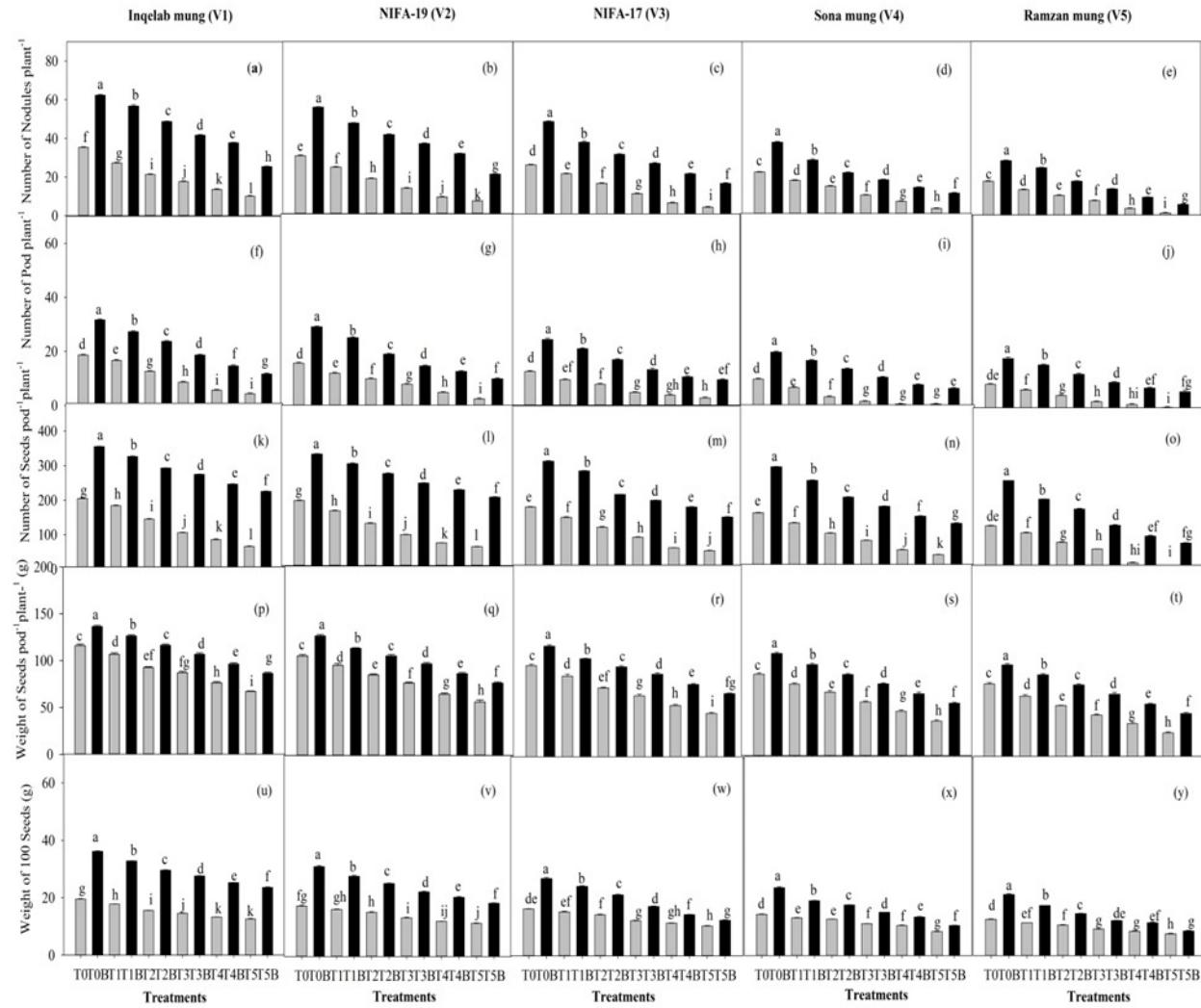


Figure 3: Effect of NaCl on different cultivars of Mung bean (*Vigna radiata* L.), Mean number of root nodules plant⁻¹ (a-e), number of pods plant⁻¹(f-j), number of seeds pod⁻¹plant⁻¹(k-o), weight of seeds pod⁻¹plant⁻¹(p-t) and weight of 100 seeds (u-y), Bars with different letter (s) exhibit significant differences from each other. Grey bars indicate NaCl treatment and black bars indicate NaCl treatment along with *B. pseudomycoïdes* inoculum.

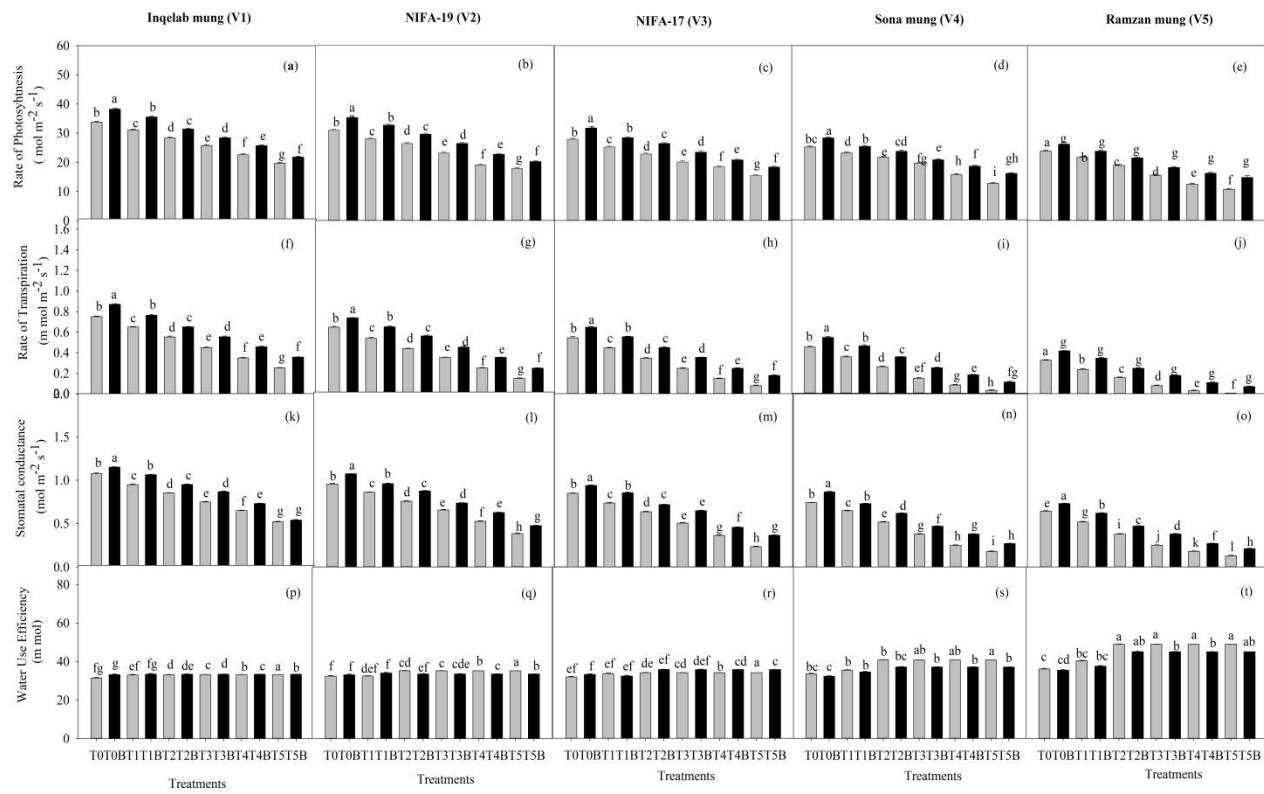


Figure 4: Effect of NaCl on different cultivars of Mung bean (*Vigna radiata* L.), Mean photosynthetic rate (a-e), transpiration rate (f-j), stomatal conductance (k-o) and water use efficiency (p-t), Bars with different letter (s) exhibit significant differences from each other. Grey bars indicate NaCl treatment and black bars indicate NaCl treatment along with *B. pseudomycoides* inoculum.

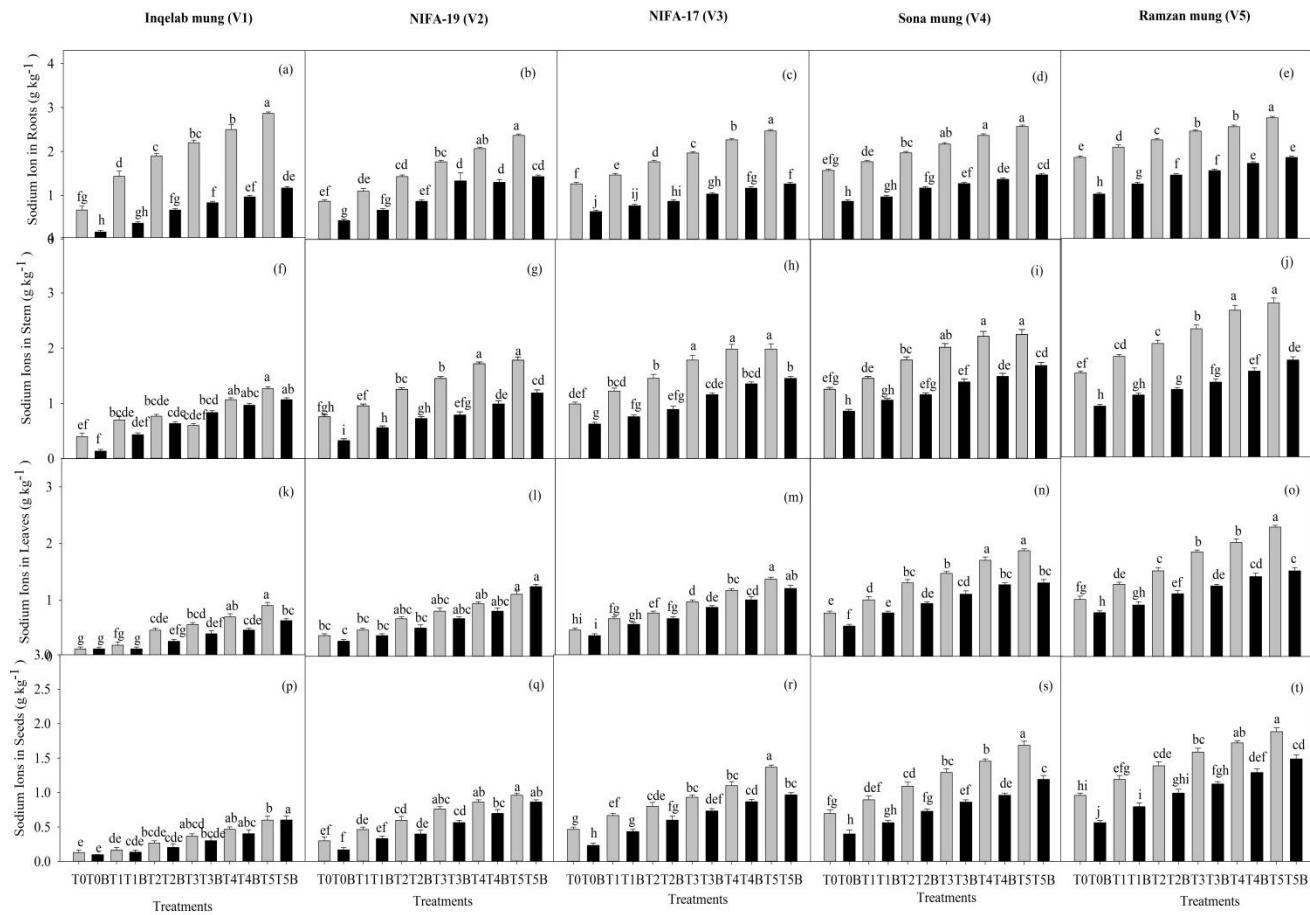


Figure 5: Effect of NaCl on different cultivars of Mung bean (*Vigna radiata* L.), Mean sodium ion uptake in root (a-e), sodium ion uptake in stem (f-j), sodium uptake in leaves (k-o) and sodium ion uptake in seeds (p-t), Bars with different letter (s) exhibit significant differences from each other. Grey bars indicate NaCl treatment and black bars indicate NaCl treatment along with *B. pseudomycoides* inoculum.

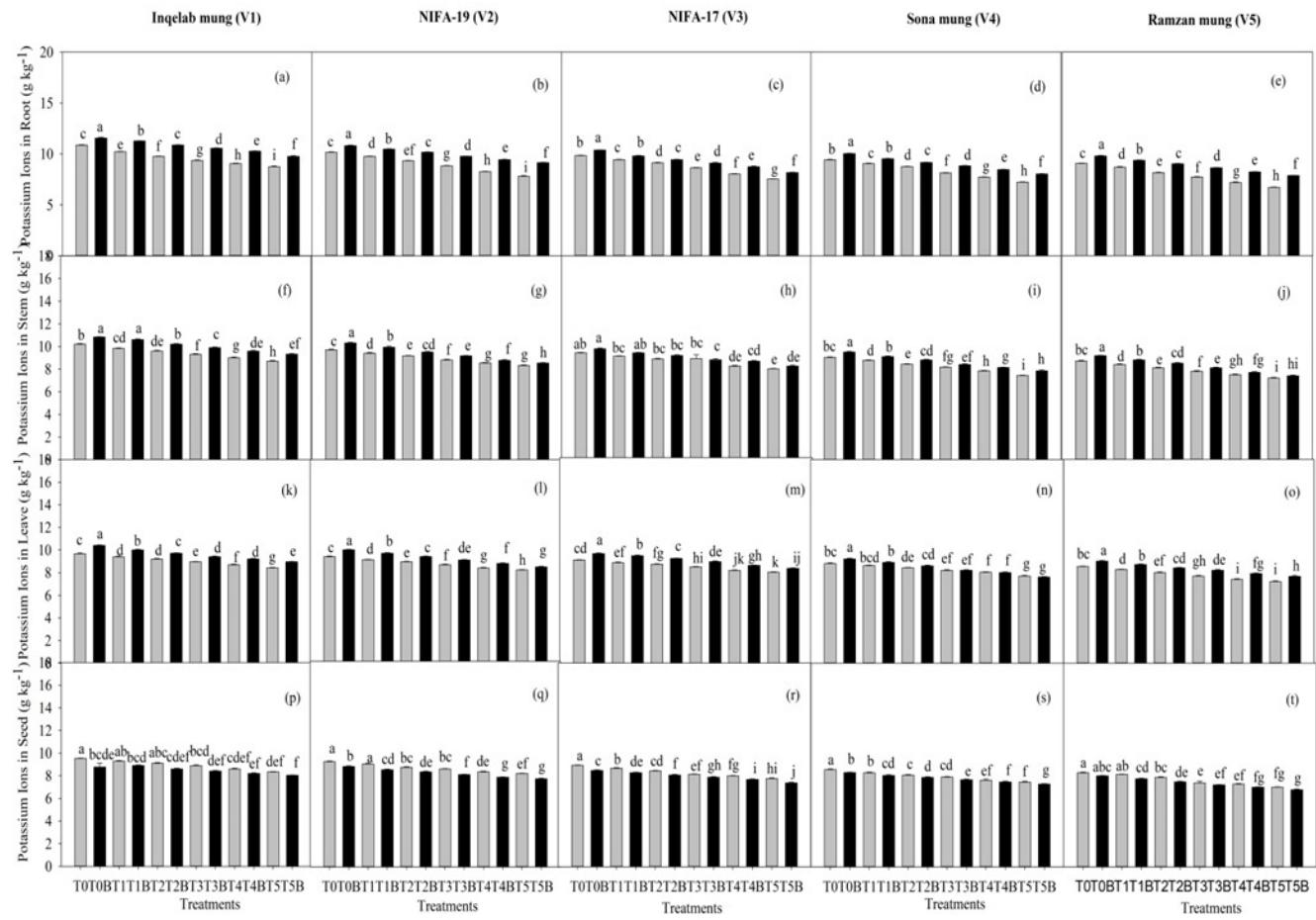


Figure 6: Effect of NaCl on different cultivars of Mung bean (*Vigna radiata* L.), Mean potassium ion uptake in root (a-e), potassium ion uptake in stem (f-j), potassium uptake in leaves (k-o) and potassium ion uptake in seeds (p-t), Bars with different letter (s) exhibit significant differences from each other. Grey bars indicate NaCl treatment and black bars indicate NaCl treatment along with *B. pseudomycoides* inoculum.

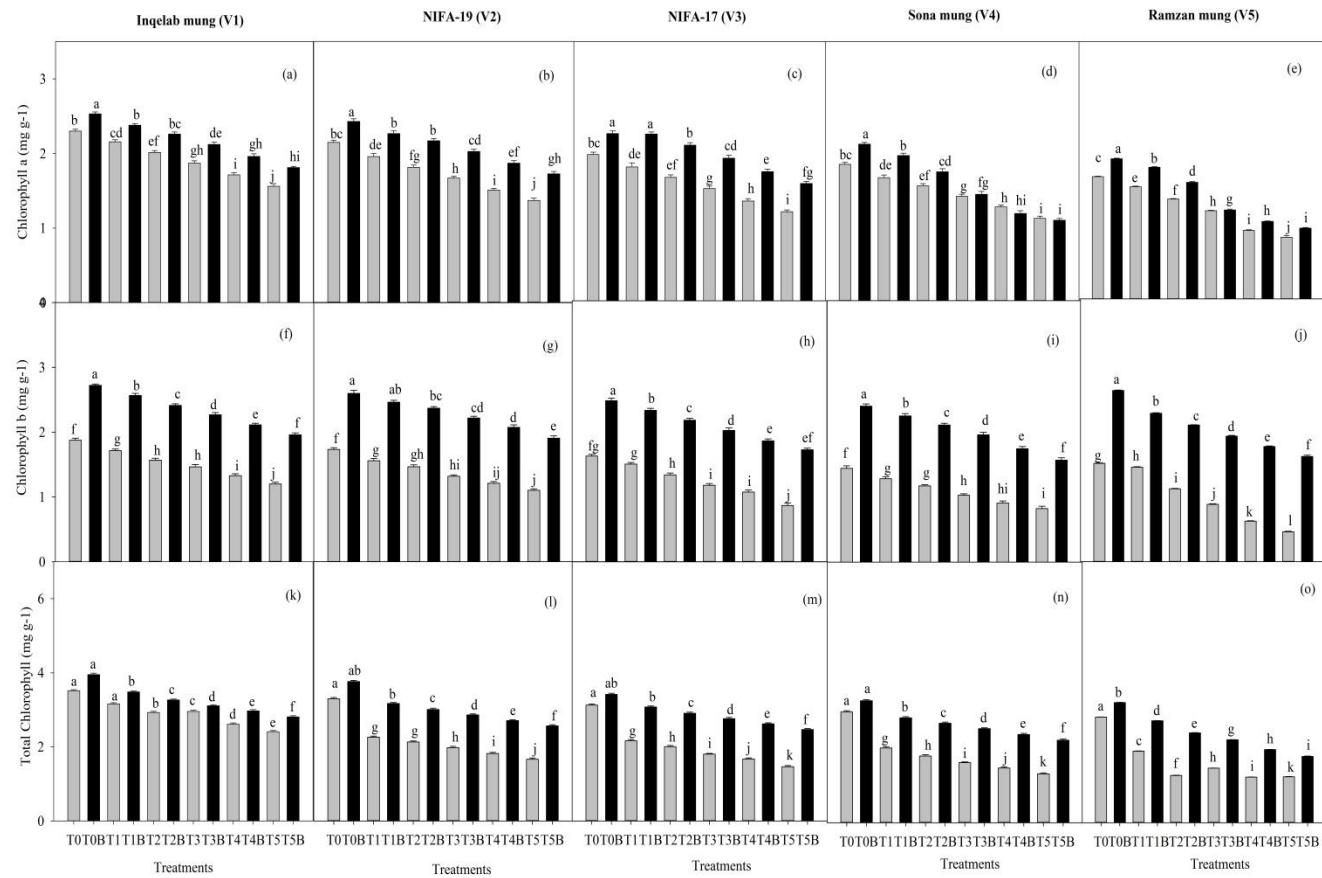


Figure-7: Effect of NaCl on different cultivars of Mung bean (*Vigna radiata* L.), Mean chlorophyll a (a-e), chlorophyll b (f-j) and total chlorophyll (k-o) contents, Bars with different letter (s) exhibit significant differences from each other. Grey bars indicate NaCl treatment and black bars indicate NaCl treatment along with *B. pseudomycooides* inoculum.

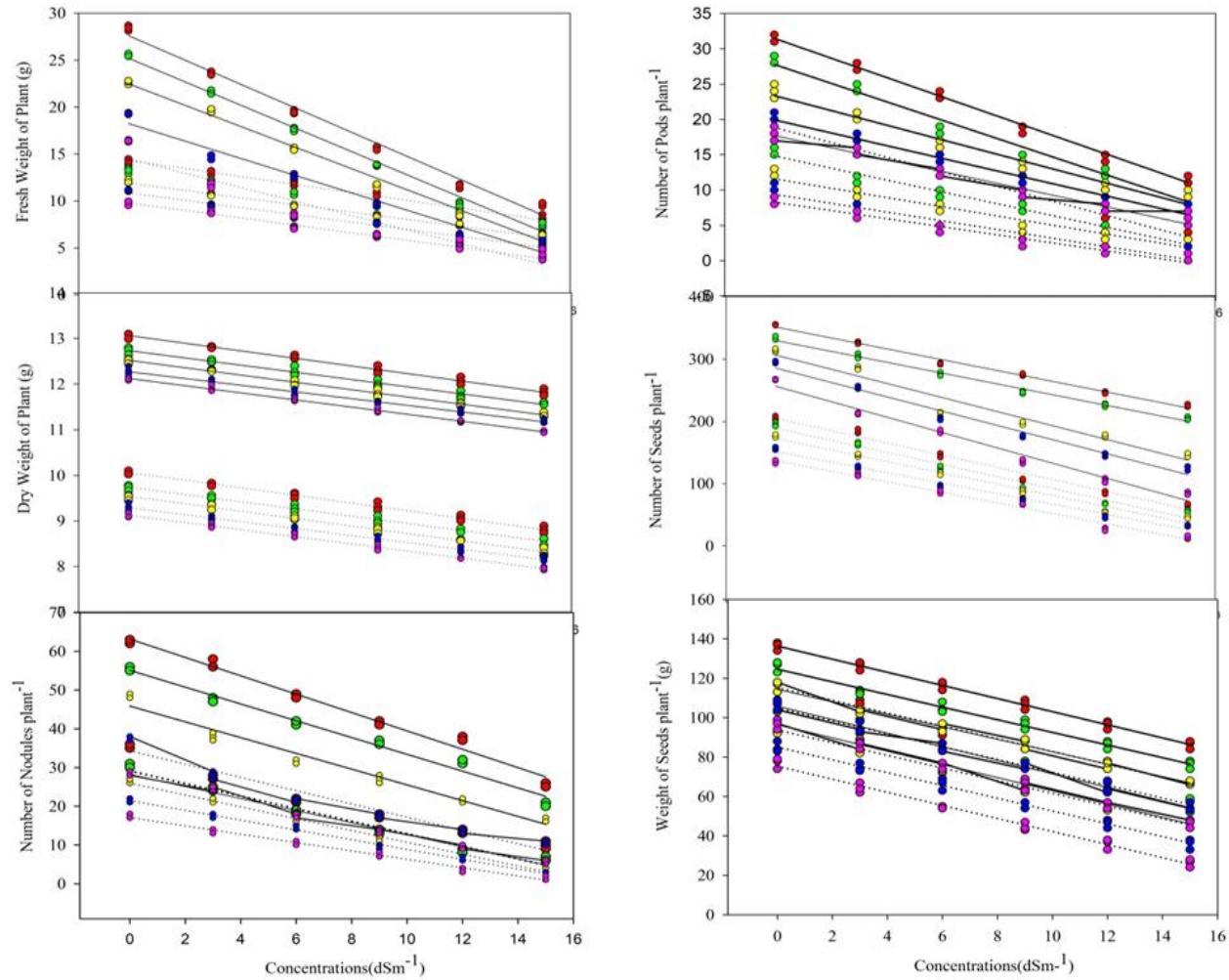


Figure 8: Effect of NaCl on different cultivars of Mung bean (*Vigna radiata* L.), Representation with dotted line = plant without inoculum of bacteria, solid line = plants with inoculum of bacteria, Red dots=V1, Green dots= V2, Yellow dots= V3, Blue dots= V4, Purple dots= V5.

Table 1(on next page)

Table

1 Tables

2 **Table 1:** Two-ANOVA model for interaction among measured parameters, salt stress and
3 *Bacillus pseudomycoides* that represent significance ($p \leq 0.05$) level

Parameters	DF	SS	MS	F	P value
Plant biomass	44	0.065	0.0015	1.22	0.0000
Weight of 100 seeds	44	596.10	13.548	123.03	0.0000
Number of seed plant ⁻¹	44	35096	798	113.00	0.0000
Number of pods plant ⁻¹	44	246.79	5.609	16.59	0.0000

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